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Advancing helminth glycomics: structural specificity and immunogenicity of schistosomal and filarial glycans

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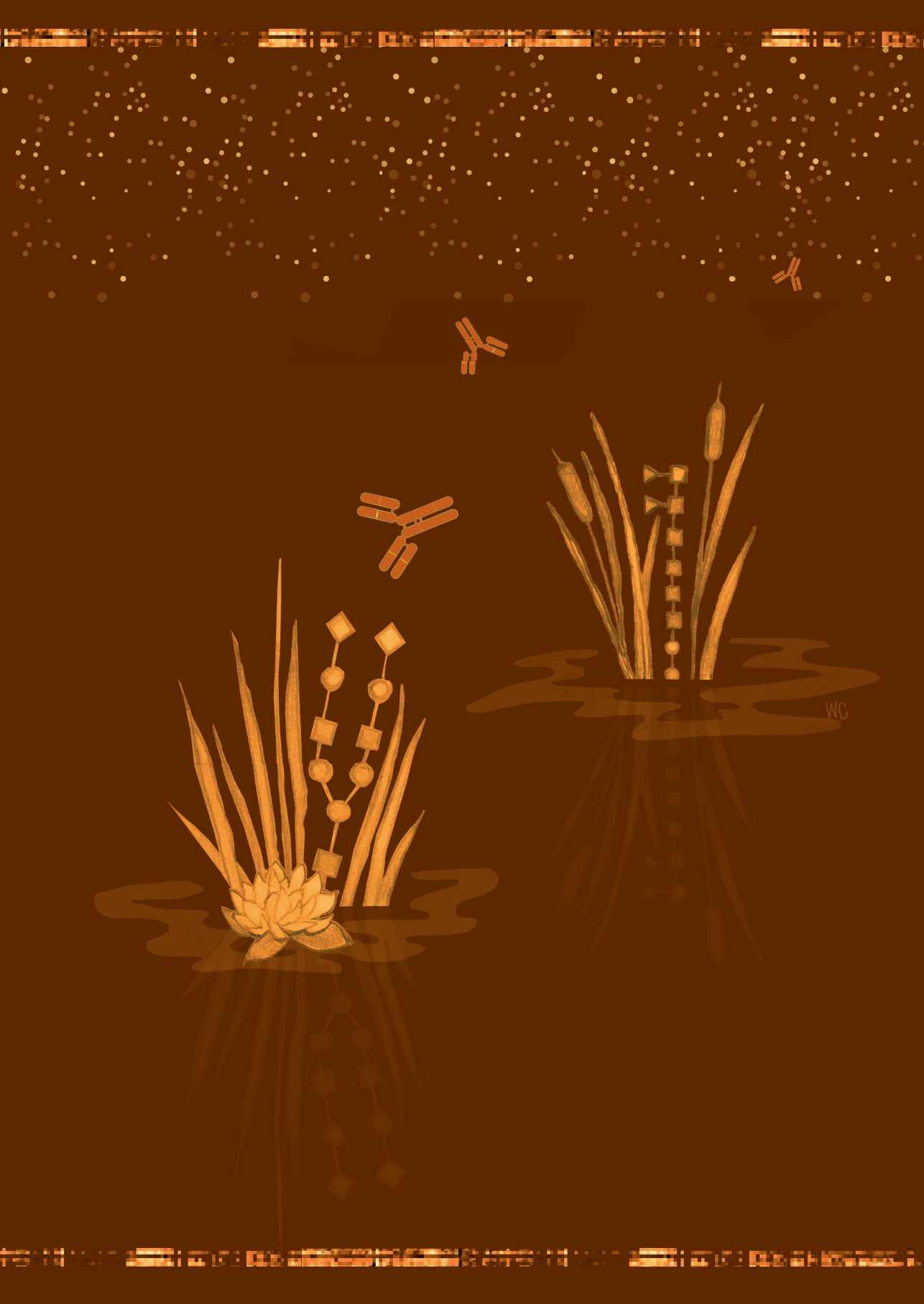
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Chapter IV.

Unraveling cross-reactivity of anti-glycan IgG responses in filarial nematode infections

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Abstract

Parasitic nematodes responsible for filarial diseases cause chronic disablement in humans worldwide. Elimination programs have substantially reduced the rate of infection in certain areas, but limitations of current diagnostics for population surveillance have been pointed out and improved assays are needed to reach the elimination targets. While serological tests detecting antibodies to parasite antigens are convenient tools, those currently available are compromised by the occurrence of antibodies cross-reactive between nematodes, as well as by the presence of residual antibodies in sera years after treatment and clearance of the infection. We recently characterized the *N*-linked and glycosphingolipid derived glycans of the parasitic nematode *Brugia malayi* and revealed the presence of various antigenic structures that triggered immunoglobulin G (IgG) responses in infected individuals. To address the specificity of IgG binding to these glycan antigens, we screened microarrays containing *Brugia malayi* glycans with plasma from uninfected individuals and from individuals infected with *Loa loa*, *Onchocerca volvulus*, *Mansonella perstans* and *Wuchereria bancrofti*, four closely related filarial nematodes. IgG to a restricted subset of cross-reactive glycans was observed in infection plasmas from all four species. In plasma from *Onchocerca volvulus* and *Mansonella perstans* infected individuals, IgG binding to many more glycans was additionally detected, resulting in total IgG responses similar to the ones of *Brugia malayi*-infected individuals. For these infection groups, *Brugia malayi*, *Onchocerca volvulus* and *Mansonella perstans*, we further studied the different IgG subclasses to *Brugia malayi* glycans. In all three infections, IgG1 and IgG2 appeared to be the major subclasses involved in response to glycan antigens. Interestingly, in *Brugia malayi*-infected individuals, we observed a marked reduction in particular in IgG2 to parasite glycans post-treatment with anthelmintic, suggesting a promising potential for diagnostic applications. Thus, we compared the IgG response to a broad repertoire of *Brugia malayi* glycans in individuals infected with various filarial nematodes. We identified broadly cross-reactive and more specific glycan targets, extending the currently scarce knowledge of filarial nematode glycosylation and host anti-glycan antibody response. We believe that our initial findings could be further exploited to develop disease-specific diagnostics as part of an integrated approach for filarial disease control.

Introduction

Parasitic filarial nematodes have been known to infect humans for many centuries¹, causing various types of diseases called filariasis². The lymphatic form, resulting from an infection with either *Brugia malayi*, *Brugia timori* or *Wuchereria bancrofti*, is the most widespread, with over 50 million people infected worldwide in 2018³. *Loa loa*, *Mansonella streptocerca* and *Onchocerca volvulus* cause subcutaneous forms of filariasis while *Mansonella ozzardi* and *Mansonella perstans* are responsible for serous cavity filariasis². Altogether, filarial nematodes represent a great burden in endemic areas, and both onchocerciasis and lymphatic filariasis (LF) are targeted for elimination as part of the World Health Organization (WHO) roadmap for 2030⁴. To achieve this goal, strategic intervention programs ranging from preventive chemotherapy to vector control (mosquitoes for LF and black flies for onchocerciasis) have to be implemented or strengthened. Stopping infection spread in populations in endemic areas relies on mass drug administration (MDA) of ivermectin for onchocerciasis⁵ and of a cocktail of ivermectin, diethylcarbamazine (DEC) and albendazole (IDA triple-therapy) for LF⁶, except in Africa, where a combination of albendazole and ivermectin is used, due to contraindications of DEC in onchocerciasis⁷. Essential to the success of these programs are diagnostic tools for confirmation of disease, mapping, screening, decision of treatment administration, monitoring and surveillance. For both LF and onchocerciasis, the current diagnostic options are deemed inadequate to complete part or all of these tasks, particularly when it comes to post-treatment surveillance which is crucial to avoid disease resurgence. Diagnosis often still relies on identification by microscopy of microfilariae in blood thick-smear for LF⁸ and in skin snip biopsies for onchocerciasis⁹. Alternative and more convenient serological methods face several issues such as non-availability in field settings, the difficulty to distinguish past from current infections due to the presence of residual antibodies after parasite clearance, and cross-reactivity between filarial infections^{10–12}.

Detection of host antibodies to parasite antigens does however have many advantages as a diagnostic strategy since antibodies have been shown to appear earlier than detectable levels of circulating filarial antigen (CFA)¹³ and greater sensitivities have been indicated^{13,14}. Infected individuals are exposed to a variety of parasite molecules of diverse nature^{15–18} to which antibodies are raised, including antibodies to carbohydrates (glycans)^{19,20}. We recently reported a comprehensive characterization of *B. malayi* N-linked and glycosphingolipid (GSL) derived glycans. Using a glycan microarray workflow, we demonstrated that many glycan motifs expressed by the parasite were antigenic and that *B. malayi*-infected individuals showed a preferential immunoglobulin (Ig) G binding to a broad subset of glycans²¹. Importantly, IgG to these glycans decreased substantially after treatment with DEC

anthelmintic, suggesting that IgG to glycan antigens could be exploited as a marker of current or recent infection in the context of LF. Certain features of nematode glycans, such as the common presence of antigenic phosphorylcholine (PC) substituents are already known to be widespread throughout the filarial nematode phylum²². However, due to the overall lack of knowledge on filarial nematode glycosylation, for PC and many other glycan epitopes identified in our previous study, their potential cross-reactivity with host antibodies induced by other filarial infections has remained poorly understood. Studies into the role of IgG to parasite crude antigens have been well documented in the past both for LF^{15,23} and onchocerciasis²⁴. Notably, it is known that nematode parasites are typically characterized by T helper 2 (Th2) immunity inducing unusually high levels of IgG4 and IgE antibodies to the parasite in infected humans²⁵. Nonetheless, with the exception of *B. malayi*²¹, the humoral immune response to filarial carbohydrate antigens has remained largely unexplored.

Thus, in addition to the previously described anti-glycan IgG response from *B. malayi*-infected individuals, we studied IgG responses in plasma of individuals infected with *L. loa*, *M. perstans*, *O. volvulus* or *W. bancrofti* to address cross-reactivity with *B. malayi* glycans. Interestingly, substantial IgG binding to a broad range of glycan structures of *B. malayi* was also observed in plasmas from *M. perstans*- and *O. volvulus*-infected individuals. In comparison, plasma from *L. loa*- and *W. bancrofti*-infected individuals exhibited an overall weaker IgG response, that was restricted to a subset of GSL glycans. Further, evaluation of the antibody subclasses indicated that the reactivity to the glycan antigens across the most reactive infection groups (*B. malayi*, *M. perstans* and *O. volvulus*) was predominantly comprised of IgG1 and IgG2. Reduction of IgG1, IgG2 and IgG4 levels was observed after treatment with DEC. In particular, IgG2 decreased significantly for all individuals tested and to a majority of glycan structures, which may be exploited further for the development of a serology-based method for diagnosis of LF.

Materials and Methods

Infection plasma and ethics statement

***B. malayi* infection plasma**

Plasma from individuals infected with *B. malayi* were described in our previous glycan array study²¹. Plasma originated from two sets of five participants obtained from two distinct studies conducted in Indonesia in accordance with the Declaration of Helsinki and the guidelines of the Indonesian Department of Health and Human services. The purposes of the studies and procedures involved were explained to all participants, and only those granting informed consent were enrolled as study participants. The first

set of individuals was involved in previously published work^{26–28}, while the second set was from another study in which infected individuals were treated with DEC anthelmintic after initial sampling in 1990^{15,29}. Thus, for each individual, we had access to plasma sampled pre-treatment and almost two-years post-infection. Details on parasitological parameters regarding the infection status of the subjects (microfilaraemia) and filarial antigen specific IgG titers were obtained from the original studies where the experimental procedures have been described^{15,29}. Briefly, blood microfilaraemia was estimated by parasitological examination of 1mL of filtered (Nuclepore) venous blood. Blood collection took place at nighttime between 8PM and 11PM, in accordance with the periodicity of the microfilariae in the area. Microfilaraemia data are shown in **Table S1.A** for each individual. Note that for the second set, the microfilaraemia pre-treatment with DEC is indicated, since all individuals were amicrofilaraemic post-treatment. Filarial antigen specific IgG1-4 titers to *B. malayi* adult somatic extract antigen (BmA) were estimated by enzyme-linked immunosorbent assay (ELISA). ELISA protocols were performed as detailed earlier^{15,29} and used monoclonal anti-isotype antibodies to detect plasma IgG1-4 (**Table S1.A**).

