

CT radiomics compared to a clinical model for predicting checkpoint inhibitor treatment outcomes in patients with advanced melanoma Maat, L.S. ter; Duin, I.A.J. van; Elias, S.G.; Leiner, T.; Verhoeff, J.J.C.; Arntz, E.R.A.N.; ...; Veta, M.

Citation

Maat, L. S. ter, Duin, I. A. J. van, Elias, S. G., Leiner, T., Verhoeff, J. J. C., Arntz, E. R. A. N., ... Veta, M. (2023). CT radiomics compared to a clinical model for predicting checkpoint inhibitor treatment outcomes in patients with advanced melanoma. *European Journal Of Cancer*, 185, 167-177. doi:10.1016/j.ejca.2023.02.017

Version: Publisher's Version

License: <u>Creative Commons CC BY 4.0 license</u>
Downloaded from: <u>https://hdl.handle.net/1887/3731638</u>

Note: To cite this publication please use the final published version (if applicable).



Available online at www.sciencedirect.com

ScienceDirect





Original Research

CT radiomics compared to a clinical model for predicting checkpoint inhibitor treatment outcomes in patients with advanced melanoma



Laurens S. ter Maat ^{a,*}, Isabella A.J. van Duin ^b, Sjoerd G. Elias ^c, Tim Leiner ^d, Joost J.C. Verhoeff ^e, Eran R.A.N. Arntz ^f, Max F. Troenokarso ^f, Willeke A.M. Blokx ^g, Ivana Isgum ^{h,i}, Geraldine A. de Wit ^j, Franchette W.P.J. van den Berkmortel ^k, Marye J. Boers-Sonderen ^l, Martijn F. Boomsma ^m, Fons J.M. van den Eertwegh ⁿ, Jan Willem B. de Groot ^o, Djura Piersma ^p, Art Vreugdenhil ^q, Hans M. Westgeest ^r, Ellen Kapiteijn ^s, Paul J. van Diest ^g, Josien P.W. Pluim ^{a,t}, Pim A. de Jong ^u, Karijn P.M. Suijkerbuijk ^{b,1}, Mitko Veta ^{t,1}

^a Image Sciences Institute, University Medical Center Utrecht, Utrecht University, Utrecht, the Netherlands

^b Department of Medical Oncology, University Medical Center Utrecht, Utrecht University, Utrecht, the Netherlands

^c Department of Epidemiology, Julius Center for Health Sciences and Primary Care, University Medical Center Utrecht, Utrecht University, Utrecht, the Netherlands

^d Department of Radiology, Mayo Clinical, Rochester, MN, USA

^e Department of Radiotherapy, University Medical Center Utrecht, Utrecht University, Utrecht, the Netherlands

f Utrecht University, Utrecht, the Netherlands

g Department of Pathology, University Medical Center Utrecht, Utrecht University, Utrecht, the Netherlands

^h Departments of Biomedical Engineering and Physics & Radiology and Nuclear Medicine, University Medical Center Amsterdam, University of Amsterdam, Amsterdam, the Netherlands

¹ Quantitative Healthcare Analysis group, Informatics Institute, Faculty of Science, University of Amsterdam, Amsterdam, the Netherlands

^j Department of Public Health, Healthcare Innovation & Evaluation and Medical Humanities, Julius Center Research Program Methodology, University Medical Center Utrecht, Utrecht University, Utrecht, the Netherlands

^k Department of Medical Oncology, Zuyderland Medical Center, Sittard-Geleen, the Netherlands

¹ Department of Medical Oncology, Radboudumc, Radboud University, Nijmegen, the Netherlands

^m Department of Radiology, Isala Zwolle, Zwolle, the Netherlands

ⁿ Department of medical oncology, Amsterdam UMC, Vrije Universiteit Amsterdam, Cancer Center Amsterdam, Boelelaan 1117, Amsterdam, the Netherlands

[°] Isala Oncology Center, Isala Zwolle, Zwolle, the Netherlands

^p Department of Medical Oncology, Medisch Spectrum Twente, Enschede, the Netherlands

^q Department of Medical Oncology, Máxima Medical Center, Veldhoven, the Netherlands

 $^{^*\} Corresponding\ author.$

E-mail address: l.s.termaat@umcutrecht.nl (L.S. ter Maat).

¹ These authors contributed equally

Received 22 December 2022; Received in revised form 10 February 2023; Accepted 17 February 2023 Available online 24 February 2023

KEYWORDS

Metastatic melanoma; Checkpoint inhibition; Computed tomography; Radiomics; Machine learning; Response prediction **Abstract** *Introduction:* Predicting checkpoint inhibitors treatment outcomes in melanoma is a relevant task, due to the unpredictable and potentially fatal toxicity and high costs for society. However, accurate biomarkers for treatment outcomes are lacking. Radiomics are a technique to quantitatively capture tumour characteristics on readily available computed tomography (CT) imaging. The purpose of this study was to investigate the added value of radiomics for predicting clinical benefit from checkpoint inhibitors in melanoma in a large, multicenter cohort.

Methods: Patients who received first-line anti-PD1 ± anti-CTLA4 treatment for advanced cutaneous melanoma were retrospectively identified from nine participating hospitals. For every patient, up to five representative lesions were segmented on baseline CT, and radiomics features were extracted. A machine learning pipeline was trained on the radiomics features to predict clinical benefit, defined as stable disease for more than 6 months or response per RECIST 1.1 criteria. This approach was evaluated using a leave-one-centre-out cross validation and compared to a model based on previously discovered clinical predictors. Lastly, a combination model was built on the radiomics and clinical model.

