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The Netherlands

High-dimensional imaging of the HPV-induced tumor microenvironment to improve clinical outcomes

Abdulrahman, Z.

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The background of the entire page is a deep blue space filled with a complex network of thin, glowing lines in various colors (blue, green, orange, purple) that represent the cosmic web. Scattered throughout this network are numerous stars and star clusters, some appearing as bright, multi-pointed starbursts in colors like orange, green, and blue, while others are smaller, distant points of light.

PART I

IMMUNOTHERAPY AND TME OF HPV-INDUCED PRECANCER

Chapter 2

The immune microenvironment in vulvar (pre)cancer: review of literature and implications for immunotherapy

Ziena Abdulrahman, Kim E. Kortekaas, Peggy J. de Vos van Steenwijk,
Sjoerd H. van der Burg, Mariette I.E. van Poelgeest

Expert opinion on biological therapy 2018

Abstract

Introduction

Vulvar squamous cell carcinoma (VSCC) develops via two different pathways: *TP53* mutations in a background of lichen sclerosus or a persistent infection with high risk human papilloma virus (HPV). The latter group of tumor responds better to treatment than the non-virally induced VSCC. This may be explained by a difference in the tumor immune microenvironment (TME).

Areas covered

This review summarizes literature on TME of VSCC and its precursors, and extrapolates this to foster the development of new therapeutic strategies.

Expert opinion

Both types of VSCC and their precursors are infiltrated with variable numbers of M2 macrophages, regulatory T cells and CD8+ T cells, indicating that they express targetable tumor antigens. Type 1 T cell immunity in precursor lesions is associated with fewer recurrences and better clinical responses to immunotherapy. Escape of these lesions and progression towards VSCC is associated with the downregulation of HLA class I, increased expression of co-inhibitory molecules, infiltration with immunosuppressive cells and the local production of immunosuppressive enzymes and cytokines.

More in-depth studies of the VSCC TME are required to fully comprehend the impact of the immune system on VSCC, and subsequently to identify patients who will benefit from immunotherapeutic strategies.

Key findings

- Two distinct types of VSCC can be identified: HPV-induced and *TP53* mutated VSCC
- VSCC and its precursor are infiltrated with M2 macrophages, Tregs and CD8+ T cells
- A type 1 immune response correlates with better clinical outcomes in vHSIL
- Analysis of the microenvironment is important for immunotherapeutic strategies in VSCC

Introduction

Vulvar cancer is a rare disease, representing 3-5% of all gynecological malignancies, with an increasing annual incidence of 1-2 per 100.000 women.^{1,2} Around 90% of all vulvar tumors are vulvar squamous cell carcinomas (VSCC).³ Two independent pathways have been identified in the pathogenesis of VSCC (**Figure 1**). Classical development of VSCC is related to *TP53* mutations (HPVnegVSCC) accounting around 80% of all VSCC.⁴ Persistent infection with high risk human papillomavirus (hrHPV) causes the other 20% (HPVposVSCC).^{5,6} The overall incidence of VSCC is rising, likely due to higher life expectancy and the overall increase in HPV infections.⁷ The standard treatment for VSCC consists of surgery and/ or radiotherapy and is associated with high morbidity rates and impaired quality of life.⁸

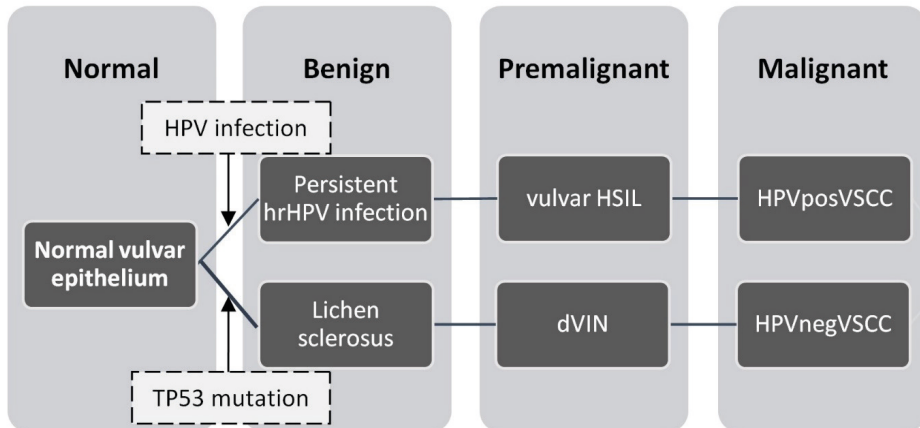


Figure 1. Overview of the multistage evolution of VSCC, depicting its two distinct pathways.

The HPV-driven pathway starts with an HPV infection, of which the lifetime risk for women is 80%. 20% of these infections persist, of which half causes vulvar high-grade squamous intraepithelial lesion (vHSIL). A small percentage of vHSIL develops into HPV positive VSCC. The HPV-independent pathway is associated with lichen sclerosus, which can develop into dVIN, commonly after TP53 mutation. DVIN is considered a direct precursor of HPV negative VSCC, which accounts for 80% of all VSCC cases.

The precursor of HPVnegVSCC is vulvar intraepithelial neoplasia of the differentiated type (dVIN) and often arises in a background of lichen sclerosus (LS).^{5,9} It is suggested that dVIN, rather than LS, is the direct precursor of HPVnegVSCC.¹⁰ HPVnegVSCC is associated with *TP53* mutations, which leads to abnormal expression of p53 protein.^{5,11} Somatic mutations in *TP53* lead to an uncontrolled cell cycle and chromosomal instability, resulting in tumor formation.¹² In HPVposVSCC ~75% of the lesions are caused by HPV type 16,

one of the most dominantly present oncogenic high risk HPV types (hrHPV).⁶ HPVposVSCC's precursor lesion is vulvar high grade squamous intra-epithelial lesions (vHSIL), formerly referred to as vulvar intraepithelial neoplasia of the usual type (uVIN). Cells infected by hrHPV express the viral oncogenes E6 and E7, which disturb the regulation of the cell cycle and promote malignant conversion. The lifetime risk of acquiring an hrHPV infection is approximately 80% for sexually active individuals. About 80% of the infections are cleared by the immune system within 18 months.¹³ Less than 10% of the infections persist and may cause vHSIL, 6.5% of which subsequently progresses towards VSCC.^{9,14} Several retrospective studies suggest that patients with HPVposVSCC display a better progression-free survival when compared to patients with HPVnegVSCC.^{11,15-17} Potentially, this difference is due to a more pronounced type 1 immune response as seen in various other malignancies.¹⁸

The adaptive cellular immune system is known to play a role in the protection against HPV-induced lesions. The group of immunosuppressed individuals (e.g. HIV, transplant patients and patients with leukemia) shows a higher incidence in persistent HPV infections and HPV-related malignancies.¹⁹⁻²¹ In addition, complete regression of HPV-induced lesions is associated with a strong influx of CD4+ and CD8+ T cells.²² Furthermore, healthy individuals display *ex-vivo* measurable frequencies of HPV-specific interferon producing T cells²³, which are largely lacking in patients with HPV-induced cancer.²⁴ Surprisingly though, the immune response in VSCC has not been studied thoroughly. Potential differences based on the distinct etiological pathways leading to HPVposVSCC and HPVnegVSCC have largely been ignored. This could be key to our understanding of the clinical behavior of VSCC. The expressed viral onco-proteins can function as tumor-specific antigens (TSAs) in HPVposVSCC, fueling the induction of a strong local type 1 T cell response and potentially leading to the better clinical outcome of HPVposVSCC when compared to HPVnegVSCC.^{11,17} Probably, the development of VSCC parallels that of oropharyngeal squamous cell carcinoma (OPSCC), which also develops via a non-viral or an hrHPV-driven pathway.²⁵ In this tumor type, the presence of HPV was found to be a strong independent determinant of progression free survival^{26,27} and associated with a dense infiltration of activated CD4+ and CD8+ T cells.^{26,28} Especially the intratumoral presence of type 1 cytokine producing HPV16-specific T cells constituted a type 1 immune contexture and improved clinical outcome. The absence of intratumoral HPV-specific T cells led to worse clinical outcome, indistinguishable from that of most HPVnegOPSCC.²⁹ The recognition that HPVposOPSCC have a better clinical outcome led to new trials

of de-intensified chemo (radio)therapy schedules, testing of novel therapeutics, including immunotherapeutic compounds in OPSCC.³⁰⁻³³ Hence, HPVposVSCC and HPVnegVSCC are different entities and they should be studied separately. Here, we summarize studies on the immune response as measured in the local tumor microenvironment (TME) in VSCC, and correlate this information to what is known about the precursors of VSCC and other HPV-induced malignancies. In addition, we will extrapolate this information to briefly delineate future therapeutic options for VSCC.

Innate immune responses in vulvar lesions

The role of the innate immune system is ambiguous, since it may promote tumor growth as well as prevent tumor progression.^{34,35} For instance, the role of tumor-associated myeloid cells is to foster tumor progression. Myeloid derived suppressor cells and type 2 macrophages can promote initiation and metastasis of tumor cells, and inhibit tumor-specific T cell responses.³⁶ On the other hand, abundantly present M1 macrophages in HPV-driven cervical cancer are an independent prognostic factor for good survival³⁷, and IFN γ -activated macrophages are important for tumor growth control in an HPV-tumor animal model.^{38,39} First, the role of macrophages will be discussed and then the focus is on professional antigen presenting cells (APCs), as these are crucial for the induction of adaptive immune responses.

