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## Genome-wide gene-sleep interaction study identifies novel lipid loci in 732,564 participants

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## HIGHLIGHTS

- Examination of gene-sleep interactions could produce information on how sleep can affect disease risk.
- We performed variant-sleep interaction studies in 732,564 participants from 55 cohorts on lipid traits.
- We identified 17 loci (9 for short sleep, 8 for long sleep) that could underpin sleep-duration-associated dyslipidemia.

## GRAPHICAL ABSTRACT

**Study Rationale**

Sleep associated with cardiovascular disease, but mechanisms remain unclear

**Research Methodology**

Variant-sleep interactions studies in **732,564** participants from **55** cohorts (**87%** European ancestry) on blood lipid levels.

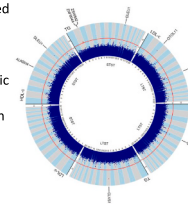


Shortest 20% > Short Total Sleep Time (STST)  
 Longest 20% > Long Total Sleep Time (LTST)

**Main results**

Multiple variants iteracted sleep sleep duration.

- Of main interest:
- *ASPH* (aspartic/succinic acid metabolism)
  - *DLEU1* (identified with both HDL-c and TG)
  - *SLC8A1* (from druggability analyses)
  - Vitamin D receptor pathway (based on bioinformatics analyses)

**Main Conclusions**

Sleep modifies the effect of some variants on lipid levels.  
 Some with biological plausibility in cardiovascular disease onset, and therefore interesting targets for follow-up studies.

## A B S T R A C T

**Background and aims:** Deviations from the population mean in sleep duration have been associated with increased risk for developing dyslipidemia and atherosclerotic cardiovascular disease, but the mechanism of effect is poorly characterized. We performed large-scale genome-wide gene-sleep interaction analyses of lipid levels to identify genetic variants underpinning the biomolecular pathways of sleep-associated lipid disturbances and to suggest possible druggable targets.

**Methods:** We collected data from 55 cohorts with a combined sample size of 732,564 participants (87% European ancestry) with data on lipid traits (high-density lipoprotein [HDL-c] and low-density lipoprotein [LDL-c] cholesterol and triglycerides [TG]). Short (STST) and long (LTST) total sleep time were defined by the extreme 20% of the age- and sex-standardized values within each cohort. Based on cohort-level summary statistics data, we performed meta-analyses for one-degree of freedom tests of interaction and two-degree of freedom joint tests of the SNP-main and -interaction effect on lipid levels.

**Results:** The one-degree of freedom variant-sleep interaction test identified 10 novel loci ( $P_{\text{int}} < 5.0 \times 10^{-9}$ ), and we additionally identify 7 loci within the two-degree of freedom analyses ( $P_{\text{joint}} < 5.0 \times 10^{-9}$  in combination with  $P_{\text{int}} < 6.6 \times 10^{-6}$ ). Multiple loci, including those mapped to *APSH* (target for aspartic and succinic acid) and *SLC8A1* showed biological plausibility and druggability potential based on literature.

<sup>1</sup> writing group.

**Conclusions:** Collectively, the 17 (9 with short and 8 with long sleep) loci provided evidence into the biomolecular mechanisms underlying sleep-associated lipid changes, including potential involvement of the vitamin D receptor pathway. Collectively, these findings may contribute developing novel interventions for treating dyslipidemia in people with sleep disturbances.

## 1. Introduction

Low levels of high-density lipoprotein cholesterol (HDL-c), and high levels of low-density lipoprotein cholesterol (LDL-c) and triglycerides (TG) are well-characterized risk factors for atherosclerotic cardiovascular disease [1–4]. High LDL-c and TG concentrations have also been shown to causally impact atherosclerotic cardiovascular disease development [5,6]. Serum lipid levels are influenced by both environmental and genetic factors [7], and large-scale efforts have identified hundreds of loci associated with increased lipid levels [8–15].

Sleep disturbances are increasingly recognized as important modifiable risk factors for various metabolic diseases including atherosclerotic cardiovascular disease and type 2 diabetes [16,17]. In 2022, sleep duration was added to the Life's Essentials by the American Heart Association, highlighting the recognition of sleep duration as an important factor in cardiovascular disease prevention [18]. However, the biological context through which sleep affects cardiovascular disease risk, needed for effective targets for interventions for cardiovascular disease prevention, is largely lacking. In epidemiological cohort studies, both short and long self-reported habitual sleep duration have been associated with adverse (atherogenic) lipid profiles [19–23], and recent Mendelian Randomization studies suggest that both short and long habitual sleep durations as potential causal risk factors for atherogenic cardiovascular disease [24–26]. However, only limited insights were so far obtained in metabolomics studies focusing on lipoprotein sub-particles potentially due to a limited sample size [23]. Examining gene-lifestyle interactions can be an important tool to identify additional genetic variants associated with the trait of interest as well as provide insights into the biomolecular mechanisms underpinning the trait-outcome association [27,28]. In previously conducted gene-lifestyle interaction projects performed within the Cohort for Heart and Aging Research in Genomic Epidemiology (CHARGE) consortium [29,30] Gene-Lifestyle Working Group [27], we identified multiple loci interacting with lifestyle exposures to lipid levels [31–34]. In particular, we performed a meta-analysis of 126,926 individuals (predominantly European-ancestry; 20 % of the participants defined as having either short or long sleep duration), which identified multiple loci associated with lipid profiles in the context of short and long sleep duration. Our results suggested that the effect of long sleep duration and short sleep duration may modify lipid profiles through distinct biological pathways [31]. In this previous study [31], we particularly focused on variants associated with lipid concentrations while accounting for variant-sleep interactions through two-degrees of freedom meta-analyses of the joint effects of the variant-lipid association and the variant-sleep interaction. At the time, we lacked sufficient statistical power to detect genome-wide significant 1df variant-sleep interaction effects on any of the studied lipid levels. The use of the two-degrees of freedom approach was able to prioritize genetic variants with potentially relevant variant-sleep interaction effects while decreasing the statistical burden for identifying variant-sleep interactions.

In recent years, more data has become available from large biobank initiatives (i.e., UK Biobank and the Million Veteran Program [35,36]). These data provide an opportunity to increase the sample size in a more diverse study population, to allow substantially improved statistical power for the detection of gene-by-sleep duration interactions on serum lipid levels as compared with our previous effort [31]. Ultimately, such efforts can improve our understanding of the biomolecular mechanisms underpinning sleep-associated lipid disturbances. Here, we conducted a new and updated multi-population gene-by-sleep duration interaction

study on serum lipid profiles in about 6 times more participants than used previously [31], which additionally creates the opportunity to perform sex-stratified analyses. In total, our new efforts were conducted using the data from 732,564 participants from 55 cohorts from five main population groups (African [AFR], East Asian [EAS], European [EUR], Hispanic/Latino [HIS] and South Asian [SAS]).

