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Mapping the maze: advancing atrial fibrillation models and therapies

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Chapter 1

The background of the page is a dark, starry night sky. In the center, there is a bright, glowing star with a soft, white-to-yellowish aura. To the right of this star, there is a faint, elongated nebula or galaxy structure. The sky is filled with numerous smaller, distant stars of varying brightness.



General introduction

General introduction

Atrial fibrillation in the clinic

Atrial fibrillation (AF) is the most common cardiac arrhythmia and an important contributor of morbidity, mortality and healthcare costs in the western world. The current prevalence of AF is estimated to be 2-4% in the adult population (1) and is expected to more than double in the coming decades as the result of population ageing (2, 3). It was recently estimated that 1 in 3 Europeans will develop AF in their lifetimes (4, 5). Individuals developing AF can (initially) remain asymptomatic, however symptoms including palpitations, dyspnoea and fatigue are common, frequently resulting in impaired quality of life (6). Additionally, the presence of AF is associated with serious complications, including a 4- to 5-fold increased risk of stroke (7, 8), increased risk of heart failure (9, 10) and 1.5- to 3.5-fold increase in all-cause mortality (5, 11, 12). Overall, it is estimated that up to 2.6% of total healthcare costs in European countries can be attributed to the management of AF and its complications (13, 14).

Presence of AF is characterised by rapid and chaotic electrical activation of the atria and is the result of a complex interaction of triggers, perpetuators and an atrial substrate (15). The most important risk factor for developing AF is age, however other factors include hypertension, heart failure, diabetes mellitus, coronary artery disease, obesity, smoking, valve disease, chronic kidney disease and more (6, 16). The pathophysiology behind the manifestation of AF remains a heavily studied topic, as the understanding of the multiform mechanisms leading to its initiation, maintenance and termination remains incomplete (17). Generally, initiation of AF is the result of rapid electrical activation that originates from the pulmonary veins, although such ectopic activity can also arise from other locations within the atria (15). This rapid electrical activation can result in the induction of re-entrant electrical activity, sustaining the arrhythmia. The stability and longevity of this re-entrant activity is in turn influenced by the severity of the atrial substrate. In clinical practise, AF is classified based on its duration, being paroxysmal when lasting <7 days, persistent when continuously present >7 days, and long-standing persistent when continuing >1 year (6). In tandem with the chronicity, the underlying mechanism driving AF generally changes, shifting from an incidental trigger-based tachyarrhythmia, to stable re-entry sustained by heavy structural and electrical remodelling of the atria (15). The progression of AF can be driven by progressive atrial remodelling due to (untreated) comorbidities, however the

presence of AF itself can also lead to remodelling in a phenomenon called “AF begets AF” (18).

The current treatment strategy for individuals with AF, based on the AF-CARE pathway, comprises a combination of stroke risk reduction, symptom management and comorbidity treatment optimization (6). Stroke prevention in the form of anticoagulation therapy is applied in individuals with an increased risk of stroke based on additional risk factors besides AF, commonly calculated using the CHA₂DS₂-VA score (6, 19). This has shown to be effective in reducing thromboembolic events, however comes with an increased risk of (major) bleeding (20, 21). In addition to stroke risk reduction, pharmacological “rate control” is frequently applied to limit the ventricular rate, with the aim of preventing tachycardia-induced cardiomyopathy and to reduce symptoms. When symptoms persist, “rhythm control” can be pursued to restore and maintain sinus rhythm, typically performed in conjunction with rate control and anticoagulation therapy. Acute restoration of sinus rhythm through electrical or pharmacological cardioversion is very effective, although limited by the need for a brief hospital admission each time and it having minimal effect on preventing AF recurrences (22, 23). Antiarrhythmic drug therapy can be used to maintain sinus rhythm, however, the efficacy is limited and is associated with significant side effects (24). Electrical isolation of the pulmonary veins, which is possible through various endovascular and surgical techniques, has shown to be effective in reducing the occurrence of AF (25), and has become an important modality in the treatment of symptomatic AF. Overall, the efficacy of these rhythm control therapies is higher in paroxysmal AF and have the benefit of slowing progression in this population (26). Additionally, there is a growing body of evidence supporting early rhythm control, suggesting a lower risk of adverse cardiovascular outcomes. Unfortunately, these treatment strategies are not effective in all patients and a large portion of patients will still progress to the persistent or long-standing persistent form of AF where restoration of sinus rhythm becomes more challenging.

