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Unravelling the mystery of migraine and cluster headache: insights into the genetics and biochemistry of these neurological disorders

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Prostaglandin-E₂ levels over the course of glyceryl trinitrate provoked migraine attacks

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Abstract

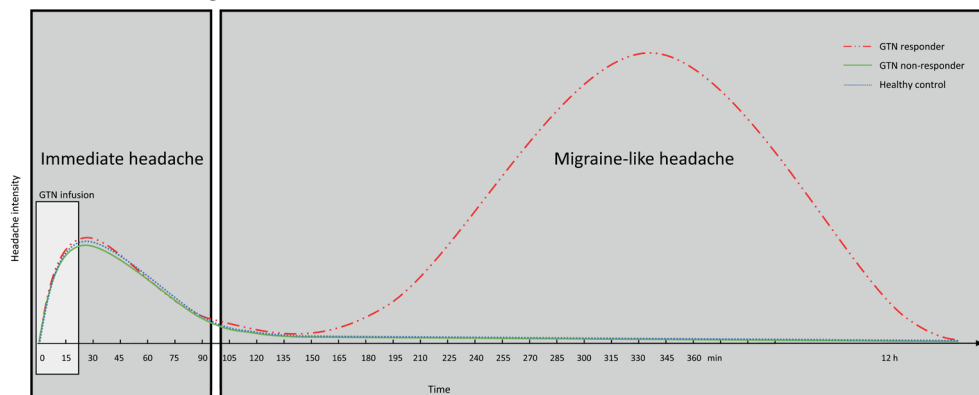
Administration of glyceryl trinitrate (GTN), a donor of nitric oxide, can induce migraine-like attacks in subjects with migraine. Provocation with GTN typically follows a biphasic pattern; it induces immediate headache in subjects with migraine, as well as in healthy controls, whereafter only subjects with migraine may develop a migraine-like headache several hours later. Interestingly, intravenous infusion with prostaglandin-E₂ (PGE₂) can also provoke a migraine-like headache, but seems to have a more rapid onset compared to GTN. The aim of the study was to shed light on the mechanistic aspect PGE₂ has in migraine attack development. Therefore, PGE₂ plasma levels were measured towards the (pre)ictal state of an attack, which we provoked with GTN. Blood samples from women with migraine (n = 37) and age-matched female controls (n = 25) were obtained before and ~140 min and ~320 min after GTN infusion. PGE₂ levels were measured using liquid chromatography tandem mass spectrometry (LC-MS/MS) analysis. Data was analyzed using a generalized linear mixed-effect model. Immediate headache after GTN infusion occurred in 85% of migraine participants and in 75% of controls. A delayed onset migraine-like attack was observed in 82% of migraine subjects and in none of the controls. PGE₂ levels were not different between the interictal and preictal state ($P = 0.527$) nor between interictal and ictal state (defined as having migraine-like headache) ($P = 0.141$). Hence, no evidence was found that a rise in PGE₂ is an essential step in the initiation of GTN-induced migraine-like attacks.

KEYWORDS: Migraine, Glyceryl trinitrate, Prostaglandin E₂, Preictal, Plasma

Introduction

Migraine is a common multifactorial paroxysmal brain disorder with a life-time prevalence of 15-20%, causing disability worldwide.^{1,2} A typical migraine attack consists of a preictal, an ictal (aura and/or headache), and a postictal (postdromal) phase.³ The pathophysiological mechanisms underlying migraine attacks, however, remain to be fully elucidated. Notably, migraine-like attacks can be induced in subjects with migraine, but not in healthy controls, by the administration of glyceryl trinitrate (GTN), a donor of nitric oxide (NO). Two types of NO-induced headaches have been reported (**Figure 1**).⁴ First, in both migraine subjects and healthy controls an immediate headache develops within the first hour of GTN infusion. This headache is of mild to medium severity and typically resolves within an hour after GTN administration. Second, only in subjects with migraine, a delayed onset migraine-like headache (moderate to severe, accompanied by associated symptoms such as nausea, vomiting, photo- and/or phonophobia) may develop within 12 hours after GTN infusion.^{5,6} This different response to GTN in cases compared to controls may provide clues for mechanisms underlying migraine attacks. Whereas the immediate headache seems related to a direct action of the NO-cGMP pathway via vasodilation by smooth muscle relaxation,⁷ independent of neuropeptide calcitonin gene-related peptide (CGRP) release,⁸ the delayed migraine-like attack is thought to be the result of trigeminovascular activation mediated via CGRP release.^{5,7,9}

Figure 1 Schematic headache pattern after the start of the GTN infusion consisting of the immediate headache and the migraine-like attack



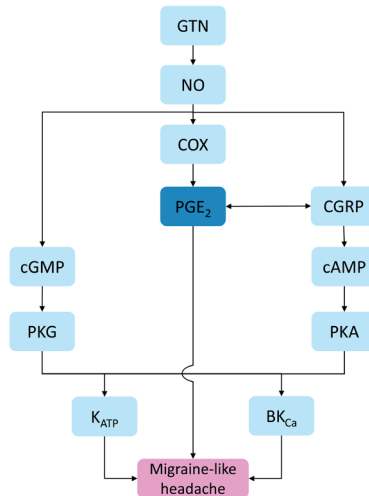
Three different response groups can be distinguished. The red two-dot chain line represents a typical headache pattern for a subject with migraine who responded to GTN (GTN responder), this is combined with typical patterns for a subject with migraine who did not respond to GTN (GTN non-responder) represented by the continuous green line, and a healthy control represented by the dotted blue line. GTN, glyceryl trinitrate. Adapted from Onderwater et al.²¹

