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Metabolism and lipid mediators as regulators of innate immune cell function: implications for inflammation and immune responses

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Hyperinflammation by Human Macrophages Induced by SARS-CoV-2 Anti-Spike IgG Is Dependent on Glucose and Fatty Acid Metabolism

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Abstract

Severe COVID-19 is an immunological disorder characterized by excessive immune activation following infection with SARS-CoV-2, which typically occurs around the time of seroconversion. Anti-spike IgG of critically ill COVID-19 patients induces excessive inflammation by activation of Fc gamma receptors (FcγRs) on human alveolar macrophages, leading to tissue damage, pulmonary edema, and coagulopathy. While metabolic reprogramming of immune cells is critical for the induction of inflammatory responses, still little is known about the metabolic pathways that are involved in COVID-19-specific hyper-inflammation. In this study, we identified that anti-spike IgG immune complexes (ICs) induce rapid metabolic reprogramming of alveolar macrophages, which is essential for induction of inflammation. Through functional inhibition, we identified that glycolysis, fatty acid synthesis and pentose phosphate pathway (PPP) activation are critical for anti-spike IgG-induced hyper-inflammation. Remarkably, while excessive pro-inflammatory cytokine production by macrophages is critically dependent on simultaneous stimulation with viral stimuli and anti-spike IgG complexes, we show that the required metabolic reprogramming is specifically driven by anti-spike IgG complexes. These findings provide new insights into the metabolic pathways driving hyper-inflammation by macrophages in the context of severe COVID-19. Targeting of these pathways may reveal new possibilities to counteract pathological inflammatory responses in severe COVID-19 and related diseases.

Introduction

The recent pandemic has prominently shown how airborne viruses such as SARS-CoV-2 can cause a significant risk to public health [1]. Even though the fast development of highly efficient vaccines has successfully dissolved the acute situation in most countries, breakthrough infections and the occurrence of new variants of concern still especially endanger risk groups such as immunocompromised, obese, elderly or diabetic individuals [1-5]. It is thus crucial to further investigate the mechanisms underlying the development of severe COVID-19 as well as to find specific drugs to treat patients experiencing severe disease progression.

Anti-spike IgG immune complexes (ICs) play a key role in the development of severe COVID-19 progression by activation of Fc receptors (FcRs) [6-8]. Patients experiencing severe COVID-19 develop a pathological antibody response characterized by high titers of anti-spike IgG with a distinct glycosylation pattern of their Fc tail at position 297, with low amounts of fucose and high amounts of galactose [8]. Decreased fucosylation of IgG antibodies is generally observed in response to surface -exposed membrane-embedded antigens [9], which has been described to occur in a variety of anti-viral immune responses such as in dengue virus [10] or HIV infected individuals [11]. However, characteristic for severe COVID-19 is the induction of a hyper-inflammatory response by human alveolar macrophages through antibody-dependent inflammation (ADI), leading to tissue damage, pulmonary edema, and coagulopathy [6-8, 12].

In recent years, it has become increasingly clear that changes in cellular metabolism are an essential mechanism actively shaping the immune response [13]. Pro-inflammatory macrophages are characterized by an increased dependency on glycolysis and pentose phosphate pathway (PPP), combined with a decreased utilization of the TCA cycle, β -oxidation and oxidative phosphorylation (OXPHOS)[14-16]. In contrast, macrophages with a wound-healing phenotype mainly depend on an intact TCA cycle and OXPHOS as main energy source [15-17].

Crosstalk of IC-induced FcR signaling and Toll-like receptor (TLR) activation plays a crucial role in boosting effector function of myeloid cells such as pro-inflammatory cytokine induction in various tissues [18, 19]. For human dendritic cells, we previously identified that FcR-mediated cytokine secretion strongly relies on IRF5 regulated metabolic reprogramming towards a highly glycolytic phenotype [20]. Furthermore, rapid upregulation of glycolytic flux is an essential factor for fueling fatty acid synthesis and driving myeloid cell activation in a pro-inflammatory milieu [21].

Several studies have already revealed the impact of metabolic changes in SARS-CoV-2 infection and replication [22-24]. However, the role of metabolic reprogramming in pathogenic antibody-mediated inflammation in the context of severe COVID-19 is not yet well defined. In this study, we investigated metabolic reprogramming of human alveolar-like macrophages upon exposure to pathological antibodies, and the role of these metabolic changes in the development of hyper-inflammation. We found that specifically anti-spike IgG, but not viral stimuli, induced pronounced changes in cellular metabolism in human alveolar-like macrophages. Moreover, we identified that inhibition of these metabolic pathways efficiently blocks anti-spike IgG induced pro-inflammatory cytokine induction, and thus may be a promising therapeutic target to counteract the effects of pathogenic antibodies in patients experiencing severe COVID-19.

Methods

Key resources table

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies		
COVA1-18 WT	P.J.M Brouwer et al. [25]	doi:10.1126/science.abc5902
Severe COVID-19 patient serum	Amsterdam UMC COVID-19 Biobank	N/A
Glut1 – Dylight 405	Heieis et al. [26]	doi:10.1038/s41467-023-41353-z
PKM – PE	Heieis et al. [26]	doi:10.1038/s41467-023-41353-z
SDHA – AF647	Heieis et al. [26]	doi:10.1038/s41467-023-41353-z
CPT1A – PE-Cy5	Heieis et al. [26]	doi:10.1038/s41467-023-41353-z
ACC1 – Pe-Cy7	Heieis et al. [26]	doi:10.1038/s41467-023-41353-z
G6PD – APC-Cy7	Heieis et al. [26]	doi:10.1038/s41467-023-41353-z
ATP5a – Dylight 488	Heieis et al. [26]	doi:10.1038/s41467-023-41353-z
CD36 – BV605	BD Biosciences	Cat# 563518
CD98 – BUV395	BD Biosciences	Cat# 744508
Human TruStain FcX™	Biolegend	Cat# 422301
HIF-1α	Invitrogen	Cat# 12-7528-80
FcIIb block	Biolegend	Cat# 398302

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Chemicals, peptides, and recombinant proteins		
Human M-CSF	Miltenyi Biotec	Cat#130-096-491
Recombinant Human IL-10 Protein	R&D Systems	Cat# 217-IL-025/CF
Recombinant SARS-CoV2-Spike Wuhan Hu-1 Protein	T.Caniels et al. [27]	GenBank accession MN908947.3
Zombie NIR™ Fixable Viability Kit	Biolegend	Cat# 423106
2-DG	MedChemExpress.com	Cat# HY-13966
BPTES	MedChemExpress.com	Cat# HY-12683
C75	MedChemExpress.com	Cat# HY-12364
Etomoxir	MedChemExpress.com	Cat# HY-50202
6-AN	MedChemExpress.com	Cat# HY-W010342
Oligomycin	Sigma-Aldrich	Cat# 75351-5MG
UK-5099	MedChemExpress.com	Cat# HY-15475
polyinosinic:polycytidylic acid	Sigma-Aldrich	Cat#P1530
MitoTracker™ Deep Red FM	ThermoFisher	Cat#M224626
Critical commercial assays		
CD14 MicroBeads, human	Miltenyi Biotec	Cat#130-050-201
MitoProbe™ TMRM Assay Kit for Flow Cytometry	ThermoFisher	Cat#M20036
2-(N-(7-Nitrobenz-2-oxa-1,3-diazol-4-yl) Amino)-2-Deoxyglucose) (2-NBDG)	ThermoFisher	Cat# N13195
4,4-Difluoro-4-bora-3a,4a-diaza-s-indacene (BODIPY)	ThermoFisher	Cat# D3821
ELISA MAX™ Standard Set Human IL-6	BioLegend	Cat#430501
Software and algorithms		
GraphPad Prism version 9.4.0	GraphPad Software	www.graphpad.com
SpectroFlo v.5	Cytek	www.cytexbio.com
FlowJo v. 10	BD Biosciences	www.flowjo.com

Monocyte-derived alveolar macrophage-like macrophages

Buffy coats from healthy anonymous donors were purchased from Sanquin blood supply (Amsterdam, NL). All donors provided written consent prior to blood donation. Briefly, monocytes were isolated via Lymphoprep isolation (Stemcell) followed by magnetic bead separation using the MACS cell separation system (Miltenyi). The purified monocytes were differentiated for 6 days Iscove's modified Dulbecco's medium (IMDM, Gibco) containing 5 % fetal calf serum (CAPRICORN) and Pencillin/Steptomycin (Thermo Fisher) supplemented with 50 ng/ml human M-CSF (Miltenyi). The culture medium was refreshed after three culture days. To generate an alveolar macrophage like phenotype the medium was replaced by culture medium containing 50 ng/ml human IL-10 (R&D) 24 h prior stimulation.

Coating

Stabilized recombinant SARS-CoV-2 spike protein and monoclonal anti-spike IgG1 (COVA1-18) were generated as previously described [25, 27]. Spike protein specific ICs were generated by coating 2 µg/ml spike protein over-night on 96-well high affinity plates (Nunc). Subsequently, the plates were blocked with 10 % FCS in PBS for 1 hour at 37 °C to prevent unspecific binding. Monoclonal anti-spike IgG1 (2ug/ml) or severe COVID-19 patient serum (1% in PBS) was added to the stimulation plates and incubated for 1 h at 37 °C followed by washing the plates with PBS to remove unbound protein.

Cell stimulation and inhibitor treatment

All inhibitors were purchased in powdered form and dissolved according to the manufacturer's instructions. Human macrophages were diluted to a concentration of 277777 cells/ml and pre-treated with the indicated inhibitors (or DMSO as control) for 30 minutes at 37 °C. After pre-incubation, macrophages were added to the stimulation plate supplemented with either culture medium or with 20 µg/ml polyinosinic:polycytidylic acid (poly(I:C), Sigma-Aldrich). For FcγRIIb block, cells were pre-incubated with 20 µg/ml anti-FcγRIIb (Biolegend Cat# 398302) for 30 minutes at 4 °C. Prior to stimulation this was adjusted to a final concentration of 5 µg/ml.

Enzyme-linked immunosorbent assay (ELISA)

To determine cytokine production, supernatants of the simulated human macrophages were harvested after 6 h or 24 h. IL-6 concentration was measured using antibody pairs from Biolegend (ELISA MAX™ Standard Set Human IL-6, 430501).

Seahorse metabolic analysis

Spike protein specific ICs were generated in an XFe96 well Seahorse plate (Agilent) as described above. Prior to the experiment the Seahorse XFe96 cartridge was hydrated with Seahorse XF calibrant (Agilent). 50000 alveolar-like macrophages per

well were added to the Seahorse plate in XF assay medium on RPMI basis (Sigma) supplemented with 2 mM glutamine (ThermoFisher), 5 % FCS and with or without 20 µg/ml poly(I:C). Injection compounds were diluted in Seahorse XF medium and added to the previously prepared cartridge and loaded to the machine for calibration. Mito Stress Test Injections (final concentrations): 10 mM Glucose (Sigma Aldrich), 1.5 µM Oligomycin A (Cayman), 3 µM FCCP (Sigma Aldrich), 1 µM Rotenone (Sigma Aldrich) and Antimycin A (Sigma Aldrich). Glycolysis stress test Injections: 10 mM Glucose (Sigma Aldrich), 1.5 µM Oligomycin A (Cayman), 10 mM 2-DG (Medchemexpress.com).

MitoTracker DR and TMRM staining

Cells were stimulated as described above. 15 min prior stimulation time end CCCP was added to the CCCP control conditions in a final concentration of 50 µM. Cells were harvested and washed with pre-warmed IMDM with-out supplements. Samples were stained with 2,5 nM MitoTracker DR and 3 nM TMRM for 15 min at 37 °C protected from light. After staining cells were once washed with IMDM + 1 % FCS and twice with cold PBS. Live cells were stained using Zombie Violet Dye (BioLegend). Cells were measured immediately after staining procedure at BD LSR Fortessa (BD Biosciences). Analysis was performed with FlowJo v.10 (BD Biosciences)

Glucose and fatty acid uptake

Alveolar-like macrophages were stimulated as mentioned above and cells were harvested after 1h of stimulation time. Cells were stained with 2-NBDG (ThermoFisher) diluted in pre-warmed 37 °C PBS in a concentration of 100 µM and BODIPY C16 (ThermoFisher) in a concentration of 20 nM. Live cells were stained using Zombie-NIR viability dye (Biolegend). Samples were acquired on a Cytex Aurora 5 L spectral flow cytometer. Spectral unmixing was performed with SpectroFlow v.5 and further analysis was executed with FlowJo v.10 (BD Biosciences).

