



Universiteit
Leiden
The Netherlands

Advanced MR image analysis in sporadic and Dutch-type hereditary Cerebral Amyloid Angiopathy

Schipper, M.R.

Citation

Schipper, M. R. (2026, June 10). *Advanced MR image analysis in sporadic and Dutch-type hereditary Cerebral Amyloid Angiopathy*. Retrieved from <https://hdl.handle.net/1887/4305152>

Version: Publisher's Version

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/4305152>

Note: To cite this publication please use the final published version (if applicable).

References

1. Charidimou A and al. e. Brain hemorrhage recurrence, small vessel disease type, and cerebral microbleeds: A meta-analysis. *Neurology* 2017; 89: 820-829. DOI: 10.1212/WNL.0000000000004259.
2. Wermer MJH and Greenberg SM. The growing clinical spectrum of cerebral amyloid angiopathy. *Curr Opin Neurol* 2018; 31: 28-35. 2017/11/10. DOI: 10.1097/WCO.0000000000000510.
3. Jäkel L, De Kort AM, Klijn CJM, et al. Prevalence of cerebral amyloid angiopathy: A systematic review and meta-analysis. *Alzheimer's & Dementia* 2022; 18: 10-28. DOI: 10.1002/alz.12366.
4. Charidimou A, Boulouis G, Frosch MP, et al. The Boston criteria version 2.0 for cerebral amyloid angiopathy: a multicentre, retrospective, MRI-neuropathology diagnostic accuracy study. *The Lancet Neurology* 2022; 21: 714-725. DOI: 10.1016/s1474-4422(22)00208-3.
5. Maat-Schieman M, Roos R and Van Duinen S. Hereditary cerebral hemorrhage with amyloidosis-Dutch type. *Neuropathology* 2005; 25: 288-297. DOI: 10.1111/j.1440-1789.2005.00631.x.
6. Bornebroek M, Haan J, Maat-Schieman ML, et al. Hereditary cerebral hemorrhage with amyloidosis-Dutch type (HCHWA-D): I--A review of clinical, radiologic and genetic aspects. *Brain pathology (Zurich, Switzerland)*. 1996, p. 111-114.
7. Van Dort R, Kaushik K, Rasing I, et al. Cognition in (pre)symptomatic Dutch-type hereditary and sporadic cerebral amyloid angiopathy. *Alzheimer's & Dementia* 2024; 20: 7518-7528. DOI: 10.1002/alz.14171.
8. Koemans EA, Chhatwal JP, Van Veluw SJ, et al. Progression of cerebral amyloid angiopathy: a pathophysiological framework. *The Lancet Neurology* 2023; 22: 632-642. DOI: 10.1016/s1474-4422(23)00114-x.
9. Strozyk D, Blennow K, White LR, et al. CSF Aβ₄₂ levels correlate with amyloid-neuropathology in a population-based autopsy study. *Neurology* 2003; 60: 652-656. 2003/02/26.
10. Charidimou A, Friedrich JO, Greenberg SM, et al. Core cerebrospinal fluid biomarker profile in cerebral amyloid angiopathy: A meta-analysis. *Neurology* 2018; 90: e754-e762. 2018/02/02. DOI: 10.1212/WNL.0000000000005030.
11. Schultz AP, Kloet RW, Sohrabi HR, et al. Amyloid imaging of dutch-type hereditary cerebral amyloid angiopathy carriers. *Annals of Neurology* 2019; 86: 616-625. DOI: 10.1002/ana.25560.
12. Chatterjee P, Tegg M, Pedrini S, et al. Plasma Amyloid-Beta Levels in a Pre-Symptomatic Dutch-Type Hereditary Cerebral Amyloid Angiopathy Pedigree: A Cross-Sectional and Longitudinal Investigation. *International Journal of Molecular Sciences* 2021; 22: 2931. DOI: 10.3390/ijms22062931.
13. van Etten ES, Verbeek MM, van der Grond J, et al. β-Amyloid in CSF: Biomarker for preclinical cerebral amyloid angiopathy. *Neurology* 2017; 88: 169-176. DOI: 10.1212/WNL.0000000000003486.
14. Keable A, Fenna K, Yuen HM, et al. Deposition of amyloid beta in the walls of human leptomenigeal arteries in relation to perivascular drainage pathways in cerebral amyloid angiopathy. *Biochim Biophys Acta* 2016; 1862: 1037-1046. 20150829. DOI: 10.1016/j.bbadis.2015.08.024.
15. Vonsattel JP, Myers RH, Hedley-Whyte ET, et al. Cerebral amyloid angiopathy without and with cerebral hemorrhages: a comparative histological study. *Ann Neurol* 1991; 30: 637-649. 1991/11/11. DOI: 10.1002/ana.410300503.

16. Vos SJ, Visser PJ, Verhey F, et al. Variability of CSF Alzheimer's disease biomarkers: implications for clinical practice. *PLoS One* 2014; 9: e100784. 20140624. DOI: 10.1371/journal.pone.0100784.
17. Dumas A, Dierksen GA, Gurol ME, et al. Functional magnetic resonance imaging detection of vascular reactivity in cerebral amyloid angiopathy. *Annals of Neurology* 2012; 72: 76-81. DOI: 10.1002/ana.23566.
18. Smith EE, Vijayappa M, Lima F, et al. Impaired visual evoked flow velocity response in cerebral amyloid angiopathy. *Neurology* 2008; 71: 1424-1430. DOI: 10.1212/01.wnl.0000327887.64299.a4.
19. Switzer AR, McCreary C, Batool S, et al. Longitudinal decrease in blood oxygenation level dependent response in cerebral amyloid angiopathy. *Neuroimage Clin* 2016; 11: 461-467. 20160303. DOI: 10.1016/j.nicl.2016.02.020.
20. van Opstal AM, van Rooden S, van Harten T, et al. Cerebrovascular function in presymptomatic and symptomatic individuals with hereditary cerebral amyloid angiopathy: a case-control study. *The Lancet Neurology* 2017; 16: 115-122. DOI: 10.1016/s1474-4422(16)30346-5.
21. Reijmer YD, Fotiadis P, Charidimou A, et al. Relationship between white matter connectivity loss and cortical thinning in cerebral amyloid angiopathy. *Hum Brain Mapp* 2017 2017/05/04. DOI: 10.1002/hbm.23629.
22. Reijmer YD, Fotiadis P, Martinez-Ramirez S, et al. Structural network alterations and neurological dysfunction in cerebral amyloid angiopathy. *Brain* 2015; 138: 179-188. DOI: 10.1093/brain/awu316.
23. Salat DH, Smith EE, Tuch DS, et al. White Matter Alterations in Cerebral Amyloid Angiopathy Measured by Diffusion Tensor Imaging. *Stroke* 2006; 37: 1759-1764. DOI: 10.1161/01.str.0000227328.86353.a7.
24. Raposo N, Zanon Zotin MC, Schoemaker D, et al. Peak Width of Skeletonized Mean Diffusivity as Neuroimaging Biomarker in Cerebral Amyloid Angiopathy. *American Journal of Neuroradiology* 2021; 42: 875-881. DOI: 10.3174/ajnr.a7042.
25. Martinez-Ramirez S, van Rooden S, Charidimou A, et al. Perivascular Spaces Volume in Sporadic and Hereditary (Dutch-Type) Cerebral Amyloid Angiopathy. *Stroke* 2018 2018/07/18. DOI: 10.1161/STROKEAHA.118.021137.
26. van Dijk SE, van der Grond J, Lak J, et al. Longitudinal Progression of Magnetic Resonance Imaging Markers and Cognition in Dutch-Type Hereditary Cerebral Amyloid Angiopathy. *Stroke* 2022; 53: 2006-2015. 20220401. DOI: 10.1161/STROKEAHA.121.035826.
27. Kozberg MG, Yi I, Freeze WM, et al. Blood-brain barrier leakage and perivascular inflammation in cerebral amyloid angiopathy. *Brain Communications* 2022; 4. DOI: 10.1093/braincomms/fcac245.
28. van Etten ES, Gurol ME, van der Grond J, et al. Recurrent hemorrhage risk and mortality in hereditary and sporadic cerebral amyloid angiopathy. *Neurology* 2016; 87: 1482-1487. 2016/09/04. DOI: 10.1212/WNL.0000000000003181.
29. Samarasekera N, Smith C and Al-Shahi Salman R. The association between cerebral amyloid angiopathy and intracerebral haemorrhage: systematic review and meta-analysis. *Journal of Neurology, Neurosurgery & Psychiatry* 2012; 83: 275-281. DOI: 10.1136/jnnp-2011-300371.
30. Wardlaw JM, Smith EE, Biessels GJ, et al. Neuroimaging standards for research into small

Appendix

vessel disease and its contribution to ageing and neurodegeneration. *The Lancet Neurology* 2013; 12: 822-838. DOI: 10.1016/S1474-4422(13)70124-8.

31. Dering M, Biessels GJ, Brodtmann A, et al. Neuroimaging standards for research into small vessel disease—advances since 2013. *The Lancet Neurology* 2023; 22: 602-618. DOI: 10.1016/s1474-4422(23)00131-x.

32. Koenecke H-C. Cerebral microbleeds on MRI. *Neurology* 2006; 66: 165-171. DOI: 10.1212/01.wnl.0000194266.55694.1e.

33. Viswanathan A and Chabriat H. Cerebral Microhemorrhage. *Stroke* 2006; 37: 550-555. DOI: 10.1161/01.str.0000199847.96188.12.

34. Cordonnier C, Al-Shahi Salman R and Wardlaw J. Spontaneous brain microbleeds: systematic review, subgroup analyses and standards for study design and reporting. *Brain* 2007; 130: 1988-2003. DOI: 10.1093/brain/awl387.

35. Agarwal A, Ajmera P, Sharma P, et al. Cerebral microbleeds: Causes, clinical relevance, and imaging approach - A narrative review. *J Neurosci Rural Pract* 2024; 15: 169-181. 20240308. DOI: 10.25259/JNRP_351_2023.

36. Rosand J, Muzikansky A, Kumar A, et al. Spatial clustering of hemorrhages in probable cerebral amyloid angiopathy. *Annals of Neurology* 2005; 58: 459-462. DOI: 10.1002/ana.20596.

37. Ni J, Auriel E, Martinez-Ramirez S, et al. Cortical localization of microbleeds in cerebral amyloid angiopathy: an ultra high-field 7T MRI study. *J Alzheimers Dis* 2015; 43: 1325-1330. DOI: 10.3233/JAD-140864.

38. Perosa V, Auger CA, Zanon Zotin MC, et al. Histopathological Correlates of Lobar Microbleeds in False-Positive Cerebral Amyloid Angiopathy Cases. *Annals of Neurology* 2023; 94: 856-870. DOI: 10.1002/ana.26761.

39. Ding J, Sigurðsson S, Jónsson PV, et al. Space and location of cerebral microbleeds, cognitive decline, and dementia in the community. *Neurology* 2017; 88: 2089-2097. DOI: 10.1212/wnl.0000000000003983.

40. Charidimou A, Shams S, Romero JR, et al. Clinical significance of cerebral microbleeds on MRI: A comprehensive meta-analysis of risk of intracerebral hemorrhage, ischemic stroke, mortality, and dementia in cohort studies (v1). *International Journal of Stroke* 2018; 13: 454-468. DOI: 10.1177/1747493017751931.

41. Charidimou A, Perosa V, Frosch MP, et al. Neuropathological correlates of cortical superficial siderosis in cerebral amyloid angiopathy. *Brain* 2020; 143: 3343-3351. DOI: 10.1093/brain/awaa266.

42. Charidimou A, Boulouis G, Greenberg SM, et al. Cortical superficial siderosis and bleeding risk in cerebral amyloid angiopathy: A meta-analysis. *Neurology* 2019; 93: e2192-e2202. 20191115. DOI: 10.1212/WNL.0000000000008590.

43. Smith EE, Charidimou A, Ayata C, et al. Cerebral Amyloid Angiopathy-Related Transient Focal Neurologic Episodes. *Neurology* 2021; 97: 231-238. 20210520. DOI: 10.1212/WNL.0000000000012234.

