



Universiteit
Leiden

The Netherlands

Healthy ageing in older adults: an exploration of interactions between lifestyle, immune, metabolic, and gut microbial health

Morwani Mangnani, J.

Citation

Morwani Mangnani, J. (2026, April 1). *Healthy ageing in older adults: an exploration of interactions between lifestyle, immune, metabolic, and gut microbial health*. Retrieved from <https://hdl.handle.net/1887/4300477>

Version: Publisher's Version

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/4300477>

Note: To cite this publication please use the final published version (if applicable).



CHAPTER 2

Design of the VOILA intervention study: A 12-week nutrition and resistance exercise intervention in metabolic or mobility compromised Dutch older adults and the response on immune-metabolic, gut and muscle health parameters

CS Kramer^{1*}, A Monsegue^{2*}, J Morwani-Mangnani^{3*}, P Grootswagers¹, M. Beekman³, PE Slagboom^{3#}, LB Verdijk^{2#}, L CPGM de Groot^{1#}

¹ Wageningen University & Research, Wageningen Campus, Agrotechnology and Food Sciences Group, Division of Human Nutrition and Health, PO Box 17, 6700 AA Wageningen, The Netherlands

² Maastricht University Medical Center+, Department of Human Biology, NUTRIM Institute of nutrition and translational research in metabolism, PO Box 616, 6200 MD Maastricht, The Netherlands

³ Leiden University Medical Centre, Department of Biomedical Data Sciences, Section of Molecular Epidemiology, Einthovenweg 20, 2333 ZC Leiden, The Netherlands

* Shared first authorship

Shared last authorship

[^] Corresponding author

Mech Ageing Dev. 2024 Dec;222:112002. doi: 10.1016/j.mad.2024.112002. Epub 2024 Oct 28. PMID: 39490538.

Supplementary files can be found online.

ABSTRACT

Background: Exercise and nutrition interventions can slow ageing-induced decline in physiology. However, effects are heterogeneous and usually studied separately per outcome domain. In the VOILA study, we simultaneously study various health outcomes relevant for older adults and the inter-individual heterogeneity in response to a lifestyle intervention.

Methods: VOILA is a 12-week lifestyle intervention in 3 groups of older adults (≥ 60 years), with compromised mobility (n=50), compromised metabolic health (n=50), or recovering from total knee replacement (TKR, n=70, of which 20 randomized to standard care only). The intervention includes high-intensity resistance exercise training thrice weekly, nutritional counselling, and nutritional supplements every morning and evening (including 20-25g whey protein and (evening only) 5.5g Biotis™ GOS). We measure immune-metabolic, gut health, muscle mass and physical functioning at baseline and after completion of the intervention/standard care. An additional reference group of healthy older adults (n=50) will undergo baseline measurements only.

Discussion: Improvements in various physiological systems are expected, but with differences between groups/individuals. This study will provide insights into how the physiological state of older adults influences the extent of lifestyle-induced health improvements to create better tailored interventions to attenuate biological ageing and improve the health span of subgroups and individuals.

Keywords

Biological age, nutrition, gerontology, intervention, resistance exercise, metabolic health, gut health

LIST OF ABBREVIATIONS

1RM: 1-Repetition Maximum
1H NMR: Proton Nuclear Magnetic Resonance
25(OH)D: 25-hydroxyvitamin D
5CST: 5-times Chair Stand Test
6MWT: 6-Minute Walking Test
ALM: Appendicular Lean Mass
DEXA: Dual Energy X-ray Absorptiometry
DNA: Deoxyribo Nucleic Acid
FRS: Framingham Risk Score
GLC: glucose
GlycA: Glycoprotein Acetyls
GOS: GalactoOligoSaccharides
GSRS: Gastrointestinal Symptom Rating Scale
HbA1c: Haemoglobin A1C
LLS: Leiden Longevity Study
MMSE: Mini Mental-State Examination
MEC: Medical Ethics Committee
QoL: Quality of Life
SCFA: Short Chain Fatty Acids
SD: Standard Deviation
SF-36: Short Form (36) Health Survey
SF-SarQoL: Short Form Sarcopenia and Quality of life
PI: Principal Investigator
SPPB: Short Physical Performance Battery
SQUASH: Short QUestionnaire to ASsess Health enhancing physical activity
TKR: Total knee replacement
TUG: Timed Up-and-Go test
VLDL-D: Very Low Density Lipoprotein Diameter
VOILA: Vitality Oriented Innovations for the Lifecourse of the Ageing society
WOMAC: Western Ontario and McMaster Universities Arthritis Index

INTRODUCTION

Life expectancy has increased twofold to threefold in the last 200 years because of better living conditions, nutrition, hygiene, and medical care (1-3). Globally, population ageing progresses as the number of people over 65 years is projected to double between 2019 and 2050, and people aged 80 years and older are projected to triple to 426 million (4). As age is the most prominent risk factor for common diseases and multi-morbidity in humans, general health declines as we get older (5), leading to loss of functional capacity in physical, cognitive, and social domains. Yet, since individuals of the same chronological age (age in units of time) display a large heterogeneity in health status, it is the biological age (i.e. age measured as a biological process or deviation from chronological age) of individuals that influences the risk of disease more than their chronological age does (3). As such, this heterogeneity in biological age is expressed as different susceptibilities to morbidity and mortality. In relation to this, adults of all ages favour a long lifespan only if accompanied by good health or vitality (6, 7). Therefore, in our ageing society, it is important to extend health span by delaying or partially reversing the ageing process. To do this we must first record the biological age for early recognition of those individuals that are at risk of faster functional decline, to then attempt to delay multiple aspects underlying the ageing process simultaneously.

During ageing, multiple physiological systems are dysregulated (8). Especially in animal studies and model systems, a range of molecular hallmark mechanisms have been identified that drive the ageing process across species (9-11). Targeting these hallmarks illustrated that health and lifespan can be extended in animal models (12). In humans, the same hallmarks are expected to underly ageing, leading to physiological deterioration and morbidity (3). Several mechanisms of ageing have already been identified to underlie multiple age-related diseases (13) and multimorbidity simultaneously (14). Consequently, prevention or delay of age-related functional decline and disease is likely possible either by lifestyle or pharmacological intervention especially when targeting these ageing processes. Experts agreed that interventions targeting hallmarks of ageing “should halt further detrimental aspects of ageing and preferably improve phenotypes associated with ageing” (11). In line with this, the “Vitality Oriented Innovations for the Lifecourse of the Ageing society” (VOILA) intervention study we present here, focuses on the improvement of phenotypical markers of three physiological systems in humans, hypothesized to be important during ageing – muscle mass and physical functioning, immune-metabolic health, and gut health - via a multimodal intervention.

The VOILA intervention we describe here focuses on resistance exercise (RE) and a healthy diet, including a high-protein component and galacto-oligosaccharides (GOS), for their combined impact on the ageing-related domains of muscle health, immune-metabolic health, and gut health. Regarding muscle health, muscle protein turnover decreases (15) and skeletal muscle strength, power, mass and quality decline with age (16) which are in turn associated with compromised mobility (17, 18) and increased chronic disease risk (19). Resistance

exercise is recommended as a countermeasure (19-23), showing increases in muscle mass and strength in healthy older adults (24-26) as well as strength and physical performance in pre-frail and frail older adults (27, 28). Remarkably, all older adults show improvements on at least one muscle modality in response to RE, although inter-individual variation is substantial (29).

RE and its aforementioned effects on muscle health in turn positively affect many physiological and metabolic ageing-related processes. For example, age-related decline of muscle mass, quality and physical activity are thought to be the main drivers of insulin resistance (30-32). Whether increasing activity levels can fully restore insulin sensitivity is not yet clear (33-35), but progressive RE has been shown to improve glycaemic control in overweight prediabetic older adults (36, 37), lower the risk of developing Diabetes Mellitus type 2 (38), and reduce mortality risk in diabetic patients (39). In addition, RE has been shown to reduce cardiovascular risk (40, 41), by improving the Framingham Risk Score (FRS) (42), decreasing blood pressure (42, 43) and improving lipid profiles (44).

Other ageing-related health benefits of RE are thought to be related to the release of myokines and other molecules by skeletal muscle upon contraction (45). Moreover, exercise reduces multiple inflammatory cytokines that are important in low-grade inflammation present with ageing-related impairments such as insulin resistance, cardiovascular disease, and atherosclerosis (46-48). Due to its extensive positive health effects throughout the body, RE is a cornerstone of interventions in older populations.

Next to exercise, adequate protein intake is essential for muscle mass maintenance as well as to support muscle hypertrophy. Protein intake increases plasma amino acid concentrations, providing the necessary building blocks for muscle and activating signalling pathways that stimulate muscle protein synthesis (49). Among older adults, relative anabolic resistance to dietary protein is prevalent (50, 51). The PROT-AGE study group proposed to increase the protein intake recommendations for adults above 65 years of age who are exercising to at least 1.2 g/kg bodyweight/d (52), and the European Society for Clinical Nutrition and Metabolism (ESPEN) guidelines advice a protein intake up to 1.5 g/kg bodyweight/d for older adults who are malnourished and/or those with acute or chronic illness (53). Currently, Dutch older adults have a lower protein intake, with means for community-dwelling, frail and institutionalized older adults ranging between 0.8-1.0 g/kg bodyweight per day (54). It remains unknown whether increasing dietary protein intake alone may counteract anabolic resistance (55-59). It might not be ageing itself, but accompanied chronic inflammation and especially reduced physical activity levels that cause anabolic resistance (60). Therefore, increasing dietary protein intake combined with RE is more likely to result in increased muscle mass and strength in older adults (61-65).

In addition to muscle and immune-metabolic health, gut health plays an important role in ageing, especially through the influence of gut microbiota on immunity. Gut microbial composition is influenced by genetics and non-genetic factors such as geographic location,

exercise, nutrition, disease, and medication (66, 67). Certain gut microbiota profiles are associated with healthy ageing (68) and diet can alter the gut microbiome in the long term (69). Increased fibre and prebiotics intake improve microbial composition and short-chain fatty acid (SCFA) production (69-71). One prebiotic of interest is GOS: carbohydrates produced from lactose, with various lengths and linkages (72). GOS enter the large intestine intact, where they serve as substrates for bacterial fermentation, producing SCFAs (73, 74). SCFAs are absorbed via the portal vein (75) and play a role in many physiological functions, ranging from energy balance and metabolism to the modulation of adipose tissue, immunity, liver tissue and skeletal muscle function (69, 76). Double-blind, placebo-controlled studies with various doses of GOS show increased gut *Bifidobacteria* count in healthy older adults (77-79), overweight adults (80) and patients with Diabetes Mellitus type 2 (81). Several specific strains of *Bifidobacteria* have been associated with positive immune and disease outcomes, potentially via the production of SCFAs, other metabolites or by improving gut barrier function (82, 83). In support, lower prevalence of *Bifidobacteria* has been associated with older age (84) and a number of diseases (82, 83). However, the health implications of increasing *Bifidobacterial* presence are less well-established, with some (77, 78, 80), but not all (77, 81, 85, 86) studies showing improved immune-metabolic profiles. Additionally, the potential immunomodulation via gut microbial alteration by GOS or fibre in general (87) might also work synergistically with protein and exercise, as specific gut bacteria, such as *Bifidobacteria*, may have anti-inflammatory effects (88) and inflammation could be an underlying factor for anabolic resistance (88, 89).

Beyond the health outcomes of specific physiological systems, overall markers of biological age have been developed in the past decade to be predictive of health decline during ageing. However, no gold standard exists yet for biological age (90). A novel biomarker indicating immune and metabolic health, the MetaboHealth score, was recently generated from blood-based metabolomics data in 44,000 people. The score includes lipids, lipoprotein particles, fatty acids, lactate, histidine, branched chain amino acids, phenylalanine, acetoacetate, albumin and glycoprotein acetyls and predicts age-related disease, frailty and mortality in prospective studies (91-93). A previous combined lifestyle intervention in older adults demonstrated the beneficial metabolomic response (94, 95), but it remains to be established to what extent the MetaboHealth score can be used as a response marker in interventions.

