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Lymphovascular space invasion in endometrial carcinoma

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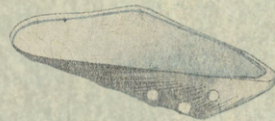
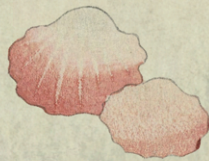
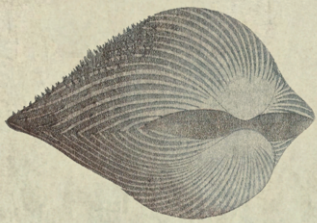
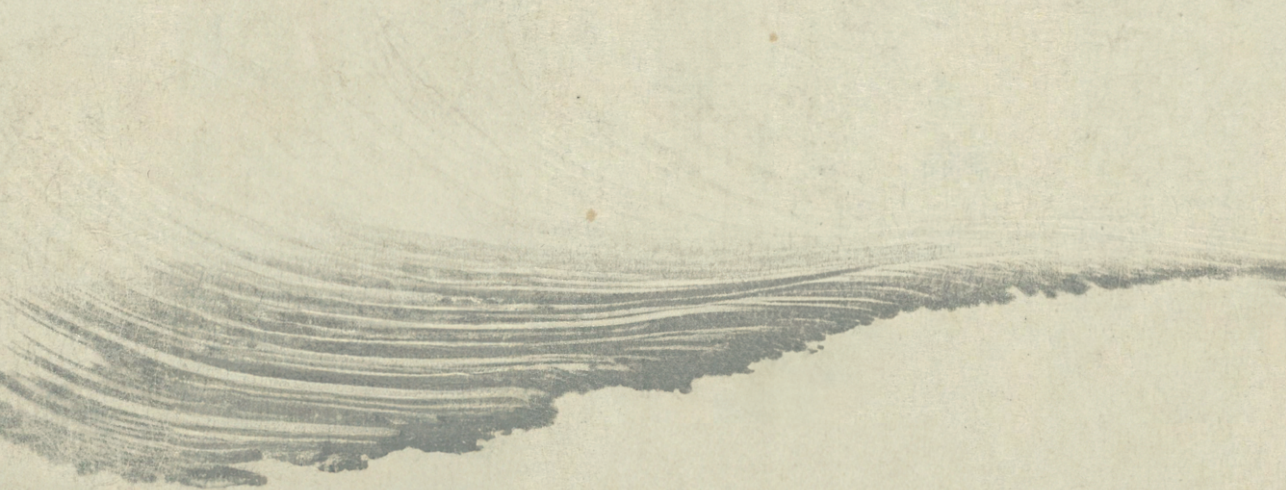
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CHAPTER 6

REPRODUCIBILITY OF LYMPHOVASCULAR SPACE INVASION (LVSI) ASSESSMENT IN ENDOMETRIAL CANCER

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Abstract

Aims: Lymphovascular space invasion (LVSI) in endometrial cancer (EC) is an important prognostic variable impacting on a patient's individual recurrence risk and adjuvant treatment recommendations. Recent work has shown that grading the extent of LVSI further improves its prognostic strength in patients with stage I endometrioid EC. Despite this, there is little information on the reproducibility of LVSI assessment in EC. Therefore, we designed a study to evaluate interobserver agreement in discriminating true LVSI from LVSI-mimics (phase 1) and reproducibility of grading extent of LVSI (phase 2).

Materials and results: Scanned haematoxylin and eosin (H&E) slides of endometrioid EC (EEC) with a predefined possible LVSI-focus were hosted on a website and assessed by a panel of six European gynaecological pathologists. In phase 1, 48 H&E slides were included for LVSI assessment and in phase 2, 42 H&E slides for LVSI grading. Each observer was instructed to apply the criteria for LVSI used in daily practice. The degree of agreement was measured using the two-way absolute agreement average-measures intra-class correlation coefficient (ICC). Reproducibility of LVSI assessment (ICC: 0.64 ($p < 0.001$)) and LVSI grading (ICC of 0.62 ($p < 0.001$)) in EEC was substantial among the observers.

Conclusions: Given the good reproducibility of LVSI, this study further supports the important role of LVSI in decision algorithms for adjuvant treatment.