44. Iliff JJ, Wang M, Liao Y, et al. A Paravascular Pathway Facilitates CSF Flow Through the Brain Parenchyma and the Clearance of Interstitial Solutes, Including Amyloid β . *Science Translational Medicine* 2012; 4: 147ra111-147ra141. DOI: 10.1126/scitranslmed.3003748.

45. Kress BT, Iliff JJ, Xia M, et al. Impairment of paravascular clearance pathways in the aging brain. *Annals of Neurology* 2014; 76: 845-861. DOI: 10.1002/ana.24271.
46. Arbel-Ornath M, Hudry E, Eikermann-Haerter K, et al. Interstitial fluid drainage is impaired in ischemic stroke and Alzheimer's disease mouse models. *Acta Neuropathologica* 2013; 126: 353-364. DOI: 10.1007/s00401-013-1145-2.
47. Perosa V, Oltmer J, Munting LP, et al. Perivascular space dilation is associated with vascular amyloid- β accumulation in the overlying cortex. *Acta Neuropathologica* 2022; 143: 331-348. DOI: 10.1007/s00401-021-02393-1.
48. Raposo N, Planton M, Payoux P, et al. Enlarged perivascular spaces and florbetapir uptake in patients with intracerebral hemorrhage. *European Journal of Nuclear Medicine and Molecular Imaging* 2019; 46: 2339-2347. DOI: 10.1007/s00259-019-04441-1.
49. Boche D, Zotova E, Weller RO, et al. Consequence of A β immunization on the vasculature of human Alzheimer's disease brain. *Brain* 2008; 131: 3299-3310. DOI: 10.1093/brain/awn261.
50. Nicoll JAR, Buckland GR, Harrison CH, et al. Persistent neuropathological effects 14 years following amyloid- β immunization in Alzheimer's disease. *Brain* 2019; 142: 2113-2126. DOI: 10.1093/brain/awz142.
51. Peng W, Achariyar TM, Li B, et al. Suppression of glymphatic fluid transport in a mouse model of Alzheimer's disease. *Neurobiology of Disease* 2016; 93: 215-225. DOI: 10.1016/j.nbd.2016.05.015.
52. Kim SH, Ahn JH, Yang H, et al. Cerebral amyloid angiopathy aggravates perivascular clearance impairment in an Alzheimer's disease mouse model. *Acta Neuropathologica Communications* 2020; 8. DOI: 10.1186/s40478-020-01042-0.
53. Van Veluw SJ, Hou SS, Calvo-Rodriguez M, et al. Vasomotion as a Driving Force for Paravascular Clearance in the Awake Mouse Brain. *Neuron* 2020; 105: 549-561.e545. DOI: 10.1016/j.neuron.2019.10.033.
54. Hawkes CA, Härtig W, Kacza J, et al. Perivascular drainage of solutes is impaired in the ageing mouse brain and in the presence of cerebral amyloid angiopathy. *Acta Neuropathologica* 2011; 121: 431-443. DOI: 10.1007/s00401-011-0801-7.
55. Simon M, Wang MX, Ismail O, et al. Loss of perivascular aquaporin-4 localization impairs glymphatic exchange and promotes amyloid β plaque formation in mice. *Alzheimer's Research & Therapy* 2022; 14. DOI: 10.1186/s13195-022-00999-5.
56. Potter GM, Chappell FM, Morris Z, et al. Cerebral perivascular spaces visible on magnetic resonance imaging: development of a qualitative rating scale and its observer reliability. *Cerebrovasc Dis* 2015; 39: 224-231. 20150319. DOI: 10.1159/000375153.
57. Lan H, Lynch KM, Custer R, et al. Weakly supervised perivascular spaces segmentation with salient guidance of Frangi filter. *Magn Reson Med* 2023; 89: 2419-2431. 20230124. DOI: 10.1002/mrm.29593.
58. Huang P, Liu L, Zhang Y, et al. Development and validation of a perivascular space segmentation method in multi-center datasets. *Neuroimage* 2024; 298: 120803. 20240823. DOI: 10.1016/j.neuroimage.2024.120803.
59. Choi Y, Nam Y, Choi Y, et al. MRI-visible dilated perivascular spaces in healthy young adults: A

Appendix

twin heritability study. *Hum Brain Mapp* 2020; 41: 5313-5324. 20200908. DOI: 10.1002/hbm.25194.

60. Lian C, Zhang J, Liu M, et al. Multi-channel multi-scale fully convolutional network for 3D perivascular spaces segmentation in 7T MR images. *Medical Image Analysis* 2018; 46: 106-117. DOI: 10.1016/j.media.2018.02.009.
61. Rashid T, Liu H, Ware JB, et al. Deep Learning Based Detection of Enlarged Perivascular Spaces on Brain MRI. *Neuroimage Rep* 2023; 3 20230307. DOI: 10.1016/j.ynrp.2023.100162.
62. Sinclair B, Pham W, Vivash L, et al. Perivascular space identification nnUNet for generalised usage (PINGU). *Med Image Anal* 2025; 109: 103903. 20251204. DOI: 10.1016/j.media.2025.103903.
63. Gokcal E, Horn MJ, van Veluw SJ, et al. Lacunes, Microinfarcts, and Vascular Dysfunction in Cerebral Amyloid Angiopathy. *Neurology* 2021; 96: e1646-e1654. 20210203. DOI: 10.1212/WNL.00000000000011631.
64. McCreary CR, Beaudin AE, Subotic A, et al. Cross-sectional and longitudinal differences in peak skeletonized white matter mean diffusivity in cerebral amyloid angiopathy. *Neuroimage Clin* 2020; 27: 102280. 20200526. DOI: 10.1016/j.nicl.2020.102280.
65. Baykara E, Gesierich B, Adam R, et al. A Novel Imaging Marker for Small Vessel Disease Based on Skeletonization of White Matter Tracts and Diffusion Histograms. *Annals of Neurology* 2016; 80: 581-592. DOI: 10.1002/ana.24758.
66. Deary IJ, Ritchie SJ, Muñoz Maniega S, et al. Brain Peak Width of Skeletonized Mean Diffusivity (PSMD) and Cognitive Function in Later Life. *Frontiers in Psychiatry* 2019; 10. DOI: 10.3389/fpsyt.2019.00524.
67. Vlegels N, van den Brink H, Kopczak A, et al. The relation between cerebral small vessel function and white matter microstructure in monogenic and sporadic small vessel disease - the ZOOM@SVDs study. *Cereb Circ Cogn Behav* 2025; 8: 100383. 20250324. DOI: 10.1016/j.cccb.2025.100383.
68. Kang BK, Na DG, Ryoo JW, et al. Diffusion-Weighted MR Imaging of Intracerebral Hemorrhage. *Korean Journal of Radiology* 2001; 2: 183. DOI: 10.3348/kjr.2001.2.4.183.
69. Ladd ME. High-Field-Strength Magnetic Resonance: Potential and Limits. *Topics in Magnetic Resonance Imaging* 2007; 18: 139-152. DOI: 10.1097/RMR.0b013e3180f612b3.
70. Yoo PE, John SE, Farquharson S, et al. 7T-fMRI: Faster temporal resolution yields optimal BOLD sensitivity for functional network imaging specifically at high spatial resolution. *NeuroImage* 2018; 164: 214-229. DOI: 10.1016/j.neuroimage.2017.03.002.
71. Voigt S, Koemans EA, Rasing I, et al. Minocycline for sporadic and hereditary cerebral amyloid angiopathy (BATMAN): study protocol for a placebo-controlled randomized double-blind trial. *Trials* 2023; 24. DOI: 10.1186/s13063-023-07371-4.
72. van Rooden S, van Opstal AM, Labadie G, et al. Early Magnetic Resonance Imaging and Cognitive Markers of Hereditary Cerebral Amyloid Angiopathy. *Stroke* 2016; 47: 3041-3044. 2016/11/12. DOI: 10.1161/STROKEAHA.116.014418.
73. Schriemer SE, Hirschler L, van Etten ES, et al. Stimulating amyloid- β clearance in cerebral amyloid angiopathy with low-sodium oxybate and/or non-invasive vagus nerve stimulation (Clear-Brain): study protocol for a randomised pre-post trial. Submitted.

74. Aguilar MI and Brott TG. Update in Intracerebral Hemorrhage. *The Neurohospitalist* 2011; 1: 148-159. DOI: 10.1177/1941875211409050.
75. Vonsattel JPG, Myers RH, Tessa Hedley???Whyte E, et al. Cerebral amyloid angiopathy without and with cerebral hemorrhages: A comparative histological study. *Annals of Neurology* 1991; 30: 637-649. DOI: 10.1002/ana.410300503.
76. Charidimou A, Linn J, Vernooij MW, et al. Cortical superficial siderosis: detection and clinical significance in cerebral amyloid angiopathy and related conditions. *Brain* 2015; 138: 2126-2139. 2015/06/28. DOI: 10.1093/brain/awv162.
77. Reijmer YD, van Veluw SJ and Greenberg SM. Ischemic brain injury in cerebral amyloid angiopathy. *J Cereb Blood Flow Metab* 2016; 36: 40-54. 2015/05/07. DOI: 10.1038/jcbfm.2015.88.
78. Greenberg SM and Charidimou A. Diagnosis of Cerebral Amyloid Angiopathy. *Stroke* 2018; 49: 491-497. DOI: 10.1161/strokeaha.117.016990.
79. Perosa V, Oltmer J, Munting LP, et al. Perivascular space dilation is associated with vascular amyloid-beta accumulation in the overlying cortex. *Acta Neuropathol* 2022; 143: 331-348. 20211220. DOI: 10.1007/s00401-021-02393-1.
80. van Veluw SJ, Reijmer YD, van der Kouwe AJ, et al. Histopathology of diffusion imaging abnormalities in cerebral amyloid angiopathy. *Neurology* 2019; 92: e933-e943. 20190130. DOI: 10.1212/WNL.00000000000007005.
81. Dumas A, Dierksen GA, Gurol ME, et al. Functional magnetic resonance imaging detection of vascular reactivity in cerebral amyloid angiopathy. *Ann Neurol* 2012; 72: 76-81. DOI: 10.1002/ana.23566.
82. Peca S, McCreary CR, Donaldson E, et al. Neurovascular decoupling is associated with severity of cerebral amyloid angiopathy. *Neurology* 2013; 81: 1659-1665. DOI: 10.1212/01.wnl.0000435291.49598.54.
83. Finsterwalder S, Vlegels N, Gesierich B, et al. Small vessel disease more than Alzheimer's disease determines diffusion MRI alterations in memory clinic patients. *Alzheimer's & Dementia* 2020; 16: 1504-1514. DOI: 10.1002/alz.12150.
84. Salat DH, Smith EE, Tuch DS, et al. White matter alterations in cerebral amyloid angiopathy measured by diffusion tensor imaging. *Stroke* 2006; 37: 1759-1764. 20060608. DOI: 10.1161/01.STR.0000227328.86353.a7.
85. Reijmer YD, Fotiadis P, Martinez-Ramirez S, et al. Structural network alterations and neurological dysfunction in cerebral amyloid angiopathy. *Brain* 2015; 138: 179-188. 20141102. DOI: 10.1093/brain/awu316.
86. Reijmer YD, Fotiadis P, Charidimou A, et al. Relationship between white matter connectivity loss and cortical thinning in cerebral amyloid angiopathy. *Hum Brain Mapp* 2017; 38: 3723-3731. 20170430. DOI: 10.1002/hbm.23629.
87. Bakker E, van Broeckhoven C, Haan J, et al. DNA diagnosis for hereditary cerebral hemorrhage with amyloidosis (Dutch type). *Am J Hum Genet* 1991; 49: 518-521. 1991/09/01.
88. Zhang-Nunes SX, Maat-Schieman ML, van Duinen SG, et al. The cerebral beta-amyloid angiopathies: hereditary and sporadic. *Brain Pathol* 2006; 16: 30-39. 2006/04/15.

