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Synthesis of an α -kojibiosyl GTA Hexamer

Introduction

Antibiotic resistance is an emerging problem in society and prevention strategies, such as vaccination with (synthetic) cell-wall antigens, have become a promising approach. As described in **chapter 1** teichoic acids (TAs) are Gram-positive cell wall components that modulate the mammalian immune system. In order to understand the molecular mode of action of these molecules, well defined fragments could be a valuable tool.¹⁻⁵ Synthetic studies to and biological evaluation of *Staphylococcus aureus* LTA fragments by Schmidt and Hartung led to more insight in the minimal structural requirements for cytokine induction.⁶⁻¹²

Enterococcus faecalis is a commensal bacterium and generally of low virulence. However, with the emergence of vancomycin-resistant strains an increasing amount of *E. faecalis* infections is reported.¹³⁻¹⁵ The TA of *E. faecalis* is antigenic and built up from 1,3-poly(glycerolphosphate), randomly decorated at the C-2 positions with D-alanine, kojibiose (α -D-glucopyranosyl-($1\rightarrow 2$)- α -D-glucose), and 6,6'-di-alanyl- α -kojibiose, as depicted in Figure 1.^{16,17}



Figure 1. General structure of E. faecalis GTA and its substituents.

This chapter deals with the synthesis of a glycerol TA hexamer, containing an α kojibiosyl substitution.¹⁸ The syntheses of the stereochemically pure (kojibiosyl) glycerol synthons and their use in the construction of a hexamer (kojibiosyl)glycerol phosphate is described. Installment of the phosphodiester bridges was performed using phosphoramidite chemistry, which has emerged in the past two decades as the method of choice for (automated) oligonucleotide synthesis.¹⁹⁻²¹

Results and Discussion

Scheme 1. Target hexamer 1 and the required phosphoramidites 2-5.



As retrosynthetically depicted in Scheme 1, TA hexamer 1 can be assembled from four synthons: dibenzyl glycerol 2, the starting building block; glycerol phosphoramidite 3 and kojibiosyl glycerol phosphoramidite 4, used for chain elongation, and aminohexylphosphoramidite 5, the terminal building block. The primary amino group of the hexyl spacer presents a chemoselective ligation handle for attachment of, for instance, carrier proteins or fluorescent tags (see **chapter 7**). The 2-cyanoethyl (CE) was chosen as a phosphate protecting group because of its robustness during the synthesis and ease of removal at the end of the synthesis. The hydroxyl functions and the amino group of the aminohexyl spacer were masked with benzyl type protecting groups, which facilitated global deprotection in the final stage of the hexamer assembly by hydrogenolysis. As a temporary protecting group the dimethoxytrityl (DMT) group has been selected because it has proven its merits in contemporary DNA and RNA synthesis protocols and can be cleaved under mildly acidic conditions.^{21,22}

The synthesis of the four building blocks is depicted in Scheme 2. Dibenzylglycerol **2** was synthesized according to literature procedures from solketal **6** (Scheme 2A).^{7,10,23,24} Solketal **6** also served as starting compound in the synthesis of glycerolphosphoramidite **3**. Allylation of the free hydroxyl function, acidic hydrolysis of the isopropylidene and subsequent selective silylation of the primary alcohol yielded partially protected glycerol **7** (Scheme 2B).⁷ The remaining hydroxyl in compound **7** was benzylated to provide the fully protected glycerol in high yield, containing a minor amount (~3%) of 1-*0*-benzyl-2-*0*-tert-butyldiphenylsilyl-3-*0*-allyl-



Scheme 2. Synthesis of (phosphoramidite) building-blocks 2-5.

sn-glycerol, which resulted from silyl migration during the basic etherification, and which could not be removed by silica gel chromatography. To liberate the C3 hydroxyl, the allyl ether was isomerized using Ir(COD)(Ph₂MeP)₂PF₆.²⁵ Subsequent enol ether cleavage using iodine and aqueous NaHCO₃ afforded glycerol **8** in 71% over 2 steps. Dimethoxytritylation of the primary alcohol furnished glycerol **9**, which was desilylated to provide alcohol **10**. At this stage the product could be separated by silica gel chromatography from its regioisomeric impurity. Finally, installment of the

phosphoramidite moiety using 2-cyanoethyl-*N*,*N*-diisopropylchlorophosphoramidite **11** provided building block **3**.

With the first two building blocks in hand the synthesis of α -kojibiosylglycerol synthon 4, containing two 1,2-cis glucosidic linkages, was undertaken. Although tremendous progress has been made in the stereoselective construction of glycosidic bonds, the installment of two 1,2-cis-glucosyl linkages still presents a significant challenge. Recently, Boons and co-workers revealed an elegant approach to this long standing problem, by installing α -directing ethyl (S)-mandelate or (1S)phenyl(thiophenyl)ethyl ethers on the 2' hydroxyl of glucose donors.²⁶⁻²⁸ Unfortunately, the assembly of the kojibiose glycerol pseudotrisaccharide requires the glycosylation of the first glucose residue at the C-2 hydroxyl and the latter ether protecting groups cannot be removed selectively while keeping the other benzyl ethers intact. Alternative methods which have been used to construct 1,2-cis glucosidic linkages include Lemieux's *in situ* anomerization protocol^{29,30} and the employment of large substituents such as the DMT group at the glucosyl C-6 hydroxyl.³¹⁻³³ Given the relatively low reactivity of the secondary alcohol function which has to be glucosylated the former approach was deemed unsuitable. The latter approach was not pursued at this point because this would exclude the use of trityltype groups in the glycerol part of the pseudotrisaccharide, which was a prerequisite in the design of our protecting group strategy (vide supra).³⁴ An alternative approach to the formation of 1,2-cis glucosidic linkages has been described by Crich and coworkers, using benzylidene protected thioglucosyl donors.³⁵ Three 4,6-O-benzylidene functionalized glucosyl donors, having either a C2 para-methoxybenzyl (PMB, 13a), a C2 para-azidobenzyl (PAB, **13b**) or a C2 trimethylsilyl (TMS, **13c**) protecting group, were explored (Scheme 2C). The installment of the para-methoxylbenzyl group in construct 12^{36} led to benzylidene glucoside 13a in 88%. This thioglycoside was then condensed with glycerol acceptor 7 using the Ph_2SO/Tf_2O activator system³⁷ to provide the intermediate pseudodisaccharide in good yield and excellent selectivity $(\alpha/\beta = 10/1)$. Subsequent treatment of the fully protected adduct with DDQ gave alcohol **14** in moderate yield (56%), resulting in a 37% total yield of α -glucosyl glycerol 14 from alcohol 12. A similar series of reactions was employed to prepare pseudodisaccharide **14** with the use of the *para*-azido benzyl ether³⁸ instead of the *para*-methoxy benzyl group as a non-participating group at C-2. In this case (13b), the benzylation reaction proceeded uneventfully and the ensuing glycosylation reaction provided the *cis*-coupled product in good selectivity (62%, α/β = 7/1). Unfortunately, liberation of the C2 hydroxyl via a reduction-oxidation sequence provided target alcohol **14** in only 39%, to complete the four step sequence in a total yield of 22% of **14** from building block **12**. The last non-participating group that was tested was the trimethylsilyl ether (TMS). Donor **13c** was obtained quantitatively from **12**, and used in the subsequent glycosylation reaction without purification. The coupling of TMS protected thioglucose 13c and glycerol 7 was immediately followed by treatment of the resulting pseudodisaccharide (~6:1 mixture of α and β -anomers) with sodium

carbonate in methanol to yield **14** in 46% over the three steps, using a single chromatographical purification step, making this the most efficient route.

With the first 1,2-*cis* glycosidic linkage in place, the synthesis was continued with the condensation of dibenzyl benzylidene thioglucoside **15** and alcohol **14**. Gratifyingly, the pseudotrisaccharide (**16**) was obtained as a single diastereoisomer in 82% yield. This trisaccharide was transformed into the phosphoramidite building block **4** following the same sequence of reactions as described for the transformation of 3-*O*-allyl-2-*O*-benzyl-1-*O*-TBDPS-*sn*-glycerol into phosphoramidite **3**. Briefly, the allyl ether in **16** was removed by iridium catalyzed isomerisation and iodine mediated cleavage of the intermediate enol ether. The DMT group was installed, after which the TBDPS ether was removed by fluoride treatment and the primary alcohol was phosphitylated to complete the synthesis of the kojibiosyl glycerol **4**. To complete the set of target synthons, benzyloxycarbonyl protected aminohexanol **20** was treated with DiPEA and chlorophosporamidite **11** to give phosphoramidite **5** (Scheme 2D).

With all necessary building blocks in hand the assembly of the hexamer was undertaken (Scheme 3). In the first step dibenzylglycerol **2** and glycerol phosphoramidite **3** were coupled with the use of 4,5-dicyanoimidazole (DCI) in acetonitrile. After oxidation of the intermediate phosphite (I_2 in THF/pyridine 4/1), the crude intermediate was detritylated using dichloroacetic acid and triethylsilane in





dry DCM, giving dimer **21** in 80% yield. Repetition of this reaction sequence led to the consecutive formation of trimer **22** in 82% from **21**, tetramer **23** in 81% from **22** and pentamer **24** in 62% from **23**. Next, kojibiosylglycerol phosphoramidite **4** was coupled to the glycerolphosphate chain and, after iodine mediated oxidation, hexamer **25** was obtained in 86% yield. Apparently, the relatively bulky kojibiosyl substituent did not have an adverse effect on the formation of the mixed phosphotriester. To avoid unwanted acidolysis of the benzylidene functionalities in **25** the mild pyridinium *para*-toluenesulphonate (PPTS)/MeOH cocktail was employed for the cleavage of the DMT group.³⁹ Alcohol **26** was obtained in 70% yield when 1 mg/ml PPTS in MeOH/DCM (9/1) was employed. Subsequent coupling of spacer phosphoramidite **5** and oxidation gave the fully protected target compound **27** in 77% yield.

The deprotection sequence started with ammonolysis of the cyanoethyl groups in **27**, giving the hexameric phosphodiester quantitatively. Finally, all benzyl ethers, the two benzylidene acetals and the benzyloxycarbamate were removed using $H_2/Pd(0)$ in water/dioxane/AcOH. After size-exclusion chromatography, repeated lyophilization, and ion exchange the target *E. faecalis* TA-hexamer (**1**) was obtained as the per sodium salt in 76% yield. Hexamer **1** was evaluated in an opsonophagocytic killing inhibition assay (OPIA), to determine its binding to antibodies raised against native (*Enterococcal*) LTA, showing some inhibitory activity (compound **22**, figure 1, **chapter three**).