Considering the above, exercise and dietary components have been shown to improve various health parameters relevant in older age, but effects on domains of ageing in different groups of older adults needs to be studied. It is important to note that especially in older adults, inter-individual variability in health status and physiology is present between individuals (44, 96-98). Understanding this variability is essential to better predict intervention responses. The VOILA intervention is therefore applied to different groups of mobility and metabolically compromised older adults (≥ 60 years of age). We will select these groups primarily on the basis of walking ability, recent total knee replacement (TKR) and MetaboHealth score to

include groups of older individuals who are at risk of fast biological ageing for different reasons, who would likely benefit from lifestyle interventions.

The intervention consists of optimized combined resistance exercise and nutritional components to improve outcomes in three major ageing related domains: muscle, immune-metabolic and gut health. The goal is to study the effect sizes on these ageing-related health domains in mobility and metabolically at-risk groups as well as the response heterogeneity to this intervention and the potential underlying actors.

METHODS

Aim

The VOILA intervention study aims to improve different health domains underlying biological ageing by a combined nutrition and RE intervention in three groups of community-dwelling older adults with varying degrees of compromised mobility or metabolism (≥ 60 years of age) and to measure the extent to which these groups respond. Three major ageing related domains will be studied: immune-metabolic health, gut-microbiome composition, muscle mass and physical functioning. Additionally, we aim to explore the factors that are responsible for the heterogeneity in responsiveness in these groups, as well as on an individual level. An additional aim is to investigate the differences between baseline variables of the health-compromised groups compared to the reference group of active “healthy” older adults and assess the extent to which the VOILA intervention can diminish this difference. The objectives are as follows:

- 1. Primary Objective:** To quantify the effect of a state-of-the-art multimodal intervention in individuals aged ≥ 60 , segmented by metabolic health status or mobility impairment on 1) muscle mass (appendicular lean mass (ALM)) and muscle function (5-times chair stand test (5CST)); 2) the gut microbiome composition (*Bifodobacteria*); and 3) immune-metabolic health (fasting blood glucose (GLC), glycoprotein acetyls (GlycA) and very low-density lipoprotein diameter (VLDL-D), within each segment.
- 2. Secondary Objective:** To determine to what extent each intervention segment compares to the active reference group, regarding muscle mass and muscle function, gut microbiome, and immune-metabolic health (using the same main parameters as the primary objective).
- 3. Exploratory Objectives:** To obtain insight in the heterogeneity of responsiveness, including the factors that may explain such heterogeneity, by comparing the effect on all primary outcomes (muscle/functional, gut microbiome and immune-metabolic markers) both in individuals within the different segments and the complete sample. Additionally, in the future we want to include comparisons of genetic and epigenetic variation with existing datasets of other lifestyle intervention studies.

Additional to these objectives, a second set of objectives includes the same 3 objectives but with the secondary outcome parameters: short physical performance battery (SPPB), timed

up-and-go test (TUG), 6-minute walking test (6MWT), 1-repetition maximums (1RM) as measured on exercise machines (muscle function), Dual X-ray Absorptiometry (DEXA) whole body and regional lean mass and fat mass (body composition), Glycated Hemoglobin (HbA1C), insulin, blood pressure (immune-metabolic health); and muscle cross-sectional area for TKR group.

Design and setting

This exploratory study is a 12-week combined exercise and nutritional intervention study (Figure 1) in the Netherlands (ClinicalTrials.gov Identifier: NCT05354310. Registered on April 29th, 2022). The intervention groups are: a metabolically compromised group, a compromised mobility group, and a specific patient group who recently received TKR. Additionally, there is a reference group of active, healthiest older adults. A separate control group receiving usual care will be included for the TKR group.

All groups will be measured at both baseline and at 12 weeks, except the active reference group, who will only be measured at baseline. The reference group is only used as a reference representing the most active and healthy older adults in our society, i.e. those ageing relatively successfully. The TKR control group is necessary because several outcome improvements are expected regardless of the intervention, due to the nature of recovery after surgery.

The study will take place at three main study sites at universities and university hospitals of Leiden (LUMC), Maastricht (MUMC+) and Wageningen (WUR). RE training will take place at the study centers, local gyms, and health centres under study supervision. The active reference group and metabolically compromised group will be recruited mainly from the vicinity of Leiden, the mobility compromised group near Wageningen, and the TKR patients from around Maastricht (the province of Limburg). The measurements and intervention activities will be organised to suit the local facilities available, but will follow standardised protocols created through collaboration of the three centres. All study procedures have been approved by the medical ethics committee (MEC) of LUMC (NL76879.058.21).

We aim to recruit fifty individuals for each of the three intervention groups and active reference group, and twenty for the TKR control group. Men and women will be included in approximately equal numbers, aiming for no more than 20% difference between the numbers of men and women within each group.

Study Population

Participants will be selected for the groups on the basis of physical functioning-related aspects (mobility compromised and TKR groups), metabolic profiles (metabolically compromised group) and physical activity and metabolic health (active reference group). Other criteria were chosen for study safety and feasibility. Only the participants for the TKR

groups will be randomized into the intervention and control group with a ratio of 5:2. TABLE 1 shows all inclusion and exclusion criteria.

Sample size

The appropriate sample size was calculated to pick up within-group changes for each intervention group, for each of the primary parameters. GPower 3.1.9.4 was used to estimate the number of participants needed for a difference between two dependent means, with a power of 0.80 and a one-sided alpha of 0.05. For all calculations mean and standard deviation (SD) of change were used when possible. If these were not available, pre and post means and SDs and a correlation coefficient from other studies were used. A dropout rate of 15% is expected and taken into account in the final sample size (103).

Based on interventions with protein and vitamin D, RE, and physiotherapy treatments respectively, physical function as measured by 5CST would require n=22 in the mobility compromised group (104), n=12 in the metabolically compromised group (105), and n=35 in the total knee replacement (TKR) group (106, 107).

Muscle mass, as measured by ALM, is estimated to require n=25 in the mobility compromised group (108), n=24 in the metabolically compromised group (109, 110), and n=55 in the TKR group (111). This is based on interventions with protein, protein and RE, or elastic-band resistance training respectively.

TABLE 1. VOILA general inclusion and exclusion criteria, and additional criteria per group. *Not applicable to the active reference group. BMI: Body mass index; COPD: chronic obstructive pulmonary disorder; LLS: Leiden longevity study; SQUASH: Short Questionnaire to Assess Health-enhancing physical activity

Inclusion criteria	Exclusion criteria
General, all groups	General, all groups
Age ≥ 60 years	Already using prebiotic fibres*
BMI 18.5-35.0 kg/m ²	Use of laxatives that interfere with outcomes, as determined by study medical doctor
Able to give written informed consent	Not willing to stop using dietary supplements in high doses (high doses: 0.10*Upper Level or more)*
Community-dwelling	Following a structured, intense exercise programme (currently or in the last year)*
MMSE of ≥ 24 points	Allergic, intolerant or hypersensitive to milk/lactose (self-reported)
Able to follow the study protocol	Dietary restrictions on milk/lactose/prebiotics/vitamin D/calcium consumption
Active reference group	
Meeting the Dutch physical activity guidelines (i.e. partaking in at least moderate exercise for at least 30 minutes 5 days per week) as determined by the SQUASH questionnaire (100).	Abnormal hepatic or renal laboratory parameters (estimated glomerular filtration rate < 30 ml/min/1.73 m ² (screening) or contra-indication by treating medical practitioner (data from hospital) (52)
	Diagnosis of disorders/diseases in which a high protein intake can be harmful, such as renal impairment or failure, liver disease, or diabetes

MetaboHealth (91) Score below the threshold for being considered metabolically-compromised (i.e. the bottom half scoring participants ≥ 60 years old from the LLS cohort).

Mobility compromised group

People able to walk but needing walking aids to walk anywhere outside their house and/or attaining a five-times chair stand test time ≥ 15 seconds (16, 101, 102).

Metabolically compromised group

MetaboHealth score in the top half scoring participants ≥ 60 years old either from the LLS cohort or the general population.

associated with nephropathy (treating medical practitioner has the decisive voice).

Diseases, conditions or disorders which may affect the ability to follow the study protocol and which cannot be overcome with help of a caregiver:

Uncontrolled hypertension (blood pressure above 160/100 mmHg)

Unstable angina

Uncontrolled dysrhythmias

Recent history of congestive heart failure that has not been evaluated and effectively treated

Severe stenotic or regurgitant valvular disease

Hypertrophic cardiomyopathy (all above: (99))

COPD (stage II/III/VI),

Peripheral artery disease (or: Fontaine III or IV),

Previously known cardiac abnormalities for which high-intensity exercise is contra-indicated (based on exercise ECG as determined by cardiologist)

Surgical intervention in past 12 weeks for which high-intensity exercise is contra-indicated (except for TKR group)

Unintentional weight loss of ≥ 3 kg in the last 6 months

Active cancer / chemotherapy or radiotherapy / end-stage malignancy in the last year

Uncontrolled diabetes mellitus

Cognitive impairments (MMSE < 24)

Recent (< 3 months prior to intervention) immobilisation for > 1 week (except for the TKR groups)

Psychiatric or behavioural problems

Use of unstable thyroid medication (i.e., changes in the last 3 months), immunosuppressive drugs (e.g. prednisone, methotrexate, biologicals (TNF-alpha antagonists))

Crohn's disease or ulcerative colitis, ileostomy or colonostomy

Any other medical condition that may interfere with the safety of the participants during training or any condition that interferes with assessment of the outcome parameters, in the investigators' judgement

Current participation in other scientific research that conflicts with this study (e.g. intervention studies or weight management studies)

Not signed up to a general practitioner

No permission to request information from the general practitioner/ treating specialist(s) about medical history, medication use.

Specific for TKR groups

Patients who have recently undergone elective knee replacement surgery (baseline testing starts ~ 6 weeks after surgery)

Specific for TKR groups

Osteoarthritis of the knee secondary to septic arthritis, osteonecrosis, fracture, osteochondritis dissecans, or malignant processes

Collagen disorders, e.g. Marfan and Ehler-Danlos syndrome

Fecal bifidobacterial count is estimated to require $n=5$ participants (77) in the mobility compromised group, and $n=15$ in the metabolically compromised group (80), based on studies with GOS.

Immune-metabolic outcomes in the mobility compromised group are estimated to require $n=23$ for GlycA and $n=38$ for VLDL-D based on a study in which participants increased their physical activity level (112). The estimated sample size for the metabolically compromised group is $n=56$ based on GLC, $n=31$ based on GlycA and $n=54$ for VLDL-D readings respectively, based on a study in which energy intake was reduced and energy expenditure was increased through physical activity (94).

Taking the above into account we chose to aim for a sample size of $n=50$ for every intervention group and the reference group. All calculated sample sizes are well below $n=50$, except for the estimation of ALM in TKR, and some metabolic markers in the metabolically compromised group. However, the studies used for those calculations applied mild interventions. Hence, we expect the intervention effects in the VOILA study to be larger due to the application of a high intensity exercise program in combination with a nutritional intervention. The choice to aim for a larger sample size than needed for most of the primary outcome measures will facilitate the exploration of heterogeneity in the adaptive responses to the intervention between individuals, and the different biological factors that may explain such heterogeneity.

Additional calculations were performed taking into account expected changes in the TKR control group (i.e., standard care) (111); $n=13$ (with dropout) patients should be included in the control group to detect differential changes in physical functioning, whereas $n=25$ patients should be included in the control group to detect differential changes in ALM. However, we expect the increase in lean mass to be greater with the proposed combined RE and nutritional intervention than in the study by Liao et al (111). Therefore, we aim to include $n=20$ patients in the TKR control group, receiving standard care only.

Participant timeline

Once identified as eligible (see below), all study participants will undergo baseline measurements at the university/university medical centre, at the training location/gym and at home up to 2 weeks before starting the intervention period. These measurements will be divided over a total of 2-4 visits depending on the logistics of each centre.

Within one week of finishing baseline measurements, the 12-week nutrition and exercise intervention will begin for the three intervention groups. These groups will undergo supervised resistance exercise training, 3 times per week, and consume nutrition supplements twice per day. Additionally, participants will choose 2 nutrition goals to work on. During this 12-week period, the TKR control group will not receive any intervention, and only receive standard care from their own health-care providers. At the end of the 12-week period, the baseline measurements will be repeated, except for the baseline questionnaire and

familiarisation session. Materials for the at-home measurements (i.e. activity and dietary measurements, questionnaires, and fecal sample) will be provided at the training locations or delivered to the participant's home (TKR control group), to be measured during the final week of the 12-week period. The final measurements will take place in the week following the end of the 12-week period, and will be spread over 1-3 measurement days, depending on the logistics of the study centre.

