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Two-dimensional Transcriptome Analysis in Chemostat Cultures

COMBINATORIAL EFFECTS OF OXYGEN AVAILABILITY AND MACRONUTRIENT LIMITATION IN *SACCHAROMYCES CEREVISIAE**[§]

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Genome-wide analysis of transcriptional regulation is generally studied by determining sets of “signature transcripts” that are up- or down-regulated relative to a reference situation when a single culture parameter or genetic modification is changed. This approach is especially relevant for defining small subsets of transcripts for use in high throughput, cost-effective diagnostic analyses. However, this approach may overlook the simultaneous control of transcription by more than one environmental parameter. This study represents the first quantitative assessment of the impact of transcriptional cross-regulation by different environmental parameters. As a model, we compared the response of aerobic as well as anaerobic chemostat cultures of the yeast *Saccharomyces cerevisiae* to growth limitation by four different macronutrients (carbon, nitrogen, phosphorus, and sulfur). The identity of the growth-limiting nutrient was shown to have a strong impact on the sets of transcripts that responded to oxygen availability and vice versa. We concluded that identification of reliable signature transcripts for specific environmental parameters can be obtained only by combining transcriptome data sets obtained under several sets of reference conditions. Furthermore, the two-dimensional approach to transcriptome analysis is a valuable new tool to study the interaction of different transcriptional regulation systems.

Recent rapid developments in DNA microarray technology have had a strong impact in research on the yeast *Saccharomyces cerevisiae*, an important industrial microorganism and model eukaryote. With the ability to study genome-wide transcriptome expression in a single microarray, large on-line transcriptome data bases obtained from different mutants and under a wide range of cultivation conditions have become available as research tools (Gene Expression Omnibus (1)

and Yeast Microarray Global Viewer (2)).

Transcript profiles contain a wealth of information that may be applied in several ways for fundamental and applied research. When clear correlations are established between cultivation conditions and transcription of subsets of genes, such correlations can be used to guide functional analysis studies of genes with as yet unknown biological functions. Furthermore, correlation of expression data with sequences of upstream regulatory elements can be applied to unravel the intricate networks of transcriptional regulation (3). In industrial biotechnology, one of the key applications of DNA microarrays lies in diagnosing industrial fermentation processes. If transcriptional responses can be directly correlated to important parameters such as nutritional status of industrial microorganisms or to the stresses to which they are exposed in industrial processes, transcriptome analysis can provide invaluable information for process optimization (4, 5). For such diagnostic purposes, it would be preferable to construct small, cost-effective microarrays that contain a limited number of “signature transcripts.” Such signature transcripts should respond uniquely to a single chemical or physical parameter that is relevant for the industrial process under study. This approach is analogous to the application of small diagnostic arrays used in clinical research for the rapid typing of tumors (6).

Hitherto, most transcriptome studies with *S. cerevisiae* have been done in shake-flask cultures (7, 8). In such cultures, it is not possible to control a number of important cultivation conditions (dissolved oxygen concentration, metabolite concentrations, pH, etc.). Therefore, shake-flask cultivation by definition involves a continuously changing environment. Consequently, interpretation of transcriptome data from shake-flask cultivation is likely to be complicated by differences in specific growth rate, carbon catabolite repression, nitrogen catabolite repression, product accumulation, acidification, etc.

Chemostat cultivation offers a number of advantages for studies with DNA microarrays because it enables cultivation of microorganisms under tightly defined environmental conditions. In a chemostat, culture broth (including biomass) is continuously replaced by fresh medium at a fixed and accurately determined dilution rate (D , h^{-1}). When the dilution rate is lower than the maximal specific growth rate of the microorganism (μ_{max} , h^{-1}), a steady-state situation will be established in which the specific growth rate equals the dilution rate ($\mu = D$). In such a steady-state chemostat culture, μ is controlled by the (low) residual concentration of a single growth-limiting nutrient. The option to accurately control and manipulate individual culture parameters (including medium composition, nature of the growth-limiting nutrient, pH, temperature, and μ) under steady-state conditions

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[§] The on-line version of this article (available at <http://www.jbc.org>) contains Supplemental Tables 1–6.

The complete data set has been submitted to the Genome Expression Omnibus Database (available at www.ncbi.nlm.nih.gov/geo/) under series GSE1723.

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makes chemostats excellent tools for studies on genome-wide transcriptional regulation. Indeed, a recent interlaboratory comparison of transcriptome data obtained in chemostat cultures demonstrated that the accuracy and reproducibility of this approach are superior to those obtained in previous studies with shake-flask cultures (9).

Chemostat cultures have recently been applied to study genome-wide transcriptional responses of *S. cerevisiae* to carbon-limited growth on different carbon sources (10); to nutrient limitation for carbon, nitrogen, phosphorus, and sulfur (4); to starvation (11); to the presence and absence of oxygen (9, 12); and to oxidative stress responses (13). In each of these studies, attempts were made to vary a single cultivation parameter while keeping all other parameters constant. This "one-dimensional" approach resulted in sets of signature transcripts that, within the experimental context, responded uniquely to a single cultivation parameter (e.g. uniquely up-regulated under nitrogen limitation, uniquely down-regulated during growth on ethanol). There is an important potential pitfall in this approach, as it does not include the possibility that expression of genes is simultaneously controlled by two or more environmental parameters. Such dual or multiple control would complicate the identification of signature transcripts and the interpretation of diagnostic transcriptome analysis.

So far, there have been no dedicated studies to investigate and quantify the way in which different transcriptional regulation responses overlap and interact. The goal of this study was to study this interaction by analyzing genome-wide transcriptional responses to four different nutrient limitation regimes under aerobic and anaerobic conditions in chemostat cultures of *S. cerevisiae*. This "two-dimensional" approach resulted in a new robust set of "anaerobic" and "aerobic" signature transcripts for *S. cerevisiae* as well as the refinement of previous reports on nutrient-responsive genes. Moreover, the identification of genes regulated both by nutrient and oxygen availability provided new insight in cross-regulated network and hierarchy in the control of gene expression. These newly defined sets of signature genes were subjected to *in silico* promoter analysis to identify consensus regulatory elements.

EXPERIMENTAL PROCEDURES

Strain and Growth Conditions—The *S. cerevisiae* prototrophic haploid reference strain CEN.PK113-7D (*MATa*) (14) was grown at 30 °C in 2-liter chemostats (Applikon) with a working volume of 1.0 liter as described (15). Cultures were fed with a defined synthetic medium that limited growth by carbon, nitrogen, phosphorus, or sulfur with all other growth requirements in excess and at a constant residual concentration (4). The dilution rate was set at 0.10 h⁻¹. The pH was measured on-line and kept constant at 5.0 by the automatic addition of 2 M KOH using an Applikon ADI 1030 Biocontroller, and the stirrer speed was set at 800 rpm. Anaerobic conditions were maintained by sparging the medium reservoir and the fermentor with pure nitrogen gas (0.5 liter min⁻¹). Furthermore, Norprene tubing and butyl septa were used to minimize oxygen diffusion into the anaerobic cultures (16). The off-gas was cooled by a condenser connected to a cryostat set at 2 °C. Oxygen and carbon dioxide were measured off-line with an NGA 2000 Rosemount gas analyzer. Steady-state samples were taken after ~10–14 volume changes to avoid strain adaptation due to long-term cultivation (17, 18). Biomass dry weight, metabolite, dissolved oxygen, and gas profiles were constant over at least three volume changes prior to sampling for RNA extraction.

Growth Media—The synthetic medium composition was based on that described (19). In all chemostats except for carbon, the residual glucose concentration was targeted to 17 g liter⁻¹ to sustain glucose repression at the same level. For anaerobic cultivations, the reservoir medium was supplemented with the anaerobic growth factors Tween 80 and ergosterol as described previously (20). These media contained the following components: for carbon-limited cultivation, 5.0 g liter⁻¹ (NH₄)₂SO₄, 3.0 g liter⁻¹ KH₂PO₄, 0.5 g liter⁻¹ MgSO₄·7H₂O, and 25 g liter⁻¹ glucose; for nitrogen-limited cultivation, 0.65 g liter⁻¹ (NH₄)₂SO₄, 5.75 g liter⁻¹ K₂SO₄, 3.0 g liter⁻¹ KH₂PO₄, 0.5 g liter⁻¹

MgSO₄·7H₂O, and 46 g liter⁻¹ glucose; for phosphorus-limited cultivation, 5.0 g liter⁻¹ (NH₄)₂SO₄, 1.9 g liter⁻¹ K₂SO₄, 0.12 g liter⁻¹ KH₂PO₄, 0.5 g liter⁻¹ MgSO₄·7H₂O, and 66 g liter⁻¹ glucose; and for sulfur-limited cultivation, 4.0 g liter⁻¹ NH₄Cl, 0.05 g liter⁻¹ MgSO₄·7H₂O, 3.0 g liter⁻¹ KH₂PO₄, 0.4 g liter⁻¹ MgCl₂, and 59 g liter⁻¹ glucose. The medium composition for the aerobic chemostat cultures was as described previously (4).

Analytical Methods—Culture supernatants were obtained after centrifugation of samples from the chemostats. For the purpose of glucose determination and carbon recovery, culture supernatants and media were analyzed by high performance liquid chromatography on an AMINEX HPX-87H ion exchange column using 5 mM H₂SO₄ as the mobile phase. Residual ammonium, phosphate, and sulfate concentrations were determined using cuvette tests from DRLANGE (Düsseldorf, Germany). Culture dry weights were determined via filtration as described by Postma *et al.* (21).

Microarray Analysis—Sampling of cells from chemostats, probe preparation, and hybridization to Affymetrix Genechip[®] microarrays were performed as described previously (9). The results for each growth condition were derived from three independently cultured replicates.

Data Acquisition and Analysis—Acquisition and quantification of array images and data filtering were performed using Affymetrix Microarray Suite Version 5.0, MicroDB Version 3.0, and Data Mining Tool Version 3.0. Before comparison, all arrays were globally scaled to a target value of 150 using the average signal from all gene features using Microarray Suite Version 5.0. To eliminate insignificant variations, genes with values below 12 were set to 12 as described (9). From the 9335 transcript features on the YG-S98 arrays, a filter was applied to extract 6383 yeast open reading frames, of which there were 6084 different genes. This discrepancy was due to several genes being represented more than once when suboptimal probe sets were used in the array design. To represent the variation in triplicate measurements, the coefficient of variation (S.D. divided by the mean) was calculated as described previously by Boer *et al.* (4).

For additional statistical analyses, Microsoft Excel running the significance analysis of microarrays (SAM Version 1.12) add-in was used (22) for pairwise comparisons. Genes were considered as being changed in expression if they were called significantly changed using SAM (expected median false discovery rate of 1%) by at least 2-fold from each other condition. Hierarchical clustering of the obtained sets of significantly changed expression levels was subsequently performed using Genespring Version 6.1 (Silicon Genetics).

Promoter analysis was performed using the web-based software Regulatory Sequence Analysis (RSA) Tools (23). The promoters (from -800 to -1) of each set of co-regulated genes were analyzed for over-represented hexanucleotides. When hexanucleotide sequences shared largely common sequences, they were aligned to form longer conserved elements. All of the individual promoter sequences contributing to these elements were then aligned, and the redundant elements were determined by counting the base representation at each position. The relative abundance of these redundant elements was determined from a new enquiry of the co-regulated gene promoters and the entire set of yeast promoters in the genome. The gene annotation was made according to the Comprehensive Yeast Genome Database at the Munich Information Center for Protein Sequence (MIPS; available at mips.gsf.de/genre/proj/yeast/index.jsp) (24), the *Saccharomyces* Genome Database (available at www.yeastgenome.org/) (25) and the Yeast Proteome Database at Incyte (available at www.incyte.com/).

RESULTS

Experimental Design and Physiology of *S. cerevisiae* in Aerobic and Anaerobic Macronutrient-limited Chemostat Cultures—To investigate the impact of transcriptional cross-regulation on the identification of signature transcripts, we designed a two-dimensional experimental approach (Fig. 1). Four nutrient limitation regimes (carbon, nitrogen, sulfur, and phosphorus) were studied. In one set of experiments, the four nutrient limitation regimes were studied in aerobic chemostat cultures. A second set of experiments was performed under the same nutrient limitation regimes, but in anaerobic chemostat cultures. The resulting set of eight fermentation conditions, each analyzed in three independent replicate cultures, enabled the identification of genes with a specific transcriptional response to one parameter only (e.g. induced under anaerobic conditions irrespective of the macronutrient limitation regime).

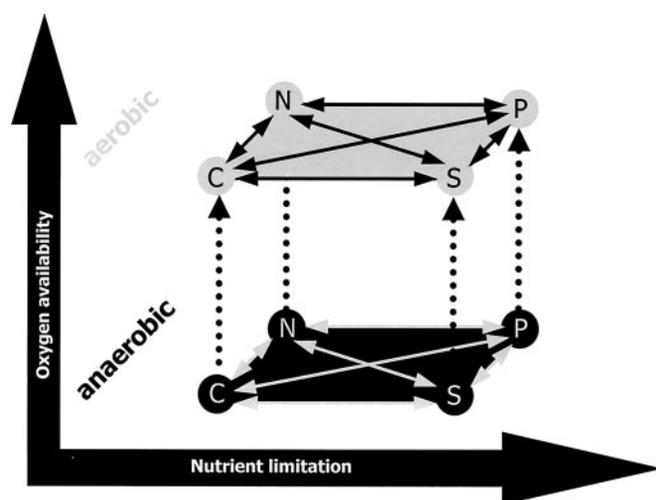


FIG. 1. **Two-dimensional experimental approach.** The experimental design for two-dimensional transcriptome analysis is shown. Each corner of the cube represents a chemostat condition. The upper horizontal surface represents the four aerobic macronutrient limitation regimes (carbon, nitrogen, phosphorus, and sulfur). The lower horizontal surface represents the same macronutrient limitation regimes analyzed under anaerobic conditions. The arrows indicate the pairwise comparisons included in the present two-dimensional transcriptome analysis.

Furthermore, genes that transcriptionally responded to multiple parameters were identified (e.g. induced under anaerobic conditions only when growth was limited by the carbon source).

To minimize experimental “noise,” the compositions of the growth media were designed such that residual concentrations of non-growth-limiting nutrients were essentially the same in all chemostat cultures (Table I). Control experiments confirmed that the concentrations of the growth-limiting nutrients were below the detection limit of the respective assay procedures (Table I). The option to control the steady-state concentrations of limiting and excess nutrients concertedly is a unique feature of chemostat cultivations.

The physiological parameters of the eight cultivation conditions are reported in Table I. Under aerobic conditions, only the glucose-limited cultures exhibited a completely respiratory glucose metabolism, without production of ethanol. This resulted in a respiratory quotient close to unity (Table I). Conversely, the aerobic cultures that were not limited by glucose exhibited a respirofermentative glucose metabolism, with simultaneous ethanol production and oxygen consumption (respiratory quotient of >1). In the anaerobic chemostat cultures, alcoholic fermentation was the sole mode of glucose dissimilation, as no oxygen was available for respiration. The ATP yield from alcoholic fermentation is much lower than that from respiratory glucose dissimilation (20), thus explaining the lower biomass yield on glucose of the anaerobic cultivations. The biomass yield on glucose in glucose-limited cultures was higher than in the non-glucose-limited cultures (Table I). Under aerobic conditions, this can be partially explained by the involvement of alcoholic fermentation in the latter cultures. However, a reduction of the biomass yield in non-glucose-limited cultures was also observed under anaerobic conditions (Table I). This may be related to the induction of energy-dependent transport systems during nitrogen-, phosphorus-, and sulfur-limited growth (26).

Microarray Reproducibility, Global Transcriptome Responses, and Data Analysis—To obtain statistically robust, reproducible transcriptome data sets (9), independent triplicate chemostat cultivations and oligonucleotide DNA microarrays were carried out for each of the eight cultivation conditions. The average coefficient of variation for the triplicate transcrip-

tome analyses (4, 9) for each of the eight conditions was <0.21 , except for the anaerobic glucose-limited chemostats (coefficient of variation of 0.27). The level of the *ACT1* transcript, a common loading standard for conventional Northern analysis, did vary by $<13\%$ over the eight growth conditions (Supplemental Table 1).

The eight different cultivation conditions would, in principle, allow for 56 different pairwise comparisons. In this study, we restricted analysis of the data to pairwise comparisons between cultivation conditions that differed in a single cultivation parameter only. Ultimately, this left 28 pairwise comparisons. Four of these were pairwise comparisons between aerobic and anaerobic cultures grown under the same macronutrient limitation regime (Fig. 1, vertical arrows). An additional 24 pairwise comparisons involved all possible combinations of the four macronutrient limitation regimes under either aerobic or anaerobic conditions (Fig. 1, horizontal surfaces).

Each pairwise comparison defined a set of genes that were significantly up- or down-regulated ($-fold$ change of >2 with a false discovery rate of 1%; see “Experimental Procedures”). In total, 3169 genes (52% of the genome) exhibited a significantly different transcript level in at least one of the 28 pairwise comparisons. 2542 genes (42%) of the genome did not exhibit a significant difference in transcript level in any of the pairwise comparisons. The remaining 373 transcripts (representing 6% of the *S. cerevisiae* genome) remained below the detection limit under all eight conditions investigated (Fig. 2 and Supplemental Table 2).

Transcripts that showed a consistent difference in the aerobic/anaerobic comparisons under all four macronutrient limitation regimes were identified by combining the four relevant pairwise comparisons (Fig. 1, vertical arrows). This set of consistently oxygen-responsive genes contained 155 genes (2.6% of the genome) (Fig. 3A and Supplemental Table 3).

To investigate transcriptional responses to macronutrient limitation, we first identified transcripts that responded to a single nutrient limitation regime under either aerobic or anaerobic conditions (sets I and V) (Fig. 3B). Combination of sets I and V for each of the four macronutrient limitation regimes yielded a subset of transcripts that showed a consistent response to macronutrient limitation irrespective of oxygen availability (set III, 152 genes) (Fig. 3B and Supplemental Table 4). In addition, this comparison yielded two sets of genes that showed a transcriptional response only to one of the four nutrient limitation conditions under either aerobic conditions (set II, 333 genes) (Fig. 3B and Supplemental Table 5) or anaerobic conditions (set IV, 302 genes) (Fig. 3B and Supplemental Table 6). The data analysis approach described above enabled us to dissect the *S. cerevisiae* genome clusters of genes that showed either a consistent, robust response to oxygen availability or macronutrient limitation or, alternatively, a more complex dual-parameter transcriptional regulation.

Signature Genes with a Consistent Transcriptional Response to Oxygen Availability or Macronutrient Limitation—Ten clusters of genes that were identified showed a specific and consistent response to anaerobiosis, glucose limitation, nitrogen limitation, phosphorus limitation, or sulfur limitation (Fig. 4). In five of these clusters, the transcriptional response was defined as “up-regulated” under the conditions indicated; in the other five clusters, the transcriptional response was defined as “down-regulated.” This terminology does not imply any mechanism of regulation. For example, down-regulation under nutrient limitation might, mechanistically, represent up-regulation under conditions of nutrient excess. In our discussion of these “consistent response” genes, we will restrict ourselves to a detailed analysis of the anaerobically up-regulated genes and

TABLE I
Nutrient concentrations and physiological parameters of chemostat cultures used in this study

Unless indicated otherwise, values represent the mean \pm S.D. of data from three independent steady-state chemostat cultivations. RQ, respiratory quotient (q_{CO_2}/q_{O_2}); BD, below detection limit of assay; NA, not applicable.

Growth-limiting nutrient	Residual nutrient measurements				Physiological parameters							Ref.
	Glc	NH ₄ ⁺	PO ₄ ³⁻	SO ₄ ²⁻	Y_{Glc-X}^a	q_{Glu}^b	$q_{ethanol}^c$	$q_{O_2}^d$	$q_{CO_2}^e$	RQ	Carbon recovery	
	g/liter	mM	mM	mM							%	
Aerobic												
Carbon	BD	58.2 \pm 1.3	19.8 \pm 0.6	38.6 ^f	0.49 \pm 0.0	1.1 \pm 0.0	0.0 \pm 0.0	2.8 \pm 0.3	2.8 \pm 0.3	1.0 \pm 0.0	98 \pm 3	4
Nitrogen	16.7 \pm 1.0	BD	18.6 \pm 1.0	40.7 \pm 1.0	0.09 \pm 0.0	5.8 \pm 0.1	8.0 \pm 0.1	2.7 \pm 0.1	12.1 \pm 0.2	4.5 \pm 0.2	96 \pm 1	4
Phosphorus	18.1 \pm 1.0	54.3 \pm 0.0	BD	47.5 \pm 1.0	0.09 \pm 0.0	6.1 \pm 0.2	7.8 \pm 0.1	4.0 \pm 0.1	13.5 \pm 0.2	3.4 \pm 0.0	95 \pm 2	4
Sulfur	17.4 \pm 0.6	53.7 \pm 2.4	18.4 \pm 0.2	BD	0.14 \pm 0.0	3.8 \pm 0.1	4.4 \pm 0.1	3.0 \pm 0.0	8.0 \pm 0.8	2.7 \pm 0.2	96 \pm 1	4
Anaerobic												
Carbon	BD ^g	68.6 \pm 2.8 ^g	22.3 \pm 0.6 ^g	42.4 \pm 1.6 ^g	0.09 \pm 0.0	6.0 \pm 0.0	9.6 \pm 0.1	NA	10.3 \pm 0.4	NA	101 \pm 2	
Nitrogen	16.2 \pm 0.6	BD	21.9 \pm 0.4	39.1 \pm 0.8	0.07 \pm 0.0	8.4 \pm 0.0	13.5 \pm 0.6	NA	14.8 \pm 0.3	NA	101 \pm 2	
Phosphorus	19.1 \pm 2.2	60.2 \pm 2.6	BD	50.9 \pm 0.9	0.06 \pm 0.0	8.7 \pm 0.2	13.9 \pm 0.6	NA	15.8 \pm 0.7	NA	101 \pm 2	
Sulfur	21.2 \pm 0.2	61.1 \pm 1.3	21.5 \pm 0.2	BD	0.07 \pm 0.0	7.9 \pm 0.2	11.9 \pm 0.4	NA	13.6 \pm 0.8	NA	98 \pm 1	

^a Biomass yield on glucose (g of biomass/g of glucose consumed).

^b mmol of glucose consumed per g of biomass/h.

^c mmol of ethanol produced per g of biomass/h.

^d mmol of oxygen consumed per g of biomass/h.

