



Universiteit
Leiden
The Netherlands

The immune divide: factors influencing immune variation and differences in vaccine responses

Pyuza, J.J.

Citation

Pyuza, J. J. (2025, November 25). *The immune divide: factors influencing immune variation and differences in vaccine responses*. Retrieved from <https://hdl.handle.net/1887/4283867>

Version: Publisher's Version

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/4283867>

Note: To cite this publication please use the final published version (if applicable).

3

Current status of schistosomiasis in school-aged children in mwanga district, Tanzania: Impact of two decades of annual mass drug administration programme.

Jeremia J. Pyuza^{1,2,3,6}, Brice Meulah^{1,4,6}, Pytsje T. Hoekstra¹, Noel Mdende³, Elizabeth Mvilli³, Lisette van Lieshout¹, Stan T. Hilt^{1,5}, Paul L. A. M. Corstjens⁵, Maria Yazdanbakhsh¹, Sia E. Msuya^{3,6} and Govert J. van Dam¹

Published: Cambridge University Press, 2024. doi:10.1017/S0031182024001045

1. Leiden University Center for Infectious Diseases (LUCID), Leiden University Medical Center, Leiden, The Netherlands;
2. Department of Pathology Kilimanjaro Christian Medical Center, Moshi, Tanzania;
3. Institute of Public Health, Kilimanjaro Christian University Medical College (KCMUCo), Moshi Tanzania;
4. Centre de Recherches Médicales des Lambaréné (CERMEL) Lambaréné, Gabon;
5. Department of Cell and Chemical Biology Leiden University Medical Center, Leiden, The Netherlands and 6 Department of Community Medicine, Kilimanjaro Christian Medical Center, Moshi, Tanzania
6. First and second authors contributed equally to this work (Shared first authorship).

Abstract

Schistosomiasis is a neglected tropical disease with significant health implications, particularly among children. A cross-sectional study was conducted among school-aged children (SAC) in Mwanga district, Tanzania, a region known to be co-endemic for *S. haematobium* and *S. mansoni* infection and where annual mass drug administration (MDA) has been conducted for 20 years. In total, 576 SAC from 5 schools provided a urine sample for the detection of *Schistosoma* circulating anodic antigen using the upconverting particle-based lateral flow (UCP-LF CAA) test. Additionally, the potential of the point-of-care circulating cathodic antigen (POC-CCA) and microhaematuria dipstick test as field-applicable diagnostic alternatives for schistosomiasis were assessed and the prevalence outcome compared to UCP-LF CAA. Risk factors associated with schistosomiasis was assessed based on UCP-LF CAA. The UCP-LF CAA test revealed an overall schistosomiasis prevalence of 20.3%, compared to 65.3% based on a combination of POC-CCA and microhaematuria dipstick. No agreement was observed between the combined POC tests and UCP-LF CAA. Factors associated with schistosomiasis included age (5–10 years), involvement in fishing, farming, swimming activities and attending 2 of the 5 primary schools. Our findings suggest a significant progress in infection control in Mwanga district due to annual MDA, although not enough to interrupt transmission. Accurate diagnostics play a crucial role in monitoring intervention measures to effectively combat schistosomiasis.

Introduction

Schistosomiasis is a major neglected tropical disease disproportionately affecting sub-Saharan African countries, with 90.0% of the global disease burden occurring in this region[1]. In Tanzania, the overall prevalence of schistosomiasis is 51.5% [2], and among school-aged children (SAC) it is reported to be 53.5%[3], reaching up to about 80.0% in the northwestern zone around Lake Victoria[4,5]. However, current prevalence estimates do not include the northern region of Tanzania, including Mwanga district in the Kilimanjaro region. This district has been known to be endemic for both *Schistosoma haematobium* and *Schistosoma mansoni* [6]. The population is at high risk of schistosomiasis possibly due to the presence of the intermediate snail host (*Bulinus* and *Biomphalaria*) as well as irrigation schemes, which are the conducive environment for the transmission the *Schistosoma* spp. The presence of the hydroelectric dam known as ‘Nyumba ya Mungu’ (Fig. 1) which ensures a constant water supply to the surrounding villages for irrigation contributes to the continues transmission of schistosomiasis in Mwanga district [5]. In Tanzania, including the Kilimanjaro region, mass drug administration (MDA) of praziquantel has been the major strategy to reduce the burden of schistosomiasis and has been organized annually since 2004 among SAC who are at the highest risk of infection[7]. The need to assess the success of MDA has been highlighted by the World Health Organization and tools to enhance strategic guidance for schistosomiasis control program in Tanzania have equally been reported[8,9]. The most recent data on the prevalence status of schistosomiasis among SAC in Mwanga district are from 2005, indicating a prevalence ranging from 33.5 to 70.0%[6]. Conventional microscopy is the reference method to diagnose schistosomiasis in endemic settings and involves the detection of *Schistosoma* eggs in feces or urine, depending on the species. However, microscopy requires experienced, well-trained technical personnel, is considered a time-consuming, laborious method and has limited sensitivity in low-intensity infection settings[10]. Furthermore, the availability and access to microscopy is challenging in many rural areas in Tanzania due to a lack of trained personnel and appropriate infrastructure. Low-cost, user-friendly rapid tests could overcome such issues, but the accuracy of available point-of-care (POC) tests to determine the prevalence of schistosomiasis in regions co-endemic for *S. haematobium* and *S. mansoni* needs to be determined. The POC test for detecting *Schistosoma* Circulating Cathodic Antigen (POC-CCA) in urine has been endorsed by the WHO as an alternative to conventional microscopy, in particular for the diagnosis of *S. mansoni* infections[9]. It requires minimal training and has been validated in several field settings endemic for intestinal schistosomiasis[11, 12]. Another