Spectral flowcytometry

Antibodies for metabolic targets were generated as previously described [26]. 600000 cells per condition were pre-stained with viability dye (Biolegend) and Fc-blocking solution (Biolegend) for 20 min on ice. Following fixation with Fixation buffer (Biolegend) for 15 min on ice, surface marker staining was performed in FACS buffer for 1h on ice. Cells were permeabilized with 1 x Perm buffer (ThermoFisher). Followed by intracellular metabolic targets staining and HIF-1α (Invitrogen Cat# 12-7528-80) staining for 2 h at RT. Samples were acquired on a Cytex Auora 5L spectral flow cytometer. Spectral unmixing was performed using SpectroFlow v.5. Samples were further analyzed with FlowJo v.10 (BD Biosciences).

Propidium iodide (PI) assay

Human macrophages were cultured in presence of the selected metabolic inhibitors for 24 h. Following the inhibitor treatment the supernatant was replaced with serum free IMDM supplemented with 3 μ M PI reagent (Sigma-Aldrich, P4170). After 30 minutes incubation under culture conditions, extracellular DNA content was measured by analyzing fluorescence intensities at $\lambda_{\text{ex}}/\lambda_{\text{em}} = 530/620$ nm.

Lactate assay

Supernatant of stimulated human macrophages was deproteinized by addition of a final concentration of 3 % w/v metaphosphoric acid (MPA). Followed by incubation with 27 mM NAD (Roche) solution in 0.5M glycine-0.4M hydrazine buffer (pH 9.0). Lactate production was measured by lactate dehydrogenase (LDH, Roche) treatment. After 30 min incubation time with LDH at RT lactate content was measured by NADH fluorescence at $\lambda_{\text{ex}}/\lambda_{\text{em}} = 340/450$.

Quantification and Statistical Analysis

Statistical significance of the data was performed using GraphPad Prism 9.4.0 (GraphPad). The statistical analysis applied for each figure is stated in the corresponding Figure legend.

Results

Individual Stimulation With Anti-Spike IgG Rapidly Boosts Glycolysis and Oxidative Phosphorylation

To assess if stimulation with anti-spike IgG ICs changes core metabolic pathways in the context of severe COVID-19, we examined changes in glycolysis and oxidative phosphorylation (OXPHOS) of monocyte-derived alveolar-like macrophages after short time exposure. Previous data has shown that *in vitro* stimulation of monocyte-derived alveolar-like human macrophages with anti-spike IgG ICs serves as a promising experimental model to study antibody-induced inflammation of alveolar macrophages in severe COVID-19 [8]. We tested the effect of the recombinant anti-spike IgG COVA1-18, which we generated previously from B cells isolated from a patient with COVID-19 [28]. In addition, we generated anti-spike-IgG ICs by incubating SARS-CoV-2 spike-coated wells with diluted serum from patients with severe COVID-19 treated in the first pandemic wave at the intensive care unit (ICU) at the Amsterdam

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University Medical Centers (UMC)[8]. In line with previous studies[8, 29], stimulation of alveolar-like macrophages with only viral stimulus poly(I:C) (reflecting the first phase of infection), did not elevate pro-inflammatory cytokine production. In contrast, upon co-stimulation of cells with a viral stimulus and anti-spike IgG ICs, mimicking the post-seroconversion phase of severe COVID-19 infection, high amounts of pro-inflammatory cytokine IL-6 were secreted (Fig. 1A,B).

Extracellular metabolic flux analysis of human macrophages treated with anti-spike IgG ICs showed an increased but not significant change in extracellular acidification rate (ECAR), indicative of a slightly enhanced glycolysis (Fig. 1C, Fig. S1 A).

In pro-inflammatory macrophages elevated glycolysis is associated with a decreased OXPHOS associated metabolism [15, 30]. Interestingly, the extracellular metabolic flux data revealed a rapid increase in oxygen consumption rate (OCR) representative of elevated OXPHOS utilization after anti-spike IgG IC stimulation (Fig. 1D,E, Fig. S1 B). Thus, our data indicates, that anti-spike IgG IC stimulation induces upregulation of both core metabolic pathways that boost ATP synthesis. Remarkably, in contrast to pro-inflammatory cytokine induction which requires a strong interplay of FcR signaling and TLR activating signal, anti-spike IgG ICs alone were sufficient to induce changes in glycolysis and basal respiration (Fig. 1C-E, Fig. S1 A,B) and poly(I:C) had no synergistic effect.

Macrophages resembling a pro-inflammatory phenotype are classically characterized by an increased dependence on glycolysis, fatty acid synthesis and the PPP [15, 17]. In this pro-inflammatory condition, pyruvate is reduced into lactate instead of being used to fuel the TCA cycle [15, 31]. Thus, lactate secretion can serve as a simple measure to determine alterations of the glycolytic pathway and reduced pyruvate metabolism in activated macrophages [32]. To further investigate the kinetics of anti-spike IgG ICs induced metabolic reprogramming we measured lactate secretion in the medium of anti-spike IgG stimulated macrophages after 1 h, 6 h and 24 h activation time. After 1 h of stimulation time, no differences between the stimulation conditions could be observed (Fig. 1F). In the next time frame, after 6 h of stimulation time, a small increase in lactate production in the samples stimulated with recombinant anti-spike IgG ICs or ICs derived from ICU patient serum could be detected (Fig. 1G). 24 h after activation the secretion of lactate in the anti-spike IgG ICs-treated cells was further elevated significantly (Fig. 1H). In line with the extracellular metabolic flux data these changes were mainly induced by anti-spike IgG stimulation, as the addition of a co-stimulatory TLR-activating agent did not further affect lactate secretion.

Since antibody stimulation significantly increased glycolytic activity and OXPHOS of alveolar-like macrophages, we assessed whether glucose and fatty acid uptake would

be increased upon stimulation as well. We visualized glucose and fatty acid uptake by tracking uptake of fluorescent glucose 2-NBDG and fluorescent BODIPY, respectively. There was a trend towards increased 2-NBDG uptake after 1 h of stimulation in the conditions stimulated with anti-spike IgG immune complexes (Fig. S1C), however, these changes did not reach statistical significance. Furthermore, no changes in fatty acid uptake were observed between the different stimulation conditions (Fig. S1D).

An important driver of glycolytic metabolism and release of pro-inflammatory cytokines is hypoxia induced factor (HIF)-1 α [33, 34]. Therefore, we investigated whether anti-spike IgG IC stimulation changed HIF-1 α expression in human alveolar-like macrophages. After 1h of stimulation time, no differences in HIF-1 α protein expression were observed between the different conditions (Fig. S1E). In contrast, after 24 h of stimulation time there was a trend towards elevated HIF-1 α expression in anti-spike IgG IC-treated conditions (Fig. S1F). Interestingly, as previously observed in the metabolic flux data, individual anti-spike IgG IC treatment was sufficient to induce this effect, and no further increase was observed in cells additionally treated with co-stimulatory signal poly(I:C).

Taken together these data indicate that anti-spike IgG IC stimulation induces upregulation of both metabolic core pathways boosting ATP synthesis, i.e. glycolysis and OXPHOS.

Anti-Spike IgG Induces Rapid Upregulation of Specific Metabolic Enzymes

To further characterize changes induced in relevant metabolic pathways upon anti-spike IgG IC stimulation, we analyzed the expression of a selection of core metabolic enzymes via Metflow analysis [26]. Briefly, human alveolar-like macrophages were stimulated with anti-spike IgG ICs as shown above (Fig. 1A,B). Following 6 h stimulation, metabolic targets were stained and measured using spectral flow cytometry (Fig. 2A). In line with the extracellular metabolic flux (Fig. 1) and 2-NBDG data (Fig S1D), stimulation of human macrophages with anti-spike IgG ICs significantly increased the expression of glucose transporter GLUT1 (Fig. 2B). Interestingly, even though stimulation with anti-spike IgG increased GLUT1 expression and elevated the glycolytic flux, PKM expression was not significantly elevated (Fig. 2C), potentially indicating an increased flux of glucose into upstream offshoots of glycolysis such as the PPP. Indeed, stimulation with anti-spike IgG ICs induced elevated expression of PPP associated enzyme G6PD (Fig. 2D). Furthermore, consistent with increased OXPHOS, anti-spike IgG ICs promoted expression of SDHA (Fig. 2E), an enzyme in

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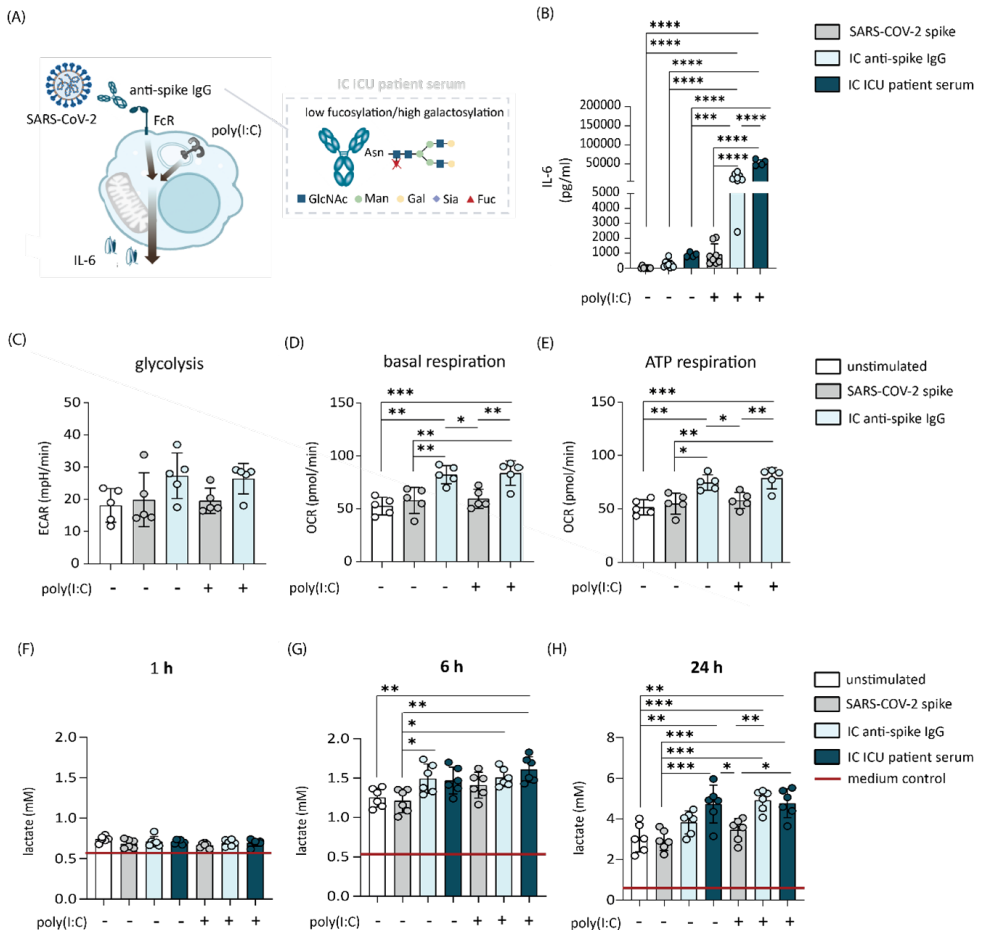


Fig. 1. Stimulation with anti-SARS-CoV-2 spike IgG rapidly boosts glycolysis and OXPHOS in human alveolar-like macrophages. (A) anti-spike IgG ICs induce pro-inflammatory cytokine induction of alveolar macrophages in combination with TLR activating signal (poly(I:C)). Antibodies derived from COVID-19 ICU patients are characterized by an aberrant glycosylation pattern (B) IL-6 production by human alveolar-like macrophages after 24 h stimulation with ICs from recombinant anti-spike IgG or serum from unvaccinated patients diagnosed with severe COVID-19 (mean + SD). (C) glycolysis, (D) basal respiration and (E) ATP production of human alveolar-like macrophages. Stimulation for 1 h followed by seahorse assay (n = 5, mean + SD). (F-H) lactate production of human alveolar-like macrophages stimulated as indicated for 1 h, 6 h and 24 h (n = 6, mean + SD). IMDM medium with-out cells was added as control (red line). Significant differences were calculated with One-way ANOVA. *P<0.05; **P<0.01; ***P<0.001; ****P<0.0001

the TCA cycle and complex II enzyme of the respiratory chain. Finally, a trend towards increased expression of amino acid transporter CD98 (Fig. 2F) was visible after anti-spike IgG IC stimulation, which became more prominent after 24 h stimulation time (Fig. S2). No significant changes in the expression of metabolic enzymes CPT1A, ACC1, CD36 and ATP5a could be detected (Fig. 2G-J). In line with the extracellular

metabolic flux data and lactate assay the changes in expression of G6PD, CD98 and SDHA were mainly induced by anti-spike IgG IC stimulation and remained elevated for at least 24 h after stimulation (Fig. S2).

