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Targeting MHC-I related proteins for cancer diagnosis and therapy

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Citation

Verhaar, E. R. (2024, July 4). *Targeting MHC-I related proteins for cancer diagnosis and therapy*. Retrieved from <https://hdl.handle.net/1887/3766089>

Version: Publisher's Version

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Note: To cite this publication please use the final published version (if applicable).

List of abbreviations

ACT – Adoptive cell transfer	MHC – Major histocompatibility complex
APC – Antigen presenting cell	MICA/B – MHC class I chain-related protein A/B
BRCA1/2 – Breast cancer gene 1 / 2	MYC – Myelocytomatosis oncogene
CAR – Chimeric antigen receptor	NFAT – Nuclear factor of activated T-cells
CMV – Cytomegalovirus	NFKB – Nuclear factor kappa-light-chain enhancer of activated B cells
CTL – Cytotoxic T lymphocyte	NK cell – Natural killer cell
DAP-10 – DNAX-activating protein 10	NKG2A/D – Natural killer group 2 member A/D
DC – Dendritic cell	NKR – NK receptor
ECM – Extracellular matrix	PBMC – Peripheral blood mononuclear cells
EGF – Epidermal growth factor	PI3K – Phosphoinositide 3-kinase
EGFR – Epidermal growth factor receptor	PTEN – Phosphatase and tensin homolog
ELISA – Enzyme-linked immunosorbent assay	RAS – Rat sarcoma virus
EMT – Epithelial-to-mesenchymal transition	scFv – Single-chain variable fragment
FACS – Fluorescence-activated cell sorting	SIV – Simian immunodeficiency virus
GFB2 – Growth factor receptor bound protein 2	STAT3/5 – Signal transducer and activator of transcription 3/5
GFP – Green fluorescent protein	TAM – Tumor associated macrophage
GrzB – Granzyme B	Tcm – Central memory T cell
GVHD – Graft-versus-host disease	Tem – Effector memory T cell
HcAb – Heavy chain-only antibody	TGF- β – Transforming growth factor- β
HIV – Human immunodeficiency virus	TME – Tumor microenvironment
HLA – Human leukocyte antigen	Tfh cell – Follicular helper T cell
HPV – Human papillomavirus	Th cell – T helper cell
IFN- γ – Interferon γ	TNF- α/β – Tumor necrosis factor α/β
Ig – Immunoglobulin	Treg cell – Regulatory T cell
IL – Interleukin	Tscm – Stem cell memory T cell
ITAM – Immunoglobulin transactivation motif	ULBP – UL-16 binding protein
ITIM – Immunoreceptor tyrosine-based inhibitory motif	VEGF – Vascular endothelial growth factor
KIR – Killer cell immunoglobulin-like receptor	VHH – Variable domain of HcAb
LPS – Lipopolysaccharides	WCL – Whole cell lysate
mAb – Monoclonal antibody	WNT – Wingless-related integration site
MAPK – Mitogen activated protein kinase	

Publications

Inge Ubink, **Elisha R. Verhaar**, Onno Kranenburg, and Roel Goldschmeding. A potential role for CCN2/CTGF in aggressive colorectal cancer. *Journal of Cell Communication and Signaling*. 2016;10(3). DOI: [10.1007/s12079-016-0347-5](https://doi.org/10.1007/s12079-016-0347-5)

Elisha R. Verhaar, Andrew Woodham, and Hidde L. Ploegh. Nanobodies in Cancer. *Seminars in Immunology*. 2020;52(363):101425. DOI: [10.1016/j.smim.2020.101425](https://doi.org/10.1016/j.smim.2020.101425)

Novalia Pishesha, Thibault J. Harmand, Paul W. Rothlauf, Patrique Praest, Ryan K. Alexander, Renate van den Doel, Mariel J. Liebeskind, Maria A. Vakaki, Nicholas McCaul, Charlotte Wijne, **Elisha R. Verhaar**, William Pinney III, Hailey Heston, Louis-Marie Bloyet, Marjorie Cornejo Pontelli, Ma. Xenia G. Llagan, Robert Jan Lebbink, William J. Buchser, Emmanuel J. H. J. Wiertz, Sean P. J. Whelan, and Hidde L Ploegh. A class II MHC-targeted vaccine elicits immunity against SARS-CoV-2 and its variants. *Proceedings of the National Academy of Sciences*. 2021;118(44):e2116147118. DOI: [10.1073/pnas.2116147118](https://doi.org/10.1073/pnas.2116147118)

Arthur W. Lambert, Christopher Fiore, Yogesh Chutake, **Elisha R. Verhaar**, Patrick C. Strasser, Mei Wei Chen, Daneyal Farouq, Sunny Das, Xin Li, Elinor Ng Eaton, Yun Zhang, Joana Liu Donaher, Ian Engstrom, Ferenc Reinhardt, Bingbing Yuan, Sumeet Gupta, Bruce Wollison, Matthew Eaton, Brian Bierie, John Carulli, Eric R. Olson, Matthew G. Guenther, Robert A. Weinberg. Δ Np63/p73 drive metastatic colonization by controlling a regenerative epithelial stem cell program in quasi-mesenchymal cancer stem cells. *Developmental Cell*. 2022;57(24):2414-2730.e8. DOI: [10.1016/j.devcel.2022.11.015](https://doi.org/10.1016/j.devcel.2022.11.015)

Elisha R. Verhaar, Anouk Knoflook, Novalia Pishesha, Xin Liu, Willemijn J.C. van Keizerswaard, Kai W. Wucherpfennig, Hidde L. Ploegh. MICA-specific nanobodies for diagnosis and immunotherapy of MICA⁺ tumors. *Frontiers in Immunology*. 2024;15. DOI: [10.3389/fimmu.2024.1368586](https://doi.org/10.3389/fimmu.2024.1368586)

Elisha R. Verhaar, Willemijn J.C. van Keizerswaard, Anouk Knoflook, Thomas Balligand, Hidde L. Ploegh. Nanobody-based CAR NK cells for possible immunotherapy of MICA⁺ tumors. *PNAS Nexus*. 2024;5(3):pgae184 DOI: [10.1093/pnasnexus/pgae184](https://doi.org/10.1093/pnasnexus/pgae184)

Elisha R. Verhaar, Jin Gan, Susan Buhl, Ziao Li, Amir Horowitz, Hidde L. Ploegh. A monoclonal antibody that recognizes a unique 13-residue epitope in the cytoplasmic tail of HLA-E. *Molecular Immunology*. 2024 article in press. Article reference: MIMM6935

Curriculum vitae

Elisha Verhaar is geboren op 17 januari, 1995 te Vlissingen. Zij behaalde haar VWO diploma aan het Dalton Lyceum in Barendrecht in 2013. In datzelfde jaar begon zij haar bachelorstudie “Biomedische Wetenschappen” aan de Universiteit Utrecht. Tijdens haar studie was ze actief bij meerdere studieverenigingen als hoofdredactrice van de redactie ter verenigingsblad “Tight Junction” der M.B.V. Mebiose, en theaterspecialist van de toneelcommissie “Produkcie”. Tijdens haar studie richtte ze zich op kanker- en stamcelonderzoek, onder meer met een onderzoeksstage in het UMC Utrecht naar de rol van connective tissue growth factor op de ontwikkeling van darmkanker, onder leiding van Onno Kranenburg en Roel Goldschmeding.

Na het behalen van haar bachelordiploma in 2016 startte ze aan de masterstudie “Cancer, Stem Cells, and Developmental Biology” aan de Universiteit Utrecht. Hier specialiseerde ze zich verder tot kankeronderzoek, onder meer met een stage aan het Hubrecht Instituut in het lab van Jacco van Rheenen, waar zij onderzoek deed naar celcompetitie in darmkanker organoids onder leiding van Saskia Suijkerbuijk. Hierna volgde ze een internationale stage in het lab van Robert Weinberg aan het MIT Whitehead Institute te Boston, Verenigde Staten. Onder leiding van Arthur Lambert deed zij onderzoek naar de rol van p63 en p73 op de epitheliale-naar-mesenchymale transitie van borstkankercellen.

Na het behalen van haar masterdiploma in 2018 werkte ze in de Weinberg groep aan hetzelfde project als onderzoeksassistent. In 2019 begon ze aan haar afstudeertraject in het lab van Hidde Ploegh, Boston Children’s Hospital, Verenigde Staten. In 2024 verdedigt ze haar proefschrift, en zal hierna waarschijnlijk werkzaam worden bij een biomedisch bedrijf in Boston, Verenigde Staten.

Acknowledgements

Beste Hidde, je gaf me alle vrijheid om zelf de teugels in handen te nemen maar stond altijd klaar om toch even een duwtje in de juiste richting te geven. Bedankt dat ik deze kans heb gekregen om mijzelf in jouw lab te ontwikkelen van “student” naar “wetenschapper”.

Beste Sjaak, bedankt voor het accepteren van deze gekke PhD situatie. Hoewel we elkaar welgeteld drie keer gezien hebben, kan ik met zekerheid zeggen dat het Neefjes lab een fantastische plek moet zijn. Bedankt voor de adviezen en feedback op mijn werk.

Dear Thomas, thank you for entertaining me every single day, not just literally (your French swearing was entertaining!) You also entertained my many, many, many questions. I could always turn to you for help and advice about lab work, cloning, PET imaging, data analysis, medical problems, personal issues... The list goes on and on. I am not sure what I would've done without you in the bay. Cool bay forever!

Dear Lotte and Claire, together we were the “three PhD’ers”. But we didn’t just bond over mutual suffering, we made an actual connection. You girls are the best lab mates anyone could ask for. Dealing with all the “llama llama holiday drama” was easy with you guys thanks to our friendship. Always up for a drink, a hangout, a dance party (cue TomM!), raclette, board games, but also a serious chat or a vent. Thank you for becoming my friends.

Lieve Willemijn and Anouk, you both came to work with me like a gift from the UVA Heaven! Your dedication to the projects was unmatched and I can’t even express how helpful the two of you have been. Your scientific mindset was a true inspiration to lift these projects to a higher level. You worked tirelessly on cloning vectors, creating virus, producing CAR NK and CAR T cells, designing and executing downstream applications, the list goes on... I am so lucky to have mentored you two incredible women and can’t wait to see what your future holds.

Dear David, my liefje. You met me the month before I started my PhD and have dealt with the ups-and-downs of #PhDLife throughout the many years that followed. David, I can’t emphasize enough how incredibly lucky I am to have you in my life and to be able to call you my husband. You have supported me in more ways than I could’ve ever hoped for. Not just by taking

over many of the household chores during my most busy times at work, but also by being there for me emotionally. You accepted when our plans had to change because I had to run experiments. You gave advice, even when I told you I wasn't looking for solutions. Liefje, you are always the solution. Coming home every day to yours and our cat's love, that's always been what's keeping me going.

Lieve papa. Wat zou ik graag willen dat je dit kon lezen... Ik weet hoe ontzettend trots je zou zijn geweest. Je vroeg zo vaak "en, ben je al bezig met het schrijven van je proefschrift?" en dan moest ik uitleggen dat ik nog steeds druk bezig was met experimenten, dus nee ik was nog niet aan het schrijven... Je stuurde me vaak linkjes naar interessante artikelen, en dan praatten we daar even over. Ik heb zo veel van jou geleerd en jij ook van mij. Hoewel jij nu niets meer van mij kunt leren, leer ik nog elke dag van jou. Papa, je meisje is nu echt klaar met haar proefschrift. Je had hem zo graag willen lezen. Ik draag hem op aan jou.

Lieve mama. Ik ben zo ontzettend blij dat ik dit moment met jou kan delen. Toen ik naar Boston verhuisde voor eens stage van 8 maanden zei je: "Meid, jij komt nooit meer terug naar Nederland". 'Onzin', dacht ik. Maar het is nu 6,5 jaar later en Boston is nog steeds mijn thuis. Ondanks de afstand hebben we elkaar zo vaak mogelijk gezien. Ik vind het zo bewonderingwaardig dat jij, zonder medische achtergrond, altijd zo geïnteresseerd was in mijn onderwerp. Je wist altijd de juiste vragen te stellen waardoor we heerlijk konden kletsen over biomedische wetenschappen en mijn onderzoek. Mama, dank je wel voor de afgelopen jaren, voor het aanhoren van mijn frustraties, voor de altijd goede raad. Ik hou zo veel van jou.

References

1. World Health Organization. Global cancer burden growing, amidst mounting need for services. (2024).
2. Fares, J., Fares, M. Y., Khachfe, H. H., Salhab, H. A. & Fares, Y. Molecular principles of metastasis: a hallmark of cancer revisited. *Signal Transduct Target Ther* **5**, (2020).
3. Neophytou, C. M., Panagi, M., Stylianopoulos, T. & Papageorgis, P. The Role of Tumor Microenvironment in Cancer Metastasis: Molecular Mechanisms and Therapeutic Opportunities. *Cancers (Basel)* **13**, 1–22 (2021).
4. McAllister, S. S. & Weinberg, R. A. The tumour-induced systemic environment as a critical regulator of cancer progression and metastasis. *Nat Cell Biol* **16**, 717–727 (2014).
5. Kurose, K. *et al.* Genetic model of multi-step breast carcinogenesis involving the epithelium and stroma: clues to tumour-microenvironment interactions. *Hum Mol Genet* **10**, 1907–1913 (2001).
6. Vogelstein, B., Lane, D. & Levine, A. J. Surfing the p53 network. *Nature* **408**, 307–310 (2000).
7. Wang, X. W. & Harris, C. C. p53 tumor-suppressor gene: clues to molecular carcinogenesis. *J Cell Physiol* **173**, 247–255 (1997).
8. Ford, D. *et al.* Genetic Heterogeneity and Penetrance Analysis of the BRCA1 and BRCA2 Genes in Breast Cancer Families. *Am J Hum Genet* **62**, 676–689 (1998).
9. Li, J. *et al.* PTEN, a Putative Protein Tyrosine Phosphatase Gene Mutated in Human Brain, Breast, and Prostate Cancer. *Science* (1979) **275**, 1943–1947 (1997).
10. Myers, M. P. *et al.* The lipid phosphatase activity of PTEN is critical for its tumor suppressor function. *Proceedings of the National Academy of Sciences* **95**, 13513–13518 (1998).
11. Tan, M. H. *et al.* Lifetime cancer risks in individuals with germline PTEN mutations. *Clinical Cancer Research* **18**, 400–407 (2012).
12. Wang, S.-C., Lin, S.-H., Su, L.-K. & Hung, M.-C. Changes in BRCA2 Expression during Progression of the Cell Cycle. *Biochem Biophys Res Commun* **234**, 247–251 (1997).
13. Fischer, M. Census and evaluation of p53 target genes. *Oncogene* **36**, 3943–3956 (2017).
14. Nusse, R., van Ooyen, A., Cox, D., Kai T. Fung, Y. & Varmus, H. Mode of proviral activation of a putative mammary oncogene (int-1) on mouse chromosome 15. *Nature* **307**, 131–136 (1984).
15. Giehl, K. Oncogenic Ras in tumour progression and metastasis. *Journal of Biological Chemistry* **386**, 193–205 (2005).
16. Barbacid, M. ras genes. *Annu Rev Biochem* **56**, 779–827 (1987).
17. Rajalingam, K., Schreck, R., Rapp, U. R. & Albert, Š. Ras oncogenes and their downstream targets. *Biochim Biophys Acta Mol Cell Res* **1773**, 1177–1195 (2007).
18. Prior, I. A., Lewis, P. D. & Mattos, C. A comprehensive survey of ras mutations in cancer. *Cancer Res* **72**, 2457–2467 (2012).
19. Madden, S. K., de Araujo, A. D., Gerhardt, M., Fairlie, D. P. & Mason, J. M. Taking the Myc out of cancer: toward therapeutic strategies to directly inhibit c-Myc. *Mol Cancer* **20**, (2021).
20. Zhan, T., Rindtorff, N. & Boutros, M. Wnt signaling in cancer. *Oncogene* **36**, 1461–1473 (2017).
21. Duffy, M. J., O'Grady, S., Tang, M. & Crown, J. MYC as a target for cancer treatment. *Cancer Treatment Reviews* vol. 94 Preprint at <https://doi.org/10.1016/j.ctrv.2021.102154> (2021).

22. Rajalingam, K., Schreck, R., Rapp, U. R. & Albert, Š. Ras oncogenes and their downstream targets. *Biochim Biophys Acta Mol Cell Res* **1773**, 1177–1195 (2007).
23. Sinn, E. et al. Coexpression of MMTV/v-Ha-ras and MMTV/c-myc genes in transgenic mice: Synergistic action of oncogenes in vivo. *Cell* **49**, 465–475 (1987).
24. Tsukamoto, A. S., Grosschedl, R., Guzman, R. C., Parslow, T. & Varmus, H. E. Expression of the int-1 gene in transgenic mice is associated with mammary gland hyperplasia and adenocarcinomas in male and female mice. *Cell* **55**, 619–625 (1988).
25. Maleno, I. et al. LOH at 6p21.3 region and HLA class I altered phenotypes in bladder carcinomas. *Immunogenetics* **58**, 503–510 (2006).
26. Maleno, I. et al. Distribution of HLA class I altered phenotypes in colorectal carcinomas: high frequency of HLA haplotype loss associated with loss of heterozygosity in chromosome region 6p21. *Immunogenetics* **56**, 244–253 (2004).
27. Hanigiri, T. et al. Prognostic implications of human leukocyte antigen class I expression in patients who underwent surgical resection for non-small-cell lung cancer. *Journal of Surgical Research* **181**, 57–63 (2013).
28. Simpson, J. A. D. et al. Intratumoral T cell infiltration, MHC class I and STAT1 as biomarkers of good prognosis in colorectal cancer. *Gut* **59**, 926–933 (2010).
29. Watson, N. F. S. et al. Immunosurveillance is active in colorectal cancer as downregulation but not complete loss of MHC class I expression correlates with a poor prognosis. *Int J Cancer* **118**, 6–10 (2006).
30. Squire, R., Fowler, C. L., Brooks, S. P., Rich, G. A. & Cooney, D. R. The relationship of class I MHC antigen expression to stage IV-S disease and survival in neuroblastoma. *J Pediatr Surg* **25**, 381–386 (1990).
31. Feenstra, M. et al. HLA class I expression and chromosomal deletions at 6p and 15q in head and neck squamous cell carcinomas. *Tissue Antigens* **54**, 235–245 (1999).
32. Garrido, M. A. et al. HLA class I alterations in breast carcinoma are associated with a high frequency of the loss of heterozygosity at chromosomes 6 and 15. *Immunogenetics* **70**, 647–659 (2018).
33. Seliger, B. et al. Immune escape of melanoma: first evidence of structural alterations in two distinct components of the MHC class I antigen processing pathway. *Cancer Res* **61**, 8647–8650 (2001).
34. Martín-Villa, J. M. et al. HLA-G: Too Much or Too Little? Role in Cancer and Autoimmune Disease. *Front Immunol* **13**, (2022).
35. Morandi, F. & Pistoia, V. Interactions between HLA-G and HLA-E in physiological and pathological conditions. *Front Immunol* **5**, (2014).
36. de Kruijff, E. M. et al. HLA-E and HLA-G Expression in Classical HLA Class I-Negative Tumors Is of Prognostic Value for Clinical Outcome of Early Breast Cancer Patients. *The Journal of Immunology* **185**, 7452–7459 (2010).
37. Zeestraten, E. C. M. et al. Combined analysis of HLA class I, HLA-E and HLA-G predicts prognosis in colon cancer patients. *Br J Cancer* **110**, 459–468 (2014).
38. Henke, E., Nandigama, R. & Ergün, S. Extracellular Matrix in the Tumor Microenvironment and Its Impact on Cancer Therapy. *Front Mol Biosci* **6**, (2020).
39. Rice, A. J. et al. Matrix stiffness induces epithelial-mesenchymal transition and promotes chemoresistance in pancreatic cancer cells. *Oncogenesis* **6**, (2017).
40. Dongre, A. & Weinberg, R. A. New insights into the mechanisms of epithelial-mesenchymal transition and implications for cancer. *Nat Rev Mol Cell Biol* **20**, 69–84 (2019).
41. Wei, S. C. et al. Matrix stiffness drives epithelial-mesenchymal transition and tumour metastasis through a TWIST1-G3BP2 mechanotransduction pathway. *Nat Cell Biol* **17**, 678–688 (2015).

