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Towards implementation of the tumour-stroma ratio in colorectal cancer

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Chapter 1

General introduction
and thesis outline

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Colorectal cancer has remained the third highest cancer type in incidence and second cause of cancer-related deaths for years all over the world [1-3]. The cornerstone of international treatment guidelines encompasses disease extent, defined by the tumour-node-metastasis (TNM) classification [4, 5], and other assessments regarding expected patient-related outcomes, such as survival and treatment benefit [6-9]. However, recurrence rates as well as heterogeneity in survival outcomes of colorectal patients with similar TNM-stages are illustrative of a currently suboptimal therapy selection, leading to high occurrences of overtreatment and undertreatment. Moreover, even a survival paradox is observed, where patients with lower, less extensive disease and theoretically thus more favourable, TNM-stages, have a conversely worse survival outcome [6, 8, 10, 11]. Therefore, improving the prediction of patient-related outcomes and thus therapy selection is imperative.

In an attempt to attain this improvement, many histopathological parameters and biological markers (biomarkers) have been discovered and developed [12, 13]. Microsatellite instability (MSI) status, tumour deposits and tumour budding were for instance relatively recently implemented in routine pathology diagnostics [6, 14-16]. Whereas these mainly pertain to the tumour epithelial compartment, i.e. the neoplastic cells, this traditional tumour epithelium-centred approach remains therefore insufficient. It is pivotal to expand our current focus to include parameters that enable more accurate predictions and a personalized treatment strategy. Hence, the paradigm is shifting, with the surrounding tumour microenvironment increasingly emerging as an important modulator and no longer deemed an innocuous substance.

Tumour microenvironment

The past decades, the complex crosstalk between the tumour epithelial compartment and tumour microenvironment has piqued the interest of researchers worldwide [17-20]. Although the exact mechanism is yet to be elucidated, it has become apparent that this dynamic entity, predominantly comprising immune cells, extracellular matrix, vasculature and fibroblasts with various forms and functions, modulates tumour behaviour [21-23]. The tumour stroma therein, especially an abundance of stroma, has furthermore been observed to be of detrimental influence and enhances aggressive characteristics, such as tumour invasion and therapy resistance [22, 24-27]. The tumour-stroma ratio (TSR), i.e. the proportion of the tumour epithelial compartment compared to the tumour stromal compartment expressed in percentages and the main focus of this thesis, captures this effect specifically.

Tumour-stroma ratio

Since the first publication of this histological parameter in 2007 by our research group [28], the TSR has been protocolized [29, 30], promoted [31, 32] and proven [33-36], as well by other researchers [24, 37-39] and in multiple cancer types [40-42]. Iteratively, stroma-high (>50% intratumoural stroma percentages) tumours indeed lead to significantly worse patient-related outcomes than their stroma-low ($\leq$ 50% intratumoural stroma) counterparts. A proposal to implement the TSR as additional biomarker in international guidelines and pathology diagnostics was thus presented to the TNM evaluation committee of the Union for International Cancer Control (UICC) and College of American Pathologists (CAP). While the advocated-to instances endorsed the potential of the TSR and despite this overwhelming evidence, however, a prospective study was still considered necessary. In a complying response, subsequent to an international consensus with support of the European Society of Pathology (ESP) and establishment of an official TSR-scoring E-learning [43], the multicentre UNITED study (Uniform Noting for International application of the Tumour-stroma ratio as Easy Diagnostic tool) was therefore initiated [44]. This thesis presents the results of the UNITED study in **Chapter 2**, validating the prognostic value of the TSR as biomarker on disease-free survival in colon cancer patients (DFS) [45].

Colon cancer vs. rectum cancer

Albeit often collectively termed, rectal cancer is a different entity than colon cancer [9, 46]. Treatment of rectal cancer varies thus from that of colon cancer, with a large focus on organ-sparing approaches [9, 47]. ‘Neoadjuvant therapy’ (Latin: *neo* = new, *adjuvare* = to help), first used to describe therapy intended for too extensive primary tumours for surgery [48], is a pillar in treatment in rectal cancer currently. Where it was initially implemented to enhance local control in addition to the total mesorectal excision (TME) [49, 50], research has shown that more intricate regimens can induce a complete response, potentially rendering surgery redundant [47, 51, 52]. However, this response is prone to variety and a large heterogeneity is seen between clinical and pathological responses (cCR and pCR, respectively) [53-56]. Upfront selection of patients standing to benefit from neoadjuvant therapy, and patients who should potentially not undergo a ‘watch-and-wait’ strategy [57], is thus essential.

