



Universiteit
Leiden
The Netherlands

The aging B cell landscape in atherosclerosis

Mol, J. de

Citation

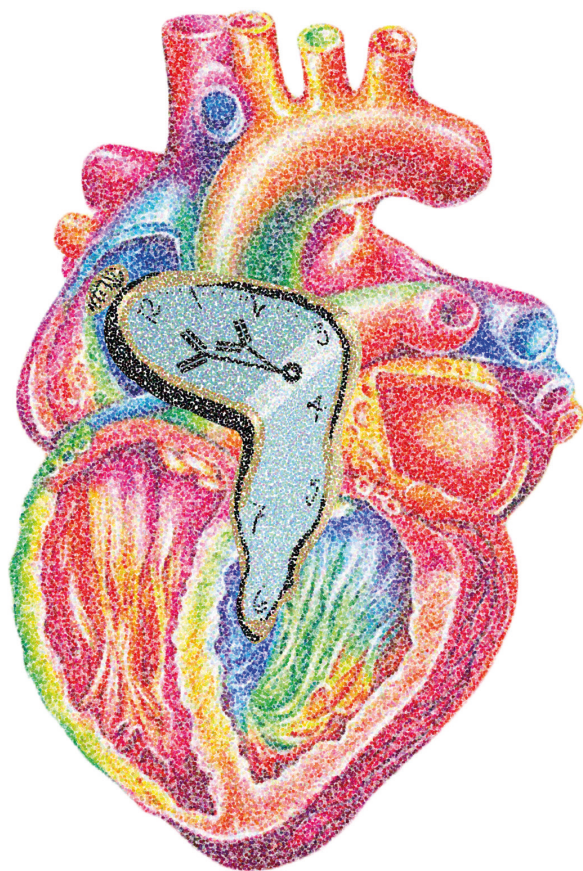
Mol, J. de. (2025, December 11). *The aging B cell landscape in atherosclerosis*. Retrieved from <https://hdl.handle.net/1887/4285092>

Version: Publisher's Version

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/4285092>

Note: To cite this publication please use the final published version (if applicable).



Chapter 1

General Introduction

CARDIOVASCULAR DISEASE

Cardiovascular disease (CVD) refers to all conditions affecting the heart and blood vessels, including ischemic heart disease, stroke, angina pectoris, and aortic aneurysm.¹ CVDs are the leading cause of death worldwide, responsible for approximately 20.5 million deaths each year.² The primary underlying pathology of most CVDs is atherosclerosis, a process marked by the progressive accumulation of lipid-rich plaques within the arterial wall.³ Expansion of the plaque can lead to narrowing of the arteries, also known as stenosis, which impedes normal blood flow and manifests in various symptoms, including intermittent claudication. Plaques eventually can rupture, or erode, resulting in thrombus formation, which can subsequently cause severe cardiovascular events, such as heart attacks or strokes.⁴

Atherosclerosis has long been viewed as a predominantly lipid-driven condition and elevated cholesterol levels generally resulting from an unhealthy diet and sedentary lifestyle are among the well-established risk factors.⁵ Over the past two decades, it has become evident that immune cells also play a significant role in the development of atherosclerosis and several clinical trials, including the CANTOS and LoDoCo trial, have highlighted the crucial role of immunotherapies in treating CVD.⁶⁻⁸ In line with these findings, individuals with chronic inflammatory or autoimmune disorders, such as systemic lupus erythematosus and rheumatoid arthritis, are at increased risk of cardiovascular events.^{9,10} In addition to chronic inflammation and genetic predisposition, unmodifiable risk factors include sex and aging. Although CVD is the primary cause of mortality in both men and women, there are notable sex differences in the prevalence and manifestation of CVD.^{2,11,12} CVD tends to develop approximately 7 to 10 years later in women compared to men. This delay is mainly attributed to the protective effects of estrogen.¹³ After menopause, women's risk of developing CVD increases, eventually catching up with or even surpassing the risk faced by men. Moreover, where men suffer from classical symptoms of chest pain, women generally experience atypical symptoms, complicating diagnosis. Once diagnosed, however, women show a poorer prognosis than men and have higher mortality rates following acute cardiovascular events. Since the mortality rate from cardiovascular disease in the 70+ age category is almost 7-fold higher than in the 50-69 year old age group¹⁴, and even 84-fold higher compared to 15-49 year old individuals, aging is considered one of the most dominant risk factors for cardiovascular death. Upon aging, several physiological and biochemical changes occur, such as arterial stiffening and cellular senescence, which can increase CVD susceptibility.¹⁵ Therefore, the ongoing demographic shift towards an older population drastically increases the global social and economic burden of CVD.

ATHEROSCLEROSIS

Healthy blood vessels are composed of three distinct layers: the intima, media, and adventitia. Each layer plays a crucial role in maintaining vascular function and integrity. The intima is the innermost layer that lines the lumen, the central space through which the blood flows. This layer is covered by a single layer of endothelial cells (ECs), which, under healthy conditions, forms a selective and protective barrier. Beneath the intima lies the media, which is primarily composed of vascular smooth muscle cells (VSMCs). These VSMCs are responsible for regulating blood pressure and flow. The outermost layer, the adventitia, is predominantly composed of fibrous connective tissue, which provides additional structural support. In atherosclerosis, the intimal endothelial cell layer becomes disrupted, enabling infiltration of lipids and immune cells and subsequent atherosclerotic plaque formation.

Initial atherosclerosis

At sites of endothelial dysfunction, atherosclerosis is initiated with the infiltration of lipid-rich lipoproteins, including chylomicrons, very low-density lipoproteins (VLDL) and low-density lipoprotein (LDL) particles into the intima of the arterial wall (**Figure 1**).¹⁶ These particles all contain apolipoprotein B on their surface, a protein which, upon transmigration, interacts with the extracellular matrix, resulting in the retention of lipoproteins in the intima. Subsequently, retained lipoproteins undergo pro-inflammatory chemical modifications¹⁷, such as oxidation, which can induce upregulation of leukocyte adhesion molecules, including VCAM-1, ICAM-1, P- and E-selectin, on endothelial cells and VSMCs.¹⁸ In addition, activated ECs and VSMCs secrete chemokines, which promote the recruitment of monocytes to the site of endothelial disruption. Consequently, integrins on recruited monocytes bind to the upregulated leukocyte adhesion molecules, allowing their transmigration into the intima. Under the influence of macrophage colony stimulating factor (M-CSF) and granulocyte-macrophage colony stimulating factor (GM-CSF), secreted by endothelial cells, monocytes transdifferentiate into macrophages. Macrophages are phagocytic cells that express scavenger receptors, enabling them to engulf oxidized LDL particles.¹⁹ The continuous uptake of oxLDL by macrophages leads to extensive lipid accumulation, a process also known as foam cell formation. These foam cells are central to the early development of atherosclerotic lesions, manifesting as fatty streaks within the arterial wall.

Advanced atherosclerosis

The presence of foam cells triggers a robust inflammatory response characterized by the release of cytokines, growth factors, and enzymes that exacerbate endothelial dysfunction and further contribute to the recruitment of inflammatory cells. Uptake of lipid particles by antigen-presenting immune cells also results in antigen presentation and subsequent T cell recruitment.^{20,21} These T cells, in turn, release various cytokines and chemokines, thereby contributing to the inflammatory plaque environment. Chemokine signaling also orchestrates migration of additional immune cells, including neutrophils, mast cells and B cells, which further contributes to the development of advanced atherosclerotic lesions. Simultaneous to immune cell infiltration, VSMCs from the media layer migrate into the intima, proliferate, and secrete extracellular matrix components such as collagen.²² These activities contribute to the formation of a fibrous cap over the lipid-rich core and accumulation of activated immune cells in the plaque. Like macrophages, VSMCs are capable of lipoprotein internalization, resulting in a VSMC-foam cell phenotype. As the plaque progresses, both VSMCs and macrophages accumulate excessive amounts of lipids, eventually surpassing the storage capacity of these cells, leading to cellular dysfunction and apoptosis. In the early stages, apoptotic cells are cleared efficiently by phagocytes through a process called efferocytosis.²³ Under normal circumstances, efferocytosis ensures that dying cells are rapidly cleared, preventing the release of their contents into the surrounding tissue, thereby maintaining plaque stability and preventing further inflammation. However, in advanced stages of plaque development, efferocytosis is impaired by several factors, including the increased volume of dying cells and reduced uptake capacity of phagocytes.²⁴ This leads to secondary necrosis, a process where the contents of the dying cells, including the accumulated lipids and cellular debris, are released into the atherosclerotic environment. This provokes additional inflammation and formation of a mass of dead cells, referred to as a necrotic core. Initially, the plaque is stabilized by a thick fibrous cap, which prevents interaction between the highly thrombogenic material within the plaque and the blood. In this

stage, patients often remain asymptomatic. However, as the inflammatory processes and necrotic core formation within the plaque continue, the integrity of the fibrous cap begins to deteriorate. Chronic inflammation leads to the activation of matrix metalloproteinases (MMPs), enzymes that degrade components of the fibrous cap. Moreover, plaque stability is further decreased by the development of calcified areas within the lesion, which introduce mechanical stress, increasing the risk of plaque erosion or rupture and subsequent clinical manifestations.

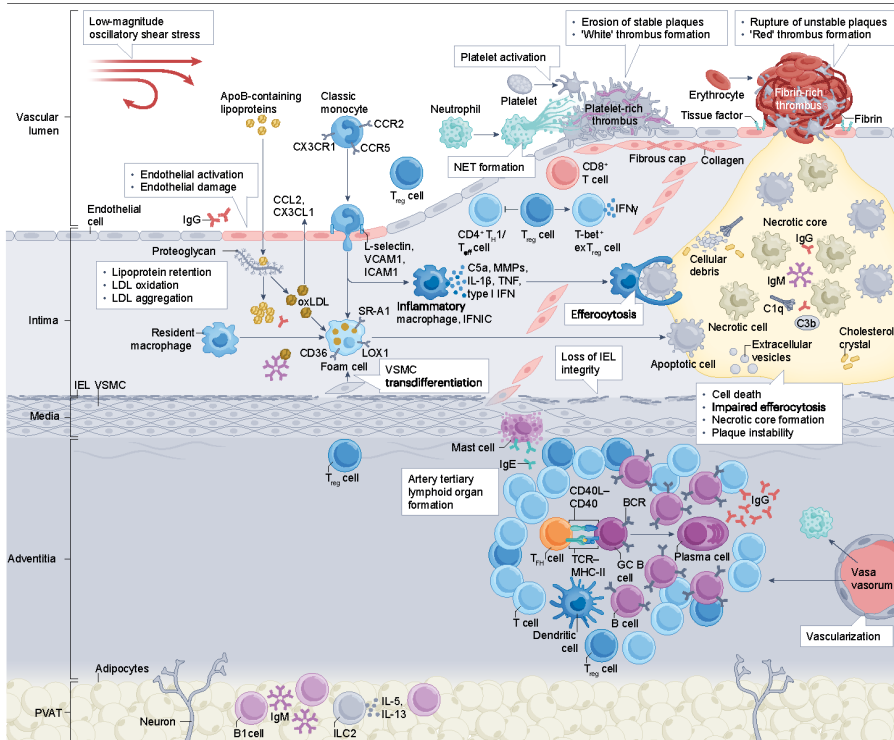


Figure 1. Atherosclerosis plaque development. Atherogenesis is initiated at sites of low shear stress, when apoB-containing lipoproteins (ApoB-LP) transmigrate through the disrupted endothelial cell layer. Consequently, ApoB-LPs are retained and modified, resulting in monocyte recruitment to the atherosclerosis-prone site. Monocytes migrate into the vessel wall and differentiate into macrophages, which can internalize ApoB-LPs. These macrophages differentiate into foam cells, which further attract immune cells of both the innate and adaptive immune system to the site of inflammation. Simultaneously, vascular smooth muscle cells (VSMCs) from the media migrate into the intima, where they can both internalize ApoB-LPs and form a stable fibrous cap over the lipid-rich core of the plaque. As the plaque progresses, excessive lipid uptake by macrophages and VSMCs causes apoptosis. Although apoptotic cells are initially cleared by phagocytosis, in later stages this process is impaired, causing secondary necrosis which leads to necrotic core formation within the lesion. As a result, more immune cells are attracted to the atherosclerosis-prone site, eventually forming artery tertiary lymphoid structures in the adventitia and aggravating inflammation. This chronic inflammation, in addition to necrotic core formation and calcification, reduces plaque stability, which can promote plaque erosion or rupture. *Adapted with permission from Porsch and Binder. (2024) Nat. Rev. Cardiol 21(11):780-807.*²⁵