89. Shirzadi Z, Yau WW, Schultz SA, et al. Progressive White Matter Injury in Preclinical Dutch Cerebral Amyloid Angiopathy. *Ann Neurol* 2022; 92: 358-363. 2022/06/08. DOI: 10.1002/ana.26429.
90. Woolrich M, Ripley B, Brady M, et al. Temporal autocorrelation in univariate linear modeling of fMRI data. *NeuroImage* 2016; 14: 1370-1386.
91. Jenkinson M, Bannister P, Brady M, et al. Improved Optimization for the Robust and Accurate Linear Registration and Motion Correction of Brain Images. *NeuroImage* 2002; 17: 825-841. DOI: 10.1006/nimg.2002.1132.
92. Smith SM. Fast robust automated brain extraction. *Hum Brain Mapp* 2002; 17: 143-155. DOI: 10.1002/hbm.10062.
93. Beckmann CF and Smith SM. Probabilistic independent component analysis for functional magnetic resonance imaging. *IEEE Trans Med Imaging* 2004; 23: 137-152. DOI: 10.1109/tmi.2003.822821.
94. Amunts K, Malikovic A, Mohlberg H, et al. Brodmann's areas 17 and 18 brought into stereotaxic space-where and how variable? *NeuroImage* 2000; 11: 66-84. DOI: 10.1006/nimg.1999.0516.
95. Jenkinson M. Non-linear registration aka Spatial normalisation. In: 2007.
96. Smith SM, Jenkinson M, Woolrich MW, et al. Advances in functional and structural MR image analysis and implementation as FSL. *NeuroImage* 2004; 23: S208-S219. DOI: 10.1016/j.neuroimage.2004.07.051.
97. MathWorks I. *MATLAB : the language of technical computing : computation, visualization, programming : installation guide for UNIX version 5*. Natick : Math Works Inc., 1996., 1996.
98. Tournier JD, Smith R, Raffelt D, et al. MRtrix3: A fast, flexible and open software framework for medical image processing and visualisation. *NeuroImage* 2019; 202: 116137. DOI: 10.1016/j.neuroimage.2019.116137.
99. Kellner E, Dhital B, Kiselev VG, et al. Gibbs-ringing artifact removal based on local subvoxel-shifts. *Magnetic Resonance in Medicine* 2016; 76: 1574-1581. DOI: 10.1002/mrm.26054.
100. Baykara E, Gesierich B, Adam R, et al. A Novel Imaging Marker for Small Vessel Disease Based on Skeletonization of White Matter Tracts and Diffusion Histograms. *Annals of Neurology* 2016; 80: 581-592. DOI: 10.1002/ana.24758.
101. Smith SM, Jenkinson M, Johansen-Berg H, et al. Tract-based spatial statistics: Voxelwise analysis of multi-subject diffusion data. *NeuroImage* 2006; 31: 1487-1505. DOI: 10.1016/j.neuroimage.2006.02.024.
102. Charidimou A, Martinez-Ramirez S, Reijmer YD, et al. Total Magnetic Resonance Imaging Burden of Small Vessel Disease in Cerebral Amyloid Angiopathy: An Imaging-Pathologic Study of Concept Validation. *JAMA Neurol* 2016; 73: 994-1001. 2016/07/02. DOI: 10.1001/jamaneurol.2016.0832.
103. Caspers S, Moebus S, Lux S, et al. Studying variability in human brain aging in a population-based German cohort-rationale and design of 1000BRAINS. *Front Aging Neurosci* 2014; 6: 149. 2014/07/30. DOI: 10.3389/fnagi.2014.00149.
104. Beaudet G, Tsuchida A, Petit L, et al. Age-Related Changes of Peak Width Skeletonized Mean Diffusivity (PSMD) Across the Adult Lifespan: A Multi-Cohort Study. *Front Psychiatry* 2020; 11: 342. 20200504. DOI: 10.3389/fpsy.2020.00342.

105. R Development Core Team. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing, 2010.
106. Johnstone T, Ores Walsh KS, Greischar LL, et al. Motion correction and the use of motion covariates in multiple-subject fMRI analysis. *Hum Brain Mapp* 2006; 27: 779-788. DOI: 10.1002/hbm.20219.
107. Harris JJ and Attwell D. The energetics of CNS white matter. *J Neurosci* 2012; 32: 356-371. 2012/01/06. DOI: 10.1523/JNEUROSCI.3430-11.2012.
108. Masuda J, Tanaka K, Ueda K, et al. Autopsy study of incidence and distribution of cerebral amyloid angiopathy in Hisayama, Japan. *Stroke* 1988; 19: 205-210. DOI: 10.1161/01.str.19.2.205.
109. Vinters HV and Gilbert JJ. Cerebral amyloid angiopathy: incidence and complications in the aging brain. II. The distribution of amyloid vascular changes. *Stroke* 1983; 14: 924-928. DOI: 10.1161/01.str.14.6.924.
110. Van Dijk SE, Van Der Grond J, Lak J, et al. Longitudinal Progression of Magnetic Resonance Imaging Markers and Cognition in Dutch-Type Hereditary Cerebral Amyloid Angiopathy. *Stroke* 2022. DOI: 10.1161/strokeaha.121.035826.
111. Van Dijk SE, Lak J, Drenth N, et al. Aging Effect, Reproducibility, and Test–Retest Reliability of a New Cerebral Amyloid Angiopathy <scp>MRI</scp> Severity Marker—Cerebrovascular Reactivity to Visual Stimulation. *Journal of Magnetic Resonance Imaging* 2023; 57: 909-915. DOI: 10.1002/jmri.28362.
112. Van Harten TW, Van Rooden S, Koemans EA, et al. Impact of region of interest definition on visual stimulation-based cerebral vascular reactivity functional MRI with a special focus on applications in cerebral amyloid angiopathy. *NMR in Biomedicine* 2023; 36. DOI: 10.1002/nbm.4916.
113. Sleight E, Stringer MS, Marshall I, et al. Cerebrovascular Reactivity Measurement Using Magnetic Resonance Imaging: A Systematic Review. *Front Physiol* 2021; 12: 643468. 20210225. DOI: 10.3389/fphys.2021.643468.
114. Baykara E, Gesierich B, Adam R, et al. A Novel Imaging Marker for Small Vessel Disease Based on Skeletonization of White Matter Tracts and Diffusion Histograms. *Ann Neurol* 2016; 80: 581-592. 20160829. DOI: 10.1002/ana.24758.
115. McCreary CR, Beaudin AE, Subotic A, et al. Cross-sectional and longitudinal differences in peak skeletonized white matter mean diffusivity in cerebral amyloid angiopathy. *NeuroImage: Clinical* 2020; 27: 102280. DOI: 10.1016/j.nicl.2020.102280.
116. Wardlaw JM, Smith EE, Biessels GJ, et al. Neuroimaging standards for research into small vessel disease and its contribution to ageing and neurodegeneration. *The Lancet Neurology* 2013; 12: 822-838. DOI: 10.1016/s1474-4422(13)70124-8.
117. Pantoni L, Basile AM, Pracucci G, et al. Impact of age-related cerebral white matter changes on the transition to disability -- the LADIS study: rationale, design and methodology. *Neuroepidemiology* 2005; 24: 51-62. 2004/10/02. DOI: 10.1159/000081050.
118. Koemans EA. Temporal ordering of biomarkers in Dutch-type hereditary cerebral amyloid angiopathy Stroke, Unpublished.
119. Zhang-Nunes SX, Maat-Schieman MLC, Duinen SG, et al. The Cerebral β -Amyloid Angiopathies: Hereditary and Sporadic. *Brain Pathology* 2006; 16: 30-39. DOI: 10.1111/j.1750-3639.2006.tb00559.x.

120. Schipper MR, Vlegels N, van Harten TW, et al. Microstructural white matter integrity in relation to vascular reactivity in Dutch-type hereditary cerebral amyloid angiopathy. *Journal of Cerebral Blood Flow & Metabolism*; 0: 0271678X231200425. DOI: 10.1177/0271678x231200425.
121. van Veluw SJ, Benveniste H, Bakker E, et al. Is CAA a perivascular brain clearance disease? A discussion of the evidence to date and outlook for future studies. *Cell Mol Life Sci* 2024; 81: 239. 20240527. DOI: 10.1007/s00018-024-05277-1.
122. Mestre H, Kostrikov S, Mehta RI, et al. Perivascular spaces, glymphatic dysfunction, and small vessel disease. *Clin Sci (Lond)* 2017; 131: 2257-2274. 20170810. DOI: 10.1042/CS20160381.
123. Rasmussen MK, Mestre H and Nedergaard M. Fluid transport in the brain. *Physiol Rev* 2022; 102: 1025-1151. 20210505. DOI: 10.1152/physrev.00031.2020.
124. Brown R, Benveniste H, Black SE, et al. Understanding the role of the perivascular space in cerebral small vessel disease. *Cardiovascular Research* 2018; 114: 1462-1473. DOI: 10.1093/cvr/cvy113.
125. Potter GM, Chappell FM, Morris Z, et al. Cerebral Perivascular Spaces Visible on Magnetic Resonance Imaging: Development of a Qualitative Rating Scale and its Observer Reliability. *Cerebrovascular Diseases* 2015; 39: 224-231. DOI: 10.1159/000375153.
126. Sepehrband F, Barisano G, Sheikh-Bahaei N, et al. Image processing approaches to enhance perivascular space visibility and quantification using MRI. *Scientific Reports* 2019; 9. DOI: 10.1038/s41598-019-48910-x.
127. Donahue EK, Bui V, Foreman RP, et al. Magnetic resonance spectroscopy shows associations between neurometabolite levels and perivascular space volume in Parkinson's disease: a pilot and feasibility study. *Neuroreport* 2022; 33: 291-296. 20220408. DOI: 10.1097/WNR.0000000000001781.
128. Ballerini L, Booth T, Valdes Hernandez MDC, et al. Computational quantification of brain perivascular space morphologies: Associations with vascular risk factors and white matter hyperintensities. A study in the Lothian Birth Cohort 1936. *Neuroimage Clin* 2020; 25: 102120. 20191209. DOI: 10.1016/j.nicl.2019.102120.
129. Fandler-Hofler S, Ambler G, Banerjee G, et al. Temporal and Spatial Clustering of Intracerebral Hemorrhage in Cerebral Amyloid Angiopathy. *Neurology* 2024; 103: e209770. 20240816. DOI: 10.1212/WNL.00000000000209770.
130. Barisano G, Sheikh-Bahaei N, Law M, et al. Body mass index, time of day and genetics affect perivascular spaces in the white matter. *Journal of Cerebral Blood Flow & Metabolism* 2021; 41: 1563-1578. DOI: 10.1177/0271678x20972856.
131. Penny WD FK, Ashburner JT, et al. *Statistical Parametric Mapping: The Analysis of Functional Brain Images*. 1st ed. 2006.
132. Lutkenhoff ES, Rosenberg M, Chiang J, et al. Optimized Brain Extraction for Pathological Brains (optiBET). *PLoS ONE* 2014; 9: e115551. DOI: 10.1371/journal.pone.0115551.
133. Diedrichsen J, Balsters JH, Flavell J, et al. A probabilistic MR atlas of the human cerebellum. *Neuroimage* 2009; 46: 39-46. 20090205. DOI: 10.1016/j.neuroimage.2009.01.045.
134. Desikan RS, Segonne F, Fischl B, et al. An automated labeling system for subdividing the human cerebral cortex on MRI scans into gyral based regions of interest. *Neuroimage* 2006; 31: 968-980. 20060310. DOI: 10.1016/j.neuroimage.2006.01.021.