In one study, myeloid cell infiltration between HPV16 driven primary vHSIL (n=43), 20 recurrent vHSIL, 21 VSCC and 26 normal vulvar tissues was compared.⁴⁰ Progression was associated with an increase in the numbers of CD14+ cells. Both in healthy controls and HPV16+ vHSIL the CD14+CD33-CD163+ mature (activated) M2 macrophages were more abundantly present than the M1 macrophages. Importantly, a higher number of both intraepithelial and stromal mature CD14+ macrophages was positively correlated to the number of intraepithelial regulatory T cells (Tregs, defined as CD4+FOXP3+, CD3+PD1+FOXP3+), negatively correlated to the number of activated CD8 T cells as well as with a decreased recurrence free survival time in vHSIL.⁴¹ The M2 macrophages are immune suppressive as reflected by their production of IL-10 and TGF- β . Potentially, M2 macrophages induce local Treg polarization.⁴² High M2 macrophage counts are associated with poor outcome in a diversity of tumors⁴³⁻⁴⁵, suggesting that they may have a similar negative impact in VSCC. Indeed a large study of 100 VSCC confirmed that these tumors were

infiltrated with variable numbers of CD68+ macrophages and T cells.⁴⁶ It also showed that increased immune infiltration of the tumors was associated with the expression of PD-L1 in 20% of the HPVnegVSCC although this was not observed in the HPVposVSCC.⁴⁶ The expression of PD-L1 was also found in a mouse tumor model³⁸ and may reflect an ongoing antitumor response in those HPVnegVSCC, suggesting that also HPVnegSCC can express antigens that are available for the immune system to react to. Unfortunately, no TME studies have been performed in dVIN, albeit that increased numbers of T cells and CD68+ macrophages were found in LS.⁴⁷

The professional APCs such as Langerhans cells (LCs) and dendritic cells (DCs) connect the innate and adaptive immune system, since they migrate from the epidermis towards the lymph node where they prime naïve T cells. A small immunohistochemistry study on VSCC with unknown HPV status (n=5) showed that VSCC were less infiltrated by CD1a+ LCs than healthy vulvar tissue.⁴⁸ What this means in the context of VSCC is unclear, but already in vHSIL the epithelial cell nests comprise lower numbers of CD1a+ and CD207+ DC than healthy tissue and this corresponds with a lower CD8 T cell infiltrate.^{49 50} The intraepithelial infiltration with CD1a+ DCs is inversely correlated with the grade of VIN.⁵¹ One explanation offered for this was the inhibition CCL20 secretion through E6 and E7 expression.⁵² Interestingly, also the dermis of LS is less infiltrated with CD1a+ DC/LCs, and this lack of infiltration is even more pronounced in VSCC that developed in the background of LS.^{48 53} The decrease in intraepithelial CD1a+ DC/LC numbers is paralleled in overall T cell infiltration.⁵⁴ Thus, non-malignant vulvar lesions and VSCC display low numbers of intraepithelial immature CD1a cells and CD8 T cells.

In addition to the lower number of APCs detected in VSCC, the function of the remaining APCs may have been altered towards a more immune suppressive phenotype. Cancer cells have been shown to inhibit DC maturation *in vitro* via the secretion of IL-6 and PgE2.^{55 56} Furthermore, both subtypes of VSCC are known to secrete receptor activator NF- κ B ligand (RANKL). RANKL can inhibit DC maturation, based on differences in the expression of CD80, CD83, CD86, and HLA-DR as well as their reduced IL-12 production and increased IL-10 production *in vitro*, suggesting that they have become tolerogenic DC. The addition of osteoprotegerin (a RANKL inhibitor) improved DC maturation, suggesting that RANKL could be an interesting therapeutic target to prevent immune escape of VSCC.⁵⁷ Furthermore, VSCC expresses indoleamine 2,3-dioxygenase (IDO).⁵⁸ This molecule not only suppresses T cell responses⁵⁹

but also blocks the maturation of DCs.⁶⁰ Notably, VSCC expressed IDO is an independent prognostic factor for worse overall survival in VSCC⁵⁸ but it remains to be investigated if this holds true for both subtypes of VSCC.

Adaptive immune responses in vulvar lesions

Two studies showed that CD4+ and CD8+ T cells co-infiltrate the cancer cells nests and stroma of primary VSCC tumors. Furthermore, the magnitude of this T cell infiltration positively correlated with the expression of HLA class 1.⁶¹ In many epithelial cancers, including HPVposOPSCC and HPVnegOPSCC, a dense infiltration with CD8+ T cells is associated with improved survival^{18 26 62 63}. However, a strong infiltration with T cells seemed not related to the prognosis of VSCC⁶⁴ which led to the hypothesis that the tumor infiltrating lymphocytes (TILs) had become anergic.⁶⁴ Indeed, the functional status of the T cells could make a difference and the expression of the T cell function suppressing enzyme IDO was shown to have a negative impact on survival in one study of VSCC⁵⁸, but this was not confirmed in another large study.⁶¹ Also the earlier reported expression of PD-L1 may restrain the function of TIL.⁴⁶ Therefore, the functional status of killer lymphocytes was analyzed using single stains for CD56 (expressed by NK/NKT cells) and granzyme B (GrB, for the cytotoxic status). CD56+ lymphocytes were mostly detected in the epithelium whereas GrB+ cells were predominantly found in the stroma. Importantly, the number of activated CD56+GrB+ intraepithelial lymphocytes significantly correlated with longer overall survival in VSCC.⁶⁵ Another study in HPVposVSCC and its precursors corroborated these findings.⁶⁵ Thus, rather than the number of infiltrating lymphocytes, their functional activity is key for clinical impact. Indeed, a detailed study on T cell infiltration, the expression of Tbet (for IFN γ producing T cells) and FOXP3 (for Tregs) and the co-inhibitory molecules programmed cell death 1 (PD1), T cell immunoglobulin mucin 3 (TIM3), and the natural killer cell lectin-like receptor A (NKG2A) to identify activated T cells, confirmed that vHSIL and HPVposVSCC are intensely surveyed by activated CD4+ and CD8+ T cells.⁴⁰ In vHSIL a dense infiltration with stromal activated CD8+TIM3+ T cells and stromal activated CD3+NKG2A+ T cells was related to the absence of recurrences and a prolonged recurrence-free survival.

It is likely that co-infiltration with Tregs blunted the activity of TILs. High numbers of Tregs in vHSIL were negatively associated with time to recurrence, especially when the stromal CD8+TIM3+ infiltration was limited.⁴⁰ In VSCC the

number of Tregs is even higher and outnumber the co-infiltrating activated T cells.⁴⁰ Potentially, these increased numbers of Tregs in VSCC^{40 58 61} are the result of tolerogenic DCs in these tumors that support the differentiation of naïve CD4⁺ T cells into Tregs.⁵⁷ Two studies that analyzed the impact of intraepithelial FOXP3⁺ Tregs in VSCC reported that their number does not influence survival rates.^{58 61} It is not clear if this result was similar for HPVnegVSCC and HPVposVSCC or what the infiltration with activated T cells was. Furthermore, both studies did not report the impact of stromal Tregs^{58 61}, which might be important in view of the presence of activated T cells in the stroma of HPVposVSCC⁴⁰ and the high number of GrB⁺ lymphocytes in the stroma of VSCC, with unknown HPV status.⁶⁵

On top of this, the impact of the infiltrating T cells may be lower due to the loss of HLA class I expression in VSCC.⁶¹ HLA-class I was found to be downregulated in 70% of vHSIL patients, 15% of which was caused by loss of heterozygosity (LOH). In the other cases HLA-class I could still be upregulated by interferon γ .⁶⁶ This will prove more difficult for HPVposVSCC, where HLA-class I downregulation through LOH is found in 25-55% of the cases.⁶⁶

Last but not least, the VSCC-infiltrating T cells may be suppressed following interaction with HLA-E, a non-classical HLA molecule expressed in at least 15% of the HPVposVSCC cases⁶⁷, and found to have a negative effect in other epithelial tumors.^{68 69} HLA-E is the ligand for the co-inhibitory molecule NKG2A, which we know is expressed by T cells in VSCC.⁶⁶ and recently has been shown to function as checkpoint for T cells in several mouse tumor models (van Montfoort et al., *in press*). Furthermore, HLA-E positive tumors show decreased numbers of T cells expressing NKG2A compared to HLA-E negative tumors, indicating a possible negative feedback loop.⁴⁰

In conclusion, the clinical impact of T cell infiltration can't be analysed outside the context of HLA expression, immune suppressive cells and regulatory mechanisms (e.g. expression of PD-1 and NKG2A).

Lichen sclerosis and dVIN

Lichen sclerosis is a chronic inflammatory skin disorder that is histologically characterized by a band-like lymphocytic infiltrate below the dermo-epidermal junction. Until now, the immune microenvironment has scarcely been studied

in LS and not at all in dVIN. In LS, the immune infiltrate is characterized by high numbers CD3+ T cells, consisting of CD4+, CD8+, and FOXP3+ T cells.⁷⁰⁻⁷⁵ The numbers of CD8+ T cells and FOXP3+ Tregs were significantly higher in the dermis of LS lesions than in healthy controls.⁷⁰ A more in depth analysis revealed the presence of high numbers of activated perforin+, GrB+, and type 1 interferon-associated CXCR3+ cytotoxic cells, suggesting that T cell mediated cytotoxicity occurs in LS.^{76,77} Increased numbers of HLA-DR+ cells in the dermal infiltrate indicates that these T cells have been activated following cognate antigen recognition.⁷⁵ Moreover, gene expression arrays showed that significantly more type 1 immune response associated pro-inflammatory cytokines are expressed in LS when compared to healthy tissue.⁷⁰ Together, these data indicate that the microenvironment of LS resembles that of an autoimmune disorder, characterized by a type 1 immune response.

Other indications for a developed T cell response comes from a recent study showing that monoclonal T cell receptor (TCR) rearrangement was identified in 47% of patients with LS.⁷⁸ Interestingly, the infiltrates of the lesions with monoclonal TCR rearrangement were dominated by CD4+ T-cells, B cells, and fascin (a protein involved in antigen presentation)-positive dendritic cells. Similarly, HPV-negative penile cancers, known to be associated with LS, and differentiated penile intraepithelial neoplasia, showed accumulation of T cells with monoclonal TCR rearrangement.⁷⁹ This suggests the presence of a specific antigen in LS and HPVnegVSCC or penile carcinoma to which the immune system can respond.

These findings suggest that LS is an immunologically active lesion, which may give rise to the development of malignancies. Future studies are needed to explore the local immune environment, in particular in relation to clinical outcomes and progression of disease.