Experimental models of atherosclerosis

Although significant progress has been made in the development of *in vitro* microvasculature models that can replicate aspects of the healthy vasculature and shear stress in atherosclerosis, these models have not yet reached the level of accuracy necessary to mimic the complex etiology of atherosclerotic plaque formation.^{26,27} Experimental animal models, therefore, remain essential for studying the multifactorial nature of plaque progression, including lipid accumulation, immune cell infiltration,

and the dynamic interactions between different cell types and extracellular matrix components. Atherosclerosis has been investigated in a variety of species, including zebrafish, rats, rabbits, pigs, and non-human primates. However, the mouse remains the most widely used animal model for atherosclerosis research, due to its rapid reproduction and short lifespan, cost-effective maintenance, ease of genetic manipulation and well-characterized genetics and physiology.²⁸ It must be noted that mice show a distinct lipoprotein profile compared to humans. Unlike humans, who transport most of their cholesterol in atherogenic LDL particles, wildtype C57Bl/6 mice primarily transport cholesterol in anti-atherogenic high-density lipoprotein (HDL) particles, making them naturally resistant to the development of atherosclerosis. To overcome these differences, mouse models with genetic and dietary modifications that better mimic human atherosclerosis are available. The two most commonly used genetically modified strains are the LDL receptor knockout (*Ldlr*^{-/-}) and the apolipoprotein E knockout (*Apoe*^{-/-}) mice, which exhibit significantly elevated levels of total plasma cholesterol, particularly in the form of VLDL and LDL, leading to the development of atherosclerotic plaques.²⁹

Deficiency in the *Apoe* gene results in plasma cholesterol levels between 400 and 600 mg/dL, allowing accelerated atherosclerosis progression upon a standard laboratory chow diet (containing 4-6% fat and <0.03% cholesterol) in *Apoe*^{-/-} mice.³⁰ When fed a Western-type diet (containing 21% fat and 0.15% cholesterol), these mice develop plasma cholesterol levels around ~1500 mg/dL, promoting atherosclerotic lesion development even more rapidly. Besides dyslipidemia, *Apoe* deficiency exacerbates inflammation. In the absence of *Apoe*, which is normally produced by macrophages, lipid uptake is increased and efferocytosis impaired³¹, contributing to a more pro-inflammatory macrophage state.³² Moreover, *Apoe* prevents excessive T and B cell activation, and inhibits the production of pro-inflammatory cytokines, such as TNF α and IL-6.^{33,34} In contrast, *Ldlr* deficient mice show less pronounced immune cell alterations. These mice lack the receptor necessary for clearing LDL particles from the bloodstream, resulting in mild hypercholesterolemia, with cholesterol levels around 200-300 mg/dL, when fed a standard diet.²⁸ To induce more severe hypercholesterolemia and accelerate atherosclerosis development in young *Ldlr*^{-/-} mice, feeding a pro-inflammatory Western-type diet is necessary. Notably, this accelerated atherosclerosis induction may not fully capture the complexity of advanced plaques, which gradually develop during the lifetime of humans. In addition to these traditional models, the overexpression of proprotein convertase subtilisin/kexin type 9 (PCSK9) in C57Bl/6 mice has emerged as a powerful model for studying atherosclerosis. PCSK9 is a protein that regulates cholesterol metabolism by promoting the degradation of LDL receptors on hepatocytes, thereby reducing the clearance of LDL cholesterol from the bloodstream.³⁵ Overexpression of PCSK9 in mice, achieved through viral vector-mediated gene transfer, complemented by a Western-type diet, leads to marked hypercholesterolemia and rapid development of atherosclerotic lesions, similar to atherosclerosis development in *Ldlr*^{-/-} mice.³⁶

THE IMMUNE SYSTEM IN ATHEROSCLEROSIS

The immune system, which can be divided in the innate and adaptive immune system, is a crucial defense mechanism that protects the body against infections and tissue damage. Immune cells mostly arise from hematopoietic stem cells in the bone marrow, which differentiate into myeloid and lymphoid lineages.³⁷ The myeloid lineage gives rise to innate immune cells, including neutrophils, monocytes,

macrophages, mast cells and dendritic cells. These cells are the first line of defense and recognize pathogen-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs) of invading and endogenous danger.³⁸ Certain innate immune cells, in particular macrophages and dendritic cells, can activate lymphoid cells of the adaptive immune system via antigen presentation. The adaptive immune system comprises of T and B cells, and although its activation upon first antigen encounter is slow, this arm has the unique ability to develop immunological memory.³⁹ Both innate and adaptive immune cells play pivotal roles in the development and progression of atherosclerosis.

INNATE IMMUNITY

Monocytes and macrophages

During homeostasis, monocytes circulate in the blood and can either return to the bone marrow or migrate into tissues. There are two main monocyte types: classical monocytes (CCR2⁺Ly6C^{high} in mice, CD14⁺CD16⁻ in humans) and non-classical monocytes (CCR2⁺Ly6C^{low} in mice, CD14^{low}CD16⁺ in humans).²¹ Classical monocytes are pro-inflammatory, producing high levels of cytokines and chemokines, and play a key role in atherosclerosis by being recruited to the vessel wall, entering the intimal layer, and differentiating into macrophages.⁴⁰ Non-classical monocytes patrol the endothelium and are generally atheroprotective, though higher levels have been linked to carotid intima-media thickening in men in a multicenter, population-based cohort.⁴¹

Macrophages are central to atherosclerosis development, with functions like phagocytosis, efferocytosis, antigen presentation, and lipid processing. Traditionally classified into pro-inflammatory M1 and anti-inflammatory M2⁴², macrophages in atherosclerosis are now known to have a broader range of subtypes, including resident-like macrophages, foamy TREM2⁺ macrophages, interferon-inducible macrophages, and inflammatory macrophages.^{43–47} Tissue resident-like macrophages are already present in healthy aortas, where they regulate collagen production by VSMCs and clear apoptotic cells. Although resident-like macrophages can take up cholesterol, thereby initiating foam cell formation during early atherogenesis, it has recently been shown that tissue resident-like macrophages exert atheroprotective properties.⁴⁸ Foamy macrophages, however, rapidly outnumber the tissue resident-like macrophages. These macrophages engulf large amounts of LDL, leading to their characteristic ‘foamy’ appearance. Recent studies have indicated that a large subset of foamy macrophages express TREM2^{43,49–51}, a surface receptor involved in lipid uptake and metabolism.⁵² Lipid-associated TREM2-expressing macrophages initially perform a protective role by processing and storing lipids. Furthermore, TREM2^{hi} macrophages limit necrotic core formation and show a low inflammatory profile, which is associated with the traditional anti-inflammatory M2 phenotype.^{53,54} Recent evidence shows that, upon toll-like receptor 2 signaling, intraplaque TREM2^{hi} macrophages can transform into more inflammatory lipid-associated PLIN2^{hi}/TREM1^{hi} macrophages.⁵⁵ These PLIN2^{hi}/TREM1^{hi} macrophages contribute to plaque progression by sustaining local inflammation and are associated with cerebrovascular events. Interferon-inducible macrophages also stimulate chronic inflammation by secreting high levels of pro-inflammatory cytokines, including IL-1 β , TNF α and CXCL2.^{51,56,57} These macrophages also express markers associated with the inflammasome and resemble a more M1-like macrophage phenotype. Their presence is particularly pronounced in advanced plaques, where they contribute to plaque instability by promoting the degradation of the extracellular matrix and thinning the fibrous cap, increasing the risk of plaque rupture and subsequent thrombus formation.

Dendritic cells

Dendritic cells (DCs) are antigen-presenting cells (APCs) that bridge the innate and adaptive immune system. In atherosclerosis, DCs are found in the intima and adventitia of arteries, where they capture and process antigens derived from modified lipoproteins, apoptotic cells, and other sources.^{58,59} Upon antigen internalization, activated DCs travel to draining lymph nodes, where these processed antigens are presented to T cells, leading to their activation, migration and differentiation. The role of DCs in atherosclerosis is complex, as they can drive both pro-atherogenic and protective immune responses depending on the context. For instance, DCs can promote the activation of pro-inflammatory T cells, contributing to plaque progression, while they also have the capacity to induce regulatory T cells (Tregs) that may help to suppress inflammation and stabilize plaques.

Mast cells

Mast cells, traditionally associated with allergic responses, also contribute to the inflammatory milieu in atherosclerosis.⁶⁰ They are found in the adventitia and perivascular tissue around atherosclerotic plaques, where they secrete a variety of pro-inflammatory mediators, including histamine, cytokines, proteases, and growth factors. These mediators can degrade the extracellular matrix, promote lipid uptake by macrophages, and enhance the recruitment of additional immune cells to the plaque. The proteases released by mast cells, such as chymase and tryptase, can also weaken the fibrous cap of the plaque, increasing the risk of rupture and subsequent thrombosis.⁶¹

ADAPTIVE IMMUNITY

T cell development

T cells are a critical component of the adaptive immune response in atherosclerosis. Their development starts with the migration of common lymphoid progenitors (CLPs) from the bone marrow to the thymus, where these cells commit to the T cell lineage under the influence of thymic stromal cells and cytokines such as IL-7.⁶² In the early stages of T cell development, known as the double negative (DN) phase, thymocytes lack the CD4 and CD8 co-receptors. These DN thymocytes go through different differentiation stages in which the T cell receptor (TCR) is developed with either α and β or γ and δ chains.⁶³ At the end of the DN phase, $\alpha\beta$ thymocytes transition to double positive (DP) cells, expressing both CD4 and CD8. In this stage, a complete and unique TCR is developed, which is then tested for its ability to recognize self-antigens.⁶⁴ TCRs with moderate affinity are positively selected, while those with high affinity receive pro-apoptosis signals. Dependent on the affinity for either major histocompatibility complex (MHC) class I or II molecules, these cells are committed to the CD8⁺ or CD4⁺ T cell lineage, respectively. These mature T cells exit the thymus and enter the peripheral circulation, where they contribute to immune surveillance and responses against pathogens.

Naïve T cells become activated primarily in secondary lymphoid organs, such as the lymph nodes and spleen, where they encounter their specific antigen, presented by APCs.⁶⁵ Antigen recognition, in combination with co-stimulatory signals from the APC, results in T cell activation. This activation prompts the T cell to proliferate and differentiate into effector T cells, which then leave the lymphoid organ to travel to sites of infection or inflammation to perform their immune functions.

