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Mitochondrial oxygen tension in critically ill patients receiving red blood cell transfusions: a multicenter observational cohort study

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Abstract

Purpose Currently, there is no marker of efficacy of red blood cell (RBC) transfusion. This study describes the impact of RBC transfusion on mitochondrial oxygen tension (mitoPO₂) and mitochondrial oxygen consumption (mitoVO₂) in critically ill patients with anemia.

Methods Critically ill patients with a hemoglobin concentration < 10 g/dL, for whom a single RBC unit had been ordered, were included. MitoPO₂ was measured with the COMET device immediately before RBC transfusion, 0.5 h, 1 h, 3 h, and 24 h after RBC transfusion. MitoVO₂ was calculated from dynamic mitoPO₂ measurements during cessation of local oxygen supply.

Results Sixty-three patients participated, median age 64.0 (interquartile range (IQR) 52.3–72.8) years, median hemoglobin concentration before transfusion 7.4 (IQR 7.1–7.7) g/dL. Median mitoPO₂ values were 55.0 (IQR 49.6–63.0) mmHg before RBC transfusion, 51.0 (IQR 41.5–61.2) directly after and 67.3 (IQR 41.6–83.7) at 24 h after RBC transfusion. Median mitoVO₂ values were 3.3 (IQR 2.1–5.9) mmHg/s before RBC transfusion, 3.7 (IQR 2.0–5.1) mmHg/s directly after, and 3.1 (IQR 2.5–4.8) mmHg/s 24 h after RBC transfusion. In the higher Hb concentration group (> 7 g/dL), we saw a dissociation of the effect of RBC transfusion on mitoPO₂ versus on mitoVO₂ values. MitoPO₂ and mitoVO₂ values were not associated with commonly used parameters of tissue perfusion and oxygenation.

Conclusion RBC transfusion did not alter mitoPO₂ and mitoVO₂ in critically ill patients with anemia. MitoPO₂ and mitoVO₂ values were not notably associated with Hb concentrations, parameters of severity of illness and markers of tissue perfusion or oxygenation. Given the high baseline value, it cannot be excluded nor confirmed whether RBC can improve low mitoPO₂.

Trial registration number NCT03092297 (registered 27 March 2017)

Keywords Anemia, Red blood cell transfusion, Tissue oxygenation, Mitochondrial oxygen tension

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Introduction

Anemia is highly prevalent in critically ill patients [1–3], with reported incidences during intensive care unit (ICU) stay as high as 66–98% [2–7]. Severe anemia can lead to diminished oxygen carrying capacity, cellular oxygen deficit and organ dysfunction contributing to organ failure, morbidity, and (cardiac) mortality [1–3]. Red blood cell (RBC) transfusions are given to solve a possible cellular oxygen deficit in critically ill patients, currently guided predominantly by hemoglobin (Hb) concentrations. However, this guidance may not be optimal since hemoglobin concentration is an indirect marker of cellular oxygenation. Commonly used surrogate markers of tissue perfusion and cellular oxygenation include mean arterial pressure (MAP), cardiac output (CO), central venous oxygen saturation (ScvO₂), lactate concentration, and venous-to-arterial carbon dioxide difference (pCO₂ gap). A limitation of these markers is that their correction is not directly correlated with improved cellular oxygenation or organ perfusion [8–10]. Markers of microcirculatory perfusion have shown that RBC transfusions result in recruitment of the microcirculation, particularly in those with low baseline values, but have not shown benefit in clinically significant outcomes [11–13]. A parameter that directly measures cellular oxygenation could provide more accurate guidance to RBC transfusions.

The Cellular Oxygen Metabolism (COMET) monitoring system, measures the mitochondrial oxygen tension (mitoPO₂) non-invasively at the bedside [14]. The system uses the protoporphyrin IX-triplet state lifetime technique (PpIX-TSLT) [15, 16]. In a hemodilution study in pigs, oxygen deficit was observed in the skin of the anterior chest wall prior to other parameters of tissue perfusion [17]. This was corroborated in two case studies in humans after clonidine administration intraoperatively and during intraoperative blood loss [14, 18]. A parameter that is indirectly calculated from mitoPO₂ values is mitochondrial oxygen consumption (mitoVO₂) [14, 19, 20]. It has been suggested that mitoVO₂ can give additional information regarding mitochondrial function and oxygen consumption [19]. We therefore postulated that the COMET device might be a valuable monitor for measurement of tissue oxygenation in critically ill patients with anemia.

The aim of this study was to describe mitoPO₂ and mitoVO₂ values as assessed with the COMET device in critically ill patients with anemia, before and after red cell transfusion, and to examine the association of these parameters with commonly used parameters of tissue perfusion and tissue oxygenation and with indicators of severity of critical illness, demographic and outcome characteristics.

Methods

Study design

A detailed overview of the design, procedure and protocol of this study was published elsewhere [21]. A concise overview regarding the study design, data collection, PpIX-TSLT technique description and study procedure is given in the supplementary material. In short, we performed an observational cohort study between March 2018 and April 2020 in two academic ICU departments in the Netherlands. Critically ill patients with anemia, defined as an Hb concentration < 10 g/dL, with an arterial catheter in situ receiving RBC transfusion were included in the study. Critically ill patients in need of RBC transfusion within 4 h were excluded from the study, as well as critically ill patients with an expected admittance in the ICU unit < 24 h. Patients younger than 18 years, with a brown plaster allergy, with photodermatosis and/or porphyria or insufficient Dutch language comprehensibility were not deemed eligible for the study, as well as pregnant women.

