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Guardians of the gut: harnessing bioinformatics to study the gut microbiome and faecal microbiota transplantation in intestinal disorders

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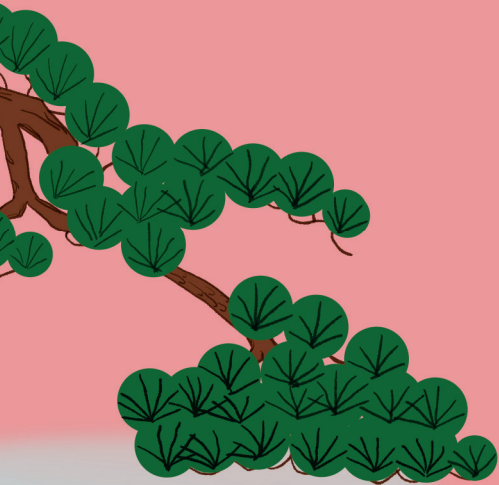
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Chapter 6

Ecological resilience in ulcerative colitis: Microbial dynamics of donor and resident species in a longitudinal fecal microbiota transplantation study

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Abstract

Fecal Microbiota Transplantation (FMT) has emerged as promising treatment for the chronic immune-mediated disease ulcerative colitis (UC). However, the ecological dynamics underlying clinical remission remain poorly understood. To investigate these dynamics, we analysed data from 24 UC patients treated with four rounds of FMT donated by two healthy donors. Microbiota samples from patients were collected at nine standardised timepoints before, during, and after treatment, covering a period of 14 weeks. Additionally, 27 donor samples were analysed. Species detected in the recipients' gut microbiota were categorised into ecological categories based on their origin and temporal dynamics: species already present in the host pre-FMT, species derived from the donor, or novel species, i.e. absent in both the pre-FMT host and the donor but detected later. Overdispersed Poisson regression models with random effects were employed to model the number of species within each category over time. Furthermore, we investigated the change in relative abundance for species present in the host pre-FMT. The results revealed that host species with higher relative abundances prior to FMT were more likely to persist following FMT. Notably, patients who achieved combined clinical and endoscopic remission at week 14 retained a significantly higher number of host species compared to non-responders. In contrast, non-responders initially exhibited a higher colonisation of donor species than responders, but their number decreased significantly over time in non-responders. These findings suggest that clinical remission following FMT is associated with a resilient patient gut community, capable of controlled incorporation of donor species, without replacing resident species.

Introduction

Fecal microbiota transplantation (FMT) is the transfer of fecal matter, including gut microorganisms, from the intestine of a healthy donor to a diseased recipient with the goal of modulating the recipient's disturbed microbiota.¹⁻³ FMT has demonstrated to be effective in recurrent *Clostridioides difficile* infection^{2,3}, but success rate is lower for more complex diseases, such as inflammatory bowel disease (IBD).^{4,5} A possible cause for the lower success rate of FMT in complex diseases is the tendency of the recipient's microbiota to revert to its original pre-FMT adverse state.⁶ Transition to a healthier state is likely helped by the successful colonisation (engraftment) of donor-derived microorganisms. Therefore, it has been suggested that the success of FMT depends on the donor's microbial diversity and composition.^{7,8} The extent to which shifts in the patient's microbiota towards the donor's microbiota are beneficial for resolving gut disturbances remains unclear.^{6,9-11} This donor-centric view has been challenged, and the importance of the recipient and procedural factors to determine FMT outcomes has been highlighted.¹²⁻¹⁵

In previous analyses of the FMT trial for ulcerative colitis (UC) we examined the engraftment of specific microbial species following FMT, and their associations with clinical remission.^{11, 16} For this, we analysed the data from a randomized controlled trial (RCT) involving 24 UC patients treated with four rounds of FMT donated by two healthy donors. Interestingly, we observed that the rate of microbial engraftment did not correlate with successful clinical remission¹⁶, a paradox also noted in a meta-analysis conducted by Schmidt et al. (2022) involving 316 FMT procedures.¹² In their study, clinical success was not correlated with donor strain colonisation or replacement of recipient species. Instead, recipient factors seemed to play a more important role in determining FMT outcomes than donor-related factors.¹² The seemingly limited role of engraftment in predicting clinical outcome of FMT defies the super-donor hypothesis and necessitates deeper investigation into the ecological changes underlying clinical remission.

In this study, the role of donor and host microbial species in determining clinical outcome of FMT is investigated further by applying the conceptual framework introduced by Schmidt et al. (2022)¹² to a longitudinal setting. We capitalize on a randomized controlled trial¹⁶ with dense repeated sampling to map the succession dynamics in the recipient's gut microbiota of UC patients following FMT treatment in relation to clinical remission. Our analysis focuses on ecological dynamics on a species level, categorising all taxa based on their origin and temporal presence: already present in the host before FMT, derived from the donor, or detected during or after the FMT therapies while absent in both the pre-FMT host and the donor.

