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Data-driven donation strategies: understanding and predicting blood donor deferral

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Citation
Vinkenoog, M. (2024, February 15). *Data-driven donation strategies: understanding and predicting blood donor deferral*. Retrieved from https://hdl.handle.net/1887/3717530

Version:	Publisher's Version
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Note: To cite this publication please use the final published version (if applicable).

CHAPTER

6

Associations between symptoms, donor characteristics and IgG antibody response in 2082 COVID-19 convalescent plasma donors

Published in: Frontiers in Immunology 13: 821721. doi:10.3389/fimmu.2022.821721

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Abstract

Background - Many studies already reported on the association between patient characteristics on the severity of COVID-19 disease outcome, but the relation with SARS-CoV-2 antibody levels is less clear.

Methods - To investigate this in more detail, we performed a retrospective observational study in which we used the IgG antibody response from 11 118 longitudinal antibody measurements of 2082 unique COVID convalescent plasma donors. COVID-19 symptoms and donor characteristics were obtained by a questionnaire. Antibody responses were modelled using a linear mixed-effects model.

Results - Our study confirms that the SARS-CoV-2 antibody response is associated with patient characteristics like body mass index and age. Antibody decay was faster in male than in female donors (average half-life of 62 versus 72 days). Most interestingly, we also found that three symptoms (headache, anosmia, nasal cold) were associated with lower peak IgG, while six other symptoms (dry cough, fatigue, diarrhoea, fever, dyspnoea, muscle weakness) were associated with higher IgG concentrations.

Introduction

Severe acute respiratory syndrome corona virus 2 (SARS-CoV-2) emerged late 2019 in China, and by March 2020 was declared a pandemic by the World Health Organization (WHO). As of September 2021, over 200 million individuals have been infected with COVID-19, which has inflicted an immense impact on the healthcare system worldwide. The virus mainly targets the respiratory tract, which can lead from mild symptoms to severe respiratory distress syndrome. Studies have shown that antibody responses against the SARS-CoV-2 spike protein can be first detected 1-3 weeks post symptom onset in most COVID-19 patients, [73, 74] and remain in circulation for up to 1 year. [75, 76, 77, 78] There is however a substantial variation in antibody levels between individuals. [77]

Many studies have reported on the association between disease severity and donor characteristics, such as sex, body mass index (BMI), age, and blood group. Males tend to be more susceptible to develop a severe course of the SARS-CoV-2 virus infection. [79, 80] In addition, age above 50 and obesity are also associated with increased risk of severe outcome. [80, 81, 82, 83] ABO blood type may also play a role in COVID-19 infection, but the exact influence remains unclear. [84, 85]

Antibody responses also seem to be associated with symptoms and clinical information. In general, SARS-CoV-2 antibody levels are higher in patients with a severe disease outcome. [86] A recent study in which COVID-19 convalescent plasma (CCP) donors were followed for three months after symptom resolution showed that greater disease severity, older age, male sex, and high BMI correlate with high SARS-CoV-2 antibody levels. [79, 87] The same study also reported that particularly the symptoms fever, body aches, and low appetite correlate with high SARS-CoV-2 antibody levels. Limitations of this study include a small number of subjects and the low number of longitudinal data points available for each subject, which restricts the possibilities to analyse trends in antibody levels over time and the association with donor characteristics and symptoms.

Here, we aimed to gain a more detailed insight into individual symptoms and donor characteristics and their association with the IgG antibody response. Therefore, we analysed a longitudinal data set of 11 118 anti-RBD antibody measurements of 2082 unique CCP donors. Interestingly, we found that three symptoms (headache, anosmia, nasal cold) were associated with lower peak IgG, while six other symptoms (dry cough, fatigue, diarrhoea, fever, dyspnoea, muscle weakness) were associated with higher IgG concentrations.

Methods

Study population samples

Between April 2020 and March 2021, Sanquin Blood Bank (Amsterdam, the Netherlands) collected samples from over 24 000 COVID-19 recovered adults who enrolled in the CCP programme. Within this programme, plasma is derived from patients that recovered from COVID-19, with the aim to help patients recover from COVID-19. Donation was voluntary and non-remunerated, and donors provided written informed consent before their first donation. Donors were included based on either a positive PCR or presence of anti-RBD IgG antibodies above 80 Arbitrary Units per ml (AU/ml) and after being free of symptoms for at least two weeks. Donors donated plasma on average every two weeks, until antibody levels were below 4 AU/ml in two consecutive donations. Only donors with at least three consecutive antibody measurements and a complete questionnaire were included in the analyses, resulting in a study population of 2082 donors. Supplementary Figure S6.1 shows the number of donors that were excluded at each step.

Questionnaire

Starting August 2020, donors that enrolled in the convalescent plasma programme were invited by e-mail to fill out an online questionnaire, programmed in Qualtrics (SAP, Walldorf, Germany). The questionnaire included questions about the possible origin of the infection, the reason why donors were tested and a list of 18 symptoms considered to be COVID-19-related according guidelines specified by the Dutch National Institute for Public Health and the Environment. [88] Participants could indicate if they experienced symptoms and, if the symptoms were present, how severe these symptoms were on a 4-point scale, from 1 (very mild) to 4 (severe). Additionally, participants were asked about the duration of their symptoms, whether they consulted a physician or were admitted to hospital and/or intensive care units. The full questionnaire is included as an online supplement. Donors were excluded from analysis if sex, age and/or date of illness was absent.