Conclusion

In summary, this chapter describes the synthesis of a kojibiose containing *Enterococcus faecalis* teichoic acid hexamer (1). The teichoic acid fragment was constructed by coupling of suitably protected glycerol and α -kojibiosyl substituted glycerol phosphoramidites. The key kojibiosyl synthon was obtained via a sequence of reactions involving two successive stereoselective α -glucosylations. The use of a benzylidene thioglucose donor, temporarily protected at the C2-hydroxyl with a TMS-group proved to be the most efficient in the synthesis of the kojibiosyl glycerol pseudotrisaccharide. The phosphoramidite chemistry used for the assembly of the teichoic acid hexamer is compatible with an automated solid-phase approach, as is illustrated in **chapter 3**.⁴⁰

Experimental section

General Procedures and Material: All chemicals (Acros, Fluka, Merck, Schleicher & Schuell, Sigma-Aldrich, Genscript) were used as received and reactions were carried out dry, under an argon atmosphere, at ambient temperature, unless stated otherwise. Tf₂O was distilled over P_2O_5 using flame-dried glass-ware. Column chromatography was performed on Screening Devices silica gel 60 (0.040-0.063 mm). TLC analysis was conducted on HPTLC aluminium sheets (Merck, silica gel 60, F245). Compounds were visualized by UV absorption (245 nm), by spraying with 20% H₂SO₄ in EtOH or with a solution of (NH₄)₆Mo₇O₂₄·4H₂O 25 g/l and (NH₄)₄Ce(SO₄)₄·2H₂O 10 g/l, in 10% aqueous H₂SO₄ followed by charring at +/- 140 °C. Some

unsaturated compounds were visualized by spraying with a solution of $KMnO_4$ (2%) and K_2CO_3 (1%) in H₂O. Optical rotation measurements ($[\alpha]_{D}^{20}$) were performed on a Propol automated polarimeter (Sodium D-line, $\lambda = 589$ nm) with a concentration of 10 mg/ml (c = 1), unless stated otherwise. Infrared spectra were recorded on a Shimadzu FT-IR 8300 and are reported in cm⁻¹. ³¹P, ¹H, and ¹³C NMR spectra were recorded with a Bruker AV 400 (161.7, 400 and 100 MHz respectively) or a Bruker DMX 600 (600 and 150 MHz respectively). NMR spectra were recorded in CDCl₃ with chemical shift (δ) relative to tetramethylsilane, unless stated otherwise. High resolution mass spectra were recorded by direct injection (2 μ l of a 2 μ M solution in water/acetonitrile; 50/50; v/v and 0.1 % formic acid) on a mass spectrometer (Thermo Finnigan LTO Orbitrap) equipped with an electrospray ion source in positive mode (source voltage 3.5 kV, sheath gas flow 10, capillary temperature 250 °C) with resolution R = 60000 at m/z 400 (mass range m/z = 150-2000) and dioctylphthalate (m/z = 391.28428) as a lock mass. The high resolution mass spectrometer was calibrated prior to measurements with a calibration mixture (Thermo Finnigan). TLC-MS (ESI) spectra of phosphoramidites 3, 4 and 5 were obtained by analysis of TLC-plates (Schleicher & Schuell, treated with Et₃N before applying the phosphoramidites) with a CAMAG TLC-MS interface coupled to a Perkin Elmer Sciex API 165 mass instrument.

1-0-(tert-Butyldiphenylsilyl)-2-0-benzyl-sn-glycerol (8)



Silyl ether **7** (22.8 g, 61.5 mmol) was dissolved in DMF (200 ml) and cooled to 0 °C. After the addition of benzyl bromide (11.1 ml, 92.5 mmol) and heptane flushed NaH (60 % dispersion in mineral oil, 3.70 g, 92.5 mmol), the mixture was allowed to stir for 1.5 h. H_2O (50 ml) was added, and after addition of Et₂O

(1.0 l), the mixture was washed with H₂O $(2 \times 200 ml)$ and brine (200 ml). The organic layer was dried over MgSO₄ and concentrated *in vacuo*. Repeated co-evaporation of the residue with H_2O (5 x 100 ml) and toluene (5 x 100 ml), followed by column chromatography (toluene/PE) resulted in the benzylated adduct (28.3 g, 61.4 mmol, 100%, containing ~3% 1-O-benzyl-2-O*tert*-butyldiphenylsilyl-3-*O*-allyl-*sn*-glycerol) as a colourless oil. (analytical data: $[\alpha]_D^{20}$ (CHCl₃): -10.4; IR: 737, 924, 1103, 1427, 2856 cm⁻¹; ¹H NMR (400 MHz): δ = 1.05 (s, 9H, *t*-Bu TBDPS), 3.55 - 3.71 (m, 3H, CH glycerol), 2 x CHH glycerol), 3.77 - 3.80 (m, 2H, 2 x CHH glycerol), 3.99 (m, 2H, CH₂ allyl), 4.64 (s, 2H, CH₂ Bn), 5.16 (m, 1H, CH*H* allyl), 5.26 (dd, 1H, 1,7 Hz, 17.3 Hz, CH*H* allyl), 5.86 - 5.93 (m, 1H, CH allyl), 7.18 - 7.43 (m, 11H, Harom), 7.66 - 7.71 (m, 4H, Harom); 13C NMR (100 MHz): $\delta = 19.2$ (C_q t-Bu), 26.8 (3 x CH₃ TBDPS), 63.5 (CH₂ glycerol), 70.2 (CH₂ glycerol), 72.2, 72.3 (CH₂ allyl, CH₂ Bn), 78.8 (CH glycerol), 116.8 (CH₂ allyl), 127.4, 127.7, 128.2, 129.6 (CH_{arom}), 133.5 (2 x Cq phenyl), 134.8 (CH allyl), 135.6 (CH_{arom}), 138.8 (Cq Bn).) A solution of the intermediate (9.21 g, 20.0 mmol) in freshly distilled THF (125 ml) was stirred under argon atmosphere for 30 min. After addition of Ir(COD)(Ph2MeP)2PF6 (423 mg, 0.50 mmol, 2.5 mol %), the solution turned red. The solution was then purged with H_2 (g) for ~30s, after which it was stirred for 2 h under argon atmosphere. The solution was diluted with THF (250 ml) and, after addition of sat. aq. NaHCO₃ (300 ml) and I₂ (7.61 g, 30.0 mmol), the mixture was allowed to stir overnight. The mixture was diluted with EtOAc (1.5 l) and washed with sat. aq. Na₂S₂O₃ (2 x 300 ml) and brine (200 ml). The organic layer was dried over MgSO₄ and concentrated in vacuo. Purification of the residue by column chromatography (EtOAc/PE), yielded 8 (6.26 g, 14.9 mmol, 75%, containing $\sim 3\%$ 1-0-benzyl-2-0-tert-butyldiphenylsilyl-sn-glycerol) as slightly yellow oil. [α]_{D²⁰} (CHCl₃): -22.0; IR: 739, 824, 1113, 1427, 2361, 2858 cm⁻¹; ¹H NMR (400 MHz): δ = 1.06 (s, 9H, *t*-Bu TBDPS), 2.05 (t, 1H, *J* = 6.2 Hz, OH), 3.60 - 3.82 (m, 5H, CH glycerol, 2 x CH₂ glycerol), 4.51 (d, 1H J = 11.7 Hz, CHH Bn), 4.63 (d, 1H, J = 11.7 Hz, CHH Bn), 7.25 - 7.45 (m, 11H, Harom), 7.66 - 7.68 (m, 4H, Harom); ¹³C NMR (100 MHz): δ = 19.2 (C_q *t*-Bu), 26.8 (3 x CH₃ TBDPS), 62.8 (CH₂ glycerol), 63.5 (CH₂ glycerol), 72.1 (CH₂ Bn), 79.6 (CH glycerol), 127.7, 128.4, 129.8 (CH_{arom}), 133.1, 133.2 (2 x C_q phenyl), 135.6 (CH_{arom}), 138.3 (C_q Bn); HRMS: $C_{26}H_{32}O_3Si + Na^+$ requires 443.2013, found 443.2013.

ODMT BNO Alcohol 8 (6.18 g, 14.7 mmol) in DCM (100 ml) was cooled to 0 °C. Added were

Et₃N (3.05 ml, 22.0 mmol) and 4,4'-dimethoxytrityl chloride (DMT-Cl, 5.47 g, OTBDPS 16.2 mmol), respectively, and the mixture was allowed to stir overnight at room temperature. The reaction was quenched by the addition of H_2O (3 ml), and stirred for 15 min. After washing with $H_{2}O(30 \text{ ml})$ and brine (30 ml), the organic layer was dried over Na_2SO_4 and concentrated *in vacuo*. The resulting oil was further purified by column chromatography (EtOAc/PE/Et₃N), giving 9 (10.3 g, 14.2 mmol, 97%, containing ~3% 1-0-benzyl-2-0-tertbutyldiphenylsilyl-3-O-(4,4'-dimethoxytrityl)-sn-glycerol) as colourless oil. [α] $_{D^{20}}$ (MeOH): +2.0; IR: 827, 1036, 1113, 1250, 1508, 1609, 2359, 2930 cm⁻¹; ¹H NMR (400 MHz): δ = 0.97 (s, 9H, t-Bu TBDPS), 3.23 - 3.31 (m, 2H, 2 x CHH glycerol), 3.71 - 3.78 (m, 9H, 2 x OMe, 2 x CHH glycerol, CH glycerol), 4.66 (2 x d, 2H, J = 12.2 Hz, CH₂ Bn), 6.74 - 6.83 (m, 4H, H_{arom}), 7.00 - 7.45 (m, 20H, H_{arom}), 7.60 - 7.62 (m, 4H, H_{arom}); ¹³C NMR (100 MHz): $\delta = 19.1$ (C_g t-Bu), 26.8 (3 x CH₃ TBDPS), 55.1 (2 x OMe) 63.7 (CH₂ glycerol), 63.9 (CH₂ glycerol), 72.3 (CH₂ Bn), 79.3 (CH glycerol), 86.0 (Cq DMT), 113.0, 113.6 (CHarom), 126.1 - 130.3 (CHarom), 133.4, 133.5 (2 x Cq phenyl), 136.3 (CH_{arom}), 136.4, 138.9, 145.1, 158.3 (Cq Bn, 5 x Cq DMT); HRMS: C47H50O5Si + Na⁺ requires 745.3320, found 745.3322.