In the context of atherosclerosis, both CD4⁺ and CD8⁺ T cells that recognize modified self-antigens, such as oxLDL, escape the thymic selection process. Upon activation, these autoreactive T cells then migrate to atherosclerotic lesions, contributing to the auto-immune like response.⁶⁶

CD4⁺ T cells

CD4⁺ T cells, also known as helper T (Th) cells, are one of the most extensively studied immune cell types in atherosclerosis. They can differentiate into various subsets, including Th1, Th2, Th9, Th17, Th22, follicular T helper (T_{FH}) cells, and regulatory T cells (Tregs), each playing distinct roles in disease progression (**Figure 2**).

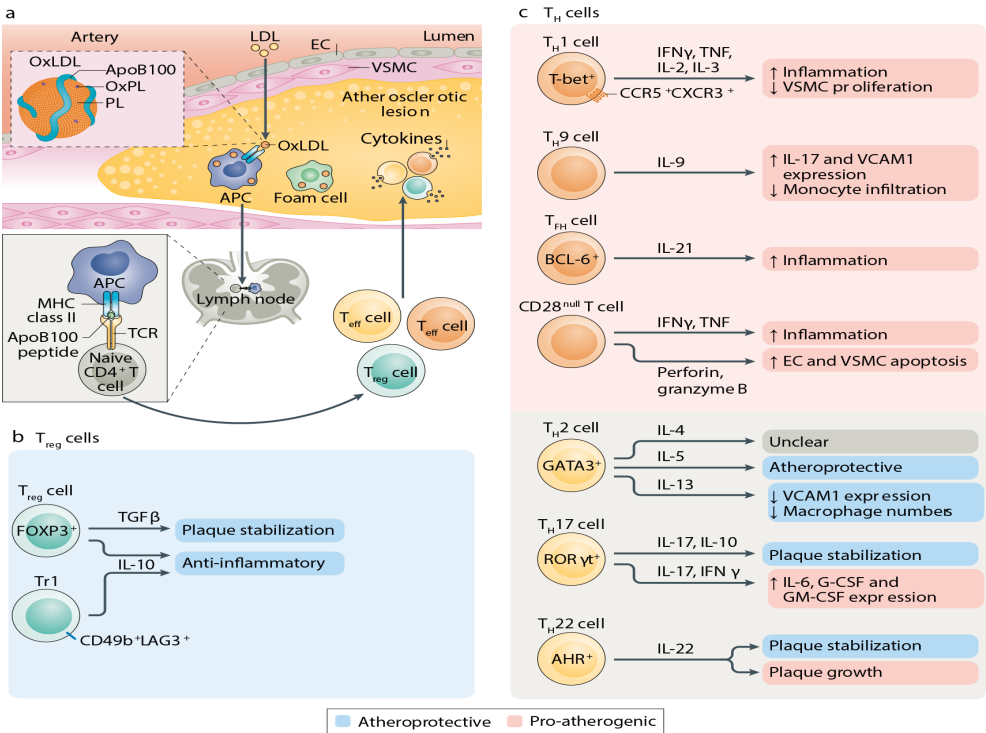


Figure 2. CD4⁺ T cell subsets in the pathology of atherosclerosis. **a.** Naïve CD4⁺ T cells are activated by presentation of oxLDL peptides on APCs in lymph nodes, after which they differentiate into effector CD4⁺ or regulatory T cells and migrate towards the atherosclerotic lesion. **b.** Regulatory T cells (FoxP3⁺) secrete anti-inflammatory cytokines and promote plaque stabilization. **c.** Effector CD4⁺ T cells can have distinct phenotypes. Atherogenic Th1 (T-bet⁺) cells are the most abundant in the plaque and promote inflammation. Th9 cells secrete IL-9 and stimulate leukocyte infiltration. T_{FH} (BCL-6⁺) cells secrete IL-21 and promote inflammation. CD28^{null} T cells promote inflammation and plaque destabilization. The role of Th2 (GATA-3⁺) cells is less clear, but they mainly exert atheroprotective functions. Th17 (RORγT⁺) promote plaque stabilization but also aggravate inflammation. IL-22, secreted by Th22 (AHR⁺) cells, increases plaque growth and stabilization. Adapted with permission from Saigusa, Winkels and Ley (2020) *Nat. Rev. Cardiol* 17(7):387-401.

Th1

Th1 cells, characterized by the transcription factor TBX21 (T-bet), are the most abundant CD4⁺ T cell subset in the human atherosclerotic plaque.^{46,67} Upon stimulation, Th1 cells secrete the pro-inflammatory cytokines IFNγ, IL-2 and TNFα, which promote macrophage activation and stimulate

inflammation. Deficiency in T-bet or IFN γ both resulted in reduced atherosclerosis development^{68,69}, underlining the pro-atherogenic nature of this subset.

Th2

In contrast, Th2 cells, defined by the transcription factor GATA-3, are generally associated with anti-inflammatory responses.⁷⁰ Th2 cells produce IL-4, IL-5 and IL-13, which can counterbalance the pro-atherogenic actions of Th1 cells. For instance, IL-4 can inhibit macrophage activation and reduce the production of IFN γ by Th1 cells.^{71,72} Deficiency of IL-5 and IL-13 accelerated atherosclerosis lesion development, further suggesting an atheroprotective role.^{73,74} An excessive Th2 response, however, can also lead to increased fibrosis, which might contribute to the thickening of the vessel wall and complicate the disease.⁷⁵

Th9

Although the function of Th9 cells in atherosclerosis remains to be elucidated, recent studies have reported pro-atherogenic effects.^{76,77} Th9 cells are the primary source of the cytokine IL-9, which is associated with increased IL-17 secretion and mast cell recruitment, thereby aggravating atherosclerosis progression.

Th17

Similar to Th2 and Th9 cells, the nature of Th17 cells in atherosclerosis development is not completely understood. Th17 cells are characterized by the transcription factor ROR γ T and a major source of IL-17. Studies have shown that IL-17 can exert pro-atherogenic functions by exacerbating immune cell infiltration and chemokine secretion^{78–80}, but might also increase plaque stability by promoting collagen production.^{81,82}

Th22

In atherosclerosis, the role of Th22 cells is complex, as they might help in tissue repair and stability in some contexts, while promoting inflammation and plaque progression in others.^{83,84} Th22 express the transcription factor AHR and secrete IL-22, which is involved in vascular repair.

T_{FH}

T_{FH} cells are a subset of pro-atherogenic CD4⁺ T cells that express the transcription factor BCL-6 and chemokine receptor CXCR5.⁸⁵ Under the influence of CXCL13, T_{FH} cells home towards B cell follicles in lymphoid organs, where they are crucial for the formation of germinal centers. Upon interaction with CD40 and IL-6 or IL-21 signals, T_{FH} cells help B cells undergo affinity maturation and class-switch recombination to produce high-affinity antibodies. In the pathogenesis of atherosclerosis, T_{FH} cell expansion has been identified in the circulation of patients and tertiary lymphoid organs in mice.^{86,87} Dysregulation of the T_{FH}- germinal center B cell axis in atherosclerosis can drive the production of pro-inflammatory autoantibodies against oxLDL, which can form immune complexes that exacerbate inflammation and contribute to the progression of the disease.⁸⁸ Moreover, T_{FH} cells can indirectly contribute to the chronic inflammation characteristic of atherosclerosis by supporting detrimental B cell responses and the formation of aortic TLOs in mice.⁸⁷ Pro-inflammatory T_{FH} cells can be counteracted

by regulatory follicular helper T cells (T_{FH}), which inhibit T_{FR} differentiation and promote regulatory B cell expansion.⁸⁹ Adoptive transfer of these T_{FR} cells in atherosclerotic mice decreased atherosclerotic plaque burden and the infiltration of pro-inflammatory macrophages.

CD28^{null} T cells

Unlike conventional $CD4^+$ T cells, $CD28^{null}$ T cells lack the CD28 co-stimulatory molecule, which is essential for typical T cell activation and survival. This loss of CD28 is associated with a senescent-like phenotype, characterized by reduced proliferative capacity and altered function.^{90,91} Notably, $CD28^{null}$ T cells exhibit increased production of pro-inflammatory cytokines, such as $IFN\gamma$ and $TNF\alpha$, contributing to vascular inflammation.⁹² They also express cytotoxic molecules like perforin and granzymes, enabling them to induce apoptosis in ECs and VSMCs, thereby destabilizing atherosclerotic plaques. Additionally, these cells demonstrate resistance to apoptosis due to the downregulation of death receptors like Fas and pro-apoptotic proteins such as Bim and Bax, leading to their accumulation in atherosclerotic lesions.⁹³ The presence of $CD28^{null}$ T cells has been correlated with increased risk of myocardial infarction and poor cardiovascular outcomes.^{94,95}

Tregs

Tregs, defined by their FoxP3 and CD25 expression, are essential for maintaining immune tolerance and preventing excessive immune responses by producing the anti-inflammatory cytokines IL-10 and $TGF\beta$.⁹⁶ These cytokines suppress the activation and proliferation of pro-inflammatory T cells, including Th1 and Th17 cells, and reduce the activation of macrophages. Although the number of Tregs in the atherosclerotic lesion is limited⁹⁷, depletion of Tregs resulted in accelerated atherosclerosis progression^{98,99}, underlining the atheroprotective function of this subset. Furthermore, Treg expansion studies have shown that Tregs support the stabilization of plaques by enhancing collagen production and inhibiting the degradation of the extracellular matrix^{100–102}, highlighting the therapeutic potential of promoting Treg responses.^{103,104}

CD8⁺ T cells

Compared to $CD4^+$ T cells, the role of cytotoxic $CD8^+$ T cells in atherosclerosis is less well characterized. Nevertheless, emerging evidence suggests a multifaceted and context-dependent role in plaque development and stability.¹⁰⁵ $CD8^+$ T cells can recognize and kill cells presenting MHC class I antigens. In the early stages of atherosclerosis, this cytotoxic activity, mediated by the release of perforin and granzyme B, contributes to endothelial dysfunction, thereby increasing vascular permeability and lipid infiltration.¹⁰⁶ Moreover, $CD8^+$ T cells secrete pro-inflammatory cytokines, such as $IFN\gamma$ and $TNF\alpha$, which further amplify the inflammatory response in atherosclerosis.¹⁰⁷ As the plaque progresses, their continued cytotoxic activity and production of pro-inflammatory cytokines can lead to the apoptosis of smooth muscle cells and degradation of the extracellular matrix, promoting plaque rupture. Nevertheless, atheroprotective functions of $CD8^+$ T cells, mostly exerted by regulatory $CD8^+$ T cells¹⁰⁸, have also been described. Depletion of $CD8^+$ T cells in advanced stages of atherosclerosis resulted in an increased pro-inflammatory Th1 response and plaque destabilization and immunization-induced $CD8^+$ T cell expansion reduced atherosclerotic lesion development.^{109,110} The distinct effects exerted by $CD8^+$ T cells might be caused by different subpopulations identified in atherosclerotic lesions.^{46,66,67,111,112}

$\gamma\delta$ T cells

$\gamma\delta$ T cells represent a small subset of T cells that express a distinct TCR composed of γ and δ chains.^{113,114} Unlike conventional $\alpha\beta$ T cells, $\gamma\delta$ T cells do not require antigen presentation by MHC molecules and can respond rapidly to stress signals, making them an important part of the innate-like immune response in atherosclerosis. Albeit in small numbers, $\gamma\delta$ T cells are found in atherosclerotic plaques, where they can produce pro-inflammatory cytokines like IL-17, contributing to local inflammation and plaque progression. Additionally, $\gamma\delta$ T cells may influence the activity of other immune cells, such as macrophages and DCs, further modulating the inflammatory environment within the plaque.