Besides CGRP there is ample evidence that prostaglandins may be pivotal in the development of GTN-induced migraine-like attacks, and possibly spontaneous migraine attacks.¹⁰ NO stimulates cyclooxygenase (COX-1 and COX-2) synthesis, which are enzymes that produce prostaglandins.¹¹ Non-steroidal anti-inflammatory drugs (NSAIDs), which inhibit prostaglandin synthesis, are a first

line treatment for migraine headaches. Cortical spreading depolarization, the underlying mechanism for the migraine aura, causes COX-2 upregulation potentially leading to increased prostaglandin levels.^{12,13} The role of prostaglandins has also been investigated in provocation experiments in migraine subjects, in most cases in those without aura, demonstrating that intravenous infusion of prostaglandin I₂ (PGI₂) and E₂ (PGE₂) induces migraine like-attacks in 75% of participants with migraine.^{14,15} Remarkably, subjects with migraine typically developed rapid onset migraine-like attacks, with a median onset of 20 minutes, in 25% (PGI₂) and 58% (PGE₂) of cases, which is in contrast to provocation with GTN, pituitary adenylate cyclase-activating peptide (PACAP) and CGRP for which the majority of cases develops a delayed onset migraine-like attack after at least a few hours.^{14,16}

It has been shown that PGE₂ is mediated via CGRP release, and *vice versa*,¹⁰ as evidenced by observations that PGE₂ stimulates the release of CGRP in rat trigeminal neurons,¹⁷ trigeminal nucleus caudalis¹⁸, and trigeminal ganglia,¹⁹ while CGRP induces secondary release of PGE₂.²⁰ All the above suggests that PGE₂ may be closely upstream of GTN-induced migraine attacks (**Figure 2**).

Figure 2 Pathway relevant to nitroglycerin (GTN)-induced migraine-like headache



Nitroglycerine (GTN) liberates nitric oxide (NO) in peripheral and cerebral structures. NO subsequently, by binding to soluble guanylyl cyclase (sGC), increases cyclic guanosine monophosphate (cGMP).²² Furthermore, NO can interact with superoxide to form peroxynitrite. Peroxynitrite (ONOO⁻) is a proinflammatory compound and has been implicated in the pathophysiology of not only stroke, but also pain and is gaining interest in the migraine field.^{23,24} Additionally, NO on the one hand stimulates COX synthesis and prostaglandin E₂ (PGE₂) production,¹¹ and on the other hand stimulates CGRP, independent of the cGMP signaling pathway.⁸ Subsequently, CGRP has been shown to induce PGE₂,²⁰ and vice versa.¹⁷⁻¹⁹ In turn, it has been shown that ONOO⁻ when inducing inflammation-derived hyperalgesia acts via the COX-to-PGE₂ pathway,²⁵ and ONOO⁻ is also implicated along the trigeminovascular migraine pathway associated with CGRP.²⁶ PKG-mediated phosphorylation opens ATP-sensitive potassium channels (K_{ATP}) channels and large (big)-conductance calcium-activated K⁺ (BK_{Ca}) channels via the NO/cGMP/PKG pathway.^{27,28} CGRP activates vascular smooth muscle K_{ATP} channels and BK_{Ca} channels via cyclic adenosine monophosphate (cAMP) and protein kinase A (PKA) phosphorylation.^{29,30} PGE₂ can also either increase or decrease the amount of cAMP depending on to which receptor it binds.³¹ Opening of K_{ATP} and BK_{Ca} channels generates outward K⁺ currents and causes vasodilation,³² and can eventually lead to a migraine-like attack.^{33,34} Provocation with PGE₂ in subjects with migraine leads to a rapid-onset migraine attack,¹⁴ which suggests that PGE₂ is closely upstream of a migraine-like attack.

We here aimed to shed light on the mechanistic aspect PGE₂ has in migraine attack development, as it might serve as a possible drug target. We measured PGE₂ plasma levels in female subjects with migraine and age-matched female healthy controls in the (pre)ictal phases of GTN provoked migraine-like attacks to assess whether PGE₂ levels change as part of GTN-induced migraine attacks.

Methods

Participants

This study was conducted as part of an extensive migraine provocation study, described in Onderwater et al.²¹ In total, 37 female subjects with migraine (without aura) and 25 age-matched female healthy controls were included. Due to the predominance of migraine in females only female subjects were included in the study. Migraine was diagnosed in accordance with the International Classification of Headache Disorders (ICHD-3).³ Participants with migraine experienced one or more migraine attacks per month during the past six months. Subjects with chronic migraine or medication-overuse headache were excluded. Healthy controls were free of (severe) headaches, neurological or psychiatric disorders and had no family history of severe primary headaches, but were allowed to occasionally have tension-type headaches. None of the participants used chronic medication other than oral contraceptives. The study was approved by the ethics committee of the Leiden University Medical Center and in accordance with the World Medical Association Declaration of Helsinki. All participants provided written informed consent prior to the study.

Study design

During the study day, each participant was subjected to detailed interviews over the course of the day and underwent three blood withdrawals. Samples were drawn by venipuncture from the medial cubital vein. Participants were attack-free at least three days prior to the investigation and had been instructed to refrain from using prophylactic medication for at least four weeks. Apart from abstaining from alcoholic beverages, caffeinated beverages, and smoking for at least 8 hours prior to and during the study day there were no dietary restrictions. Before GTN infusion, all participants underwent a baseline assessment consisting of a neurological examination, headache assessment, and a blood withdrawal in ethylenediaminetetraacetic acid (EDTA)-containing tubes was performed for baseline measurement [T0]. Following the baseline measurement, participants received an intravenous infusion of GTN (0.5 µg/kg/min over 20 minutes) between 9:45 and 10:45 AM, in supine position. After GTN infusion, blood was again drawn from participants at two time points, namely ~140 minutes [T1] and ~320 minutes after the start of GTN infusion [T2]. To avoid biochemical interference in the processes related to the initiation and onset of a migraine-like headache, participants were requested to abstain from using acute migraine attack medication until after the 3rd and final blood measurement [T2]. Blood was centrifuged at room

temperature for 20 minutes (2,000 rpm, 622 g). The supernatant was transferred to a 15-mL polypropylene tube (Greiner Bio-One CELLSTAR®), inverted several times, and divided in 0.5-mL aliquots (1.0 mL Nunc™ cryotubes). Plasma samples were stored at -80°C until further use; no extra freeze-thaw cycles were allowed.