The TSR has been shown previously to not only be a prognostic biomarker, but also to predict response to treatment [58-60]. Since neoadjuvant therapy alters the tumour microenvironment [61-63], resulting in e.g. fibrosis, this leads to inconclusive TSR scores after treatment. Scoring the TSR in the biopsies has previously proven to be representative for the TSR score of the primary tumour and, moreover, a predictive biomarker [64, 65], although large, multicentre studies evaluating the potential of the TSR in

predicting neoadjuvant response in rectal cancer are lacking [66-68]. In **Chapter 3** therefore, we integrate two large, clinical trials (RAPIDO [53] and PROCTOR-SCRIPT [69]) with our LUMC cohort and aimed to validate the predictive value of the TSR in rectal cancer.

Stromal fibroblasts and lymph node metastasis

Initial cancer diagnosis and staging, especially lymph nodes, continues to be challenging with the current CT and MRI scanning per protocol [6, 70]. PET scanning, a modality where anatomical and physiological functions are combined by using radioactive fluor-labelled fluorodeoxyglucose ([18-F]-FDG), is not commonly used in routine work-up and has pitfalls as well [6, 71, 72]. Cancer-specific tracers are increasingly researched, for which, targets pertain to the tumour stroma as well. Despite the large heterogeneity tumours are prone to, the tumour stroma generally contains quiescent fibroblasts, which can be activated through many pathways, giving rise to cancer-associated fibroblasts (CAFs) [73, 74]. Due to their tumour promoting functions, these CAFs are considered intriguing targets, and as such, a universal and high potential CAF marker has been identified: fibroblast activation protein (FAP) [75-77]. Coupled with a radioactive label, the fibroblast activation protein-inhibitor (FAPI) is exponentially subject to research in radiological imaging as tracer with FAPI-PET/CT scans, aiming to ascertain improvement to current imaging protocols [78-81]. Ultimately, tumour-targeted tracers can even be applied in a theragnostic sense [82-85]. Much research is done currently on the FAPI-tracer, although correlation studies with the golden standard, i.e. pathology, are lacking [86, 87].

As the importance of the TSR in lymph nodes metastases has been proven previously [88, 89], first, a theoretical framework for lymph node metastasis, specifically the stromal assessment, was established (**Chapter 4**). Here, we describe the tumour stroma in lymph node metastases in detail and expand on our TSR measurements, to support future correlation research between FAPI uptake on PET/CT scanning with the pathological lymph node assessments [90]. Moreover, as the CAFs had been identified as crucial modulators in the tumour behaviour of CRC, in **Chapter 5** we performed a large histological comparison between the traditional haematoxylin and eosin (H&E)-stained slides and immunohistochemical staining for FAP in CRC, biopsies and resected material with primary tumour and lymph nodes. As FAP constitutes an attractive marker for improved diagnosis through FAPI imaging, and translational studies are scarce, we aimed to describe the patterns to form a biological background for future clinical studies and ascertain correlation to the TSR.

Digitalisation and artificial intelligence

Historically, as the breakthrough that shaped the modern pathology from the mid-nineteenth century onward, the microscope has been the golden standard modality and trusted technique for the specialty [91, 92]. Glass slides containing tissue are visualized and analysed through Bright-field light microscopy in routine diagnostics, however, digitalisation is gaining interest in pathology [93]. Slides are increasingly scanned, resulting in digital whole slide images (WSIs). Initially intended to decrease workload and enable remote working for pathologists, these WSIs can also be analysed by computers, for instance by using artificial intelligence algorithms [94-96]. Often criticized and characterised as a ‘black box’, **Chapter 6** attempts to elucidate this phenomenon, where, in contrast to training on time-consuming annotations like in supervised deep learning to automate e.g. TSR scoring [97-99], we trained a state-of-art self-supervised deep learning model on unannotated WSIs [100]. Small image patches (tiles) from these WSIs were grouped based on similar histological patterns by this model, forming histomorphological phenotype clusters (HPCs). We subsequently analysed the HPCs and correlated these to patient-related outcomes, providing novel insights in potential histological aspects with the emphasis on tumour stroma, and advocate their clinical relevance.