Methods

The study population

A total of 24 adult patients experiencing mild to moderate exacerbations of UC were included in a double-blind randomized controlled trial conducted at LUMC.¹⁶ Written informed consent was obtained from all study participants prior to their participation. Demographic variables and subject characteristics are provided in Supplementary Table S1, with further details on the study population and clinical characteristics provided by van Lingem et al. (2024) and in Supplementary Information S1.¹⁶

Following pretreatment with either budesonide (n=12) or placebo (n=12), patients received four fecal transplants at weekly intervals. Donors (D07 and D08) were randomly assigned. FMTs were infused in the duodenum via a nasoduodenal tube or gastroscope.¹⁷ Stool samples were obtained before and after the pretreatment phase, four times before every FMT, and 1 week, 4 weeks, and 8 weeks after treatment. At the end of the study, at week 14, a sigmoidoscopy was performed to assess the endoscopic MAYO score. Clinical remission (i.e. response) was defined as no symptoms (partial MAYO score of 2 with no individual sub score of >2) and an endoscopic MAYO score 0-1. A total of ten patients achieved combined clinical and endoscopic remission (n=9) or partial remission (n=1). Of the 14 non-responders, 10 patients left the study early (in total 2 patients did not finish all 4 FMT treatments) because their symptoms worsened.^{11, 16} For this study we defined a responder as a patient in combined clinical and endoscopic remission at week 14 (n=9).

Microbiota data

DNA was extracted from the donor and recipient stool samples and shotgun sequenced with 100 bp single-end reads to a median depth of 2.9m reads by Diversigen (New Brighton, Minneapolis, USA) using the Illumina NovaSeq platform. Raw reads mapping to the human genome were removed using bowtie2 (version 2.4.2)¹⁸ and the GRCh37 reference genome and reads were quality-trimmed using fastq (version 0.20.1)¹⁹, both of which are part of an in-house workflow (<https://git.lumc.nl/snooij/metagenomics-preprocessing>). The mOTUs3 workflow (version 3.0.1) was used to generate taxonomic profiles.^{20, 21} Unassigned, human-derived, archaeal, and low-quality reads were removed from the data, which resulted in 1552 unique mOTUs. For the sake of simplicity, we use the term 'species' to refer to unique mOTUs throughout. The results table was then imported into R for analyzing the data, visualizing the results and performing the statistical tests (R version 4.2.2). R code is available via GitHub (https://github.com/susannepinto/FECBUD_microbiome.git).

Mapping ecological categories

Respectively 13 and 14 samples were available for donor D07 and donor D08. Note that every recipient received FMT material from only one of the donors. We could not

match every recipient sample to a specific donor sample used for the FMT, because not every donor sample used for FMT was sequenced. Therefore, we created a dataset with the core microbiota for each donor. The core donor microbiota was defined as having its relative abundance higher than the detection limit of 0.1% in at least one sample. The core donor microbiota yielded 120 and 84 unique species for donors D07 and D08, respectively.

Subsequently, we created a presence/absence dataset of all species per recipient and per timepoint, and every species was assigned to an ecological category per recipient and per timepoint based on its origin and presence over time, according to the decision tree presented in Figure 1 (detailed explanation Supplementary Information S2). Per recipient, for every species ever present at any timepoint in the recipient, or present in the microbiota of the associated donor, a comparison was made with the recipient's pre-FMT sample and with the microbiota of the corresponding donor. All species present in the recipient's pre-FMT sample were placed into a host category (Resident, Host transient, or Species loss), depending on the pattern of presence over time. If species were unique for the donor relative to the recipient's pre-FMT samples, species were placed into a donor category (Colonisation, Donor transient, or Rejection). If species were not present in the host pre-FMT or in the microbiota of the donor, they were classified as a novel species (Novel, Novel transient, or Novel loss). Within these broad categories, a species was further categorised as a stable (Resident, Colonisation, or Novel), intermittent (Host transient, Donor transient or Novel transient), or previous occupant (Species loss, Rejection, or Novel loss) in the microbiota, depending on the presence at that moment and at the previous timepoints. Because absence in microbiota data can also mean that the abundance was under the detection limit, in the base case we allowed for each species the occurrence of one single absence without direct consequences for categorisation in the rest of the timeseries. Due to the way the categories are defined, some categories cannot occur at the first timepoints. For example, a donor-derived species first had to colonise the gut (colonisation), then be absent for at least two timepoints (NA and Rejection), and then be detected again to be categorised as a Donor transient species (Supplementary Information S2).