Modulation of the local microenvironment

Until now, the cornerstone of the treatment for VSCC, vHSIL and dVIN is surgery. This treatment is associated with significant morbidity due to lymph oedema, wound infections, and psychological and sexual problems.⁸⁰ However, recurrence rates after these treatments occur in up to 50% of patients.⁸¹ Non-surgical treatments for vHSIL include topical creams and gels, HPV vaccines, and photodynamic therapy (PDT). Understanding of the modulation of the local microenvironment is important as this might unveil clues to novel therapy. An overview of previous clinical trials can be found in **Table 1**.

Table 1. Overview immunotherapeutic clinical studies in vHSIL.

Author	Treatment	Study design	Phase trial	Patient number	Primary endpoint	Main immunological outcome	Main clinical outcome
Jayne 2002	Imiquimod	RS	2	13	Clinical response	Not investigated.	8/13 CR, 4/7 75% regression
Van Seters 2002	Imiquimod	PS	2	15	Clinical response	Not investigated.	27% CR, 60% PR
Todd 2002	Imiquimod	PS	2	15	Clinical response	Not investigated.	4/15 showed clinical improvement of which 3 had CR
Wendling 2004	Imiquimod	PS	2	12	Clinical response	Not investigated.	7/12 CR or PR
van Poelgeest 2005* (ref 93)	Imiquimod	PS	2	17	Immunological assay of responders to imiquimod	Systemic HPV16-specific IFN γ -associated CD4+ T cell immunity associated with CR.	Not investigated.
Le 2007	Imiquimod	PS	2	39	Clinical response	Not investigated.	64% showed CR after 16 weeks, 27% had clinically significant shrinkage.
Mathiesen 2007	Imiquimod vs placebo	RCT	2	32	Clinical response	Not investigated.	81% in the imiquimod group showed CR, 10% PR and no responses in the placebo-group
van Seters 2008 (ref 92)	Imiquimod vs placebo	RCT	2	52	Clinical response	Higher numbers of intralesional CD1a+ DCs, CD8+ T cells and CD94+ NK cells with imiquimod.	81% of all patients had >25% reduction of the primary lesion
Terlou 2010 (ref 45)	Imiquimod vs placebo	RCT	2	25	HPV presence (based on p16 IHC)	Lower intralesional CD208+, CD8+, CD4+, CD94+, CD14+ and CD68+ in the HPV cleared patients.	19/25 were HPV+ before treatment, of which 9 became HPV- after treatment.
Westermann 2013	Imiquimod	RS	2	62	Clinical response	Not investigated.	76% CR, 19% PR, 3% weak PR, 1 non-responder. Recurrent disease in 27%.

Author	Treatment	Study design	Phase trial	Patient number	Primary endpoint	Main immunological outcome	Main clinical outcome
Frega 2013	Imiquimod vs cold-knife excision	PS	2	80	Complete response, recurrence rate	Not investigated.	Recurrence rate was lower for imiquimod, relapse rate was higher. The overall CR was inferior in this group.
Kim 2015	Imiquimod	PS	2	9	Clinical response	Not investigated.	4/9 patients had CR, 2/9 PR and 3 were lost to FU.
Winters 2008 (ref 95)	Imiquimod + PDT	PS	2	20	Clinical response	Responders to imiquimod had higher Tregs before treatment, and more CD8+ T cells after treatment.	Overall response rate 55% by intention treat and 64% per protocol.
Kenter 2009 (ref 103)	HPV16 SLP vaccine	PS	2	20	Clinical response	Not investigated.	25% CR and histologic response, 35% PR at 3 months.
Daayana 2010 (ref 94)	Imiquimod + HPV16 vaccine	PS	2	19	Clinical response	After treatment with imiquimod + vaccination more local infiltration of CD4/8+ T cells was seen in the responders with lower Tregs.	32% complete response at week 10, 58% at week 20 and 63% at week 52.
Van Poelgeest 2016 (ref 105)	HPV16 SLP vaccine +/- imiquimod	RCT	2	43	Ex vivo detectable HPV16 specific CD8+ T cell response	84% showed systemic vaccine-induced HPV16-IFN γ associated CD4+ and CD8+ T cell response. No difference was found between the groups.	53% vaccine-induced CR/PR at 3 months. No difference was found between the groups.

An overview of previously performed clinical trials with imiquimod, PDT and/or vaccination in vHSIL.

RCT = randomised controlled trial, PS = prospective cohort study, RS = retrospective cohort study, CR = complete response, PR = partial response. *van Poelgeest 2005 (reference 93) and Terlou 2010 (reference 45) performed immunological analyses on data from the RCT of van Seters 2008 (reference 92).

Imiquimod

Imiquimod activates the innate immune response by interacting with toll-like receptor (TLR)7, expressed by DC, monocytes and macrophages. TLR7 activation induces secretion of proinflammatory cytokines, particularly IFN γ and also enhances DC maturation and antigen presentation.⁸² These cytokines induce polarisation towards a type 1 immune response.⁸³ Imiquimod is FDA approved for the treatment of basal cell carcinoma (BCC) and external genital warts^{84 85} but it can also be effective for the treatment of HPV-induced HSIL of the cervix, vagina and vulva.⁸⁶ In a retrospective study imiquimod was found to be effective, with complete responses between 20-81%, and partial responses of 10-75%, as well as and safe for treatment of vHSIL, although long-term response and progression of disease are unknown. The side effects of imiquimod treatment are mainly local adverse effects (itching, burning), which occur in the majority of patients, and flu-like symptoms.⁸⁷ Importantly, a randomized controlled trial (RCT) showed objective clinical responses in 81% of vHSIL patients compared to none in the placebo group.⁸⁸ In this trial, the clinical response, defined as lesion regression and HPV clearance, was associated with a pre-existing systemic HPV-specific Th1 cell response.⁸⁹ Several studies in vHSIL have shown that imiquimod induced T cell infiltration⁹⁰⁻⁹², including CD8+ T cells.^{90,93} This was associated with expression of CXCR3 and the ubiquitous cytoplasmic protein MxA, indicative of type 1 IFN signalling.⁸⁴ Non-responsiveness to imiquimod was associated with higher levels of intralesional Tregs.⁹² Another study showed that clinical responses after imiquimod were associated with increased numbers of intraepithelial CD1a+ and CD8+ cells and a decrease in Tregs in the stroma.⁹⁰ In addition, the elevated CD14+ cells found in vHSIL lesions returned to control levels in HPV-cleared patients, in contrast to non-cleared patients. A phase II study of imiquimod followed by therapeutic HPV16 E6E7L2 vaccination indicated that increased CD4+ and CD8+ was associated with clinical response to the vaccine and that relatively elevated Treg levels were observed in the group not responding to the vaccine.⁹¹

These findings suggest that a combination therapy in which the number of HPV 16 specific type 1 T cells are amplified by vaccination and the attraction of immune cells to the lesion site is fostered by local administration of imiquimod will be more successful than monotherapy with either one of the treatments.

Photodynamic therapy

Topical photodynamic therapy (PDT) is an effective and safe nonsurgical treatment option⁹⁴ that involves topical application of a non-toxic

photosensitizer, which is absorbed into neoplastic tissue. This leads to the production of reactive oxygen species and tumor cell death.⁹⁵ The acute inflammation following the clearance of destructed cells is known to contribute to prolonged recurrence free survival in advanced cancers.⁹⁵ In addition, the pro-inflammatory effects of PDT might increase DC migration, antigen uptake and maturation. PDT might also increase the immunogenicity of dead tumor cells by exposing or creating new antigens or by the induction of more effective cytotoxic T cells. Clinical studies in vHSIL have shown varying clinical and histological responses from 20-67%^{95 96}, however long-term clinical outcomes are not known.

Immunological studies of PDT in vHSIL have shown a significant CD8+ infiltration in posttreatment vHSIL responders compared to non-responders.⁹⁷ In contrast, loss or downregulation of HLA class I was associated with a lack of response to PDT. A phase II trial of sequential imiquimod and PDT in 20 women with vHSIL showed significantly higher numbers of Tregs in the lesions after imiquimod treatment in non-responders, whereas no differences in the infiltration of CD4+, CD8+, CD68+, and CD1a+ cells were observed between responders and non-responders before or after PDT treatment.⁹² Interestingly, also in this study a systemic pre-existing HPV-specific immune response was significantly correlated with response to PDT. Together, these studies suggest that the efficacy of PDT is related to immune promoting changes in the microenvironment of treated lesions.

Therapeutic vaccination

The aim of therapeutic vaccination against cancer is to launch an immune mediated attack against cancers by amplifying an anti-tumor T cell response. Therapeutic vaccines stimulate T cells against TSA created by oncogenic viruses or DNA mutations.⁹⁸

VHSIL and HPVposVSCC express the viral oncoproteins E6 and E7, and expose therefore ideal TSAs for a therapeutic vaccine.⁹⁹ In two independent clinical trials it was shown that HPV16 synthetic long peptide (SLP) vaccination of patients with HPV16-induced vHSIL resulted in complete and partial regression of the lesion(s), especially in patients who showed a stronger vaccine-induced HPV16-induced T cell response, irrespective of the immune status of these patients. The first study showed a complete response rate of 47% at 3 months.¹⁰⁰ Post hoc analyses suggested that complete responders had significantly stronger type 1 T cell response. Importantly, a subsequent RCT

confirmed that the clinical efficacy of HPV16-SLP vaccination was related to the strength of the vaccine-induced HPV-specific immune response.⁸⁹

Successful treatment of HPV16-induced vHSIL was also achieved with an HPV16 E6E7L2 fusion protein. This phase II study of combination immunotherapy of topical imiquimod followed by HPV16 E6E7L2 vaccine in women with vHSIL showed that there was significantly increased local infiltration of CD4+ and CD8+ T cells in responders, whereas non-responders showed an increased density of Tregs.⁹¹ Similarly, clinical responses after recombinant vaccinia virus encoding HPV16/18 E6E7 vaccination were associated with pre-treatment higher levels of intralesional CD4+, CD8+, and CD1a+ cells compared to non-responders. Although non-responders showed an increase in CD4+ and CD8+, but not CD1a+ cells post-vaccination, their numbers were still lower than the numbers observed in the pre-treatment samples of responders.¹⁰¹ These data suggest that therapeutic vaccines can boost the specific T cell response but are only effective in settings where the lesions have the capacity to attract substantial T cell infiltration. The identification of suitable target antigens in HPVnegVSCC is key when vaccination is considered as treatment option.