2-O-Benzyl-3-O-(4,4'-dimethoxytrityl)-sn-glycerol (10)



To a solution of glycerol derivative **9** (25.5 g, 35.3 mmol) in THF (200 ml), was added TBAF (1.0 M solution in THF, 53 ml). After stirring for 3 h at room temperature, the solvent was removed *in vacuo*. The residue was purified by column chromatography (EtOAc/PE/Et₃N), affording primary alcohol **10** (16.4 g,

33.8 mmol, 96%) as colourless oil. $[\alpha]_{p^{20}}$ (MeOH): +10.6; IR: 829, 1034, 1177, 1250, 1508, 1607, 2359 cm⁻¹; ¹H NMR (400 MHz): $\delta = 2.02$ (bs, 1H, OH), 3.21 - 3.31 (m, 2H, 2 x CHH glycerol), 3.63 - 3.67 (m, 2H, CH glycerol, CHH glycerol), 3.70 - 3.74 (m, 7H, 2 x OMe, CHH glycerol), 4.53 (d, 1H, *J* = 11.6 Hz, CHH Bn), 4.66 (d, 1H, *J* = 12.0 Hz, CHH Bn), 6.80 (ad, 4H, *J* = 8.8 Hz, H_{arom}), 7.16 - 7.38 (m, 12H, H_{arom}), 7.44 (d, 2H, *J* = 7.6, H_{arom}); ¹³C NMR (100 MHz): $\delta = 55.0$ (2 x OMe), 63.0 (CH₂ glycerol), 63.3 (CH₂ glycerol), 72.0 (CH₂ Bn), 78.9 (CH glycerol), 86.4 (C_q DMTr), 113.2 (CH_{arom}), 126.7 - 129.9 (CH_{arom}), 135.9, 138.2, 144.7, 158.4 (C_q Bn, 5 x C_q DMTr); HRMS: C₃₁H₃₂Os + Na⁺ requires 507.3142, found 507.3135.



1-([*N*,*N*-Diisopropylamino]-2-cyanoethylphosphite)-2-*0*-benzyl-3-*0*-(4,4'-dimethoxytrityl)-sn-glycerol (3)

To a solution of **10** (3.19 g, 6.59 mmol) and DIPEA (1.72 ml, 9.88 mmol) in DCM (65 ml) were added activated MS3Å and (*N*,*N*-diisopropylamino)-2-cyanoethyl-chlorophosphite (**11**, 1.82 g, 7.90 mmol). After stirring for 2 h, the reaction was quenched with H₂O (5.0 ml) and washed with H₂O (50 ml) and brine (50 ml) respectively. The organic layer was dried over

Na₂SO₄ and concentrated *in vacuo*. Column chromatography (EtOAc/heptane/Et₃N) gave phosphoramidite **3** (3.74 g, 5.34 mmol, 81%, mixture of diastereoisomers) as colourless oil. ³¹P NMR (161.7 MHz, CD₃CN): δ = 149.2; ¹H NMR (400 MHz, CD₃CN): δ = 1.08 (t, 6H, *J* = 6.5 Hz, 2 x CH₃ isopropylamino), 1.14 (dd, 6H, *J* = 1.8 Hz, 6.8 Hz, 2 x CH₃ isopropylamino), 2.51 - 2.56 (m, 2H, CH₂ cyanoethyl), 3.17 - 3.24 (m, 2H, 2 x CH isopropylamino), 3.50 - 3.58 (m, 2H, 2 x CHH glycerol), 3.64 - 3.81 (m, 11H, CH glycerol, 2 x CHH glycerol, CH₂ cyanoethyl, 2 x OMe), 4.61 -

4.68 (m, 2H, CH₂ Bn), 6.83 (d, 4H, J = 8.9 Hz, H_{arom}), 7.19 - 7.39 (m, 12H, H_{arom}), 7.45 - 7.47 (m, 2H, H_{arom}); TLC-MS: C₄₀H₄₉N₂O₆P + H⁺ requires 685.34, found 685.5.



Phenyl 2-0-(4-methoxybenzyl)-3-0-benzyl-4,6-0-benzylidene-1thio-β-D-glucopyranoside (13a)

To a cooled (0 °C) solution of thioglucose **12** (18.3 g, 40.5 mmol) in THF (250 ml) were added, respectively, *p*-methoxybenzyl chloride (6.03 ml,

44.5 mmol) and NaH (60 % dispersion in mineral oil, 1.78 g, 44.5 mmol). After stirring for 5 h, the mixture was quenched with MeOH (5.0 ml), diluted with EtOAc (1.0 l) and washed with, sat. aq. NaHCO₃ (400 ml) and brine (400 ml). The organic layer was dried with MgSO₄ and concentrated *in vacuo*, after which crystallization (EtOAc/PE) yielded fully protected thioglucose derivative **13a** (20.4 g, 35.8 mmol, 88%) as white solid. M.p.: 146-147 °C; $[\alpha]_{D^{20}}$ (CHCl₃): -11.2; IR: 748, 818, 1030, 1088, 1250, 1516 cm⁻¹; ¹H NMR (400 MHz): δ = 3.45 (m, 1H, H-5), 3.50 (dd, 1H, *J* = 8.4 Hz, 9.7 Hz, H-2), 3.69 (at, 1H, *J* = 9.4 Hz, H4), 3.76 - 3.84 (m, 5H, OMe, H-3, H-6), 4.37 (dd, 1H, *J* = 5.0 Hz, 10.5 Hz, H-6), 4.73 - 4.81 (m, 4H, 3 x CH*H* Bn, H-1), 4.94 (d, 1H, *J* = 11.2 Hz, CH*H* Bn), 5.57 (s, 1H, CH benzylidene), 6.85 - 6.88 (m, 2H, H_{arom}), 7.26 - 7.39 (m, 13H, H_{arom}), 7.46 - 7.54 (m, 4H, H_{arom}); ¹³C NMR (100 MHz): δ = 55.3 (OMe), 68.7 (C-6), 70.2 (C-5), 75.3, 75.5 (CH₂ Bn, CH₂ PMB), 80.1 (C-2), 81.4 (C-4), 83.0 (C-3), 88.3 (C-1), 101.1 (CH benzylidene), 113.8 (CH_{arom}), 126.0 - 129.9 (CH_{arom}), 130.2 (Cq *S*-phenyl), 132.2 (CH_{arom}), 133.2, 137.2, 138.3 (Cq Bn, Cq PMB, Cq benzylidene), 159.4 (Cq PMB); HRMS: C₃₄H₃₄O₆S + H⁺ requires 571.2149, found 571.2147.



Phenyl 2-0-(4-azidobenzyl)-3-0-benzyl-4,6-0-benzylidene-1-thio-β-D-glucopyranoside (13b)

To a cooled (0 °C) solution of thioglucose **12** (895 mg, 1.99 mmol) in DMF (20 ml) were added *p*-azidobenzyl bromide (634 mg, 2.99 mmol) and NaH (60 % dispersion in mineral oil, 127 mg, 3.18 mmol) respectively. After stirring for 1.5 h, MeOH (2.0 ml) was added before the mixture was diluted with Et₂O (80 ml) and washed with H₂O (20 ml) and brine (20 ml). The organic layer was dried with MgSO₄ and concentrated *in vacuo*.

The residue was taken up in EtOAc (5 ml) and crystallized upon addition of PE, yielding **13b** (1.07 g, 1.84 mmol, 93%) as ochreous solid. M.p.: 150-155 °C (decomp.); $[\alpha]_D^{20}$ (CHCl₃): +3.0; IR: 991, 1088, 1508, 2112 cm⁻¹; ¹H NMR (400 MHz): δ = 3.44 - 3.51 (m, 2H, H-2, H-5), 3.70 (at, 1H, *J* = 9.4 Hz, H-4), 3.78 - 3.84 (m, 2H, H-3, H-6), 4.39 (dd, 1H, *J* = 5.2 Hz, 10.4 Hz, H-6), 4.73 - 4.82 (m, 4H, 3 x CH*H* Bn, H-1), 4.95 (d, 1H, *J* = 11.2 Hz, CH*H* Bn), 5.59 (s, 1H, CH benzylidene), 6.96 - 7.00 (m, 2H, H_{arom}), 7.26 - 7.40 (m, 13H, H_{arom}), 7.45 - 7.53 (m, 4H, H_{arom}); ¹³C NMR (100 MHz): δ = 68.7 (C-6), 70.2 (C-5), 75.1, 75.3 (CH₂ Bn, CH₂ *p*-N₃Bn), 80.3 (C-2), 81.5 (C-4), 82.9 (C-3), 88.2 (C-1), 101.1 (CH benzylidene), 118.9 (CH_{arom}), 126.0 - 132.2 (CH_{arom}), 133.0, 134.8, 137.2, 138.2, 139.5 (C_q *S*-phenyl, C_q Bn, 2 x C_q *p*-N₃Bn, C_q benzylidene); HRMS: C₃₄H₃₁N₃O₅S + H⁺ requires 582.2057, found 582.2056.



1-0-(tert-Butyldiphenylsilyl)-2-0-(3-0-benzyl-4,6-0-benzylidene-α-D-glucopyranosyl)-3-0-allyl-sn-glycerol (14) From **13a**:

A solution of donor **13a** (2.85 g, 4.99 mmol), TTBP (2.86 g, 11.5 mmol) and Ph_2SO (1.11 g, 5.50 mmol) in freshly distilled DCM (100 ml), together with activated MS3Å, was stirred under argon at RT

for 30 min. Subsequently, the mixture was cooled to -75 °C and stirred for another 15 min. After the addition of Tf₂O (0.93 ml, 5.5 mmol), the mixture was stirred for 45 min at -75 °C and,