Introduction

Classic histopathological parameters are the cornerstone of the current risk-assessment and guide adjuvant treatment for patients with early stage (stage I/II) endometrial carcinoma (EC). Tumour factors included in the risk assessment of early-stage disease are histological type, tumour grade, cervical stromal involvement, depth of myometrial invasion and lymphovascular space invasion (LVSI). Combinations of these factors stratify early-stage EC patient into low-risk (LR), high-intermediate risk (HIR) and high-risk (HR) for recurrence with differential adjuvant treatment choices [1-3].

Currently, significant advances in our understanding of molecular alterations in EC are reshaping the risk-assessment by incorporating molecular features. Novel models in which molecular factors are integrated to further refine the risk assessment are being developed [4, 5]. These integrated approaches still rely on the most relevant histological variables mentioned above. The Achilles heel of those histological variables, however, is the reproducibility among pathologists. One of the strongest prognostic variables in this context is the presence (or absence) of LVSI.

LVSI has gained a prominent position in most of the risk stratification systems for EC [5-7]. Adjuvant radiation treatment for patients with grade 1 or 2 stage I EEC is recommended in the presence of LVSI, independent of the depth of myometrial invasion [7]. It is interesting that the adjective “*unequivocal*” is used for LVSI in the most recent ESMO-ESTRO-ESGO clinical guidelines[7], as it advises to report LVSI only when there is no other interpretation possible. This immediately evokes the question “how reproducible among pathologists is unequivocal LVSI”. In addition, recent work shows that substantial LVSI in EC may have a stronger prognostic significance than focal LVSI [52, 78]; similar effects are reported for LVSI grading in breast cancer [10].

A diversity of LVSI definitions can be found in the EC literature, reflecting different ways to approach its assessment. Irrespective of the exact formulation, all these refinements are aimed to help distinguish LVSI from LVSI-mimics. The most frequently encountered LVSI-mimic is artefactual displacement of tumour within myometrial clefts or large endothelial lined vessels. These displacements are likely the result of manipulation of the uterus by an intrauterine balloon during surgery [11] or an artefact induced by inappropriate grossing of a friable tumour [12]. Artefactual displacement is more likely to occur in cases with poor fixation or in EC with abundant necrosis. Another frequent artefact that mimics LVSI, is stromal retraction around invading tumour glands. Furthermore, “emboli” in vascular spaces are not always clearly composed of viable tumour cells. There may be degenerative changes and infiltration of inflammatory cells may obscure the presence of tumour cells in these emboli. A specific type of myometrial invasion referred to as microcystic elongated and fragmented (MELF)-type invasion [13], may also be confused with LVSI, but importantly is also associated with true LVSI.

Additional histological criteria, such as proximity to a venous and arterial vessel [10] or perivascular lymphocytes, have been proposed to favour true LVSI [14].

The reported prevalence of LVSI in stage I EC varies widely (3.2-35%), indicating there may be local differences in how LVSI assessment is conducted and reported [15, 16]; however, interobserver variability studies focusing on LVSI in EC are sparse. Given the significance of LVSI evaluation in risk allocation of EC, and the widely accepted difficulties in LVSI assessment, this study was initiated to examine interobserver agreement on the presence of LVSI and LVSI grading. To our knowledge, this is the first study to assess the reproducibility of the recently proposed grading system for LVSI.

Materials and methods

In a previous study [8] haematoxylin and eosin (H&E) slides of EEC from 926 patients derived from the PORTEC 1 and 2 trials [2, 17] were locally re-reviewed for the presence of LVSI by the study-pathologists (EEMP, TB and VTHBMS). At review, the presence of LVSI-mimics was also noted.

In phase 1, to determine agreement of LVSI assessment, 48 cases were selected by the study pathologists, composed of challenging LVSI-mimics (n = 29) and cases with convincing true LVSI (n = 19). The LVSI-mimics were composed of MELF (n = 8); retraction artefact (no endothelial lining) (n = 10); artefactual tumour displacement (n = 5) and LVSI-mimics of emboli without tumour cells (n = 6). H&E slides were scanned and hosted on a website designed for this purpose. To ensure all observers evaluated the same focus, they were guided to the predefined, digitally annotated putative LVSI focus. It remained possible for the observers to view the whole section and not just the preselected focus, by scrolling through the complete scanned slide. In this phase observers were asked to indicate if the selected focus was true LVSI, using the LVSI definition they used in daily practice. When observers did not consider the marked focus as true LVSI, they were asked to specify what type of LVSI-mimic was present (supplementary table 1A). In this phase we also asked the observers to explain their choice. We also asked the observers for the definition of LVSI, they used in everyday practice.