Results: A total of 620 patients were included, of which 59.2% experienced clinical benefit. The radiomics model achieved an area under the receiver operator characteristic curve (AUROC) of 0.607 [95% CI, 0.562–0.652], lower than that of the clinical model (AUROC=0.646 [95% CI, 0.600–0.692]). The combination model yielded no improvement over the clinical model in terms of discrimination (AUROC=0.636 [95% CI, 0.592–0.680]) or calibration. The output of the radiomics model was significantly correlated with three out of five input variables of the clinical model (p < 0.001).

Discussion: The radiomics model achieved a moderate predictive value of clinical benefit, which was statistically significant. However, a radiomics approach was unable to add value to a simpler clinical model, most likely due to the overlap in predictive information learned by both models. Future research should focus on the application of deep learning, spectral CT-derived radiomics, and a multimodal approach for accurately predicting benefit to checkpoint inhibitor treatment in advanced melanoma.

© 2023 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).

1. Introduction

Survival of patients with advanced melanoma has improved dramatically after the introduction of immunotherapy. The survival of patients with unresectable stage III and stage IV melanoma has historically been very poor with a 1-year overall survival of 25% in phase II trials up to 2007 [1]. This changed with the introduction of anti-cytotoxic t-lymphocyte antigen-4 (CTLA4) therapy in 2011 [2] and anti-programmed death-1 (PD1) therapy in 2014 [3,4]. In patients treated with anti-PD1 antibodies, real-world 1-year overall survival is now 67%, with 40% of patients achieving remissions of several years [5]. For patients treated with anti-PD1 plus anti-CTLA4 therapy, 5-year overall survival is reported to be as high as 52% [6].

However, not all patients benefit from checkpoint inhibitors. At 6 months after start of anti-PD1 treatment, 43% of patients experience progression or death. Furthermore, overall survival of patients with progression at 6 months was shown to be only 16% at 30 months. This is in contrast to a 30-month overall survival of 60%, 79%, and 96% for patients with stable disease, partial response, and complete response at 6 months of follow-up, respectively, in real-world data [5]. Similar results were reported in patients treated with anti-PD1 plus anti-CTLA4 therapy [7].

Accurate prediction of treatment benefit is an important topic for several reasons. First, treatment with checkpoint inhibitors is associated with severe and potentially fatal or irreversible toxicity. Severe toxicity occurs in 10–15% of patients treated with anti-PD1

^r Department of Medical Oncology, Amphia Hospital, Breda, the Netherlands

s Department of Medical Oncology, Leiden University Medical Center, Leiden University, Leiden, the Netherlands

^t Medical Image Analysis, Department of Biomedical Engineering, Eindhoven University of Technology, Eindhoven, the Netherlands

^u Department of Radiology, University Medical Center Utrecht, Utrecht University, Utrecht, the Netherlands

monotherapy [5,8–10], and in as much as 60% of patients treated with anti-PD1 plus anti-CTLA4 combination therapy [11]. Second, checkpoint inhibition therapy is very costly. Depending on country and setting, estimates of additional costs per gained quality-adjusted life year range from 25,000 to 81,000 United States Dollars [12,13]. Lastly, if patients who will not benefit are identified before start of treatment, alternative or experimental therapies can be started without delay.

Previously identified predictors for treatment outcomes are not yet sufficient to guide clinical decisions. Known clinical predictors of poor outcome include high tumour load, presence of liver metastases and symptomatic brain metastases, increased lactate dehydrogenase (LDH), and worse Eastern Cooperative Oncology Group (ECOG) performance status [14]. In addition, other biomarkers have been explored, such as PD-L1 expression, tumour mutational burden, and histopathology features. Thus far, however, these predictors are not strong enough to predict treatment outcomes with high certainty [15], or the results remain to be validated in future studies [16].

Radiomics are by now an established modality for diagnosis, prognosis, and prediction. Radiomics capture information about shape, intensity, and texture of lesions in imaging and thereby form a reflection of tumour characteristics, such as necrosis or vascularisation. These extracted features can subsequently be correlated to a clinical outcome [17]. This makes radiomics a cheap and non-invasive modality to, for example, discern benign from malignant lung nodules [18], estimate prognosis in non-small cell lung cancer (NSCLC) patients [19], and assess mutation status in glioblastoma [20]. Regarding prediction of checkpoint inhibitor treatment outcomes, promising findings have been published, particularly in NSCLC patients [21].

The added value of computed tomography (CT) radiomics for predicting clinical benefit of checkpoint inhibitors in melanoma remains to be determined in large multicenter studies. Three previous smaller studies have investigated radiomics for this purpose, with conflicting findings. The studies by Trebeschi et al. [22] and Peisen et al. [23] report a significant discriminative value of radiomics for treatment outcomes (area under the receiver operator characteristic curve [AUROC]=0.78 on a dataset of 80 patients, and AUROC=0.64 on a dataset of 262 patients, respectively). In contrast, Brendlin et al. [24] reported a non-discriminative performance, despite using a similar methodology (AUROC=0.50 in 140 patients). These differences in results highlight the importance of a large dataset to determine the value of radiomics. Furthermore, only the study by Peisen et al. investigated the added value over a simpler clinical model, with varying results across different outcomes. Lastly, none of the previous studies evaluated their model on data from other centres, although variability in scanner protocol may add significant noise [25]. In this study, we aimed to address these limitations and determine the added value of radiomics for predicting checkpoint inhibitor outcomes in a multicenter study in advanced melanoma.