^e mmol of carbon dioxide produced per g of biomass/h.

^f Single assay measurement.

^g Mean \pm S.D. of two separate chemostat steady states.

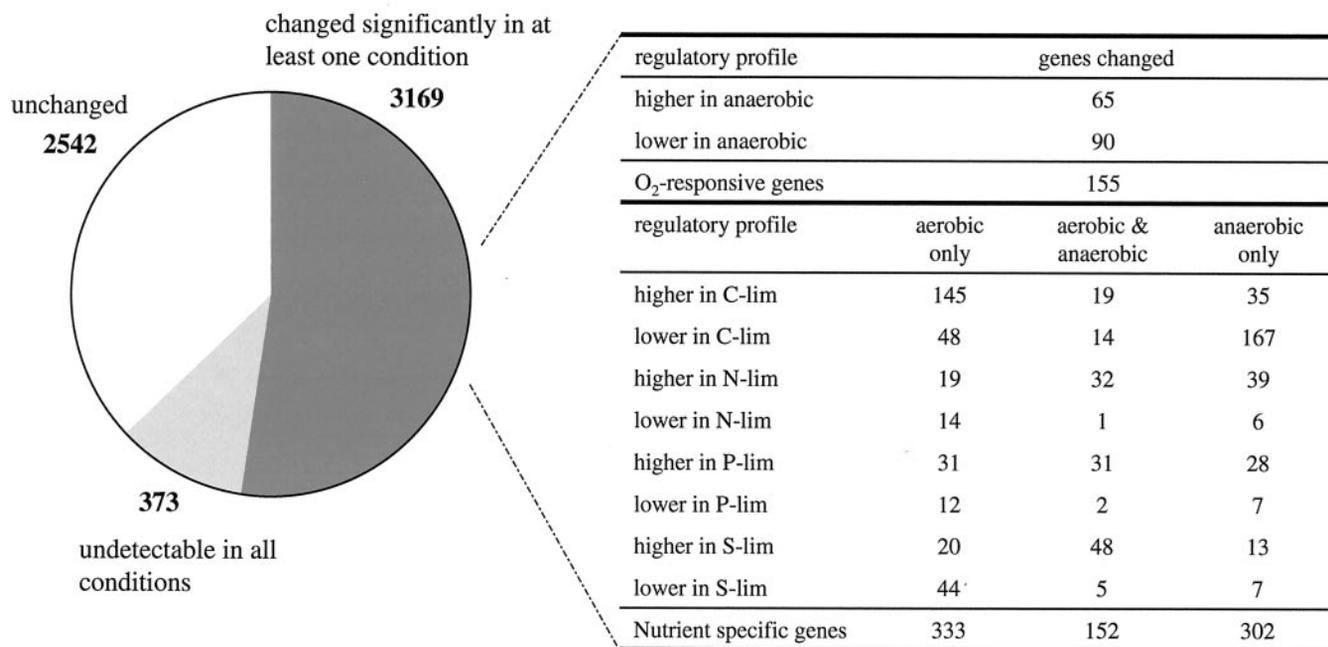


FIG. 2. **Global transcriptional responses to growth in aerobic and anaerobic macronutrient-limited chemostats.** The genome-wide transcript profiles of *S. cerevisiae* grown under different oxygen availability conditions and limitations (*lim*) for carbon, nitrogen, phosphorus, or sulfur are compared, and the classes of expression profiles are scored. About half of the predicted genome (48%) was either unchanged or not measurable across all eight conditions. The remaining significantly changed genes (3169) were categorized into oxygen-responsive genes (155); genes that responded to macronutrient limitation under solely aerobic conditions (333), solely anaerobic conditions (302), and irrespective of the presence of oxygen (152); and genes with a more complex transcription profile.

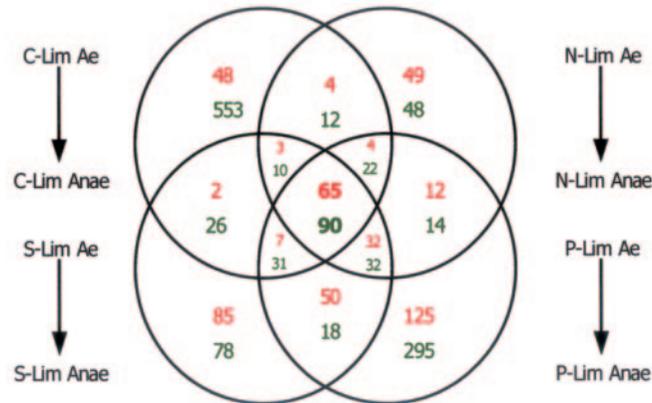
some specific observations on the macronutrient limitation-responsive genes.

Anaerobically Up-regulated Genes—Based on a statistically robust, two-laboratory transcriptome analysis of glucose-limited chemostat cultures of *S. cerevisiae*, Piper *et al.* (9) identified 877 transcripts that were differentially expressed in anaerobic and aerobic cultures. These genes were distributed in 133 anaerobically up-regulated and 744 anaerobically down-regulated genes. In our two-dimensional approach, the transcriptional response to oxygen availability of 722 of these genes (82%) depended on the macronutrient limitation regime, and only 155 genes showed a consistent response to anaerobiosis under all four macronutrient limitation regimes (65 up-regu-

lated and 90 down-regulated) (Fig. 3A).

Of the 65 anaerobically up-regulated genes, 20 have an as yet poorly defined or unknown biological function. The 45 genes with known function were distributed over the functional categories as follows: metabolism and energy (21 genes), transport (four genes), cell rescue and defense (11 genes), protein synthesis (three genes), and cell wall and organization (six genes) according to the MIPS Database (Fig. 4) (24). A closer inspection reflected the biosynthetic role of molecular oxygen in *S. cerevisiae* (27). Under anaerobic conditions, *S. cerevisiae* is not capable of *de novo* biosynthesis of sterols and unsaturated fatty acids, and therefore, these compounds are required as growth factors under anaerobic conditions (28, 29). Although the an-

A



B

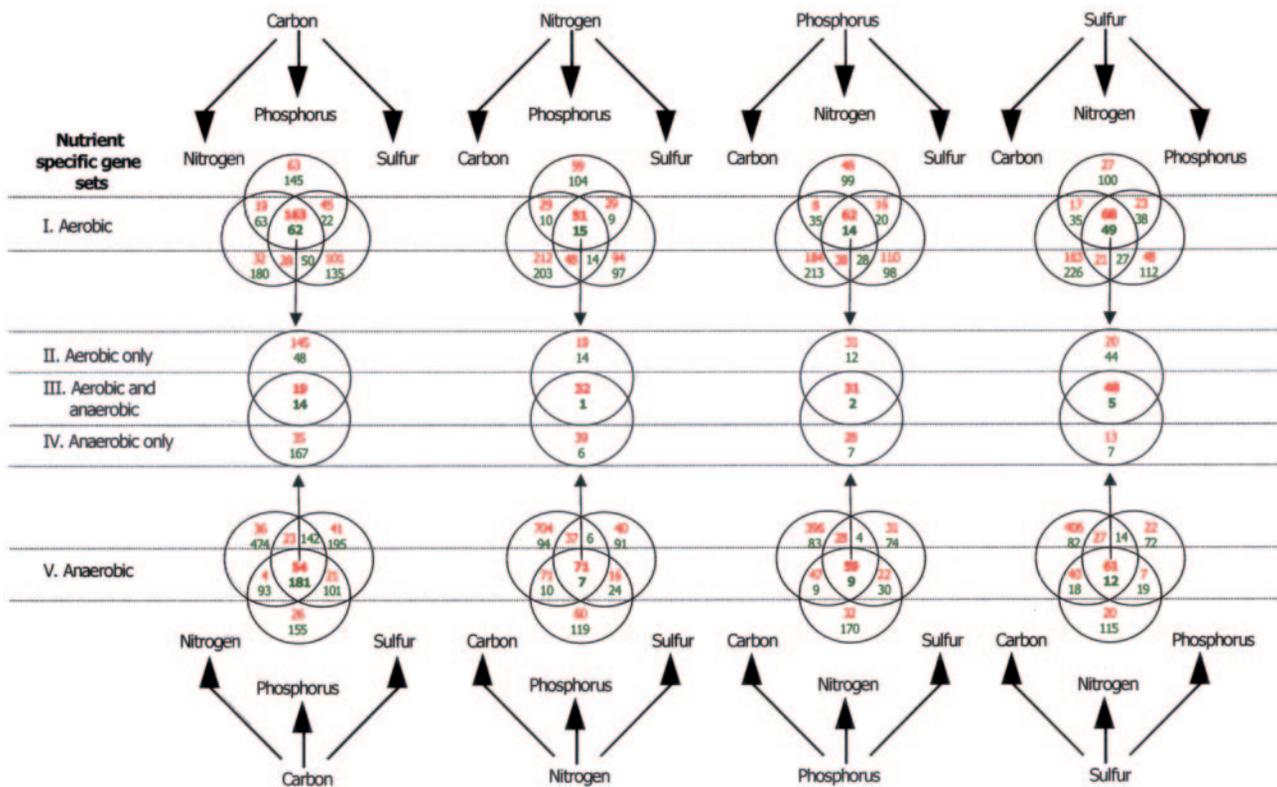


FIG. 3. Data mining strategy: dissection of the transcriptome response with respect to nutrient limitation and oxygen availability. A, Venn diagram of signature anaerobic genes. Red and green represent up-regulation and down-regulation, respectively, under anaerobic conditions. Each of the four circles corresponds to a cluster of genes that showed a transcriptional response to oxygen availability under one of the four macronutrient limitation regimes. The overlap of the four clusters represents genes that showed a consistent response to oxygen availability irrespective of the nutrient limitation regime. *Lim Ae*, limited aerobic; *Lim Anae*, limited anaerobic. B, Venn diagram of macronutrient limitation-responsive genes. The diagram shows pairwise transcriptome comparisons (see Fig. 1) of each macronutrient limitation regime against the other three macronutrient limitation regimes for aerobic and anaerobic cultures. Each circle represents the cluster of genes up-regulated (red) or down-regulated (green) for the reference macronutrient limitation. Sets I and V contain the genes that showed a consistent response to each of the four macronutrient limitation regimes in the three pairwise comparisons under aerobic and anaerobic conditions, respectively. Combination of sets I and V yielded three new subsets of macronutrient limitation-responsive genes. Set III represents signature genes that showed a consistent response to each of the macronutrient limitation regimes under aerobic and anaerobic conditions, respectively. Sets II and IV represent genes whose transcriptional response to a single macronutrient limitation regime was specific for aerobic and anaerobic conditions, respectively.

aerobic chemostat cultures were supplied with ergosterol and oleate, 22 of the consistently anaerobically up-regulated genes have been implicated in or associated with sterol or lipid metabolism. Of these genes, *UPC2* and *SUT1* are transcription factors for sterol uptake in yeast, and *PDR11* and *AUS1* (members of the ABC membrane transporter family) have been

shown to be involved in sterol uptake for anaerobic growth (30, 31). 13 members of the seripauperin family of possible cell wall mannoproteins (*DAN1*, *DAN2*, *DAN3*, *DAN4*, *TIR1*, *TIR2*, *TIR3*, *TIR4*, *PAU1*, *PAU3*, *PAU4*, *PAU5*, and *PAU6*) that were consistently up-regulated in anaerobic cultures encode mannoproteins. These important determinants of cell wall permeabil-



FIG. 4. Signature genes that respond to a single environmental parameter. The three independent transcriptome data sets for each condition were averaged and then compared. Green (relatively low expression) and red (relatively high expression) squares are used to represent the transcription profiles of genes deemed specifically changed. The signature genes were sorted by functional categories according to the Comprehensive Yeast Genome Database (24) and the *Saccharomyces* Genome Database (25).

ity during anaerobiosis (32) may be involved in sterol uptake, as recently shown for *DAN1* (30). The *MGA2* gene product regulates the transcription of *OLE1*, which is involved in the biosynthesis of unsaturated fatty acids (33). *HES1*, *ARE1*, *YSR3*, and *PLB2* encode a putative oxysterol-binding protein, an acyl-CoA acetyltransferase, a putative regulator of sphingo-

lipid metabolism, and phospholipase B₂, respectively (34–38). In addition to genes involved in sterol and fatty acid metabolism, *COX5B* and *HEM13* displayed a consistent up-regulation in all anaerobic cultures. *COX5B* encodes the “anoxic subunit” of cytochrome *c* oxidase, which is proposed to be involved in oxygen sensing (39). *HEM13* encodes a cytosolic coproporphyr-

TABLE II
Gene coverage of over-represented sequences retrieved from promoters of co-regulated genes

Unless stated otherwise, elements were counted present in a gene promoter only if they occurred at least twice. NS, no significant patterns retrieved by RSA Tools.

Regulatory cluster	Promoter element ^a		Putative-binding protein	Gene coverage			Genome coverage ^b			Range ^c		
	Forward	Reverse										
Specifically higher in anaerobiosis	AAGGCAC ^d	GTGCCTT		%			%					
				38			8					0–3
	ATTGTTC ^d	GAACAAT	Rox1p	26			12					0–3
	ddACGAGG ^d	CCTCGT ^{hy}	Upc2p	40			18					0–2
	TCGT ^{wy} AG ^d	CT ^{rw} ACGA	Upc2p	38			7					0–3
				Set II ^e	Set III ^f	Set IV ^g		Set II	Set III	Set IV		
Specifically higher in C limitation	dCCCCdh	dhGGGGh	Mig1p	43	65	25	28	0–5	0–6	0–5		
Specifically higher in N limitation	rGATAAs	sTTATCy	Gln3p/Gat1p/Dal80p/Gzf3p	11	61	14	6	0–3	0–5	0–3		
Specifically higher in P limitation	CAATGA	TCATTG	Dal82p	11	23	3	13	0–4	0–3	0–2		
	mACGTGs	sCACGTk	Pho4p	13	58	13	3	0–2	0–6	0–6		
Specifically higher in S limitation	GCCACA	TGTGGC	Cbf1p/Met4p/Met28p	5	33	NS	3	0–4	0–5	0–1		
	CACGTGA	TCACGTG	Met31p/Met32p	NS	10	NS	2	0–1	0–3	0–1		

^a Redundant nucleotides are as follows: *r* = A or G; *y* = C or T; *s* = G or C; *k* = G or T; *m* = A or C; *w* = A or T; *d* = A, G, or T; and *h* = A, C, or T.

^b Relative to 6451 open reading frame upstream promoters in the yeast genome according to RSA Tools.

^c Range of motifs present in each promoter of the specific gene cluster.

^d Elements counted present in a gene promoter when occurring at least once.

^e Set I, containing aerobic-only nutrient-specific genes, as in Fig. 3B.

^f Set II, containing aerobic and anaerobic nutrient-specific genes, as in Fig. 3B.

^g Set III, containing anaerobic-only nutrient-specific genes, as in Fig. 3B.

nitrogen III oxidase and has been described as the first, molecular oxygen-dependent and rate-controlling step of heme biosynthesis (27).

As a further approach to assess the biological significance of the consistent transcriptional responses identified via the two-dimensional approach, we analyzed the enrichment of regulatory motifs in promoter sequences of the oxygen-responsive genes (Figs. 3A and 4 and Table II). Four over-represented sequences were recovered from the 65 anaerobically up-regulated gene promoter regions (Table II). At least one of the two overlapping sequences (TCGT^{wy}AG or CCTCGT^{wy}) was recovered from 34 genes (52%) in the cluster. These sequences are similar to the previously described binding site for Upc2p (CGTTT) (40), a transcription factor whose structural gene itself was consistently up-regulated in the anaerobic cultures. 17 genes (26%) share the element ATTGTTC, which is the known binding site for the anaerobic transcription factor Rox1p (41). We also identified a new motif (AAGGCAC) within this cluster of genes for which no DNA-binding protein has yet been identified. The Upc2p and AAGGCAC motifs showed a remarkable coincidence in the promoters of 12 genes of the cluster (Fig. 5). In the upstream regions of these genes, the Upc2p-binding site is present at –450 to –380, and the AAGGCAC element is present at –360 to –300 (Fig. 5). The conservation of both the distance to the coding region and the distance between the elements strongly suggests biological relevance. 70% of the promoter sequences of the genes that were consistently up-regulated in the anaerobic cultures contain at least one of the three elements discussed above.

Transcriptional Responses to Macronutrient Limitation: Genes Up-regulated upon Phosphate Limitation—The four clusters of genes that were consistently (under aerobic as well as anaerobic conditions) up-regulated in response to growth limitation by a single macronutrient share some conserved features. These involve induction of high affinity uptake systems for the limiting macronutrient, excretion of nutrient-scavenging enzymes to the extracellular medium, induction of sys-

tems for mobilization and utilization of intracellular reserves, and induction of systems for transport and assimilation of alternative sources of the limiting nutrient (Fig. 4). This is exemplified by the transcriptional response to phosphorus limitation.

Previous comparison identified 62 up-regulated signature transcripts for aerobic phosphate-limited growth (4). Introducing a second dimension (anaerobic phosphate limitation) resulted in a 50% decrease in the genes composing this cluster. Indeed, 31 genes showed a consistent up-regulation relative to the other macronutrient limitation regimes in aerobic and anaerobic phosphate-limited chemostat cultures. Among these genes, seven are involved in transport, 14 in metabolism, one in protein fate, and one in transcription, and eight have an as yet unknown function according to the MIPS Database (24) and the *Saccharomyces* Genome Database (25). 23 of these phosphate limitation-induced genes (74%) could be directly related to phosphorus metabolism. All seven genes classified in the transport category were associated with phosphate transport (*PHO84*, high affinity inorganic phosphate/proton symporter; *PHO89*, high affinity sodium-dependent phosphate transporter (42); *PHO86*, protein associated with the phosphate transport complex (43); *GITI1*, glycerophosphoinositol transporter belonging to the major facilitator superfamily (44); and *VTC1*, *VTC3*, and *VTC4*, subunits of the vacuolar membrane polyphosphate transporter complex (45)). Of the remaining genes in this cluster, several are involved in phosphate mobilization: *PHO11* and *PHO3* encode phosphatases; *HOR2* encodes a glycerol-3-phosphate phosphatase (46, 47); *INM1* encodes an inositol monophosphatase (48); YNL217W encodes a putative metallophosphatase (49); YPL110C encodes a putative glycerophosphoryl-diester phosphodiesterase; *DDP1* encodes a diadenosine-hexaphosphate hydrolase (50); *PLB3* encodes phospholipase B (38); and *PYK2* encodes a glucose-repressed pyruvate kinase (51). The proteins encoded by *PHM6* and *PHM8* are likely to encode proteins involved in phosphate metabolism (45) as well, and their promoter regions exhibit a Pho4p-binding

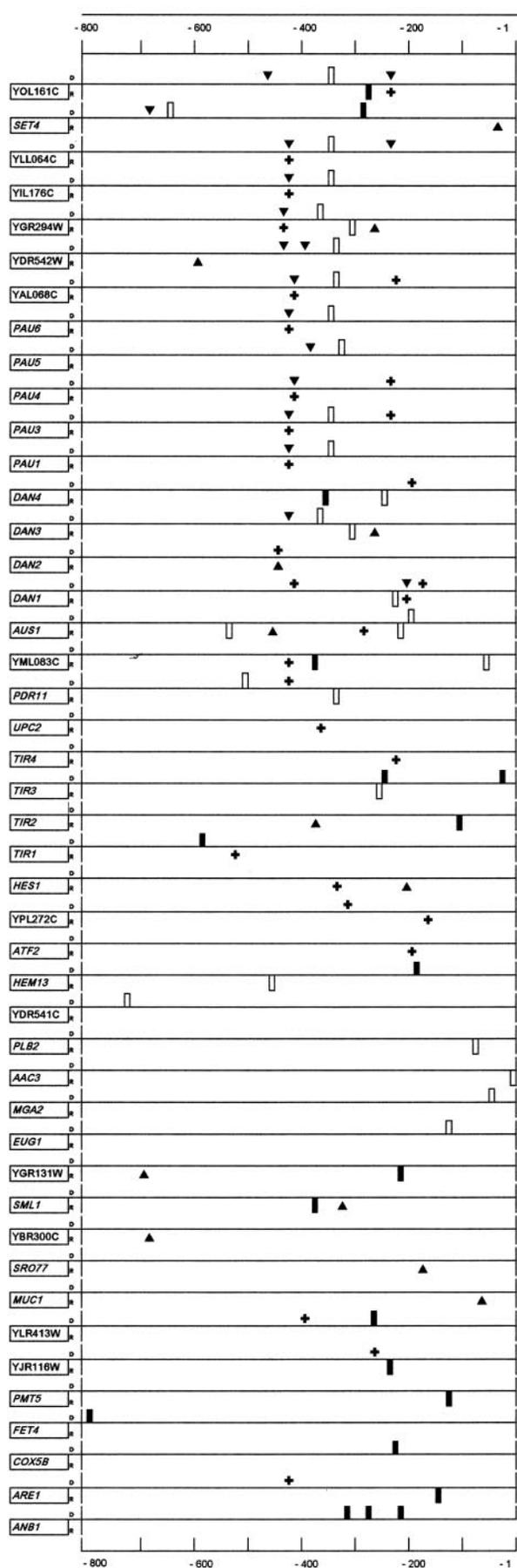


FIG. 5. Localization of the consensus binding site in promoter sequences of up-regulated genes in the absence of oxygen irrespective of nutrient limitation. The promoter regions of genes from

site. *PHO81* and *SPL2* are presumed inhibitors of the Pho80p-Pho85p cyclin-dependent protein-kinase complex and positive regulators of phosphate-related genes (52). Furthermore, *KCS1*, an inositol-1,2,3,4,5,6-hexaphosphate kinase involved in inositol metabolism (53), was up-regulated. The remaining eight genes in the cluster (25%) could not be directly associated with phosphate metabolism. Interestingly, two of these genes are involved in transcriptional regulation: *ZAP1* encodes a zinc-responsive transcriptional activator (54), and *MAF1* encodes a putative repressor of RNA polymerase III transcription and a common component of multiple signaling pathways in *S. cerevisiae* that sense changes in the cellular environment (55).

In silico promoter analysis of the genes that were consistently up-regulated upon phosphate limitation revealed an over-represented *mACGTGs* motif (present in 58% of the genes in the cluster as opposed to 3% in the *S. cerevisiae* genome). This sequence shows strong similarity to the CACGTG consensus sequence for the binding site of Pho4p (56), the main transcription factor required for expression of the phosphate-related genes (Table II).