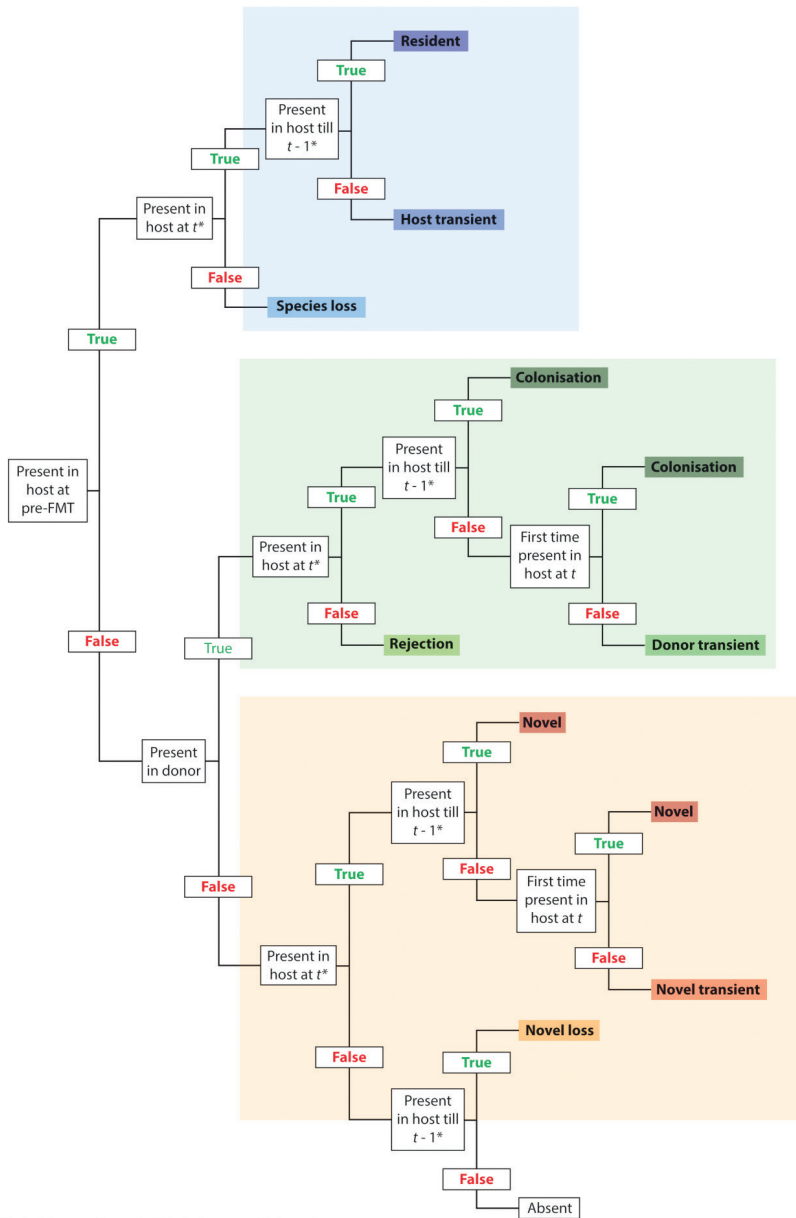
In sensitivity analyses, we tested some variations to the base case criteria regarding the temporal information used for categorising the species. In Sensitivity 1 we did not allow the occurrence of any absence when categorising species into either of the host, donor, or novel categories (Fig. 1). In Sensitivity 2 we only considered the presence/absence at the previous timepoint instead of all the previous timepoints (Supplementary Figure S3). In contrast, in Sensitivity 3 the presence of species at all timepoints is considered in categorisation of species at a particular timepoint (Supplementary Figure S4). Sensitivity 4 is the same as Sensitivity 3 but with the added criterion of not allowing the occurrence of any absence (Supplementary Figure S4). In Supplementary Information S2 examples on categorisation of species and the differences between the sensitivity analyses are illustrated.

Modelling the number of species across ecological categories

We modelled the number of species across ecological categories by means of overdispersed Poisson regression models with random effects to accommodate correlation between repeated measurements per recipient. For this, we employed a generalized linear mixed-effects model (GLMM) for the negative binomial family with log-link using the `glmer.nb` function from the `lme4` R package.²² The temporal evolution of the expected log-number of species in each category was modelled with a spline transformation of the original time variable (in weeks since start of FMT treatment). Estimates from the spline model were compared to those from a linear growth model in sensitivity analysis, by modelling the expected log-number of species as a simple linear function of time. Possible differences in succession dynamics between responders and non-responders were investigated by adding the treatment response variable as a covariate to the model, and through specification of interaction terms with time and ecological category. Patient specific variables, namely, donor (donor D07 vs. D08), pretreatment (budesonide vs. placebo), age and sex (female vs. male), were included based upon their role as possible confounders.

Change in population abundances of host-derived species

To explore the dynamics of host-derived species in response to FMT in more detail, we investigated the relative abundance over time for the species that were already present in the host pre-FMT. Results reveal the distribution of abundance differences at particular timepoints across subjects per ecological category for the species that were already present pre-FMT. In addition, we compared the baseline distributions among species that were later categorized as resident, host transient, and lost among both responders and non-responders. Finally, we also calculated the differences in microbial abundance before and after FMT for all species that were present in the recipients' pre-FMT samples. Because several non-responder patients quitted early during the study, we only included patients who completed all four rounds of FMT ($n = 22$ patients, of whom 9 were defined as responders) and used the last available post-FMT measurement when calculating the difference in relative abundance before and after FMT. Because the abundance distributions were right-skewed, we used a natural log transformation of the abundances. Consequently, the abundance differences on the log scale can be interpreted as proportional differences on the original scale (in percentage difference). To assess the significance of these differences between responders and non-responders, linear mixed-effects models (LMM) were applied, accounting for the correlation of repeated observations within each patient (using the `lmer` function from the `lme4` R package).²²



*The first absence of a species (after being present) is ignored

Figure 1. Decision tree used to assign species to ecological categories. The categories are based on the origin and presence of a species over time. First, the species was compared to the pre-FMT host samples, then to the core donor microbiota. Next, the presence/absence at all previous timepoints was considered to assign the species to an ecological category. Note that we ignored the first absence of species when categorising species as lost or as transient upon re-detection. In Sensitivity 1 we evaluated whether this choice had an impact on the results (Supplementary Information S2).

Results

Succession of host-derived, donor-derived, and novel species following FMT
 To study the succession dynamics of species during and after FMT in our UC cohort, we modelled the number of species across ecological categories and investigated differences between responders and non-responders (Figure 2). In these models, donor and sex were included as covariates, while pretreatment and age were not relevant as confounders. Supplementary Figure S1 shows the specific parameter estimates of the model depicted in Figure 2.