Study procedures and data collection

Recruitment, consent and inclusion

Recruitment strategies of the study will differ per study group. The metabolically compromised and active reference groups will be recruited firstly from the Leiden Longevity Study (LLS) (113). If they consent, their previously collected immune-metabolic parameters will be used to determine if they are suitable for the metabolically compromised group, or active reference group. Additionally, participants for these groups will be recruited from the general public via advertisements. The mobility compromised intervention group will be recruited via the university research volunteer database, as well as through local advertisements. The patients undergoing elective total knee arthroplasty will be recruited via the outpatient orthopaedic clinic at Maastricht University Medical Center (MUMC+) or other hospitals in the surrounding areas, physiotherapy practices, and advertisements.

Potential participants will contact or be contacted by the researchers. The full written study information and an example informed consent (IC) form will be sent to those who are interested (group specific), and they will be given at least 1 week to consider participation. They will be invited for either an on-site information session or a telephone call during which the entire experimental trial will be explained and questions will be answered. Potential participants who want to join will then sign the IC form, before any measurements can be taken (Supplement 1).

Screening

After both the study participant and the local researcher sign the IC form, participants will be assigned a unique study number in the electronic participant database and continue to the screening. The screening questionnaire created for this study contains questions on living situation, age, mobility status, disease details and comorbidities, medication (type, frequency, duration, dose), recent vaccination status and types, Corona Virus Disease-19 history, involvement in other research, weight loss, allergies and diets, nutritional supplement and prebiotic use, following exercise programmes and having a general practitioner (Dutch questionnaire available upon request). FRAIL questions (114), and Mini Nutritional Assessment short version questions are also included(115). Additionally, participants will complete the Short QUestionnaire to ASsess Health-enhancing physical activity (SQUASH), a validated questionnaire including duration, intensity, and type of physical activities, to

assess adherence to the Dutch guidelines for physical activity (100). The MMSE will be applied to exclude participants with cognitive problems, using a cut-off of ≥ 24 (116).

For participant safety, kidney function will be assessed by calculating the estimated glomerular filtration rate (eGFR) using the Chronic Kidney Disease Epidemiology Collaboration prediction equation. This calculation is based on age, sex, and serum creatinine levels (117). Therefore, a non-fasted blood sample will be drawn via venepuncture and levels determined locally within 24 hours. Participants with an eGFR below 30ml/min/m² will be excluded.

The 5CST (mobility compromised group only) will be carried out in the way described in the SPPB (118). Height, weight and blood pressure will be carried out as described in “Physical functioning and muscle strength”.

Study measurement procedures

All study sites will follow the same standard operating procedures and biweekly meetings are held for alignment. For an overview of the measurements, see TABLE 2.

TABLE 2. Overview of VOILA outcomes and collection methods per timepoint. ^aReference group does not have any measurements after baseline; ^bMobility compromised group only; ^cTotal knee replacement groups only; ^dMetabolically compromised and reference group only; ¹Measured in week 12. 1H NMR: Proton Nuclear Magnetic Resonance; 25(OH)D: 25-hydroxyvitamin D; 5CST: 5-times Chair Stand Test; 6MWT: 6-Minute Walking Test; ALM: Appendicular Lean Mass; CT: Computed Tomography; DEXA: Dual X-ray Absorptiometry; DNA: DeoxyriboNucleic Acid; GRS: Gastrointestinal Symptom Rating Scale; MMSE: Mini Mental-State Examination; RNA: RiboNucleic Acid; SQUASH: Short QUestionnaire to ASsess Health enhancing physical activity; SF-36: Short Form (36) Health Survey; SF-SarQol: Short Form; Sarcopenia and Quality of life; SPPB: Short Physical Performance Battery; TUG: Timed Up-and-Go test; WOMAC: Western Ontario and McMaster Universities Arthritis Index

Outcome	Collection method	Screening	Baseline ^a		Intervention	Endline
			Week 4 & 8	Week 6		
Primary study parameters						
Muscle mass	ALM in kg, DEXA			✓		✓
Muscle function	5CST in seconds	✓ ^a	✓			✓
Gut microbiota	<i>Bifidobacteria</i> (stool sample), in log ₁₀ cells per gram dried feces		✓			✓ ¹
Immune-metabolic health	Fasting blood glucose in mmol/L, glycoprotein acetyls in mmol/L very low-density lipoprotein diameter in nm (from Nightingale 1H NMR platform)		✓			✓

Secondary parameters				
Muscle function	SPPB, TUG test, 6MWT		✓	✓
	1 repetition maximum test as measured on exercise machines		✓	✓
Body composition	Whole body and regional lean and fat mass, DEXA		✓	✓
Immune-metabolic health	Fasting haemoglobin A1c, Fasting insulin,		✓	✓
	Blood pressure (systolic, diastolic)	✓	✓	✓
Muscle mass post-surgery	single-slice CT-scans midway the upper legs		✓ ^b	✓ ^b
Physical functioning post-surgery	WOMAC		✓ ^b	✓ ^b
Other study parameters				
Physical activity	7-day accelerometry (Actigraph)		✓	✓ ¹
	SQUASH questionnaire	✓		
Hand grip strength	JAMAR Dynamometer		✓	✓
Anthropometrics	Height	✓	✓	✓
	Bodyweight	✓	✓	✓
	Waist circumference		✓	✓
<i>Questionnaires</i>				
Screening questionnaire	Demographics, medical history, supplement use, diet, medication, oral health	✓		
Baseline questionnaire	Education, alcohol intake, smoking, mini-nutritional assessment, FRAIL score		✓	
Gut health	Bristol Stool Chart		✓	✓ ¹
	GSRS		✓	✓ ¹
Sleep quality	Pittsburgh Sleep Quality Index questionnaire		✓	✓ ¹
General quality of life	SF-36		✓	✓ ¹

Muscle related quality of life	SF-SarQol		✓		✓ ¹
Vaccination status	Sars-Cov-2 vaccination status, recent vaccinations (<1y)	✓			
Global cognition	MMSE	✓			
Nutrient and food intake	3-day pre-structured dietary records on paper		✓		✓ ¹
Dutch Healthy Diet Index adherence	Digital EETSCORE		✓	✓	✓
<i>Biological measurements and scores</i>					
Gut microbiota composition and function	shotgun metagenomics in fecal samples		✓		✓ ¹
Vitamin D status	Fasted serum 25(OH)D levels		✓		✓
Kidney health	Estimated Glomerular Filtration rate from creatinine	✓			
Biological age	MetaboHealth Score (from Nightingale 1H NMR in serum)	✓ ^c	✓		✓
	MetaboAge from Nightingale 1H NMR in serum		✓		✓
Cardiometabolic risk	Framingham Risk Score		✓		✓
Intervention response predictors and measures of response heterogeneity	Algorithms will be created with Nightingale platform metabolites, predicting relevant primary and secondary endpoints		✓		✓
Intervention responses in RNA expression and DNA methylation, and the influence of genetic variation on the response heterogeneity	In reference and metabolically compromised groups for comparison with other lifestyle interventions in the future.		✓ ^c		✓ ^c

Questionnaires

Health related quality of life will be assessed with the short-form health survey 36 (SF-36) (119), a measure of functional health and wellbeing with good reliability in community-dwelling older adults (120). Specific muscle-related quality of life will be measured by the short version Sarcopenia Quality of Life (SF-SarQoL) questionnaire (121), which was created to measure sarcopenia-related quality of life (122, 123).

Sleep quality over the past month will be measured with the Pittsburgh Sleep Quality Index (124). It measures several aspects of sleep quality (125, 126) and has been validated in older adults (124, 127).

Gastrointestinal health will be measured by the Gastrointestinal Symptom Rating Scale (GSRS) questionnaire, which evaluates a wide range of gastrointestinal symptoms by letting people rate the severity of symptoms over the past week and is sensitive to change (128). It consists of 15 questions covering 5 common symptom clusters (abdominal pain, reflux, indigestion, constipation and diarrhoea) on a 7-point Likert scale (129).

In the TKR groups, the Western Ontario and McMaster Universities Arthritis Index (WOMAC) will be applied. The WOMAC-index is specifically developed for patients with knee or hip osteoarthritis and represents the gold standard to assess functional outcome following TKR. It consists of three domains that assess pain, stiffness and physical functioning (130).

Physical activity

At baseline and during week 12, physical activity levels will be assessed with a three-axial accelerometer (Actigraph GT3x-BT) with a sampling rate of 30Hz, which will be worn on the waist for 7 days during waking hours (131, 132). In parallel to wearing the accelerometer, general physical activity will be recorded in an activity diary in order to account for potential extreme values recorded by the accelerometer.

Blood measurements

All baseline and endline blood samples will be collected in a fasted state via venepuncture. Whole blood will be collected in an EDTA tube for the measurement of HbA1c, locally at each centre. Serum samples will be collected in serum gel tubes, processed locally, and aliquoted and stored at -80C. These serum aliquots will be used to assess Vitamin 25(OH)D levels and Nightingale metabolomics at the end of the study. Plasma samples will be collected in EDTA tubes and processed locally. Plasma as well as buffy coat will be aliquoted and frozen at -80C. At the end of the study insulin levels will be determined in plasma and deoxyribonucleic acid (DNA) extracted from the buffy coat. Lastly, blood will be collected into PAXgene tubes, processed, and stored at -20 locally for later mRNA extraction.

Biological age and metabolic health

The primary immune-metabolic read-outs of fasting GLC, GlycA and VLDL-D have been chosen for their association with three aspects of metabolic health; blood glucose homeostasis, inflammation and lipid profiles, respectively (133-135). These will be measured as part of the high-throughput Proton NMR metabolomics platform Nightingale Health (136). Thirty-seven of the 250 measured biomarkers are clinically validated, which include GLC and GlycA. The MetaboHealth score will be calculated from 14 of the Nightingale biomarkers: total lipids in chylomicrons and extremely large VLDL, total lipids in small HDL, mean diameter for VLDL particles, ratio of polyunsaturated fatty acids to total fatty acids (%), GLC, lactate, histidine, isoleucine, leucine, valine, phenylalanine, acetoacetate, albumin and glycoprotein acetyls. The MetaboHealth score predicts 5- and 10-year mortality as well as several other health outcomes(91).

Other metabolomics-based scores will be investigated for exploratory studies into baseline and response heterogeneity, such as the MetaboAge score. MetaboAge is associated with chronological age and is calculated using 56 measurements of the Nightingale platform alongside total cholesterol, HbA1c, fasted insulin level, systolic and diastolic blood pressure (137). Additionally, the FRS score will be investigated, as it predicts cardiovascular disease by using metabolic data (total cholesterol, HDL cholesterol), blood pressure, current smoking, diabetes status, antihypertensive medication use, age, sex and systolic blood pressure (138).

Fecal measurements

Fecal samples are collected by the participants at home at a chosen time in the baseline/endline week with the materials provided. Participants will be provided with a cooling element (for cooled-transport of the fecal samples), which they will freeze >24 hours before sample collection in their home freezer (-20C). With a “feces catchers” (Tag Hemi) stool is caught, and participants are instructed to fill two-thirds of each of two 3ml tubes (Micronic MP52961, NBS Scientific Holding BV) using small spatulas (BIDFOOD 231-0416 or VWR International BV 848930). The tubes will then be immediately stored in the frozen cooling element (SARSTEDT 951123) in their freezer. Within one week, the tubes will be transported, via the cooling element placed inside a styrofoam container, to the university and stored at -80C. Participants will complete a short questionnaire at the time of stool collection, containing the Bristol stool chart on stool consistency (139), date and time of defecation, storage time, recent antibiotic use (<12 months), probiotic use (<3 days), current laxative use, milk/lactose tolerance and travel abroad (<1 month). Fecal *Bifidobacteria* numbers will be used as main gut outcome (78-80, 140). Additionally, when all data is collected, additional analyses (shotgun metagenomics (141)) will be carried out to determine the microbiota composition, gene diversity and functional pathways.

