

Monitoring immune responsiveness: novel assays to explore immune system dynamics in health and disease

Veld, A.E. in 't

Citation

Veld, A. E. in 't. (2024, November 14). *Monitoring immune responsiveness: novel assays to explore immune system dynamics in health and disease*. Retrieved from https://hdl.handle.net/1887/4109182

Version: Publisher's Version License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](https://hdl.handle.net/1887/license:5) Downloaded from: <https://hdl.handle.net/1887/4109182>

Note: To cite this publication please use the final published version (if applicable).

Chapter 6 Hydroxychloroquine effects on TLR signalling: underexposed but unneglectable in COVID-19 Journal of Immunology Research. 2021 Mar 9, 2021:6659410. Doi: 10.1155/2021/6659410

Aliede E. in 't Veld 1,2 , Manon A.A. Jansen 1 , Luuk C.A. Ciere 1 , Matthijs Moerland 1,2

Abstract

The main basis for hydroxychloroquine (HCQ) treatment in HCQ is the compound's ability to inhibit viral replication in vitro. HCQ also suppresses immunity, mainly by interference in TLR signalling, but reliable clinical data on the extent and nature of HCQ-induced immunosuppression are lacking. Here we discuss the mechanistic basis for the use of HCQ against HCQ in a prophylactic setting and in a therapeutic setting, at different stages of the disease. We argue that the clinical effect of prophylactic or therapeutic HCQ treatment in HCQ depends on the balance between inhibition of viral replication, immunosuppression, and off-target side effects, and that the outcome is probably dependent on disease stage and disease severity. This is supported by the initial outcomes of the well-designed randomized controlled trials: so far evidence for a beneficial effect of HCQ treatment for HCQ is weak and conflicting.

Introduction

Hydroxychloroquine sulfate (HCQ, Figure 1) is a less toxic derivative of the antimalarial drug chloroquine (CQ). Besides the use as antimalarial drug, HCQ is also prescribed for the treatment of several different auto-immune diseases such as rheumatoid arthritis, juvenile idiopathic arthritis, and systemic lupus erythematosus. The compound has been evaluated extensively in an ever-increasing number of clinical trials as treatment modality to fight HCQ infection, also in a prophylactic setting.

Figure 1 Chemical structure of hydroxychloroquine sulfate

The insight we are aiming to provide in this paper is whether the effects of HCQ use on HCQ infection align with the predicted effects of HCQ. Can the molecular activities of the drug, in particular its direct immunosuppressive activities, predict the effect on HCQ infection? We advocate that especially these immunosuppressive effects ultimately determine the clinical outcome, while so far they have remained largely underexposed in clinical trials evaluating HCQ effects on HCQ.

The main reason why HCQ initially emerged as potential treatment in HCQ was because of its in vitro antiviral properties against several RNA viruses, including SARS-CoV-1 and -2.1-4 In addition, HCQ possesses immunosuppressive properties that may be beneficial in dampening the derailed immune response in later stages of HCQ infection.5 Based on these pharmacological activities, HCQ was considered to be a promising drug to combat HCQ, at least until the moment an effective vaccine would become available. In spring 2020, this even urged some governments to recommend prophylactic HCQ use, for example by the Indian Council of Medicinal Research⁶ and repeatedly by President Trump in White House briefings. This was remarkable, because at that moment in time conclusive data from large, randomized and well-monitored clinical trials on the preventive or therapeutic efficacy of HCQ in HCQ were pending. The outcomes of initial clinical studies evaluating HCQ effects in HCQ were not convincing, also because many studies suffered from major methodological limitations and decent peer review of study reports was complicated by time constraints. This has been extensively outlined in earlier reviews⁷ and was breaking news when two papers in The Lancet and New England Journal of Medicine were retracted.8-9 Six months later, the results of the first randomized controlled trials have been published, and overall they are disappointing. So far, there is no convincing proof for beneficial effects of HCQ, neither in a therapeutic setting nor in a postexposure prophylactic setting.10

A complicating factor for the evaluation of HCQ's effects on HCQ has been the highly variable pathophysiology, within an individual patient over time, but also between patients.¹¹ HCQ's inhibiting effect on HCQ replication, based on in vitro evidence, would be beneficial at any stage of the disease, in any population (being it non-infected subjects, asymptomatic patients, or severe patients). However, this is not equally self-evident for the compound's immunosuppressive effects, as we will outline later in this

manuscript. Importantly, despite extensive mechanistic evidence based on in vitro experiments, reliable clinical data on the extent and nature of HCQinduced immunosuppression are lacking.

This article discusses the mechanistic basis for the use of HCQ against HCQ in a prophylactic setting and in a therapeutic setting, at different stages of the disease. The focus lies on HCQ's immunosuppressive effects, since we advocate that especially this aspect is largely underexposed in recent clinical trials evaluating HCQ effects on HCQ. A non-systematic review of published literature was performed, mainly PubMed-based, to build this mechanistic basis. This article only discusses HCQ, since this compound suffers less from side effects, drug-drug interactions, and toxicity than its parent compound chloroquine, while their pharmacological activities are well comparable.¹²

Immunosuppressive effects of HCQ

The basis for HCQ's use in autoimmune diseases is its wide range of immunosuppressive properties (Figure 2). HCQ accumulates in the lysosomes where it increases the pH and inhibits the enzymatic activity in both lysosomes and autophagosomes. Since these organelles play an important role in antigen processing and MHC class II presentation, a rise in lysosomal pH indirectly inhibits the immune response to both intracellular and extracellular antigens.¹³