135. Jenkinson MaS, S. A global optimisation method for robust affine registration of brain images. 2001. DOI: 10.1016/s1361-8415(01)00036-6.
136. Mevislab.de. MeVisLab Documentation, <https://www.mevislab.de/developer/documentation> (2004).
137. Yoo TS AM, Lorensen WE, et al. . Engineering and algorithm design for an image processing Api: a technical report on ITK--the Insight Toolkit. *Stud Health Technol Inform* 2002; 85: 586-592. DOI: 10.3233/978-1-60750-929-5-586.
138. Frangi AF, Niessen WJ, Vincken KL, et al. Multiscale vessel enhancement filtering. Springer Berlin Heidelberg, 1998, pp.130-137.
139. Schipper MR, Vlegels N, van Harten TW, et al. Microstructural white matter integrity in relation to vascular reactivity in Dutch-type hereditary cerebral amyloid angiopathy. *J Cereb Blood Flow Metab* 2023; 43: 2144-2155. 20230914. DOI: 10.1177/0271678X231200425.
140. Charidimou A, Boulouis G, Pasi M, et al. MRI-visible perivascular spaces in cerebral amyloid angiopathy and hypertensive arteriopathy. *Neurology* 2017; 88: 1157-1164. 2017/02/24. DOI: 10.1212/WNL.0000000000003746.
141. Fazekas FK, R.; Roob, G.; Kleinert, G.; Kapeller, P.; Schmidt, R.; Hartung, H.P. Histopathological analysis of foci in signal loss on gradient-echo T2*-weighted MR images in patients with spontaneous intracerebral hemorrhage: evidence of microangiopathy-related microbleeds. *AJNR Am J Neuroradiol* 1999; 20: 637-642.
142. Knudsen KA, Rosand J, Karluk D, et al. Clinical diagnosis of cerebral amyloid angiopathy: validation of the Boston criteria. *Neurology* 2001; 56: 537-539. 2001/02/27.
143. Tsai H-H, Tsai L-K, Chen Y-F, et al. Correlation of Cerebral Microbleed Distribution to Amyloid Burden in Patients with Primary Intracerebral Hemorrhage. *Scientific Reports* 2017; 7: 44715. DOI: 10.1038/srep44715.
144. Dierksen GA, Skehan ME, Khan MA, et al. Spatial relation between microbleeds and amyloid deposits in amyloid angiopathy. *Ann Neurol* 2010; 68: 545-548. 2010/09/25. DOI: 10.1002/ana.22099.
145. Van Veluw SJ, Scherlek AA, Freeze WM, et al. Different microvascular alterations underlie microbleeds and microinfarcts. *Annals of Neurology* 2019; 86: 279-292. DOI: 10.1002/ana.25512.
146. Treaba CA, Granberg TE, Sormani MP, et al. Longitudinal Characterization of Cortical Lesion Development and Evolution in Multiple Sclerosis with 7.0-T MRI. *Radiology* 2019; 291: 740-749. DOI: 10.1148/radiol.2019181719.
147. Krijnen EA, Jelgerhuis J, Van Dam M, et al. Evolution of Cortical Lesions and Function-Specific Cognitive Decline in People With Multiple Sclerosis. *Neurology* 2025; 104: e213650. 20250512. DOI: 10.1212/WNL.00000000000213650.
148. Howell OW, Reeves CA, Nicholas R, et al. Meningeal inflammation is widespread and linked to cortical pathology in multiple sclerosis. *Brain* 2011; 134: 2755-2771. DOI: 10.1093/brain/awr182.
149. Rohr SO, Greiner T, Joost S, et al. Aquaporin-4 Expression during Toxic and Autoimmune Demyelination. *Cells* 2020; 9: 2187. DOI: 10.3390/cells9102187.
150. Aoki-Yoshino K, Uchihara T, Duyckaerts C, et al. Enhanced expression of aquaporin 4 in human

Appendix

brain with inflammatory diseases. *Acta Neuropathologica* 2005; 110: 281-288. DOI: 10.1007/s00401-005-1052-2.

151. Gabr RE, Lincoln JA, Hasan KM, et al. Functional assessment of the dural lymphatic vessels using dynamic contrast MRI in multiple sclerosis. *Brain and Behavior* 2023; 13. DOI: 10.1002/brb3.3042.

152. Low A, Mak E, Rowe JB, et al. Inflammation and cerebral small vessel disease: A systematic review. *Ageing Research Reviews* 2019; 53. DOI: 10.1016/j.arr.2019.100916.

153. Panteleienko L, Banerjee G, Mallon DH, et al. Sulcal Hyperintensity as an Early Imaging Finding in Cerebral Amyloid Angiopathy-Related Inflammation. *Neurology* 2024; 103: e210084. 20241125. DOI: 10.1212/WNL.0000000000210084.

154. Charidimou A. Diagnosing Cerebral Amyloid Angiopathy-Related Inflammation. *Neurology* 2024; 103: e209647. 20240620. DOI: 10.1212/WNL.0000000000209647.

155. Martucci M, Sarria S, Toledo M, et al. Cerebral amyloid angiopathy-related inflammation: imaging findings and clinical outcome. *Neuroradiology* 2014; 56: 283-289. DOI: 10.1007/s00234-014-1330-6.

156. Traschutz A, Tzaridis T, Penner AH, et al. Reduction of microbleeds by immunosuppression in a patient with Abeta-related vascular inflammation. *Neurol Neuroimmunol Neuroinflamm* 2015; 2: e165. 20151015. DOI: 10.1212/NXI.0000000000000165.

157. Koemans EA, Voigt S, Rasing I, et al. Cerebellar Superficial Siderosis in Cerebral Amyloid Angiopathy. *Stroke* 2022; 53: 552-557. DOI: 10.1161/strokeaha.121.035019.

158. Koemans EA, Van Harten TW, Voigt S, et al. One Size Does Not Fit All: Micro-, Meso-, and Macrobleeds in Cerebral Amyloid Angiopathy. *Cerebrovascular Diseases* 2024: 1-8. DOI: 10.1159/000540899.

159. Fischl B. Automatically Parcellating the Human Cerebral Cortex. *Cerebral Cortex* 2004; 14: 11-22. DOI: 10.1093/cercor/bhg087.

160. Fischl B, Sereno MI, Tootell RBH, et al. High-resolution intersubject averaging and a coordinate system for the cortical surface. *Human Brain Mapping* 1999; 8: 272-284. DOI: 10.1002/(sici)1097-0193(1999)8:4<272::aid-hbm10>3.0.co;2-4.

161. Fischl BS, M. I., Dale., A. M. Cortical surface-based analysis, II inflation, flattening, and a surface-based coordinate system. *NeuroImage* 1999; 9: 195-207. DOI: 10.1006/nimg.1998.0396.

162. D. Louis Collins CJH, T. M. Peters, A. C. Evans. Automatic 3-D model-based neuroanatomical segmentation. *Human Brain Mapping* 1995; 3: 190-208. DOI: 10.1002/hbm.460030304.

163. Mazziotta J, Toga A, Evans A, et al. A probabilistic atlas and reference system for the human brain: International Consortium for Brain Mapping (ICBM). *Philosophical Transactions of the Royal Society of London Series B: Biological Sciences* 2001; 356: 1293-1322. DOI: 10.1098/rstb.2001.0915.

164. Frazier JA, Chiu S, Breeze JL, et al. Structural Brain Magnetic Resonance Imaging of Limbic and Thalamic Volumes in Pediatric Bipolar Disorder. *American Journal of Psychiatry* 2005; 162: 1256-1265. DOI: 10.1176/appi.ajp.162.7.1256.

165. Goldstein JM, Seidman LJ, Makris N, et al. Hypothalamic abnormalities in schizophrenia: sex effects and genetic vulnerability. *Biol Psychiatry* 2007; 61: 935-945. 20061013. DOI: 10.1016/j.

biopsych.2006.06.027.

166. Makris N, Goldstein JM, Kennedy D, et al. Decreased volume of left and total anterior insular lobule in schizophrenia. *Schizophr Res* 2006; 83: 155-171. 20060131. DOI: 10.1016/j.schres.2005.11.020.
167. R Core Team (2023). R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria. <https://www.R-project.org>.
168. Bates D, Mächler M, Bolker B, et al. Fitting Linear Mixed-Effects Models Using lme4. *Journal of Statistical Software* 2015; 67: 1 - 48. DOI: 10.18637/jss.v067.i01.
169. Szalardy L, Fakan B, Maszlag-Torok R, et al. Identifying diagnostic and prognostic factors in cerebral amyloid angiopathy-related inflammation: A systematic analysis of published and seven new cases. *Neuropathology and Applied Neurobiology* 2024; 50. DOI: 10.1111/nan.12946.
170. Eide PK and Ringstad G. Functional analysis of the human perivascular subarachnoid space. *Nature Communications* 2024; 15. DOI: 10.1038/s41467-024-46329-1.
171. Li Y, Rusinek H, Butler T, et al. Decreased CSF clearance and increased brain amyloid in Alzheimer's disease. *Fluids and Barriers of the CNS* 2022; 19. DOI: 10.1186/s12987-022-00318-y.
172. Kim J, Barcus RA, Lipford ME, et al. The Role of Cerebrospinal Fluid Flow in the Subarachnoid Space in Beta-Amyloid Clearance. *Alzheimer's & Dementia* 2023; 19. DOI: 10.1002/alz.071202.
173. Simon MJ and Iliff JJ. Regulation of cerebrospinal fluid (CSF) flow in neurodegenerative, neurovascular and neuroinflammatory disease. *Biochim Biophys Acta* 2016; 1862: 442-451. 20151022. DOI: 10.1016/j.bbadis.2015.10.014.
174. Kameya N, Sakai I, Saito K, et al. Evolutionary changes leading to efficient glymphatic circulation in the mammalian brain. *Nat Commun* 2024; 15: 10048. 20241204. DOI: 10.1038/s41467-024-54372-1.
175. Ringstad G, Vatnehol SAS and Eide PK. Glymphatic MRI in idiopathic normal pressure hydrocephalus. *Brain* 2017; 140: 2691-2705. DOI: 10.1093/brain/awx191.
176. Bèchet NB, Shanbhag NC and Lundgaard I. Glymphatic pathways in the gyrencephalic brain. *Journal of Cerebral Blood Flow & Metabolism* 2021; 41: 2264-2279. DOI: 10.1177/0271678x21996175.
177. Lin HY, Huang CC, Chou KH, et al. Differential Patterns of Gyral and Sulcal Morphological Changes During Normal Aging Process. *Front Aging Neurosci* 2021; 13: 625931. 20210203. DOI: 10.3389/fnagi.2021.625931.
178. Maat-Schieman ML, van Duinen SG, Bornebroek M, et al. Hereditary cerebral hemorrhage with amyloidosis-Dutch type (HCHWA-D): II--A review of histopathological aspects. *Brain Pathol* 1996; 6: 115-120. 1996/04/01.
179. Bornebroek M, Haan J, Maat-Schieman ML, et al. Hereditary cerebral hemorrhage with amyloidosis-Dutch type (HCHWA-D): I--A review of clinical, radiologic and genetic aspects. *Brain Pathol* 1996; 6: 111-114. 1996/04/01.
180. Koemans EA, Chhatwal JP, van Veluw SJ, et al. Progression of cerebral amyloid angiopathy: a pathophysiological framework. *The Lancet Neurology* 2023; 22: 632-642. 20230523. DOI: 10.1016/s1474-4422(23)00114-x.