2. Materials and methods

2.1. Patient selection

Eligible patients were retrospectively identified from high-quality registry data [26] from nine participating centres in The Netherlands (Amphia Ziekenhuis, Isala Zwolle, Leids Universitair Medisch Centrum, Máxima MC, Medisch Spectrum Twente, Radboudumc, UMC Utrecht, Amsterdam UMC, Zuyderland MC). Patients over the age of 18 were included if they received first-line treatment with anti-PD1 ± anti-CTLA4 checkpoint inhibition for irresectable stage IIIC or stage IV cutaneous melanoma after 01–01–2016. Exclusion criteria were (i) unavailability of baseline contrast-enhanced CT imaging (CE-CT), (ii) lack of eligible target lesions, and (iii) less than 6 months of follow-up. Clinical characteristics were collected for the included patients and compared to those of the excluded patients. CT acquisition characteristics were extracted for included patients.

2.2. Lesion selection and segmentation

For every patient, one to five lesions were selected on baseline CT imaging and segmented. We aimed to make this selection of lesions as informative and representative as possible by using the following protocol: first, the five largest lesions were selected with a maximum of two per organ. If more lesions remained after segmenting a maximum of two per organ, the largest remaining lesions were segmented up to a total of five. Lesion selections were made without knowledge of the outcome. Lesions were excluded if they were not welldemarcated, affected by imaging artifacts or if the maximum diameter was less than 5 mm. Segmentations were performed in 3D Slicer [27] on the series with the lowest slice thickness by authors LSM and IAJD, under supervision of board-certified radiologists with 17 and 18 years of experience (PJ and TL, respectively).

2.3. Feature extraction

Features were extracted from the segmented volumes using PyRadiomics [28]. For every volume, 1874 features were extracted at five different levels of detail, resulting in a total of 9370 features. An overview of the extracted features is given in the Supplementary Methods. Interobserver agreement of segmentations and features was calculated using Dice scores and

intraclass correlation coefficient (ICC), respectively, based on 16 scans segmented by both observers (LSM, IAJD).

2.4. Outcome definition

The primary outcome was clinical benefit, defined as the best overall response of partial or complete response, or stable disease for a minimum of 6 months after start of treatment; response was determined by the treating physician in line with Response Evaluation Criteria in Solid Tumours (RECIST) 1.1 criteria [29]. The secondary outcome was objective response, defined as the best overall response of partial or complete response. Clinical benefit was used as the primary outcome, as the intended use of the model was to identify patients who would quickly progress despite treatment and therefore not derive any benefit from treatment. Individual lesion response was assessed using maximum diameter recordings at baseline and at 3, 6 and 9 months, or until treatment was changed. Given the possibility of pseudoprogression, the last available follow-up was used to determine lesion outcomes. If the maximum diameter at the last follow-up was less than 120% of the baseline diameter, the lesion was labelled as 'does benefit', and 'does not benefit' otherwise. In parallel, the lesion was labelled as 'responsive' if the maximum diameter was less than 70% of the baseline diameter at the last followup, and 'non-responsive' otherwise. These lesion-level cut-offs were chosen in correspondence with the patient-level cut-offs used in RECIST 1.1 [29].

2.5. Evaluated models

Three predictive models were compared: a model based on radiomics, a model based on baseline clinical characteristics and an ensemble model that combined the predictions of these models. The radiomics model consisted of a machine learning pipeline that automatically selected optimal components and hyperparameters for feature selection, dimensionality reduction, and classification (Fig. 1). This pipeline was trained to predict outcomes per lesion; these outputs per lesion were then aggregated to a patient-level prediction. The clinical model used the same machine learning pipeline, which was fitted on five clinical variables that were consistently shown to be predictive of checkpoint inhibitor treatment outcomes in previous literature [5,14,30,31]. These predictors were (i) ECOG performances status, (ii) LDH level, presence of (iii) brain and (iv) liver metastases, and (v) number of affected organs. All variables were onehot encoded; missing values were encoded as a separate label. The ensemble model consisted of a logistic regression fitted on the output of the radiomics and clinical model. All three models were evaluated using a nested cross validation. The inner loop was used for optimal model selection and hyperparameter tuning; the outer loop was used to evaluate predictive performance

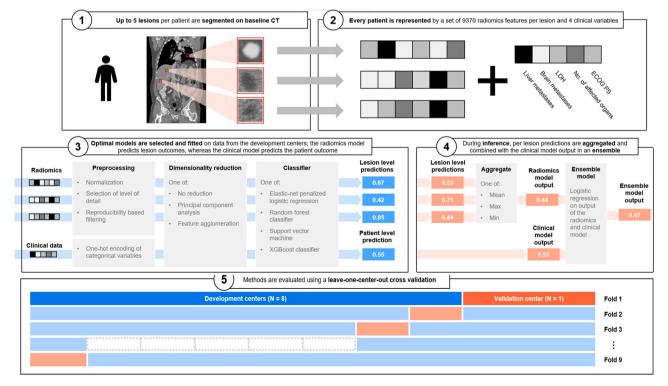


Fig. 1. Overview of methodology.

on unseen data and was conducted in a leave-one-centre-out manner. Further details are supplied in the Supplementary Methods.

2.6. Statistical analysis

The discriminative performance of the models was evaluated using the AUROC and corresponding 95% confidence interval. The cross-validated AUROC and confidence interval were calculated using the cvAUC R package [32]. Methods for comparing cross-validated AUROCs between models are detailed in the Supplementary Methods. Subgroup analyses were conducted for patients treated with anti-PD1 therapy and anti-PD1 plus anti-CTLA4 therapy by evaluating the fitted model only on patients from the respective groups. The output of the radiomics model for predicting clinical benefit was correlated to the input variables of the clinical model to determine if the radiomics model learned features that were already represented in the baseline clinical model.

2.7. Adherence to quality standards

The TRIPOD checklist [33] was completed and is available in Supplementary Table 1. The study design was reviewed by the Medical Ethics Committee and not considered subject to the Medical Research Involving

Human Subjects Act in compliance with Dutch regulations; informed consent was waived.