42. Greenburg, G. & Hay, E. D. Epithelia Suspended in Collagen Gels Can Lose Polarity and Express Characteristics of Migrating Mesenchymal Cells. *J Cell Biol* **95**, 333–339 (1982).
43. Ribatti, D., Tamma, R. & Annese, T. Epithelial-Mesenchymal Transition in Cancer: A Historical Overview. *Transl Oncol* **13**, (2020).
44. Hallmann, R. *et al.* The regulation of immune cell trafficking by the extracellular matrix. *Current Opinions in Cell Biology* **36**, 54–61 (2015).
45. Zhu, X. & Zhu, J. CD4 T helper cell subsets and related human immunological disorders. *Int J Mol Sci* **21**, 1–26 (2020).
46. Knutson, K. L. & Disis, M. L. Tumor antigen-specific T helper cells in cancer immunity and immunotherapy. *Cancer Immunology, Immunotherapy* **54**, 721–728 (2005).
47. Knutson, K. L. & Disis, M. L. Tumor antigen-specific T helper cells in cancer immunity and immunotherapy. *Cancer Immunology, Immunotherapy* **54**, 721–728 (2005).
48. Nonaka, K. *et al.* Th1 polarization in the tumor microenvironment upregulates the myeloid-derived suppressor-like function of macrophages. *Cell Immunol* **369**, (2021).
49. Laheurte, C. *et al.* Distinct prognostic value of circulating anti-telomerase CD4+ Th1 immunity and exhausted PD-1+/TIM-3+ T cells in lung cancer. *Br J Cancer* **121**, 405–416 (2019).
50. Aue, G. *et al.* Activation of Th1 Immunity within the Tumor Microenvironment Is Associated with Clinical Response to Lenalidomide in Chronic Lymphocytic Leukemia. *The Journal of Immunology* **201**, 1967–1974 (2018).
51. Haabeth, O. A. W. *et al.* Inflammation driven by tumour-specific Th1 cells protects against B-cell cancer. *Nat Commun* **2**, (2011).
52. Walker, J. A. & McKenzie, A. N. J. TH2 cell development and function. *Nat Rev Immunol* **18**, 121–133 (2018).
53. Steinke, J. W. & Borish, L. Th2 cytokines and asthma Interleukin-4: its role in the pathogenesis of asthma, and targeting it for asthma treatment with interleukin-4 receptor antagonists. *Respir Res* **2**, 66–70 (2001).
54. Wang, N., Liang, H. & Zen, K. Molecular mechanisms that influence the macrophage M1-M2 polarization balance. *Front Immunol* **5**, (2014).
55. Yao, Y., Xu, X. H. & Jin, L. Macrophage polarization in physiological and pathological pregnancy. *Front Immunol* **10**, (2019).
56. Murray, P. J. Macrophage Polarization. *Annu Rev Physiol* **79**, 541–566 (2017).
57. Chraa, D., Naim, A., Olive, D. & Badou, A. T lymphocyte subsets in cancer immunity: Friends or foes. *J Leukoc Biol* **105**, 243–255 (2019).
58. Jacenik, D., Karagiannidis, I. & Beswick, E. J. Th2 cells inhibit growth of colon and pancreas cancers by promoting anti-tumorigenic responses from macrophages and eosinophils. *Br J Cancer* **128**, 387–397 (2023).
59. Protti, M. P. & De Monte, L. Cross-talk within the tumor microenvironment mediates Th2-type inflammation in pancreatic cancer. *Oncoimmunology* **1**, 89–91 (2012).
60. Mattes, J. *et al.* Immunotherapy of cytotoxic T cell-resistant tumors by T helper 2 cells: An eotaxin and STAT6-dependent process. *Journal of Experimental Medicine* **197**, 387–393 (2003).
61. Boieri, M. *et al.* CD4+ T helper 2 cells suppress breast cancer by inducing terminal differentiation. *Journal of Experimental Medicine* **219**, (2022).
62. Nishimura, T. *et al.* Distinct Role of Antigen-specific T Helper Type 1 (Th1) and Th2 Cells in Tumor Eradication In Vivo. *Journal of Experimental Medicine* **190**, 617–627 (1999).
63. Jiang, Y. *et al.* TNF- α enhances Th9 cell differentiation and antitumor immunity via TNFR2-dependent pathways. *J Immunother Cancer* **7**, (2019).

64. Lu, Y. *et al.* Th9 cells promote antitumor immune responses in vivo. *Journal of Clinical Investigation* **122**, 4160–4171 (2012).
65. Kim, I. K. *et al.* GM-CSF promotes antitumor immunity by inducing Th9 cell responses. *Cancer Immunol Res* **7**, 498–509 (2019).
66. Végrán, F. *et al.* The transcription factor IRF1 dictates the IL-21-dependent anticancer functions of T H9 cells. *Nat Immunol* **15**, 758–766 (2014).
67. Xue, G., Jin, G., Fang, J. & Lu, Y. IL-4 together with IL-1 β induces antitumor Th9 cell differentiation in the absence of TGF- β signaling. *Nat Commun* **10**, (2019).
68. Humblin, E. *et al.* IRF8-dependent molecular complexes control the Th9 transcriptional program. *Nat Commun* **8**, (2017).
69. Kim, I. K. *et al.* GM-CSF promotes antitumor immunity by inducing Th9 cell responses. *Cancer Immunol Res* **7**, 498–509 (2019).
70. You, F.-P. *et al.* Th9 cells promote antitumor immunity via IL-9 and IL-21 and demonstrate atypical cytokine expression in breast cancer. *Int Immunopharmacol* **52**, 163–167 (2017).
71. Wei, L., Laurence, A., Elias, K. M. & O'shea, J. J. IL-21 is produced by TH17 cells and drives IL-17 production in a STAT3-dependent manner. *Journal of Biological Chemistry* **282**, 34605–34610 (2007).
72. Yang, L. *et al.* IL-21 and TGF- β are required for differentiation of human TH17 cells. *Nature* **454**, 350–352 (2008).
73. Miossec, P., Korn, T. & Kuchroo, V. K. Mechanisms of Disease Interleukin-17 and Type 17 Helper T Cells. *New England Journal of Medicine* **361**, 888–98 (2009).
74. Noack, M. & Miossec, P. Th17 and regulatory T cell balance in autoimmune and inflammatory diseases. *Autoimmun Rev* **13**, 668–677 (2014).
75. Yang, L. *et al.* Expression of Th17 Cells in Breast Cancer Tissue and Its Association with Clinical Parameters. *Cell Biochem Biophys* **153**–159 (2012) doi:10.1007/s12013-011-9276-3.
76. Baharloo, R. *et al.* Reduced levels of T-helper 17-associated cytokines in the serum of patients with breast cancer: Indicators for following the course of disease. *Central European Journal of Immunology* **41**, 78–85 (2016).
77. Chen, W.-C. *et al.* Interleukin-17-producing cell infiltration in the breast cancer tumour microenvironment is a poor prognostic factor. *Histopathology* **63**, 225–233 (2013).
78. Avalos-Navarro, G. *et al.* Circulating soluble levels of MIF in women with breast cancer in the molecular subtypes: relationship with Th17 cytokine profile. *Clin Exp Med* **19**, 385–391 (2019).
79. Eyerich, S. *et al.* Th22 cells represent a distinct human T cell subset involved in epidermal immunity and remodeling. *Journal of Clinical Investigation* **119**, 3573–3585 (2009).
80. Trifari, S., Kaplan, C. D., Tran, E. H., Crellin, N. K. & Spits, H. Identification of a human helper T cell population that has abundant production of interleukin 22 and is distinct from TH-17, TH1 and TH2 cells. *Nat Immunol* **10**, 864–871 (2009).
81. Duhen, T., Geiger, R., Jarrossay, D., Lanzavecchia, A. & Sallusto, F. Production of interleukin 22 but not interleukin 17 by a subset of human skin-homing memory T cells. *Nat Immunol* **10**, 857–863 (2009).
82. Jiang, R. *et al.* Interleukin-22 promotes human hepatocellular carcinoma by activation of STAT3. *Hepatology* **54**, 900–909 (2011).
83. Khosravi, N. *et al.* IL22 promotes kras-mutant lung cancer by induction of a protumour immune response and protection of stemness properties. *Cancer Immunol Res* **6**, 788–797 (2018).
84. Cui, G. TH9, TH17, and TH22 Cell Subsets and Their Main Cytokine Products in the Pathogenesis of Colorectal Cancer. *Front Oncol* **9**, (2019).

85. Crotty, S. T Follicular Helper Cell Differentiation, Function, and Roles in Disease. *Immunity* **41**, 529–542 (2014).
86. King, C. New insights into the differentiation and function of T follicular helper cells. *Nat Rev Immunol* **9**, 757–766 (2009).
87. Pandey, S. *et al.* IL-4/CXCL12 loop is a key regulator of lymphoid stroma function in follicular lymphoma. *Blood* **129**, 2507–2518 (2017).
88. Rawal, S. *et al.* Cross Talk between Follicular Th Cells and Tumor Cells in Human Follicular Lymphoma Promotes Immune Evasion in the Tumor Microenvironment. *The Journal of Immunology* **190**, 6681–6693 (2013).
89. Amé-Thomas, P. *et al.* CD10 delineates a subset of human IL-4 producing follicular helper T cells involved in the survival of follicular lymphoma B cells. *Blood* **125**, (2015).
90. Zhanshan Cha *et al.* Circulating CXCR5+CD4+ T cells assist in the survival and growth of primary diffuse large B cell lymphoma cells through interleukin 10 pathway. *Exp Cell Res* **350**, 154–160 (2017).
91. Goubet, A. G. *et al.* Escherichia coli-Specific CXCL13-Producing TFH Are Associated with Clinical Efficacy of Neoadjuvant PD-1 Blockade against Muscle- Invasive Bladder Cancer. *Cancer Discov* **12**, 2280–2307 (2022).
92. Gu-Trantien, C. *et al.* CD4+ follicular helper T cell infiltration predicts breast cancer survival. *Journal of Clinical Investigation* **123**, 2873–2892 (2013).
93. Noël, G. *et al.* Functional Th1-oriented T follicular helper cells that infiltrate human breast cancer promote effective adaptive immunity. *Journal of Clinical Investigation* **131**, (2021).
94. Niogret, J. *et al.* Follicular helper-T cells restore CD8+ -dependent antitumor immunity and anti-PD-L1/PD-1 efficacy. *J Immunother Cancer* **9**, (2021).
95. Phanthunane, C. *et al.* Intratumoral Niches of B Cells and Follicular Helper T Cells, and the Absence of Regulatory T Cells, Associate with Longer Survival in Early-Stage Oral Tongue Cancer Patients. *Cancers (Basel)* **14**, (2022).
96. Walunas, T. L., Akker, C. Y. B. & Bluestone, J. A. CTLA-4 Ligation Blocks CD28-dependent T Cell Activation. *Journal of Experimental Medicine* **183**, 2541–2550 (1996).
97. Walunas, T. L. *et al.* CTLA-4 can function as a negative regulator of T cell activation. *Immunity* **1**, 405–413 (1994).
98. Ruella, M. *et al.* Overcoming the immunosuppressive tumor microenvironment of Hodgkin lymphoma using chimeric antigen receptor T cells. *Cancer Discov* **7**, 1154–1167 (2017).
99. Crane, C. A. *et al.* TGF- β downregulates the activating receptor NKG2D on NK cells and CD8+ T cells in glioma patients. *Neuro Oncol* **12**, 7–13 (2010).
100. Huang, J. J. & Blobe, G. C. Dichotomous roles of TGF- β in human cancer. *Biochem Soc Trans* **44**, 1441–1454 (2016).
101. Bjorkman, P. J. *et al.* The foreign antigen binding site and T cell recognition regions of class I histocompatibility antigens. *Nature* **329**, 512–518 (1987).
102. Xing, Y. & Hogquist, K. A. T-Cell tolerance: Central and peripheral. *Cold Spring Harb Perspect Biol* **4**, 1–15 (2012).
103. Sigal, L. J. Activation of CD8 T Lymphocytes during Viral Infections. *Encyclopedia of Immunobiology* **4**, 286–290 (2016).
104. De Araujo-Souza, P. S., Hanschke, S. C. H. & Viola, J. P. B. Epigenetic control of interferon-gamma expression in CD8 T cells. *J Immunol Res* **2015**, (2015).
105. Araki, Y., Fann, M., Wersto, R. & Weng, N.-P. Histone Acetylation Facilitates Rapid and Robust Memory CD8 T Cell Response through Differential Expression of Effector Molecules (Eomesodermin and Its Targets: Perforin and Granzyme B). *The Journal of Immunology* **180**, 8102–8108 (2008).

106. Haring, J. S., Badovinac, V. P. & Harty, J. T. Inflaming the CD8+ T Cell Response. *Immunity* **25**, 19–29 (2006).
107. De Araujo-Souza, P. S., Hanschke, S. C. H. & Viola, J. P. B. Epigenetic control of interferon-gamma expression in CD8 T cells. *J Immunol Res* **2015**, (2015).
108. Araki, Y., Fann, M., Wersto, R. & Weng, N.-P. Histone Acetylation Facilitates Rapid and Robust Memory CD8 T Cell Response through Differential Expression of Effector Molecules (Eomesodermin and Its Targets: Perforin and Granzyme B) 1. *The Journal of Immunology* **180**, 8102–8108 (2008).
109. Djenidi, F. et al. CD8+CD103+ Tumor-Infiltrating Lymphocytes Are Tumor-Specific Tissue-Resident Memory T Cells and a Prognostic Factor for Survival in Lung Cancer Patients. *The Journal of Immunology* **194**, 3475–3486 (2015).
110. Noh, B. J., Kwak, J. Y. & Eom, D. W. Immune classification for the PD-L1 expression and tumour-infiltrating lymphocytes in colorectal adenocarcinoma. *BMC Cancer* **20**, (2020).
111. Kawai, O. et al. Predominant infiltration of macrophages and CD8+ T cells in cancer nests is a significant predictor of survival in stage IV nonsmall cell lung cancer. *Cancer* **113**, 1387–1395 (2008).
112. Zhul, Y. et al. Impact of Cytotoxic T Lymphocytes Immunotherapy on Prognosis of Colorectal Cancer Patients. *Clinics in Oncology* **7**, (2023).
113. Pal, A. & Kundu, R. Human Papillomavirus E6 and E7: The Cervical Cancer Hallmarks and Targets for Therapy. *Front Microbiol* **10**, (2020).
114. Peng, S. et al. Identification of human MHC-I HPV18 E6/E7-specific CD8 + T cell epitopes and generation of an HPV18 E6/E7-expressing adenosquamous carcinoma in HLA-A2 transgenic mice. *J Biomed Sci* **29**, (2022).
115. Hong, J. Y. et al. Claudin 18.2 expression in various tumor types and its role as a potential target in advanced gastric cancer. *Transl Cancer Res* **9**, 3367–3374 (2020).
116. Siehl, J. M. et al. Expression of Wilms' tumor gene 1 at different stages of acute myeloid leukemia and analysis of its major splice variants. *Ann Hematol* **83**, 745–750 (2004).
117. Kramarzova, K. et al. Real-time PCR quantification of major Wilms' tumor gene 1 (WT1) isoforms in acute myeloid leukemia, their characteristic expression patterns and possible functional consequences. *Leukemia* **26**, 2086–2095 (2012).
118. Busse, A. et al. Wilms' tumor gene 1 (WT1) expression in subtypes of acute lymphoblastic leukemia (ALL) of adults and impact on clinical outcome. *Ann Hematol* **88**, 1199–1205 (2009).
119. Strickland, K. C. et al. Association and prognostic significance of BRCA1/2-mutation status with neoantigen load, number of tumor-infiltrating lymphocytes and expression of PD-1/PD-L1 in high grade serous ovarian cancer. *Oncotarget* **7**, (2016).
120. Xu, K., Yang, S. & Zhao, Y. Prognostic significance of BRCA mutations in ovarian cancer: an updated systematic review with meta-analysis. *Oncotarget* **8**, 285–302 (2017).
121. Bailur, J. K., Gueckel, B., Derhovanessian, E. & Pawelec, G. Presence of circulating Her2-reactive CD8 + T-cells is associated with lower frequencies of myeloid-derived suppressor cells and regulatory T cells, and better survival in older breast cancer patients. *Breast Cancer Research* **17**, (2015).
122. Mitri, Z., Constantine, T. & O'Regan, R. The HER2 Receptor in Breast Cancer: Pathophysiology, Clinical Use, and New Advances in Therapy. *Chemother Res Pract* **2012**, 1–7 (2012).
123. Le, K. et al. Overexpression of mesothelin in pancreatic ductal adenocarcinoma (PDAC). *Int J Med Sci* **17**, 422–427 (2020).
124. Wang, K., Wei, G. & Liu, D. CD19: a biomarker for B cell development, lymphoma diagnosis and therapy. *Exp Hematol Oncol* **1**, (2012).