Appendix

181. van der Plas MC, Koemans EA, Schipper MR, et al. One-Year Radiologic Progression in Sporadic and Hereditary Cerebral Amyloid Angiopathy. *Neurology* 2025; 104: e213546. 20250408. DOI: 10.1212/wnl.00000000000213546.
182. Rosand J, Muzikansky A, Kumar A, et al. Spatial clustering of hemorrhages in probable cerebral amyloid angiopathy. *Annals of neurology* 2005; 58: 459-462. DOI: 10.1002/ana.20596.
183. Perosa V, Auger CA, Zanon Zotin MC, et al. Histopathological Correlates of Lobar Microbleeds in False-Positive Cerebral Amyloid Angiopathy Cases. *Annals of neurology* 2023; 94: 856-870. 20230906. DOI: 10.1002/ana.26761.
184. van Veluw SJ, Scherlek AA, Freeze WM, et al. Different microvascular alterations underlie microbleeds and microinfarcts. *Annals of neurology* 2019; 86: 279-292. 20190614. DOI: 10.1002/ana.25512.
185. Voigt S, Amlal S, Koemans EA, et al. Spatial and temporal intracerebral hemorrhage patterns in Dutch-type hereditary cerebral amyloid angiopathy. *International Journal of Stroke* 2021: 174749302110570. DOI: 10.1177/17474930211057022.
186. Attems J, Jellinger KA and Lintner F. Alzheimer's disease pathology influences severity and topographical distribution of cerebral amyloid angiopathy. *Acta neuropathologica* 2005; 110: 222-231. 20050825. DOI: 10.1007/s00401-005-1064-y.
187. Johnson KA, Gregas M, Becker JA, et al. Imaging of amyloid burden and distribution in cerebral amyloid angiopathy. *Annals of neurology* 2007; 62: 229-234. DOI: 10.1002/ana.21164.
188. McCarter SJ, Lesnick TG, Lowe V, et al. Cerebral Amyloid Angiopathy Pathology and Its Association With Amyloid- β PET Signal. *Neurology* 2021; 97: e1799-e1808. 20210909. DOI: 10.1212/wnl.00000000000012770.
189. Regenhardt RW, Thon JM, Das AS, et al. Association Between Immunosuppressive Treatment and Outcomes of Cerebral Amyloid Angiopathy-Related Inflammation. *JAMA Neurology* 2020; 77: 1261. DOI: 10.1001/jamaneurol.2020.1782.
190. Duering M, Biessels GJ, Brodtmann A, et al. Neuroimaging standards for research into small vessel disease-advances since 2013. *The Lancet Neurology* 2023; 22: 602-618. 20230523. DOI: 10.1016/s1474-4422(23)00131-x.
191. Koemans EA, van Harten TW, Voigt S, et al. One Size Does Not Fit All: Micro-, Meso-, and Macrobleeds in Cerebral Amyloid Angiopathy. *Cerebrovascular diseases (Basel, Switzerland)* 2024: 1-8. 20240819. DOI: 10.1159/000540899.
192. MeVisLab.de. MeVisLab Documentation, <https://www.mevislab.de/development/documentation> (accessed 25-02-2025 2025).
193. Mutsaerts HJ, van Dalen JW, Heijtel DF, et al. Cerebral Perfusion Measurements in Elderly with Hypertension Using Arterial Spin Labeling. *PLoS One* 2015; 10: e0133717. 20150804. DOI: 10.1371/journal.pone.0133717.
194. Jenkinson M, Bannister P, Brady M, et al. Improved optimization for the robust and accurate linear registration and motion correction of brain images. *Neuroimage* 2002; 17: 825-841. DOI: 10.1016/s1053-8119(02)91132-8.
195. Jenkinson M and Smith S. A global optimisation method for robust affine registration of brain

- images. *Med Image Anal* 2001; 5: 143-156. DOI: 10.1016/s1361-8415(01)00036-6.
196. Jesper L. R. Andersson MJaSS. *Non-linear registration aka Spatial normalisation*. 2007. FMRIB Centre, Oxford, United Kingdom.
197. Edelman D and Goeman J. A regression perspective on generalized distance covariance and the Hilbert–Schmidt independence criterion. *Statistical Science* 2022; 37: 562-579.
198. Hemerik J and Goeman J. Exact testing with random permutations. *Test (Madr)* 2018; 27: 811-825. 20171130. DOI: 10.1007/s11749-017-0571-1.
199. Smith EE, Vijayappa M, Lima F, et al. Impaired visual evoked flow velocity response in cerebral amyloid angiopathy. *Neurology* 2008; 71: 1424-1430. DOI: 10.1212/01.wnl.0000327887.64299.a4.
200. Mesker DJ, Poels MM, Ikram MA, et al. Lobar distribution of cerebral microbleeds: the Rotterdam Scan Study. *Archives of neurology* 2011; 68: 656-659. DOI: 10.1001/archneurol.2011.93.
201. Pettersen JA, Sathiyamoorthy G, Gao FQ, et al. Microbleed topography, leukoaraiosis, and cognition in probable Alzheimer disease from the Sunnybrook dementia study. *Archives of neurology* 2008; 65: 790-795. DOI: 10.1001/archneur.65.6.790.
202. van Laar PJ, Hendrikse J, Golay X, et al. In vivo flow territory mapping of major brain feeding arteries. *Neuroimage* 2006; 29: 136-144. 20050810. DOI: 10.1016/j.neuroimage.2005.07.011.
203. Tschoe C, Bushnell CD, Duncan PW, et al. Neuroinflammation after Intracerebral Hemorrhage and Potential Therapeutic Targets. *J Stroke* 2020; 22: 29-46. 20200131. DOI: 10.5853/jos.2019.02236.
204. Keep RF, Zhou N, Xiang J, et al. Vascular disruption and blood-brain barrier dysfunction in intracerebral hemorrhage. *Fluids Barriers CNS* 2014; 11: 18. 20140810. DOI: 10.1186/2045-8118-11-18.
205. Alten B, Gokcal E, Warren A, et al. Cerebrospinal Fluid Beta-Amyloid Concentration and Clinical and Radiographic Manifestations of Cerebral Amyloid Angiopathy. *J Am Heart Assoc* 2025; 14: e040025. 20250303. DOI: 10.1161/jaha.124.040025.
206. Kuo PY, Tsai HH, Lee BC, et al. Differences in lobar microbleed topography in cerebral amyloid angiopathy and hypertensive arteriopathy. *Sci Rep* 2024; 14: 3774. 20240215. DOI: 10.1038/s41598-024-54243-1.
207. Switzer AR, Cheema I, McCreary CR, et al. Cerebrovascular reactivity in cerebral amyloid angiopathy, Alzheimer disease, and mild cognitive impairment. *Neurology* 2020; 95: e1333-e1340. 20200708. DOI: 10.1212/wnl.0000000000010201.
208. Jäkel L, De Kort AM, Klijn CJM, et al. Prevalence of cerebral amyloid angiopathy: A systematic review and meta-analysis. *Alzheimers Dement* 2022; 18: 10-28. 20210531. DOI: 10.1002/alz.12366.
209. Cordonnier C, Al-Shahi Salman R and Wardlaw J. Spontaneous brain microbleeds: systematic review, subgroup analyses and standards for study design and reporting. *Brain* 2007; 130: 1988-2003. 20070224. DOI: 10.1093/brain/awl387.
210. Greenberg SM, Vernooij MW, Cordonnier C, et al. Cerebral microbleeds: a guide to detection and interpretation. *Lancet Neurol* 2009; 8: 165-174. 2009/01/24. DOI: 10.1016/s1474-4422(09)70013-4.
211. Cordonnier C. Brain microbleeds: more evidence, but still a clinical dilemma. *Curr Opin Neurol* 2011; 24: 69-74. DOI: 10.1097/WCO.0b013e328341f8c0.

Appendix

212. Greenberg SM, Vernooij MW, Cordonnier C, et al. Cerebral microbleeds: a guide to detection and interpretation. *Lancet Neurol* 2009; 8: 165-174. DOI: 10.1016/S1474-4422(09)70013-4.
213. Boulouis G, Charidimou A and Greenberg SM. Sporadic Cerebral Amyloid Angiopathy: Pathophysiology, Neuroimaging Features, and Clinical Implications. *Semin Neurol* 2016; 36: 233-243. 2016/05/24. DOI: 10.1055/s-0036-1581993.
214. Gatti L, Tinelli F, Scelzo E, et al. Understanding the Pathophysiology of Cerebral Amyloid Angiopathy. *Int J Mol Sci* 2020; 21: 20200513. DOI: 10.3390/ijms21103435.
215. De Cocker LJ, Lindenholz A, Zwanenburg JJ, et al. Clinical vascular imaging in the brain at 7T. *Neuroimage* 2018; 168: 452-458. 20161118. DOI: 10.1016/j.neuroimage.2016.11.044.
216. Koemans EA, van Etten ES, van Opstal AM, et al. Innovative Magnetic Resonance Imaging Markers of Hereditary Cerebral Amyloid Angiopathy at 7 Tesla. *Stroke* 2018; 49: 1518-1520. 2018/04/27. DOI: 10.1161/STROKEAHA.117.020302.
217. Penckofer M, Kazmi KS, Thon J, et al. Neuro-imaging in intracerebral hemorrhage: updates and knowledge gaps. *Front Neurosci* 2024; 18: 1408288. 20240509. DOI: 10.3389/fnins.2024.1408288.
218. Kang BK, Na DG, Ryoo JW, et al. Diffusion-weighted MR imaging of intracerebral hemorrhage. *Korean J Radiol* 2001; 2: 183-191. DOI: 10.3348/kjr.2001.2.4.183.
219. Boulouis G, Charidimou A, Van Veluw S, et al. Imaging the Acute Formation of a Cortical Microbleed in Cerebral Amyloid Angiopathy. *JAMA Neurol* 2017; 74: 120-121. 2016/11/29. DOI: 10.1001/jamaneurol.2016.3445.
220. Koemans EA, Rasing I, Voigt S, et al. Temporal Ordering of Biomarkers in Dutch-Type Hereditary Cerebral Amyloid Angiopathy. *Stroke* 2024; 55: 954-962. 20240306. DOI: 10.1161/strokeaha.123.044688.
221. Greenberg SM and Charidimou A. Diagnosis of Cerebral Amyloid Angiopathy: Evolution of the Boston Criteria. *Stroke* 2018; 49: 491-497. 2018/01/18. DOI: 10.1161/STROKEAHA.117.016990.
222. Wardlaw JM, Smith EE, Biessels GJ, et al. Neuroimaging standards for research into small vessel disease and its contribution to ageing and neurodegeneration. *Lancet Neurol* 2013; 12: 822-838. DOI: 10.1016/s1474-4422(13)70124-8.
223. Benjamin P, Viessmann O, MacKinnon AD, et al. 7 Tesla MRI in cerebral small vessel disease. *Int J Stroke* 2015; 10: 659-664. 20150406. DOI: 10.1111/ijvs.12490.
224. Perosa V, Rotta J, Yakupov R, et al. Implications of quantitative susceptibility mapping at 7 Tesla MRI for microbleeds detection in cerebral small vessel disease. *Front Neurol* 2023; 14: 1112312. 20230315. DOI: 10.3389/fneur.2023.1112312.
225. Brundel M, Heringa SM, de Bresser J, et al. High prevalence of cerebral microbleeds at 7Tesla MRI in patients with early Alzheimer's disease. *Journal of Alzheimer's disease : JAD* 2012; 31 2: 259-263.
226. de Bresser J, Brundel M, Conijn MM, et al. Visual cerebral microbleed detection on 7T MR imaging: reliability and effects of image processing. *AJNR Am J Neuroradiol* 2013; 34: E61-64. 20120216. DOI: 10.3174/ajnr.A2960.
227. Conijn MM, Geerlings MI, Biessels GJ, et al. Cerebral microbleeds on MR imaging: comparison between 1.5 and 7T. *AJNR Am J Neuroradiol* 2011; 32: 1043-1049. 2011/05/07. DOI: 10.3174/ajnr.A2450.