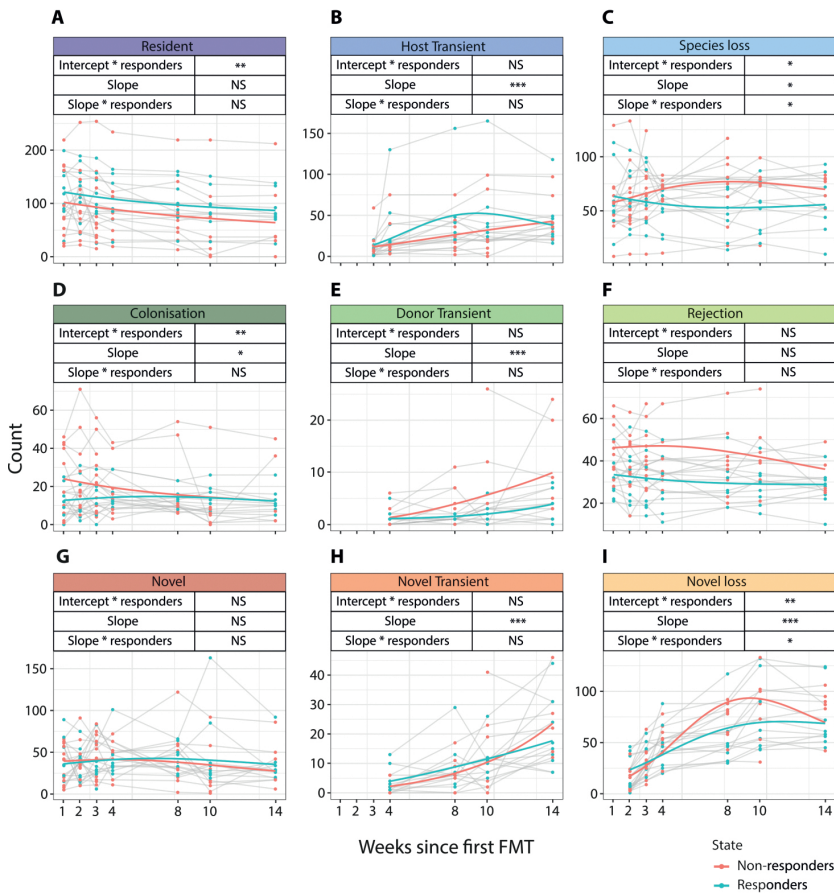


Figure 2. Changes in the number of species per ecological category over time. Average trajectories among responders to the treatment are indicated with blue lines, average trajectories among non-responders with red lines. Individual patient trajectories are shown with grey lines. Note the different scaling of the y-axis per category. The model contained a random intercept per patient to account for repeated measurements. Time was modelled with a spline. The levels of significance are reported above each plot and are indicated by asterisks (***: $p < 0.01$; **: $p < 0.05$; NS: not significant).

At the start of the study, we observed a significantly higher number of host species in the resident categories (species that were present in the patient's gut pre-FMT) among responders compared to non-responders, and this difference persisted over time (Figure 2A). Although the number of resident species declined over time in both responders and non-responders this decrease was not statistically significant. In contrast, the number of host transient species increased significantly over time in both patient groups (Figure 2B). Of note, this increase may be partly attributable to the definition of host-derived species being transient upon re-detection after temporary absence. Non-responder patients exhibited a significantly greater loss of host species over time compared to responders, in whom, the number of host species lost decreased significantly over time (Figure 2C).

Conversely, non-responders were initially colonised by a significantly higher number of donor species compared to responders. However, the number of colonising species in non-responders significantly declined over time, whereas it remained constant in responders (Figure 2D). The number of donor transient species was similar between the two patient groups at the start of the study and showed a significant increase over time, especially in non-responders. However, this category remained relatively small and differences according to treatment response were not significant (Figure 2E). The number of rejected donor species was higher at baseline and over time for non-responders compared to responders, however this difference also did not reach statistical significance (Figure 2F).

The number of novel species detected post-FMT was similar for both responders and non-responders and remained constant in time (Figure 2G). The number of novel transient species increased significantly over time, this increase was more or less similar for both the responders and non-responders (Figure 2H). Initially, the responders lost significantly more novel species than the non-responders, but over time the latter group lost significantly more novel species than the responders (Figure 2, panel I).

We also found significant differences between responders and non-responders in the host transient and novel transient categories when applying a linear growth model instead of splines for the temporal evolution of number of species in each category (Supplementary Figure S2). It should be noted that these categories contained relatively few species, and the lack of statistical significance when using splines is likely explained by a reduced statistical power. Importantly, all differences between responders and non-responders identified by the spline model were retained in the linear growth model for category size (Supplementary Figure S2).

Sensitivity analyses

We conducted four different sensitivity analyses concerning the categorisation of the species. To illustrate the effect of categorisation on the rates of change over time, we

generated a plot of the average slope estimates according to each sensitivity analysis (Supplementary Figures S5-S9). Sensitivity analysis 1 resulted in a slightly stronger decline in the number of species for the resident, colonisation, and novel categories (Supplementary Figures S5, S9, and S10). This outcome is a logical consequence of the criterion that a species can no longer be absent for a single time point. Consequently, the likelihood of a species moving to a different category (transient or loss) increased, since it was by definition not possible to return to the categories denoting stable presence over time. This resulted in transient categories having higher intercepts, but the average slopes remained unchanged for all other categories (Supplementary Figures S5, S9, and S10). Similarly, for Sensitivity analysis 2, no substantial differences from the base case were found (Supplementary Figures S6, S9, and S10). The most profound differences were noted in the slopes of the resident and transient categories. The slopes of the transient categories were smaller, especially for the host-derived species among non-responders (Supplementary Figures S6, S9, and S10). Sensitivity analyses 3 and 4 led to more stable patterns over time, especially for the resident category, as compared to both the base case scenario and the other sensitivity analyses (Supplementary Figures S7-S10). This stability can be attributed to the modifications in the category assignment criteria in Sensitivity analyses 3 and 4, where stable presence is defined on all timepoints. Consequently, fewer species were assigned to the resident, colonisation and novel categories and more to the transient categories (Supplementary Information S2).