Lysosomal accumulation of HCQ does not only result in a pH increase, but also directly affects endosomal TLR signalling triggered by nucleic acids. The endosomal TLRs (i.e. TLR3, TLR7, TLR8 and TLR9) play an important role in the innate immune response by recognizing double-stranded RNA, single-stranded RNA and CpG motifs in viral DNA. ¹⁴ HCQ can bind nucleic acids within the endosome, thereby preventing interaction of the endosomal TLRs with their ligands, inhibiting subsequent TLR activation. Downstream innate immune responses are dampened, such as IFN- and TNF production by plasmacytoid dendritic cells.¹⁵⁻¹⁶ In addition, the adaptive immune response is impaired by HCQ effects on B cell differentiation and cytokine production.17-18 Moreover, HCQ inhibits T cell activation, proliferation and cytokine production by inhibiting intracellular calcium and mobilization and subsequent NFAT signalling¹⁹⁻²⁰, and apoptosis in CD45RO+ memory and effector T cells by inhibiting autophagy.²¹

Figure 2 Immunosuppressive effects of HCQ. Hydroxychloroquine affects both the innate and adaptive immune system. By accumulating in the lysosome and autophagosome, the pH is increased causing an inhibition of MHC-II antigen presentation and subsequent T cell activation. In addition, HCQ accumulation abrogates viral recognition by endosomal TLRs, resulting in a decrease of the anti-viral innate immune response (i.a. IFN-I production). Moreover, HCQ can also directly affect the adaptive immune system through inhibition of T and B cell differentiation and activation.

The majority of the mechanistic work on HCQ's immunosuppressive activity has been performed in cell lines. Experimental evidence for immune suppression by HCQ in primary human cells is scarce. Some publications are available describing HCQ effects on innate immune responses in human whole blood, peripheral blood mononuclear cells, or T cells, with TLR-mediated cytokine production, or T cell activation and proliferation as endpoint.²²⁻²⁷ Most experiments used HCQ concentrations largely exceeding expected circulating concentrations in vivo after prophylactic or therapeutic dosing. Moreover, with one exception, none of the papers provides a decent HCQ concentrationeffect relationship, so an IC50 for HCQ's immunosuppressive activities cannot be estimated. Interestingly, HCQ's IC50 for inhibition of

HCQ replication $(4-17 \mu M)^{28}$ appears to exceed HCQ concentrations effectively inhibiting TLR responses in vitro $(3 \mu M)^{24,27}$, which means that it will be difficult to inhibit viral replication without impairing the immune system.

Mechanistic support for HCQ use in COVID-19 PROPHYLACTIC SETTING

Cell entry by HCQ is thought to be similar to SARS-CoV entry, being mediated by spike (S) protein binding to angiotensin-converting enzyme 2 (ACE2).29- ³⁰ In silico predictions showed that HCQ prevents the cellular binding and entering of HCQ virus particles, by interfering with sialic acids and surface gangliosides.31 Based on this pharmacological activity, prophylactic HCQ treatment could theoretically be beneficial and prevent HCQ infection in vulnerable populations or populations professionally exposed to HCQ patients.

Upon cell entry, HCQ is likely recognized by TLR3, TLR4, TLR7, TLR8 and RIG-132, resulting in a type I IFN response which is crucial for an efficient adaptive antiviral response.33 HCQ suppresses parts of the immune system that are essential in fighting infections, including TLR signalling and type I IFN production. In previous SARS-CoV and MERS-CoV outbreaks, downregulation of IFNs by coronavirus proteins strongly correlated with worse disease progression and increased lethality.34 Cell and animal models of HCQ infection, and transcriptional and serum profiling of HCQ patients, revealed an imbalanced host response with low levels of type I and III IFNs.35 Early IFN signalling was protective in SARS-CoV-1 infected mice, whereas delayed IFN signalling was detrimental leading to severe disease progression and related lethal pneumonia.³⁶

The importance of TLR signalling in viral defence has been well established in SARS-CoV-1 mouse models. Both TLR3 and TLR4 deficient mice are more susceptible to SARS-CoV-1 infections.37 Murine MyD88 or TRIF deficiency, which are downstream signalling molecules shared by multiple TLRs, resulted in a mortality rate of over 90% upon experimental infection with SARS-COV-1, which is usually non-lethal in immunocompetent mice.³⁷⁻ ³⁸ HCQ abrogates endosomal acidification thereby reducing endosomal TLR activation²², but interestingly enough data confirming this HCQ effect on endosomal TLRs in primary human cells are scarce. Since the relationship between HCQ dose/concentration and level of immunosuppression remains largely unexplored in primary human immune cells, it is difficult to

estimate the effect of prophylactic HCQ treatment regimens on the innate immune response. If HCQ's immunosuppressive IC50s would fall in the concentration range reached after prophylactic HCQ treatment, endosomal TLR responses, type I IFN production, and T and B cell activation and proliferation could be impaired in vivo. Theoretically this could result in an increased viral infection risk, including HCQ infection. On the other hand, HCQ use in rheumatoid arthritis patients is not associated with an increased infection risk.39-40 So far, prophylactic HCQ studies did not show clinical benefit of HCQ administration.41-42

Next to mechanistic arguments, the fact that long-term HCQ use comes with side-effects further fuels doubts about prophylactic use of HCQ. Retinal toxicity, cardiac disease, (reversible) neuromyopathy, dermatological manifestations, gastrointestinal and hematological changes, and hearing abnormalities have been reported upon long-term HCQ treatment, amongst others.43-45 Such side effects could be avoided by local HCQ administration, for example by inhalation.