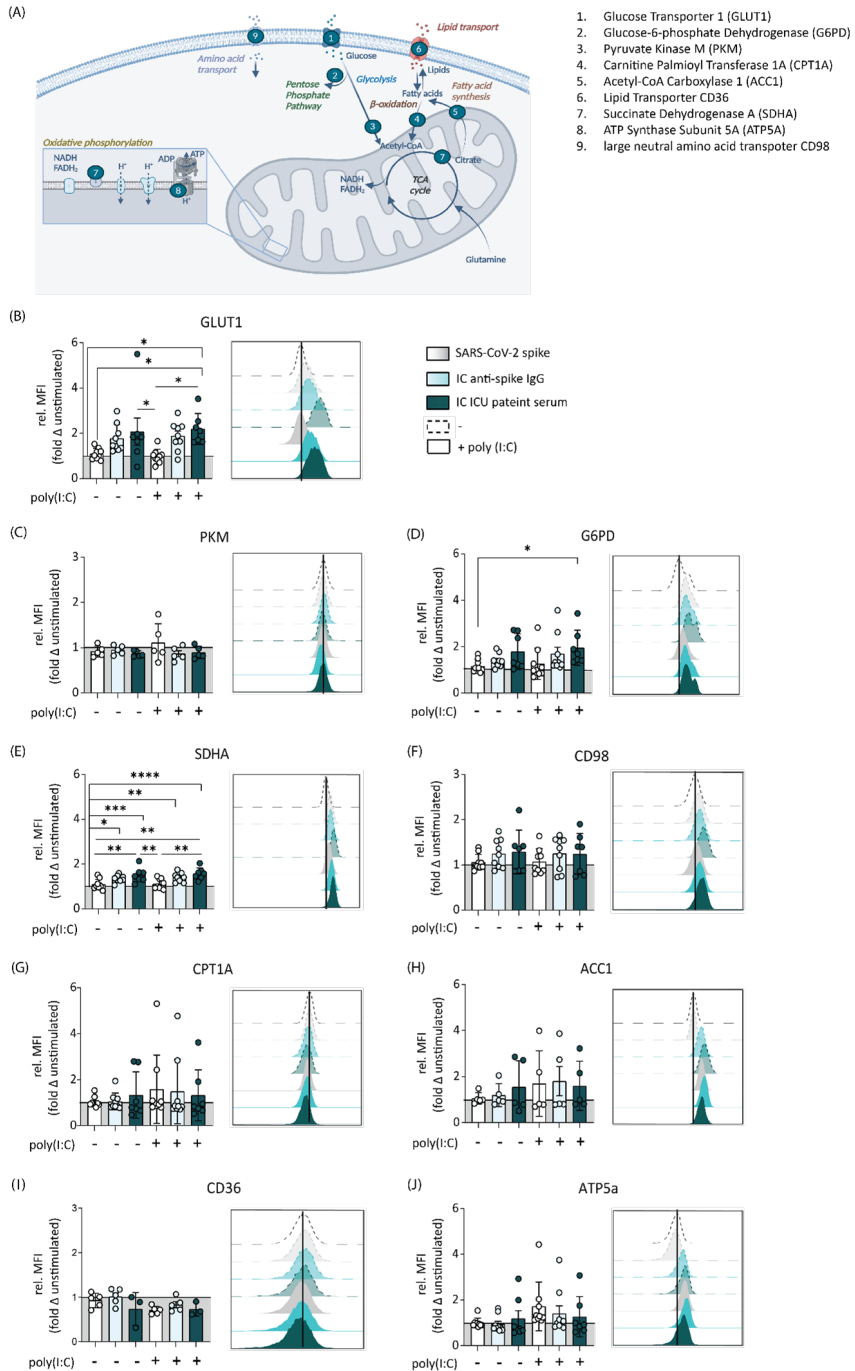
Since Fc-mediated antibody effector functions correlate with expression and type of FcR, we measured expression levels of the different FcγR on human alveolar-like macrophages. As shown in Figure S4A FcγRI (CD64) and FcγRIII (CD64) were expressed at similar levels, while FcγRIIb/c (CD32 b/c) showed a slightly lower and FcγRIIa/b (CD32a/b) increased expression on human alveolar-like macrophages (Fig. S4A). In contrast to other receptors of the FcR family expressed on human alveolar-like macrophages, FcγRIIb (CD32b) cross-linking inhibits activating signals by phosphorylation of its immunoreceptor tyrosine-based inhibitory motif (ITIM) in the cytoplasmic domain [35]. We therefore determined whether block of FcγRIIb would further amplify the metabolic changes induced by anti-spike IgG IC. Notably, FcγRIIb inhibition indeed further increased GLUT-1 expression of alveolar-like macrophages, especially when additionally treated with costimulatory signal poly(I:C) (Fig. S4B).

Anti-Spike IgG Triggers Rapid Changes in Macrophage Mitochondria

To further understand effect of pathogenic anti-spike IgG on mitochondrial function of human alveolar-like macrophages in severe COVID-19, we analyzed mitochondrial mass and membrane potential in response to anti-spike IgG IC stimulation, using Mito Tracker Deep Red [36, 37] and Tetramethylrhodamine methyl ester (TMRM) probes [38], respectively. After 1 h of stimulation, human alveolar-like macrophages stimulated with recombinant anti-spike IgG and ICs derived from severe COVID-19 ICU patient serum showed a significantly increased Mito Tracker Deep Red (DR) signal (Fig. 3A), as well as an elevated mitochondrial membrane potential (Fig. 3B). After 6 h of stimulation time the changes in Mito Tracker DR accumulation remained stable while the TMRM signal was decreased (Fig. 3A,B).

Taken together, these data suggest that anti-spike IgG stimulation induces rapid changes in mitochondrial membrane potential and mass, suggesting first signs of mitochondrial damage, such as mitochondrial membrane depolarization.

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(legend on next page)

Fig. 2. Anti-spike IgG induces upregulation of specific metabolic enzymes. (A) Schematic representation of Metflow analysis targets. Created in BioRender. Geyer, C. (2024) BioRender.com/h10r049 (B-J) Changes in relative MFI of metabolic enzymes of human macrophages after 6 h stimulation with ICs of recombinant IgG or ICU patient serum with or without viral stimulus poly(I:C). Representative histograms (unstimulated indicated as black line) and data points from individual donors (n = 5-8, mean + SD). Gating strategy is show in Figure S3. Significant differences were calculated with One-way ANOVA. *P<0.05; **P<0.01; ***P<0.001; ****P<0.0001

Anti-Spike IgG-Induced Hyper-Inflammation Is Functionally Dependent on Glycolysis, PPP, and Fatty Acid Synthesis

To assess if the identified changes in metabolic pathway activity are functionally required for anti-spike IgG IC-induced hyper-inflammation, we tested the effect of metabolic pathway inhibitors on the pro-inflammatory cytokine induction of human alveolar-like macrophages stimulated with anti-spike IgG ICs with or without poly(I:C). In line with the extracellular metabolic flux and metabolic enzyme expression data (Fig. 1,2), IL-6 production induced by co-stimulation of human alveolar-like macrophages was blocked by inhibition of glycolysis (by 2-DG) and PPP (by 6-AN) (Fig.4 A,B).

Interestingly, even though most donors did not show changes in ACC1 expression (Fig. 2H) blocking fatty acid synthesis with C75 efficiently reduced IL-6 production to the level of unstimulated macrophages, suggesting an important functional role from fatty acid synthesis in anti-spike IgG IC-induced inflammation. On other hand, neither inhibition of the mitochondrial pyruvate carrier by UK-5099 nor β -oxidation using Etomoxir affected the antibody induced pro-inflammatory cytokine production (Fig. 4 A,B). Despite the strong upregulation of basal respiration seen in the extracellular flux assay (Fig. 1D,E) blocking OXPHOS with oligomycin, only induced a small but still significant decrease in IL-6 induction (Fig. 4). This may be explained by the kinetics of the anti-spike IgG IC-induced metabolic reprogramming, which was dependent on OXPHOS mainly early after stimulation (Fig. 1 D,E). Of note, none of the inhibitors induced cell death at the indicated concentrations (Fig. S5).

Finally, we set out to validate these findings with anti-spike IgG from severely ill patients, by stimulating cells with serum obtained from first-wave COVID-19 ICU patients. Importantly, ICU patient serum displayed a similar dependency on glycolysis, fatty acid synthesis and PPP to promote elevated IL-6 induction (Fig. 4B). Moreover, when we compared the percentage of inhibition, the metabolic inhibitors suppressed cytokine induction by recombinant and patient-derived anti-spike IgG in a strikingly similar manner (Fig. 4C). This suggests that the higher cytokine induction by patient-derived IgG (Fig. 1B) results from quantitative, but not qualitative differences in metabolic reprogramming.

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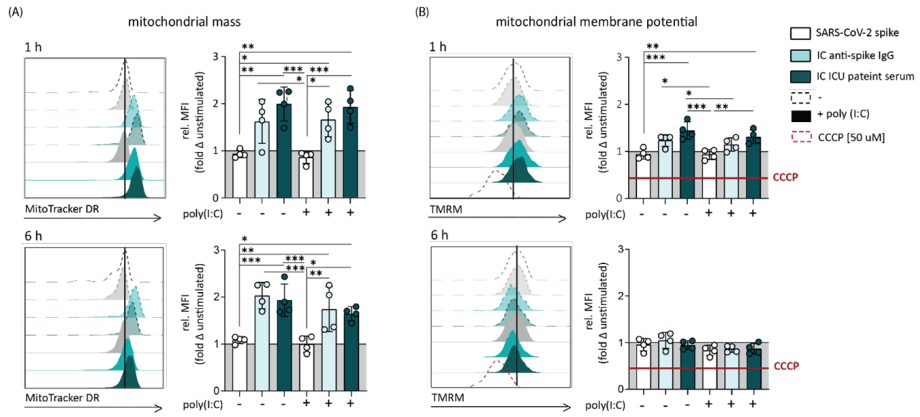


Fig. 3: Anti-spike IgG IC rapidly induces rapid changes in human macrophage mitochondria. Representative data of independent experiments (left, unstimulated indicated as black line) MitoTracker DR (A), TMRM (B). Data points from individual donors (right, n = 4, mean + SD). Human alveolar-like macrophages were stimulated as indicated and immediately stained with MitoTracker DR (3 nM) and TMRM (3 nM). 15 min prior stimulation end 50 uM CCCP was added to induce mitochondrial depolarization in TMRM samples as negative control. Gating strategy is show in Figure S3. Significant differences were calculated with One-way ANOVA. *P<0.05; **P<0.01; ***P<0.001; ****P<0.0001