MitoPO₂ and mitoVO₂ measurements

MitoPO₂ measurements consisted of two phases: first we did dynamic measurements, after which we performed static measurements. During the dynamic phase, local pressure was applied on the measurement probe resulting in occlusion of the underlying microcirculation, resulting in an immediate drop in mitoPO₂ values as well as a fast recovery after release of the pressure [14, 21]. The mitoPO₂ value was measured every second for 120 s. The mitoPO₂ values before and during local pressure were used to fit a sigmoid function, after which a linear function was used to calculate the mitoVO₂ on the steepest part of the sigmoid curve. After the dynamic phase, mitoPO₂ was measured once per minute for five minutes, to obtain a mean mitoPO₂ at each time point. More details regarding the study procedure is given in the supplementary material.

Statistical analyses

Descriptive statistics were used to describe the characteristics of the study population. Quantitative data were shown as means with a standard deviation (SD) or median with an interquartile range (IQR), as appropriate. Categorical variables were presented as number (percentage). The number of observations that were missing were visualized and described. Number of mitoPO₂ measurement moments without a valid value are presented in Supplementary Materials-Table 2 [22].

The mitoPO₂ measurements and signal quality were described per measurement moment in the total population, and in subgroups of participating study centers. MitoPO₂ and mitoVO₂ values were described for the total

study population and stratified according to pre-transfusion hemoglobin concentration of ≤ 7 g/dL and > 7 g/dL. We calculated the change in mitoPO₂ at various time-points after transfusion compared with before transfusion, for all individuals, as well as per pre-transfusion subgroups, and presented mean differences with 95% confidence intervals using non-missing mitoPO₂ values at each measurement time point.

The non-missing mitoPO₂ values before RBC transfusion were used to assess the association between mitoPO₂ and mitoVO₂ with demographic and outcome characteristics. Univariate analyses were performed using ANOVA to calculate the associations between mitoPO₂ and demographic and outcome characteristics. To examine the association between hemodynamic characteristics and mitoPO₂, mitoPO₂ values of all measurement moments were used. Univariate analyses with ANOVA were used to calculate the associations between mitoPO₂ and mitoVO₂ with markers of tissue perfusion and oxygenation. It has been suggested that normal mitoPO₂ values are between 40 and 70 mmHg [23, 24]. Therefore, mitoPO₂ values were categorized into three subgroups: < 40 mmHg, 40–70 mmHg, and > 70 mmHg. Within these subgroups, markers of tissue perfusion and oxygenation were described, including a lactate concentration measured one measurement time later. Furthermore, the mitoPO₂ values before RBC transfusion were categorized into the same subgroups to describe the course over time in these subgroups.

We used mitoPO₂ values with a signal quality of at least 20% for our analyses. The cut-off value of the signal quality of a mitoPO₂ measurement to ascertain a valid measurement is around 20%. We performed sensitivity analyses post hoc to evaluate the effect of less strict signal quality of measurements in our results. During the sensitivity analyses, all aforementioned analyses were performed with mitoPO₂ values based on a signal quality of at least 10%.

MitoVO₂ calculations were performed using an automated MATLAB (Mathworks, R2022b Update 3) algorithm [25]. All other statistical analyses were performed using R (R foundation for Statistical Computing, Vienna, Austria) [26].

Results

Characteristics of the study population

Of the 475 critically ill patients planned to receive RBC transfusion during the study period, 63 patients were included in the analyses, as depicted in Supplementary Materials-Fig. 1, corresponding to 378 observation moments (six measurements per included patient). Table 1 shows characteristics of the study population consisting of mostly male (76%), with a median age of

Table 1 Characteristics of the study population of all 63 critically ill patients with anemia

Characteristic	Overall cohort (n = 63)
Age in years, median (IQR)	64.0 (53.0; 73.0)
Female sex, n (%)	15 (24%)
BMI in kg/m ² , median (IQR)	27.9 (23.9; 32.6)
Admission reason, n (%)	
Surgical	31 (49.0%)
Cardiovascular	15 (48.4%)
Digestive	6 (19.4%)
Neurosurgery	2 (6.5%)
Genitourinary	1 (3.2%)
Trauma	3 (9.7%)
Miscellaneous	4 (12.9%)
Infectious process in throat	1 (3.2%)
Liver transplantation	1 (3.2%)
Arthrotomy	1 (3.2%)
Fasciitis necroticans	1 (3.2%)
Non-surgical	32 (51.0%)
Cardiovascular	2 (6.3%)
Respiratory	12 (37.5%)
Digestive	7 (21.9%)
Genitourinary tract	1 (3.1%)
Sepsis	8 (25.0%)
Neurological	1 (3.1%)
Chronic comorbidities, n (%)	
No comorbidity	22 (35.0%)
Chronic cardiovascular insufficiency	5 (8.0%)
Chronic obstructive lung disease	1 (2.0%)
Chronic renal insufficiency	2 (3.0%)
Chronic dialysis	1 (2.0%)
Cirrhosis	5 (8.0%)
Hematologic malignancy	3 (5.0%)
Immunologic deficiency	2 (3.0%)
Diabetes mellitus	22 (35.0%)
SOFA score before transfusion, median (IQR)	10 (8–12)
APACHE IV score, median (IQR)	70.0 (58.0; 88.0)
	Missing: 2 (3.2%)
Hemoglobin concentration before transfusion in g/dL, median (IQR)	7.4 (7.1; 7.7)
Hematocrit before transfusion in L/L, median (IQR)	0.23 (0.22; 0.24)
Days admitted to ICU at inclusion, median (IQR)	6.0 (3.0; 11.0)

APACHE Acute Physiologic and Chronic Health Evaluation, BMI body mass index, IQR interquartile range, SOFA Sequential Organ Failure Assessment

64.0 (IQR 53.0–73.0) years. The median Hb concentration before RBC transfusion was 7.4 (IQR 7.1; 7.7) g/dL. The median Hb concentration one hour after RBC transfusion was 8.2 (IQR 7.9; 8.9) g/dL, and it was 8.4 (IQR 7.9; 9.0) g/dL after 24 h.