Ablation of symptomatic long-standing persistent atrial fibrillation

Patients who have progressed to long-standing persistent AF (LSPAF) are particularly difficult to treat due to the extensive arrhythmogenic substrate present. Where in paroxysmal AF the majority of triggers can be localised to the pulmonary veins, triggers in LSPAF are also frequently located in other parts of the left and right atrium, particularly in the left atrial posterior wall and left atrial

appendage (15). As a result, minimally invasive catheter ablation of the pulmonary veins alone has shown poor outcomes in LSPAF, resulting in around 20-24% AF freedom after a single intervention and 41-47% AF freedom after multiple (up to 5) interventions at 5-year follow-up (27, 28). Performing more extensive endocardial ablation in this population, by for instance targeting complex fractionated electrograms and/or adding linear lesions to isolate the left atrial posterior wall or left atrial appendage, has thus far not yielded improved outcomes (29). This likely results from lesion transmural and contiguity issues associated with the current point-by-point endocardial ablation techniques and technologies applied for linear lesions (30).

Conversely, surgical ablation using the Cox-maze IV technique, widely considered the gold standard for AF ablation, has shown to be effective in the LSPAF population with rates of AF freedom as high as 67% at 7-year follow-up (31). This technique, introduced in its current form in 2002, comprises an extensive set of lesions in the left and right atrium, including amputation of the left atrial appendage. The aim of this lesion set is prohibiting AF to establish and perpetuate, while allowing impulses from the sinus node to reach the atrioventricular node (32). The advantages of this surgical approach include better access to both the endo- and epicardial side of the atrial myocardium, elimination of the blood cooling effect and use of linear ablation tools and clamps, all aiding better lesion transmural and contiguity. Despite the high efficacy, the applicability of the stand-alone Cox-maze IV is severely limited by its invasiveness, required expertise and need for cardiopulmonary bypass. As a result, the Cox-maze IV has a more prominent place as a concomitant procedure during cardiac surgery of patients with AF (33).

For stand-alone surgical ablation to serve as an alternative to catheter ablation in LSPAF, minimal invasiveness, ease of performing the procedure and high efficacy are important. With the advent of thoracoscopic surgery techniques, minimally invasive thoracoscopic ablation on the beating heart has become commonplace over the past decade. This has resulted in a wide array of minimally invasive thoracoscopic ablation devices and techniques applied in clinical practice to treat AF. The efficacy of these thoracoscopic ablation procedures, however, has (thus far) mostly been studied in patients with paroxysmal and early-persistent AF and not in the more difficult to treat LSPAF patients. As a result, literature on thoracoscopic ablation efficacy in LSPAF is sparse and has mostly been limited to short- to medium-term follow-up studies (34-36). In addition to the variability in devices and techniques in these studies, the modality of rhythm follow-up differs frequently, ranging from

incidental electrocardiography up to continuous rhythm analysis through implantable loop recorders. To provide more clarity on the applicability of thoracoscopic ablation as alternative to catheter ablation in LSPAF patients, more (preferably long-term) comparative data is needed on the different ablation techniques and rhythm follow-up strategies.

Preclinical research and models of atrial fibrillation

On the other side of the spectrum, development of novel therapeutics effectively targeting the underlying pathophysiology of AF has been arduous. In the last decade, no new antiarrhythmic drugs have been approved, and only a few compounds are currently in the pipeline (37, 38). One of the factors limiting preclinical safety and efficacy studies has been the lack of sufficient numbers of well-differentiated human atrial cardiomyocytes. Together with the (associated) absence of clinically relevant human-based *in vitro* models of AF, preclinical research into AF mechanisms and novel therapeutic strategies has been mainly limited to animal studies. Given the principal differences in (patho)physiology between humans and animals, these new therapeutics frequently fail due to limited efficacy or strong adverse effects in patients (39).