Transcriptional Cross-regulation Identified by Two-dimensional Transcriptome Analysis—By combining the transcriptional responses to (an)aerobiosis in cultures subjected to four different macronutrient limitation regimes, it was possible to identify gene clusters that were subjected to transcriptional regulation by two environmental parameters. Identification of such clusters is not possible in conventional one-dimensional pairwise comparisons between cultivation conditions. Eight such clusters (sets II and IV) (Fig. 3B) could be assigned. To explore the biological significance of defining these clusters, we will discuss one of these clusters in more detail.

Of the 428 genes that showed a transcriptional response to carbon limitation in our analysis (sets II–IV) (Fig. 3B), only 33 genes showed a consistent response to carbon limitation irrespective of the availability of oxygen (set III) (Fig. 3B). 193 genes (set II) (Fig. 3B) showed a significant transcriptional response only under aerobic conditions. Of the remaining 202 genes (set IV) (Fig. 3B), which responded only to carbon limitation in the anaerobic cultures, 167 genes were down-regulated in anaerobic carbon-limited cultures, and 35 genes were up-regulated.

Of the 35 genes that were uniquely up-regulated in anaerobic carbon-limited chemostat cultures at the level of transcription, 21 genes are related to mitochondrial function (Fig. 6, upper panel), even though glucose dissimilation in these cultures was completely fermentative. 15 of these mitochondrial function-related genes are involved in oxidative phosphorylation and respiration: *QCR2*, *QCR6*, *QCR7*, and *RIP1* as core subunits of ubiquinol-cytochrome *c* reductase (complex III); *COX4*, *COX5A*, *COX6*, *COX8*, *COX12*, and *COX13* as core subunits of cytochrome *c* oxidase (complex IV); *ATP4*, *ATP15*, and *ATP20* as core subunits of the F_0 subunit of the mitochondrial ATP synthase; *INH1* as the inhibitory subunit of the mitochondrial ATP synthase; and finally, *CYC1* as the predominant aerobic isoform of cytochrome *c*. In addition, three of the four subunits of succinate dehydrogenase (*SDH1*, *SDH2*, and *SDH4*) were significantly up-regulated in the anaerobic carbon-limited cul-

–800 to –1 were based on the sequences obtained from RSA Tools (23). ▲, Upc2p consensus sequence TCGTwyAG; ▼, Upc2p consensus sequence TCGTwyAG found on the Crick strand; +, Upc2p consensus sequence CCTCGThh; ■, Rox1p consensus sequence (ATTGTTC); □, AAGGCAC consensus sequence. Consensus sequences on both strands are indicated. The open reading frames on the Watson strand and on the Crick strand are indicated by *D* and *R*, respectively. ^a Redundant nucleotides are as follows: *r* = A or G; *y* = C or T; *s* = G or C; *k* = G or T; *m* = A or C; *w* = A or T; *d* = A, G, or T; and *h* = A, C, or T.

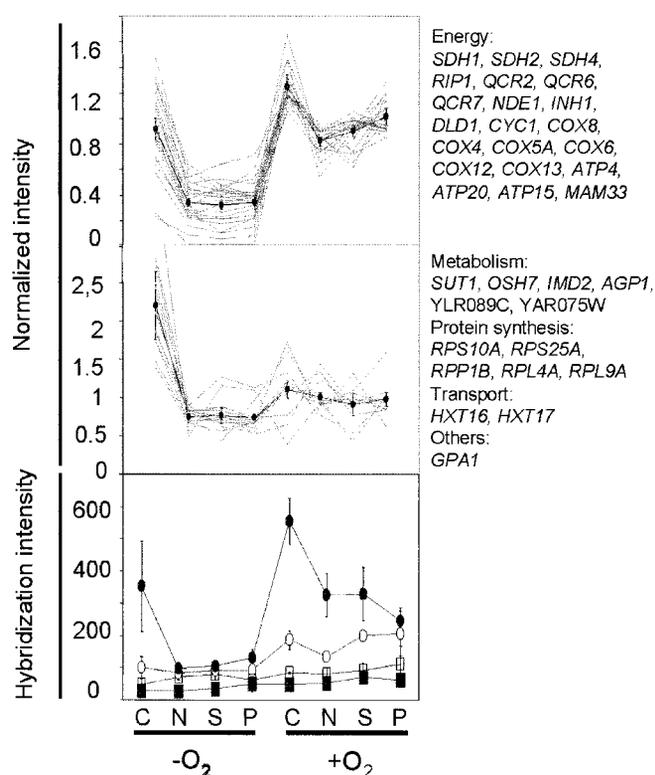


FIG. 6. Hierarchical regulatory control of gene expression. *Upper panel*, normalized transcript profiles of mitochondrial function-related genes that clustered with a set of genes that showed a specific transcriptional up-regulation upon carbon limitation in anaerobic cultures. The transcript level across the four conditions for each gene was normalized relative to mean and variance. The data represent the normalized abundance of all transcripts for each cultivation condition, as well as the normalized average abundance of all transcripts of the cluster (thick black line). Error bars represent S.D. *Middle panel*, normalized transcript profiles of genes belonging to the cluster described for A that do not have a (known) relation to mitochondrial function. The data are presented as described for the upper panel. *Lower panel*, transcript profiles of the structural genes for the components of the Hap2/3/4/5p complex. Error bars represent S.D. ■, HAP2; ○, HAP3; ●, HAP4; □, HAP5.

tures. *DLD1* encodes mitochondrial D-lactate-ferricytochrome *c* oxidoreductase (57); *MAM33* encodes a mitochondrial protein required for normal respiratory growth (58); and *NDE1* encodes a mitochondrial, cytosolically directed NADH dehydrogenase (59). The remaining 14 genes of the 35 genes of the discussed cluster were composed of two hexose transporter genes (*HXT16* and *HXT17*), five genes encoding ribosomal proteins (*RPS10A*, *RPS25A*, *RPP1B*, *RPL4A*, and *RPL9A*), and seven genes belonging to different metabolic routes (*SUT1*, *OSH7*, *AGP1*, *IMD2*, *YLR089C*, *YAR075W*, and *GPA1*) (Fig. 6, middle panel).

The low mRNA levels of these genes in anaerobic chemostat cultures that were limited by nitrogen, phosphorus, or sulfur and thus had a high residual glucose concentration strongly suggest that glucose catabolite plays an important role in their transcriptional regulation. Conversely, under aerobic conditions, the identity of the growth-limiting nutrient did not significantly affect transcription. In fact, closer inspection indicated that, in the aerobic cultures, high transcript levels were observed in all four macronutrient limitation regimes (Fig. 6, upper panel). Furthermore, also in the aerobic cultures, the combined expression patterns of these genes suggest a moderate induction under glucose-limited conditions (Fig. 6, upper panel). However, the statistical criteria used for the definition of the cluster did not identify this induction as significant.

A simple verbal model to explain these observations is that, for this particular subset of *S. cerevisiae* genes, induction by oxygen supersedes glucose catabolite repression. It is beyond the scope of this work to analyze the molecular basis for this apparent hierarchy in transcriptional regulation. However, several genes of this cluster such as *DLD1*, *QCR2*, *QCR7*, and *CYC1* are known targets of the Hap2/3/4/5p complex (Fig. 6, upper panel) (57, 60–62). *In silico* promoter analysis of the 35 genes of this subset revealed a significant over-representation (3-fold) of the ACCAATnA sequence, which overlaps the CCAAT core of the Hap2/3/4/5p-binding site. Furthermore, the transcript level of *HAP4*, known as the regulatory subunit of the Hap2/3/4/5p complex, correlated with the expression pattern within this subset of genes (Fig. 6, lower panel). Interestingly, *HAP4* expression is reported to be glucose-repressible, being up-regulated after the diauxic shift and during growth on respiratory carbon sources (63). Further research is required to investigate which factors, in addition to regulation by the Hap2/3/4/5p complex, are involved in oxygen regulation of these genes and which factors determine the relative impact of glucose repression and oxygen induction.

DISCUSSION

DNA Microarrays as a Diagnostic Tool for Biotechnology—A detailed understanding of the environmental stimuli to which microorganisms are exposed in industrial fermentation processes is invaluable for rational design and optimization of such processes. DNA microarrays provide an interface that allows, in principle, the use of the microorganisms themselves as the ultimate “biosensor.” An unequivocal coupling between relevant environmental parameters and transcriptional responses is essential for this application of DNA microarrays. An important concept in this context is that of the signature transcript, a transcript whose levels specifically increase (or decrease) in response to a single environmental stimulus.

This study indicates that, in general, robust signature transcripts cannot be identified by varying the process parameter of interest against a single constant experimental background (one-dimensional transcriptome analysis). Instead, identification of robust signature transcripts requires that transcriptional responses to an environmental parameter be analyzed against multiple experimental backgrounds. For example, the sets of signature transcripts for (an)aerobiosis and growth limitation by four macronutrients that were previously established in one-dimensional transcriptome comparisons (4, 9, 11, 12) were considerably reduced in size by the two-dimensional approach followed in this study. Chemostat cultivation is an indispensable tool for this combinatorial approach, as, in contrast to batch cultivation, it allows the manipulation of individual culture parameters while other relevant parameters, including the specific growth rate, are kept constant (4, 9, 11, 13).

Although this study covers only a minute fraction of the staggering diversity of environmental conditions to which *S. cerevisiae* may be exposed in nature and in industry, it clearly demonstrates the complexity of transcriptional regulation. In real life, transcriptional responses of cells are influenced by hundreds of extracellular signals. The interplay of these signals results in a multidimensional space in which each possible combination of signals results in a unique transcriptome. It is therefore to be anticipated that the number of robust signature transcripts will decrease further when, in addition to nutrient limitation and oxygen availability, other chemical or physical process parameters are included.

The significance of the combinatorial nature of the regulation of gene expression extends beyond *S. cerevisiae* and industrial biotechnology. For example, in the medical field, it is to be ex-

pected that the transcriptional profiles coupled to a disease or pharmacological efficacy will be equally sensitive to other stimuli and variance received by the cells. Although, in a statistical sense, such effects may be averaged out when the identification of disease-correlated signature transcripts is based on large numbers of healthy and ill individuals (64), this does not exclude a strong impact of transcriptional “cross-talk” in individual patients that have been exposed to special circumstances.

Unraveling Transcriptional Regulation—Despite the combinatorial nature of transcriptional regulation, identification of unequivocal signature transcripts should be possible when mechanisms of transcriptional regulation are fully understood. Ideally, signature transcripts should be encoded by genes that respond to a single transcriptional regulator protein whose expression and activity are uniquely dependent on a single environmental stimulus. Identification of such genes and regulators requires detailed knowledge of the regulons and recognition sequences of all relevant transcriptional regulators. Such knowledge is also essential for rational and predictable reprogramming of transcriptional regulation to improve the performance of industrial microorganisms.

Even for well studied organisms like *S. cerevisiae*, the physiological roles of many transcriptional regulators, as well as the sequence motifs they recognize, remain to be identified. The two-dimensional chemostat-based approach proposed in this study provides a powerful new tool for unraveling transcriptional regulation networks. This is exemplified by the enrichment of regulatory motifs in the consistently anaerobically induced transcripts (Table II). Clearly, regulation by known transcriptional regulators (relief of *ROX1* repression and transcriptional activation by *UPC2*) (30, 65, 66) is not sufficient to account for the transcriptional response of all 65 genes that were consistently up-regulated under anaerobic conditions. Indeed, our study strongly suggests that at least a third factor, which recognizes an AAGGCAC motif, is involved in transcriptional regulation by oxygen availability. This motif had gone unnoticed in a previous one-dimensional aerobic/anaerobic comparison (12). In general, a combinatorial analysis of the transcriptional responses to environmental stimuli is likely to increase enrichment of relevant regulatory elements and facilitate their identification.

The approach used in this study also allows statements on the hierarchy of transcriptional regulation. This is exemplified by a subset of genes related to mitochondrial function. Under anaerobic conditions, these genes were regulated primarily by glucose repression/derepression. However, under aerobic conditions, a high transcript level was observed even under excess glucose conditions. Together, these data indicate that, in the aerobic cultures, oxygen regulation supersedes glucose repression (Fig. 6). By expanding data sets and combining them with an *in silico* analysis of promoter structure, combinatorial analysis of transcriptomes can accelerate the unraveling of transcriptional regulation networks.

Functional Analysis—Assigning physiological functions to “unknown function” genes still poses a major challenge in the post-genomic era. By identifying groups of genes that appear to be coexpressed (67), DNA microarrays can guide functional analysis. Indeed, many studies have correlated mRNA levels to cultivation conditions. However, even when chemostat cultivation is used to change only a single environmental parameter, pairwise comparisons characteristically lead to large numbers of target genes, complicating functional analysis (4, 9, 10, 12). Moreover, in a recent study on the genome-wide transcriptional responses to low temperature (68), a very poor correlation was observed between transcriptional responses of genes and the phenotype of the corresponding null mutants at low temperature.

Compared with previous one-dimensional studies, the combinatorial approach followed in this study led to a clear enrichment in our “robust response sets” of (i) genes with known function related to the environmental status under study (Fig. 4) and/or (ii) genes with relevant regulatory elements (Table II). By implication, also the unknown function genes found in the corresponding data sets are more likely to have a direct functional relationship to the corresponding nutritional/environmental status. We are currently testing this hypothesis for the subset of genes that showed a consistent up-regulation under anaerobic conditions.

Among the robust response signature genes identified in this study, 38% do not have a clearly established biological function (Fig. 4). It is noteworthy that some of these (YJL118C, YAR069C, and YGR190C) belong to a group of open reading frames for which it has recently been proposed that they should be discarded from the yeast genome directory based on genomic comparison of *S. cerevisiae*, *Saccharomyces bayanus*, *Saccharomyces mikatae*, and *Saccharomyces paradoxus* (69). The observation that three of these genes showed a consistent response to phosphate limitation (YJL118C and YAR069C) or nitrogen limitation (YGR190C) strongly suggests they are *bona fide*, biologically relevant genes.

Provided that yeast strains and cultivation procedures are standardized, DNA microarray analysis of chemostat cultures is well reproducible in different laboratories (9). We propose that a multi-laboratory effort to build an extensive, chemostat-based, “multidimensional” gene expression data base is an invaluable research tool for functional analysis of the *S. cerevisiae* genome and for yeast system biology. Obviously, such a data base should not necessarily be confined to transcriptome data, but could also cover other levels of information.

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Supplementary data Table 1

Culture limiting nutrient	Oxygen availability	Average coefficient of variation	ACT1
Carbon	Aerobic	0.18	2489 ± 81
Nitrogen	Aerobic	0.14	2265 ± 106
Phosphorus	Aerobic	0.21	2314 ± 266
Sulfur	Aerobic	0.13	2172 ± 249
Carbon	Anaerobic	0.27	3118 ± 319
Nitrogen	Anaerobic	0.19	2287 ± 363
Phosphorus	Anaerobic	0.17	2517 ± 101
Sulfur	Anaerobic	0.15	2346 ± 249

Up-regulated under anaerobiosis (FC = fold change, ANA = anaerobic, A = aerobic)			cellular function	FC-C-Lim ANA vs C-Lim A																FC-N-Lim ANA vs N-Lim A																FC-S-Lim ANA vs S-Lim A																FC-P-Lim ANA vs P-Lim A															
Protein Set	Gene name	systematic name		functional description	C-Lim ANA Average	C-Lim ANA S-Lim ANA	N-Lim ANA Average	N-Lim ANA S-Lim ANA	P-Lim ANA Average	P-Lim ANA S-Lim ANA	S-Lim ANA Average	S-Lim ANA P-Lim ANA	C-Lim A Average	C-Lim A S-Lim A	N-Lim A Average	N-Lim A S-Lim A	P-Lim A Average	P-Lim A S-Lim A	S-Lim A Average	S-Lim A P-Lim A	C-Lim A Average	C-Lim A S-Lim A	N-Lim A Average	N-Lim A S-Lim A	P-Lim A Average	P-Lim A S-Lim A	S-Lim A Average	S-Lim A P-Lim A	C-Lim A Average	C-Lim A S-Lim A	N-Lim A Average	N-Lim A S-Lim A	P-Lim A Average	P-Lim A S-Lim A	S-Lim A Average	S-Lim A P-Lim A																															
4069_at	MG42	YR033W	YR033W may be involved in the remodeling chromatin structure	103.3	56.4	102.3	27.1	7.2	8.7	2.9	24.5	4.7	2.3	3.7	4.3	3.1	2.3	3.7	4.3	3.1	2.3	3.7	4.3	3.1	2.3	3.7	4.3	3.1	2.3	3.7	4.3	3.1	2.3	3.7	4.3	3.1	2.3	3.7	4.3																												
7480_at	SR077	YBL100C	YBL100C yeast homolog of the Drosophila tumor suppressor, lethal giant vein	204.2	150.3	243.7	61.7	255.5	45.1	258.8	47.7	83.9	19.2	78	13.8	56.9	15.1	37.1	8.9	4.5	6.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9																											
11248_at	PAU1	YR041W	YR041W member of the serpinogen proteoglycan family	1432.5	497.2	1566.6	43.1	356.4	43.8	188	160.3	109.8	42.8	56	41.6	19.6	67.4	8.4	4.1	6.9	16.4	14.4	14.4	14.4	14.4	14.4	14.4	14.4	14.4	14.4	14.4	14.4	14.4	14.4	14.4	14.4	14.4	14.4	14.4	14.4																											
4114_at	TR43	YLI011W	YLI011W strong similarity to members of the Srp17p/Tlp1p family	387.6	117.9	543.7	592.6	3335.7	80.3	3306.2	390.3	252.3	93.3	88.8	6	84.2	13.7	174	17.3	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9																											
4563_at	PAU5	YHL046C	YHL046C similar to members of the Srp17p/Tlp1p family	1527.1	414.8	739.2	120.6	1128.9	73.3	2915.5	867	153.2	61.1	88.3	7.6	74.2	10	137.1	20.1	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6																											
5410_at	TR1	YR010W	YR010W member of the serpinogen proteoglycan family	690.2	206.1	445.5	74.2	510.8	39.5	801	339.9	92.2	30.4	30.7	13.4	27.8	8.3	45.7	20.9	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8																											
5711_at	PAU1	YR014W	YR014W member of the serpinogen proteoglycan family	4568.3	3906.3	3953.1	103.2	4155.5	210.5	4857.4	1028.3	1497	28.1	74.4	11.6	87.4	4.2	137	17.7	30.5	30.5	30.5	30.5	30.5	30.5	30.5	30.5	30.5	30.5	30.5	30.5	30.5	30.5	30.5	30.5	30.5	30.5	30.5	30.5	30.5																											
6791_at	PAU3	YR016W	YR016W member of the serpinogen proteoglycan family	361	204.7	216.1	12.2	462.3	42.3	247.7	139.3	116.5	24.3	6.5	15.6	4.5	28.4	6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6																												
6826_at	TR4	YR009W	YR009W similar to Tlp2 and Tlp2p	3446.6	1088.8	2031.6	474	2781.5	14	2998.7	421.0	100.8	44.4	48.7	7.1	42.5	6.1	93.4	29.3	34.2	34.2	34.2	34.2	34.2	34.2	34.2	34.2	34.2	34.2	34.2	34.2	34.2	34.2	34.2	34.2	34.2	34.2	34.2	34.2																												
8527_at	TR2	YR010C	YR010C Cold shock induced protein of the Srp17p/Tlp1p family of sense-amine rich proteins	1863.1	194.4	999.7	71.5	1602.3	137.5	778.1	58.2	159.4	23.5	129.8	14.9	147.6	20.5	207	113.8	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6	8.6																												
8721_at	PAU3	YR010W	YR010W member of the serpinogen proteoglycan family	460.2	206.1	445.5	74.2	510.8	39.5	801	339.9	92.2	30.4	30.7	13.4	27.8	8.3	45.7	20.9	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8																													
8786_at	PAU6	YR010W	YR010W member of the serpinogen proteoglycan family	554.4	175	435.7	65.4	483.5	3.1	1011.8	389.7	64.2	22	61.4	4.9	4.5	6.6	73.3	14.7	10.6	10.6	10.6	10.6	10.6	10.6	10.6	10.6	10.6	10.6	10.6	10.6	10.6	10.6	10.6	10.6	10.6	10.6	10.6	10.6																												
8825_at	PAU4	YR041W	YR041W member of the serpinogen proteoglycan family	1218.8	292	825	123.6	1620.1	165.2	2023.3	214.2	117	114.3	12.7	87.2	6.2	140.6	30.9	11.8	11.8	11.8	11.8	11.8	11.8	11.8	11.8	11.8	11.8	11.8	11.8	11.8	11.8	11.8	11.8	11.8	11.8	11.8	11.8																													
4876_at	TR2	YR013W	YR013W similar to Noc2p	214.6	88.9	310.6	58.9	316.2	24.8	508.3	124.8	28.4	5.8	35.3	3.2	18.4	4.4	22.9	8.2	7.6	7.6	7.6	7.6	7.6	7.6	7.6	7.6	7.6	7.6	7.6	7.6	7.6	7.6	7.6	7.6	7.6	7.6	7.6																													
4565_at	ECM34	YHL045W	YHL045W Extracellular Matrix	32.5	18.8	43.5	14.1	81.7	20.5	76.6	9.5	12	2.9	14.1	3.4	12	2.7	12.2	0.6	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7																													
4200_at	CO058	YLI111W	YLI111W Cytochrome c-oxidase chain Vb	149.8	32.6	150.7	15.3	117.9	10.8	169.3	19.9	15.7	0.6	64.5	6.5	39.1	8.1	57.8	18.8	9.5	9.5	9.5	9.5	9.5	9.5	9.5	9.5	9.5	9.5	9.5	9.5	9.5	9.5	9.5	9.5	9.5	9.5	9.5																													
5387_at	GSY1	YFR015C	YFR015C Cytochrome XDPase (UDP-glucose-4-epimerase)	358	60.1	236.9	29	88.5	13.2	192.7	42.9	92	13.3	68.4	29.9	25.5	3.4	53.1	6.6	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9																														
10525_at	YR53	YR0053C	YR0053C DHP-1 P-phosphatase	852.8	235.7	604.7	60.4	539.8	76.9	1048.4	98.2	43.5	39	56.3	7.5	58	5.8	79.3	21.4	10.7	10.7	10.7	10.7	10.7	10.7	10.7	10.7	10.7	10.7	10.7	10.7	10.7	10.7	10.7	10.7	10.7	10.7	10.7																													
10662_at	BN42	YR078W	YR078W similar to mammalian indoleamine 2,3-dioxygenase	371.3	62.8	194.4	28.8	293.9	38.8	322.6	31.6	36.3	156.2	42.5	8.3	2.8	78.4	9.8	54.3	17	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1																													
11234_at	MUC1	YAL028W	YAL028W strong similarity to E. coli galactosyl O-acetyltransferase	113.5	34.8	76.3	23	106.4	15.8	123.1	15.3	30.3	7.6	21.8	4.1	17.6	2	41.6	12.5	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6																													
4100_at	AT2	YR019C	YR019C Nucleic acid sulfate lyase with structure similar to sense-amine rich GPI-anchored LCCOLATION	290.4	209.2	119.4	109.6	128.9	82.8	2356.4	119	30.7	6.8	56.5	5.1	74.3	26.1	84.8	8.6	24.7	24.7	24.7	24.7	24.7	24.7	24.7	24.7	24.7	24.7	24.7	24.7	24.7	24.7	24.7	24.7	24.7	24.7	24.7	24.7																												
4832_at	ATF2	YGR177C	YGR177C Steroid O-acetyltransferase, acylates certain toxic steroids such as pregnenolone	384.6	16.4	467.8	50.9	415.5	22.4	369.2	105.4	79.8	43.5	34.1	4.5	3.1	1.8	46.1	23.5	4.8	4.8	4.8	4.8	4.8	4.8	4.8	4.8	4.8	4.8	4.8	4.8	4.8	4.8	4.8	4.8	4.8	4.8	4.8																													
5596_at	PAU4	YGL039W	YGL039W similar to V. vinifera dihydroflavonol reductase	211.1	47.4	291.9	33.7	396.7	65.9	414.9	12.2	103.7	19.1	5.9	7.3	22.8	11.1	78.4	11.9	3.2	3.2	3.2	3.2	3.2	3.2	3.2	3.2	3.2	3.2	3.2	3.2	3.2	3.2	3.2	3.2	3.2	3.2	3.2																													
5173_at	SUT1	YGL162W	YGL162W involved in sterol uptake	168.4	14.5	67.4	7.3	75.6	19.2	67.4	4.4	37.5	8.7	20.6	3.8	37.3	1.2	17.3	2.4	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6	2.6																													
5654_at	SPK3	YER081W	YER081W strong similarity to phosphoglycerate dehydrogenase	123.1	24.3	73.4	13.6	58.1	26.7	136.7	32.8	33.5	5.9	31.8	1.4	22.4	0.4	36.1	4	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7	2.7																														
5920_at	UCR2	YDR132W	YDR132W regulatory protein involved in control of sterol uptake	34.3	26.5	48.9	21.9	92.2	15.4	90.3	15.7	13.2	5.7	12	3.3	12	2	5.4	2.6	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2																														
5940_at	HER3	YDR518C	YDR518C strong similarity to glucosylase	896.9	178.9	528	42.1	369.1	4.1	603.4	64.3	305.5	43.6	220.7	43.1	118.7	1.3	182.8	2.4	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9																														
6270_at	UPC2	YDR132W	YDR132W regulatory protein involved in control of sterol uptake	34.3	26.5	48.9	21.9	92.2	15.4	90.3	15.7	13.2	5.7	12	3.3	12	2	5.4	2.6	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2	4.2																														
6411_at	HEM13	YDR044W	YDR044W Coproporphyrinogen III oxidase	387.8	385.8	699	23.1	703.4	165.5	918.4	176.4	5.4	10.3	49.7	6.8	45.9	12.1	35.7	1.9	5.5	5.5	5.5	5.5	5.5	5.5	5.5	5.5	5.5	5.5	5.5	5.5	5.5	5.5	5.5	5.5	5.5	5.5																														
6837_at	ARE1	YR040W	YR040W Acyl-CoA cholesterol acyltransferase (sterol ester synthetase)	385.8	17.2	412.2</																																																													