3. Results

3.1. Patient characteristics

Out of 1191 eligible patients, 620 patients with a total of 2352 lesions were included. A flowchart of the selection process is shown in Fig. 2. The rate of clinical benefit was 59.2% (367 patients); the objective response rate was 51.3% (318 patients): Lesion level outcomes were available for 75.2% of lesions. Lesion-level outcomes could not be recorded for patients from the Radboudumc (327 lesions, 13.9%) due to local regulations. In addition, follow-up imaging was unavailable due to patient death or clinical progression before the first follow-up moment for 185 lesions (7.9%), due to the lesion falling outside the field of view in 27 lesions (1.1%) and due to technical issues in 44 lesions (1.9%). Rate of benefit was 79.4% among lesions with available labels, whereas response rate was 54.8% (Supplementary Table 5). Of all eligible patients, 490 patients were excluded because of the unavailability of contrast-enhanced pretreatment CT. In most of these cases, an 18-fluorodeoxyglucose positron emission tomography (FDG-PET) with low-dose CT was made. Characteristics for the included patients are shown in Table 1 and compared to those of excluded patients in

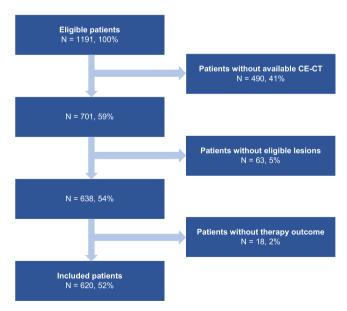


Fig. 2. Flowchart of patient selection.

Table 1 Characteristics of included patients.

		Missing	Overall
n			620
Age, median [Q1, Q3]		0	67.5 [58.0,75.0]
Sex, n (%)	Female	0	239 (38.5)
	Male		381 (61.5)
Stage, n (%)	IIIC	4	25 (4.1)
	M1a		49 (8.0)
	M1b		94 (15.3)
	M1c		296 (48.1)
	M1d		152 (24.7)
Eastern Cooperative Oncology Group performance status,	0	26	287 (48.3)
n (%)	1		247 (41.6)
	2–4		60 (10.1)
Primary tumour location, n (%)	Acral	10	15 (2.5)
	Extremity		167 (27.4)
	Head, neck		66 (10.8)
	Trunk		247 (40.5)
	Unknown		115 (18.9)
Brain metastases, n (%)	Absent	45	423 (73.6)
	Asymptomatic		76 (13.2)
	Symptomatic		76 (13.2)
Liver metastases, n (%)	Absent	30	398 (67.5)
	Present		192 (32.5)
No. of affected organs, n (%)	<3	0	338 (54.5)
	>2		282 (45.5)
Lactate dehydrogenase, n (%)	Normal	9	381 (62.4)
	1–2x upper limit of		177 (29.0)
	normal		` '
	>2x upper limit of		53 (8.7)
	normal		` ′
Clinical benefit, n (%)	No benefit	0	253 (40.8)
	Benefit		367 (59.2)
Objective response, n (%)	No response	0	302 (48.7)
	Response		318 (51.3)
Therapy, n (%)	Anti-PD1	0	370 (59.7)
	Ipilimumab &		250 (40.3)
	Nivolumab		` ′

Supplementary Table 2. The subgroups of patient treated with anti-PD1 and combination therapy consisted of 370 and 250 patients, respectively. Supplementary Tables 3 and 4 show patient characteristics per centre, and for the subgroups treated with monotherapy and combination therapy, respectively. CT acquisition characteristics per centre are displayed in Supplementary Table 6.

3.2. Interobserver variability

52 lesions in 16 scans were segmented by two observers. Segmentations corresponded with a median Dice score of 0.88 (IQI 0.82–0.92). For the extracted features, the median ICC was 0.97 (IQI 0.92–0.99).

3.3. Treatment outcome prediction

For predicting clinical benefit, the radiomics model achieved an AUROC of 0.607 [95% CI, 0.562–0.652],

the clinical model an AUROC of 0.646 [95% CI, 0.600-0.692], and the ensemble model an AUROC of 0.636 [95% CI, 0.592-0.680] (Fig. 3). The difference in AUROC between the ensemble and clinical model was not statistically significant (Supplementary Figure 1). Calibration curves showed adequate calibration of the three models with no evidence of poor fit (Hosmer-Lemeshow p > 0.07). The range of predicted probabilities was comparable between models (IOI 0.56-0.65, 0.53-0.67, and 0.52-0.69 for the radiomics, clinical and ensemble model, respectively). Results were similar for predicting objective response (Supplementary Figure 2-3). Predictive performance for both outcomes was comparable in subgroups of patients treated with monotherapy and combination therapy, with a trend of better discrimination in the subgroup of patients treated with combination therapy (Supplementary Figures 4–7). Details of the selected models and hyperparameters per fold are shown in Supplementary Table 7.