Other filarial nematode infection plasma

Plasma from individuals infected with either *L. loa*, *M. perstans*, *O. volvulus* or *W. bancrofti* were all obtained from the same study (n = 6 for each infection). Plasma samples from Cameroonian patients were collected and archived as part of the Bill & Melinda Gates Foundation consortium project "Rapid and high throughput diagnosis of *Onchocerca volvulus* infections" (RADIO; OPP1083888; **Table S1.B**). Ethical clearance for the collection of blood samples for biomarker research was obtained from the National Ethics Committee for Human Health Research (Ref: N°2015/09/641/CE/CNERSH/S). In addition, administrative authorization was received from the Ministry of Public Health (Ref. No48/L/MINSANTE/ SG/DLMEP/ PNLO). All work was conducted in compliance with the Helsinki Declaration on the use of humans in biomedical research. Volunteers received detailed information about the study and how the samples would be biobanked and used for biomarker research to discover new biomarkers for their infections. All volunteers provided written informed consent before donating blood. Infected individuals were identified by palpation of onchocercomata (*O. volvulus*) and identification of microfilariae by microscopy of skin biopsies (*O. volvulus*) or blood (*L. loa*, *M. perstans*, *W. bancrofti*) as described^{11,30,31}.

Control plasma

Plasma from six non-infected African donors from Ghana collected in parallel to the Cameroon study were used as controls, in addition to plasma from five non-endemic European donors, that were obtained from Sanquin (Dutch blood bank).

Glycan microarray

Unless specified otherwise, reagents mentioned in the experimental section were obtained from Sigma-Aldrich.

Construction of a B. malayi N-linked and GSL glycan microarray

A glycan microarray of native parasite *N*-linked and GSL glycans was previously constructed and validated²¹. Briefly, glycans were isolated from adult female worm proteins and lipids using enzymatic treatment with *N*-Glycosidase (PNGase) F (#P0709, New England Biolabs (NEB)) and endoglycoceramidase I (EGCase I, cat. #P0773, NEB) respectively. Released glycans were labelled with 2-aminobenzoic acid (2-AA) and purified using a two-dimensional ultra-high performance liquid chromatography (UHPLC) procedure consisting of an initial separation using hydrophilic interaction chromatography followed by a reverse-phase purification on an octadecylsilane (C18) column. Glycan structures in UHPLC fractions were determined using matrix-assisted laser desorption/ionization-time of flight-mass spectrometry (MALDI-TOF-MS) analysis as detailed previously²¹ and summarized below. Fractions containing isolated individual - or small pools of mixed - glycan structures with a minimum glycan content of 20 pmol were selected for array printing. This amounted to a total of 35 *N*-glycan-containing fractions and 17 GSL glycan-containing fractions, largely covering the defined *N*-linked and GSL derived glycan repertoires of the parasite in terms of structural diversity. In addition, to evaluate the effect of PC substituents and fucose residues in antibody binding, aliquots of 4 GSL glycan fractions were treated with hydrofluoric acid (HF), a reagent removing α 1-2,3,4 linked fucoses and PC, leaving only unsubstituted GSL glycan backbones. HF treatment was performed using the conditions detailed below (see Glycan sequencing section) and fractions were cleaned-up by reverse-phase UHPLC before being added to the array selection, amounting to 21 GSL glycan fractions on the array. A detailed overview of the printed glycan fractions has been published as supplementary data (**Table S5**) in our previous study²¹.

Printing was performed as described previously^{32,33}. Briefly, fractions were aliquoted to a 384-well V-bottom polypropylene plate (cat. #784201, Greiner Bio-One) to create a glycan library plate containing 1, 3, 10 or 30 μ M printing solutions in 20 μ L of 1x spotting buffer (Nexterion Spot, cat. #1066029, Nexterion, Schott AG) with 10% DMSO. Additionally, 11 wells filled solely with spotting buffer were included for control of non-specific binding to the array. Samples and controls were printed in triplicate to epoxysilane-coated glass slides (Slide E, cat. #1066643, Nexterion, Schott AG), using the Microgrid 600 microarrayer (Genomic Solutions) equipped with SMP3 pins that deposit 0.7 nL upon each contact. Each array was printed eight times per glass slide

with a 0.245 mm spacing between spots and 4.60 mm spacing between the printing areas of each array.

Glycan array screening

Binding assay

The panel of control and infected human plasma samples described above was used to screen the glycan microarrays for anti-glycan antibody binding, following the procedures described previously for slide incubation, scanning and data handling^{32–35} with slight modifications where applicable. First, array blocking was performed with a solution of 2% BSA and 50 mM ethanolamine in PBS to cap residual epoxides on the slides. Next, plasma samples were diluted 1:100 in PBS, 0.01% Tween20 with 1% BSA and incubated on the array. Anti-glycan IgG from plasma was detected using goat anti-human IgG (Fc-specific) Cy3-conjugate (cat. #C-2571, Sigma-Aldrich) as secondary antibody. Specific binding of the different IgG subclasses was studied using mouse anti-human isotype antibodies purchased from SouthernBiotech. Alexa Fluor® 555-conjugates were used for detection of IgG1 and IgG2 (cat. #9052-32 and cat. #9070-32) and Alexa Fluor® 647-conjugates for IgG3 and IgG4 (cat. #9210-31 and cat. #9190-31). Secondary antibody used for the detection of total plasma IgG was diluted in PBS, 0.01% Tween20 with 1% BSA in a 1:1000 ratio while a 1:400 dilution was used for the isotype specific antibodies, with the exception of anti-human IgG4 where a 1:250 dilution was applied. Successive rinses were performed between each incubation step with PBS containing 0.05% Tween20 and with PBS. Finally, slides were washed with MilliQ water (MQ) prior to drying and scanning.

Scanning and data analysis

Fluorescence was detected using a G2565BA scanner (Agilent Technologies) at 10 µm resolution using two laser channels at 532 nm and 633 nm. Image analysis was processed with GenePix Pro 7.0 software (Molecular Devices). Spots were aligned and resized according to published methods³⁶. Next, data were exported to Excel, corrected for background using average fluorescence intensity of the control blank spots as a baseline, and intensities were averaged for each triplicate.

GLycan Array Dashboard (GLAD)³⁷, from the glycotoolkit website (glycotoolkit.com/Tools/GLAD/) hosted by the National Center for Functional Glycomics (NCFG), was used for data visualization, data mining, generation of heatmaps and boxplot graphs.

Statistical analysis

Statistical data analysis was performed under the publicly available statistical programming language R (<http://CRAN.R-project.org/>, version 3.5.0) using Bayesian statistics. Significant differences between log₂ normalized fluorescence intensities of

the various groups were assessed using the R/Bioconductor software package *limma*^{38,39} as described previously²¹.

Structural characterization of *O. volvulus* glycans

Enzymatic release of *N*-glycans and GSL glycans from *O. volvulus*

O. volvulus adult female worms that were pulverized in liquid N₂ were used for the extraction of *N*-linked and GSL glycans, following the procedure mentioned above for *B. malayi*. 2-AA-labeled *N*-linked and GSL glycan profiles were measured using MALDI-TOF-MS. Next, glycans were similarly purified using the two-dimensional UHPLC workflow. For structural elucidation, selected glycan samples – total *N*-glycans, *N*-glycan and GSL glycan-containing UHPLC-fractions – were subjected to glycan sequencing and analyzed by MALDI-TOF-MS analysis.

Glycan sequencing

A selection of glycan samples was subjected to selective degradation of glycans using treatment with HF or exoglycosidases to elucidate the main structures expressed by *O. volvulus*. The following enzymes were all obtained from NEB: α1-2,3,6 Mannosidase (cat. #P0768), β-N-Acetylglucosaminidase S (cat. #P0744), β-N-Acetylhexosaminidase_T (cat. #P0721), α1-3,6 Galactosidase (cat. #P0731) and α1-3,4,6 Galactosidase (cat. #P0747). Exoglycosidases were used alone or in combination to digest selected glycan fractions. Small aliquots of 2-AA-labeled glycans (typically 1-2 μL) were mixed with 1-2 μL of the exoglycosidase(s) in recommended buffer (GlycoBuffer 1 or 4, NEB). According to the manufacturer's instructions, the reaction was supplemented with BSA (0.1 μg/mL final concentration) when α-Galactosidases were used, and zinc (2 mM Zn²⁺ final concentration) was added to reactions with α1-2,3,6 Mannosidase. Final reactions were adjusted with MQ to 10 μL total volume and incubated overnight at 37°C. An undigested control consisting of all reagents except enzyme(s) was performed in parallel to each exoglycosidase digestion.

HF was used by mixing small aliquots of 2-AA-labeled glycans suspended in MQ with cold 48% HF in a 1:100 ratio. Samples were incubated at 4°C for 48h and HF was then removed by evaporation under nitrogen (N₂) flow. Multiple washes were performed by addition and subsequent N₂ evaporation of MeOH. Finally, the HF treated 2-AA-labeled glycan samples were re-dissolved in MQ.

After treatment with exoglycosidase(s) and HF, enzyme removal and clean-up was performed using C18 Millipore® Zip-Tips (cat. #Z720046-960EA) as described previously⁴⁰.

MALDI-TOF-MS and MALDI-TOF-MS/MS analysis

2-AA-labeled glycans from *O. volvulus* were analyzed using MALDI-TOF-MS as described previously²¹ using a Bruker rapifleX® instrument. Briefly, glycans were spotted onto a 384 well steel polished target plate. 2-AA-labeled glycans solubilized in MQ were mixed on the plate with 20 mg/ml 2,5-dihydroxybenzoic acid (DHB) matrix (cat. #8201346, Bruker Daltonics) in 30% ACN, while products of glycan sequencing (HF treatments, exoglycosidase digestions and corresponding undigested controls) were directly eluted onto the plate in 50% ACN, 0.1% TFA mixed with DHB (10 mg/ml) at the end of the enzyme removal with C18 Millipore® Zip-Tips. All spectra were obtained in negative-ion reflectron mode after external calibration with Bruker® peptide calibration mix (cat. #8206195, Bruker Daltonics). Spectra were obtained over a mass window of m/z 700 – 3500 with ion suppression below m/z 700. A minimum of 20,000 shots (2000 Hz) were obtained by manual selection of “sweet spots”. The software FlexAnalysis (Version 3.4, Build 50, Bruker Daltonics) was used for data processing including smoothing of the spectra (Savitzky Golay algorithm, peak width: m/z 0.06, 1 cycle), baseline subtraction (Tophat algorithm) and manual peak picking. Peaks with a signal-to-noise ratio below 5 were excluded as well as known non-glycan peaks such as glucose polymers. Glycan compositions correlating with the deprotonated masses of the selected peaks were assigned using the GlycoPeakfinder® (GlycoWorkBench, Version 3, 29 June 2007⁴¹). A deviation of 300 ppm was allowed and the 2-AA label was taken into account as a fixed reducing-end modification. Tandem MS (MS/MS) was performed for structural elucidation via fragmentation ion analysis by MALDI-TOF/TOF on selected ions using the rapifleX® mass spectrometer in negative-ion mode. Most probable glycan compositions were assigned based on knowledge from literature, analogy with *B. malayi* and results of MS/MS and glycan sequencing procedures.

Results

Cross-reactivity of anti-glycan IgG responses in human filarial infections

Comparison of total IgG responses to *B. malayi* glycans

Given that several glycan features found in *B. malayi* are shared with other filarial nematodes, e.g. presence of PC⁴² or terminal α -linked galactose (α -Gal) in GSLs⁴³, we evaluated the potential IgG cross-reactivity in plasma derived from other filarial nematode infections – *L. loa*, *M. perstans*, *O. volvulus* and *W. bancrofti* ($n = 6$ for each infection) – to *B. malayi* N-linked and GSL-derived glycans printed on the array. Results

were compared to those obtained for *B. malayi*-infected individuals (n = 10), Ghanaian individuals not infected with filarial nematodes (n = 6) and uninfected European donors (n = 5).

Substantial IgG binding to *B. malayi* glycans was detected in plasma from individuals infected with any of the other filarial nematodes when compared to plasma from uninfected individuals. In particular, for *O. volvulus*-infected individuals very high fluorescence levels were observed as glycan microarray read-out, and to a lesser extent for *M. perstans*-infected individuals (**Figure 1**). Comparatively, *L. loa* and *W. bancrofti* infection plasma showed lower IgG binding, with clear differences with non-endemic control plasma only apparent for a limited number of GSL containing fractions.