Topical treatments for dVIN and LS

Interestingly, topical treatment with pimecrolimus induces a clinical response rate in 80% of the LS patients. The response was associated with a significant downregulation of CD3+, CD8+ T cells, CD57+ NK cells, and CD68+ macrophages.¹⁰²

Intriguingly, a recent study in 507 LS patients showed that regular long-term use of topical glucocorticosteroids reduced the risk of developing dVIN or VSCC.¹⁰³ Glucocorticosteroids are steroid hormones that inhibit the NF- κ B pathway, resulting in decreased numbers of transcription factors which regulate the expression of pro-inflammatory genes (e.g. TNF α and IL-1 β) and anti-apoptotic genes.¹⁰⁴ Glucocorticoids have pro-apoptotic properties in lymphoid cells¹⁰⁵, and can selectively inhibit Tregs. Keeping in mind chronic inflammation as a hallmark of cancer¹⁰⁶, inhibition of chronic inflammation by topical steroids may underlie the reduced risk for malignant transformation in LS.

Conclusion

In this review, we summarized studies that describe the TME in VSCC, and its precursors LS/dVIN and vHSIL (**Figure 2**). The immune infiltrate in VSCC and its precursors is characterized by variable numbers of tumor infiltrating innate and adaptive immune cells. The lesions show an increase in M2 macrophages and Tregs and a decrease in activated CD8+ T cells compared to healthy controls.^{40 41} In patients with vHSIL, the presence of type 1 immunity is associated with fewer recurrences after treatment and also with better response to topical treatment with imiquimod, PDT, or therapeutic vaccination.^{40 89 97 107} Although the number of studies in vHSIL is limited and immune correlates for a treatment response need to be defined, patients with HLA-class I positive lesions, high numbers of infiltrating T cells with low numbers of Tregs, or pre-existing HPV-specific immunity, may help identify patients more likely to respond to immune therapy. Moreover, in LS high numbers of CD4+, CD8+, and Tregs are found indicating that these lesions are immunologically active and suggest that also the HPVnegVSCC might be potential candidate for future immune therapies.⁷⁰⁻⁷⁵

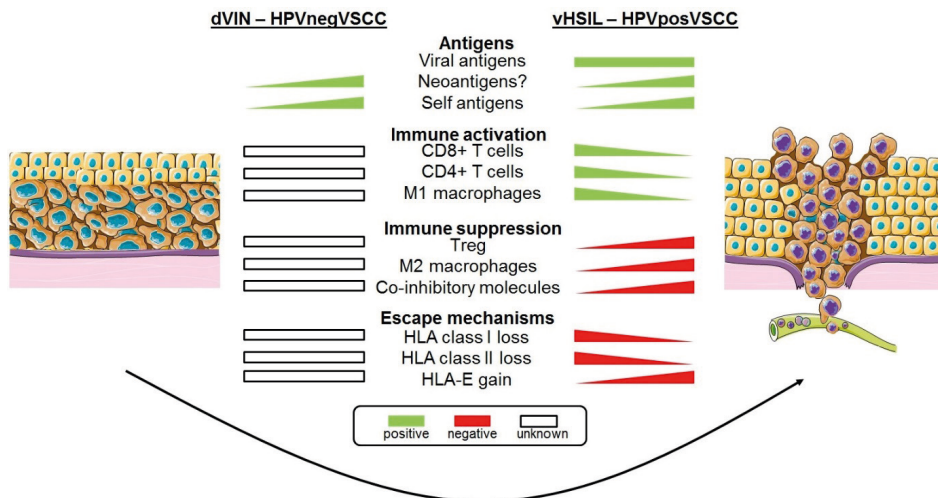


Figure 2. Overview of current literature about tumor microenvironment in VSCC and its precursors.

The immune infiltrate in VSCC and its precursors is characterized by variable numbers of tumor infiltrating innate and adaptive immune cells. HPVposVSCC's precursor lesion is vHSIL, and as progression occurs these lesions show an increase in M2 macrophages, and Tregs and a decrease in activated CD4 and CD8+ T cells. Cancer may escape the immune response due to downregulation of HLA Class I and II molecules on the cell surface and increased expression of co-inhibitory molecules. Literature on HPVnegVSCC and its precursor dVIN is scarce. Illustrations of lesions created from adapted images of Servier Medical Art, licensed under a Creative Commons Attribution 3.0 Unported License.

Expert opinion

Tumors can exploit several immunosuppressive mechanisms to escape from the immune system, by HLA class I loss, or by creating an immune-suppressive TME, by attracting Tregs and suppressive myeloid cells as well as by secretion of immune suppressive cytokines such as TGF- β or IL-10.⁹⁸ In addition, ligation of co-inhibitory molecules expressed by T cells, including cytotoxic T-lymphocyte associated antigen-4 (CTLA-4), programmed death-1 (PD-1) and NKG2a, may occur. Overexpression of the ligand of PD-1 (PD-L1), is associated with poor clinical outcomes in many immunogenic cancers.¹⁰⁸ Monoclonal-antibody-based therapies targeting CTLA-4 and/or PD-1 (checkpoint blockade) have yielded significant clinical benefits including durable responses to patients with different malignancies. Blocking of PD-1/PD-L1 rebalances the host-tumor interaction and is successful in the treatment of many tumor types.¹⁰⁹

Cancers responsive to checkpoint inhibitors may be those with abundant tumor specific target antigens, generated by a high frequency of mutations that are recognized as non-self-peptides by CD4+ and CD8+ T cells.¹¹⁰⁻¹¹² Studies on the expression of immune checkpoint molecules in VSCC are scarce. A recent study in OPSCC showed that the percentage of CTLA-4 and PD1/PD-L1 expressing T cells was higher in TILs compared to peripheral blood mononuclear cells from patients.¹¹³ Recurrent HPV-positive tumors showed a similar composition of T cell subsets and checkpoint molecule expression as found in HPV-negative tumors. Not to our surprise, blockade of PD-1 /PD-L1 gave similar results in HPVpos- and HPVnegOPSCC.³² The absence of a better result in recurrent HPV-induced OPSCC is likely because these are the ones lacking an HPV-specific T cell response.¹¹⁴ Therapeutic HPV16 vaccination induced an immune response which was associated with clinical response in patients with HPV16-induced vHSIL.^{100 107} However, it is unlikely that therapeutic vaccines will be able to overcome the immune-suppressed microenvironment in established and advanced cancers without co-treatment. Indeed, a recent phase I/II study in HPV16-positive cancers showed that HPV16-SLP vaccination in combination with the PD1 antibody nivolumab resulted in a response rate of 36% in OPSCC patients, compared to 16% for nivolumab monotherapy in p16+ OSCC patients.^{32 33} Few studies showed the expression of PD-L1 in a subset of HPVnegVSCC.^{46 115} The presence of CD8+ T cells has been shown to be pivotal for the prediction of response to PD-L1 checkpoint inhibitors in melanoma.¹¹⁶ Although VSCC is highly infiltrated by CD8+ T cells, high numbers of just CD8+ T cells were not associated with survival.⁶¹ However,

three different studies showed that these cells needed to be activated before a positive impact on clinical outcome was detected in early stage VSCC.⁶⁵ PD-L1 expression is also found on cervical cancer cells, but not of prognostic value.¹¹⁷ Besides by tumor cells, PD-L1 is also expressed by M2 macrophages⁵⁶, which are frequently found in HPVposVSCC⁴¹, melanoma and several HPV-positive tumors.¹¹⁸⁻¹²¹ Importantly, PD-L1 positivity of peritumoral immune cells is an independent favorable prognostic factor for survival¹²², probably because it's expression is a telltale for the local production of IFN γ by activated T cells. Further effort must be made to determine the value of this and other checkpoints as predictive biomarkers in both types of VSCC.

As previously mentioned, Tregs are an immunosuppressive cell type in the TME, and some of them are shown to react to tumor antigens.¹²³ Tregs may be inactivated by treatment of patients with cyclophosphamide, or by inhibition of IDO¹²⁴ but also by antibodies against CTLA-4.¹²⁵

Recent clinical data have clearly demonstrated that human cancer cells express antigens that are newly created by mutations in the DNA encoding normal proteins (neoantigens). These mutated cancer proteins can be processed into peptides and presented on the surface of tumor cells leading to immune recognition in vivo as “non-self” or foreign by autologous tumor-specific T cells.

A few studies reported a higher incidence of somatic mutations in HPVnegVSCC compared to HPVposVSCC, correlating with increasing grades of neoplasia.¹²⁶ *TP53*, *CDKN2A* and *HRAS* were most frequently mutated, but the full range of mutations still needs to be established as current studies focused on these known hot-spots.^{11 127} In some cancers, the expression of foreign/mutated antigens correlates with increased numbers of TILs and better with clinical outcome.^{128 129} Moreover, high mutational load in tumors is associated with sensitivity to immune checkpoint inhibition and better clinical outcome.¹³⁰ These developments are highly promising and open the opportunities for precision medicine in patients with VSCC, including vaccination. On the basis of landmark preclinical studies that identified immunogenic mutations and showed survival benefit upon neoantigen vaccination in prophylactic and therapeutic settings¹⁰⁶, clinical phase I/II neoantigen cancer vaccine trials are currently ongoing in metastatic melanoma (NCT 03300843, NCT 01970358, NCT 02897765), glioblastoma (NCT 02287428), and pancreatic cancer (NCT 03122106).¹⁰⁷⁻¹⁰⁹

Combination treatment of vaccination with checkpoint inhibitors, chemotherapy, or modulation of the microenvironment by PDT or radiotherapy has already shown clinical success in different cancer types.¹³¹ Adoptive cell therapy (ACT) makes use of infusion of tumor-specific T cells into patients with advanced cancers. Currently, phase 1 clinical trials have been initiated with neoantigen-based ACT in patients with myelodysplastic syndrome (NCT03258359) or in patients with glioblastoma, non-small cell lung cancer, ovarian, and breast cancer (NCT03412877). In the light of these developments, it is to be expected that HPV16 oncoprotein-based and neoantigen-based vaccination, in combination with chemo-or radiotherapy, checkpoint blockade and/or ACT will become available for the treatment of advanced VSCC.^{132 133}

By unraveling the VSCC mutational and immune landscape, current treatments can be improved, and new immunotherapeutic interventions can be designed for HPVnegVSCC and HPVposVSCC and its precursors. Future studies should aim at whole genome sequencing in combination with in depth analysis of the immune microenvironment, not only with respect to type and numeric distribution of innate and adaptive immune cell types, but also to their functional characteristics.