Summary and discussion

The presented research in this thesis is ultimately summarized and followed by a discussion where future perspectives of research and more specifically, concrete and necessary steps are described towards the imperative implementation of the TSR as additional biomarker in guidelines and pathology routine diagnostics (**Chapter 7**). This thesis is concluded by the summary and discussion translated in Dutch and other appendices.

References

1. Sung, H., et al., Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin*, 2021.
2. Siegel, R.L., et al., Cancer statistics, 2022. *CA Cancer J Clin*, 2022. 72(1): p. 7-33.
3. Bray, F., et al., Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin*, 2024. 74(3): p. 229-263.
4. Brierley, J.D., Gospodarowicz, M.K., Wittekind, C., The TNM Classification of Malignant Tumours. 8 ed. 2016: Wiley Blackwell.
5. Weiser, M.R., AJCC 8th Edition: Colorectal Cancer. *Ann Surg Oncol*, 2018. 25(6): p. 1454-1455.
6. Argiles, G., et al., Localised colon cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol*, 2020. 31(10): p. 1291-1305.
7. Cervantes, A., et al., Metastatic colorectal cancer: ESMO Clinical Practice Guideline for diagnosis, treatment and follow-up. *Ann Oncol*, 2023. 34(1): p. 10-32.
8. Baxter, N.N., et al., Adjuvant Therapy for Stage II Colon Cancer: ASCO Guideline Update. *J Clin Oncol*, 2022. 40(8): p. 892-910.
9. Glynne-Jones, R., et al., Rectal cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol*, 2018. 29(Suppl 4): p. iv263.
10. Kim, H.S., et al., Clinicopathological and biomolecular characteristics of stage IIB/IIC and stage IIIA colon cancer: Insight into the survival paradox. *J Surg Oncol*, 2019. 120(3): p. 423-430.
11. Li, H., et al., Re-Evaluation of the Survival Paradox Between Stage IIB/IIC and Stage IIIA Colon Cancer. *Front Oncol*, 2020. 10: p. 595107.
12. Chuang, J.P., et al., Comprehensive Review of Biomarkers for the Treatment of Locally Advanced Colon Cancer. *Cells*, 2022. 11(23).
13. Chen, K., et al., Pathological Features and Prognostication in Colorectal Cancer. *Curr Oncol*, 2021. 28(6): p. 5356-5383.
14. Lugli, A., et al., Recommendations for reporting tumour budding in colorectal cancer based on the International Tumour Budding Consensus Conference (ITBCC) 2016. *Mod Pathol*, 2017. 30(9): p. 1299-1311.
15. Roth, A.D., et al., Integrated analysis of molecular and clinical prognostic factors in stage II/III colon cancer. *J Natl Cancer Inst*, 2012. 104(21): p. 1635-46.
16. Ueno, H., et al., Optimal colorectal cancer staging criteria in TNM classification. *J Clin Oncol*, 2012. 30(13): p. 1519-26.
17. Fidler, I.J., Seed and soil revisited: contribution of the organ microenvironment to cancer metastasis. *Surg Oncol Clin N Am*, 2001. 10(2): p. 257-69, vii-viii.
18. Ouahoud, S., et al., Bidirectional tumour/stroma crosstalk promotes metastasis in mesenchymal colorectal cancer. *Oncogene*, 2020. 39(12): p. 2453-2466.
19. Sudhakar, M., H. Vignesh, and K.N. Natarajan, Crosstalk between tumour and microenvironment: Insights from spatial transcriptomics. *Adv Cancer Res*, 2024. 163: p. 187-222.
20. Werb, Z. and P. Lu, The Role of Stroma in Tumour Development. *Cancer J*, 2015. 21(4): p. 250-3.
21. Balkwill, F.R., M. Capasso, and T. Hagemann, The tumour microenvironment at a glance. *J Cell Sci*, 2012. 125(Pt 23): p. 5591-6.
22. Bremnes, R.M., et al., The role of tumour stroma in cancer progression and prognosis: emphasis on carcinoma-associated fibroblasts and non-small cell lung cancer. *J Thorac Oncol*, 2011. 6(1): p. 209-17.
23. Sugai, T., et al., Microenvironmental markers are correlated with lymph node metastasis in invasive submucosal colorectal cancer. *Histopathology*, 2021. 79(4): p. 584-598.
24. Park, J.H., et al., The relationship between tumour stroma percentage, the tumour microenvironment and survival in patients with primary operable colorectal cancer. *Ann Oncol*, 2014. 25(3): p. 644-651.
25. Tsujino, T., et al., Stromal myofibroblasts predict disease recurrence for colorectal cancer. *Clin Cancer Res*, 2007. 13(7): p. 2082-90.
26. Zhuyan, J., et al., Critical steps to tumour metastasis: alterations of tumour microenvironment and extracellular matrix in the formation of pre-metastatic and metastatic niche. *Cell Biosci*, 2020. 10: p. 89.
27. Strous, M.T.A., et al., A high tumour-stroma ratio (TSR) in colon tumours and its metastatic lymph nodes predicts poor cancer-free survival and chemo resistance. *Clin Transl Oncol*, 2022. 24(6): p. 1047-1058.
28. Mesker, W.E., et al., The carcinoma-stromal ratio of colon carcinoma is an independent factor for survival compared to lymph node status and tumour stage. *Cell Oncol*, 2007. 29(5): p. 387-98.
29. van Pelt, G.W., et al., Scoring the tumour-stroma ratio in colon cancer: procedure and recommendations. *Virchows Arch*, 2018. 473(4): p. 405-412.
30. van Pelt, G.W., et al., The tumour-stroma ratio in colon cancer: the biological role and its prognostic impact. *Histopathology*, 2018. 73(2): p. 197-206.
31. Smit, M.A. and W.E. Mesker, A Stromal Solution, in *The Pathologist*. 2018, Texere Publishing.
32. Mesker, W. Uniform Noting for International application of the Tumour-stroma ratio as Easy Diagnostic Tool - website. 2018 10-10-2024]; Available from: <http://watchstroma.com/>.
33. Mesker, W.E., et al., Presence of a high amount of stroma and downregulation of SMAD4 predict for worse survival for stage I-II colon cancer patients. *Cell Oncol*, 2009. 31(3): p. 169-78.
34. Huijbers, A., et al., The value of additional bevacizumab in patients with high-risk stroma-high colon cancer. A study within the QUASAR2 trial, an open-label randomized phase 3 trial. *J Surg Oncol*, 2018. 117(5): p. 1043-1048.