Anthropometrics

Height (to nearest cm; SECA stadiometer) and body weight (to nearest 0.1 kg; on SECA or Welch Allyn – HOM599) will be measured with light clothing on and used to determine body mass index (weight/m²).

Waist circumference will be measured using non-stretching measuring tape, midway between the superior iliac crest and lowest rib in standing position with feet ~30cm apart, on exhalation and recorded twice. The mean of the two measurements will be rounded to 0.5 cm or repeated if the difference is >1cm (142).

Blood pressure will be recorded 3 times consecutively on the right arm once the participant has been seated with uncrossed legs for at least 10 minutes, with a minute rest in between measurements (OMRON HEM-907 or Welch Allyn – ProBP3400). If systolic measures differ >12 mmHg, or diastolic >10 mmHg, one (if only the first is much different) or three extra measurements are taken.

Physical functioning and muscle strength

Several aspects of physical performance will be measured. The 5CST, the Balance Test and the Gait Speed test together make up the SPPB, measuring lower extremity function (143). The total SPPB score has been shown to predict disability and mortality (118).

The TUG test will be applied to measure basic functional mobility (107, 144). During this test, the amount of time needed for the participant to rise from an arm chair, walk 3 meters and return to back to the chair in the seated position, is recorded (107, 144, 145).

In the 6MWT two cones are placed 30 metres apart (Wageningen: tape crosses 14 meters apart) and participants walk around these. Using walking aids or sitting down to rest is allowed. Total distance is measured to the nearest metre after 6 minutes (146).

Maximal handgrip strength will be measured with a handheld dynamometer (JAMAR hydraulic dynamometer)(147). Participants are seated on a chair without armrest, holding their elbow at 90, separate from their body. The dynamometer is set so that the middle knuckle of the middle finger is at a 90° angle around the dynamometer. Per hand, three maximal effort measurements will be taken with at least 30 seconds rest in between, alternating between left and right hands. Participants are encouraged verbally. The maximum of all 6 repetitions will be used. Handgrip strength is an indicator of overall health as it has predictive validity for mobility, functional status, cognitive decline and mortality (148).

Muscle strength will be measured by 1RM tests on weightlifting machines (leg press, leg extension, chest press, horizontal row, shoulder press and vertical pull) (149). The 1RM testing procedure has been shown to be valid, reliable, and safe in older adults (149). 1RM strength determines the maximum amount of weight that can be lifted in a single repetition. Before the baseline assessment of 1RM strength, participants will be familiarized with the

exercise equipment, ensuring proper lifting technique. An estimation of 1RM will be obtained using the multiple-repetitions testing procedure and Brzycki formula(150). After warming up, weight is increased until a weight is reached that participants can move ≤ 6 times. In a second session (at least 2 days after the estimation), the actual 1RM will be determined as previously described (149).

Muscle mass and body composition parameters

DEXA whole body scan (GE Lunar Prodigy advance, Hologic Horizon or Hologic Discovery A) will be used to measure ALM as a measure of muscle mass (151). Participants will be fasted and wearing light clothing. Concomitantly, whole body lean mass, whole body and regional fat mass and bone mineral density will be recorded in the same scan using the systems' software package.

In the TKR group, CT-scans will be additionally performed to assess thigh muscle cross-sectional area in both the operated and non-operated leg simultaneously, to compare legs. Participants will undergo a CT scan (IDT 8000; Philips Medical Systems, Best Netherlands) at mid-thigh level of both legs performed at the radiology department of MUMC+ to determine (anatomical) cross sectional area of all thigh muscles, as well as the quadriceps femoris and hamstring muscles separately. While the participants are supine with their legs extended and their feet secured, a 3-mm thick axial image will be taken 15 cm proximal to the top of the patella. The scanning characteristics will be as follows: 120kV, 300mA, rotation time of 0.75 sec, and a field of view of 500 mm. The exact scanning position will be measured and marked for replication 12 weeks later.

Nutrition measurements

Nutritional intake will be measured with a structured 3-day food diary. Days are randomized over a week to obtain a good distribution of the different days. Participants are instructed to fill out as much detail as possible on type, brand and portion size in household measures. Food diaries are checked for completeness upon handing in, and standardized glass models of cups, glasses, and bowls are used for estimating portion sizes of protein-rich food if description is insufficient. Data will be processed in Compl-eat™ (152)(Human Nutrition WUR, Wageningen, NL), which uses the Dutch Food Composition Database (153) and Dutch portion sizes (154).

Additionally, participants will fill in the EETSCORE digitally for monitoring and guidance. This is a 34-item food frequency questionnaire that tests adherence to the Dutch Healthy Diet guidelines (155, 156). An advice module is attached that informs people how well their intake matches the guidelines and informs them how they could be improved.

Intervention

Nutrition intervention

The aim of the nutritional intervention is to reach sufficient intake of dietary protein while improving compliance to the general guidelines for healthy eating later in life (Dutch Healthy Diet guidelines (157)). The intervention is aimed at a total protein intake of 1.5 grams per kg bodyweight per day, with a sufficient amount (20-25 g) at the three main meals and pre-sleep (158), and additional GOS fibre intake of 5.5 g per day on top of the regular intake. The targets will be reached with dietary counselling and a protein-rich supplement (FrieslandCampina, the Netherlands) which will be consumed twice a day; at breakfast and pre-sleep. Breakfast is chosen because protein intake of older adults is especially low during breakfast (54, 159). The pre-sleep moment is chosen to introduce an additional moment of protein ingestion as regular protein peaks throughout the day are proposed to be especially important for supporting muscle protein synthesis in older adults (52). The morning powder contains 25g whey protein, 5.5g Biotis™ GOS, 800 IU vitamin D, and 250 mg calcium per portion. The evening powder contains 20g whey protein and 116mg calcium per portion (160). The powders have a mild vanilla flavour.

At baseline, participants will meet with trained researchers/dieticians to discuss their diet and how to adapt this towards the Dutch Healthy Diet guidelines by choosing 2 goals based on the EETSCORE filled out digitally (161) which will show the participant how well they adhere to the different components of the Dutch Healthy Diet Index (156).

During the 12 weeks intervention, regular contact will take place at the training location to increase compliance. Adherence to the intervention powders will be checked and stimulated. At week 6 of the intervention, each individual participant of the intervention groups fills in the EETSCORE again to monitor change, support compliance and choose a new goal if desired. Compliance to the intake of 'breakfast' and 'before sleep' products will be checked with intake logs on which participants fill in their daily supplement intake.

Resistance exercise intervention

Participants will go to the training location three days per week for 12 weeks of whole body RE training under experienced supervision, with always at least 1 rest day in between training days. Seven resistance type exercises will be performed using regular weightlifting machines and weight-bearing exercises: leg press, leg extension, calf raises, chest press, horizontal row, shoulder press and vertical pull. Focus in sessions 1 and 3 of each week are on 1-2 seconds of concentric and 1-2 seconds of eccentric contractions as to specifically train for muscle mass and strength. Session 2 of each week focusses on speed of contraction as to specifically train for muscular power, by making concentric contractions quick and explosive. Before and after each session, a 5-10 min warm up / cooling down at low intensity will be performed on a cycle ergometer/stepper/cross-trainer or equivalent.

Intensity and workload of the program will gradually increase from 3 sets of 10-15 repetitions at 50-70% of one-repetition maximum (1RM) in week 1, to 4 sets of 8-12 repetitions at 70-80% of 1RM in week 4 and beyond for the leg press and leg extension. For the remaining exercises, 2 sets will be performed throughout the entire training period. All sets and exercises will be separated by 1.5 - 2.5 minutes of rest, resulting in a total session time of approximately 45-60 min.

All final sets of each exercise (excluding calf raises) will be performed until voluntary failure. Training intensity (weight lifted) will be adapted based on 1RM assessment in week 4 and 8, as well as when more than 10 repetitions can be performed in the final set until failure. Apart from these general characteristics, several group specific aspects apply for each intervention group separately. The metabolically compromised group will train in groups of 3-5 participants with at least one trainer. The mobility compromised group will start training in groups of 2 participants with one trainer in week 1-3, which can be increased to 2-4 participants per trainer from week 4 and beyond. The intervention group recovering from total knee replacement will have supervised training on top of standard care and/or any physiotherapy sessions that patients follow. Training will commence with one-on-one supervision in week 1-2, increasing to groups of 2-4 participants when possible in week 3-4 and beyond, depending on inclusion rate. Intensity, workload and progressiveness of the program will be designed on an individual patient level (i.e., taking post-surgery trainability and pain of the individual patient into account) and in consultation with the treating physician and physiotherapist when appropriate.

Statistical analysis

Both intention-to-treat, as well as per protocol analyses will be carried out. For the primary analysis, linear mixed models will be used to analyse within-group changes of the primary study parameters (5CST, ALM, *Bifidobacteria*, GLC, GlycA, VLDL-D). Individuals will be added as random factors. To study the intervention in the TKR group, the TKR intervention group will be modelled with control group (both fixed factors). Secondary analyses will compare the primary study parameters of each group to the active reference group values with linear models, using age and sex as covariates.

Data-driven exploratory analyses will be carried out to study heterogeneity in response within each group and in the total sample post-hoc. The within-group heterogeneity will be analysed by studying the changes in the primary outcomes for the concerning group, through methods such as Principal Component Analysis (PCA) or a penalized linear regression model. Using this approach we investigate which secondary/exploratory measurements associate the strongest with the primary outcome measurements, at baseline as well as when comparing the deltas imposed by the intervention study, i.e. to explore which factors play a role in the response heterogeneity. The heterogeneous changes between the different groups will be explored by the construction of a composite score consisting of all the primary outcomes. We

will use this, along with a PCA and regression approach to investigate the differences between the individuals at baseline and heterogeneity of the different intervention groups.

Future analyses may include the influence of genetic variation on the response and changes in mRNA expression and DNA methylation in comparison to other existing lifestyle studies in older adults.

Compliance and withdrawal

Training participation will be deemed as ‘compliant’ when 80% or more of the training sessions have been followed. An intake of 90% or more of the powders will be taken as ‘compliant’. Participants can leave the study at any time for any reason if they wish to do so, without any consequences. The investigator can decide to withdraw a participant from the study for urgent medical reasons. Participants dropping out of the study will be asked for permission to contact them for inclusion in the endline measurements as to enable intention-to-treat analyses, if the reason for dropout does not limit the possibility for such post-measurements. Drop-outs are not replaced because the sample size calculations include an expected 15% dropout.

Logging, data collection and management

A data management plan is available at DMP online and can be shared upon reasonable request (162). All measurements are carried out by trained researchers according to the standard operating procedures created for this study. The privacy of the potential participants that are screened and the participants of the study will be protected according to the European Union General Data Protection Regulation and the Dutch Act on Implementation of the General Data Protection Regulation.

All necessary personal information collected material and study results will be related to the unique participant number. Measurements are entered directly into Castor when possible and include range checks when possible (163). Personal information such as name and address are encrypted. Measurements recorded on paper or measuring device are stored in locked locations at each university or at university network drives. Biological material will be stored locally. Data collected and biological materials will be stored for 20 years. If participants revoke their consent, the information stored up to that point in time will be kept, but no new data will be collected or stored. Biological materials will be destroyed. We will archive and save all data that underlie publications, including data collection, cleaning and analysis methods. All research data will be pseudonymized and brought together in a data warehouse, so that researchers of each participating centre have access to all generated research data.

Adverse events

All adverse events related to the study protocol (measurements and intervention) reported spontaneously by the participant or observed by the investigators will be recorded. Severe adverse events are reported to the coordinating investigator and MEC digitally within a week.

Oversight and monitoring

The coordinating investigator at LUMC is the sponsor of this investigator-initiated research. The principal investigators (PI) at LUMC, MUMC+ and WUR are responsible for the trial at each of the respective locations. Yearly updates are provided to the MEC. A data management plan was agreed upon beforehand and approved by the MEC and data steward at LUMC. Data managers at LUMC built and maintain the Castor database used at all locations. PIs and other researchers involved have met monthly to create the protocol, discuss progress and decide on issues and changes in the protocol. Protocol amendments need to be approved by the original MEC before implementation. New IC forms will be provided if necessary.

The study has been approved by the MEC as having a negligible risk, therefore no data safety monitoring board needs to be installed. A monitor is appointed by LUMC who checks data integrity at every location.