125. Tomiyama, H., Matsuda, T. & Takiguchi, M. Differentiation of Human CD8+ T Cells from a Memory to Memory/Effector Phenotype. *The Journal of Immunology* **168**, 5538–5550 (2002).
126. Cui, W. & Kaech, S. M. Generation of effector CD8+ T cells and their conversion to memory T cells. *Immunol Rev* **236**, 151–166 (2010).
127. Youngblood, B. *et al.* Effector CD8 T cells dedifferentiate into long-lived memory cells. *Nature* **552**, 404–409 (2017).
128. Akondy, R. S. *et al.* Origin and differentiation of human memory CD8 T cells after vaccination. *Nature* **552**, 362–367 (2017).
129. Turner, D. L. *et al.* Lung niches for the generation and maintenance of tissue-resident memory T cells. *Mucosal Immunol* **7**, 501–510 (2014).
130. Corgnac, S., Boutet, M., Kfouri, M., Naltet, C. & Mami-Chouaib, F. The emerging role of CD8+ tissue resident memory T (TRM) cells in antitumor immunity: A unique functional contribution of the CD103 integrin. *Front Immunol* **9**, (2018).
131. Farber, D. L., Yudanin, N. A. & Restifo, N. P. Human memory T cells: Generation, compartmentalization and homeostasis. *Nat Rev Immunol* **14**, 24–35 (2014).
132. Edwards, J. *et al.* CD103+ tumor-resident CD8+ T cells are associated with improved survival in immunotherapy-naïve melanoma patients and expand significantly during anti-PD-1 treatment. *Clinical Cancer Research* **24**, 3036–3045 (2018).
133. Park, S. L. *et al.* Tissue-resident memory CD8+ T cells promote melanoma-immune equilibrium in skin. *Nature* **565**, 366–371 (2019).
134. Malik, B. T. *et al.* Resident memory T cells in the skin mediate durable immunity to melanoma. *Sci Immunol* **2**, (2017).
135. Ganeshan, A. P. *et al.* Tissue-resident memory features are linked to the magnitude of cytotoxic T cell responses in human lung cancer. *Nat Immunol* **18**, 940–950 (2017).
136. Koh, J. *et al.* Prognostic implications of intratumoral CD103+ tumor-infiltrating lymphocytes in pulmonary squamous cell carcinoma. *Oncotarget* **8**, 13762–13769 (2017).
137. Wang, Z. Q. *et al.* CD103 and intratumoral immune response in breast cancer. *Clinical Cancer Research* **22**, 6290–6297 (2016).
138. Peter Savas *et al.* Single-cell profiling of breast cancer T cells reveals a tissue-resident memory subset associated with improved prognosis. *Nat Med* **24**, 986–993 (2018).
139. Webb, J. R. *et al.* Profound elevation of CD8+ T cells expressing the intraepithelial lymphocyte marker CD103 ($\alpha E/\beta 7$ Integrin) in high-grade serous ovarian cancer. *Gynecologic Oncology* **118**, 228–236 (2010).
140. Duhen, T. *et al.* Co-expression of CD39 and CD103 identifies tumor-reactive CD8 T cells in human solid tumors. *Nat Commun* **9**, (2018).
141. Atri, C., Guerfali, F. Z. & Laouini, D. Role of human macrophage polarization in inflammation during infectious diseases. *Int J Mol Sci* **19**, (2018).
142. Shapouri-Moghaddam, A. *et al.* Macrophage plasticity, polarization, and function in health and disease. *J Cell Physiol* **233**, 2425–6440 (2018).
143. Mantovani, A. *et al.* The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol* **25**, 677–686 (2004).
144. Italiani, P. *et al.* Transcriptomic profiling of the development of the inflammatory response in human monocytes in vitro e8768o. *PLoS One* **9**, (2014).
145. Martinez, F. O., Gordon, S., Locati, M. & Mantovani, A. Transcriptional Profiling of the Human Monocyte-to-Macrophage Differentiation and Polarization: New Molecules and Patterns of Gene Expression. *The Journal of Immunology* **177**, 7303–7311 (2006).
146. Kadomoto, S., Izumi, K. & Mizokami, A. Macrophage Polarity and Disease Control. *Int J Mol Sci* **23**, (2022).

147. Zhang, F. *et al.* TGF- β induces M2-like macrophage polarization via SNAIL-mediated suppression of a pro-inflammatory phenotype. *Oncotarget* **7**, 52294–52306 (2016).
148. Mantovani, A., Sozzani, S., Locati, M., Allavena, P. & Sica, A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol* **23**, 534–555 (2002).
149. Murray, P. J. & Wynn, T. A. Protective and pathogenic functions of macrophage subsets. *Nat Rev Immunol* **11**, 723–737 (2011).
150. Sumitomo, R. *et al.* PD-L1 expression on tumor-infiltrating immune cells is highly associated with M2 TAM and aggressive malignant potential in patients with resected non-small cell lung cancer. *Lung Cancer* **136**, 136–144 (2019).
151. Cao, L. *et al.* M2 macrophage infiltration into tumor islets leads to poor prognosis in non-small-cell lung cancer. *Cancer Manag Res* **11**, 6125–6138 (2019).
152. Sumitomo, R. *et al.* M2 tumor-associated macrophages promote tumor progression in non-small-cell lung cancer. *Exp Ther Med* (2019) doi:10.3892/etm.2019.8068.
153. Hughes, R. *et al.* Perivascular M2 macrophages stimulate tumor relapse after chemotherapy. *Cancer Res* **75**, 3479–3491 (2015).
154. Kim, O. H. *et al.* Proangiogenic TIE2+/CD31+ macrophages are the predominant population of tumor-associated macrophages infiltrating metastatic lymph nodes. *Mol Cells* **36**, 432–438 (2013).
155. Chen, Y. *et al.* Tumor-associated macrophages: An accomplice in solid tumor progression. *J Biomed Sci* **26**, (2019).
156. Yeo, E. J. *et al.* Myeloid wnt7b mediates the angiogenic switch and metastasis in breast cancer. *Cancer Res* **74**, 2962–2973 (2014).
157. Palazon, A. *et al.* An HIF-1 α /VEGF-A Axis in Cytotoxic T Cells Regulates Tumor Progression. *Cancer Cell* **32**, 669–683.e5 (2017).
158. Kadowaki, I. *et al.* Accelerated lymphangiogenesis in malignant lymphoma: Possible role of VEGF-A and VEGF-C. *Br J Haematol* **130**, 869–877 (2005).
159. De Maria, A., Bozzano, F., Cantoni, C. & Moretta, L. Revisiting human natural killer cell subset function revealed cytolytic CD56dimCD16+ NK cells as rapid producers of abundant IFN- γ on activation. *Proceedings of the National Academy of Sciences* **108**, 728–732 (2011).
160. Kärre, K. NK cells, MHC class I molecules and the missing self. *Scand J Immunol* **55**, 221–228 (2002).
161. Vivier, E. *et al.* Innate or adaptive immunity? The example of natural killer cells. *Science* (1979) **331**, 44–49 (2011).
162. Paul, S. & Lal, G. The molecular mechanism of natural killer cells function and its importance in cancer immunotherapy. *Front Immunol* **8**, (2017).
163. Kumar, S. Natural killer cell cytotoxicity and its regulation by inhibitory receptors. *Immunology* **154**, 383–393 (2018).
164. Sivori, S. *et al.* Inhibitory Receptors and Checkpoints in Human NK Cells, Implications for the Immunotherapy of Cancer. *Front Immunol* **11**, (2020).
165. Dębska-Zielkowska, J. *et al.* Kir receptors as key regulators of nk cells activity in health and disease. *Cells* **10**, (2021).
166. Slamon, D. *et al.* Adjuvant Trastuzumab in HER2-Positive Breast Cancer. *New England Journal of Medicine* **344**, 1273–83 (2001).
167. Maloney, D. G. *et al.* IDEC-C2B8 (Rituximab) Anti-CD20 Monoclonal Antibody Therapy in Patients With Relapsed Low-Grade Non-Hodgkin's Lymphoma. *Blood* **90**, 2189–2195 (1997).
168. Mendelsohn, J. & Baselga, J. Status of Epidermal Growth Factor Receptor Antagonists in the Biology and Treatment of Cancer. *Journal of Clinical Oncology* **21**, 2787–2799 (2003).

169. Shah, M. H. *et al.* Phase I study of IMGN901, a CD56-targeting antibody-drug conjugate, in patients with CD56-positive solid tumors. *Invest New Drugs* **34**, 290–299 (2016).
170. Gordon, M. S. *et al.* Phase I Safety and Pharmacokinetic Study of Recombinant Human Anti-Vascular Endothelial Growth Factor in Patients With Advanced Cancer. *Journal of Clinical Oncology* **19**, (2001).
171. Fu, Z., Li, S., Han, S., Shi, C. & Zhang, Y. Antibody drug conjugate: the “biological missile” for targeted cancer therapy. *Signal Transduct Target Ther* **7**, (2022).
172. Diamantis, N. & Banerji, U. Antibody-drug conjugates - An emerging class of cancer treatment. *Br J Cancer* **114**, 362–367 (2016).
173. Peters, C. & Brown, S. Antibody-drug conjugates as novel anti-cancer chemotherapeutics. *Biosci Rep* **35**, (2015).
174. Issell, B. F. & Crooke, S. T. Maytansine. *Cancer Treat Rev* **5**, 199–207 (1978).
175. Kupchan, S. M. *et al.* Maytansine, a novel antileukemic ansa macrolide from Maytenus ovatus. *J Am Chem Soc* **94**, 1354–1356 (1972).
176. Sieber, S. M. *et al.* Experimental studies with maytansine--a new antitumor agent. *Bibl Haematol* **43**, 495–500 (1975).
177. Bhattacharyya, A. & Wolff, J. Maytansine binding to the vinblastine sites of tubulin. *FEBS Lett* **75**, 159–162 (1977).
178. Krop, I. E. *et al.* Phase I study of trastuzumab-DM1, an HER2 antibody-drug conjugate, given every 3 weeks to patients with HER2-positive metastatic breast cancer. *Journal of Clinical Oncology* **28**, 2698–2704 (2010).
179. Geller, J. I. *et al.* ADVL1522: A phase 2 study of lorotuzumab mertansine (IMGN901) in children with relapsed or refractory wilms tumor, rhabdomyosarcoma, neuroblastoma, pleuropulmonary blastoma, malignant peripheral nerve sheath tumor, or synovial sarcoma—A Children’s Oncology Group study. *Cancer* **126**, 5303–5310 (2020).
180. Woll, P. J. *et al.* Efficacy results from a phase I study of lorotuzumab mertansine (IMGN901) in patients with CD56-positive solid tumors. *Journal of Clinical Oncology* **29**, (2011).
181. Staudacher, A. H. & Brown, M. P. Antibody drug conjugates and bystander killing: is antigen-dependent internalisation required. *Br J Cancer* **117**, 1736–1742 (2017).
182. Han, Y., Liu, D. & Li, L. PD-1/PD-L1 pathway: current researches in cancer. *Am J Cancer Res* **10**, 727–742 (2020).
183. Alsaab, H. O. *et al.* PD-1 and PD-L1 checkpoint signaling inhibition for cancer immunotherapy: mechanism, combinations, and clinical outcome. *Front Pharmacol* **8**, (2017).
184. Pardoll, D. M. The blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer* **12**, 252–264 (2012).
185. Lee, H. T., Lee, S. H. & Heo, Y. S. Molecular interactions of antibody drugs targeting PD-1, PD-L1, and CTLA-4 in immuno-oncology. *Molecules* **24**, (2019).
186. Buchbinder, E. I. & Desai, A. CTLA-4 and PD-1 pathways similarities, differences, and implications of their inhibition. *American Journal of Clinical Oncology: Cancer Clinical Trials* **39**, 98–106 (2016).
187. Rotte, A. Combination of CTLA-4 and PD-1 blockers for treatment of cancer. *Journal of Experimental and Clinical Cancer Research* **38**, (2019).
188. Rohaan, M. W. *et al.* Tumor-Infiltrating Lymphocyte Therapy or Ipilimumab in Advanced Melanoma. *New England Journal of Medicine* **387**, 2113–2125 (2022).
189. Völkel, T., Korn, T., Bach, M., Mü, R. & Kontermann, R. E. Optimized Linker Sequences for the Expression of Monomeric and Dimeric Bispecific Single-Chain Diabodies. *Protein Engineering* vol. 14 (2001).

190. Shah, N. N. & Fry, T. J. Mechanisms of resistance to CAR T cell therapy. *Nat Rev Clin Oncol* **16**, 372–385 (2019).
191. Gorovits, B. & Koren, E. Immunogenicity of Chimeric Antigen Receptor T-Cell Therapeutics. *BioDrugs* **33**, 275–284 (2019).
192. Klee, G. G. Human anti-mouse antibodies. *Arch Pathol Lab Med* **124**, 921–923 (2000).
193. Gruber, R. The human antimouse immunoglobulin response and the anti-idiotypic network have no influence on clinical outcome in patients with minimal residual colorectal cancer treated with monoclonal antibody CO17-1A. *Cancer Res* **60**, 1921–1926 (2000).
194. DeNardo, G. L., Bradt, B. M., Mirick, G. R. & DeNardo, S. J. Human antiglobulin response to foreign antibodies: therapeutic benefit? *Cancer Immunology, Immunotherapy* **52**, 309–316 (2003).
195. Muyldermans, S. Nanobodies: Natural Single-Domain Antibodies. *Annual Reviews of Biochemistry* **82**, 775–797 (2013).
196. Ackaert, C. *et al.* Immunogenicity Risk Profile of Nanobodies. *Front Immunol* **12**, (2021).
197. Rossotti, M. A., Bélanger, K., Henry, K. A. & Tanha, J. Immunogenicity and humanization of single-domain antibodies. *Federation of European Biochemical Societies Journal* **289**, 4304–4327 (2022).
198. Brocker, T. & Karjalainen, K. Signals through T Cell Receptor- ζ Chain Alone Are Insufficient to Prime Resting T Lymphocytes. *Journal of Experimental Medicine* **181**, 1653–1659 (1995).
199. Imai, C. *et al.* Chimeric receptors with 4-1BB signaling capacity provoke potent cytotoxicity against acute lymphoblastic leukemia. *Leukemia* **18**, 676–684 (2004).
200. Maher, J., Brentjens, R. J., Gunset, G., Riviere, I. & Sadelain, M. Human T-lymphocyte cytotoxicity and proliferation directed by a single chimeric TCR ζ /CD28 receptor. *Nat Biotechnol* **20**, 70–75 (2002).
201. Van Der Stegen, S. J. C., Hamieh, M. & Sadelain, M. The pharmacology of second-generation chimeric antigen receptors. *Nat Rev Drug Discov* **14**, 499–509 (2015).
202. Chen, Y. J., Abila, B. & Mostafa Kamel, Y. CAR-T: What Is Next? *Cancers (Basel)* **15**, (2023).
203. Subklewe, M., Von Bergwelt-Bailedon, M. & Humpe, A. Chimeric Antigen Receptor T Cells: A Race to Revolutionize Cancer Therapy. *Transfusion Medicine and Hemotherapy* **46**, 15–24 (2019).
204. Li, H., Song, W., Li, Z. & Zhang, M. Preclinical and clinical studies of CAR-NK-cell therapies for malignancies. *Front Immunol* **13**, (2022).
205. Liu, S. *et al.* NK cell-based cancer immunotherapy: from basic biology to clinical development. *J Hematol Oncol* **14**, (2021).
206. Lin, X. *et al.* iPSC-derived CAR-NK cells for cancer immunotherapy. *Biomedicine and Pharmacotherapy* **165**, (2023).
207. Goldenson, B. H., Hor, P. & Kaufman, D. S. iPSC-Derived Natural Killer Cell Therapies - Expansion and Targeting. *Front Immunol* **13**, (2022).
208. Cichocki, F. *et al.* iPSC-derived NK cells maintain high cytotoxicity and enhance in vivo tumor control in concert with T cells and anti-PD-1 therapy. *Sci Transl Med* **12**, (2020).
209. Li, Y., Hermanson, D. L., Moriarity, B. S. & Kaufman, D. S. Human iPSC-Derived Natural Killer Cells Engineered with Chimeric Antigen Receptors Enhance Anti-tumor Activity. *Cell Stem Cell* **23**, 181–192.e5 (2018).
210. You, F. *et al.* A novel CD7 chimeric antigen receptor-modified NK-92MI cell line targeting T-cell acute lymphoblastic leukemia. *Am J Cancer Res* **9**, 64–78 (2019).