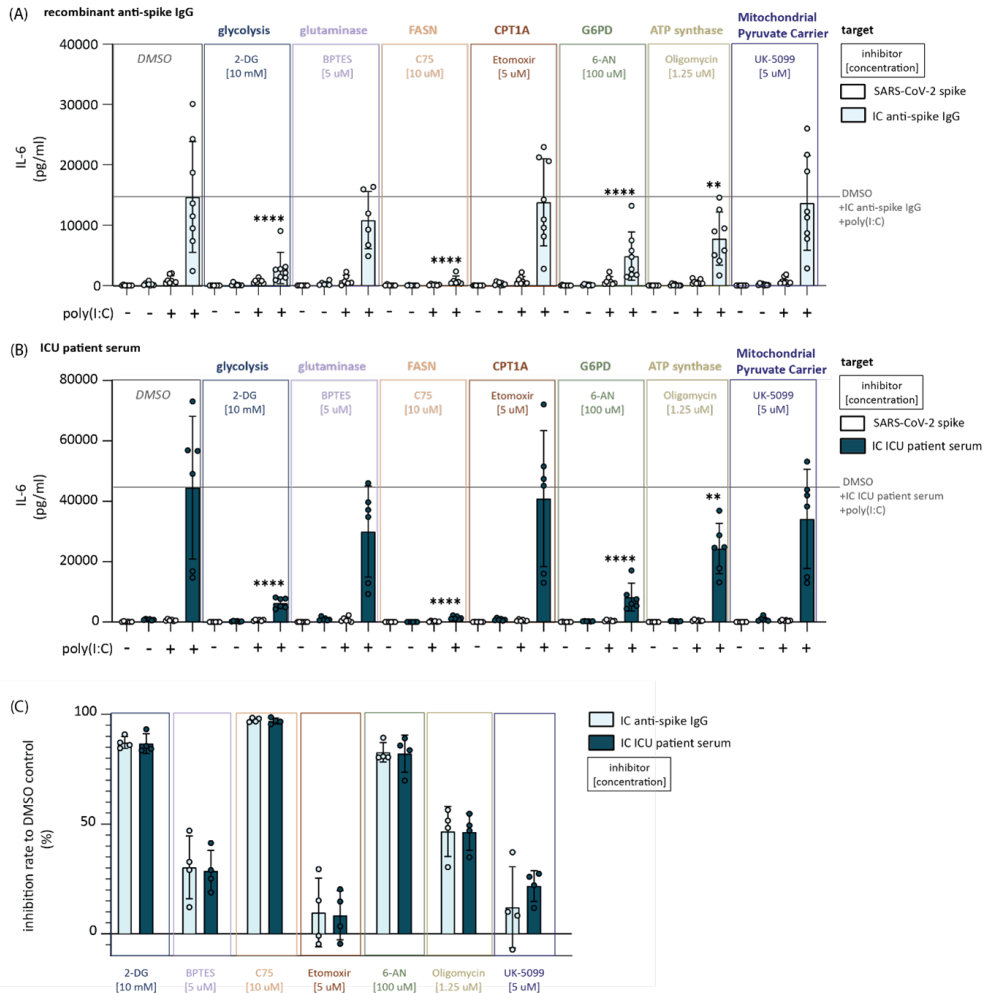


Fig. 4: Anti-spike IgG-induced pro-inflammatory cytokine induction is functionally depended on glycolysis, fatty acid synthesis and PPP (A) IL-6 secretion of human alveolar-like macrophages stimulated with anti-spike IgG IC and poly(i:C) after treatment with the indicated metabolic inhibitors. Each data point represents one individual donor (n = 4-8, mean + SD). stimulation with immune complexes derived from recombinant anti-spike IgG (A) or ICU patient serum (B) inhibition rate compared to DMSO control (C). Significant differences were calculated with one-way ANOVA. *P<0.05; **P<0.01; ***P<0.001; ****P<0.0001. Statistics indicate changes between inhibitor treated stimulation condition compared to identical stimulation condition in DMSO control sample.

Discussion

Severe COVID-19 is a immunological disorder, characterized by hyper-inflammation of macrophages and monocytes that cause tissue destruction [8, 39] edema, and coagulopathy [12, 40]. While metabolic reprogramming of immune cells is critical for immune activation [13], the metabolic pathways that enable hyper-inflammation in the context of severe COVID-19 are still largely unknown. In this study, we set out to investigate the role of antibody-induced metabolic reprogramming in severe disease progression to identify potential therapeutic targets for treatment of severe COVID-19.

In general, metabolic reprogramming towards high glycolysis provides important building blocks for transcription and promotes generation of reactive oxygen species which are beneficial for the host immune system leading to a rapid and effective pathogen clearance [15, 41]. In contrast, in severe COVID-19 patients alteration of glucose metabolism, such as increased glycolysis in BALF macrophages [42, 43] are associated with pathogenic over-activation of the immune response and correlated with a poor disease prognosis [43-46].

Here we show that anti-spike IgG ICs, as present upon seroconversion, may contribute to severe disease outcome by rapidly triggering metabolic reprogramming of alveolar macrophages towards a highly glycolytic phenotype. Blocking this glycolytic reprogramming by 2-DG treatment, counteracted the hyper-inflammatory immune reaction. Thus, 2-DG may be an efficient target to control the pathogenic over-activation in severe COVID-19. In phase II and III clinical trials performed in India in patients with moderate-to-severe COVID-19 progression, treatment with 2-DG showed alleviation of symptoms [47, 48]. These studies thus provide first proof of principle that reducing glycolytic activity in severe COVID-19 patients could have therapeutic potential. However, previous studies done in cancer patients have shown that 2-DG can induce potential side effects including hypoglycemia, nausea and fatigue. However, these side effects were non-life-threatening if the doses were limited to a maximum of 63 mg/kg [49, 50]. Nevertheless, it is yet unclear whether 2-DG treatment would interfere with the standard treatment protocol of severe COVID-19 including antiviral drugs and systematic corticosteroids [51]. Therefore, while 2-DG is a potentially promising treatment for severe COVID-19, further evaluation of the optimal treatment doses, administration time and target patient group needs to be performed in additional randomized clinical trials.

Our experiments also show that anti-spike IgG IC strongly upregulates PPP-associated enzyme G6PD in human macrophages. In addition, our data demonstrate that antibody-induced cytokine secretion crucially depends on PPP activity, as inhibition

of this pathway via 6-AN treatment efficiently blocks anti-spike IgG IC-induced IL-6. Polidatin, a molecule naturally occurring in *Polygonum cuspidatum*, directly inhibits G6PD [52]. Phase II clinical studies testing supportive treatment with Polydatin in the context of inflammatory bowel disease and endometriosis-related pain indicated no toxic effects in humans for a treatment dose of 20-40 mg [53, 54]. Given the importance of the PPP for the generation of biosynthesis precursor molecules and NADPH, this pathway may be an additional promising therapeutic target to counterbalance the excessive cytokine induction in severe COVID-19 patients [23]. Even though the PPP inhibitor Polidatin is a promising drug candidate due to its low toxicity in humans, further *in vitro* studies followed by randomized clinical trials would be required to evaluate efficiency and specificity in severe COVID-19 disease treatment.

It has been established that inflammatory macrophages tend to rely on *de novo* fatty acid synthesis to maintain cellular functions, instead of increasing fatty acid uptake [55-60]. Indeed, several studies have identified fatty acid synthesis as an essential link of metabolic reprogramming and cytokine secretion in myeloid cells [21, 57, 61, 62]. By fostering ER and Golgi expansion, fatty acid synthesis supports cytokine induction of myeloid cells in a pro-inflammatory milieu [21, 63]. Inhibition via C75 furthermore blocks IL-1 β secretion in murine macrophages in the context of sepsis [62]. This is further supported by recent studies where both ACC and fatty acid synthase (FASN) have been implicated in the metabolic rewiring of macrophages and in the subsequent synthesis of pro-inflammatory cytokines, such as IL-6 and IL-1 β [64, 65]. In line with this, our data reveal no role for the process of lipid uptake in the inflammatory response (as evidenced by the lack of differences in BODIPY C16 staining), and instead show that inhibition of FASN via C75 effectively blocks anti-spike IgG IC-induced IL-6 cytokine secretion, suggesting an important role for *de novo* fatty acid synthesis in antibody-induced pro-inflammatory cytokine induction. Combined with the increased glycolysis and PPP enhancement, these data suggest a similar molecular mechanism underlying cytokine secretion as described for dendritic cells [21], thereby linking increased glycolytic flux and fatty acid synthesis as essential key pathways for pro-inflammatory cytokine production. Enhanced fatty acid synthesis was shown to induce NLRP3 inflammasome activation and pro-inflammatory cytokine synthesis [65]. Additionally, it has been suggested that fatty acid synthesis may also trigger increased membrane synthesis, supporting the ER and Golgi expansion required for the FcR-triggered increase of pro-inflammatory cytokine transcription [8, 20, 21]. The increased *de novo* fatty acid synthesis is fueled by NADPH produced by the elevated glycolytic flux enhancing PPP activation [21]. Interestingly, in our dataset, ACC1 expression showed a mixed pattern of increased expression in some donors upon anti-spike IgG IC stimulation, while in other donors the expression levels remained stable. ACC1 expression is reported to mainly reflect the cell's ability for fatty acid synthesis [66]. Since obese severely ill COVID-19 patients show aberrant levels of

blood lipids [67] and altered ACC1 levels have been associated with the development of obesity [68], the role of ACC1 expression in the development of severe COVID-19 disease progression may be of interest for further studies.

High levels of the pro-inflammatory cytokine IL-6 is a key factor in the progression of severe COVID-19, and is associated with increased mortality [69, 70]. Even though antibody-induced pro-inflammatory cytokine induction requires a strong cross-talk between FcR and TLR signaling [8], our data indicate that anti-spike IgG IC-mediated metabolic reprogramming is mainly controlled by FcR signaling. This is in line with a recent study that demonstrated that FcR-mediated increase in glycolysis boosts inflammation in lupus nephritis [71]. This FcR-mediated glycolytic switch, independent of a costimulatory signal, may therefore be relevant for inflammatory responses beyond the context of COVID-19, including rheumatoid arthritis (RA) [72], Sjögren's Syndrome [73] or systemic lupus erythematosus (SLE) [71].

IgG affinity towards FcRs and affiliated induction of effector functions are strongly dependent on changes in the antibody glycosylation pattern [74, 75]. Afucosylated antibodies from patients amplify inflammation by increased binding to FcγRIII [76], while recombinant anti-spike IgG (with conventional Fc glycosylation) predominantly activates FcγRIIa signaling [8]. Interestingly, in contrast to cytokine secretion, our data indicate that metabolic reprogramming induced by anti-spike IgG is not influenced by alterations in antibody glycosylation pattern (Fig. 4). This may relate to overlapping signaling pathways downstream of FcRs. Both FcγRIIa and FcγRIII activate Syk and PI3K signaling upon antibody crosslinking [77, 78], which has been identified as key molecules that mediate anti-spike IgG IC-induced hyper-inflammatory responses by alveolar macrophages [8, 29]. Data by Jing et al. [71] indicate that FcR-mediated glycolytic activation is functionally dependent on Syk and PI3K activation, suggesting a potential molecular link between FcγR activation and metabolic reprogramming via Syk and PI3K activation. Interestingly, as opposed to the activating FcγRs that promote metabolic rewiring, we find the inhibitory receptor FcγRIIb does the opposite, suggesting that the net degree and possible nature of FcγR-driven metabolic alterations are shaped by balance between engagement of activating and inhibitory FcγRs.

Mitochondrial dysfunction plays a role in increased stress and inflammation contributing to the worsening of symptom severity in COVID-19 patients [79-82]. In addition, risk groups such as elderly patients are characterized by generally reduced mitochondrial health [83]. Our data show that rapidly in the kinetics of the anti-spike IgG-induced inflammatory response, alveolar-like macrophage mitochondria show first signs of alternated membrane potential and function. After 1 h of stimulation, the TMRM signal and respiration in anti-spike IgG IC stimulated cells was increased, indicating that

the initial response is characterized by increased mitochondrial activity. However, this appeared not to be functionally relevant for heightened cytokine production as oligomycin treatment has minimal effects on this. Interestingly, after 6 h of stimulation the first signs of a loss of TMRM staining were visible, indicating the presence of depolarized mitochondria, and an indication of a loss of mitochondrial activity and potentially damage [84]. Of note, the Mito Tracker staining was increased at time points 1 h and 6 h upon stimulation with anti-spike IgG ICs. Given that mitochondrial biosynthesis in macrophages is reported to be occurring in a time frame of several hours [85], these changes are probably not induced by increased mitochondrial biosynthesis but may rather reflect changes in mitochondrial morphology. Since mitochondrial damage has been associated with symptoms persisting even after the virus has been cleared [86-88], further investigation of the role of anti-spike IgG ICs in the development of mitochondrial dysfunction may be of interest for further studies even beyond macrophages. For instance, platelets of severe COVID-19 patients have also been reported to display signs of damaged mitochondria and bioenergetics failure [89]. Given that platelets express FcγRIIIa and that pathogenic antibodies of severe COVID-19 patients can directly activate platelets [90] to contribute to thrombosis in severe COVID-19 [12], it is conceivable that metabolic reprogramming by IgG IC during severe COVID-19 applies to a wider range of cells.

A limitation of our study is that the *in vitro* conditions of our model are not able to completely mimic the metabolic niche of alveolar macrophages in severe COVID-19 patients. Furthermore, since it was not possible to analyze bronchoalveolar lavage (BAL) macrophages derived from severe COVID-19 patients, we used a model closely mimicking alveolar macrophages based on transcriptomic signature [91] as well as on alveolar macrophage surface marker expression (data not shown). However, the use of healthy donors for the generation of alveolar-like macrophages is not taking into account potential pre-existing metabolic deficiencies in alveolar macrophages of severe COVID-19 risk groups individuals.

Taken together, we here show that the hyper-inflammatory response induced in the context of severe COVID-19 critically depends on anti-spike IgG IC-induced metabolic reprogramming of macrophages towards a highly glycolytic phenotype. This mechanism is mainly dependent on FcγR signaling and characterized by rapid kinetics. Moreover, we show that IgG IC-mediated inflammation can be counteracted by inhibiting glycolysis, fatty acid synthesis, and the pentose phosphate pathway. Therefore, these metabolic pathways may be promising targets to counteract the effects of pathological antibodies in patients suffering from severe COVID-19.