Migraine-like headache and criteria

Participants were notified that GTN could potentially induce a headache, without any information regarding the expected onset or course. Questionnaires were performed, as described in Onderwater et al.²¹ In short, during the 20-minute GTN infusion, headache characteristics and associated symptoms were documented every 5 minutes. After the infusion period, the occurrence of premonitory symptoms, headache, and associated symptoms was documented every 15 minutes until 5 hours after GTN infusion. After the study day (6 hours after GTN infusion), to determine GTN responder status, participants filled in a headache diary and were asked for headache fitting migraine-like attack onset in a telephone follow-up ~3 days after participation. Headache intensity was scored with a verbal rating scale (VRS) from 0 to 10 (0 indicating no headache, 1 indicating a very mild headache and 10 indicating the worst possible headache pain imaginable). In addition, the response form included the type of pain, localization, associated symptoms, premonitory symptoms, and adverse events. Furthermore, subjects with migraine were asked whether the reported headache resembled their usual migraine attacks. Despite the resemblance with spontaneous attacks, induced attacks are referred to as ‘migraine-like headaches’, as they cannot fulfil all criteria of a migraine without aura attack; for this the attack needs to be spontaneous and last (untreated) at least 4 hours.³ Therefore, in accordance with earlier provocation studies,¹⁶ migraine-like attack onset (ictal) was determined as either (1) a moderate to severe headache (VRS ≥ 4) fulfilling ICHD-3 criteria C and D for migraine without aura or (2) a headache described as mimicking the subject’s usual migraine attack and treated with acute migraine medication.

PGE₂ quantification

PGE₂ was quantified in EDTA plasma using a method analogue for the quantification of 8-iso-PGF₂α, previously described.³⁵ In short, 250 mL EDTA plasma was diluted with 2.0 mL sodium acetate buffer (0.1 M, pH 3.5) and 3 mL PGE₂-d₄ (50 ng/mL) in methanol (MeOH) was added. The samples were loaded onto C18 SPE cartridges (200 mg, 3cc; Waters, Sep-Pak, Milford, MA) that had been conditioned and equilibrated with MeOH and water. After a wash with water and n-hexane samples were eluted using methyl formate. Eluates were then dried under a gentle stream of nitrogen at 40°C and reconstituted in 150 mL 40% MeOH.

Samples were measured by Liquid Chromatography (Shimadzu SIL-30AC autosampler, two Shimadzu LC-30AD pumps and a Shimadzu CTO-20AC column oven) coupled to a Sciex Qtrap 6500 mass spectrometer. Forty-mL samples were injected and separated on a C18 column

(Phenomenex, 50 × 2.1 mm, 1.7 μm). A gradient of 0.01% acetic acid in water (A) and 0.01% acetic acid in MeOH (B) was used to elute the components of interest from the column. The total flow rate was 400 mL/min. The column oven was set to 50°C. The mass spectrometer (MS) was equipped with an ESI source and operated in negative scheduled MRM mode. The needle voltage was set to -4,500 V, the drying temperature to 450°C, ion source gas 1/nebulizer gas (air) at 40 psi, ion source gas 2/drying gas (air) at 30 psi and the nebulizer gas (nitrogen) at 30 psi. For PGE₂ the transition used was 351/271, for PGE₂-d4 355/193. PGE₂ was identified based on its tandem MS transition and relative retention time and, quantified using external calibration.

Statistical analysis

We aimed to investigate the role of PGE₂ over the course of a provoked migraine attack, healthy controls were included to ensure that direct pharmacological effects of the provocation substance itself is not incorrectly labelled as a marker for provoked attacks. As we were primarily interested in the effect of different phases on PGE₂ levels in blood, we distinguished three phases: interictal (outside a migraine-like attack), preictal (before a migraine-like headache of which the onset is ≤ 12 hours after GTN infusion), and ictal (migraine-like headache). To account for repeated measurements within each subject, we used a linear mixed model with a random effect per person and unstructured correlation, the same model was used previously.³⁶ The outcome (dependent variable) was the measured PGE₂ concentration. Predictors (independent variables) were age, diagnosis (migraine or control), time point (T0, T1, T2) and migraine phase (interictal, preictal, ictal). Controls were coded as “interictal” at all time points. Furthermore, we added the interaction between time point and diagnosis to account for subjects with migraine possibly reacting differently to GTN than controls, irrespective of migraine phase. Statistical analyses were performed using SPSS (version 25.0, IBM SPSS Statistics for Windows, IBM Corp, Armonk, NY). In this study, data was collected as part of an extensive larger study and, therefore, no *a priori* power calculations were performed for this sub-study.

Results

Clinical characteristics

We initially included n = 37 participants with migraine and n = 25 healthy controls, of which five participants were excluded for further analyses. Two cases were removed as GTN infusion was not performed, both participants withdrew from participation after the baseline measurement. Two cases were excluded, because we were unable to classify the provoked headache attack (one not fully fulfilling a migraine-like headache nor classifying as a non-responder and the other developed a migraine-like attack, but already proceeded to a postdrome state during the study day). One healthy control was excluded due to a (first) provoked migraine-like headache. In total, data from n = 33 participants with migraine and n = 24 healthy controls were included in the

analyses. The demographic and clinical characteristics of cases and controls are shown in **Table 1**. There were no adverse events reported.