In phase 2, we set out to determine agreement of LVSI grading. For this, a new selection of 42 cases was put together by the study pathologists. All 42 cases were considered positive for true-LVSI on re-review and were graded as either focal (n = 20) or substantial LVSI (n = 22). Cases were presented to the same group of observers on the same website, asking them first to confirm LVSI and next to grade LVSI positive cases as either focal LVSI or substantial LVSI. Focal LVSI was defined semi quantitatively as “the presence of a single focus of LVSI around the tumour”. Substantial LVSI was defined as “diffuse or multifocal LVSI around the tumour” (supplementary table 1B) [18]. Free text comments were optional.

Six experienced gynaecologic pathologists (observers) were recruited via the ENITEC network. We aimed to include pathologists of different nationalities and from different European institutes, in order to assure differing training backgrounds.

Statistics

Raw data was stored on the website, downloaded and processed prior to analysis. Agreement among observers was measured using the two-way absolute agreement average-measures intra-class correlation coefficient (ICC). Due to the lack of a gold standard for true-LVSI, this method results in a measure of intra-observer and inter-observer variability[19]. SPSS 23.0 package was used for statistical analyses. An ICC value reflects slight (0 – 0.19); fair (0.2 – 0.39); moderate (0.4 – 0.59; substantial (0.6 – 0.79) or almost perfect (>0.8) agreement. Additionally, agreement was qualitatively expressed as: “full agreement” when all observers agreed; “partial agreement” when 4 or 5 observers agreed and “no agreement” when 3 or less observers agreed [20].

Results

Table 1 lists the LVSI definitions provided by the gynaecologic pathologists (observers). These definitions all capture the key element of the consensus definition of LVSI, namely the presence of tumour cells in a vessel lined by endothelial cells. Some observers also include exclusion criteria or components such as: adherence to the vessel wall, and the presence of erythrocytes.

Table 1. Definitions of LVSI as used by the observers.

Observer	What definition of LVSI do you use in daily practice?
A	Cohesive aggregates of tumour cells located inside a vascular space (defined by the presence of an endothelial lining) and preferentially juxtaposed to the vessel wall.
B	Carcinoma cells adherent to vessel wall (with endothelial cells).
C	Definite tumour cells within an endothelial lined channel and no features to suggest artefactual vascular invasion.
D	Presence of tumour cells in lymphatics or vessels, which is not caused by artefacts (such as smears, retraction).
E	Tumour cells usually as a group or nest within a space that is covered by endothelial cells and does not contain a significant number of erythrocytes.
F	The presence of a tumour embolus within a vessel (capillary or lymphatic), usually well defined, rounding up to the contour of the vessel, may or may not be attached to the inner surface, may include red cells or fibrin; absence of marked autolysis.

Phase 1: Reproducibility of LVSI assessment

Full agreement about the presence or absence of LVSI was found in 10 out of 48 cases (21%); partial agreement in 23 cases (48%) and no agreement in 15 cases (31%) (table 2). Individual scores are presented in supplementary table 2. One observer was a noted outlier and appeared to have a low threshold for diagnosing true-LVSI. Overall, these outcomes resulted in substantial agreement (ICC of 0,64 (p < 0,001)) in LVSI assessment.

Table 2. Qualitative level of agreement in LVSI assessment (phase 1) and LVSI grading (phase 2), according to initial central review.

Level of agreement	Phase 1		Phase 2	
	Initial review LVSI positive (n = 19)	Initial review LVSI negative (n = 29)	Initial review focal LVSI (n = 20)	Initial review substantial LVSI (n = 22)
Full	5	5	3	3
Partial	10	13	11	13
No	4	11	6	6

Some representative examples of LVSI mimics from the study are illustrated in figure 1. Interestingly, there was little agreement upon the various reasons to score the focus as negative for LVSI. There were 26 cases in which at least two observers stated there was no LVSI. In just eight of these cases (31%) the same explanation was given. In the remaining 18 cases (69%) at least two different reasons for “no LVSI” were given. This is illustrated in figure 2, a case in which mimics co-exist resulting in more than one reason to reject true LVSI.