subsequently, glycerol acceptor 7 (2.22 g, 5.99 mmol) in DCM (10 ml) was added. After stirring for 60 minutes at -75 °C, the mixture was allowed to, gradually, warm to room temperature (~3 h). The reaction was guenched upon addition of Et_3N (2.0 ml, 14 mmol) and stirred for 30 min. After washing the mixture with sat, aq. NaHCO₃ (30 ml) and brine (30 ml), the organic layer was dried over MgSO₄ and concentrated *in vacuo*. The resulting oil was dissolved in pyridine (30 ml) and, after the addition of Ac₂O (7 ml), stirred for 2 h. After removal of the solvents in vacuo, the residue was dissolved in EtOAc (200 ml) and washed with sat. aq. NaHCO₃ (2×60 ml) and brine (60 ml). The organic layer was dried over MgSO₄ and the solvent removed under reduced pressure. Purification of the resulting oil by column chromatography (EtOAc/PE) gave the fully protected product (3.16 g, 3.80 mmol, 76%, α/β mixture, ~10/1), as colourless oil. IR: 737, 1088, 1369, 1454, 1751, 2855, 2924 cm⁻¹; ¹H NMR α -product (400 MHz): δ = 1.05 (s, 9H, *t*-Bu), 3.51 - 3.65 (m, 4H, CH glycerol, CHH glycerol, H-2, H-6), 3.69 - 3.80 (m, 6H, OMe, 3 x CHH glycerol), 3.89 (m, 1H, H-5), 3.96 - 4.03 (m, 5H, CH₂ allyl, H-3, H-4, H-6), 4.66 (d, 1H, *J* = 11.7 Hz, CH*H* Bn), 4.71 (d, 1H, *J* = 11.7 Hz, CH*H* Bn), 4.79 (d, 1H, *J* = 11.2 Hz, CH*H* Bn), 4.85 (d, 1H, *J* = 11.3 Hz, CHH Bn), 5.17 (dd, 1H, J = 1.6 Hz, 10.4 Hz, CHH allyl), 5.20 (d, 1H, J = 3.8 Hz, H-1), 5.26 (dd, 1H, J = 1.7 Hz, 17.2 Hz, CHH allyl), 5.49 (s, 1H, CH benzylidene), 5.89 (ddd, 1H, J = 5.5 Hz, 10.7 Hz, 22.7 Hz, CH allyl), 6.83 (d, 2H, J = 8.6 Hz, H_{arom}), 7.24 - 7.47 (m, 18H, H_{arom}), 7.65 - 7.68 (m, 4H, H_{arom}); ¹³C NMR α-product (100 MHz): δ = 19.2 (C_q *t*-Bu), 26.8 (3 x CH₃ TBDPS), 55.2 (OMe), 62.4 (C-5), 63.8 (CH₂ glycerol), 68.9 (C-6), 70.7 (CH₂ glycerol), 72.3 (CH₂ allyl), 72.3, 75.2 (CH₂ Bn, CH₂ PMB), 76.6, 78.2 (C-3, C-4), 78.6 (CH glycerol), 82.1 (C-2), 97.2 (C-1), 101.2 (CH benzylidene), 113.7 (CHarom), 116.9 (CH2 allyl), 126.1 - 129.7 (CHarom), 130.4, 133.2, 133.2, 134.7, 135.5, 137.5, 138.9 (CH_{arom}, CH allyl, 2 x Cq phenyl, Cq PMB, Cq Bn, Cq benzylidene), 159.2 (Cq PMB); HRMS: C₅₀H₅₈O₉Si + Na⁺ requires 853.3742, found 853.3745.) A combination of batches of the fully protected pesudodisaccharide (8.81 g, 10.6 mmol) was dissolved in DCM (100 ml) and cooled to 0 °C. After adding H₂O (5.0 ml) and 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDO, 2.88 g, 12.7 mmol), respectively, the mixture was stirred vigorously for 4 h, before it was quenched by the addition of sat. aq. NaHCO₃ (25 ml). The mixture was diluted with EtOAc (500 ml) and washed with sat. aq. NaHCO₃ (3 x 200 ml) and brine (3 x 150 ml), respectively, in order to remove the bulk of DDOH₂. The organic layer was dried over MgSO₄ and concentrated in vacuo. The crude product was purified by column chromatography (EtOAc/PE). Alcohol 14 (4.23 g, 5.95 mmol, 56%) was obtained as pale yellow oil.

From **13b**:

A solution of donor 13b (145 mg, 0.250 mmol), TTBP (155 mg, 0.625 mmol) and Ph₂SO (60.7 mg, 0.300 mmol) in freshly distilled DCM (8.0 ml), together with activated MS 3Å, was stirred under argon at RT for 30 min. Subsequently, the mixture was cooled to -78 °C and stirred for another 15 min. After the addition of Tf_2O (50.5 µl, 0.300 mmol), the mixture was stirred for 45 min at -50 °C before it was cooled down to -78 °C. After the addition of a solution of glycerol acceptor 7 (111 mg, 0.300 mmol) in DCM (2.5 ml), the mixture was stirred at -75 °C for 1h, before it was allowed to gradually $(\sim 3 h)$ warm to room temperature. The reaction was quenched by the addition of Et₃N (0.2 ml, 1.4 mmol) and stirred for 30 min. The mixture was diluted with DCM (30 ml) and after washing with sat. aq. NaHCO₃ (15 ml) and brine (15 ml), the organic layer was dried over MgSO4 and concentrated in vacuo. Purification of the resulting oil by size-exclusion chromatography (sephadex LH-20, MeOH/DCM 1/1), followed by column chromatography (EtOAc/PE) gave the fully protected intermediate (131 mg, 0.156 mmol, 62%, α/β mixture, ~7/1), as pale yellow oil. (Analytical data: ¹H NMR α -product (400 MHz): δ = 1.06 (s, 9H, t-Bu), 3.49 - 4.04 (m, 13H, CH glycerol, 2 x CH₂ glycerol, H-2, H-3, H-4, H-5, 2 x H-6, CH₂ allyl), 4.68 (d, 1H, J = 12.0 Hz, CHH Bn), 4.74 (d, 1H, J = 12.0 Hz, CHH Bn), 4.80 (d, 1H, J = 12.0 Hz, CH*H* Bn), 4.88 (d, 1H, *J* = 11.6 Hz, CH*H* Bn), 5.17 (dd, 1H, *J* = 1.2 Hz, 10.4 Hz, CH*H* allyl), 5.23 -5.28 (m, 2H, H-1, CH*H* allyl), 5.50 (s, 1H, CH benzylidene), 5.88 (ddd, 1H, *J* = 5.6 Hz, 10.6 Hz, 22.8

Hz, CH allyl), 6.93 (d, 2H, J = 8.4 Hz, Harom), 7.22 - 7.46 (m, 18H, Harom), 7.64 - 7.68 (m, 4H, Harom); ¹³C NMR α-product (100 MHz): δ = 19.2 (C_q t-Bu, 26.8 (3 x CH₃ TBDPS), 62.4 (C-5), 63.9 (CH₂ glycerol), 68.8 (C-6), 70.9 (CH₂ glycerol), 71.8, 72.3, 75.2 (CH₂ allyl, CH₂ p-N₃Bn, CH₂ Bn), 76.7, 78.2 (C-3, C-4), 79.0 (CH glycerol), 82.2 (C-2), 97.2 (C-1), 101.2 (CH benzylidene), 116.9 (CH₂ allyl), 118.8 (CHarom) 126.1 - 129.7 (CHarom), 133.1, 133.2 (2 x Cq phenyl), 134.6 (CH allyl), 135.1 (Cq *p*-N₃Bn), 135.5 (CH_{arom}), 137.5, 138.8, 139.3 (Cq Bn, Cq benzylidene Cq *p*-N₃Bn)). A part of the intermediate (58 mg, 0.069 mmol) was dissolved in a mixture of dioxane/water (9/1, 10 ml) and treated with PMe₃ (1M solution in toluene, 0.34 ml). After stirring for 1h, the mixture was concentrated *in vacuo* and coevaporated three times with dioxane (portions of 5 ml) before the mixture was redissolved in moist DCM (10 ml) and DDQ (24 mg, 0.10 mmol) was added. After stirring vigorously for 4 h, the reaction was quenched by the addition of sat. aq. $NaHCO_3$ (2.5) ml). The mixture was then diluted with EtOAc (40 ml) and washed with, respectively, sat. aq. NaHCO₃ (3 x 15 ml) and brine (3 x 15 ml) in order to remove the bulk of DDQH₂. The organic layer was dried over MgSO₄ and concentrated *in vacuo*. The crude product was purified by column chromatography (EtOAc/PE), giving alcohol **14** (19.0 mg, 26.7 µmol, 39%) as pale yellow oil.

From 12 (via 13c):

To a solution of thioglucose **12** (225 mg, 0.500 mmol) in DCM (5.0 ml) were added Et₃N (0.69 ml, 5.0 mmol), TMS-Cl (0.32 ml, 2.50 mmol) and a drop of pyridine. After stirring for 2 h, the reaction was diluted with EtOAc (40 ml) and washed with H_2O (15 ml) and brine (15 ml) respectively. The organic layer was dried with MgSO₄ and concentrated in vacuo. Crude 13c, together with Ph₂SO (121 mg, 0.600 mmol) and TTBP (323 mg, 1.30 mmol) were coevaporated with toluene (3 x 5 ml) and together with activated MS3Å dissolved in dry DCM (10 ml). After stirring for 15 min, the mixture was cooled to -78 °C and stirred for 5 min. After the addition of Tf₂O (101 µl, 0.600 mmol), the mixture was allowed to slowly warm up to -60 °C (\sim 30 min), after which the mixture was cooled down to -78 °C and glycerol acceptor 7 (278 mg, 0.750 mmol, dissolved in 2.0 ml DCM) was added. After stirring for 1 h at -78 °C, the mixture was allowed to warm up to RT overnight. After the addition of a few drops of H₂O the mixture was diluted with MeOH (20 ml). K₂CO₃ (10 g) was subsequently added and, after stirring for 5 hrs, the mixture was diluted with EtOAc (100 ml). The organic layer was washed with H_2O (40 ml) and brine (40 ml) and dried with MgSO₄. After removal of the solvents in vacuo, the residue was purified by column chromatography (EtOAc/PE), affording 14 (165 mg, 0.232 mmol, 46%, only α) as pale yellow oil. [α]_D²⁰ (CHCl₃): +33.6; IR: 741, 1040, 1072, 2343, 2361 cm⁻¹; ¹H NMR (400 MHz): δ = 1.06 (s, 9H, *t*-Bu), 2.92 (bs, 1H, OH), 3.54 - 3.79 (m, 8H, 2 x CH₂ glycerol, H-2, H-3, H-4, H-6), 3.85 - 3.95 (m, 2H, H-5, CH glycerol), 4.00 (ad, 2H, J = 5.6 Hz, CH₂ allyl), 4.05 (dd, 1H, J = 4.9 Hz, 10.2 Hz, H-6), 4.85 (d, 1H, J = 11.8 Hz, CHH Bn), 4.91 (d, 1H, J = 11.8 Hz, CHH Bn), 5.06 (d, 1H, *J* = 3.8 Hz, H-1), 5.20 (dd, 1H, *J* = 1.1 Hz, 10.4 Hz, CH*H* allyl), 5.28 (dd, 1H, *J* = 1.5 Hz, 17.2 Hz, CH*H* allyl), 5.51 (s, 1H, CH benzylidene), 5.88 (ddd, 1H, J = 5.5 Hz, 10.8 Hz, 22.8 Hz, CH allyl), 7.23 -7.47 (m, 16H, H_{arom}), 7.64 - 7.68 (m, 4H, H_{arom}); 13 C NMR (100 MHz): δ = 19.2 (C_q t-Bu), 26.8 (3 x CH3 TBDPS), 63.0 (C-5), 63.8 (CH2 glycerol), 68.8 (C-6), 69.8 (CH2 glycerol), 72.3 (CH2 allyl), 73.1 (C-2), 74.6 (CH₂ Bn), 79.0 (CH glycerol), 79.1, 81.5 (C-3, C-4), 99.8 (C-1), 101.2 (CH benzylidene), 117.5 (CH₂ allyl), 126.1 - 129.8 (CH_{arom}), 133.0, 133.1 (2 x C_q phenyl), 134.2 (CH allyl), 135.5 (CHarom), 137.5, 138.8 (Cq Bn, Cq benzylidene); HRMS: C42H50O8Si + Na⁺ requires 734.3201, found 734.3203.