9724_at	HMG1	YML079C	YML079C 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase isozyme	STEROL METABOLISM	258.7	56.8	346.4	36.9	221.3	53.2	240.2	24.7	1301.6	135.2	850.1	55.2	995.8	75.1	901	247.2	-5	-2.5	-4.5	-3.8	
10148_at		0 YLR168C	YLR168C probably involved in intramitochondrial protein sorting	UNKNOWN	49.1	24.4	47.7	7.1	33.3	4.9	38.6	8.3	260.6	71.9	279.4	37.6	316.4	2.9	252.3	37.7	-5.3	-5.9	-6.5	-5.1	
10290_at		CDX12	YLR028C	YLR028C subunit Vta of cytochrome c oxidase	OXIDATIVE PHOSPHORYLATION	1168.6	252.6	386.2	105	264.7	44.8	436	94	2507.4	46.2	1268.2	205	2062.8	59.2	1966.4	168.5	-2.1	-5	-5.7	-4.5
4759_at		PEQ21	YGR029C	YGR029C weak similarity to hypothetical protein YHR160c	PEROXISOME BIOGENESIS (PUTATIVE)	34.4	10.4	43.8	8.2	42	3.5	32.4	11.5	193.4	7.3	129.7	20.7	226.4	25.1	105.7	32.4	-2.9	-2.9	-3.3	-3.3
7543_at		CCQ2	YPR191W	YPR191W 43 kDa diaphenol cytochrome c reductase core protein 2	OXIDATIVE PHOSPHORYLATION	321.6	78.6	113.4	10.7	106.9	16.1	137.6	3.8	808	99.5	571.6	97.7	709.4	31.9	986.2	22.7	-5	-6	-6.6	-7.2
10591_at		ATV7	YKL016C	YKL016C ATP synthase subunit	ATP SYNTHESIS	409.5	104.1	226.3	22.1	162.7	23.4	134.9	6.8	835.5	145.4	562	32.3	555.2	49.3	497.1	47.7	-2	-2	-2.5	-3.4
4768_at		SOL4	YGR248W	YGR248W similar to SOL3	UNKNOWN	47.9	27.2	87.1	8	48.3	18.5	61.2	18.6	568.3	184.4	363.7	153	110.6	4.9	528.4	7.5	-11.9	-4.2	-2.3	-8.6
9008_at		PRP12	YMR022C	YMR022C integral membrane mitochondrial protein	RRNA PROCESSING	40.5	12.1	66.5	13.4	55.9	4.1	42.4	3.2	150.2	19.6	178.5	6.7	196.2	21.3	145.8	28.9	-3.7	-2.7	-3.5	-3.4
9555_at		MOT3	YMR070W	YMR070W Cys2-His2 zinc fingers at C-terminus; glutamine and asparagine rich	MATING	12	3.8	12	3	12	2.2	12	5.4	25.9	4.6	25.3	7	29.8	13.1	29.5	1.6	-2.1	-2.1	-2.1	-2.1
10377_at		AQY2	YLL053C	YLL053C similar to water channel proteins	UNKNOWN	75	23.3	157.4	12.9	172.9	33.2	171.5	47.5	1248.4	136.8	348.8	56.7	2716.5	272.8	2129.6	758.2	-16.6	-15.7	-16.6	-12.4
10378_at		AQY2	YLL052C	YLL052C member of msp family transmembrane channels	TRANSPORT WATER	23.9	6.8	56.1	7.2	77.3	10	86	18.2	418.7	78.5	130.1	5.7	1355	120.9	1078.8	594.3	-17.5	-17.5	-22.3	-12.5
6190_at		CCQ2	YDR270W	YDR270W Copper-transporting P-type ATPase with similarity to human Menkes and Wilson OXIDATIVE PHOSPHORYLATION	OXIDATIVE PHOSPHORYLATION	29.4	8.6	32	2.5	39.8	8.6	41.7	8.5	135.7	10	100	13.4	165.6	40.5	137.3	12.3	-4.6	-4.2	-3.3	-3.3
7831_at		PRP12	YPL088C	YPL088C multidrug resistance transporter	DRUG RESISTANCE	70.4	31.6	114.3	19.8	81.7	6.9	65.9	22.6	637.9	155.7	523.1	92.3	217.5	15.3	467.6	55.3	-9.1	-4.6	-3.5	-7.1
8132_at		PUJ14	YOR044C	YOR044C putative proline-specific permease	TRANSPORT	16.9	6.6	176	26.5	28.1	6.5	21.7	8.9	1202.8	181	2267.3	294	138.5	44.6	516.2	179.5	-14.5	-14.5	-14.5	-23.9
8669_at		MCH4	YOL119C	YOL119C similar to monocarboxylate transporter proteins	UNKNOWN	27.9	7.5	56.8	20.5	42.6	3.4	50	4.4	73.3	8.3	272.8	50.5	243.1	37.5	111.9	48.3	-2.6	-4.8	-5.7	-2.2
9596_at		AAC1	YMR055C	YMR055C mitochondrial ADP/ATP translocase	TRANSPORT	60.1	1.7	117.5	14.6	102.6	22	72.4	10	528.7	75.7	462.7	67	352.6	26	439.8	233.8	-8.8	-4.1	-6.1	-6.1
10027_at		0 YLR121C	YLR121C hypothetical protein	UNKNOWN	27.3	21.2	81.1	32.5	37.4	11.8	32	6.7	289.2	10.8	173.3	27.2	124.6	6.1	66.5	15.4	-10.8	-2.1	-10.8	-10.8	
10227_at		0 YLR108C	YLR108C strong similarity to YDR132c	UNKNOWN	12	2.9	23.8	3.9	21.5	2.8	23.1	0.7	38	10.6	107.9	12	166	17.4	112.3	3.8	-3.2	-4.2	-4.9	-7.7	
11095_at		0 YAL046C	YAL046C similarity to hypothetical protein YBR273c	UNKNOWN	103.1	23.2	137.5	14.8	115.6	2.2	143.8	3.2	544.7	56.2	459.6	67.8	473	42	397.5	39.8	-5.3	-3.3	-4.1	-2.8	
11102_at		LSB8	YAL100W	YAL100W similar to hypothetical C. elegans protein C56A3.8	UNKNOWN	19.1	8.7	19.8	1.3	32.2	2.8	15	2.3	279.3	127.1	170	51.4	265.6	14.8	192.8	8.7	-14.6	-8.6	-14.6	-14.6
11135_at		MDV1	YAL112W	YAL112W similar to Mel5p and N. crassa sulfur control-2	UNKNOWN	28.5	17.6	75.2	14.3	58.8	4.9	41.2	3.7	154.1	42.7	210.1	19.7	194.3	34.1	148.7	13.2	-2.7	-3.3	-3.6	-3.6
11359_at		0 YAL046C	YAL046C weak similarity to Legionella small basic protein sbpA	UNKNOWN	124.6	21.4	302.3	10.2	169.2	38.1	15.2	609.9	45.1	731.6	57.4	610.5	81.3	642.9	242.4	-5	-2.4	-3.6	-2.9	-5.6	
4131_at		0 YLD04W	YLD04W weak similarity to T. brucei NADH dehydrogenase	UNKNOWN	51.1	23.9	58.7	3.8	54.8	1.9	52.8	19.2	123.7	10.3	125.9	20	126.5	14.1	125.1	36.7	-2.4	-2.4	-2.4	-2.4	
4462_at		0 YHR090C	YHR090C similarity to hypothetical protein YOR056c, YLR042c and YLR072w	UNKNOWN	23.4	13.8	39.7	5.3	31.9	4.1	25.1	4.2	86.4	21.4	109.8	14.6	75.7	3.9	95	25.5	-4.1	-2.8	-2.4	-3.8	
4742_at		0 YGR266W	YGR266W hypothetical protein	UNKNOWN	20.8	5.4	24.2	4.9	21.7	5	21.4	2.6	67.2	9.6	76.5	7.3	125.7	21.1	57.1	11.5	-4.7	-3.2	-5.8	-2.7	
4763_at		0 YGR248W	YGR248W strong similarity to hypothetical protein YHR162w	UNKNOWN	384.2	140.3	68	1.2	59.6	5.8	77	21.3	179.2	97.7	155.6	9.6	178.2	41.4	203.3	45.6	-4.6	-2.3	-2.6	-3	
5050_at		0 YGL057C	YGL057C hypothetical protein	UNKNOWN	26.5	8.2	33.3	10.5	37	6.5	31.1	7.2	70.4	8.8	70.2	5	81.8	7.6	71.2	21.2	-2.7	-2.7	-2.7	-2.7	
5096_at		0 YGL101W	YGL101W strong similarity to hypothetical protein YBR242w	UNKNOWN	12.7	6.4	25.7	7.1	21	5.3	24.2	3.5	77.8	7.6	122.9	23.1	127.6	6.2	85.7	8.1	-6.1	-4.8	-6.1	-6.1	
5183_at		0 YGL196W	YGL196W hypothetical protein	UNKNOWN	24.3	10.6	90.4	25.3	81.6	3.1	91	6.6	148.6	29.2	284.4	37	123.9	7.8	113.6	20.4	-3.1	-2.4	-2.2	-2.2	
6555_at		0 YDL086W	YDL086W similarity to hypothetical Synchocystis protein	UNKNOWN	461.6	183	327.9	7.5	333.5	95.2	265.5	22.9	1051.6	137.6	903	156	963.8	74.1	1006.1	221.7	-2.3	-2.8	-2.9	-3.5	
6576_at		0 YDL115C	YDL115C hypothetical protein	UNKNOWN	180.7	14.9	150.4	14.4	112.6	38.7	140.1	10.5	952.2	90.4	953.7	43.2	207.1	38.3	441.3	86.6	-5.1	-3.9	-2.9	-3.2	
6846_at		0 YCR061W	YCR061W hypothetical protein	UNKNOWN	12	1.5	12	2.3	12	2	22.7	4.6	61.4	13.3	49	10	50.2	5.1	129.5	90.4	-5.1	-4.1	-4.2	-4.2	
7140_at		0 YBR230C	YBR230C hypothetical protein	UNKNOWN	136.7	24.5	76.1	17.8	53.4	1.2	50	3.1	1635.1	197.6	364.2	56.9	262.7	25.7	353.5	38.4	-1.2	-4.8	-4.9	-7.1	
7315_at		0 YBR047W	YBR047W hypothetical protein	UNKNOWN	12	4.1	14.7	3.6	19.8	1.4	15.5	4.1	51.4	54.3	41.7	2.4	68.5	8.7	39.4	2.9	-10.1	-2.8	-3.5	-2.5	
7445_at		0 YBL095W	YBL095W similarity to C. albicans hypothetical protein	UNKNOWN	29	14.7	62	6.7	52.8	8.6	46.2	4.7	141.8	57.5	457.7	77.1	381.6	17	353.3	70.4	-4.9	-7.4	-7.2	-7.7	
7594_at		0 YPR151C	YPR151C weak similarity to YPL150c	UNKNOWN	12	2.7	12	0.7	12	1.6	12	2.1	721.4	197.9	385.8	102.5	334.9	25	322.3	22.5	-60.1	-27.9	-26.9	-26.9	
7681_at		0 YPR061C	YPR061C weak similarity to Synchococcus sp. DnaJ protein	UNKNOWN	12	4.1	12	0.9	12	0.6	12	3.7	48.2	2.9	44.7	5.8	44.3	1.5	35.4	8.2	-4	-3.7	-3.4	-3	
7749_at		0 YPL044C	YPL044C strong similarity to YGR086c	UNKNOWN	403.2	122.3	454.1	69.3	375	74.2	369.7	63.8	1734.4	299.2	1344.8	173.2	868.2	55.7	1241.9	65.8	-4.3	-3	-2.3	-3.4	
7873_at		0 YPL101W	YPL101W hypothetical protein	UNKNOWN	16.8	8.2	36.3	3.6	30.1	1.6	26.3	8.3	95	3.5	108.3	11.9	107.8	3.9	106.4	7.5	-3.6	-4	-4	-4	
8326_at		0 YOR215C	YOR215C similarity to M. xanthus hypothetical protein	UNKNOWN	62.2	22.7	69.9	5.6	62.7	16.3	48.8	7.8	322.5	40.6	226.4	6	157.1	23.5	169.1	39.5	-5.2	-5.2	-5.2	-5.2	
8406_at		0 YOR161C	YOR161C similarity to C. elegans coxIII F35C6	UNKNOWN	12	4.7	66.7	6.5	27.2	2.1	51.8	10.6	125.6	57.5	164	34.3	59	7	138	29.2	-10.5	-10.5	-10.5	-10.5	
8659_at		PHM7	YOL084W	YOL084W similarity to A. thaliana hsp1 protein	UNKNOWN	12	5.5	13.1	3	12	0.8	12	3.6	53.6	13.7	47	23.3	39.1	4	33.8	0.7	-4.6	-3.8	-3.3	-2.8
8724_at		0 YOL155C	YOL155C similarity to glucan 1,4-alpha-glucosidase Sta1p and YAR066w	UNKNOWN	264.4	136.8	73	40.6	300.2	29.4	64.3	29.6	1103.3	588.4	214	144.1	1827.5	111.2	2198	343.1	-4.3	-2.9	-3.1	-3.2	
8977_at		0 YNL100W	YNL100W hypothetical protein	UNKNOWN	37.2	41.8	91.6	24.6	93.9	14.4	87	7.6	604.8	151.2	348.8	40.3	320.4	6.2	253.7	18	-6.2	-3.8	-3.5	-2.9	
9024_at		0 YMR002W	YMR002W similarity to hypothetical S. pombe and C. elegans proteins	UNKNOWN	327.2	48.4	244.1	42.7	269.3	46.1	219.7	46.2	2217.2	364.2	1192.4	59.3	1940.1	222.8	2047.2	805.9	-6.8	-4.9	-7.2	-9.3	
9631_at		0 YMR009W	YMR009W weak similarity to P. aeruginosa regulatory protein mmsR	UNKNOWN	73.5	29.4	117.3	33.1	296	71.6	73.8	18.4	776.5	105.3	357.3	45.4	1442.4	54.3	291.5	39.1	-10.6	-4.9	-4.8	-4.8	

