



Universiteit
Leiden
The Netherlands

Under construction: improving arteriovenous fistula maturation

Laboyrie, S.L.

Citation

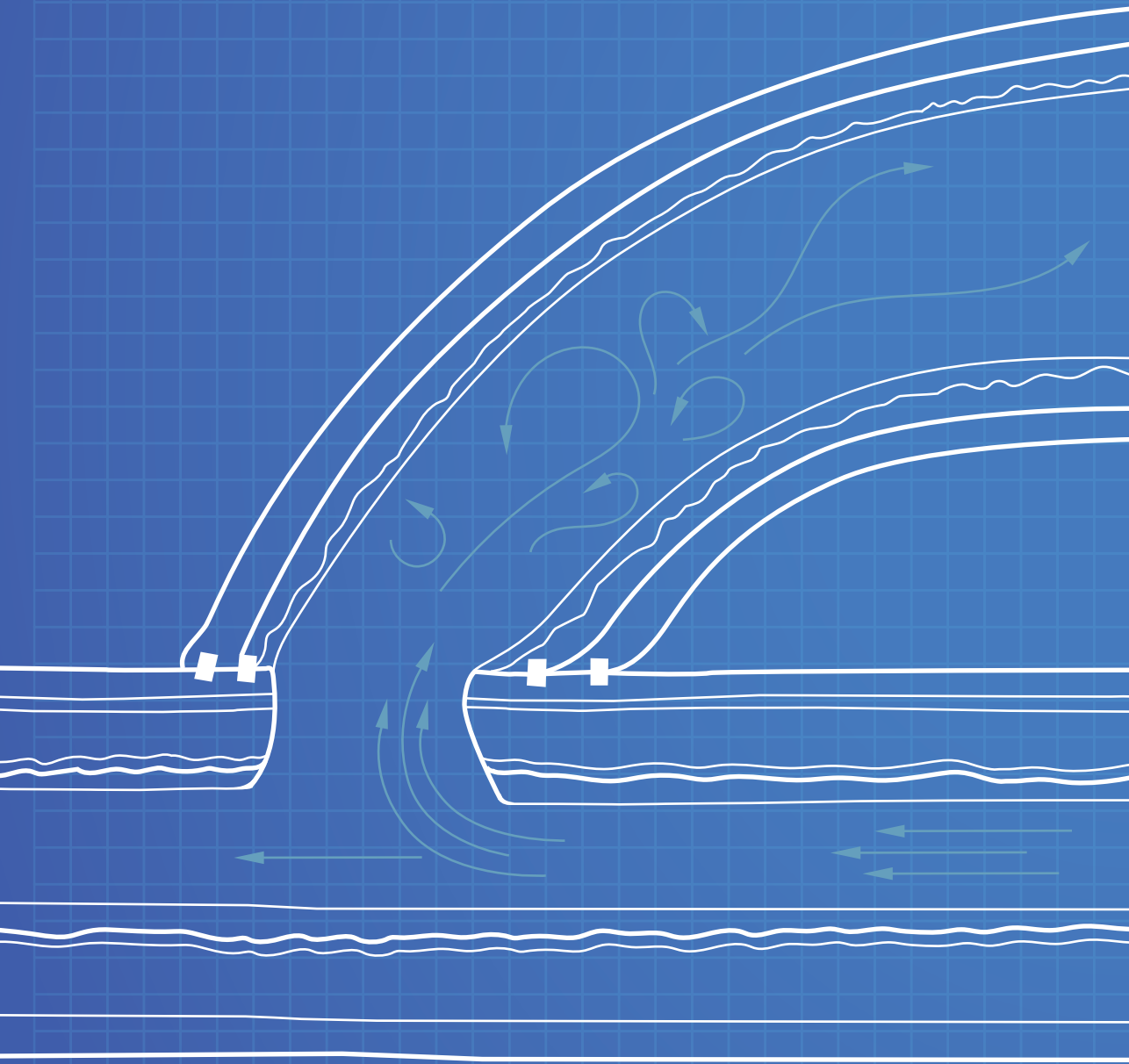
Laboyrie, S. L. (2025, October 17). *Under construction: improving arteriovenous fistula maturation*. Retrieved from <https://hdl.handle.net/1887/4270719>

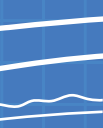
Version: Publisher's Version

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/4270719>

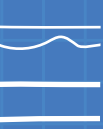
Note: To cite this publication please use the final published version (if applicable).





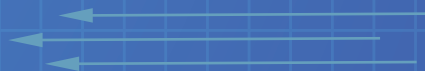
Chapter 3

Von Willebrand factor: a central regulator of arteriovenous fistula maturation through smooth muscle cell proliferation and outward remodelling



Suzanne L. Laboyrie, Margreet R. de Vries, Alwin de Jong, Hetty C. de Boer, Reshma A. Lalai, Laisel Martinez, Roberto I. Vazquez-Padron and Joris I. Rotmans

Journal of the American Heart Association. 2022;11:e024581



Abstract

Background

Arteriovenous fistula (AVF) maturation failure is a main limitation of vascular access. Maturation is determined by the intricate balance between outward remodeling and intimal hyperplasia, whereby endothelial cell dysfunction, platelet aggregation, and vascular smooth muscle cell (VSMC) proliferation play a crucial role. von Willebrand Factor (VWF) is an endothelial cell-derived protein involved in platelet aggregation and VSMC proliferation. We investigated AVF vascular remodeling in VWF-deficient mice and VWF expression in failed and matured human AVFs.

Methods and Results

Jugular-carotid AVFs were created in wild-type and VWF^{-/-} mice. AVF flow was determined longitudinally using ultrasonography, whereupon AVFs were harvested 14 days after surgery. VSMCs were isolated from vena cavae to study the effect of VWF on VSMC proliferation. Patient-matched samples of the basilic vein were obtained before brachio-basilic AVF construction and during superficialization or salvage procedure 6 weeks after AVF creation. VWF deficiency reduced VSMC proliferation and macrophage infiltration in the intimal hyperplasia. VWF^{-/-} mice showed reduced outward remodeling (1.5-fold, $P=0.002$) and intimal hyperplasia (10.2-fold, $P<0.0001$). AVF flow in wild-type mice was incremental over 2 weeks, whereas flow in VWF^{-/-} mice did not increase, resulting in a two-fold lower flow at 14 days compared with wild-type mice ($P=0.016$). Outward remodeling in matured patient AVFs coincided with increased local VWF expression in the media of the venous outflow tract. Absence of VWF in the intimal layer correlated with an increase in the intima-media ratio.

Conclusions

VWF enhances AVF maturation because its positive effect on outward remodeling outweighs its stimulating effect on intimal hyperplasia.

Introduction

The arteriovenous fistula (AVF) is considered the gold standard of vascular access (VA) for hemodialysis [1]. Compared to central venous catheters (CVC) and arteriovenous grafts (AVGs), AVFs have better longevity and less complications [2-4]. However, unassisted maturation rates range from 60 to 79% [5]. This results in frequent VA-related interventions and hospitalization in order to achieve, maintain or restore patency. Therefore, there is an urgent need to improve AVF maturation.

AVF maturation depends on the ability of the venous outflow tract to adapt to the rapid increase in pressure and flow upon AVF creation. Concordantly, the luminal diameter of the vein needs to expand after AVF placement in order to facilitate the increase in blood flow created at the arteriovenous anastomosis. This process is determined by the intricate balance between outward remodeling (OR) of the vessel and the degree of obstruction of the lumen by intimal hyperplasia (IH). One key player in both OR and IH is the vascular smooth muscle cell (VSMC) [6-9]. However, the interplay between favourable VSMC proliferation that enables OR, and detrimental VSMC proliferation that causes IH, is an unknown process.

Von Willebrand Factor (VWF) is a multimeric glycoprotein that is essential for blood clotting by binding platelets and as carrier protein of circulating Factor VIII [10]. VWF is synthesized by megakaryocytes located in the bone marrow and by endothelial cells (ECs) lining the intima, but upon vascular damage, VWF can localize subintimal in vicinity of VSMCs [11, 12]. VWF^{-/-} mice have reduced angiogenesis, arteriogenesis, wound healing and leukocyte infiltration [13, 14], and it has been established in several animal models that VWF is involved in intimal thickening [11, 15, 16]. Moreover, previous studies revealed that VWF is a potent inducer of VSMC proliferation [15, 17] and inhibits gene expression of mature non-mitotic VSMCs [18]. Hence, VWF might play an important role in the proliferation of VSMCs in AVFs and influence the degree of OR and IH. We hypothesized that VWF deficient mice will present with reduced OR and IH, affecting vascular remodeling and thereby AVF maturation. In the present study we investigated the effect of VWF deficiency on murine AVF vascular remodeling and VWF expression levels in the intima and medial layer of native veins and patient-matched AVFs obtained during two-stage brachiobasilic surgery. We hereby assessed whether VWF deficiency tips the scale in the intricate balance between OR and IH to achieve AVF maturation.