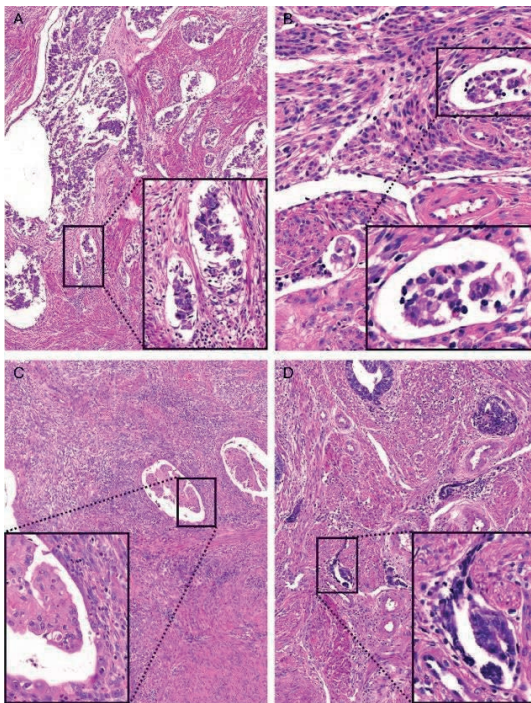


Figure 1. Representative examples of LVSI mimics presented in phase I. **A:** Retraction artefact around poorly preserved invading tumour. **B:** A cluster of inflammatory cells within a vessel, mimicking tumour cells. **C:** A microcyst aligned by flattened epithelial cells with a cluster of tumour cells in the centre, mimicking true LVSI. **D:** A cluster of tumour cells trapped within a myometrial cleft without an endothelial lining. Note the lack of perivascular infiltrate in all LVSI-mimics.

Phase 2: Reproducibility of LVSI grading

Full agreement was achieved in six cases (14%); partial agreement in 23 cases (55%) and no agreement in 13 cases (31%). Figure 3 is an example of a case with full agreement on focal LVSI. Figure 4 illustrates a case with partial agreement on substantial LVSI. The overall reproducibility in this phase was moderate (ICC 0,54 ($p < 0.001$)). However, one pathologist consistently scored

cases as negative for LVSI, whereas two pathologists had a noted tendency to diagnose substantial LVSI. Individual scores are presented in supplementary table 3. LVSI grading in cases recognized by the observers as true-LVSI resulted in substantial agreement (ICC 0.62 ($p < 0.001$)) using the predefined semi quantitative definitions for grading LVSI.

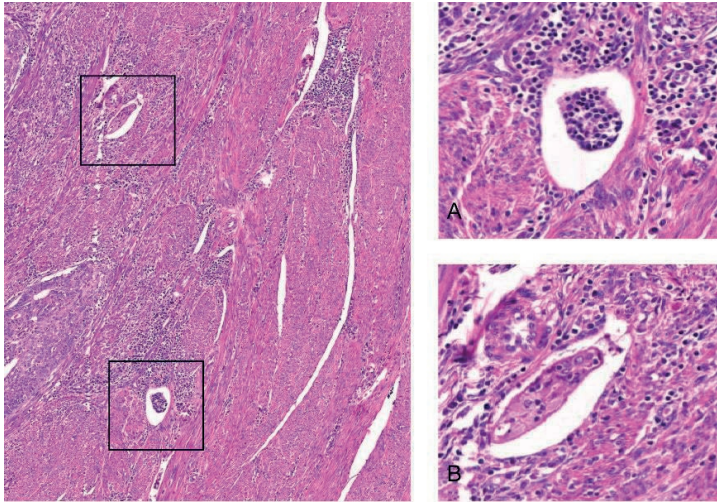


Figure 2. A representative example of a case with no consensus on LVSI assessment. This case shows two suspected foci of LVSI close to each other. The lower focus (detail A) shows the presence of endothelial cells indicating that this is a vessel, however the cell cluster within this vessel does not unequivocally contain tumour cells. The upper focus (detail B) shows a vessel with a cluster of epithelioid cells infiltrated by a few lymphocytes. Three observers scored this case as LVSI positive, two scored negative arguing the lack of tumour cells, one scores negative because of lack of endothelial cells. In this case subsequent IHC would likely result in higher level of agreement.