Statistical analysis using limma *F*-test for multiple-groups comparison confirmed these visual observations (**Table S2**). IgG binding was significantly higher to 37 glycan fractions for *B. malayi*-infected plasma, to 35 fractions for *O. volvulus* and to 24 fractions for *M. perstans* when compared to uninfected individuals. For the *L. loa*- and *W. bancrofti*-infected individuals, median fluorescence intensity (MFI) values significantly higher than both uninfected groups were only observed for 5 glycan fractions. Between the *B. malayi*, *M. perstans* and *O. volvulus* groups, significant differences in IgG binding were observed for only a limited number of fractions (5 glycan fractions for *B. malayi* vs *M. perstans*, 7 fractions for *B. malayi* vs *O. volvulus* and 4 fractions for *M. perstans* vs *O. volvulus*). This indicated highly similar anti-glycan IgG responses in these infections with a broad (cross-)reactivity to many glycan fractions and a similar absence of recognition of a restricted subset of glycans, clearly visible on the heatmap in **Figure 1**. Many differences were apparent when cross-comparing *B. malayi*-, *M. perstans*- or *O. volvulus*- to *L. loa*- or *W. bancrofti*-infected individuals, confirming a weaker IgG response from the two latter groups to the *B. malayi* glycans. Little IgG binding was observed for the two populations of uninfected individuals, with IgG binding in the Ghanaian control group only slightly but significantly higher than the European group for 8 fractions. The presence of IgG against *B. malayi* glycans in these uninfected Ghanaian individuals might be explained by possible previous infection with cross-reactive helminths or other pathogens, not endemic in Europe.

Identification of cross-reactive glycan motifs

The highest MFI values were observed for GSL glycan containing fractions. IgG binding was significantly higher ($p < 0.05$) for 16 out of the 21 printed GSL glycan fractions in the *B. malayi*-infected group than for both uninfected groups. Similarly, MFI values were significantly higher ($p < 0.05$) for 20 fractions in the *O. volvulus*-infected group and for 14 fractions in the *M. perstans* group compared to uninfected ones. This indicated that both *O. volvulus*- and *M. perstans*-infected individuals elicit IgG towards various glycan motifs present in *B. malayi* GSL glycans. Only a subset of the *N*-glycan

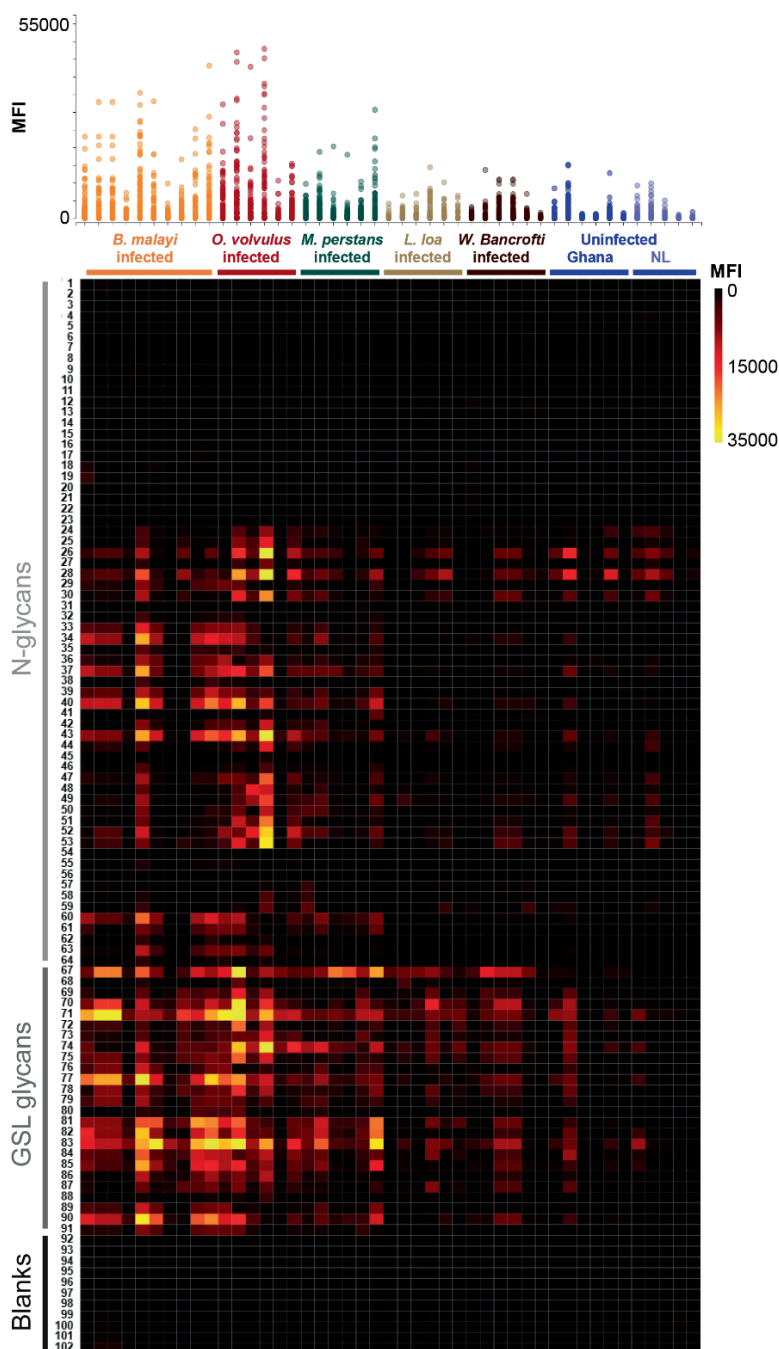


Figure 1. IgG responses to *B. malayi* N-linked and GSL glycans in plasma from various filarial-infected and uninfected individuals (caption next page)

Figure 1. IgG responses to *B. malayi* N-linked and GSL glycans in plasma from various filarial-infected and uninfected individuals

B. malayi glycan microarrays were screened with plasma from individuals infected with *B. malayi* (n = 10), *O. volvulus* (n = 6), *M. perstans* (n = 6), *L. loa* (n = 6), *W. bancrofti* (n = 6) and with plasma from uninfected individuals from Africa (Ghana, n = 6) and Europe (The Netherlands, NL, n = 5). Background corrected median fluorescence intensities (MFIs) resulting from IgG binding are shown on the upper graph for each donor (X-axis). Each spot on the Y-axis corresponds to a different glycan fraction or blank spot. The heatmap generated from these data allows the visualization of IgG from each donor (X-axis) to each array fraction (Y-axis) indicated by their ID numbers²¹. The type of glycan in the fractions is indicated on the left: N-glycans, GSL glycans or Blanks (negative controls, no glycan). Raw data can be found in **Table S2**.

structures printed on the array appeared to be antigenic and MFI levels were generally lower than observed for GSL glycans (**Figure 1** and **Figure S1**). To further identify the specific antigenic motifs targeted by IgG in the different filarial infection groups, we analyzed IgG binding in relation to the glycan structures printed on the array. Our previous work²¹ revealed that N-glycans of *B. malayi* contained terminal N-acetylglucosamine (GlcNAc), terminal glucuronic acid (GlcA) residues and mannosidic glycans (Man₂₋₉) as well as PC substituents (either GlcNAc or mannose-linked). GSL glycans contained terminal α-Gal, terminal N-acetylgalactosamine (GalNAc), terminal GlcNAc, terminal GlcA, fucosylated terminal α-Gal and fucosylated terminal N-acetylhexosamine (HexNAc, either GalNAc or GlcNAc). Fractions containing the listed glycan motifs were grouped into corresponding categories and MFI values of all fractions in a particular category were averaged for each individual.

As shown in **Figure 2A** and **Table S3**, *B. malayi*-infected individuals showed significantly higher MFI values ($p < 0.05$) to a limited subset of N-glycans, including high-mannosidic- (both Man5-7 and Man8-9) and GlcA-containing structures, than *L. loa*, *W. bancrofti* and both uninfected groups, for all of which no IgG binding to *B. malayi* derived N-glycans was apparent. Interestingly, while *M. perstans*-infected plasmas mimicked the reactivity seen in the *B. malayi*-infected ones, IgG binding in plasma from *O. volvulus* individuals yielded strikingly higher MFI values to all N-glycans assayed compared to the other infection groups, including the *B. malayi* group.

Notwithstanding these N-glycan associated observations, cross-reactivity of the IgG responses of *B. malayi*-, *O. volvulus*- and *M. perstans*-infected individuals was particularly apparent for the GSL fractions, with no significant differences between these three groups for all GSL glycan categories (**Figure 2B**, **Table S3**). For GlcA, terminal fucosylated α-Gal and fucosylated HexNAc-containing GSL glycans, all 3 infections gave significantly higher MFI values than *L. loa* and *W. bancrofti* groups which showed MFI levels comparable to the controls. This indicated that *B. malayi*, *O. volvulus* and *M. perstans* express these antigenic glycan motifs in the human host but that they may be absent in *L. loa* and *W. bancrofti* glycans. To the terminal α-Gal and terminal GlcNAc-containing GSL glycans however, significantly higher IgG reactivity was measured for all 5 infection groups, including *L. loa*- and *W. bancrofti*-infected

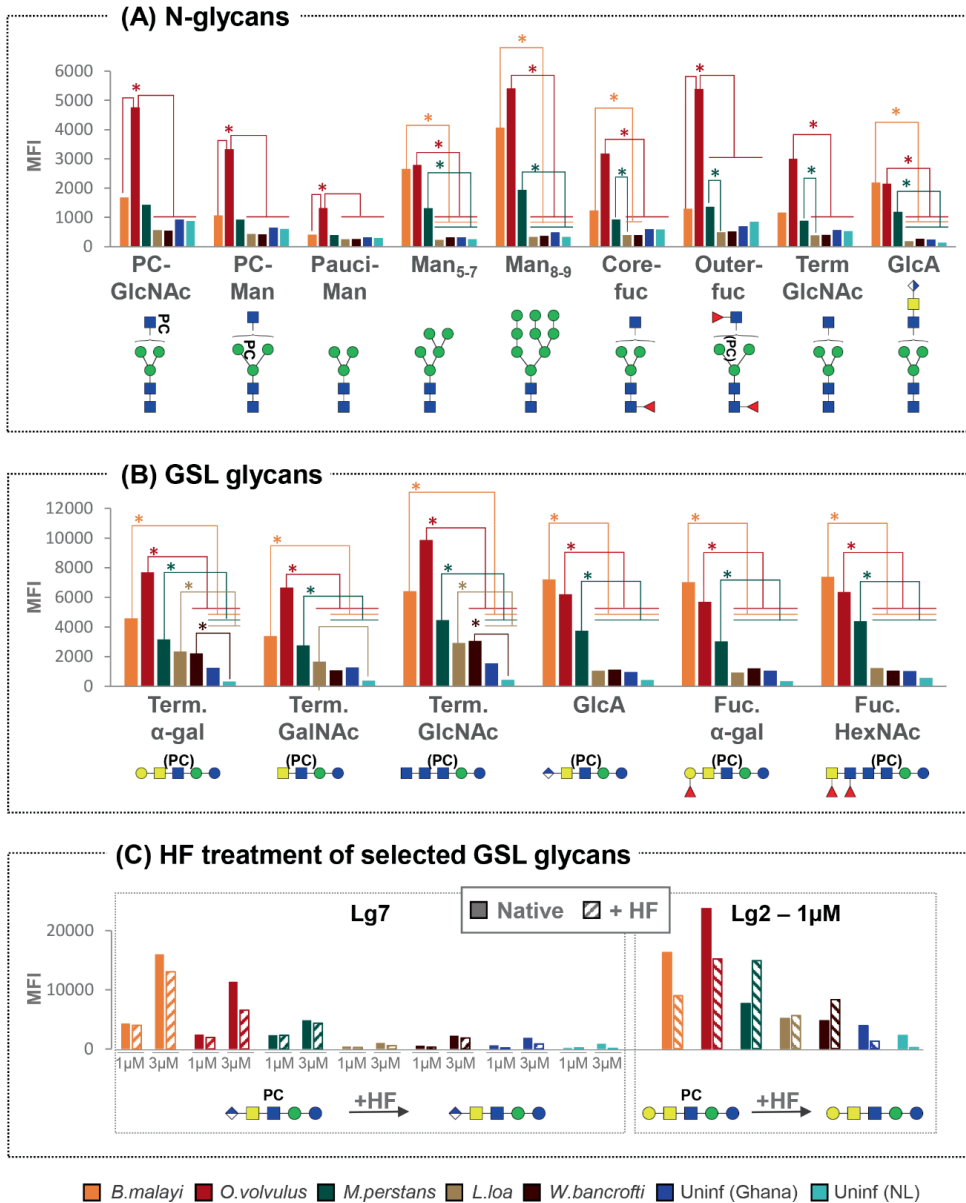


Figure 2. Cross-reactivity of IgG in various filarial infection and control plasmas to *B. malayi* N-linked and GSL glycan motifs

IgG binding to specific glycan motifs was determined for each individual by averaging MFI values of fractions belonging to the same category based on their glycan contents. Fraction groups are shown on the X-axis and named after the corresponding glycan motif present in the fractions: PC-GlcNAc = PC-substituted GlcNAc, PC-Man = PC-substituted mannose, Pauci-Man = Pauci-mannosidic N-glycans, Man_{x-y} = high-mannosidic N-glycans carrying 5 to 7 or 8 to 9 mannose residues, Core-fuc = α1-6 core fucosylated N-glycans, Out-fuc = α1-3 fucose attached to terminal GlcNAc, Term. GlcNAc = Terminal N-

Figure 2. (continued) acetylglucosamine, GlcA = terminal GlcA, Term. α -Gal = terminal α 1-4 Gal, Term. GalNAc = Terminal N-acetylglactosamine, Fuc. α -Gal = Fucosylated (terminal) α 1-4 Gal, Fuc-HexNAc = Fucosylated HexNAc(s). A representative glycan structure for each category is shown below the X-axis using the Consortium for Functional Glycomics (CFG) nomenclature (see symbol key inset in **Figure 3**).