Reporting and dissemination

Results of the study will be made public through peer-reviewed international scientific journals, according to the Dutch “Centrale Committee on Research Involving Human Subjects” statement of publication policy and disclosed via their register.

DISCUSSION

This paper describes the design of the VOILA human intervention study that explores the effects of a healthy lifestyle intervention with nutrition and RE in selected groups of older adults at risk for adverse ageing outcomes. This intervention study has been designed to better understand the response in several groups of older adults with different health statuses to a lifestyle intervention, as well as the heterogeneity of response, with the response measured by physical function, muscle, immune-metabolic and gut health parameters and novel biomarkers of ageing.

The intervention was designed using a combination of lifestyle interventions likely to improve health by targeting multiple physiological systems relevant to ageing. The novelty in the approach of this study is represented by i) preselection of three segments of older adults based on their health status to allow exploration of heterogeneous responses and ii) simultaneously measuring outcomes in three major ageing related domains including immune-metabolic health, gut-microbiome composition, and muscle mass/physical functioning.

This study will give us the opportunity to study the effect of the intervention on a range of markers of biological ageing possibly enabling improved and individual response monitoring of interventions in older adults. Currently, no gold standard biomarkers are available in human ageing research, neither for defining the rate of biological ageing nor for recording the response to interventions to slow down that rate. Despite the lack of consensus on the most informative biomarkers (164), it is largely (>80%) agreed upon that ageing is

heterogeneous, cannot be measured by only a single metric, and proceeds differently in different tissues. Furthermore, the type of intervention that received the highest agreement (91%) in terms of its potential to attenuate biological ageing was the lifestyle intervention (164), in line with our approach. Very few interventions combining nutrition and exercise have been carried out in which biological age is measured, but exploratory first results mainly using epigenetic clocks look promising (165, 166). In the VOILA intervention we will largely focus on metabolomics and proteomics biomarkers of biological age in addition to the major outcomes of muscle, immune-metabolic, and gut health. The components of the composite score reflecting biological age, such as those in the MetaboHealth and proteomics scores, are biologically well understood and will allow mechanistic explorations into the response heterogeneity in VOILA.

Our VOILA intervention study has the potential to improve our current understanding of the positive changes induced by a healthy ageing lifestyle intervention in different subgroups of older adults with varying degrees of compromised mobility or metabolism. This allows for exploration of the heterogeneity in individual responses within and across the groups. Better understanding of this heterogeneity should enable further tailoring of intervention efforts towards specific needs of (subgroups of) older individuals as to maximize the beneficial health effects. The VOILA intervention study we report on here is embedded within the larger Dutch Society for Research on Ageing (DuSRA)-VOILA project, in which different disciplines of nine institutes and eight private partners collaborate (167). The overarching project aim is to improve the recording of the current and forecast the future biological age of the population aged ≥ 50 years and reinforce physiological reserves that help maintain functional ability. Data and samples from the intervention study we report on here will allow for interdisciplinary research within the DuSRA-VOILA project to advance the field in new ways. At the time of writing, no ageing biomarkers have been approved for clinical application (90), but there is a need for biological age markers responsive to intervention (90, 168). With VOILA we plan to create biomarkers to predict intervention response, contributing one step towards preclinical or clinical use of biological age biomarkers.

In the intervention study, the main research questions focus on the three age-related outcome domains of muscle/physical function, immune-metabolic and gut health. Several additional measurements will allow for better phenotypic characterisation of individuals in exploring the heterogeneity in ageing and intervention response, including objective measurement of activity levels via accelerometry, and detailed 3-day food diaries from which intake of most nutrients can be extracted. Moreover, the questionnaires on gut complaints, sleep quality, as well as general and sarcopenia-related quality of life add information on patient characteristics potentially contributing to differences in outcomes, as well as measuring well-being which is a very relevant but often overlooked paradigm in intervention studies focusing on a single specific domain. Comprehensive blood samples are collected so that future analyses may include the influence of genetic variation on the response, and changes in

mRNA expression and DNA methylation in comparison to other existing lifestyle studies in older adults.

The VOILA intervention study is not a randomized controlled trial, as our aim is not to study the effectiveness of specific subparts of the intervention. In the within-group changes, no distinction can be made between the separate components of the intervention (RE, nutritional powders and their components, and dietary advice). Instead, we use a positive reference group consisting of a sample of the healthiest older adults based on activity level and immune-metabolic profile. The health of the intervention participants can be compared to the reference group to assess in which ways these groups differ at baseline, and to assess to what extent the outcomes of the health-compromised groups can shift towards the profile of the healthiest counterparts following the intervention. Of main importance is that the intervention groups all receive the same intervention and are measured according to the standardized protocol so we can study response heterogeneity both between and within groups.

This human lifestyle intervention is innovative in that we target domains hypothesized to underly overall biological ageing, as opposed to a single domain or disease as outcome. Additionally, this study will provide valuable insight into the heterogeneity in effect of a healthy diet and exercise intervention aimed at improving healthy ageing in various different groups of health compromised community-dwelling older adults.

Results will provide information on the usefulness of markers of biological ageing in an intervention study and pave the way to develop improved response biomarkers for interventions. Insights from this intervention study can be used as the basis to further our understanding of factors underlying the heterogeneous ageing process, to better tailor future preclinical interventions and to fuel novel measurements such as biological ageing markers and intervention response markers. The results of this VOILA intervention study will be relevant for improving health in older adults and recording and forecasting which interventions effectively improve biological ageing and wellbeing.

DECLARATIONS

Ethics approval and consent to participate

All procedures are performed in accordance with the World Medical Association Declaration of Helsinki and in compliance with relevant laws and institutional guidelines, including national and European privacy rights. Ethical approval was obtained from the Medical Research Ethical Committee Leiden Den Haag Delft, the Netherlands, on 20th October 2021 (registration: “P21.049” locally and “NL76879.058.21” nationally at the Central Committee on Research Involving Human Subjects). Additional amendments were approved on 27th May 2022, 21st July 2022, 11th November 2022, 21st July 2023 and 27th of February 2024. Written consent is obtained from all participants before starting.

FUNDING

This work was supported by ZonMW (457001001). As part of this, FrieslandCampina supported financially and in kind by providing the nutritional powders. The private partner had no influence on the study design and will not be involved in data collection, analysis and interpretation. Manuscripts of the results will be published irrespective of the results and open access within the constraints of the consortium agreement.

AUTHORS' CONTRIBUTIONS

ES, MB, LdG and LV created the study and obtained funding. CK, LV, LdG, ES, MB, SM, AM and JM designed and wrote the protocol and acquired ethical approval. LV, ES, PG and LdG are responsible for the overall study coordination and JM, AM and CK are running the day-to-day trial, recruitment and sample collection. CK drafted the manuscript and all authors revised it. All authors read and approved the final manuscript.

ACKNOWLEDGEMENTS

We would like to thank the supporting staff from LUMC, MUMC+ and WUR for their valuable assistance and flexibility in creating and setting up the VOILA study in each centre. We also thank Frans van der Ouderaa for thinking along in the early stages of designing the VOILA intervention study and Arjen Nauta and Sjors Verlaan for providing the supplement.

REFERENCES

1. Riley JC. Estimates of regional and global life expectancy, 1800–2001. *Population and development review*. 2005;31(3):537-43.
2. Roser M, Ortiz-Ospina E, Ritchie H. Life expectancy. *Our World in Data*. 2013.
3. Partridge L, Deelen J, Slagboom PE. Facing up to the global challenges of ageing. *Nature*. 2018;561(7721):45-56.
4. United Nations DoE, Affairs S. *World Population Ageing*, 2019. 2019.
5. Vos T, Flaxman AD, Naghavi M, Lozano R, Michaud C, Ezzati M, et al. Years lived with disability (YLDs) for 1160 sequelae of 289 diseases and injuries 1990-2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet*. 2012;380(9859):2163-96.
6. Karppinen H, Laakkonen M-L, Strandberg TE, Huohvanainen EA, Pitkala KH. Do you want to live to be 100? Answers from older people. *Age and Ageing*. 2016;45(4):543-9.
7. Bowen CE, Skirbekk V. Old age expectations are related to how long people want to live. *Ageing & Society*. 2017;37(9):1898-923.
8. Li Q, Wang S, Milot E, Bergeron P, Ferrucci L, Fried LP, et al. Homeostatic dysregulation proceeds in parallel in multiple physiological systems. *Ageing Cell*. 2015;14(6):1103-12.
9. Lopez-Otin C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. *Cell*. 2013;153(6):1194-217.
10. López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. Hallmarks of aging: An expanding universe. *Cell*. 2023.
11. Schmauck-Medina T, Molière A, Lautrup S, Zhang J, Chlopicki S, Madsen HB, et al. New hallmarks of ageing: a 2022 Copenhagen ageing meeting summary. *Ageing (Albany NY)*. 2022;14(16):6829-39.
12. Fontana L, Partridge L, Longo VD. Extending healthy life span—from yeast to humans. *Science*. 2010;328(5976):321-6.
13. Aunan JR, Watson MM, Hagland HR, Soreide K. Molecular and biological hallmarks of ageing. *Br J Surg*. 2016;103(2):e29-46.
14. Fraser HC, Kuan V, Johnen R, Zwierzyna M, Hingorani AD, Beyer A, et al. Biological mechanisms of aging predict age-related disease co-occurrence in patients. *Ageing Cell*. 2022;21(4):e13524.
15. Short KR, Vittone JL, Bigelow ML, Proctor DN, Nair KS. Age and aerobic exercise training effects on whole body and muscle protein metabolism. *Am J Physiol Endocrinol Metab*. 2004;286(1):E92-101.
16. Cruz-Jentoft AJ, Bahat G, Bauer J, Boirie Y, Bruyère O, Cederholm T, et al. Sarcopenia: revised European consensus on definition and diagnosis. *Age Ageing*. 2019;48(1):16-31.
17. Visser M, Goodpaster BH, Kritchevsky SB, Newman AB, Nevitt M, Rubin SM, et al. Muscle mass, muscle strength, and muscle fat infiltration as predictors of incident mobility limitations in well-functioning older persons. *The journals of gerontology Series A, Biological sciences and medical sciences*. 2005;60(3):324-33.
18. Visser M, Kritchevsky SB, Goodpaster BH, Newman AB, Nevitt M, Stamm E, et al. Leg muscle mass and composition in relation to lower extremity performance in men and women aged 70 to 79: the health, aging and body composition study. *Journal of the American Geriatrics Society*. 2002;50(5):897-904.
19. Pedersen BK, Saltin B. Exercise as medicine - evidence for prescribing exercise as therapy in 26 different chronic diseases. *Scand J Med Sci Sports*. 2015;25 Suppl 3:1-72.
20. Pahor M, Blair SN, Espeland M, Fielding R, Gill TM, Guralnik JM, et al. Effects of a physical activity intervention on measures of physical performance: Results of the lifestyle interventions and independence for Elders Pilot (LIFE-P) study. *The journals of gerontology Series A, Biological sciences and medical sciences*. 2006;61(11):1157-65.
21. Booth FW, Roberts CK, Laye MJ. Lack of exercise is a major cause of chronic diseases. *Compr Physiol*. 2012;2(2):1143-211.
22. Stamatakis E, Lee IM, Bennie J, Freeston J, Hamer M, O'Donovan G, et al. Does Strength-Promoting Exercise Confer Unique Health Benefits? A Pooled Analysis of Data on 11 Population Cohorts With All-Cause, Cancer, and Cardiovascular Mortality Endpoints. *American journal of epidemiology*. 2018;187(5):1102-12.
23. Barker K, Holland AE, Skinner EH, Lee AL. Clinical Outcomes Following Exercise Rehabilitation in People with Multimorbidity: A Systematic Review. *J Rehabil Med*. 2023;55:jrm00377.
24. Peterson MD, Rhea MR, Sen A, Gordon PM. Resistance exercise for muscular strength in older adults: a meta-analysis. *Ageing research reviews*. 2010;9(3):226-37.
25. Stewart VH, Saunders DH, Greig CA. Responsiveness of muscle size and strength to physical training in very elderly people: a systematic review. *Scand J Med Sci Sports*. 2014;24(1):e1-10.