211. Sheng, L. *et al.* Cytotoxicity of Donor Natural Killer Cells to Allo-Reactive T Cells Are Related With Acute Graft-vs.-Host-Disease Following Allogeneic Stem Cell Transplantation. *Front Immunol* **11**, (2020).
212. Olson, J. A. *et al.* NK cells mediate reduction of GVHD by inhibiting activated, alloreactive T cells while retaining GVT effects. *Blood* **115**, 4293–4301 (2010).
213. Asai, O. *et al.* Suppression of Graft-Versus-Host Disease and Amplification of Graft-Versus-Tumor Effects by Activated Natural Killer Cells after Allogeneic Bone Marrow Transplantation. *J Clin Invest* **101**, 1835–1842 (1998).
214. Simonetta, F., Alvarez, M. & Negrin, R. S. Natural killer cells in graft-versus-host-disease after allogeneic hematopoietic cell transplantation. *Front Immunol* **8**, (2017).
215. Ruggeri, L. *et al.* Effectiveness of Donor Natural Killer Cell Alloreactivity in Mismatched Hematopoietic Transplants. *Science* (1979) **295**, 2097–2100 (2002).
216. Zamai, L. *et al.* Understanding the Synergy of NKp46 and Co-Activating Signals in Various NK Cell Subpopulations: Paving the Way for More Successful NK-Cell-Based Immunotherapy. *Cells* **9**, (2020).
217. Zhang, L., Meng, Y., Feng, X. & Han, Z. CAR-NK cells for cancer immunotherapy: from bench to bedside. *Biomark Res* **10**, (2022).
218. Bauer, S. *et al.* Activation of NK Cells and T Cells by NKG2D, a Receptor for Stress-Inducible MICA. *Science* (1979) **285**, 727–729 (1999).
219. Fuertes, M. B., Domaica, C. I. & Zwirner, N. W. Leveraging NKG2D Ligands in Immuno-Oncology. *Front Immunol* **12**, (2021).
220. Zingoni, A. *et al.* NKG2D and its ligands: ‘One for all, all for one’. *Front Immunol* **9**, (2018).
221. Raulet, D. H., Gasser, S., Gowen, B. G., Deng, W. & Jung, H. Regulation of ligands for the NKG2D activating receptor. *Annu Rev Immunol* **31**, 413–441 (2013).
222. Boivin, W. A., Cooper, D. M., Hiebert, P. R. & Granville, D. J. Intracellular versus extracellular granzyme B in immunity and disease: Challenging the dogma. *Laboratory Investigation* **89**, 1195–1220 (2009).
223. Ewen, C. L., Kane, K. P. & Bleackley, R. C. A quarter century of granzymes. *Cell Death Differ* **19**, 28–35 (2012).
224. Agaue, S., Hargreaves, A., De Sousa, P., De Waele, P. & Gilham, D. The high expression of NKG2D ligands on tumor and the lack of surface expression on healthy tissues provides a strong rationale to support NKG2D-based therapeutic approaches for cancer. *Annals of Oncology* **29**, viii420 (2018).
225. McGilvray, R. W. *et al.* NKG2D ligand expression in human colorectal cancer reveals associations with prognosis and evidence for immunoediting. *Clinical Cancer Research* **15**, 6993–7002 (2009).
226. Li, K. *et al.* Clinical significance of the NKG2D ligands, MICA/B and ULBP2 in ovarian cancer: high expression of ULBP2 is an indicator of poor prognosis. *Cancer Immunology, Immunotherapy* **58**, 641–652 (2009).
227. Cho, H. *et al.* MICA/B and ULBP1 NKG2D ligands are independent predictors of good prognosis in cervical cancer. *BMC Cancer* **14**, (2014).
228. de Kruijf, E. M. *et al.* NKG2D ligand tumor expression and association with clinical outcome in early breast cancer patients: an observational study. *BMC Cancer* **12**, (2012).
229. Chen, J., Xu, H. & Zhu, X. X. Abnormal expression levels of sMICA and NKG2D are correlated with poor prognosis in pancreatic cancer. *Ther Clin Risk Manag* **12**, 11–18 (2015).
230. Vetter, C. S. *et al.* Expression of Stress-induced MHC Class I Related Chain Molecules on Human Melanoma. *J Invest Dermatol* **118**, 600–605 (2002).

231. Tsukagoshi, M. *et al.* Overexpression of natural killer group 2 member D ligands predicts favorable prognosis in cholangiocarcinoma. *Cancer Sci* **107**, 116–122 (2016).
232. Liu, G., Atteridge, C. L., Wang, X., Lundgren, A. D. & Wu, J. D. Cutting Edge: The Membrane Type Matrix Metalloproteinase MMP14 Mediates Constitutive Shedding of MHC Class I Chain-Related Molecule A Independent of A Disintegrin and Metalloproteinases. *The Journal of Immunology* **184**, 3346–3350 (2010).
233. Salih, H. R., Rammensee, H.-G. & Steinle, A. Cutting Edge: Down-Regulation of MICA on Human Tumors by Proteolytic Shedding. *The Journal of Immunology* **169**, 4098–4102 (2002).
234. Kaiser, B. K. *et al.* Disulphide-isomerase-enabled shedding of tumour-associated NKG2D ligands. *Nature* **447**, 482–486 (2007).
235. Xing, S. & Ferrari de Andrade, L. NKG2D and MICA/B shedding: a ‘tag game’ between NK cells and malignant cells. *Clin Transl Immunology* **9**, (2020).
236. Waldhauer, I. *et al.* Tumor-associated MICA is shed by ADAM proteases. *Cancer Res* **68**, 6368–6376 (2008).
237. Vyas, M. *et al.* Soluble NKG2D ligands in the ovarian cancer microenvironment are associated with an adverse clinical outcome and decreased memory effector T cells independent of NKG2D downregulation. *Oncoimmunology* **6**, (2017).
238. Groh, V., Wu, J., Yee, C. & Spies, T. Tumour-derived soluble MIC ligands impair expression of NKG2D and T-cell activation. *Nature* **419**, 734–738 (2002).
239. Klöß, S. *et al.* Increased sMICA and TGF β 1 levels in HNSCC patients impair NKG2D-dependent functionality of activated NK cells. *Oncoimmunology* **4**, (2015).
240. Jinushi, M., Hodi, F. S. & Dranoff, G. Therapy-induced antibodies to MHC class I chain-related protein A antagonize immune suppression and stimulate antitumor cytotoxicity. *Proceedings of the National Academy of Sciences* **103**, 9190–9195 (2006).
241. Hodi, F. S. *et al.* Immunologic and clinical effects of antibody blockade of cytotoxic T lymphocyte-associated antigen 4 in previously vaccinated cancer patients. *Proceedings of the National Academy of Sciences* **105**, 3005–3010 (2008).
242. Badrinath, S. *et al.* A vaccine targeting resistant tumours by dual T cell plus NK cell attack. *Nature* **606**, 992–998 (2022).
243. De Andrade, L. F. *et al.* Antibody-mediated inhibition of MICA and MICB shedding promotes NK cell-driven tumor immunity. *Science (1979)* **359**, 1537–1542 (2018).
244. Wieczorek, M. *et al.* Major histocompatibility complex (MHC) class I and MHC class II proteins: Conformational plasticity in antigen presentation. *Front Immunol* **8**, (2017).
245. Stern, L. J. & Wiley, D. C. Antigenic peptide binding by class I and class II histocompatibility proteins. *Structure* **2**, 245–251 (1994).
246. Engelhard, V. H. Structure of Peptides Associated with Class I and Class II MHC Molecules. *Annu Rev Immunol* **12**, 181–207 (1994).
247. Bjorkman, P. J. & Parham, P. Structure, Function, and diversity of Class I Major Histocompatibility Complex Molecules. *Annual Reviews of Biochemistry* **59**, 253–288 (1990).
248. van der Merwe, P. A. & Davis, S. J. Molecular Interactions Mediating T Cell Antigen Recognition. *Annu Rev Immunol* **21**, 659–684 (2003).
249. Klein, J. & Sato, A. The HLA System. First of two parts. *N Engl J Med* **343**, 702–709 (2000) doi:10.1056/NEJM200009073431006.
250. Pamer, E. & Cresswell, P. Mechanisms of MHC Class I-Restricted Antigen Processing. *Annu Rev Immunol* **16**, 323–358 (1998).
251. Pugliese, A. Central and peripheral autoantigen presentation in immune tolerance. *Immunology* **111**, 138–146 (2004).

252. Walters, L. C. *et al.* Pathogen-derived HLA-E bound epitopes reveal broad primary anchor pocket tolerability and conformationally malleable peptide binding. *Nat Commun* **9**, (2018).
253. Walters, L. C., McMichael, A. J. & Gillespie, G. M. Detailed and atypical HLA-E peptide binding motifs revealed by a novel peptide exchange binding assay. *Eur J Immunol* **50**, 2075–2091 (2020).
254. Kraemer, T. *et al.* HLA-E: Presentation of a broader peptide repertoire impacts the cellular immune response - Implications on HSCT outcome. *Stem Cells Int* **2015**, (2015).
255. Ruibal, P. *et al.* Peptide Binding to HLA-E Molecules in Humans, Nonhuman Primates, and Mice Reveals Unique Binding Peptides but Remarkably Conserved Anchor Residues. *The Journal of Immunology* **205**, 2861–2872 (2020).
256. Romagnani, C. *et al.* Identification of HLA-E-specific alloreactive T lymphocytes: A cell subset that undergoes preferential expansion in mixed lymphocyte culture and displays a broad cytolytic activity against allogeneic cells. *Proceedings of the National Academy of Sciences* **99**, 11328–11333 (2002).
257. Braud, V., Jones, E. Y. & McMichael, A. The human major histocompatibility complex class Ib molecule HLA-E binds signal sequence-derived peptides with primary anchor residues at positions 2 and 9. *Eur J Immunol* **27**, 1164–1169 (1997).
258. Michaëlsson, J. *et al.* A signal peptide derived from hsp60 binds HLA-E and interferes with CD94/NKG2A recognition. *Journal of Experimental Medicine* **196**, 1403–1414 (2002).
259. Pietra, G., Romagnani, C., Moretta, L. & Mingari, M. C. HLA-E and HLA-E-Bound Peptides: Recognition by Subsets of NK and T Cells. *Current Pharmaceutical Design* **15**, 3336–3344 (2009).
260. Braud, V. M., Allan, D. S. J., Wilson, D. & McMichael, A. J. TAP-and tapasin-dependent HLA-E surface expression correlates with the binding of an MHC class I leader peptide. *Current Biology* **8**, 1–10 (1997).
261. Ishido, S., Wang, C., Lee, B.-S., Cohen, G. B. & Jung, J. U. Downregulation of Major Histocompatibility Complex Class I Molecules by Kaposi's Sarcoma-Associated Herpesvirus K3 and K5 Proteins. *J Virol* **74**, 5300–5309 (2000).
262. Gainey, M. D., Rivenbark, J. G., Cho, H., Yang, L. & Yokoyama, W. M. Viral MHC class I inhibition evades CD8+ T-cell effector responses in vivo but not CD8+ T-cell priming. *Proc Natl Acad Sci U S A* **109**, (2012).
263. Cohen, G. B. *et al.* The Selective Downregulation of Class I Major Histocompatibility Complex Proteins by HIV-1 Protects HIV-Infected Cells from NK Cells. *Immunity* **10**, 661–671 (1999).
264. Petersen, J. L., Morris, C. R. & Solheim, J. C. Virus Evasion of MHC Class I Molecule Presentation. *The Journal of Immunology* **171**, 4473–4478 (2003).
265. Koutsakos, M. *et al.* Downregulation of MHC class I expression by influenza A and B viruses. *Front Immunol* **10**, (2019).
266. Wang, E. C. Y. *et al.* UL40-mediated NK evasion during productive infection with human cytomegalovirus. *Proceedings of the National Academy of Sciences* **99**, (2002).
267. Ulbrecht, M. *et al.* Cutting Edge: The Human Cytomegalovirus UL40 Gene Product Contains a Ligand for HLA-E and Prevents NK Cell-Mediated Lysis. *The Journal of Immunology* **164**, 5019–5022 (2000).
268. Tomasec, P. *et al.* Surface Expression of HLA-E, an Inhibitor of Natural Killer Cells, Enhanced by Human Cytomegalovirus gpUL40. *Science (1979)* **287**, 1031–1033 (2000).
269. Borrego, F., Ulbrecht, M., Weiss, E. H., Coligan, J. E. & Brooks, A. G. Recognition of Human Histocompatibility Leukocyte Antigen (HLA)-E Complexed with HLA Class I Signal Sequence-Derived Peptides by CD94/NKG2 Confers Protection from Natural Killer

- Cell-Mediated Lysis. The Journal of Experimental Medicine* • vol. 187 <http://www.jem.org> (1998).
270. Brooks, A. G. *et al.* Specific Recognition of HLA-E, But Not Classical, HLA Class I Molecules by Soluble CD94/NKG2A and NK Cells. *The Journal of Immunology* **162**, 305–313 (1999).
271. Huisman, B. D. *et al.* An unbiased characterization of the HLA-E and CD94/NKG2x peptide repertoire reveals 1 peptide ligands that skew NK cell activation 2 3. *BioRxiv* (2022) doi:10.1101/2022.08.03.502719.
272. Lee, N. I. *et al.* HLA-E is a major ligand for the natural killer inhibitory receptor CD94NKG2A. *Proceedings of the National Academy of Sciences* **95**, 5199–5204 (1998).
273. Braud, V. M. *et al.* HLA-E binds to natural killer cell receptors CD94/NKG2A, B and C. *Nature* **391**, 795–799 (1998).
274. Io Monaco, E. *et al.* Human leukocyte antigen E contributes to protect tumor cells from lysis by natural killer cells. *Neoplasia* **13**, 822–830 (2011).
275. Gooden, M. *et al.* HLA-E expression by gynecological cancers restrains tumor-infiltrating CD8+ T lymphocytes. *Proc Natl Acad Sci U S A* **108**, 10656–10661 (2011).
276. Ferns, D. M. *et al.* Classical and non-classical HLA class I aberrations in primary cervical squamous- and adenocarcinomas and paired lymph node metastases. *J Immunother Cancer* **4**, (2016).
277. Van Esch, E. M. G. *et al.* Alterations in classical and nonclassical HLA expression in recurrent and progressive HPV-induced usual vulvar intraepithelial neoplasia and implications for immunotherapy. *Int J Cancer* **135**, 830–842 (2014).
278. Le Dréan, E. *et al.* Inhibition of antigen-induced T cell response and antibody-induced NK cell cytotoxicity by NKG2A: Association of NKG2A with SHP-1 and SHP-2 protein-tyrosine phosphatases. *Eur J Immunol* **28**, 264–276 (1998).
279. Wang, X., Xiong, H. & Ning, Z. Implications of NKG2A in immunity and immune-mediated diseases. *Front Immunol* **13**, (2022).
280. Vega, M. A. & Strominger, J. L. Cell Biology Constitutive endocytosis of HLA class I antigens requires a specific portion of the intracytoplasmic tail that shares structural features with other endocytosed molecules. *Proceedings of the National Academy of Sciences* **86**, 2688–2692 (1989).
281. He, W. *et al.* Intracellular trafficking of HLA-E and its regulation. *J Exp Med* **220**, (2023).
282. Barber, C. *et al.* Structure-guided stabilization of pathogen-derived peptide-HLA-E complexes using non-natural amino acids conserves native TCR recognition. *Eur J Immunol* **52**, 618–632 (2022).
283. Hoare, H. L. *et al.* Subtle Changes in Peptide Conformation Profoundly Affect Recognition of the Non-Classical MHC Class I Molecule HLA-E by the CD94-NKG2 Natural Killer Cell Receptors. *J Mol Biol* **377**, 1297–1303 (2008).
284. Strong, R. K. *et al.* HLA-E Allelic Variants: Correlating differential expression, peptide affinities, crystal structures, and thermal stabilities. *Journal of Biological Chemistry* **278**, 5082–5090 (2003).
285. Walters, L. C. *et al.* Primary and secondary functions of HLA-E are determined by stability and conformation of the peptide-bound complexes. *Cell Rep* **39**, (2022).
286. O'Callaghan, C. A. *et al.* Structural Features Impose Tight Peptide Binding Specificity in the Nonclassical MHC Molecule HLA-E. *Mol Cell* **1**, 531–541 (1998).
287. Yazdi, M. T. *et al.* The positive prognostic effect of stromal CD8+ tumor-infiltrating T cells is restrained by the expression of HLA-E in non-small cell lung carcinoma. *Oncotarget* **7**, (2015).
288. Wu, Z. *et al.* HLA-E expression in diffuse glioma: Relationship with clinicopathological features and patient survival. *BMC Neurol* **20**, (2020).

289. Wolpert, F. *et al.* HLA-E contributes to an immune-inhibitory phenotype of glioblastoma stem-like cells. *J Neuroimmunol* **250**, 27–34 (2012).
290. Seliger, B. *et al.* HLA-E expression and its clinical relevance in human renal cell carcinoma. *Oncotarget* **7**, 67360–67372 (2016).
291. Eugène, J. *et al.* The inhibitory receptor CD94/NKG2A on CD8+ tumor-infiltrating lymphocytes in colorectal cancer: a promising new druggable immune checkpoint in the context of HLA-E/β2m overexpression. *Modern Pathology* **33**, 468–482 (2020).
292. Bossard, C. *et al.* HLA-E/β2 microglobulin overexpression in colorectal cancer is associated with recruitment of inhibitory immune cells and tumor progression. *Int J Cancer* **131**, 855–863 (2012).
293. Levy, E. M. *et al.* Human leukocyte antigen-E protein is overexpressed in primary human colorectal cancer. *Int J Oncol* **32**, 633–641 (2008).
294. Zheng, H. *et al.* Human leukocyte antigen-E alleles and expression in patients with serous ovarian cancer. *Cancer Sci* **106**, 522–528 (2015).
295. Andersson, E. *et al.* Non-classical HLA-class I expression in serous ovarian carcinoma: Correlation with the HLA-genotype, tumor infiltrating immune cells and prognosis. *Oncoimmunology* **5**, (2016).
296. Borst, L. *et al.* NKG2A is a late immune checkpoint on CD8 T cells and marks repeated stimulation and cell division. *Int J Cancer* **150**, 688–704 (2022).
297. Hamid, M. A. *et al.* Enriched HLA-E and CD94/NKG2A interaction limits antitumor CD8+ tumor-infiltrating T lymphocyte responses. *Cancer Immunol Res* **7**, 1293–1306 (2019).
298. Sun, C. *et al.* High NKG2A expression contributes to NK cell exhaustion and predicts a poor prognosis of patients with liver cancer. *Oncoimmunology* **6**, (2017).
299. André, P. *et al.* Anti-NKG2A mAb Is a Checkpoint Inhibitor that Promotes Anti-tumor Immunity by Unleashing Both T and NK Cells. *Cell* **175**, 1731–1743.e13 (2018).
300. Greenberg, A. S. *et al.* A new antigen receptor gene family that undergoes rearrangement and extensive somatic diversification in sharks. *Nature* **374**, (1995).
301. Hamers-Casterman, C. *et al.* Naturally occurring antibodies devoid of light chains. *Nature* **363**, 446–448 (1993).
302. Davies, J. & Riechmann, L. Antibody VH domains as small recognition units. *Biotechnology* **13**, 475–479 (1995).
303. Reiter, Y., Schuck, P., Boyd, L. F. & Plaksin, D. An antibody single-domain phage display library of a native heavy chain variable region: Isolation of functional single-domain VH molecules with a unique interface. *J Mol Biol* **290**, 685–698 (1999).
304. Feng, M. *et al.* Therapeutically targeting glycan-3 via a conformation-specific single-domain antibody in hepatocellular carcinoma. *Proceedings of the National Academy of Sciences* **110**, 1–9 (2013).
305. Li, N., Fu, H., Hewitt, S. M., Dimitrov, D. S. & Ho, M. Therapeutically targeting glycan-2 via single-domain antibody-based chimeric antigen receptors and immunotoxins in neuroblastoma. *Proceedings of the National Academy of Sciences* **114**, E6623–E6631 (2017).
306. Chen, W., Zhu, Z., Feng, Y., Xiao, X. & Dimitrov, D. S. Construction of a large phage-displayed human antibody domain library with a scaffold based on a newly identified highly soluble, stable heavy chain variable domain. *J Mol Biol* **382**, 779–789 (2008).
307. Ward, E. S., Guusow, D., Griffiths, A. D., Jones, P. T. & Winter, G. Binding activities of a repertoire of single immunoglobulin variable domains secreted from Escherichia coli. *Nature* **341**, 544–546 (1989).
308. Holt, L. J., Herring, C., Jespers, L. S., Woolven, B. P. & Tomlinson, I. M. Domain antibodies: proteins for therapy. *Trends Biotechnol* **21**, 484–490 (2003).