Antibody measurements

IgG to RBD was measured essentially as described before. [77, 78] In brief, plates were coated with recombinant RBD, incubated with samples, and bound IgG antibodies

were detected using an anti-human IgG antibody (MH16, Sanquin); quantification was done relative to a plasma pool consisting of CCP donors and expressed as AU/mL.

Statistical model

Longitudinal trends in antibody levels were analysed with a linear mixed-effects model, using log-transformed anti-RBD IgG levels as outcome variable. Timepoint 0 corresponds to 20 days post onset of symptoms. [73] As such, the estimated intercept of the model corresponds to a donor's estimated peak IgG level. [89] The estimated slope of the model is used to calculate a donor's IgG half-life, in days: $t_{1/2} = log(\frac{1}{2})/slope$.

Only measurements within six months post onset of symptoms were included, as in later stages of recovery antibody decline is expected to slow down and no longer expected to follow a loglinear decline. [77]

A three-step approach was used to analyse the effects of the covariates. In the first step, a null-model was fit to the data, using time as the only predictor variable and allowing a random intercept and slope to be estimated for each donor. In the second step, we tried to explain the variance in random intercepts and slopes by including fixed effects for donor characteristics, i.e., sex, age, height, weight, BMI, and blood group (ABO and RhD), in addition to the random intercept and slope per donor. In the third step, fixed effects that were statistically insignificant in the second step were removed and additional donor information variables obtained from the questionnaires were added as fixed effects. This information concerned data on hospitalisation, ICU admission, co-morbidities, and the presence of 18 symptoms as shown in Table 6.1. This approach allowed separate estimation of the proportion of variance explained by donor characteristics and clinical information.

Significance levels of individual variables were estimated using Satterthwaite's approximation, as degrees of freedom cannot be calculated exactly in models that include both random and fixed effects. [90] Because this approximation is slightly anti-conservative, an alpha-level of 0.01 was chosen to determine statistical significance. Non-significant predictors were excluded after each step. Relative quality, taking into account both goodness of fit and model complexity, of the models was assessed by comparing the Akaike information criteria (AIC) after each step.

Data were processed and analysed with the R programming language and environment for statistical computing (version 4.0.3), using packages lme4 and lmerTest for analyses and ggplot2 for generating graphs.

Results

Study population characteristics

We used 11 118 antibody measurements of 2082 unique donors to study the associations between symptoms, donor characteristics, and IgG antibody response. The number of available antibody measurements per donor ranged from 3 to 18 measurements. In addition, each donor completed a questionnaire, which gave insight into symptoms and donor characteristics. Table 6.1 shows the distributions of donor and COVID-19 related disease characteristics in the study population.

Compared to all active whole-blood and plasma donors in 2020, donors in our study population are slightly older (46 vs 42 years for women, 52 vs 48 years for men). Median weight and height, as well as proportion of female donors and rhesus D blood group are similar to those of the active donor population. Blood group A is overrepresented in our study population (47% vs 39% for women, 45% vs 39% for men), while blood group O is underrepresented (39% vs 47% for women, 42% vs 47% for men).

Table 6.1: Study population characteristics. Continuous variables are represented by their median and interquartile range (IQR), categorical variables by absolute count and percentage.

	Women		Men	
	Median or count	IQR or percentage	Median or count	IQR or percentage
Number of donors (proportion of total)	1236	59.4%	846	40.6%
Number of donations per donor	6	4 – 8	6	4 – 10
Days POS at first donation	48	33 – 77	47	32 – 77
Days POS at last donation*	122	97 – 151	126	103 – 157
Age (years)	45.9	28.0 – 55.3	51.8	39.6 – 59.3
Height (cm)	171	167 – 176	184	180 – 189
Weight (kg)	73	65 – 83	88	80 – 97
BMI (kg/m2)	24.8	22.6 – 28.4	26.4	24.0 – 28.2

Blood group ABO				
A	581	47%	381	45%
B	120	9.7%	84	9.9%
O	484	39%	352	42%
AB	51	4.1%	29	3.4%
Blood group RhD				
Positive	1024	83%	691	82%
Negative	212	17%	155	18%
Hospital admission	19	1.5%	50	5.9%
Intensive care	4	0.3%	8	0.9%
Symptoms				
Asymptomatic	*8*	*0.6%*	*7*	*0.8%*
Fatigue	979	79%	597	71%
Anosmia/ageusia	853	69%	471	56%
Headache	820	66%	467	55%
Myalgia	705	57%	445	53%
Nasal cold	692	56%	424	50%
Fever	621	50%	507	60%
Dry cough	560	45%	396	47%
Sore throat	519	42%	307	36%
Chills	499	40%	356	42%
Sneezing	461	37%	381	45%
Dyspnoea	461	37%	297	35%
Muscle weakness	426	34%	260	31%
Diarrhoea	221	18%	102	12%
Nausea	184	15%	72	8.5%
Sputum production	178	14%	152	18%
Altered mental status	127	10%	80	9.5%
Skin rash	69	5.6%	27	3.2%
Vomiting	49	4.0%	28	3.3%