Replacing the animal-derived cells in the *in vitro* models of AF with human cells, aiming to increase the translational value, has been difficult. Despite good accessibility of the human heart during routine cardiac surgery, (atrial) cardiomyocytes isolated from surgical waste material rapidly dedifferentiate in culture and vary in terms of quality (40). Attempts to prolong the lifespan of these cells by saving the extracellular matrix and culturing cardiac tissue slices are promising (41), although still only applied on small scale, given that they are most commonly derived from scarcely available explanted human hearts.

Since the early 2010s, an alternative source of human atrial cardiomyocytes is provided by human pluripotent stem cells (hPSCs), where the development of differentiation protocols utilizing retinoic acid have allowed the creation of cardiomyocytes with atrial features (42, 43). However, in spite of certain attractive features and unique applications in personalized disease modelling, current hPSC-derived atrial cardiomyocytes (hPSC-AMs) are associated with laborious workflows, high phenotypic variation, and overall immaturity. Attempts to create *in vitro* models of human AF using hPSC-AMs have been reported (44, 45), however they were unable to represent the rapid activation frequencies of 6-8 Hz (46-48) seen during human AF.

The creation of permanent cardiac cell lines from primary material to create a standardised and unlimited source of cardiomyocytes has been pursued on several occasions. Currently, three cell lines are commercially available and frequently cited in literature (49). These include the H9c2 cell line, generated in 1976 through selective serial passage of embryonic BDIX rat heart tissue (50). These cells have a myoblast origin and display properties of both skeletal muscle and, to some extent, cardiac muscle (51). Two decades later, the HL-1 cell line was established (52), originating from adult mice where the expression of simian virus 40 large T antigen (SV40-LT) was targeted to atrial cardiomyocytes via the atrial natriuretic factor promoter. Later, the AC16 line was generated through the fusion of primary human ventricular cardiomyocytes with SV40-transformed human fibroblasts (53). Despite their widespread use and popularity in literature, these three cell lines show limited resemblance to mature cardiac cells (49), likely due to the fact that continuing proliferation inhibits differentiation in most cell types (54). To address this limitation, efforts to generate conditionally immortalized cardiomyocyte cell lines that can switch between proliferative and differentiated phenotypes have been undertaken (55-59). Among these conditionally immortalized lines, the iAM-1 line (59) demonstrated the most effective transition between a proliferative state and a differentiated phenotype. This cell line, functioning through doxycycline-induced expression of SV40-LT in neonatal rat atrial cardiomyocytes, could even be applied to model atrial arrhythmias in vitro. Although the aforementioned lines are of great value in reducing the need for animals in cardiovascular research, their differences with respect to human (patho)physiology remain significant, reducing their clinical relevance as previously mentioned. Successfully applying this technology to human atrial cardiomyocytes could lead to more accurate models for human heart diseases and reduce reliance on animal testing, ultimately accelerating the development of effective therapies, not just for atrial fibrillation, but for cardiovascular conditions in general.

Aims and outline of this thesis

Part One. Outcomes of thoracoscopic ablation in long-standing persistent atrial fibrillation

The first part of this thesis, is aimed at investigating the efficacy of thoracoscopic ablation in the LSPAF population as an intermediate option between minimally invasive (but limited efficacious) catheter ablation and highly effective (but highly invasive) surgical ablation. In chapter 2, the efficacy of the two most common ablation tools used for isolating the posterior left atrium during thoracoscopic ablation are compared on perioperative performance and short-term follow-up. In chapter 3, the long-term efficacy of thoracoscopic ablation in LSPAF is studied, including the role of catheter ablation in patients with recurrences. Also, recurrence rates characterized by incidental 24h-Holter monitoring and continuous rhythm monitoring using implantable loop recorders are compared.