35. Huijbers, A., et al., The proportion of tumour-stroma as a strong prognosticator for stage II and III colon cancer patients: validation in the VICTOR trial. *Ann Oncol*, 2013. 24(1): p. 179-85.
36. Zunder, S.M., et al., Predictive potential of tumour-stroma ratio on benefit from adjuvant bevacizumab in high-risk stage II and stage III colon cancer. *Br J Cancer*, 2018. 119(2): p. 164-169.
37. Strous, M.T., et al., Node-negative colon cancer: histological, molecular, and stromal features predicting disease recurrence. *Molecular Medicine*, 2023. 29(1): p. 77.
38. Park, J.H., et al., Tumour invasiveness, the local and systemic environment and the basis of staging systems in colorectal cancer. *Br J Cancer*, 2017. 116(11): p. 1444-1450.
39. Sullivan, L., et al., Tumour Stroma Ratio and Its Significance in Locally Advanced Colorectal Cancer. *Curr Oncol*, 2022. 29(5): p. 3232-3241.
40. Smit, M.A., et al., The prognostic value of the tumour-stroma ratio in squamous cell lung cancer, a cohort study. *Cancer Treat Res Commun*, 2020. 25: p. 100247.
41. Vangangel, K.M.H., et al., The prognostic value of the tumour-stroma ratio is most discriminative in patients with grade III or triple-negative breast cancer. *Int J Cancer*, 2020. 146(8): p. 2296-2304.
42. Hagenaars, S.C., et al., Standardization of the tumour-stroma ratio scoring method for breast cancer research. *Breast Cancer Res Treat*, 2022. 193(3): p. 545-553.
43. Smit, M.A., et al., e-Learning for Instruction and to Improve Reproducibility of Scoring Tumour-Stroma Ratio in Colon Carcinoma: Performance and Reproducibility Assessment in the UNITED Study. *JMIR Form Res*, 2021. 5(3): p. e19408.
44. Smit, M., et al., Uniform Noting for International Application of the Tumour-Stroma Ratio as an Easy Diagnostic Tool: Protocol for a Multicenter Prospective Cohort Study. *JMIR Res Protoc*, 2019. 8(6): p. e13464.
45. Polack, M., et al., Results from the UNITED study: a multicenter study validating the prognostic effect of the tumour-stroma ratio in colon cancer. *ESMO Open*, 2024. 9(4): p. 102988.
46. Wei, E.K., et al., Comparison of risk factors for colon and rectal cancer. *International journal of cancer*, 2004. 108(3): p. 433-442.
47. Beets, G.L., et al., A new paradigm for rectal cancer: Organ preservation: Introducing the International Watch & Wait Database (IWWD). *European Journal of Surgical Oncology (EJSO)*, 2015. 41(12): p. 1562-1564.
48. Wood, W.C., Neoadjuvant chemotherapy, in *Adjuvant Therapy of Breast Cancer*, I.C. Henderson, Editor. 1992, Springer US: Boston, MA. p. 279-291.
49. Peeters, K.C., et al., The TME trial after a median follow-up of 6 years: increased local control but no survival benefit in irradiated patients with resectable rectal carcinoma. *Ann Surg*, 2007. 246(5): p. 693-701.
50. Kapiteijn, E., et al., Preoperative radiotherapy combined with total mesorectal excision for resectable rectal cancer. *New England Journal of Medicine*, 2001. 345(9): p. 638-646.
51. Garcia-Aguilar, J., et al., Organ Preservation in Patients With Rectal Adenocarcinoma Treated With Total Neoadjuvant Therapy. *J Clin Oncol*, 2022. 40(23): p. 2546-2556.