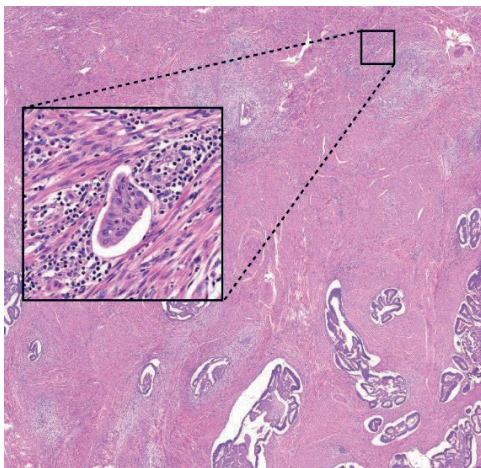


Figure 3. A case derived from phase II with full agreement on focal LVSI. The overview shows infiltrating tumour glands surrounded by an extensive stromal reaction. Some glands are surrounded by retraction artefacts. There is a focus top right (detail shown left) suspected for LVSI. The focus contains a perivascular lymphocytic infiltrate and is adjacent to a venule. This was the only LVSI focus on this H&E. All observers graded this as focal LVSI.

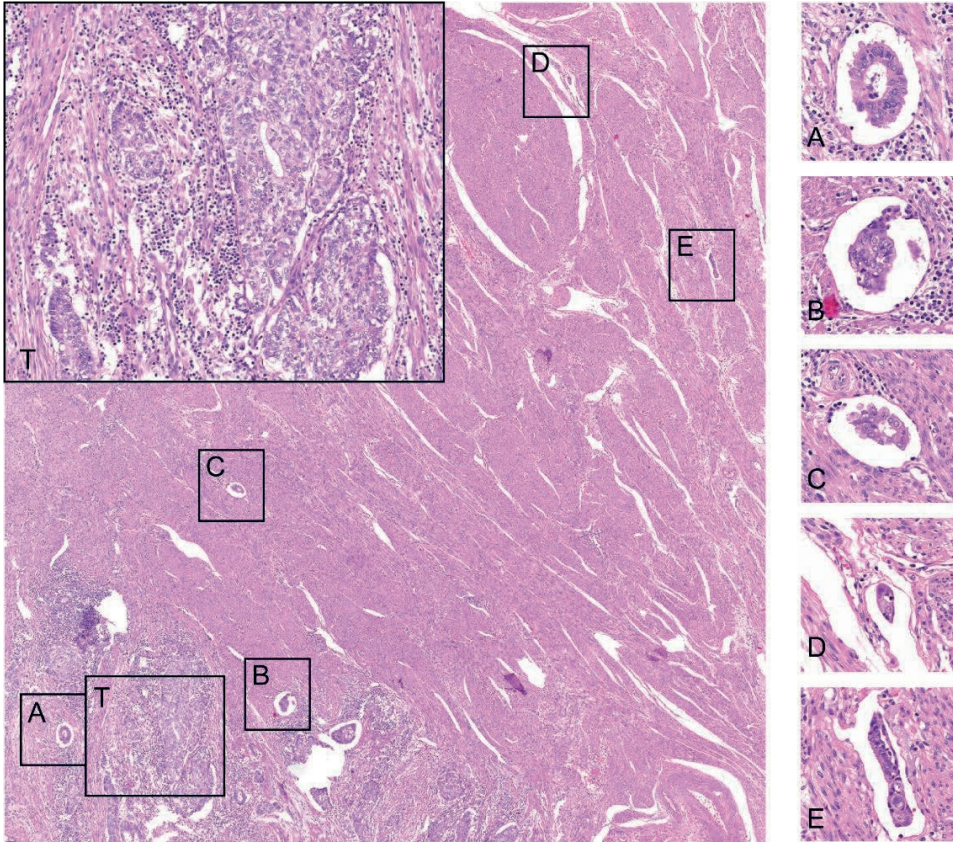


Figure 4. A case derived from phase II with partial agreement on substantial LVSI. Box T shows a detail of the EEC with a prominent peritumoral infiltrate. Inset A-E show details of putative LVSI foci that were annotated for this case that was called substantial LVSI by the study pathologist. Five observers diagnosed this case as positive for LVSI and four agreed to grade this as substantial LVSI.