MFI values obtained for each category were averaged for individuals from the same infection or control group: *B. malayi* (n = 10), *O. volvulus* (n = 6), *M. perstans* (n = 6), *L. loa* (n = 6), *W. bancrofti* (n = 6) infected individuals, uninfected (uninf.) Africans (Ghana, n = 6) and uninfected Europeans (The Netherlands, NL, n = 5). The upper graph shows results for N-glycan categories (A) and the middle graph for GSL glycan categories (B). Statistical differences between groups were assessed using Bayesian statistics and obtained *p*-values can be found in **Table S3**. Significant differences (with threshold of significance $\alpha = 0.05$) are represented on the graphs using asterisks and connecting lines.

The lower graphs (C) compare the averaged MFI values for selected native or HF treated GSL fractions, *i.e.* with or without PC substituents as illustrated below the X-axis, where structures contained in the corresponding fractions are represented.

individuals compared to the controls, indicating the widespread occurrence of these antigenic glycan motifs among the filarial nematodes (**Figure 2B**).

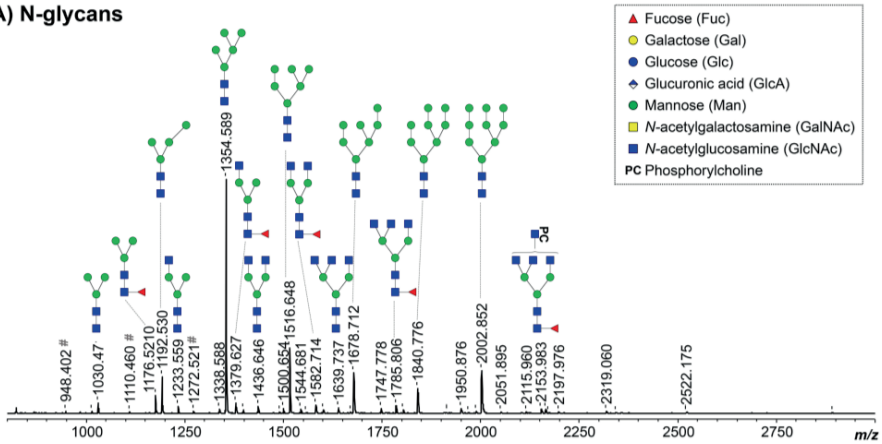
Since most of the *B. malayi* GSL glycans were substituted with PC, we treated certain fractions with HF to remove the PC substituent and evaluate its influence on IgG binding. As shown in **Figure 2C**, presence or absence of PC did not appear to strongly impact IgG binding to fraction Lg7 that contains a glycan of composition GlcA(β)-GalNAc(β 1-4)[PC-6]GlcNAc(β 1-3)Man(β 1-4)Glc1-R across all the groups. This suggests that the IgG reactivity is directed to a motif generated by the terminal GlcA of the GSL backbone structure. For the HF treated version of fraction Lg2 (putative structure Gal(α 1-4)GalNAc(β 1-4)[PC-6]GlcNAc(β 1-3)Man(β 1-4)Glc1-R) a slight decrease in IgG binding was observed for the *B. malayi* and *O. volvulus* infection plasma, and an increase for the *M. perstans* and *W. bancrofti* groups. However, none of these differential reactivities were statistically significant (**Figure 2C**), confirming that the PC substituent was of only minor relevance for the specific IgG binding to this structure.

Structural study of *O. volvulus* N-linked and GSL glycans

Since *O. volvulus* infection plasmas showed IgG binding to a large number of *B. malayi* glycans, we characterized the N-linked and GSL derived glycans from *O. volvulus* adult female worms using the glycomic workflow previously applied to *B. malayi* to determine whether the *O. volvulus* glycan repertoire would explain the observed cross-reactivity. Glycans were enzymatically released, fluorescently labeled with 2-AA and measured using MALDI-TOF-MS (**Figure 3**). MALDI-TOF-MS profiles of *O. volvulus* adult worms were strikingly similar to those obtained for *B. malayi* for both classes of glycoconjugates²¹.

As for *B. malayi*, mannosidic N-glycans were present in abundance in *O. volvulus*, as confirmed using digestion with α (1-2,3,6) Mannosidase (**Figure S2.A**). Comparative analysis by digestion with β -N-Acetylglucosaminidase S and β -N-

(A) N-glycans



(B) GSL glycans

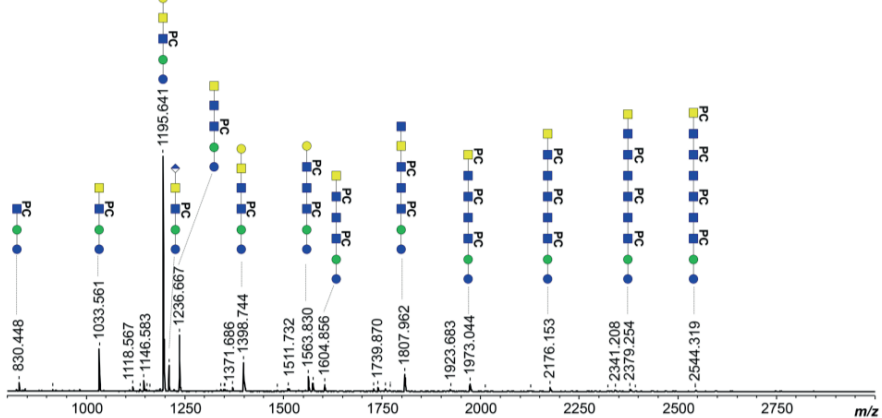


Figure 3. MALDI-TOF-MS of *O. volvulus* adult female worm N-linked and GSL glycans

O. volvulus glycoproteins were treated with PNGase F to release N-glycans (A) while EGCase I was used on glycolipids (B). Enzymatically released glycans were then labelled with 2-AA and analyzed using MALDI-TOF-MS in negative-ion reflectron mode. Monoisotopic masses of measured signals are indicated and proposed glycan structures for ions with signal-to-noise ratios superior to 5 are depicted using the CFG nomenclature (see symbol key insert). # symbols signal m/z of glucose polymers from unknown origin. Compositions and structures were deduced using a panel of glycan sequencing techniques in combination with MALDI-TOF-MS/MS fragmentation, analogy with *B. malayi* and information from literature. See **Table S4** for MALDI-TOF-MS spectra raw data and complete structural assignment.

Acetylhexosaminidase_r highlighted many structures extended with GlcNAc antennae and indicated the presence of β 1-2-linked GlcNAc residues (**Figure S2.B, E**). The infrequent presence of LacdiNAc (GalNAc β 1-4GlcNAc) terminal units, that are resistant to the β -N-Acetylglucosaminidase but sensitive to the β -N-Acetylhexosaminidase_r, was also observed (**Figure S2.B**). In addition, HF treatment

revealed PC-substitutions in abundance and outer-arm fucoses, most likely α 1-3 linked to GlcNAc, that were sensitive to the treatment, while many HF-resistant α 1-6 core-fucoses were also observed (**Figure S2.C, D**). Digestion with β -N-Acetylglucosaminidase S showed the presence of PC both on the antennae - substituting GlcNAc residues - as well as on the *N*-glycan core. The enzyme could digest a subset of structures up to a tri-mannosylated core of composition $H_3N_2F_1PC_1$ (m/z 1341.665 [$M - H$]⁻) containing both a core-fucose and a PC substituent (**Figure S2.E**). Most likely, this substituent would be linked to one of the terminal mannose residues, as seen in *B. malayi*. Finally, a feature identified in *B. malayi* was the presence of GlcA-containing *N*-glycans. However, we did not detect any in *O. volvulus* adult parasites.

In line with literature⁴³, the major GSL glycan structure in *O. volvulus* was the zwitterionic glycan of composition $H_3N_2PC_1$ (m/z 1195.641 [$M - H$]⁻). Using α 1-3,6 Galactosidase and α 1-3,4,6 Galactosidase, we determined the terminal hexose to be an α 1-4 linked galactose (**Figure 4.A-1** and **Figure S3.A**), as for *B. malayi*. HF treatment proved the presence of PC-substitution in the structure backbone (**Figure S3.B**) that can also be observed by MS/MS analysis of the ion with m/z 1195.4287 (**Figure 4.A-2**). The HF-sensitive fucosylated version of this glycan (m/z 1341.569 [$M - H$]⁻) was also detected in *O. volvulus* (**Figure S3.C**). Interestingly, we noted several structures of composition $H_3N_3PC_{1-3}$, with an additional HexNAc and one to three PC substituent(s) (m/z 1398.744; 1563.830 and 1728.619 [$M - H$]⁻). MS/MS analysis of ion with m/z 1398.744 (**Figure 4.B-1**) and resistance to digestion with both β -N-Acetylhexosaminidase_r and β -N-Acetylglucosaminidase S suggested the presence of the terminal α -Gal epitope in those structures as well (**Figure S3.D, G**). The presence of hexuronic acid-containing GSL glycans was detected in *O. volvulus*, as shown by MS/MS analysis of the ion with m/z 1209.408 [$M - H$]⁻ (**Figure 4.B-2**). As for *B. malayi*, this residue was found to occupy the terminal position on the glycan chain and by analogy, is expected to be β -linked GlcA. Interestingly, structures of compositions $H_3N_{2-3}A_1PC_{1-2}$ containing both α -Gal and GlcA residues, that were not detected in *B. malayi*, were found in *O. volvulus* (m/z 1371.686; 1574.798; 1739.596 [$M - H$]⁻). As shown by MS/MS analysis and sensitivity to HF treatment, these structures contained PC substituent(s) (**Figure 4.B** and **Figure S3.D, F**). Masses corresponding to structures of similar backbone compositions but with a fucose residue ($H_3N_{2-4}A_1F_1$) were also observed (m/z 1352.463 and 1758.622 [$M - H$]⁻) (**Table S4**). When analyzing the m/z 1371.461 [$M - H$]⁻ ion species using MS/MS, both a loss of a hexose and of a hexuronic acid were observed from the parent ion suggesting a possible branching of those residues at the terminal position (**Figure 4.B-3**). Other terminal residues included GlcNAc and GalNAc which were discriminated by comparing sensitivity to digestions with β -N-Acetylglucosaminidase S and β -N-Acetylhexosaminidase_r (**Figure S3.D, E, G, H**).