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Statement of Ethics

This study protocol was reviewed and approved by Commissie Toetsing Biobanken (CTB, Amsterdam UMC, The Netherlands), approval number 2023.0978. All participants provided written informed consent to Amsterdam UMC COVID-19 Biobank. Buffy coats from healthy donors were purchased from Sanquin blood supply (Amsterdam, The Netherlands). Donors provided written informed consent prior blood donation to Sanquin.



Conflict of Interest

The authors have no conflicts of interest to declare.

Funding Sources

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Author Contributions

Conceptualization RHH, BE, and JdD; Methodology CEG, LA, GH, RHH, BE and JdD; Validation CEG; Formal analysis CEG; Investigation CEG, LA, LM and HJC; Resources LA, GH, JV, SWdT, TGC, TPLB, MJvG, MdW, Amsterdam UMC COVID-19 Biobank, JVdB Writing – Original Draft CEG; Writing – Review and Editing LA, LM, GAH, SWdT, HJC, TGC, TPLB, MJvG, MdW, JVdB, RHH, BE and JdD. Visualization CEG; Supervision RHH, BE and JdD; Funding Acquisition BE and JdD.

Data Availability Statement

All data associated with this study are presented in the paper or supplementary materials. The recombinant anti-Spike IgG1 antibody COVA1-18 is available upon request to the corresponding authors through a materials transfer agreement. Further information and requests for resources, and reagents should be directed to and will be fulfilled by the corresponding author, Jeroen den Dunnen (j.dendunnen@amsterdamumc.nl)

Supplementary Materials

Table S1. Members and affiliation of the Amsterdam UMC COVID-19 Biobank

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Chapter III

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M.R. Smit	Department of Intensive Care
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W. Stilma	Department of Intensive Care
C.E. Teunissen	Neurochemical Laboratory
P. Thorat	Department of Intensive Care
A.M. Tsonas	Department of Intensive Care
P.R. Tuinman	Department of Intensive Care
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Hyperinflammation by Human Macrophages Induced by SARS-CoV-2 Anti-Spike IgG Is Dependent on Glucose and Fatty Acid Metabolism

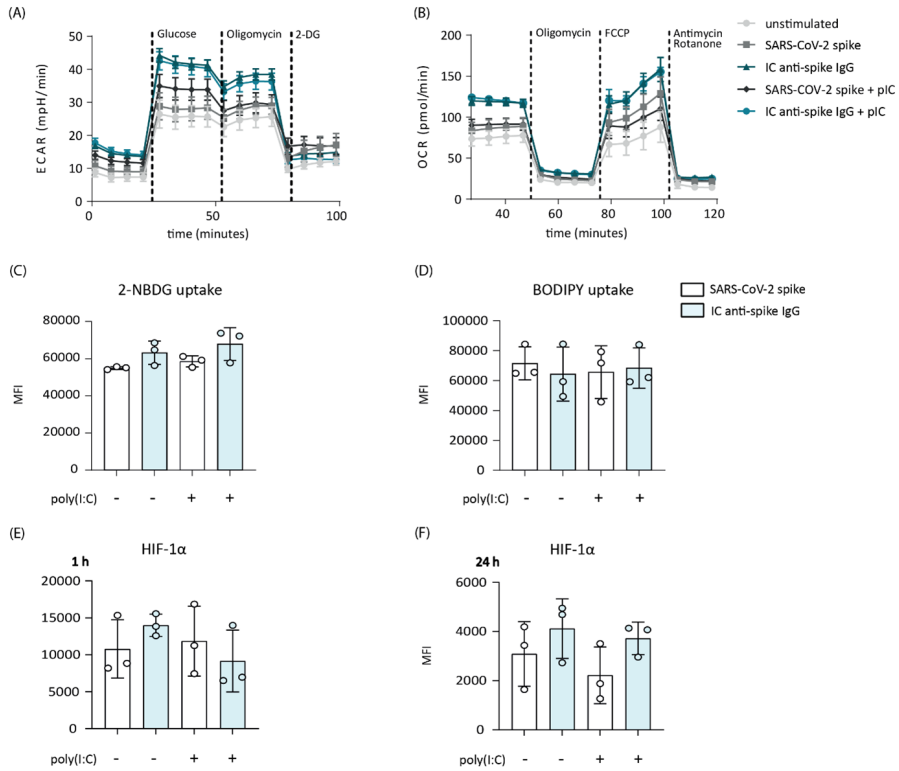


Figure S1: Metabolic changes of alveolar-like macrophages induced by anti-spike IgG stimulation (A) Real-time changes in ECAR (A) and OCR (B) of human alveolar-like macrophages stimulated with ICs from recombinant anti-spike IgG as indicated for a stimulation time of 1 h (Representative donor, mean + SEM). (C) glucose uptake quantified by glucose analogue 2-NBDG uptake (C), and fatty acid uptake, quantified by BODIPY uptake (D) (n=3, mean + SD) of human alveolar-like macrophages after 1 h of stimulation time. (E) changes of HIF-1 α expression in alveolar-like macrophages upon stimulation after 1 h and 24 h (n=3, mean + SD).

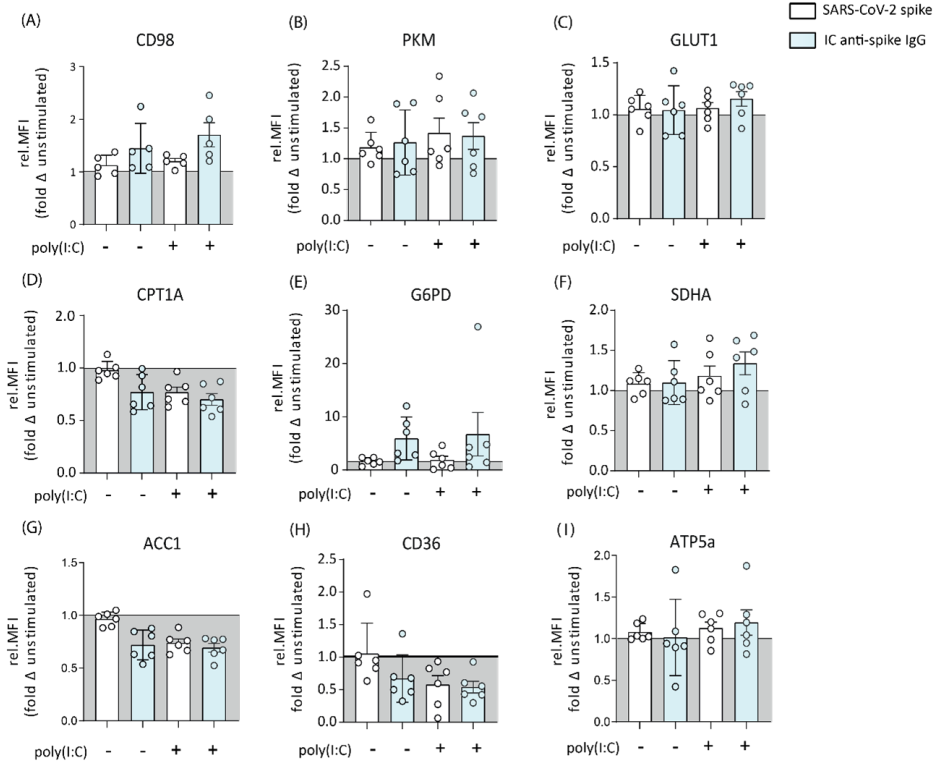


Figure S2. Anti-spike IgG induced change in metabolic enzymes after 24 h stimulation. (A-I) Changes in relative MFI of metabolic enzymes of human macrophages after 24 h stimulation with ICs of recombinant with or without viral stimulus poly(I:C). Data points from individual donors (n = 5, mean + SD).

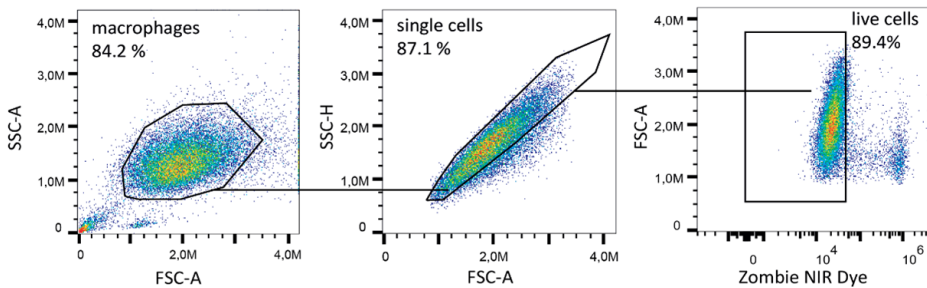


Figure S3: Gating strategy FACS. We selected human-like alveolar macrophages by applying a forward- and side-scatter gate. From this gate singles cells were selected followed by exclusion of dead cells by a live cell gate (Zombie NIR negative population).

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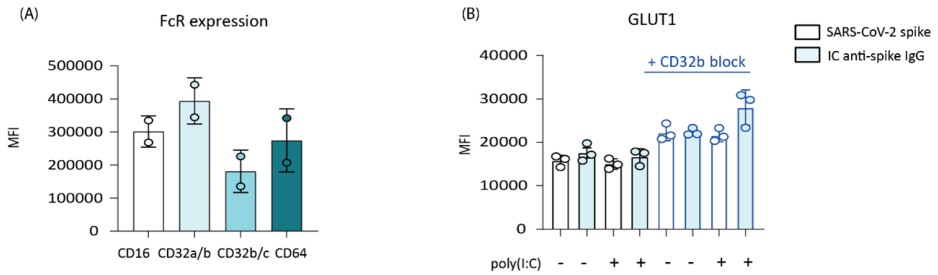


Figure S4. Fc γ R expression levels on human alveolar-like macrophages and effect of Fc γ RIIb blockage on anti-spike IgG IC induced changes in GLUT1 expression. (A) Human alveolar-like macrophages were harvested and stained with CD16-PE, CD32a/b-PerCP, CD32b/c-APC-A700, CD64-ECD to determine FcR expression levels. Representative data from $n = 2$ individual experiments (mean + SD). (B) GLUT1 expression of alveolar-like macrophages treated with FcRIIb blocking antibody (blue) compared to untreated cells (black). Samples were stimulated for 24 h stimulation with ICs of recombinant anti-spike IgG with or without viral stimulus poly(I:C) ($n = 3$, mean + SD).

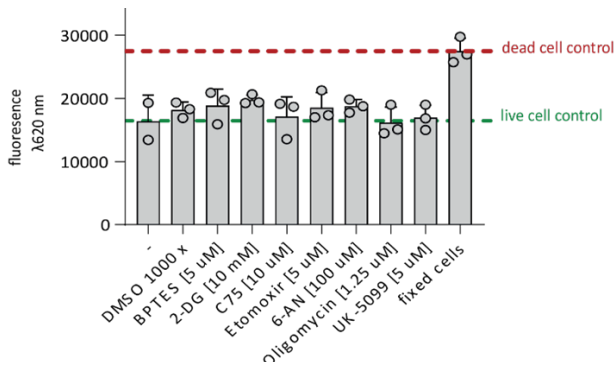


Figure S5. Effect of metabolic inhibitors on cell viability of human alveolar-like macrophages. Human macrophages were incubated with the indicated inhibitor concentration under culture conditions for 24 h. Afterwards, extracellular DNA content was determined via PI staining. Increased extracellular DNA content leads to elevated fluorescence at 620 nm. Representative data from $n = 4$ individual experiments (mean + SD).