Null-model fit (step 1)

In the first step we estimated an intercept and slope for each individual donor using the null model, describing the linear relationship between log-transformed IgG levels and time post onset of symptoms. [73] The residuals, i.e., the difference between measured IgG and predicted IgG as estimated by the null model, follow a normal distribution with mean 0 and standard deviation of 0.21 log (AU/ml). This distribution is independent of time post onset symptoms, supporting the assumption that the relationship is linear after log-transformation. Given this assumption, the estimated peak IgG level set at 20 days POS is most likely an accurate extrapolation and allows for comparisons between donors. Supplementary Figure S6.2A shows the fitted line and actual measurements for four randomly selected donors (donors A to D). Supplementary Figure S6.2B shows the distribution of the residuals over all observations for all donors.

After analysing all samples, we found a median peak IgG concentration of 38.8 AU/ml (IQR 20.9-78.6) and a median half-life of 66 days (IQR 50-94) (Figure 6.1). For the majority of donors, the estimated slope corresponds to a plausible antibody half-life. However, for 80 donors (3.8%), the fitted slope was positive, which results in a negative estimated half-life estimate. For an additional 59 donors (2.8%), the estimated half-life is extremely long (defined here as more than 365 days, but estimates ranged up to 16 000 days). This occurs when the estimated slope is very close to zero (but still negative), which may happen when IgG levels barely decrease between measurements and no decay in antibody levels are measured. Examples of donors with a negative half-life and very long half-life are given in Supplementary Figure S6.3. These donors were not excluded from the study in order not to overstate accuracy, and because there was no reason to assume the IgG measurements were incorrect.

Associations with predictor variables

The results of step 2, where individual donor characteristics were added to the model as predictor variables, are shown in Figures 6.2A–C and Table 6.2. Sex was associated with the slope (Figure 6.2A), as the rate of antibody decay is faster in men: the median slope for men corresponds to a half-life of 62 days, while this is 72 days for women. Men displayed higher peak IgG levels than women, but this difference was not statistically significant ($p = 0.68$). Age (Figure 6.2B) and BMI (Figure 6.2C) were both positively correlated with peak IgG concentration. A one-year increase in age corresponds to a 0.013 increase in the log-transformed IgG level, an increase of one BMI point corresponds to a 0.024 increase in log-transformed IgG level. No

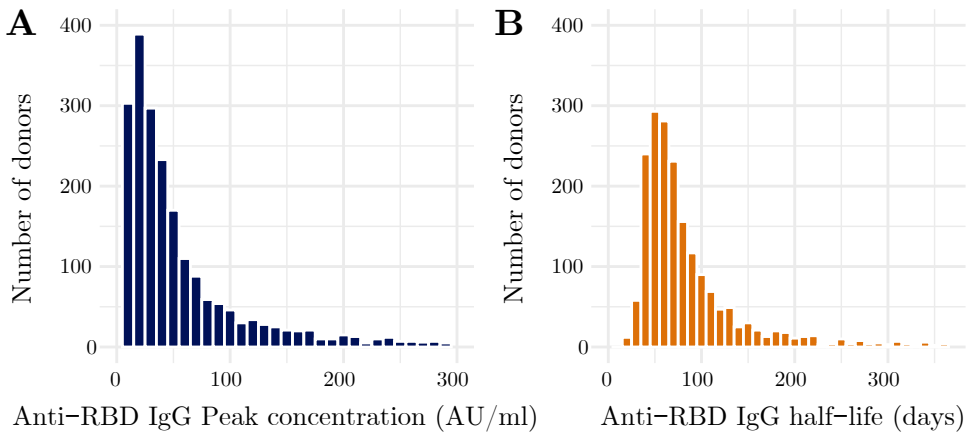

Figure 6.1: Anti-RBD IgG peak and half-life. (A) Distribution of estimated peak IgG concentration (at 20 days POS) and (B) estimated half-life of 2082 COVID convalescent plasma donors, as estimated by the null model. Please note that since both distributions have an extremely long right tail, the horizontal axes are truncated at (A) 300 AU/ml and (B) 365 days, excluding 70 and 139 donors from left and right histograms, respectively.

significant associations with antibody titres were found for variables blood group, height, and weight. Random effects for peak IgG level and antibody half-life are positively correlated with a correlation coefficient of 0.29, indicating that higher peak IgG is moderately associated with higher (less negative) slope, and therefore with a longer half-life.

Associations with clinical information

After adding clinical information significant associations with peak IgG concentration were found for hospital admission and various clinical symptoms (Figures 6.2D, 6.3, 6.4 and Table 6.2). Hospital admission was significantly associated with both higher peak IgG level and shorter half-life (Figure 6.2D). Nasal cold, headache, and anosmia were associated with lower peak IgG levels, while dry cough, fatigue, fever, dyspnoea, diarrhoea, and muscle weakness were associated with higher peak IgG levels. Figure 6.3 shows the estimated peak IgG level when these symptoms are present. Note that values on the y-axis are the predicted peak IgG levels when all continuous variables are equal to their average value, and all binary variables (hospital admission and all other symptoms) equal zero.