easy-to-use rapid test is the microhaematuria dipstick test for the detection of haematuria, which has been shown to be strongly associated with urogenital schistosomiasis, although it is considered nonspecific[13]. A quantitative Up-Converting reporter Particle Lateral Flow (UCP-LF) test detecting the genus-specific *Schistosoma* Circulating Anodic Antigen (CAA) is a highly accurate test to detect active infection of all *Schistosoma* species in urine or serum[14]. [14]. Although it requires a more advanced laboratory infrastructure, it has been shown to be 100% specific and can reach a sensitivity to detect single-worm infections[14,15,16]. This study aimed to determine the current prevalence of schistosomiasis in the Mwanga district Tanzania after approximately twenty years of MDA using the UCP-LF CAA test and to explore the potential of using the POC-CCA and microhaematuria dipstick as a combined POC test for diagnosing schistosomiasis in co-endemic settings in comparison to the laboratory-based UCP-LF CAA test. Furthermore, we investigated *Schistosoma* infection risk factors and associated parameters.

Materials and methods

Ethical considerations

Ethical approval for this study was obtained from Kilimanjaro Christian Medical University College (KCMUCo) research and ethical committee board (reference number: 2588). Administrative authorization was obtained from the district education officer and Mwanga District Medical Officer. Children were enrolled based on their availability, and those willing to participate were given a consent form to be signed by guardians and/or parents. Immediately after sample collection all children, including those who participated in our study, were provided with praziquantel under the yearly MDA program at school at the recommended dose in the presence of a local clinician.

Study area and population

The study was conducted in Mwanga district, one of the seven districts of Kilimanjaro region in Tanzania. Farming, fishing, sand collection, pebble making, soil bricking and animal keeping are the major economic activities. The study was conducted in five schools, of which two were selected based on a previous study[6]. MDA of praziquantel had occurred in these schools more than 6 months prior to our study.