1-0-(*tert*-Butyldiphenylsilyl)-2-0-(2-[2,3-di-0-benzyl-4,6-0-benzylidene- α -D-glucopyranosyl]-3-0-benzyl-4,6-0-benzylidene- α -D-glucopyranosyl)-3-0-allyl-sn-glycerol (16)

A solution of glucose donor **15** (130 mg, 0.240 mmol), TTBP (149 mg, 0.600 mmol) and Ph_2SO (57 mg, 0.28 mmol) in freshly distilled DCM (5.0 ml), together with activated MS3Å, was stirred under

argon at RT for 30 min. The mixture was cooled to -78 °C and stirred for another 15 min before Tf₂O (47 μ l, 0.28 mmol) was added and the mixture stirred for another 45 min at -70 °C. Subsequently, alcohol 14 (142 mg, 0.200 mmol, dissolved in 2.5 ml DCM) was added and after stirring for 60 min at -75 °C, the mixture was allowed to, gradually, warm to room temperature $(\sim 3 \text{ h})$. The reaction was quenched by the addition of Et₃N (0.14 ml, 1.0 mmol) and stirred for 30 min. The mixture was diluted with EtOAc (30 ml) and washed with sat. aq. NaHCO₃ (10 ml) and brine (10 ml). The organic layer was dried over MgSO4 and concentrated in vacuo. The resulting oil was purified by size-exclusion chromatography (sephadex LH-20, MeOH/DCM 1/1) and, subsequently, column chromatography (EtOAc/pentane). Kojibiose derivative 16 (187 mg, 0.164 mmol, 82%, pure α) was obtained as white foam. [α]_D²⁰ (CHCl₃): +34.4; IR: 746, 997, 1028, 1074, 1369, 1454, 2860 cm⁻¹; ¹H NMR (400 MHz): δ = 1.05 (s, 9H, t-Bu), 3.54 (dd, 1H, J = 5.9 Hz, 10.5 Hz, CHH glycerol), 3.57 - 3.69 (m, 7H, 2 x CHH glycerol, H-2, 2 x H-4, 2 x H-6), 3.74 (dd, 1H, J = 5.3 Hz, 10.4 Hz, CHH glycerol), 3.80 (dd, 1H, J = 3.7 Hz, 9.4 Hz, H-2), 3.88 (m, 2H, CH₂ allyl), 3.91 - 4.00 (m, 2H, CH glycerol, H-5), 4.04 - 4.21 (m, 5H, 2 x H-3, H-5, 2 x H-6), 4.70 (d, 1H, *I* = 11.8 Hz, CH*H* Bn), 4.74 - 4.83 (m, 4H, CH₂ Bn), 4.90 (d, 1H, *I* = 11.8 Hz, CH*H* Bn), 5.12 (dd, 1H, *J* = 1.1 Hz, 10.4 Hz, CH*H* allyl), 5.19 (dd, 1H, *J* = 1.6 Hz, 17.3 Hz, CH*H* allyl), 5.27 (d, 1H, *J* = 3.5 Hz, H-1), 5.40 (d, 1H, J = 3.6 Hz, H-1), 5.52 (s, 1H, CH benzylidene), 5.54 (s, 1H, CH benzylidene), 5.82 (ddd, 1H, J = 5.6 Hz, 10.7 Hz, 22.7 Hz, CH allyl), 7.06 - 7.11 (m, 2H, H_{arom}), 7.19 - 7.39 (m, 25H, H_{arom}), 7.44 - 7.48 (m, 4H, H_{arom}), 7.64 - 7.68 (m, 4H, H_{arom}); ¹³C NMR (100 MHz): δ = 19.1 (Cq t-Bu), 26.8 (3 x CH3 TBDPS), 62.5 (C-5), 62.8 (C-5), 63.5 (CH2 glycerol), 68.8 (C-6), 68.9 (C-6), 69.7 (CH₂ glycerol), 72.1 (CH₂ allyl), 72.4 (CH₂ Bn), 75.0 (CH₂ Bn), 75.3 (C-2), 75.3 (CH₂ Bn), 75.9 (CH glycerol), 76.7 (C-3), 78.1 (C-3), 79.0 (C-2), 82.2 (2 x C-4), 95.4 (C-1), 95.5 (C-1), 101.2 (2 x CH benzylidene), 117.0 (CH₂ allyl), 126.1 - 129.7 (CH_{arom}), 133.2 (2 x Cq phenyl), 134.5 (CH allyl), 135.5 (CHarom), 137.5, 137.6, 138.3, 138.6 (3 x Cq Bn, 2 x Cq benzylidene); HRMS: C₆₉H₇₆O₁₃Si + Na⁺ requires 1163.4947, found 1163.4962.



1-*O*-(*tert*-Butyldiphenylsilyl)-2-*O*-(2-[2,3-di-*O*-benzyl-4,6-*O*-benzylidene- α -D-glucopyranosyl]-3-*O*-benzyl-4,6-*O*-benzylidene- α -D-glucopyranosyl)-sn-glycerol (17)

A solution of **16** (3.18 g, 2.79 mmol) and freshly activated MS3Å in freshly distilled THF (35 ml) was stirred under argon for 30 min. After the addition of $Ir(COD)(Ph_2MeP)_2PF_6$ (236 mg, 10 mol %) the

solution turned red and the mixture was purged with H₂ (g) for ~ 1 min. After stirring under argon for 4 h, the solution was diluted with THF (30 ml) and sat. aq. NaHCO₃ (100 ml). After the addition of I₂ (1.06 g, 4.19 mmol), the mixture was allowed to stir overnight at room temperature. The mixture was diluted with EtOAc (250 ml) and washed with sat. aq. Na₂S₂O₃ (2 x 80 ml) and brine (80 ml), respectively. The organic layer was dried over MgSO₄ and concentrated *in vacuo*. Column chromatography (EtOAc/toluene) afforded **17** (2.05 g, 1.86 mmol, 67%) as pale yellow foam. $[\alpha]_{\rm D^{20}}$ (CHCl₃): +29.4; IR: 748, 997, 1028, 1074, 1371, 2360, 2858 cm⁻¹; ¹H NMR (400 MHz): δ = 1.07 (s, 9H, *t*-Bu), 3.44 (dd, 1H, *J* = 5.3 Hz, 8.1 Hz, OH), 3.50 - 3.68 (m, 8H, 2 x CH*H* glycerol, 2 x H-2, 2 x H-4, 2 x H-6), 3.76 - 3.91 (m, 5H, 2 x CH*H* glycerol, CH glycerol, H-5, H-6), 3.96 (dd, 1H, *J* = 4.9 Hz, 10.2 Hz, H-6), 4.02 (at, 1H, *J* = 9.3 Hz, H-3), 4.16 (at, 1H, *J* = 9.3 Hz, H-3), 4.13 - 4.22 (m, 1H, H-5), 4.73 - 4.86 (m, 5H, H-1, 2 x CH₂ Bn), 4.94 (d, 1H, *J* = 11.1 Hz, CH*H* Bn), 4.97 (d, 1H, *J* = 10.7 Hz, CH*H* Bn), 5.09 (d, 1H, *J* = 3.6 Hz, H-1), 5.48 (s, 1H, CH

benzylidene), 5.50 (s, 1H, CH benzylidene), 7.12 - 7.15 (m, 3H, H_{arom}), 7.27 - 7.47 (m, 28H, H_{arom}), 7.66 - 7.68 (m, 4H, H_{arom}); ¹³C NMR (100 MHz): δ = 19.1 (C_q *t*-Bu), 26.8 (3 x CH₃ TBDPS), 62.4 (CH₂ glycerol), 62.5 (C-5), 62.8 (C-5), 63.7 (CH₂ glycerol), 68.7 (2 x C-6), 74.0 (CH₂ Bn), 75.1 (CH₂ Bn), 75.2 (CH₂ Bn), 76.4 (C-3), 77.2 (C-2), 77.7 (C-2), 78.8 (C-3), 80.7 (CH glycerol), 78.1 (C-3), 79.0 (C-2), 82.5 (C-4), 82.8 (C-4), 97.6 (C-1), 98.6 (C-1), 101.2 (CH benzylidene), 101.3 (CH benzylidene), 126.0 - 129.8 (CH_{arom}), 133.0, 133.1 (2 x C_q phenyl), 135.5 (CH_{arom}), 137.2, 137.5, 137.6, 138.0, 138.5 (3 x C_q Bn, 2 x C_q benzylidene); HRMS: C₆₆H₇₂O₁₃Si + Na⁺ requires 1123.4634, found 1123.4648.



$\label{eq:a-def} 1-O-(tert-Butyldiphenylsilyl)-2-O-(2-[2,3-di-O-benzyl-4,6-O-benzylidene-\alpha-D-glucopyranosyl]-3-O-benzyl-4,6-O-benzylidene-\alpha-D-glucopyranosyl)-3-O-(4,4'-dimethoxytrityl)-sn-glycerol (18)$

Alcohol **17** (480 mg, 0.436 mmol) was dissolved in DCM (20 ml) and cooled to 0 °C. After the addition of DIPEA (0.23 ml, 1.3 mmol)

and DMT-Cl (295 mg, 0.872 mmol) respectively, the mixture was allowed to stir at room temperature overnight. The reaction was quenched upon addition of H_2O (0.2 ml), and stirred for 15 min. The mixture was diluted with DCM (20 ml) and, after washing with H₂O (10 ml) and brine (10 ml), the organic layer was dried over Na₂SO₄ and concentrated *in vacuo*. The resulting oil was purified by size-exclusion chromatography (sephadex LH-20, MeOH/DCM 1/1) and, subsequently, column chromatography (EtOAc/pentane/Et₃N), giving DMTr-ether 18 (601 mg, 0.428 mmol, 98%) as white foam. [α]_{D²⁰} (CHCl₃): + 35.6; IR: 752, 1030, 1074, 1250, 1508, 2361, 2858 cm⁻¹; ¹H NMR (400 MHz): $\delta = 0.97$ (s, 9H, t-Bu), 3.08 (dd, 1H, I = 5.7 Hz, 10.1 Hz, CHH glycerol), 3.44 (dd, 1H, J = 3.4 Hz, 9.2 Hz, H-2), 3.51 (dd, 1H, J = 2.9 Hz, 10.1 Hz, CHH glycerol), 3.55 - 3.71 (m, 12H, 2 x OMe, 2 x CHH glycerol, 2 x H-4, 2 x H-6), 3.77 (dd, 1H, J = 3.4 Hz, 9.4 Hz, H-2), 3.93 - 4.02 (m, 3H, CH glycerol, H-3, H-5), 4.07 - 4.17 (m, 4H, H-3, H-5, 2 x H-6), 4.30 (d, 1H, *J* = 11.5 Hz, CH*H* Bn), 4.36 (d, 1H, *J* = 11.4 Hz, CH*H* Bn), 4.56 (d, 1H, *J* = 11.4 Hz, CH*H* Bn), 4.72 (d, 1H, J = 11.4 Hz, CHH Bn), 4.78 (d, 1H, J = 11.0 Hz, CHH Bn), 4.88 (d, 1H, J = 11.0 Hz, CHH Bn), 4.98 (d, 1H, / = 3.4 Hz, H-1), 5.39 (d, 1H, / = 3.4 Hz, H-1), 5.53 (s, 1H, CH benzylidene), 5.55 (s, 1H, CH benzylidene), 6.74 (d, 4H, J = 8.7 Hz, Harom), 7.08 - 7.48 (m, 40H, Harom), 7.55 (d, 2H, J = 7.1 Hz, Harom), 7.59 (d, 2H, J = 6.7 Hz, Harom); ¹³C NMR (100 MHz): δ = 19.1 (C_q t-Bu), 26.8 (3 x CH₃) TBDPS), 55.1 (2 x OMe), 62.5 (C-5), 62.8 (C-5), 63.3 (CH₂ glycerol), 64.3 (CH₂ glycerol), 68.8 (C-6), 68.9 (C-6), 72.4 (CH₂ Bn), 75.0 (CH₂ Bn), 75.4 (CH₂ Bn), 75.7 (C-2), 76.8, 76.9 (C-3, CH glycerol), 78.1 (C-3), 79.4 (C-2), 82.1 (C-4), 82.3 (C-4), 86.4 (Cq DMT), 95.2 (C-1), 95.6 (C-1), 101.2 (CH benzylidene), 101.3 (CH benzylidene), 113.1 (CH_{arom}), 126.1 - 130.0 (CH_{arom}), 133.1, 133.2 (2 x Cq phenyl), 135.5 (CHarom), 137.5, 137.7, 138.1, 138.2, 138.7, 144.9, 158.4 (3 x Cq Bn, 2 x Cq benzylidene, 5 x Cq DMT); HRMS: C87H90O15Si + Na⁺ requires 1425.5941, found 1425.5961.