Relative abundances of host resident species pre- and post-FMT

We further assessed changes in the relative abundance of species present in the gut prior to treatment to investigate whether the relative abundance pre-FMT is indicative of the category that a species will reach post-FMT. Host transient species displayed significantly lower relative abundances at all timepoints compared to resident species (Figure 3A and Supplementary Table S2). In both responders and non-responders host species with higher pre-FMT relative abundances were more likely to become resident species compared to host transient or lost species in both recipient groups (Figure 3B, Supplementary Figure S11, and Supplementary Table S2). Therefore, our findings show that initial microbiota composition is associated with post-FMT composition. The differences in relative abundance of host resident species between the pre-FMT measurement and the last available post-FMT measurement were centered around zero (Figure 3C). A positive difference indicates an increase in the relative abundance of resident species following FMT, while a negative difference denotes a decrease. Thus, approximately equal numbers of resident species showed either a positive or negative response to FMT. No significant differences were found between responders and non-responders in relative abundances of resident species in response to FMT (Figure 3C, Supplementary Figure S12, and Supplementary Table S2).

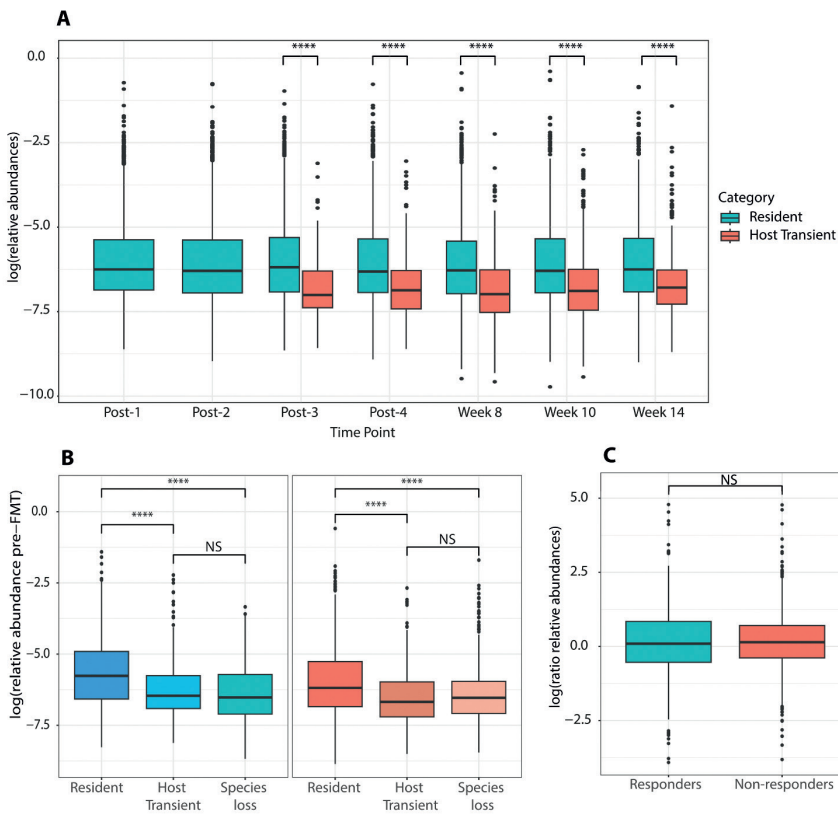


Figure 3. Comparison of relative abundances of species in different categories. A) Relative abundances of Resident and Host Transient species over time. Here, no distinction has been made between responders and non-responders. B) Relative abundance of host species at pre-FMT measurement. The relative abundances in resident, host transient, and species loss species between responders (blue) and non-responders (red) are not significant (Supplementary Table S2). C) Difference in relative abundance in resident species between pre-FMT and last available post-FMT measurement. Significance was tested with linear mixed-models and shown in the plots (***: $p < 0.01$; **: $p < 0.01$; *: $p < 0.05$; NS: not significant).

Discussion

The success of FMT for UC is ultimately determined by whether the patient achieves clinical and endoscopic remission after treatment. It has been suggested that treatment success is related to the extent to which the recipient's microbiota composition shifts towards that of the donor.^{7, 23} However, we found no evidence supporting this link, in line with several other studies.^{10-13, 16}

We used an ecological framework of succession to investigate microbiota dynamics associated with clinical success of FMT. Microbial species were categorised as pre-existing in the host before FMT, donor-derived, or newly detected. We found that responders retained a higher number of host species compared to non-responders. Although non-responders initially exhibited colonisation by more donor species than responders, this colonisation in non-responders declined over time and eventually became equal to the levels observed in responders. These findings suggest that a successful clinical response to FMT may be facilitated by a microbiota receptive to colonisation without compromising the resident microbiota. Additionally, non-responders lost substantially more novel species over time compared to responders, indicating that newly detected species failed to establish stably within the non-responder gut microbiota. This finding suggests less robust alterations in gut microbiota composition among non-responders. A successful FMT may induce a shift in which the recipient's microbiota integrates donor and novel species, achieving a balanced coexistence to restore the gut microbial ecosystem. This observation aligns with earlier research.^{12,24} Our study expands upon previous analyses by using longitudinal analysis of UC patients, thereby providing a fine-grained view of the ecological dynamics over time of donor and host species following FMT.