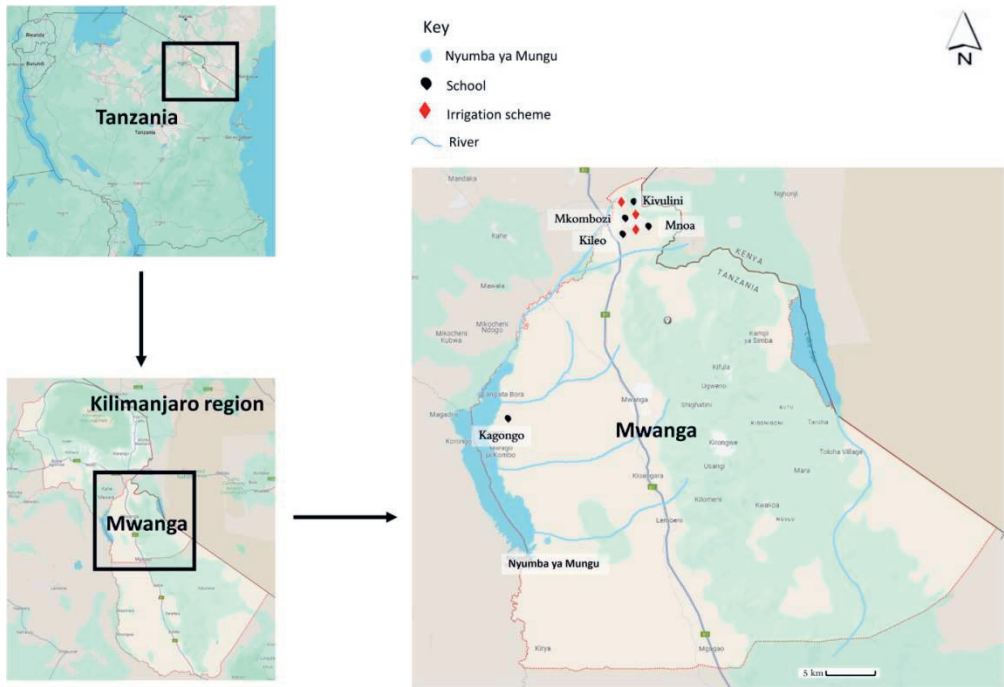


Figure 1. Map of Mwanga district Tanzania showing water sources, five primary schools and irrigation schemes

Sample/ data collection and processing

Enrolled study participants provided consent forms from parents and were given sterile containers with unique identifiers to provide fresh urine samples. For each sample, POC-CCA and microhaematuria dipstick was done in the field for the diagnosis of *S. mansoni* and *S. haematobium*, respectively. An aliquot of 2 mL of urine was conserved (at -20°C) per participant and was shipped on ice to Leiden University Medical Center in the Netherlands for retrospective UCP-LF CAA analysis. Following urine sample collection, a face-to-face interview using a closed-ended questionnaire in English and Swahili was conducted.

Field and laboratory analysis

The POC-CCA test (batch 180817091) was obtained from Rapid Medical Diagnostics, South Africa (SA), and analysis was done according to the manufacturer's instructions. Briefly, two drops of urine were transferred into the sample window of the test cassette. The readout of the cassette was done in 20 minutes. Results were scored as negative, trace or positive. Microhaematuria dipstick (Mission Urinalysis, Lot no: URS8090018) test was performed by

placing the strip on a flat surface and a drop of urine applied to the reagent pad. Readouts were done in 1 minute as either negative or positive according to the manufacturer's instructions. The *Schistosoma* genus-specific UCP-LF CAA test was employed to detect CAA in urine samples and to confirm active infection with *Schistosoma* spp[14]. All urine samples were subjected to the UCAAhT417 wet format of the test. Briefly, 500 μ L of each urine sample was mixed with 100 μ L of 12% trichloroacetic acid, then incubated at room temperature for 5 minutes and centrifuged. The clear supernatant was then concentrated to 20 μ L using a 0.5 mL centrifugal device (Amicon Ultra-0.5, Millipore, Merck Chemicals B.V., Amsterdam, The Netherlands). The resulting concentrate was then applied to the lateral flow test strip. To quantify CAA concentrations and to validate the assay cutoff (0.6 pg mL⁻¹), reference standards with known CAA-levels were included. A CAA concentration above 0.6 pg mL⁻¹ was considered positive. Infection intensity was categorized as low positive (>0.6–10 pg mL⁻¹), moderate positive (>10–100 pg mL⁻¹) and high positive (>100 pg mL⁻¹).

Statistical analysis

The agreement between the combination of POC-CCA and microhaematuria dipstick (combined POC test) and UCP-LF CAA was performed using Kappa (K) statistics. For POC-CCA, trace results were considered negative. Furthermore, the association between socio-demographic characteristics and risk factors associated with *Schistosoma* infection, based on the UCP-LF CAA test, was performed using chi-square statistics, binary and multiple logistic regression analyses. Statistical analysis was performed using IBM Statistical Package for Social Sciences version 29 (SPSS Inc., Chicago, United States of America). For generation of plots, GraphPad Prism version 9.3.1 for Windows (GraphPad Software, San Diego, California USA, www.graphpad.com) was used.

Results

Socio-demographic characteristics

A total of 576 children provided informed consent and subsequently provided a urine sample and were therefore included in the final analysis. In Table 1 socio-demographic characteristics of the study population are given. The children's age ranged from 5 to 16 years, with a mean age of 9.8 years (S.D. 2.4) and 50.7% were females. Furthermore, the majority of the children were in class range 1 to 3 (50.3%). Farming was the most common father's profession

(46.7%) followed by fishing (23.4%). The most common mother's profession was farming (44.8%), followed by small businesses (34.2%).