References

- 1 Harris, M., Hart, J., Bhattacharya, O. and Russell, F. M., Risk factors for SARS-CoV-2 infection during the early stages of the COVID-19 pandemic: a systematic literature review. *Front Public Health* 2023. 11: 1178167.
- 2 Alberca, R. W., Oliveira, L. d. M., Branco, A. C. C. C., Pereira, N. Z. and Sato, M. N., Obesity as a risk factor for COVID-19: an overview. *Critical Reviews in Food Science and Nutrition* 2021. 61: 2262-2276.
- 3 Sawadogo, W., Tsegaye, M., Gizaw, A. and Adera, T., Overweight and obesity as risk factors for COVID-19-associated hospitalisations and death: systematic review and meta-analysis. *BMJ Nutr Prev Health* 2022. 5: 10-18.
- 4 Evans, R. A., Dube, S., Lu, Y., Yates, M., Arnetorp, S., Barnes, E., Bell, S., Carty, L., Evans, K., Graham, S., Justo, N., Moss, P., Venkatesan, S., Yokota, R., Ferreira, C., McNulty, R., Taylor, S. and Quint, J. K., Impact of COVID-19 on immunocompromised populations during the Omicron era: insights from the observational population-based INFORM study. *Lancet Reg Health Eur* 2023. 35: 100747.
- 5 Liu, B., Spokes, P., He, W. and Kaldor, J., High risk groups for severe COVID-19 in a whole of population cohort in Australia. *BMC Infect Dis* 2021. 21: 685.
- 6 Chakraborty, S., Gonzalez, J., Edwards, K., Mallajosyula, V., Buzzanco, A. S., Sherwood, R., Buffone, C., Kathale, N., Providenza, S., Xie, M. M., Andrews, J. R., Blish, C. A., Singh, U., Dugan, H., Wilson, P. C., Pham, T. D., Boyd, S. D., Nadeau, K. C., Pinsky, B. A., Zhang, S., Memoli, M. J., Taubenberger, J. K., Morales, T., Schapiro, J. M., Tan, G. S., Jagannathan, P. and Wang, T. T., Proinflammatory IgG Fc structures in patients with severe COVID-19. *Nat Immunol* 2021. 22: 67-73.
- 7 Chakraborty, S., Gonzalez, J. C., Sievers, B. L., Mallajosyula, V., Chakraborty, S., Dubey, M., Ashraf, U., Cheng, B. Y., Kathale, N., Tran, K. Q. T., Scallan, C., Sinnott, A., Cassidy, A., Chen, S. T., Gelbart, T., Gao, F., Golan, Y., Ji, X., Kim-Schulze, S., Prahl, M., Gaw, S. L., Gnjatich, S., Marron, T. U., Merad, M., Arunachalam, P. S., Boyd, S. D., Davis, M. M., Holubar, M., Khosla, C., Maecker, H. T., Maldonado, Y., Mellins, E. D., Nadeau, K. C., Pulendran, B., Singh, U., Subramanian, A., Utz, P. J., Sherwood, R., Zhang, S., Jagannathan, P., Tan, G. S. and Wang, T. T., Early non-neutralizing, afucosylated antibody responses are associated with COVID-19 severity. *Sci Transl Med* 2022. 14: eabm7853.
- 8 Hoepel, W., Chen, H. J., Geyer, C. E., Allahverdiyeva, S., Manz, X. D., de Taeye, S. W., Aman, J., Mes, L., Steenhuis, M., Griffith, G. R., Bonta, P. I., Brouwer, P. J. M., Caniels, T. G., van der Straten, K., Golebski, K., Jonkers, R. E., Larsen, M. D., Linty, F., Nouta, J., van Roomen, C., van Baarle, F., van Drunen, C. M., Wolbink, G., Vlaar, A. P. J., de Bree, G. J., Sanders, R. W., Willemsen, L., Neele, A. E., van de Beek, D., Rispens, T., Wuhrer, M., Bogaard, H. J., van Gils, M. J., Vidarsson, G., de Winther, M. and den Dunnen, J., High titers and low fucosylation of early human anti-SARS-CoV-2 IgG promote inflammation by alveolar macrophages. *Sci Transl Med* 2021. 13.
- 9 Larsen, M. D., de Graaf, E. L., Sonneveld, M. E., Plomp, H. R., Nouta, J., Hoepel, W., Chen, H.-J., Linty, F., Visser, R., Brinkhaus, M., Šuštić, T., de Taeye, S. W., Bentlage, A. E. H., Toivonen, S., Koeleman, C. A. M., Sainio, S., Kootstra, N. A., Brouwer, P. J. M., Geyer, C. E., Derksen, N. I. L., Wolbink, G., de Winther, M., Sanders, R. W., van Gils, M. J., de Bruin, S., Vlaar, A. P. J., COVID-19, A. U., group†, b. s., Rispens, T., den Dunnen, J., Zaaier, H. L., Wuhrer, M., Ellen van der Schoot, C., Vidarsson, G., van Aagtmael, M., Algera, A. G., van Baarle, F., Bax, D., van de Beek, D., Beudel, M., Bogaard, H. J., Bonta, P. I., Bomers, M., Bos, L., Botta, M., de Bree, G., Brouwer, M. C., Brabander, J., de Bruin, S., Bugiani, M., Bulle, E., Chouchane, O., Cloherty, A., Elbers, P., Fleuren, L., Geerlings, S., Geerts, B., Geijtenbeek, T., Girbes, A., Goorhuis, B., Grobusch, M. P., Hafkamp, F., Hagens, L., Hamann, J., Harris, V., Hemke, R., Hermans, S. M., Heunks, L., Hollmann, M. W., Horn, J., Hovius, J. W., de Jong, M., Koning, R., van Mourik, N., Nellen, J., Nossent, E. J., Paulus, F., Peters, E., van der Poll, T., Preckel, B., Prins, J. M.,

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- Raasveld, J., Rijnders, T., Schinkel, M., Schultz, M., Schuurmans, A. R., Sigaloff, K., Smit, M., Stijnis, C. S., Stilma, W., Teunissen, C., Thorat, P., Tsonas, A., van der Valk, M., Veelo, D., Vlaar, A. P. J., de Vries, H., van Vugt, M., Wiersinga, W. J., Wouters, D., Afucosylated IgG characterizes enveloped viral responses and correlates with COVID-19 severity. *Science* 2021. 371: eabc8378.
- 10 Wang, T. T., Sewatanon, J., Memoli, M. J., Wrammert, J., Bournazos, S., Bhaumik, S. K., Pinsky, B. A., Chokephaibulkit, K., Onlamoon, N., Pattanapanyasat, K., Taubenberger, J. K., Ahmed, R. and Ravetch, J. V., IgG antibodies to dengue enhanced for FcγRIIIA binding determine disease severity. *Science* 2017. 355: 395-398.
- 11 Ackerman, M. E., Crispin, M., Yu, X., Baruah, K., Boesch, A. W., Harvey, D. J., Dugast, A. S., Heizen, E. L., Ercan, A., Choi, I., Streeck, H., Nigrovic, P. A., Bailey-Kellogg, C., Scanlan, C. and Alter, G., Natural variation in Fc glycosylation of HIV-specific antibodies impacts antiviral activity. *J Clin Invest* 2013. 123: 2183-2192.
- 12 Bye, A. P., Hoepel, W., Mitchell, J. L., Jégouic, S., Loureiro, S., Sage, T., Vidarsson, G., Nouta, J., Wuhrer, M., de Taeye, S., van Gils, M., Kriek, N., Cooper, N., Jones, I., den Dunnen, J. and Gibbins, J. M., Aberrant glycosylation of anti-SARS-CoV-2 spike IgG is a prothrombotic stimulus for platelets. *Blood* 2021. 138: 1481-1489.
- 13 O'Neill, L. A. and Pearce, E. J., Immunometabolism governs dendritic cell and macrophage function. *J Exp Med* 2016. 213: 15-23.
- 14 Newsholme, P., Curi, R., Gordon, S. and Newsholme, E. A., Metabolism of glucose, glutamine, long-chain fatty acids and ketone bodies by murine macrophages. *Biochem J* 1986. 239: 121-125.
- 15 Viola, A., Munari, F., Sánchez-Rodríguez, R., Scolaro, T. and Castegna, A., The Metabolic Signature of Macrophage Responses. *Front Immunol* 2019. 10: 1462.
- 16 Jha, A. K., Huang, S. C., Sergushichev, A., Lampropoulou, V., Ivanova, Y., Loginicheva, E., Chmielewski, K., Stewart, K. M., Ashall, J., Everts, B., Pearce, E. J., Driggers, E. M. and Artyomov, M. N., Network integration of parallel metabolic and transcriptional data reveals metabolic modules that regulate macrophage polarization. *Immunity* 2015. 42: 419-430.
- 17 Liu, Y., Xu, R., Gu, H., Zhang, E., Qu, J., Cao, W., Huang, X., Yan, H., He, J. and Cai, Z., Metabolic reprogramming in macrophage responses. *Biomark Res* 2021. 9: 1.
- 18 Vogelpoel, L. T. C., Hansen, I. S., Rispens, T., Muller, F. J. M., van Capel, T. M. M., Turina, M. C., Vos, J. B., Baeten, D. L. P., Kapsenberg, M. L., de Jong, E. C. and den Dunnen, J., Fc gamma receptor-TLR cross-talk elicits pro-inflammatory cytokine production by human M2 macrophages. *Nature Communications* 2014. 5: 5444.
- 19 Hansen, I. S., Krabbendam, L., Bernink, J. H., Loayza-Puch, F., Hoepel, W., van Burgsteden, J. A., Kuijper, E. C., Buskens, C. J., Bemelman, W. A., Zaat, S. A. J., Agami, R., Vidarsson, G., van den Brink, G. R., de Jong, E. C., Wildenberg, M. E., Baeten, D. L. P., Everts, B. and den Dunnen, J., FcαRI co-stimulation converts human intestinal CD103+ dendritic cells into pro-inflammatory cells through glycolytic reprogramming. *Nature Communications* 2018. 9: 863.
- 20 Hoepel, W., Newling, M., Vogelpoel, L. T. C., Sritharan, L., Hansen, I. S., Kapsenberg, M. L., Baeten, D. L. P., Everts, B. and den Dunnen, J., FcγR-TLR Cross-Talk Enhances TNF Production by Human Monocyte-Derived DCs via IRF5-Dependent Gene Transcription and Glycolytic Reprogramming. *Frontiers in Immunology* 2019. 10.
- 21 Everts, B., Amiel, E., Huang, S. C.-C., Smith, A. M., Chang, C.-H., Lam, W. Y., Redmann, V., Freitas, T. C., Blagih, J., van der Windt, G. J. W., Artyomov, M. N., Jones, R. G., Pearce, E. L. and Pearce, E. J., TLR-driven early glycolytic reprogramming via the kinases TBK1-IKKε supports the anabolic demands of dendritic cell activation. *Nature Immunology* 2014. 15: 323-332.
- 22 Mullen, P. J., Garcia, G., Purkayastha, A., Matulionis, N., Schmid, E. W., Momcilovic, M., Sen, C., Langerman, J., Ramaiah, A., Shackelford, D. B., Damoiseaux, R., French, S. W., Plath, K., Gomperts, B. N., Arumugaswami, V. and Christofk, H. R., SARS-CoV-2 infection rewires host cell metabolism and is potentially susceptible to mTORC1 inhibition. *Nature Communications* 2021. 12: 1876.
- 23 Bojkova, D., Costa, R., Reus, P., Bechtel, M., Jaborek, M. C., Olmer, R., Martin, U., Ciesek,