Up-regulated under C-Lim under aerobic and anaerobic conditions (FC = fold change, ANA = anaerobic, A = aerobic)		Gene name		systematic name		functional description		cellular function														
C-Lim ANA	C-Lim A	N-Lim ANA	N-Lim A	P-Lim ANA	P-Lim A	S-Lim ANA	S-Lim A	C-Lim ANA	C-Lim A	N-Lim ANA	N-Lim A	P-Lim ANA	P-Lim A	S-Lim ANA	S-Lim A	FC C-Lim ANA vs C-Lim A	FC C-Lim ANA vs N-Lim A	FC C-Lim ANA vs P-Lim A	FC C-Lim ANA vs S-Lim A	FC C-Lim A vs N-Lim A	FC C-Lim A vs P-Lim A	FC C-Lim A vs S-Lim A
6663	at	MRK1	YDL070C	YDL070C	MDJ1 related protein kinase	UNKNOWN											10.9	10.1	10.9	3.7	5.6	4.6
7072	at	MDJ2	YDL070C	YDL070C	MDJ2 related protein kinase	UNKNOWN											10.9	10.1	10.9	3.7	5.6	4.6
8173	at	ALD4	YOR034W	YOR034W	aldehyde dehydrogenase (EC 1.2.1.5) (iso1 by SIGMA under the catalytic ETHANOL UTILIZATION	ALDEHYDE DEHYDROGENASE											10.9	10.1	10.9	3.7	5.6	4.6
10756	at	MA11	YOR030C	YOR030C	alpha-glucosyl transferase protein homolog	GLUCANASE											10.9	10.1	10.9	3.7	5.6	4.6
4043	at	SUC2	YIL025W	YIL025W	invertase (sucrose hydrolyzing enzyme)	SUCROSE UTILIZATION											14.7	14.8	14.7	8.4	10.7	8.4
4376	at	MA11	YOR030C	YOR030C	alpha-glucosyl transferase protein homolog	GLUCANASE											14.7	14.8	14.7	8.4	10.7	8.4
5307	at	HXK1	YPR053C	YPR053C	Hexokinase 1 (PH) (also called Hexokinase A)	GLYCOLYSIS											14.7	14.8	14.7	8.4	10.7	8.4
9242	at	YR244C	YR244C	unc119	YR244C zinc finger protein factor of the Zfp2-Cyfl1b structural cluster domain homologous to METACONJUGAL	UNKNOWN											14.7	14.8	14.7	8.4	10.7	8.4
9507	at	ISF1	YMR081C	YMR081C	Major regulate RNA polymerase II activity, possibly at level of RNA polymerase	RNA SPLICING MITOCHONDRIAL											14.7	14.8	14.7	8.4	10.7	8.4
9508	at	GAL4	YJL0227C	YJL0227C	transcription factor of the Zfp2-Cyfl1b structural cluster domain homologous to METACONJUGAL	UNKNOWN											14.7	14.8	14.7	8.4	10.7	8.4
9788	at	SO11	YMR044W	YMR044W	YJL0227C strong similarity to YJL0227C	UNKNOWN											14.7	14.8	14.7	8.4	10.7	8.4
4180	at	YJL0227C	YJL0227C	transcription factor of the Zfp2-Cyfl1b structural cluster domain homologous to METACONJUGAL	UNKNOWN												14.7	14.8	14.7	8.4	10.7	8.4
4753	at	YMR044W	YMR044W	YMR044W	YJL0227C strong similarity to YJL0227C	UNKNOWN											14.7	14.8	14.7	8.4	10.7	8.4
5350	at	YPR071C	YPR071C	hypothetical protein	UNKNOWN												14.7	14.8	14.7	8.4	10.7	8.4
5640	at	DAL2	YOR027W	YOR027W	strong similarity to hypothetical protein YOR027W	UNKNOWN											14.7	14.8	14.7	8.4	10.7	8.4
7738	at	CRB2	YPR020W	YPR020W	weak similarity to YEL191C	UNKNOWN											14.7	14.8	14.7	8.4	10.7	8.4
8432	at	YMR020W	YMR020W	weak similarity to hypothetical protein YMR020W	UNKNOWN												14.7	14.8	14.7	8.4	10.7	8.4
9357	at	SUC4	SUC4	SUC4	invertase (sucrose hydrolyzing enzyme)	SUCROSE UTILIZATION											12.7	12.7	12.7	6.4	10.1	6.4
7754	at	POH1	YPR020W	YPR020W	strong similarity to S. cerevisiae POH1 protein	UNKNOWN											12.7	12.7	12.7	6.4	10.1	6.4
Down-regulated under C-Lim under aerobic and anaerobic conditions (FC = fold change, ANA = anaerobic, A = aerobic)																						
C-Lim ANA	C-Lim A	N-Lim ANA	N-Lim A	P-Lim ANA	P-Lim A	S-Lim ANA	S-Lim A	C-Lim ANA	C-Lim A	N-Lim ANA	N-Lim A	P-Lim ANA	P-Lim A	S-Lim ANA	S-Lim A	FC C-Lim ANA vs C-Lim A	FC C-Lim ANA vs N-Lim A	FC C-Lim ANA vs P-Lim A	FC C-Lim ANA vs S-Lim A	FC C-Lim A vs N-Lim A	FC C-Lim A vs P-Lim A	FC C-Lim A vs S-Lim A
4605	at	RIM3	YHR021C	YHR021C	Rim3p1, mitochondrial DNA repair and recombination protein	TRANSCRIPTION											-3.8	-2.2	-3.4	-2.3	-2.2	-3.2
9654	at	SBT2	YLR042C	SBT2	regulatory factor of C-to-pyruvate kinase that negatively regulates	REGULATION											-2.7	-2.4	-2.7	-1.6	-1.4	-3.6
4880	at	WDC4	YJL025W	YJL025W	Putative integral membrane protein containing novel cysteine motif. Similarity CELL WALL MAINTENANCE	UNKNOWN											-8.6	-7.9	-8.6	-4.6	-1.6	-8.3
4884	at	TPO2	YOR138C	YOR138C	transcription factor	UNKNOWN											-15.9	-14.9	-15.9	-2.6	-2.4	-14.9
4428	at	HXT4	YHR022C	YHR022C	low to moderate-affinity glucose transporter	TRANSPORT											-40.4	-40.8	-40.4	-3.5	-3.4	-40.4
4430	at	HXT1	YHR040C	YHR040C	high-affinity glucose transporter	TRANSPORT											-32.2	-32.9	-32.2	-3.7	-3.7	-32.2
5178	at	HXT3	YOL157W	YOL157W	similar to Y. lipolytica dihydroxyacetone 4-epimerase	UNKNOWN											-9.7	-9.7	-9.7	-4.2	-4.2	-9.7
6131	at	YOR046C	YOR046C	glucose repressor	UNKNOWN												-20.9	-20.9	-20.9	-3.9	-3.9	-20.9
5216	at	YOL209W	YOL209W	YOL209W	transcription factor involved in glucose repression of SUC2, contains GLUCOSE REPRESSION	UNKNOWN											-7.3	-7.3	-7.3	-4.2	-4.2	-7.3
6033	at	TUP1	YOR046C	YOR046C	glucose repressor protein. Exhibits similarity to basic subunits of GTF-2/TFIIID	TRANSCRIPTION											-2.3	-2.3	-2.3	-2.3	-2.3	-2.3
8520	at	STD1	YOR047C	YOR047C	dosage-dependent modulator of protein repressor. Increased dosage represses CARBOHYDRATE METABOLISM	UNKNOWN											-8.2	-8.2	-8.2	-4.9	-4.9	-8.2
7892	at	TPO3	YHR156C	YHR156C	transcription factor	UNKNOWN											-25.5	-25.5	-25.5	-4.4	-4.4	-25.5
5517	at	O YER188W	YER188W	dosage-dependent modulator of protein repressor. Increased dosage represses CARBOHYDRATE METABOLISM	UNKNOWN												-4.2	-4.2	-4.2	-2.1	-2.1	-4.2
8025	at	O YPR124W	YPR124W	weak similarity to human mtm1 protein homolog	UNKNOWN												-3.2	-3.2	-3.2	-2.3	-2.3	-3.2
Down-regulated under N-Lim under aerobic and anaerobic conditions (FC = fold change, ANA = anaerobic, A = aerobic)																						
C-Lim ANA	C-Lim A	N-Lim ANA	N-Lim A	P-Lim ANA	P-Lim A	S-Lim ANA	S-Lim A	C-Lim ANA	C-Lim A	N-Lim ANA	N-Lim A	P-Lim ANA	P-Lim A	S-Lim ANA	S-Lim A	FC C-Lim ANA vs C-Lim A	FC C-Lim ANA vs N-Lim A	FC C-Lim ANA vs P-Lim A	FC C-Lim ANA vs S-Lim A	FC C-Lim A vs N-Lim A	FC C-Lim A vs P-Lim A	FC C-Lim A vs S-Lim A
8249	at	SPR4	YOR131C	YOR131C	sporulation-specific protein	SPOROULATION											2.2	2.2	2.2	5.9	4.6	5.9
9483	at	ALD2	YMR170C	YMR170C	aldehyde dehydrogenase (NAD(P)+) illytic cyclase	ETHANOL UTILIZATION											9.9	9.9	9.9	4.5	4.1	9.9
10214	at	MDJ8	YOR030C	YOR030C	alpha-glucosyl transferase homolog (NAD(P)+) illytic cyclase	GLUCANASE											2.8	2.8	2.8	6.5	6.5	2.8
10751	at	PUT1	YLR142W	YLR142W	(PUT1) Putative oxidase/protein dehydrogenase, first step in synthesis of glu/PURINE METABOLISM	PURINE METABOLISM											10.3	10.3	10.3	4.5	4.5	10.3
10511	at	QAP1	YOR030W	YOR030W	QAP1 General amino acid permease, proton symport transporter for all net TRANSPORT	TRANSPORT											22.2	22.2	22.2	10.1	10.1	22.2
10121	at	DAL8	YOR030W	YOR030W	YOR030W (putative) regulator of multiple nitrogen catabolic genes	NITROGEN CATABOLISM											37.6	37.6	37.6	20.9	20.9	37.6
4063	at	DAL2	YOR027W	YOR027W	alpha-glucosyl transferase	GLUCANASE											15.2	15.2	15.2	8.4	8.4	15.2
4063	at	DAL1	YOR027C	YOR027C	invertase	SUCROSE UTILIZATION											22.2	22.2	22.2	10.1	10.1	22.2
4063	at	DAL2	YOR027W	YOR027W	alpha-glucosyl transferase	GLUCANASE											15.2	15.2	15.2	8.4	8.4	15.2
4063	at	DCO1	YHR030C	YHR030C	involved in nitrogen-catabolite metabolism	UNKNOWN											6.6	6.6	6.6	5.6	5.6	6.6
4067	at	YOR027W	YOR027W	alpha-glucosyl transferase	GLUCANASE												15.2	15.2	15.2	8.4	8.4	15.2
4068	at	DAL3	YHR030C	YHR030C	ureidoglycolytic hydrolase	PURINE METABOLISM											8.4	8.4	8.4	2.3	2.3	8.4
4284	at	BSL1	YOR030C	YOR030C	beta-1,4-galactosyl transferase, converts sucrose to pyruvate and ammonia for PROLINE METABOLISM	PURINE METABOLISM											4.3	4.3	4.3	2.4	2.4	4.3
4511	at	PUR2	YHR037W	YHR037W	beta-1,4-galactosyl transferase, converts sucrose to pyruvate and ammonia for PROLINE METABOLISM	PURINE METABOLISM											2.8	2.8	2.8	2.2	2.2	2.8
4511	at	MDJ2	YOR030C	YOR030C	beta-1,4-galactosyl transferase, converts sucrose to pyruvate and ammonia for PROLINE METABOLISM	PURINE METABOLISM											2.8	2.8	2.8	2.2	2.2	2.8
8599	at	MLS1	YHL117C	YHL117C	MLS1 Maltase system 1, functions in glycoprotein cycle, has near identity to GLYCOYLATE CYCLE	UNKNOWN											24.7	24.7	24.7	10.1	10.1	24.7
7230	at	DURS	YOR156C	YOR156C	Urea transporter	TRANSPORT											2.1	2.1	2.1	2.7	2.7	2.1
4068	at	DURS	YOR156C	YOR156C	Urea transporter	TRANSPORT											2.4	2.4	2.4	10.1	10.1	2.4
10554	at	MLS2	YOR156C	YOR156C	Urea transporter	TRANSPORT											2.4	2.4	2.4	10.1	10.1	2.4
4068	at	DAL4	YHR030C	YHR030C	ureidoglycolytic hydrolase	PURINE METABOLISM											24.7	24.7	24.7	10.1	10.1	24.7
7845	at	YOR030C	YOR030C	alpha-glucosyl transferase	GLUCANASE												15.2	15.2	15.2	8.4	8.4	15.2
8194	at	PUR4	YOR034C	YOR034C	putative proteinase-specific permease	TRANSPORT											10.4	10.4	10.4	8.3	8.3	10.4
8194	at	MEP2	YOR142W	YOR142W	MEP2 membrane protein involved in protein capacity and high affinity, involved genes	TRANSPORT											10.4	10.4	10.4	8.3	8.3	10.4
9530	at	YHR086C	YHR086C	similarity to multidrug resistance protein	UNKNOWN												11.6	11.6	11.6	9.9	9.9	11.6
9530	at	YHR086C	YHR086C	similarity to multidrug resistance protein	UNKNOWN												11.6	11.6	11.6	9.9	9.9	11.6
4068	at	DURS	YOR156C	YOR156C	Urea transporter	TRANSPORT											2.4	2.4	2.4	10.1	10.1	2.4
4068	at</																					

Gene Set	Gene Name	Function	C ₁ Lim ANA Average	C ₁ Lim ANA Stdev	N ₁ Lim ANA Average	N ₁ Lim ANA Stdev	P ₁ Lim ANA Average	P ₁ Lim ANA Stdev	S ₁ Lim ANA Average	S ₁ Lim ANA Stdev	C ₂ Lim ANA Average	C ₂ Lim ANA Stdev	N ₂ Lim ANA Average	N ₂ Lim ANA Stdev	P ₂ Lim ANA Average	P ₂ Lim ANA Stdev	S ₂ Lim ANA Average	S ₂ Lim ANA Stdev	FC S-Lim ANA vs C-Lim ANA	FC S-Lim ANA vs N-ANA	FC S-Lim ANA vs P-Lim ANA	FC S-Lim ANA vs C-Lim ANA	FC S-Lim ANA vs N-AE	FC S-Lim ANA vs P-Lim ANA	
10404_at	MET1	YKR090W	YKR090W serine synthase	477.4	63.2	476.5	72.1	1861.9	151.2	328.5	36.4	548.4	111.1	626.8	70.1	2150.3	32.1	889.9	384.9	3.9	3.9	5.7	3.9	4.1	2.4
11032_at	MET3	YJR010W	ATF sulfuryase	922.1	103.5	703.1	54.8	2051.2	191.6	318.2	54.9	784.5	112.8	924.8	62.7	2175.3	165.3	699.8	6.6	2.2	2.2	2.6	2.4	3.1	2.4
11037_at	DMB3	YAL060W	YAL060W similar to tyrosine aminotransferase and glutamine phenylpyruvate transaminase	153.6	20.2	255.4	48.3	635	54.9	134.4	20.7	188.7	17.1	302.2	82.2	687	116	202.1	15.9	4.1	2.5	4.7	4.6	2.3	4.3
11355_at	CVY3	YAL012W	YAL012W cystathionine gamma-lyase	1004.9	264.7	148.9	3240.4	512.4	614.2	84.5	688.9	151.3	800.6	214.4	2020.7	361.1	1378.1	644.4	4.4	5.3	4.5	2.8	3.7	2.1	3.1
4059_at	MET28	YR017C	YR017C Transaminase activator of sulfur amino acid metabolism	275.8	28.6	232.3	10.9	1010.4	97.9	181.3	34.7	289.9	28.3	271.7	38.3	1141.1	85.9	231.5	14.3	3.7	4	5.6	3.9	4.2	4
4190_at	SEK33	YL074C	YL074C strong similarity to E. coli phosphoglycerate dehydrogenase	369.2	11.7	306.5	35.4	754	22.1	182.4	46.8	181.4	21.8	218.6	11.8	712.5	104.8	288.9	58.4	4.1	2.5	2.8	3.4	2.8	3.4
4331_at	YMR15W	YMR15W Diethylmalonate methyltransferase	33.7	10.6	66.1	9.5	699.8	66.7	33.2	6.6	42.3	6.3	119.8	20.9	700.8	35.4	83.2	25.8	6.5	8.4	16.9	6.4	8.4	8.4	
4572_at	MUP3	YHL030W	YHL030W (MUP3) very low affinity methionine permease	62.2	18.9	144.8	31.6	432.0	66.4	63.9	14	66.3	9.2	206.3	38.6	620.3	87.2	116.3	10.5	7	3	8	3.9	3.3	2.4
4583_at	FOG6	YOR097C	YOR097C Thiol methyl acetyltransferase	18.6	6.6	71.5	18.2	1140.3	254.5	24	6.1	96	43.9	181.1	20.3	874.3	236.7	37.4	7.3	65.8	48.4	26.4	11.6	60.1	60.1
4636_at	MUP1	YOR025W	YOR025W high affinity methionine permease	417.7	20.3	48.1	5.1	896.7	113.3	29.5	8.5	104.8	33.9	151.8	28.3	1544.3	221.3	237.4	30.6	2.8	2.8	14.7	10.2	6.5	5.1
5136_at	ETH3	YOL184C	YOL184C strong similarity to Emericella nidulans cystathionine beta-lyase	50	15.8	86.7	14.3	546.8	13.9	48.7	7.4	248.2	73.9	180.8	18.9	839	88.1	184.6	8.6	11.2	3.8	5.2	5.1	11.2	11.2
5327_at	MET10	YFR030W	YFR030W subunit of assimilatory sulfate reductase	368.2	53.1	441.9	32.2	941.6	176.2	142.6	5.4	329.2	35.8	391.9	24.9	1038.3	109.9	336.4	89.6	2.6	2.1	16.4	2.1	3.6	3.1
6218_at	MET22	YH025C	YH025C zinc finger DNA binding factor in the expression of FAPs reductase and sulfate reductase	100.9	26.1	104.9	46.9	443	76.1	29.5	84	217.2	47	142.8	13.8	86.4	97.7	80.9	13.6	4.4	1.5	3.9	5.9	9.6	9.6
7189_at	MET8	YBR210W	YBR210W Ethanol: S-sulfate lyase	30.8	2.4	29.5	10.9	402	73.7	23.8	3.2	42.7	17.8	51.3	12.2	663	69.4	120.1	58.5	15.1	16.9	15.6	15.6	15.6	15.6
7955_at	MET16	YPR167C	YPR167C 3-oxopropionyl-CoA thioesterase	247.7	44	364.9	31.9	917	238.7	175.6	10	242.4	39.7	386.6	43.1	1072.4	136.6	338.5	78.4	3.7	2.5	5.2	2.8	2.9	2.2
8021_at	SAM3	YPR274W	YPR274W strong similarity to amino acid transport protein	52.1	8.4	74	11.9	158.4	26.1	56.4	17.8	58.9	5	61	19.5	235.1	54.6	118.7	17.7	3	2.1	4	2.9	2	4
9152_at	MET2	YNL277W	YNL277W homoserine O-transacetylase	501.2	41.5	640.7	107.1	1468.7	158.2	363	42.3	389.5	42	614.2	81.5	1922.7	53.3	709.3	109.5	2.9	2.9	2.9	2.9	2.2	2.2
10256_at	SUL2	YLR002W	YLR002W high affinity sulfate permease	506.6	66	498.3	52.1	1396.3	89.8	91.1	22.2	349.9	43.6	433.3	30.5	1193.7	102	344.7	26	3.4	2.6	3.4	3.5	2.6	3.5
10210_at	YHL136C	YHL136C zinc finger containing homolog of mammalian TIS11	12	3.4	15.2	6.2	63.6	18	1.9	10.2	5.7	1.2	3.3	3.5	12.2	12.2	44.4	3.9	2.4	4.5	2.9	2.4	4.5	2.4	
10375_at	TIS11	YLL059W	YLL059W similar to Gsp9	93.9	18.6	321.3	23.6	1862.5	245.1	75.2	8.2	174.2	69.1	454	22.4	2003.7	187.4	146.6	35.1	6.2	2.6	11.6	4.6	6.2	11.6
10415_at	MMP1	YLL061W	YLL061W (MMP1) high affinity S-methylmethionine permease. has similarity to Gsp19	110.2	50.6	153.5	48.7	794.4	102.3	28.5	6.6	324.9	16.2	23.8	2.3	827.9	54.4	17.6	4.9	7.2	5.5	27.8	4.7	7.2	
4585_at	APN1	YLR166C	YLR166C similar to glutathione permease Dsp9	12	6.3	20.6	5.7	245.5	141.1	25.3	3.4	22.3	2.9	21.7	2.2	1650.4	271.2	24.9	4.4	2.7	16.8	8.6	7.2	7.2	
4668_at	APN1	YHL040C	YHL040C Protein involved in the transport of ferrioxamine. member of the yeast-specific UNKOWN	23.1	10.9	70.3	5.1	175.4	21.8	42.2	11.9	97	18.4	43.4	5.7	325.9	68.8	53.8	13.5	7.6	2.5	2.2	5.8	6.1	
5452_at	AGP3	YFR059W	YFR059W Amino acid permease	12	1.9	12.1	2.7	400.2	54.5	12	4.6	31.6	4.8	12	0.2	773.8	111.3	12	2.3	3.3	24.7	35	64.5	35	
7059_at	APN1	YBR205W	YBR205W Probable multidrug resistance protein	99.8	14.3	126.3	18.3	546.7	105.4	137.1	11.7	113.4	14.2	121.1	11.7	113.4	14.2	121.1	11.7	5.5	4	4	8.2	9	
8177_at	D	YFR059W	YFR059W Amino acid permease	12	1.9	12.1	2.7	400.2	54.5	12	4.6	31.6	4.8	12	0.2	773.8	111.3	12	2.3	3.3	24.7	35	64.5	35	
8718_at	D	YFR059W	YFR059W Amino acid permease	12	1.9	12.1	2.7	400.2	54.5	12	4.6	31.6	4.8	12	0.2	773.8	111.3	12	2.3	3.3	24.7	35	64.5	35	
9207_at	ATM1	YMR031C	YMR031W strong similarity to aminobenzoyl transporter	12	5	55.9	16	440.5	94.6	22.1	1.7	12	2.8	21.9	3.6	638	10.6	12	1.7	5.6	4.3	8.2	9		
10074_at	D	YMR031C	YMR031W strong similarity to aminobenzoyl transporter	12	5	55.9	16	440.5	94.6	22.1	1.7	12	2.8	21.9	3.6	638	10.6	12	1.7	5.6	4.3	8.2	9		
10374_at	D	YMR031C	YMR031W strong similarity to aminobenzoyl transporter	12	5	55.9	16	440.5	94.6	22.1	1.7	12	2.8	21.9	3.6	638	10.6	12	1.7	5.6	4.3	8.2	9		
10414_at	MHT1	YLL026C	YLL026C weak similarity to Y. pseudotuberculosis CDP-3,6-dioxey-D-glycero-L-glycero-L-glycerate UNKOWN	25.1	10.4	52.8	7.6	208.8	50.7	41.3	9.7	45.5	9.3	40.1	7.4	588.3	26.5	53.7	10.9	8.2	3.9	11.1	5.9	11	
4855_at	D	YGR154C	YGR154C weak similarity to M. luteus metC protein	17.4	21.6	55.6	14.9	956.4	160.2	19.5	2.7	103.3	17.5	19.5	4.3	603.8	17.9	12	4.4	7.7	10.7	30.4	3.4	56.3	
5423_at	D	YGR154C	YGR154C weak similarity to M. luteus metC protein	17.4	21.6	55.6	14.9	956.4	160.2	19.5	2.7	103.3	17.5	19.5	4.3	603.8	17.9	12	4.4	7.7	10.7	30.4	3.4	56.3	
5423_at	D	YGR154C	YGR154C weak similarity to M. luteus metC protein	17.4	21.6	55.6	14.9	956.4	160.2	19.5	2.7	103.3	17.5	19.5	4.3	603.8	17.9	12	4.4	7.7	10.7	30.4	3.4	56.3	
7999_at	ICY2	YPR250C	YPR250C weak similarity to YMR156W	184.4	37	163.7	27.8	955.5	138.9	234.3	2.8	364.7	122.5	126.4	33.5	1705.5	181.7	486.7	23.5	4.9	5.8	5.8	6.8	13.5	
8719_at	D	YHL191W	YHL191W strong similarity to hypothetical proteins YKR075W and YMR215W	14.7	10.7	12	4.9	424	3	19.8	6.5	12	2.1	18.5	7.6	42.7	10.5	17.4	6.8	3.9	2.5	2.3	2.5	2.3	
9068_at	D	YHL191W	YHL191W strong similarity to hypothetical proteins YKR075W and YMR215W	14.7	10.7	12	4.9	424	3	19.8	6.5	12	2.1	18.5	7.6	42.7	10.5	17.4	6.8	3.9	2.5	2.3	2.5	2.3	
9648_at	D	YHL191W	YHL191W strong similarity to hypothetical proteins YKR075W and YMR215W	14.7	10.7	12	4.9	424	3	19.8	6.5	12	2.1	18.5	7.6	42.7	10.5	17.4	6.8	3.9	2.5	2.3	2.5	2.3	
9648_at	D	YHL191W	YHL191W strong similarity to hypothetical proteins YKR075W and YMR215W	14.7	10.7	12	4.9	424	3	19.8	6.5	12	2.1	18.5	7.6	42.7	10.5	17.4	6.8	3.9	2.5	2.3	2.5	2.3	
9643_at	D	YLR364W	YLR364W hypothetical protein	61.3	22	70.8	15.9	441.2	60.3	29.7	3.7	100.7	21	124.7	7.5	714	64.5	188.1	28.8	7.2	6.2	14.8	6.8	7.1	
10039_at	CHP1	YKL095W	YKL095W cell wall maintenance	155.6	54.7	98.4	8.9	421.8	110.6	116.7	35.2	282.3	99.3	100.8	17.5	1222	68.6	69.5	24.3	2.2	4.3	5.7	4.2	12.1	

Down-regulated under S-Lim anaerobic and aerobic conditions (FC < 0.5 change, ANA + anaerobic, A + aerobic)