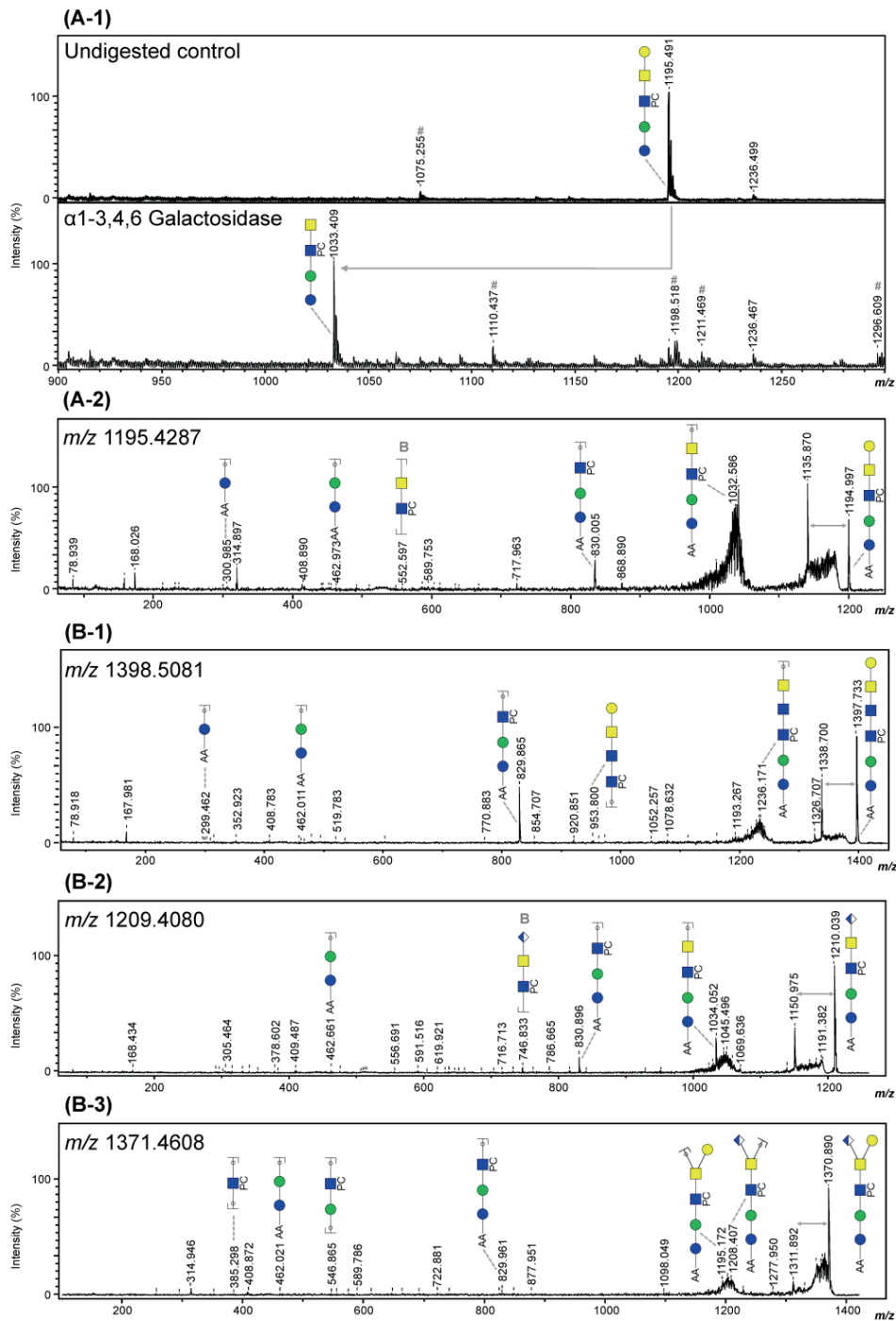


Figure 4. Evidence of terminal α -Gal and GlcA-containing epitopes in the GSL glycans of *O. volvulus* (caption next page)

Figure 4. Evidence of terminal α -Gal and GlcA-containing epitopes in the GSL glycans of *O. volvulus* MALDI-TOF-MS analysis of 2-AA-labeled *O. volvulus* major GSL glycan species subjected to digestion with α 1-3,4,6 galactosidase (A-1) Digestion product showing the loss of a terminal α -Gal ($\Delta m/z = 162$) is highlighted using a grey arrow.

Negative ion reflectron mode MALDI-TOF-MS/MS of ions with theoretical m/z 1195.4287 (A-2), m/z 1398.5081 (B-1), m/z 1209.4080 (B-2), and m/z 1371.4608 (B-3). Resulting spectra are labeled with graphic representation of Y-type ions, unless indicated otherwise (B = B-type). Loss of mass 59 Da from the parent ion is indicative of a PC loss^{62,63} and is highlighted by grey double arrows.

For both MALDI-TOF-MS/MS and MALDI-TOF-MS spectra monoisotopic masses of 2-AA-labeled glycans are indicated and glycans are represented using the CFG nomenclature (see symbol key insert in **Figure 3**).

Despite some notable differences between species, overall, the structural analysis of the *O. volvulus* N-linked and GSL glycans confirmed the presence of many shared epitopes with *B. malayi*. A detailed list of the N-glycan and GSL glycan structures identified in *O. volvulus* is provided in **Table S4**. Thus, the observed cross-reactivity of IgG from *O. volvulus*-infected plasma to *B. malayi* glycans could be attributed to the overlapping similarity of glycan structures present in *O. volvulus*, and supports the conclusion that (partial) glycome overlaps with the other species also occur.

IgG subclass responses to *B. malayi* glycan antigens in filarial nematode infections

In view of the particularly strong cross-reactivity observed between *B. malayi*, *O. volvulus* and *M. perstans* for total IgG and the general lack of data with respect to IgG subclasses and reactivity to helminth glycans, we next investigated the specific IgG subclass responses in these infection groups.

Study of plasma IgG subclasses in *B. malayi*-infected individuals

Screening of the glycan microarrays with the *B. malayi* infection plasma yielded strong fluorescence signals for IgG1 and IgG2 subclasses (**Figure 5**). A similar pattern of glycan recognition to that seen for total IgG was observed, with a major reactivity toward the GSL glycans. Comparatively, lower MFI values were obtained for IgG3 and very low signals were observed for IgG4 (**Figure 5**, **Table S5** and **Figure S4**) indicating that IgG to *B. malayi* antigenic N-linked and GSL glycans is mainly of the IgG1 and IgG2 subclasses.

During previous studies^{15,29}, IgG subclasses to *B. malayi* crude antigen (BmA) were measured by ELISA. Those titers of IgG1-4 to BmA (in $\mu\text{g/mL}$) are indicated in **Figure 5** below the boxplots showing IgG subclass binding to the *B. malayi* glycan fractions for each individual. When comparing the ELISA data to the glycan microarray results, we noticed a clear correlation between the array MFI values and the BmA-

specific IgG titers, with individuals having higher titers of IgG1-4 to BmA (in $\mu\text{g/mL}$) also showing higher binding to the glycan microarray.

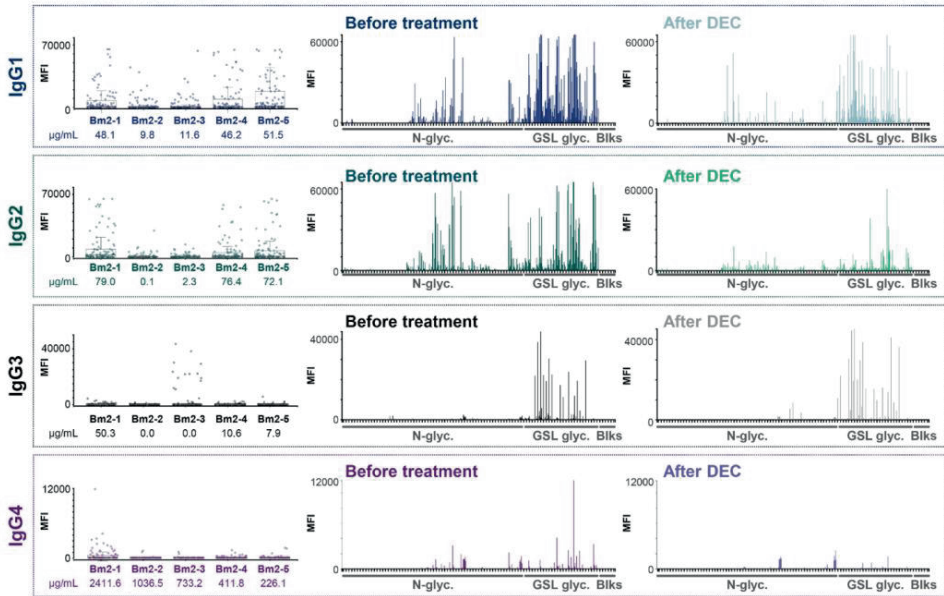


Figure 5. IgG subclass responses to *B. malayi* glycans in plasmas from infected individuals before and after DEC anthelmintic treatment

B. malayi glycan microarrays were screened for binding of IgG subclasses in plasma of *B. malayi*-infected individuals during infection and after treatment with DEC anthelmintic when individuals were all amicrofilaraemic. Subclass specific fluorescent secondary antibodies were used for detection of IgG subclass binding. Obtained background corrected MFI values are shown on the Y-axis of boxplots and graphs.

Boxplots on the left show IgG subclass binding for all 5 individuals (ID numbers displayed on the X-axis, see **Table S1.A**) to each glycan fraction on the array ($n = 100$ with various printing concentrations and including buffer spots). BmA-specific IgG1-4 titers from the original study^{15,29} are depicted in $\mu\text{g/mL}$ for each individual below their corresponding ID number underneath the boxplots.

Graphs show overall IgG binding to the various glycan array fractions (X-axis, $n = 100$) for plasma from all 5 individuals either during infection (Before treatment, left) or after DEC treatment (After DEC, right).

We previously reported that total IgG response to *B. malayi* glycan antigens dropped markedly after DEC treatment²¹. Focusing on IgG subclass responses in the same sample set of plasma ($n = 5$, second set, see Materials and Methods), a decrease in MFI levels on the glycan microarray after DEC treatment was clearly visible for IgG2 as well as for IgG1 and IgG4 – although less pronounced – but no changes were apparent for IgG3 (**Figure 5** and **Figure S4**). Using Bayesian statistics for paired samples (a moderated paired *t*-test allowing for sib-pair effects in the linear model), we determined that differences pre- and post-treatment were only significant for 4 fractions for IgG1 binding while IgG2 binding to 19 fractions was significantly decreased

post-DEC treatment. Differences in binding to specific glycan fractions of IgG3 and IgG4 between the two time-points were not significant (**Table S5**). MFI values for IgG1 towards parasite glycans decreased for 4 of the 5 individuals tested, whereas IgG2 MFI values were clearly lower for all of them (**Figure S4**). In an earlier study, a decrease of IgG1 after DEC treatment was observed also to crude BmA, however with a substantial individual-to-individual variability²⁹. The consistent and clear reduction of IgG2 to *B. malayi* glycans post-treatment was however not reflected by IgG2 to BmA²⁹, indicating that specific *B. malayi* glycans form an antigen subset with differential characteristics.

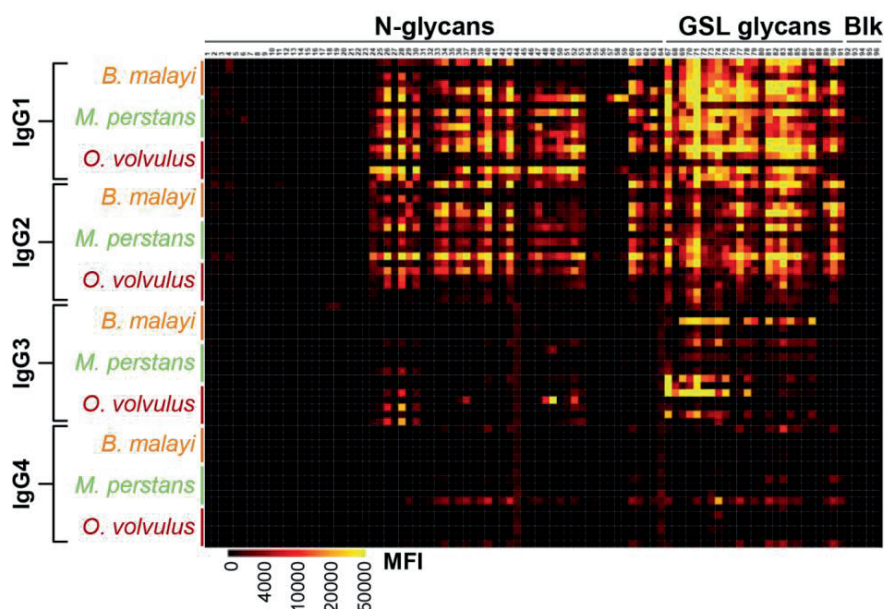


Figure 6. IgG subclass responses in plasma from *B. malayi*, *O. volvulus* and *M. perstans* infected individuals to *B. malayi* glycans

B. malayi glycan microarrays were screened for binding of IgG subclasses with plasma of *B. malayi* (n = 5), *O. volvulus* (n = 6) and *M. perstans* (n = 6) infected individuals. Subclass specific fluorescent secondary antibodies were used for detection of IgG subclass binding. Heatmap was generated from obtained background corrected MFIs (see intensity key at the bottom) and shows binding of all IgG subclasses for each infected individual as indicated on the Y-axis. X-axis displays the various glycan fractions printed on the array as detailed in²¹. Type of fraction content (N-glycan and GSL glycan) or control (Blk) is indicated above the fraction IDs.