Table 1. The prevalence of schistosomiasis across all five schools based on UCP-LF CAA, POC-CCA, microhaematuria dipstick and a combination of the POC-CCA and microhaematuria dipstick

			Diagnostic test			
			UCP-LF CAA	POC-CCA	Microhaematuria dipstick	Combined Test ^a
Number of children			Positive (%)	Positive (%)	Positive (%)	Positive (%)
School	Kagongo	279	35 (12.5)	174 (62.4)	84 (30.1)	209 (74.9)
	Kileo	59	14 (23.7)	37 (62.7)	12 (20.3)	41 (69.5)
	Kivulini	57	19 (33.3)	30 (52.6)	12 (21.1)	36 (63.2)
	Mkombozi	106	31 (22.2)	35 (33.1)	23 (21.6)	43 (40.6)
	Mnoa	75	18 (24.0)	38 (50.6)	17 (22.7)	47 (62.7)
Total		576	117 (20.3)	314 (54.5)	148 (25.7)	376 (65.3)

^a Combination of POC-CCA and/or microhaematuria dipstick positive outcome.

Prevalence and intensity of *Schistosoma* infection

In total 117 (20.3%) of children were found to be CAA positive (Table 1). The highest proportion of positives was observed among children attending Kivulini primary school (33.3%), followed by Mkombozi primary school (29.2%). The lowest proportion was 12.5% and was found among school children at Kagongo primary school. The majority of moderate to high intensity infections based on CAA-levels was observed in the schools in Kivulini and Mkombozi (Fig. 2a). Furthermore, SAC within the age category 5–10 years were found to be more often CAA positive than those aged 11–16 years (Fig. 2b). The outcome of the point-of-care tests are also summarized in Table 1. Based on POC-CCA and microhaematuria dipstick tests the prevalence of schistosomiasis were, 54.5% and 25.7% respectively. Assuming that the combination of two tests will give more clearer prevalence, combining POC-CCA and microhaematuria dipstick, the prevalence was 65.3%.

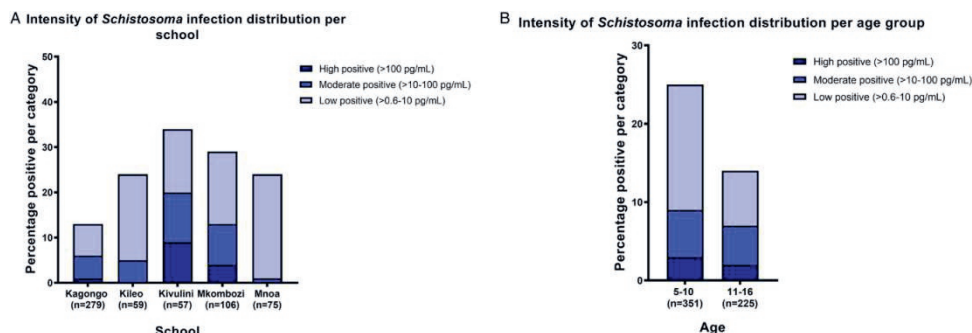


Figure 2. Prevalence and intensity of *Schistosoma* infection based on UCP-LF CAA amongst (A), five selected schools (B) the age categories.

The agreement between the combination of POC-CCA and microhaematuria dipstick and the reference test UCP-LF CAA

The combination of POC-CCA and microhaematuria dipstick showed no agreement with the UCP-LF CAA test (Table 2). Furthermore the *P* value indicates that this lack of agreement is not statistically significant, suggesting that the disagreement between tests is likely due to chance at significance level of 0.05. Analysis of the individual POC-CCA and microhaematuria dipstick tests also showed no agreement with the UCP-LF CAA test.

Table 2. The level of agreement between point-of-care circulating cathodic antigen (POC-CCA) test, microhaematuria dipstick and upconverting particle lateral flow circulating anodic antigen (UCP-LF CAA) urine test by Cohen's kappa coefficient in 576 school-aged children from Mwanza Tanzania

Test		UCP-LF CAA				Interpretation
		Positive	Negative	K-value	P Value	
POC-CCA	Positive	86	367	0.011	0.128	Poor
	Negative	31	92			
Microhaematuria dipstick	Positive	37	110	0.013	0.090	Poor
	Negative	80	349			
Combined (Microhaematuria and POC-CCA)	Positive	79	297	0.015	0.569	Poor
	Negative	38	162			

Risk factors associated with schistosomiasis

Using multivariate logistic regression analysis (adjusted odd ratio), the potential risk factors associated with *Schistosoma* infection, based on the presence of CAA, showed that children in class level 1–3 had two times higher odds of having schistosomiasis than children in higher classes. Children involved in farming and swimming activities had respectively 5.6 and 3.6

odds of being infected than those who did not farm nor swim. Furthermore, children attending Kileo, Kivulini, Mkombozi and Mnoa primary schools had 2.2, 2.6, 2.4 and 2.7 times higher odds of CAA positive results respectively, when compared to those attending Kagongo primary school. More details can be found in Supplementary Table 1.