Gene Set	Gene Name	Function	C ₁ Lim ANA Average	C ₁ Lim ANA Stdev	N ₁ Lim ANA Average	N ₁ Lim ANA Stdev	P ₁ Lim ANA Average	P ₁ Lim ANA Stdev	S ₁ Lim ANA Average	S ₁ Lim ANA Stdev	C ₂ Lim ANA Average	C ₂ Lim ANA Stdev	N ₂ Lim ANA Average	N ₂ Lim ANA Stdev	P ₂ Lim ANA Average	P ₂ Lim ANA Stdev	S ₂ Lim ANA Average	S ₂ Lim ANA Stdev	FC S-Lim ANA vs C-Lim ANA	FC S-Lim ANA vs N-ANA	FC S-Lim ANA vs P-Lim ANA	FC S-Lim ANA vs C-Lim ANA	FC S-Lim ANA vs N-AE	FC S-Lim ANA vs P-Lim ANA
7843_at	SSU1	YPL092W	YPL092W sensitive to sulfate	217.5	41	226	62.2	86.5	63	265.5	30.4	449.7	149.5	109	61.1	4.7	552.8	21.3	-2.6	-2.6	-4.8	-2.6	-4.8	-3.2
5116_at	SCB3	YGL120W	YGL120W required for inositol prototrophy	419.3	47.3																			

Up-regulated under C-Lim Aerobic only (FC = fold change; ANA = anaerobic; A = aerobic)	Probe Set	Gene name	systematic name	functional description	cellular function	C-Lim ANA		C-Lim A		N-Lim ANA		N-Lim A		P-Lim ANA		P-Lim A		S-Lim ANA		S-Lim A		C-Lim A		N-Lim A		P-Lim A		S-Lim A		FC C-Lim AE vs N-Lim AE	FC C-Lim AE vs S-Lim AE	FC C-Lim AE vs P-Lim AE		
						Average	Sidev	Average	Sidev	Average	Sidev	Average	Sidev	Average	Sidev	Average	Sidev				Average	Sidev												
10022_at	CD1A	YLR307W	YLR307W	Chitin Deacetylase	CELLULOSE DEGRADATION	45.1	19.5	49.3	12.8	43	8.3	110.6	2.8	26.6	7.2	12	3.6	12	0.1	12	4.2	2.2	2.2	2.2	2.2	2.2	2.2	2.2	2.2	2.2	2.2	2.2	2.2	
10043_at	EC11	YLR284W	YLR284W	lysine-dependent acyl-CoA hydratase	CELLULOSE DEGRADATION	36.6	16.3	71.2	7.9	64.8	12.4	38.6	12.7	39.3	62.7	12.7	39.3	62.7	12.7	39.3	62.7	12.7	39.3	62.7	12.7	39.3	62.7	12.7	39.3	62.7	12.7	39.3	62.7	
10120_at	ICD2	YLR194C	YLR194C	hypothetical protein	CELLULOSE DEGRADATION	301.9	42.3	216.4	17.8	236.2	43.1	259.1	28	458.6	114.3	88.9	63	168.8	65	148.2	35.5	5.2	5.2	5.2	5.2	5.2	5.2	5.2	5.2	5.2	5.2	5.2	5.2	
10154_at	ICP1	YLR174W	YLR174W	Cytosolic form of NADP-dependent isocitrate dehydrogenase	TCA CYCLE	12	2.6	39.2	2.7	21.9	3.7	19.8	7.4	69.9	86.9	51.8	9.5	4.1	5.4	34.3	14.7	12.9	12.9	12.9	12.9	12.9	12.9	12.9	12.9	12.9	12.9	12.9	12.9	
10470_at	SR33	YK091W	YK091W	Suppressor of rad53 lethality	CELLULOSE DEGRADATION	107.8	22.2	104.2	7.4	121.9	22.5	153.1	17.4	89.8	8	34.5	5.4	29.8	16	27.7	10.4	2.6	3	3	3	3	3	3	3	3	3	3	3	3
10492_at	GP72	YK067W	YK067W	strong similarity to Sclp1	CELLULOSE DEGRADATION	187.8	27.2	179.8	36	173	54.5	148.2	42	543.6	59.4	237	62	171.8	32.9	180.7	65.7	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3
10500_at	ECM4	YK075C	YK075C	weak similarity to negative regulator Reg1p	CELLULOSE DEGRADATION	38.6	13.8	64	12.1	39.5	1.1	31.1	5	186.9	26.6	65.5	3.4	51.7	5	37.5	6.4	2.9	3	3	3	3	3	3	3	3	3	3	3	3
10501_at	ECM4	YK075C	YK075C	weak similarity to negative regulator Reg1p	CELLULOSE DEGRADATION	46.1	14.2	63.9	22.2	44.7	7.3	23	6	215.1	48.8	83.9	14.2	72.2	10.5	51.9	5.8	3	3	3	3	3	3	3	3	3	3	3	3	3
10521_at	YK049C	YK049C	YK049C	hypothetical protein	CELLULOSE DEGRADATION	377.2	86.2	344.9	47.3	243.1	67.2	258	9.7	1064.8	301.7	362.1	21.6	20.4	19.4	19.6	452.9	147.4	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5
10572_at	FOX2	YK029C	YK029C	peroxisomal multifunctional beta-oxidation protein	FATTY ACID METABOLISM	30.7	3.4	102.4	8.7	63.3	8.6	54.7	14.4	894.6	254.8	104.8	24.4	75.6	17.4	67.6	12.1	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9
10581_at	CPK1	YK102C	YK102C	strong similarity to glutamine peptidase	CELLULOSE DEGRADATION	92.9	53.8	99.6	23.2	45.5	14.2	58.8	6.3	763.5	191.7	134.5	23.8	69	5.9	70.9	4.9	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	
10646_at	MBR1	YK1093W	YK1093W	MIR1 protein precursor	MITOCHONDRIAL BIOGENESIS	19.2	7.7	16	6.2	17.3	2.5	12	1.6	302	76.8	21.5	4.3	52.8	3.2	16.7	4.5	14	14	14	14	14	14	14	14	14	14	14	14	14
10677_at	MRP13	YK107W	YK107W	weak similarity to S-ambiosin protease oxidoreductase	MITOCHONDRIAL BIOGENESIS	12	2.6	12	0.2	12	1.8	12	0.9	46.9	12	12	3.7	12	2.1	12	1	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	
10735_at	YK138C	YK138C	YK138C	15.8 kDa mitochondrial ribosomal protein Yml31	PROTEIN SYNTHESIS	110.2	31.9	72.7	17.5	60.7	6.3	59.8	6.9	195.5	41.4	92.2	21.1	85.4	1.9	79.5	14.7	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	
10748_at	GP72	YK067W	YK067W	strong similarity to Sclp1	CELLULOSE DEGRADATION	187.8	27.2	179.8	36	173	54.5	148.2	42	543.6	59.4	237	62	171.8	32.9	180.7	65.7	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	
10500_at	ECM4	YK075C	YK075C	weak similarity to negative regulator Reg1p	CELLULOSE DEGRADATION	38.6	13.8	64	12.1	39.5	1.1	31.1	5	186.9	26.6	65.5	3.4	51.7	5	37.5	6.4	2.9	3	3	3	3	3	3	3	3	3	3	3	
10521_at	YK049C	YK049C	YK049C	hypothetical protein	CELLULOSE DEGRADATION	377.2	86.2	344.9	47.3	243.1	67.2	258	9.7	1064.8	301.7	362.1	21.6	20.4	19.4	19.6	452.9	147.4	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	
10572_at	FOX2	YK029C	YK029C	peroxisomal multifunctional beta-oxidation protein	FATTY ACID METABOLISM	30.7	3.4	102.4	8.7	63.3	8.6	54.7	14.4	894.6	254.8	104.8	24.4	75.6	17.4	67.6	12.1	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	
10581_at	CPK1	YK102C	YK102C	strong similarity to glutamine peptidase	CELLULOSE DEGRADATION	92.9	53.8	99.6	23.2	45.5	14.2	58.8	6.3	763.5	191.7	134.5	23.8	69	5.9	70.9	4.9	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	
10646_at	MBR1	YK1093W	YK1093W	MIR1 protein precursor	MITOCHONDRIAL BIOGENESIS	19.2	7.7	16	6.2	17.3	2.5	12	1.6	302	76.8	21.5	4.3	52.8	3.2	16.7	4.5	14	14	14	14	14	14	14	14	14	14	14	14	14
10677_at	MRP13	YK107W	YK107W	weak similarity to S-ambiosin protease oxidoreductase	MITOCHONDRIAL BIOGENESIS	12	2.6	12	0.2	12	1.8	12	0.9	46.9	12	12	3.7	12	2.1	12	1	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	
10735_at	YK138C	YK138C	YK138C	15.8 kDa mitochondrial ribosomal protein Yml31	PROTEIN SYNTHESIS	110.2	31.9	72.7	17.5	60.7	6.3	59.8	6.9	195.5	41.4	92.2	21.1	85.4	1.9	79.5	14.7	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	
10748_at	GP72	YK067W	YK067W	strong similarity to Sclp1	CELLULOSE DEGRADATION	187.8	27.2	179.8	36	173	54.5	148.2	42	543.6	59.4	237	62	171.8	32.9	180.7	65.7	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	
10500_at	ECM4	YK075C	YK075C	weak similarity to negative regulator Reg1p	CELLULOSE DEGRADATION	38.6	13.8	64	12.1	39.5	1.1	31.1	5	186.9	26.6	65.5	3.4	51.7	5	37.5	6.4	2.9	3	3	3	3	3	3	3	3	3	3	3	
10521_at	YK049C	YK049C	YK049C	hypothetical protein	CELLULOSE DEGRADATION	377.2	86.2	344.9	47.3	243.1	67.2	258	9.7	1064.8	301.7	362.1	21.6	20.4	19.4	19.6	452.9	147.4	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	
10572_at	FOX2	YK029C	YK029C	peroxisomal multifunctional beta-oxidation protein	FATTY ACID METABOLISM	30.7	3.4	102.4	8.7	63.3	8.6	54.7	14.4	894.6	254.8	104.8	24.4	75.6	17.4	67.6	12.1	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	
10581_at	CPK1	YK102C	YK102C	strong similarity to glutamine peptidase	CELLULOSE DEGRADATION	92.9	53.8	99.6	23.2	45.5	14.2	58.8	6.3	763.5	191.7	134.5	23.8	69	5.9	70.9	4.9	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	5.7	
10646_at	MBR1	YK1093W	YK1093W	MIR1 protein precursor	MITOCHONDRIAL BIOGENESIS	19.2	7.7	16	6.2	17.3	2.5	12	1.6	302	76.8	21.5	4.3	52.8	3.2	16.7	4.5	14	14	14	14	14	14	14	14	14	14	14	14	
10677_at	MRP13	YK107W	YK107W	weak similarity to S-ambiosin protease oxidoreductase	MITOCHONDRIAL BIOGENESIS	12	2.6	12	0.2	12	1.8	12	0.9	46.9	12	12	3.7	12	2.1	12	1	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	3.9	
10735_at	YK138C	YK138C	YK138C	15.8 kDa mitochondrial ribosomal protein Yml31	PROTEIN SYNTHESIS	110.2	31.9	72.7	17.5	60.7	6.3	59.8	6.9	195.5	41.4	92.2	21.1	85.4	1.9	79.5	14.7	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	2.1	
10748_at	GP72	YK067W	YK067W	strong similarity to Sclp1	CELLULOSE DEGRADATION	187.8	27.2	179.8	36	173	54.5	148.2	42	543.6	59.4	237	62	171.8	32.9	180.7	65.7	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	2.3	
10500_at	ECM4	YK075C	YK075C	weak similarity to negative regulator Reg1p	CELLULOSE DEGRADATION	38.6	13.8	64	12.1	39.5	1.1	31.1	5	186.9	26.6	65.5	3.4	51.7	5	37.5	6.4	2.9	3	3	3	3	3	3	3	3	3	3	3	
10521_at	YK049C	YK049C	YK049C	hypothetical protein	CELLULOSE DEGRADATION	377.2	86.2	344.9	47.3	243.1	67.2	258	9.7	1064.8	301.7	362.1	21.6	20.4	19.4	19.6	452.9	147.4	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	
10572_at	FOX2	YK029C	YK029C	peroxisomal multifunctional beta-oxidation protein	FATTY ACID METABOLISM	30.7	3.4	102.4	8.7	63.3	8.6	54.7	14.4	894.6	254.8	104.8	24.4	75.6	17.4	67.6	12.1	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	2.9	
10581_at	CPK1	YK102C	YK102C	strong similarity to glutamine peptidase	CELLULOSE DEGRADATION	92.9	53.8	99.6	23.2	45.5	14.2	58.8																						

Probe Set	Gene name	systematic name	functional description	cellular function	C-Lim ANA Average	C-Lim ANA Stdev	N-Lim ANA Average	N-Lim ANA Stdev	P-Lim ANA Average	P-Lim ANA Stdev	S-Lim ANA Average	S-Lim ANA Stdev	C-Lim AE Average	C-Lim AE Stdev	N-Lim AE Average	N-Lim AE Stdev	P-Lim AE Average	P-Lim AE Stdev	S-Lim AE Average	S-Lim AE Stdev	FC N-Lim AE vs C-Lim AE	FC N-Lim AE vs S-Lim AE	FC N-Lim AE vs P-Lim AE	
6905_at	FUS1	YCL027W	YCL027W serine/threonine-rich membrane protein	MATING, CELL FUSION	25.6	10.2	74.8	9.5	72.7	3.7	117.1	19.4	23.9	8.1	75	19.2	37.5	7.4	33.6	3.3	3.1	2	2.2	
6917_at	MRC1	YCL061C	YCL061C similarity to myosin heavy chain form b from Chicken and Xenopus	UNKNOW	20.8	9.3	37.9	4.4	34.6	15.3	32.7	5.2	23	5.7	50.3	13.4	23.8	10.9	24.7	8.6	2.2	2	2.1	
7820_at		YPL150W	YPL150W similarity to ser/Thr protein kinases	UNKNOW	36.5	8.5	66.8	22.6	52.4	8.3	41.1	6.5	48.9	22.1	140.3	32.6	60.7	3	53.7	11.3	2.9	2.3	2.6	
7827_at	MFJALPHA1	YPL187AC	YPL187AC similarity to Rho GTPase	UNKNOW	45.8	10.8	70.3	26.5	62.1	14.6	104.1	110.3	24.5	45	11.1	30.5	45	11.1	30.5	45	11.1	30.5	45	
8505_at	HMS1	YOR032C	YOR032C myc-family transcription factor homolog	PSEUDOHYPHAL GROWTH	53.5	26.6	72.2	4.9	19.9	1.8	67	12	2.5	63.5	13.9	12	0.1	31.3	3.2	2.1	3.3	5.3	2	
8780_at		YOR086C	YOR086C similarity to Bu1p	UNKNOW	29.3	23.2	146.7	39.6	47.9	3.3	62.8	10.7	61.4	4.8	126.8	20	38.1	0.2	45.2	4.1	5.3	2.8	2.8	
8993_at		YNL129W	YNL129W similarity to M.pneumoniae uridine kinase ukl	UNKNOW	59.9	30.6	145.7	24.8	18.2	13.4	103.1	18.1	62.6	24	289.1	12.8	77.3	5.2	42.9	8.2	4.3	3.5	6.3	
9284_at	FT4	YMR319C	YMR319C Low-affinity Fe(II) transport protein	TRANSPORT	156.8	41.4	334.3	87.5	315.7	28.2	293.2	18.9	12	1.9	122.9	30.2	17.4	2.8	55.1	4.4	10.2	7.1	2.2	2.2
Down-regulated under P-Lim aerobic only (FC = fold change, ANA = anaerobic, AEA = aerobic)																								
10377_at		YLL052C	YLL052C similarity to walter channel proteins	cellular function	C-Lim ANA Average	C-Lim ANA Stdev	N-Lim ANA Average	N-Lim ANA Stdev	P-Lim ANA Average	P-Lim ANA Stdev	S-Lim ANA Average	S-Lim ANA Stdev	C-Lim AE Average	C-Lim AE Stdev	N-Lim AE Average	N-Lim AE Stdev	P-Lim AE Average	P-Lim AE Stdev	S-Lim AE Average	S-Lim AE Stdev	FC N-Lim AE vs C-Lim AE	FC N-Lim AE vs S-Lim AE	FC N-Lim AE vs P-Lim AE	
10378_at	AQZY	YLL052C	YLL052C member of myc family transmembrane channels	TRANSPORT, WATER	23.9	6.8	56.1	7.2	77.3	10	86	18.2	41.87	78.5	130.1	5.7	135.6	120.9	107.8	594.3	-3.6	-7.8	-6.1	
11133_at	NCA3	YAL116C	YAL116C With NCA2, regulates proper expression of subunits 6 (Atp6p) and 8 (Atp8p) of ATP SYNTHESIS	TRANSPORT	27	11.8	44.1	6.8	47.6	5	49.1	15.7	192.3	79.6	13.8	5.7	87.5	11.9	90.3	14.8	-13.9	-10.3	-8.3	
4891_at	HIP1	YGR191W	YGR191W histidine permease	TRANSPORT	123.6	18.6	69.3	9.5	133.9	23.4	189.2	17.3	95.8	15.2	37.7	5.7	105.5	3.6	195.1	6.8	-2.6	-2.9	-4.9	
4846_at		YGR146C	YGR146C nuclear localization sequence	UNKNOW	78.3	25.5	74.7	14.4	165.8	26.1	169.9	82.7	148	36.3	13.8	2.8	53.7	11	137.4	28.3	-10.7	-3.9	-9.9	
4860_at	NSR1	YGR159C	YGR159C nuclear localization sequence binding protein	NUCLEAR PROTEIN TARGETING	394.4	54.1	224.9	44.7	307.9	126.6	412.6	65.9	518.4	127.2	137.3	76.4	367.8	238	408.4	111	-3.8	-2.7	-3	
6854_at	HSP20	YCR021C	YCR021C Heat shock protein located in the plasma membrane with a role in cellular pH homeostasis	STRESS	55.1	24.8	113.9	7.3	98.3	20.4	114.3	13.7	498.8	334.9	62	10.1	246.8	19.9	157.6	77.1	-4	-2.5	-4	
7291_at	BAP2	YBR088C	YBR088C probable amino acid permease for leucine, valine, and isoleucine	TRANSPORT	102	9.4	26.7	4.8	99.4	9.7	45.1	9.4	336.3	77.4	20.3	3.8	119.3	14.3	44.2	5	-16.6	-5.9	-2.2	
7295_at	HSP26	YBR072W	YBR072W Heat shock protein of 26 kDa, expressed during entry to stationary phase and in STRESS	UNKNOW	148.2	41.1	281.1	37.2	352.2	81.9	258.4	31.7	1769.5	317.3	222.6	27.3	519	74.1	734.3	121.1	-7.9	-2.3	-3.3	
7981_at	GRE1	YPL223C	YPL223C induced by osmotic stress	UNKNOW	13.3	4.4	24.9	1.5	47.3	12.6	20.1	7.4	190.4	818.6	12	3.1	846.6	181	29.5	3.9	-11.4	-70.6	-11.4	
7999_at	ICY2	YPL250C	YPL250C weak similarity to YMR195W	UNKNOW	185.4	37	163.7	27.8	90.5	138.9	234.3	2.6	304.7	125.3	128.4	33.5	170.5	181.7	498.7	23.5	-2.4	-3.5	-3.9	
8724_at		YPL155C	YPL155C similarity to glucan 1,4-alpha-glucosidase St1p and YAR066w	UNKNOW	264.4	136.8	73	40.8	300.2	29.4	64.3	28.8	1133.3	588.4	214	144.1	1627.5	111.2	2198	343.1	-5.3	-18.5	-10.3	
9038_at	FE23	YMR069W	YMR069W multicopper oxidase	TRANSPORT	13.8	6	15.2	2.8	45.5	23.1	12	4.4	126.3	43.3	289.9	2.8	109.5	35.8	136.2	18.8	-4.4	-3.8	-4.7	
9633_at	HXT2	YMR011W	YMR011W high affinity hexose transporter-2	TRANSPORT	457.3	283.3	1169.8	50.9	934.1	138.4	1117.7	117.4	1744.9	308.4	92.1	21.5	597.5	69.5	202.6	49.9	-1.9	-6.5	-2.2	-2.2
Up-regulated under P-Lim aerobic only (FC = fold change, ANA = anaerobic, AEA = aerobic)																								
10310_at	PPR1	YLR014C	YLR014C zinc-finger transcription factor of the Zn(2)-Cys(6) binuclear cluster domain type	PYRIMIDINE BIOSYNTHESIS	40.2	19.5	85.8	12.2	88.1	9	161.6	19.2	39.4	13	63.8	5.3	65.7	7.8	186.3	7.7	4.7	2.9	2.8	
10522_at	TRK2	YKR050W	YKR050W membrane protein, low affinity potassium transport	TRANSPORT	15.4	11.1	68.5	9.1	46.1	8.3	63.7	10.2	44.4	3.9	48	8.6	46.6	5.5	116.5	6.1	2.6	2.4	2.5	
11119_at		YLL030W	YLL030W similarity to hypothetical protein YKR019c	UNKNOW	42	16.8	17.9	4.8	16.2	8.2	23.6	1.7	16.8	3.4	15.8	1.8	12	1.9	47.3	20.6	2.3	2.6	3.9	
1250_at	COS10	YNR075W	YNR075W Protein with strong similarity to subtelomericly encoded proteins such as Cos10	UNKNOW	52.6	39	124	64	211.9	37	279.2	33.3	12	1.7	12.2	4.4	12	1.7	34.5	1	2.9	2.9	2.9	
3938_at	AS1	YBL069W	YBL069W Protein involved in targeting of plasma membrane H+ATPase	PLASMA MEMBRANE PROTEIN TARGETING	12	3.3	23.7	3	18.8	3.9	35.1	6.2	16.7	1.9	1.9	0.9	22.1	2.3	71.2	19.7	4.3	3.2	3.2	
4235_at	GOT1	YIL120W	YIL120W similarity to antibiotic resistance proteins	UNKNOW	24.6	5	28.2	3.1	34.9	4	49.4	1.8	23.8	4.7	7	20.7	4.7	13.5	3.4	64.0	17.5	3.1	4.8	
4322_at		YHR210C	YHR210C UDP-glucose-4-epimerase (GAL10, galE)	UNKNOW	300.3	77.5	109.1	13.1	75	4	338.3	78.4	22.8	1	61.2	4.5	47.7	12	219	89.3	9.6	4.6	3.6	
4506_at		YHR032W	YHR032W ethionine resistance protein	UNKNOW	27	5.1	53.1	12.9	37.8	4.5	93.5	4.1	33.9	16.2	35.2	5.1	34.8	19.2	138.5	79.8	4.1	4	4	
4507_at		YGR070W	YGR070W membrane protein	UNKNOW	28.1	12.5	41.3	6.7	48.1	5.5	82.3	41.9	5.5	64.7	9	78.9	19.8	296.1	50.3	3.9	4	4		
5246_at	SDT1	YGL224C	YGL224C strong similarity to hypothetical protein YMR037w	UNKNOW	66.8	24.3	257.7	46	148.4	28.8	476.9	117	140.7	28.2	269.9	21	155.6	16.7	688.8	139	2.9	4.4	4.4	
5535_at		YER186C	YER186C weak similarity to hypothetical protein YMR319c	UNKNOW	121.9	58	251.9	15.8	218.8	59.5	391.6	63.4	135	29.8	184.3	70.2	169.3	21.9	743.2	111.5	5.5	4.4	4.4	
6290_at	SHE9	YDR039W	YDR039W identified by a library screen) that causes growth arrest when overexpressed	UNKNOW	23.5	4.6	21.1	6.8	18.1	5.8	24.4	5.2	10.8	2.1	1.8	2.0	1.6	2.0	1.6	2.0	1.6	2.0	1.6	
6577_at		YDL109C	YDL109C strong similarity to thiamine-repressed protein Th4p	UNKNOW	12	2.3	14.6	2	12	2.7	21.3	15.2	13.3	1.6	1.2	1.4	1.2	3.2	3.1	15.2	2.6	2.6	2.6	
6835_at	BUD23	YCR047C	YCR047C Protein carboxyl methylase	UNKNOW	74.6	4	228.3	149.9	109.2	13.6	193.5	11.8	85.8	57.2	87.3	41.1	56.3	8.7	258.9	120	3	4.6	4.6	
6894_at	AUT4	YOL086W	YOL086W transmembrane transporter	UNKNOW	15.3	45.2	159.6	67.3	147.3	18.3	348.2	18.5	164	25.2	187.3	20	175.9	17.4	700.3	85.5	4.3	4	4	
7361_at	NTH2	YBR001C	YBR001C Neutral trehalase, highly homologous to Nth1p	TREHALOSE METABOLISM	55.6	12.5	122.7	36.6	71.6	2.8	103.7	9.3	126.5	15.3	146.5	9.6	75.2	4.3	26.6	10.9	2.2	2.2	2.2	
7427_at		YBL070C	YBL070C questionator ORF	UNKNOW	12	2.8	12	1	12	3.4	12	3.1	12	0.9	1.2	1.1	1.2	1.3	68.4	9	5.7	5.7	5.7	
7428_at	PRS4	YBL069W	YBL069W ribose-phosphate pyrophosphokinase 4	PEPTIDE PHOSPHATE CYCLE	131.8	33.9	195.9	41.4	147.3	18.3	348.2	18.5	164	25.2	187.3	20	175.9	17.4	700.3	85.5	4.3	4	4	
7611_at		YPR125W	YPR125W suppressor of mrs2-1 mutation	UNKNOW	149.2	24	139.9	23.8	146.7	21.6	185.5	47.7	185.4	27.3	188.3	16	193.5	9.4	443.2	133.7	2.4	2.3	2.4	
8187_at		YOR343C	YOR343C hypothetical protein	UNKNOW	74.4	29	49.1	15.2	57.8	3.9	146.9	21.1	74.1	13.7	29.6	6.9	13.8	9.2	212.5	111.4	2.9	2.9	15.4	
8726_at		YIL154W	YIL154W similarity to S. pombe asp1+	UNKNOW	54	17.7	319.6	9	54	17.7	319.6	9	54	17.7	319.6	9	54	17.7	319.6	9	2.1	2.1	2.1	
8862_at		YNR014W	YNR014W weak similarity to hypothetical protein YMR206w	UNKNOW	87.3	26.5	52.9	4.8	34.7	7.6	47.1	14.4	14	3	19.3	3.5	1.8	2.9	40.3	16.7	2.9	2.2	2.2	
8896_at		YNL040W	YNL040W hypothetical protein	UNKNOW	227.2	54.9	336.7	42.1	218.7	44	289	24.1	188.1	19.8	228.3	50.6	195.6	12.2	474.2	184.9	2.5	2.1	2.1	
9072_at		YNL224C	YNL224C hypothetical protein	UNKNOW	15	2.3	12.2	1.7	13.2	1.7	13.2	1.7	13.2	1.7	13.2	1.7	13.2	1.7	13.2	1.7	37	3.7	3.7	
9107_at	APFG2	YNL242W	YNL242W similarity to human hypothetical protein KIAA0404	UNKNOW	14.5	7	27.4	3.2	18.4	4.5	19.5	5.1	40.7	8	37.7	17	39.3	2.8	135	45	2.4	2.4	2.4	
9292_at		YMR194C	YMR194C hypothetical protein identified by SAGE	UNKNOW	12	4.1</																		