THERAPEUTIC SETTING

Although our understanding of the pathophysiology continues to increase on a daily basis, it is clear that HCQ is a highly heterogenous disease. With increased disease severity, the complexity of the pathophysiology grows.32,46 Since many excellent reviews are available in the public domain, this manuscript does not revisit HCQ pathophysiology and disease progression. Instead, it discusses the alignment between HCQ's mechanism of action and disease stage: how could specific pharmacological activities of HCQ theoretically affect HCQ's pathophysiology at a particular disease stage? As guidance, the disease progression has been separated into three stages: stage 1 - virus entry and replication in the airway cells (day 0-2), stage 2 - activation of innate immunity in the lung (maladaptive inflammatory response, day 3-7), and stage 3 - acute respiratory distress syndrome (ARDS, >day 7).47 Obviously, the clinical presentation of HCQ varies between patients from asymptomatic to mild, moderate and severe, and not all patients develop advanced disease stages.

When discussing potential effects of HCQ treatment in a therapeutic setting, most papers focus on the off-target side effects of HCQ, specifically potentially severe cardiac disorders such as or segment prolongation. However, safety concerns related to the short-term use of HCQ (i.e. regimens of 1 month) are probably limited, as demonstrated by a recently

published (non-peer reviewed) international study in more than 900,000 HCQ-treated patients.48 We advocate that one of HCQ's pharmacological activities, namely its immunosuppressive effect, is critical when considering HCQ as potential treatment modality for HCQ. Surprisingly, HCQ's exact molecular mechanism of action has remained largely neglected in considerations on therapeutic HCQ use for HCQ. Therefore, we discuss in the next sections how HCQ's pharmacological activities could be beneficial, or detrimental, at different disease stages (stage 1-3, see above) and in different disease severities (asymptomatic, mild, moderate-severe) (Figure 3).

Figure 3 Theoretical effects of HCQ at different stages of SARS-COV-2 infection. Potential HCQ effects on COVID-19 are schematically presented over the course of the disease, ranging from prophylactic use in uninfected subjects to therapeutic use in acute respiratory distress syndrome (ARDS) in severe patients. A beneficial HCQ effect is indicated with '+' and a detrimental HCQ effect with '-'. The stages of SARS-CoV-2 infection are indicated in green. Stage 0 – no infection, stage 1 – virus entry and replication in the airway cells, stage 2 – activation of innate and adaptive immune system, stage 3 – ARDS.

For therapeutic treatment, the first stage (day 0-2 of infection) is irrelevant, since patients are asymptomatic and viral titers may be low⁴⁹, so patients in this stage of the disease are untreated or fall in the prophylactic treatment category (see previous section). HCQ treatment theoretically could be beneficial in the next stages of the disease (stage 2; day 3-7, and stage 3; >day 7), when the innate immune response in the lungs starts to evolve, and ultimately culminates in respiratory impairment and multi-organ failure. The drug may not only inhibit virus replication, but also suppress TLR-mediated cytokine responses and over-activation and apoptosis of lymphocytes, processes that are observed in severe HCQ. 50-51 Especially prevention of a cytokine storm is critical since this is a major factor driving multi-organ failure, ARDS, disseminated intravascular coagulation, and the resulting high mortality. Taken together, HCQ treatment in progressed HCQ is mechanistically supported by HCQ's pharmacological activities.

Obviously, progressed disease as outlined above (stage 2 and 3) only applies to moderate to severe HCQ patients. The large majority of HCQ patients only suffers from mild disease, or even remains asymptomatic.52 These patients have a low viral load, develop an efficient type I IFN response, produce virus-neutralizing antibodies, and do not develop a maladaptive inflammatory response.53 Since it is especially the latter response that could be targeted by HCQ's immunosuppressive activity, the question arises whether HCQ treatment is rational in asymptomatic or mild patients. On one hand, one could argue that HCQ-dependent inhibition of viral replication (though not clinically proven) is important, independent of disease stage. Moreover, HCQ-dependent immunosuppression may prevent mild disease turning into inflammation-driven moderate/severe disease. On the other hand, in the early disease stage it is important that the virus-specific anti-HCQ response is driven by an efficient antiviral innate immune response, and especially this response may be significantly impaired upon HCQ treatment. The net result of HCQ treatment will depend on the balance between these two pharmacological activities. The outcome of therapeutic studies have shown that HCQ treatment overall does not seem to reduce mortality, improve clinical scores, or suppress viral load in moderate to severe HCQ patients.54-56 However, low dose HCQ treatment (< 2.5 g in total) was associated with a reduced risk of intensive care unit admission and lower mortality rates.57-58 HCQ's clinical beneficial effects may depend on the inflammatory status of the patient: chronic low-dose HCQ treatment of a large cohort of rheumatic patients coincided with reduced mortality following HCQ infection59, and another study reported a therapeutic benefit of HCQ treatment in patients with elevated C-reactive protein levels.⁶⁰ These reports are mechanistically in line with the immunosuppressive activities of HCQ, as outlined above.