- S., Michaelis, M. and Cinatl, J., Jr., Targeting the Pentose Phosphate Pathway for SARS-CoV-2 Therapy. *Metabolites* 2021. 11.
- 24 Steenblock, C., Schwarz, P. E. H., Ludwig, B., Linkermann, A., Zimmet, P., Kulebyakin, K., Tkachuk, V. A., Markov, A. G., Lehnert, H., de Angelis, M. H., Rietzsch, H., Rodionov, R. N., Khunti, K., Hopkins, D., Birkenfeld, A. L., Boehm, B., Holt, R. I. G., Skyler, J. S., DeVries, J. H., Renard, E., Eckel, R. H., Alberti, K., Geloneze, B., Chan, J. C., Mbanya, J. C., Onyegbutulem, H. C., Ramachandran, A., Basit, A., Hassanein, M., Bewick, G., Spinass, G. A., Beuschlein, F., Landgraf, R., Rubino, F., Mingrone, G. and Bornstein, S. R., COVID-19 and metabolic disease: mechanisms and clinical management. *Lancet Diabetes Endocrinol* 2021. 9: 786-798.
- 25 Brouwer, P. J. M., Caniels, T. G., van der Straten, K., Snitselaar, J. L., Aldon, Y., Bangaru, S., Torres, J. L., Okba, N. M. A., Claireaux, M., Kerster, G., Bentlage, A. E. H., van Haaren, M. M., Guerra, D., Burger, J. A., Schermer, E. E., Verheul, K. D., van der Velde, N., van der Kooij, A., van Schooten, J., van Breemen, M. J., Bijl, T. P. L., Sliepen, K., Aartse, A., Derking, R., Bontjer, I., Kootstra, N. A., Wiersinga, W. J., Vidarsson, G., Haagmans, B. L., Ward, A. B., de Bree, G. J., Sanders, R. W. and van Gils, M. J., Potent neutralizing antibodies from COVID-19 patients define multiple targets of vulnerability. *Science* 2020. 369: 643-650.
- 26 Heieis, G. A., Patente, T. A., Almeida, L., Vrieling, F., Tak, T., Perona-Wright, G., Maizels, R. M., Stienstra, R. and Everts, B., Metabolic heterogeneity of tissue-resident macrophages in homeostasis and during helminth infection. *Nature Communications* 2023. 14: 5627.
- 27 Caniels, T. G., Bontjer, I., van der Straten, K., Poniman, M., Burger, J. A., Appelman, B., Lavell, A. H. A., Oomen, M., Godeke, G. J., Valle, C., Mögling, R., van Willigen, H. D. G., Wynberg, E., Schinkel, M., van Vught, L. A., Guerra, D., Snitselaar, J. L., Chaturbhuj, D. N., Cuella Martin, I., Moore, J. P., de Jong, M. D., Reusken, C., Sikkens, J. J., Bomers, M. K., de Bree, G. J., van Gils, M. J., Eggink, D. and Sanders, R. W., Emerging SARS-CoV-2 variants of concern evade humoral immune responses from infection and vaccination. *Sci Adv* 2021. 7: eabj5365.
- 28 Brouwer Philip, J. M., Caniels Tom, G., van der Straten, K., Snitselaar Jonne, L., Aldon, Y., Bangaru, S., Torres Jonathan, L., Okba Nisreen, M. A., Claireaux, M., Kerster, G., Bentlage Arthur, E. H., van Haaren Marlies, M., Guerra, D., Burger Judith, A., Schermer Edith, E., Verheul Kirsten, D., van der Velde, N., van der Kooij, A., van Schooten, J., van Breemen Mariëlle, J., Bijl Tom, P. L., Sliepen, K., Aartse, A., Derking, R., Bontjer, I., Kootstra Neeltje, A., Wiersinga, W. J., Vidarsson, G., Haagmans Bart, L., Ward Andrew, B., de Bree Godelieve, J., Sanders Rogier, W. and van Gils Marit, J., Potent neutralizing antibodies from COVID-19 patients define multiple targets of vulnerability. *Science* 2020. 369: 643-650.
- 29 Geyer, C. E., Chen, H.-J., Bye, A. P., Manz, X. D., Guerra, D., Caniels, T. G., Bijl, T. P., Griffith, G. R., Hoepel, W., de Taeye, S. W., Veth, J., Vlaar, A. P., Vidarsson, G., Bogaard, H. J., Aman, J., Gibbins, J. M., van Gils, M. J., de Winther, M. P. and den Dunnen, J., Identification of new drugs to counteract anti-spike IgG-induced hyperinflammation in severe COVID-19. *Life Science Alliance* 2023. 6: e202302106.
- 30 Hu, C., Xuan, Y., Zhang, X., Liu, Y., Yang, S. and Yang, K., Immune cell metabolism and metabolic reprogramming. *Mol Biol Rep* 2022. 49: 9783-9795.
- 31 Russo, S., Kwiatkowski, M., Govorukhina, N., Bischoff, R. and Melgert, B. N., Meta-Inflammation and Metabolic Reprogramming of Macrophages in Diabetes and Obesity: The Importance of Metabolites. *Front Immunol* 2021. 12: 746151.
- 32 Verberk, S. G. S., de Goede, K. E., Gorki, F. S., van Dierendonck, X., Argüello, R. J. and Van den Bossche, J., An integrated toolbox to profile macrophage immunometabolism. *Cell Rep Methods* 2022. 2: 100192.
- 33 Corcoran, S. E. and O'Neill, L. A., HIF1 α and metabolic reprogramming in inflammation. *J Clin Invest* 2016. 126: 3699-3707.
- 34 Tang, Y.-Y., Wang, D.-C., Wang, Y.-Q., Huang, A.-F. and Xu, W.-D., Emerging role of hypoxia-inducible factor-1 α in inflammatory autoimmune diseases: A comprehensive review. *Frontiers in Immunology* 2023. Volume 13 - 2022.
- 35 Smith, K. G. and Clatworthy, M. R., Fc γ RIIB in autoimmunity and infection: evolutionary

Hyperinflammation by Human Macrophages Induced by SARS-CoV-2 Anti-Spike IgG Is Dependent on Glucose and Fatty Acid Metabolism

- and therapeutic implications. *Nat Rev Immunol* 2010. 10: 328-343.
- 36 Clutton, G., Mollan, K., Hudgens, M. and Goonetilleke, N., A Reproducible, Objective Method Using MitoTracker® Fluorescent Dyes to Assess Mitochondrial Mass in T Cells by Flow Cytometry. *Cytometry A* 2019. 95: 450-456.
- 37 Neikirk, K., Marshall, A. G., Kula, B., Smith, N., LeBlanc, S. and Hinton, A., Jr., MitoTracker: A useful tool in need of better alternatives. *Eur J Cell Biol* 2023. 102: 151371.
- 38 Vianello, C., Dal Bello, F., Shin, S. H., Schiavon, S., Bean, C., Magalhães Rebelo, A. P., Knedlík, T., Esfahani, E. N., Costiniti, V., Lacruz, R. S., Covello, G., Munari, F., Scolaro, T., Viola, A., Rampazzo, E., Persano, L., Zumerle, S., Scorrano, L., Gianelle, A. and Giacomello, M., High-Throughput Microscopy Analysis of Mitochondrial Membrane Potential in 2D and 3D Models. *Cells* 2023. 12.
- 39 Merad, M. and Martin, J. C., Pathological inflammation in patients with COVID-19: a key role for monocytes and macrophages. *Nature Reviews Immunology* 2020. 20: 355-362.
- 40 Mucha, S. R., Dugar, S., McCrae, K., Joseph, D. E., Bartholomew, J., Sacha, G. and Militello, M., Coagulopathy in COVID-19. *Cleveland Clinic Journal of Medicine* 2020.
- 41 Koo, S. J. and Garg, N. J., Metabolic programming of macrophage functions and pathogens control. *Redox Biol* 2019. 24: 101198.
- 42 Zhao, Q., Yu, Z., Zhang, S., Shen, X. R., Yang, H., Xu, Y., Liu, Y., Yang, L., Zhang, Q., Chen, J., Lu, M., Luo, F., Hu, M., Gong, Y., Xie, C., Zhou, P., Wang, L., Su, L., Zhang, Z. and Cheng, L., Metabolic modeling of single bronchoalveolar macrophages reveals regulators of hyperinflammation in COVID-19. *iScience* 2022. 25: 105319.
- 43 Codo, A. C., Davanzo, G. G., Monteiro, L. B., de Souza, G. F., Muraro, S. P., Virgilio-da-Silva, J. V., Prodonoff, J. S., Carregari, V. C., de Biagi Junior, C. A. O., Crunfli, F., Jimenez Restrepo, J. L., Vendramini, P. H., Reis-de-Oliveira, G., Bispo Dos Santos, K., Toledo-Teixeira, D. A., Parise, P. L., Martini, M. C., Marques, R. E., Carmo, H. R., Borin, A., Coimbra, L. D., Boldrini, V. O., Brunetti, N. S., Vieira, A. S., Mansour, E., Ulaf, R. G., Bernardes, A. F., Nunes, T. A., Ribeiro, L. C., Palma, A. C., Agrela, M. V., Moretti, M. L., Sposito, A. C., Pereira, F. B., Velloso, L. A., Vinolo, M. A. R., Damasio, A., Proença-Módena, J. L., Carvalho, R. F., Mori, M. A., Martins-de-Souza, D., Nakaya, H. I., Farias, A. S. and Moraes-Vieira, P. M., Elevated Glucose Levels Favor SARS-CoV-2 Infection and Monocyte Response through a HIF-1 α /Glycolysis-Dependent Axis. *Cell Metab* 2020. 32: 437-446.e435.
- 44 Zhang, Z. L., Hou, Y. L., Li, D. T. and Li, F. Z., Laboratory findings of COVID-19: a systematic review and meta-analysis. *Scand J Clin Lab Invest* 2020. 80: 441-447.
- 45 Chen, P., Wu, M., He, Y., Jiang, B. and He, M.-L., Metabolic alterations upon SARS-CoV-2 infection and potential therapeutic targets against coronavirus infection. *Signal Transduction and Targeted Therapy* 2023. 8: 237.
- 46 Ardestani, A. and Azizi, Z., Targeting glucose metabolism for treatment of COVID-19. *Signal Transduction and Targeted Therapy* 2021. 6: 112.
- 47 Sahu, K. K. and Kumar, R., Role of 2-Deoxy-D-Glucose (2-DG) in COVID-19 disease: A potential game-changer. *Journal of Family Medicine and Primary Care* 2021. 10: 3548-3552.
- 48 Bhatt, A. N., Shenoy, S., Munjal, S., Chinnadurai, V., Agarwal, A., Vinoth Kumar, A., Shanavas, A., Kanwar, R. and Chandna, S., 2-deoxy-d-glucose as an adjunct to standard of care in the medical management of COVID-19: a proof-of-concept and dose-ranging randomised phase II clinical trial. *BMC Infectious Diseases* 2022. 22: 669.
- 49 Pajak, B., Siwiak, E., Sołtyka, M., Priebe, A., Zieliński, R., Fokt, I., Ziemniak, M., Jaśkiewicz, A., Borowski, R., Domoradzki, T. and Priebe, W., 2-Deoxy-d-Glucose and Its Analogs: From Diagnostic to Therapeutic Agents. *Int J Mol Sci* 2019. 21.
- 50 Raez, L. E., Papadopoulos, K., Ricart, A. D., Chiorean, E. G., DiPaola, R. S., Stein, M. N., Rocha Lima, C. M., Schlesselman, J. J., Tolba, K., Langmuir, V. K., Kroll, S., Jung, D. T., Kurtoglu, M., Rosenblatt, J. and Lampidis, T. J., A phase I dose-escalation trial of 2-deoxy-d-glucose alone or combined with docetaxel in patients with advanced solid tumors. *Cancer Chemotherapy and Pharmacology* 2013. 71: 523-530.

- 51 Jirjees, F., Saad, A. K., Al Hano, Z., Hatahet, T., Al Obaidi, H. and Dallal Bashi, Y. H., COVID-19 Treatment Guidelines: Do They Really Reflect Best Medical Practices to Manage the Pandemic? *Infect Dis Rep* 2021. 13: 259-284.
- 52 Mele, L., Paino, F., Papaccio, F., Regad, T., Boocock, D., Stiuso, P., Lombardi, A., Liccardo, D., Aquino, G., Barbieri, A., Arra, C., Coveney, C., La Noce, M., Papaccio, G., Caraglia, M., Tirino, V. and Desiderio, V., A new inhibitor of glucose-6-phosphate dehydrogenase blocks pentose phosphate pathway and suppresses malignant proliferation and metastasis in vivo. *Cell Death & Disease* 2018. 9: 572.
- 53 Cremon, C., Stanghellini, V., Barbaro, M. R., Cogliandro, R. F., Bellacosa, L., Santos, J., Vicario, M., Pigrau, M., Alonso Cotoner, C., Lobo, B., Azpiroz, F., Bruley des Varannes, S., Neunlist, M., DeFilippis, D., Iuvone, T., Petrosino, S., Di Marzo, V. and Barbara, G., Randomised clinical trial: the analgesic properties of dietary supplementation with palmitoylethanolamide and polydatin in irritable bowel syndrome. *Aliment Pharmacol Ther* 2017. 45: 909-922.
- 54 Indraccolo, U., Indraccolo, S. R. and Mignini, F., Micronized palmitoylethanolamide/trans-polydatin treatment of endometriosis-related pain: a meta-analysis. *Ann Ist Super Sanita* 2017. 53: 125-134.
- 55 Infantino, V., Iacobazzi, V., Palmieri, F. and Menga, A., ATP-citrate lyase is essential for macrophage inflammatory response. *Biochem Biophys Res Commun* 2013. 440: 105-111.
- 56 Infantino, V., Convertini, P., Cucci, L., Panaro, M. A., Di Noia, M. A., Calvello, R., Palmieri, F. and Iacobazzi, V., The mitochondrial citrate carrier: a new player in inflammation. *Biochem J* 2011. 438: 433-436.
- 57 Wei, X., Song, H., Yin, L., Rizzo, M. G., Sidhu, R., Covey, D. F., Ory, D. S. and Semenovich, C. F., Fatty acid synthesis configures the plasma membrane for inflammation in diabetes. *Nature* 2016. 539: 294-298.
- 58 Miki, H., Han, K. H., Scott, D., Croft, M. and Kang, Y. J., 4-1BBL Regulates the Polarization of Macrophages, and Inhibition of 4-1BBL Signaling Alleviates Imiquimod-Induced Psoriasis. *J Immunol* 2020. 204: 1892-1903.
- 59 Johnson, A. R., Qin, Y., Cozzo, A. J., Freemerman, A. J., Huang, M. J., Zhao, L., Sampey, B. P., Milner, J. J., Beck, M. A., Damania, B., Rashid, N., Galanko, J. A., Lee, D. P., Edin, M. L., Zeldin, D. C., Fueger, P. T., Dietz, B., Stahl, A., Wu, Y., Mohlke, K. L. and Makowski, L., Metabolic reprogramming through fatty acid transport protein 1 (FATP1) regulates macrophage inflammatory potential and adipose inflammation. *Mol Metab* 2016. 5: 506-526.
- 60 Klein-Wieringa, I. R., Andersen, S. N., Kwekkeboom, J. C., Giera, M., de Lange-Brokaar, B. J., van Osch, G. J., Zuurmond, A. M., Stojanovic-Susulic, V., Nelissen, R. G., Pijl, H., Huizinga, T. W., Kloppenburg, M., Toes, R. E. and Ioan-Facsinay, A., Adipocytes modulate the phenotype of human macrophages through secreted lipids. *J Immunol* 2013. 191: 1356-1363.
- 61 Almeida, L. and Everts, B., Fa(c)t checking: How fatty acids shape metabolism and function of macrophages and dendritic cells. *Eur J Immunol* 2021. 51: 1628-1640.
- 62 Moon, J. S., Lee, S., Park, M. A., Siempos, I., Haslip, M., Lee, P. J., Yun, M., Kim, C. K., Howrylak, J., Ryter, S. W., Nakahira, K. and Choi, A. M., UCP2-induced fatty acid synthase promotes NLRP3 inflammasome activation during sepsis. *J Clin Invest* 2015. 125: 665-680.
- 63 Ecker, J., Liebisch, G., Englmaier, M., Grandl, M., Robenek, H. and Schmitz, G., Induction of fatty acid synthesis is a key requirement for phagocytic differentiation of human monocytes. *Proceedings of the National Academy of Sciences* 2010. 107: 7817-7822.
- 64 Yeudall, S., Upchurch, C. M., Seegren, P. V., Pavelec, C. M., Greulich, J., Lemke, M. C., Harris, T. E., Desai, B. N., Hoehn, K. L. and Leitinger, N., Macrophage acetyl-CoA carboxylase regulates acute inflammation through control of glucose and lipid metabolism. *Sci Adv* 2022. 8: eabq1984.
- 65 Leishman, S., Aljadeed, N. M., Qian, L., Cockcroft, S., Behmoaras, J. and Anand, P. K., Fatty acid synthesis promotes inflammasome activation through NLRP3 palmitoylation. *Cell Rep* 2024. 43: 114516.
- 66 Seton-Rogers, S., Building up and breaking down fatty acids. *Nature Reviews Cancer* 2016. 16: 677-677.