10262_at	YLR053C	YLR053C hypothetical protein	UNKNOW	12	6.9	73.2	8.2	12	3.7	26.3	11.1	81.8	23.4	335.2	21.8	28	3	104.1	37.9	-2.9	-12	-3.7	
10795_at	JEN1	YKL217W	TRANSPORT	68.4	20.6	27.7	8.4	12.6	3.4	16	8.9	1943.4	142.8	39.5	4.3	18.2	8.3	42	12.8	-106.8	-2.2	-2.3	
10865_at	BNM1	YJR025C	NICOTINIC ACID BIOSYNTHESIS	224.1	26	218.8	47	148.4	37.9	414.5	29	114.9	6.1	63.4	2.6	21.5	3.4	53.1	19.8	-5.4	-2	-2.5	
10908_at	YJR115W	YJR115W similarity to hypothetical protein YBL043w	UNKNOW	83.5	48.6	122.3	8.9	35.4	1.5	63.8	27.6	237.7	91.2	57.2	27.4	20.7	5.6	117.5	54.4	-11.5	-2.8	-5.7	
11052_at	YJL016W	YJL016W weak similarity to hypothetical protein YNL278w and YLR187w	UNKNOW	194.2	41.9	483.8	85.6	288.6	34.4	378.9	64.7	233.7	82.4	305.3	2.3	103.1	14.7	275.6	41.4	-2.3	-3	-2.7	
11060_at	TDH1	YJL052W	YJL052W Glyceraldehyde-3-phosphate dehydrogenase 1	2717.7	728	1568.6	287.6	1001.5	80.7	2508.8	272.9	1453	285.6	1417.2	82.6	447.6	23.3	1798.3	288.6	-3.2	-3.2	-4	
4388_at	SPS100	YKR139C	YKR139C sporulation-specific wall maturation protein	12.1	5.2	31.7	2.7	26.4	4.8	30.3	2.8	310.8	232	42.1	8.5	18.5	6.2	49	13.3	-16.8	-2.3	-2.6	
4732_at	GMD2	YGR256W	YGR256W 6-phosphogluconate dehydrogenase	21.1	10.5	40.2	2.9	24.5	5.4	40.9	7.3	316.4	69.2	26.9	5.7	12	3.1	78.8	10.7	-26.4	-2.2	-6.6	
4786_at	SOL4	YGR248W	YGR248W similar to SOL3	47.9	27.2	87.1	8	48.3	18.5	61.2	18.6	568.3	184.4	363.7	153	110.6	4.9	528.4	75	-5.1	-3.3	-4.8	
5121_at	YGL121C	YGL121C hypothetical protein	UNKNOW	345.8	6.6	337.5	34	180.8	10.4	195.5	82.5	884.9	113.3	473.1	37.8	125	17.9	330.2	77.3	-7.1	-3.8	-2.6	
5357_at	GSY1	YFR015C	YFR015C Glycogen synthase (UDP-glucose-starch glucosyltransferase)	358	60.1	236.9	29	98.5	13.2	192.7	42.9	92	13.3	68.4	29.9	25.5	3.4	53.1	6.6	-3.6	-2.7	-2.1	
5372_at	HSP12	YFL014W	YFL014W Heat shock protein of 12 kDa, induced by heat, osmotic stress, oxidative stress	681.6	335	1316.4	85.4	708.3	209.7	855.5	208.1	3088.1	342	2276.7	194.2	926.4	100.3	2357.1	345.4	-3.3	-2.5	-2.5	
5544_at	SPH1	YER150W	YER150W strong similarity to putative cell surface glycoprotein Sed5p	170.3	74.3	286.1	21.8	243.4	58.7	291.6	65.7	326.9	55.7	263.1	36.7	117.7	10.3	333.9	30.4	-2.8	-2.4	-2.8	
5690_at	EDC2	YER035W	YER035W hypothetical protein	48.4	23.2	145.3	12.4	59.2	8.1	74.4	20.8	121.2	33.8	140.1	9.4	60	5.4	140.7	58.8	-2	-2.3	-2.3	
5735_at	GLC3	YEL011W	YEL011W 1,4-glucan-6-(1,4-glucano)transferase	415.2	97.9	317.1	36.8	241.7	14	200.5	56.9	323.1	48.7	351.6	144.2	108.7	44.5	329.3	100.1	-3	-3.2	-3	
5786_f_at	PAU2	YEL040W	YEL040W member of the spo10/pepin protein/gene family (see Gene_class PAU)	614.1	107.1	276.6	42.1	305.5	8.7	1577.5	589.1	329.7	291.6	80.5	12.9	38.8	2.7	78.9	18.9	-8.5	-2	-2	
6128_f_at	HX17	YDR342C	YDR342C Hexose transporter	3228.6	606	1272.9	87.8	649.7	17.8	1460.7	173.8	2254.9	201.2	1340.2	110.7	621.9	131.8	1420.4	88.5	-3.6	-2.2	-2.3	
6129_f_at	HX16	YDR343C	YDR343C Hexose transporter	3297.3	1036.2	1591.8	124.5	784.4	38.3	1839.1	138	2086.8	527.7	1770.1	131.9	816.6	88.8	1844.5	251.5	-2.6	-2.2	-2.3	
6171_at	SLR2	YDR297W	YDR297W Syringomycin response protein 2	439.8	56.6	302.1	59.8	311.2	47.4	428.6	34.5	176.4	51.1	250.6	30.2	83	4	172.5	50.2	-2.1	-3.1	-2.1	
6384_at	YDR070C	YDR070C hypothetical protein	UNKNOW	45.5	36.7	39.4	0.8	24.7	9.1	23.7	4	1056	188.3	112.5	30.2	16.3	3.1	73.1	4.4	-64.8	-6.9	-4.5	
6483_at	DM3	YDL024C	YDL024C strong similarity to acid phosphatase	19.9	4.9	54	15.7	15.2	1.6	84.1	10.5	42.9	7.7	42.8	2.1	19	6.2	38	4.7	-2.4	-2	-2	
6486_at	CPM2	YDL021W	YDL021W Similar to CPM1 (phosphoglycerate mutase)	174.6	60.6	203.3	30.7	103.5	11.1	269.8	29.3	66.2	5.6	125.7	7.6	26.2	9.2	80.8	12.9	-4.6	-3.5	-3.5	
7324_at	YBR058W	YBR058W Homolog to glucan-1,3-glucosidase (EC 3.2.1.5); S. cerevisiae) 2	UNKNOW	49.9	19.8	128.8	27.6	40.6	10.5	57.4	2.6	68.2	12	74	15.4	31.8	13.1	65.4	7.9	-2.1	-2.3	-2.1	
7604_at	CPH1	YPR180W	YPR180W Glycogen phosphorylase	450.9	71.7	441.3	43.3	77.1	9.8	267.1	44.9	303.6	44.3	431.6	100.8	35	2.4	393.6	33.3	-8.7	-12.3	-11.2	
7610_at	CTR1	YPR124W	YPR124W High affinity copper transporter into the cell, probable integral membrane protein	114.2	26.1	76.1	9.7	67.8	15.7	64.9	5.7	277.3	42.6	358.2	81.8	80.8	8.6	183	20.7	-4.6	-3	-3	
7831_at	PDR12	YPL058C	YPL058C multidrug resistance transporter	70.4	31.6	114.3	19.8	61.7	6.9	65.9	22.6	637.9	155.7	523.1	92.3	217.5	15.3	467.6	55.3	-2.9	-2.4	-2.1	
7843_at	SSU1	YPL092W	YPL092W sensitive to sulfite	217.5	41	238	45.2	84.5	8.3	308.5	30.4	448.7	149.5	287.5	10.9	60.1	4.7	193.8	21.3	-7.5	-4.8	-3.2	
7926_at	UUP4	YPL186C	YPL186C weak similarity to Xenopus protein xlp7	34.9	24.1	69.9	22.7	24.2	4.9	29.2	4.6	316.2	73.7	143.8	13.5	23.2	0.2	47.8	4.7	-13.7	-6.2	-2.1	
7974_at	USV1	YPL230W	YPL230W Up n Star/Vation	51.9	33.8	43.9	8.1	23.9	5	13	2.4	157.5	30.4	53.1	21.8	21.2	2.2	50.8	12.7	-7.4	-2.4	-2.4	
8186_at	TYE7	YDR344C	YDR344C TYE7, a 33 kDa serine-rich protein, is a potential member of the basic region/helix-loop-helix protein family	33.5	14.9	38.6	14.8	35.4	7.7	93.8	26.2	41.2	2	36.4	5	12	1.8	59	18.8	-3.4	-3	-4.9	
8192_at	PUT4	YOR348C	YOR348C putative protease-specific peptidase	18.9	6.6	176	26.5	28.1	6.5	21.7	8.9	1282.8	181	2557.3	204	138.5	44.6	518.2	179.5	-9.1	-16.5	-3.7	
8375_at	YOR173W	YOR173W strong similarity to YLR270W	UNKNOW	148.7	33.2	183	22.9	123.7	24.5	114	16.1	452.6	55.2	280.8	48.9	107	9.3	226	29.3	-4.2	-2.6	-2.1	
8599_at	YOL053C	YOL053C DNA Damage Responsive	UNKNOW	678.1	212.9	817.1	71.6	352	51.2	419	65.2	2502.8	1023.5	1288.4	124.2	274.9	17.6	2762.2	730.5	-10.6	-4.7	-10	
8693_at	MDH2	YOL126C	YOL126C cytosolic malate dehydrogenase	69.4	31.3	108	17.5	79.8	5.2	73.5	17	725.4	124.9	150.6	12.9	56.2	4.7	136.3	18.4	-2.4	-2.4	-2.4	
8727_at	SOL1	YOL153C	YOL153C strong similarity to Cps1p	12	2.2	26.4	4.9	12	2.3	12	1.7	31.9	2.9	27.9	3.1	12.5	1.8	29.6	11.5	-2.5	-2.4	-2.4	
8796_at	YMR034W	YMR034W hypothetical protein	UNKNOW	928.9	166.7	465.9	61.1	144.5	16.2	362.3	76.4	835.7	234.4	233.6	20.4	42.3	3.1	92.4	6.7	-19.8	-5.5	-2.2	
9341_at	PGM2	YMR291W	YMR291W similarity to serTfr protein kinase	32.5	18	59.6	5.5	46.5	3.4	70.5	13.1	135.3	14	83	5.7	18.1	2.4	51.5	9.3	-4.6	-2.6	-2.6	
9380_at	CAD1	YMR251W	YMR251W similarity to serTfr protein kinase	112.9	38.5	62.1	5.6	44.3	0.7	106.2	22.9	104.2	33.6	42.5	14.8	13.7	1	89.2	13.5	-7.6	-3.1	-6.5	
9547_at	PGM2	YMR250W	YMR250W similarity to glutamate decarboxylases	6	20.3	6	56	14	22.5	3.2	28	4.9	47.8	16.7	64.5	18.3	13.4	1.5	105.7	9.3	-3.6	-7.9	-4.8
9743_at	TSL1	YMR105C	YMR105C Phosphoglucomutase	326.4	156.4	237.1	42	120.5	17.4	197.9	57.9	325.8	4.9	214.1	84	47.5	6	221	28.4	-6.9	-4.5	-4.7	
9802_at	CTR3	YML100W	YML100W 123 kD regulatory subunit of trehalose-6-phosphate synthase/phosphatase con	92.9	13	221.5	2.6	116.5	25.2	147.7	18.8	281.8	25.3	322.8	108.1	115.2	9.7	276.9	15.2	-2.3	-2.8	-2.4	
		YLR411W	YLR411W Copper Transporter	61.7	22	40.9	9.2	23.4	5.5	34.9	12	1039.9	139.1	439	29.3	26.6	5.5	345.3	57.2	-39.1	-16.5	-13	