MitoPO₂ and mitoVO₂ before and after RBC transfusion

Figure 1 presents all observed valid (signal quality > 20%) mitoPO₂ values as assessed with the COMET measurement device in all 63 critically ill patients with anemia before and at the predefined timepoints during the first 24 h after RBC transfusion. MitoPO₂ values prior to RBC transfusion showed large variation and were largely within normal limits. The overall median mitoPO₂ before RBC transfusion was 55.0 (IQR 49.6; 63.0) mmHg, it was 51.0 (IQR 41.5; 61.2) mmHg at the end of the transfusion and 67.3 (IQR 41.6; 83.7) mmHg 24 h after RBC transfusion (Table 2). Stratification according to baseline mitoPO₂ also yielded a heterogeneous response immediately following RBC transfusion (Fig. 1). After 24 h, those with a low baseline mitoPO₂ < 40 mmHg tended to increase, whereas those with a mitoPO₂ baseline value > 70 mmHg tended to decrease. However, the number of missing values at t = 24 h hamper statistical interpretation of this

observation (supplementary materials-Table 3). Also, only two patients had mitoPO₂ values below 30 mmHg.

Supplementary Materials-Table 2 presents the number of patient-moments at which we did not obtain a valid mitoPO₂ value along with reasons for it. Absence of a valid mitoPO₂ value was associated with severity of illness. The median APACHE IV score was 86.5 (IQR 59.3; 97.3) in critically ill patients without a valid mitoPO₂, whereas the median APACHE IV score was 70.0 (IQR 58.0; 83.0) in the critically ill patients with a valid mitoPO₂ value (Supplementary Materials-Table 4). In patients with a high SOFA score, mitoPO₂ values tended to increase after RBC over time, whereas in those with a low SOFA score, mitoPO₂ tended to decrease over time (Supplementary Table 6).

Twelve patients had a pre-transfusion Hb concentration ≤ 7 g/dL and their median mitoPO₂ before RBC transfusion was 61.3 (IQR 51.0; 69.3) mmHg, while in patients with an Hb concentration > 7 g/dL it was 55.0 (IQR 48.9;