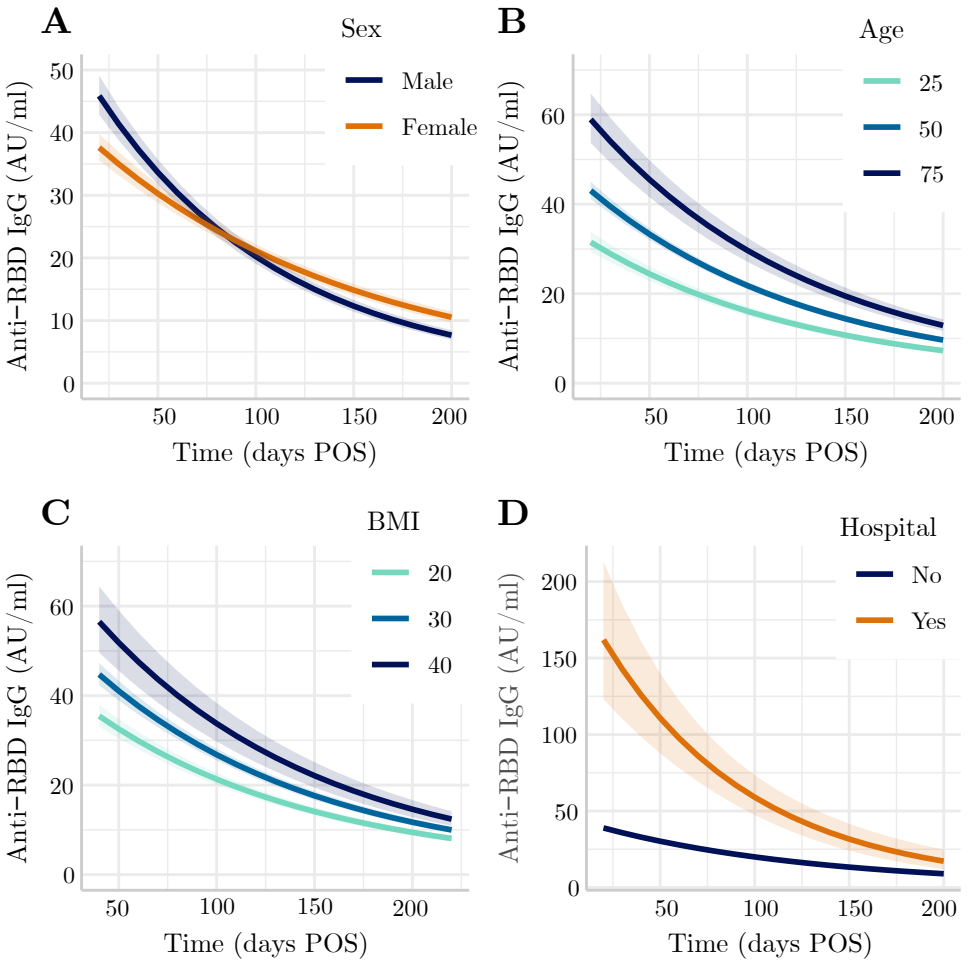

Figure 6.2: Associations between donor/clinical characteristics and antibody levels. The effects of variables (A) sex, (B) age, (C) BMI, and (D) hospital admission on predicted antibody decline. Note that age and BMI are included in the model as continuous predictors; for clarity, the associations are only plotted for three values. Light-coloured bands represent 95% confidence intervals.

Term	Estimate	95% CI
Random effects		
Intercept [log(peak IgG)]	2.382	2.274 – 2.490
Slope [delta log(IgG) per day]	-0.010	-0.011 – -0.101
Fixed effects on the intercept		
*Sex: female	-0.017	-0.063 – 0.096
Age (per 10 years)	0.128	0.100 – 0.157
BMI (per 5 points)	0.119	0.097 – 0.164
Hospital admission: yes	1.156	0.934 – 1.379
Headache: yes	-0.113	-0.193 – -0.032
Anosmia: yes	-0.111	-0.189 – -0.033
Nasal cold: yes	-0.101	-0.177 – -0.025
Dry cough: yes	0.095	0.019 – 0.171
Fatigue: yes	0.140	0.044 – 0.236
Diarrhoea: yes	0.148	0.043 – 0.252
Muscle weakness: yes	0.172	0.083 – 0.261
Shortness of breath: yes	0.196	0.111 – 0.280
Fever: yes	0.228	0.149 – 0.308
Fixed effects on the slope		
Sex: female	0.003	0.002 – 0.004
Hospital admission: yes	-0.004	-0.007 – -0.001

Table 6.2: Point estimates and 95% confidence intervals of fixed effects on log-transformed IgG levels.
* The effect of sex on the intercept (peak IgG) was not statistically significant, but the variable is not excluded due to its effect on the slope.