Up-regulated under C-Lim Anaerobic only (FC = fold change; ANA = anaerobic; A = aerobic)			FC C-Lim ANA vs N-Lim ANA C-Lim ANA vs S-Lim ANA FC C-Lim ANA vs P-Lim ANA																		
Probe Set	Gene name	systematic name	cellular function	C-Lim ANA Average	C-Lim ANA S/Slide	N-Lim ANA Average	N-Lim ANA S/Slide	P-Lim ANA Average	P-Lim ANA S/Slide	S-Lim ANA Average	S-Lim ANA S/Slide	C-Lim A Average	C-Lim A S/Slide	N-Lim A Average	N-Lim A S/Slide	P-Lim A Average	P-Lim A S/Slide	S-Lim A Average	S-Lim A S/Slide		
10253_at	ATP15	YL0909C	YL0909C strong similarity to alanine transaminases	12552	256.5	534.6	33.3	448.5	77	442.6	62.8	805.2	11.4	1008.2	67.2	582.6	50.5	633.7	59.1	2.3	
10290_at	C0X12	YL0308C	YL0308C subunit VI of cytochrome c oxidase	11686	252.6	386.2	105	367.7	44.8	436	54	2607.4	46.2	1928.2	20.5	2092.8	29.2	1956.4	168.5	3	
10390_at	SDH2	YL0416C	YL0416C Succinate dehydrogenase (ubiquinone) iron-sulfur protein subunit	11013	227	480.5	49.1	523.8	106.6	503.4	2.14	2171.7	48.4	1282.2	37.7	1016.4	208.4	1875.7	206.8	2.3	
10725_at	SDH1	YL0415C	YL0415C Flavoprotein subunit of succinate dehydrogenase	273.8	52.2	117.1	24.7	82.7	12.9	108.5	103.9	200.5	35.5	207.4	20.5	312.5	49.8	20.5	312.5	49.8	2.3
10860_s_at	CY116	YJR158W	YJR158W hexose transporter	84.2	38.3	12	2.3	12	6.8	12	2	13.5	6.1	12	3.1	12	3.6	15.7	3.5	7	
10977_at	CYC1	YJ0484W	YJ0484W subunit I of cytochrome c oxidase	179.8	52.8	12	2.5	12	1.2	12	5.6	1608.9	203.9	290.9	33.8	1248.8	17.4	1611.3	155	15	
11264_at	CYC21	YAR075W	YAR075W strong similarity to IMP dehydrogenases	1465.7	396.1	477.3	70.2	512.7	69.1	433.9	162.4	348.5	109.7	556.1	24.7	416.8	55.8	484.9	97.2	3.1	
4149_at	MAM33	YJL070C	YJL070C mitochondrial oxidase matrix protein	523.1	5.9	192.6	6.2	230.1	4.2	254.2	85	488.7	3.9	385.9	61	304.8	17.4	488.1	59.8	2.7	
43521_at	IMD2	YHR216W	YHR216W IMP dehydrogenase; low affinity P1UR5 gene	1390.9	164.9	379.3	28.1	398.3	62.3	261.7	28	262.2	50.9	596.4	30.9	387.6	15.6	411.4	62	3.7	
4481_at	C0X6	YHR051W	YHR051W subunit VI of cytochrome c oxidase	663.7	42.4	277.4	47.9	292	31.8	290.3	93	1484.6	252.7	1031.6	84.3	1191.2	69.4	1234.6	140.3	2.3	
4516_at	OSH7	YHR010W	YHR010W 8.5 kDa subunit of the ubiquinol-cytochrome c oxidoreductase complex	305.6	52.1	94.6	23.4	77.3	5.1	83	9.4	799	60.7	629.9	21.1	766.1	91.3	699.3	114.9	3.2	
4522_at	SPA11	YHR035C	YHR035C May act cooperatively with Mds30 in mitochondrial protein import or other related essential MITOCHONDRIAL PROTEIN TARGETING	853	86	392.2	78.9	400.7	11.9	338.3	30	775.9	63.7	614.8	60.5	626.6	8.7	544.2	55.8	2.1	
5000_s_at	RP525A	YGR027C	YGR027C Ribosomal protein S25A (S21A) (p45) (Y523)	801.3	25.3	319.9	11	341.5	92.7	312.8	44.3	562.6	39.7	485.1	12.7	476.4	51.7	412.9	99.6	2.6	
5142_s_at	RP51A	YGL147C	YGL147C Ribosomal protein L5A (L5A) (p24) (Y111)	414.8	96.8	186.8	39.4	186.3	47.2	162.4	23.4	234	24.1	42.5	194.4	44.8	223.8	59.8	197.3	64.6	2.2
5178_at	SUT11	YGL162W	YGL162W Involved in sterol uptake	188.4	14.5	73.3	7.5	75.0	19.2	67.4	9.4	37.5	8.7	20.6	3.8	37.3	12	17.3	2.4	2.2	
5183_at	C0X13	YGL191W	YGL191W subunit VIa of cytochrome c oxidase; may specifically interact with ATP	644.9	49.7	172.2	23.3	280.1	41.3	309.5	66.5	945.9	65.6	786.9	24.8	801.4	17.2	750.7	34.3	3.7	
5192_at	C0X4	YGL197C	YGL197C subunit W of cytochrome c oxidase	598.5	44.6	142.4	16.2	138.7	9.9	151.3	41.2	1225.6	116.1	888.9	48.7	938.7	40.1	1026.2	78.3	4	
5332_at	ORP8	YFR033C	YFR033C ubiquinol-cytochrome c oxidoreductase subunit 6 (17 kDa)	885.8	87.6	314.8	46.9	386.9	13.9	307.5	53.4	1371.8	40.7	1101.7	113.9	41.6	1271	89.6	2.8	2.3	
5786_at	OCR7	YJL024W	YJL024W Rieske iron-sulfur protein of the mitochondrial cytochrome bc1 complex	692.1	66.5	201.9	78.4	287.7	3.4	334.3	121	1615	197.5	1136.3	59.8	1323.2	69	1270.6	205.3	3.4	
6052_at	HKT7	YDR342C	YDR342C hexose transporter	1450.8	374.7	609.6	68.7	516.7	84.4	474.8	34.2	1894.1	277.6	1475.6	38.9	1930.8	61.2	1555.9	9	2.4	
6128_at	SDH4	YDR178W	YDR178W succinate dehydrogenase membrane anchor subunit	3229.6	606	1229.9	97.8	649.7	17.8	546.0	173.6	2254.9	201.2	1340.2	110.7	621.9	131.8	1420.4	88.5	2.5	
6098_at	PRP1B	YOL130W	YOL130W ATPase stabilizing factor	1131	91.6	534.1	18.9	387.3	22	532.6	16.9	1688.7	62.2	972.8	110.1	747.1	137.1	1317.1	29	2.1	
6646_at	DL11	YOL174C	YOL174C mitochondrial enzyme D-lactate ferrioxalicytochrome c oxidoreductase	381.5	102.1	436.3	43	394.9	12.6	441.7	41	145	198.8	596.2	43.4	410.7	18.8	747.3	79.5	2.4	
6854_at	NH1	YOL191W	YOL191W ATPase inhibitor	608.7	6.4	209.1	8.8	188.9	35.8	198.2	13.2	921	117.8	345.6	24.8	583.3	78.7	682.4	116.1	2.9	
6907_at	AGP1	YOL025C	YOL025C (AGP1) Broad substrate range amino acid permease with high affinity for asparagine and arginine	352.9	102.6	102	17.3	41.1	25	83.2	4.4	798.8	152.7	516.7	60.5	370.8	113.2	706.3	164.1	3.5	
7344_at	RLP4A	YBR013W	YBR013W Ribosomal protein L4A (L2A) (p52) (Y2)	782.9	56.7	155.9	23.1	363.4	68.5	346.5	31.2	821	28.6	208.6	17.3	204.4	116.9	353.1	52.1	2.2	
7345_at	OCR2	YBR019W	YBR019W 8.5 kDa ubiquinol-cytochrome c oxidoreductase core protein 2	389.7	84.4	157.9	53.4	109.2	36.8	95.9	207	196.9	61.3	194.2	15.6	155.8	35.3	159.8	51.5	3.4	
7727_at	ATP2D	YPR020W	YPR020W hypothetical protein	191.6	19.6	78.6	10.8	78.6	10.8	30.8	6.1	28.8	11.3	17.3	16	184.6	21.4	187.4	40.4	2.3	
7812_at	ATP4	YPL078C	YPL078C (F1F0)-ATPase complex delta subunit, mitochondrial	539.2	100.2	240.5	67	252.3	15.3	298	131	813.3	143.5	800.3	28.2	653.3	61.8	551.6	38.2	2.1	
8024_at	ATP15	YJL271W	YJL271W ATPase for ATP synthase epsilon subunit	174	21.7	114.5	11.6	116.5	8.7	116.5	18.2	1209.8	64.3	804.3	43.4	809.7	110.3	929	37.3	2.4	
8289_s_at	RP510A	YOR029W	YOR029W Ribosomal protein S10A	1923.9	133.3	912.9	220.7	930.2	187.1	736.3	183.7	1396	206.2	1008.3	212.9	1119	154.4	975.5	242.1	2.3	
8554_at	ND1	YLR036C	YLR036C mitochondrial cytochrome c oxidase chain Va	321	14.8	61.6	7.2	66.2	11.6	61.6	18.2	162.2	16.2	162.2	16.2	162.2	16.2	162.2	16.2	162.2	16.2
9800_at	ND1	YMR145C	YMR145C mitochondrial cytochromically reduced NADH dehydrogenase	115.7	23.4	45.7	6.1	28.7	4.4	40.9	3.4	1257.3	68.1	633.8	99.7	808.8	62.4	1153.7	124.9	2.5	
9830_at	C0X8	YLR395C	YLR395C Cytochrome c oxidase chain VIII	1223.3	130.2	517.3	59.9	475	56.4	494.2	65.7	2121	160.1	1631.7	19.3	1785	71.4	1665	112.4	2.6	
Down-regulated under C-Lim Anaerobic only (FC = fold change; ANA = anaerobic; A = aerobic)			FC C-Lim ANA vs N-Lim ANA C-Lim ANA vs S-Lim ANA FC C-Lim ANA vs P-Lim ANA																		
Probe Set	Gene name	systematic name	cellular function	C-Lim ANA Average	C-Lim ANA S/Slide	N-Lim ANA Average	N-Lim ANA S/Slide	P-Lim ANA Average	P-Lim ANA S/Slide	S-Lim ANA Average	S-Lim ANA S/Slide	C-Lim A Average	C-Lim A S/Slide	N-Lim A Average	N-Lim A S/Slide	P-Lim A Average	P-Lim A S/Slide	S-Lim A Average	S-Lim A S/Slide		
10012_at	URC2	YLR297W	YLR297W weak similarity to Vibrio vulnificus VvpC protein	53.1	17.2	143.9	15	115.5	8.5	161	60.1	119.8	43.3	83.1	11	75.2	7.3	107.5	17.9	-2.7	
10048_at	UBF12	YLR338W	YLR338W Ubiquitin-conjugating enzyme	28.3	15	75.3	6.1	75.3	6.1	80	7.3	75.3	6.1	75.3	6.1	75.3	6.1	75.3	6.1	75.3	-3.2
10103_at	FRE1	YLR211C	YLR211C ERG1 Protein involved in palmitoylation and localization of Ras2p	30.4	27.8	212.5	38.1	96.3	13.1	87.8	7.8	8	10.6	17.7	27	10.3	12.4	97.5	11.5	-4.9	
10118_at	BUR2	YLR228W	YLR228W hypothetical protein	30.2	12.5	73.5	12.8	112.4	19.1	95.8	24.9	68.8	5.3	68.5	16.2	69.2	7.5	52.4	3.3	-3.7	
10129_at	ADY4	YLR227C	YLR227C hypothetical protein	17.8	12.2	38.3	15.8	53.8	8.9	43.7	14.7	34.1	3.9	46	4.7	62.5	12	57.3	14.8	-3	
10147_at	SRN2	YLR119W	YLR119W suppressor of mat1 mutation	21	8.5	78.1	17.8	42.8	7	44.7	2.2	44.8	4.7	60.6	7	36.5	5.2	36.6	13.9	-2	
10217_at	CH4A	YLR088C	YLR088C DNA-binding transcriptional activator or CHA1	17.5	7.5	49.6	19.1	44.3	4.4	36.8	2	26.6	3	44.5	19.7	38	4.7	35	29	-2.5	
10280_at	YLR051C	YLR051C similarity to human acidic 62 kDa protein	UNKNOWN	51.1	32.7	104.7	44	133.9	14.6	121.4	13.3	102.5	25	157.8	38.8	178	34.8	125.9	24	-2.1	
10377_at	YLR053C	YLR053C similarity to C. elegans and M. marneffei hypothetical proteins	UNKNOWN	40.7	19	114.5	21	116.5	28.4	107.6	47.7	176	47.7	176	47.7	176	47.7	176	47.7	-2.6	
10444_s_at	MMN4	YK0210C	YK0210C regulates the mannosyl phosphorylation	7.5	23.3	157.4	12.9	172.9	33.2	171.5	47.5	142.4	136.8	348.8	86.7	2716.5	272.8	2125.6	758.2	-2.3	
10465_s_at	PRP16	YKR086W	YKR086W putative ATP-binding protein	21	6.8	56	7	56	7	56	7	56	7	56	7	56	7	56	7	56	-2.3
10485_at	LAS21	YKR104W	YKR104W similarity to regulating expression genes	22.1	10.9	46.2	7.8	52.5	13.4	63	28.4	50.1	16.9	34.1	3.1	32.1	2.9	30.7	9.7	-2.8	
10488_at	LAS1	YKR036C	YKR036C similarity to multiple resistance genes involved in bud formation and morphogenesis	40.1	21	199.2	53.2	161.6	45	313.7	45.3	106.7	37.4	284.8	89.8	176.2	103	168	47.8	-4.7	
10522_at	TKC2	YKR060W	YKR060W membrane protein; low affinity potassium transport	25.2	13.3	58.6	17.7	65.4	11.3	56	11.3	56	11.3	56	11.3	56	11.3	56	11.3	56	-2.8
10531_at	YKR060W	YKR060W hypothetical protein	UNKNOWN	12	6.1	42	8.9	32.1	8.4	31.4	2.4	51.4	5.6	47.1	9.1	2.4	52	45.6	6.3	-2.7	
10540_at	SPK190	YKR029W	YKR029W membrane protein; low affinity potassium transport	38.7	14.3	82	17.3	115.5	20.2	89.5	27.7	57.7	5.5	101.6	46	117.7	47				

Accession	Gene name	Systematic name	Cellular function	C-Lim ANA	C-Lim ANA	N-Lim ANA	N-Lim ANA	P-Lim ANA	P-Lim ANA	S-Lim ANA	S-Lim ANA	C-Lim ANA	C-Lim ANA	N-Lim ANA	N-Lim ANA	P-Lim ANA	P-Lim ANA	S-Lim ANA	S-Lim ANA	FC N-Lim ANA vs C-Lim ANA	FC N-Lim ANA vs S-Lim ANA	FC N-Lim ANA vs P-Lim ANA
				Average	Stdev																	
4866_at	MEP1	YGR121C	YGR121C ammonia permease	204.5	19.2	508.7	16.8	91	37.9	153.4	23.9	308.3	59.2	236	14.7	153.5	6.5	238.2	29.1	2.5	5.6	3.3
4870_at	ROM1	YGR070W	YGR070W similarity to S.pombe hypothetical protein SPAC24H6.11c	106.6	17.9	606.8	109.5	178.9	24.3	140.8	17.6	112.9	10.2	724.6	96.1	415.8	69.5	236.1	46.9	5.7	3.4	2.7
4951_at		YGR070W	YGR070W GDP-GTP Exchange Protein (GEP) for the Rho p Small GTP-binding Protein	84.2	38.7	225	89.3	75.3	12.8	72.9	20.5	93.1	15.4	19.9	12.2	51.7	1.9	74.5	22.2	5.7	3.8	4.3
5146_c		YLR041C	YLR041C weak similarity to Lacisobas guanine histidine kinase Sypk	25	10.3	108.2	20.1	108.2	4.3	108.2	4.3	108.2	4.3	108.2	4.3	108.2	4.3	108.2	4.3	2.7	2.7	2.7
5537_at		YER185W	YER185W strong similarity to Rlm1p	12	3.2	35.7	1.8	17.4	1.5	14.8	6.2	22.1	2.6	19.9	7.7	20.6	2.1	17	8.6	3	2.1	2.4
5634_at	BR073	YR0404W	YR0404W hypothetical protein	52.7	20.8	107.7	28.4	5.1	5.6	41.9	11.6	75.9	17.2	61.1	8.4	80.4	9.7	37.8	9.6	2	2.1	2.6
5616_at	HPA3	YLR069W	YLR069W Histone and other Protein Acetyltransferase; Has sequence homology to known HATs	447.6	73.6	1050.2	121.2	447.6	87.6	361.4	51.2	522.7	31.3	1048.7	111.3	362.7	12.4	526.3	151.7	3	2.1	2.4
5944_at		YDR252C	YDR252C weak similarity to transcription factors of the zinc finger class	37.7	16.2	165.8	39.7	74.6	14.2	73.6	15.4	53.3	10.6	189.2	22.3	79.7	11.7	101.8	19.3	4.4	2.2	2.3
6093_at		YDR311C	YDR311C strong similarity to hypothetical protein YGR012w	47.7	11.1	139	14.5	65.3	7.5	51.2	7.1	52.6	3.5	114.4	19.4	47.1	13.6	81.7	20.9	2.9	2.1	2.1
6722_at	BR4	YDL231C	YDL231C (BR4) Protein of unknown function. Has a putative zinc finger domain	69.9	26	264	3.4	108.3	4.7	101.4	10.4	101.1	14.5	264.4	19	165.1	10.5	153.3	7.3	2.6	2.6	2.6
7138_at	SLX1	YBR228W	YBR228W similarity to hypothetical A.thaliana protein	17.9	6.2	38.4	12.2	18.6	2.6	15	4.8	21.3	1.9	41.9	4.6	38.1	5.4	30.3	9.8	2.1	2.1	2.6
7145_at		YBR225W	YBR225W similarity to sumatrade-sensitive Ndc10 cotransport protein	45.8	7.9	178.4	29.6	7.7	3.9	55.7	11.7	71	4.3	191.1	6	98.8	10.2	91.6	17.5	3.9	2.2	3.2
7219_at	APG14	YBR128C	YBR128C required for autophagy	12	5.8	31.4	4.5	15.6	2.1	13.2	3.3	21.9	0.9	36.9	1.7	19.2	2.2	24.5	1.7	2.6	2.4	2.4
7324_at		YBR056W	YBR056W Homolog to glucan-1,3-glycosylase (EC 3.2.1.5); S. cerevisiae 2	49.9	19.8	128.8	27.6	40.6	10.5	57.4	2.6	68.2	12	74	15.4	31.8	13.1	65.4	7.9	2.6	3.2	2.2
7528_at	UP4	YLR186C	YLR186C weak similarity to Xenopus protein xip7	34.9	24.1	89.9	22.7	24.2	4.9	29.2	4.6	316.2	73.7	143.8	13.5	23.2	0.2	47.8	4.7	2.5	3.6	3.3
7891_at	LEA1	YPL213W	YPL213W (LEA1) Component of the U2 snRNP complex similar to human U2a' protein. Involved in mRNA SPLICING	37.2	13.5	138	35.4	57.9	2.8	56	11.2	39	12.9	148.7	20.3	90.4	4.6	68.1	14.8	2.7	2.4	2.5
8030_at	DPS	YPL265W	YPL265W isocitobate amino acid permease	598.1	48.4	1576.8	191.1	497.3	36.9	332.6	62.2	28.1	9.7	998.8	44.4	446.0	71.1	13.2	2.6	2.8	2.9	3.2
8037_at	THI2	YPL258C	YPL258C similar to B.subtilis transcriptional activator tenA, and strong similarity to hypothetical protein THIAMINE METABOLISM (PUTATIVE)	40.1	16.9	177.9	26.4	48.7	6.2	84.7	10.1	45.1	9.9	77.7	6.2	44.3	4.7	58.2	16.8	4.4	3.4	2.1
8207_at	FAA1	YOR317W	YOR317W long chain fatty acyl-CoA synthetase	25.6	17.5	86.6	39.5	19	1	22.6	3.8	370.9	64.3	516.9	93.3	308.7	47.1	311	45	4.4	4.6	3.8
8551_at	PHO80	YOL001W	YOL001W negative transcriptional regulator	42.2	15.7	113.1	24.8	51.6	5.6	54.9	6.6	51.8	3.5	79.9	10.4	54.2	2.1	110.8	33	2.2	2.1	2.1
8727_at		YOL153C	YOL153C strong similarity to Csp1p	12	2.2	26.4	4.9	12	2.3	12	1.7	31.9	2.9	27.9	3.1	12.5	1.8	29.6	11.5	2.2	2.2	2.2
9114_at		YNL244W	YNL244W weak similarity to mouse hemoglobin zeta chain	12	2	26	5.6	12	4.1	12	2.4	12	1.2	52.7	6.8	20.3	1.6	40.5	1.6	2.2	2.2	2.2
9388_at		YMR250W	YMR250W similarity to glutamate decarboxylase	20.3	6	56	4.4	22.5	3.2	28	4.9	47.8	16.7	64.5	18.3	13.4	1.5	105.7	9.3	2.8	2.5	2.1
9466_at		YMR155W	YMR155W weak similarity to E.coli hypothetical protein f402	18.2	9.3	74.7	13.2	33.4	5	27.4	3.6	45.6	2.6	95.6	1.5	64.4	4.9	32.1	9.9	4.1	2.7	2.7
9482_at	ALD3	YMR186C	YMR186C Aldehyde Dehydrogenase (NAD(P)+)	63.1	28.6	352.1	90.7	141	31.7	147	22.6	228.7	62.7	307.4	27.1	127.7	17.4	145.6	3.8	5.6	2.5	2.4
9515_at		YMR118C	YMR118C strong similarity to succinate dehydrogenase	16.5	11.2	56.3	12.4	23.9	3.5	22.7	4.7	325.4	181	17.4	2.4	12	3.7	12	6.2	3.4	2.4	2.4
9518_at	AS1	YMR119W	YMR119W questionable ORF	34.9	10.1	71.9	19.4	34.3	2.2	28.2	3.4	55.3	9.3	66	9.3	38.8	2.3	39.1	8.7	2.1	2.5	2.5
9549_at		YMR107W	YMR107W hypothetical protein	18.8	15	73.2	10.5	21.7	7.2	18.4	4.5	262.7	320.3	12	1.5	15.9	1.3	12	4.1	3.9	3.1	3.8
Down regulated under N-Lim anaerobic only (FC > fold change, ANA = anaerobic, A = aerobic)																						
Probe Set	Gene name	Systematic name	Cellular function	C-Lim ANA	C-Lim ANA	N-Lim ANA	N-Lim ANA	P-Lim ANA	P-Lim ANA	S-Lim ANA	S-Lim ANA	C-Lim ANA	C-Lim ANA	N-Lim ANA	N-Lim ANA	P-Lim ANA	P-Lim ANA	S-Lim ANA	S-Lim ANA	FC N-Lim ANA vs C-Lim ANA	FC N-Lim ANA vs S-Lim ANA	FC N-Lim ANA vs P-Lim ANA
10658_at	TEF4	YKL051W	YKL051W Translation elongation factor EF-1gamma	83.3	30	29.5	13.8	112.9	12.9	190.1	36	97.2	34.6	51.4	21	85.3	7.6	122.3	47.4	-3.8	-6.4	-4.4
10613_at	DLP3	YEL071W	YEL071W strong similarity to Rho1p	321.2	44.8	527.4	53.7	811.6	64.1	591.7	21.4	223.7	4.6	184.7	34.7	735.5	102.1	289.4	82	2.8	2.4	2.4
6415_at	BAP3	YDR046C	YDR046C Valine transporter	113.1	34.6	12	2.6	357.2	86.6	127.1	3.9	17.0	2	0.6	281	51.2	161	25.2	-9.4	-10.6	-10.6	
6502_at	ASG1	YDR025C	YDR025C similar to E.coli broad specific range amino acid permease with high affinity for asparagine and arginine	56.7	18.9	306.5	42.1	365.5	8.7	362.6	10.1	362.6	10.1	362.6	10.1	362.6	10.1	362.6	10.1	4.0	-2.3	-2.3
7845_at		YPL170W	YPL170W similarity to C.elegans Lim homeobox protein	714.7	106.1	322.3	105.8	695.5	57.4	657.9	44.3	539	11.2	200.4	24.7	294	16.9	340.6	30.2	-2.2	-2	-2
8587_at	TAT2	YOL020W	YOL020W Tryptophan permease. high affinity	68.7	30.4	12	1.2	77.2	6.7	84.8	7.3	48.1	5.5	36.3	2.3	78.2	10.5	105.6	21.3	-6.2	-6.4	-6.4
Up-regulated under P-Lim anaerobic only (FC > fold change, ANA = anaerobic, A = aerobic)																						
Probe Set	Gene name	Systematic name	Cellular function	C-Lim ANA	C-Lim ANA	N-Lim ANA	N-Lim ANA	P-Lim ANA	P-Lim ANA	S-Lim ANA	S-Lim ANA	C-Lim ANA	C-Lim ANA	N-Lim ANA	N-Lim ANA	P-Lim ANA	P-Lim ANA	S-Lim ANA	S-Lim ANA	FC P-Lim ANA vs C-Lim ANA	FC P-Lim ANA vs S-Lim ANA	FC P-Lim ANA vs N-Lim ANA
10022_at	GAD1	YLR037W	YLR037W Chain decarboxylase	45.1	19.5	49.3	12.8	48.3	10.6	2.8	25.6	7.2	3.8	12	0.1	7.2	2.5	2.2	2.6	2.2	2.6	2.6
10294_at		YLR042C	YLR042C hypothetical protein	27	6.1	29.6	2.4	23.2	4.5	65.1	7.6	30.9	11.3	16	3.4	12.2	3.1	12	1.7	2.4	2.8	2.8
10442_s_at		YOL200C	PROTEIN GLYCOSYLATION	12.3	6.0	12	3.8	12	3.2	5.9	2.2	3.2	2.2	6.8	2.2	6.8	2.2	6.8	2.2	2.4	2.4	2.4
10443_s_at	FLO10	YKR102W	YKR102W protein with similarity to flocculation protein Flo1p	16.5	9.4	65.4	6.1	30.8	5.9	380	4.7	25.9	6.8	37.2	6.9	23.3	1.9	35.3	5.6	5.8	12.3	12.3
10853_at	DNA1	YJR151C	YJR151C similarity to mucin proteins. YNL224c Stp1p	560.6	298.8	701.2	22.9	618.5	55.3	1551.8	341.6	111.6	29.8	115.2	27.7	96.3	6.4	132.6	58.3	2.8	2.2	2.5
10855_at	FOU4	YJR155W	YJR155W Endo-polygalacturonase	12	3.2	19	1.7	12	1.2	21.9	1.2	1.4	3.1	1.4	1.2	1.5	1.2	1.5	1.5	2.6	2.6	2.6
11186_at		YAL152W	YAL152W questionable ORF	23.2	5.3	25.5	6.6	38.8	6.1	15.2	18.7	6.9	15.5	1.8	24.3	4.3	18.2	8.4	3.7	2.5	2.5	2.5
11304_at		YAR061W	YAR061W putative pseudogene	12	4.5	13	2.2	12	3.2	38.8	11.9	12	2	12	5.7	12	0.8	12	4.7	3.2	3.2	3.2
11305_s_at		YAR061W	YAR061W putative pseudogene	12	0.9	14	6.9	12	1.3	76.1	19.3	12	1.7	12	0.4	12	0.7	12	2.2	6.3	6.3	6.3
4386_at	AR09	YHR137W	YHR137W aromatic amino acid aminotransferase II	13.8	6.7	23.2	2.8	21.5	5.7	147.2	17	314.6	21	19.6	6	12	3.9	172.6	36.2	3.2	6.4	6.8
4430_at		YHR128C	YHR128C hypothetical protein	25.9	10.7	15.8	4.7	17.3	2.3	54.7	16.7	87.8	65.9	12.2	12.8	7.4	20.1	12.8	2.8	3.2	3.2	3.2
4593_at		YHL046C	YHL046C weak similarity to members of the Stp1p/Tip1 family	1321.7	414.8	737.2	126.9	1128.7	78.3	2915.5	897	153.2	61.1	88.3	7.5	74.2	10	137.1	10.1	2.2	2.6	2.6
5537_at		YER188W	YER188W hypothetical protein	101.8	30.2	44.8	2.7	212	28.7	95.6	71.3	16.2	6.6	47.2	12.8	43.8	6.2	3				

**Two-dimensional Transcriptome Analysis in Chemostat Cultures:
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