26. Borde R, Hortobágyi T, Granacher U. Dose-Response Relationships of Resistance Training in Healthy Old Adults: A Systematic Review and Meta-Analysis. *Sports Med.* 2015;45(12):1693-720.
27. Jadcak AD, Makwana N, Luscombe-Marsh N, Visvanathan R, Schultz TJ. Effectiveness of exercise interventions on physical function in community-dwelling frail older people: an umbrella review of systematic reviews. *JBIG Database System Rev Implement Rep.* 2018;16(3):752-75.
28. Liu CJ, Latham NK. Progressive resistance strength training for improving physical function in older adults. *Cochrane Database Syst Rev.* 2009;2009(3):Cd002759.
29. Churchward-Venne TA, Tieland M, Verdijk LB, Leenders M, Dirks ML, de Groot LC, et al. There Are No Nonresponders to Resistance-Type Exercise Training in Older Men and Women. *J Am Med Dir Assoc.* 2015;16(5):400-11.
30. DeFronzo RA, Tripathy D. Skeletal muscle insulin resistance is the primary defect in type 2 diabetes. *Diabetes Care.* 2009;32 Suppl 2(Suppl 2):S157-63.
31. Thiebaut D, Jacot E, DeFronzo RA, Maeder E, Jequier E, Felber JP. The effect of graded doses of insulin on total glucose uptake, glucose oxidation, and glucose storage in man. *Diabetes.* 1982;31(11):957-63.
32. Srikanthan P, Karlamangla AS. Relative muscle mass is inversely associated with insulin resistance and prediabetes. Findings from the third National Health and Nutrition Examination Survey. *J Clin Endocrinol Metab.* 2011;96(9):2898-903.
33. McGlory C, von Allmen MT, Stokes T, Morton RW, Hector AJ, Lago BA, et al. Failed Recovery of Glycemic Control and Myofibrillar Protein Synthesis With 2 wk of Physical Inactivity in Overweight, Prediabetic Older Adults. *The Journals of Gerontology: Series A.* 2017;73(8):1070-7.
34. Reidy PT, McKenzie AI, Mahmassani Z, Morrow VR, Yonemura NM, Hopkins PN, et al. Skeletal muscle ceramides and relationship with insulin sensitivity after 2 weeks of simulated sedentary behaviour and recovery in healthy older adults. *J Physiol.* 2018;596(21):5217-36.
35. Handy RM, Holloway GP. Insights into the development of insulin resistance: Unraveling the interaction of physical inactivity, lipid metabolism and mitochondrial biology. *Front Physiol.* 2023;14:1151389.
36. Davy BM, Winett RA, Savla J, Marinik EL, Baugh ME, Flack KD, et al. Resist diabetes: A randomized clinical trial for resistance training maintenance in adults with prediabetes. *PLOS ONE.* 2017;12(2):e0172610.
37. Iglay HB, Thyfault JP, Apolzan JW, Campbell WW. Resistance training and dietary protein: effects on glucose tolerance and contents of skeletal muscle insulin signaling proteins in older persons. *The American journal of clinical nutrition.* 2007;85(4):1005-13.
38. Grøntved A, Rimm EB, Willett WC, Andersen LB, Hu FB. A prospective study of weight training and risk of type 2 diabetes mellitus in men. *Arch Intern Med.* 2012;172(17):1306-12.
39. Lee DH, Luo X, Rezende LFM, Joh HK, Keum N, Rimm EB, et al. Long-term Weight Training and Mortality in U.S. Male Health Professionals With and Without Type 2 Diabetes. *Diabetes Care.* 2023;46(1):138-48.
40. Tanasescu M, Leitzmann MF, Rimm EB, Willett WC, Stampfer MJ, Hu FB. Exercise type and intensity in relation to coronary heart disease in men. *Jama.* 2002;288(16):1994-2000.
41. Shiroma EJ, Cook NR, Manson JE, Moorthy MV, Buring JE, Rimm EB, et al. Strength Training and the Risk of Type 2 Diabetes and Cardiovascular Disease. *Med Sci Sports Exerc.* 2017;49(1):40-6.
42. Roberson KB, Potiaumpai M, Widdowson K, Jaghab AM, Chowdhari S, Armitage C, et al. Effects of high-velocity circuit resistance and treadmill training on cardiometabolic risk, blood markers, and quality of life in older adults. *Appl Physiol Nutr Metab.* 2018;43(8):822-32.
43. MacDonald HV, Johnson BT, Huedo-Medina TB, Livingston J, Forsyth KC, Kraemer WJ, et al. Dynamic Resistance Training as Stand-Alone Antihypertensive Lifestyle Therapy: A Meta-Analysis. *J Am Heart Assoc.* 2016;5(10).
44. Ihalainen JK, Inglis A, Mäkinen T, Newton RU, Kainulainen H, Kyröläinen H, et al. Strength training improves metabolic health markers in older individual regardless of training frequency. *Frontiers in physiology.* 2019;10:32.
45. Pedersen BK, Febbraio MA. Muscles, exercise and obesity: skeletal muscle as a secretory organ. *Nat Rev Endocrinol.* 2012;8(8):457-65.
46. Severinsen MCK, Pedersen BK. Muscle-Organ Crosstalk: The Emerging Roles of Myokines. *Endocr Rev.* 2020;41(4):594-609.
47. Gleeson M, Bishop NC, Stensel DJ, Lindley MR, Mastana SS, Nimmo MA. The anti-inflammatory effects of exercise: mechanisms and implications for the prevention and treatment of disease. *Nature reviews Immunology.* 2011;11(9):607-15.
48. Lancaster GI, Febbraio MA. The immunomodulating role of exercise in metabolic disease. *Trends Immunol.* 2014;35(6):262-9.

49. Guimarães-Ferreira L, Cholewa JM, Naimo MA, Zhi XI, Magagnin D, de Sá RB, et al. Synergistic effects of resistance training and protein intake: practical aspects. *Nutrition* (Burbank, Los Angeles County, Calif). 2014;30(10):1097-103.
50. Moore DR, Churchward-Venne TA, Witard O, Breen L, Burd NA, Tipton KD, et al. Protein ingestion to stimulate myofibrillar protein synthesis requires greater relative protein intakes in healthy older versus younger men. *The journals of gerontology Series A, Biological sciences and medical sciences*. 2015;70(1):57-62.
51. Wall BT, Gorissen SH, Pennings B, Koopman R, Groen BB, Verdijk LB, et al. Aging Is Accompanied by a Blunted Muscle Protein Synthetic Response to Protein Ingestion. *PLoS One*. 2015;10(11):e0140903.
52. Bauer J, Biolo G, Cederholm T, Cesari M, Cruz-Jentoft AJ, Morley JE, et al. Evidence-based recommendations for optimal dietary protein intake in older people: a position paper from the PROT-AGE Study Group. *J Am Med Dir Assoc*. 2013;14(8):542-59.
53. Deutz NE, Bauer JM, Barazzoni R, Biolo G, Boirie Y, Bosy-Westphal A, et al. Protein intake and exercise for optimal muscle function with aging: recommendations from the ESPEN Expert Group. *Clin Nutr*. 2014;33(6):929-36.
54. Tieland M, Borgonjen-Van den Berg KJ, Van Loon LJ, de Groot LC. Dietary Protein Intake in Dutch Elderly People: A Focus on Protein Sources. *Nutrients*. 2015;7(12):9697-706.
55. Campbell WW, Trappe TA, Wolfe RR, Evans WJ. The recommended dietary allowance for protein may not be adequate for older people to maintain skeletal muscle. *The Journals of Gerontology Series A: Biological Sciences and Medical Sciences*. 2001;56(6):M373-M80.
56. Houston DK, Nicklas BJ, Ding J, Harris TB, Tylavsky FA, Newman AB, et al. Dietary protein intake is associated with lean mass change in older, community-dwelling adults: the Health, Aging, and Body Composition (Health ABC) Study. *The American journal of clinical nutrition*. 2008;87(1):150-5.
57. Volpi E, Campbell WW, Dwyer JT, Johnson MA, Jensen GL, Morley JE, et al. Is the optimal level of protein intake for older adults greater than the recommended dietary allowance? *Journals of Gerontology Series A: Biomedical Sciences and Medical Sciences*. 2013;68(6):677-81.
58. Labata-Lezaun N, Llorca-Almuzara L, López-de-Celis C, Rodríguez-Sanz J, González-Rueda V, Hidalgo-García C, et al. Effectiveness of Protein Supplementation Combined with Resistance Training on Muscle Strength and Physical Performance in Elderly: A Systematic Review and Meta-Analysis. *Nutrients*. 2020;12(9).
59. Ten Haaf DSM, Nuijten MAH, Maessen MFH, Horstman AMH, Eijsvogels TMH, Hopman MTE. Effects of protein supplementation on lean body mass, muscle strength, and physical performance in nonfrail community-dwelling older adults: a systematic review and meta-analysis. *The American journal of clinical nutrition*. 2018;108(5):1043-59.
60. Shad BJ, Thompson JL, Breen L. Does the muscle protein synthetic response to exercise and amino acid-based nutrition diminish with advancing age? A systematic review. *Am J Physiol Endocrinol Metab*. 2016;311(5):E803-e17.
61. Cermak NM, Res PT, de Groot LC, Saris WH, van Loon LJ. Protein supplementation augments the adaptive response of skeletal muscle to resistance-type exercise training: a meta-analysis. *The American journal of clinical nutrition*. 2012;96(6):1454-64.
62. Finger D, Goltz FR, Umpierre D, Meyer E, Rosa LH, Schneider CD. Effects of protein supplementation in older adults undergoing resistance training: a systematic review and meta-analysis. *Sports Med*. 2015;45(2):245-55.
63. Hidayat K, Chen GC, Wang Y, Zhang Z, Dai X, Szeto IMY, et al. Effects of Milk Proteins Supplementation in Older Adults Undergoing Resistance Training: A Meta-Analysis of Randomized Control Trials. *The journal of nutrition, health & aging*. 2018;22(2):237-45.
64. Hengeveld LM, Wijnhoven HAH, Olthof MR, Brouwer IA, Simonsick EM, Kritchevsky SB, et al. Prospective Associations of Diet Quality With Incident Frailty in Older Adults: The Health, Aging, and Body Composition Study. *Journal of the American Geriatrics Society*. 2019;67(9):1835-42.
65. Kirwan RP, Mazidi M, Rodríguez García C, Lane KE, Jafari A, Butler T, et al. Protein interventions augment the effect of resistance exercise on appendicular lean mass and handgrip strength in older adults: a systematic review and meta-analysis of randomized controlled trials. *The American journal of clinical nutrition*. 2022;115(3):897-913.
66. Goodrich JK, Waters JL, Poole AC, Sutter JL, Koren O, Blekhan R, et al. Human genetics shape the gut microbiome. *Cell*. 2014;159(4):789-99.
67. Rothschild D, Weissbrod O, Barkan E, Kurilshikov A, Korem T, Zeevi D, et al. Environment dominates over host genetics in shaping human gut microbiota. *Nature*. 2018;555(7695):210-5.
68. Claesson MJ, Jeffery IB, Conde S, Power SE, O'Connor EM, Cusack S, et al. Gut microbiota