B cells

Next to T cells, the adaptive immune system consists of B lymphocytes, which are traditionally known for their role in humoral immunity and antibody production. B cells also arise from common lymphoid progenitors. In the bone marrow, CLP differentiate into pre-pro, pro- and pre-B cells, during which a unique B cell receptor (BCR) is developed.¹¹⁵ Similar to TCRs, BCRs undergo selection procedures before immature B cells are approved to leave the bone marrow, after which they enter the peripheral circulation as transitional B cells. Transitional B cells migrate to secondary lymphoid organs, such as the spleen and lymph nodes, where they further mature and undergo additional selection processes to ensure tolerance. Transitional B cells that pass through the selection processes become mature, naive B cells.¹¹⁶ Mature B cells express both IgM and IgD on their surface and circulate through the blood and lymphoid tissues, ready to encounter their specific antigen. Follicular (FO) B cells are the most common subset of mature B cells, which reside in the follicles of lymph nodes and the spleen.¹¹⁷ They participate in T cell-dependent immune responses, where they can be activated by CD4⁺ T cells upon encountering their specific antigen. Located mainly in the spleen, marginal zone (MZ) B cells respond rapidly to blood-borne pathogens and are involved in T cell-independent immune responses.¹¹⁸ Predominantly found in the peritoneal and pleural cavities, B1 cells are a unique subset that arise from distinct B cell progenitors, produce natural antibodies and play a role in early defense against pathogens.^{119–121} B1 cells are also involved in T cell-independent responses and can self-renew, maintaining a steady state of natural antibodies in the body. Upon activation, mature B cells can differentiate into plasma cells, which are specialized for antibody production and secretion.¹²² Some activated B cells differentiate into memory B cells, which persist in lymphoid organs to provide a rapid and robust immune response upon subsequent encounter with the same antigen.

Different B cell subsets contribute differently to atherosclerosis.¹²³ B1 cells, which in mice can be subdivided in the B1a and B1b subpopulations, are generally considered protective in atherosclerosis. The natural IgM antibodies they produce can recognize and bind to oxLDL-specific epitopes (OSEs) and apoptotic cells, facilitating their clearance through opsonization and promoting their uptake by phagocytes.¹²⁴ This process helps prevent the formation and progression of atherosclerotic plaques. Studies have shown that adoptive transfer of B1 cells or their IgM antibodies decreased susceptibility to atherosclerosis.^{125,126} Other protective B cell subsets include marginal zone B cells, which exert atheroprotective functions by the production of natural IgM and the inhibition of pro-inflammatory Tfh responses^{127,128}, and regulatory B cells (Bregs), which secrete the anti-inflammatory IL-10 cytokine.¹²⁹ However, the majority of mature B cells comprise the FO B cell subset. Multiple studies demonstrated

that FO B cells and FO B cell-derived plasma cells aggravate atherosclerosis by their production of IgG and release of pro-inflammatory cytokines, such as IL-6.^{88,130,131}

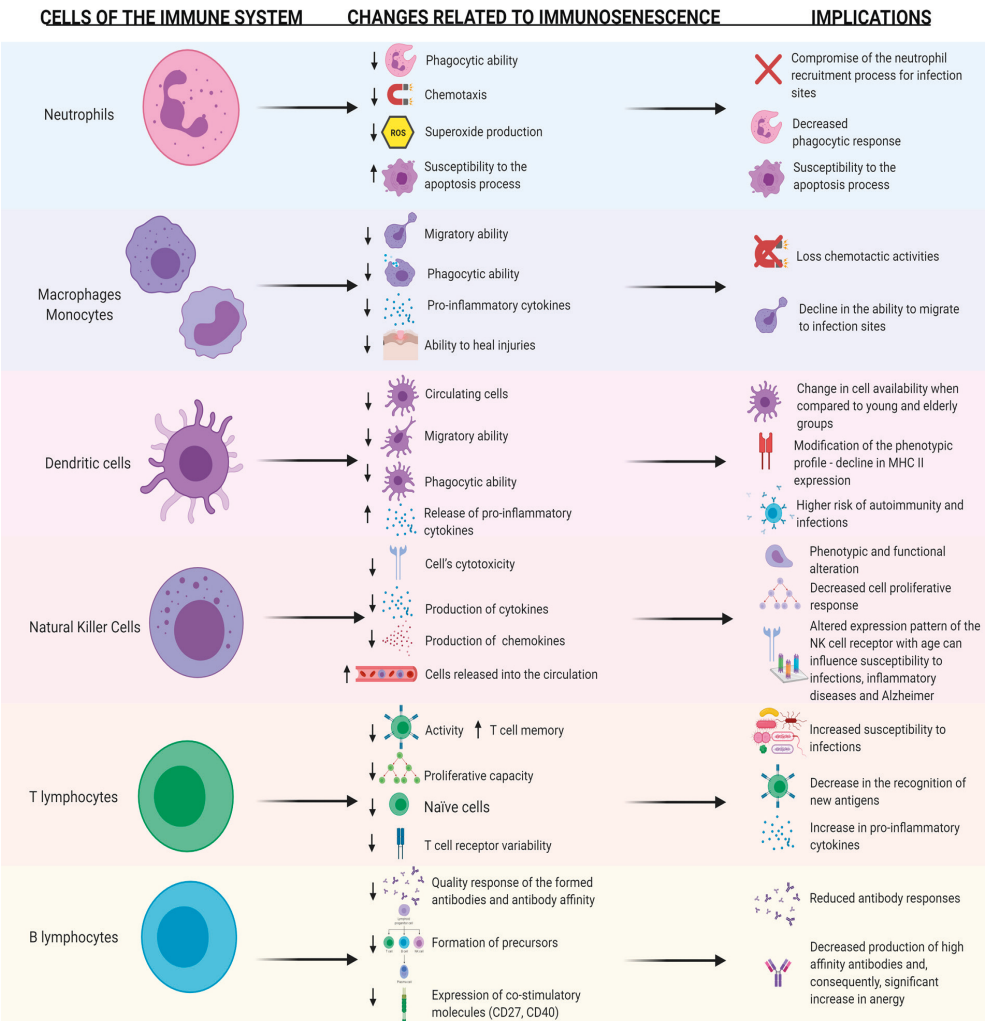


Figure 3. Immunosenescence affects both innate as adaptive immune cells. During aging, neutrophils lose their phagocytic and chemotactic ability, resulting in compromised neutrophil recruitment and increased neutrophil apoptosis. Monocytes and macrophages also show reduced migratory and phagocytic capacities, leading to a decline in chemotaxis. In addition to reduced migration and phagocytosis, dendritic cells also become more pro-inflammatory upon aging. Natural Killer cells become less cytotoxic, thereby increasing susceptibility to inflammatory diseases. T lymphocytes show reduced activity, receptor variability and proliferative capacity, resulting in a decreased recognition of new antigens. Moreover, B lymphocyte, and subsequent antibody, production is dysfunctional. *Adapted from Rodrigues et al (2021) Cytokine Growth Factor Rev. 59:9-21.*¹³²

AGING OF THE IMMUNE SYSTEM: IMMUNOSENESCENCE

Aging profoundly impacts the immune system, leading to a decline in immune function, increased susceptibility to infections, and a higher risk of chronic inflammatory diseases such as atherosclerosis. This phenomenon, referred to as immunosenescence, involves changes in both the innate and adaptive arms of the immune system (**Figure 3**).¹³² Additionally, aging is associated with inflammaging, a chronic

low-grade inflammation that further contributes to age-related pathologies. Key processes involved in aging of the immune system include myeloid skewing in the bone marrow, thymic involution, and the development of the senescence-associated secretory phenotype (SASP).¹³³

Hematopoietic stem cells (HSCs) in the bone marrow give rise to all blood cells, including immune cells. In young individuals, HSCs can differentiate into both myeloid and lymphoid lineages, maintaining a balance between innate and adaptive immune cells. However, aging disrupts this balance, leading to a phenomenon known as myeloid skewing.¹³⁴ The increased production of myeloid cells, such as neutrophils, monocytes, and macrophages, occurs at the expense of lymphoid cells, particularly B and T cells. This shift results in a reduced output of lymphoid progenitors and a decline in the diversity and function of adaptive immune cells. Moreover, the diversity of T cells is declined upon aging due to thymic involution¹³⁵, leading to a reliance on memory T cells. This shift impairs the ability to respond to novel infections and may contribute to the accumulation of dysfunctional, pro-inflammatory, senescent T cells. Besides diminished T cell responses, B cells in the aged immune system produce antibodies with lower affinity, less diversity, and impaired class-switching, thereby reducing the effectiveness of humoral immunity.¹³² Although the number of myeloid cells generally increases upon aging, the chemotactic, cytotoxic and phagocytotic capability of neutrophils, macrophages, dendritic cells and natural killer cells, is declined.

In addition to structural and functional changes, aging leads to an accumulation of senescent cells. Cellular senescence is a state of irreversible cell cycle arrest, marked by the high expression of cell cycle suppressors p16, p21 and p53, that occurs in response to various stressors, including DNA damage, oxidative stress, and telomere shortening.¹³⁶ Although senescence prevents the proliferation of damaged cells, the upregulation of the pro-survival kinase mTOR and anti-apoptotic Bcl-2 proteins promote senescent cells to remain metabolically active and develop resistance to apoptosis.¹³⁷ Senescent cells develop a distinct phenotype characterized by the secretion of a variety of pro-inflammatory cytokines, chemokines, growth factors, and proteases, collectively known as the SASP.¹³⁸ Chronic SASP expression contributes to tissue dysfunction, inflammation, and the progression of age-related diseases.

CELLULAR AGING AND ATHEROSCLEROSIS

It has become clear that aging is a major risk factor for atherosclerosis through various mechanisms at the cellular level. In addition to immunosenescence, studies have demonstrated that aging influences endothelial cells, VSMCs, and other components of the vascular system.¹³⁹ With age, endothelial cells lose their ability to produce nitric oxide¹⁴⁰, a molecule that helps dilate blood vessels and maintain a healthy vascular tone. This leads to reduced vasodilation, increased oxidative stress, and heightened vulnerability to damage from circulating lipids and immune cells. Moreover, aged endothelial cells exhibit increased permeability¹⁴¹, allowing for easier infiltration of LDL and increased atherogenesis.¹⁴² In response to aging, VSMCs become more prone to proliferation and migration into the intima.¹⁴³ This shift is associated with changes in the production of extracellular matrix components, such as increased collagen deposition and reduced elastin, resulting in arterial stiffness. This stiffening increases blood pressure, which can further damage the endothelium and promote atherosclerosis. In contrast, when VSMCs become senescent, they lose their ability to proliferate and produce matrix components.¹⁴⁴

Senescent VSMCs may also undergo osteogenic differentiation, leading to vascular calcification, a hallmark of advanced atherosclerotic lesions. Calcified plaques are more rigid and prone to rupture, which can augment acute cardiovascular events. Like endothelial and immune cells, senescent VSMCs also exhibit the pro-inflammatory SASP, thereby further aggravating atherosclerosis progression.