Table 1 Demographic and clinical characteristics of the study population

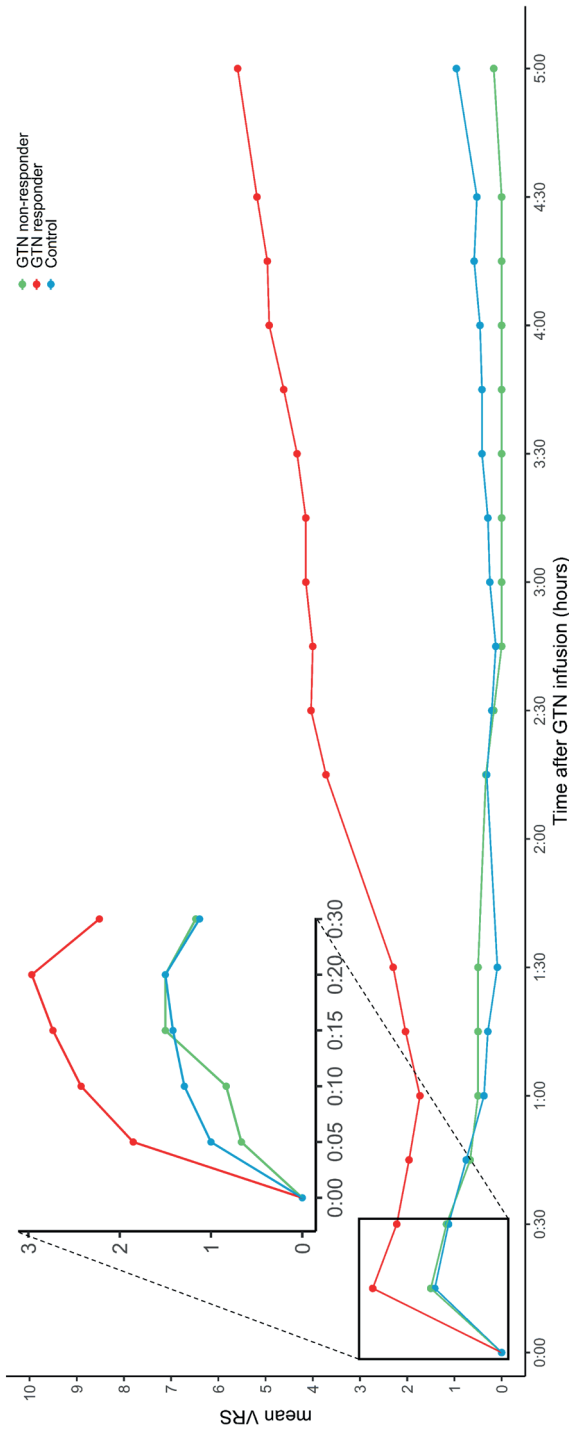
Participants Characteristics	Migraine cases (n = 33)	Healthy controls (n = 24)	P value	GTN responders (n = 27)	GTN non-responders (n = 6)
General characteristics					
Age	34.3 ± 8.2	35.2 ± 9.1	0.709 [†]	35.2 ± 8.4	30.3 ± 6.4
BMI	22.9 ± 2.6	23.2 ± 2.7	0.714 [†]	23.3 ± 2.7	21.6 ± 1.4
Smoking (n, %)	5 (15.1%)	3 (12.5%)	1.000 [‡]	5 (18.5%)	0 (0%)
Migraine characteristics					
Age of onset	16.3 ± 5.6		-	17.4 ± 4.7	11.2 ± 6.7
Migraine days (attack/month)	4.7 ± 2.7		-	5.1 ± 2.8	2.7 ± 0.8

Values are expressed as absolute values and percentage or mean ± SD., *P* values are calculated with [†] Student's *t*-test, [‡] Fisher's Exact Test. GTN, glyceryl trinitrate, BMI, body mass index.