Discussion

Using a highly accurate diagnostic approach (UCP-LF CAA), this study indicated that after nearly two decades of MDA schistosomiasis remains highly prevalent (20%) among school-aged children in Mwanga district, Tanzania. Although POC-CCA and microhaematuria dipstick test showed an even higher prevalence than the UCP-LF CAA test, no agreement was found between these tests and the UCP-LF CAA results nor any association was observed between these tests and known risk factors for schistosomiasis, highlighting the limitation of these currently available rapid diagnostics tests (POC-CCA and microhaematuria) in accurately determining the true prevalence in this specific setting that is known to be co-endemic for *S. mansoni* and *S. haematobium*.

Different prevalence's have been observed throughout Tanzania[17,18,19,20,21]. As commonly known as well as shown in the current study, measurement of prevalence highly depends on the diagnostic method used. Since we have used a highly accurate diagnostic method in our study, i.e. the UCP-LF CAA test, it is difficult to directly compare our results to previously published results based on microscopy and/or POC-CCA as these methods have limited sensitivity/ specificity. Our data confirm regional variation in the burden of schistosomiasis in Mwanga district, which would, extrapolated to Tanzania as a whole, argue for a more focally oriented schistosomiasis control approach. A significant difference in infection rates among different age groups was identified. Younger children (5–10 years) exhibited a higher prevalence of schistosomiasis than the older age group (11–16 years), indicating early exposure to the infection[20]. The possible reason for older children having low prevalence is through acquired immunity due repeated infections as indicated by other previous studies for example possible presence of IgE antibodies[22, 23]. A significant association was found between schistosomiasis and children who swim in water bodies, which may be attributed to playful behavioural activities common among children[24]. The children involved in swimming activities had 3.6 times more risk of being infected, in line with recent systematic review demonstrated by Reitzug and colleagues[25]. Children's involvement in farming was found to be associated with an increased risk of being infected with

schistosomiasis. Finally, children attending Kileo, Kivulini, Mkombozi and Mnoa primary schools were identified to have higher rates of *Schistosoma* infection compared to those attending Kagongo primary school. These findings are likely due to the proximity of irrigation schemes/rivers to these schools, which are perceived as safer water sources for young children and so most likely to visit compared to larger water bodies like the dams located closer to Kagongo, where parents have concerns about the risk of children drowning and so cautioned not to go there for water.

Despite of providing crucial updates on the prevalence of schistosomiasis among school children in the area, the study has several limitations. Firstly, the laboratory UCP-LF CAA test detects all *Schistosoma* species, but it does not provide any species-specific information[14]. In control settings, where species information would be relevant, other measures can provide this, e.g. determining the presence of specific snail species, or performing egg microscopy or species-specific PCR. For treatment, species information as such is not needed, and CAA has been demonstrated to be an excellent marker for monitoring treatment efficacy[26,16,27,28]. Although POC-CCA and microhaematuria rapid tests are user-friendly, kappa statistics revealed a poor agreement between these tests and the UCP-LF CAA. It was expected that the combined positivity rates of POC-CCA and microhaematuria dipstick test would reflect the UCP-LF CAA results, however this was not the case (Table 1). The poor agreement may be due to production batch differences, sexually transmitted infections (STIs), low-intensity infections, and subjectivity to test readouts, which might also affect results[29]. Furthermore, more accurate result with both tests might have been possible if the test was scored in a more quantitative manner. For example, following the recently described G-score scoring method for POC-CCA, an inclusion of control samples as a way to standardize the readout and to determine the cut-off for positivity. The microhaematuria test can be scored semi-quantitatively based on colour intensity linked to the level of red blood cells. However, registering of more quantitative results was not foreseen in this study.

In conclusion, this study demonstrates a moderate prevalence of schistosomiasis in Mwanga district Tanzania, implicating that the ~20 years annual MDA of praziquantel in this region may have had an effect on reducing the schistosomiasis burden, but transmission is still ongoing. To improve the efficacy of MDA strategies, diagnosis at acute stages of the disease in combination with treatment could be extended not only to higher risk groups but also to all persons above 2 years of age as recommended by WHO[30, 9]. Apart from that, integrated approaches including improved access to WASH infrastructure, political willingness, and

production of reliable data are important for controlling schistosomiasis in Tanzania[31]. The presence of persistent hotspots in countries like Tanzania where provision of MDA program failed to provide long terms solution in some villages shows the need for such an integrated approach[32]. Furthermore, a combination of the POC-CCA and microhaematuria dipstick did not prove to be useful as a screening tool for schistosomiasis in this *S. haematobium* and *S. mansoni* co-endemic setting. However, efforts are ongoing to make CAA detection generally available, with a recent initiative focusing on the development of a more easy-to-use, accurate, affordable and visually scored CAA-RDT[33,34]. The CAA-RDT could be of great potential in resource-poor endemic settings and assist in the development of targeted control measures and interventions to effectively combat schistosomiasis.