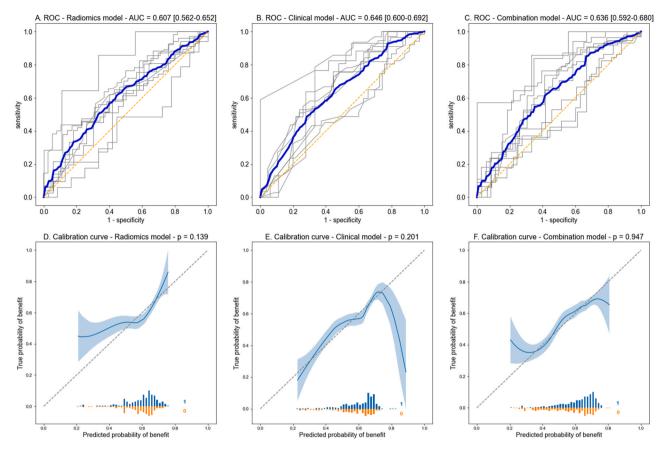


Fig. 3. Receiver operator characteristic (ROC) curves and calibration curves for predicting clinical benefit. (A–C) ROC curves for predicting durable clinical benefit in patients with melanoma treated with anti-PD1 ± anti-CTLA4 checkpoint inhibition for the radiomics model (A), clinical model (B) and combination model (C). Grey curves correspond to results per fold; blue curves are the weighted average of the results per fold. The area under the curve (AUC) with corresponding 95% confidence intervals is displayed. (D–F) Locally estimated scatterplot smoothing (LOESS) fitted calibration curves for predicting durable clinical benefit in the radiomics model (D), clinical model (E), and combination model (F); the shaded area corresponds to ± 1 standard deviation. Histograms of the predictions for positive (blue) and negative (orange) samples are provided below the curves, the x-axis displays the predicted values for these histograms. P-values of the Hosmer-Lemeshow goodness-of-fit test are shown in the plot titles.

3.4. Comparison of radiomics and clinical model

The predicted probability of clinical benefit by the radiomics model was significantly lower in patients in whom liver metastases were absent (Mann-Whitney U, p < 0.001, Fig. 4A), in patients with higher LDH (Kruskal-Wallis p < 0.001, Fig. 4D) and who had more affected organs (Mann-Whitney U p < 0.001, Fig. 4E). The output of radiomics model was not significantly different in patients with and without brain metastases), and for different categories of ECOG performance status (Fig. 4B–C). The output of radiomics and clinical models were significantly and positively correlated (Spearman's correlation coefficient = 0.369, p < 0.001, Fig. 4F).

4. Discussion

The present work shows that radiomics are moderately predictive of checkpoint inhibitor treatment outcomes

in patients with advanced melanoma. The results were consistent for both clinical benefit and objective response rate, and are most in line with the findings of the earlier study by Peisen et al. A recent work by Dercle et al. allows for comparison to a model that also incorporates radiomics from on-treatment CT scans [34]. This model reached an AUROC of 0.92 for predicting overall survival at 6 months, indicating that on-treatment radiomics are strongly predictive. However, most toxicity occurs in the first 3 months, and long-term outcomes can already be accurately predicted using ontreatment information without the use of radiomics [6,35]. Predicting response using the 3-month on-treatment scan therefore appears to be of limited clinical relevance.

Addition of radiomics to known clinical predictors, however, did not yield improvement in predictive value. The combination model was not superior to the clinical model in either discrimination or calibration. This lack of improvement can be explained due to an overlap in the

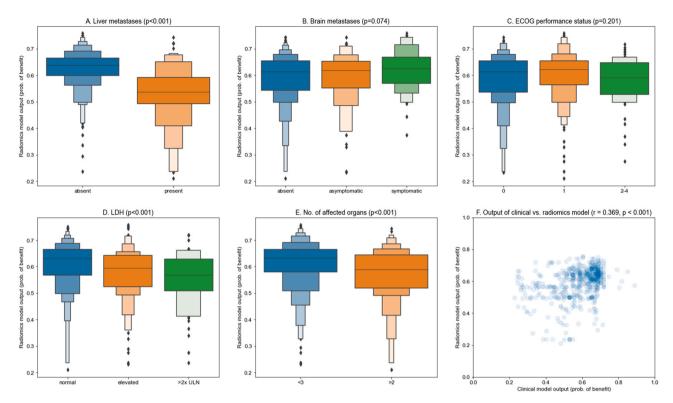


Fig. 4. Correspondence between the output of the clinical and radiomics models for predicting clinical benefit. Graphical overview of correspondence between the output of the radiomics and clinical models. (A–E) Boxenplots of the output of the radiomics model, compared across different values for clinical predictors. (A) The output of the radiomics model is significantly lower in patients with liver metastases than in patients without (Mann-Whitney U p < 0.001). (B) No statistical difference was found in the output of the radiomics model between patients without or with asymptomatic or symptomatic brain metastases (Kruskal-Wallis p = 0.074) and Eastern Cooperative Oncology Group performance status (Kruskal-Wallis p = 0.201). D) The output of the radiomics model is significantly lower in patients with higher levels of lactate dehydrogenase (Kruskal-Wallis p < 0.001) and with more affected organs (Mann-Whitney U p < 0.001). (F) The outputs of the clinical and radiomics models (predicted probability of response) are positively correlated (Spearman's rank correlation coefficient = 0.369, p < 0.001).

information learned by the radiomics model, and the information that is already represented in clinical variables. As demonstrated, the radiomics model indirectly learns to detect the presence of liver metastases and the amount of tumour burden, as reflected in LDH and number of affected organs. The fact that this information is indeed learned as expected is a strong argument for the validity of the present work. Furthermore, this indicates that such overlap is likely to be present in any radiomics model which is investigated for clinical purposes.

Studies on radiomics should assess the added value over simpler predictors. Many smaller exploratory studies have been conducted into the predictive value of radiomics for checkpoint inhibitor outcomes across different malignancies [21]. Their findings are almost exclusively positive, but the added value over clinical predictors was seldomly assessed. The present work demonstrates that clinical predictors can be captured by radiomics, and that the added value of radiomics should therefore always be investigated, even in exploratory studies.