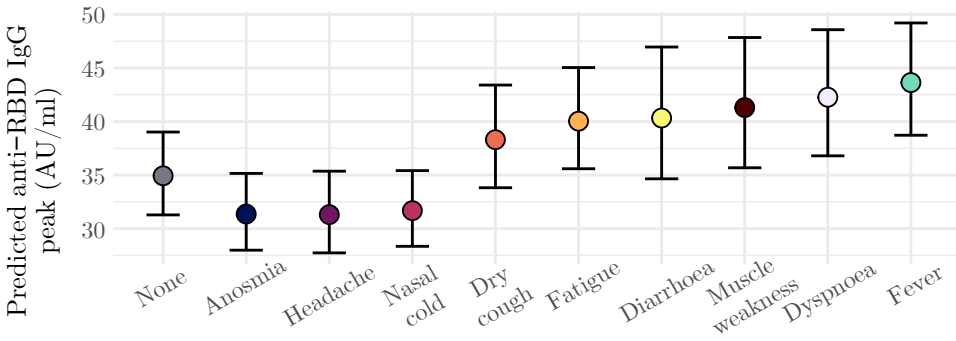

Figure 6.3: Predicted impact of various symptoms on anti-RBD IgG peak concentration. Estimated peak IgG concentrations when different symptoms are displayed. For each of the symptoms here, the difference in peak IgG as compared to the group without this symptom is statistically significant with $p < 0.001$.

The largest difference was found for the variable *hospital admission*. Donors admitted to the hospital had considerably higher antibody levels, with an estimated difference of 77.8 AU/ml on the peak IgG concentration. These donors also have a faster rate of antibody decay, corresponding to an estimated half-life of 48 days (95% CI: 40-58 days) for men, and 60 days (95% CI: 49-80 days) for women.

Variance explained by model

In the null-model that was fitted in step 1 (without any fixed effects), all variation in peak IgG and half-life was attributed to the individual variation per donor. As fixed effects were added in step 2, part of this variation was now explained by these fixed effects, and the variation explained by the random effects decreased. Table 6.3 shows the variance of the random effects per donor in the null-model, as well the variance of the random effects as after adding donor characteristics as covariates (step 2), and after adding the clinical information (step 3). The variance reduction relative to the null-model (step 1) by the addition of extra explanatory variables in each step is also provided. Model fit was compared using the Akaike Information Criterion (AIC) and tested for statistical significance using a nested ANOVA, results of which are shown in Table 6.3.

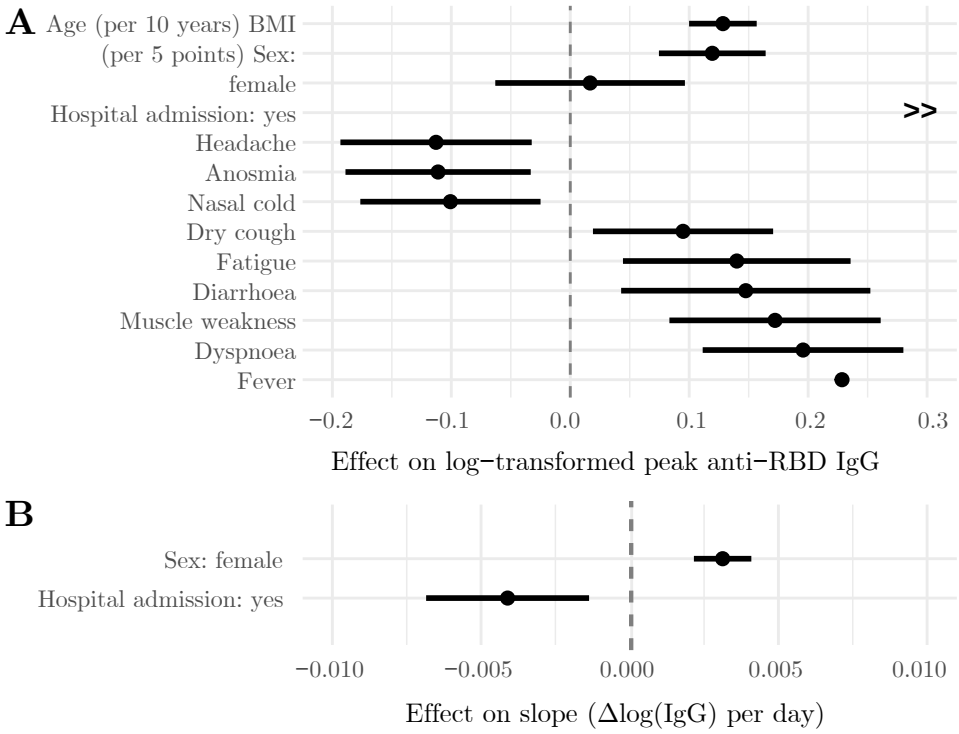

Figure 6.4: Effect size and 95% confidence intervals of fixed effects on anti-RBD IgG peak concentration (log-transformed) and the slope.

	Variance of random effect on peak log(IgG)	Variance of random effect on slope Δlog(IgG)/day	AIC
Step 1: null-model	0.8814	0.0497	11886
Step 2: donor characteristics	0.7758 (-12%)	0.0485 (-2.4%)	11615 ($p < 0.001$)
Step 3: donor characteristics + clinical information	0.6610 (-25%)	0.0481 (-3.2%)	11290 ($p < 0.001$)

Table 6.3: Variance of random effects in models of all three steps. Percentual variance decrease relative to the null-model is given in brackets. P-values are relative to the previous step, obtained with ANOVA.