Supplementary material

The supplementary material for this article can be found at <https://doi.org/10.1017/S0031182024001045>.

Data availability statement

The authors confirm that the data supporting the findings of this study are available within the article and its supplementary and that the raw data is available upon request to the corresponding author.

Acknowledgements

We gratefully acknowledge Claudia de Dood (LUMC) for her assistance in performing the CAA analysis, Dieuwke Kornelis (LUMC) for technical assistance with the POC-CCA, and George A. Masisila for assisting in data entry and POC-CCA processing.

Authors' contributions

Conceptualization of the study: J.J.P, S.E.M, G.J.v.D, M.Y. Data collection, analysis interpretation of study findings: J.J.P, B.M, P.T.H, L.v.L, E.M, N.M, S.H, P.L.A.M.C, S.E.M, G.J.v.D, Editing: J.J.P, B.M, P.T.H, L.v.L, P.L.A.M.C, G.J.v.D; Original draft: J.J.P,B.M

Financial support

This study was funded by LUMC Global PhD fellowship fund which supported Jeremia J.Pyuza, LUMC-SF-GLOBAL and partly by NWO-WOTRO Science for Global Development program, Grant Number W 07.30318.009 (INSPIRED – Inclusive diagnostics for Poverty Related parasitic Diseases in Nigeria and Gabon) which supported Brice Meulah. This work is part of the EDCTP2 program supported by the European Union.

Competing interests

None.

Reference:

1. World Health Organization (WHO) (2023) Schistosomiasis facts 2023. Available at <https://www.who.int/news-room/fact-sheets/detail/schistosomiasis>. (accessed 1 July 2023).
2. Mazigo HD, Nuwaha F, Kinung'hi SM, Morona D, de Moira AP, Wilson S, Heukelbach J and Dunne DW (2012) Epidemiology and control of human schistosomiasis in Tanzania. *Parasites & Vectors* 5, 274.
3. Kavana NJ (2018) Prevalence of schistosomiasis infection among young children aged 5 to 17 years in Kilosa District, Tanzania: a 3 year retrospective review. *Journal of Tropical Diseases* 6, 255
4. Munisi DZ, Buza J, Mpolya EA and Kinung'hi SM (2016) Intestinal schistosomiasis among primary schoolchildren in two on-shore communities in Rorya district, northwestern Tanzania: prevalence, intensity of infection and associated risk factors. *Journal of Parasitology Research* 2016, 1–11.
5. Bakuza JS, Denwood MJ, Nkwengulila G and Mable BK (2017) Estimating the prevalence and intensity of *Schistosoma mansoni* infection among rural communities in Western Tanzania: the influence of sampling strategy and statistical approach. *PLOS Neglected Tropical Diseases* 11, e0005937.
6. Poggensee G, Krantz I, Nordin P, Mtweve S, Ahlberg B, Mosha G and Freudenthal S (2005) A six-year follow-up of schoolchildren for urinary and intestinal schistosomiasis and soil-transmitted helminthiasis in Northern Tanzania. *Acta Tropica* 93, 131–140.
7. Mwanga JR, Kinung'hi SM, Mosha J, Angelo T, Maganga J and Campbell CH (2020) Village response to mass drug administration for schistosomiasis in Mwanza Region, Northwestern Tanzania: are we missing socioeconomic, cultural, and political dimensions? *The American Journal of Tropical Medicine and Hygiene* 103, 1969–1977.
8. Clements ACA, Lwambo NJS, Blair L, Nyandindi U, Kaatano G, Kinung'hi S, Webster JP, Fenwick A and Brooker S (2006) Bayesian spatial analysis and disease mapping: tools to enhance planning and implementation of a schistosomiasis control programme in Tanzania. *Tropical Medicine & International Health* 11, 490–503..
9. World Health Organization (WHO) (2022) WHO guideline on Control and Elimination of Human Schistosomiasis. World Health Organization. Available at <https://www.who.int/publications/i/item/9789240041608>
10. Hoekstra PT, Madinga J, Lutumba P, van Grootveld R, Brienens EAT, Corstjens PLAM, van Dam GJ, Polman K and van Lieshout L (2022c) Diagnosis of schistosomiasis without a microscope: evaluating circulating antigen (CCA, CAA) and DNA detection methods on banked samples of a community-based survey from DR Congo. *Tropical Medicine and Infectious Disease* 7, 315.
11. Ochodo EA, Gopalakrishna G, Spek B, Reitsma JB, van Lieshout L, Polman K, Lamberton P, Bossuyt PM and Leeftang MM (2015) Circulating antigen tests and urine reagent strips for diagnosis of active schistosomiasis in endemic areas. *Cochrane Database of Systematic Reviews* 2015, CD009579.
12. Bärenbold O, Garba A, Colley DG, Fleming FM, Haggag AA, Ramzy RMR, Assaré RK,