Future works should aim to improve on radiomics through deep learning or spectral CT-derived radiomics. Deep learning has a significant advantage over hand-crafted radiomics, as this method is not limited by predefined features in what information can be captured. Instead, a deep learning approach is given the raw data as input and learns informative features on the fly [36]. Furthermore, spectral CT-derived radiomics were shown to be superior over single energy radiomics for predicting response to checkpoint inhibition in patients with melanoma by Brendlin et al. [24]. As spectral CT scanners become increasingly available, this approach may be tested more thoroughly in future research.

Lastly, the multimodal approach should be extended with other data sources. Accurately predicting checkpoint inhibitor treatment outcomes in melanoma remains challenging. It is possible that individual biomarkers are insufficient to guide clinical decisions. An approach that combines different data sources may therefore prove to be superior. A possible modalities that may be explored for this purpose is histopathology

imaging [16], which will be investigated in this cohort in a future work.

The strengths of this work are the large sample size, the multicenter design and extensive hyperparameter optimisation. This is the largest work published on radiomics for prediction of checkpoint inhibitor treatment outcomes in any malignancy [21]. This large size adds to the weight of the presented conclusion. Furthermore, the dataset in this work includes patients from nine different centres. As stability of radiomics features across scanner types and protocols is far from certain, external validation is essential for determining the practical value of a radiomics approach. Lastly, the proposed pipeline systematically explores design choices, from the extraction of radiomics to the final prediction. This approach should maximise potential performance by avoiding arbitrary and therefore possibly suboptimal design choices.

A potential limitation is the exclusion of a large fraction of patients due to unavailability of CE-CT imaging. Comparison of patient characteristics between the included and excluded groups showed minor differences overall, with a trend towards more progressed disease in the included patients. Our hypothesis for this is that patients with more progressed disease are more likely to directly present to medical oncology, instead of being referred after having undergone imaging by a different specialty where FDG-PET is the preferred modality. Although this selection may theoretically have influenced the presented results, this risk is arguably limited as the characteristics of in- and excluded patients are overall very comparable.

Furthermore, patients with stable disease for a minimum of 6 months were labelled as having clinical benefit. This group could therefore theoretically include patients with indolent tumour progression (less than 120% of original diameters in 6 months), without effect from checkpoint inhibition therapy. However, given the consistent results across outcomes and small proportion of patients for which this may be the case, the impact on eventual results is likely limited.

In conclusion, radiomics are predictive of checkpoint inhibition treatment outcomes in patients with advanced melanoma, but did not improve predictive value over a simpler clinical model. A radiomics model can predict both clinical benefit and response from checkpoint inhibitor therapy with moderate discriminative performance. However, the predictive value of this radiomics model overlaps with that of a clinical model, which is evident from the lack of improvement of a combined model. The added value of a radiomics approach therefore appears to be limited. Future research should focus on related techniques, such as deep learning or radiomics on dual energy CT images. In addition, an approach that combines radiomics and clinical data with other modalities may provide a next step towards accurate prediction of checkpoint inhibitor treatment outcomes in melanoma.

Funding

This research was funded by The Netherlands Organization for Health Research and Development (ZonMW, project number 848101007) and Philips.

CRediT authorship contribution statement

Concept and design: LStM, IAJvD, TL, JJCV, II, GAdW, WAMB, PJvD, JPWP, KPMS, MV. Acquisition, analysis, and interpretation of data: LStM, IAJvD, TL, ERANA, MFT, FWPJvdB, MJBS, MFB, AJMvdE, JWBdG, DP, GV, HMW, EK, PAdJ, KPMS. Drafting of the manuscript: LStM, IAJvD, KPMS, MV. Critical revision of the manuscript: LStM, IAJvD, SGE, TL, JJCV, ERANA, MFT, WAMB, II, GAdW, FWPJvdB, MJBS, MFB, AJMvdE, JWBdG, DP, GV, HMW, EK, PJvD, JPWP, PAdJ, KPMS, MV. Analysis: LStM, IAJvD, SGE, TL, JPWP, PAdJ, KPMS, MV. Supervision: SGE, TL, JJCV, PJvD, JPWP, PAdJ, KPMS, MV.

Data Availability

Due to confidentiality agreements, clinical and imaging data cannot be made available.

Conflict of interest statement

The authors declare the following financial interests/ personal relationships which may be considered as potential competing interests: AvdE has advisory relationships with Amgen, Bristol Myers Squibb, Roche, Novartis, MSD, Pierre Fabre, Sanofi, Pfizer, Ipsen, Merck and has received research study grants not related to this paper from Sanofi, Roche, Bristol Myers Squibb, Idera, and TEVA and has received travel expenses from MSD Oncology, Roche, Pfizer, and Sanofi and has received speaker honoraria from BMS and Novartis. JdG has consultancy/advisory relationships with Bristol Myers Squibb, Pierre Fabre, Servier, MSD, Novartis. PJ has a research collaboration with Philips Healthcare and Vifor Pharma. MBS has consultancy/ advisory relationships with Pierre Fabre, MSD, and Novartis. EK has consultancy/advisory relationships with Bristol Myers Squibb, Novartis, Merck, Pierre Fabre, Lilly, Bayer, EISAI, and Ipsen, and received research grants not related to this paper from Bristol Myers Squibb and Pierre Fabre. PD has consultancy/ advisory relationships with Paige, Pantarei, and Samantree paid to the institution, and research grants from Pfizer, none related to current work, and paid to institute. KS has advisory relationships with Bristol Myers Squibb, Novartis, MSD, Pierre Fabre, AbbVie and received honoraria from Novartis, MSD, and Roche and research funding from Bristol Myers Squibb, TigaTx, and Philips. TL has received research funding from Philips. All remaining authors have declared no conflicts of interest.

Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at doi:10.1016/j.ejca.2023. 02.017.