Part Two. Development of clinically relevant human in vitro models of atrial fibrillation

The second part of this thesis is aimed at investigating the improvement of access to well-differentiated human atrial cardiomyocytes and to develop a clinically relevant in vitro model of AF. In chapter 4, the electrophysiological properties of excised human atrial tissue are studied, and the feasibility of creating culturable atrial tissue slices is assessed. In chapter 5, the conditional immortalization of human atrial cardiomyocytes is explored, including analysis of the success rate, extensive characterisation of the phenotype of these cells and their applicability for modelling AF in vitro.

Finally, in chapter 6 the main conclusions and their implications, as well as future perspectives, are discussed.

References

1. Benjamin EJ, Muntner P, Alonso A, Bittencourt MS, Callaway CW, Carson AP, et al. Heart Disease and Stroke Statistics-2019 Update A Report From the American Heart Association. *Circulation*. 2019;139(10):E56-E528.
2. Colilla S, Crow A, Petkun W, Singer DE, Simon T, Liu XC. Estimates of Current and Future Incidence and Prevalence of Atrial Fibrillation in the US Adult Population. *Am J Cardiol*. 2013;112(8):1142-7.
3. Krijthe BP, Kunst A, Benjamin EJ, Lip GYH, Franco OH, Hofman A, et al. Projections on the number of individuals with atrial fibrillation in the European Union, from 2000 to 2060. *Eur Heart J*. 2013;34(35):2746-51.
4. Staerk L, Wang BQ, Preis SR, Larson MG, Lubitz SA, Ellinor PT, et al. Lifetime risk of atrial fibrillation according to optimal, borderline, or elevated levels of risk factors: cohort study based on longitudinal data from the Framingham Heart Study. *Bmj-Brit Med J*. 2018;361.
5. Magnussen C, Niiranen TJ, Ojeda FM, Gianfagna F, Blankenberg S, Njolstad I, et al. Sex Differences and Similarities in Atrial Fibrillation Epidemiology, Risk Factors, and Mortality in Community Cohorts Results From the BiomarCaRE Consortium (Biomarker for Cardiovascular Risk Assessment in Europe). *Circulation*. 2017;136(17):1588-+.
6. Van Gelder IC, Rienstra M, Bunting KV, Casado-Arroyo R, Caso V, Crijns HJGM et al. 2024 ESC Guidelines for the management of atrial fibrillation developed in collaboration with the European Association for Cardio-Thoracic Surgery (EACTS). *Eur Heart J*. 2024;45(36):3314-3414
7. Wolf PA, Dawber TR, Thomas HE, Kannel WB. Epidemiologic Assessment of Chronic Atrial-Fibrillation and Risk of Stroke - Framingham Study. *Neurology*. 1978;28(10):973-7.
8. Andrew NE, Thrift AG, Cadilhac DA. The Prevalence, Impact and Economic Implications of Atrial Fibrillation in Stroke: What Progress Has Been Made? *Neuroepidemiology*. 2013;40(4):227-39.
9. Anter E, Jessup M, Callans DJ. Atrial Fibrillation and Heart Failure Treatment Considerations for a Dual Epidemic. *Circulation*. 2009;119(18):2516-25.
10. Kotecha D, Piccini JP. Atrial fibrillation in heart failure: what should we do? *Eur Heart J*. 2015;36(46):3250-U58.
11. Andersson T, Magnuson A, Bryngelsson IL, Frobert O, Henriksson KM, Edvardsson N, et al. All-cause mortality in 272 186 patients hospitalized with incident atrial fibrillation 1995-2008: a Swedish nationwide long-term case-control study. *Eur Heart J*. 2013;34(14):1061-7.