- composition correlates with diet and health in the elderly. *Nature*. 2012;488(7410):178-84.
69. Valdes AM, Walter J, Segal E, Spector TD. Role of the gut microbiota in nutrition and health. *Bmj*. 2018;361:k2179.
70. Zhao L, Zhang F, Ding X, Wu G, Lam YY, Wang X, et al. Gut bacteria selectively promoted by dietary fibers alleviate type 2 diabetes. *Science*. 2018;359(6380):1151-6.
71. Sasaki D, Sasaki K, Ikuta N, Yasuda T, Fukuda I, Kondo A, et al. Low amounts of dietary fibre increase in vitro production of short-chain fatty acids without changing human colonic microbiota structure. *Sci Rep*. 2018;8(1):435.
72. Niittynen L, Kajander K, Korpela R. Galactooligosaccharides and bowel function. *Scandinavian Journal of Food and Nutrition*. 2007;51(2):62-6.
73. Blaut M. Relationship of prebiotics and food to intestinal microflora. *European Journal of Nutrition*. 2002;41(1):i11-i6.
74. Bouhnik Y, Flourié B, D'Agay-Abensour L, Pochart P, Gramet Gv, Durand ML, et al. Administration of transgalacto-oligosaccharides increases fecal bifidobacteria and modifies colonic fermentation metabolism in healthy humans. *The Journal of nutrition*. 1997;127(3):444-8.
75. Chambers ES, Preston T, Frost G, Morrison DJ. Role of Gut Microbiota-Generated Short-Chain Fatty Acids in Metabolic and Cardiovascular Health. *Curr Nutr Rep*. 2018;7(4):198-206.
76. Canfora EE, Jocken JW, Blaak EE. Short-chain fatty acids in control of body weight and insulin sensitivity. *Nat Rev Endocrinol*. 2015;11(10):577-91.
77. Vulevic J, Drakoularakou A, Yaqoob P, Tzortzis G, Gibson GR. Modulation of the fecal microflora profile and immune function by a novel trans-galactooligosaccharide mixture (B-GOS) in healthy elderly volunteers. *The American journal of clinical nutrition*. 2008;88(5):1438-46.
78. Vulevic J, Juric A, Walton GE, Claus SP, Tzortzis G, Toward RE, et al. Influence of galactooligosaccharide mixture (B-GOS) on gut microbiota, immune parameters and metabonomics in elderly persons. *The British journal of nutrition*. 2015;114(4):586-95.
79. Walton GE, van den Heuvel EG, Kosters MH, Rastall RA, Tuohy KM, Gibson GR. A randomised crossover study investigating the effects of galactooligosaccharides on the faecal microbiota in men and women over 50 years of age. *The British journal of nutrition*. 2012;107(10):1466-75.
80. Vulevic J, Juric A, Tzortzis G, Gibson GR. A mixture of trans-galactooligosaccharides reduces markers of metabolic syndrome and modulates the fecal microbiota and immune function of overweight adults. *The Journal of nutrition*. 2013;143(3):324-31.
81. Gonai M, Shigehisa A, Kigawa I, Kurasaki K, Chonan O, Matsuki T, et al. Galactooligosaccharides ameliorate dysbiotic Bifidobacteriaceae decline in Japanese patients with type 2 diabetes. *Benef Microbes*. 2017;8(5):705-16.
82. Arboleya S, Watkins C, Stanton C, Ross RP. Gut Bifidobacteria Populations in Human Health and Aging. *Frontiers in microbiology*. 2016;7:1204.
83. Ohland CL, Macnaughton WK. Probiotic bacteria and intestinal epithelial barrier function. *Am J Physiol Gastrointest Liver Physiol*. 2010;298(6):G807-19.
84. Brooks CN, Wight ME, Azeez OE, Bleich RM, Zwetsloot KA. Growing old together: What we know about the influence of diet and exercise on the aging host's gut microbiome. *Front Sports Act Living*. 2023;5:1168731.
85. Liu F, Li P, Chen M, Luo Y, Prabhakar M, Zheng H, et al. Fructooligosaccharide (FOS) and Galactooligosaccharide (GOS) Increase Bifidobacterium but Reduce Butyrate Producing Bacteria with Adverse Glycemic Metabolism in healthy young population. *Sci Rep*. 2017;7(1):11789.
86. Canfora EE, van der Beek CM, Hermes GDA, Goossens GH, Jocken JWE, Holst JJ, et al. Supplementation of Diet With Galactooligosaccharides Increases Bifidobacteria, but Not Insulin Sensitivity, in Obese Prediabetic Individuals. *Gastroenterology*. 2017;153(1):87-97.e3.
87. Morwani-Mangnani J, Rodriguez-Girondo M, Singh-Povel C, Verlaan S, Beekman M, Slagboom PE. Physical activity and fiber intake beneficial for muscle mass and strength preservation during aging: A comprehensive cross-sectional study in the UK biobank cohort. *Exp Gerontol*. 2024;193:112474.
88. Prokopicis K, Chambers E, Ni Lochlainn M, Witard OC. Mechanisms Linking the Gut-Muscle Axis With Muscle Protein Metabolism and Anabolic Resistance: Implications for Older Adults at Risk of Sarcopenia. *Front Physiol*. 2021;12:770455.
89. Narici MV, Maffulli N. Sarcopenia: characteristics, mechanisms and functional significance. *Br Med Bull*. 2010;95:139-59.
90. Moqri M, Herzog C, Poganik JR, Justice J, Belsky DW, Higgins-Chen A, et al. Biomarkers of aging for the identification and evaluation of longevity interventions. *Cell*. 2023;186(18):3758-75.
91. Deelen J, Kettunen J, Fischer K, van der Spek A, Trompet S, Kastenmuller G, et al. A metabolic

- profile of all-cause mortality risk identified in an observational study of 44,168 individuals. *Nat Commun.* 2019;10(1):3346.
92. Kuiper LM, Polinder-Bos HA, Bizzarri D, Vojinovic D, Vallerga CL, Beekman M, et al. Epigenetic and Metabolomic Biomarkers for Biological Age: A Comparative Analysis of Mortality and Frailty Risk. *The journals of gerontology Series A, Biological sciences and medical sciences.* 2023;78(10):1753-62.
 93. Zonneveld MH, Kuhaili NA, Mooijaart SP, Slagboom PE, Jukema JW, Noordam R, et al. Increased 1 H-NMR metabolomics-based health score associates with declined cognitive performance and functional independence in older adults at risk of cardiovascular disease. *medRxiv.* 2023:2023.12.21.23300037.
 94. van de Rest O, Schutte BA, Deelen J, Stassen SA, van den Akker EB, van Heemst D, et al. Metabolic effects of a 13-weeks lifestyle intervention in older adults: The Growing Old Together Study. *Aging (Albany NY).* 2016;8(1):111-26.
 95. Bogaards FA, Gehrman T, Beekman M, van den Akker EB, van de Rest O, Hangelbroek RWJ, et al. PLIS: A metabolomic response monitor to a lifestyle intervention study in older adults. *The FASEB Journal.* 2022;36(11):e22578.
 96. Biagi E, Nylund L, Candela M, Ostan R, Bucci L, Pini E, et al. Through ageing, and beyond: gut microbiota and inflammatory status in seniors and centenarians. *PLoS One.* 2010;5(5):e10667.
 97. Kresovich JK, Park YM, Keller JA, Sandler DP, Taylor JA. Healthy eating patterns and epigenetic measures of biological age. *The American journal of clinical nutrition.* 2022;115(1):171-9.
 98. Pollock RD, Carter S, Velloso CP, Duggal NA, Lord JM, Lazarus NR, et al. An investigation into the relationship between age and physiological function in highly active older adults. *J Physiol.* 2015;593(3):657-80; discussion 80.
 99. Pollock ML, Franklin BA, Balady GJ, Chaitman BL, Fleg JL, Fletcher B, et al. Resistance Exercise in Individuals With and Without Cardiovascular Disease Benefits, Rationale, Safety, and Prescription An Advisory From the Committee on Exercise, Rehabilitation, and Prevention, Council on Clinical Cardiology, American Heart Association. *Circulation.* 2000;101(7):828-33.
 100. Wendel-Vos GW, Schuit AJ, Saris WH, Kromhout D. Reproducibility and relative validity of the short questionnaire to assess health-enhancing physical activity. *Journal of clinical epidemiology.* 2003;56(12):1163-9.
 101. Rossi AP, Micciolo R, Rubele S, Fantin F, Caliari C, Zoico E, et al. Assessing the Risk of Sarcopenia in the Elderly: The Mini Sarcopenia Risk Assessment (MSRA) Questionnaire. *The journal of nutrition, health & aging.* 2017;21(6):743-9.
 102. Malmstrom TK, Miller DK, Simonsick EM, Ferrucci L, Morley JE. SARC-F: a symptom score to predict persons with sarcopenia at risk for poor functional outcomes. *J Cachexia Sarcopenia Muscle.* 2016;7(1):28-36.
 103. Tieland M, Dirks ML, van der Zwaluw N, Verdijk LB, van de Rest O, de Groot LC, et al. Protein supplementation increases muscle mass gain during prolonged resistance-type exercise training in frail elderly people: a randomized, double-blind, placebo-controlled trial. *J Am Med Dir Assoc.* 2012;13(8):713-9.
 104. Bauer JM, Verlaan S, Bautmans I, Brandt K, Donini LM, Maggio M, et al. Effects of a vitamin D and leucine-enriched whey protein nutritional supplement on measures of sarcopenia in older adults, the PROVIDE study: a randomized, double-blind, placebo-controlled trial. *J Am Med Dir Assoc.* 2015;16(9):740-7.
 105. Hsieh PL, Tseng CH, Tseng YJ, Yang WS. Resistance Training Improves Muscle Function and Cardiometabolic Risks But Not Quality of Life in Older People With Type 2 Diabetes Mellitus: A Randomized Controlled Trial. *J Geriatr Phys Ther.* 2018;41(2):65-76.
 106. French HP, Fitzpatrick M, FitzGerald O. Responsiveness of physical function outcomes following physiotherapy intervention for osteoarthritis of the knee: an outcome comparison study. *Physiotherapy.* 2011;97(4):302-8.
 107. van Dongen EJI, Haveman-Nies A, Doets EL, Dorhout BG, de Groot L. Effectiveness of a Diet and Resistance Exercise Intervention on Muscle Health in Older Adults: ProMuscle in Practice. *J Am Med Dir Assoc.* 2020.
 108. Beaudart C, Bruyere O, Geerinck A, Hajaoui M, Scafoglieri A, Perikias S, et al. Equation models developed with bioelectric impedance analysis tools to assess muscle mass: A systematic review. *Clin Nutr ESPEN.* 2020;35:47-62.
 109. Bhasin S, Apovian CM, Travison TG, Pencina K, Moore LL, Huang G, et al. Effect of Protein Intake on Lean Body Mass in Functionally Limited Older Men: A Randomized Clinical Trial. *JAMA Intern Med.* 2018;178(4):530-41.
 110. Tieland M, van de Rest O, Dirks ML, van der Zwaluw N, Mensink M, van Loon LJ, et al. Protein supplementation improves physical performance in frail elderly people: a randomized, double-blind, placebo-controlled trial. *J Am Med Dir Assoc.* 2012;13(8):720-6.