FMT can be seen as a perturbation experiment on the gut microbiota, creating a dynamic interplay between donor and recipient communities, which may open ecological niches for other microorganisms.^{12,25} The balance between the engraftment of beneficial microorganisms and competition with deleterious microorganisms in the recipient gut, combined with systemic host processes, such as the modulation of immune responses and the interaction with (external) environmental factors and genetic characteristics, could initiate clinical remission.⁴ The process of microbial invasion involves various challenges that incoming microorganisms need to overcome to establish colonisation and influence the existing microbial community. It is important for the invading species to achieve sufficient metabolic activity in the gut to interact with the resident community. This interaction may also be achieved by transient species, indicating that permanent colonisation is not always necessary.²⁵ Analogous to nurturing an ecosystem such as a crop field through biological control, FMT necessitates the introduction of donor species with healthy functional properties to modify the recipient's system rather than inducing wholesale changes that might lead to the extinction and the replacement of existing microbial inhabitants. Therefore, the recipient microbiota must exhibit a degree of resilience, allowing it to integrate donor species without completely altering its composition. FMTs may also strengthen recipient species by introducing beneficial spores or metabolites, thereby enhancing the stability and functionality of the recipient's own microbiota.²⁶ The stability of the microbiota is maintained through controlled species loss, ensuring that introduced organisms integrate harmoniously with the pre-existing ecosystem.

The outcome of FMT is influenced by a range of ecological processes, spanning from neutral or stochastic factors (e.g. donor propagule pressure) to adaptive or selective factors (e.g. niche competition and differentiation).^{12, 27} This indicates a complex mechanism of action of FMT in patients with UC, necessitating the establishment of a novel homeostasis between the donor and recipient microbiota. This complexity may also explain why prolonged FMT treatment with multiple donor infusions appears necessary in UC, as repeated exposition may be required to achieve an optimal balance between recipient and donor microbiota. This approach contrasts with the FMT treatment of rCDI, which is characterised by a depleted microbiota that can be effectively restored with a single infusion, with a cure rate of about 80%.¹

The success of FMT may not be reliant on resembling the donor's microbiota, but rather on establishing a complementary relationship, emphasizing the importance of selecting donors whose microbiota optimally aligns with the recipient's specific needs.⁷ Unlike the developmental stages of a child's microbiota, the gut microbiota of a UC patient is already an established, independent microbial community. This pre-existing microbiota makes the introduction of new species and the induction of change considerably more challenging.^{28, 29} Tailoring the selection of FMT donors to those enriched in taxa capable of restoring disturbed metabolic pathways in the recipient might enhance the effectiveness of FMT, particularly in metabolic dysfunction associated diseases.^{6, 7, 10} For example, incoming species that are metabolically complementary to the recipient's community, by introducing novel functions or by occupying previously unfilled niches, may be more likely to colonise the resident community.^{30, 31} In addition, a high diversity in the donor and low diversity in the recipient may further influence the success of colonisation.^{10, 32}

From an ecological perspective, our findings suggest that donor and recipient species can coexist. We might hypothesise that they occupy distinct metabolic niches. Moreover, we observed that species with a higher abundance prior to FMT (the main 'founders') are more likely to persist during the FMT than species with a lower abundance. This implies that the competitive strength of the resident species is related to their abundance, indicating that within each metabolic niche, communities are built by random winners, driven by stochastic colonisation.³³ This is in line with ecological studies showing that functional differences create opportunities for coexistence (niche theory). However, within each niche functionally similar species can coexist, and communities are structured to random stochastic rules (neutral theory).³⁴ Within the gut microbiota, species often have overlapping functions, allowing them to replace each other and take over specific functional traits if one species is perturbed or removed.³⁵

This study has several limitations. The first concerns the classification of patients into responders and non-responders. Patients that dropped out early due to worsening symptoms were classified as non-responders. Microbiota data were not collected for

these patients, as a result, this potentially introduces bias into the results for the non-responder group. Moreover, the study concerns only 24 UC patients and the time series up to 14 weeks represents only a snapshot of the dynamic process of microbial succession. This sample size is too small to draw definite conclusions and further investigation into longer-term outcomes is necessary to gain a more comprehensive understanding.³⁶ A third limitation is the sequencing depth (2.9 M 100bp single-end Illumina), which does not allow for definitive determination whether an absent species was actually absent in the host or donor, or simply undetected.³¹ Also, the low sequencing resolution makes it impossible to determine whether the same strain present in the donor sample successfully colonised the recipient's gut microbiota or whether the donor and host strains coexisted or were replaced following FMT. Lastly, we did not have data to directly link the unique donor sample used for FMT to the corresponding recipient samples. Therefore, we used the combined microbiota data, which may have led to the misclassification of some low abundant colonising species from the donor as novel species.

By applying an ecological perspective to FMT, our study sheds new light on the importance of ecological principles, such as succession of microorganisms and the resilience of the recipient's system, in shaping therapeutic outcomes. Our study reveals the ecological dynamics of the gut microbiota during and after FMT in patients with UC, with a particular focus on the dynamics of recipient, donor, and novel species. Contrary to some previous studies, the overall engraftment of the donor microbiota did not emerge as the most important factor for FMT success in this study.^{7, 13} The key factor influencing the response may not be the overall engraftment of donor species, but rather the recipient's ability to retain resident species while simultaneously enriching specific novel and donor species. Thus, successful FMT hinges on fostering a microbiota shift that complements rather than compromises the existing ecosystem. This ecological interpretation aids in understanding the mechanism through which FMT may induce clinical remission and also underscores the nuanced interplay between donor and recipient microbiota essential for therapeutic efficacy.

Ethics approval and consent to participate

This research project was reviewed and approved by the Medical Ethical Committee of the LUMC, with reference number NL 65976.098.18. The study was registered in the Netherlands Trial Register, with reference number NL9858.