52. Custers, P.A., et al., Long-term Quality of Life and Functional Outcome of Patients With Rectal Cancer Following a Watch-and-Wait Approach. *JAMA Surgery*, 2023. 158(5): p. e230146-e230146.
53. Bahadoer, R.R., et al., Short-course radiotherapy followed by chemotherapy before total mesorectal excision (TME) versus preoperative chemoradiotherapy, TME, and optional adjuvant chemotherapy in locally advanced rectal cancer (RAPIDO): a randomised, open-label, phase 3 trial. *The Lancet Oncology*, 2021. 22(1): p. 29-42.
54. Lambregts, D.M.J., T.N. Boellaard, and R.G.H. Beets-Tan, Response evaluation after neoadjuvant treatment for rectal cancer using modern MR imaging: a pictorial review. *Insights Imaging*, 2019. 10(1): p. 15.
55. Temmink, S.J., et al., Complete response rates in rectal cancer: Temporal changes over a decade in a population-based nationwide cohort. *European Journal of Surgical Oncology*, 2023. 49(11): p. 106991.
56. Nahas, S.C., et al., Diagnostic performance of magnetic resonance to assess treatment response after neoadjuvant therapy in patients with locally advanced rectal cancer. *Abdom Radiol (NY)*, 2019. 44(11): p. 3632-3640.
57. Temmink, S.J.D., et al., Watch and wait after neoadjuvant treatment in rectal cancer: comparison of outcomes in patients with and without a complete response at first reassessment in the International Watch & Wait Database (IWWD). *Br J Surg*, 2023. 110(6): p. 676-684.
58. van Pelt, G.W., et al., The value of tumour-stroma ratio as predictor of pathologic response after neoadjuvant chemoradiotherapy in esophageal cancer. *Clin Transl Radiat Oncol*, 2020. 20: p. 39-44.
59. Hagenaars, S.C., et al., Tumour-stroma ratio is associated with Miller-Payne score and pathological response to neoadjuvant chemotherapy in HER2-negative early breast cancer. *Int J Cancer*, 2021.
60. Hansen, T.F., et al., Tumour-stroma ratio predicts recurrence in patients with colon cancer treated with neoadjuvant chemotherapy. *Acta Oncol*, 2018. 57(4): p. 528-533.
61. Nagtegaal, I., et al., Morphological changes in tumour type after radiotherapy are accompanied by changes in gene expression profile but not in clinical behaviour. *J Pathol*, 2004. 204(2): p. 183-92.
62. Stone, H.B., et al., Effects of radiation on normal tissue: consequences and mechanisms. *Lancet Oncol*, 2003. 4(9): p. 529-36.
63. Miller, K.E., J.L. Ostrowski, and C.M. Quinn, Effects of chemotherapy on breast cancer tissue. *Histopathology*, 1997. 30(4): p. 397-8.
64. Park, J.H., et al., Preoperative, biopsy-based assessment of the tumour microenvironment in patients with primary operable colorectal cancer. *J Pathol Clin Res*, 2020. 6(1): p. 30-39.
65. Courrech Staal, E.F., et al., Reproducibility and validation of tumour stroma ratio scoring on oesophageal adenocarcinoma biopsies. *Eur J Cancer*, 2011. 47(3): p. 375-82.
66. Scheer, R., et al., Tumour-stroma ratio as prognostic factor for survival in rectal adenocarcinoma: A retrospective cohort study. *World J Gastrointest Oncol*, 2017. 9(12): p. 466-474.