12. Benjamin EJ, Wolf PA, D'Agostino RB, Silbershatz H, Kannel WB, Levy D. Impact of atrial fibrillation on the risk of death. *Circulation*. 1998;98(10):946-52.
13. Ringborg A, Nieuwlaat R, Lindgren P, Jonsson B, Fidan D, Maggioni AP, et al. Costs of atrial fibrillation in five European countries: results from the Euro Heart Survey on atrial fibrillation. *Europace*. 2008;10(4):403-11.
14. Cotte FE, Chaize G, Gaudin AF, Samson A, Vainchtock A, Fauchier L. Burden of stroke and other cardiovascular complications in patients with atrial fibrillation hospitalized in France. *Europace*. 2016;18(4):501-7.
15. Lip GYH, Fauchier L, Freedman SB, Van Gelder I, Natale A, Gianni C, et al. Atrial fibrillation. *Nat Rev Dis Primers*. 2016;2.
16. Chung MK, Refaat M, Shen WK, Kuttyifa V, Cha YM, Di Biase L, et al. Atrial Fibrillation JACC Council Perspectives. *J Am Coll Cardiol*. 2020;75(14):1689-713.
17. Nattel S, Heijman J, Zhou LP, Dobrev D. Molecular Basis of Atrial Fibrillation Pathophysiology and Therapy A Translational Perspective. *Circ Res*. 2020;127(1):51-72.
18. Wijffels MCEF, Kirchhof CJHJ, Dorland R, Allesie MA. Atrial-Fibrillation Begets Atrial-Fibrillation - a Study in Awake Chronically Instrumented Goats. *Circulation*. 1995;92(7):1954-68.
19. Lip GYH, Nieuwlaat R, Pisters R, Lane DA, Crijns HJGM. Refining Clinical Risk Stratification for Predicting Stroke and Thromboembolism in Atrial Fibrillation Using a Novel Risk Factor-Based Approach The Euro Heart Survey on Atrial Fibrillation. *Chest*. 2010;137(2):263-72.
20. McBride R. Stroke Prevention in Atrial-Fibrillation Study - Final Results. *Circulation*. 1991;84(2):527-39.
21. Connolly SJ, Ezekowitz MD, Yusuf S, Eikelboom J, Oldgren J, Parekh A, et al. Dabigatran versus Warfarin in Patients with Atrial Fibrillation. *New Engl J Med*. 2009;361(12):1139-51.
22. Fetsch T, Bauer P, Engberding R, Koch HP, LUKL J, et al. Prevention of atrial fibrillation after cardioversion: results of the PAFAC trial. *Eur Heart J*. 2004;25(16):1385-94.
23. Tieleman RG, Van Gelder IC, Crijns HJ, De Kam PJ, Van Den Berg MP, et al. Early recurrences of atrial fibrillation after electrical cardioversion: a result of fibrillation-induced electrical remodeling of the atria? *J Am Coll Cardiol*. 1998;31(1):167-73.
24. Zimetbaum P. Antiarrhythmic drug therapy for atrial fibrillation. *Circulation*. 2012;125(2):381-9.
25. Asad ZU, Yousif A, Khan MS, Al-Khatib SM, Stavrakis S. Catheter Ablation Versus Medical Therapy for Atrial Fibrillation A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Circ-Arrhythmia Elec*. 2019;12(9).