## 2. Methods

### 2.1. Overall study design

The study was designed to include cohorts that collected questionnaire-based data on habitual total sleep time and measured blood lipids levels (TG, LDL-c and/or HDL-c). Genome-wide gene  $\times$  sleep interaction analyses were performed separately by each participating study (and separately for each population group: (AFR, EAS, EUR, HIS, and SAS) following a standardized analysis protocol. Participants 18 years and older were included if they reported a total sleep time between 3 and 14 h. For studies having habitual total sleep time and lipid levels collected at multiple rounds of visits, the visit with the largest sample size was selected for analysis. Statistical analyses were performed for men and women combined as well as separately for men and women to observe potential effect modification of the variant-sleep interaction effect by sex. Data were subsequently aggregated centrally for quality control and meta-analyses. When applicable, the analysis protocol was reviewed and approved by institutional review boards. Each contributing study was approved by local medical ethics committees and each participant provided written informed consent, in line with the declaration of Helsinki. More information on the individual cohorts is presented in the **Online Supplement**, and on the overall study design and analytical pipeline in **Fig. 1**.

### 2.2. Harmonization of exposure variables

Data on habitual total sleep time were collected through questionnaires using questions like “On an average day, how long do you sleep?”. Given the previously observed U-shaped association between sleep duration and disease risk, we defined the dichotomous exposures short total sleep time (STST) and long total sleep time (LTST). To prevent misclassification of the exposure caused by the known associations between age and sex with sleep duration and to adjust for potential cohort-level differences in sleep duration reporting, we first performed cohort-specific linear regression analyses regressing sleep duration on age, sex, and age  $\times$  sex, or as indicated otherwise (**Table S2**). Based on the model-derived residuals, the 20th and 80th percentiles were used as cutoffs: STST = 1 if  $\leq$  20th percentile (otherwise “0”); LTST = 1 if  $\geq$  80th percentile (otherwise “0”).

### 2.3. Harmonization of outcome variables

We considered 3 lipid traits as outcome variables that can be easily measured in high quality in large samples: LDL-c, HDL-c and TG. For most cohorts, fasting ( $\geq$ 8 h) LDL-c and TG were used. In UK Biobank (N = 359,962 for the combined sample; 49.1 % of the total sample) participants were not asked to fast prior to blood samples, and therefore the vast majority (>90 %) had no  $\geq$  8 h fasting time. For the purpose of data harmonization between cohorts, for LDL-c and TG, analyses in UK Biobank were done separately for those meeting the fasting criteria and those who did not, and considered as separate cohorts in subsequent

meta-analyses. LDL-c was either directly assayed or derived using the Friedewald equation (the latter restricted to those with TG  $\leq$  400 mg/dL) [37]. LDL-c was corrected for the use of lipid-lowering drugs, defined as any use of a statin drug or any unspecified lipid-lowering drug after the year 1994 (when statin use became common in general clinical practice). As done previously [31,33,34], if LDL-c was directly assayed, the concentration of LDL-c was corrected by dividing the LDL-c concentration by 0.7. Otherwise (i.e. if LDL-c was derived using the Friedewald equation), we first divided the concentration of total cholesterol by 0.8 before LDL-c calculation. Due to the skewed distribution of HDL-c and TG, we natural log-transformed the concentration prior to the analyses. No transformation for LDL-c was required. All lipid levels were winsorized at 6 standard deviations from the (transformed, if applicable) mean.

#### 2.4. Individual cohort statistical data analyses

Genotype data were restricted to autosomal chromosomes, imputation quality  $R^2 \geq 0.3$  and minor allele frequency (MAF)  $\geq 0.001$  (Table S1). After data harmonization, each population-group specific cohort ran 2 regression models for 18 phenotype-exposure-sex combinations (3 phenotypes  $\times$  2 exposures  $\times$  All/Men/Women). Below E denotes the sleep exposure (STST or LTST), Y denotes the lipid trait (LDL-c, HDL-c, TG), and C denotes the vector of covariates mentioned above specific to E. Analyses were preferably conducted by each cohort using

either of the three software: LinGxEscanR v1.0 (<https://github.com/USCbiostats/LinGxEscanR>), GEM v1.4.1 (<https://github.com/large-scale-gxe-methods/GEM>), or MMAP (<https://github.com/MMAP/MMAP.github.io>) with robust standard errors (SEs) enforced [38] (Table S1). LinGxEscanR and GEM can be used in cohorts with unrelated participants, but were different in the format of required genomic input files. In cohorts with related participants, the program MMAP was used, and null model residuals (regressing lipid traits on a kinship matrix/genetic covariance matrix) were formulated as the lipid outcome.

The two regression models performed included one-degree of freedom (df) tests for examining the variant-sleep interaction effects (denoted below as ExSNP), and the two-df-joint test that simultaneously assesses the variant-main and variant-sleep interaction effects on the lipid levels (denoted below as Y) [39]. Both results are extracted using the following regression formula:  $Y = \beta_0 + \beta_E + \beta_{\text{SNP}} + \beta_{\text{E} \times \text{SNP}} + \beta_C + \epsilon$ . Covariates (denoted above as C) for which we adjusted in the regression models included population-specific principal components of the genotype matrix, cohort-specific confounders (e.g., study center), age, age [2], sex, and for their potential interaction with the exposures (notably, age  $\times$  S/LTST, age [2]  $\times$  S/LTST, and sex  $\times$  S/LTST). Finally, for a fair comparison of our results with the previous work (e.g., standard genome-wide association model for comparison [15]), we also conducted a standard marginal genetic effect model without the consideration of STST or LTST within the same study sample