Discussion

In this study we explored the interobserver reproducibility in both diagnosing LVSI and in the application of a recently introduced LVSI grading system[8]. As presence of LVSI is considered one of the strongest predictors of recurrence in early-stage EC, it is critical to assess reproducibility and identify problematic areas to further improve LVSI assessment. Here, we show that gynaecological pathologists reach substantial agreement in LVSI assessment.

We did not provide the observers with a LVSI definition, because in literature a consensus definition for LVSI is lacking. A variety of elements in the definition of LVSI can be found in the literature, such as: the presence of an endothelial lining [21], use of ancillary studies [22-24], position of the LVSI focus relative to the tumour [25], attachment of the embolus to the vessel wall or not [26, 27], nature of the vessel (lymphatic, vascular, “capillary-like”) [25, 28, 29], vitality and shape of the embolus [30] and presence of surrounding erythrocytes [31] or perivascular

infiltrates [14]. We did however ask our observers to provide the LVSI definition they use in their daily practice. These definitions showed significant overlap, and all LVSI to be defined as “tumour cells” located in a “vessel”. The minor differences in refinements to this definition are unlikely a source of varying interpretations.

With this study we add to previous studies regarding reproducibility of pathological reporting of other EC specific characteristics like histological typing, tumour grading assessment of cervical involvement and assessment of myometrial invasion [32-35]. Levels of reproducibility of these tumour characteristics are similar to our results for LVSI assessment. None of the previous studies specifically focused on LVSI assessment, but there are two studies that report on reproducibility of LVSI in EC [33, 36]. LVSI and other tumour characteristics were reviewed as part of upfront pathology review before randomisation in the PORTEC-3 trial [36]. A high rate of inter-observer agreement between the original pathology report and central pathology review was found for LVSI ($\kappa = 0,72$). In the study by Guan et al., LVSI assessment was part of an alternative binary grading system in EC [33]. Here, LVSI was defined as clusters of malignant epithelial cells within vascular spaces located outside the main tumour. Assessment was performed on H&E slides and CD31 was used to identify the endothelial lining in indeterminate or suspicious cases. Assessment of 254 EC by four pathologists resulted in a disappointing κ -value of 0.23 for LVSI. Several explanations may be considered as to why our study resulted in much higher κ -values. First, LVSI was one parameter among three others, making observers less focussed on one particular parameter. Second, in our study observers were guided to a predefined focus, ensuring that all observers looked at the same area of interest. Last, the observers in our study were selected based on their special interest in gynaecological pathology with an assumption that they are familiar with common LVSI-mimics in EC.

Some of the observers in our study commented that they would have used immunohistochemistry (IHC) to prove the presence of endothelial cells in a subset of the presented cases. Although, the role of adding IHC to LVSI assessment was not part of the study design, it seems obvious that difficult cases may benefit from the use of IHC. Appropriate IHC to help demonstrate LVSI are pan endothelial (CD31) or lymph vessel specific (podoplanin/D2-40) antibodies. Weber et al. found D2-40 IHC increases the proportion of LVSI positive cases in EC compared to H&E evaluation alone. Interestingly, all D2-40 positive cases could be retrospectively identified on H&E [37]. Alexandre-Sefre et al. compared routine H&E LVSI detection with dual pancytokeratin and CD31 staining and found a threefold increase in the LVSI detection rate from 18% with H&E to 54% using IHC in stage I EC [25]. However, both studies fail to illustrate how the increased detection with IHC would affect the clinical relevance/prognostic strength of LVSI detection. There may also be reasons to be reluctant to apply IHC universally. Cancer-associated fibroblasts surrounding adenocarcinoma of the lung [38], and breast [39] have been shown to express podoplanin. Although nonspecific fibroblastic reactivity was not described in the studies of Weber et al. and Alexandre-Sefre et al., it is

possible that an extensive fibroblastic reaction in EC (e.g., in MELF-infiltrative growth pattern) could exhibit podoplanin positivity and results in an incorrect diagnosis of LVSI. Furthermore, Harris et al, showed that the assessment of both small and large vessel involvement in colorectal carcinoma could not be not improved by application of D2-40 and CD31[40]. We acknowledge however that the use of IHC can be useful in selected difficult cases (e.g., cases with extensive retraction artefact), and when used in the correct context will likely further improve interobserver agreement.