Conclusion

Immunosuppression by HCQ, via interference in endosomal TLR signalling, has remained largely underexposed in the public debate, while it may be a critical factor for the (lack of?) clinical efficacy of HCQ in HCQ. Experimental evidence for immune suppression by HCQ in primary human cells is scarce,

REFERENCES

- Savarino, A.; Boelaert, J. R.; Cassone, A.; Majori, G.; Cauda, R., Effects of chloroquine on viral infections: an old drug against today's diseases? The Lancet. Infectious diseases 2003, 3 (11), 722-7.
- 2 Mele, T. S.; Halloran, P. F., The use of mycophenolate mofetil in transplant recipients. Immunopharmacology 2000, 47 (2-3), 215-45.
- 3 Keyaerts, E.; Vijgen, L.; Maes, P.; Neyts, J.; Van Ranst, M., In vitro inhibition of severe acute respiratory syndrome coronavirus by chloroquine. Biochemical and biophysical research communications 2004, 323 $(1), 264-8.$
- 4 Vincent, M. J.; Bergeron, E.; Benjannet, S.; Erickson, B. R.; Rollin, P. E.; Ksiazek, T. G.; Seidah, N. G.; Nichol, S. T., Chloroquine is a potent inhibitor of SARS coronavirus infection and spread. Virology journal 2005, 2, 69.
- 5 Yao, X.; Ye, F.; Zhang, M.; Cui, C.; Huang, B.; Niu, P.; Liu, X.; Zhao, L.; Dong, E.; Song, C.; Zhan, S.; Lu, R.; Li, H.; Tan, W.; Liu, D., In Vitro Antiviral Activity and Projection of Optimized Dosing Design of Hydroxychloroquine for the Treatment of Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2). Clinical infectious diseases : an official publication of the Infectious Diseases Society of America 2020.
- 6 Indian National Taskforce for COVID19, Advisory on the use of hydroxy-chloroquine as prophylaxis for SARS-CoV-2 infection. March 21, 2020.
- Das, S.; Bhowmick, S.; Tiwari, S.; Sen, S., An Updated Systematic Review of the Therapeutic Role of Hydroxychloroquine in Coronavirus Disease-19 (COVID-19). Clinical drug investigation 2020, 1-11.
- 8 Mehra, M. R.; Ruschitzka, F.; Patel, A. N., Retraction-21 Hydroxychloroquine or chloroquine with or without a macrolide for treatment of COVID-19: a multinational registry analysis. The Lancet 2020.
- 9 Mehra, M. R.; Desai, S. S.; Kuy, S.; Henry, T. D.; Patel, A. N., Retraction: Cardiovascular Disease, Drug Therapy, and Mortality in Covid-19. N Engl J Med. DOI: 10.1056/NEJMoa2007621. The New England journal of medicine 2020.
- 10 Rughiniş, C.; Dima, L.; Vasile, S., Hydroxychloroquine and COVID-19: Lack of Efficacy and the Social Construction of Plausibility. American journal of therapeutics 2020, 27 (6), e573-e583.
- 11 Zhang, X.; Tan, Y.; Ling, Y.; Lu, G.; Liu, F.; Yi, Z.; Jia, X.; Wu, M.; Shi, B.; Xu, S.; Chen, J.; Wang, W.; Chen, B.; Jiang, L.; Yu, S.; Lu, J.; Wang, J.; Xu, M.; Yuan, Z.; Zhang, Q.; Zhang, X.; Zhao, G.; Wang, S.; Chen, S.; Lu, H., Viral and host factors related to the clinical outcome of COVID-19. Nature 2020.
- 12 Gasmi, A.; Peana, M.; Noor, S.; Lysiuk, R.; Menzel, A.; Gasmi Benahmed, A.; Bjørklund, G., Chloroquine and hydroxychloroquine in the treatment of COVID-19: the never-ending story. Applied microbiology and biotechnology 2021, 105 (4), 1333-1343.
- 13 Schrezenmeier, E.; Dörner, T., Mechanisms of action of hydroxychloroquine and chloroquine: implications for rheumatology. Nature reviews. Rheumatology 2020, 16 (3), 155-166.
- 14 Kawasaki, T.; Kawai, T., Toll-like receptor signaling pathways. Frontiers in immunology 2014, 5, 461.
- 15 Sacre, K.; Criswell, L. A.; McCune, J. M., Hydroxychloroquine is associated with impaired interferon-alpha and tumor necrosis factor-alpha production by plasmacytoid dendritic cells in

systemic lupus erythematosus. Arthritis research & therapy 2012, 14 (3), R155.