Methods

The data that support the findings of this study are available from the corresponding author upon reasonable request.

Animals and study design

This study was performed in compliance with Dutch government guidelines, the Directive 2010/63/EU of the European Parliament and approved by the Institutional Committee for Animal Welfare at the LUMC. Adult C57BL/6J and B6.129S2-VWF^{tm1Wgr} (VWF knockout) mice [19], aged 8-12 weeks and bred in our own facility, were used for the *in vivo* studies. For histological analysis, 9 VWF^{-/-} mice (4 females) and 9 WT mice (4 females) were included. Ultrasound analysis was performed on 9 VWF^{-/-} and six WT male mice, as female VWF^{-/-} mice had a higher exsanguination rate during surgery and were less likely to survive subsequent anesthesia during ultrasound analysis. Prior to the surgery, the mice were anesthetized via intraperitoneal injection of midazolam (5 mg/kg, Roche), medetomidine (0.5 mg/kg, Orion) and fentanyl (0.05 mg/kg, Janssen) and received unilateral AVFs, as previously described [20] between the dorsomedial branch of the external jugular vein and the common carotid artery (CCA). To prevent massive blood loss, human plasma derived VWF (Wilfactin 150 IU/kg, Sanquin) was administered intravenously to the AVF during surgery. After surgery, mice were antagonized with atipamezole (2.5 mg/kg, Orion) and flumazenil (0.5 mg/kg, Fresenius Kabi). Buprenorphine (0.1 mg/kg, MSD Animal Health) was given after surgery to relieve pain.

Ultrasound analysis

The Vevo 3100 LAZR-X Imaging System (FUJIFILM Visual Sonics) coupled to a 50-MHz linear-frequency transducer was used to longitudinally follow blood velocity and vessel diameter of the CCA of the AVF. The mice were anesthetized with 4% isoflurane and placed on the animal imaging platform where temperature, heart rate, and respiration rate were monitored in real time. During the experiments, anesthesia was maintained using a vaporized isoflurane gas system (1 l/min of oxygen 0.3 l/min air and 1.5 - 2.5% isoflurane). The concentration of isoflurane was adjusted accordingly to the pedal reflex and respiration rate to ensure adequate anaesthesia. Measurements were performed at baseline one day before surgery, at t=0 immediately following surgery, 7 and 14 days. The number of animals included varied per time point due to swelling and wound crust formation hindering ultrasound measurement (minimum n=3).

Data was obtained using B-mode to identify the anatomical region, EKV (ECG-gated Kilohertz Visualization) to visualize vessel wall characteristics such as diameter of the CCA, and PV color mode to quantify the pulse wave velocity. Quantification was performed with measurements that were obtained in between breathing cycles. The data was analysed using Vevo lab software. Flow rate was calculated from pulse-wave velocity measurements and vessel diameter of the CCA afferent to the anastomosis.

Murine tissue harvesting and processing

14 days after AVF surgery, the mice were anesthetized using an anaesthetic-mixture containing midazolam (5 mg/kg, Roche), medetomidine (0.5 mg/kg, Orion) and fentanyl (0.05 mg/kg, Janssen) through intraperitoneal injection. The thorax was opened and flushed with 4% formalin through intracardiac perfusion, whereafter the AVF was dissected and submerged in 4% formalin overnight, then embedded in paraffin. Tissue sections of 5 μ m perpendicular to the venous outflow tract of the AVF were collected with an interval of 150 μ m per slide as described before [20].

Plasma and the superior vena cava were collected from mice 6-8 weeks of age, who were anesthetized as described above and were sacrificed by cervical dislocation after tissue collection. Retro-orbital blood was collected in EDTA tubes for the Luminex assay and in eppendorfs supplemented with 9:1 sodium citrate for cell culture purposes. The tubes were tumbled to mix the blood and anti-coagulant and spun down for 20 minutes at 2000 x *g*, after which the plasma was collected and frozen until further use. The superior vena cava was flushed with PBS and connective tissue was removed *in vivo*, after which the vein was dissected and put in PBS on ice.

Histology murine tissue

As most AVF occlusions occur in the venous outflow tract of the AVF, the first three venous-cross sections downstream of the anastomosis with 150 μ m interval were used for morphological and immunohistochemical analysis. Morphometric analysis of the murine AVFs was performed using Weigert's elastin staining and α SMA (DAKO M0851) as a VSMC marker. Outward remodeling was studied by measuring the length of the internal elastic lamina (IEL) in Caseviewer (3DHISTECH) in Weigert's Elastin-stained samples. Tissue within the IEL was defined as intimal hyperplasia (IH) and determined using histoquant software (3DHISTECH). Luminal area was calculated by subtracting IH from the area within the IEL. α SMA positive tissue within the IH was determined using histoquant software. Thrombi were

defined as anuclear α SMA negative tissue within the IH and calculated by manual tracing using annotations in Caseviewer.

Nuclei of Mac3⁺ cells (BD Pharmingen, 550292) were counted manually in three random fields of view (80x magnification) from which the mean was calculated. Proliferative VSMCs were detected by counting Ki67 (BD Pharmingen, 550609) and α SMA positive cells within the IEL and normalized to the α SMA positive area, which was defined using histoquant.

Luminex assay

To verify the effect of VWF deficiency on Weibel-Palade body (WPB)-stored proteins Angiopoietin-2 (Ang-2), Interleukin-6 (IL-6), osteoprotegerin and P-selectin, we performed a magnetic luminex assay (BioTechne) on EDTA-plasma obtained from nine WT and five VWF^{-/-} mice according to manufacturer's protocol. Dilutions of 1:3, 1:6 and 1:9 were analysed in duplicate on a Bio-Rad Bio-Plex analyser using the Bioplex Manager 6.0 software. Analyte concentrations (pg/mL) were calculated using a 5-PL logistic regression model based on a 6-point standard calibration curve.

Murine VSMC cell culture and proliferation assay

Primary venous VSMCs were isolated from the superior vena cava of WT and VWF^{-/-} mice without an AVF. In a sterile flow cabinet, 12-well plates were coated with 1% gelatin/PBS. The vena cava was cut longitudinally and the endothelial monolayer scraped off using sterile surgical forceps. Subsequently, the tissue was dissected into explants of around 1-3 mm. By placing a sterile coverslip on top of the explants a vacuum was created, keeping the explants in close contact with the culture surface.

Cells were cultured in DMEM (Gibco) supplemented with 20% fetal calf serum (FCS), 2 mM L-glutamine, 100 U/mL penicillin and 100 mg/mL streptomycin (penicillin/streptomycin, Gibco) and new medium was added after 7 days. Around two weeks after explant isolation, the explants were removed and the cells were trypsinized, replated and cultured until 80-90% confluence, whereafter the cells were cultured overnight in DMEM supplemented with 1% FCS and used to measure proliferative capacity.

Murine VSMC proliferation assay

Venous VSMCs were trypsinized, counted and resuspended in DMEM. Ten-thousand cells were plated per well in a 96-well plate in a total volume of 100 μ L DMEM supplemented with 10% murine plasma of either WT or VWF^{-/-} mice. To verify the direct effect of VWF on VSMCs, 10% VWF^{-/-} plasma supplemented with 100 ng/mL Wifactin (Sanquin, The Netherlands) was added to VWF^{-/-} and WT VSMCs. After 40 hours incubation at 37°C with 5% CO₂, proliferation was quantified using the CyQuant Direct Cell Proliferation Assay according to the manufacturer's protocol (Thermo Fisher Scientific). The experiment was performed in biological n = 3-6, in technical duplicates.