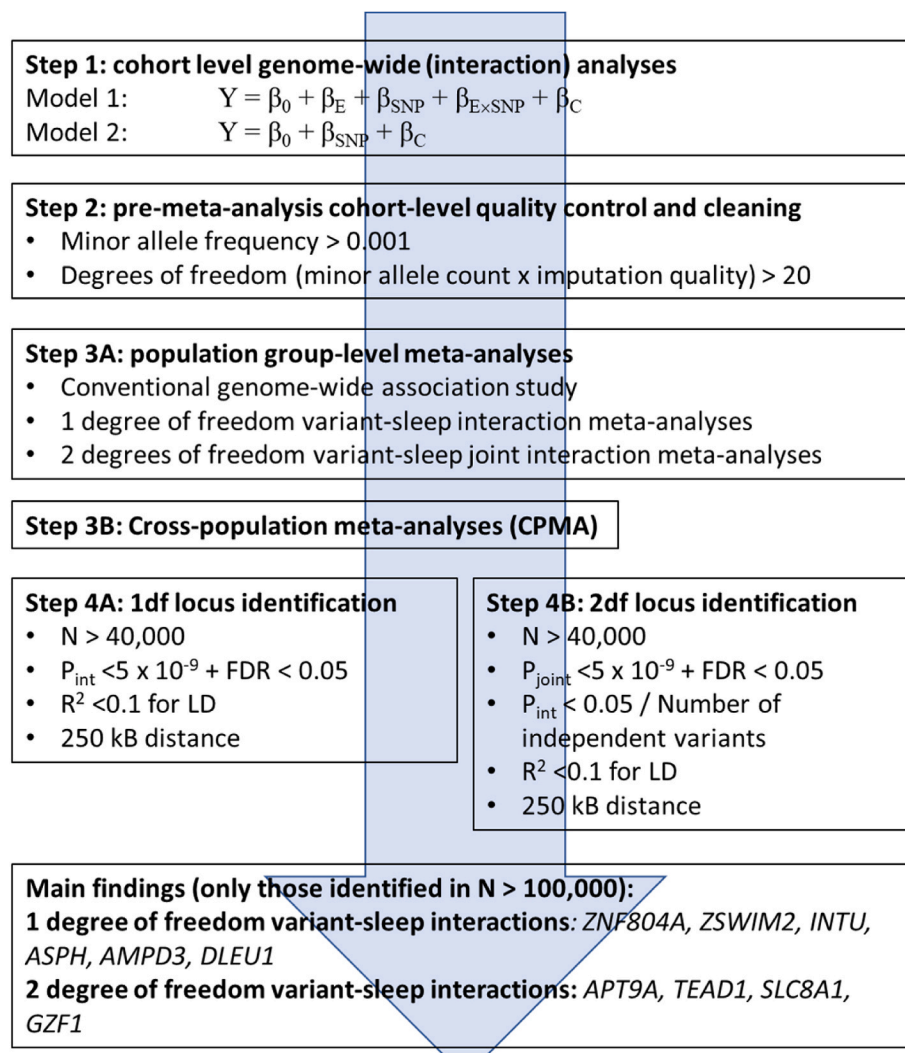


Fig. 1. Study design and analytical pipeline of the project.

with the following regression formula:  $Y = \beta_0 + \beta_{\text{SNP}}\text{SNP} + \beta_{\text{C}}\text{C} + \varepsilon$ . For these analyses, we only adjusted for age, age [2], and sex. Sex-stratified analyses were performed using similar statistical models without the adjustment for sex and sex  $\times$  S/LTST.

## 2.5. Centralized cohort-level and meta-level quality control

Cohort-level summary statistics were processed centrally. For quality control (QC), we used the EasyQC2 software ([www.genepi-regensburg.de/easyqc2](http://www.genepi-regensburg.de/easyqc2)) package in R [40]. Data were filtered for degrees of freedom  $\geq 20$  calculated as minor allele count  $\times$  imputation quality within the unexposed, the exposed, and the total sample. When required, hg37 genomic coordinates were lifted over to hg38 genomic coordinates. Allele frequency discrepancies relative to population-matched TOPMed-imputed 1000G reference panels (Trans-Omics for Precision Medicine imputed 1000Genomes) were assessed, along with genomic control (GC) lambda inflation. Next, meta-level quality control was conducted within population groups (AFR: 13 cohorts, EAS: 5 cohorts, EUR: 30 cohorts, HIS: 7 cohorts, SAS: 1 cohort), with the evaluation of the improper transformation of the outcome variables, unstable numerical computation, or alarming inflation.

## 2.6. Meta-analyses

Meta-analyses were performed for each population group separately and further combined in a cross-population meta-analyses (CPMA). This resulted in a total of 18 meta-analyses per combination of sleep exposure and lipid trait: five population groups (EUR, HIS, EAS, AFR, SAS) and CPMA, and 3 sex groups (all, women, men). Four tests were considered: the marginal genetic effect ( $B_{M2,G}$ ), the main genetic effect from the interaction model ( $B_{M1,G}$ ), the interaction effect ( $B_{M1,G \times E}$ ), and the joint main and interaction effects ( $B_{M1,G,G \times E}$ ) with cohort-level GC correction to correct for possible inflation [41]. METAL software for meta-analysis with inverse-variance weights [28] was used to combine evidence across studies for each of the four tests. CPMA was subsequently executed on the resultant population-specific METAL output results, with population-level GC correction. Due to the low numbers of participants contributing to the HIS, EAS and SAS analyses, these population-specific results were not interpreted separately, but only as a part of the CPMA. Furthermore, to minimize the identification of spurious findings as much as possible and based on careful examination of QQ plots and -log(p) plots, we only considered variants analyzed in at least 40,000 participants in the main analysis for discovery.

## 2.7. Identification of independent genomic loci

We used EasyStrata2 software in R to prioritize top loci from significant results identified in the one-df interaction and two-df joint tests [42]. We excluded variants within 1 Mb distance of the major histocompatibility complex (MHC) region. Significant variants were identified using the threshold criteria detailed below. (1) Variants with significant one-df interaction effect ( $P_{\text{int}} < 5 \times 10^{-9}$ ,  $\text{FDR} < 0.05$ ) and (2) variants with significant two-df joint effect ( $P_{\text{joint}} < 5 \times 10^{-9}$  with  $\text{FDR} < 0.05$ ) were selected as top variants. To prioritize lead variants from the two-df joint analysis with evidence for having variant-sleep interaction, we evaluated the two-df joint lead variants for one-df interactions and used a Bonferroni correction for the number of two-df joint variants identified in the respective population-specific group (CPMA, EUR, AFR) [43]. Note that the two-df joint test and the one-df interaction effect tests are correlated, so the former procedure does not offer formal statistical evidence of interaction. Nevertheless, it provides a fast and easy prioritization of variants most likely to be involved in interaction with the sleep variables. All such variants were narrowed down to loci based on a 250 kb distance. Finally, within these regions, independent loci were identified by linkage disequilibrium (LD)

$r^2$  threshold  $< 0.1$  using TOPMed-imputed 1000G reference panels. If variants were missing in the LD panels, then the most significant variant within each 500 kb region was retained. From the lead variants identified, we additionally extracted the variant information from the sex-stratified analyses to test for heterogeneity of the interaction effects by sex. The heterogeneity of the variant-sleep interaction effect between men and women was tested by performing two-sample Z-tests assuming independence, which were conducted for each interaction loci in the meta-analysis of men and women combined [44].

## 2.8. Gene mapping, functional annotation, and follow-up phenotypic annotations

For the lead variants identified, variant mapping was primarily performed using Functional Mapping and Annotation of Genome-wide Association Studies v1.6.0 (FUMA) [45], and Locuszoom (<https://my.locuszoom.org>) [46,47]. At the genomic region level, FUMA's SNP2GENE pipeline was used to annotate a comprehensive list of genes for each top locus, incorporating genomic position, chromatin interaction ( $\text{FDR} \leq 1 \times 10^{-6}$ , 250bp upstream - 500 bp downstream of the transcription startsite [TSS]), and GTExv8 eQTL evidence with the top variant or its variants in LD ( $r^2 > 0.1$  within 500 kb) [45,48]. At the variant level, PheWeb and Open Target Genetics were queried for significant trait associations ( $p < 5 \times 10^{-8}$ ) from past GWAS analyses [49, 50]. At the gene level, we explored the International Mouse Phenotyping Consortium release 19.1 (IMPC), Online Mendelian Inheritance in Man (OMIM; <https://omim.org/>), PheWeb, Phenotype-Genotype Integrator (PheGenI), Open Target Genetics, and the online drugbank for retrieving information on the genes as potential drug targets (<https://go.drugbank.com>) [49–52]. All identified mapped protein-coding genes were then queried using FUMA's GENE2FUNC pipeline to identify significant (adjusted p-value  $< 0.05$ ) pathways and traits [45].