Discussion

In this retrospective observational study, we investigated potential associations between SARS-CoV-2 specific antibody kinetics and various donor characteristics and COVID-19 symptoms. To our knowledge, this is currently the largest study that describes such associations. Individual antibody responses were modelled using a linear mixed-effects model, from which peak IgG concentration and antibody half-life were determined. Symptoms and donor characteristics were obtained from a questionnaire. Our study shows that the SARS-CoV-2 antibody response is associated with patient characteristics like sex, age, and BMI. Of note, we also found that specific COVID-19 symptoms are associated with antibody levels.

As reported earlier, we found a large variation in anti-RBD antibody peak levels. A strength of our study are the longitudinal measurements, which enabled us to reliably estimate the peak level of each individual donor independent on the timing of the first antibody measurement. Only a quarter of the variation in peak IgG concentration between patients can be explained by associations with donor characteristics and disease symptoms. To a lesser degree, donor characteristics were also associated with differences in antibody half-life, which was also variable between donors, albeit less than the peak level. The antibody half-life was not correlated to peak levels. Whether these differences in antibody half-life reflect differences in protection for re-infection will be investigated, and this thoroughly characterised donor cohort can serve as bench mark for those studies.

Six symptoms (dry cough, fatigue, diarrhoea, fever, dyspnoea, muscle weakness) were associated with higher IgG concentrations and three symptoms (headache, anosmia, nasal cold) were associated with lower peak IgG concentrations against RBD. This association between symptoms and antibody levels may possibly reflect the fact that the SARS-CoV-2 virus frequently initiates infection in the upper airways (mild symptoms and low IgG levels) before spreading through the body (severe symptoms and high IgG levels). Headache, anosmia and nasal cold were common symptoms, each present in at least 50% of patients in our population. Fatigue was present in more than 70% of patients and was associated with higher peak IgG concentration, suggesting more severe illness. A previous study in a hospital cohort found that fatigue and dyspnoea are prognostic for severe infection, and a stuffed nose (comparable to nasal cold) for mild infection, which is in line with our findings. [91]

Furthermore, we found higher age and BMI to be associated with higher peak IgG concentrations. Sex was not associated with peak IgG concentration, but men

had significantly shorter antibody half-lives than women (62 vs 72 days respectively). The small group of patients that had been admitted to hospital displayed both higher peak IgG concentrations and shorter half-lives. Probably this effect is the result of the presence of short-lived plasmablasts that produce high levels of antibodies. Previous studies found sex differences in COVID-19 immune responses, with higher IgG concentrations associated with male sex, older age, and hospitalisation. [92, 93, 94] Although our results are consistent with these findings for age and hospitalisation, we found that the association between male sex and higher peak IgG concentration was not significant after correction for age and BMI. This suggests that the previously found association with male gender was possibly due to the increased risk of severe disease in men. Most studies on differences in antibody response are performed in hospital cohorts, our study population consisted mainly of recovered patients that were not admitted to hospital (96.7%), and therefore disease severity is expected to be lower. Consistently, BMI in the non-hospitalised group was 25.9 compared to 28.8 in hospitalised patients.

A strength of our study is the large number of recovered patients included in our study population. The status of Sanquin as the only blood bank in the Netherlands, combined with well-established connections with municipal health services, allowed us to invite people with a positive PCR test to become CCP donors after recovery. This allowed us to both include non-hospitalised and hospitalised patients in the cohort. However, we could only include donors who were healthy enough to regularly donate plasma, which means that our results are mainly applicable towards patients with a mild outcome. As a result our study is more representative of the total COVID-19 patient population than studies on hospitalised patient cohorts. It should also be noted that some bias may be present in our data, as symptoms are self-reported by patients after recovery. Relatively mild symptoms, such as nasal cold, may therefore be underreported by patients who at the same time experienced more severe symptoms, such as fever or dyspnoea. However, this explanation is unlikely to negate the association we found, as all symptoms associated with lower peak IgG were present in more than 50% of patients.

In conclusion, our study indicates that several COVID-19 symptoms are associated with SARS-CoV-2 antibody levels in addition to the previously described association with sex, age, and BMI. Discovery of these associations aids us in understanding why antibody responses differ between patients. The predictive value of IgG concentrations could also be used by blood banks to pre-select individuals with high and/or stable antibody levels as potential CCP donors.

Appendix

Questionnaire anti-SARS-CoV-2 donors

Note: the original questionnaire was in Dutch, this is a translated version.

Q1 What is your donor ID? You can find this in the accompanying email.

Q36 What is your date of birth?

Day _____ - Month _____ - Year _____

Q2 What is your sex?

- ❍ Male
- ❍ Female

Q3 How would you describe your COVID-19 status?

- ❍ I suspect I have had COVID-19 because I have had a positive PCR test.
- ❍ I suspect I have had COVID-19 because antibodies have been detected in my blood.
- ❍ Other: _____

Q4 Where did you contract the infection?