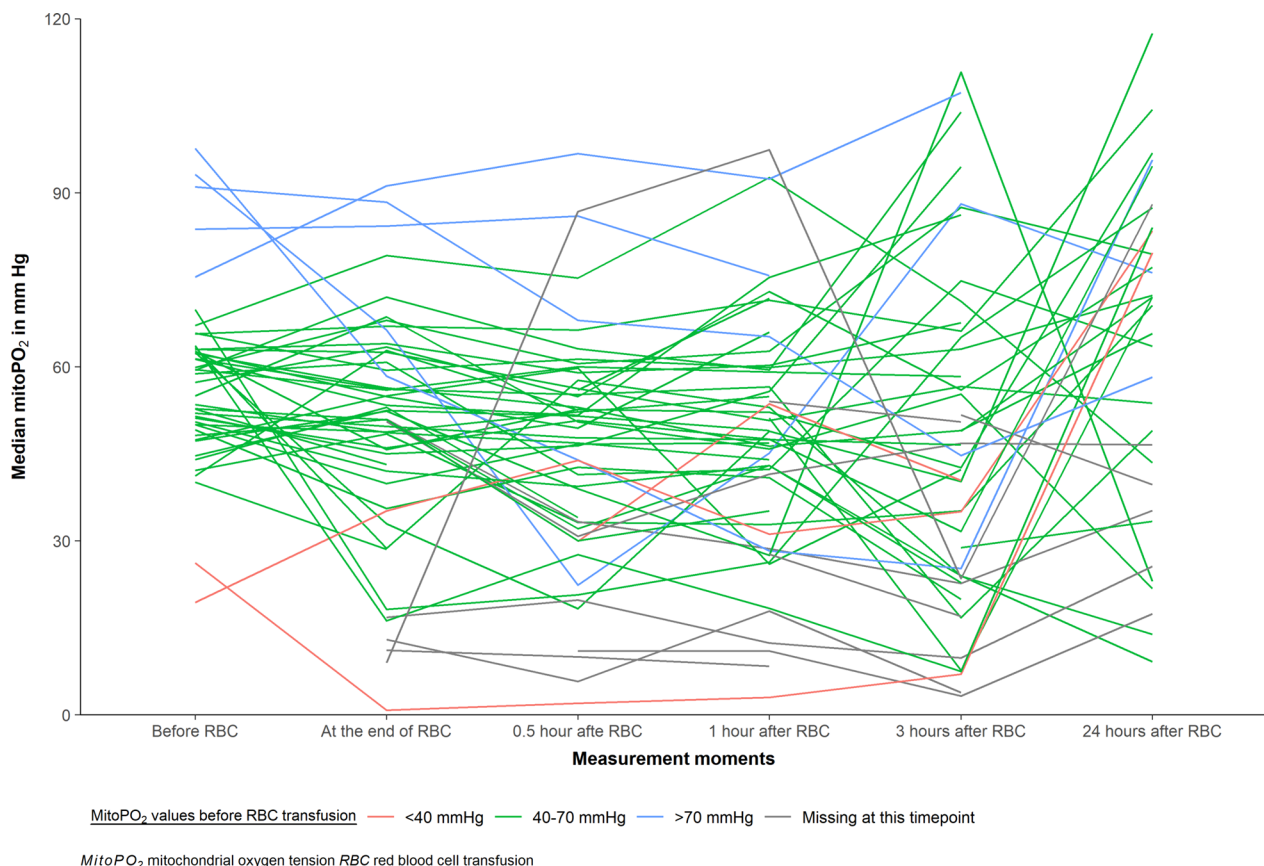


Fig. 1 Spaghetti plot showing all observed valid (signal quality > 20%) mitoPO₂ values measured with the COMET measurement device in all 63 critically ill patients with anemia, before and at various timepoints during the first 24 h after RBC transfusion. The range of mitoPO₂ values is approximately 40–70 mmHg before RBC transfusion, while 24 h after RBC transfusion a wider range in mitoPO₂ values is seen (15–110 mmHg). Red lines represent patients whose first valid mitoPO₂ value was < 40 mmHg; green lines represent patients whose first valid mitoPO₂ value was between 40 and 70 mmHg and blue line represents patients whose first valid mitoPO₂ was above 70 mmHg

Table 2 Mitochondrial oxygenation tension measured with COMET in critically ill patients receiving RBC transfusion in the total study population and stratified according to pre-transfusion hemoglobin concentration

Measurement time	MitoPO ₂ ^a in mmHg in all critically ill patients, median (IQR); n	MitoPO ₂ ^a in mmHg in strata of critically ill patients with anemia based on Hb concentration before RBC transfusion, median (IQR), n	
		Hb concentration ≤ 7 g/dL, n = 12	Hb concentration > 7 g/dL, n = 51
Before RBC transfusion	55.0 (49.6; 63.0); 51	61.3 (51.0; 69.3); 8	55.0 (48.9; 62.8); 42
End of RBC transfusion	51.0 (41.5; 61.2); 54	48.8 (23.1; 61.4); 11	51.0 (43.6; 60.4); 43
0.5 h after RBC transfusion	49.4 (33.2; 57.7); 55	51.6 (41.2; 79.3); 11	47.8 (32.6; 56.1); 44
1 h after RBC transfusion	47.5 (33.4; 59.4); 55	58.0 (44.5; 75.6); 11	47.1 (32.4; 55.3); 44
3 h after RBC transfusion	45.6 (23.8; 63.6); 50	59.6 (23.3; 88.3); 9	43.7 (24.0; 58.4); 41
24 h after RBC transfusion	67.3 (41.6; 83.7); 44	39.7 (25.6; 56.2); 6	71.3 (46.7; 83.9); 38

^a MitoPO₂ mitochondrial oxygen tension measured with the COMET system

62.8) mmHg (Table 2). This is contrary to our hypothesis. The course of median mitoPO₂ over time, stratified to pre-transfusion Hb concentration, suggests a decrease in those with low pre-transfusion Hb level (≤ 7 g/dL) and an increase in those with a higher pre-transfusion Hb level (> 7 g/dL) (Table 2 and Supplemental Materials-Fig. 3). Mean differences between mitoPO₂ before RBC transfusion and mitoPO₂ values at each measurement moment after RBC transfusion are depicted in Supplementary Materials-Table 5.

Supplementary Materials-Tables 8–16 present mitoPO₂ values over time in a number of other subgroups, including stratification according to age or sex. The estimates were all consistent with our primary analysis.

If we look at O₂ consumption, patients with a pre-transfusion Hb concentration of ≤ 7 g/dL had a median mitoVO₂ before RBC transfusion of 7.6 (IQR 4.6; 10.5) mmHg/s. In patients with an Hb concentration > 7 g/dL pre-transfusion mitoVO₂ was 3.3 (IQR 2.2; 5.7) mmHg/s (Supplementary Materials-Table 17).

Following RBC transfusion both mitoPO₂ and mitoVO₂ decreased in the low Hb concentration group (≤ 7 g/dL) whereas in the higher Hb concentration group (> 7 g/dL), we saw that after RBC transfusion, mitoPO₂ increased while mitoVO₂ did not change, i.e., a dissociation of the effect of RBC transfusion on mitoPO₂ versus on mitoVO₂ (Table 2, Supplementary Materials-Table 17).

Association with clinical characteristics and with commonly used markers of tissue perfusion or oxygenation

Table 3 shows similar mitoPO₂ and mitoVO₂ values according to different demographic and clinical outcome characteristics. MitoPO₂ per APACHE IV score: median mitoPO₂ was 61.1 (IQR 59.0; 83.9) mmHg in the APACHE IV score < 50 subgroup; it was 53.1 (IQR 46.3; 62.5) mmHg in the APACHE IV score 50–80 subgroup, and 55.0 (50.5; 63.6) mmHg in the APACHE score > 80 subgroup (p-value for the trend = 0.137). Of all 378 mitoPO₂ measurements 79 values were < 40 mmHg, and 169 mitoPO₂ measurements were between 40 and 70 mmHg, and 53 mitoPO₂ measurement > 70 mmHg (Table 4). Very low mitoPO₂ (< 40 mmHg) was not convincingly associated with any of the measured markers of tissue perfusion and oxygenation (Table 4). Supplementary Materials-Table 19 similarly illustrates the absence of clear association between mitoPO₂ or mitoVO₂ values and the other measured markers of tissue perfusion and oxygenation. A statistically non-significant increase was seen in mitoVO₂ values in patients with higher FiO₂ values as depicted by a median mitoVO₂ of 2.92 (IQR 2.01; 4.25) mmHg/s with FiO₂ ≤ 30% to median 3.47 (IQR 2.30; 4.95) mmHg/s with FiO₂ > 30% (p-value for trend = 0.074).