309. Tanha, J. *et al.* Optimal Design Features of Camelized Human Single-domain Antibody Libraries. *Journal of Biological Chemistry* **276**, 24774–24780 (2001).
310. Van Der Linden, R. H. J. *et al.* Comparison of physical chemical properties of llama VH antibody fragments and mouse monoclonal antibodies. *Biochim Biophys Acta* **1431**, 37–46 (1999).
311. Kijanka, M., Dorresteijn, B., Oliveira, S. & Van Bergen En Henegouwen, P. M. P. Nanobody-based cancer therapy of solid tumors. *Nanomedicine* **10**, 161–174 (2015).
312. Fang, T. *et al.* Nanobody immunostaining for correlated light and electron microscopy with preservation of ultrastructure. *Nat Methods* **15**, 1029–1032 (2018).
313. Tijink, B. M. *et al.* Improved tumor targeting of anti-epidermal growth factor receptor Nanobodies through albumin binding: Taking advantage of modular Nanobody technology. *Mol Cancer Ther* **7**, 2288–2297 (2008).
314. Rashidian, M. *et al.* Immuno-PET identifies the myeloid compartment as a key contributor to the outcome of the antitumor response under PD-1 blockade. *Proceedings of the National Academy of Sciences* **116**, 16971–16980 (2019).
315. Altintas, I. *et al.* Nanobody-albumin nanoparticles (NANAPs) for the delivery of a multikinase inhibitor 17864 to EGFR overexpressing tumor cells. *Journal of Controlled Release* **165**, 110–118 (2013).
316. Iezzi, M. E., Policasro, L., Werbjah, S., Podhajcer, O. & Canziani, G. A. Single-Domain Antibodies and the Promise of Modular Targeting in Cancer imaging and Treatment. *Front Immunol* **9**, 1–11 (2018).
317. Bannas, P., Hambach, J. & Koch-Nolte, F. Nanobodies and nanobody-based human heavy chain antibodies as antitumor therapeutics. *Front Immunol* **8**, 1–13 (2017).
318. Rahbarizadeh, F., Ahmadvand, D. & Sharifzadeh, Z. Nanobody; an Old Concept and New Vehicle for Immunotargeting. *Immunol Invest* **40**, 299–338 (2011).
319. Wesolowski, J. *et al.* Single domain antibodies: promising experimental and therapeutic tools in infection and immunity. *Med Microbiol Immunol* **198**, 157–174 (2009).
320. Hu, Y., Liu, C. & Muyldermans, S. Nanobody-Based Delivery Systems for Diagnosis and Targeted Tumor Therapy. *Front Immunol* **8**, (2017).
321. De Meyer, T., Muyldermans, S. & Depicker, A. Nanobody-based products as research and diagnostic tools. *Trends Biotechnol* **32**, 263–270 (2014).
322. Chanier, T. & Chames, P. Nanobody Engineering: Toward Next Generation Immunotherapies and Immunoimaging of Cancer. *Antibodies* **8**, (2019).
323. Lecocq, Q. *et al.* Theranostics Theranostics in immuno-oncology using nanobody derivatives. *Theranostics* **9**, 7772–7791 (2019).
324. Ingram, J. R., Schmidt, F. I. & Ploegh, H. L. Exploiting Nanobodies' Singular Traits. *Annu Rev Immunol* **36**, (2018).
325. Woodham, A. W. *et al.* Nanobody-antigen conjugates elicit HPV-specific antitumor immune responses. *Cancer Immunol Res* **6**, 870–880 (2018).
326. Cheloha, R. W. *et al.* Improved GPCR ligands from nanobody tethering. *Nat Commun* **11**, 1–11 (2020).
327. Huang, L. *et al.* SPECT imaging with ^{99m}Tc -labeled EGFR-specific nanobody for in vivo monitoring of EGFR expression. *Mol Imaging Biol* **10**, 167–175 (2008).
328. Vaneycken, I. *et al.* Preclinical screening of anti-HER2 nanobodies for molecular imaging of breast cancer. *The FASEB Journal* **25**, 2433–2446 (2011).
329. Jailkhani, N. *et al.* Noninvasive imaging of tumor progression, metastasis, and fibrosis using a nanobody targeting the extracellular matrix. *Proceedings of the National Academy of Sciences* **116**, 14181–14190 (2019).

330. Oda, K., Matsuoka, Y., Funahashi, A. & Kitano, H. A comprehensive pathway map of epidermal growth factor receptor signaling. *Mol Syst Biol* 1–17 (2005) doi:10.1038/msb4100014.
331. Gibson, T. B., Ranganathan, A. & Grothey, A. Randomized Phase III Trial Results of Panitumumab, a Fully Human Anti-Epidermal Growth Factor Receptor Monoclonal Antibody, in Metastatic Colorectal Cancer. *Clin Colorectal Cancer* 6, 29–31 (2006).
332. Roovers, R. C. *et al.* Efficient inhibition of EGFR signalling and of tumour growth by antagonistic anti-EGFR Nanobodies. *Cancer Immunology, Immunotherapy* 56, 303–317 (2007).
333. Gainkam, L. O. T. *et al.* Comparison of the biodistribution and tumor targeting of two 99mTc-labeled anti-EGFR nanobodies in mice, using pinhole SPECT/micro-CT. *Journal of Nuclear Medicine* 49, 788–795 (2008).
334. Roovers, R. C. *et al.* A biparatopic anti-EGFR nanobody efficiently inhibits solid tumour growth. *Int J Cancer* 129, 2013–2024 (2011).
335. Omidfar, K. *et al.* Efficient growth inhibition of EGFR over-expressing tumor cells by an anti-EGFR nanobody. *Mol Biol Rep* 40, 6737–6745 (2013).
336. Kijanka, M. *et al.* Rapid optical imaging of human breast tumour xenografts using anti-HER2 VHJs site-directly conjugated to IRDye 800CW for image-guided surgery. *Eur J Nucl Med Mol Imaging* 40, 1718–1729 (2013).
337. Pruszynski, M. *et al.* Targeting breast carcinoma with radioiodinated anti-HER2 Nanobody. *Nucl Med Biol* 40, 52–59 (2013).
338. Olsson, A.-K., Dimberg, A., Kreuger, J. & Claesson-Welsh, L. VEGF receptor signalling – in control of vascular function. *Nat Rev Mol Cell Biol* 7, 359–371 (2006).
339. Holmes, K., Roberts, O. L., Thomas, A. M. & Cross, M. J. Vascular endothelial growth factor receptor-2: Structure, function, intracellular signalling and therapeutic inhibition. *Cell Signal* 19, 2003–2012 (2007).
340. Behdani, M. *et al.* Generation and characterization of a functional Nanobody against the vascular endothelial growth factor receptor-2; angiogenesis cell receptor. *Mol Immunol* 50, 35–41 (2012).
341. Ma, L. *et al.* Generation and characterization of a human nanobody against VEGFR-2. *Acta Pharmacol Sin* 37, 857–864 (2016).
342. Kazemi-Lomedasht, F. *et al.* Inhibition of angiogenesis in human endothelial cell using VEGF specific nanobody. *Mol Immunol* 65, 58–67 (2015).
343. Kazemi-Lomedasht, F., Muyldermans, S., Habibi-Anbouhi, M. & Behdani, M. Design of a humanized anti vascular endothelial growth factor nanobody and evaluation of its in vitro function. *Iran J Basic Med Sci* 21, 260–266 (2018).
344. Ebrahimizadeh, W., Mousavi Gargari, S. L., Javidan, Z. & Rajabibazl, M. Production of Novel VHH Nanobody Inhibiting Angiogenesis by Targeting Binding Site of VEGF. *Appl Biochem Biotechnol* 176, 1985–1995 (2015).
345. Bottaro, D. P. *et al.* Identification of the hepatocyte growth factor receptor as the c-met proto-oncogene product. *Science (1979)* 251, 802–804 (1991).
346. Gherardi, E., Birchmeier, W., Birchmeier, C. & Vande Woude, G. Targeting MET in cancer: rationale and progress. *Nat Rev Cancer* 12, 89–103 (2012).
347. Boccaccio, C. & Comoglio, P. M. Invasive growth: a MET-driven genetic programme for cancer and stem cells. *Nat Rev Cancer* 6, 637–645 (2006).
348. Wallenius, V. *et al.* Overexpression of the hepatocyte growth factor (HGF) receptor (Met) and presence of a truncated and activated intracellular HGF receptor fragment in locally aggressive/malignant human musculoskeletal tumors. *American Journal of Pathology* 156, 821–829 (2000).

349. Heukers, R. *et al.* Targeting hepatocyte growth factor receptor (Met) positive tumor cells using internalizing nanobody-decorated albumin nanoparticles. *Biomaterials* **35**, 601–610 (2014).
350. Schmidt Slørdahl, T. *et al.* Anti-c-MET Nanobody® - A new potential drug in multiple myeloma treatment. *Eur J Haematol* **91**, 399–410 (2013).
351. Vosjan, M. J. W. D. *et al.* Nanobodies targeting the hepatocyte growth factor: Potential new drugs for molecular cancer therapy. *Mol Cancer Ther* **11**, 1017–1025 (2012).
352. Balkwill, F. Cancer and the chemokine network. *Nat Rev Cancer* **4**, 540–550 (2004).
353. Bradley, M. E. *et al.* Potent and efficacious inhibition of CXCR2 signaling by biparatopic nanobodies combining two distinct modes of action. *Mol Pharmacol* **87**, 251–262 (2015).
354. Jähnichen, S. *et al.* CXCR4 nanobodies (VHH-based single variable domains) potently inhibit chemotaxis and HIV-1 replication and mobilize stem cells. *Proceedings of the National Academy of Sciences* **107**, 20565–20570 (2010).
355. De Wit, R. H. *et al.* CXCR4-specific nanobodies as potential therapeutics for WHIM syndrome. *Journal of Pharmacology and Experimental Therapeutics* **363**, 35–44 (2017).
356. Van Hout, A. *et al.* CXCR4-targeting nanobodies differentially inhibit CXCR4 function and HIV entry. *Biochem Pharmacol* **158**, 402–412 (2018).
357. Maussang, D. *et al.* Llama-derived single variable domains (nanobodies) directed against chemokine receptor CXCR7 reduce head and neck cancer cell growth in vivo. *Journal of Biological Chemistry* **288**, 29562–29572 (2013).
358. Blanchetot, C. *et al.* Neutralizing nanobodies targeting diverse chemokines effectively inhibit chemokine function. *Journal of Biological Chemistry* **288**, 25173–25182 (2013).
359. Burg, J. S. *et al.* Structural basis for chemokine recognition and activation of a viral G protein-coupled receptor John. *Science (1979)* **347**, 1113–1117 (2015).
360. Heukers, R. *et al.* The constitutive activity of the virally encoded chemokine receptor US28 accelerates glioblastoma growth. *Oncogene* **37**, 4110–4121 (2018).
361. De Groof, T. W. M. *et al.* Nanobody-Targeted Photodynamic Therapy Selectively Kills Viral GPCR-Expressing Glioblastoma Cells. *Mol Pharm* **16**, 3145–3156 (2019).
362. Cortez-Retamozo, V. *et al.* Efficient Cancer Therapy with a Nanobody-Based Conjugate. *Cancer Res* **64**, 2853–2857 (2004).
363. Wang, H., Meng, A.-M., Li, S.-H. & Zhou, X.-L. A nanobody targeting carcinoembryonic antigen as a promising molecular probe for non-small cell lung cancer. *Mol Med Rep* **16**, 625–630 (2017).
364. Kaliberov, S. A. *et al.* Adenoviral targeting using genetically incorporated camelid single variable domains. *Laboratory Investigation* **94**, 893–905 (2014).
365. Chatalic, K. L. S. *et al.* A novel mIn-labeled anti-prostate-specific membrane antigen nanobody for targeted SPECT/CT imaging of prostate cancer. *Journal of Nuclear Medicine* **56**, 1094–1099 (2015).
366. Evazalipour, M. *et al.* Generation and characterization of nanobodies targeting PSMA for molecular imaging of prostate cancer. *Contrast Media Mol Imaging* **9**, 211–220 (2014).
367. Zare, H. *et al.* Production of nanobodies against prostate-specific membrane antigen (PSMA) recognizing LnCaP cells. *International Journal of Biological Markers* **29**, 169–179 (2014).
368. Fan, X. *et al.* Ultrasonic nanobubbles carrying anti-PSMA nanobody: Construction and application in prostate cancer-targeted imaging. *PLoS One* **10**, 1–13 (2015).
369. Saerens, D. *et al.* Single domain antibodies derived from dromedary lymph node and peripheral blood lymphocytes sensing conformational variants of prostate-specific antigen. *Journal of Biological Chemistry* **279**, 51965–51972 (2004).

370. Movahedi, K. *et al.* Nanobody-based targeting of the macrophage mannose receptor for effective in vivo imaging of tumor-associated macrophages. *Cancer Res* **72**, 4165–4177 (2012).
371. Blykers, A. *et al.* PET imaging of macrophage mannose receptor-expressing macrophages in tumor stroma using ¹⁸F-radiolabeled camelid single-domain antibody fragments. *Journal of Nuclear Medicine* **56**, 1265–1271 (2015).
372. Tang, J. *et al.* Novel CD7-specific nanobody-based immunotoxins potently enhanced apoptosis of CD7-positive malignant cells. *Oncotarget* **7**, 34070–34083 (2016).
373. Yu, Y. *et al.* Humanized CD7 nanobody-based immunotoxins exhibit promising anti-T-cell acute lymphoblastic leukemia potential. *Int J Nanomedicine* **12**, 1969–1983 (2017).
374. Wan, R. *et al.* Screening and antitumor effect of an anti-CTLA-4 nanobody. *Oncol Rep* **39**, 511–518 (2018).
375. Ingram, J. R. *et al.* Anti-CTLA-4 therapy requires an Fc domain for efficacy. *Proceedings of the National Academy of Sciences* **115**, 3912–3917 (2018).
376. Zhang, F. *et al.* Structural basis of a novel PD-L1 nanobody for immune checkpoint blockade. *Cell Discov* **3**, 1–12 (2017).
377. Broos, K. *et al.* Evaluating a single domain antibody targeting human PD-L1 as a nuclear imaging and therapeutic agent. *Cancers (Basel)* **11**, 1–19 (2019).
378. Broos, K. *et al.* Non-invasive assessment of murine PD-L1 levels in syngeneic tumor models by nuclear imaging with nanobody tracers. *Oncotarget* **8**, 41932–41946 (2017).
379. Xing, Y. *et al.* Early phase I study of a ^{99m}Tc-labeled anti-programmed death ligand-1 (PD-L1) single-domain antibody in SPECT/CT assessment of PD-L1 expression in non-small cell lung cancer. *Journal of Nuclear Medicine* **60**, 1213–1220 (2019).
380. Ingram, J. R. *et al.* PD-L1 is an activation-independent marker of brown adipocytes. *Nat Commun* **8**, (2017).
381. Rashidian, M. *et al.* Predicting the response to CTLA-4 blockade by longitudinal noninvasive monitoring of CD8 T cells. *Journal of Experimental Medicine* **214**, 2243–2255 (2017).
382. Rossotti, M. *et al.* Streamlined method for parallel identification of single domain antibodies to membrane receptors on whole cells. *Biochim Biophys Acta* **1850**, 1397–1404 (2015).
383. Rashidian, M. *et al.* Noninvasive imaging of immune responses. *Proceedings of the National Academy of Sciences* **112**, 6146–6151 (2015).
384. Krasniqi, A. *et al.* Theranostic radiolabeled anti-CD20 sdAb for targeted radionuclide therapy of non-hodgkin lymphoma. *Mol Cancer Ther* **16**, 2828–2839 (2017).
385. Fumey, W. *et al.* Nanobodies effectively modulate the enzymatic activity of CD38 and allow specific imaging of CD38+ tumors in mouse models in vivo. *Sci Rep* **7**, 1–13 (2017).
386. Bachran, C. *et al.* The activity of myeloid cell-specific VHH immunotoxins is target-, epitope-, subset- and organ dependent. *Sci Rep* **7**, 2–11 (2017).
387. Van Elssen, C. H. M. J. *et al.* Noninvasive imaging of human immune responses in a human xenograft model of graft-versus-host disease. *Journal of Nuclear Medicine* **58**, 1003–1008 (2017).
388. Rashidian, M. *et al.* Use of ¹⁸F-2-fluorodeoxyglucose to label antibody fragments for immuno-positron emission tomography of pancreatic cancer. *ACS Cent Sci* **1**, 142–147 (2015).
389. Samec, N. *et al.* Glioblastoma-specific anti-TUFM nanobody for in-vitro immunoimaging and cancer stem cell targeting. *Oncotarget* **9**, 17282–17299 (2018).
390. Van Impe, K. *et al.* A nanobody targeting the F-actin capping protein CapG restrains breast cancer metastasis. *Breast Cancer Research* **15**, 1–15 (2013).