- ❍ In the Netherlands
- ❍ Abroad

Q5 (if Q4 answered with 'abroad') In which country did you contract the infection?

Q6 Why were you tested for presence of the coronavirus?

- ❍ Because I was ill/had symptoms
- ❍ Because I was in contact with a (possibly) infected person
- ❍ Because of my occupation (health care, contact profession)
- ❍ Other: _____

Q7 Did you experience the following symptoms?

- ❑ Nasal cold/coryza
- ❑ Sore throat
- ❑ Dry cough
- ❑ Fatigue
- ❑ Sputum production
- ❑ Muscle or joint ache
- ❑ Headache
- ❑ Fever
- ❑ Shortness of breath
- ❑ Diarrhoea
- ❑ Nausea
- ❑ Vomiting
- ❑ Chills
- ❑ Sneezing
- ❑ Skin rash
- ❑ Feeling confused
- ❑ Muscle weakness
- ❑ Loss of/less smell or taste

Q8-25 (for each symptom answered with 'yes' in Q7) How much were you affected by this symptom?

- ❍ Very mildly affected
- ❍ Mildly affected
- ❍ Moderately affected
- ❍ Severely affected

Q37 Did you have pneumonia?

- ❍ Yes
- ❍ No

Q26 When did your symptoms start? If you don't remember exactly, please make an estimate.

Day _____ - Month _____ - Year _____

Q27 When did your symptoms end? If you don't remember exactly, please make an estimate.

Day _____ - Month _____ - Year _____

Q28 Were you admitted to hospital for these symptoms?

- ❍ Yes
- ❍ No

Q29 (if Q28 is 'yes') On which date were you admitted to hospital?

Day _____ - Month _____ - Year _____

Q30 (if Q28 is 'yes') Were you admitted to intensive care?

- ❍ Yes
- ❍ No

Q31 (if Q30 is 'yes') How many days were you in intensive care in total?

Q32 (if Q28 is 'yes') Were you given extra oxygen?

- ❍ Yes
- ❍ No

Q33 (if Q28 is 'yes') Did you receive artificial ventilation?

- ❍ Yes
- ❍ No

Q34 (if Q28 is 'yes') On which date were you discharged from the hospital?

Day _____ - Month _____ - Year _____

Q35 Are you in one or more of the following risk groups?

- ❑ People aged 70 or older
- ❑ People with chronic airway or lung disease and under treatment by pulmonologist
- ❑ Chronic heart disease patients under treatment by cardiologist
- ❑ People with diabetes that is not well regulated and/or with complications
- ❑ People with kidney disease who need dialysis or are waiting for a kidney transplantation
- ❑ People with lowered immunity to infection due to medication use for autoimmune disease
- ❑ People who have had an organ or stem cell transplantation
- ❑ People without a spleen or without a functioning spleen
- ❑ People with a blood disease
- ❑ People with lowered immunity due to immunity-lowering medication
- ❑ Cancer patients who have had chemotherapy and/or radiation in the past 3 months
- ❑ People with severe immune disorder that requires medical treatment
- ❑ People with HIV infection who are not (yet) under treatment, or with HIV infection with CD4 under 200/mm2
- ❑ People with severe liver disease
- ❑ People with a BMI over 40

Q38 If there is anything you would like to add, or explain an answer further, please do so here.

Supplemental figures and tables

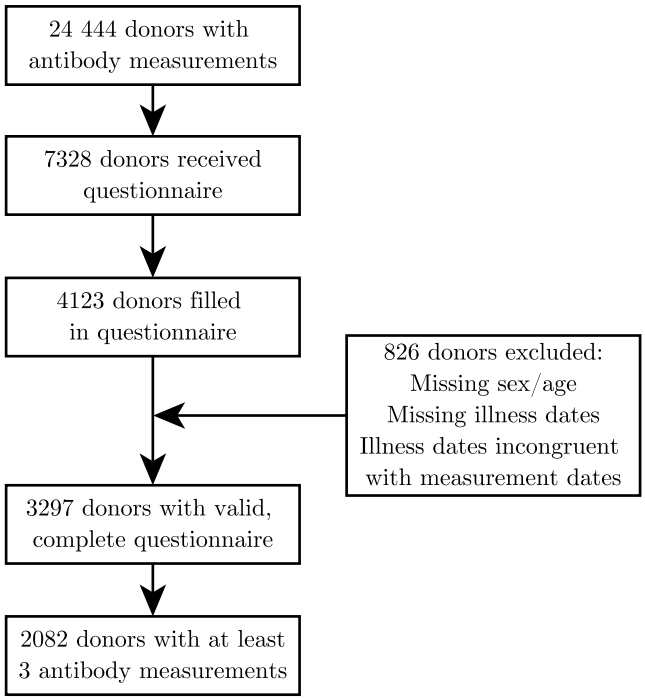

Figure S6.1: Flowchart showing the criteria for inclusion and exclusion of donors.