### 2.8.1. Druggability analysis

We investigated the potential druggability of the sleep duration-lipid trait candidate interacting gene targets as previously described [53]. In short, we first used the Drug-Gene Interaction database (DGIdb; v4.2.0) to query high or medium priority sleep-lipid interacting genes to determine the potential druggability of the candidate gene targets. We annotated genes for implicated pathways and functions using the Kyoto Encyclopedia of Genes and Genomes (KEGG) database. We annotated the druggability target categories and queried all interacting drugs reported in 43 databases (BaderLabGenes, CarisMolecularIntelligence, dGene, FoundationOneGenes, GO, HingoraniCasas, HopkinsGroom, HumanProteinAtlas, IDG, MskImpact, OncoPrint, Pharos, RussLampel, Tempus, CGI, CIViC, COSMIC, CancerCommons, ChEMBLDrugs, ChEMBLInteractions, ClarityFoundationBiomarkers, ClarityFoundationClinicalTrial, DTC, DoCM, DrugBank, Ensembl, Entrez, FDA, GuideToPharmacology, JACX-CKB, MyCancerGenome, MyCancerGenomeClinicalTrial, NCI, OncoKB, PharmGKB, TALC, TEND, TTD, TdgClinicalTrial, Wikidata). We queried protein targets for available active ligands in ChEMBL. We queried gene targets in the druggable genome using the most recent druggable genome list established from the NIH Illuminating the Druggable Genome Project (<https://github.com/druggablegenome/IDGTARGETS>) available through the Pharos web platform (<https://pharos.nih.gov/targets>). We also queried FDA-approved drugs, late-stage clinical trials and disease indications in the DrugBank, ChEMBL, and [ClinicalTrials.gov](http://ClinicalTrials.gov) databases. We provided results for the top MESH and DrugBank indications and clinical trials.

## 3. Results

### 3.1. Study overview

Data from 55 cohorts including five population groups were included: AFR (13 cohorts,  $N = 48,851$  [7 %]), EAS (4 cohorts,  $N = 8097$  [1 %]), EUR (30 cohorts,  $N = 637,166$  [87 %]), HIS (7 cohorts,  $N =$

32,508 [4 %]), and SAS (1 cohort, N = 7619 [1 %]). The total sample size was 732,564 participants in the CPMA with 149,210 participants with STST and 147,603 participants with LTST. Additional information on the characteristics of each of study sample as well as the study sample combined is presented in [Tables S1-3](#).

### 3.2. Findings from the one-df variant-sleep interaction analyses

One-df interaction CPMA identified 10 loci displaying evidence for genetic associations with the lipid traits modified by either STST or LTST ( $P_{\text{int}} < 5 \times 10^{-9}$  in combination with an FDR < 0.05) ([Fig. 1](#); [Table 1](#); [Figs. S1-3](#) for  $-\log(P_{\text{int}})$  and QQ plots). Of these, we identified 5 variants for TG, 2 variants for LDL-c and 3 variants for HDL-c. These variants have not been observed before in studies on lipid levels (i.e., [15]) nor did we find evidence of potential variant main effects in the same study sample ([Table S4](#)).

Of the lead variants identified, rs14172636 (Minor Allele Frequency [MAF] = 0.0087), mapped to the *DLEU1* gene, interacted with STST in its association with both TG ( $P_{\text{int}} = 2.40 \times 10^{-16}$ ) and HDL-c ( $P_{\text{int}} = 4.10 \times 10^{-12}$ ). In more detail, among those exposed to STST, the rs14172636-C allele was associated with 0.26 units higher log-transformed TG (equivalent to an approximate additive increase of 22.9 %) and 0.132 units lower log-transformed HDL-c (equivalent to an approximate additive decrease of 14.1 %) compared to those without exposure to STST. This variant was not identified in the analyses on LDL-c, also not at a nominal significant level ( $P_{\text{int}} = 0.07$ ).

For TG, we identified the variant rs147261056 (MAF: 0.0048, mapped to *ASPH/CLVS1*) with interaction with LTST ( $P_{\text{int}} = 2.78 \times 10^{-13}$ ), and we identified the variant rs6760240 (*ZSWIM2*) with interaction with STST ( $P_{\text{int}} = 1.47 \times 10^{-9}$ ) in the CPMA analysis. For LDL-c, we identified the variants rs1847639939 (*AMPD3*), rs190975828 (*ZNF804A*), and rs162278901 (*OTOL1*) with interaction with LTST ( $P_{\text{int}} = 4.72 \times 10^{-9}$ ,  $P_{\text{int}} = 4.72 \times 10^{-9}$ , and  $P_{\text{int}} = 2.78 \times 10^{-13}$  respectively), and no variants with STST. And for HDL-c, we identified rs573762901 (*CALN1*) with interaction with LTST ( $P_{\text{int}} = 1.43 \times 10^{-10}$ ), and variant rs543672875 (*ALKBH4*) with interaction with STST ( $P_{\text{int}} = 1.51 \times 10^{-9}$ ). However, the rs162278901 (*OTOL1*), rs573762901 (*CALN1*) and rs543672875 (*ALKBH4*) variants were, although identified in a study sample of above our set threshold of 40,000 (to prevent false-positive associations to be identified as much as possible), and the rs162278901 was identified with an extremely large effect size, and should therefore be interpreted with caution, especially in combination

with their relatively low allele frequency.

One additional variant was identified in the EUR only meta-analysis. The variant rs192018195 (*INTU/SLC25A31/HSPA4L*;  $P_{\text{int}} = 4.81 \times 10^{-11}$ , MAF = 0.0151) identified in the STST analysis on TG, and was just outside the significance boundaries in the CPMA ( $P_{\text{int}} = 5.03 \times 10^{-9}$ ). Some of the more rare variants identified in these efforts were unable to be investigated further in the population-specific subgroup analyses as variants did not pass post-meta-analysis QC ([Fig. 2](#)). Of the remaining variants, we only found evidence that rs1847639939 was associated with LDL-c in the EUR sample ( $P_{\text{int}} = 1.61 \times 10^{-8}$ ), and not in the AFR meta-analysis ( $P_{\text{int}} = 0.74$ ) ([Fig. 2](#)) (see [Fig. 3](#)).

An extensive summary of the primary results, including reporting of the results in the sex-specific and population-specific analyses when passing post meta-analysis QC, are presented in [Table S4](#); additional information on the region of the identified variants is presented in regional plots presented in [Fig. S4](#), and forest plots presenting the individual cohort results are presented in [Fig. S5](#). With the exception of the lead variants mapped to *ASPH* and *DLEU1*, none were noncoding. No additional variants were identified in the sex-stratified analyses nor did we observe evidence for sex differences ( $P_{\text{sex-int}} > 0.05$ ) for variants identified with the one-df interaction test.