Most experimental research on atherosclerosis therapies has been conducted in relatively young animals, which correspond to a human equivalent age around 20-30 years. This presents a challenge, as cardiovascular disease patients who are in need for such treatments are often of advanced age with an aged vascular and immune system, making it difficult to directly apply these findings to clinical settings. As evidenced by Simo *et al.* aging is accompanied with a decreased expression of membrane cholesterol transporters on macrophages, resulting in cholesterol accumulation and increased susceptibility to atherosclerosis progression.¹⁴⁵ Furthermore, multiple studies demonstrated that age-associated mitochondrial and vascular dysfunction promote inflammation and aggravate atherosclerosis development in mice.^{142,146,147} In addition, senescent cells have been identified in the atherosclerotic plaque, and their presence has been associated with accelerated atherosclerosis progression and increased inflammation.¹⁴⁸ Incorporating aging in atherosclerosis studies is therefore crucial to better understand immune cell behavior and responses in the cardiovascular disease patient. Moreover, recent studies have demonstrated that CVD patients exhibit signs of accelerated biological aging, including shortened telomere length in leukocytes and increased T cell senescence compared to healthy age-matched individuals.^{149,150} Therapeutic anti-aging strategies and the elimination of senescent cells have therefore emerged as promising anti-atherosclerotic approaches.^{151–153} Senolytics selectively induce apoptosis in senescent cells, whereas senomorphics suppress SASP production.¹⁵⁴ Various classes of senolytics have been developed and senolytic mechanisms of action include disruption of anti-apoptotic proteins Bcl-2 and Bcl-x, inhibition of mTOR-induced cell survival, nuclear exclusion of p53, PI3K inhibition and proteasomal degradation.^{155–158}

SINGLE-CELL APPROACHES TO STUDY THE CELLULAR LANDSCAPE IN ATHEROSCLEROSIS

In order to develop new anti-atherosclerotic immune therapies for the aged CVD patient, it is important to gain more insights into immune cell heterogeneity, crosstalk and function during atherosclerosis progression. Historically, immune cell diversity within plaques was explored using immunostaining techniques in the 1980s¹⁵⁹, which allowed researchers to detect and visualize only two markers simultaneously. Over time, this approach improved, and by combining various staining methods, it is now possible to examine up to 16 markers concurrently.^{160,161} However, these methods are limited in scope and provide only a partial view of the complex cellular environment within plaques. When flow cytometry was introduced into atherosclerosis research, it quickly became a powerful tool for studying heterogeneous immune cell populations at the single-cell level. Flow cytometry offers significant advantages, including its speed, cost-effectiveness, and ability to analyze large numbers of cells.¹⁶² Moreover, it provides insights into immune cell phenotypes by allowing the simultaneous analysis of multiple markers, keeping it still a widely used method in atherosclerosis research today. Limitations in flow cytometry stem from the overlap of fluorescence signals, making it nowadays still difficult to analyze more than 20 markers without signal interference or autofluorescence. Spectral

flow cytometry technology reduces signal interference and autofluorescence, extending the number of markers that can be distinguished simultaneously to >40. The introduction of mass cytometry, or Cytometry by Time of Flight (CyTOF), with heavy metal-tagged antibodies in 2009 allowed to detect up to 50 markers simultaneously, far surpassing the capabilities of traditional flow cytometry.¹⁶³ This increased dimensionality offered a more detailed and comprehensive view of the cellular landscape in atherosclerotic plaques.

In parallel, single-cell RNA sequencing (scRNA-seq) was introduced, bringing an entirely new level of resolution to study gene expression in individual cells. Unlike bulk transcriptomics, which averages gene expression across large populations of cells or tissues, scRNA-seq enabled measuring the gene expression profiles of individual cells, allowing for the identification of distinct cellular subpopulations without prior knowledge of specific markers. Single-cell transcriptomics thus provides an unbiased method to characterize the diversity of cells in heterogeneous environments, offering insights into the molecular mechanisms driving disease progression. Although single-cell omics are still relatively new in cardiovascular research, the rapid development of these technologies has enabled the integration of multiple omics layers, including genome, epigenome, transcriptome, proteome, metabolome, and immune repertoire, thereby already providing invaluable insights into cellular function and identity in atherosclerosis. In 2018, the immune transcriptome in young *Ldlr*^{-/-} and *Apoe*^{-/-} mice was mapped using single-cell RNA sequencing analysis, revealing heterogeneity in macrophage subsets and the abundance of T cells inside the atherosclerotic lesion.^{43,49} This murine data was followed by the single-cell immune landscape in human carotid plaques, confirming the high percentage of lesional T cells.^{46,67} In the past years, additional transcriptomic data of the atherosclerotic lesion has become available, allowing the integration of different atherosclerosis models and further identification of myeloid and lymphoid subsets.^{164,165} Nevertheless, the effects of aging on the immune transcriptome in atherosclerosis remain to be investigated.

THESIS OUTLINE

In this thesis, single-cell approaches were applied to characterize the impact of aging on the immunological B cell landscape in different models of atherosclerotic cardiovascular disease, with the goal to identify and study new B cell-associated biomarkers and therapeutic targets to halt atherosclerosis progression.

In **chapter 2** we provide an overview of the aged B cell in health and disease. We describe how aging affects the proliferation and function of distinct B cell subsets, and their possible effect in atherosclerosis development. In **chapter 3**, we characterized the impact of aging on atherosclerosis progression in *Ldlr*^{-/-} mice and illustrated that aging promotes more advanced atherosclerotic lesions, enriched in collagen, cholesterol crystals, and calcification. In addition, we identified age-associated immune cells, such as age-associated B cells (ABCs), and revealed increased immunosenescence in the aged atherosclerotic environment, using single-cell RNA sequencing and flow cytometry. We compared the morphology and immune landscape of atherosclerotic lesions between aged female and aged male *Ldlr*^{-/-} mice in **chapter 4**, and show that age-associated B cells are more pronounced in females and that female B cells exhibit a more activated phenotype. In **chapter 5**, we induced atherosclerosis in non-atherosclerotic aged C57Bl/6 mice, providing insight into how immunosenescence influences disease development.

We demonstrate that the immune system of aged mice shows immunosenescent features, including the emergence of ABCs and increased antibody production, which was accompanied with aggravated atherosclerosis development in aged mice compared to young mice. To investigate the role of age-associated B cells in atherosclerosis development, we further characterized these cells in **chapter 6**. We show that ABCs predict coronary events in humans and are clonally expanded in aged atherosclerotic mice. Upon adoptive transfer, we reveal that ABCs differentiate into plasma cells, thereby exacerbating lesion development. In **chapter 7**, we identified a new anti-atherosclerotic strategy with IFN γ -stimulated B cells by significant upregulation of the inhibitory ligand PD-L1. We demonstrated that these cells inhibit follicular T helper cell responses and halt atherosclerosis progression. To further explore anti-atherogenic therapies in the aged CVD patient, we aimed to reduce advanced atherosclerosis with rapamycin treatment in **chapter 8**. We show that rapamycin was able to restrict inflammation in the aged atherosclerotic lesion and reduced the frequency of ABCs, which might stabilize the atherosclerotic lesion and reduce the risk for a cardiovascular event. **Chapter 9** provides a summary and discussion of all the data presented in this thesis, along with concluding remarks and future perspectives.

REFERENCES

1. Tsao, C. W. et al. Heart Disease and Stroke Statistics-2023 Update: A Report From the American Heart Association. *Circulation* 147, E93–E621 (2023).
2. Timmis, A. et al. European Society of Cardiology: cardiovascular disease statistics 2021. *European heart journal* 43, 716–799 (2022).
3. Lusis AJ. Atherosclerosis. *Nature*. 2000;407:233-241
4. Sary HC. Macrophage foam cells in the coronary artery intima of human infants. *Ann NY Acad Sci*. 1985;454:5-8
5. Sary HC. Evolution and progression of atherosclerotic lesions in coronary arteries of children and young adults. *Arteriosclerosis*. 1989;9:119-32
6. Libby P. Inflammation in atherosclerosis. *Nature*. 2002;420:868-874
7. Jonasson L, Holm J, Skalli O, Bondjers G, Hansson GK. Regional accumulations of t cells, macrophages, and smooth muscle cells in the human atherosclerotic plaque. *Arteriosclerosis*. 1986;6:131-138
8. Davignon J, Ganz P. Role of endothelial dysfunction in atherosclerosis. *Circulation*. 2004;109:III27-32
9. Bobryshev YV, Lord RS. Co-accumulation of dendritic cells and natural killer t cells within rupture-prone regions in human atherosclerotic plaques. *J Histochem Cytochem*. 2005;53:781-785
10. Dai G, Kaazempur-Mofrad MR, Natarajan S, Zhang Y, Vaughn S, Blackman BR, Kamm RD, Garcia-Cardena G, Gimbrone MA, Jr. Distinct endothelial phenotypes evoked by arterial waveforms derived from atherosclerosis-susceptible and -resistant regions of human vasculature. *Proc Natl Acad Sci U S A*. 2004;101:14871-14876
11. Nakashima Y, Raines EW, Plump AS, Breslow JL, Ross R. Upregulation of vcam-1 and icam-1 at atherosclerosis-prone sites on the endothelium in the apoe-deficient mouse. *Arteriosclerosis, thrombosis, and vascular biology*. 1998;18:842-851
12. Ross R. Atherosclerosis--an inflammatory disease. *The New England journal of medicine*. 1999;340:115-126
13. Ginsberg HN. Lipoprotein physiology. *Endocrinol Metab Clin North Am*. 1998;27:503-519
14. Steinberg D. Low density lipoprotein oxidation and its pathobiological significance. *The Journal of biological chemistry*. 1997;272:20963-20966
15. Hansson GK. Inflammation, atherosclerosis, and coronary artery disease. *N Engl J Med*. 2005;352:1685-1695
16. Drexler H, Hornig B. Endothelial dysfunction in human disease. *J Mol Cell Cardiol*. 1999;31:51-60
17. Vestweber D, Blanks JE. Mechanisms that regulate the function of the selectins and their ligands. *Physiol Rev*. 1999;79:181-213
18. Cybulsky MI, Gimbrone MA, Jr. Endothelial expression of a mononuclear leukocyte adhesion molecule during atherogenesis. *Science*. 1991;251:788-791
19. Collins RG, Velji R, Guevara NV, Hicks MJ, Chan L, Beaudet AL. P-selectin or intercellular adhesion molecule (icam)-1 deficiency substantially protects against atherosclerosis in apolipoprotein e-deficient mice. *The Journal of experimental medicine*. 2000;191:189-194
20. Nageh MF, Sandberg ET, Marotti KR, Lin AH, Melchior EP, Bullard DC, Beaudet AL. Deficiency of inflammatory cell adhesion molecules protects against atherosclerosis in mice. *Arteriosclerosis, thrombosis, and vascular biology*. 1997;17:1517-1520
21. Reape TJ, Groot PH. Chemokines and atherosclerosis. *Arteriosclerosis*. 1999;147:213-225
22. Griending KK, Alexander RW. Oxidative stress and cardiovascular disease. *Circulation*. 1997;96:3264-3265
23. Han J, Hajjar DP, Febbraio M, Nicholson AC. Native and modified low density lipoproteins increase the functional expression of the macrophage class b scavenger receptor, cd36. *The Journal of biological chemistry*. 1997;272:21654-21659
24. Li AC, Glass CK. The macrophage foam cell as a target for therapeutic intervention. *Nat Med*. 2002;8:1235-1242
25. Davies MJ. The pathophysiology of acute coronary syndromes. *Heart*. 2000;83:361-366
26. van der Wal AC, Becker AE, van der Loos CM, Das PK. Site of intimal rupture or erosion of thrombosed coronary atherosclerotic plaques is characterized by an inflammatory process irrespective of the dominant plaque morphology. *Circulation*. 1994;89:36-44
27. Dollery CM, Libby P. Atherosclerosis and proteinase activation. *Cardiovasc Res*. 2006;69:625-635
28. Rauch U, Osende JI, Fuster V, Badimon JJ, Fayad Z, Chesebro JH. Thrombus formation on atherosclerotic plaques: Pathogenesis and clinical consequences. *Ann Intern Med*. 2001;134:224-238
29. Farb A, Burke AP, Tang AL, Liang TY, Mannan P, Smialek J, Virmani R. Coronary plaque erosion without rupture into a lipid core. A frequent cause of coronary thrombosis in sudden coronary death. *Circulation*. 1996;93:1354-1363
30. Schoenhagen P, Ziada KM, Kapadia SR, Crowe TD, Nissen SE, Tuzcu EM. Extent and direction of arterial remodeling in stable versus unstable coronary syndromes: An intravascular ultrasound study. *Circulation*. 2000;101:598-603
31. Takano M, Mizuno K, Okamatsu K, Yokoyama S, Ohba T, Sakai S. Mechanical and structural characteristics of vulnerable plaques: Analysis by coronary angiography and intravascular ultrasound. *J Am Coll Cardiol*. 2001;38:99-104
32. von Birgelen C, Klinkhart W, Mintz GS, Papatheodorou A, Herrmann J, Baumgart D, Haude M, Wieneke H, Ge J, Erbel R. Plaque distribution and vascular remodeling of ruptured and nonruptured coronary plaques in the same vessel: An intravascular ultrasound study in vivo. *J Am Coll Cardiol*. 2001;37:1864-1870
33. Goncalves I, Stenstrom K, Skog G, Mattsson S, Nitulescu M, Nilsson J. Short communication: Dating components of human atherosclerotic plaques. *Circ Res*. 2010;106:1174-1177
34. Hansson GK. Immune mechanisms in atherosclerosis. *Arterioscler Thromb Vasc Biol*. 2001;21:1876-1890
35. Silverstein RL. Inflammation, atherosclerosis, and arterial thrombosis: Role of the scavenger receptor cd36. *Cleve Clin J Med*. 2009;76 Suppl 2:S27-30