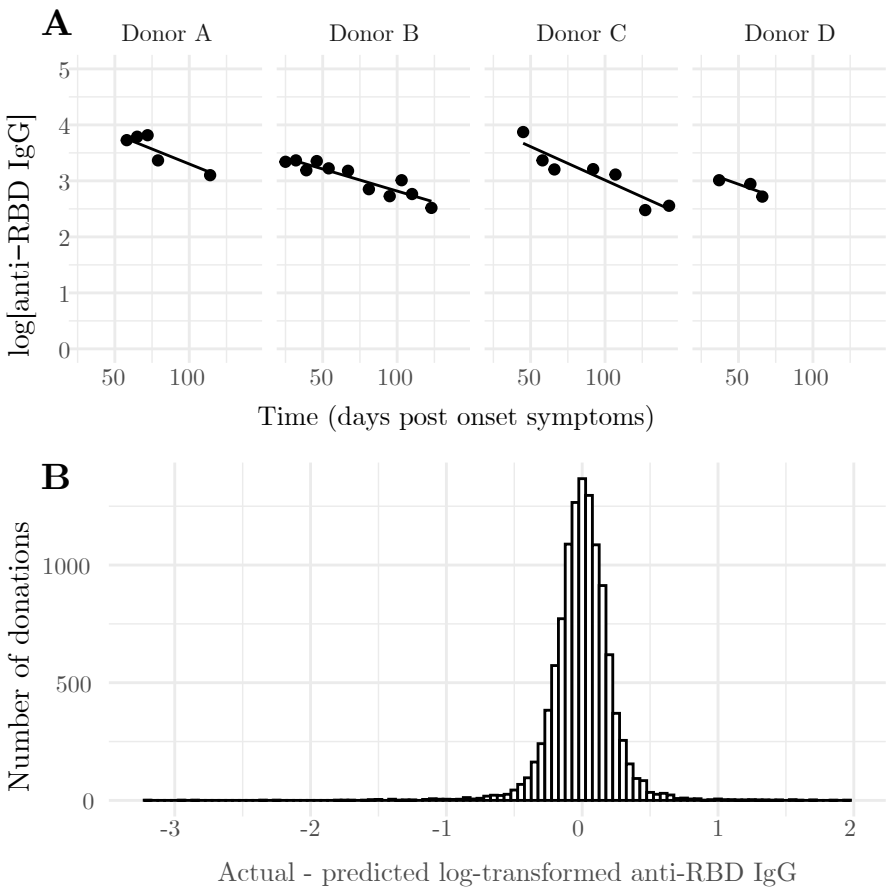

Figure S6.2: Null model fit (step 1). (A) Measured anti-RBD IgG levels (points) and fitted line as estimated by the linear model for four randomly selected donors, and (B) distribution of residuals over all observations, for all donors.

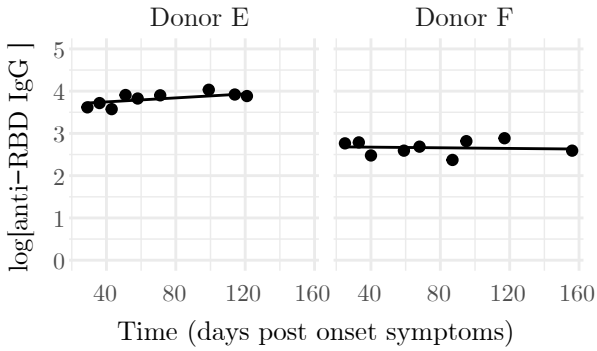

Figure S6.3: No decay in antibody levels. Example of a donor with increasing IgG levels (left panel) and one with near-constant IgG levels (right panel). Estimated slopes for these donors are 0.0024 and -0.0151, corresponding to estimated half-lives of -292 and 1843 days, respectively.

Fixed effect on intercept	Sum of squares	P-value
Weight	0.0070	0.762
Blood group ABO	0.4456	0.121
Height	0.3630	0.030
Blood group RhD	0.4638	0.014
BMI	3.888	<0.001 *
Age	8.752	<0.001 *
Fixed effect on slope	**Sum of squares**	**P-value**
BMI	0.002	0.890
Age	0.004	0.831
Height	0.009	0.735
Weight	0.035	0.500
Blood group ABO	0.188	0.483
Blood group RhD	0.085	0.294
Sex	3.403	<0.001 *

Table S6.1: Sum of squares and p-values of fixed effects after step 2, calculated by backward stepwise reduction

Fixed effect on intercept	Sum of squares	P-value
Sneezing	0.000	0.971
Vomiting	0.004	0.812
Confusion	0.011	0.700
Coughing up mucus	0.084	0.296
Throat ache	0.120	0.210
Joint/muscle ache	0.251	0.070
Nausea	0.247	0.072
Intensive care admission	0.274	0.059
Shivers	0.345	0.034
Skin rash	0.451	0.015
Anosmia	0.645	0.004 *
Fatigue	0.567	0.003 *
Nasal cold	0.449	0.002 *
Dry cough	0.394	0.002 *
Diarrhoea	0.462	0.001 *
BMI	2.000	<0.001 *
Hospital admission	8.555	<0.001 *
Fever	1.836	<0.001 *
Shortness of breath	1.510	<0.001 *
Fixed effect on slope	Sum of squares	P-value
Sex	3.470	<0.001 *

Table S6.2: Sum of squares and p-values of fixed effects after step 3, calculated by backward stepwise reduction.