67. Strous, M.T.A., et al., Tumour-stroma ratio to predict pathological response to neo-adjuvant treatment in rectal cancer. *Surg Oncol*, 2022. 45: p. 101862.
68. Zhu, Y., et al., Prognostic Value of Tumour-Stroma Ratio in Rectal Cancer: A Systematic Review and Meta-analysis. *Front Oncol*, 2021. 11: p. 685570.
69. Breugom, A.J., et al., Adjuvant chemotherapy for rectal cancer patients treated with preoperative (chemo)radiotherapy and total mesorectal excision: a Dutch Colorectal Cancer Group (DCCG) randomized phase III trial. *Ann Oncol*, 2015. 26(4): p. 696-701.
70. Brouwer, N.P.M., et al., Clinical lymph node staging in colorectal cancer; a flip of the coin? *Eur J Surg Oncol*, 2018. 44(8): p. 1241-1246.
71. Godefroy, J., et al., Perceptual omission errors in Positron emission tomography/Computed tomography reporting. *Q J Nucl Med Mol Imaging*, 2021.
72. Lu, Y.Y., et al., A systematic review and meta-analysis of pretherapeutic lymph node staging of colorectal cancer by 18F-FDG PET or PET/CT. *Nucl Med Commun*, 2012. 33(11): p. 1127-33.
73. Chen, Y., K.M. McAndrews, and R. Kalluri, Clinical and therapeutic relevance of cancer-associated fibroblasts. *Nat Rev Clin Oncol*, 2021. 18(12): p. 792-804.
74. Kobayashi, H., et al., Cancer-associated fibroblasts in gastrointestinal cancer. *Nat Rev Gastroenterol Hepatol*, 2019. 16(5): p. 282-295.
75. Henrich, L.M., et al., The Impact of Cancer-Associated Fibroblasts on the Biology and Progression of Colorectal Carcinomas. *Genes (Basel)*, 2024. 15(2).
76. Liu, F., et al., Fibroblast activation protein overexpression and clinical implications in solid tumours: a meta-analysis. *PLoS One*, 2015. 10(3): p. e0116683.
77. Janani, M., et al., Association of future cancer metastases with fibroblast activation protein- α : a systematic review and meta-analysis. *Front Oncol*, 2024. 14: p. 1339050.
78. Kratochwil, C., et al., (68)Ga-FAPI PET/CT: Tracer Uptake in 28 Different Kinds of Cancer. *J Nucl Med*, 2019. 60(6): p. 801-805.
79. Mori, Y., et al., FAPI PET: Fibroblast Activation Protein Inhibitor Use in Oncologic and Nononcologic Disease. *Radiology*, 2023. 306(2): p. e220749.
80. Dendl, K., et al., The Role of Fibroblast Activation Protein Ligands in Oncologic PET Imaging. *PET Clin*, 2021. 16(3): p. 341-351.
81. Giesel, F.L., et al., (68)Ga-FAPI PET/CT: Biodistribution and Preliminary Dosimetry Estimate of 2 DOTA-Containing FAP-Targeting Agents in Patients with Various Cancers. *J Nucl Med*, 2019. 60(3): p. 386-392.
82. Parisi, A., et al., What Is Known about Theragnostic Strategies in Colorectal Cancer. *Biomedicines*, 2021. 9(2).
83. Lindner, T., et al., Targeting of activated fibroblasts for imaging and therapy. *EJNMMI Radiopharm Chem*, 2019. 4(1): p. 16.
84. Langbein, T., W.A. Weber, and M. Eiber, Future of Theranostics: An Outlook on Precision Oncology in Nuclear Medicine. *J Nucl Med*, 2019. 60(Suppl 2): p. 13s-19s.
85. Mori, Y., et al., Fibroblast Activation Protein Inhibitor Theranostics: Early Clinical Translation. *PET Clin*, 2023. 18(3): p. 419-428.