Reproducibility of LVSI assessment has also been studied in the context of other tumours like hepatocellular carcinoma (HCC)[41], colorectal cancer [40] and squamous cell carcinoma of the floor of the mouth [42]. In the HCC study [41] inter- and intraobserver reproducibility of six pathologists were analysed. LVSI definitions were not provided and 126 slides and 26 images circulated among the observers twice. There was moderate overall agreement in both attempts (first round $\kappa = 0.50$, second round $\kappa = 0.43$), with slightly lower agreement among non-hepatopathologists compared to hepatopathologists. A study in colorectal cancer [40] included 50 cases from which one H&E slide circulated among six gastrointestinal pathologists assessing small and large vessel invasion using the individual pathologists own criteria. The agreement for small vessel invasion on H&E slides was fair ($\kappa = 0.28$). Agreement was not improved with the use of CD31 ($\kappa = 0.26$) or D2-40 ($\kappa = 0.32$). LVSI assessment in squamous carcinoma of the floor of the mouth [42] was performed on H&E slides from 58 cases by three pathologists using their own criteria. This resulted in substantial agreement for LVSI ($\kappa = 0.64$), comparable to our findings. The variation in levels of agreement between these studies shows that reproducibility of LVSI assessment is likely tumour type specific.

A three-tiered LVSI grading system for EEC (no, focal, substantial) has only recently been proposed [8]. Despite its novelty, this study showed that the observers were able to apply the semiquantitative system with good agreement. Focal LVSI was defined as “a single focus of LVSI around a tumour” and substantial LVSI was defined as “diffuse or multifocal LVSI around a tumour”. Given the considerable reproducibility of this system, this seems a very reasonable approach in daily practice. We do, however, recognize that problematic cases exist, in which this semiquantitative approach may not suffice. For example, cases with 2-5 involved vessels, clustered in a small focus, may be regarded as “focal” by some (if assumed that all the foci of LVSI involve a single vessel) and “substantial” by others. Although this scenario is rare and therefore will be a minor problem in practice, the grading system may benefit from more precise cut-off values. One would anticipate that this would result in further improvement of the reproducibility. At the time of this study, no evidence based cut-off values were available.

Like all interobserver studies, this study is not without its limitations. Importantly, given the lack of a gold standard, we had to rely on the assessment of the study-pathologists for case selection. The study cohort was enriched for cases with potential LVSI, including a selection of LVSI artefacts and mimics, and therefore represents a selected and diagnostically difficult

cohort. The level of interobserver agreement in this study, therefore, likely represents an underestimation of the true agreement for LVSI assessment in EC. A more realistic unselected routine cohort would include many LVSI-negative cases without artefacts or mimics, which would likely result in a much higher agreement. Furthermore, we did not provide serial sections or additional stains to the observers, which in selected cases may have improved agreement.

In summary, this study shows that gynaecological pathologists are able to adequately discriminate unequivocal LVSI from LVSI-mimics. LVSI grading using a recently proposed 3-tiered system (no, focal, substantial) was reproducible. Given the prognostic relevance [8], this study further supports the implementation of this LVSI grading system to routine clinical practice.