111. Liao CD, Tsao JY, Chiu YS, Ku JW, Huang SW, Liou TH. Effects of Elastic Resistance Exercise After Total Knee Replacement on Muscle Mass and Physical Function in Elderly Women With Osteoarthritis: A Randomized Controlled Trial. *Am J Phys Med Rehabil.* 2020;99(5):381-9.
112. Vroeghe DP, Wijsman CA, Broekhuizen K, de Craen AJ, van Heemst D, van der Ouderaa FJ, et al. Dose-response effects of a Web-based physical activity program on body composition and metabolic health in inactive older adults: additional analyses of a randomized controlled trial. *J Med Internet Res.* 2014;16(12):e265.
113. Schoenmaker M, de Craen AJ, de Meijer PH, Beekman M, Blauw GJ, Slagboom PE, et al. Evidence of genetic enrichment for exceptional survival using a family approach: the Leiden Longevity Study. *Eur J Hum Genet.* 2006;14(1):79-84.
114. Morley JE, Malmstrom TK, Miller DK. A simple frailty questionnaire (FRAIL) predicts outcomes in middle aged African Americans. *The journal of nutrition, health & aging.* 2012;16(7):601-8.
115. Rubenstein LZ, Harker JO, Salvà A, Guigoz Y, Vellas B. Screening for undernutrition in geriatric practice: developing the short-form mini-nutritional assessment (MNA-SF). *The journals of gerontology Series A, Biological sciences and medical sciences.* 2001;56(6):M366-72.
116. Mitchell AJ. The Mini-Mental State Examination (MMSE): update on its diagnostic accuracy and clinical utility for cognitive disorders. *Cognitive screening instruments: Springer;* 2017. p. 37-48.
117. Björk J, Bäck SE, Ebert N, Evans M, Grubb A, Hansson M, et al. GFR estimation based on standardized creatinine and cystatin C: a European multicenter analysis in older adults. *Clin Chem Lab Med.* 2018;56(3):422-35.
118. Guralnik JM, Simonsick EM, Ferrucci L, Glynn RJ, Berkman LF, Blazer DG, et al. A short physical performance battery assessing lower extremity function: association with self-reported disability and prediction of mortality and nursing home admission. *Journal of gerontology.* 1994;49(2):M85-M94.
119. Aaronson NK, Muller M, Cohen PD, Essink-Bot M-L, Fekkes M, Sanderman R, et al. Translation, validation, and norming of the Dutch language version of the SF-36 Health Survey in community and chronic disease populations. *Journal of clinical epidemiology.* 1998;51(11):1055-68.
120. Haywood KL, Garratt AM, Fitzpatrick R. Quality of life in older people: a structured review of generic self-assessed health instruments. *Qual Life Res.* 2005;14(7):1651-68.
121. Beaudart C, Biver E, Reginster JY, Rizzoli R, Rolland Y, Bautmans I, et al. Development of a self-administrated quality of life questionnaire for sarcopenia in elderly subjects: the SarQoL. *Age Ageing.* 2015;44(6):960-6.
122. Geerincx A, Alekna V, Beaudart C, Bautmans I, Cooper C, De Souza Orlandi F, et al. Standard error of measurement and smallest detectable change of the Sarcopenia Quality of Life (SarQoL) questionnaire: An analysis of subjects from 9 validation studies. *PLoS One.* 2019;14(4):e0216065.
123. Geerincx A, Bruyere O, Locquet M, Reginster JY, Beaudart C. Evaluation of the Responsiveness of the SarQoL((R)) Questionnaire, a Patient-Reported Outcome Measure Specific to Sarcopenia. *Adv Ther.* 2018;35(11):1842-58.
124. Buysse DJ, Reynolds CF, 3rd, Monk TH, Berman SR, Kupfer DJ. The Pittsburgh Sleep Quality Index: a new instrument for psychiatric practice and research. *Psychiatry research.* 1989;28(2):193-213.
125. Devine EB, Hakim Z, Green J. A systematic review of patient-reported outcome instruments measuring sleep dysfunction in adults. *Pharmacoeconomics.* 2005;23(9):889-912.
126. Frohnhofen H, Popp R, Stieglitz S, Netzer N, Danker-Hopfe H. Assessment of sleep and sleep disorders in geriatric patients. *Z Gerontol Geriatr.* 2020;53(2):100-4.
127. Mollayeva T, Thurairajah P, Burton K, Mollayeva S, Shapiro CM, Colantonio A. The Pittsburgh sleep quality index as a screening tool for sleep dysfunction in clinical and non-clinical samples: A systematic review and meta-analysis. *Sleep Med Rev.* 2016;25:52-73.
128. Revicki DA, Wood M, Wiklund I, Crawley J. Reliability and validity of the Gastrointestinal Symptom Rating Scale in patients with gastroesophageal reflux disease. *Qual Life Res.* 1998;7(1):75-83.
129. Svedlund J, Sjodin I, Dotevall G. GRSR--a clinical rating scale for gastrointestinal symptoms in patients with irritable bowel syndrome and peptic ulcer disease. *Dig Dis Sci.* 1988;33(2):129-34.
130. Bellamy N, Buchanan WW, Goldsmith CH, Campbell J, Stitt LW. Validation study of WOMAC: a health status instrument for measuring clinically important patient relevant outcomes to antirheumatic drug therapy in patients with osteoarthritis of the hip or knee. *J Rheumatol.* 1988;15(12):1833-40.

131. Heesch KC, Hill RL, Aguilar-Farias N, Van Uffelen JG, Pavey T. Validity of objective methods for measuring sedentary behaviour in older adults: a systematic review. *International Journal of Behavioral Nutrition and Physical Activity*. 2018;15(1):1-17.
132. Bammann K, Thomson NK, Albrecht BM, Buchan DS, Easton C. Generation and validation of ActiGraph GT3X+ accelerometer cut-points for assessing physical activity intensity in older adults. The OUTDOOR ACTIVE validation study. *PLoS one*. 2021;16(6):e0252615.
133. Adiels M, Olofsson S-O, Taskinen M-R, Borén J. Overproduction of very low-density lipoproteins is the hallmark of the dyslipidemia in the metabolic syndrome. *Arteriosclerosis, thrombosis, and vascular biology*. 2008;28(7):1225-36.
134. Connelly MA, Otvos JD, Shalurova I, Playford MP, Mehta NN. GlycA, a novel biomarker of systemic inflammation and cardiovascular disease risk. *Journal of translational medicine*. 2017;15(1):1-5.
135. Tirosh A, Shai I, Tekes-Manova D, Israeli E, Pereg D, Shochat T, et al. Normal fasting plasma glucose levels and type 2 diabetes in young men. *New England Journal of Medicine*. 2005;353(14):1454-62.
136. Nightingale Health [Available from: <https://nightingalehealth.com>].
137. van den Akker EB, Trompet S, Wolf JHH, Beekman M, Suchiman HED, Deelen J, et al. Predicting biological age based on the BBMRI-NL ¹H-NMR metabolomics repository. *bioRxiv*. 2019:632919.
138. D'Agostino RB, Sr., Vasan RS, Pencina MJ, Wolf PA, Cobain M, Massaro JM, et al. General cardiovascular risk profile for use in primary care: the Framingham Heart Study. *Circulation*. 2008;117(6):743-53.
139. Lewis SJ, Heaton KW. Stool form scale as a useful guide to intestinal transit time. *Scand J Gastroenterol*. 1997;32(9):920-4.
140. Ito M, Deguchi Y, Miyamori A, Matsumoto K, Kikuchi H, Kobayashi Y, et al. Effects of Administration of Galactooligosaccharides on the Human Faecal Microflora, Stool Weight and Abdominal Sensation. *Microbial Ecology in Health and Disease*. 1990;3(6):285-92.
141. Allaband C, McDonald D, Vázquez-Baeza Y, Minich JJ, Tripathi A, Brenner DA, et al. Microbiome 101: Studying, Analyzing, and Interpreting Gut Microbiome Data for Clinicians. *Clin Gastroenterol Hepatol*. 2019;17(2):218-30.
142. Organization WH. Measuring obesity - classification and description of anthropometric data. Report on a WHO consultation of the epidemiology of obesity. Warsaw 21-23 October 1987. Copenhagen: WHO, 1989. Nutrition Unit document, EUR/ICP/NUT. 1987;123.
143. Cruz-Jentoft AJ, Landi F, Schneider SM, Zúñiga C, Arai H, Boirie Y, et al. Prevalence of and interventions for sarcopenia in ageing adults: a systematic review. Report of the International Sarcopenia Initiative (EWG SOP and IWGS). *Age Ageing*. 2014;43(6):748-59.
144. Watson SL, Weeks BK, Weis LJ, Harding AT, Horan SA, Beck BR. High-Intensity Resistance and Impact Training Improves Bone Mineral Density and Physical Function in Postmenopausal Women With Osteopenia and Osteoporosis: The LIFTMOR Randomized Controlled Trial. *J Bone Miner Res*. 2018;33(2):211-20.
145. Podsiadlo D, Richardson S. The timed "Up & Go": a test of basic functional mobility for frail elderly persons. *Journal of the American Geriatrics Society*. 1991;39(2):142-8.
146. Enright PL, McBurnie MA, Bittner V, Tracy RP, McNamara R, Arnold A, et al. The 6-min walk test: a quick measure of functional status in elderly adults. *Chest*. 2003;123(2):387-98.
147. Roberts HC, Denison HJ, Martin HJ, Patel HP, Syddall H, Cooper C, et al. A review of the measurement of grip strength in clinical and epidemiological studies: towards a standardised approach. *Age Ageing*. 2011;40(4):423-9.
148. Rijk JM, Roos PR, Deckx L, van den Akker M, Buntinx F. Prognostic value of handgrip strength in people aged 60 years and older: A systematic review and meta-analysis. *Geriatr Gerontol Int*. 2016;16(1):5-20.
149. Verdijk LB, van Loon L, Meijer K, Savelberg HH. One-repetition maximum strength test represents a valid means to assess leg strength in vivo in humans. *J Sports Sci*. 2009;27(1):59-68.
150. Brzycki M. Strength testing - predicting a one-rep max from reps-to-fatigue. *Journal of physical education, recreation & dance*. 1993;64(1):88-90.
151. Lustgarten MS, Fielding RA. Assessment of analytical methods used to measure changes in body composition in the elderly and recommendations for their use in phase II clinical trials. *The journal of nutrition, health & aging*. 2011;15(5):368-75.
152. Meijboom S, van Houts-Streppel MT, Perenboom C, Siebelink E, van de Wiel AM, Geelen A, et al. Evaluation of dietary intake assessed by the Dutch self-administered web-based dietary 24-h recall tool (Compl-eat™) against interviewer-administered telephone-based 24-h recalls. *Journal of nutritional science*. 2017;6:e49.

153. Rijksinstituut voor Volksgezondheid en Milieu (RIVM). Dutch Food Composition Database (NEVO online) 2019 [Available from: <https://nevo-online.rivm.nl/>].
154. Rijksinstituut voor Volksgezondheid en Milieu (RIVM). Portiegrootte voedingsmiddelen 2019 [Available from: <https://www.rivm.nl/portiegrootte-voedingsmiddelen>].
155. van Lee L, Feskens EJ, Meijboom S, Hoofd van Huysduynen EJ, van't Veer P, de Vries JH, et al. Evaluation of a screener to assess diet quality in the Netherlands. *The British journal of nutrition*. 2016;115(3):517-26.
156. de Rijk MG, Slotegraaf AI, Brouwer-Brolsma EM, Perenboom CWM, Feskens EJM, de Vries JHM. Development and evaluation of a diet quality screener to assess adherence to the Dutch food-based dietary guidelines. *The British journal of nutrition*. 2021:1-11.
157. Looman M, Feskens EJM, Rijk dMG, Meijboom S, Briesbroek S, Temme EHM, et al. Development and evaluation of the Dutch Healthy Diet index 2015. *Public health nutrition*. 2017;20(13):2289-99.
158. Paddon-Jones D, Rasmussen BB. Dietary protein recommendations and the prevention of sarcopenia. *Current opinion in clinical nutrition and metabolic care*. 2009;12(1):86-90.
159. Ocke MC B-RE, de Boer EJ, Wilson-van den Hooven C, Etemad-Ghameslou Z, Drijvers JJMM, van Rossum CTM. Diet of community-dwelling older adults: Dutch National Food Consumption Survey Older adults 2010-2012. RIVM Rapport National Institute for Public Health and the Environment; 2013. p. 127.
160. Berridge MJ. Vitamin D, reactive oxygen species and calcium signalling in ageing and disease. *Philos Trans R Soc Lond B Biol Sci*. 2016;371(1700).
161. Rollnick S, Butler CC, Kinnersley P, Gregory J, Mash B. Motivational interviewing. *Bmj*. 2010;340.
162. Donnelly M, Jones S, Pattenden-Fail JW. DMP online: The digital curation centre's Web-based tool for creating, maintaining and exporting data management plans. *International Journal of Digital Curation*. 2010;5(1):187-93.
163. Castor Electronic Data Capture [Available from: <https://www.castoredc.com/>].
164. Cohen AA, Kennedy BK, Anglas U, Bronikowski AM, Deelen J, Dufour F, et al. Lack of consensus on an aging biology paradigm? A global survey reveals an agreement to disagree, and the need for an interdisciplinary framework. *Mechanisms of ageing and development*. 2020;191:111316.
165. Fitzgerald KN, Hodges R, Hanes D, Stack E, Cheishvili D, Szyf M, et al. Correction for: Potential reversal of epigenetic age using a diet and lifestyle intervention: a pilot randomized clinical trial. *Aging (Albany NY)*. 2022;14(14):5959.
166. Fitzgerald KN, Campbell T, Makarem S, Hodges R. Potential reversal of biological age in women following an 8-week methylation-supportive diet and lifestyle program: a case series. *Aging (Albany NY)*. 2023;15(6):1833-9.
167. The Dutch Society for Research on Ageing (DuSRA). DUSRA VOILA [cited 2023. Available from: <https://dusra.nl/voila/?lang=en>].
168. Justice JN, Ferrucci L, Newman AB, Aroda VR, Bahnson JL, Divers J, et al. A framework for selection of blood-based biomarkers for geroscience-guided clinical trials: report from the TAME Biomarkers Workgroup. *Geroscience*. 2018;40(5-6):419-36.