86. Mona, C.E., et al., Correlation of (68)Ga-FAPI-46 PET Biodistribution with FAP Expression by Immunohistochemistry in Patients with Solid Cancers: Interim Analysis of a Prospective Translational Exploratory Study. *J Nucl Med*, 2022. 63(7): p. 1021-1026.
87. Strating, E., et al., Fibroblast activation protein identifies Consensus Molecular Subtype 4 in colorectal cancer and allows its detection by (68)Ga-FAPI-PET imaging. *Br J Cancer*, 2022. 127(1): p. 145-155.
88. van Pelt, G.W., et al., Stroma-high lymph node involvement predicts poor survival more accurately for patients with stage III colon cancer. *J Med Surg Pathol*, 2016. 1(02).
89. Vangangelt, K.M.H., et al., The prognostic value of tumour-stroma ratio in tumour-positive axillary lymph nodes of breast cancer patients. *Int J Cancer*, 2018. 143(12): p. 3194-3200.
90. Polack, M., et al., Characteristics of tumour stroma in regional lymph node metastases in colorectal cancer patients: a theoretical framework for future diagnostic imaging with FAPI PET/CT. *Clin Transl Oncol*, 2022.
91. van den Tweel, J.G. and C.R. Taylor, A brief history of pathology: Preface to a forthcoming series that highlights milestones in the evolution of pathology as a discipline. *Virchows Arch*, 2010. 457(1): p. 3-10.
92. Loughrey, M.B., et al., Dataset for Pathology Reporting of Colorectal Cancer: Recommendations From the International Collaboration on Cancer Reporting (ICCR). *Ann Surg*, 2022. 275(3): p. e549-e561.
93. Pallua, J.D., et al., The future of pathology is digital. *Pathol Res Pract*, 2020. 216(9): p. 153040.
94. Hinton, G.E., 20 - CONNECTIONIST LEARNING PROCEDURES11This chapter appeared in Volume 40 of Artificial Intelligence in 1989, reprinted with permission of North-Holland Publishing. It is a revised version of Technical Report CMU-CS-87-115, which has the same title and was prepared in June 1987 while the author was at Carnegie Mellon University. The research was supported by contract N00014-86-K-00167 from the Office of Naval Research and by grant IST-8520359 from the National Science Foundation, in Machine Learning, Y. Kodratoff and R.S. Michalski, Editors. 1990, Morgan Kaufmann: San Francisco (CA). p. 555-610.
95. Hamet, P. and J. Tremblay, Artificial intelligence in medicine. *Metabolism*, 2017. 69s: p. S36-s40.
96. Chen, R.J., et al., Towards a general-purpose foundation model for computational pathology. *Nature Medicine*, 2024. 30(3): p. 850-862.
97. Firmbach, D., et al., Tumour-Stroma Ratio in Colorectal Cancer-Comparison between Human Estimation and Automated Assessment. *Cancers (Basel)*, 2023. 15(10).
98. Geessink, O.G.F., et al., Computer aided quantification of intratumoural stroma yields an independent prognosticator in rectal cancer. *Cell Oncol (Dordr)*, 2019. 42(3): p. 331-341.

99. Smit, M.A., et al., Deep learning based tumour-stroma ratio scoring in colon cancer correlates with microscopic assessment. *J Pathol Inform*, 2023. 14: p. 100191.
100. Zbontar, J., Barlow Twins: Self-Supervised Learning via Redundancy Reduction. 2021.