- 26.** Camm AJ, Naccarelli GV, Mittal S, Crijns H, Hohnloser SH, Ma CS, et al. The Increasing Role of Rhythm Control in Patients With Atrial Fibrillation: JACC State-of-the-Art Review. *J Am Coll Cardiol.* 2022;79(19):1932-48.
- 27.** Tilz RR, Rillig A, Thum AM, Arya A, Wohlmuth P, Metzner A, et al. Catheter ablation of long-standing persistent atrial fibrillation: 5-year outcomes of the Hamburg Sequential Ablation Strategy. *J Am Coll Cardiol.* 2012;60(19):1921-9.
- 28.** Teunissen C, Kassenberg W, van der Heijden JF, Hassink RJ, van Driel VJHM, et al. Five-year efficacy of pulmonary vein antrum isolation as a primary ablation strategy for atrial fibrillation: a single-centre cohort study. *Europace.* 2016;18(9):1335-42.
- 29.** Scott PA, Silberbauer J, Murgatroyd FD. The impact of adjunctive complex fractionated atrial electrogram ablation and linear lesions on outcomes in persistent atrial fibrillation: a meta-analysis. *Europace.* 2016;18(3):359-67.
- 30.** McCarthy PM, Cox JL, Kislitsina ON, Kruse J, Churyla A, Malaisrie SC, et al. Surgery and Catheter Ablation for Atrial Fibrillation: History, Current Practice, and Future Directions. *J Clin Med.* 2022;11(1).
- 31.** Lapenna E, De Bonis M, Giambuzzi I, Del Forno B, Ruggeri S, Cireddu M, et al. Long-term Outcomes of Stand-Alone Maze IV for Persistent or Long-standing Persistent Atrial Fibrillation. *Ann Thorac Surg.* 2020;109(1):124-31.
- 32.** Ruaengsri C, Schill MR, Khiabani AJ, Schuessler RB, Melby SJ, Damiano RJ, Jr. The Cox-maze IV procedure in its second decade: still the gold standard? *Eur J Cardiothorac Surg.* 2018;53(suppl_1):i19-i25.
- 33.** Ruaengsri C, Schill MR, Khiabani AJ, Schuessler RB, Melby SJ, Damiano RJ. The Cox-maze IV procedure in its second decade: still the gold standard? *Eur J Cardio-Thorac.* 2018;53:19-25.
- 34.** Haldar S, Khan HR, Boyalla V, Kralj-Hans I, Jones S, Lord J, et al. Catheter ablation vs. thoracoscopic surgical ablation in long-standing persistent atrial fibrillation: CASA-AF randomized controlled trial. *Eur Heart J.* 2020;41(47):4471-80.
- 35.** Ohtsuka T, Nonaka T, Hisagi M, Ninomiya M, Stewart JR. En Bloc Left Pulmonary Vein and Appendage Isolation in Thoracoscopic Surgery for Atrial Fibrillation. *Annals of Thoracic Surgery.* 2018;106(5):1340-8.
- 36.** van Laar C, Bentala M, Weimar T, Doll N, Swaans MJ, Molhoek SG, et al. Thoracoscopic ablation for the treatment of atrial fibrillation: a systematic outcome analysis of a multicentre cohort. *Europace.* 2019;21(6):893-9.
- 37.** Geng M, Lin A, Nguyen TP. Revisiting Antiarrhythmic Drug Therapy for Atrial Fibrillation: Reviewing Lessons Learned and Redefining Therapeutic Paradigms. *Front Pharmacol.* 2020;11.

- 38.** Peyronnet R, Ravens U. Atria-selective antiarrhythmic drugs in need of alliance partners. *Pharmacol Res.* 2019;145.
- 39.** Robinson NB, Krieger K, Khan FM, Huffman W, Chang M, Naik A, et al. The current state of animal models in research: A review. *Int J Surg.* 2019;72:9-13.
- 40.** Voigt N, Pearman CM, Dobrev D, Dibb KM. Methods for isolating atrial cells from large mammals and humans. *J Mol Cell Cardiol.* 2015;86:187-98.
- 41.** Amez JH, de Groot NMS, Langmuur SJJ, el Azzouzi HE, Tiggeloven VPC, van Rooij MMM, et al. Biomimetic cultivation of atrial tissue slices as novel platform for in-vitro atrial arrhythmia studies. *Sci Rep-Uk.* 2023;13(1).
- 42.** Devalla HD, Schwach V, Ford JW, Milnes JT, El-Haou S, Jackson C, et al. Atrial-like cardiomyocytes from human pluripotent stem cells are a robust preclinical model for assessing atrial-selective pharmacology. *EMBO Mol Med.* 2015;7(4):394-410.
- 43.** Zhang Q, Jiang J, Han P, Yuan Q, Zhang J, Zhang X, et al. Direct differentiation of atrial and ventricular myocytes from human embryonic stem cells by alternating retinoid signals. *Cell Res.* 2011;21(4):579-87.
- 44.** Goldfracht I, Protze S, Shiti A, Setter N, Gruber A, Shaheen N, et al. Generating ring-shaped engineered heart tissues from ventricular and atrial human pluripotent stem cell-derived cardiomyocytes. *Nat Commun.* 2020;11(1):75.
- 45.** Laksman Z, Wauchop M, Lin E, Protze S, Lee J, Yang W, et al. Modeling Atrial Fibrillation using Human Embryonic Stem Cell-Derived Atrial Tissue. *Sci Rep.* 2017;7(1):5268.
- 46.** Sanders P, Berenfeld O, Hocini M, Jais P, Vaidyanathan R, Hsu LF, et al. Spectral analysis identifies sites of high-frequency activity maintaining atrial fibrillation in humans. *Circulation.* 2005;112(6):789-97.
- 47.** Schuessler RB, Kay MW, Melby SJ, Branham BH, Boineau JP, Damiano RJ, Jr. Spatial and temporal stability of the dominant frequency of activation in human atrial fibrillation. *J Electrocardiol.* 2006;39(4 Suppl):S7-12.
- 48.** Yoshida K, Ulfarsson M, Oral H, Crawford T, Good E, Jongnarangsin K, et al. Left atrial pressure and dominant frequency of atrial fibrillation in humans. *Heart Rhythm.* 2011;8(2):181-7.
- 49.** Onodi Z, Visnovitz T, Kiss B, Hambalko S, Koncz A, Agg B, et al. Systematic transcriptomic and phenotypic characterization of human and murine cardiac myocyte cell lines and primary cardiomyocytes reveals serious limitations and low resemblances to adult cardiac phenotype. *J Mol Cell Cardiol.* 2022;165:19-30.
- 50.** Kimes BW, Brandt BL. Properties of a clonal muscle cell line from rat heart. *Exp Cell Res.* 1976;98(2):367-81.