391. Araste, F., Ebrahimizadeh, W., Rasooli, I., Rajabibazl, M. & Mousavi Gargari, S. L. A novel VHH nanobody against the active site (the CA domain) of tumor-associated, carbonic anhydrase isoform IX and its usefulness for cancer diagnosis. *Biotechnol Lett* **36**, 21–28 (2014).
392. van Brussel, A. S. A. *et al.* Hypoxia-Targeting Fluorescent Nanobodies for Optical Molecular Imaging of Pre-Invasive Breast Cancer. *Mol Imaging Biol* **18**, 535–544 (2016).
393. Romão, E. *et al.* Identification of nanobodies against the acute myeloid leukemia marker CD33. *Int J Mol Sci* **21**, (2020).
394. Ma, L. *et al.* Preclinical development of a novel CD47 nanobody with less toxicity and enhanced anti-cancer therapeutic potential. *J Nanobiotechnology* **18**, 1–15 (2020).
395. Sockolosky, J. T. *et al.* Durable antitumor responses to CD47 blockade require adaptive immune stimulation. *Proceedings of the National Academy of Sciences* **113**, E2646–E2654 (2016).
396. Koch-Nolte, F. *et al.* Single domain antibodies from llama effectively and specifically block T cell ecto-ADP-ribosyltransferase ART2.2 in vivo. *The FASEB Journal* **21**, 3490–3498 (2007).
397. Ji, X. *et al.* Neutralization of TNF α in tumor with a novel nanobody potentiates paclitaxel-therapy and inhibits metastasis in breast cancer. *Cancer Lett* **386**, 24–34 (2017).
398. Weissleder, R. Molecular imaging in cancer. *Science (1979)* **312**, 1168–1171 (2006).
399. Hong, H., Zhang, Y., Sun, J. & Cai, W. Molecular imaging and therapy of cancer with radiolabeled nanoparticles. *Nano Today* **4**, 399–413 (2009).
400. de Vos, J., Devoogdt, N., Lahoutte, T. & Muyldermans, S. Camelid single-domain antibody-fragment engineering for (pre)clinical in vivo molecular imaging applications: adjusting the bullet to its target. *Expert Opin Biol Ther* **13**, 1149–1160 (2013).
401. Knowles, S. M. & Wu, A. M. Advances in immuno-positron emission tomography: Antibodies for molecular imaging in oncology. *Journal of Clinical Oncology* **30**, 3884–3892 (2012).
402. Vosjan, M. J. W. D. *et al.* Facile labelling of an anti-epidermal growth factor receptor Nanobody with 68Ga via a novel bifunctional desferal chelate for immuno-PET. *Eur J Nucl Med Mol Imaging* **38**, 753–763 (2011).
403. Oliveira, S. *et al.* Rapid visualization of human tumor xenografts through optical imaging with a near-infrared fluorescent anti-epidermal growth factor receptor nanobody. *Mol Imaging* **11**, 33–46 (2012).
404. Hernot, S. *et al.* Nanobody-coupled microbubbles as novel molecular tracer. *Journal of Controlled Release* **158**, 346–353 (2012).
405. Berger, A. How does it work? Positron emission tomography. *The British Medical Journal Preprint at https://doi.org/10.1136/bmj.326.7404.1449* (2003).
406. Vaidyanathan, G., Bigner, D. D. & Zalutsky, M. R. Fluorine-18-labeled monoclonal antibody fragments: A potential approach for combining radioimmunoscintigraphy and positron emission tomography. *Journal of Nuclear Medicine* **33**, 1535–1541 (1992).
407. Cai, W. *et al.* PET imaging of colorectal cancer in xenograft-bearing mice by use of an 18F-labeled T84.66 anti-carcinoembryonic antigen diabody. *Journal of Nuclear Medicine* **48**, 304–310 (2007).
408. Xavier, C. *et al.* Synthesis, preclinical validation, dosimetry, and toxicity of 68Ga-NOTA-anti-HER2 nanobodies for iPET imaging of HER2 receptor expression in cancer. *Journal of Nuclear Medicine* **54**, 776–784 (2013).
409. Keyaerts, M. *et al.* Phase I study of 68Ga-HER2-Nanobody for PET/CT assessment of HER2 expression in breast carcinoma. *Journal of Nuclear Medicine* **57**, 27–33 (2016).

410. Xavier, C. *et al.* Clinical Translation of [68Ga]Ga-NOTA-anti-MMR-sdAb for PET/CT Imaging of Protumorigenic Macrophages. *Mol Imaging Biol* **21**, 898–906 (2019).
411. Banerjee, S. R. & Pomper, M. G. Clinical applications of Gallium-68. *Applied Radiation and Isotopes* **76**, 2–13 (2013).
412. Garcia-Torano, E. & Ibarra, M. R. The half-life of ¹⁸F. *Applied Radiation and Isotopes* **68**, 1561–1565 (2010).
413. Xavier, C. *et al.* ¹⁸F-nanobody for PET imaging of HER2 overexpressing tumors. *Nucl Med Biol* **43**, 247–252 (2016).
414. Leach, D. R., Krummel, M. F. & Allison, J. P. Enhancement of Antitumor Immunity by CTLA-4 Blockade. *Science* (1979) **271**, 1734–1736 (1996).
415. Lipson, E. J. *et al.* Antagonists of PD-1 and PD-L1 in Cancer Treatment. *Semin Oncol* **42**, 587–600 (2015).
416. Zhang, Y. *et al.* Myeloid cells are required for PD-1/PD-L1 checkpoint activation and the establishment of an immunosuppressive environment in pancreatic cancer. *Gut* **66**, 124–136 (2017).
417. Bocanegra, A. *et al.* PD-L1 expression in systemic immune cell populations as a potential predictive biomarker of responses to PD-L1/PD-1 blockade therapy in lung cancer. *Int J Mol Sci* **20**, 1–13 (2019).
418. Pico De Coaña, Y., Masucci, G., Hansson, J. & Kiessling, R. Myeloid-derived suppressor cells and their role in CTLA-4 blockade therapy. *Cancer Immunology, Immunotherapy* **63**, 977–983 (2014).
419. Dedman, J. R., Gracy, R. W. & Harris, B. G. A method for estimating sequence homology from amino acid compositions. The evolution of Ascaris employing aldolase and glyceraldehyde-3-phosphate dehydrogenase. *Comparative Biochemistry and Physiology B, Basic Health* **49**, 715–731 (1974).
420. Fang, T. *et al.* Targeted antigen delivery by an anti-class II MHC VHH elicits focused α mUC1(Tn) immunity. *Chem Sci* **8**, 5591–5597 (2017).
421. Vaneycken, I. *et al.* In vitro analysis and in vivo tumor targeting of a humanized, grafted Nanobody in mice using pinhole SPECT/micro-CT. *Journal of Nuclear Medicine* **51**, 1099–1106 (2010).
422. Van Driel, P. B. A. A. *et al.* Intraoperative fluorescence delineation of head and neck cancer with a fluorescent Anti-epidermal growth factor receptor nanobody. *Int J Cancer* **134**, 2663–2673 (2014).
423. Debie, P. *et al.* Improved Debunking of Peritoneal Tumor Implants by Near-Infrared Fluorescent Nanobody Image Guidance in an Experimental Mouse Model. *Mol Imaging Biol* **20**, 361–367 (2018).
424. Bannas, P. *et al.* Molecular imaging of tumors with nanobodies and antibodies: Timing and dosage are crucial factors for improved in vivo detection. *Contrast Media Mol Imaging* **10**, 367–378 (2015).
425. Debie, P. *et al.* Effect of dye and conjugation chemistry on the biodistribution profile of near-infrared-labeled nanobodies as tracers for image-guided surgery. *Mol Pharm* **14**, 1145–1153 (2017).
426. Zhang, J. *et al.* The optimized fabrication of a novel nanobubble for tumor imaging. *Front Pharmacol* **10**, 1–15 (2019).
427. Kogan, P., Gessner, R. C. & Dayton, P. A. Microbubbles in imaging: Applications beyond ultrasound. *Bubble Sci Eng Technol* **2**, 3–8 (2010).
428. Yin, T. *et al.* Nanobubbles for enhanced ultrasound imaging of tumors. *Int J Nanomedicine* **7**, 895–904 (2012).

429. Gainkam, L. O. T. *et al.* Localization, mechanism and reduction of renal retention of technetium-99m labeled epidermal growth factor receptor-specific nanobody in mice. *Contrast Media Mol Imaging* **6**, 85–92 (2011).
430. Zhou, Z., Devoogdt, N., Zalutsky, M. R. & Vaidyanathan, G. An Efficient Method for Labeling Single Domain Antibody Fragments with ¹⁸F Using Tetrazine-Trans-Cyclooctene Ligation and a Renal Brush Border Enzyme-Cleavable Linker. *Bioconjug Chem* **29**, 4090–4103 (2018).
431. Baumeister, S. H., Freeman, G. J., Dranoff, G. & Sharpe, A. H. Coinhibitory Pathways in Immunotherapy for Cancer. *Annu Rev Immunol* **34**, 539–573 (2016).
432. Curran, M. A., Montalvo, W., Yagita, H. & Allison, J. P. PD-1 and CTLA-4 combination blockade expands infiltrating T cells and reduces regulatory T and myeloid cells within Bi6 melanoma tumors. *Proceedings of the National Academy of Sciences* **107**, 4275–4280 (2010).
433. Ingram, J. R. *et al.* Localized CD47 blockade enhances immunotherapy for murine melanoma. *Proceedings of the National Academy of Sciences* **114**, 2–7 (2017).
434. Peters, C. & Brown, S. Antibody-drug conjugates as novel anti-cancer therapeutics. *Biosci Rep* **35**, (2015).
435. Thomas, A., Teicher, B. A. & Hassan, R. Antibody-drug conjugates for cancer therapy. *Lancet Oncology* **17**, e254–e262 (2016).
436. Fang, T. *et al.* Structurally-defined α MHC-II nanobody-drug conjugates: Therapeutic and imaging platforms for B-cell lymphoma. *Angewandte Chemie International Edition* **55**, 2416–2420 (2016).
437. Helft, J. *et al.* GM-CSF Mouse Bone Marrow Cultures Comprise a Heterogeneous Population of CD11c+MHCII+ Macrophages and Dendritic Cells. *Immunity* **42**, 1197–1211 (2015).
438. Becher, B. *et al.* High-dimensional analysis of the murine myeloid cell system. *Nat Immunol* **15**, 1181–1189 (2014).
439. Knox, S. J. *et al.* Yttrium-90-labeled Anti-CD20 Monoclonal Antibody Therapy of Recurrent B-Cell Lymphoma. *Clinical Cancer Research* **2**, 457–470 (1996).
440. Witzig, T. E. *et al.* Randomized controlled trial of yttrium-90-labeled ibritumomab tiuxetan radioimmunotherapy versus rituximab immunotherapy for patients with relapsed or refractory low-grade, follicular, or transformed B-cell non-Hodgkin's lymphoma. *Journal of Clinical Oncology* **20**, 2453–2463 (2002).
441. Witzig, T. E. *et al.* Phase I/II Trial of IDEC-Y2B8 Radioimmunotherapy for Treatment of Relapsed or Refractory CD20+ B-Cell Non-Hodgkin's Lymphoma. *Journal of Clinical Oncology* **17**, 3793–3803 (1999).
442. Vose, J. M. *et al.* Phase II Trial of ¹³¹Iodine Tositumomab with High-Dose Chemotherapy and Autologous Stem Cell Transplantation for Relapsed Diffuse Large B Cell Lymphoma. *Biology of Blood and Marrow Transplantation* **19**, 123–128 (2013).
443. Jurcic, J. G. *et al.* Targeted α particle immunotherapy for myeloid leukemia. *Blood* **100**, 1233–1239 (2002).
444. Sgouros, G. *et al.* Pharmacokinetics and dosimetry of an α -particle emitter labeled antibody: ²¹³Bi-HuM195 (anti-CD33) in patients with leukemia. *Journal of Nuclear Medicine* **40**, 1935–1946 (1999).
445. Mattes, M. J., Sharkey, R. M., Karacay, H., Czuczman, M. S. & Goldenberg, D. M. Therapy of advanced B-lymphoma xenografts with a combination of ⁹⁰Y-anti-CD22 IgG (epratuzumab) and unlabeled anti-CD20 IgG (veltuzumab). *Clinical Cancer Research* **14**, 6154–6160 (2008).

446. Sharkey, R. M., Press, O. W. & Goldenberg, D. M. A re-examination of radioimmunotherapy in the treatment of non-Hodgkin lymphoma: prospects for dual-targeted antibody/radioantibody therapy. *Blood* **113**, 3891–3895 (2009).
447. Verheijen, R. H. *et al.* Phase III trial of intraperitoneal therapy with yttrium-90-labeled HMFG1 murine monoclonal antibody in patients with epithelial ovarian cancer after a surgically defined complete remission. *Journal of Clinical Oncology* **24**, 571–578 (2006).
448. Wong, J. Y. C. *et al.* A Phase I Radioimmunotherapy Trial Evaluating 90 Yttrium-labeled Anti-Carcinoembryonic Antigen (CEA) Chimeric T84.66 in Patients with Metastatic CEA-producing Malignancies. *Clinical Cancer Research* **6**, 3855–3863 (2000).
449. Liersch, T. *et al.* Phase II trial of carcinoembryonic antigen radioimmunotherapy with ^{131}I -labetuzumab after salvage resection of colorectal metastases in the liver: five-year safety and efficacy results. *Journal of Clinical Oncology* **23**, 6763–6770 (2005).
450. Myers, R. *et al.* Toxicology study of repeat intracerebral administration of a measles virus derivative producing carcinoembryonic antigen in rhesus macaques in support of a phase I/II clinical trial for patients with recurrent gliomas. *Hum Gene Ther* **19**, 690–698 (2008).
451. Meredith, R. F. *et al.* Phase II Study with Interferon of Dual ^{131}I -labeled Monoclonal Antibody Therapy in Patients with Metastatic Colorectal. *Clinical Cancer Research* **2**, 1811–1818 (1996).
452. D’Huyvetter, M. *et al.* Targeted radionuclide therapy with A ^{177}Lu -labeled anti-HER2 nanobody. *Theranostics* **4**, 708–720 (2014).
453. D’Huyvetter, M. *et al.* ^{131}I -labeled anti-HER2 camelid sdAb as a theranostic tool in cancer treatment. *Clinical Cancer Research* **23**, 6616–6628 (2017).
454. Stein, R. *et al.* Advantage of a residualizing iodine radiolabel in the therapy of a colon cancer xenograft targeted with an anticarcinoembryonic antigen monoclonal antibody. *Clinical Cancer Research* **11**, 2727–2734 (2005).
455. Pruszynski, M. *et al.* Improved tumor targeting of anti-her2 nanobody through n-succinimidyl 4-guanidinomethyl-3-iodobenzoate radiolabeling. *Journal of Nuclear Medicine* **55**, 650–656 (2014).
456. Torchilin, V. P. Multifunctional nanocarriers. *Adv Drug Deliv Rev* **58**, 1532–1555 (2006).
457. Oliveira, S. *et al.* Downregulation of EGFR by a novel multivalent nanobody-liposome platform. *Journal of Controlled Release* **145**, 165–175 (2010).
458. Talelli, M. *et al.* Nanobody - Shell functionalized thermosensitive core-crosslinked polymeric micelles for active drug targeting. *Journal of Controlled Release* **151**, 183–192 (2011).
459. Liu, Y. *et al.* EGFR-Targeted Nanobody Functionalized Polymeric Micelles Loaded with mTHPC for Selective Photodynamic Therapy. *Mol Pharm* **17**, 1276–1292 (2020).
460. Xu, R. *et al.* Extracellular vesicles in cancer — implications for future improvements in cancer care. *Nat Rev Clin Oncol* **15**, 617–638 (2018).
461. Kooijmans, S. A. A. *et al.* Display of GPI-anchored anti-EGFR nanobodies on extracellular vesicles promotes tumour cell targeting. *J Extracell Vesicles* **5**, 1–11 (2016).
462. Breckpot, K., Aerts, J. L. & Thielemans, K. Lentiviral vectors for cancer immunotherapy: transforming infectious particles into therapeutics. *Gene Ther* **14**, 847–862 (2007).
463. Gennari, F., Lopes, L., Verhoeven, E., Marasco, W. & Collins, M. K. Single-Chain Antibodies That Target Lentiviral Vectors to MHC Class II on Antigen-Presenting Cells. *Hum Gene Ther* **20**, 554–562 (2009).
464. Goyvaerts, C. *et al.* Development of the Nanobody display technology to target lentiviral vectors to antigen-presenting cells. *Gene Ther* **19**, 1133–1140 (2012).
465. Eichhoff, A. M. *et al.* Nanobody-Enhanced Targeting of AAV Gene Therapy Vectors. *Mol Ther Methods Clin Dev* **15**, 211–220 (2019).

466. Duarte, J. N. *et al.* Generation of Immunity against Pathogens via Single-Domain Antibody-Antigen Constructs. *Journal of Immunology* **197**, 4838–4847 (2016).
467. Chinnasamy, D. *et al.* Gene therapy using genetically modified lymphocytes targeting VEGFR-2 inhibits the growth of vascularized syngenic tumors in mice. *Journal of Clinical Investigation* **120**, 3953–3968 (2010).
468. Davila, M. L. *et al.* Efficacy and Toxicity Management of 19-28z CAR T Cell Therapy. *Sci Transl Med* **6**, (2014).
469. Maude, S. L. *et al.* Chimeric antigen receptor T cells for sustained remissions in leukemia. *New England Journal of Medicine* **371**, 1507–1517 (2014).
470. Lee, D. W. *et al.* T cells expressing CD19 chimeric antigen receptors for acute lymphoblastic leukaemia in children and young adults: a phase 1 dose-escalation trial. *Lancet Oncology* **385**, 517–528 (2015).
471. Gorovits, B. & Koren, E. Immunogenicity of Chimeric Antigen Receptor T-Cell Therapeutics. *BioDrugs* **33**, 275–284 (2019).
472. Albert, S. *et al.* A novel nanobody-based target module for retargeting of T lymphocytes to EGFR-expressing cancer cells via the modular UniCAR platform. *Oncoimmunology* **6**, 1–17 (2017).
473. Albert, S. *et al.* From mono- to bivalent: Improving theranostic properties of target modules for redirection of UniCAR T cells against EGFR-expressing tumor cells in vitro and in vivo. *Oncotarget* **9**, 25597–25616 (2018).
474. Hajari Taheri, F. *et al.* T cell engineered with a novel nanobody-based chimeric antigen receptor against VEGFR2 as a candidate for tumor immunotherapy. *IUBMB Life* **71**, 1259–1267 (2019).
475. De Munter, S. *et al.* Nanobody based dual specific CARs. *Int J Mol Sci* **19**, 1–11 (2018).
476. Xie, Y. J. *et al.* Nanobody-based CAR T cells that target the tumor microenvironment inhibit the growth of solid tumors in immunocompetent mice. *Proceedings of the National Academy of Sciences* **116**, 7624–7631 (2019).
477. Xie, Y. J. *et al.* Improved Antitumor Efficacy of Chimeric Antigen Receptor T Cells that Secrete Single-Domain Antibody Fragments. *Cancer Immunol Res* **8**, 518–530 (2020).
478. Hambach, J. *et al.* Targeting CD38-Expressing Multiple Myeloma and Burkitt Lymphoma Cells In Vitro with Nanobody-Based Chimeric Antigen Receptors (Nb-CARs). *Cells* **9**, 1–14 (2020).
479. Koch-Nolte, F. *et al.* Single domain antibodies from llama effectively and specifically block T cell ecto-ADP-ribosyltransferase ART2.2 in vivo. *The FASEB Journal* **21**, 3490–3498 (2007).
480. Araste, F., Ebrahimizadeh, W., Rasooli, I., Rajabibazl, M. & Mousavi Gargari, S. L. A novel VHH nanobody against the active site (the CA domain) of tumor-associated, carbonic anhydrase isoform IX and its usefulness for cancer diagnosis. *Biotechnol Lett* **36**, 21–28 (2014).
481. van Brussel, A. S. A. *et al.* Hypoxia-Targeting Fluorescent Nanobodies for Optical Molecular Imaging of Pre-Invasive Breast Cancer. *Mol Imaging Biol* **18**, 535–544 (2016).
482. Van Impe, K. *et al.* A nanobody targeting the F-actin capping protein CapG restrains breast cancer metastasis. *Breast Cancer Research* **15**, 1–15 (2013).
483. Rossotti, M. *et al.* Streamlined method for parallel identification of single domain antibodies to membrane receptors on whole cells. *Biochim Biophys Acta* **1850**, 1397–1404 (2015).
484. Woodham, A. W. *et al.* Nanobody-antigen conjugates elicit HPV-specific antitumor immune responses. *Cancer Immunol Res* **6**, 870–880 (2018).
485. Krasniqi, A. *et al.* Theranostic radiolabeled anti-CD20 sdAb for targeted radionuclide therapy of non-hodgkin lymphoma. *Mol Cancer Ther* **16**, 2828–2839 (2017).