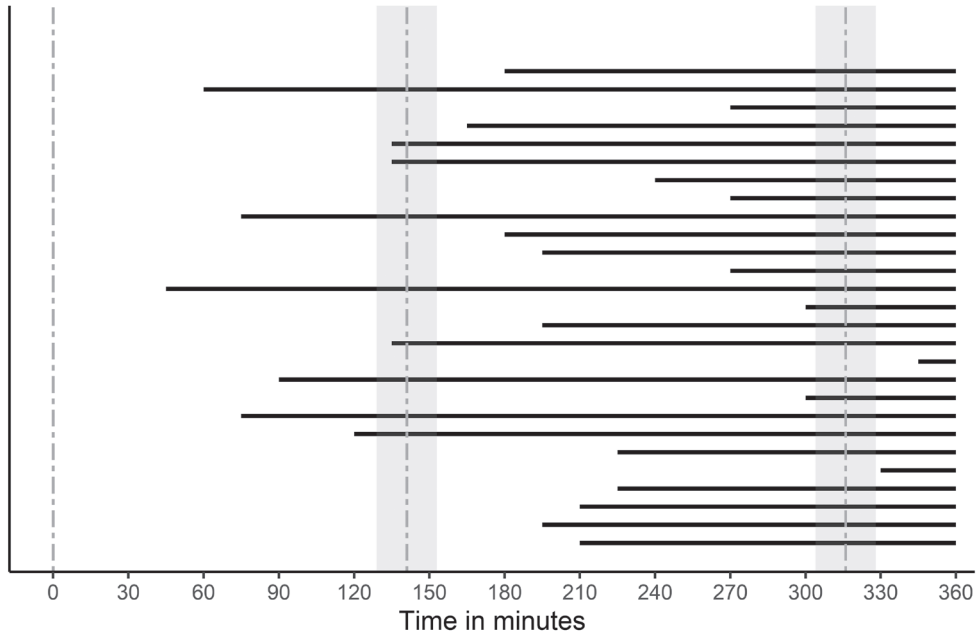
GTN response

In total, *n* = 28 subjects with migraine (85%) and *n* = 18 healthy controls (75%) developed an immediate headache (VRS ≥ 1) during the GTN infusion. At 5 minutes after the start of GTN infusion, the mean VRS value was 1.6 for those with migraine (1.8 for responders and 0.7 for non-responders) and 1 for controls. In total, *n* = 20 subjects with migraine (61%) and *n* = 10 healthy controls (42%) had an immediate headache. The mean VRS value increased until the end of the GTN infusion to 2.6 (3 for responders and 1.5 for non-responders) and 1.5, for subjects with migraine and controls, respectively. At 20 minutes, 26 subjects with migraine (79%) and 13 controls (54%) experienced a headache. Overall, the immediate headache was mild to moderate in severity and generally resolved rapidly after termination of the infusion (**Figure 3**, **Figure S1**). In some subjects with migraine a "headache-free" interval was absent (**Figure S1**), in those subjects the headache continued after infusion and eventually became more severe with characteristics of a migraine-like attack. The mean VRS for those who responded to GTN (responders) continued to increase, as the headache became more severe although only at a later stage met the criteria of migraine and in those who classified as non-responders the headache severity decreased. Generally, the immediate phase is considered to be 0-90 minutes post infusion. Four subjects developed migraine within this timeframe. One subject with migraine developed a headache fulfilling the migraine-like criteria within one hour after the start of GTN infusion, one at 60 minutes, and two at 75 minutes. Eventually, 27 (82%) subjects with migraine receiving GTN experienced a migraine-like attack (**Figure 4**) during the study day and 6 (18%) did not experience such an attack, hence they were labelled as GTN responders and GTN non-responders, respectively (**Table 1**). Migraine-like attack onset ranged between 45 and 345 minutes (mean 192 ± 84 minutes) (**Figure 4**).

Figure 3 Mean verbal rating scale (VRS) for headache severity per responder group



The X-axis represents time after GTN infusion and the Y-axis the mean VRS. In red, participants with migraine who responded to GTN (GTN responders), participants with migraine who did not respond to GTN (GTN non-responders) in green and healthy controls in blue. GTN, glyceryl trinitrate; VRS, verbal rating scale. All subjects (including those without headache) were used in the calculation of the average. For some time points there were many missing values, this resulted in exclusion of these time points from the figure. Whiskers represent the standard deviation from the mean.

Figure 4 Timing migraine onset in GTN responders

The onset of migraine is plotted for each glyceryl trinitrate (GTN) responder with respect to time after GTN infusion. The start of the black continuous line represents the timing of onset of migraine attack per individual. The dotted line represents the blood draw timepoints T0, T1 and T2 at 0, ~140 and ~320 minutes, respectively, after the start of the GTN infusion.

Table 2 Median PGE₂ concentrations over time independent of migraine phase

Group	[T0]	[T1]	[T2]
GTN responders	0.044 (0.02-0.10)	0.053 (0.03-0.10)	0.049 (0.03-0.08)
GTN non-responders	0.052 (0.01-0.09)	0.031 (0.01-0.07)	0.040 (0.02-0.07)
Controls	0.044 (0.02-0.08)	0.043 (0.03-0.09)	0.060 (0.03-0.09)

Values are the uncorrected medians of absolute concentrations in ng/mL with their interquartile range.

[T0] = baseline, [T1] = ~140 minutes after the start of GTN infusion, [T2] = ~320 minutes after GTN infusion. GTN responder, migraine patients who responded to GTN; GTN non-responder, migraine patients who did not respond to GTN. GTN, glyceryl trinitrate.

PGE₂ in relation to migraine-like attack onset

The level of PGE₂ per individual varied per time point (**Table 2**). To determine whether PGE₂ levels were linked to the various phases (baseline, preictal and ictal) of a migraine attack, a generalized linear mixed model was used. The transition from an interictal state towards a migraine-like attack had no influence on PGE₂ concentration (F (2, 69.70) = 1.235, $P = 0.297$). Both the transition from “interictal to preictal” ($P = 0.527$) and “interictal to ictal” ($P = 0.141$) phase of GTN-induced migraine-like attacks had no influence on PGE₂ concentration (Table S1).

Discussion

We performed a GTN provocation study in subjects with migraine and healthy controls and found that 82% of migraine participants developed a delayed onset migraine-like attack. We prospectively assessed PGE₂ levels at three time points selected over the course of provoked migraine-like attacks and compared these to those without provoked attacks and controls. We found no evidence that GTN-induced migraine-like headaches are characterized by changes in plasma PGE₂ levels towards the (pre)ictal state. This suggests that a rise in PGE₂ is not an essential step in the initiation of GTN-induced migraine-like attacks.

PGE₂ is able to induce rapid-onset migraine-like attacks in subjects with migraine within 90 minutes,¹⁴ in contrast to provocation with substances such as PACAP, CGRP and GTN that result in a delayed (after a few hours) onset of a migraine-like attack.^{14,16} Thus, we hypothesized that PGE₂ could be one of the molecules involved in a(n experimentally induced) migraine attack. Given that administration of PGE₂ can cause a rapid-onset migraine-like attack, in contrast to the other provocative substances, PGE₂ may perhaps serve as a marker for upcoming migraine attacks, albeit that the timing of blood sampling is important. In our study, we used the GTN provocation model to assess the role of PGE₂. It has been hypothesized that the time it takes to develop delayed migraine-like attack is due to various processes that include the regulation of gene expression and proteins ultimately resulting in migraine-like attacks in subjects with migraine with a median attack onset of 3 to 6 hours, after infusion of the provocation substance. After all, in animal models of migraine, GTN activates the COX-2-PGE₂ pathway in the brainstem not before 4 hours after GTN administration.³⁷ However, based on our proposed mechanism and the PGE₂ human provocation studies with rapid onset of provoked migraine-like headaches, we expected a rise in PGE₂ to be close to the start of a migraine attack as an early marker of migraine, which would fit our time points of blood withdrawal. The alternative explanation that we did not find a rise in PGE₂ levels might indicate that the pathway activated by GTN towards a migraine-like attack does not primarily act via PGE₂. One can envisage that pathways, independent of PGE₂ via for instance cGMP or cAMP, are more strongly activated than the PGE₂-pathway when GTN is administered. Another explanation might be that a rise in PGE₂ is very locally and hence not measurable in blood.