References

- [1] Korn EL, Liu P-Y, Lee SJ, Chapman J-AW, Niedzwiecki D, Suman VJ, et al. Meta-analysis of phase II cooperative group trials in metastatic stage IV melanoma to determine progressionfree and overall survival benchmarks for future phase II trials. J Clin Oncol 2008;26:527–34. https://doi.org/10.1200/JCO.2007.12. 7837.
- [2] Sondak VK, Smalley KSM, Kudchadkar R, Grippon S, Kirkpatrick P. Ipilimumab. Nat Rev Drug Discov 2011;10:411–3.
- [3] Barone A, Hazarika M, Theoret MR, Mishra-Kalyani P, Chen H, He K, et al. FDA approval summary: pembrolizumab for the treatment of patients with unresectable or metastatic melanoma. Clin Cancer Res 2017;23:5661–5. https://doi.org/10.1158/1078-0432.CCR-16-0664.
- [4] Beaver JA, Theoret MR, Mushti S, He K, Libeg M, Goldberg K, et al. FDA approval of nivolumab for the first-line treatment of patients with BRAFV600 wild-type unresectable or metastatic melanoma. Clin Cancer Res 2017;23:3479–83. https://doi.org/10.1158/1078-0432.CCR-16-0714.
- [5] van Zeijl MCT, Haanen JBAG, Wouters MWJM, de Wreede LC, Jochems A, Aarts MJB, et al. Real-world outcomes of first-line anti-PD-1 therapy for advanced melanoma: a nationwide population-based study. J Immunother 2020;43:256–64. https://doi. org/10.1097/CJI.0000000000000334.
- [6] Larkin J, Chiarion-Sileni V, Gonzalez R, Grob J-J, Rutkowski P, Lao CD, et al. Five-year survival with combined nivolumab and Ipilimumab in advanced melanoma. N Engl J Med 2019;381:1535–46. https://doi.org/10.1056/NEJMoa1910836.
- [7] Asher N, Ben-Betzalel G, Lev-Ari S, Shapira-Frommer R, Steinberg-Silman Y, Gochman N, et al. Real world outcomes of Ipilimumab and Nivolumab in patients with metastatic melanoma. Cancers 2020;12:2329. https://doi.org/10.3390/ cancers12082329.
- [8] Robert C, Long GV, Brady B, Dutriaux C, Maio M, Mortier L, et al. Nivolumab in previously untreated melanoma without BRAF mutation. N Engl J Med 2015;372:320–30. https://doi.org/10.1056/NEJMoa1412082.
- [9] Robert C, Schachter J, Long GV, Arance A, Grob JJ, Mortier L, et al. Pembrolizumab versus Ipilimumab in advanced melanoma. N Engl J Med 2015;372:2521–32. https://doi.org/10.1056/ NEJMoa1503093.
- [10] Verheijden RJ, May AM, Blank CU, van der Veldt AAM, Boers-Sonderen MJ, Aarts MJB, et al. Lower risk of severe checkpoint inhibitor toxicity in more advanced disease. ESMO Open 2020;5:e000945. https://doi.org/10.1136/esmoopen-2020-000945.
- [11] Wolchok JD, Chiarion-Sileni V, Gonzalez R, Grob J-J, Rutkowski P, Lao CD, et al. Long-term outcomes with Nivolumab plus Ipilimumab or Nivolumab alone versus Ipilimumab in patients with advanced melanoma. J Clin Oncol 2022;40:127–37. https://doi.org/10.1200/jco.21.02229.
- [12] Leeneman B, Uyl-de Groot CA, Aarts MJB, van Akkooi ACJ, van den Berkmortel FWPJ, van den Eertwegh AJM, et al. Healthcare costs of metastatic cutaneous melanoma in the era of immunotherapeutic and targeted drugs. Cancers 2020;12:E1003. https://doi.org/10.3390/cancers12041003.