228. Conijn MM, Geerlings MI, Luijten PR, et al. Visualization of cerebral microbleeds with dual-echo T2*-weighted magnetic resonance imaging at 7.0 T. *J Magn Reson Imaging* 2010; 32: 52-59. DOI: 10.1002/jmri.22223.
229. Voigt S, Amlal S, Koemans EA, et al. Spatial and temporal intracerebral hemorrhage patterns in Dutch-type hereditary cerebral amyloid angiopathy. *Int J Stroke* 2022; 17: 793-798. 20211118. DOI: 10.1177/17474930211057022.
230. Charidimou A, Boulouis G, Frosch MP, et al. The Boston criteria version 2.0 for cerebral amyloid angiopathy: a multicentre, retrospective, MRI-neuropathology diagnostic accuracy study. *Lancet Neurol* 2022; 21: 714-725. DOI: 10.1016/s1474-4422(22)00208-3.
231. van Veluw SJ, Biessels GJ, Klijn CJ, et al. Heterogeneous histopathology of cortical microbleeds in cerebral amyloid angiopathy. *Neurology* 2016; 86: 867-871. 2016/02/05. DOI: 10.1212/WNL.0000000000002419.
232. van Veluw SJ, Charidimou A, van der Kouwe AJ, et al. Microbleed and microinfarct detection in amyloid angiopathy: a high-resolution MRI-histopathology study. *Brain* 2016; 139: 3151-3162. 2016/09/21. DOI: 10.1093/brain/aww229.
233. Jolink WMT, van Veluw SJ, Zwanenburg JJM, et al. Histopathology of Cerebral Microinfarcts and Microbleeds in Spontaneous Intracerebral Hemorrhage. *Transl Stroke Res* 2023; 14: 174-184. 20220406. DOI: 10.1007/s12975-022-01016-5.
234. Harris JJ and Attwell D. The Energetics of CNS White Matter. *The Journal of Neuroscience* 2012; 32: 356-371. DOI: 10.1523/jneurosci.3430-11.2012.
235. Zhou Y, Wang Y, Wang J, et al. Inflammation in intracerebral hemorrhage: from mechanisms to clinical translation. *Prog Neurobiol* 2014; 115: 25-44. 20131126. DOI: 10.1016/j.pneurobio.2013.11.003.
236. Loan JJ, Kirby C, Emelianova K, et al. Secondary injury and inflammation after intracerebral haemorrhage: a systematic review and meta-analysis of molecular markers in patient brain tissue. *Journal of Neurology, Neurosurgery & Psychiatry* 2022; 93: 126-132. DOI: 10.1136/jnnp-2021-327098.
237. Zimmer JA, Ardayfio P, Wang H, et al. Amyloid-Related Imaging Abnormalities With Donanemab in Early Symptomatic Alzheimer Disease. *JAMA Neurology* 2025; 82: 461. DOI: 10.1001/jamaneurol.2025.0065.
238. Doran SJ and Sawyer RP. Risk factors in developing amyloid related imaging abnormalities (ARIA) and clinical implications. *Front Neurosci* 2024; 18: 1326784. 20240119. DOI: 10.3389/fnins.2024.1326784.
239. Greenberg SM. CAA and ARIA: Emerging Concepts and Opportunities for Translation. *Alzheimer's & Dementia* 2025; 20. DOI: 10.1002/alz.085576.
240. Hampel H, Elhage A, Cho M, et al. Amyloid-related imaging abnormalities (ARIA): radiological, biological and clinical characteristics. *Brain* 2023; 146: 4414-4424. DOI: 10.1093/brain/awad188.
241. Beaudin AE, McCreary CR, Mazerolle EL, et al. Cerebrovascular Reactivity Across the Entire Brain in Cerebral Amyloid Angiopathy. *Neurology* 2022; 98: e1716-e1728. 20220224. DOI: 10.1212/WNL.0000000000200136.
242. Park L, Koizumi K, El Jamal S, et al. Age-Dependent Neurovascular Dysfunction and

Appendix

Damage in a Mouse Model of Cerebral Amyloid Angiopathy. *Stroke* 2014; 45: 1815-1821. DOI: 10.1161/strokeaha.114.005179.

243. Shin HK, Jones PB, Garcia-Alloza M, et al. Age-dependent cerebrovascular dysfunction in a transgenic mouse model of cerebral amyloid angiopathy. *Brain* 2007; 130: 2310-2319. DOI: 10.1093/brain/awm156.

244. Roefs ECA, Schipper MR, van Haren G, et al. Vasomotor dysfunction in cerebral amyloid angiopathy characterized by VASO and BOLD fMRI *ISMRM 2026*.

245. Hirschler L, Runderkamp BA, Decker A, et al. Region-specific drivers of CSF mobility measured with MRI in humans. *Nature Neuroscience* 2025; 28: 2392-2401. DOI: 10.1038/s41593-025-02073-3.

246. Mukherjee S, Mirzaee M and Tithof J. Quantifying the relationship between spreading depolarization and perivascular cerebrospinal fluid flow. *Sci Rep* 2023; 13: 12405. 20230731. DOI: 10.1038/s41598-023-38938-5.

247. Lynch KM, Sepehrband F, Toga AW, et al. Brain perivascular space imaging across the human lifespan. Cold Spring Harbor Laboratory, 2022.

248. Boutet A, Son HJ, Malik M, et al. Enlarging and shrinking focal perivascular spaces. *Neuroradiol J* 2025; 38: 224-229. 20240402. DOI: 10.1177/19714009241242642.

249. Menze I, Bernal J, Kaya P, et al. Perivascular space enlargement accelerates in ageing and Alzheimer's disease pathology: evidence from a three-year longitudinal multicentre study. *Alzheimer's Research & Therapy* 2024; 16. DOI: 10.1186/s13195-024-01603-8.

250. Barisano G, Iv M, Choupan J, et al. Robust, fully-automated assessment of cerebral perivascular spaces and white matter lesions: a multicentre MRI longitudinal study of their evolution and association with risk of dementia and accelerated brain atrophy. *EBioMedicine* 2025; 111: 105523. 20241224. DOI: 10.1016/j.ebiom.2024.105523.

251. Fultz NE, Ringstad G, Debiase M, et al. Non-invasive Characterization of Perivascular Subarachnoid Spaces. *Submitted*.

252. Vijayakrishnan Nair V, Kish BR, Inglis B, et al. Human CSF movement influenced by vascular low frequency oscillations and respiration. *Frontiers in Physiology* 2022; 13. DOI: 10.3389/fphys.2022.940140.

253. Renard D, Tatu L, Collombier L, et al. Cerebral Amyloid Angiopathy and Cerebral Amyloid Angiopathy-Related Inflammation: Comparison of Hemorrhagic and DWI MRI Features. *J Alzheimers Dis* 2018; 64: 1113-1121. DOI: 10.3233/JAD-180269.

254. Puy L, Barus R, Pasi M, et al. Distinct neuroinflammatory patterns between cerebral microbleeds and microinfarcts in cerebral amyloid angiopathy. *Annals of Clinical and Translational Neurology* 2024; 11: 3328-3332. DOI: 10.1002/acn3.52226.

Portfolio

PhD trajectory – an overview

During my PhD, I engaged in a variety of activities and efforts beyond those presented in the scientific chapters of this thesis, a selection is highlighted in this chapter.

The first year of my PhD started during the COVID pandemic, however, the LUMC's (Dutch-type) Cerebral Amyloid Angiopathy ((D-)CAA) studies I was involved in quickly resumed their research visits, provided that participants and investigators had no COVID symptoms. I learned how to perform or assist with every aspect of the research visits as presented in [Chapter 1](#), Table 1.2, under the supervision of fellow PhD-students Thijs van Harten, Sabine Voigt, Kanishk Kaushik, Emma Koemans, and Ingeborg Rasing. Together with fellow PhD-students Reinier van der Zwet, Rosemarie van Dort, and Sanne Schriemer, we later took over these visits for all CAA studies. I obtained scan brevets for 3 Tesla (T) and 7T magnetic resonance imaging (MRI) and was responsible for assessing participants' MRI safety profiles, under the guidance of medical physicist Nick de Jong and senior scientist Wouter Teeuwisse. My contribution to the research visits focused on MRI data acquisition and curation and MRI post-processing, as I took this over from Thijs van Harten. Thijs also introduced me to and mentored me in various post-processing steps and software, including functional (f)MRI post-processing and MeVisLab.

During my first year, I worked on a side project assessing the robustness of fMRI measures across two 3T MR systems and visual stimulation setups. This project was published as conference abstract and demonstrated limited bias in longitudinal and cross-sectional comparisons. This was of importance as the TRACK (AURORA-PLUS) study was planned to be performed on a different MR system than the already running AURORA study. The first main project I worked on is presented in [Chapter 2](#), and includes two MRI analyses – diffusion tensor imaging and visually stimulated fMRI analysis – revealing the association between microstructural white matter integrity and vasoreactivity in (D-)CAA. Also, the project presented in [Chapter 7](#), identifying the occurrence of acute cerebral microbleeds (CMBs) on 7T MRI in (D-)CAA, was initiated.

A project that I focused on during the second year, involved development and validation of a pipeline to quantify perivascular spaces (PVS) in the presence of large, disruptive pathology, as observed in (D-)CAA ([Chapter 4](#)). With this project we identified PVS volume fraction as early marker in (D-)CAA. Later this year, I presented the project at the international CAA conference, where I was awarded the prize for best oral presentation.

During the third year, my role in checking MRI participant safety profiles transitioned into a supervisory role, as research assistants (Yasmine Alladin, Nikki Hengeveld and Isabel Kohlmann) took over this task. In this year, I was awarded best poster and best oral presentation prizes at the LUMC Neuroscience Symposium and the CAA early career researchers conference, respectively. Also, I performed the fMRI baseline and one-year follow-up analysis in the (D-)CAA cohorts, revealing variation in vasoreactivity over time, which was included in van der Plas et al., 2025 (<https://doi.org/10.1212/WNL.0000000000213546>) and resulted in [Chapter 3](#). In addition, the project presented in [Chapter 5](#) was initiated, showing sulcal predominance of CMBs. Lastly, together with Ellen Stijl – 't Hart, Rosemarie van Dort, and Sanne Schriemer, I co-organized a (D-)CAA information day for patients, families, and caregivers, which was

attended by over 250 participants. This also included an item and follow-up session on early discussions regarding potential inclusion of D-CAA as a disease subgroup in the cAPPricorn clinical trial.

During the later stages of my PhD, the project presented in [Chapter 6](#) was initiated. Here we identified interindividual CMB clustering in (D-)CAA using a flow territory-based analysis.

Over the course of my PhD, as a team we have acquired roughly 900 MRI scans, reflecting the substantial time investment and coordination required for large-scale data collection. In parallel, I have supervised five students; Arie-Tjerk Razoux-Schultz (bachelor's internship), Lex Otto and Roelof Schraa (BSc gap-year students), Abel Thissen (master's student), and Maria Andrikopoulou (master's student who worked as research assistant). Also, I was involved in the organization and management of four recurring meetings, of which two meetings were new initiatives. One was established for focused sessions and workshops for clinical neuro-focused researchers and the other was a radiology-wide initiative to promote interdisciplinary contact and collaboration. Furthermore, I contributed to annually distributed (D-)CAA newsletters, which communicate research and recent publications to people with (D-)CAA, family members, research participants, and health care professionals.

Characterizing for my PhD has been its collaborative nature. A major study I worked on was the multicenter TRACK study; a collaboration between researchers from Leiden, Perth, and Boston. Furthermore, working within the C.J. Gorter MRI Center and closely collaborating with the neurology department, fostered a multidisciplinary team and research environment – facilitating further collaborations for post-processing support of the headache and neurovascular (non-CAA) subgroups of the neurology department.

CRediT statement for the chapters in this thesis

CRediT table for the thesis of Manon R. Schipper																				
Ch.	Type*	Short Title	Conceptualization	Data Curation	Formal Analysis	Funding Acquisition	Investigation	Methodology	Project Administration	Resources	Software	Supervision	Validation	Visualization	Writing – Original Draft	Writing – Review & Editing	Preregistered	Preprinted	Published with Peer Review	
1	Introduction																			
2	PhD project chapter	DTI vs. fMRI in D-CAA																		<input checked="" type="checkbox"/>
3	PhD project chapter	fMRI follow-up in (D-)CAA																		<input type="checkbox"/>
4	PhD project chapter	PVS in D-CAA																		<input checked="" type="checkbox"/>
5	PhD project chapter	CMB sulcus vs. gyrus																		<input type="checkbox"/>
6	PhD project chapter	CMB flow territories																		<input type="checkbox"/>
7	PhD project chapter	Acute CMB on 7T MRI																		<input checked="" type="checkbox"/>
8	Discussion																			<input type="checkbox"/>

*PhD project chapters are the direct result of the PhD project of the PhD candidate. Some theses also include Collaboration Chapters, to which the PhD candidate has contributed but fall outside the PhD project.