Hyperinflammation by Human Macrophages Induced by SARS-CoV-2 Anti-Spike IgG Is Dependent on Glucose and Fatty Acid Metabolism

- 67 Shang, L., Wang, L., Zhou, F., Li, J., Liu, Y. and Yang, S., Long-term effects of obesity on COVID-19 patients discharged from hospital. *Immun Inflamm Dis* 2021. 9: 1678-1685.
- 68 Garrido-Sánchez, L., Vendrell, J., Fernández-García, D., Ceperuelo-Mallafré, V., Chacón, M. R., Ocaña-Wilhelmi, L., Alcaide, J., Tinahones, F. J. and García-Fuentes, E., De Novo Lipogenesis in Adipose Tissue Is Associated with Course of Morbid Obesity after Bariatric Surgery. *PLOS ONE* 2012. 7: e31280.
- 69 Nikkhoo, B., Mohammadi, M., Hasani, S., Sigari, N., Borhani, A., Ramezani, C., Charajoo, A., Badri, S., Rostami, F., Etemadi, M. and Rahmani, K., Elevated interleukin (IL)-6 as a predictor of disease severity among Covid-19 patients: a prospective cohort study. *BMC Infect Dis* 2023. 23: 311.
- 70 McGonagle, D., Sharif, K., O'Regan, A. and Bridgewood, C., The Role of Cytokines including Interleukin-6 in COVID-19 induced Pneumonia and Macrophage Activation Syndrome-Like Disease. *Autoimmun Rev* 2020. 19: 102537.
- 71 Jing, C., Castro-Dopico, T., Richoz, N., Tuong, Z. K., Ferdinand, J. R., Lok, L. S. C., Loudon, K. W., Banham, G. D., Mathews, R. J., Cader, Z., Fitzpatrick, S., Bashant, K. R., Kaplan, M. J., Kaser, A., Johnson, R. S., Murphy, M. P., Siegel, R. M. and Clatworthy, M. R., Macrophage metabolic reprogramming presents a therapeutic target in lupus nephritis. *Proc Natl Acad Sci U S A* 2020. 117: 15160-15171.
- 72 Shi, J., Knevel, R., Suwannalai, P., van der Linden, M. P., Janssen, G. M., van Veelen, P. A., Levarht, N. E., van der Helm-van Mil, A. H., Cerami, A., Huizinga, T. W., Toes, R. E. and Trouw, L. A., Autoantibodies recognizing carbamylated proteins are present in sera of patients with rheumatoid arthritis and predict joint damage. *Proc Natl Acad Sci U S A* 2011. 108: 17372-17377.
- 73 Fayyaz, A., Kurien, B. T. and Scofield, R. H., Autoantibodies in Sjögren's Syndrome. *Rheum Dis Clin North Am* 2016. 42: 419-434.
- 74 Damelang, T., Brinkhaus, M., van Osch, T. L. J., Schuurman, J., Labrijn, A. F., Rispens, T. and Vidarsson, G., Impact of structural modifications of IgG antibodies on effector functions. *Front Immunol* 2023. 14: 1304365.
- 75 Wada, R., Matsui, M. and Kawasaki, N., Influence of N-glycosylation on effector functions and thermal stability of glycoengineered IgG1 monoclonal antibody with homogeneous glycoforms. *MAbs* 2019. 11: 350-372.
- 76 Šuštić, T., Van Coillie, J., Larsen, M. D., Derksen, N. I. L., Szittner, Z., Nouta, J., Wang, W., Damelang, T., Rebergen, I., Linty, F., Visser, R., Mok, J. Y., Geerdes, D. M., van Esch, W. J. E., de Teye, S. W., van Gils, M. J., van de Watering, L., van der Schoot, C. E., Wuhrer, M., Rispens, T. and Vidarsson, G., Immunoassay for quantification of antigen-specific IgG fucosylation. *EBioMedicine* 2022. 81: 104109.
- 77 Kiener, P. A., Rankin, B. M., Burkhardt, A. L., Schieven, G. L., Gilliland, L. K., Rowley, R. B., Bolen, J. B. and Ledbetter, J. A., Cross-linking of Fc gamma receptor I (Fc gamma RI) and receptor II (Fc gamma RII) on monocytic cells activates a signal transduction pathway common to both Fc receptors that involves the stimulation of p72 Syk protein tyrosine kinase. *J Biol Chem* 1993. 268: 24442-24448.
- 78 Daëron, M., Fc receptor biology. *Annu Rev Immunol* 1997. 15: 203-234.
- 79 Guarnieri, J. W., Dybas, J. M., Fazelinia, H., Kim, M. S., Frere, J., Zhang, Y., Soto Albrecht, Y., Murdock, D. G., Angelin, A., Singh, L. N., Weiss, S. L., Best, S. M., Lott, M. T., Zhang, S., Cope, H., Zaksas, V., Saravia-Butler, A., Meydan, C., Fook, J., Mozsary, C., Bram, Y., Kidane, Y., Priebe, W., Emmett, M. R., Meller, R., Demharter, S., Stentoft-Hansen, V., Salvatore, M., Galeano, D., Enguita, F. J., Grabham, P., Trovao, N. S., Singh, U., Haltom, J., Heise, M. T., Moorman, N. J., Baxter, V. K., Madden, E. A., Taft-Benz, S. A., Anderson, E. J., Sanders, W. A., Dickmander, R. J., Baylin, S. B., Wurtele, E. S., Moraes-Vieira, P. M., Taylor, D., Mason, C. E., Schisler, J. C., Schwartz, R. E., Beheshti, A. and Wallace, D. C., Core mitochondrial genes are down-regulated during SARS-CoV-2 infection of rodent and human hosts. *Sci Transl Med* 2023. 15: eabq1533.
- 80 Shoraka, S., Samarasinghe, A. E., Ghaemi, A. and Mohebbi, S. R., Host mitochondria: more than an organelle in SARS-CoV-2 infection. *Frontiers in Cellular and Infection Microbiology* 2023. 13.

- 81 Scozzi, D., Cano, M., Ma, L., Zhou, D., Zhu, J. H., O'Halloran, J. A., Goss, C., Rauseo, A. M., Liu, Z., Sahu, S. K., Peritore, V., Rocco, M., Ricci, A., Amodeo, R., Aimati, L., Ibrahim, M., Hachem, R., Kreisel, D., Mudd, P. A., Kulkarni, H. S. and Gelman, A. E., Circulating mitochondrial DNA is an early indicator of severe illness and mortality from COVID-19. *JCI Insight* 2021. 6.
- 82 Cortese, M., Lee, J. Y., Cerikan, B., Neufeldt, C. J., Oorschot, V. M. J., Köhrer, S., Hennies, J., Schieber, N. L., Ronchi, P., Mizzon, G., Romero-Brey, I., Santarella-Mellwig, R., Schorb, M., Boermel, M., Mocaer, K., Beckwith, M. S., Templin, R. M., Gross, V., Pape, C., Tischer, C., Frankish, J., Horvat, N. K., Laketa, V., Stanifer, M., Boulant, S., Ruggieri, A., Chatel-Chaix, L., Schwab, Y. and Bartenschlager, R., Integrative Imaging Reveals SARS-CoV-2-Induced Reshaping of Subcellular Morphologies. *Cell Host Microbe* 2020. 28: 853-866.e855.
- 83 Moreno Fernández-Ayala, D. J., Navas, P. and López-Lluch, G., Age-related mitochondrial dysfunction as a key factor in COVID-19 disease. *Exp Gerontol* 2020. 142: 111147.
- 84 Sadeghian, M., Mastrolia, V., Rezaei Haddad, A., Mosley, A., Mullali, G., Schiza, D., Sajic, M., Hargreaves, I., Heales, S., Duchon, M. R. and Smith, K. J., Mitochondrial dysfunction is an important cause of neurological deficits in an inflammatory model of multiple sclerosis. *Sci Rep* 2016. 6: 33249.
- 85 Wang, C., Zhao, M., Bin, P., Ye, Y., Chen, Q., Tang, Z. and Ren, W., Serine synthesis controls mitochondrial biogenesis in macrophages. *Sci Adv* 2024. 10: eadn2867.
- 86 Chen, T. H., Chang, C. J. and Hung, P. H., Possible Pathogenesis and Prevention of Long COVID: SARS-CoV-2-Induced Mitochondrial Disorder. *Int J Mol Sci* 2023. 24.
- 87 Noonong, K., Chatatikun, M., Surinkaew, S., Kotepui, M., Hossain, R., Bunluepuech, K., Noothong, C., Tedasen, A., Klangbud, W. K., Imai, M., Kawakami, F., Kubo, M., Kitagawa, Y., Ichikawa, H., Kanekura, T., Sukati, S., Somsak, V., Udomwech, L., Ichikawa, T., Nissapatorn, V., Tangpong, J., Indo, H. P. and Majima, H. J., Mitochondrial oxidative stress, mitochondrial ROS storms in long COVID pathogenesis. *Frontiers in Immunology* 2023. 14.
- 88 Chang, Y.-Y. and Wei, A.-C., Transcriptome and machine learning analysis of the impact of COVID-19 on mitochondria and multiorgan damage. *PLOS ONE* 2024. 19: e0297664.
- 89 Léopold, V., Chouchane, O., Butler, J. M., Schuurman, A. R., Michels, E. H. A., de Brabander, J., Schomakers, B. V., van Weeghel, M., Picavet-Havik, D. I., Grootemaat, A. E., Douma, R. A., Reijnders, T. D. Y., Klarenbeek, A. M., Appelman, B., Wiersinga, W. J., van der Wel, N. N., den Dunnen, J., Houtkooper, R. H., Van't Veer, C. and van der Poll, T., Platelets of COVID-19 patients display mitochondrial dysfunction, oxidative stress, and energy metabolism failure compatible with cell death. *Res Pract Thromb Haemost* 2023. 7: 102213.
- 90 Boylan, B., Gao, C., Rathore, V., Gill, J. C., Newman, D. K. and Newman, P. J., Identification of FcγRIIIa as the ITAM-bearing receptor mediating αIIbβ3 outside-in integrin signaling in human platelets. *Blood* 2008. 112: 2780-2786.
- 91 Chen, H. J., Li Yim, A. Y. F., Griffith, G. R., de Jonge, W. J., Mannens, M., Ferrero, E., Henneman, P. and de Winther, M. P. J., Meta-Analysis of in vitro-Differentiated Macrophages Identifies Transcriptomic Signatures That Classify Disease Macrophages in vivo. *Front Immunol* 2019. 10: 2887.