To our knowledge no other study measured PGE₂ levels over the course of GTN-induced migraine-like attack in subjects with migraine. Still, few studies reporting measurements of PGE₂ levels during spontaneous migraine attacks suggested those to be elevated in blood,^{38,39} and saliva.⁴⁰ More specifically, in contrast to our study, a much smaller study of only five subjects with migraine reported an increase in PGE₂ levels in jugular venous blood peaking between 2 and 6 hours after the start of a spontaneous migraine attack and normalizing towards the end of the attack.³⁸ In our study the mean attack onset was ~192 minutes, hence many cases were over 2 hours into their delayed migraine-like attack at the ~320-minute time point, which suggests that our timing was

not different from the spontaneous migraine attack study and thus could have picked up a similar rise in PGE₂ levels. In addition, two studies found that PGE₂ levels in plasma,³⁹ (18 cases and 12 controls) and saliva⁴⁰ (6 cases and 9 controls) in subjects with migraine were lower compared to controls outside attacks and increased during a spontaneous attack surpassing the levels found in controls. Although this was not our primary question, we tested this and did not find a difference in baseline PGE₂ levels between cases and controls. Giving the small number of participants in previous studies, our larger study should have been able to reveal differences in PGE₂ levels during GTN-induced migraine-like attacks. Another reason for the discrepancy with earlier studies might be in the measuring techniques used and/or the matching and correction of data. For our study we used a highly reliable, standardized technique for measuring PGE₂ levels and additionally have minimized external effects on PGE₂ levels, by careful matching and correcting for multiple factors to single out the effect of PGE₂ on a migraine attack. Whereas such external effects do not seem to have affected our results, they might have played a role in earlier studies. Another possibility is that spontaneous attacks are not always the same as provoked attacks (e.g. GTN provocation in migraine patients with aura leads to a migraine-like attack, but not an aura). This may indicate that in spontaneous attacks different pathways may be initiated depending on headache (sub)type, none the less these pathways ultimately lead to the same migraine headache.

We envisage several possible explanations why we found no evidence for a change in PGE₂ levels over the course of a GTN-induced attack. PGE₂ acts via four distinct G protein-coupled receptors EP1, EP2, EP3 and EP4. Ligand binding to the different EP receptors leads to the activation of distinct downstream signaling pathways, resulting in distinct biological outcomes,^{31,41} one of these second messengers being cyclic adenosine monophosphate (cAMP).³¹ Via its receptors, PGE₂ is known to play a role in nociceptive pain processing and inflammation,^{42,43} exerting both damaging pro-inflammatory and protective anti-inflammatory effects in the brain.⁴⁴⁻⁴⁶ Thus, the PGE₂ response is dependent on the array of receptors cells express as well as on intracellular pathways to which they are coupled^{46,47}. Hence, any involvement of PGE₂ in the pathogenesis of migraine may be very complex.

As mentioned previously, the immediate headache is thought to be the result of vasodilation via the NO-cGMP pathway,⁷ independent of CGRP release⁸, whereas the delayed migraine-like attack is thought to be the result of trigeminovascular activation mediated via CGRP.^{5,7,9} However, there likely is extensive cross talk between both pathways (for details see **Figure 2**). For instance, on a cellular level multiple components in the migraine pathway are known to be vasodilators, but can also lead to migraine attacks. As exemplified by ATP-sensitive potassium (K_{ATP}) channel openers (levcromakalim) and (big)-conductance calcium-activated K⁺ (BK_{Ca}) channel opener (MaxiPost), both activated via the NO-cGMP pathway, which is known to play a role in the immediate headache, but activation of these channels can also induce migraine-like attacks.^{33,34} However, the rather long delay of several hours between infusion of levcromakalim/MaxiProst and the occurrence of a migraine-like attack (with a median time of 3 hours) indicates

that various mediators must be involved in slower cascades of events leading to a migraine-like attack. Evidence for cross talk is that both the administration of CGRP (pathway via cAMP) and sildenafil (pathway via cGMP) can lead to a migraine-like attack, suggesting that convergence to a common cellular determinant seem to exist ultimately triggering similar attacks.⁴⁸ Given that the median time until an attack for CGRP is ~165 minutes, so much shorter than the ~285 minutes for sildenafil, one can envisage that CGRP acts more downstream in the generation of a migraine attack. Such sequential actions, given the cross talk between CGRP and PGE₂, especially in GTN-induced attacks, seems in line with a chain-of-events-pathway (**Figure 2**).

Our study has several limitations. We collected a large number of blood samples of migraine participants and healthy controls. Each participant was sampled at three fixed times during the study day in an attempt to measure PGE₂ concentrations during attack development and during the attack itself. We find that the 95% confidence interval of the change in PGE₂ levels from interictal to another phase extends from -0.02 to 0.05 ng/mL. Of course, the onset of the attack varies between subjects and did not align perfectly with the measurement times. Moreover, we must account for a possible temporal effect of the GTN infusion on PGE₂ concentrations. Combined with within and between subject measurement variation, we must acknowledge that not finding a statistically significant difference in PGE₂ levels over the course of an induced migraine-like attack does not prove the absence of such an effect. There may yet exist subtle, short-duration, variations in PGE₂ levels that we could not detect. Furthermore, we have used LC-MS/MS which is distinct from the more often used ELISA kits to measure PGE₂, this might make it difficult to compare absolute concentrations between studies. However, by using this method we were able to detect very low levels of PGE₂ with good accuracy, despite the short half-life of PGE₂. Furthermore, whereas we did not observe changes in PGE₂ levels in blood it is conceivable that levels may be different in cerebrospinal fluid, as increased PGE₂ levels have been reported indicative for probable Alzheimer's disease.⁴⁹ However, we deem it too unethical and unlikely that subjects with migraine (and controls) are willing to participate in a provocation study with, logically, repeated lumbar punctures to get information on PGE₂ levels over time. Finally, our study only consists of females to prevent any sex effects, which may limit the generalizability of our findings to male migraine patients. Additionally, although we have performed our study in a female only population to account for the most notable sex hormone differences, small differences in cycle and use of contraceptives might be of influence in the downstream provocation pathways.