Publication and dissemination table

1. Schipper, M. R.*, van Harten, T. W., van Bronkhorst, M., van der Plas, M. C. E., Wermer, M. J. H., van Walderveen, M. A. A., and van Osch, M. J. P. (2022). Vascular reactivity measurements are insensitive to changes in visual stimulus presentation method. Conference abstract – poster presentation at ISMRM Benelux 2022. Not part of this thesis.
2. Schipper, M. R.*, Vlegels, N.* , Voigt, S., van Harten, T. W., de Luca, A., Rasing, I., Biessels, G.J., van Osch, M.J.P., van Walderveen, M.A.A., and Wermer M.J.H. (2022). The relationship between vascular reactivity and microstructural white matter integrity in (pre-)symptomatic Dutch-type cerebral amyloid angiopathy. Conference abstract – power pitch at World Stroke Conference 2022. Not part of this thesis.
3. Schipper, M. R.*, Vlegels, N.* , van Harten, T. W., Voigt, S., Koemans, E. A., Rasing, I., de Luca, A., Kaushik, K., van Etten, E.S., van Buchem, M.A., Terwindt, G.M., Biessels, G.J., van Osch, M.J.P., van Walderveen, M.A.A., and Wermer, M.J.H. (2022). The relationship between vascular reactivity and microstructural white matter integrity in (pre-)symptomatic Dutch-type cerebral amyloid angiopathy. Conference abstract – poster presentation at International CAA conference 2022. Not part of this thesis.
4. Schipper, M. R., Razoux-Schultz, A., van Harten, T. W., van der Grond, J., van Buchem, M. A., Greenberg, S. M., Wermer, M.J.H., van Osch, M.J.P., van Walderveen, M.A.A., and van Rooden, S. (2022). Total white matter perivascular space volume: An early marker for Dutch-type Cerebral Amyloid Angiopathy. Conference abstract – oral presentation at International CAA conference 2022. Not part of this thesis.
5. Schipper, M. R., Razoux-Schultz, A., van Harten, T. W., van der Grond, J., van Buchem, M. A., Greenberg, S. M., Wermer, M.J.H., van Osch, M.J.P., van Walderveen, M.A.A., and van Rooden, S. (2023). Total white matter perivascular space volume: An early marker for Dutch-type Cerebral Amyloid Angiopathy. Conference abstract – poster presentation at ISMRM Benelux 2023. Not part of this thesis.
6. Schipper, M. R., Razoux-Schultz, A., van Harten, T. W., van der Grond, J., van Buchem, M. A., Greenberg, S. M., Wermer, M.J.H., van Osch, M.J.P., van Walderveen, M.A.A., and van Rooden, S. (2023). Total white matter perivascular space volume: An early marker for Dutch-type Cerebral Amyloid Angiopathy. Conference abstract – oral presentation at ISMRM 2023. Not part of this thesis.
7. Schipper, M.R., Vlegels, N., Voigt, S., van Harten, T.W., de Luca, A., Rasing, I., Biessels, G.J., van Osch, M.J.P., van Walderveen, M.A.A., and Wermer, M.J.H. (2023). The relationship between vascular reactivity and microstructural white matter integrity in Dutch-type Cerebral Amyloid Angiopathy. Conference abstract – poster presentation at ISMRM 2023. Not part of this thesis.
8. Schipper, M. R., Razoux-Schultz, A., van Harten, T. W., Wermer, M. J. H., van Osch, M. J. P., van Walderveen, M. A. A., and van Rooden, S. (2023). Does a manual determined threshold bias quantitative perivascular spaces measurements based on the Frangi vesselness filter? Meeting abstract – oral presentation at ICP 2023. Not part of this thesis.
9. Schipper, M. R., van der Plas, M. C., Koemans, E. A., Kaushik, K., van Dort, R., van der Zwet, R. G. J., Schriemer, S.E., van Harten, T.W., Voigt, S., Rasing, I., van Walderveen, M.A.A., van Osch, M.J.P., and Wermer, M.J.H. (2024). One-year follow-up of visually stimulated task-based fMRI in Dutch-type and sporadic Cerebral Amyloid Angiopathy. Meeting abstract – poster presentation at ISMRM Benelux 2024. Part of this thesis (Chapter 3).
- 10 van der Zwet, R. G. J., Koemans, E. A., Voigt, S., van Dort, R., Rasing, I., Kaushik, K., van Harten T. W., Schipper, M. R., Terwindt, G.M., van Osch, M.J.P., van Walderveen, M.A.A., van Etten, E.S., and Wermer, M.J.H. (2024). Sensitivity of the Boston Criteria Version 2.0 in Dutch-Type Hereditary Cerebral Amyloid Angiopathy. <https://doi.org/10.1177/17474930241239801>. Not part of this thesis.

11. [Schipper, M. R.*](#), Vlegels, N.* , van Harten, T. W., Rasing, I., Koemans, E. A., Voigt, S., de Luca, A., Kaushik, K., van Etten, E.S., van Zwet, E.W., Terwindt, G.M., Biessels, G.J., van Osch, M.J.P., van Walderveen, M.A.A., and Wermer, M.J.P. (2023). Microstructural white matter integrity in relation to vascular reactivity in Dutch-type hereditary cerebral amyloid angiopathy. <https://doi.org/10.1177/0271678X231200425>. Part of this thesis (Chapter 2)
 12. van Dort, R., Kaushik, K., Rasing, I. van der Zwet, R. G. J., [Schipper, M. R.](#), van der Grond, J., van Rooden, S., van Zwet, E.W., Terwindt, G.M., Middelkoop, H.A.M., Hart, E.P., van Osch, M.J.P., van Walderveen, M.A.A., and Wermer, M.J.H. (2024). Cognition in (pre)symptomatic Dutch-type hereditary and sporadic cerebral amyloid angiopathy. <https://doi.org/10.1002/alz.14171>. Not part of this thesis.
 13. Rasing, I., Vlegels, N., [Schipper, M. R.](#), Voigt, S., Koemans, E. A., Kaushik, K., van Dort, R., van Harten, T.W., de Luca, A., van Etten, E.S., van Zwet, E.W., van Buchem, M.A., Middelkoop, H.A.M., Biessels, G.J., Terwindt, G.M., van Osch, M.J.P., van Walderveen, M.A.A., and Wermer, M.J.H. (2024). Microstructural white matter damage on MRI is associated with disease severity in Dutch-type cerebral amyloid angiopathy. <https://doi.org/10.1177/0271678X241261771>. Not part of this thesis.
 14. Koemans, E. A., Rasing I., Voigt, S., van Harten, T. W., van der Zwet, R. G. J., Kaushik, K., [Schipper, M. R.](#), van der Weerd, N., van Zwet, E.W., van Etten, E.S., van Osch, M.J.P., Kuiperij, B., Verbeek, M.M., Terwindt, G.M., Greenberg, S.M., van Walderveen, M.A.A., and Wermer, M.J.H. (2024). Temporal ordering of biomarkers in Dutch-type hereditary cerebral amyloid angiopathy. <https://doi.org/10.1161/STROKEAHA.123.044688>. Not part of this thesis.
 15. [Schipper, M. R.](#), van Harten, T. W., Razoux-Schultz, A., Otto, A. L. D., Kaushik, K., Hirschler, L., Voigt, S., Rasing, I., Koemans, E.A., van Dort, R., van der Zwet, R.G.J., Schriemer, S.E., van Zwet, E.W., van der Grond, J., van Buchem, M.A., Greenberg, S.M., Wermer, M.J.H., van Osch, M.J.P., van Walderveen, M.A.A., and van Rooden, S. (2025). Quantification of Total White Matter Perivascular Space Volume: Follow-up in Dutch-type Hereditary Cerebral Amyloid Angiopathy. Meeting abstract – poster presentation at ISMRM 2025. Not part of this thesis.
 16. [Schipper, M. R.](#), van Harten, T. W., Razoux-Schultz, A., Kaushik, K., Hirschler, L., Voigt, S., Rasing, I., Koemans, E.A., van Dort, R., van der Zwet, R.G.J., Schriemer, S.E., van Zwet, E.W., van der Grond, J., van Buchem, M.A., Greenberg, S.M., Wermer, M.J.H., van Osch, M.J.P., van Walderveen, M.A.A., and van Rooden, S. (2025). Cross-sectional and longitudinal quantification of Total white matter perivascular space volume fraction in Dutch-type Cerebral Amyloid Angiopathy. <https://doi.org/10.1016/j.nicl.2025.103778>. Part of this thesis (Chapter 4).
 17. van der Plas, M. C., Koemans, E. A., [Schipper, M. R.](#), Voigt, S., Rasing, I., van der Zwet, R. G. J., Kaushik, K., van Dort, R., Schriemer, S.E., van Harten, T.W., van Zwet, E.W., van Etten, E.S., van Osch, M.J.P., Terwindt, G.M., van Walderveen, M.A.A., and Wermer, M.J.H. (2025). One-Year Radiologic Progression in Sporadic and Hereditary Cerebral Amyloid Angiopathy. <https://doi.org/10.12.12/WNL.000000000213546>. Not part of this thesis.
 18. de Bruin, O. F.* , [Schipper, M. R.*](#), Koemans, E. A., van Harten, T. W., Rasing, I., Kaushik, K., Hirschler, L., van Dort, R., van der Zwet, R.G.J., van Osch, M.J.P., van Walderveen, M.A.A., Wermer, M.J.H., and Voigt, S. (2025). Hyperintense lesions suspect for acute cerebral microbleeds on ultra-high field T1 weighted 7 Tesla MRI in patients with cerebral amyloid angiopathy. <https://doi.org/10.1016/j.ejrad.2025.112428>. Part of this thesis (Chapter 7)
 19. Schipper, M. R.* , van der Zwet, R. G. J.* , Goeman, J. J., van Harten, T. W., Koemans, E. A., Wermer, M. J. H., Walderveen, M.A.A., and van Osch, M.J.P. (in submission). Clustering of Cerebral Microbleeds in Cerebral Amyloid Angiopathy: A Flow Territory-Based Analysis. Part of this thesis (Chapter 6). Open code is available: R-code for statistical analysis is added as supplementary materials to the submission
 20. [Schipper, M. R.*](#), van Harten, T. W., Koemans, E. A., Goeman, J. J., Wermer, M. J. H., van Osch, M.J.P., and van Walderveen M.A.A. (in submission). Following the curvature: sulcal versus gyral cortical localization of cerebral microbleeds in Cerebral Amyloid Angiopathy. Part of this thesis (Chapter 5).
-

Completed courses and other training

Mandatory activities

Year	Title (+ description if needed)	Hours
2021	Leiden University Onboarding Program Inform & Connect (2 activities), LUMC	5
2021	Basic Methods and Reasoning in Biostatistics, LUMC	42
2021	Responsible Research (2 activities), LUMC	42

Scientific courses, workshops, and other training activities

Year	Title (+ description if needed)	Hours
2021	European Stroke Organization Conference (ESOC) conference, online	27
2022	International Society for Magnetic Resonance in Medicine (ISMRM) Benelux conference, Maastricht	9
2022	ISMRM conference, London	36
2022	Getting a @Handle on Big Neuroimaging Datasets, educational ISMRM 2022	4
2022	Microstructure: Relaxation, Magnetization Transfer & Susceptibility, educational ISMRM 2022	4
2022	The Circadian Clock & Its Effect on the Human Brain, educational ISMRM 2022	4
2022	MRI Artifacts & Corrections, educational ISMRM 2022	4
2022	Physics for Clinicians I and II, educational ISMRM 2022	4
2022	World Stroke Conference (WSC), Singapore	34
2022	International CAA conference (ICAA), Perth	25.5
2022-2025	Organization of multiple meeting (LUMC)	36
2023	ISMRM Benelux conference, Brussel	9
2023	School of MRI (organized by ESMRM-B)	18
2023	ISMRM conference, Toronto	36
2023	MR Physics I, educational ISMRM 2023	4
2023	MR Physics II, educational ISMRM 2023	4
2023	Key Contrast Mechanisms for Imaging Neuroinflammation, educational ISMRM 2023	4
2023	International CAA ECR conference, online	4
2023	Imaging Cerebral Physiology (ICP), Utrecht	27
2023	LUMC Neuroscience Symposium with presentation	6
2024	ISMRM Benelux, 's Hertogenbosch	11
2024	7T Philips User meeting with presentation, Amsterdam	16
2024	ICAA conference, München	25
2024	Leducq Consortium meeting, München	16
2025	Leducq Consortium meeting, Seattle	24
2025	ISMRM Benelux, Hilversum	9
2025	ISMRM conference, Honolulu	48