References

1. Sankaranarayanan R, Ferlay J. Worldwide burden of gynaecological cancer: the size of the problem. *Best practice & research Clinical obstetrics & gynaecology* 2006;20(2):207-25.
2. Judson PL, Habermann EB, Baxter NN, et al. Trends in the incidence of invasive and in situ vulvar carcinoma. *Obstetrics & Gynecology* 2006;107(5):1018-22.
3. Kurman RJ, Carcangiu ML, Herrington CS, et al. WHO classification of tumours of female reproductive organs. *Lyon: International Agency for Research on Cancer* 2014;172
4. Pilotti S, Donghi R, D'Amato L, et al. Papillomavirus, p53 alteration and primary carcinoma of the vulva. *European journal of cancer (Oxford, England : 1990)* 1993;29a(6):924-5. [published Online First: 1993/01/01]
5. van der Avoort IA, Shirango H, Hoevenaars BM, et al. Vulvar squamous cell carcinoma is a multifactorial disease following two separate and independent pathways. *International journal of gynecological pathology : official journal of the International Society of Gynecological Pathologists* 2006;25(1):22-9. [published Online First: 2005/11/25]
6. Santos M, Landolfi S, Olivella A, et al. p16 overexpression identifies HPV-positive vulvar squamous cell carcinomas. *The American journal of surgical pathology* 2006;30(11):1347-56. doi: 10.1097/01.pas.0000213251.82940.bf [published Online First: 2006/10/26]
7. Schuurman MS, van den Einden LC, Massuger LF, et al. Trends in incidence and survival of Dutch women with vulvar squamous cell carcinoma. *European journal of cancer (Oxford, England : 1990)* 2013;49(18):3872-80. doi: 10.1016/j.ejca.2013.08.003 [published Online First: 2013/09/10]
8. de Hullu JA, van der Zee AG. Surgery and radiotherapy in vulvar cancer. *Critical reviews in oncology/hematology* 2006;60(1):38-58. doi: 10.1016/j.critrevonc.2006.02.008 [published Online First: 2006/07/11]
9. van de Nieuwenhof HP, van der Avoort IA, de Hullu JA. Review of squamous premalignant vulvar lesions. *Critical reviews in oncology/hematology* 2008;68(2):131-56. doi: 10.1016/j.critrevonc.2008.02.012 [published Online First: 2008/04/15]
10. Kokka F, Singh N, Faruqi A, et al. Is differentiated vulvar intraepithelial neoplasia the precursor lesion of human papillomavirus-negative vulvar squamous cell carcinoma? *International journal of gynecological cancer : official journal of the International Gynecological Cancer Society* 2011;21(7):1297-305. doi: 10.1097/IGC.0b013e31822dbe26 [published Online First: 2011/09/29]
11. Nooij LS, Ter Haar NT, Ruano D, et al. Genomic Characterization of Vulvar (Pre) cancers Identifies Distinct Molecular Subtypes with Prognostic Significance. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2017;23(22):6781-89. doi: 10.1158/1078-0432.ccr-17-1302 [published Online First: 2017/09/14]
12. Olivier M, Hollstein M, Hainaut P. TP53 mutations in human cancers: origins, consequences, and clinical use. *Cold Spring Harbor perspectives in biology* 2010;2(1):a001008. doi: 10.1101/cshperspect.a001008 [published Online First: 2010/02/26]
13. Baseman JG, Koutsky LA. The epidemiology of human papillomavirus infections. *Journal of clinical virology : the official publication of the Pan American Society for Clinical Virology* 2005;32 Suppl 1:S16-24. doi: 10.1016/j.jcv.2004.12.008 [published Online First: 2005/03/09]
14. van Seters M, van Beurden M, de Craen AJ. Is the assumed natural history of vulvar intraepithelial neoplasia III based on enough evidence? A systematic review of 3322 published patients. *Gynecologic oncology* 2005;97(2):645-51. doi: 10.1016/j.ygyno.2005.02.012 [published Online First: 2005/05/03]
15. Wakeham K, Kavanagh K, Cuschieri K, et al. HPV status and favourable outcome in vulvar squamous cancer. *International journal of cancer* 2017;140(5):1134-46. doi: 10.1002/ijc.30523 [published Online First: 2016/11/20]
16. Rasmussen CL, Sand FL, Hoffmann Frederiksen M, et al. Does HPV status influence survival after vulvar cancer? *International journal of cancer* 2018;142(6):1158-65. doi: 10.1002/ijc.31139 [published Online First: 2017/11/02]
17. Hinten F, Molijn A, Eckhardt L, et al. Vulvar cancer: Two pathways with different localization and prognosis. *Gynecologic oncology* 2018 doi: 10.1016/j.ygyno.2018.03.003 [published Online First: 2018/03/21]

18. Fridman WH, Pagès F, Sautès-Fridman C, et al. The immune contexture in human tumours: impact on clinical outcome. *Nat Rev Cancer* 2012;12(4):298-306. doi: 10.1038/nrc3245 [published Online First: 2012/03/16]
19. van der Burg SH, Palefsky JM. Human Immunodeficiency Virus and Human Papilloma Virus - why HPV-induced lesions do not spontaneously resolve and why therapeutic vaccination can be successful. *Journal of translational medicine* 2009;7:108. doi: 10.1186/1479-5876-7-108 [published Online First: 2009/12/22]
20. Petry KU, Kochel H, Bode U, et al. Human papillomavirus is associated with the frequent detection of warty and basaloid high-grade neoplasia of the vulva and cervical neoplasia among immunocompromised women. *Gynecologic oncology* 1996;60(1):30-4. doi: 10.1006/gyno.1996.0007 [published Online First: 1996/01/01]
21. Jamieson DJ, Paramsothy P, Cu-Uvin S, et al. Vulvar, vaginal, and perianal intraepithelial neoplasia in women with or at risk for human immunodeficiency virus. *Obstetrics and gynecology* 2006;107(5):1023-8. doi: 10.1097/01.AOG.0000210237.80211.ff [published Online First: 2006/05/02]
22. Coleman N, Birley HD, Renton AM, et al. Immunological events in regressing genital warts. *American journal of clinical pathology* 1994;102(6):768-74. [published Online First: 1994/12/01]
23. Welters MJ, van der Logt P, van den Eeden SJ, et al. Detection of human papillomavirus type 18 E6 and E7-specific CD4+ T-helper 1 immunity in relation to health versus disease. *International journal of cancer* 2006;118(4):950-6. doi: 10.1002/ijc.21459 [published Online First: 2005/09/10]
24. de Jong A, van Poelgeest MI, van der Hulst JM, et al. Human papillomavirus type 16-positive cervical cancer is associated with impaired CD4+ T-cell immunity against early antigens E2 and E6. *Cancer research* 2004;64(15):5449-55. doi: 10.1158/0008-5472.can-04-0831 [published Online First: 2004/08/04]
25. Snijders PJ, Steenbergen RD, Top B, et al. Analysis of p53 status in tonsillar carcinomas associated with human papillomavirus. *The Journal of general virology* 1994;75 (Pt 10):2769-75. doi: 10.1099/0022-1317-75-10-2769 [published Online First: 1994/10/01]
26. Wansom D, Light E, Worden F, et al. Correlation of cellular immunity with human papillomavirus 16 status and outcome in patients with advanced oropharyngeal cancer. *Archives of otolaryngology--head & neck surgery* 2010;136(12):1267-73. doi: 10.1001/archoto.2010.211 [published Online First: 2010/12/22]
27. Ang KK, Harris J, Wheeler R, et al. Human papillomavirus and survival of patients with oropharyngeal cancer. *The New England journal of medicine* 2010;363(1):24-35. doi: 10.1056/NEJMoa0912217 [published Online First: 2010/06/10]
28. Badoual C, Hans S, Rodriguez J, et al. Prognostic value of tumor-infiltrating CD4+ T-cell subpopulations in head and neck cancers. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2006;12(2):465-72. doi: 10.1158/1078-0432.ccr-05-1886 [published Online First: 2006/01/24]
29. Welters MJP, Ma W, Santegoets SJ, et al. Intratumoral HPV16-specific T-cells constitute a type 1 oriented tumor microenvironment to improve survival in HPV16-driven oropharyngeal cancer. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2017 doi: 10.1158/1078-0432.ccr-17-2140 [published Online First: 2017/10/12]
30. Bhatia A, Burtneess B. Human Papillomavirus-Associated Oropharyngeal Cancer: Defining Risk Groups and Clinical Trials. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2015;33(29):3243-50. doi: 10.1200/jco.2015.61.2358 [published Online First: 2015/09/10]
31. Sacco AG, Cohen EE. Current Treatment Options for Recurrent or Metastatic Head and Neck Squamous Cell Carcinoma. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2015;33(29):3305-13. doi: 10.1200/jco.2015.62.0963 [published Online First: 2015/09/10]
32. Ferris RL, Blumenschein G, Jr., Fayette J, et al. Nivolumab for Recurrent Squamous-Cell Carcinoma of the Head and Neck. *The New England journal of medicine* 2016;375(19):1856-67. doi: 10.1056/NEJMoa1602252 [published Online First: 2016/10/11]
33. Glisson B, Massarelli E, William W, et al. 1136ONivolumab and ISA 101 HPV vaccine in incurable HPV-16+ cancer. *Annals of Oncology* 2017;28(suppl_5)
34. Hagerling C, Casbon AJ, Werb Z. Balancing the innate immune system in tumor development. *Trends in cell biology* 2015;25(4):214-20. doi: 10.1016/j.tcb.2014.11.001 [published Online First: 2014/12/03]