Patient samples: pre-access native vein and AVF venous outflow tract and plasma

Five μ m paraffin-embedded cross-sections of matched pre-access veins and AVF tissue pairs of end-stage renal disease (ESRD) patients were obtained from the University of Miami (UM) Human Vascular Biobank. The study was performed according to the ethical principles of the Declaration of Helsinki and regulatory requirements at Jackson Memorial Hospital (JMH) and UM. The ethics committee and UM Institutional Review Board approved the study. Thirty-eight patients who underwent two-stage AVF creation at JMH or the UM Hospital (19 with successful maturation and 19 with anatomic maturation failure) were selected from a total of 273 patients with tissue pairs in the biorepository (189 matured, 84 failed) [21, 22]. Anatomic maturation failure was defined as an AVF with an internal cross-sectional luminal diameter $\leq$ 6 mm as determined intraoperatively using intravascular probes. Random selection was performed using propensity score matching while adjusting for age, sex, ethnicity, diabetes, and pre-dialysis status. Fifteen mature AVFs and 16 failed AVFs were available for matched-pair analysis. Patient characteristics can be found in the table below.

Table 1: Characteristics of ESRD patients who underwent two-stage brachiobasilic AVF surgery and were included for histological analysis. ASCVD = Atherosclerotic cardiovascular disease.

	Average age (years $\pm$ SD)	Sex (% female)	Ethnicity (% Black)	Diabetes (%)	ASCVD (%)	Pre-dialysis (%)	Previous AVF (%)	Smoking (%)
Failed (N=19)	57.45 $\pm$ 11.95	40	60	50	50	35	15	40
Mature (N=19)	58.80 $\pm$ 15.72	40	40	70	45	35	30	25

Human samples were stained with VWF (DAKO A0082), CD31 (DAKO M0823) and α SMA (DAKO M0851). The medial layer was defined as the area between the IEL and external elastic lamina (EEL) and determined using Caseviewer. Positively stained tissue was defined using Caseviewer's Histoquant. Matched pair analysis of native veins and AVFs (15 mature and 16 failed) was performed for IH area, OR (IEL perimeter), area of the medial layer and luminal area. Intima/media (I/M) area ratio was calculated by dividing the IH by the medial layer. The I/M ratio of 25 out of 67 samples was previously analysed [22]. VWF and CD31 co-staining was performed on five patient-matched native veins and AVFs of both outcome groups – 20 samples in total. Plasma samples from 8 patients with AVF failure and 10 with AVF maturation were collected during surgery at the time of AVF creation and at superficialization through transposition. VWF antigen levels in patient plasma were determined as previously described [23].

Statistical analysis

Graphpad Prism 8 was used to perform statistical analysis. Normally distributed data is presented as mean value $\pm$ standard deviation (SD). Non-parametric data is presented as the median + interquartile range (IQR). Unpaired t-test, one-way ANOVA, Wilcoxon matched-pairs signed rank test and Mann-Witney U-test (two-tailed) were used when applicable. $p < 0.05$ is considered significant (*), ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$.

Results

VWF enhances outward remodeling and intimal hyperplasia

To investigate how the von Willebrand factor (VWF) influences AVF maturation, we created an end-to-side carotid-jugular AVF in VWF deficient (VWF^{-/-}) and wild-type (WT) mice. Two weeks post-surgery, the AVFs were collected and morphometric analysis of the murine AVF venous outflow tract was performed using Weigert's elastin staining for vessel circumference (left figure 1B and 1C) and α SMA as a VSMC marker (right figure 1B and 1C). α SMA-negative and anuclear lesions were defined as thrombi. The IEL (internal elastic lamina) perimeter of unoperated external jugular veins was comparable between WT and VWF^{-/-} mice at $\pm$ 1200 μ m (figure 1A). At 14 days after AVF surgery, VWF^{-/-} mice had a 1.5-fold smaller IEL perimeter compared to WT mice (figure 1D, $p = 0.002$). Total IH surface area in VWF^{-/-} mice was reduced 10.2-fold compared to WT mice ($p < 0.0001$). Within the IEL, the α SMA positive area in VWF^{-/-} mice was 8.1-fold smaller than in WT mice ($p = 0.0002$). Almost no thrombi were present in the VWF^{-/-} AVF, with a mean area

of $4 \mu\text{m}^2$, whereas thrombi in WT mice occupied $2868 \mu\text{m}^2$ ($p < 0.0001$). There was a 1.4-fold reduction in luminal area of $\text{VWF}^{-/-}$ mice, although this finding did not reach statistical significance ($p=0.10$).

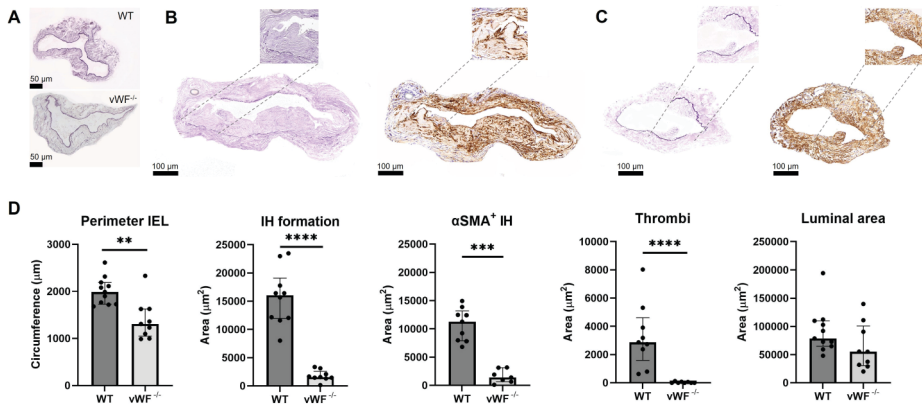


Figure 1: VWF deficiency leads to reduced outward remodeling and intimal hyperplasia following AVF surgery. Representative images of baseline vena jugularis, scale bar 50 μm (A) and AVFs stained using Weigert's Elastin (left) and αSMA staining (right) from a WT (B) and $\text{VWF}^{-/-}$ (C) mouse AVF. Inlays illustrate IH in closer detail, including a thrombus in figure 1B. The perimeter was measured by lining the internal elastic lamina (IEL), thrombi were defined as αSMA -negative anuclear tissue. Stainings were quantified and presented as median $\pm$ IQR (interquartile range) (D). OR was calculated using the IEL perimeter in Weigert's Elastin-stained coupes, IH was defined as the tissue within the IEL. αSMA -positive tissue within the IH was determined using histoquant software. Thrombi were manually traced. Luminal area was calculated by subtracting the IH from the area within the IEL. Scale bar is 100 μm . N=9 per group. Statistical significance between groups was determined using the Mann-Whitney U test ** $p=0.002$, *** $p<0.001$, **** $p<0.0001$.

VWF deficient mice do not show incremental AVF flow

Ultrasonography was performed on the AVFs at baseline, post-surgery at 0-, 7- and 14-days post AVF-creation to determine AVF functionality in terms of blood flow in the common carotid artery (CCA), proximal to the site of anastomosis (figure 2A). WT and $\text{VWF}^{-/-}$ mice had comparable flow at baseline. Flow rate in the WT mice increased over two weeks post-AVF creation, with a 1.3-fold increase at 7 days ($p=0.057$) and 2.55-fold at 14 days ($p=0.016$) compared to the AVF flow volume at baseline (figure 2B). AVF flow remained constant in $\text{VWF}^{-/-}$ mice over two weeks post-AVF creation, congruent with the low degree of OR. At t=14 days, flow in the AVFs of $\text{VWF}^{-/-}$ mice was 2.0-fold lower than in WT mice ($p=0.008$).

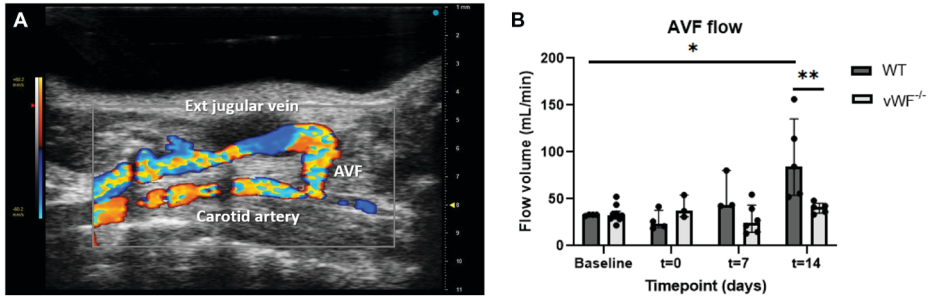


Figure 2: VWF deficiency leads to reduced AVF flow 14 days post-AVF creation. Effect of VWF deficiency on AVF blood flow was determined in the common carotid artery (CCA) afferent to the anastomosis (A). AVF blood flow in the CCA was calculated with the diameter of the CCA using EKV obtained data and blood velocity obtained in PV color mode (B). Ultrasound measurements were performed one day before surgery (baseline), immediately post-surgery (t=0) and at t=7 and t=14 days. Data is presented as median $\pm$ IQR, n=6 WT and 9 VWF^{-/-} mice. Statistical significance between groups was determined at timepoint t=14 days, and between baseline and t=14 days in WT mice using the Mann-Whitney U test * p=0.016, ** p=0.008.