- [13] Verma V, Sprave T, Haque W, Simone CB, Chang JY, Welsh JW, et al. A systematic review of the cost and cost-effectiveness studies of immune checkpoint inhibitors. J Immunother Cancer 2018;6(128):1–5. https://doi.org/10.1186/s40425-018-0442-7.
- [14] da Silva IP, Ahmed T, McQuade JL, Nebhan CA, Park JJ, Versluis JM, et al. Clinical models to define response and survival with anti-PD-1 antibodies alone or combined with Ipilimumab in metastatic melanoma. J Clin Oncol 2022;40:1068–80. https://doi. org/10.1200/JCO.21.01701.
- [15] Morrison C, Pabla S, Conroy JM, Nesline MK, Glenn ST, Dressman D, et al. Predicting response to checkpoint inhibitors in melanoma beyond PD-L1 and mutational burden. J Immunother Cancer 2018;6(32):1–12. https://doi.org/10.1186/ s40425-018-0344-8.
- [16] Johannet P, Coudray N, Donnelly DM, Jour G, Illa-Bochaca I, Xia Y, et al. Using machine learning algorithms to predict immunotherapy response in patients with advanced melanoma. Clin Cancer Res Off J Am Assoc Cancer Res 2021;27:131–40. https:// doi.org/10.1158/1078-0432.CCR-20-2415.
- [17] Gillies RJ, Kinahan PE, Hricak H. Radiomics: images are more than pictures, they are data. Radiology 2016;278:563–77. https:// doi.org/10.1148/radiol.2015151169.
- [18] Way TW, Hadjiiski LM, Sahiner B, Chan H-P, Cascade PN, Kazerooni EA, et al. Computer-aided diagnosis of pulmonary nodules on CT scans: Segmentation and classification using 3D active contours. Med Phys 2006;33:2323–37. https://doi.org/10. 1118/1.2207129.
- [19] Aerts HJWL, Velazquez ER, Leijenaar RTH, Parmar C, Grossmann P, Carvalho S, et al. Decoding tumour phenotype by noninvasive imaging using a quantitative radiomics approach. Nat Commun 2014;5:4006. https://doi.org/10.1038/ncomms5006.
- [20] Gutman DA, Dunn WD, Grossmann P, Cooper LAD, Holder CA, Ligon KL, et al. Somatic mutations associated with MRIderived volumetric features in glioblastoma. Neuroradiology 2015;57:1227–37. https://doi.org/10.1007/s00234-015-1576-7.
- [21] ter Maat LS, van Duin IAJ, Elias SG, van Diest PJ, Pluim JPW, Verhoeff JJC, et al. Imaging to predict checkpoint inhibitor outcomes in cancer. A systematic review. Eur J Cancer 2022;175:60–76. https://doi.org/10.1016/j.ejca.2022.07.034.
- [22] Trebeschi S, Drago SG, Birkbak NJ, Kurilova I, Călin AM, Delli Pizzi A, et al. Predicting response to cancer immunotherapy using noninvasive radiomic biomarkers. Ann Oncol Off J Eur Soc Med Oncol 2019;30:998–1004. https://doi.org/10.1093/annonc/mdz108.
- [23] Peisen F, Hänsch A, Hering A, Brendlin AS, Afat S, Nikolaou K, et al. Combination of whole-body baseline CT radiomics and clinical parameters to predict response and survival in a stage-IV melanoma cohort undergoing immunotherapy. Cancers 2022;14:2992. https://doi.org/10.3390/cancers14122992.
- [24] Brendlin AS, Peisen F, Almansour H, Afat S, Eigentler T, Amaral T, et al. A Machine learning model trained on dual-energy CT radiomics significantly improves immunotherapy response prediction for patients with stage IV melanoma. J Immunother Cancer 2021;9:e003261. https://doi.org/10.1136/jitc-2021-003261.
- [25] Mackin D, Fave X, Zhang L, Fried D, Yang J, Taylor B, et al. Measuring CT scanner variability of radiomics features. Investig Radiol 2015;50:757–65. https://doi.org/10.1097/RLI. 00000000000000180.
- [26] Jochems A, Schouwenburg MG, Leeneman B, Franken MG, van den Eertwegh AJM, Haanen JBAG, et al. Dutch melanoma treatment registry: quality assurance in the care of patients with metastatic melanoma in the Netherlands. Eur J Cancer 2017;72:156–65. https://doi.org/10.1016/j.ejca.2016.11.021.
- [27] Pieper S, Halle M, Kikinis R: 3D Slicer, in: Proceedings of the 2004 2nd IEEE Int. Symp. Biomed. Imaging Nano Macro IEEE Cat No 04EX821. 2004; Vol. 1:p. 632-635. (https://doi.org/10.1109/ISBI.2004.1398617).

- [28] van Griethuysen JJM, Fedorov A, Parmar C, Hosny A, Aucoin N, Narayan V, et al. Computational radiomics system to decode the radiographic phenotype. Cancer Res 2017;77:e104–7. https://doi.org/10.1158/0008-5472.CAN-17-0339.
- [29] Eisenhauer EA, Therasse P, Bogaerts J, Schwartz LH, Sargent D, Ford R, et al. New response evaluation criteria in solid tumours: Revised RECIST guideline (version 1.1). Eur J Cancer 2009;45:228–47. https://doi.org/10.1016/j.ejca.2008.10.026.
- [30] van Not OJ, de Meza MM, van den Eertwegh AJM, Haanen JB, Blank CU, Aarts MJB, et al. Response to immune checkpoint inhibitors in acral melanoma: a nationwide cohort study. Eur J Cancer 2022;167:70–80. https://doi.org/10.1016/j.ejca.2022.02. 026
- [31] van Zeijl MCT, de Wreede LC, van den Eertwegh AJM, Wouters MWJM, Jochems A, Schouwenburg MG, et al. Survival outcomes of patients with advanced melanoma from 2013 to 2017: results of a nationwide population-based registry. Eur J Cancer 2021;144:242–51. https://doi.org/10.1016/j.ejca.2020.11.028.
- [32] LeDell E, Petersen M, van der Laan M. Computationally efficient confidence intervals for cross-validated area under the ROC

- curve estimates. Electron J Stat 2015;9:1583–607. https://doi.org/10.1214/15-EJS1035.
- [33] Collins GS, Reitsma JB, Altman DG, Moons KGM. Transparent Reporting of a multivariable prediction model for Individual Prognosis Or Diagnosis (TRIPOD): the TRIPOD statement. Br J Surg 2015;102:148–58. https://doi.org/10.1002/bjs.9736.
- [34] Dercle L, Zhao B, Gönen M, Moskowitz CS, Firas A, Beylergil V, et al. Early readout on overall survival of patients with melanoma treated with immunotherapy using a novel imaging analysis. JAMA Oncol 2022;8:385–92. https://doi.org/10.1001/jamaoncol.2021.6818.
- [35] Robert C, Long GV, Brady B, Dutriaux C, Giacomo AMD, Mortier L, et al. Five-year outcomes with Nivolumab in patients with wild-type BRAF advanced melanoma. J Clin Oncol 2020;38:3937–46. https://doi.org/10.1200/JCO.20.00995.
- [36] Afshar P, Mohammadi A, Plataniotis KN, Oikonomou A, Benali H. From handcrafted to deep-learning-based cancer radiomics: challenges and opportunities. IEEE Signal Process Mag 2019;36:132–60. https://doi.org/10.1109/MSP.2019.2900993.