35. Berraondo P, Minute L, Ajona D, et al. Innate immune mediators in cancer: between defense and resistance. *Immunological reviews* 2016;274(1):290-306. doi: 10.1111/immr.12464 [published Online First: 2016/10/27]
36. Yang L, Zhang Y. Tumor-associated macrophages: from basic research to clinical application. *Journal of hematology & oncology* 2017;10(1):58. doi: 10.1186/s13045-017-0430-2 [published Online First: 2017/03/01]
37. de Vos van Steenwijk PJ, Ramwadhoebe TH, Goedemans R, et al. Tumor-infiltrating CD14-positive myeloid cells and CD8-positive T-cells prolong survival in patients with cervical carcinoma. *Int J Cancer* 2013;133(12):2884-94. doi: 10.1002/ijc.28309 [published Online First: 2013/06/07]
38. van der Sluis TC, Sluijter M, van Duiker S, et al. Therapeutic Peptide Vaccine-Induced CD8 T Cells Strongly Modulate Intratumoral Macrophages Required for Tumor Regression. *Cancer immunology research* 2015;3(9):1042-51. doi: 10.1158/2326-6066.cir-15-0052 [published Online First: 2015/04/19]
39. Thoreau M, Penny HL, Tan K, et al. Vaccine-induced tumor regression requires a dynamic cooperation between T cells and myeloid cells at the tumor site. *Oncotarget* 2015;6(29):27832-46. doi: 10.18632/oncotarget.4940 [published Online First: 2015/09/05]
40. van Esch EM, van Poelgeest MI, Kouwenberg S, et al. Expression of coinhibitory receptors on T cells in the microenvironment of usual vulvar intraepithelial neoplasia is related to proinflammatory effector T cells and an increased recurrence-free survival. *International journal of cancer* 2015;136(4):E95-106. doi: 10.1002/ijc.29174 [published Online First: 2014/09/16]
41. van Esch EM, van Poelgeest MI, Trimboos JB, et al. Intraepithelial macrophage infiltration is related to a high number of regulatory T cells and promotes a progressive course of HPV-induced vulvar neoplasia. *International journal of cancer* 2015;136(4):E85-94. doi: 10.1002/ijc.29173 [published Online First: 2014/09/16]
42. Gajewski TF, Schreiber H, Fu YX. Innate and adaptive immune cells in the tumor microenvironment. *Nature immunology* 2013;14(10):1014-22. doi: 10.1038/ni.2703 [published Online First: 2013/09/21]
43. Fujii N, Shomori K, Shiomi T, et al. Cancer-associated fibroblasts and CD163-positive macrophages in oral squamous cell carcinoma: their clinicopathological and prognostic significance. *Journal of oral pathology & medicine : official publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology* 2012;41(6):444-51. doi: 10.1111/j.1600-0714.2012.01127.x [published Online First: 2012/02/03]
44. Komohara Y, Ohnishi K, Kuratsu J, et al. Possible involvement of the M2 anti-inflammatory macrophage phenotype in growth of human gliomas. *The Journal of pathology* 2008;216(1):15-24. doi: 10.1002/path.2370 [published Online First: 2008/06/17]
45. Kurahara H, Shinchi H, Mataka Y, et al. Significance of M2-polarized tumor-associated macrophage in pancreatic cancer. *The Journal of surgical research* 2011;167(2):e211-9. doi: 10.1016/j.jss.2009.05.026 [published Online First: 2009/09/22]
46. Hecking T, Thiesler T, Schiller C, et al. Tumoral PD-L1 expression defines a subgroup of poor-prognosis vulvar carcinomas with non-viral etiology. *Oncotarget* 2017;8(54):92890-903. doi: 10.18632/oncotarget.21641 [published Online First: 2017/12/02]
47. Farrell AM, Marren P, Dean D, et al. Lichen sclerosis: evidence that immunological changes occur at all levels of the skin. *The British journal of dermatology* 1999;140(6):1087-92. [published Online First: 1999/06/03]
48. Rotsztejn H, Trznadel-Budzko E, Jesionek-Kupnicka D. Langerhans cells in vulvar lichen sclerosis and vulvar squamous cell carcinoma. *Archivum immunologiae et therapiae experimentalis* 2006;54(5):363-6. doi: 10.1007/s00005-006-0039-6 [published Online First: 2006/10/13]
49. van Seters M, Beckmann I, Heijmans-Antonissen C, et al. Disturbed patterns of immunocompetent cells in usual-type vulvar intraepithelial neoplasia. *Cancer research* 2008;68(16):6617-22. doi: 10.1158/0008-5472.can-08-0327 [published Online First: 2008/08/15]
50. Mulvany NJ, Allen DG. Differentiated intraepithelial neoplasia of the vulva. *International journal of gynecological pathology : official journal of the International Society of Gynecological Pathologists* 2008;27(1):125-35. doi: 10.1097/pgp.0b0318134ea34 [published Online First: 2007/12/25]

51. Singh K, Yeo Y, Honest H, et al. Antigen processing and correlation with immunological response in vulval intraepithelial neoplasia--a study of CD1a, CD54 and LN3 expression. *Gynecologic oncology* 2006;102(3):489-92. doi: 10.1016/j.ygyno.2006.01.036 [published Online First: 2006/03/07]
52. Jiang B, Xue M. Correlation of E6 and E7 levels in high-risk HPV16 type cervical lesions with CCL20 and Langerhans cells. *Genetics and molecular research : GMR* 2015;14(3):10473-81. doi: 10.4238/2015.September.8.8 [published Online First: 2015/09/25]
53. Raspollini MR, Baroni G, Taddei GL. Langerhans cells in lichen sclerosus of the vulva and lichen sclerosus evolving in vulvar squamous cell carcinoma. *Histology and histopathology* 2009;24(3):331-6. doi: 10.14670/hh-24.331 [published Online First: 2009/01/09]
54. Scrimin F, Rustja S, Radillo O, et al. Vulvar lichen sclerosus: an immunologic study. *Obstetrics and gynecology* 2000;95(1):147-50. [published Online First: 2000/01/15]
55. Pahne-Zeppenfeld J, Schroer N, Walch-Ruckheim B, et al. Cervical cancer cell-derived interleukin-6 impairs CCR7-dependent migration of MMP-9-expressing dendritic cells. *International journal of cancer* 2014;134(9):2061-73. doi: 10.1002/ijc.28549 [published Online First: 2013/10/19]
56. Heusinkveld M, de Vos van Steenwijk PJ, Goedemans R, et al. M2 macrophages induced by prostaglandin E2 and IL-6 from cervical carcinoma are switched to activated M1 macrophages by CD4+ Th1 cells. *Journal of immunology (Baltimore, Md : 1950)* 2011;187(3):1157-65. doi: 10.4049/jimmunol.1100889 [published Online First: 2011/06/29]
57. Demoulin SA, Somja J, Duray A, et al. Cervical (pre)neoplastic microenvironment promotes the emergence of tolerogenic dendritic cells via RANKL secretion. *Oncoimmunology* 2015;4(6):e1008334. doi: 10.1080/2162402x.2015.1008334 [published Online First: 2015/07/15]
58. Sznurkowski JJ, Zawrocki A, Emerich J, et al. Expression of indoleamine 2,3-dioxygenase predicts shorter survival in patients with vulvar squamous cell carcinoma (vSCC) not influencing on the recruitment of FOXP3-expressing regulatory T cells in cancer nests. *Gynecologic oncology* 2011;122(2):307-12. doi: 10.1016/j.ygyno.2011.04.050 [published Online First: 2011/05/28]
59. Puccetti P, Grohmann U. IDO and regulatory T cells: a role for reverse signalling and non-canonical NF-kappaB activation. *Nature reviews Immunology* 2007;7(10):817-23. doi: 10.1038/nri2163 [published Online First: 2007/09/04]
60. Lee WS, Lee SM, Kim MK, et al. The tryptophan metabolite 3-hydroxyanthranilic acid suppresses T cell responses by inhibiting dendritic cell activation. *International immunopharmacology* 2013;17(3):721-6. doi: 10.1016/j.intimp.2013.08.018 [published Online First: 2013/09/14]
61. de Jong RA, Toppen NL, Ten Hoor KA, et al. Status of cellular immunity lacks prognostic significance in vulvar squamous carcinoma. *Gynecologic oncology* 2012;125(1):186-93. doi: 10.1016/j.ygyno.2011.12.416 [published Online First: 2011/12/14]
62. Jung AC, Guihard S, Krugell S, et al. CD8-alpha T-cell infiltration in human papillomavirus-related oropharyngeal carcinoma correlates with improved patient prognosis. *International journal of cancer* 2013;132(2):E26-36. doi: 10.1002/ijc.27776 [published Online First: 2012/08/15]
63. Nasman A, Romanitan M, Nordfors C, et al. Tumor infiltrating CD8+ and Foxp3+ lymphocytes correlate to clinical outcome and human papillomavirus (HPV) status in tonsillar cancer. *PloS one* 2012;7(6):e38711. doi: 10.1371/journal.pone.0038711 [published Online First: 2012/06/16]
64. Sznurkowski JJ, Zawrocki A, Emerich J, et al. Prognostic significance of CD4+ and CD8+ T cell infiltration within cancer cell nests in vulvar squamous cell carcinoma. *International journal of gynecological cancer : official journal of the International Gynecological Cancer Society* 2011;21(4):717-21. doi: 10.1097/IGC.0b013e3182131f36 [published Online First: 2011/05/06]
65. Sznurkowski JJ, Zawrocki A, Biernat W. Subtypes of cytotoxic lymphocytes and natural killer cells infiltrating cancer nests correlate with prognosis in patients with vulvar squamous cell carcinoma. *Cancer immunology, immunotherapy : CII* 2014;63(3):297-303. doi: 10.1007/s00262-013-1511-x [published Online First: 2013/12/26]
66. van Esch EM, Tummers B, Baartmans V, et al. Alterations in classical and nonclassical HLA expression in recurrent and progressive HPV-induced usual vulvar intraepithelial neoplasia and implications for immunotherapy. *International journal of cancer* 2014;135(4):830-42. doi: 10.1002/ijc.28713 [published Online First: 2014/01/15]