VWF deficiency affects Weibel Palade Body proteins

Weibel-Palade bodies (WPBs) are the storage granules of ECs, of which VWF is a pivotal component. To verify whether deficiency of VWF affects the systemic release of WPB-stored proteins Angiopoietin-2 (Ang-2), Interleukin-6 (IL-6), Osteoprotegerin (OPG) and P-selectin, we performed a Luminex assay on plasma obtained from unoperated WT and VWF^{-/-} mice. IL-6 was undetectable in the plasma of both groups (data not shown) and no difference was observed in Ang-2 levels (figure 3A). Systemic levels of P-selectin (3.0-fold, p=0.007) and OPG (2.9-fold, p=0.001) (figures 3B and 3C) expression were significantly enhanced in VWF^{-/-} mouse plasma.

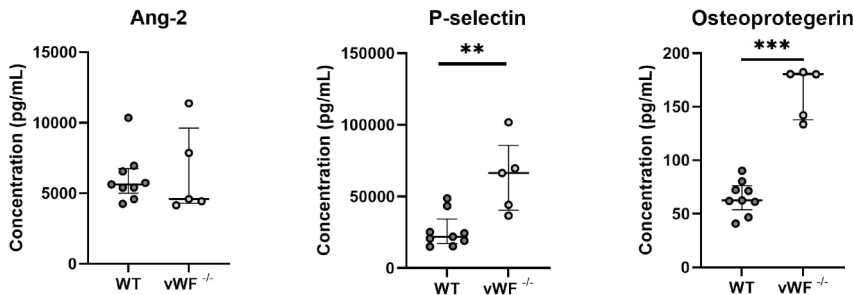


Figure 3: Luminex analysis of systemic WPB protein expression. The effect of VWF deficiency on Weibel-Palade body (WPB)-proteins Angiopoietin-2 (A), P-selectin (B) and Osteoprotegerin (C) was determined in 1:3 diluted EDTA-plasma from n=9 WT and 5 VWF^{-/-} mice using a 5-PL logistic regression model. Data is presented as median $\pm$ IQR. Statistical significance between groups was determined using the Mann-Whitney U test. ** p=0.007, *** p=0.001.

VWF^{-/-} mice exhibit less Mac3⁺ cells in the intimal hyperplasia

Similar to VWF, transmembrane protein P-selectin is involved in leukocyte recruitment and adhesion. To investigate the effect of VWF deficiency combined with increased systemic levels of P-selectin on local recruitment of inflammatory cells and IH formation, we performed a double-staining for macrophage marker Mac3 and α SMA. Compared to WT mice (figure 4A), AVFs of VWF^{-/-} mice (figure 4B) have a 37% reduction in expression of Mac3⁺ cells in the IH (figure 4C), $p=0.006$.

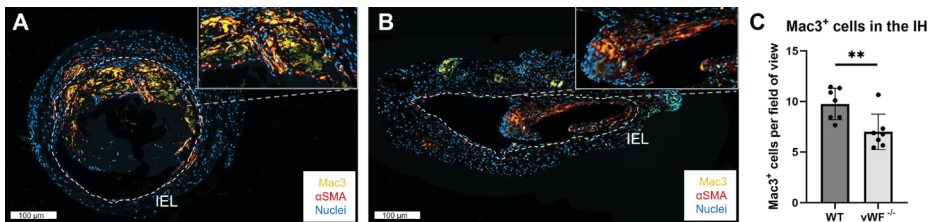


Figure 4: Intimal hyperplasia in the AVF of VWF^{-/-} mice displays fewer Mac3⁺ cells. Immunofluorescent staining for nuclei (blue), α SMA (red) and Mac3 (yellow) of an AVF of a WT (A) and VWF^{-/-} (B) mouse, with a higher magnification inlay of part of the IH. The IEL is traced in white, scale bar represents 100 μ m. Nuclei of Mac3⁺ cells were counted manually per field of view and quantified for $n=7$ per group (C). Data is presented as mean $\pm$ SD, ** $p = 0.006$, statistical significance between groups was determined using unpaired t-test.

VWF enhances VSMC proliferation

To assess the effect of VWF expression on VSMC proliferation, sections from the venous outflow tract of AVFs from WT (figure 5A) and VWF^{-/-} mice (figure 5B) were stained for Ki67 and α SMA. Double-positive cells within the IH were counted and normalized to the α SMA-positive area. Compared to WT mice, VWF^{-/-} mice showed an 82% decrease in proliferating Ki67⁺ VSMCs in the IH (figure 5C, $p=0.0002$).

To verify the direct effect of VWF on VSMC proliferation, VSMCs were grown from explants of the superior vena cava and stimulated with murine plasma to simulate *in vivo* conditions. Compared to WT VSMCs exposed to WT plasma, VWF deficient plasma induced less proliferation in both WT and VWF^{-/-} VSMCs (0.6-fold and 0.5-fold, figure 5D), which was rescued by addition of human plasma derived VWF (Wilfactin). A non-significant 1.5-fold upregulation was observed when VWF^{-/-} VSMCs were stimulated with Wilfactin in addition to VWF deficient plasma alone ($p=0.09$), indicating a trend of VSMC proliferation due to rescue of VWF deficiency

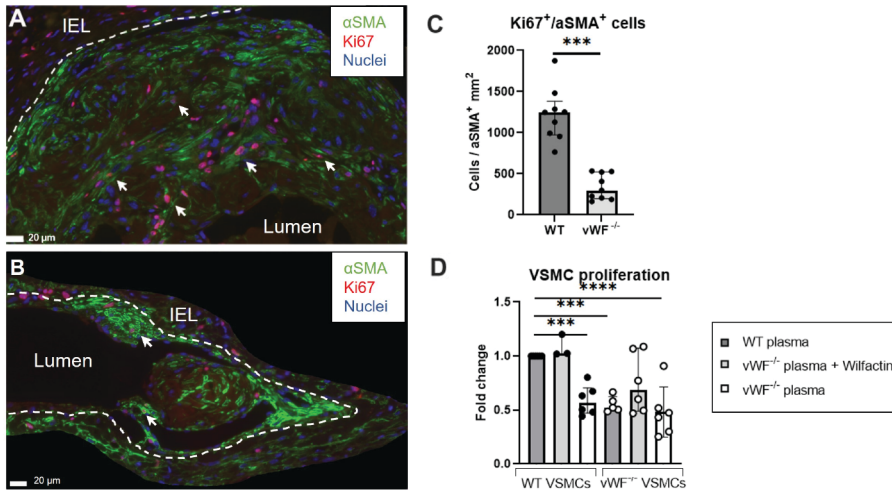


Figure 5: Characterization of proliferating VSMCs in the IH at two weeks post AVF creation and in vitro. Immunofluorescent staining for nuclei (blue), Ki67 (red) and αSMA (green) in a WT (A) and VWF^{-/-} mouse (B). The IEL is traced in white. Ki67⁺/αSMA⁺ cells, examples indicated with an arrow, within the IH were counted manually and normalized to the αSMA-positive area within the IH. Scale bar represents 20 μm. Data is analysed by the Mann-Whitney U test and presented as median + IQR, n=9 per group (C). VSMCs from WT (left three bars) and VWF^{-/-} mice (right three bars) were stimulated with 10% murine plasma of WT or VWF^{-/-} mice ± 100 ng/mL Wilfactin (see legend) for 40 hours, after which proliferation was measured. Data is presented as mean ± SD, n = 3-6. Statistical significance between WT VSMCs stimulated with WT plasma (control) and other experimental conditions was determined using one-way ANOVA and Dunnett's multiple comparisons test *** p<0.001, **** p=0.0001 (D).

VWF resides in the IH and vasa vasorum of ESRD patients

Human pre-access native veins and pair-matched venous AVF samples were obtained from ESRD patients undergoing two-stage brachio-basilic AVF surgery. We observed VWF expression in the IH of both murine (figure 6A) and human AVFs (figure 6B). Human AVFs show VWF and CD31 co-expression in the vasa vasorum in the tunica media (medial layer) and tunica adventitia (asterisk figure 6B). Expression of VWF not in the vicinity of CD31-positive ECs was also observed in the tunica media of a mature AVF (arrow in figure 6B). No difference in systemic VWF antigen levels were observed in samples obtained during the first procedure - AVF surgery - (1.31±0.69 IU/mL in patients with an AVF that would mature; 1.58±0.69 IU/mL in patients with an AVF that would fail) and during the second procedure - superficialization of the basilic vein - (mature AVF: 1.36±0.55 IU/mL; failed AVF: 1.35±0.55 IU/mL).