Supplementary information

Table S1 <https://ars.els-cdn.com/content/image/1-s2.0-S2452073X22000290-mmc2.docx>

Figure S1 <https://ars.els-cdn.com/content/image/1-s2.0-S2452073X22000290-mmc1.pdf>



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Author's contributions: Aster V.E. Harder: Conceptualization, Data curation, Formal analysis, Investigation, Writing original draft, Methodology, Project Administration. Gerrit L.J. Onderwater: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Project Administration, Writing – Review & Editing. Robin M. van Dongen: Conceptualization, Investigation, Writing – Review & Editing. Marieke Heijink: Methodology, Validation, Writing – Review & Editing. Erik W. van Zwet: Formal analysis, Methodology, Writing – Review & Editing. Martin Giera: Conceptualization, Resources, Validation, Writing – Review & Editing. Arn M.J.M. van den Maagdenberg: Conceptualization, Funding Acquisition, Resources, Supervision, Writing – Review & Editing. Gisela M. Terwindt: Conceptualization, Funding Acquisition, Resources, Supervision, Writing – Review & Editing.

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References

1. GBD 2019 Diseases and Injuries Collaborators. Global burden of 369 diseases and injuries in 204 countries and territories, 1990-2019: a systematic analysis for the Global Burden of Disease Study 2019. *Lancet*. 2020;396(10258):1204-1222.
2. Launer LJ, Terwindt GM, Ferrari MD. The prevalence and characteristics of migraine in a population-based cohort: the GEM study. *Neurology*. 1999;53(3):537-542.
3. Headache Classification Committee of the International Headache Society (IHS). The International Classification of Headache Disorders, 3rd edition. *Cephalalgia*. 2018;38(1):1-211.
4. Olesen J. The role of nitric oxide (NO) in migraine, tension-type headache and cluster headache. *Pharmacol Ther*. 2008;120(2):157-171.
5. Bagdy G, Riba P, Kecskemeti V, Chase D, Juhasz G. Headache-type adverse effects of NO donors: vasodilation and beyond. *Br J Pharmacol*. 2010;160(1):20-35.
6. Iversen HK, Olesen J, Tfelt-Hansen P. Intravenous nitroglycerin as an experimental model of vascular headache. Basic characteristics. *Pain*. 1989;38(1):17-24.
7. Akerman S, Williamson DJ, Kaube H, Goadsby PJ. Nitric oxide synthase inhibitors can antagonize neurogenic and calcitonin gene-related peptide induced dilation of dural meningeal vessels. *Br J Pharmacol*. 2002;137(1):62-68.
8. Bellamy J, Bowen EJ, Russo AF, Durham PL. Nitric oxide regulation of calcitonin gene-related peptide gene expression in rat trigeminal ganglia neurons. *Eur J Neurosci*. 2006;23(8):2057-2066.
9. Juhasz G, Zsombok T, Modos EA, et al. NO-induced migraine attack: strong increase in plasma calcitonin gene-related peptide (CGRP) concentration and negative correlation with platelet serotonin release. *Pain*. 2003;106(3):461-470.
10. Davis RJ, Murdoch CE, Ali M, et al. EP4 prostanoid receptor-mediated vasodilatation of human middle cerebral arteries. *Br J Pharmacol*. 2004;141(4):580-585.
11. Mollace V, Muscoli C, Masini E, Cuzzocrea S, Salvemini D. Modulation of prostaglandin biosynthesis by nitric oxide and nitric oxide donors. *Pharmacol Rev*. 2005;57(2):217-252.
12. Eikermann-Haerter K, Ayata C. Cortical spreading depression and migraine. *Curr Neurol Neurosci Rep*. 2010;10(3):167-173.
13. Yokota C, Inoue H, Kuge Y, et al. Cyclooxygenase-2 expression associated with spreading depression in a primate model. *J Cereb Blood Flow Metab*. 2003;23(4):395-398.
14. Antonova M, Wienecke T, Olesen J, Ashina M. Prostaglandin E(2) induces immediate migraine-like attack in migraine patients without aura. *Cephalalgia*. 2012;32(11):822-833.
15. Wienecke T, Olesen J, Ashina M. Prostaglandin I2 (epoprostenol) triggers migraine-like attacks in migraineurs. *Cephalalgia*. 2010;30(2):179-190.
16. Ashina M, Hansen JM, Olesen J. Pearls and pitfalls in human pharmacological models of migraine: 30 years' experience. *Cephalalgia*. 2013;33(8):540-553.
17. Jenkins DW, Feniuk W, Humphrey PP. Characterization of the prostanoid receptor types involved in mediating calcitonin gene-related peptide release from cultured rat trigeminal neurones. *Br J Pharmacol*. 2001;134(6):1296-1302.
18. Jenkins DW, Langmead CJ, Parsons AA, Strijbos PJ. Regulation of calcitonin gene-related peptide release from rat trigeminal nucleus caudalis slices in vitro. *Neurosci Lett*. 2004;366(3):241-424.
19. Neeb L, Hellen P, Boehnke C, et al. IL-1beta stimulates COX-2 dependent PGE(2) synthesis and CGRP release in rat trigeminal ganglia cells. *PLoS One*. 2011;6(3):e17360.