Cross-comparison of plasma IgG subclass responses from filaria-infected individuals and uninfected controls

Next, we examined *B. malayi* glycan reactive IgG subclasses in the highly cross-reactive *O. volvulus* and *M. perstans* infection plasmas. As for *B. malayi*, highest MFI

signals were obtained for IgG1 and IgG2 while IgG3 and IgG4 binding to the array yielded lower MFI values (**Figure 6** and **Figures S5-6**). Statistical analysis confirmed the similarity of the IgG subclass responses to the glycans between the three infections (**Table S5**). In each case, MFI values were significantly higher than in uninfected individuals to a large proportion of the glycans on the array. This was particularly clear for IgG1 that was detected to virtually all GSL glycans and to many *N*-glycans in *O. volvulus* and *M. perstans* infection plasmas. Similar observations were made for IgG2, however with the *O. volvulus*-infected group showing lower MFI values for IgG2 than for IgG1, yielding less significant differences with the control plasmas than the *B. malayi*- and *M. perstans*-infected groups (**Table S5**).

MFI values resulting from IgG3 binding to the microarray were lower than the ones observed for IgG1 and IgG2 for all three infection groups but were still significantly higher than the uninfected groups for several GSL glycan fractions. IgG3 to *N*-glycans however, was very limited in *B. malayi* and *M. perstans* infection plasmas but was detected in *O. volvulus* infection plasmas. Study of IgG subclasses to specific glycan motifs (**Figure 7**) reflected this observation, with *O. volvulus* infection plasma showing significantly higher MFI values than uninfected individuals and *B. malayi*-infected individuals for several *N*-glycan categories (**Table S6**). Interestingly, 2 individuals in each uninfected group showed relatively high levels of plasma IgG3 to GSL glycans (**Figure S7**). This resulted in the absence of significant differences between infected and uninfected individuals when averaging MFI for the various GSL glycan categories, although some statistical differences were observed for IgG3 binding to individual fractions (**Table S5**).

Although markedly weaker, IgG4 was detected to a few fractions. When compared with uninfected groups, IgG4 to 8 and 11 glycan fractions was significantly higher for the *B. malayi* and *M. perstans* groups, respectively, but only 2 fractions gave significantly higher MFI values for the *O. volvulus* group (**Table S5**). Thus, mirroring observations made for *B. malayi* infections, glycans appear not to be the primary targets of IgG4 in *M. perstans* and *O. volvulus* infections either, despite elevated plasma IgG4 titers⁴⁴. Nonetheless, MFI values averaged to assess IgG4 to specific glycan motifs were still significantly higher for the infected groups than the ones measured for the uninfected groups for certain glycan categories (**Figure 7** and **Table S6**).

Overall, the analysis focusing on IgG subclasses to specific glycan motifs (**Figure 7**) confirmed and summarized the similarities observed between the three infections, with few statistical differences between the three groups (**Table S6**). IgG1 was comparable for all, with significantly higher MFI values obtained for each GSL glycan category when compared to uninfected controls. Similarly, the IgG2 response in *B. malayi*- and *M. perstans*-infected individuals was significantly higher to all GSL glycans than in the uninfected groups. However, as mentioned above, IgG2 binding from the *O.*

volvulus-infected individuals yielded lower MFI values that were only significantly higher than the uninfected controls for the α -Gal and terminal GlcNAc-containing GSL subsets (Figure 7).

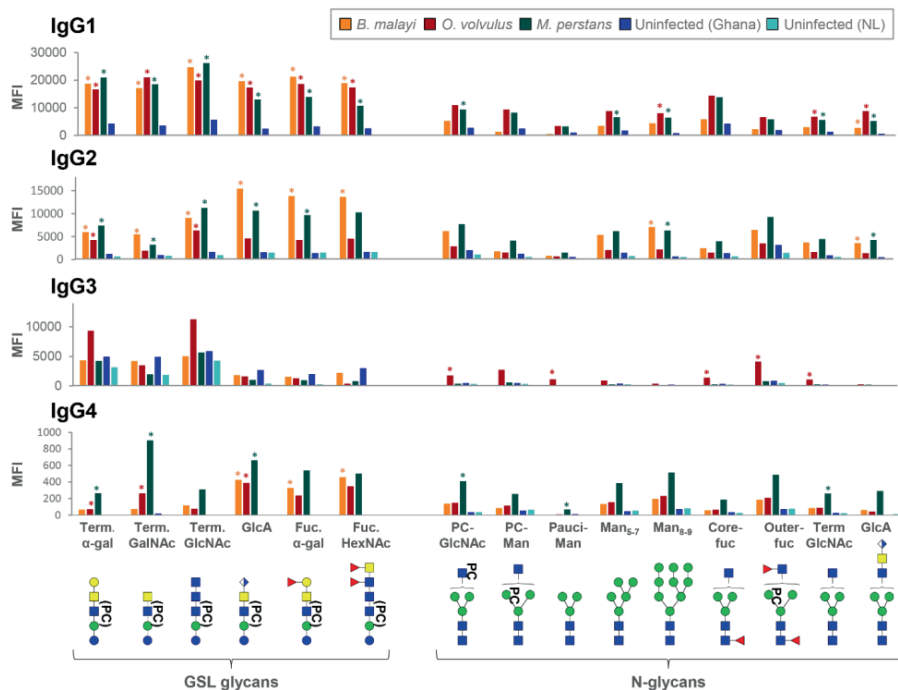


Figure 7. Binding of IgG subclasses from *B. malayi*, *O. volvulus* and *M. perstans* infection and control plasmas to *B. malayi* N-linked and GSL glycan motifs

IgG subclass binding to specific glycan motifs was calculated for each individual by averaging MFIs of fractions belonging to the same category based on their glycan content. Fraction groups are shown on the X-axis and named after the corresponding glycan motif present in the fractions: PC-GlcNAc = PC-substituted GlcNAc, PC-Man = PC-substituted mannose, Pauci-Man = Pauci-mannosidic N-glycans, Man x-y = High-mannosidic N-glycans carrying 5 to 7 or 8 to 9 mannose residues, Core-fuc = α 1-6 core fucosylated N-glycans, Out-fuc = α 1-3 fucose attached to terminal GlcNAc, Term. GlcNAc = Terminal N-acetylglucosamine, GlcA = terminal GlcA, Term. α -Gal = terminal α 1-4 Gal, Term. GalNAc = Terminal N-acetylgalactosamine, Fuc. α -Gal = Fucosylated (terminal) α 1-4 Gal, Fuc-HexNAc = Fucosylated HexNAc(s). A representative glycan structure for each category is shown below the X-axis using the CFG nomenclature (see symbol key inset in Figure 3).

Category MFIs thus obtained were averaged for individuals from the same infection or control group: *B. malayi*-infected individuals (before DEC treatment, n = 5), *O. volvulus*-infected individuals (n = 6), *M. perstans*-infected individuals (n = 6), uninfected (uninf.) Africans (Ghana, n = 6) and uninfected Europeans (The Netherlands, NL, n = 5). Significant differences between groups were assessed using Bayesian statistics. MFIs significantly higher for infection plasma IgG than for both uninfected groups are indicated with asterisks. Obtained p-values can be found in Table S6.

Discussion

We have shown previously that the filarial nematode *B. malayi* expresses a variety of *N*-glycans and highly antigenic GSL glycans, to which individuals infected with the parasite develop IgG²¹. Here, we showed that individuals infected with *O. volvulus* and *M. perstans* also produce IgG to antigenic motifs contained in a broad range of *B. malayi* glycans, while *W. bancrofti*- and *L. loa*-infected individuals appeared to produce IgG reactive with only a restricted subset of the *B. malayi* GSL-derived glycans. This suggests that *B. malayi*, *O. volvulus* and *M. perstans* express highly similar *N*-linked and GSL glycans that are not present in *L. loa* and *W. bancrofti*.

Structural characterization of the *N*-linked and GSL glycans of *O. volvulus* adult worms supported this hypothesis. Many (antigenic) glycan elements reported for *B. malayi* were identified in *O. volvulus*, including terminal α -Gal and GlcA in the GSL glycans (**Figure 4**). We also noted the presence of an epitope containing both α -Gal and GlcA in several GSL glycans of *O. volvulus* that was not detected in *B. malayi* (**Figure 4.B**). The glycan microarray used in this study contained exclusively glycans isolated from *B. malayi*, but it would be of interest to study antibody binding to this combined α -Gal and GlcA motif, which may perhaps represent an *O. volvulus* specific glycan epitope. *N*-glycans from both species were very similar, with the presence of mannosidic structures and structures extended with GlcNAc antennae that were fucosylated and/or substituted with PC. These findings were overall in line with previous work conducted on the *N*-glycans of *O. volvulus*⁴⁵. There too, structures of composition $H_3N_{3-6}F_{0-1}PC_1$ were detected, as well as the presence of both GlcNAc and, to a lesser extent, GalNAc residues within the antennae. We did not detect GlcA as part of *O. volvulus* *N*-glycans, while we clearly observed IgG to the *B. malayi*-derived GlcA containing *N*-glycans on the microarray in *O. volvulus* infection plasmas. This suggests that the GlcA-containing epitope present in the *O. volvulus* GSL glycans - and possibly in other classes of glyconjugates not studied here - induce the IgG that also binds to an identical or similar GlcA-containing epitope in the context of *N*-glycans. Specifically, we have found the GlcA residue to be present as a terminal residue on the LacdiNAc (*i.e.* GalNAc β 1-4GlcNAc) motif in the *B. malayi* *N*-linked and GSL glycans²¹.

In that regard, glycan microarray screening is highly informative in indicating exposure to, and therefore presence of, a specific glycan motif during parasitic or bacterial infections. This is particularly useful when structural glycomic studies of a specific pathogen have not been conducted yet or are not possible due to unavailability of the source materials. Here, performing a comprehensive glycomic study and native glycan release from two species, *B. malayi* and *O. volvulus*, indirectly allowed us to gather information on the glycomes of three other related nematodes. We indeed detected the presence of IgG to GSL glycans containing terminal α -Gal and stretches of GlcNAc residues in the plasma of all infection groups (**Figure 2**) allowing us to infer

the expression of these types of structures by all species of nematodes, although they might be present in diverse glycoconjugate classes. The presence of IgG to terminal α -Gal, in particular, is consistent with indications of the zwitterionic glycolipid of composition Gal(α 1-4)GalNAc(β 1-4)[PC-6]GlcNAc(β 1-3)Man(β 1-4)Glc(1-1)ceramide being highly conserved in the nematode phylum⁴³. On the other hand, of the species examined, it is likely that only *B. malayi*, *O. volvulus* and *M. perstans* express other motifs, based on terminal GlcA or fucosylated α -Gal, since IgG to these types of glycans were only measured in plasma from these infections. We cannot completely exclude that *L. loa* and *W. bancrofti* also harbor these glycan epitopes but, if so, they appear to either express them in a much lower amount or in a less antigenic context or in non-mammalian life stages. In that regard, it is also interesting to note the high IgG MFI levels observed with *O. volvulus* infection plasma to virtually all *B. malayi* *N*-glycans while *B. malayi*-infected individuals only showed high MFI levels to a restricted subset of *N*-glycans (**Figure 2**). Perhaps higher titers of IgG to *N*-glycans are elicited in onchocerciasis either as a consequence of more abundant excretion/secretion of antigenic *N*-glycosylated proteins or of a more efficient and immunogenic presentation on different protein carriers.

Comparative glycomic studies are useful alternatives when structural glycomic studies are not possible or required. It would be complementary to our work to study whether certain glycotransferase genes and activities are absent from *L. loa* and *W. bancrofti* compared to the other filarial species. However, despite recent improvements for several species of filarial nematodes⁴⁶, the genome quality of the five species of filarial nematodes included in this study is uneven. In addition, the insufficiency of functional genomic annotations and the absence of well-characterized glycosylation pathways for non-mammalian organisms, particularly where they diverge from human or mouse pathways, make this type of study challenging.