36. Medzhitov R, Janeway C, Jr. Innate immunity. *The New England journal of medicine*. 2000;343:338-344
37. Gosling J, Slaymaker S, Gu L, Tseng S, Zlot CH, Young SG, Rollins BJ, Charo IF. Mcp-1 deficiency reduces susceptibility to atherosclerosis in mice that overexpress human apolipoprotein b. *The Journal of clinical investigation*. 1999;103:773-778
38. Boring L, Gosling J, Cleary M, Charo IF. Decreased lesion formation in ccr2^{-/-} mice reveals a role for chemokines in the initiation of atherosclerosis. *Nature*. 1998;394:894-897
39. Swirski FK, Pittet MJ, Kircher MF, Aikawa E, Jaffer FA, Libby P, Weissleder R. Monocyte accumulation in mouse atherogenesis is progressive and proportional to extent of disease. *Proc Natl Acad Sci U S A*. 2006;103:10340-10345
40. Swirski FK, Libby P, Aikawa E, Alcaide P, Luscinskas FW, Weissleder R, Pittet MJ. Ly-6chi monocytes dominate hypercholesterolemia-associated monocytosis and give rise to macrophages in atheromata. *The Journal of clinical investigation*. 2007;117:195-205
41. Tacke F, Alvarez D, Kaplan TJ, Jakubzick C, Spanbroek R, Llodra J, Garin A, Liu J, Mack M, van Rooijen N, Lira SA, Habenicht AJ, Randolph GJ. Monocyte subsets differentially employ ccr2, ccr5, and cx3cr1 to accumulate within atherosclerotic plaques. *The Journal of clinical investigation*. 2007;117:185-194
42. Ylitalo C, Oksala O, Yla-Herttuala S, Ylitalo P. Effects of clodronate (dichloromethylene bisphosphonate) on the development of experimental atherosclerosis in rabbits. *J Lab Clin Med*. 1994;123:769-776
43. Combadiere C, Poteaux S, Rodero M, Simon T, Pezard A, Esposito B, Merval R, Proudfoot A, Tedgui A, Mallat Z. Combined inhibition of ccl2, cx3cr1, and ccr5 abrogates ly6c(hi) and ly6c(lo) monocytosis and almost abolishes atherosclerosis in hypercholesterolemic mice. *Circulation*. 2008;117:1649-1657
44. Saederup N, Chan L, Lira SA, Charo IF. Fractalkine deficiency markedly reduces macrophage accumulation and atherosclerotic lesion formation in ccr2^{-/-} mice: Evidence for independent chemokine functions in atherogenesis. *Circulation*. 2008;117:1642-1648
45. Lesnik P, Haskell CA, Charo IF. Decreased atherosclerosis in cx3cr1^{-/-} mice reveals a role for fractalkine in atherogenesis. *The Journal of clinical investigation*. 2003;111:333-340
46. Endemann G, Stanton LW, Madden KS, Bryant CM, White RT, Protter AA. Cd36 is a receptor for oxidized low density lipoprotein. *J Biol Chem*. 1993;268:11811-11816
47. Steinberg D. Atherogenesis in perspective: Hypercholesterolemia and inflammation as partners in crime. *Nat Med*. 2002;8:1211-1217
48. Melian A, Geng YJ, Sukhova GK, Libby P, Porcelli SA. Cd1 expression in human atherosclerosis. A potential mechanism for t cell activation by foam cells. *Am J Pathol*. 1999;155:775-786
49. Smith JD, Trogan E, Ginsberg M, Grigaux C, Tian J, Miyata M. Decreased atherosclerosis in mice deficient in both macrophage colony-stimulating factor (op) and apolipoprotein e. *Proc Natl Acad Sci U S A*. 1995;92:8264-8268
50. Stoneman V, Braganza D, Figg N, Mercer J, Lang R, Goddard M, Bennett M. Monocyte/macrophage suppression in cd11b diphtheria toxin receptor transgenic mice differentially affects atherogenesis and established plaques. *Circ Res*. 2007;100:884-893
51. Gordon S, Taylor PR. Monocyte and macrophage heterogeneity. *Nature reviews. Immunology*. 2005;5:953-964
52. Kadl A, Meher AK, Sharma PR, Lee MY, Doran AC, Johnstone SR, Elliott MR, Gruber F, Han J, Chen W, Kensler T, Ravichandran KS, Isakson BE, Wamhoff BR, Leitinger N. Identification of a novel macrophage phenotype that develops in response to atherogenic phospholipids via nr1f2. *Circ Res*. 2010;107:737-746
53. Gleissner CA, Shaked I, Little KM, Ley K. Cxc chemokine ligand 4 induces a unique transcriptome in monocyte-derived macrophages. *Journal of immunology*. 2010;184:4810-4818
54. Mosser DM, Edwards JP. Exploring the full spectrum of macrophage activation. *Nature reviews. Immunology*. 2008;8:958-969
55. Ley K, Miller YI, Hedrick CC. Monocyte and macrophage dynamics during atherogenesis. *Arteriosclerosis, thrombosis, and vascular biology*. 2011;31:1506-1516
56. Wolfs IM, Donners MM, de Winther MP. Differentiation factors and cytokines in the atherosclerotic plaque micro-environment as a trigger for macrophage polarisation. *Thromb Haemost*. 2011;106:763-771
57. Murray PJ, Wynn TA. Protective and pathogenic functions of macrophage subsets. *Nature reviews. Immunology*. 2011;11:723-737
58. Martinez FO, Helming L, Gordon S. Alternative activation of macrophages: An immunologic functional perspective. *Annu Rev Immunol*. 2009;27:451-483
59. Gordon S, Martinez FO. Alternative activation of macrophages: Mechanism and functions. *Immunity*. 2010;32:593-604
60. Bouhrel MA, Derudas B, Rigamonti E, Dievart R, Brozek J, Haulon S, Zawadzki C, Jude B, Torpier G, Marx N, Staels B, Chinetti-Gbaguidi G. Ppargamma activation primes human monocytes into alternative m2 macrophages with anti-inflammatory properties. *Cell Metab*. 2007;6:137-143
61. Chase AJ, Bond M, Crook MF, Newby AC. Role of nuclear factor-kappa b activation in metalloproteinase-1, -3, and -9 secretion by human macrophages in vitro and rabbit foam cells produced in vivo. *Arteriosclerosis, thrombosis, and vascular biology*. 2002;22:765-771
62. Gallardo-Soler A, Gomez-Nieto C, Campo ML, Marathe C, Tontonoz P, Castrillo A, Corraliza I. Arginase i induction by modified lipoproteins in macrophages: A peroxisome proliferator-activated receptor-gamma/delta-mediated effect that links lipid metabolism and immunity. *Mol Endocrinol*. 2008;22:1394-1402
63. Johnson JL, Newby AC. Macrophage heterogeneity in atherosclerotic plaques. *Curr Opin Lipidol*. 2009;20:370-378