- 16 Bodewes, I. L. A.; Gottenberg, J.-E.; van Helden-Meeuwsen, C. G.; Mariette, X.; Versnel, M. A., Hydroxychloroquine treatment downregulates systemic interferon activation in primary Sjögren's syndrome in the JOQUER randomized trial. Rheumatology 2019, 59 (1), 107-111.
- 17 Torigoe, M.; Sakata, K.; Ishii, A.; Iwata, S.; Nakayamada, S.; Tanaka, Y., Hydroxychloroquine efficiently suppresses inflammatory responses of human class-switched memory B cells via Toll-like receptor 9 inhibition. Clinical immunology (Orlando, Fla.) 2018, 195, 1-7.
- 18 Brauner, S.; Folkersen, L.; Kvarnström, M.; Meisgen, S.; Petersen, S.; Franzén-Malmros, M.; Mofors, J.; Brokstad, K. A.; Klareskog, L.; Jonsson, R.; Westerberg, L. S.; Trollmo, C.; Malmström, V.; Ambrosi, A.; Kuchroo, V. K.; Nordmark, G.; Wahren-Herlenius, M., H1N1 vaccination in Sjögren's syndrome triggers polyclonal B cell activation and promotes autoantibody production. Annals of the rheumatic diseases 2017, 76 (10), 1755-1763.
- 19 Goldman, F. D.; Gilman, A. L.; Hollenback, C.; Kato, R. M.; Premack, B. A.; Rawlings, D. J., Hydroxychloroquine inhibits calcium signals in T cells: a new mechanism to explain its immunomodulatory properties. Blood 2000, 95 (11), 3460-6.
- 20 Wu, S. F.; Chang, C. B.; Hsu, J. M.; Lu, M. C.; Lai, N. S.; Li, C.; Tung, C. H., Hydroxychloroquine inhibits CD154 expression in CD4(+) T lymphocytes of systemic lupus erythematosus through NFAT, but not STAT5, signaling. Arthritis research & therapy 2017, 19 (1), 183.
- van Loosdregt, J.; Spreafico, R.; Rossetti, M.; Prakken, B. J.; Lotz, M.; Albani, S., Hydroxychloroquine preferentially induces apoptosis of CD45RO+ effector T cells by inhibiting autophagy: a possible mechanism for therapeutic modulation of T cells. The Journal of allergy and clinical immunology 2013, 131 (5), 1443-6. $_{\mathsf{P1}}$
- 22 Kuznik, A.; Bencina, M.; Svajger, U.; Jeras, M.; Rozman, B.; Jerala, R., Mechanism of endosomal TLR inhibition by antimalarial drugs and imidazoquinolines. Journal of immunology (Baltimore, Md. : 1950) 2011, 186 (8), 4794-804.
- 23 Gardet, A.; Pellerin, A.; McCarl, C. A.; Diwanji, R.; Wang, W.; Donaldson, D.; Franchimont, N.; Werth, V. P.; Rabah, D., Effect of in vivo Hydroxychloroquine and ex vivo Anti-BDCA2 mAb Treatment on pDC IFN Production From Patients Affected With Cutaneous Lupus Erythematosus. Frontiers in immunology 2019, 10, 275.
- 24 Zeidi, M.; Kim, H. J.; Werth, V. P., Increased Myeloid Dendritic Cells and TNF- Expression Predicts Poor Response to Hydroxychloroquine in Cutaneous Lupus Erythematosus. The Journal of investigative dermatology 2019, 139 (2), 324-332.
- 25 Silva, J. C.; Mariz, H. A.; Rocha, L. F., Jr.; Oliveira, P. S.; Dantas, A. T.; Duarte, A. L.; Pitta Ida, R.; Galdino, S. L.; Pitta, M. G., Hydroxychloroquine decreases Th17 related cytokines in systemic lupus erythematosus and rheumatoid arthritis patients. Clinics (Sao Paulo, Brazil) 2013, 68 (6), 766-71.
- 26 Schmidt, R. L.; Jutz, S.; Goldhahn, K.; Witzeneder, N.; Gerner, M. C.; Trapin, D.; Greiner, G.; Hoermann, G.; Steiner, G.; Pickl, W. F.; Burgmann, H.; Steinberger, P.; Ratzinger, F.; Schmetterer, K. G., Chloroquine inhibits

which is surprising for such an old drug. Clinical trials evaluating HCQ as HCQ treatment did not include readout measures to study this immunosup pressive effect of HCQ. As a result, the extent of immunosuppression by HCQ cannot be reliably estimated in vivo. If systemic or local HCQ concentrations would be sufficiently high to suppress key components of the innate immune response, this could translate into a clinical benefit. The other side of the coin is that in mild HCQ patients, or in a prophylactic setting, immunosuppression by HCQ could have a detrimental effect, since an efficient virus-specific an ti-HCQ response depends on a robust antiviral innate immune response. We argue that ultimately the clinical effect of HCQ treatment in HCQ depends on the balance between inhibition of viral replication, immunosuppression, and off-target side effects (which have been extensively evaluated recently, within and outside the setting of HCQ treatment or prevention $61-62$, and are as such not the topic of this article). The outcome of this balance is probably dependent on disease stage and disease severity (**Figure 3**). This is supported by the initial outcomes of the well-designed randomized controlled tri als: so far evidence for a beneficial effect of HCQ treatment for HCQ is weak and conflicting.

human CD4(+) T-cell activation by AP-1 signaling modulation. Scientific reports 2017, 7, 42191.