- Tukahebwa EM, Mbonigaba JB, Bucumi V, Kebede B, Yibi MS, Meité A, Coulibaly JT, N'Goran EK, Tchuem Tchuenté L-A, Mwinzi P, Utzinger J and Vounatsou P (2018) Translating preventive chemotherapy prevalence thresholds for *Schistosoma mansoni* from the Kato-Katz technique into the point-of-care circulating cathodic antigen diagnostic test. *PLOS Neglected Tropical Diseases* 12, e0006941.
13. Emukah E, Gutman J, Eguagie J, Miri ES, Yinkore P, Okocha N, Jibunor V, Nebe O, Nwoye AI and Richards FO (2012) Urine heme dipsticks are useful in monitoring the impact of Praziquantel treatment on *Schistosoma haematobium* in sentinel communities of Delta State, Nigeria. *Acta Tropica* 122, 126–131.
 14. Corstjens PLAM, de Dood CJ, Knopp S, Clements MN, Ortu G, Umulisa I, Ruberanziza E, Wittmann U, Kariuki T, LoVerde P, Secor WE, Atkins L, Kinung'hi S, Binder S, Campbell CH, Colley DG and van Dam GJ (2020). Circulating anodic antigen (CAA): a highly sensitive diagnostic biomarker to detect active *Schistosoma* infections—improvement and use during SCORE. *The American Journal of Tropical Medicine and Hygiene* 103(1_Suppl), 50–57.
 15. Langenberg MCC, Hoogerwerf M-A, Koopman JPR, Janse JJ, Kos-van Oosterhoud J, Feijt C, Jochems SP, de Dood CJ, van Schuijlenburg R, Ozir-Fazalalikhani A, Manurung MD, Sartono E, van der Beek MT, Winkel BMF, Verbeek-Menken PH, Stam KA, van Leeuwen FWB, Meij P, van Diepen A, van Lieshout L, van Dam GJ, Corstjens PLAM, Hokke CH, Yazdanbakhsh M, Visser LG and Roestenberg M (2020) A controlled human *Schistosoma mansoni* infection model to advance novel drugs, vaccines and diagnostics. *Nature Medicine* 26, 326–332.
 16. Hoekstra PT, van Esbroeck M, de Dood CJ, Corstjens PLAM, Cnops L, van Zeijl-van der Ham CJG, Wammes LJ, van Dam GJ, Clerinx J and van Lieshout L (2021) Early diagnosis and follow-up of acute schistosomiasis in a cluster of infected Belgian travellers by detection of antibodies and circulating anodic antigen (CAA): a diagnostic evaluation study. *Travel Medicine and Infectious Disease* 41, 102053.
 17. Mnkugwe RH, Minzi OS, Kinung'hi SM, Kamuhabwa AA and Aklillu E (2020) Prevalence and correlates of intestinal schistosomiasis infection among school-aged children in North-Western Tanzania. *PLOS ONE* 15, e0228770
 18. Mazigo HD, Uisso C, Kazyoba P, Nshala A and Mwingira UJ (2021) Prevalence, infection intensity and geographical distribution of schistosomiasis among pre-school and school aged children in villages surrounding Lake Nyasa, Tanzania. *Scientific Reports* 11, 295.
 19. Kajembe VR, Gasarasi DB, Tarimo DS, Lushina M and Sylvester B (2022) Prevalence and factors associated with persistent transmission of *Schistosoma haematobium* among primary school children after five rounds of mass drug administration using praziquantel: a cross sectional study in Mkuranga district, Tanzania. *Tropical Doctor* 52, 526–531.
 20. Ogwenyo G, Mushi V, Silvestri V, Bonaventura W, Justine NC, Noah M, Yoram F, Mohamed H and Tarimo D (2023) Burden and risk factors for *Schistosoma mansoni* infection among primary school children: a quantitative school-based cross-sectional survey in Busega district, Northern Tanzania. *PLOS ONE* 18, e0280180.
 21. Maganga JK, Campbell CH, Angelo T, Masha J, Mwanga JR and Kinung'hi SM (2023) Test-Treat-Track-Test-Treat strategy for control of schistosomiasis in two low-prevalence villages in Northwestern Tanzania. *The American*