Transferable skills courses, workshops, and other training activities

Year	Title (+ description if needed)	Hours
2022	Teaching – Bachelor’s internship supervision	28
2022	Committee member of Teaching Radiology Exchange (T-REx)	52
2023	Teaching – gap-year internship supervision	28
2023	7 Tesla MRI tour assistance during Leiden International (Bio)Medical Student Conference (LIMSC)	5
2023	Teaching – one-day psychology BSc student supervision	4
2023	Organizing team (D-)CAA information day	58
2023	Teaching – supervising research assistant	40.5
2023	Careers outside academia, educational ISMRM 2023	4
2023	More success with less stress, Leiden University	28
2024	Career orientation - LAP day focused with meetings focused on career opportunities	8
2024	Teaching – Master’s internship supervision	16
2024	Teaching – gap-year internship supervision	16
2021-2025	Total of mandatory activities, scientific skills and transferable skills	825

Dissemination, acknowledgement, esteem and other relevant scientific activities table**Presentations at conferences and different type of meetings**

Year	Description	Linked to chapter(s)
2022	Poster presentation at ISMRM Benelux 2022, titled “Vascular reactivity measurements are insensitive to changes in visual stimulus presentation method.”	-
2022	Power pitch at World Stroke Conference 2022, titled “The relationship between vascular reactivity and microstructural white matter integrity in (pre-)symptomatic Dutch-type cerebral amyloid angiopathy.”	Chapter 2
2022	Poster presentation at International CAA conference 2022 “The relationship between vascular reactivity and microstructural white matter integrity in (pre-)symptomatic Dutch-type cerebral amyloid angiopathy.”	Chapter 2
2022	Oral presentation at International CAA conference 2022, titled “Total white matter perivascular space volume: An early marker for Dutch-type Cerebral Amyloid Angiopathy.”	Chapter 4
2023	Poster presentation at ISMRM Benelux 2023, titled “Total white matter perivascular space volume: An early marker for Dutch-type Cerebral Amyloid Angiopathy.”	Chapter 4
2023	Oral presentation at ISMRM 2023, titled “Total white matter perivascular space volume: An early marker for Dutch-type Cerebral Amyloid Angiopathy.”	Chapter 4

2023	Poster presentation at ISMRM 2023, titled “The relationship between vascular reactivity and microstructural white matter integrity in Dutch-type Cerebral Amyloid Angiopathy”	Chapter 2
2023	Oral presentation at ICP 2023, titled “Does a manual determined threshold bias quantitative perivascular spaces measurements based on the Frangi vesselness filter?”	Chapter 4
2023	Poster presentation at Neuroscience Symposium LUMC 2023, titled “Total white matter perivascular space volume: An early marker for Dutch-type Cerebral Amyloid Angiopathy.”	Chapter 4
2023	Oral presentation at CAA ECR online conference 2023, titled “Quantification of Total White Matter Perivascular Space Volume in Dutch-type Hereditary Cerebral Amyloid Angiopathy: Longitudinal Preliminary Data.”	Chapter 4
2024	Poster presentation at ISMRM Benelux 2024, titled “One-year follow-up of visually stimulated task-based fMRI in Dutch-type and sporadic Cerebral Amyloid Angiopathy.”	Chapter 3
2025	Poster presentation at ISMRM 2025, titled “Quantification of Total White Matter Perivascular Space Volume: Follow-up in Dutch-type Hereditary Cerebral Amyloid Angiopathy.”	Chapter 4

Awards

Year	Description	Linked to chapter(s)
2022	Best oral presentation award at International CAA conference 2022	Chapter 4
2023	Best poster presentation award at Neuroscience Symposium LUMC 2023	Chapter 4
2023	Best oral presentation award at CAA ECR online conference 2023	Chapter 4

Grant applications

Year	Description	Linked to chapter(s)
2025	Alzheimer Nederland travel grant application: Small Grants - Financial support for conference attendance	Chapter 4

Curriculum Vitae

Manon Roxanne Schipper was born on March 11th, 1997, in Hardinxveld-Giessendam, the Netherlands. After graduating from high school in 2015, she started the bachelor in Psychology with the Honours track ‘science and society’ at the social faculty of Leiden University. She completed her bachelor’s internship under supervision of Dr. Laura Steenbergen at Leiden University and wrote a bachelor’s thesis titled “The relationship between transcutaneous vagal nerve stimulation, decision-making, happiness, and heart rate variability”. In 2018 she started the Cognitive Neuroscience track of the Psychology research master at Leiden University. She completed her master’s internship at the Amsterdam UMC under supervision of Dr. Marieke van der Pluijm and Dr. Anne Trutti and wrote a master’s thesis titled “The use of neuromelanin-sensitive MRI to assess neuromelanin differences in the substantia nigra between treatment resistant and respondent patients with schizophrenia”. She graduated from the master’s program in 2020. During her studies, she worked as a care assistant for demented elderly at a restricted psychogeriatric department of caring home ‘Rijn en Vliet’ in Leiden.

In May 2021 Manon started her PhD research at the C.J. Gorter MRI Center of the Radiology department at the Leiden University Medical Center (LUMC) under the supervision of Dr. Marianne van Walderveen, Prof. Dr. Marieke Wermer, and Prof. dr. ir. Matthias van Osch. She worked in a multidisciplinary team; working closely with the LUMC Neurology department and researchers in both Boston, USA, and Perth, Australia. The focus of her PhD was on advanced MR image analyses in sporadic and Dutch-type Cerebral Amyloid Angiopathy.

Manon is currently continuing her research as a post-doctoral researcher at the C.J. Gorter MRI Center of the Radiology department at the LUMC, further focusing on advanced MR image analyses in sporadic and Dutch-type Cerebral Amyloid Angiopathy.

Acknowledgements

Door het beperkte aantal woorden toegewezen aan het dankwoord, is het onmogelijk iedereen te bedanken. Van collega's tot coauteurs, van vrienden tot familie; naast jullie steun en hulp, hebben jullie mijn PhD een mooie en bijzondere tijd gemaakt. Bedankt voor jullie steun, geduld en alle gezelligheid.

Zonder alle deelnemers waren de ontwikkelingen op het gebied van CAA en mijn PhD niet mogelijk geweest. Jullie hart voor het welzijn van lotgenoten is een inspiratie.

Thijs, Marianne en Marieke, bedankt voor de kans die jullie mij hebben gegeven. Thijs, jouw voortdurende betrokkenheid waardeer ik enorm. Jouw pragmatische en ondernemende houding heeft mij enorm geholpen en neem ik de rest van mijn carrière met mij mee. Marianne, bedankt voor jouw kritische blik op zowel de wetenschappelijke inhoud als op de manier waarop ik mezelf neerzet. Dit hield mij scherp en liet mij als onderzoeker, maar ook als persoon verder ontwikkelen. Marieke, bedankt voor je voortdurende vertrouwen en optimistische houding. Wanneer ik door de bomen het bos niet meer zag, bleef je altijd hoopvol en motiverend.

Lieve paranimfen, Roos en Sanne, woorden schieten te kort om jullie te bedanken voor jullie steun en vriendschap. Ik had mijn promotie niet anders willen afsluiten dan met jullie aan mijn zijde.

Thijs_(vH), dankjewel voor jouw mentor- en vriendschap. Jouw geduld en hulp hebben een groter aandeel gehad aan mijn rol als onderzoeker dan je waarschijnlijk denkt.

Reinier, Roos en Sanne, het was me een waar genoegen om mijn PhD tegelijkertijd met jullie te mogen doen, bedankt voor alles! Sabine, Kanishk, Emma en Ingeborg, bedankt voor jullie 'opvoeding' binnen het CAA-onderzoek. Yasmine, bedankt voor al jouw inzet, wat fijn dat je nu onderdeel bent van het 'vaste' onderzoeksteam. Ellen_(S-H), dankjewel voor de fijne en veilige plek die jij vanaf dag één al wist te creëren. Bedankt dat ik onderdeel mocht zijn van de neurologie-groep, en de altijd gezellige borrels en congressen.

Dear Gorter center, or rather, Gorter family, you have opened my eyes for how warm, inclusive, and supportive the academic world can be. Dear all, I cannot thank you enough for the amazing conferences, dinners, and much more.

Dear (former) officemates; Danielle_(T), Ernst, Fieke, Jasmin, Lennart, Lydiane, Nadieh, and Thijs_(vH), thank you for the deep conversations, support, and advice.

Dear MR-physiology; Balazs, Barbara, Danielle_(vD), Ellen_(v), Emiel, Eva, Helena, Ingmar, Jasmin, Leonie, Lena, Lydiane, Maaïke, Madda, Martijn, Max, Navid, Nina, Simone, Thijs_(vH), and Yiming, thank you for being such a warm and inclusive group, I am grateful for being and having been part of this group and for the friendships that have formed.

Emiel, Fieke, Helena, Madda, Nina, and Yiming, thank you for the Hawaiian post-conference trip. From star gazing at the Mauna Kea to our very adventurous trip to see that one manta ray, I treasure these memories.

Danielle^(VD), met jou mocht ik de ISMRM-wereld ontdekken, ik waardeer de band die we hier hebben opgebouwd. Danielle^(T), zo fijn om jou als kantoorgenoot te hebben, dankjewel voor de goede gesprekken en de ruimte om te klagen. Ernst, dankjewel voor je luisterend oor en alle gezelligheid. Ik hoop nog veel koffiemomenten te delen, ook nu we geen kantoor meer delen. Fieke, dankjewel voor jouw mooie vriendschap en alle diepe gesprekken, ik hoop dat onze paden blijven kruisen. Jasmin, thank you for the countless talks and laughs, it never is a dull moment with you. Lena, dankjewel voor de veilige omgeving die jij altijd weet te creëren. Ik kijk op naar jou als onderzoeker en persoon. Lydiane, thank you for our scientific discussions, but also for your endless hospitality. Martijn, dankjewel voor jouw gezelligheid en nuchterheid. Ik waardeer onze gesprekken, je hulp en je mening. Navid, even though we only got close near the end of my PhD, I am happy that we did. Thank you for your friendship. Vesna, jouw vriendschap kwam uit een onverwachte hoek, maar wat fijn om jou als vriendin te hebben.

Lieve vrienden, bedankt voor jullie constante liefde, steun en afleiding. In het bijzonder, Aletta, Angela en Sarah, dank voor het ondersteunen van mijn persoonlijke groei tijdens dit proces en jullie geduld en respect wanneer het soms minder ging. Dewi, dankjewel voor jouw warme vriendschap en diepe interesse, je begrijpt mij op een heel bijzonder niveau. Mirre, dankjewel voor je voortdurende goede zorgen, betrokkenheid en dat ik altijd bij je terecht kan.

Lieve pap, mam, Laura, Elise en Jaap, bedankt voor jullie vertrouwen in mijn keuzes en geduld wanneer ik weinig tijd en een vol hoofd had. Elise, dankjewel voor de prachtige omslag van dit proefschrift.

Lieve Merlin, dankjewel voor jouw liefde en steun, jouw geduld en begrip wanneer alles te veel was en jouw blijdschap wanneer er iets te vieren was. Je weet mij uit te balanceren wanneer dat nodig is en maakt mijn leven zoveel mooier en rijker.