Table 3 Median mitoPO₂ and mitoVO₂ values, using the measurement before RBC transfusion, according to demographic and outcome characteristics of the study population

Characteristics	Number of participants with mitoPO ₂ measurement (total 51) ^a	MitoPO ₂ in mmHg, median (IQR)	Mean difference (95% CI)	p-value for trend	Number of participants with mitoVO ₂ measurement (total 26) ^b	MitoVO ₂ in mmHg/s, median (IQR)	Mean difference (95% CI)	p-value for trend
Demographic characteristics								
Sex								
Female	12	52.3 (33.5; 68.9)	Reference	0.742	4	1.99 (1.60; 3.00)	Reference	0.210
Male	39	50.9 (40.2; 63.0)	− 1.6 (− 11.3; 8.1)		22	3.70 (2.34; 7.03)	2.56 (− 1.53; 6.65)	
Age								
< 50 years	7	56.9 (47.4; 62.4)	Reference	0.743	4	2.83 (1.92; 4.94)	Reference	0.673
50–64 years	21	52.8 (41.6; 66.3)	− 1.5 (− 14.4; 11.5)		11	3.87 (2.23; 6.70)	0.97 (− 3.64; 5.58)	
65–75 years	15	47.0 (27.3; 58.1)	− 4.9 (− 18.6; 8.7)		8	3.98 (2.54; 7.13)	1.58 (− 3.26; 6.42)	
> 75 years	8	51.0 (42.3; 62.5)	− 6.6 (− 22.0; 8.8)		3	2.88 (1.93; 3.40)	− 1.44 (− 7.47; 4.60)	
BMI								
< 25 kg/m ²	19	56.1 (41.3; 66.3)	Reference	0.232	9	3.91 (2.12; 5.09)	Reference	0.547
25–30 kg/m ²	14	49.4 (28.8; 60.1)	− 8.8 (− 19.0; 1.4)		7	3.09 (2.85; 4.74)	− 0.51 (− 4.43; 3.40)	
> 30 kg/m ²	18	49.1 (40.4; 60.6)	− 3.1 (− 12.7; 6.3)		10	2.92 (1.81; 10.32)	1.41 (− 2.16; 4.98)	
APACHE IV score								
< 50	6	61.1 (59.0; 83.9)	Reference	0.137	3	10.70 (7.81; 11.25)	Reference	0.102
50–80	26	53.1 (46.3; 62.5)	− 13.3 (− 26.5; − 0.1)		14	2.88 (2.21; 3.90)	− 4.91 (− 9.59; − 0.24)	
> 80	17	55.0 (50.5; 63.6)	− 10.0 (− 23.9; 3.8)		8	3.39 (1.59; 6.36)	− 4.92 (− 9.90; 0.06)	
Missing	2	55.1 (52.7; 57.5)			1	3.53 (3.53; 3.53)		
Reason of admission to ICU								
Medical	26	52.5 (43.9; 63.0)	Reference	0.536	10	2.24 (1.80; 4.67)	Reference	0.230
Surgical	25	49.8 (28.6; 63.5)	− 2.6 (− 10.8; 5.7)		16	3.70 (2.87; 7.20)	1.82 (− 1.23; 4.86)	
Hemoglobin concentration before RBC transfusion								
< 7 g/dL	7	52.1 (39.5; 75.7)	Reference	0.380	2	7.59 (4.64; 10.55)	Reference	0.269
≥ 7 g/dL	44	50.9 (39.3; 62.8)	− 5.2 (− 17.2; 6.7)		24	3.31 (2.16; 5.72)	− 3.06 (− 8.65; 2.52)	
Days admitted to ICU before inclusion								
< 6	22	50.5 (39.0; 63.1)	Reference	0.675	12	2.98 (1.55; 5.99)	Reference	0.670
≥ 6	29	52.3 (39.6; 63.4)	1.7 (− 6.6; 10.1)		14	3.89 (2.43; 5.92)	0.64 (− 2.42; 3.69)	

Table 3 (continued)