- Journal of Tropical Medicine and Hygiene 108, 1167–1174.
22. Woolhouse MEJ (1998) Patterns in parasite epidemiology: the peak shift. *Parasitology Today* 14, 428–434.
 23. Oettle R and Wilson S (2017) The interdependence between schistosome transmission and protective immunity. *Tropical Medicine and Infectious Disease* 2, 42.
 24. Munisi DZ, Buza J, Mpolya EA, Angelo T and Kinung'hi SM (2017) Knowledge, attitude, and practices on intestinal schistosomiasis among primary schoolchildren in the Lake Victoria basin, Rorya District, northwestern Tanzania. *BMC Public Health* 17, 731.
 25. Reitzug F, Ledien J and Chami GF (2023) Associations of water contact frequency, duration, and activities with schistosome infection risk: a systematic review and meta-analysis. *PLOS Neglected Tropical Diseases* 17, e0011377.
 26. Sousa MS, van Dam GJ, Pinheiro MCC, de Dood CJ, Peralta JM, Peralta RHS, Daher E de F, Corstjens PLAM and Bezerra FSM (2019) Performance of an ultra-sensitive assay targeting the circulating anodic antigen (CAA) for detection of *Schistosoma mansoni* infection in a low endemic area in Brazil. *Frontiers in Immunology* 10, 682.
 27. Hoekstra PT, Casacuberta-Partal M, van Lieshout L, Corstjens PLAM, Tsonaka R, Assaré RK, Silué KD, N'Goran EK, N'Gbesso YK, Brienens EAT, Roestenberg M, Knopp S, Utzinger J, Coulibaly JT and van Dam GJ (2022a) Limited efficacy of repeated praziquantel treatment in *Schistosoma mansoni* infections as revealed by highly accurate diagnostics, PCR and UCP-LF CAA (RePST trial). *PLOS Neglected Tropical Diseases* 16, e0011008.
 28. Hoekstra PT, Chernet A, de Dood CJ, Brienens EAT, Corstjens PLAM, Labhardt ND, Nickel B, Wammes LJ, van Dam GJ, Neumayr A and van Lieshout L (2022b) Sensitive diagnosis and post-treatment follow-up of *Schistosoma mansoni* infections in asymptomatic Eritrean refugees by circulating anodic antigen detection and polymerase chain reaction. *The American Journal of Tropical Medicine and Hygiene* 106, 1240–1246.
 29. Assaré RK, Tra-Bi MI, Coulibaly JT, Corstjens PLAM, Ouattara M, Hürlimann E, van Dam GJ, Utzinger J and N'Goran EK (2021). Accuracy of two circulating antigen tests for the diagnosis and surveillance of *Schistosoma mansoni* infection in low-endemicity settings of Côte d'Ivoire. *The American Journal of Tropical Medicine and Hygiene* 105, 677–683.
 30. Faust CL, Osakunor DNM, Downs JA, Kayuni S, Stothard JR, Lamberton PHL, Reinhard-Rupp J and Rollinson D (2020) Schistosomiasis control: leave no age group behind. *Trends in Parasitology* 36, 582–591.
 31. Abe EM, Tambo E, Xue J, Xu J, Ekpo UF, Rollinson D, Yang K, Li S-Z and Zhou X-N (2020) Approaches in scaling up schistosomiasis intervention towards transmission elimination in Africa: leveraging from the Chinese experience and lessons. *Acta Tropica* 208, 105379.
 32. Kittur N, King CH, Campbell CH, Kinung'hi S, Mwinzi PNM, Karanja DMS, N'Goran EK, Phillips AE, Gazzinelli-Guimaraes PH, Olsen A, Magnussen P, Secor WE, Montgomery SP, Utzinger J, Walker JW, Binder S and Colley DG (2019) Persistent hotspots in schistosomiasis consortium for operational research and evaluation studies for gaining and sustaining control of schistosomiasis after four years of mass drug administration of

praziquantel. *The American Journal of Tropical Medicine and Hygiene* 101, 617–627.

33. FIND (2020) Rapid test for precision mapping, and monitoring and evaluation of schistosomiasis control programmes. Available at <https://www.finddx.org/what-we-do/projects/rapid-test-for-precision-mapping-and-monitoring-and-evaluation-of-schistosomiasis-control-programmes/> (accessed 12 June 2024).
34. GHIT (2020) Schistosomiasis rapid diagnostic test to support control programmes in monitoring treatment impact and reassessment mapping. Available at <https://www.ghitfund.org/investment/portfoliodetail/detail/167/en> (accessed 12 June 2024).