Consent for publication

Not applicable

Availability of data and material

R code is available via GitHub (https://github.com/susannepinto/FECBUD_microbiome.git) and the in-house preprocessing workflow is available via <https://git.lumc.nl/snooij/metagenomics-preprocessing>. We have uploaded the metagenomic sequences to NCBI with: SRA Bioproject PRJNA1071720.

Competing interests

The authors (SP, EB, SN, ES, and JAB) declare that they have no competing interests. EMT, AEM, and JJK report a research grant and consulting fee from Vedanta Biosciences (Boston, MA, USA US). JJK serves in the scientific advisory board of Microviable Therapeutics (Gyon, Spain).

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Authors' contributions

The authors confirm contribution to the paper as follows: study conception and design: SP, EB, JAB; funding acquisition: JAB, EB, ES, EMT, AEM, JJK; data collection: SN, EMT, JJK, AEvdM-dj; data analysis and interpretation of results of microbiota analysis: SP, EB, JAB; draft manuscript preparation: SP, EB, JAB. All authors reviewed the results and approved the final version of the manuscript.

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List of Supplementary files

Information S1. Clinical and laboratory procedures

Information S2. Examples illustrating the categorisation of the species in the base case and in the four sensitivity analyses.

Table S1. Clinical and demographic information of non-responders and responders.

Table S2. Model estimates and p-values for the differences in relative abundances.

Figure S1. Results of modelling the number of species per ecological category for the base case.

Figure S2. Results of modelling the number of species per ecological category for the base case.

Figure S3. Decision tree used in Sensitivity 2 to assign species to ecological categories according to different inclusion criteria.

Figure S4. Decision tree to assign species to ecological categories according to different criteria as in the base case analysis (Sensitivity 3 and 4).

Figure S5. Changes in the number of species per ecological category over time for Sensitivity 1.

Figure S6. Changes in the number of species per ecological category over time for Sensitivity 2.

Figure S7. Changes in the number of species per ecological category over time for Sensitivity 3.

Figure S8. Changes in the number of species per ecological category over time for Sensitivity 4.

Figure S9. Average changes in the number of species per ecological category over time for the base case (BC) and all Sensitivity (S1, S2, S3, and S4) analyses.

Figure S10. Distribution of the number of species per ecological category for the base case and all sensitivity analyses estimated by overdispersed Poisson regression models with random effects.

Figure S11. Histograms showing the absolute abundance of host species (Resident, Host transient and Species loss) pre-FMT.

Figure S12. Histograms showing the distribution of the differences in relative abundance of resident species (pre- and post FMT).

Supplementary documents are available online on the publisher's website:



<https://academic.oup.com/ismecommun/advance-article/doi/10.1093/ismeco/ycaf119/8203245?searchresult=1#supplementary-data>

References

1. van Nood E, Vrieze A, Nieuwdorp M, Fuentes S, Zoetendal EG, de Vos WM, et al. Duodenal infusion of donor feces for recurrent *Clostridium difficile*. *N Engl J Med*. 2013;368(5):407-415.
2. Debast SB, Bauer MP, Kuijper EJ, Diseases ESoCMal. European Society of Clinical Microbiology and Infectious Diseases: update of the treatment guidance document for *Clostridium difficile* infection. *Clin Microbiol Infect*. 2014;20 Suppl 2:1-26.
3. Ianiro G, Puncoschar M, Karcher N, Porcari S, Armanini F, Asnicar F, et al. Variability of strain engraftment and predictability of microbiome composition after fecal microbiota transplantation across different diseases. *Nat Med*. 2022;28(9):1913-1923.
4. Hanssen NMJ, de Vos WM, Nieuwdorp M. Fecal microbiota transplantation in human metabolic diseases: From a murky past to a bright future? *Cell Metab*. 2021;33(6):1098-1110.
5. El Hage Chehade N, Ghoneim S, Shah S, Chahine A, Mourad FH, Francis FF, et al. Efficacy of Fecal Microbiota Transplantation in the Treatment of Active Ulcerative Colitis: A Systematic Review and Meta-Analysis of Double-Blind Randomized Controlled Trials. *Inflamm Bowel Dis*. 2023;29(5):808-817.
6. Sommer F, Anderson JM, Bharti R, Raes J, Rosenstiel P. The resilience of the intestinal microbiota influences health and disease. *Nat Rev Microbiol*. 2017;15(10):630-638.
7. Wilson BC, Vatanen T, Cutfield WS, O'Sullivan JM. The Super-Donor Phenomenon in Fecal Microbiota Transplantation. *Front Cell Infect Microbiol*. 2019;9:2.
8. Vermeire S, Joossens M, Verbeke K, Wang J, Machiels K, Sabino J, et al. Donor Species Richness Determines Faecal Microbiota Transplantation Success in Inflammatory Bowel Disease. *J Crohns Colitis*. 2016;10(4):387-394.
9. Rees NP, Shaheen W, Quince C, Tselepis C, Horniblow RD, Sharma N, et al. Systematic review of donor and recipient predictive biomarkers of response to faecal microbiota transplantation in patients with ulcerative colitis. *EBioMedicine*. 2022;81:104088.
10. He R, Li P, Wang J, Cui B, Zhang F, Zhao F. The interplay of gut microbiota between donors and recipients determines the efficacy of fecal microbiota transplantation. *Gut Microbes*. 2022;14(1):2100197.
11. Pinto S, Sajbenova D, Beninca E, Nooij S, Terveer EM, Keller JJ, et al. Dynamics of Gut Microbiota after Fecal Microbiota Transplantation in Ulcerative Colitis: Success Linked to Control of Prevotellaceae. *J Crohns Colitis*. 2024.
12. Schmidt TSB, Li SS, Maistrenko OM, Akanni W, Coelho LP, Dolai S, et al. Drivers and determinants of strain dynamics following fecal microbiota transplantation. *Nat Med*. 2022;28(9):1902-1912.
13. Olesen SW, Gerardin Y. Re-Evaluating the Evidence for Faecal Microbiota Transplantation 'Super-Donors' in Inflammatory Bowel Disease. *J Crohns Colitis*. 2021;15(3):453-461.
14. Danne C, Rolhion N, Sokol H. Recipient factors in faecal microbiota transplantation: one stool does not fit all. *Nat Rev Gastroenterol Hepatol*. 2021;18(7):503-513.
15. Peri R, Aguilar RC, Tuffers K, Erhardt A, Link A, Ehlermann P, et al. The impact of technical and clinical factors on fecal microbiota transfer outcomes for the treatment of recurrent *Clostridioides difficile* infections in Germany. *United European Gastroenterol J*. 2019;7(5):716-722.