A well-known shared feature of filarial nematodes is the presence of PC substituents, most of the time present in a phosphodiester linkage with C6 of the GlcNAc residues of *N*-linked and GSL glycans⁴². Immunomodulatory properties of PC-containing epitopes have been extensively studied over the years^{47,48}, particularly in relation to the ES-62 glycoprotein of *Acanthocheilonema viteae*^{49,50}. Notably, PC-containing glycoproteins are thought to contribute to low antibody levels and poor lymphocyte responses observed in some filariasis patients⁵¹. In our study, HF treatment of selected GSL containing fractions showed that highly antigenic elements were still present after removal of PC and that these are major IgG targets. Thus, the contribution of PC substituents to IgG binding to glycans was not dominant. This is consistent with previous observations in both brugian²¹ and bancroftian filariasis¹⁹ and it appeared that PC substituents do not elicit specific IgG in the five filarial infections studied here, although they might have influenced the overall antibody levels. The presence of PC is known to generate “background” cross-reactivity of antibodies to

parasite antigens, both in filarial nematode infections but also with non-endemic serum since PC is present in many bacteria and fungi^{44,52}. In our study, averaged MFI values obtained for the control groups to PC-containing *N*-glycans (both to PC-substituted GlcNAc and PC-substituted mannose categories) indeed tend to be higher than for the other *N*-glycan categories (**Figure 2**). IgG binding to PC-substituted glycans from individuals not infected with filarial nematodes, is likely to be a consequence of exposure to PC from various other pathogens⁵³. In the past, cross-reactivity attributed to PC and other determinants has been deemed responsible for hampering the development of serological diagnosis for filariasis⁴⁴. However, in our work, the background possibly caused by these shared epitopes was negligible when compared to the strong signals generated by specific IgG recognizing other, potentially more specific and antigenic glycans.

Another consequence of glycan cross-reactivity issues, has been the increased interest towards the IgG4 subclass for serological diagnostic applications since IgG4 was known to recognize restricted subsets of antigens⁴⁴ of non-glycan nature⁵⁴ compared to the other IgG subclasses. Despite the unusually high IgG4 titers to filarial antigens typical of the immune response in LF^{15,25}, we observed very little or no IgG4 binding to glycans in infected plasmas (**Figure 5**), indicating that IgG4 response in LF would mainly target non-carbohydrate epitopes in BmA, presumably protein components. This is consistent with previous work where humans were found to be “genetically restricted” from making IgG4 to PC, polysaccharides and streptococcal carbohydrate antigens^{54,55}, although *N*-glycans and GSL-glycans may have different molecular properties with respect to generating an immune response in the host. The low-intensity IgG4 signals measured in our study were highly variable between individuals (**Figure S4**) but were still significantly higher in *B. malayi*-infected individuals than in uninfected controls to a few glycans such as GlcA-containing fractions (**Table S5**). In *Schistosoma mansoni*-infected individuals³⁵ IgG4 binding has also been found to be very restricted to specific glycans, in that case, those containing α 1-3 core-fucoses and core-xylose. There too, variability between individuals was substantial, which, in our study, is reflected in the low significance levels observed between the different groups (**Figure 7** and **Table S6**).

In contrast to IgG4, a strong IgG1 and IgG2 binding to our glycan microarrays was measured in the filarial infection plasmas when compared to uninfected controls (**Figure 6**). IgG2 is known to play a role in responses to carbohydrate antigens in the context of bacterial and parasitic infections^{19,56}, although it is now clear that anti-glycan responses are not restricted to this subclass^{35,57}. Plasma IgG subclasses differ highly from each other in terms of antigen specificity and functional properties⁵⁸. Their distribution also varies as a function of age with the IgG1 and IgG3 isotype levels reaching adult levels in sera sooner than IgG2 and IgG4 subclasses^{59,60}, which is an important consideration to take into account for diagnosis of children. Nonetheless, in

other respects, IgG2 to glycan antigens in plasmas from *B. malayi*-infected individuals, and particularly to GSL glycans, showed promising features for potential diagnostic applications. High MFI values were observed in infected plasma, that were significantly decreased in the plasma sampled post-DEC treatment, less than 2 years later. Moreover, IgG2 to the *B. malayi* glycans was also detected in the plasma of *M. perstans*-infected individuals but to a lesser extent in the plasma of *O. volvulus*-infected individuals (**Figure 7**), suggesting that IgG2 might allow a gain in species-specificity compared to the other subclasses. Additionally, cross-reactivity between these filarial nematodes might not be a major concern since *B. malayi* is not co-endemic with *O. volvulus*⁴ or *M. perstans*⁶¹.

An important consideration in LF is the association of specific IgG subclasses with the various subpopulations in endemic areas where asymptomatic microfilariaemic carriers have higher ratios of IgG4 to IgE while individuals with chronic disease have elevated IgG1-3¹⁵. Further research should aim to address how IgG subclasses to filarial glycan antigens are distributed within the endemic subpopulations, since it is a crucial point to evaluate for considering a further diagnostic potential of our findings. Moreover, it would be of great interest to determine if symptomatic or putatively protected individuals exhibit differences in anti-glycan antibodies, which could relate to protection or be involved in pathology. Another aspect to consider, is whether the presence of circulating microfilariae is necessary for development of anti-glycan IgG. In our previous study, we demonstrated comparable profiles for the *N*-glycans and GSL glycans from *B. malayi* human stages - infective third-stage larvae, adult worms (males and females) and microfilariae²¹. Thus, it is not known from which parasite life-stage(s) the host immune system gets exposed to glycan antigens. In *B. malayi*-infected rhesus macaques, IgG to glycan epitopes is apparent five weeks post-infection, prior to microfilarial production²¹. Whether the same happens in humans and whether the mere presence of adult worms, as occurs in amicrofilariaemic infections, is enough to drive and maintain the anti-glycan antibody response was not addressed in this study, since all individuals were microfilariaemic (**Table S1**).

In conclusion, research on anti-glycan antibodies in the context of parasitic infections is still limited, particularly at the subclass level. With this study, we investigated the plasma IgG response to *B. malayi* glycans from individuals infected with filarial nematodes. We addressed cross-reactivity and subclass specificity of the anti-glycan IgG responses for five major filarial nematode infections. Many crucial aspects should be further investigated on the basis of this work, such as possible differences in IgG responses between different subpopulations in endemic areas, *i.e.* chronic and asymptomatic infections. Also, it will be important to validate our observations on a larger cohort, including individuals from diverse geographic areas.

Nonetheless, our data indicates that IgG responses to glycan antigens in filariasis might offer unexplored and valuable alternatives for use in diagnostic applications.

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Data availability

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found below: GPST000299 (<https://glycopost.glycosmos.org/>), and in Github (<https://github.com/lmcpetralia/Front-Immunol-doi-10.3389-fimmu.2023.1102344.git/>).

Ethics statement

The studies involving human participants were reviewed and approved by the Indonesian Department of Health and Human Services and the National Ethics Committee for Human Health Research. The patients/participants provided their written informed consent to participate in this study.

Supplementary data

This article contains supplementary data. All supplementary material can be found online with the published article ([10.3389/fimmu.2023.1102344](https://doi.org/10.3389/fimmu.2023.1102344)). A selection of supplementary tables is appended at the end of this chapter.

Author contributions

LMCP: Conceptualization, Investigation, Formal analysis, Writing – Original draft, Reviewing & Editing, Visualization; **AvD:** Conceptualization, Writing – Reviewing & Editing, Supervision; **DLN, LAL:** Investigation; **ES, SB, TBN, KP, AH, SW:** Resources; **JMF, CHH:** Conceptualization, Writing – Reviewing & Editing, Supervision, Funding acquisition. All authors contributed to revision of the article and approved the submitted version.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Abbreviations

α-Gal	α-linked galactose
2-AA	2-aminobenzoic acid
BmA	<i>B. malayi</i> crude antigen
C18	Octadecylsilane
CFA	Circulating filarial antigen
DEC	Diethylcarbamazine citrate
DHB	2,5-dihydroxybenzoic acid
EGCase I	Endoglycoceramidase I
ELISA	Enzyme-linked immunosorbent assay
GalNAc	N-acetylgalactosamine
GlcA	Glucuronic acid
GlcNAc	N-acetylglucosamine
GSL	Glycosphingolipid
HexNAc	N-acetylhexosamine
HF	Hydrofluoric acid
Ig	Immunoglobulin
LF	Lymphatic filariasis
MALDI-TOF-MS	Matrix-assisted laser desorption/ionization-Time of flight-MS
Man₂₋₉	Mannosidic glycans
MDA	Mass drug administration
MFI	Median Fluorescence Intensity
MQ	MilliQ water
MS	Mass spectrometry
MS/MS	Tandem mass spectrometry
PC	Phosphorylcholine
PNGase F	N-Glycosidase F
Th2	T helper 2
UHPLC	Ultra high performance liquid chromatography
WHO	World Health Organization

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Supplementary material

Supplementary Tables

Table S1: Information on infection and control sera

A] Information on *B. malayi*-infected individuals

Set #	Subject ID #	Microfilaremia (Mfs/mL) ¹	IgG1 (µg/mL)	IgG2 (µg/mL)	IgG3 (µg/mL)	IgG4 (µg/mL)
1	Bm1-1	1696	30	35	19	594
	Bm1-2	- ²	39	76	10	1669
	Bm1-3	1	56	23	39	109
	Bm1-4	755	22	13	10	243
	Bm1-5	6	289	89	46	538
2	Bm2-1	950	48	79	50	2412
	Bm2-2	5	10	0	0	1037
	Bm2-3	150	12	2	0	733
	Bm2-4	290	46	76	11	412
	Bm2-5	15	52	72	8	226

¹Blood microfilaremia was estimated by filtration (Nuclepore) of a 1 mL nighttime sample of venous blood. Blood collection for parasitologic examination took place between 20:00 h and 23:00 h, in accordance with the periodicity of the MF in this area²⁸. For Set #1, plasma from blood sampling from April 1990 was used in our study, and thus, microfilariaemia observed at this time-point is presented here. For Set #2, blood sampling was carried out on two occasions, 3 months apart, before treatment with DEC. The mean of the two counts is presented. Details of the experimental procedure have been reported in¹⁵.

²Data not available (no sampling done) for this individual for the particular time-point (April 1990) used in our study. However, in the original study, this individual was microfilariaemic in the follow-up samplings from July and November 1990.

B] Information on *L. loa*-, *M. perstans*-, *O. volvulus*- and *W. bancrofti*-infected individuals

Subject ID #	Infection	Microfilaremia (Mfs/mL) ¹	Nodule
Ov-1	<i>O. volvulus</i>	0	1
Ov-2	<i>O. volvulus</i>	2	0
Ov-3	<i>O. volvulus</i>	0	1
Ov-4	<i>O. volvulus</i>	2	0
Ov-5	<i>O. volvulus</i>	17	0
Ov-6	<i>O. volvulus</i>	1	0
Mp-1	<i>M. perstans</i>	60	N/A
Mp-2	<i>M. perstans</i>	1	N/A
Mp-3	<i>M. perstans</i>	80	N/A

Mp-4	<i>M. perstans</i>	20	N/A
Mp-5	<i>M. perstans</i>	40	N/A
Mp-6	<i>M. perstans</i>	2	N/A
Ll-1	<i>L. loa</i>	740	N/A
Ll-2	<i>L. loa</i>	700	N/A
Ll-3	<i>L. loa</i>	34	N/A
Ll-4	<i>L. loa</i>	17	N/A
Ll-5	<i>L. loa</i>	25	N/A
Ll-6	<i>L. loa</i>	320	N/A
Wb-1	<i>W. bancrofti</i>	100	N/A
Wb-2	<i>W. bancrofti</i>	9670	N/A
Wb-3	<i>W. bancrofti</i>	110	N/A
Wb-4	<i>W. bancrofti</i>	3880	N/A
Wb-5	<i>W. bancrofti</i>	50	N/A
Wb-6	<i>W. bancrofti</i>	0	N/A

¹Microfilariaemia was estimated by microscopy of skin biopsies (*O. volvulus*) or blood (*L. loa*, *M. perstans*, *W. bancrofti*) as described in^{11,30,31}.