2-0-(2-[2,3-Di-0-benzyl-4,6-0-benzylidene-α-D-glucopyranosyl]-3-0-benzyl-4,6-0-benzylidene-α-D-glucopyranosyl)-3-0-(4,4'-dimethoxytrityl)-sn-glycerol (19)

To a solution of kojibiose derivative **18** (575 mg, 0.410 mmol) in THF (10 ml), was added TBAF (1.00 M solution in THF, 1.20 ml). After stirring for 48 h at RT, the solvent was removed *in vacuo*. The

residue was purified by column chromatography (EtOAc/pentane/Et₃N), affording compound **19** (435 mg, 0.373 mmol, 91%) as white foam. $[\alpha]_{D^{20}}$ (CHCl₃): + 38.0; IR: 1030, 1074, 1508, 2341, 2361 cm⁻¹; ¹H NMR (400 MHz): δ = 2.52 (bs, 1H, OH), 3.02 (dd, 1H, *J* = 4.1 Hz, 9.9 Hz, CH*H* glycerol), 3.44 - 3.49 (m, 2H, CH*H* glycerol, H-2), 3.55 - 3.78 (m, 14H, 2 x OMe, 2 x CH*H* glycerol, CH glycerol, H-2, 2 x H-4, 2 x H-6), 3.96 (at, 1H, *J* = 9.3 Hz, H-3), 4.04 - 4.16 (m, 4H, H-3, 2 x H-5), H-6), 4.31 (dd, 1H, *J* = 4.8 Hz, 10.2 Hz, H-6), 4.49 - 4.55 (m, 3H, 3 x CH*H* Bn), 4.73 (d, 1H, *J* = 11.4

Hz, CH*H* Bn), 4.83 (d, 1H, *J* = 11.0 Hz, CH*H* Bn), 4.90 (d, 1H, *J* = 3.3 Hz, H-1), 4.93 (d, 1H, *J* = 11.0 Hz, CH*H* Bn), 5.17 (d, 1H, *J* = 3.5 Hz, H-1), 5.52 (s, 1H, CH benzylidene), 5.58 (s, 1H, CH benzylidene), 6.75 (d, 2H, *J* = 8.8 Hz, H_{arom}), 6.76 (d, 2H, *J* = 8.8 Hz, H_{arom}), 7.16 - 7.50 (m, 34H, H_{arom}); ¹³C NMR (100 MHz): δ = 55.1 (2 x OMe), 63.0 (2 x C-5), 63.8 (CH₂ glycerol), 64.3 (CH₂ glycerol), 68.7 (C-6), 68.9 (C-6), 73.5 (CH₂ Bn), 75.0 (CH₂ Bn), 75.4 (CH₂ Bn), 76.9 (C-3), 77.0 (C-2), 78.4 (C-3), 79.0 (C-2), 79.8 (CH glycerol), 82.3 (C-4), 82.5 (C-4), 86.3 (C_q DMTr), 97.1 (C-1), 97.2 (C-1), 101.2 (CH benzylidene), 101.3 (CH benzylidene), 113.1 (CH_{arom}), 126.0 - 129.9 (CH_{arom}), 135.7, 135.9, 137.2, 137.6, 137.8, 138.1, 138.6, 144.7, 158.4 (3 x C_q Bn, 2 x C_q benzylidene, 5 x C_q DMTr); HRMS: C₇₁H₇₂O₁₅ + Na⁺ requires 1187.4763, found 1187.4782.



1-([*N*,*N*-Diisopropylamino]-2-cyanoethyl-phosphite)-2-*O*-(2-[2,3-di-*O*-benzyl-4,6-*O*-benzylidene-α-D-glucopyranosyl]-3-*O*-benzyl-4,6-*O*-benzylidene-α-D-glucopyranosyl)-3-*O*-(4,4'-dimethoxytrityl)-sn-glycerol (4)

To a cooled (0 °C) solution of alcohol **19** (1.37 g, 1.17 mmol) and DIPEA (0.31 ml, 1.75 mmol) in freshly distilled DCM (12 ml) was added chlorophosphite **11** (333 mg, 1.41 mmol). After stirring for 4 h, the reaction was quenched with H_2O (1.0 ml),

diluted with DCM (50 ml) and washed with H₂O (20 ml) and brine (20 ml) respectively. The organic layer was dried over Na₂SO₄ and concentrated *in vacuo*. After purification of the residue by column chromatography (EtOAc/pentane/Et₃N), phosphoramidite **4** (1.15 g, 0.842 mmol, 72%) was obtained as colourless oil. ³¹P NMR (161.7 MHz, CD₃CN): δ = 148.5, 148.7 (diastereoisomers); ¹H NMR (400 MHz, CD₃CN): δ = 1.10 - 1.17 (m, 12H, CH₃ isopropylamino), 2.46 - 2.52 (m, 2H, CH₂ cyanoethyl), 3.12 - 3.16 (m, 1H, CH isopropylamino), 3.32 - 3.39 (m, 1H, CH isopropylamino), 3.51 - 4.15 (m, 24H, 2 x OMe, 2 x CH₂ glycerol, CH glycerol, 2 x H-2, 2 x H-3, 2 x H-4, 2 x H-5, 3 x H-6, CH₂ cyanoethyl), 4.29 (dd, 1H, *J* = 4.2 Hz, 10.0 Hz, H-6), 4.39 (d, 1H, *J* = 11.4 Hz, CH*H* Bn), 4.46 (d, 1H, *J* = 11.3 Hz, CH*H* Bn), 4.60 - 4.70 (m, 2H, 2 x CH*H* Bn), 4.81 (s, 2H, 2 x CH*H* Bn), 5.12 (d, 1H, *J* = 3.3 Hz, H-1), 5.30 (d, 0.5H, *J* = 3.3 Hz, H-1 diastereomer 1), 5.34 (d, 0.5H, *J* = 3.3 Hz, H-1 diastereomer 2), 5.60 (s, 1H, CH benzylidene), 5.66 (s, 1H, CH benzylidene), 6.82 (d, 4H, *J* = 8.6 Hz, Harom), 7.16 - 7.50 (m, 34H, Harom); TLC-MS: C₈₀H₈₉N₂O₁₆P + H⁺ requires 1365.60, found 1365.6.



benzyl 6-([*N*,*N*-diisopropylamino]-2-cyanoethyl-phosphite)-hexyl-1-carbamate (5)

To a cooled (0 °C) solution of 6-(benzyloxycarbonylamino)-1hexanol⁴¹ (3.02 g, 12.0 mmol) and DiPEA (2.51 ml, 14.4 mmol) in DCM (60 ml) was added chlorophosphite **11** (2.90 g, 12.2 mmol). After stirring for 4 h, the reaction was washed with H₂O

(20 ml) and brine (20 ml), respectively. The organic layer was dried over Na₂SO₄ and concentrated *in vacuo*. After purification of the residue by column chromatography (EtOAc/PE/Et₃N), phosphoramidite **5** (3.97 g, 8.79 mmol, 73%) was obtained as colourless oil. ³¹P NMR (161.7 MHz, CD₃CN): δ = 148.2; ¹H NMR (400 MHz, CD₃CN): δ = 1.18 (d, 6H, *J* = 2.7 Hz, 2 x CH₃ isopropylamino), 1.19 (d, 6H, *J* = 2.7 Hz, 2 x CH₃ isopropylamino), 1.30 - 1.43 (m, 4H, 2 x CH₂ aminohexanol), 1.48 (m, 2H, CH₂ aminohexanol), 1.59 (m, 2H, CH₂ aminohexanol), 2.65 (t, 2H, *J* = 6.0 Hz, CH₂ cyanoethyl), 3.10 (dd, 2H, *J* = 6.7 Hz, 13.2 Hz, CH₂-N aminohexanol), 3.57 - 3.68 (m, 4H, CH₂-O aminohexanol), 2 x CH isopropylamino), 3.73 - 3.85 (m, 2H, CH₂ cyanoethyl), 5.06 (bs, 2H, CH₂ benzylcarbamate), 5.64 (bs, 1H, NH), 7.31 - 7.42 (m, 5H, H_{arom}); TLC-MS: C₂₃H₃₈N₃O₄P + H⁺ requires 452.27, found 452.1.