### 3.3. Loci identified through the two-df variant-sleep interaction meta-analyses

Additional analyses were performed to prioritize potential variant-sleep interactions identified by the two-df joint main and interaction effect meta-analyses. Using the conventional GWAS, we identified ( $P < 5 \times 10^{-9}$  and FDR < 0.05) 709 lead variants for TG, 674 lead variants for HDL-c, and 831 lead variants for LDL-c ([Table S5](#)). In the two-df CPMA ([Tables S4 and S6](#); [Fig. S6](#)), we identified ( $P_{2\text{df}} < 5 \times 10^{-9}$  and FDR < 0.05) a total of 1190 lead variants for the TG-LTST analysis (covering 371 genomic regions), 1156 lead variants for the TG-STST analysis (covering 312 genomic regions), 1185 lead variants for the HDL-c-LTST analyses (covering 362 genomic regions), 1178 lead variants for the HDL-c-STST analyses (covering 358 genomic regions), 1433 lead variants for the LDL-c-LTST analyses (covering 264 genomic regions), and 1431 lead variants for the LDL-c-STST analyses.

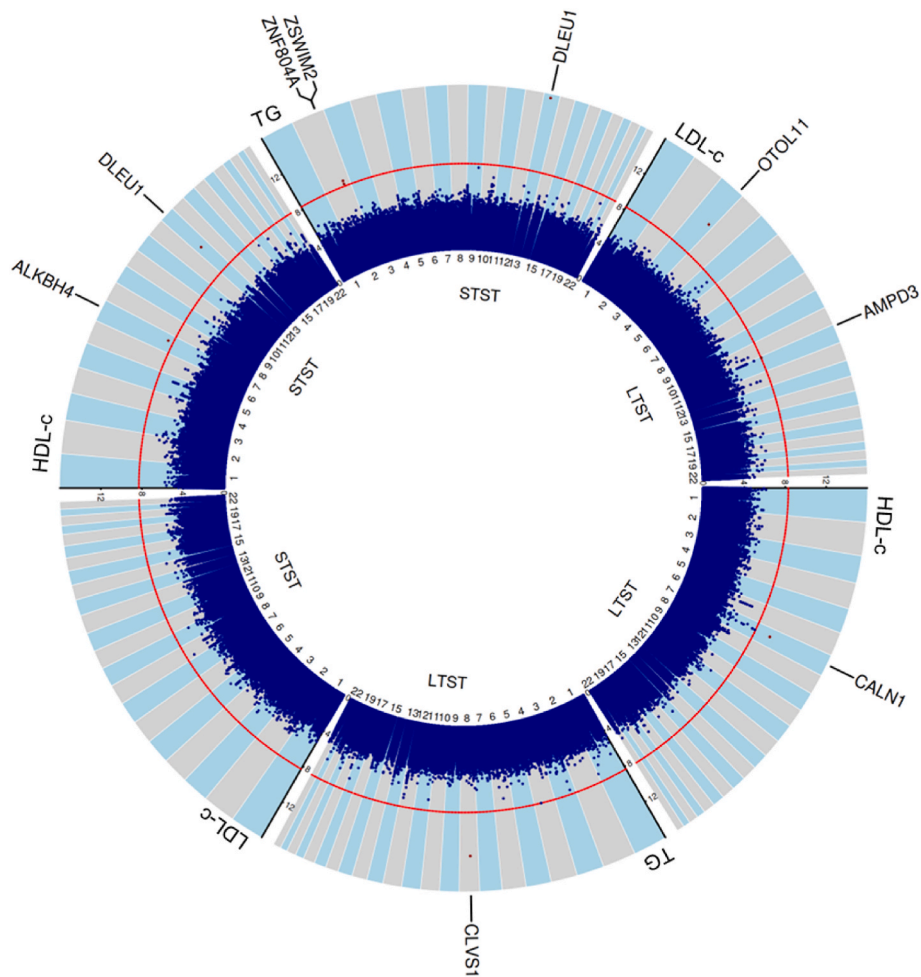
These lead variants were subsequently tested for one-df interaction. Here, we used a less stringent *P*-value cut off for one-df interactions based on the total number of lead variants identified in the CPMA sample for the three traits and two exposure groups combined ( $P_{\text{int}} <$

**Table 1**

Nine variants identified through the 1 degree of freedom interaction analyses in the meta-analyses of men and women combined.

Variant	RSid	Effect allele	Exposure	Trait	EAF	Sample Size	Sample	Mapped gene	Interaction Beta	SE	1df p-value
2:184828292: C.T	rs190975828	T	STST	TG	0.9909	557,910	CPMA	<i>ZNF804A</i>	-0.102	0.0171	2.99E-09
2:186808058: G.T	rs6760240	T	STST	TG	0.0075	188,049	CPMA	<i>ZSWIM2</i>	0.184	0.0304	1.47E-09
3:162278901: A.T	rs162278901	A	LTST	LDL-c	0.0062	41,379	CPMA	<i>OTOL1</i> [1]	25.360	3.5968	1.78E-12
4:127678773: C.G	rs192018195	C	LTST	TG	0.9849	208,087	EUR	<i>INTU, SLC25A31, HSPA4L</i>	-0.0956	0.0145	4.81E-11
7:72156448:A.G	rs573762901	A	LTST	HDL-c	0.0033	42,445	CPMA	<i>CALN1</i>	0.148	0.0230	1.43E-10
7:102460277: G.T	rs543672875	T	STST	HDL-c	0.0024	42,445	CPMA	<i>ALKBH4</i>	0.140	0.0231	1.51E-09
8:61617696:C.T	rs147261056	C	LTST	TG	0.0071	212,110	CPMA	<i>CLVS1, ASPH</i>	0.185	0.0253	2.78E-13
11:10411707: C.CT	rs1847639939	CT	LTST	LDL-c	0.4588	654,182	CPMA	<i>AMPD3</i>	0.934	0.1594	4.72E-09
13:50374420: C.T	rs14172636	T	STST	HDL-c	0.9919	196,379	CPMA	<i>DLEU1</i>	0.132	0.0190	4.10E-12
13:50374420: C.T	rs14172636	T	STST	TG	0.9913	188,528	CPMA	<i>DLEU1</i>	-0.266	0.0325	2.40E-16

Abbreviations: CPMA, Cross Population Meta-Analysis; EAF, Effect Allele Frequency; EUR, European; HDL-c, High-Density Lipoprotein Cholesterol; LDL-c, Low-Density Lipoprotein Cholesterol; SE, standard error; TG, Triglycerides. 1) Identified variant not found in FUMA or in LocusZoom, *OTOL1* gene is closest gene.



**Fig. 2.** Circular  $-\log_{10}(P_{int})$  plot of all the 6 main analyses in the cross-population meta-analysis of men and women combined. ASPH (TG and LTST) maps also at the *CLVS1* locus.

$6.60 \times 10^{-6} = 0.05/7,573$ , Bonferroni-corrected, see **Methods**). Through this process we identified seven additional genetic lead variants showing evidence for one-df interaction (**Table 2**); of these, five variants were identified for TG (one with LTST, four for STST), one variant for HDL-c (for LTST), and one variant for LDL-c (for LTST) not previously identified for lipid levels nor associated with the lipid trait in the model when not incorporating sleep duration in the same study sample (**Table 2** and **Tables S4 and S6** for detailed information).