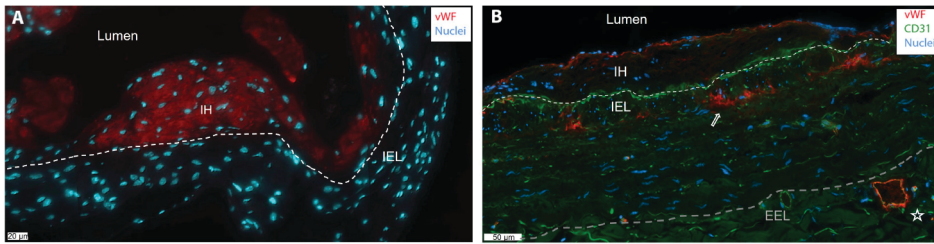


Figure 6: Von Willebrand Factor resides in the intima and medial layer. Representative immunofluorescent staining for VWF (red) in a wild-type murine AVF (A) and in a matured AVF of an end-stage renal disease (ESRD) patient at superficialization (B). Endothelial cell marker CD31 is stained in the humane AVF in green, nuclei in blue. The white dashed line signifies the IEL, the grey line indicates the EEL (external elastic lamina). Scale bar indicates 20 μm in A and 50 μm in B.

Matured AVFs show increased expression of VWF in the venous tunica media

Matured AVFs (figure 7A) have a thicker tunica media compared to failed AVFs (figure 7B). VWF colocalized with CD31^+ cells in the vasa vasorum of both failed and mature AVFs (figure 6A, indicated with an asterisk), or with αSMA , indicated by arrows in a mature AVF sample (figure 7A). When studying maturation outcome parameters, the median increase in intimal area was not significantly different in the pair-matched samples of patients with matured versus failed AVFs (figure 7C; IH 2.1 mm^2 versus 1.7 mm^2 respectively, $p = 0.67$). The increase in IEL perimeter, medial area as well as luminal area in patients with matured AVFs was significantly larger when compared to patients with failed AVFs (median perimeter 5.4 mm versus 2.7 mm , $p=0.022$; medial area: 2.7 mm^2 versus 0.6 mm^2 , $p = 0.009$; luminal area: 1.8 mm^2 versus 0.7 mm^2 , $p = 0.016$). In addition, the increase in intima/media (I/M) ratio was smaller in patients with matured AVFs (0.2) when compared to patients with failed AVFs (0.7, $p= 0.01$), due to reduced thickening of the medial area in failed AVFs, indicating the importance of OR in human AVF maturation. The wall thickening and OR in matured AVFs coincided with 167% ($p=0.03$) increase in VWF expression in the medial layer of pre-access veins to mature venous AVF samples, but not in patient samples of failed AVFs (figure 7D).

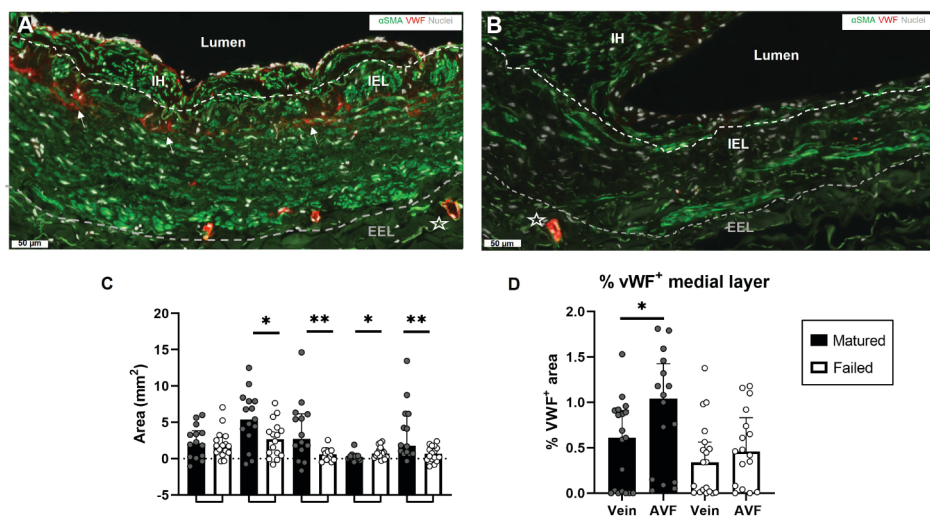


Figure 7: Morphometric and VWF expression analysis of ESRD patient samples. Analysis of ESRD pair-matched samples of pre-access native veins and venous AVF outflow tracts obtained during two-stage brachio-basilic AVF surgery. AVF failure was defined as having an internal cross-sectional luminal diameter ≤ 6 mm. Representative sample of a mature (A) and failed AVF (B). Scale bar indicates 50 μ m, asterisk indicates a vessel in the vasa vasorum, α SMA $^+$ /VWF $^+$ staining is indicated with an arrow. The inner white line signifies the IEL, the outer grey line signifies the external elastic lamina (EEL). Pair-matched analysis of increase in IH area, delta (Δ) OR (perimeter in mm), area of the medial layer, the intima/media (I/M) ratio and luminal size was calculated by subtracting the pre-access venous parameters from the patient-matched AVF. N=15 pair-matched samples from matured AVFs and 16 pair-matched failed AVFs. Statistical significance between groups was determined using the Mann-Whitney U test (C). VWF $^+$ tissue in the medial layer was quantified in immunofluorescently stained samples and normalized to the area of the medial layer. Statistical significance was determined using Wilcoxon matched pairs signed rank test (D). Data is presented as median $\pm$ IQR. * $p < 0.05$, ** $p < 0.01$.

Failed AVFs express less VWF in the intimal layer

In both samples from mature and failed AVFs, absence of VWF lining the IH was observed. This was often at sites with increased IH (figure 8A), whereas little IH was observed at sites with VWF lining the intima (figure 8B). We observed a reduction of VWF $^+$ intima from the native vein to the venous AVF outflow tract in both patient groups (figure 8D), with a significant 3.6-fold reduction in failed AVF samples compared to mature AVFs ($p = 0.04$). The intimal hyperplasia to medial layer (I/M) ratio increased from the native vein to the venous AVF sample (figure 8C) with 2.6-fold in mature AVFs ($p = 0.02$) and 4.0-fold in failed AVFs ($p = 0.0001$). With no significant difference in IH between the two groups, and significant wall thickening in matured AVFs (figure 7C), the high fold increase in I/M ratio in native veins to failed AVFs is caused by inefficient wall thickening. We hypothesized that VWF lining the intima might correlate with decreased I/M ratio due to VWF

production to induce thickening of the tunica media. The I/M and VWF⁺ intima of AVFs were negatively correlated using linear regression (figure 8E) and showed that when the VWF⁺ intimal layer is disrupted, the I/M ratio increases ($p=0.0017$ for matured AVFs and $p=0.0264$ for failed AVFs).