486. Romão, E. *et al.* Identification of nanobodies against the acute myeloid leukemia marker CD33. *Int J Mol Sci* **21**, (2020).
487. Li, T. *et al.* Immuno-targeting the multifunctional CD38 using nanobody. *Nature Publishing Group* **6**, 1–11 (2016).
488. Fumey, W. *et al.* Nanobodies effectively modulate the enzymatic activity of CD38 and allow specific imaging of CD38+ tumors in mouse models in vivo. *Sci Rep* **7**, 1–13 (2017).
489. Sockolosky, J. T. *et al.* Durable antitumor responses to CD47 blockade require adaptive immune stimulation. *PNAS* **113**, E2646–54 (2016).
490. Ingram, J. R. *et al.* Localized CD47 blockade enhances immunotherapy for murine melanoma. *PNAS* **114**, 2–7 (2017).
491. Ma, L. *et al.* Preclinical development of a novel CD47 nanobody with less toxicity and enhanced anti-cancer therapeutic potential. *J Nanobiotechnology* **18**, 1–15 (2020).
492. Tang, J. *et al.* Novel CD7-specific nanobody-based immunotoxins potently enhanced apoptosis of CD7-positive malignant cells. *Oncotarget* **7**, 34070–34083 (2016).
493. Yu, Y. *et al.* Humanized CD7 nanobody-based immunotoxins exhibit promising anti-T-cell acute lymphoblastic leukemia potential. *Int J Nanomedicine* **12**, 1969–1983 (2017).
494. Rashidian, M. *et al.* Predicting the response to CTLA-4 blockade by longitudinal noninvasive monitoring of CD8 T cells. *Journal of Experimental Medicine* **214**, 2243–2255 (2017).
495. Cortez-Retamozo, V. *et al.* Efficient Cancer Therapy with a Nanobody-Based Conjugate. *Cancer Res* **64**, 2853–2857 (2004).
496. Kaliberov, S. A. *et al.* Adenoviral targeting using genetically incorporated camelid single variable domains. *Laboratory Investigation* **94**, 893–905 (2014).
497. Wang, H., Meng, A.-M., Li, S.-H. & Zhou, X.-L. A nanobody targeting carcinoembryonic antigen as a promising molecular probe for non-small cell lung cancer. *Mol Med Rep* **16**, 625–630 (2017).
498. Schmidt Slørdahl, T. *et al.* Anti-c-MET Nanobody® - A new potential drug in multiple myeloma treatment. *Eur J Haematol* **91**, 399–410 (2013).
499. Heukers, R. *et al.* Targeting hepatocyte growth factor receptor (Met) positive tumor cells using internalizing nanobody-decorated albumin nanoparticles. *Biomaterials* **35**, 601–610 (2014).
500. Wan, R. *et al.* Screening and antitumor effect of an anti-CTLA-4 nanobody. *Oncol Rep* **39**, 511–518 (2018).
501. Ingram, J. R. *et al.* Anti-CTLA-4 therapy requires an Fc domain for efficacy. *PNAS* **115**, 3912–3917 (2018).
502. Blanchetot, C. *et al.* Neutralizing nanobodies targeting diverse chemokines effectively inhibit chemokine function. *Journal of Biological Chemistry* **288**, 25173–25182 (2013).
503. Bradley, M. E. *et al.* Potent and efficacious inhibition of CXCR2 signaling by biparatopic nanobodies combining two distinct modes of action. *Mol Pharmacol* **87**, 251–262 (2015).
504. Jähnichen, S. *et al.* CXCR4 nanobodies (VHH-based single variable domains) potently inhibit chemotaxis and HIV-1 replication and mobilize stem cells. *PNAS* **107**, 20565–20570 (2010).
505. De Wit, R. H. *et al.* CXCR4-specific nanobodies as potential therapeutics for WHIM syndrome. *Journal of Pharmacology and Experimental Therapeutics* **363**, 35–44 (2017).
506. Van Hout, A. *et al.* CXCR4-targeting nanobodies differentially inhibit CXCR4 function and HIV entry. *Biochem Pharmacol* **158**, 402–412 (2018).
507. Maussang, D. *et al.* Llama-derived single variable domains (nanobodies) directed against chemokine receptor CXCR7 reduce head and neck cancer cell growth in vivo. *Journal of Biological Chemistry* **288**, 29562–29572 (2013).

508. Omidfar, K. *et al.* Production of a novel camel single-domain antibody specific for the type III mutant EGFR. *Tumor Biology* **25**, 296–305 (2004).
509. Roovers, R. C. *et al.* Efficient inhibition of EGFR signalling and of tumour growth by antagonistic anti-EGFR Nanobodies. *Cancer Immunology, Immunotherapy* **56**, 303–317 (2007).
510. Hofman, E. G. *et al.* EGF induces coalescence of different lipid rafts. *J Cell Sci* **121**, 2519–2528 (2008).
511. Huang, L. *et al.* SPECT imaging with 99mTc-labeled EGFR-specific nanobody for in vivo monitoring of EGFR expression. *Mol Imaging Biol* **10**, 167–175 (2008).
512. Tijink, B. M. *et al.* Improved tumor targeting of anti-epidermal growth factor receptor Nanobodies through albumin binding: Taking advantage of modular Nanobody technology. *Mol Cancer Ther* **7**, 2288–2297 (2008).
513. Gainkam, L. O. T. *et al.* Comparison of the biodistribution and tumor targeting of two 99mTc-labeled anti-EGFR nanobodies in mice, using pinhole SPECT/micro-CT. *Journal of Nuclear Medicine* **49**, 788–795 (2008).
514. Roovers, R. C. *et al.* A biparatopic anti-EGFR nanobody efficiently inhibits solid tumour growth. *Int J Cancer* **129**, 2013–2024 (2011).
515. Omidfar, K. *et al.* Efficient growth inhibition of EGFR over-expressing tumor cells by an anti-EGFR nanobody. *Mol Biol Rep* **40**, 6737–6745 (2013).
516. Jailkhani, N. *et al.* Noninvasive imaging of tumor progression, metastasis, and fibrosis using a nanobody targeting the extracellular matrix. *PNAS* **116**, 14181–14190 (2019).
517. Vaneycken, I. *et al.* Preclinical screening of anti-HER2 nanobodies for molecular imaging of breast cancer. *The FASEB Journal* **25**, 2433–2446 (2011).
518. D'Huyvetter, M. *et al.* Targeted radionuclide therapy with A 177Lu-labeled anti-HER2 nanobody. *Theranostics* **4**, 708–720 (2014).
519. Kijanka, M. *et al.* Rapid optical imaging of human breast tumour xenografts using anti-HER2 VHJs site-directly conjugated to IRDye 800CW for image-guided surgery. *Eur J Nucl Med Mol Imaging* **40**, 1718–1729 (2013).
520. Pruszynski, M. *et al.* Targeting breast carcinoma with radioiodinated anti-HER2 Nanobody. *Nucl Med Biol* **40**, 52–59 (2013).
521. Vosjan, M. J. W. D. *et al.* Nanobodies targeting the hepatocyte growth factor: Potential new drugs for molecular cancer therapy. *Mol Cancer Ther* **11**, 1017–1025 (2012).
522. Bachran, C. *et al.* The activity of myeloid cell-specific VHH immunotoxins is target-, epitope-, subset- and organ dependent. *Sci Rep* **7**, 2–11 (2017).
523. Rashidian, M. *et al.* Use of 18F-2-fluorodeoxyglucose to label antibody fragments for immuno-positron emission tomography of pancreatic cancer. *ACS Cent Sci* **1**, 142–147 (2015).
524. Van Elssen, C. H. M. J. *et al.* Noninvasive imaging of human immune responses in a human xenograft model of graft-versus-host disease. *Journal of Nuclear Medicine* **58**, 1003–1008 (2017).
525. Movahedi, K. *et al.* Nanobody-based targeting of the macrophage mannose receptor for effective in vivo imaging of tumor-associated macrophages. *Cancer Res* **72**, 4165–4177 (2012).
526. Blykers, A. *et al.* PET imaging of macrophage mannose receptor-expressing macrophages in tumor stroma using 18F-radiolabeled camelid single-domain antibody fragments. *Journal of Nuclear Medicine* **56**, 1265–1271 (2015).
527. Zhang, Y. *et al.* Myeloid cells are required for PD-1/PD-L1 checkpoint activation and the establishment of an immunosuppressive environment in pancreatic cancer. *Gut* **66**, 124–136 (2017).

528. Broos, K. *et al.* Non-invasive assessment of murine PD-L1 levels in syngeneic tumor models by nuclear imaging with nanobody tracers. *Oncotarget* **8**, 41932–41946 (2017).
529. Broos, K. *et al.* Evaluating a single domain antibody targeting human PD-L1 as a nuclear imaging and therapeutic agent. *Cancers (Basel)* **11**, 1–19 (2019).
530. Xing, Y. *et al.* Early phase I study of a ^{99m}Tc -labeled anti-programmed death ligand-1 (PD-L1) single-domain antibody in SPECT/CT assessment of PD-L1 expression in non-small cell lung cancer. *Journal of Nuclear Medicine* **60**, 1213–1220 (2019).
531. Saerens, D. *et al.* Single domain antibodies derived from dromedary lymph node and peripheral blood lymphocytes sensing conformational variants of prostate-specific antigen. *Journal of Biological Chemistry* **279**, 51965–51972 (2004).
532. Zare, H. *et al.* Production of nanobodies against prostate-specific membrane antigen (PSMA) recognizing LnCaP cells. *International Journal of Biological Markers* **29**, 169–179 (2014).
533. Evazalipour, M. *et al.* Generation and characterization of nanobodies targeting PSMA for molecular imaging of prostate cancer. *Contrast Media Mol Imaging* **9**, 211–220 (2014).
534. Chatalic, K. L. S. *et al.* A novel ^{111}In -labeled anti-prostate-specific membrane antigen nanobody for targeted SPECT/CT imaging of prostate cancer. *Journal of Nuclear Medicine* **56**, 1094–1099 (2015).
535. Fan, X. *et al.* Ultrasonic nanobubbles carrying anti-PSMA nanobody: Construction and application in prostate cancer-targeted imaging. *PLoS One* **10**, 1–13 (2015).
536. Ji, X. *et al.* Neutralization of TNF α in tumor with a novel nanobody potentiates paclitaxel-therapy and inhibits metastasis in breast cancer. *Cancer Lett* **386**, 24–34 (2017).
537. Samec, N. *et al.* Glioblastoma-specific anti-TUFM nanobody for in-vitro immunoimaging and cancer stem cell targeting. *Oncotarget* **9**, 17282–17299 (2018).
538. Behdani, M. *et al.* Generation and characterization of a functional Nanobody against the vascular endothelial growth factor receptor-2; angiogenesis cell receptor. *Mol Immunol* **50**, 35–41 (2012).
539. Kazemi-Lomedasht, F. *et al.* Inhibition of angiogenesis in human endothelial cell using VEGF specific nanobody. *Mol Immunol* **65**, 58–67 (2015).
540. Kazemi-Lomedasht, F., Muyldermans, S., Habibi-Anbouhi, M. & Behdani, M. Design of a humanized anti vascular endothelial growth factor nanobody and evaluation of its in vitro function. *Iran J Basic Med Sci* **21**, 260–266 (2018).
541. Ma, L. *et al.* Generation and characterization of a human nanobody against VEGFR-2. *Acta Pharmacol Sin* **37**, 857–864 (2016).
542. Ebrahimizadeh, W., Mousavi Gargari, S. L., Javidan, Z. & Rajabibazl, M. Production of Novel VHH Nanobody Inhibiting Angiogenesis by Targeting Binding Site of VEGF. *Appl Biochem Biotechnol* **176**, 1985–1995 (2015).
543. Heukers, R. *et al.* The constitutive activity of the virally encoded chemokine receptor US28 accelerates glioblastoma growth. *Oncogene* **37**, 4110–4121 (2018).
544. De Groof, T. W. M. *et al.* Nanobody-Targeted Photodynamic Therapy Selectively Kills Viral GPCR-Expressing Glioblastoma Cells. *Mol Pharm* **16**, 3145–3156 (2019).
545. Burg, J. S. *et al.* Structural basis for chemokine recognition and activation of a viral G protein-coupled receptor John. *Science (1979)* **347**, 1113–1117 (2015).
546. Rajabzadeh, A., Ahmadvand, D., Salmani, M. K., Rahbarizadeh, F. & Hamidieh, A. A. A VHH-based anti-MUC1 chimeric antigen receptor for specific retargeting of human primary T cells to MUC1-positive cancer cells. *Cell J* **22**, 502–513 (2021).
547. Hajari Taheri, F. *et al.* T cell engineered with a novel nanobody-based chimeric antigen receptor against VEGFR2 as a candidate for tumor immunotherapy. *JUBMB Life* **71**, 1259–1267 (2019).

548. Jamnani, F. R. *et al.* T cells expressing VHH-directed oligoclonal chimeric HER2 antigen receptors: Towards tumor-directed oligoclonal T cell therapy. *Biochimica et Biophysica Acta (BBA) - General Subjects* **1840**, 378–386 (2014).
549. Rahbarizadeh, F., Ahmadvand, D. & Moghimi, S. CAR T-cell bioengineering: Single variable domain of heavy chain antibody targeted CARs. *Adv Drug Deliv Rev* **141**, 41–46 (2019).
550. Bao, C. *et al.* The application of nanobody in CAR-T therapy. *Biomolecules* **11**, 1–18 (2021).
551. Zajc, C. U. *et al.* Driving CARs with alternative navigation tools – the potential of engineered binding scaffolds. *FEBS Journal* **288**, 2103–2118 (2021).
552. Pardon, E. *et al.* A general protocol for the generation of Nanobodies for structural biology. *Nat Protoc* **9**, 674–693 (2014).
553. Jeong, H. J., Abhiraman, G. C., Story, C. M., Ingram, J. R. & Dougan, S. K. Generation of Ca²⁺-independent sortase A mutants with enhanced activity for protein and cell surface labeling. *PLoS One* **12**, (2017).
554. Beatty, J. D., Beatty, B. G. & Vlahos, W. G. Measurement of monoclonal antibody affinity by non-competitive enzyme immunoassay. *J Immunol Methods* **100**, 173–179 (1987).
555. Truong, T. T. T., Huynh, V. Q., Vo, N. T. & Nguyen, H. D. Studying the characteristics of nanobody CDR regions based on sequence analysis in combination with 3D structures. *Journal of Genetic Engineering and Biotechnology* **20**, (2022).
556. Klarenbeek, A. *et al.* Camelid Ig V genes reveal significant human homology not seen in therapeutic target genes, providing for a powerful therapeutic antibody platform. *MAbs* **7**, 693–706 (2015).
557. Klussmeier, A. *et al.* High-Throughput MICA/B Genotyping of Over Two Million Samples: Workflow and Allele Frequencies. *Front Immunol* **11**, (2020).
558. Fang, T. *et al.* Structurally Defined αMHC-II Nanobody-Drug Conjugates: A Therapeutic and Imaging System for B-Cell Lymphoma. *Angewandte Chemie - International Edition* **55**, 2416–2420 (2016).
559. Hervier, B. *et al.* Increased Concentrations of Circulating Soluble MHC Class I-Related Chain A (sMICA) and sMICB and Modulation of Plasma Membrane MICA Expression: Potential Mechanisms and Correlation With Natural Killer Cell Activity in Systemic Lupus Erythematosus. *Front Immunol* **12**, (2021).
560. Li, J. J. *et al.* Prognostic value of soluble MICA levels in the serum of patients with advanced hepatocellular carcinoma. *Chin J Cancer* **32**, 141–148 (2013).
561. Arai, J. *et al.* Baseline soluble MICA levels act as a predictive biomarker for the efficacy of regorafenib treatment in colorectal cancer. *BMC Cancer* **22**, (2022).
562. Henry Dunand, C. J. & Wilson, P. C. Restricted, canonical, stereotyped and convergent immunoglobulin responses. *Philos Trans R Soc Lond B Biol Sci* **370**, (2015).
563. Tian, C. *et al.* Genome-wide association and HLA region fine-mapping studies identify susceptibility loci for multiple common infections. *Nat Commun* **8**, (2017).
564. Tsuji, I. *et al.* Somatic Hypermutation and Framework Mutations of Variable Region Contribute to Anti-Zika Virus-Specific Monoclonal Antibody Binding and Function. *J Virol* **96**, (2022).
565. Klein, F. *et al.* Somatic mutations of the immunoglobulin framework are generally required for broad and potent HIV-1 neutralization. *Cell* **153**, 126–138 (2013).
566. Briney, B. S., Willis, J. R. & Crowe, J. E. Location and length distribution of somatic hypermutation-associated DNA insertions and deletions reveals regions of antibody structural plasticity. *Genes Immun* **13**, 523–529 (2012).
567. Wilson, P. C. *et al.* Somatic Hypermutation Introduces Insertions and Deletions into Immunoglobulin V Genes. *Journal of Experimental Medicine* **187**, 59–70 (1998).