20. Kress M, Guthmann C, Averbeck B, Reeh PW. Calcitonin gene-related peptide and prostaglandin E2 but not substance P release induced by antidromic nerve stimulation from rat skin in vitro. *Neuroscience*. 1999;89(1):303-310.
21. Onderwater GLJ, Dool J, Ferrari MD, Terwindt GM. Premonitory symptoms in glyceryl trinitrate triggered migraine attacks: a case-control study. *Pain*. 2020;161(9):2058-2067.
22. Arnold WP, Mittal CK, Katsuki S, Murad F. Nitric oxide activates guanylate cyclase and increases guanosine 3':5'-cyclic monophosphate levels in various tissue preparations. *Proc Natl Acad Sci U S A*. 1977;74(8):3203-3207.
23. Taffi R, Vignini A, Lanciotti C, et al. Platelet membrane fluidity and peroxynitrite levels in migraine patients during headache-free periods. *Cephalalgia*. 2005;25(5):353-358.
24. Bredt DS. Endogenous nitric oxide synthesis: biological functions and pathophysiology. *Free Radic Res*. 1999;31(6):577-596.
25. Ndengele MM, Cuzzocrea S, Esposito E, et al. Cyclooxygenases 1 and 2 contribute to peroxynitrite-mediated inflammatory pain hypersensitivity. *EASEB J*. 2008;22(9):3154-3164.
26. Akerman S, Salvemini D, Romero-Reyes M. Targeting reactive nitroxidative species in preclinical models of migraine. *Cephalalgia*. 2021;41(11-12):1187-1200.
27. Schubert R, Nelson MT. Protein kinases: tuners of the BKCa channel in smooth muscle. *Trends Pharmacol Sci*. 2001;22(10):505-512.
28. Murphy ME, Brayden JE. Nitric oxide hyperpolarizes rabbit mesenteric arteries via ATP-sensitive potassium channels. *J Physiol*. 1995;486 (Pt 1)(Pt 1):47-58.
29. Miyoshi H, Nakaya Y. Calcitonin gene-related peptide activates the K⁺ channels of vascular smooth muscle cells via adenylate cyclase. *Basic Res Cardiol*. 1995;90(4):332-336.
30. Hosokawa S, Endoh T, Shibukawa Y, et al. Calcitonin gene-related peptide- and adrenomedullin-induced facilitation of calcium current by different signal pathways in nucleus tractus solitarius. *Brain Res*. 2010;1327:47-55.
31. Markovič T, Jakopin Ž, Dolenc MS, Mlinarič-Raščan I. Structural features of subtype-selective EP receptor modulators. *Drug discovery today*. 2017;22(1):57-71.
32. Chrissobolis S, Sobey CG. Inwardly rectifying potassium channels in the regulation of vascular tone. *Curr Drug Targets*. 2003;4(4):281-289.
33. Al-Karagholi MA, Hansen JM, Guo S, Olesen J, Ashina M. Opening of ATP-sensitive potassium channels causes migraine attacks: a new target for the treatment of migraine. *Brain*. 2019;142(9):2644-2654.
34. Al-Karagholi MA, Ghanizada H, Waldorff Nielsen CA, et al. Opening of BKCa channels causes migraine attacks: a new downstream target for the treatment of migraine. *Pain*. 2021;162(10):2512-2520.
35. Dekker J, Martherus T, Lopriore E, et al. The Effect of Initial High vs. Low FiO₂ on Breathing Effort in Preterm Infants at Birth: A Randomized Controlled Trial. *Clinical Trial. Front Pediatr*. 2019;7504.:
36. Onderwater GLJ, Wijnen JP, Najac C, et al. Cortical glutamate and gamma-aminobutyric acid over the course of a provoked migraine attack, a 7 Tesla magnetic resonance spectroscopy study. *Neuroimage Clin*. 2021;32:102889.
37. Tassorelli C, Greco R, Armentero MT, et al. A role for brain cyclooxygenase-2 and prostaglandin-E2 in migraine: effects of nitroglycerin. *Int Rev Neurobiol*. 2007;82:373-382.
38. Sarchielli P, Alberti A, Codini M, Floridi A, Gallai V. Nitric oxide metabolites, prostaglandins and trigeminal vasoactive peptides in internal jugular vein blood during spontaneous migraine attacks. *Cephalalgia*. 2000;20(10):907-918.
39. Nattero G, Allais G, De Lorenzo C, et al. Relevance of prostaglandins in true menstrual migraine. *Headache*. 1989;29(4):233-238.
40. Vardi J, Flechter S, Alguati A, Regev I, Ayalon D. Prostaglandin--E2 levels in the saliva of common migrainous women. *Headache*. 1983;23(2):59-61.

41. Negishi M, Sugimoto Y, Ichikawa A. Molecular mechanisms of diverse actions of prostanoid receptors. *Biochim Biophys Acta*. 1995;1259(1):109-119.
42. Kawabata A. Prostaglandin E2 and pain--an update. *Biol Pharm Bull*. 2011;34(8):1170-3117.
43. Uda R, Horiguchi S, Ito S, Hyodo M, Hayaishi O. Nociceptive effects induced by intrathecal administration of prostaglandin D2, E2, or F2 alpha to conscious mice. *Brain Res*. 1990;510(1):26-32.
44. Pradhan SS, Salinas K, Garduno AC, et al. Anti-Inflammatory and Neuroprotective Effects of PGE(2) EP4 Signaling in Models of Parkinson's Disease. *J Neuroimmune Pharmacol*. 2017;12(2):292-304.
45. Liang X, Wang Q, Shi J, et al. The prostaglandin E2 EP2 receptor accelerates disease progression and inflammation in a model of amyotrophic lateral sclerosis. *Ann Neurol*. 2008;64(3):304-314.
46. Andreasson K. Emerging roles of PGE2 receptors in models of neurological disease. *Prostaglandins Other Lipid Mediat*. 2010;91(3-4):104-112.
47. Tilley SL, Coffman TM, Koller BH. Mixed messages: modulation of inflammation and immune responses by prostaglandins and thromboxanes. *J Clin Invest*. 2001;108(1):15-23.
48. Younis S, Christensen CE, Toft NM, et al. Investigation of distinct molecular pathways in migraine induction using calcitonin gene-related peptide and sildenafil. *Cephalalgia*. 2019;39(14):1776-1788.
49. Montine TJ, Sidell KR, Crews BC, et al. Elevated CSF prostaglandin E2 levels in patients with probable AD. *Neurology*. 1999;53(7):1495-1498.