- 27 Alves, P.; Bashir, M. M.; Wysocka, M.; Zeidi, M.; Feng, R.; Werth, V. P., Quinacrine Suppresses Tumor Necrosis Factor- and IFN- in Dermatomyositis and Cutaneous Lupus Erythematosus. The journal of investigative dermatology. Symposium proceedings 2017, 18 (2), S57-s63.
- 28 Liu, J.; Cao, R.; Xu, M.; Wang, X.; Zhang, H.; Hu, H.; Li, Y.; Hu, Z.; Zhong, W.; Wang, M., Hydroxychloroquine, a less toxic derivative of chloroquine, is effective in inhibiting SARS-CoV-2 infection in vitro. Cell discovery 2020, 6, 16.
- 29 Hoffmann, M.; Kleine-Weber, H.; Schroeder, S.; Krüger, N.; Herrler, T.; Erichsen, S.; Schiergens, T. S.; Herrler, G.; Wu, N. H.; Nitsche, A.; Müller, M. A.; Drosten, C.; Pöhlmann, S., SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. Cell 2020, 181 (2), 271-280.e8.
- 30 Li, W.; Moore, M. J.; Vasilieva, N.; Sui, J.; Wong, S. K.; Berne, M. A.; Somasundaran, M.; Sullivan, J. L.; Luzuriaga, K.; Greenough, T. C.; Choe, H.; Farzan, M., Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. Nature 2003, 426 (6965), 450-4.
- 31 Fantini, J.; Di Scala, C.; Chahinian, H.; Yahi, N., Structural and molecular modelling studies reveal a new mechanism of action of chloroquine and hydroxychloroquine against SARS-CoV-2 infection. International journal of antimicrobial agents 2020, 55 (5), 105960.
- 32 Vabret, N.; Britton, G. J.; Gruber, C.; Hegde, S.; Kim, J.; Kuksin, M.; Levantovsky, R.; Malle, L.; Moreira, A.; Park, M. D.; Pia, L.; Risson, E.; Saffern, M.; Salomé, B.; Selvan, M. E.; Spindler, M. P.; Tan, J.; van der Heide, V.; Gregory, J. K.; Alexandropoulos, K.; Bhardwaj, N.; Brown, B. D.; Greenbaum, B.; Gümüş, Z. H.; Homann, D.; Horowitz, A.; Kamphorst, A. O.; Curotto de Lafaille, M. A.; Mehandru, S.; Merad, M.; Samstein, R. 44 Kwon, J. B.; Kleiner, A.; Ishida, K.; Godown, J.; M.; The Sinai Immunology Review, P., Immunology of COVID-19: current state of the science. Immunity 2020.
- 33 Samuel, C. E., Antiviral actions of interferons. Clinical microbiology reviews 2001, 14 (4), 778-809, table of contents.
- 34 de Wit, E.; van Doremalen, N.; Falzarano, D.; Munster, V. J., SARS and MERS: recent insights into emerging coronaviruses. Nature Reviews Microbiology 2016, 14 46 Azkur, A. K.; Akdis, M.; Azkur, D.; Sokolowska, M.; (8), 523-534.
- 35 Blanco-Melo, D.; Nilsson-Payant, B. E.; Liu, W. C.; Uhl, S.; Hoagland, D.; Møller, R.; Jordan, T. X.; Oishi, K.; Panis, M.; Sachs, D.; Wang, T. T.; Schwartz, R. E.; Lim, J. K.; Albrecht, R. A.; tenOever, B. R., Imbalanced 47 Host Response to SARS-CoV-2 Drives Development of COVID-19. Cell 2020, 181 (5), 1036-1045.e9.
- 36 Channappanavar, R.; Fehr, A. R.; Vijay, R.; Mack, M.; Zhao, J.; Meyerholz, D. K.; Perlman, S., Dysregulated Type I Interferon and Inflammatory Monocyte-Macrophage Responses Cause Lethal Pneumonia in SARS-CoV-Infected Mice. Cell host & microbe 2016, 19 (2), 181-93.
- 37 Totura, A. L.; Whitmore, A.; Agnihothram, S.; Schäfer, A.; Katze, M. G.; Heise, M. T.; Baric, R. S., Toll-Like Receptor 3 Signaling via TRIF Contributes to a Protective Innate Immune Response to Severe Acute Respiratory Syndrome Coronavirus Infection. mBio 2015, 6 (3), e00638-15.
- 38 Sheahan, T.; Morrison, T. E.; Funkhouser, W.;

Uematsu, S.; Akira, S.; Baric, R. S.; Heise, M. T., MyD88 is required for protection from lethal infection with a mouse-adapted SARS-CoV. PLoS pathogens 2008, 4 (12), e1000240.