568. Bemark, M. & Neuberger, M. S. By-products of immunoglobulin somatic hypermutation. *Genes Chromosomes Cancer* **38**, 32–39 (2003).
569. Koskela, S. *et al.* MICA and MICB allele assortment in Finland. *HLA Immune Response Genetics* **102**, 52–61 (2023).
570. Fuentes-Antrás, J., Genta, S., Vijenthira, A. & Siu, L. L. Antibody-drug conjugates: in search of partners of choice. *Trends Cancer* **9**, 339–354 (2023).
571. Fu, Y. & Ho, M. DNA damaging agent-based antibody-drug conjugates for cancer therapy. *Antib Ther* **1**, 43–53 (2018).
572. Fuentes-Antrás, J., Genta, S., Vijenthira, A. & Siu, L. L. Antibody-drug conjugates: in search of partners of choice. *Trends Cancer* **9**, 339–354 (2023).
573. Milenic, D. E., Brady, E. D. & Brechbiel, M. W. Antibody-targeted radiation cancer therapy. *Nat Rev Drug Discov* **3**, 488–498 (2004).
574. Lin, M., Paolillo, V., Le, D. B., Macapinlac, H. & Ravizzini, G. C. Monoclonal antibody based radiopharmaceuticals for imaging and therapy. *Curr Probl Cancer* **45**, (2021).
575. Verhaar, E. R. *et al.* MICA-specific nanobodies for diagnosis and immunotherapy of MICA+ tumors. *Front Immunol* **15**, (2024).
576. Verhaar, E. R., Woodham, A. W. & Ploegh, H. L. Nanobodies in cancer. *Semin Immunol* **52**, (2021).
577. Pleiner, T., Bates, M. & Görlich, D. A toolbox of anti-mouse and anti-rabbit IgG secondary nanobodies. *Journal of Cell Biology* **217**, 1143–1154 (2018).
578. Zhong, P. *et al.* CRGD-installed docetaxel-loaded mertansine prodrug micelles: Redox-triggered ratiometric dual drug release and targeted synergistic treatment of Bi6F10 melanoma. *Nanotechnology* **28**, (2017).
579. Sievers, N. M., Dörrie, J. & Schaft, N. CARs: Beyond t cells and t cell-derived signaling domains. *Int J Mol Sci* **21**, (2020).
580. Guedan, S., Calderon, H., Posey, A. D. & Maus, M. V. Engineering and Design of Chimeric Antigen Receptors. *Mol Ther Methods Clin Dev* **12**, 145–156 (2019).
581. Jayaraman, J. *et al.* CAR-T design: Elements and their synergistic function. *EBioMedicine* **58**, (2020).
582. Ajina, A. & Maher, J. Strategies to address chimeric antigen receptor tonic signaling. *Mol Cancer Ther* **17**, 1795–1815 (2018).
583. Gil, D. & Schrum, A. G. Strategies to stabilize compact folding and minimize aggregation of antibody-based fragments. *Advances in Bioscience and Biotechnology* **04**, 73–84 (2013).
584. Wörn, A. & Plückthun, A. Stability engineering of antibody single-chain Fv fragments. *J Mol Biol* **305**, 989–1010 (2001).
585. Martin, T. *et al.* Ciltacabtagene Autoleucel, an Anti-B-cell Maturation Antigen Chimeric Antigen Receptor T-Cell Therapy, for Relapsed/Refractory Multiple Myeloma: CARTITUDE-1 2-Year Follow-Up. *Journal of Clinical Oncology* **41**, 1265–1274 (2022).
586. Raje, N. *et al.* Anti-BCMA CAR T-Cell Therapy bb2121 in Relapsed or Refractory Multiple Myeloma. *New England Journal of Medicine* **380**, 1726–1737 (2019).
587. Chen, Y. J., Abila, B. & Mostafa Kamel, Y. CAR-T: What Is Next? *Cancers (Basel)* **15**, (2023).
588. Oelsner, S. *et al.* Continuously expanding CAR NK-92 cells display selective cytotoxicity against B-cell leukemia and lymphoma. *Cytotherapy* **19**, 235–249 (2017).
589. You, F. *et al.* A novel CD7 chimeric antigen receptor-modified NK-92MI cell line targeting T-cell acute lymphoblastic leukemia. *Am J Cancer Res* **9**, 64–78 (2019).
590. Imai, C., Iwamoto, S. & Campana, D. Genetic modification of primary natural killer cells overcomes inhibitory signals and induces specific killing of leukemic cells. *Blood* **106**, 376–383 (2005).

591. Muller, T. *et al.* Expression of a CD20-specific chimeric antigen receptor enhances cytotoxic activity of NK cells and overcomes NK-resistance of lymphoma and leukemia cells. *Cancer Immunology, Immunotherapy* **57**, 411–423 (2007).
592. Boissel, L. *et al.* Retargeting NK-92 cells by means of CD19- and CD20-specific chimeric antigen receptors compares favorably with antibody-dependent cellular cytotoxicity. *Oncoimmunology* **2**, e26527 (2013).
593. Boissel, L., Betancur, M., Wels, W. S., Tuncer, H. & Klingemann, H. Transfection with mRNA for CD19 specific chimeric antigen receptor restores NK cell mediated killing of CLL cells. *Leuk Res* **33**, 1255–1259 (2009).
594. Li, L. *et al.* Expression of chimeric antigen receptors in natural killer cells with a regulatory-compliant non-viral method. *Cancer Gene Ther* **17**, 147–154 (2010).
595. Shimasaki, N. *et al.* A clinically adaptable method to enhance the cytotoxicity of natural killer cells against B-cell malignancies. *Cytotherapy* **14**, 830–840 (2012).
596. Battula, V. L. *et al.* Ganglioside GD2 identifies breast cancer stem cells and promotes tumorigenesis. *Journal of Clinical Investigation* **122**, 2066–2078 (2012).
597. Altvater, B. *et al.* 2B4 (CD244) signaling by recombinant antigen-specific chimeric receptors costimulates natural killer cell activation to leukemia and neuroblastoma cells. *Clinical Cancer Research* **15**, 4857–4866 (2009).
598. Esser, R. *et al.* NK cells engineered to express a GD 2-specific antigen receptor display built-in ADCC-like activity against tumour cells of neuroectodermal origin. *J Cell Mol Med* **16**, 569–581 (2012).
599. Zhang, C. *et al.* ErbB2/HER2-Specific NK Cells for Targeted Therapy of Glioblastoma. *J Natl Cancer Inst* **108**, (2016).
600. Morgan, R. A. *et al.* Case report of a serious adverse event following the administration of t cells transduced with a chimeric antigen receptor recognizing ERBB2. *Molecular Therapy* **18**, 843–851 (2010).
601. Wouters, Y. *et al.* VHVs as tools for therapeutic protein delivery to the central nervous system. *Fluids Barriers CNS* **19**, (2022).
602. Chekol Abebe, E., Yibeltal Shiferaw, M., Tadele Admasu, F. & Asmamaw Dejenie, T. Ciltacabtagene autoleucel: The second anti-BCMA CAR T-cell therapeutic armamentarium of relapsed or refractory multiple myeloma. *Front Immunol* **13**, (2022).
603. Zhang, X. *et al.* Cytokine Release Syndrome After Modified CAR-NK Therapy in an Advanced Non-small Cell Lung Cancer Patient: A Case Report. *Cell Transplant* **31**, (2022).
604. Groh, V. *et al.* Cell stress-regulated human major histocompatibility complex class I gene expressed in gastrointestinal epithelium. *Immunology* **93**, 12445–12450 (1996).
605. Ghadially, H. *et al.* MHC class i chain-related protein A and B (MICA and MICB) are predominantly expressed intracellularly in tumour and normal tissue. *Br J Cancer* **116**, 1208–1217 (2017).
606. Kim, Y., Born, C., Bléry, M. & Steinle, A. MICAgene Mice Recapitulate the Highly Restricted but Activation-Inducible Expression of the Paradigmatic Human NKG2D Ligand MICA. *Front Immunol* **11**, (2020).
607. Tam, Y. K., Martinson, J. A., Doligosa, K. & Klingemann, H.-G. Ex vivo expansion of the highly cytotoxic human natural killer-92 cell-line under current good manufacturing practice conditions for clinical adoptive cellular immunotherapy. *Cytotherapy* **5**, 259–272 (2003).
608. Yeap, W. H. *et al.* CD16 is indispensable for antibody-dependent cellular cytotoxicity by human monocytes. *Sci Rep* **6**, (2016).
609. Sondel, P. M. & Alderson, K. L. Clinical cancer therapy by NK cells via antibody-dependent cell-mediated cytotoxicity. *J Biomed Biotechnol* **2011**, (2011).

610. Zhu, H. *et al.* Pluripotent stem cell-derived NK cells with high-affinity noncleavable CD16a mediate improved antitumor activity. *Blood* **135**, 399–410 (2020).
611. Christodoulou, I. *et al.* Engineering CAR-NK cells to secrete IL-15 sustains their anti-AML functionality but is associated with systemic toxicities. *J Immunother Cancer* **9**, (2021).
612. Xu, D. L. *et al.* A Novel Sushi-IL15-PD1 CAR-NK92 Cell Line With Enhanced and PD-L1 Targeted Cytotoxicity Against Pancreatic Cancer Cells. *Front Oncol* **12**, (2022).
613. López-Cantillo, G., Urueña, C., Camacho, B. A. & Ramírez-Segura, C. CAR-T Cell Performance: How to Improve Their Persistence? *Front Immunol* **13**, (2022).
614. Zhang, J. *et al.* Generation of anti-GD2 CAR macrophages from human pluripotent stem cells for cancer immunotherapies. *Stem Cell Reports* **18**, 585–596 (2023).
615. Su, S. *et al.* Induced CAR-Macrophages as a Novel Therapeutic Cell Type for Cancer Immune Cell Therapies. *Cells* **11**, (2022).
616. Sloas, C., Gill, S. & Klichinsky, M. Engineered CAR-Macrophages as Adoptive Immunotherapies for Solid Tumors. *Front Immunol* **12**, (2021).
617. Klichinsky, M. *et al.* Human chimeric antigen receptor macrophages for cancer immunotherapy. *Nat Biotechnol* **38**, 947–953 (2020).
618. Wang, S. *et al.* CAR-macrophage: An extensive immune enhancer to fight cancer. *EBioMedicine* **76**, 103873 (2022).
619. Pierini, S. *et al.* Chimeric antigen receptor macrophages (CAR-M) sensitize solid tumors to anti-PD1 immunotherapy. in *Journal for ImmunoTherapy of Cancer* A390–A390 (BMJ, 2022). doi:10.1136/jitc-2022-sitc2022.0371.
620. Verhaar, E. R., van Keizerswaard, W. J. C., Knoflook, A., Balligand, T. & Ploegh, H. L. Nanobody-based CAR NK cells for possible immunotherapy of MICA+ tumors. *PNAS nexus* **3**, pgaer184 (2024).
621. Naviaux, R. K., Costanzi, E., Haas, M. & Verma, I. M. The pCL Vector System: Rapid Production of Helper-Free, High-Titer, Recombinant Retroviruses. *J Virol* **70**, 5701–5705 (1996).
622. Xie, G. *et al.* CAR-NK cells: A promising cellular immunotherapy for cancer. *EBioMedicine* **59**, (2020).
623. Hensel, J. A., Khattar, V., Ashton, R. & Ponnazhagan, S. Characterization of immune cell subtypes in three commonly used mouse strains reveals gender and strain-specific variations. *Laboratory Investigation* **99**, 93–106 (2019).
624. Kim, Y. *et al.* Optimized conditions for gene transduction into primary immune cells using viral vectors. *Sci Rep* **13**, (2023).
625. Dufait, I. *et al.* Retroviral and Lentiviral Vectors for the Induction of Immunological Tolerance. *Scientifica (Cairo)* **2012**, 1–14 (2012).
626. Pishesha, N., Harmand, T. J. & Ploegh, H. L. A guide to antigen processing and presentation. *Nat Rev Immunol* **22**, 751–764 (2022).
627. Ravindranath, M. H., Pham, T., El-Awar, N., Kaneko, H. & Terasaki, P. I. Anti-HLA-E mAb 3D12 mimics MEM-E/o2 in binding to HLA-B and HLA-C alleles: Web-tools validate the immunogenic epitopes of HLA-E recognized by the antibodies. *Mol Immunol* **48**, 423–430 (2011).
628. McMaster, M. *et al.* HLA-G Isoforms Produced by Placental Cytotrophoblasts and Found in Amniotic Fluid Are Due to Unusual Glycosylation. *The Journal of Immunology* **160**, 5922–5928 (1998).
629. Stam, N. J., Spits, H. & Ploegh, H. L. Monoclonal antibodies raised against denatured HLA-B locus heavy chains permit biochemical characterization of certain HLA-C locus products. *The Journal of Immunology* **137**, 2299–2306 (1986).

630. Seitz, C., Uchanska-Ziegler, B., Zank, A. & Ziegler, A. The monoclonal antibody HCA2 recognises a broadly shared epitope on selected classical as well as several non-classical HLA class I molecules. *Mol Immunol* **35**, 819–827 (1998).
631. Barnstable, C. J. *et al.* Production of monoclonal antibodies to group A erythrocytes, HLA and other human cell surface antigens-new tools for genetic analysis. *Cell* **14**, 9–20 (1978).
632. Stam, N. J., Vroom, Th. M., Peters, P. J., Pastoors, E. B. & Ploegh, H. L. HLA-A- and HLA-B-specific monoclonal antibodies reactive with free heavy chains in Western blots, in formalin-fixed, paraffin-embedded tissue sections and in cryo-immuno-electron microscopy. *Int Immunol* **2**, 113–125 (1990).
633. Grimsley, C. *et al.* Definitive high resolution typing of HLA-E allelic polymorphisms: Identifying potential errors in existing allele data. *Tissue Antigens* **60**, 306–312 (2002).
634. Geraghty, D. E., Stockscheleider, M., Ishitani, A. & Hansen, J. A. Polymorphism at the HLA-E locus predates most HLA-A and -B polymorphism. *Hum Immunol* **33**, 174–184 (1992).
635. Guimaraes, C. P. *et al.* Site-specific C-terminal and internal loop labeling of proteins using sortase-mediated reactions. *Nat Protoc* **8**, 1787–1799 (2013).
636. French, D., Fischberg, E., Buhl, S. & Scharff, M. D. The production of more useful monoclonal antibodies I. Modifications of the basic technology. *Immunol Today* **7**, 344–346 (1986).
637. Zhao, Y., Long, M. J. C., Wang, Y., Zhang, S. & Aye, Y. Ube2V2 Is a Rosetta Stone Bridging Redox and Ubiquitin Codes, Coordinating DNA Damage Responses. *ACS Cent Sci* **4**, 246–259 (2018).
638. Tsang, M., Gantchev, J., Ghazawi, F. M. & Litvinov, I. V. Protocol for adhesion and immunostaining of lymphocytes and other non-adherent cells in culture. *Biotechniques* **63**, 230–233 (2017).
639. Salomé, B. *et al.* NKG2A and HLA-E define an alternative immune checkpoint axis in bladder cancer. *Cancer Cell* **40**, 1027–1043.e9 (2022).
640. Kwon, S. *et al.* Targeted Delivery of Cyclotides via Conjugation to a Nanobody. *ACS Chem Biol* **13**, 2973–2980 (2018).
641. Ovchinnikov, V., Louveau, J. E., Barton, J. P., Karplus, M. & Chakraborty, A. K. Role of framework mutations and antibody flexibility in the evolution of broadly neutralizing antibodies. *eLife* (2018) doi:10.7554/eLife.33038.001.
642. Verma, S. *et al.* Trastuzumab Emtansine for HER2-Positive Advanced Breast Cancer. *New England Journal of Medicine* **367**, 1783–1791 (2012).
643. Matulonis, U. A. *et al.* Efficacy and Safety of Mirvetuximab Soravtansine in Patients With Platinum-Resistant Ovarian Cancer With High Folate Receptor Alpha Expression: Results From the SORAYA Study. *Journal of Clinical Oncology* **41**, 2436–2445 (2023).
644. Bouchard, H., Viskov, C. & Garcia-Echeverria, C. Antibody-drug conjugates - A new wave of cancer drugs. *Bioorg Med Chem Lett* **24**, 5357–5363 (2014).
645. Khantasup, K., Chantima, W., Sangma, C., Poomputsa, K. & Dharakul, T. Design and generation of humanized single-chain Fv derived from mouse hybridoma for potential targeting application. *Monoclon Antib Immunodiagn Immunother* **34**, 404–417 (2015).
646. Bell, M. & Gottschalk, S. Engineered Cytokine Signaling to Improve CAR T Cell Effector Function. *Front Immunol* **12**, (2021).
647. Pei, K. *et al.* A Comparison Study of Anti-CLL1 CART Cells Equipped with Different Co-Stimulatory Domains in the Treatment of Children with Refractory/Relapsed Acute Myeloid Leukemia. *Blood* **138**, 824–824 (2021).
648. Kagoya, Y. *et al.* A novel chimeric antigen receptor containing a JAK–STAT signaling domain mediates superior antitumor effects. *Nat Med* **24**, 352–359 (2018).

649. Boucher, J. C. *et al.* CD28 costimulatory domain-targeted mutations enhance chimeric antigen receptor T-cell function. *Cancer Immunol Res* **9**, 62–74 (2021).
650. Feucht, J. *et al.* Calibration of CAR activation potential directs alternative T cell fates and therapeutic potency. *Nat Med* **25**, 82–88 (2019).
651. Llames, S., García-Pérez, E., Meana, Á., Larcher, F. & Del Río, M. Feeder Layer Cell Actions and Applications. *Tissue Eng Part B Rev* **21**, 345–353 (2015).
652. Borst, L., van der Burg, S. H. & van Hall, T. The NKG2A-HLA-E axis as a novel checkpoint in the tumor microenvironment. *Clinical Cancer Research* **26**, 5549–5556 (2021).
653. Ravindranath, M. H., Pham, T., El-Awar, N., Kaneku, H. & Terasaki, P. I. Anti-HLA-E mAb 3D12 mimics MEM-E/o2 in binding to HLA-B and HLA-C alleles: Web-tools validate the immunogenic epitopes of HLA-E recognized by the antibodies. *Mol Immunol* **48**, 423–430 (2011).