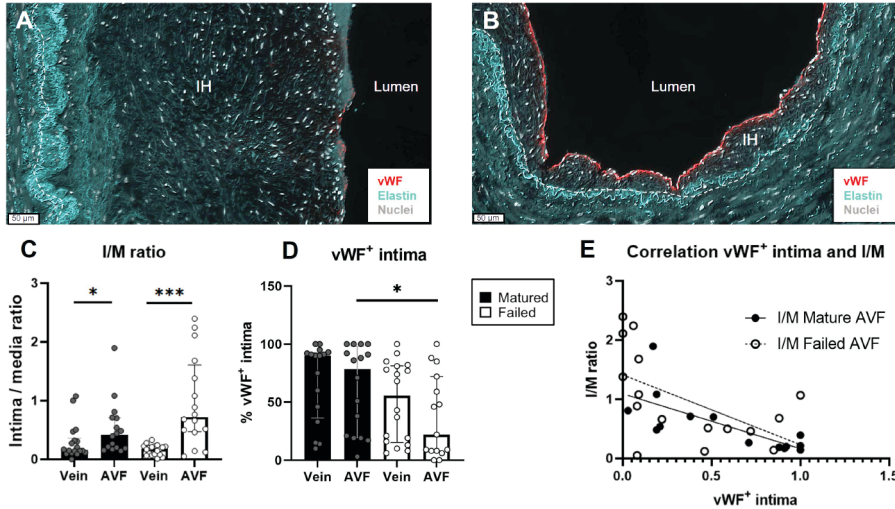


Figure 8: Relationship between disruption of VWF lining the intima and an enhanced I/M ratio. Analysis of ESRD samples of pre-access native veins and venous AVF outflow tracts obtained during two-stage brachio-basilic AVF surgery. Absence of VWF lining the intimal layer was observed at sites with increased IH (A) while an intact VWF⁺ intimal layer was observed at sites with little IH (B). The dashed line signifies the IEL, VWF is stained in red, elastin in turquoise, nuclei in white. Scale bar indicates 50 μ m. The ratio between intimal area and the medial layer (I/M) (C) and the percentage of VWF⁺ intimal layer in native veins and AVFs was quantified (D) and correlated using linear regression (E). N=15 pair-matched samples from matured AVFs and 16 pair-matched failed AVFs, data is represented as median $\pm$ IQR. Statistical significance between groups was determined using the Mann–Whitney U test * $p<0.05$, *** $p=0.0001$

Discussion

In this study we investigated the role of VWF in the process of AVF maturation and observed that VWF deficiency in mice impaired VSMC proliferation, macrophage infiltration and both OR as well as IH, which resulted in the inability to increase AVF flow. Human AVF samples showed no difference in systemic VWF plasma levels, but we did observe a local increase in VWF⁺ tissue in the medial layer of matured AVFs, which did not occur in failed AVFs. Secondly, we observed a correlation between disruption of a VWF expression in the intimal layer and increase in the I/M ratio, signifying the importance of VWF production to promote OR. This suggests a potential role for VWF in AVF maturation in ESRD patients as well.

Our study is the first to directly investigate the effect of VWF depletion on murine AVF maturation. VWF is well-known for its function in platelet aggregation and venous thrombosis [24-26]. Indeed, previous studies illustrated that platelet inhibition causes reduction of IH in mice [27]. We observed that in our VWF deficient mouse model, systemic plasma levels of WPB-proteins P-selectin and OPG were significantly higher than in WT mice. OPG is known to be atheroprotective and anticalcific by inhibiting calcium deposition in VSMCs [28], possibly positively affecting AVF function in VWF^{-/-} mice by reducing AVF stiffness. VWF deficient mice are known to exhibit altered local regulation of P-selectin through a reduction in both intracellular storage and expression at the endothelial cell surface, along with impaired leukocyte recruitment [14, 29]. This reduction in inflammatory cells might also be due to the reduction in vascular permeability and leukocyte docking after loss of VWF and thereby VWF-platelet interaction [30, 31]. We indeed observed a reduction of thrombi and Mac3⁺ cells in the IH of VWF^{-/-} mice, possibly affecting production of MMPs [32] and AVF wall thickening [33].

Besides inflammation and thrombus formation, VWF is also involved in the development of α SMA⁺ IH and OR of the venous outflow tract of murine AVFs, with an increase in expression. To verify that VWF promotes VSMC proliferation in our murine model, we performed an α SMA/Ki67 co-staining, which showed VWF^{-/-} mice to have a significant 82% decrease in proliferating α SMA⁺ cells in the IH. This is reinforced by recent work showing that VWF induces proliferation and migration of human arterial VSMCs through binding via its A2 domain to the LRP4-receptor, causing α V β 3 integrin signaling [17]. α V β 3 integrin expression is upregulated in the rabbit AVF, compared to control vessels [34], and inhibition of α V β 3 reduced SMC invasion of a collagen type I lattice [35]; similar to migration through vessel layers. These findings suggest that α V β 3 inhibition results in a reduction of IH and possibly thickening of the medial layer. Furthermore, as VWF can bind to several growth factors through its A1 heparin binding domain, VWF^{-/-} mice present with decreased proliferation of α SMA⁺/CD45⁻ cells [13]. These growth factors include VEGF-A, which promotes VSMC proliferation [36, 37], PDGF-BB which induces VSMC phenotypic switching, proliferation, migration and IH [38, 39], and CXCL-12 which helps recruit VSMCs [40]. Furthermore, arteries from patients with CADASIL (Cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy) show VWF expression extending into the arterial wall, where VWF inhibits expression of mature contractile smooth muscle cell markers such as SM22, SM-actin, Calponin and SM-MHC, indicating that VWF causes phenotypic VSMC switching towards a proliferative profile [18, 41]. This

provides support for the role of VWF in inducing VSMC proliferation in the tunica intima and tunica media of the AVF.

Besides apical secretion into the circulation, endothelial cells can also secrete VWF basally into the subendothelial matrix [42]. We observed VWF expression in α SMA⁺ tissue in the tunica media, indicating that VWF might directly influence VSMCs and switch them to a proliferative state to enhance OR and wall thickening. This illustrates the importance of the interplay between VWF produced by the endothelial layer and VSMCs in AVF maturation. Indeed, impaired endothelial functioning pre-AVF creation is associated with decreased AVF remodeling and function in ESRD patients [43, 44]. This corresponds to our observed correlation between loss of VWF expression in the AVF endothelial intima and an increase in the I/M ratio in ESRD patient samples.

A previous study in mice [6] observed high numbers of dedifferentiated Ki67⁺ VSMCs in the IH four weeks after AVF creation. Furthermore, matured MYH11⁺/Ki67⁻ VSMCs were mainly detected in the vascular wall, inciting the statement that VSMCs have a dual function in AVF remodeling: proliferative VSMCs aggravate IH, while mature VSMCs facilitate wall thickening. We postulate however that vsmc proliferation is also required in the medial layer to facilitate OR but occurs primarily in the first two weeks after murine AVF creation, whereafter proliferation of VSMCs in the intima is still ongoing [45]. With both IH and OR reduction in our VWF^{-/-} mice resulting in a worse outcome in AVF functionality, this reinforces the current hypothesis that the degree of OR is of greater importance than minimizing the degree of IH. Indeed, patient matched data did not indicate differential increase in IH between matured and failed AVFs, while matured AVFs showed more OR and wall thickening than failed AVFs. This was accompanied by a significant local upregulation of VWF expression in the medial layer of mature AVFs, signifying the clinical translation of our murine findings that VWF might enhance OR in ESRD patients.

Some aspects of our study however require further discussion. Unlike the murine AVFs, the patient samples have not been perfused with formalin as the murine AVFs, possibly affecting the analysis of OR. Secondly, human plasma derived VWF was administered during AVF surgery to rescue VWF deficiency and prevent excessive blood loss. Human plasma-derived VWF has a mean residence time of about 2.2 hours in mice, thus affecting our model temporarily [46]. Moreover, we used a constitutive VWF knockout model, which does not reflect biological variations in humans, affects storage of WPB-proteins and could have a broader

impact on endothelial cell function. Lastly, since the surgical procedure and ultrasound analysis combined with VWF deficiency puts a strain on the mice, our experiments have been performed in a model without renal failure. Previous studies revealed that chronic kidney disease significantly accelerates wall thickening and IH in mice [47, 48], but the composition of the stenotic lesions in these mice seems comparable with mice with normal kidney function [47]. However, it is unknown to which degree the murine IH process exactly resembles human IH formation in the early phase after AVF surgery.

In conclusion, we have shown the importance of VWF in VSMC proliferation and the AVF maturation process in mice, with human data enforcing the importance of an intact VWF⁺ intima and medial VWF expression to facilitate OR and wall thickening.

To apply this knowledge clinically, further research should be conducted on the origin of VSMCs that cause OR, IH and wall thickening, as currently there is no consensus where the VSMCs populating the IH originate from [7, 49-51]. Secondly, future studies with inducible VWF expression in a VWF-knockout model would enhance the understanding of the direct effect of VWF on vascular remodeling in AVF, besides its effect on endothelial cell function. Moreover, as systemic VWF will greatly enhance the risk of thrombosis, local delivery such as a slow releasing gel or targeted nanoparticles should be further investigated. Most importantly, the time frame of VSMC proliferation that is involved in OR and wall thickening in ESRD mice and patients should be studied to determine the optimal treatment delivery. Knowledge of timing and delivery of VWF will provide a solid background to develop a promising VWF therapy to enhance AVF maturation, thereby reducing the complication- and intervention burden on the already fragile ESRD patient group.