64. El Hadri K, Mahmood DF, Couchie D, Jguirim-Souissi I, Genze F, Diderot V, Syrovets T, Lunov O, Simmet T, Rouis M. Thioresdixin-1 promotes anti-inflammatory macrophages of the m2 phenotype and antagonizes atherosclerosis. *Arteriosclerosis, thrombosis, and vascular biology*. 2012;32:1445-1452
65. Gordon S. Alternative activation of macrophages. *Nature reviews. Immunology*. 2003;3:23-35
66. Mills CD. Macrophage arginine metabolism to ornithine/urea or nitric oxide/citrulline: A life or death issue. *Crit Rev Immunol*. 2001;21:399-425
67. Chinetti-Gbaguidi G, Baron M, Boulhel MA, Vanhoutte J, Copin C, Sebti Y, Derudas B, Mayi T, Bories G, Tailleux A, Haulon S, Zawadzki C, Jude B, Staels B. Human atherosclerotic plaque alternative macrophages display low cholesterol handling but high phagocytosis because of distinct activities of the ppargamma and lxralpha pathways. *Circ Res*. 2011;108:985-995
68. Weber C, Zernecke A, Libby P. The multifaceted contributions of leukocyte subsets to atherosclerosis: Lessons from mouse models. *Nature reviews. Immunology*. 2008;8:802-815
69. Bobryshev YV. Dendritic cells and their role in atherogenesis. *Lab Invest*. 2010;90:970-984
70. Bobryshev YV. Dendritic cells in atherosclerosis: Current status of the problem and clinical relevance. *Eur Heart J*. 2005;26:1700-1704
71. Nakai Y, Iwabuchi K, Fujii S, Ishimori N, Dashtsoodol N, Watano K, Mishima T, Iwabuchi C, Tanaka S, Bezbradica JS, Nakayama T, Taniguchi M, Miyake S, Yamamura T, Kitabatake A, Joyce S, Van Kaer L, Onoe K. Natural killer t cells accelerate atherogenesis in mice. *Blood*. 2004;104:2051-2059
72. Hansson GK, Hellstrand M, Rymo L, Rubbia L, Gabbiani G. Interferon gamma inhibits both proliferation and expression of differentiation-specific alpha-smooth muscle actin in arterial smooth muscle cells. *J Exp Med*. 1989;170:1595-1608
73. Amento EP, Ehsani N, Palmer H, Libby P. Cytokines and growth factors positively and negatively regulate interstitial collagen gene expression in human vascular smooth muscle cells. *Arterioscler Thromb*. 1991;11:1223-1230
74. Galkina E, Ley K. Immune and inflammatory mechanisms of atherosclerosis (*). *Annu Rev Immunol*. 2009;27:165-197
75. Manthey HD, Zernecke A. Dendritic cells in atherosclerosis: Functions in immune regulation and beyond. *Thromb Haemost*. 2011;106:772-778
76. Wu H, Gower RM, Wang H, Perrard XY, Ma R, Bullard DC, Burns AR, Paul A, Smith CW, Simon SI, Ballantyne CM. Functional role of cd11c+ monocytes in atherogenesis associated with hypercholesterolemia. *Circulation*. 2009;119:2708-2717
77. Paulson KE, Zhu SN, Chen M, Nurmohamed S, Jongstra-Bilen J, Cybulsky MI. Resident intimal dendritic cells accumulate lipid and contribute to the initiation of atherosclerosis. *Circ Res*. 2010;106:383-390
78. Gautier EL, Huby T, Saint-Charles F, Ouzilleau B, Pirault J, Deswaerte V, Ginhoux F, Miller ER, Witztum JL, Chapman MJ, Lesnik P. Conventional dendritic cells at the crossroads between immunity and cholesterol homeostasis in atherosclerosis. *Circulation*. 2009;119:2367-2375
79. Geissmann F, Manz MG, Jung S, Sieweke MH, Merad M, Ley K. Development of monocytes, macrophages, and dendritic cells. *Science*. 2010;327:656-661
80. Habets KL, van Puijvelde GH, van Duivenvoorde LM, van Wanrooij EJ, de Vos P, Tervaert JW, van Berkel TJ, Toes RE, Kuiper J. Vaccination using oxidized low-density lipoprotein-pulsed dendritic cells reduces atherosclerosis in ldl receptor-deficient mice. *Cardiovasc Res*. 2010;85:622-630
81. Cheong C, Matos I, Choi JH, Dandamudi DB, Shrestha E, Longhi MP, Jeffrey KL, Anthony RM, Kluger C, Nchinda G, Koh H, Rodriguez A, Idoyaga J, Pack M, Velinzon K, Park CG, Steinman RM. Microbial stimulation fully differentiates monocytes to dc-sign/cd209(+) dendritic cells for immune t cell areas. *Cell*. 2010;143:416-429
82. Naik SH, Metcalf D, van Nieuwenhuijze A, Wicks I, Wu L, O'Keeffe M, Shortman K. Intrasplenic steady-state dendritic cell precursors that are distinct from monocytes. *Nature immunology*. 2006;7:663-671
83. Onai N, Obata-Onai A, Tussiwand R, Lanzavecchia A, Manz MG. Activation of the flt3 signal transduction cascade rescues and enhances type i interferon-producing and dendritic cell development. *The Journal of experimental medicine*. 2006;203:227-238
84. Liu K, Vitoria GD, Schwickert TA, Guernonprez P, Meredith MM, Yao K, Chu FF, Randolph GJ, Rudensky AY, Nussenzweig M. In vivo analysis of dendritic cell development and homeostasis. *Science*. 2009;324:392-397
85. Choi JH, Cheong C, Dandamudi DB, Park CG, Rodriguez A, Mehndru S, Velinzon K, Jung IH, Yoo JY, Oh GT, Steinman RM. Flt3 signaling-dependent dendritic cells protect against atherosclerosis. *Immunity*. 2011;35:819-831
86. Hermansson A, Johansson DK, Ketelhuth DF, Andersson J, Zhou X, Hansson GK. Immunotherapy with tolerogenic apolipoprotein b-100-loaded dendritic cells attenuates atherosclerosis in hypercholesterolemic mice. *Circulation*. 2011;123:1083-1091
87. Jego G, Palucka AK, Blanck JP, Chalouni C, Pascual V, Banchereau J. Plasmacytoid dendritic cells induce plasma cell differentiation through type i interferon and interleukin 6. *Immunity*. 2003;19:225-234
88. Daisormont IT, Christ A, Temmerman L, Sampedro Millares S, Seijkens T, Manca M, Rousch M, Poggi M, Boon L, van der Loos C, Daemen M, Lutgens E, Halvorsen B, Aukrust P, Janssen E, Biessen EA. Plasmacytoid dendritic cells protect against atherosclerosis by tuning t-cell proliferation and activity. *Circ Res*. 2011;109:1387-1395
89. Bonneville M, O'Brien RL, Born WK. Gammadelta t cell effector functions: A blend of innate programming and acquired plasticity. *Nature reviews. Immunology*. 2010;10:467-478
90. Bluestone JA, Mackay CR, O'Shea JJ, Stockinger B. The functional plasticity of t cell subsets. *Nature reviews. Immunology*. 2009;9:811-816
91. Wan YY, Flavell RA. How diverse--cd4 effector t cells and their functions. *J Mol Cell Biol*. 2009;1:20-36

92. Andersen MH, Schrama D, Thor Straten P, Becker JC. Cytotoxic t cells. *J Invest Dermatol*. 2006;126:32-41
93. Roselaar SE, Kakkannathu PX, Daugherty A. Lymphocyte populations in atherosclerotic lesions of apoe^{-/-} and ldl receptor^{-/-} mice. Decreasing density with disease progression. *Arteriosclerosis, thrombosis, and vascular biology*. 1996;16:1013-1018
94. Zhou X, Stemme S, Hansson GK. Evidence for a local immune response in atherosclerosis. Cd4⁺ t cells infiltrate lesions of apolipoprotein-e-deficient mice. *The American journal of pathology*. 1996;149:359-366
95. Kolbus D, Ramos OH, Berg KE, Persson J, Wigren M, Bjorkbacka H, Fredrikson GN, Nilsson J. Cd8⁺ t cell activation predominate early immune responses to hypercholesterolemia in apoe^{-/-}(/)(-) mice. *BMC Immunol*. 2010;11:58
96. Gewaltig J, Kummer M, Koella C, Cathomas G, Biedermann BC. Requirements for cd8 t-cell migration into the human arterial wall. *Hum Pathol*. 2008;39:1756-1762
97. Fyfe AI, Qiao JH, Lusis AJ. Immune-deficient mice develop typical atherosclerotic fatty streaks when fed an atherogenic diet. *The Journal of clinical investigation*. 1994;94:2516-2520
98. Chyu KY, Zhao X, Dimayuga PC, Zhou J, Li X, Yano J, Lio WM, Chan LF, Kirzner J, Trinidad P, Cercek B, Shah PK. Cd8⁺ t cells mediate the athero-protective effect of immunization with an apob-100 peptide. *PLoS ONE*. 2012;7:e30780
99. Elhage R, Gourdy P, Bouchet L, Jawien J, Fouque MJ, Fievet C, Huc X, Barreira Y, Couloumiers JC, Amal JF, Bayard F. Deleting tcr alpha beta⁺ or cd4⁺ t lymphocytes leads to opposite effects on site-specific atherosclerosis in female apolipoprotein e-deficient mice. *The American journal of pathology*. 2004;165:2013-2018
100. Hansson GK, Jonasson L, Lofsted B, Stemme S, Kocher O, Gabbiani G. Localization of t lymphocytes and macrophages in fibrous and complicated human atherosclerotic plaques. *Atherosclerosis*. 1988;72:135-141
101. Hansson GK, Holm J, Jonasson L. Detection of activated t lymphocytes in the human atherosclerotic plaque. *The American journal of pathology*. 1989;135:169-175
102. Song L, Leung C, Schindler C. Lymphocytes are important in early atherosclerosis. *J Clin Invest*. 2001;108:251-259
103. Reardon CA, Blachowicz L, White T, Cabana V, Wang Y, Lukens J, Bluestone J, Getz GS. Effect of immune deficiency on lipoproteins and atherosclerosis in male apolipoprotein e-deficient mice. *Arteriosclerosis, thrombosis, and vascular biology*. 2001;21:1011-1016
104. Zhou X, Nicoletti A, Elhage R, Hansson GK. Transfer of cd4(+) t cells aggravates atherosclerosis in immunodeficient apolipoprotein e knockout mice. *Circulation*. 2000;102:2919-2922
105. Elhage R, Gourdy P, Bouchet L, Jawien J, Fouque MJ, Fievet C, Huc X, Barreira Y, Couloumiers JC, Amal JF, Bayard F. Deleting tcr alpha beta⁺ or cd4⁺ t lymphocytes leads to opposite effects on site-specific atherosclerosis in female apolipoprotein e-deficient mice. *Am J Pathol*. 2004;165:2013-2018
106. Gupta S, Pablo AM, Jiang X, Wang N, Tall AR, Schindler C. Ifn-gamma potentiates atherosclerosis in apoe knock-out mice. *The Journal of clinical investigation*. 1997;99:2752-2761
107. Whitman SC, Ravisankar P, Elam H, Daugherty A. Exogenous interferon-gamma enhances atherosclerosis in apolipoprotein e^{-/-} mice. *The American journal of pathology*. 2000;157:1819-1824
108. Buono C, Binder CJ, Stavrakis G, Witztum JL, Glimcher LH, Lichtman AH. T-bet deficiency reduces atherosclerosis and alters plaque antigen-specific immune responses. *Proc Natl Acad Sci U S A*. 2005;102:1596-1601
109. Whitman SC, Ravisankar P, Daugherty A. Ifn-gamma deficiency exerts gender-specific effects on atherogenesis in apolipoprotein e^{-/-} mice. *J Interferon Cytokine Res*. 2002;22:661-670
110. Branan L, Hovgaard L, Nitulescu M, Bengtsson E, Nilsson J, Jovinge S. Inhibition of tumor necrosis factor-alpha reduces atherosclerosis in apolipoprotein e knockout mice. *Arteriosclerosis, thrombosis, and vascular biology*. 2004;24:2137-2142
111. Kirii H, Niwa T, Yamada Y, Wada H, Saito K, Iwakura Y, Asano M, Moriaki H, Seishima M. Lack of interleukin-1beta decreases the severity of atherosclerosis in apoe-deficient mice. *Arteriosclerosis, thrombosis, and vascular biology*. 2003;23:656-660
112. Davenport P, Tipping PG. The role of interleukin-4 and interleukin-12 in the progression of atherosclerosis in apolipoprotein e-deficient mice. *The American journal of pathology*. 2003;163:1117-1125
113. Elhage R, Jawien J, Rudling M, Ljunggren HG, Takeda K, Akira S, Bayard F, Hansson GK. Reduced atherosclerosis in interleukin-18 deficient apolipoprotein e-knockout mice. *Cardiovasc Res*. 2003;59:234-240
114. Emeson EE, Shen ML, Bell CG, Qureshi A. Inhibition of atherosclerosis in cd4 t-cell-ablated and nude (nu/nu) c57bl/6 hyperlipidemic mice. *The American journal of pathology*. 1996;149:675-685
115. Huber SA, Sakkinen P, David C, Newell MK, Tracy RP. T helper-cell phenotype regulates atherosclerosis in mice under conditions of mild hypercholesterolemia. *Circulation*. 2001;103:2610-2616
116. Binder CJ, Hartvigsen K, Chang MK, Miller M, Broide D, Palinski W, Curtiss LK, Corr M, Witztum JL. Il-5 links adaptive and natural immunity specific for epitopes of oxidized ldl and protects from atherosclerosis. *The Journal of clinical investigation*. 2004;114:427-437
117. Miller AM, Xu D, Asquith DL, Denby L, Li Y, Sattar N, Baker AH, McInnes IB, Liew FY. Il-33 reduces the development of atherosclerosis. *The Journal of experimental medicine*. 2008;205:339-346
118. Cardilo-Reis L, Gruber S, Schreier SM, Drechsler M, Papac-Milicevic N, Weber C, Wagner O, Stangl H, Soehnlein O, Binder CJ. Interleukin-13 protects from atherosclerosis and modulates plaque composition by skewing the macrophage phenotype. *EMBO Mol Med*. 2012;4:1072-1086
119. King VL, Szilvassy SJ, Daugherty A. Interleukin-4 deficiency decreases atherosclerotic lesion formation in a site-specific manner in female ldl receptor^{-/-} mice. *Arteriosclerosis, thrombosis, and vascular biology*. 2002;22:456-461