Characteristics	Number of participants with mitoPO ₂ measurement (total 51) ^a	MitoPO ₂ in mmHg, median (IQR)	Mean difference (95% CI)	p-value for trend	Number of participants with mitoVO ₂ measurement (total 26) ^b	MitoVO ₂ in mmHg/s, median (IQR)	Mean difference (95% CI)	p-value for trend
Outcome characteristics								
SOFA score before RBC transfusion								
< 10	17	47.0 (27.8; 56.5)	Reference	0.365	9	5.61 (1.69; 10.70)	Reference	0.289
≥ 10	34	54.9 (42.8; 65.9)	3.9 (− 4.7; 12.6)		17	3.09 (2.30; 4.93)	− 1.65 (− 4.78; 1.49)	
SOFA score change in 24 h								
≥ 0 (decrease in SOFA score)	16	50.5 (28.7; 61.9)	Reference	0.297	13	3.53 (2.18; 6.03)	Reference	0.842
< 0 (increase in SOFA score)	28	53.0 (42.0; 53.5)	5.0 (− 14.7; 4.6)		11	2.88 (1.90; 5.35)	0.30 (− 2.78; 3.37)	
Missing	7	50.5 (48.7; 52.8)			2	7.32 (5.07; 9.56)		
Total ICU admission duration								
< 5 days	7	56.2 (43.1; 66.2)	Reference	0.547	2	6.56 (3.94; 9.18)	Reference	0.824
5–10 days	8	46.8 (27.6; 59.4)	1.7 (− 13.6; 16.9)		6	4.35 (2.90; 8.29)	− 1.10 (− 7.65; 5.44)	
11–20 days	15	50.8 (43.0; 63.2)	7.4 (− 6.1; 20.9)		5	2.88 (2.30; 4.93)	− 1.86 (− 8.57; 4.85)	
> 20 days	21	52.5 (35.1; 63.4)	7.5 (− 5.4; 20.3)		13	3.53 (2.12; 5.09)	− 2.36 (− 8.45; 3.73)	
Total hospital admission duration								
< 10 days	6	59.4 (51.4; 67.3)	Reference	0.648	2	6.56 (3.94; 9.18)	Reference	0.926
10–20 days	10	45.8 (33.0; 52.8)	0.4 (− 14.8; 15.7)		6	4.35 (2.50; 8.29)	− 1.19 (− 7.91; 5.52)	
21–30 days	9	56.3 (44.2; 67.6)	7.3 (− 8.3; 22.8)		5	2.18 (1.69; 4.93)	− 2.26 (− 9.13; 4.62)	
31–50 days	10	59.2 (49.3; 63.8)	4.7 (− 10.6; 20.0)		4	2.85 (2.65; 4.00)	− 2.76 (− 9.89; 4.36)	
> 50 days	16	51.2 (41.5; 62.7)	7.9 (− 6.2; 22.1)		9	3.87 (2.88; 5.09)	− 1.90 (− 8.33; 4.53)	
In-hospital or ICU mortality								
No	37	51.7 (40.1; 63.5)	Reference	0.878	16	3.31 (2.01; 5.33)	Reference	0.925
Yes	14	50.9 (35.0; 62.9)	0.7 (− 8.5; 10.0)		10	3.90 (2.21; 6.92)	0.14 (− 3.00; 3.29)	

APACHE Acute Physiology and Chronic Health Evaluation, BMI body mass index, CI confidence interval, Hb hemoglobin, ICU intensive care unit, MitoPO₂ mitochondrial oxygen tension measured with the COMET system, MitoVO₂ mitochondrial oxygen consumption calculated with a linear function on a fitted sigmoid curve of the mitoPO₂ measurements

^a Number of participants with mitoPO₂ measurements before RBC transfusion

^b Number of participants with mitoVO₂ measurements before RBC transfusion

Sensitivity analyses

The results of the sensitivity analyses with mitoPO₂ value with a signal quality of at least 10% showed

similar results and trends as our main analyses where the signal quality used was at least 20% (Supplementary Materials-Tables 20–25).

Table 4 Characteristics of clinically used surrogate markers of tissue perfusion and oxygenation in critically ill patients in ICU within subgroups of mitoPO₂ values of < 40, 40–70, and > 70 mmHg

Clinical characteristics ^a	MitoPO ₂		
	< 40 mmHg (n = 79)	40–70 mmHg (n = 169)	> 70 mmHg (n = 53)
MitoPO ₂ in mmHg, median (IQR)	23.5 (13.5; 31.0)	52.8 (47.5; 59.9)	86.2 (75.7; 94.6)
MitoVO ₂ in mmHg/s, median (IQR)	2.44 (1.54; 3.75)	3.15 (2.20; 4.91)	4.13 (3.00; 5.36)
Missing, n (%)	26 (32.9%)	86 (50.9%)	15 (28.3%)
MAP in mmHg, median (IQR)	79.0 (69.2; 87.7)	80.0 (72.7; 89.0)	77.0 (71.0; 87.7)
Vasopressor use, n (%)	39 (49%)	93 (55%)	19 (36%)
Missing, n (%)	0 (0.0%)	1 (0.6%)	0 (0.0%)
Lactate in mmol/L, median (IQR)	1.5 (1.1; 1.9)	1.4 (1.0; 1.7)	1.5 (1.2; 1.9)
Missing, n (%)	3 (3.8%)	12 (7.1%)	6 (11.3%)
Lactate one measurement later in mmol/L, median (IQR)	1.5 (1.2; 1.9)	1.4 (1.1; 1.7)	1.4 (1.0; 1.8)
Missing, n (%)	17 (21.5%)	35 (20.7%)	32 (60.4%)
ScvO ₂ in %, median (IQR)	70.0 (64.5; 75.0)	71.0 (64.0; 76.8)	69.7 (59.3; 80.5)
Missing, n (%)	32 (40.5%)	79 (46.7%)	23 (43.4%)
pCO ₂ gap in mmHg, median (IQR)	4.5 (2.3; 6.0)	4.5 (2.4; 6.6)	4.5 (3.0; 6.0)
Missing, n (%)	41 (51.9%)	95 (56.2%)	28 (52.8%)
Cardiac index in L/min/m ² , median (IQR)	3.8 (3.1; 4.3)	3.9 (3.0; 5.2)	3.6 (3.1; 4.2)
Missing, n (%)	48 (60.8%)	108 (63.9%)	32 (60.4%)
Fractional inspired oxygen in %, median (IQR)	35.0 (30.0; 40.0)	35.0 (28.0; 45.0)	35.0 (25.0; 40.0)
Missing, n (%)	15 (19%)	20 (11.8%)	8 (15.1%)
Fluid balance in L, median (IQR)	0.749 (− 0.295; 1.927)	0.742 (− 0.018; 1.423)	0.582 (− 0.541; 1.105)
Missing, n (%)	7 (8.9%)	37 (21.9%)	14 (26.4%)