67. van Esch EM, Dam MC, Osse ME, et al. Clinical characteristics associated with development of recurrence and progression in usual-type vulvar intraepithelial neoplasia. *International journal of gynecological cancer : official journal of the International Gynecological Cancer Society* 2013;23(8):1476-83. doi: 10.1097/IGC.0b013e3182a57fd6 [published Online First: 2013/11/22]
68. Talebian Yazdi M, van Riet S, van Schadewijk A, et al. The positive prognostic effect of stromal CD8+ tumor-infiltrating T cells is restrained by the expression of HLA-E in non-small cell lung carcinoma. *Oncotarget* 2016;7(3):3477-88. doi: 10.18632/oncotarget.6506 [published Online First: 2015/12/15]
69. Gooden M, Lampen M, Jordanova ES, et al. HLA-E expression by gynecological cancers restrains tumor-infiltrating CD8(+) T lymphocytes. *Proceedings of the National Academy of Sciences of the United States of America* 2011;108(26):10656-61. doi: 10.1073/pnas.1100354108 [published Online First: 2011/06/15]
70. Terlouw A, Santegoets LA, van der Meijden WJ, et al. An autoimmune phenotype in vulvar lichen sclerosis and lichen planus: a Th1 response and high levels of microRNA-155. *The Journal of investigative dermatology* 2012;132(3 Pt 1):658-66. doi: 10.1038/jid.2011.369 [published Online First: 2011/11/25]
71. Regauer S. Immune dysregulation in lichen sclerosis. *European journal of cell biology* 2005;84(2-3):273-7. doi: 10.1016/j.ejcb.2004.12.003 [published Online First: 2005/04/12]
72. Ben-Hur H, Ashkenazi M, Huszar M, et al. Lymphoid elements and apoptosis-related proteins (Fas, Fas ligand, p53 and bcl-2) in lichen sclerosis and carcinoma of the vulva. *European journal of gynaecological oncology* 2001;22(2):104-9. [published Online First: 2001/07/12]
73. Farrell AM, Dean D, Millard PR, et al. Cytokine alterations in lichen sclerosis: an immunohistochemical study. *The British journal of dermatology* 2006;155(5):931-40. doi: 10.1111/j.1365-2133.2006.07414.x [published Online First: 2006/10/13]
74. Hunger RE, Bronnimann M, Kappeler A, et al. Detection of perforin and granzyme B mRNA expressing cells in lichen sclerosis. *Experimental dermatology* 2007;16(5):416-20. doi: 10.1111/j.1600-0625.2007.00543.x [published Online First: 2007/04/18]
75. Carlson JA, Grabowski R, Chichester P, et al. Comparative immunophenotypic study of lichen sclerosis: epidermotropic CD57+ lymphocytes are numerous--implications for pathogenesis. *The American journal of dermatopathology* 2000;22(1):7-16. [published Online First: 2000/03/04]
76. Gross T, Wagner A, Ugurel S, et al. Identification of TIA-1+ and granzyme B+ cytotoxic T cells in lichen sclerosis et atrophicus. *Dermatology (Basel, Switzerland)* 2001;202(3):198-202. doi: 10.1159/000051636 [published Online First: 2001/06/01]
77. Wenzel J, Wiechert A, Merkel C, et al. IP10/CXCL10 - CXCR3 interaction: a potential self-recruiting mechanism for cytotoxic lymphocytes in lichen sclerosis et atrophicus. *Acta dermato-venereologica* 2007;87(2):112-7. doi: 10.2340/00015555-0194 [published Online First: 2007/03/07]
78. Regauer S, Reich O, Beham-Schmid C. Monoclonal gamma-T-cell receptor rearrangement in vulvar lichen sclerosis and squamous cell carcinomas. *The American journal of pathology* 2002;160(3):1035-45. [published Online First: 2002/03/14]
79. Mannweiler S, Sygulla S, Beham-Schmid C, et al. Penile carcinogenesis in a low-incidence area: a clinicopathologic and molecular analysis of 115 invasive carcinomas with special emphasis on chronic inflammatory skin diseases. *The American journal of surgical pathology* 2011;35(7):998-1006. doi: 10.1097/PAS.0b013e3182147e59 [published Online First: 2011/06/18]
80. Gaarenstroom K, Kenter G, Trimbos J, et al. Postoperative complications after vulvectomy and inguinofemoral lymphadenectomy using separate groin incisions. 2003;13(4):522-27.
81. Pepas L, Kaushik S, Nordin A, et al. Medical interventions for high-grade vulval intraepithelial neoplasia. 2015;8(8):CD007924.
82. Adams S, Kozhaya L, Martiniuk F, et al. Topical TLR7 agonist imiquimod can induce immune-mediated rejection of skin metastases in patients with breast cancer. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2012;18(24):6748-57. doi: 10.1158/1078-0432.ccr-12-1149 [published Online First: 2012/07/07]
83. Schön MP, Schön M. Imiquimod: mode of action. *Br J Dermatol* 2007;157 Suppl 2:8-13. doi: 10.1111/j.1365-2133.2007.08265.x [published Online First: 2007/12/11]

84. Wenzel J, Uerlich M, Haller O, et al. Enhanced type I interferon signaling and recruitment of chemokine receptor CXCR3-expressing lymphocytes into the skin following treatment with the TLR7-agonist imiquimod. *J Cutan Pathol* 2005;32(4):257-62.
85. Beutner KR, Geisse JK, Helman D, et al. Therapeutic response of basal cell carcinoma to the immune response modifier imiquimod 5% cream. *Journal of the American Academy of Dermatology* 1999;41(6):1002-7. [published Online First: 1999/11/26]
86. de Witte CJ, van de Sande AJ, van Beekhuizen HJ, et al. Imiquimod in cervical, vaginal and vulvar intraepithelial neoplasia: a review. *Gynecologic oncology* 2015;139(2):377-84. doi: 10.1016/j.ygyno.2015.08.018 [published Online First: 2015/09/04]
87. Westermann C, Fischer A, Clad AJJoG, et al. Treatment of vulvar intraepithelial neoplasia with topical 5% imiquimod cream. 2013;120(3):266-70.
88. van Seters M, van Beurden M, ten Kate FJ, et al. Treatment of vulvar intraepithelial neoplasia with topical imiquimod. *The New England journal of medicine* 2008;358(14):1465-73. doi: 10.1056/NEJMoa072685 [published Online First: 2008/04/04]
89. van Poelgeest MI, van Seters M, van Beurden M, et al. Detection of human papillomavirus (HPV) 16-specific CD4+ T-cell immunity in patients with persistent HPV16-induced vulvar intraepithelial neoplasia in relation to clinical impact of imiquimod treatment. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2005;11(14):5273-80. doi: 10.1158/1078-0432.ccr-05-0616 [published Online First: 2005/07/22]
90. Terlou A, van Seters M, Kleinjan A, et al. Imiquimod-induced clearance of HPV is associated with normalization of immune cell counts in usual type vulvar intraepithelial neoplasia. *International journal of cancer* 2010;127(12):2831-40. doi: 10.1002/ijc.25302 [published Online First: 2011/02/26]
91. Daayana S, Elkord E, Winters U, et al. Phase II trial of imiquimod and HPV therapeutic vaccination in patients with vulvar intraepithelial neoplasia. *British journal of cancer* 2010;102(7):1129-36. doi: 10.1038/sj.bjc.6605611 [published Online First: 2010/03/18]
92. Winters U, Daayana S, Lear JT, et al. Clinical and immunologic results of a phase II trial of sequential imiquimod and photodynamic therapy for vulvar intraepithelial neoplasia. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2008;14(16):5292-9. doi: 10.1158/1078-0432.CCR-07-4760 [pii] 10.1158/1078-0432.CCR-07-4760 [published Online First: 2008/08/14]
93. Urošević M, Maier T, Benninghoff B, et al. Mechanisms underlying imiquimod-induced regression of basal cell carcinoma in vivo. *Archives of dermatology* 2003;139(10):1325-32. doi: 10.1001/archderm.139.10.1325 [published Online First: 2003/10/22]
94. Brown SB, Brown EA, Walker I. The present and future role of photodynamic therapy in cancer treatment. *The Lancet Oncology* 2004;5(8):497-508. doi: 10.1016/s1470-2045(04)01529-3 [published Online First: 2004/08/04]
95. Castano AP, Mroz P, Hamblin MR. Photodynamic therapy and anti-tumour immunity. *Nature reviews Cancer* 2006;6(7):535-45. doi: 10.1038/nrc1894 [published Online First: 2006/06/24]
96. Dougherty TJ, Gomer CJ, Henderson BW, et al. Photodynamic therapy. *Journal of the National Cancer Institute* 1998;90(12):889-905. [published Online First: 1998/06/24]
97. Abdel-Hady ES, Martin-Hirsch P, Duggan-Keen M, et al. Immunological and viral factors associated with the response of vulvar intraepithelial neoplasia to photodynamic therapy. *Cancer research* 2001;61(1):192-6. [published Online First: 2001/02/24]
98. van der Burg SH, Arens R, Ossendorp F, et al. Vaccines for established cancer: overcoming the challenges posed by immune evasion. *Nature reviews Cancer* 2016;16(4):219-33. doi: 10.1038/nrc.2016.16 [published Online First: 2016/03/12]
99. Welters MJ, Kenter GG, Piersma SJ, et al. Induction of tumor-specific CD4+ and CD8+ T-cell immunity in cervical cancer patients by a human papillomavirus type 16 E6 and E7 long peptides vaccine. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2008;14(1):178-87. doi: 10.1158/1078-0432.ccr-07-1880 [published Online First: 2008/01/04]
100. Kenter GG, Welters MJ, Valentijn AR, et al. Vaccination against HPV-16 oncoproteins for vulvar intraepithelial neoplasia. *N Engl J Med* 2009;361(19):1838-47. doi: 10.1056/NEJMoa0810097 [published Online First: 2009/11/06]