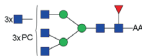
C] List of non-infected sera (control groups)

Subject ID #	Geographic area of origin
Un-G1	Africa, Ghana
Un-G2	Africa, Ghana
Un-G3	Africa, Ghana
Un-G4	Africa, Ghana
Un-G5	Africa, Ghana
Un-G6	Africa, Ghana
Un-NL1	Europe, The Netherlands
Un-NL2	Europe, The Netherlands
Un-NL3	Europe, The Netherlands
Un-NL4	Europe, The Netherlands
Un-NL5	Europe, The Netherlands

Table S4 (simplified): *O. volvulus* glycansA] *O. volvulus* N-glycans

N-GP number ¹	Theoretical m/z ²	Composition ³	HF ⁴	Exoglycosidase digestions ⁵			In figure ⁶	Most probable structural assignment ⁷
				α Mannosidase	β -GlcNAcAse	β -HexNAcAse		
N-GP1	868.32	H2N2-AA	ns	-1H	ns	ns	-	
N-GP2	1014.38	H2N2F1-AA	ns	-1H	ns	ns	-	
N-GP3	1030.37	H3N2-AA	ns	-2H	ns	ns	S2.A	
N-GP4	1176.43	H3N2F1-AA	ns	-2H	ns	ns	S2.A	
N-GP5	1192.43	H4N2-AA	ns	-3H	ns	ns	S2.A	
N-GP6	1233.43	H3N3-AA	ns	ns	-1N	ns	S2.B	
N-GP7	1338.48	H4N2F1-AA	ns	-2H	ns	ns	S2.A	
N-GP8	1354.48	H5N2-AA	ns	-4H	ns	ns	S2.A	
N-GP9	1379.51	H3N3F1-AA	ns	ns	-1N	ns	-	
N-GP10	1398.51	H3N3PC1-AA	-1PC	ns	ns	ns	S2B, C	
N-GP11	1436.53	H3N4-AA	ns	ns	-2N	ns	S2.B	
N-GP12	1500.54	H5N2F1-AA	ns	-4H	ns	ns	S2.A	
N-GP13	1516.53	H6N2-AA	ns	-5H	ns	ns	S2.A	
N-GP14	1544.48	H3N3PC1F1-AA	-1PC	ns	ns	ns	S2.C	
N-GP15	1557.56	H5N3-AA	ns	-2H	-1N	ns	-	
N-GP16	1582.59	H3N4F1-AA	ns	ns	-2N	ns	S2.B	

N-GP17	1601.59	H3N4PC1-AA	-1PC	ns	-1N	ns	S2B, C	
N-GP18	1639.61	H3N5-AA	ns	ns	-3N	-1N	S2.B	
N-GP19	1678.58	H7N2-AA	ns	-6H	ns	ns	S2.A	
N-GP20	1747.65	H3N4PC1F1-AA	-1PC	ns	-1N	ns	S2.C	
N-GP21	1785.67	H3N5F1-AA	ns	ns	-3N	-1N	S2.B	
N-GP22	1804.67	H3N5PC1-AA	-1PC	ns	-2N	-1N	S2.C	
N-GP23	1840.64	H8N2-AA	ns	-7H	ns	ns	S2.A	
N-GP24	1912.70	H3N4PC2F1-AA	-2PC	ns	ns	ns	S2.D, E	
N-GP25	1950.72	H3N5PC1F1-AA	-1PC	ns	-2N	-1N	S2.C	
N-GP26	1969.72	H3N5PC2-AA	-2PC	ns	-2N	-1N	S2.D	
N-GP27	1988.75	H3N6F1-AA	ns	ns	-4N	-2N	S2.C	
N-GP28	2002.69	H9N2-AA	ns	-8H	ns	ns	S2.A	
N-GP29	2007.75	H3N6PC1-AA	-1PC	ns	-3N	-2N	S2.D	
N-GP30	2115.78	H3N5PC2F1-AA	-2PC	ns	ns	ns	-	
N-GP31	2153.80	H3N6PC1F1-AA	-1PC	ns	-3/4N	-1/2N	S2.D, E	
N-GP32	2164.74	H10N2-AA	ns	-5H	ns	ns	-	
N-GP33	2172.80	H3N6PC2-AA	-2PC	ns	-1N	-1N	S2.D, E	
N-GP34	2318.86	H3N6PC2F1-AA	-2PC	ns	-2N	-1/2N	-	
N-GP35	2356.88	H3N7PC1F1-AA	-1PC	ns	-2/3/4N	-1/2N	S2.D, E	
N-GP36	2521.94	H3N7PC2F1-AA	-2PC	ns	-2/3N	-1/2N	-	

N-GP37	2890.07	H3N8PC3F1-AA	-3PC	ns	-2/3N	-1/2N	-	
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¹**N-GP number:** “glycopeak” number was associated to each m/z for which a *N*-glycan structure could be assigned and confirmed using listed glycan sequencing procedure

²**Theoretical m/z :** theoretical monoisotopic mass in negative-ion mode (m/z [M-H]⁻)

³**Compositions** were initially deduced using the glycopeakfinder tool of Glycoworkbench™, literature data and analogy with *B. malayi*. They were further confirmed using glycan sequencing (exoglycosidase digestions and MS/MS analysis).

H = hexose, N = N-acetylhexosamine, F = fucose, PC = phosphorylcholine, AA = 2-aminobenzoic acid

⁴**HF:** effect of treatment with HF on corresponding *N*-glycan, -nPC/F = loss of n PC/F, ns = non-sensitive

⁵**Exoglycosidase digestions:** effect of exoglycosidase treatment on corresponding *N*-glycan, -nH/N/A = loss of n H/N, ns = non-sensitive. αMannosidase = α1-2,3,6 Mannosidase, β-GlcNAcAse = β-N-Acetylglucosaminidase S, β-HexNAcAse = β-N-Acetylhexosaminidase (see M&M section).

Gray filling indicates that the treatment has not actually been performed, and the column is filled with its putative effect.

⁶**In figure:** Figures in which experimental data are shown (Fig n = main figures, Figure Sn = supplementary figures). Cells are filled with a dash = data not shown in this manuscript.

⁷**Most probable structural assignment** drawn using the CFG nomenclature (see symbol key below).

B] *O. volvulus* GSL glycans

GSL-GP number ¹	Theoretical m/z ²	Composition ³	HF ⁴	Exoglycosidase digestions ⁵			In figure ⁶	Most probable structural assignment ⁷
				αGalactosidase	β-GlcNAcAse	β-HexNAcAse		
GSL-GP1	830.30	H2N1PC1-AA	-1PC	ns	ns	-1N	-	
GSL-GP2	1033.38	H2N2PC1-AA	-1PC	ns	ns	-1N	-	
GSL-GP3	1195.43	H3N2PC1-AA	-1PC	-1H	ns	ns	4.A S3.A	
GSL-GP4	1209.41	H2A1N2PC1-AA	-1PC	ns	ns	ns	S3.C	
GSL-GP5	1236.46	H2N3PC1-AA	-1PC	ns	ns	ns	S3.A-B	
GSL-GP6	1341.49	H3N2PC1F1-AA	-1F -1PC	ns	ns	ns	S3.A-B	
GSL-GP7	1352.46	H3A1N2F1-AA	-1F	ns	ns	ns	-	
GSL-GP8	1371.46	H3A1N2PC1-AA	-1PC	ns	ns	ns	-	
GSL-GP9	1398.51	H3N3PC1-AA	-1PC	ns	ns	ns	S3.A-B	

GSL-GP10	1563.56	H3N3PC2-AA	-2PC	-1H	ns	ns	S3.D	
GSL-GP11	1574.54	H3A1N3PC1-AA	-1PC	ns	ns	ns	S3.D	
GSL-GP12	1604.59	H2N4PC2-AA	-2PC	ns	ns	-1N	S3.D	
GSL-GP13	1728.62	H3N3PC3-AA	-2PC	-1H	ns	ns	S3.G	
GSL-GP14	1739.60	H3A1N3PC2-AA	-2PC	ns	ns	ns	S3.F	
GSL-GP15	1758.62	H3A1N4F1-AA	-1F	ns	ns	ns	-	
GSL-GP16	1769.67	H2N4PC3-AA	-2PC	ns	ns	ns	S3.G	
GSL-GP17	1807.67	H2N5PC2-AA	-2PC	ns	ns	-1N	S3.F	
GSL-GP18	1972.75	H2N5PC3-AA	-3PC	ns	ns	ns	S3.G, H	
GSL-GP19	2010.75	H2N6PC2-AA	-2PC	ns	ns	-1N	S3.F	
GSL-GP20	2175.39	H2N6PC3-AA	-2PC	ns	ns	-1N	S3.G, H	
GSL-GP21	2321.89	H2N6PC3F1-AA	-1F -3PC	ns	ns	ns	S3.G	
GSL-GP22	2340.86	H2N6PC4-AA	-4PC	ns	ns	-2N	S3.G	
GSL-GP23	2378.91	H2N7PC3-AA	-3PC	ns	ns	-1N	S3.G, H	
GSL-GP24	2543.94	H2N7PC4-AA	-4PC	ns	ns	ns	-	

¹**GSL-GP number:** “glycopeak” number was associated to each *m/z* for which a GSL glycan structure could be assigned and confirmed using listed glycan sequencing procedure

²**Theoretical *m/z*:** theoretical monoisotopic mass in negative-ion mode (*m/z* [M-H]⁻)

³**Compositions** were initially deduced using the glycopeakfinder tool of Glycoworkbench™, literature data and analogy with *B. malayi*. They were further confirmed using glycan sequencing (exoglycosidase digestions and MS/MS analysis).

H = hexose, N = N-acetylhexosamine, F = fucose, A = glucuronic acid, PC = phosphorylcholine, AA = 2-aminobenzoic acid

⁴**HF:** effect of treatment with HF on corresponding GSL glycan, -nPC/F = loss of n PC/F, ns = non-sensitive

⁵**Exoglycosidase digestions:** effect of exoglycosidase treatment on corresponding GSL glycan, -nH/N/A = loss of n H/N, ns = non-sensitive. αGalactosidase = α1-3,4,6 Galactosidase, β-GlcNAcAse = β-N-Acetylglucosaminidase S, β-HexNAcAse = β-N-Acetylhexosaminidase_i (see M&M section).

Gray filling indicates that the treatment has not actually been performed, and the column is filled with its putative effect.

⁶**In figure:** Figures in which experimental data are shown (Fig n = main figures, Figure Sn = supplementary figures). Cells are filled with a dash = data not shown in this manuscript.

⁷**Most probable structural assignment** drawn using the CFG nomenclature (see symbol key below).

An extended version of **Table S4** is available online.

 Glucose (Glc)	 Galactose (Gal)
 <i>N</i> -acetylglucosamine (GlcNAc)	 <i>N</i> -acetylgalactosamine (GalNAc)
 Glucuronic acid (GlcA)	 Fucose (Fuc)
 Mannose (Man)	PC Phosphorylcholine (PC)

Additional supplementary material online

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Additional supplementary tables online:

- **Table S2:** Glycan array and statistical analysis raw data - IgG responses to *B. malayi* N-linked and GSL glycans in plasma from various filarial-infected and uninfected individuals
- **Table S3:** Glycan array and statistical analysis raw data - IgG responses to categories of *B. malayi* glycans in plasma from various filarial-infected and uninfected individuals
- Extended version of **Table S4:** *O. volvulus* N-linked and GSL glycans
- **Table S5:** Glycan array and statistical analysis raw data - IgG subclasses to *B. malayi* N-linked and GSL glycans in plasma from *B. malayi*, *O. volvulus* or *M. perstans*-infected and uninfected individuals
- **Table S6:** Glycan array and statistical analysis raw data - IgG subclasses to categories of *B. malayi* glycans in plasma from *B. malayi*, *O. volvulus* or *M. perstans*-infected and uninfected individuals

Additional supplementary figures online:

- **Figure S1:** Average plasma IgG responses to *B. malayi* glycans of individuals infected with *B. malayi*, *O. volvulus*, *M. perstans*, *L. loa* or *W. bancrofti* and of uninfected individuals
- **Figure S2:** Glycan sequencing of selected *O. volvulus* N-glycans
- **Figure S3:** Glycan sequencing of selected *O. volvulus* GSL glycans
- **Figure S4:** IgG subclass response to *B. malayi* glycans from infected individuals before and after DEC anthelmintic treatment
- **Figure S5:** IgG subclass response to *B. malayi* glycans from *O. volvulus*-infected individuals
- **Figure S6:** IgG subclass response to *B. malayi* glycans from *M. perstans*-infected individuals