There were 77 mitoPO₂ measurements in 37 patients missing, which were not used in the subgroups

MitoPO₂ mitochondrial oxygen tension measured with the COMET system; MitoVO₂ mitochondrial oxygen consumption calculated with a linear function on a fitted sigmoid curve of the mitoPO₂ measurements; pCO₂ gap venous-to-arterial carbon dioxide difference; ScvO₂ central venous oxygen saturation; SOFA Sequential Organ Function Assessment

^a These are median values over the overall study period

Discussion

We performed a study in which we assessed mitoPO₂ and mitoVO₂ in critically ill patients with anemia before and after RBC transfusion. MitoPO₂ values in critically ill patients with anemia were not substantially lower than values previously observed in other critically ill patients and did not significantly change during the first 24 h after RBC transfusion. MitoPO₂ and mitoVO₂ values were not notably associated with Hb concentrations, parameters of severity of illness, and markers of tissue perfusion or cellular oxygenation in our study population. In patients with a pre-transfusion concentration > 7 g/dL we saw a dissociation between mitoPO₂ and mitoVO₂ with respect to the effect of RBC transfusion. In patients with a pre-transfusion Hb concentration ≤ 7 g/dL, both mitoPO₂ and mitoVO₂ did not increase after RBC transfusion but rather both decreased over time (24 h).

Main findings in relation to what is already known about the topic

Our study showed relatively normal mitoPO₂ values, in critically ill patients with anemia, that did not increase after a RBC transfusion. An absence of effect of RBC

transfusion on mitoPO₂ is in line with a previous trial showing no benefit of guiding RBC transfusion according to a marker of tissue oxygenation, although some observational studies have suggested benefit of this approach [27, 36, 37]. Theoretically one would expect an increase in tissue oxygenation after RBC transfusion in patients who benefit from transfusion, especially in the critically ill patient with markers of low tissue perfusion and low Hb concentration before RBC transfusion [11, 27]. However, most of our study participants had a transfusion trigger above 7 g/dL, as well as normal mitoPO₂, lactate, MAP, ScvO₂, and pCO₂ gap values before RBC transfusion. These ‘normal’ indices of tissue perfusion before RBC transfusion, suggest that a too liberal transfusion trigger was used, which may possibly explain the absence of an increase in mitoPO₂ values after RBC transfusion. This lack of effect caused by a too liberal transfusion trigger is supported by the fact that low mitoPO₂ values are expected with hematocrit values of 0.14 L/L or lower [23], which none of our study participants had. Just one of our study participants received more than one RBC transfusion units before mitoPO₂ measurements, thereby limiting the interpretation of our results to only critically ill patients receiving RBC transfusion. Furthermore, it

might be suggested that the critical illness has not (yet) led to low mitoPO₂ values. An alternative explanation may be that RBC transfusion is ineffective at improving tissue oxygenation in this cohort of critically ill patients which are not actively resuscitated. Pre-transfusion baseline values have previously been shown important in predicting the response to transfusion on a micro-circulatory level. A recent study suggested that a critically low mitoPO₂ value of 30 mmHg or lower would be indicative of tissue hypoxia [23]. Of note, only 2 of the 63 critically ill patients with anemia pre-transfusion had a mitoPO₂ < 30 mmHg. All the above considerations raise the question if RBC transfusions were needed in most of our critically ill patients with anemia [11, 12]. A final explanation of an absence of effect of a single RBC transfusion on mitoPO₂ might be that the effect of RBC transfusion might be too small to increase mitoPO₂ value [21]. Being a new monitoring technique, multiple studies have already been performed with the COMET measurement device. Studies with the COMET measurement device in healthy volunteers and critically ill patients have shown a normal mitoPO₂ ranging between 40 and 70 mmHg [24, 31–34]. A recent study into the mitochondrial oxygen measurement with the COMET measurement device has determined that a normal range of mitoPO₂ in physiological steady state is between 40 and 60 mmHg in the skin [23]. Indeed, our median mitoPO₂ values correspond with normal mitoPO₂ values.

The mitoVO₂ in our population ranged from 2.8 to 3.7 mmHg/s corresponding with mitoVO₂ values between 3.3 and 4.6 mmHg/s in critically ill patients that have been described in other studies [31, 32]. This is lower than the mitoVO₂ values found in healthy volunteers ranging from 5.8 to 6.7 mmHg/s [14, 33], suggesting a decreased cellular respiration in critically ill patients with anemia. Since mitoVO₂ is not directly measured by the COMET measurement device, the mitoVO₂ needs to be calculated from the mitoPO₂ values during application of pressure on the COMET probe. Different mechanisms have been described to calculate the mitoVO₂, using the Michaelis–Menten kinetics [14], fitting a sigmoid curve [32], or using a linear function [31]. These different approaches could lead to different results, therefore mitoVO₂ comparison should be done cautiously.