101. Davidson EJ, Boswell CM, Sehr P, et al. Immunological and clinical responses in women with vulval intraepithelial neoplasia vaccinated with a vaccinia virus encoding human papillomavirus 16/18 oncoproteins. *Cancer research* 2003;63(18):6032-41.
102. Kauppila S, Kotila V, Knuuti E, et al. The effect of topical pimecrolimus on inflammatory infiltrate in vulvar lichen sclerosis. *American journal of obstetrics and gynecology* 2010;202(2):181.e1-4. doi: 10.1016/j.ajog.2009.09.030 [published Online First: 2009/11/06]
103. Lee A, Bradford J, Fischer G. Long-term Management of Adult Vulvar Lichen Sclerosis: A Prospective Cohort Study of 507 Women. *JAMA dermatology* 2015;151(10):1061-7. doi: 10.1001/jamadermatol.2015.0643 [published Online First: 2015/06/13]
104. Yamamoto Y, Gaynor RB. Therapeutic potential of inhibition of the NF- κ B pathway in the treatment of inflammation and cancer. *The Journal of clinical investigation* 2001;107(2):135-42.
105. Mattern J, Büchler MW, Herr I. Cell cycle arrest by glucocorticoids may protect normal tissue and solid tumors from cancer therapy. *Cancer biology & therapy* 2007;6(9):1341-50.
106. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011;144(5):646-74.
107. van Poelgeest MI, Welters MJ, Vermeij R, et al. Vaccination against Oncoproteins of HPV16 for Noninvasive Vulvar/Vaginal Lesions: Lesion Clearance Is Related to the Strength of the T-Cell Response. *Clin Cancer Res* 2016;22(10):2342-50. doi: 10.1158/1078-0432.Ccr-15-2594 [published Online First: 2016/01/28]
108. Wang X, Teng F, Kong L, et al. PD-L1 expression in human cancers and its association with clinical outcomes. *OncoTargets and therapy* 2016;9:5023-39. doi: 10.2147/ott.s105862 [published Online First: 2016/08/31]
109. Sunshine J, Taube JM. PD-1/PD-L1 inhibitors. *Current opinion in pharmacology* 2015;23:32-8. doi: 10.1016/j.coph.2015.05.011 [published Online First: 2015/06/07]
110. Verdegaaal EME, van der Burg SH. The Potential and Challenges of Exploiting the Vast But Dynamic Neoepitope Landscape for Immunotherapy. *Frontiers in immunology* 2017;8:1113. doi: 10.3389/fimmu.2017.01113 [published Online First: 2017/09/30]
111. Ilyas S, Yang JC. Landscape of Tumor Antigens in T Cell Immunotherapy. *Journal of immunology (Baltimore, Md : 1950)* 2015;195(11):5117-22. doi: 10.4049/jimmunol.1501657 [published Online First: 2015/11/22]
112. Yarchoan M, Hopkins A, Jaffee EM. Tumor Mutational Burden and Response Rate to PD-1 Inhibition. *The New England journal of medicine* 2017;377(25):2500-01. doi: 10.1056/NEJMc1713444 [published Online First: 2017/12/21]
113. Lechner A, Schlosser H, Rothschild SI, et al. Characterization of tumor-associated T-lymphocyte subsets and immune checkpoint molecules in head and neck squamous cell carcinoma. *Oncotarget* 2017;8(27):44418-33. doi: 10.18632/oncotarget.17901 [published Online First: 2017/06/03]
114. Welters MJ, Ma W, Santegoets S, et al. Intratumoral HPV16-Specific T Cells Constitute a Type I-Oriented Tumor Microenvironment to Improve Survival in HPV16-Driven Oropharyngeal Cancer. *Clin Cancer Res* 2018;24(3):634-47. doi: 10.1158/1078-0432.Ccr-17-2140 [published Online First: 2017/10/12]
115. Howitt BE, Sun HH, Roemer MG, et al. Genetic Basis for PD-L1 Expression in Squamous Cell Carcinomas of the Cervix and Vulva. *JAMA oncology* 2016;2(4):518-22. doi: 10.1001/jamaoncol.2015.6326 [published Online First: 2016/02/26]
116. Tumeh PC, Harview CL, Yearley JH, et al. PD-1 blockade induces responses by inhibiting adaptive immune resistance. *Nature* 2014;515(7528):568-71. doi: 10.1038/nature13954 [published Online First: 2014/11/28]
117. Karim R, Jordanova ES, Piersma SJ, et al. Tumor-expressed B7-H1 and B7-DC in relation to PD-1+ T-cell infiltration and survival of patients with cervical carcinoma. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2009;15(20):6341-7. doi: 10.1158/1078-0432.ccr-09-1652 [published Online First: 2009/10/15]
118. Obeid JM, Erdag G, Smolkin ME, et al. PD-L1, PD-L2 and PD-1 expression in metastatic melanoma: Correlation with tumor-infiltrating immune cells and clinical outcome. *Oncoimmunology* 2016;5(11):e1235107. doi: 10.1080/2162402x.2016.1235107 [published Online First: 2016/12/22]

119. Heeren AM, Punt S, Bleeker MC, et al. Prognostic effect of different PD-L1 expression patterns in squamous cell carcinoma and adenocarcinoma of the cervix. *Modern pathology: an official journal of the United States and Canadian Academy of Pathology, Inc* 2016;29(7):753-63. doi: 10.1038/modpathol.2016.64 [published Online First: 2016/04/09]
120. Yu GT, Bu LL, Huang CF, et al. PD-1 blockade attenuates immunosuppressive myeloid cells due to inhibition of CD47/SIRPalpha axis in HPV negative head and neck squamous cell carcinoma. *Oncotarget* 2015;6(39):42067-80. doi: 10.18632/oncotarget.5955 [published Online First: 2015/11/18]
121. Ottenhof SR, Djajadiningrat RS, de Jong J, et al. Expression of Programmed Death Ligand 1 in Penile Cancer is of Prognostic Value and Associated with HPV Status. *The Journal of urology* 2017;197(3 Pt 1):690-97. doi: 10.1016/j.juro.2016.09.088 [published Online First: 2016/10/05]
122. Sznurkowski JJ, Zawrocki A, Sznurkowska K, et al. PD-L1 expression on immune cells is a favorable prognostic factor for vulvar squamous cell carcinoma patients. *Oncotarget* 2017;8(52):89903-12. doi: 10.18632/oncotarget.20911 [published Online First: 2017/11/23]
123. van der Burg SH, Piersma SJ, de Jong A, et al. Association of cervical cancer with the presence of CD4+ regulatory T cells specific for human papillomavirus antigens. *Proceedings of the National Academy of Sciences of the United States of America* 2007;104(29):12087-92. doi: 10.1073/pnas.0704672104 [published Online First: 2007/07/07]
124. Prendergast GC, Malachowski WP, DuHadaway JB, et al. Discovery of IDO1 Inhibitors: From Bench to Bedside. *Cancer research* 2017;77(24):6795-811. doi: 10.1158/0008-5472.can-17-2285 [published Online First: 2017/12/17]
125. Matheu MP, Othy S, Greenberg ML, et al. Imaging regulatory T cell dynamics and CTLA4-mediated suppression of T cell priming. *Nature communications* 2015;6:6219. doi: 10.1038/ncomms7219 [published Online First: 2015/02/06]
126. Trietsch MD, Spaans VM, ter Haar NT, et al. CDKN2A(p16) and HRAS are frequently mutated in vulvar squamous cell carcinoma. *Gynecologic oncology* 2014;135(1):149-55. doi: 10.1016/j.ygyno.2014.07.094 [published Online First: 2014/07/30]
127. Spaans VM, Trietsch MD, Crobach S, et al. Designing a high-throughput somatic mutation profiling panel specifically for gynaecological cancers. *PLoS one* 2014;9(3):e93451. doi: 10.1371/journal.pone.0093451 [published Online First: 2014/03/29]
128. Jung YS, Najy AJ, Huang W, et al. HPV-associated differential regulation of tumor metabolism in oropharyngeal head and neck cancer. *Oncotarget* 2017;8(31):51530-41. doi: 10.18632/oncotarget.17887 [published Online First: 2017/09/09]
129. Haratani K, Hayashi H, Tanaka T, et al. Tumor immune microenvironment and nivolumab efficacy in EGFR mutation-positive non-small-cell lung cancer based on T790M status after disease progression during EGFR-TKI treatment. *Annals of oncology: official journal of the European Society for Medical Oncology* 2017;28(7):1532-39. doi: 10.1093/annonc/mdx183 [published Online First: 2017/04/14]
130. Rizvi NA, Hellmann MD, Snyder A, et al. Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science* 2015;348(6230):124-8. doi: 10.1126/science.aaa1348 [published Online First: 2015/03/15]
131. Hodge JW, Ardiani A, Farsaci B, et al. The tipping point for combination therapy: cancer vaccines with radiation, chemotherapy, or targeted small molecule inhibitors. *Seminars in oncology* 2012;39(3):323-39. doi: 10.1053/j.seminoncol.2012.02.006 [published Online First: 2012/05/19]
132. Stevanovic S, Pasetto A, Helman SR, et al. Landscape of immunogenic tumor antigens in successful immunotherapy of virally induced epithelial cancer. *Science (New York, NY)* 2017;356(6334):200-05. doi: 10.1126/science.aak9510 [published Online First: 2017/04/15]
133. van Poelgeest MI, Visconti VV, Aghai Z, et al. Potential use of lymph node-derived HPV-specific T cells for adoptive cell therapy of cervical cancer. *Cancer immunology, immunotherapy: CII* 2016;65(12):1451-63. doi: 10.1007/s00262-016-1892-8 [published Online First: 2016/09/14]