Glycerol phosphate dimer (21)



To a solution of dibenzylglycerol **2** (163 mg, 0.600 mmol) in acetonitrile (4.0 ml), containing freshly activated MS3Å, were added DCI (0.25 M in acetonitrile, 9.6 ml) and phosphoramidite **3** (0.2 M in acetonitrile, 3.9 ml). After stirring for 2 h, I₂ (0.2 M in THF/pyridine 4/1, 9.0 ml) and H₂O (1.0 ml) were added and the

mixture was allowed to stir for 1 h. EtOAc (80 ml) was added and the mixture was washed with sat. aq. Na₂S₂O₃ (30 ml), aqueous KHSO₄ (0.5 M, 30 ml), sat. aq. NaHCO₃ (30 ml) and brine (30 ml) respectively. The organic layer was dried with Na₂SO₄ and concentrated *in vacuo*, after which the residue was redissolved in DCM (30 ml), containing freshly activated MS3Å. Et₃SiH (0.97 ml, 6.0 mmol) and dichloroacetic acid (0.49 ml, 6.0 mmol) were added and the mixture stirred until the orange colour disappeared (~ 1 h), after which the mixture was diluted with DCM (20 ml) and washed with sat. aq. $NaHCO_3$ (20 ml) and brine (20 ml), respectively. The organic layer was dried with Na₂SO₄, concentrated *in vacuo* and subsequently purified by column chromatography (MeOH/DCM), yielding dimer 21 (272 mg, 0.478 mmol, 80%, mixture of diastereoisomers) as colourless oil. ³¹P NMR (161.7 MHz): $\delta = -0.6, -0.7; ^{1}H$ NMR (400 MHz): δ = 2.50 (t, 1H, *J* = 6.2 Hz, CH₂ cyanoethyl), 2.54 (t, 1H, *J* = 6.2 Hz, CH₂ cyanoethyl), 2.66 (bs, 1H, OH), 3.54 - 3.73 (m, 5H, CH glycerol, 2 x CH₂ glycerol), 3.77 - 3.82 (m, 1H, CH glycerol), 4.03 -4.32 (m, 6H, CH₂ cyanoethyl, 2 x CH₂ glycerol), 4.51 - 4.66 (m, 6H, 3 x CH₂ Bn), 7.25 - 7.35 (m, 15H, H_{arom}); ¹³C NMR (100 MHz): δ = 19.2, 19.3, 19.3 (CH₂ cyanoethyl), 60.6, 60.7 (CH₂ glycerol), 61.9 (CH₂ cyanoethyl), 66.3, 66.4 (CH₂ glycerol), 67.4, 67.5, 67.6 (CH₂ glycerol), 68.6 (CH₂ glycerol), 71.9, 72.0 (CH₂ Bn), 72.1 (CH₂ Bn), 73.4 (CH₂ Bn), 76.2, 76.3 (CH glycerol), 77.3, 77.4, 77.5 (CH glycerol), 116.4 (Cq cyanoethyl), 127.6 - 128.4 (CHarom), 137.7, 137.8, 137.8 (Cq Bn); HRMS: C₃₀H₃₇NO₈P + Na⁺ requires 592.2071, found 592.2070.



Glycerol phosphate trimer (22)

To a solution of dimer **21** (229 mg, 0.402 mmol) in acetonitrile (4.0 ml), containing freshly activated MS3Å, were added DCI (0.25 M in acetonitrile, 6.4 ml) and phosphoramidite **3** (0.2 M in acetonitrile, 2.6 ml). After stirring for 2 h, more **3** (0.2 M in acetonitrile, 0.8 ml) was added and the mixture was allowed to

react for another 2 hrs. I₂ (0.2 M in THF/pyridine 4/1, 6.0 ml) and H₂O (1.0 ml) were added and the mixture was stirred for 1 h. EtOAc (50 ml) was added and the mixture was washed with sat. aq. Na₂S₂O₃ (20 ml), aqueous KHSO₄ (0.5 M, 20 ml), sat. aq. NaHCO₃ (20 ml) and brine (20 ml) respectively. The organic layer was dried with Na₂SO₄ and concentrated in vacuo, after which the residue was redissolved in DCM (20 ml), containing freshly activated MS3Å. Et₃SiH (0.65 ml, 4.0 mmol) and dichloroacetic acid (0.32 ml, 4.0 mmol) were added and the mixture stirred until the orange colour disappeared (\sim 1.5 h), after which the mixture was diluted with DCM (30 ml) and washed with sat. aq. $NaHCO_3$ (20 ml) and brine (20 ml) respectively. The organic layer was dried with Na₂SO₄, concentrated *in vacuo* and the residue purified by column chromatography (MeOH/DCM), yielding trimer 22 (286 mg, 0.330 mmol, 82%) as colourless oil. ³¹P NMR (161.7 MHz): δ = -1.2, -1.2, -1.1, -1.0, -0.8, -0.8; ¹H NMR (400 MHz): δ = 2.47 - 2.58 (m, 4H, 2 x CH₂) cyanoethyl), 2.97 (bs, 1H, OH), 3.53 - 3.80 (m, 7H, 3 x CH glycerol, 2 x CH₂ glycerol), 4.03 - 4.32 (m, 12H, 2 x CH₂ cyanoethyl, 4 x CH₂ glycerol), 4.48 - 4.66 (m, 8H, 4 x CH₂ Bn), 7.25 - 7.33 (m, 20H, H_{arom}); ¹³C NMR (100 MHz): δ = 19.2 - 19.4 (CH₂ cyanoethyl), 60.5, 60.6 (CH₂ glycerol), 61.8 - 62.1 (2 x CH₂ cyanoethyl), 65.5 - 65.9 (2 x CH₂ glycerol), 66.5 - 66.7 (CH₂ glycerol), 67.4 - 67.7 (CH₂ glycerol), 68.6 - 68.7 (CH₂ glycerol), 71.9 - 72.1 (3 x CH₂ Bn), 73.4 (CH₂ Bn), 75.3 - 75.5 (CH glycerol), 76.3 - 76.4 (CH glycerol), 77.4 - 77.6 (CH glycerol), 116.4 - 116.5 (2 x C_q cyanoethyl), 127.6 - 128.4 (CHarom), 137.2, 137.7 - 137.8 (4 x Cq Bn); HRMS: C43H52N2O13P2 + Na⁺ requires 889.2837, found 889.2843.

Glycerol phosphate tetramer (23)



To a solution of trimer **22** (280 mg, 0.323 mmol) in acetonitrile (3.0 ml), containing freshly activated MS3Å, were added DCI (0.25 M in acetonitrile, 5.2 ml) and phosphoramidite **3** (0.2 M in acetonitrile, 2.6 ml). After stirring for 2 h, more **3** (0.2 M in acetonitrile, 1.3 ml) was added and the mixture was allowed to

react for another 2 h. I₂ (0.2 M in THF/pyridine 4/1, 6.5 ml) and H₂O (1.0 ml) were added and the mixture was stirred for 1 h. EtOAc (50 ml) was added and the mixture was washed with sat. aq. Na₂S₂O₃ (20 ml), aqueous KHSO₄ (0.5 M, 20 ml), sat. aq. NaHCO₃ (20 ml) and brine (20 ml) respectively. The organic layer was dried with Na₂SO₄ and concentrated *in vacuo*, after which the residue was redissolved in DCM (15 ml), containing freshly activated MS3Å. Et₃SiH (0.52 ml, 3.23 mmol) and dichloroacetic acid (0.27 ml, 3.23 mmol) were added and the mixture stirred until the orange colour disappeared (~ 2 h), after which the mixture was diluted with DCM (20 ml) and washed with sat. aq. NaHCO₃ (15 ml) and brine (15 ml) respectively. The organic layer was dried with Na₂SO₄, concentrated in vacuo and, subsequently, purified by column chromatography (MeOH/DCM), vielding tetramer 23 (305 mg, 0.262 mmol, 81%) as colourless oil. ³¹P NMR (161.7 MHz): δ = -1.3 - 0.8; ¹H NMR (400 MHz): δ = 2.47 - 2.61 (m, 6H, 3 x CH₂) cyanoethyl), 3.20 (bs, 1H, OH), 3.56 - 3.58 (m, 2H, CH₂ glycerol) 3.62 - 3.71 (m, 3H, CH glycerol, CH₂ glycerol), 3.78 - 3.81 (m, 3H, 3 x CH glycerol), 4.03 - 4.30 (m, 18H, 3 x CH₂ cyanoethyl, 6 x CH₂ glycerol), 4.50 - 4.65 (m, 10H, 5 x CH₂ Bn), 7.26 - 7.34 (m, 25H, H_{arom}); ¹³C NMR (100 MHz): $\delta = 19.2 - 19.4$ (3 x CH₂ cyanoethyl), 60.4, 60.5 (CH₂ glycerol), 61.8 - 62.2 (3 x CH₂ cyanoethyl), 65.5 - 66.0 (4 x CH₂ glycerol), 66.5 - 66.7 (CH₂ glycerol), 67.4 - 67.7 (CH₂ glycerol), 68.6 (CH₂ glycerol), 71.9 - 72.1 (4 x CH2 Bn), 73.4 (CH2 Bn), 75.2 - 75.5 (2 x CH glycerol), 76.2 - 76.4 (CH glycerol), 77.4 - 77.6 (CH glycerol), 116.5 (3 x Cq cyanoethyl), 127.6 - 128.4 (CH_{arom}), 137.2, 137.7 - 137.9 (5 x Cq Bn); HRMS: C56H68N3O18P3 + Na⁺ requires 1186.3603, found 1186.3614.

Glycerol phosphate pentamer (24)

To a solution of tetramer **23** (283 mg, 0.243 mmol) in acetonitrile (3.0 ml), containing freshly activated MS3Å, were added DCI (0.25 M in acetonitrile, 4.0 ml) and phosphoramidite **3** (0.2 M in acetonitrile, 2.5 ml). After stirring for 4 h, more **3** (0.2 M in acetonitrile, 1.2 ml) was added and the mixture was

allowed to react overnight. I₂ (0.2 M in THF/pyridine 4/1, 5.0 ml) and H₂O (1.0 ml) were added and the mixture was stirred for 1 h. EtOAc (30 ml) was added and the mixture was washed with sat. aq. Na₂S₂O₃ (15 ml), aqueous KHSO₄ (0.5 M, 15 ml), sat. aq. NaHCO₃ (15 ml) and brine (15 ml) respectively. The organic layer was dried with Na₂SO₄ and concentrated in vacuo, after which the residue was redissolved in DCM (15 ml), containing freshly activated MS3Å. Et₃SiH (0.39 ml, 2.4 mmol) and dichloroacetic acid (0.20 ml, 2.4 mmol) were added and the mixture stirred until the orange colour disappeared (\sim 3 h), after which the mixture was diluted with DCM (20 ml) and washed with, respectively, sat. aq. NaHCO₃ (15 ml) and brine (15 ml). The organic layer was dried with Na₂SO₄, concentrated *in vacuo* and, subsequently, purified by column chromatography (MeOH/DCM), yielding pentamer 24 (221 mg, 0.151 mmol, 62%) as colourless oil. ³¹P NMR (161.7 MHz): δ = -1.3 - -0.8; ¹H NMR (400 MHz): δ = 1.67 (bs, 1H, OH), 2.45 - 2.64 (m, 8H, 4 x CH₂ cyanoethyl), 3.55 - 3.58 (m, 2H, CH₂ glycerol) 3.62 - 3.71 (m, 3H, CH glycerol, CH₂ glycerol), 3.75 - 3.80 (m, 4H, 4 x CH glycerol), 4.04 - 4.28 (m, 24H, 4 x CH₂ cyanoethyl, 8 x CH₂ glycerol), 4.49 - 4.51 (m, 2H, CH₂ Bn), 4.56 - 4.66 (m, 10H, 5 x CH₂ Bn), 7.26 -7.35 (m, 30H, H_{arom}); ¹³C NMR (100 MHz): δ = 19.2 - 19.4 (4 x CH₂ cyanoethyl), 60.2 - 60.4 (CH₂ glycerol), 62.0 - 62.5 (4 x CH₂ cyanoethyl), 65.6 - 66.1 (6 x CH₂ glycerol), 66.7 - 66.9 (CH₂ glycerol), 67.6 - 67.9 (CH₂ glycerol), 68.5 (CH₂ glycerol), 71.9 - 72.2 (5 x CH₂ Bn), 73.4 (CH₂ Bn), 75.2 - 75.4 (3 x CH glycerol), 76.3 - 76.4 (CH glycerol), 77.4 - 77.5 (CH glycerol), 116.6 - 116.8 (4 x C_q cyanoethyl), 127.6 - 128.5 (CH_{arom}), 137.2 - 137.3, 137.7 - 137.8 (6 x C_q Bn); HRMS: $C_{69}H_{84}N_4O_{23}P_4$ + Na⁺ requires 1483.4369, found 1483.4377.