- 51.** Hescheler J, Meyer R, Plant S, Krautwurst D, Rosenthal W, Schultz G. Morphological, biochemical, and electrophysiological characterization of a clonal cell (H9c2) line from rat heart. *Circ Res.* 1991;69(6):1476-86.
- 52.** Claycomb WC, Lanson NA, Jr., Stallworth BS, Egeland DB, Delcarpio JB et al. HL-1 cells: a cardiac muscle cell line that contracts and retains phenotypic characteristics of the adult cardiomyocyte. *Proc Natl Acad Sci U S A.* 1998;95(6):2979-84.
- 53.** Davidson MM, Nesti C, Palenzuela L, Walker WF, Hernandez E, Protas L, et al. Novel cell lines derived from adult human ventricular cardiomyocytes. *J Mol Cell Cardiol.* 2005;39(1):133-47.
- 54.** Ruijtenberg S, van den Heuvel S. Coordinating cell proliferation and differentiation: Antagonism between cell cycle regulators and cell type-specific gene expression. *Cell Cycle.* 2016;15(2):196-212.
- 55.** Jahn L, Sadoshima J, Greene A, Parker C, Morgan KG, Izumo S. Conditional differentiation of heart- and smooth muscle-derived cells transformed by a temperature-sensitive mutant of SV40 T antigen. *J Cell Sci.* 1996;109 (Pt 2):397-407.
- 56.** Goldman BI, Amin KM, Kubo H, Singhal A, Wurzel J. Human myocardial cell lines generated with SV40 temperature-sensitive mutant tsA58. *In Vitro Cell Dev Biol Anim.* 2006;42(10):324-31.
- 57.** Rybkin, II, Markham DW, Yan Z, Bassel-Duby R, Williams RS, Olson EN. Conditional expression of SV40 T-antigen in mouse cardiomyocytes facilitates an inducible switch from proliferation to differentiation. *J Biol Chem.* 2003;278(18):15927-34.
- 58.** Zhang Y, Nuglozeh E, Toure F, Schmidt AM, Vunjak-Novakovic G. Controllable expansion of primary cardiomyocytes by reversible immortalization. *Hum Gene Ther.* 2009;20(12):1687-96.
- 59.** Liu J, Volkers L, Jangsangthong W, Bart CI, Engels MC, Zhou G, et al. Generation and primary characterization of iAM-1, a versatile new line of conditionally immortalized atrial myocytes with preserved cardiomyogenic differentiation capacity. *Cardiovasc Res.* 2018;114(14):1848-59.