Interestingly, when looking in patients with lower (≤ 7 g/dL) versus a higher (> 7 g/dL) pre-transfusion Hb concentration, we observed different baseline values of mitoPO₂ and mitoVO₂ and different effects of RBC on mitoPO₂ and mitoVO₂, that were unexpected. Contrary to our expectations, the patients with a pre-transfusion Hb concentration < 7 g/dL had somewhat higher mitoPO₂ values compared to patients with Hb ≥ 7 g/dL. The higher mitoPO₂ value could have

been the result of mitochondrial adaptation for an optimal mitochondrial energy metabolism [9, 28, 29]. It has been described that oxygen consumption in the mitochondria can be reduced in response to mitochondrial hypoxia, leading to excess oxygen in the mitochondria. Inflammatory mediators, e.g., nitric oxide, in sepsis and shock have been described causing this mitochondrial adaptation [9, 28, 30]. However, this is contradicted by our finding that the calculated mitochondrial oxygen consumption was higher in critically ill patients with a pre-transfusion Hb concentration < 7 g/dL compared to patients with Hb ≥ 7 g/dL. It would be interesting to study the activity of mitochondrial adaptation mechanisms and their influence on the mitoPO₂ in future studies.

In the patients with a higher pre-transfusion Hb concentration, a dissociation between the effect of RBC on mitoPO₂ and mitoVO₂ was observed, i.e., mitoPO₂ increased and mitoVO₂ did not change after RBC. Concomitantly, we observed a decrease both in mitoPO₂ and mitoVO₂ after RBC in patients with a low pre-transfusion Hb concentration. A possible explanation that has been offered before may be the nitric oxide-dependent vasodilatation effect of RBC transfusions due to plasma-free Hb [27]. Furthermore, this may suggest an inability of cells to use oxygen, previously referred to as cellular dysoxia, as shown before in sepsis patients [28].

A relatively large part of the mitoPO₂ values had a low signal quality, which persisted until one hour after the end of RBC transfusion. This has been reported in other studies using the COMET measurement device [31, 32, 35]. Importantly, this is one of the first studies describing the characteristics of the patients with missing mitoPO₂ values due to signal quality below a protocol-set threshold. It seems that overall, these patients were more critically ill compared to the critically ill patients with valid mitoPO₂ measurements, which may have led to an overestimation of mitoPO₂. The critical illness might have influenced the ALA-plaster absorption or PpIX formation, resulting in a sub-par signal quality after four hours ALA plaster induction. Our data suggest that more than 4 h ALA plaster induction may be needed to guarantee adequate upregulation of PpIX in critically ill patients for a qualitative mitoPO₂ measurement with the COMET measurement device.

Strength and limitations

Strengths of our study entailed the prospective nature of our study in multiple study sites, as well as the gathering of data at multiple timepoints. The data gathering was made as complete as possible to gain as much insight into the critically ill patients with anemia. Furthermore,

the study design mimics clinical practice, making it more applicable to the daily practice.

Despite the high protocol adherence, missing data could not be prevented. Overall, most missing data were due to logistical issues, i.e., measurement in a weekend day or night time when no one of the study team was available. Therefore, missing not at random could not be ruled out, hence missing data could not be handled with imputation methods. We therefore interpreted our data cautiously, keeping in mind the large confidence intervals of mitoPO₂ values, while looking into the mean and median mitoPO₂ values.

Clinical implications

This study is one of the first studies looking into bedside cellular oxygenation in patients receiving RBC transfusion and the effect of this RBC transfusion on the cellular oxygenation. It shows that in critically ill patients, overall mitoPO₂ values are normal, and that when administered based on an Hb trigger, RBC transfusion does not result in an increase in mitoPO₂ or mitoVO₂. Findings are in line with other studies trying to determine the efficacy of RBC transfusion on the level of tissue oxygenation. Whether results are due to a too liberal RBC transfusion policy, or to an inability to utilize oxygen, or to a decrease in perfusion, or to another cause, cannot be dissected from our findings. In follow-up studies on the utility of mitoPO₂ to guide interventions to improve tissue oxygenation, it should be noted that signal quality is impaired in the most severely ill patients.

Conclusion

MitoPO₂ and mitoVO₂ in critically ill patients in the ICU with anemia were similar to previously observed in critically ill patients and did not significantly change during the first 24 h after RBC transfusion. MitoPO₂ and mitoVO₂ values were not notably associated with Hb concentrations, parameters of severity of illness, markers of tissue perfusion or cellular oxygenation in these moderately ill ICU patients with anemia. Given the high baseline value, it cannot be excluded nor confirmed whether RBC can improve low mitoPO₂ values.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s40635-024-00646-3>.

Additional file 1.

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Author contributions

All authors contributed to the study conception and design. Material preparation, data collection and analysis were performed by Meryem Baysan, Bashar Hilderink, Camila Caram-Deelder, and Lisa van Manen. The first draft of the manuscript was written by Meryem Baysan and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate

All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards. The study was approved by the Bioethics Committee of the Medical University of Leiden, the Hague and Delft (P16.303).

Consent for publication

Informed consent was obtained from all individual participants included in the study. However, in case of patients not able to consent, informed consent was obtained from their legal representatives. From February 2019 onwards, deferred consent procedure was used during the study due to logistical problems regarding informed consent for inclusion.

Competing interests

EG Mik is founder and shareholder of Photonic Healthcare. NP Juffermans is editor-in-chief for *Intensive Care Medicine Experimental*. The other authors have no conflicts of interest to declare.

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