Hexamer (25)

To a solution of pentamer **24** (150 mg, 0.103 mmol) and kojibiose-glycerol phosphoramidite **5** (350 mg, 0.257 mmol) in acetonitrile (3.0 ml), containing freshly activated MS3Å, was added DCI (0.25 M in acetonitrile, 2.0 ml). After stirring the mixture overnight at room temperature, I_2 (0.2 M in THF/pyridine 4/1, 2.5 ml) and H₂O (1.0 ml) were added and the mixture was stirred for 1 h. The mixture was diluted with EtOAc (30 ml) and subsequently washed with sat. aq. Na₂S₂O₃ (10 ml), aqueous KHSO₄ (0.5 M, 10 ml), sat. aq. NaHCO₃ (10

ml) and brine (10 ml). The organic layer was dried with Na₂SO₄ and concentrated *in vacuo*, after which the residue was purified by size-exclusion chromatography (sephadex LH-20, THF), followed by column chromatography (MeOH/DCM), yielding dimethoxytritylated intermediate **25** (243 mg, 88.6 µmol, 86%) as white foam. ³¹P NMR (161.7 MHz): δ = -1.3 - -0.9; ¹H NMR (400 MHz): δ = 2.29 - 2.56 (m, 10H, 5 x CH₂ cyanoethyl), 3.10 - 4.36 (m, 58H, 5 x CH₂ cyanoethyl, 6 x CH glycerol, 12 x CH₂ glycerol, 2 x OMe, 2 x H-2, 2 x H-3, 2 x H-4, 2 x H-5, 4 x H-6), 4.45 - 4.89 (m, 18H, 9 x CH₂ Bn), 4.92 - 4.94 (m, 1H, H-1), 5.10 - 5.13 (m, 1H, H-1), 5.52 (s, 1H, CH benzylidene), 5.56 (s, 1H, CH benzylidene), 6.74 (d, 4H, *J* = 8.6 Hz, H_{arom}), 7.14 - 7.48 (m, 64H, H_{arom}); HRMS: [C₁₄₃H₁₅₈N₅O₄₀P₅ + Na]²⁺ requires 1392.9478, found 1392.9489.



Hexamer (26)

Intermediate **25** (122 mg, 44.5 µmol) was dissolved in a mixture of DCM (2.5 ml) and MeOH (25 ml). After the addition of pyridinium *para*-toluenesulphonate (PPTS, 25 mg, 0.099 mmol), the solution was allowed to stir overnight. Toluene (50 ml) was added and the mixture was concentrated partially (leaving ~30 ml) under reduced pressure. Column chromatography of the solution (MeOH/DCM) yielded hexamer **26** (76.0 mg, 31.1 µmol, 70%) as colourless oil. ³¹P NMR (161.7 MHz): δ = -1.3 - 0.8; ¹H NMR (400 MHz): δ = 1.79 (bs, 1H, OH),

2.44 - 2.57 (m, 10H, 5 x CH₂ cyanoethyl), 3.38 - 4.34 (m, 52H, 5 x CH₂ cyanoethyl, 6 x CH glycerol, 12 x CH₂ glycerol, 2 x H-2, 2 x H-3, 2 x H-4, 2 x H-5, 4 x H-6), 4.51 - 4.65 (m, 12H, 6 x CH₂ Bn), 4.76 - 4.99 (m, 7H, 3 x CH₂ Bn, H-1), 5.11 - 5.13 (m, 1H, H-1), 5.50 (s, 1H, CH benzylidene), 5.55 (s, 1H, CH benzylidene), 7.15 - 7.17 (m, 3H, H_{arom}), 7.26 - 7.40 (m, 48H, H_{arom}), 7.42 - 7.47 (m, 4H, H_{arom}); HRMS: [C₁₂₂H₁₄₀N₅O₃₈P₅ + Na]²⁺ requires 1241.8824, found 1241.8837.



Hexamer-spacer (27)

To a solution of kojibiose substituted hexamer **26** (124 mg, 51.0 μ mol) and Zaminohexylphosphoramidite **5** (184 mg, 0.408 mmol) in acetonitrile (3.0 ml), containing freshly activated MS3Å, was added DCI (0.25 M in acetonitrile, 2.0 ml). After stirring the mixture for 4 h, acetonitrile (5.0 ml), I₂ (0.2 M in THF/pyridine 4/1, 3.1 ml) and H₂O (1.0 ml) were added and the mixture stirred for 1 h. The mixture was diluted with EtOAc (30 ml) and subsequently washed with sat. aq. Na₂S₂O₃ (10 ml), aqueous KHSO₄ (0.5 M, 10 ml), sat. aq. NaHCO₃ (10 ml) and brine (10 ml). The organic layer was dried with Na₂SO₄ and concentrated *in vacuo*, after which the residue was purified by size-exclusion chromatography (sephadex LH-20, MeOH/DCM), followed by column chromatography (MeOH/DCM), yielding compound **27** (111 mg, 39.5 µmol, 77%) as colourless oil. ³¹P NMR (161.7 MHz): δ = -1.3 - -0.9; ¹H NMR (400 MHz): δ = 1.18 - 1.32 (m, 4H, 2 x CH₂ hexylspacer), 1.35 - 1.44 (m, 2H, CH₂ hexylspacer), 1.53 - 1.62 (m, 2H, CH₂ hexylspacer), 2.44 - 2.58 (m, 12H, 6 x CH₂ cyanoethyl), 3.05 - 3.13 (m, 2H, CH₂-N hexylspacer), 3.56 - 4.33 (m, 56H, 6 x CH₂ cyanoethyl, 6 x CH glycerol, 12 x CH₂ glycerol, CH₂-O hexylspacer, 2 x H-2, 2 x H-3, 2 x H-4, 2 x H-5, 4 x H-6), 4.50 - 4.59 (m, 12H, 6 x CH₂ Bn), 4.74 - 5.15 (m, 10H, 4 x CH₂ Bn, 2 x H-1), 5.53 (s, 1H, CH benzylidene), 7.16 - 7.18 (m, 3H, H_{arom}), 7.26 - 7.39 (m, 53H, H_{arom}), 7.43 - 7.48 (m, 4H, H_{arom}); HRMS: [C₁₃₉H₁₆₃N₇O₄₃P₆ + Na]²⁺ requires 1424.9497 found 1424.9512.



Teichoic acid (1)

A solution of protected hexamer **27** (51.0 mg, 18.2 μ mol) in dioxane (5.0 ml) was diluted with ammonia (25% in H₂O, 5.0 ml) and stirred overnight. The solution was concentrated *in vacuo* and the residue redissolved in H₂O (2.0 ml). The solution was then eluted through a small column containing Amberlite Na⁺ resin. The volatiles were removed under reduced

pressure and the residue lyophilized, yielding the partially protected intermediate (47.6 mg, 100%) as amorphous white solid. A part of the intermediate (23.6 mg, 9.01 μ mol) was dissolved in a mixture of dioxane (2.0 ml), H_2O (2.0 ml) and AcOH (three drops). After the addition of Palladium black (~50 mg, 0.5 mmol), the mixture was allowed to stir for 5 days under a hydrogen atmosphere. The mixture was filtered, concentrated in vacuo, and purified by size exclusion chromatography (Sephadex HW40, 0.15M NH₄HCO₃). The purified residue was lyophilized twice before it was dissolved in H_2O (2.0 ml) and eluted through a small column containing Amberlite Na⁺ resin. Cryodesiccation of the filtrate yielded deprotected teichoic acid **1** (10.6 mg, 6.81 μ mol, 76%) as amorphous white solid. ³¹P NMR (161.7 MHz, D₂O): δ = 0.9 (1P), 1.1 (1P), 1.2 (3P), 1.3 (1P); ¹H NMR (600 MHz, D_2O): $\delta = 1.36 - 1.40$ (m, 4H, 2 x CH₂ hexylspacer), 1.59 - 1.66 (m, 4H, 2 x CH₂ hexylspacer), 2.95 (t, 2H, *J* = 7.5 Hz, CH₂-N hexylspacer), 3.37 - 3.41 (m, 2H, 2 x H-5), 3.51 - 3.56 (m, 2H, CHH glycerol, H-2), 3.61 - 3.64 (m, 2H, CHH glycerol, H-2), 3.70 - 4.01 (m, 37H, 5 x CH glycerol, 11 x CH₂ glycerol, CH₂-O hexylspacer, 2 x H-3, 2 x H-4, 4 x H-6), 4.11 - 4.14 (m, 1H, CH glycerol), 5.11 (d, 1H, / = 3.7 Hz, H-1), 5.39 (d, 1H, / = 3.5 Hz, H-1); ¹³C NMR (150 MHz, D₂O): δ = 25.4, 26.1, 27.6 (3 x CH₂ hexylspacer), 30.4 (d, J = 6.7 Hz, CH₂ hexylspacer), 40.4 (CH₂-N hexylspacer), 61.4, 61.4 (2 x C-6), 63.0 (CH₂ glycerol), 65.5 (d, J = 5.1 Hz, CH₂ glycerol), 66.0 (d, *J* = 4.4 Hz, CH₂ glycerol), 67.0 - 67.3 (9 x CH₂ glycerol, CH₂-O hexylspacer), 70.4 - 70.6 (4 x CH glycerol, 2 x C-5), 71.7 (d, J = 7.8 Hz, CH glycerol), 72.1 (C-2, C-3), 72.7 (2 x C-4), 73.3 (C-3), 75.1 (C-2), 76.0 (t, J = 8.3 Hz, CH glycerol), 95.6 (C-1), 96.4 (C-1). HRMS: C₃₆H₇₇NO₄₁P₆ + H⁺ requires 1366.2469 found 1366.2483.

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- 34. In **chapter five**, the glycosylation of glycerol **7** using the 6-*0*-FMOc protected glucosyl imidate donor from reference 33 is described.
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