- 39 Wolfe, F.; Caplan, L.; Michaud, K., Treatment for rheumatoid arthritis and the risk of hospitalization for pneumonia: associations with prednisone, disease-modifying antirheumatic drugs, and anti-tumor necrosis factor therapy. Arthritis and rheumatism 2006, 54 (2), 628-34.
- 40 Bernatsky, S.; Hudson, M.; Suissa, S., Anti-rheumatic drug use and risk of serious infections in rheumatoid arthritis. Rheumatology (Oxford, England) 2007, 46 (7), 1157-60.
- 41 Abella, B. S.; Jolkovsky, E. L.; Biney, B. T.; Uspal, J. E.; Hyman, M. C.; Frank, I.; Hensley, S. E.; Gill, S.; Vogl, D. T.; Maillard, I.; Babushok, D. V.; Huang, A. C.; Nasta, S. D.; Walsh, J. C.; Wiletyo, E. P.; Gimotty, P. A.; Milone, M. C.; Amaravadi, R. K., Efficacy and Safety of Hydroxychloroquine vs Placebo for Pre-exposure SARS-CoV-2 Prophylaxis Among Health Care Workers: A Randomized Clinical Trial. JAMA internal medicine 2020.
- 42 Boulware, D. R.; Pullen, M. F.; Bangdiwala, A. S.; Pastick, K. A.; Lofgren, S. M.; Okafor, E. C.; Skipper, C. P.; Nascene, A. A.; Nicol, M. R.; Abassi, M.; Engen, N. W.; Cheng, M. P.; LaBar, D.; Lother, S. A.; MacKenzie, L. J.; Drobot, G.; Marten, N.; Zarychanski, R.; Kelly, L. E.; Schwartz, I. S.; McDonald, E. G.; Rajasingham, R.; Lee, T. C.; Hullsiek, K. H., A Randomized Trial of Hydroxychloroquine as Postexposure Prophylaxis for Covid-19. The New England journal of medicine 2020, 383 (6), 517-525.
- Yogasundaram, H.; Putko, B. N.; Tien, J.; Paterson, D. I.; Cujec, B.; Ringrose, J.; Oudit, G. Y., Hydroxychloroquine-Induced Cardiomyopathy: Case Report, Pathophysiology, Diagnosis, and Treatment. Canadian Journal of Cardiology 2014, 30 (12), 1706-1715.
- Ciafaloni, E.; Looney, R. J., Jr., Hydroxychloroquineinduced myopathy. Journal of clinical rheumatology : practical reports on rheumatic & musculoskeletal diseases 2010, 16 (1), 28-31.
- 45 Ben-Zvi, I.; Kivity, S.; Langevitz, P.; Shoenfeld, Y., Hydroxychloroquine: from malaria to autoimmunity. Clinical reviews in allergy & immunology 2012, 42 (2), 145-53.
	- van de Veen, W.; Brüggen, M. C.; O'Mahony, L.; Gao, Y.; Nadeau, K.; Akdis, C. A., Immune response to SARS-CoV-2 and mechanisms of immunopathological changes in COVID-19. Allergy 2020.
- Risitano, A. M.; Mastellos, D. C.; Huber-Lang, M.; Yancopoulou, D.; Garlanda, C.; Ciceri, F.; Lambris, J. D., Complement as a target in COVID-19? Nature reviews. Immunology 2020, 20 (6), 343-344.
- 48 Lane, J. C. E.; Weaver, J.; Kostka, K.; Duarte-Salles, T.; Abrahao, M. T. F.; Alghoul, H.; Alser, O.; Alshammari, T. M.; Biedermann, P.; Burn, E.; Casajust, P.; Conover, M.; Culhane, A. C.; Davydov, A.; DuVall, S. L.; Dymshyts, D.; Fernández Bertolín, S.; Fišter, K.; Hardin, J.; Hester, L.; Hripcsak, G.; Kent, S.; Khosla, S.; Kolovos, S.; Lambert, C. G.; ver der Lei, J.; Lynch, K. E.; Makadia, R.; Margulis, A. V.; Matheny, M. E.; Mehta, P.; Morales, D. R.; Morgan-Stewart, H.; Mosseveld, M.; Newby, D.; Nyberg, F.; Ostropolets, A.; Park, R. W.; Prats-Uribe, A.; Rao, G. A.; Reich, C.; Reps, J.; Rijnbeek, P.; Sathappan, S. M.

K.; Schuemie, M.; Seager, S.; Sena, A.; Shoaibi, A.; Spotnitz, M.; Suchard, M. A.; Swerdel, J.; Torre, C. O.; Vizcaya, D.; Wen, H.; de Wilde, M.; You, S. C.; Zhang, L.; Zhuk, O.; Ryan, P.; Prieto-Alhambra, D., Safety of hydroxychloroquine, alone and in combination with azithromycin, in light of rapid wide-spread use for COVID-19: a multinational, network cohort and self-controlled case series study. medRxiv 2020, 2020.04.08.20054551.

- 49 Sethuraman, N.; Jeremiah, S. S.; Ryo, A., Interpreting Diagnostic Tests for SARS-CoV-2. Jama 2020.
- 50 Wang, F.; Nie, J.; Wang, H.; Zhao, Q.; Xiong, Y.; Deng, L.; Song, S.; Ma, Z.; Mo, P.; Zhang, Y., Characteristics of Peripheral Lymphocyte Subset Alteration in COVID-19 Pneumonia. J Infect Dis 2020, 221 (11), 1762-1769.
- 51 Prompetchara, E.; Ketloy, C.; Palaga, T., Immune responses in COVID-19 and potential vaccines: Lessons learned from SARS and MERS epidemic. Asian Pacific journal of allergy and immunology 2020, $38(1), 1-9.$
- 52 World Health Organization, Report of the WHO-China Joint Mission on Coronavirus Disease 2019 (COVID-19). 2020.
- 53 Wang, F.; Hou, H.; Luo, Y.; Tang, G.; Wu, S.; Huang, M.; Liu, W.; Zhu, Y.; Lin, Q.; Mao, L.; Fang, M.; Zhang, H.; Sun, Z., The laboratory tests and host immunity of COVID-19 patients with different severity of illness. JCI insight 2020, 5 (10).
- 54 Ulrich, R. J.; Troxel, A. B.; Carmody, E.; Eapen, J.; Bäcker, M.; DeHovitz, J. A.; Prasad, P. J.; Li, Y.; Delgado, C.; Jrada, M.; Robbins, G. A.; Henderson, B.; Hrycko, A.; Delpachitra, D.; Raabe, V.; Austrian, J. S.; Dubrovskaya, Y.; Mulligan, M. J., Treating COVID-19 With Hydroxychloroquine (TEACH): A Multicenter, Double-Blind Randomized Controlled Trial in Hospitalized Patients. Open Forum Infectious Diseases 2020, 7 (10), ofaa446.
- 55 Fiolet, T.; Guihur, A.; Rebeaud, M. E.; Mulot, M.; Peiffer-Smadja, N.; Mahamat-Saleh, Y., Effect of hydroxychloroquine with or without azithromycin on the mortality of coronavirus disease 2019 (COVID-19) patients: a systematic review and meta-analysis. Clinical microbiology and infection : the official publication of the European Society of Clinical Microbiology and Infectious Diseases 2020.
- 56 Faíco-Filho, K. S.; Conte, D. D.; de Souza Luna, L. K.; Carvalho, J. M. A.; Perosa, A. H. S.; Bellei, N., No benefit of hydroxychloroquine on SARS-CoV-2 viral load reduction in non-critical hospitalized patients with COVID-19. Brazilian journal of microbiology : [publication of the Brazilian Society for Microbiology] 2020, 1-5.
- 57 Dauby, N.; Bottieau, E., The unfinished story of hydroxychloroquine in COVID-19: the right anti-inflammatory dose at the right moment? International Journal of Infectious Diseases 2020.
- 58 Lammers, A. J. J.; Brohet, R. M.; Theunissen, R. E. P.; Koster, C.; Rood, R.; Verhagen, D. W. M.; Brinkman, K.; Hassing, R. J.; Dofferhoff, A.; el Moussaoui, R.; Hermanides, G.; Ellerbroek, J.; Bokhizzou, N.; Visser, H.; van den Berge, M.; Bax, H.; Postma, D. F.; Groeneveld, P. H. P., Early hydroxychloroquine but not chloroquine use reduces ICU admission in COVID-19 patients. International Journal of Infectious Diseases 2020, 101, 283-289.
- 59 Gentry, C. A.; Humphrey, M. B.; Thind, S. K.; Hendrickson, S. C.; Kurdgelashvili, G.; Williams, R. J., 2nd, Long-term hydroxychloroquine use in patients