120. van Wanrooij EJ, van Puijvelde GH, de Vos P, Yagita H, van Berkel TJ, Kuiper J. Interruption of the tnfrsf4/tnfsf4 (ox40/ox40l) pathway attenuates atherogenesis in low-density lipoprotein receptor-deficient mice. *Arteriosclerosis, thrombosis, and vascular biology*. 2007;27:204-210
121. Shimoda K, van Deursen J, Sangster MY, Sarawar SR, Carson RT, Tripp RA, Chu C, Quelle FW, Nosaka T, Vignali DA, Doherty PC, Grosveld G, Paul WE, Ihle JN. Lack of il-4-induced th2 response and ige class switching in mice with disrupted stat6 gene. *Nature*. 1996;380:630-633
122. Zheng W, Flavell RA. The transcription factor gata-3 is necessary and sufficient for th2 cytokine gene expression in cd4 t cells. *Cell*. 1997;89:587-596
123. Kurowska-Stolarska M, Kewin P, Murphy G, Russo RC, Stolarski B, Garcia CC, Komai-Koma M, Pitman N, Li Y, Niedbala W, McKenzie AN, Teixeira MM, Liew FY, Xu D. Il-33 induces antigen-specific il-5+ t cells and promotes allergic-induced airway inflammation independent of il-4. *J Immunol*. 2008;181:4780-4790
124. Bettelli E, Carrier Y, Gao W, Korn T, Strom TB, Oukka M, Weiner HL, Kuchroo VK. Reciprocal developmental pathways for the generation of pathogenic effector th17 and regulatory t cells. *Nature*. 2006;441:235-238
125. Veldhoen M, Hocking RJ, Atkins CJ, Locksley RM, Stockinger B. Tgfbeta in the context of an inflammatory cytokine milieu supports de novo differentiation of il-17-producing t cells. *Immunity*. 2006;24:179-189
126. Mangan PR, Harrington LE, O'Quinn DB, Helms WS, Bullard DC, Elson CO, Hatton RD, Wahl SM, Schoeb TR, Weaver CT. Transforming growth factor-beta induces development of the t(h)17 lineage. *Nature*. 2006;441:231-234
127. Park H, Li Z, Yang XO, Chang SH, Nurieva R, Wang YH, Wang Y, Hood L, Zhu Z, Tian Q, Dong C. A distinct lineage of cd4 t cells regulates tissue inflammation by producing interleukin 17. *Nature immunology*. 2005;6:1133-1141
128. Steinman L. A brief history of t(h)17, the first major revision in the t(h)1/t(h)2 hypothesis of t cell-mediated tissue damage. *Nature medicine*. 2007;13:139-145
129. Erbel C, Dengler TJ, Wangler S, Lasitschka F, Bea F, Wambsgans N, Hakimi M, Bockler D, Katus HA, Gleissner CA. Expression of il-17a in human atherosclerotic lesions is associated with increased inflammation and plaque vulnerability. *Basic Res Cardiol*. 2011;106:125-134
130. Eid RE, Rao DA, Zhou J, Lo SF, Ranjbaran H, Gallo A, Sokol SI, Pfau S, Pober JS, Tellides G. Interleukin-17 and interferon-gamma are produced concomitantly by human coronary artery-infiltrating t cells and act synergistically on vascular smooth muscle cells. *Circulation*. 2009;119:1424-1432
131. Patel S, Chung SH, White G, Bao S, Celermajer DS. The "atheroprotective" mediators apolipoprotein a-i and foxp3 are over-abundant in unstable carotid plaques. *Int J Cardiol*. 2010;145:183-187
132. Gao Q, Jiang Y, Ma T, Zhu F, Gao F, Zhang P, Guo C, Wang Q, Wang X, Ma C, Zhang Y, Chen W, Zhang L. A critical function of th17 proinflammatory cells in the development of atherosclerotic plaque in mice. *Journal of immunology*. 2010;185:5820-5827
133. Smith E, Prasad KM, Butcher M, Dobrian A, Kolls JK, Ley K, Galkina E. Blockade of interleukin-17a results in reduced atherosclerosis in apolipoprotein e-deficient mice. *Circulation*. 2010;121:1746-1755
134. van Es T, van Puijvelde GH, Ramos OH, Segers FM, Joosten LA, van den Berg WB, Michon IM, de Vos P, van Berkel TJ, Kuiper J. Attenuated atherosclerosis upon il-17r signaling disruption in ldlr deficient mice. *Biochem Biophys Res Commun*. 2009;388:261-265
135. Erbel C, Chen L, Bea F, Wangler S, Celik S, Lasitschka F, Wang Y, Bockler D, Katus HA, Dengler TJ. Inhibition of il-17a attenuates atherosclerotic lesion development in apoe-deficient mice. *Journal of immunology*. 2009;183:8167-8175
136. Madhur MS, Funt SA, Li L, Vinh A, Chen W, Lob HE, Iwakura Y, Blinder Y, Rahman A, Quyyumi AA, Harrison DG. Role of interleukin 17 in inflammation, atherosclerosis, and vascular function in apolipoprotein e-deficient mice. *Arteriosclerosis, thrombosis, and vascular biology*. 2011;31:1565-1572
137. Taleb S, Romain M, Ramkhalawon B, Uyttenhove C, Pasterkamp G, Herbin O, Esposito B, Perez N, Yasukawa H, Van Snick J, Yoshimura A, Tedgui A, Mallat Z. Loss of socs3 expression in t cells reveals a regulatory role for interleukin-17 in atherosclerosis. *The Journal of experimental medicine*. 2009;206:2067-2077
138. von Boehmer H. Mechanisms of suppression by suppressor t cells. *Nat Immunol*. 2005;6:338-344
139. Nakamura K, Kitani A, Strober W. Cell contact-dependent immunosuppression by cd4(+)cd25(+) regulatory t cells is mediated by cell surface-bound transforming growth factor beta. *J Exp Med*. 2001;194:629-644
140. Hori S, Nomura T, Sakaguchi S. Control of regulatory t cell development by the transcription factor foxp3. *Science*. 2003;299:1057-1061
141. Bluestone JA, Abbas AK. Natural versus adaptive regulatory t cells. *Nat Rev Immunol*. 2003;3:253-257
142. Maggi E, Cosmi L, Liotta F, Romagnani P, Romagnani S, Annunziato F. Thymic regulatory t cells. *Autoimmun Rev*. 2005;4:579-586
143. Sakaguchi S. Regulatory t cells: Key controllers of immunologic self-tolerance. *Cell*. 2000;101:455-458
144. Jordan MS, Boesteanu A, Reed AJ, Petrone AL, Holenbeck AE, Lerman MA, Naji A, Caton AJ. Thymic selection of cd4+cd25+ regulatory t cells induced by an agonist self-peptide. *Nat Immunol*. 2001;2:301-306
145. Kronenberg M, Rudensky A. Regulation of immunity by self-reactive t cells. *Nature*. 2005;435:598-604
146. Bacchetta R, Gregori S, Roncarolo MG. Cd4+ regulatory t cells: Mechanisms of induction and effector function. *Autoimmun Rev*. 2005;4:491-496
147. Groux H, O'Garra A, Bigler M, Rouleau M, Antonenko S, de Vries JE, Roncarolo MG. A cd4+ t-cell subset inhibits antigen-specific t-cell responses and prevents colitis. *Nature*. 1997;389:737-742
148. O'Garra A, Vieira P. Regulatory t cells and mechanisms of immune system control. *Nat Med*. 2004;10:801-805

149. Vieira PL, Christensen JR, Minaae S, O'Neill EJ, Barrat FJ, Boonstra A, Barthlott T, Stockinger B, Wraith DC, O'Garra A. IL-10-secreting regulatory t cells do not express foxp3 but have comparable regulatory function to naturally occurring cd4+cd25+ regulatory t cells. *J Immunol.* 2004;172:5986-5993
150. Chen Y, Kuchroo VK, Inobe J, Hafler DA, Weiner HL. Regulatory t cell clones induced by oral tolerance: Suppression of autoimmune encephalomyelitis. *Science.* 1994;265:1237-1240
151. Weiner HL. Induction and mechanism of action of transforming growth factor-beta-secreting th3 regulatory cells. *Immunol Rev.* 2001;182:207-214
152. Marson A, Kretschmer K, Frampton GM, Jacobsen ES, Polansky JK, MacIsaac KD, Levine SS, Fraenkel E, von Boehmer H, Young RA. Foxp3 occupancy and regulation of key target genes during t-cell stimulation. *Nature.* 2007;445:931-935
153. Sakaguchi S, Sakaguchi N, Asano M, Itoh M, Toda M. Immunologic self-tolerance maintained by activated t cells expressing il-2 receptor alpha-chains (cd25). Breakdown of a single mechanism of self-tolerance causes various autoimmune diseases. *J Immunol.* 1995;155:1151-1164
154. McHugh RS, Whitters MJ, Piccirillo CA, Young DA, Shevach EM, Collins M, Byrne MC. Cd4(+)cd25(+) immunoregulatory t cells: Gene expression analysis reveals a functional role for the glucocorticoid-induced tnfr receptor. *Immunity.* 2002;16:311-323
155. Sakaguchi S. Naturally arising foxp3-expressing cd25+cd4+ regulatory t cells in immunological tolerance to self and non-self. *Nat Immunol.* 2005;6:345-352
156. Mor A, Planer D, Luboshits G, Afek A, Metzger S, Chajek-Shaul T, Keren G, George J. Role of naturally occurring cd4+cd25+ regulatory t cells in experimental atherosclerosis. *Arteriosclerosis, thrombosis, and vascular biology.* 2007;27:893-900
157. Ait-Oufella H, Salomon BL, Potteaux S, Robertson AK, Gourdy P, Zoll J, Merval R, Esposito B, Cohen JL, Fisson S, Flavell RA, Hansson GK, Klatzmann D, Tedgui A, Mallat Z. Natural regulatory t cells control the development of atherosclerosis in mice. *Nat Med.* 2006;12:178-180
158. van Es T, van Puijvelde GH, Foks AC, Habets KL, Bot I, Gilboa E, Van Berkel TJ, Kuiper J. Vaccination against foxp3(+) regulatory t cells aggravates atherosclerosis. *Atherosclerosis.* 209:74-80
159. van Puijvelde GH, Hauer AD, de Vos P, van den Heuvel R, van Herwijnen MJ, van der Zee R, van Eden W, van Berkel TJ, Kuiper J. Induction of oral tolerance to oxidized low-density lipoprotein ameliorates atherosclerosis. *Circulation.* 2006;114:1968-1976
160. van Puijvelde GH, van Es T, van Wanrooij EJ, Habets KL, de Vos P, van der Zee R, van Eden W, van Berkel TJ, Kuiper J. Induction of oral tolerance to hsp60 or an hsp60-peptide activates t cell regulation and reduces atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2007;27:2677-2683
161. Binder CJ, Chang MK, Shaw PX, Miller YI, Hartvigsen K, Dewan A, Witztum JL. Innate and acquired immunity in atherogenesis. *Nat Med.* 2002;8:1218-1226
162. Tedgui A, Mallat Z. Cytokines in atherosclerosis: Pathogenic and regulatory pathways. *Physiol Rev.* 2006;86:515-581
163. Mallat Z, Gojova A, Marchiol-Fournigault C, Esposito B, Kamate C, Merval R, Fradelizi D, Tedgui A. Inhibition of transforming growth factor-beta signaling accelerates atherosclerosis and induces an unstable plaque phenotype in mice. *Circ Res.* 2001;89:930-934
164. Robertson AK, Rudling M, Zhou X, Gorelik L, Flavell RA, Hansson GK. Disruption of tgf-beta signaling in t cells accelerates atherosclerosis. *The Journal of clinical investigation.* 2003;112:1342-1350
165. de Boer OJ, van der Meer JJ, Teeling P, van der Loos CM, van der Wal AC. Low numbers of foxp3 positive regulatory t cells are present in all developmental stages of human atherosclerotic lesions. *PLoS ONE.* 2007;2:e779