16. van Lingen E, Nooij S, Terveer E, Crossette E, Prince A, Bhattarai S, et al. Fecal Microbiota Transplantation engraftment after budesonide or placebo in patients with active ulcerative colitis using pre-selected donors: a randomized pilot study. *J Crohns Colitis*. 2024;jjae043:1381-1393.
17. Terveer EM, Vendrik KE, Ooijevaar RE, Lingen EV, Boeije-Koppenol E, Nood EV, et al. Faecal microbiota transplantation for *Clostridioides difficile* infection: Four years' experience of the Netherlands Donor Feces Bank. *United European Gastroenterol J*. 2020;8(10):1236-1247.
18. Langmead B, Salzberg SL. Fast gapped-read alignment with Bowtie 2. *Nat Methods*. 2012;9(4):357-359.
19. Chen S, Zhou Y, Chen Y, Gu J. fastp: an ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics*. 2018;34(17):i884-i890.
20. Milanese A, Mende DR, Paoli L, Salazar G, Ruscheweyh HJ, Cuenca M, et al. Microbial abundance, activity and population genomic profiling with mOTUs2. *Nat Commun*. 2019;10(1):1014.
21. Ruscheweyh HJ, Milanese A, Paoli L, Karcher N, Clayssen Q, Keller MI, et al. Cultivation-independent genomes greatly expand taxonomic-profiling capabilities of mOTUs across various environments. *Microbiome*. 2022;10(1):212.
22. Bates D, Mächler M, Bolker B, Walker S. Fitting Linear Mixed-Effects Models Using lme4. *Journal of Statistical Software*. 2015;67(1).
23. Xiao Y, Angulo MT, Lao S, Weiss ST, Liu YY. An ecological framework to understand the efficacy of fecal microbiota transplantation. *Nat Commun*. 2020;11(1):3329.
24. Li SS, Zhu A, Benes V, Costea PI, Hercog R, Hildebrand F, et al. Durable coexistence of donor and recipient strains after fecal microbiota transplantation. *Science*. 2016;352(6285):586-589.
25. Walter J, Maldonado-Gomez MX, Martinez I. To engraft or not to engraft: an ecological framework for gut microbiome modulation with live microbes. *Curr Opin Biotechnol*. 2018;49:129-139.
26. Gonze D, Lahti L, Raes J, Faust K. Multi-stability and the origin of microbial community types. *ISME J*. 2017;11(10):2159-2166.
27. Schmidt TSB, Raes J, Bork P. The Human Gut Microbiome: From Association to Modulation. *Cell*. 2018;172(6):1198-1215.
28. Bosch A, Levin E, van Houten MA, Hasrat R, Kalkman G, Biesbroek G, et al. Development of Upper Respiratory Tract Microbiota in Infancy is Affected by Mode of Delivery. *EBioMedicine*. 2016;9:336-345.
29. van Best N, Hornef MW, Savelkoul PH, Penders J. On the origin of species: Factors shaping the establishment of infant's gut microbiota. *Birth Defects Res C Embryo Today*. 2015;105(4):240-251.
30. Grinspan AM, Kelly CR. Fecal Microbiota Transplantation for Ulcerative Colitis: Not Just Yet. *Gastroenterology*. 2015;149(1):15-18.
31. Darcy JL, Washburne AD, Robeson MS, Prest T, Schmidt SK, Lozupone CA. A phylogenetic model for the recruitment of species into microbial communities and application to studies of the human microbiome. *ISME J*. 2020;14(6):1359-1368.

32. Lewin R. Complexity: life at the edge of chaos. 2nd ed: The university of Chicago Press; 1999.
33. Verster AJ, Borenstein E. Competitive lottery-based assembly of selected clades in the human gut microbiome. *Microbiome*. 2018;6(1):186.
34. Scheffer M, van Nes EH. Self-organized similarity, the evolutionary emergence of groups of similar species. *Proc Natl Acad Sci U S A*. 2006;103(16):6230-6235.
35. Lozupone CA, Stombaugh JI, Gordon JI, Jansson JK, Knight R. Diversity, stability and resilience of the human gut microbiota. *Nature*. 2012;489(7415):220-230.
36. Fuentes S, Rossen NG, van der Spek MJ, Hartman JH, Huuskonen L, Korpela K, et al. Microbial shifts and signatures of long-term remission in ulcerative colitis after faecal microbiota transplantation. *ISME J*. 2017;11(8):1877-1889.