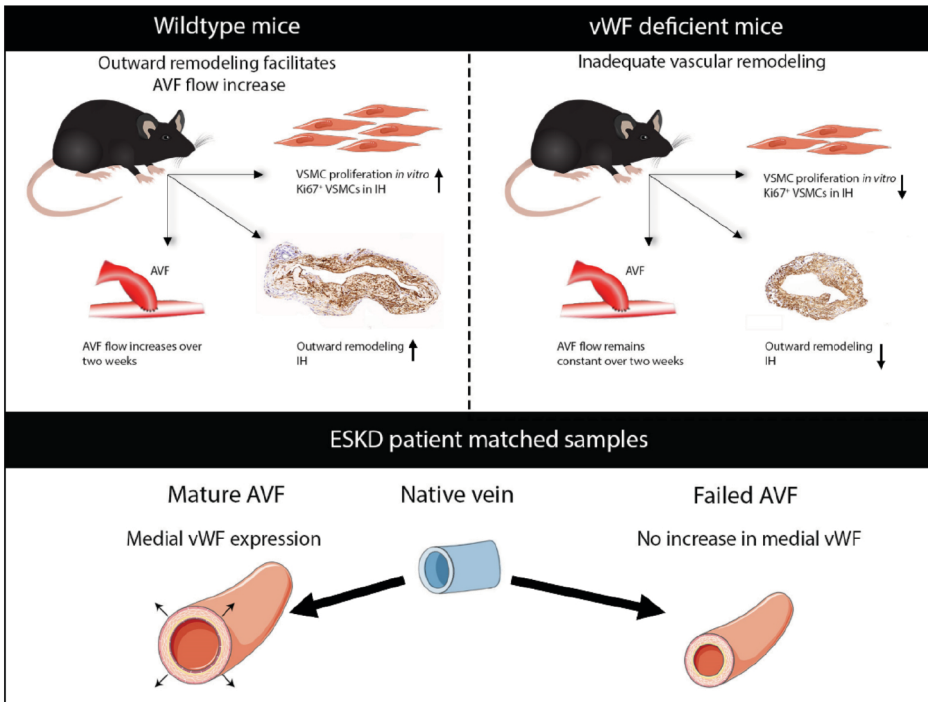


Figure 9. Overview of the findings.

AVF indicates arteriovenous fistula; ESKD, end-stage kidney disease; IH, intimal hyperplasia; VSMC, vascular smooth muscle cell; and VWF, von Willebrand Factor.

References

- Schmidli J, Widmer MK, Basile C, de Donato G, Gallieni M, Gibbons CP, Haage P, Hamilton G, Hedin U, Kamper L, et al. Vascular access: 2018 clinical practice guidelines of the european society for vascular surgery (esvs). *European Journal of Vascular and Endovascular Surgery*. 2018. **55**:757-818
- Astor BC, Eustace JA, Powe NR, Klag MJ, Fink NE, Coresh J, Study ftC. Type of vascular access and survival among incident hemodialysis patients: The choices for healthy outcomes in caring for esrd (choice) study. 2005. **16**:1449-1455
- Banerjee T, Kim SJ, Astor B, Shafi T, Coresh J, Powe NR. Vascular access type, inflammatory markers, and mortality in incident hemodialysis patients: The choices for healthy outcomes in caring for end-stage renal disease (choice) study. *American Journal of Kidney Diseases*. 2014. **64**:954-961
- Dhingra RK, Young EW, Hulbert-Shearon TE, Leavey SF, Port FK. Type of vascular access and mortality in u.S. Hemodialysis patients. *Kidney International*. 2001. **60**:1443-1451
- Voorzaat BM, Janmaat CJ, van der Bogt KEA, Dekker FW, Rotmans JI. Patency outcomes of arteriovenous fistulas and grafts for hemodialysis access: A trade-off between nonmaturation and long-term complications. 2020. **1**:916-924
- Zhao J, Jourdeuil FL, Xue M, Conti D, Lopez-Soler RI, Ginnan R, Asif A, Singer HA, Jourdeuil D, Long X. Dual function for mature vascular smooth muscle cells during arteriovenous fistula remodeling. *J Am Heart Assoc*. 2017. **6**
- Liang M, Wang Y, Liang A, Mitch WE, Roy-Chaudhury P, Han G, Cheng J. Migration of smooth muscle cells from the arterial anastomosis of arteriovenous fistulas requires notch activation to form neointima. *Kidney International*. 2015. **88**:490-502
- Roy-Chaudhury P, Wang Y, Krishnamoorthy M, Zhang J, Banerjee R, Munda R, Heffelfinger S, Arend L. Cellular phenotypes in human stenotic lesions from haemodialysis vascular access. *Nephrology Dialysis Transplantation*. 2009. **24**:2786-2791
- De Mey JG, Schiffers PM, Hilgers RH, Sanders MM. Toward functional genomics of flow-induced outward remodeling of resistance arteries. *American journal of physiology. Heart and circulatory physiology*. 2005. **288**:H1022-1027
- Ruggeri ZM, Ware J. Von willebrand factor. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology*. 1993. **7**:308-316
- Giddings JC, Banning AP, Ralis H, Lewis MJ. Redistribution of von willebrand factor in porcine carotid arteries after balloon angioplasty. *Arteriosclerosis, thrombosis, and vascular biology*. 1997. **17**:1872-1878
- Bosmans JM, Kockx MM, Vrints CJ, Bult H, De Meyer GR, Herman AG. Fibrin(ogen) and von willebrand factor deposition are associated with intimal thickening after balloon angioplasty of the rabbit carotid artery. *Arteriosclerosis, thrombosis, and vascular biology*. 1997. **17**:634-645
- Ishihara J, Ishihara A, Starke RD, Peghaire CR, Smith KE, McKinnon TAJ, Tabata Y, Sasaki K, White MJV, Fukunaga K, et al. The heparin binding domain of von willebrand factor binds to growth factors and promotes angiogenesis in wound healing. *Blood*. 2019. **133**:2559-2569
- de Vries MR, Peters EAB, Quax PHA, Nossent AY. Von willebrand factor deficiency leads to impaired blood flow recovery after ischaemia in mice. *Thromb Haemost*. 2017. **117**:1412-1419
- Qin F, Impeduglia T, Schaffer P, Dardik H. Overexpression of von willebrand factor is an independent risk factor for pathogenesis of intimal hyperplasia: Preliminary studies. *Journal of Vascular Surgery*. 2003. **37**:433-439
- De Meyer GR, Hoylaerts MF, Kockx MM, Yamamoto H, Herman AG, Bult H. Intimal deposition of functional von willebrand factor in atherogenesis. *Arteriosclerosis, thrombosis, and vascular biology*. 1999. **19**:2524-2534
- Lagrange J, Worou ME, Michel JB, Raoul A, Didelot M, Muczynski V, Legendre P, Plénat F, Gauchotte G, Lourenco-Rodrigues MD, et al. The vwf/lrp4/avβ3-axis represents a novel pathway regulating proliferation of human vascular smooth muscle cells. *Cardiovascular research*. 2021