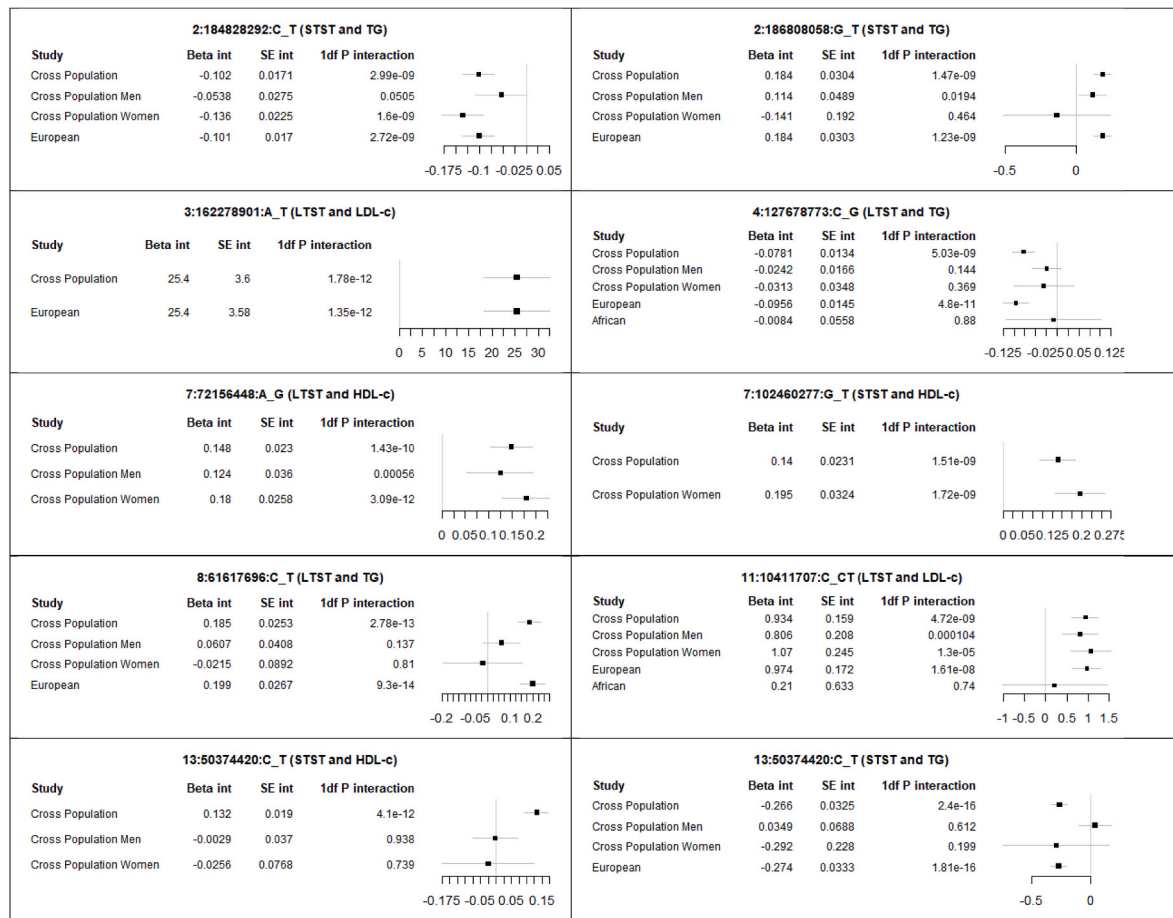
Regional plots of the one-df interaction results of these variants are presented in **Fig. S9**, and forest plots of the individual cohort findings are presented in **Fig. S10**. In particular, we identified for TG variant rs59374498 (*TEAD1/RASSF10*,  $P_{int} = 5.71 \times 10^{-8}$ ), with interaction with LTST, and identified variants rs150607032 (*ATP9A/NFATC2/SALL4*,  $P_{int} = 3.59 \times 10^{-8}$ ), rs114083565 (*RUNX1*,  $P_{int} = 8.40 \times 10^{-7}$ ), rs34771893 (*SLC8A1*,  $P_{int} = 4.12 \times 10^{-6}$ ), and rs73319497 (*GZF1/NPAB/CASTL1/CAST11/NXT1*,  $P_{int} = 4.47 \times 10^{-6}$ ) with interaction with STST. Furthermore, for HDL-c, we identified variant rs9949541 (*TCF4*,  $P_{int} = 1.92 \times 10^{-6}$ ) with interaction with LTST, with no variants being identified with interaction with STST. No evidence was observed that the interaction terms differed for men and women (sex-difference  $P_{sex-int} > 0.05$ ) (**Supplementary Table 4**). We identified no additional variants among the two-df joint findings showing evidence for one-df interaction ( $P_{int} > 1.10 \times 10^{-5}$  and  $> 1.36 \times 10^{-4}$ , respectively; **Tables S7 and S8** and **Figs. S7 and S8**).

### 3.4. Follow-up analyses

Based on the findings identified in the TG-STST analyses (the lipid-sleep combination with most identified variants in the one-df and two-df interaction analyses), and using the GTEx v8 databases, we did not observe evidence for eQTLs enrichment in any particular tissue ( $P > 0.05$ ). Some evidence (p-value = 0.01) was found for enrichment of the Vitamin D receptor pathway (based on the *SLC8A1*, *NFATC2* and *SALL4* genes; based on Wikpathways using the GENE2FUNC in FUMA [45]) (**Fig. S11**). No evidence for tissue and pathway enrichment was observed for the other loci identified in the exposure-trait combinations.

### 3.5. Druggability analyses

We first queried mapped gene targets from the different analyses using the Drug-Gene Interaction database (DGIdb), which identified seven genes annotated as clinically actionable or members of the druggable genome (**Table S9a**). Several of these gene targets are implicated in calcium signaling (*SLC25A31*, *SLC8A1*, *ASPH*), amino acid or purine metabolism (*PYROXD2*, *AMPD3*), and regulation of gene transcription (*TEAD1*, *NFATC2*, *RUNX1*). We identified seven gene targets of FDA-approved drugs evaluated in late-stage clinical trials using DrugBank and [ClinicalTrials.gov](https://clinicaltrials.gov) databases (**Table S9b**). *SLC8A1* is a target of the nutraceutical icosapent (a modified version of omega-3 fatty acid ethyl eicosapentaenoic acid (EPA)), which is used to treat patients with hypertriglyceridemia. *SLC8A1* is also a target of the small molecule



**Fig. 3.** Main results from the 1-degree of freedom interaction analyses in different subgroups. Presented results are the additive variant-interaction effects (log units for TG and HDL-c; mg/dL). Only variants passing all post meta-analysis QC steps were presented. Abbreviations: HDL-c, high-density lipoprotein cholesterol. LDL-c, low-density lipoprotein cholesterol; LTST, long total sleep time; STST, short total sleep time; TG, triglycerides.