with rheumatic conditions and development of SARS-CoV-2 infection: a retrospective cohort study. The Lancet Rheumatology 2020, 2 (11), e689-e697.

- 60 Castelnuovo, A. D.; Costanzo, S.; Antinori, A.; Berselli, N.; Blandi, L.; Bruno, R.; Cauda, R.; Guaraldi, G.; Menicanti, L.; My, I.; Parruti, G.; Patti, G.; Perlini, S.; Santilli, F.; Signorelli, C.; Spinoni, E.; Stefanini, G. G.; Vergori, A.; Ageno, W.; Agodi, A.; Aiello, L.; Agostoni, P.; Moghazi, S. A.; Astuto, M.; Aucella, F.; Barbieri, G.; Bartoloni, A.; Bonaccio, M.; Bonfanti, P.; Cacciatore, F.; Caiano, L.; Cannata, F.; Carrozzi, L.; Cascio, A.; Ciccullo, A.; Cingolani, A.; Cipollone, F.; Colomba, C.; Crosta, F.; Pra, C. D.; Danzi, G. B.; D'Ardes, D.; Donati, K. d. G.; Giacomo, P. D.; Gennaro, F. D.; Di Tano, G.; D'Offizi, G.; Filippini, T.; Fusco, F. M.; Gentile, I.; Gialluisi, A.; Gini, G.; Grandone, E.; Grisafi, L.; Guarnieri, G.; Lamonica, S.; Landi, F.; Leone, A.; Maccagni, G.; Maccarella, S.; Madaro, A.; Mapelli, M.; Maragna, R.; Marra, L.; Maresca, G.; Marotta, C.; Mastroianni, F.; Mazzitelli, M.; Mengozzi, A.; Menichetti, F.; Meschiari, M.; Minutolo, F.; Montineri, A.; Mussinelli, R.; Mussini, C.; Musso, M.; Odone, A.; Olivieri, M.; Pasi, E.; Petri, F.; Pinchera, B.; Pivato, C. A.; Poletti, V.; Ravaglia, C.; Rinaldi, M.; Rognoni, A.; Rossato, M.; Rossi, I.; Rossi, M.; Sabena, A.; Salinaro, F.; Sangiovanni, V.; Sanrocco, C.; Scorzolini, L.; Sgariglia, R.; Simeone, P. G.; Spinicci, M.; Trecarichi, E. M.; Venezia, A.; Veronesi, G.; Vettor, R.; Vianello, A.; Vinceti, M.; Vocciante, L.; De Caterina, R.; Iacoviello, L., Use of hydroxychloroquine in hospitalised COVID-19 patients is associated with reduced mortality: Findings from the observational multicentre Italian CORIST study. European journal of internal medicine 2020.
- 61 Maraolo, A. E.; Grossi, A., Safety of hydroxychloroquine for treatment or prevention of SARS-CoV-2 infection: A rapid systematic review and meta-analysis of randomized clinical trials. Immunity, inflammation and disease 2021, 9 (1), 31-36.
- 62 Celotto, S.; Veronese, N.; Barbagallo, M.; Ometto, F.; Smith, L.; Pardhan, S.; Barnett, Y.; Ilie, P. C.; Soysal, P.; Lagolio, E.; Kurotschka, P. K.; Tonelli, R.; Demurtas, J., An umbrella review of systematic reviews with metaanalyses evaluating positive and negative outcomes of Hydroxychloroquine and chloroquine therapy. International journal of infectious diseases : IJID : official publication of the International Society for Infectious Diseases 2021, 103, 599-606.