18. Meng H, Zhang X, Lee SJ, Wang MM. Von willebrand factor inhibits mature smooth muscle gene expression through impairment of notch signaling. *PLOS ONE*. 2013. **8**:e75808
19. Denis C, Methia N, Frenette PS, Rayburn H, Ullman-Cullere M, Hynes RO, Wagner DD. A mouse model of severe von willebrand disease: Defects in hemostasis and thrombosis. *Proceedings of the National Academy of Sciences of the United States of America*. 1998. **95**:9524-9529
20. Wong CY, de Vries MR, Wang Y, van der Vorst JR, Vahrmeijer AL, van Zonneveld A-J, Hamming JF, Roy-Chaudhury P, Rabelink TJ, Quax PHA, et al. A novel murine model of arteriovenous fistula failure: The surgical procedure in detail. *J Vis Exp*. 2016. e53294-e53294
21. Tabbara M, Duque JC, Martinez L, Escobar LA, Wu W, Pan Y, Fernandez N, Velazquez OC, Jaimes EA, Salman LH, et al. Pre-existing and postoperative intimal hyperplasia and arteriovenous fistula outcomes. *American Journal of Kidney Diseases*. 2016. **68**:455-464
22. Martinez L, Duque JC, Tabbara M, Paez A, Selman G, Hernandez DR, Sundberg CA, Tey JCS, Shiu YT, Cheung AK, et al. Fibrotic venous remodeling and nonmaturation of arteriovenous fistulas. *J Am Soc Nephrol*. 2018. **29**:1030-1040
23. de Jong A, Dirven RJ, Oud JA, Tio D, van Vlijmen BJM, Eikenboom J. Correction of a dominant-negative von willebrand factor multimerization defect by small interfering rna-mediated allele-specific inhibition of mutant von willebrand factor. 2018. **16**:1357-1368
24. Brill A, Fuchs TA, Chauhan AK, Yang JJ, De Meyer SF, Kollnberger M, Wakefield TW, Lammle B, Massberg S, Wagner DD. Von willebrand factor-mediated platelet adhesion is critical for deep vein thrombosis in mouse models. *Blood*. 2011. **117**:1400-1407
25. Chauhan AK, Kisucka J, Lamb CB, Bergmeier W, Wagner DD. Von willebrand factor and factor viii are independently required to form stable occlusive thrombi in injured veins. *Blood*. 2007. **109**:2424-2429
26. Receveur N, Nechipurenko D, Knapp Y, Yakusheva A, Maurer E, Denis CV, Lanza F, Panteleev M, Gachet C, Mangin PH. Shear rate gradients promote a bi-phasic thrombus formation on weak adhesive proteins, such as fibrinogen in a vwf-dependent manner. *Haematologica*. 2019. 235754
27. Drosopoulos JHF, Kraemer R, Shen H, Upmacis RK, Marcus AJ, Musi E. Human solcd39 inhibits injury-induced development of neointimal hyperplasia. *Thromb Haemost*. 2010. **103**:426-434
28. Callegari A, Coons ML, Ricks JL, Rosenfeld ME, Scatena M. Increased calcification in osteoprotegerin-deficient smooth muscle cells: Dependence on receptor activator of nf-kb ligand and interleukin 6. *Journal of Vascular Research*. 2014. **51**:118-131
29. Denis CV, André P, Saffaripour S, Wagner DD. Defect in regulated secretion of p-selectin affects leukocyte recruitment in von willebrand factor-deficient mice. *Proceedings of the National Academy of Sciences*. 2001. **98**:4072-4077
30. Petri B, Broermann A, Li H, Khandoga AG, Zarbock A, Krombach F, Goerge T, Schneider SW, Jones C, Nieswandt B, et al. Von willebrand factor promotes leukocyte extravasation. *Blood*. 2010. **116**:4712-4719
31. Gagnano F, Sperlongano S, Golia E, Natale F, Bianchi R, Crisci M, Fimiani F, Pariggiano I, Diana V, Carbone A, et al. The role of von willebrand factor in vascular inflammation: From pathogenesis to targeted therapy. *Mediators of Inflammation*. 2017. 2017:5620314
32. Hanania R, Song Sun H, Xu K, Pustynnik S, Jeganathan S, Harrison RE. Classically activated macrophages use stable microtubules for matrix metalloproteinase-9 (mmp-9) secretion *Journal of Biological Chemistry*. 2012. **287**:8468-8483
33. Guo X, Fereydooni A, Isaji T, Gorecka J, Liu S, Hu H, Ono S, Alozie M, Lee SR, Taniguchi R, et al. Inhibition of the akt1-mtorc1 axis alters venous remodeling to improve arteriovenous fistula patency. *Scientific Reports*. 2019. **9**:11046

34. Cai W-J, Li MB, Wu X, Wu S, Zhu W, Chen D, Luo M, Eitenmüller I, Kampmann A, Schaper J, et al. Activation of the integrins $\alpha 5\beta 1$ and $\alpha v\beta 3$ and focal adhesion kinase (fak) during arteriogenesis. *Molecular and Cellular Biochemistry*. 2009. **322**:161-169
35. Kanda S, Kuzuya M, Ramos MA, Koike T, Yoshino K, Ikeda S, Iguchi A. Matrix metalloproteinase and $\alpha v\beta 3$ integrin-dependent vascular smooth muscle cell invasion through a type I collagen lattice. *Arteriosclerosis, thrombosis, and vascular biology*. 2000. **20**:998-1005
36. Liao XH, Xiang Y, Li H, Zheng DL, Xu Y, Xi Yu C, Li JP, Zhang XY, Xing WB, Cao DS, et al. Vegf-a stimulates stat3 activity via nitrosylation of myocardin to regulate the expression of vascular smooth muscle cell differentiation markers. *Scientific Reports*. 2017. **7**:2660
37. Parenti A, Bellik L, Brogelli L, Filippi S, Ledda F. Endogenous vegf-a is responsible for mitogenic effects of mcp-1 on vascular smooth muscle cells. 2004. **286**:H1978-H1984
38. Lu QB, Wan MY, Wang PY, Zhang CX, Xu DY, Liao X, Sun HJ. Chicoric acid prevents pdgf-bb-induced vsmc dedifferentiation, proliferation and migration by suppressing ros/nfkb/mtor/p70s6k signaling cascade. *Redox biology*. 2018. **14**:656-668
39. Kingsley K, Huff JL, Rust WL, Carroll K, Martinez AM, Fitchmun M, Plopper GE. Erk1/2 mediates pdgf-bb stimulated vascular smooth muscle cell proliferation and migration on laminin-5. *Biochemical and biophysical research communications*. 2002. **293**:1000-1006
40. Stratman AN, Burns MC, Farrelly OM, Davis AE, Li W, Pham VN, Castranova D, Yano JJ, Goddard LM, Nguyen O, et al. Chemokine mediated signalling within arteries promotes vascular smooth muscle cell recruitment. *Communications Biology*. 2020. **3**:734
41. Zhang X, Meng H, Blaivas M, Rushing EJ, Moore BE, Schwartz J, Lopes MBS, Worrall BB, Wang MMJTSR. Von willebrand factor permeates small vessels in cadasil and inhibits smooth muscle gene expression. *Translational stroke research*. 2012. **3**:138-145
42. Lopes da Silva M, Cutler DF. Von willebrand factor multimerization and the polarity of secretory pathways in endothelial cells. *Blood*. 2016. **128**:277-285
43. MacRae JM, Ahmed S, Hemmelgarn B, Sun Y, Martin BJ, Roifman I, Anderson T. Role of vascular function in predicting arteriovenous fistula outcomes: An observational pilot study. *Canadian journal of kidney health and disease*. 2015. **2**:19
44. Owens CD, Wake N, Kim JM, Hentschel D, Conte MS, Schanzer A. Endothelial function predicts positive arterial-venous fistula remodeling in subjects with stage IV and V chronic kidney disease. *The journal of vascular access*. 2010. **11**:329-334
45. Wong C-Y, de Vries MR, Wang Y, van der Vorst JR, Vahrmeijer AL, van Zonneveld AJ, Roy-Chaudhury P, Rabelink TJ, Quax PHA, Rotmans JJ. Vascular remodeling and intimal hyperplasia in a novel murine model of arteriovenous fistula failure. *Journal of Vascular Surgery*. 2014. **59**:192-201.e191
46. Groeneveld DJ, van Bekkum T, Cheung KL, Dirven RJ, Castaman G, Reitsma PH, van Vlijmen B, Eikenboom J. No evidence for a direct effect of von willebrand factor's ABO blood group antigens on von willebrand factor clearance. *Journal of Thrombosis and Haemostasis*. 2015. **13**:592-600
47. Kang L, Grande JP, Hillestad ML, Croatt AJ, Barry MA, Katusic ZS, Nath KA. A new model of an arteriovenous fistula in chronic kidney disease in the mouse: Beneficial effects of upregulated heme oxygenase-1. *Am J Physiol Renal Physiol*. 2016. **310**:F466-F476
48. Kokubo T, Ishikawa N, Uchida H, Chasnoff SE, Xie X, Mathew S, Hruska KA, Choi ET. CKD accelerates development of neointimal hyperplasia in arteriovenous fistulas. 2009. **20**:1236-1245
49. Liang M, Liang A, Wang Y, Jiang J, Cheng J. Smooth muscle cells from the anastomosed artery are the major precursors for neointima formation in both artery and vein grafts. *Basic research in cardiology*. 2014. **109**:431
50. Skartsis N, Manning E, Wei Y, Velazquez OC, Liu Z-J, Goldschmidt-Clermont PJ, Salman LH, Asif A, Vazquez-Padron RI. Origin of neointimal cells in arteriovenous fistulae: Bone marrow, artery, or the vein itself? 2011. **24**:242-248
51. Vazquez-Padron RI, Martinez L, Duque JC, Salman LH, Tabbara M. The anatomical sources of neointimal cells in the arteriovenous fistula. *The journal of vascular access*. 2021:11297298211011875