**Table 2**

Additional 7 variants identified through the 2 degree of freedom interaction analyses after prioritization for joint effects in the meta-analyses of men and women combined.

Variant	RSid	Exposure	Effect allele	Trait	EAF	Sample Size	Sample	Mapped gene	2df joint p-value	Interaction Beta	SE	1df interaction p-value
20:51830403: A_G	rs150607032	STST	G	TG	0.0066	532,172	CPMA	<i>APT9A, NFATC2, SALL4</i>	1.77E-17	0.098	0.0178	3.59E-08
11:13058160: C_T	rs59374498	LTST	T	TG	0.9752	661,725	CPMA	<i>TEAD1, RASSF10</i>	3.34E-38	-0.0358	0.0066	5.71E-08
10:97769146: A_G	rs191757273	LTST	A	LDL-c	0.0023	52,159	CPMA	<i>PYROXD2</i>	8.51E-16	-23.9315	4.4475	7.41E-08
21:35272725: A_T	rs114083564	STST	A	TG	0.9855	43,202	CPMA	<i>RUNX1</i>	4.45E-29	0.1284	0.0261	8.40E-07
18:55378517: A_T	rs9949541	LTST	A	HDL-c	0.9748	71,290	CPMA	<i>TCF4</i>	2.64E-73	0.0478	0.01	1.92E-06
2:40094191: A_T	rs34771893	STST	A	TG	0.0055	557,910	CPMA	<i>SLC8A1</i>	1.97E-18	0.1018	0.0221	4.12E-06
20:23353740: A_G	rs73319497	STST	A	TG	0.9781	666,234	CPMA	<i>GZF1, NAPB, CASTL1, CST11, NXT1</i>	4.31E-32	-0.036	0.0078	4.47E-06

Abbreviations: CPMA, Cross Population Meta-analysis; EAF, Effect Allele Frequency; HDL-c, High-Density Lipoprotein Cholesterol; LDL-c, Low-Density Lipoprotein Cholesterol; SE, standard error; TG, Triglycerides.

inhibitor caldaret, which was investigated for preventing acute myocardial infarction and treating patients with congestive heart failure. *SLC8A1* is also a target of FDA-approved antiarrhythmic dronedarone to treat patients with atrial fibrillation or atrial flutter. We also identified *SLC25A31*, *ASPH*, and *PYROXD2* as targets of commonly

prescribed drugs: beta-blocker metoprolol, anticoagulant warfarin, and the attention deficit hyperactivity disorder (ADHD) drug methylphenidate, respectively, all drugs with indications that are frequently observed in people with sleep disorders [54–56].

#### 4. Discussion

This large-scale effort identified several variant-lipid trait associations that were modified by either STST or LTST, without overlap, including 10 loci previously-unidentified in relation to lipid levels that interact with either STST or LTST to blood lipid levels. Using joint meta-analyses, in which the main effect of the variant and the variant-sleep interaction effects are tested jointly, 7 additional genetic lead variants were identified that also showed evidence for interaction with STST or LTST. One of the variants mapped to *DLEU1* and was identified for 2 traits (HDL-c and TG). Moreover, we found distinct variants for STST and LTST interactions— a pattern we also previously reported in a smaller sample for generally higher frequency alleles [31]— suggesting that short and long sleep duration affect the lipid traits through distinct biomolecular mechanisms. Although the identified variants should be externally replicated and effect sizes for a number of the variants was relatively small, a number of the identified genes (most notably *SLC8A1*, *SLC25A31* and *ASPH*) were previously identified as targets for the prevention or treatment of cardiovascular disease and, therefore show promise as future targets for further validation and clinical translation. Collectively, these findings could, when further validated and associated to incident atherosclerotic cardiovascular disease outcomes, initiate novel directions for future clinical translation of targeting these genes to decrease atherosclerotic cardiovascular disease risk in people with disturbances in sleep duration.

The variants identified in the present study have not been associated previously with sleep duration [57], other sleep phenotypes (i.e., chronotype, insomnia symptoms or daytime napping) [58–60], or the blood lipid levels that were considered in the present study [15]. The majority of the previously unreported findings in this study are low-frequency variants, with the notable exception of rs1847639939 (mapped to *AMPD3*), that were unlikely to be found in previous studies because they were either not included in the used imputation panels or there was insufficient statistical power. The identification of mainly low-frequency variants could have also resulted in insufficient power to produce significant results in the GTEx analyses for the identification of relevant tissue-specific eQTLs. Furthermore, the observed heterogeneity in variant-sleep interaction effects of multiple of our observations between contributing studies might be influenced by cohort-specific characteristics, and additional interaction effects with other (unmeasured) factors. Of the variants identified in the one-df interaction analyses, only the lead variants identified mapped to *ASPH* and *DLEU1* were upstream/downstream transcript variants; all other variants were intronic variants. Nevertheless, these findings support the importance of gene-phenotype interaction testing in large studies to explore mechanisms and potential health preventive targets.

A number of the variants identified in the present effort are supported by biological follow-up analyses. Interestingly, we identified *DLEU1* (Deleted In Lymphocytic Leukemia 1), a gene originally identified as a possible tumour suppressor gene and often deleted in patients with B-cell chronic lymphocytic leukemia [61], in both the variant-STST analyses on HDL-c and TG (and not LDL-c). Previously, genome-wide association studies have also increasingly identified this gene with, amongst others, lipid levels [62], fatty acid [63], anthropometrics [64, 65], immune markers [66], and blood pressure [67]. Furthermore, epigenetic changes in peripheral blood in this gene have been identified in acute myocardial infarction [68]. Although *DLEU1* has not been identified with the habitual sleep variables [57–59, 69], *DLEU1* has been related to sleep apnea, which is often associated with poor sleep quality and altered sleep duration [70]. We found that the rs14172636 C-allele in *DLEU1* was associated with lower TG and higher HDL-c in individuals reporting short sleep duration, indicative of a lower atherogenic profile. Whether short sleep duration is protective of *DLEU1*-related dyslipidemia, or this variant modifies adverse effects of short sleep duration on lipid levels, cannot be sorted out. The *ASPH* gene was found to be a target for the supplemental Aspartic acid and Succinic acid. Succinate

metabolism has been hypothesized as a novel target for myocardial reperfusion injury [71], and elevated plasma succinate levels have been associated with higher levels of cardiovascular risk factors [72]. Other variants, such as the variant identified mapped to *ALKBH4*, showed no clear evidence from the literature in relation to either sleep or cardiometabolic health. For example, the *ALKBH4* gene is described as a hypoxia-responsive tumor suppressor [73], and *ZSWIM2* is predicted to be involved in the apoptosis signaling pathway. Variants as these require additional mechanistic studies to provide insights into their potential biological function in cardiometabolic health.

Our druggability analysis results suggest there are potential drug repurposing opportunities to intervene in common signaling and metabolic pathways implicated in sleep behaviour and lipid metabolism, which could help attenuate serious cardiovascular complications in high-risk patients. One of our top plausible gene targets identified in the 2-degree interaction analyses, *SLC8A1*, is targeted by nutraceutical icosapent. Furthermore, *SLC8A1* has previously been described as a target for the investigational drug caldaret. Caldaret, which acts as a cardioprotective drug modulating intracellular calcium levels, has been previously investigated to reduce infarct sizes in patients with acute myocardial infarction, although did not show positive results [74, 75]. Of interest, *SLC8A1* is affected by the renin angiotensin system [76], which is altered by different sleep conditions [77, 78]. These might present an effective strategy to reduce elevated triglycerides in patients with short sleep duration at risk for cardiovascular complications (e.g., acute myocardial infarction or atrial fibrillation). We also identified several FDA-approved compounds with decades of safe use, which could be evaluated in future preclinical or clinical studies. It is also worth noting the limitations of these predicted drug interactions, which could potentially reflect medication side effects on sleep duration and lipid traits and thus should be interpreted with caution.

We found preliminary evidence for the involvement of the Vitamin D receptor pathway in the association between STST and TG. Although vitamin D itself has not been shown to play any significant role in the onset of cardiovascular disease based on data from randomized clinical trials and Mendelian randomization [79, 80], the vitamin D receptor appears to be involved in lipid metabolism [81]. Furthermore, genetic variation in the vitamin D receptor gene (*VDR*) has been associated with cardiovascular disease [82]. Accelerated atherosclerosis was observed in *VDR* knock-out mice [83], suggesting that vitamin D receptor signaling inhibits atherosclerosis development. Finally, vitamin D levels have been reported to vary with various sleep outcomes [84], and vitamin D supplementation has been hypothesized to improve sleep [85]. Nevertheless, the role of the vitamin D receptor in the association between sleep and lipid disturbances should be explored in greater detail.

Sleep is increasingly considered as a risk factor driving cardiometabolic disease risk [24, 25]. The present study identified several variant-sleep interactions mapped to genes with biological plausibility in cardiometabolic traits, with evidence for some of them to be potential drug targets, which could therefore be of considerable interest for further validation and clinical testing. While the current evidence for a number of these targets is mixed (e.g., limited effectiveness in randomized clinical trials), it is possible, based on the findings from the present study on the different potential drug targets, that the drug response is dependent on the sleep duration. Of particular interest, although currently used lipid-modifying drugs, such as statins, are not associated with sleep disorders [86], it can be hypothesized that the effect of lipid modification is dependent on the sleep. However, to the best of our knowledge, no such studies have been performed to date investigating whether the response to drugs like statins is modified by the sleep of the patient. In the quest to precision medicine, these are additional leads to further explore in the context of sleep-associated dyslipidemia.

The present study used the largest study sample currently possible, by considering as many cohorts as possible with available data on genomics, self-reported sleep duration, and concurrent lipid levels.

Furthermore, we attempted to standardize the self-reported dichotomous sleep-exposure variables as much as possible by first taking the age- and sex-adjusted residuals of total sleep time. In addition, we excluded the HLA region from the meta-analysis given its complex structure between ancestries. Despite our efforts to increase sample size in combination with increased ancestry diversity compared with our previous effort [31], the vast majority of our study still consisted of cohorts with mainly EUR participants. It is very likely that population-specific variant-sleep interactions were missed in the meta-analyses of the non-European populations due to a lack of sufficient statistical power; furthermore, because of low statistical power, we did not present the results from the Hispanic and Asian specific meta-analyses as separate analyses. Future efforts in non-European cohorts, when more data become available, should be further expanded. Although some of the identified loci had some evidence of biological plausibility, they should be further explored in independent samples as we did not have the power to separate cohorts into discovery and independent replication analyses, and there was still considerable heterogeneity between cohorts for a number of the findings. Current initiatives, including OurFutureHealth [87], that focus on extremely large sample sizes and overrepresent participant inclusion from non-European ancestry, will likely facilitate the further discovery and validation of variant-sleep interactions, including on the X chromosome which was not considered due to methodological limitations in the present study. The present study used information on habitual sleep duration collected through self-report, which may have measurement error, possibly resulting in lower statistical power. Note that phenotypic and genetic correlations between sleep duration assessed through questionnaire and accelerometry are low to modest at most [57,69,88], which suggests phenotypes derived by these methodologies reflect different sleep aspects. Currently available sample sizes of populations with accelerometer data are unfortunately too low for use in variant-sleep interaction studies as the accelerometer data collection in UK Biobank (currently the largest) is not performed at the same time as the measurements on the blood lipid levels. Furthermore, although standard practice in many studies – including many of the main genomics analyses [31,33,34] – the correction of the use of cholesterol-lowering therapy (dividing the level by 0.7 when using a cholesterol-lowering drug) assumes all participants taking cholesterol-lowering therapy show the same reduction. This type of correction could have potentially resulted in some level of measurement error, although other correction methods (e.g., exclusion) would have resulted in forms of selection bias [89]. Finally, the present study considered sleep as a single dimension, whereas sleep is largely acknowledged to be highly dimensional and complex [90]. Indeed, joint associations between sleep duration and sleep quality have been observed in relation to atherosclerotic cardiovascular disease [26,91,92]. However, detailed data on self-reported sleep quality measures were not available in many cohorts, nor was it possible to harmonize these measures when available. Identified variants should, therefore, also be explored in independent samples, as they become available with other sleep variables.

In summary, the present study identified several novel genetic loci associated with lipid traits that were modified by self-reported short- and long total sleep time. The findings yield novel insights into the biology underpinning the observed association between sleep duration and atherosclerotic cardiovascular disease. Collectively, these findings may contribute to developing novel interventions for treating dyslipidemia in people with sleep disturbances, and therefore lead to new opportunities for the prevention of atherosclerotic cardiovascular disease.

#### Authors' contributions

Conception and design: RN, WW, HW, DCR, SR, DvH. Writing group: RN, WW, PN, HW, MRB, ARB, HC, CLM, TWW, DCR, SR, DvH. All

authors were responsible for cohort-level data analyses, critical commenting on the initial versions of the manuscript, and final approval of the manuscript submitted to Genome Medicine.

#### Ethics approval and consent to participate

When applicable, the analysis protocol was reviewed and approved by institutional review boards. Each contributing study was approved by local medical ethics committees and each participant provided written informed consent, in line with the declaration of Helsinki.

#### Consent for publication

All authors approved the final version of the manuscript for publication.

#### Availability of data and materials

Summary-level data of the genomics analyses is open upon request. Access to individual-level data from UK Biobank is arranged to approval of an analysis protocol and payment of a data access fee.

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#### Declaration of competing interest

HJG has received travel grants and speakers honoraria from Neuraxpharm, Servier, Indorsia and Janssen Cilag. LMR is a consultant for the TOPMed Administrative Coordinating Center (through Westat). TDS is co-founder and shareholder of ZOE Ltd. HC receives consulting fees from Character Biosciences. All other co-authors declare to have no conflicts of interest.

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#### Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.atherosclerosis.2026.120603>.

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