References

1. Keys, H.M., Roberts, J.A., Brunetto, V.L., et al., *A phase III trial of surgery with or without adjunctive external pelvic radiation therapy in intermediate risk endometrial adenocarcinoma: a Gynecologic Oncology Group study*. *Gynecol Oncol*, 2004;**92**(3):744-51.
2. Creutzberg, C.L., van Putten, W.L., Koper, P.C., et al., *Surgery and postoperative radiotherapy versus surgery alone for patients with stage-1 endometrial carcinoma: multicentre randomised trial. PORTEC Study Group. Post Operative Radiation Therapy in Endometrial Carcinoma*. *Lancet*, 2000;**355**(9213):1404-11.
3. Kong, A., Johnson, N., Kitchener, H.C., et al., *Adjuvant Radiotherapy for Stage I Endometrial Cancer: An Updated Cochrane Systematic Review and Meta-analysis*. *JNCI: Journal of the National Cancer Institute*, 2012;**104**(21):1625-34.
4. Talhouk, A., McConechy, M.K., Leung, S. et al., *A clinically applicable molecular-based classification for endometrial cancers*. *Br J Cancer*, 2015;**113**(2):299-310.
5. Stelloo, E., Nout, R.A., Osse, E.M., et al., *Improved Risk Assessment by Integrating Molecular and Clinicopathological Factors in Early-stage Endometrial Cancer-Combined Analysis of the PORTEC Cohorts*. *Clin Cancer Res*, 2016;**22**(16):4215-24.
6. Morrow, C.P., Bundy, B.N., Kurman, R.J., et al., *Relationship between surgical-pathological risk factors and outcome in clinical stage I and II carcinoma of the endometrium: a Gynecologic Oncology Group study*. *Gynecol Oncol*, 1991;**40**(1):55-65.
7. Colombo, N., Creutzberg, C.L., Amant, F., et al., *ESMO-ESGO-ESTRO Consensus Conference on Endometrial Cancer: diagnosis, treatment and follow-up*. *Ann Oncol*, 2016;**27**(1):16-41.
8. Bosse, T., Peters, E.E.M., Creutzberg, C.L., et al., *Substantial lymph-vascular space invasion (LVSI) is a significant risk factor for recurrence in endometrial cancer--A pooled analysis of PORTEC 1 and 2 trials*. *Eur J Cancer*, 2015;**51**(13):1742-50.
9. Winer, I., Ahmed, Q.F., Mert, I. et al., *Significance of lymphovascular space invasion in uterine serous carcinoma: what matters more; extent or presence?* *Int J Gynecol Pathol*, 2015;**34**(1):47-56.
10. Dekker, T.J., van de Velde C.J., van Bruggen D., et al., *Quantitative assessment of lymph vascular space invasion (LVSI) provides important prognostic information in node-negative breast cancer*. *Ann Oncol*, 2013;**24**(12):2994-8.
11. Logani, S., Herdman A.V., Little J.V., et al., *Vascular "pseudo invasion" in laparoscopic hysterectomy specimens: a diagnostic pitfall*. *Am J Surg Pathol*, 2008;**32**(4):560-5.
12. Kitahara, S., Walsh, C., Frumovitz, M., et al., *Vascular pseudoinvasion in laparoscopic hysterectomy specimens for endometrial carcinoma: a grossing artifact?* *Am J Surg Pathol*, 2009;**33**(2):298-303.
13. Murray, S.K., Young R.H., Scully, R.E. *Unusual epithelial and stromal changes in myoinvasive endometrioid adenocarcinoma: a study of their frequency, associated diagnostic problems, and prognostic significance*. *Int J Gynecol Pathol*, 2003;**22**(4):324-33.
14. Ambros, R.A., Kurman, R.J. *Combined assessment of vascular and myometrial invasion as a model to predict prognosis in stage I endometrioid adenocarcinoma of the uterine corpus*. *Cancer*, 1992;**69**(6):1424-31.
15. Geels, Y.P., Pijnenborg, J.M., van den Berg-van Erp, S.H., et al., *Endometrioid endometrial carcinoma with atrophic endometrium and poor prognosis*. *Obstet Gynecol*, 2012. **120**(5):1124-31.
16. Rasool, N., Fader, A.N., Saemon L. et al., *Stage I, grade 3 endometrioid adenocarcinoma of the endometrium: an analysis of clinical outcomes and patterns of recurrence*. *Gynecol Oncol*, 2010. **116**(1):10-4.
17. Nout, R.A., Smit, V.T.H.B.M., Putter H., et al., *Vaginal brachytherapy versus pelvic external beam radiotherapy for patients with endometrial cancer of high-intermediate risk (PORTEC-2): an open-label, non-inferiority, randomised trial*. *Lancet*, 2010. **375**(9717):816-23.
18. Hachisuga, T., Kaku, T., Fukuda, K., et al., *The grading of lymphovascular space invasion in endometrial carcinoma*. *Cancer* 1999;**86**(10):2090-7.

19. Hallgren, K.A. *Computing Inter-Rater Reliability for Observational Data: An Overview and Tutorial*. Tutor Quant Methods Psychol, 2012;**8**(1):23-34.
20. Lim, D., Alvarez, T., Nucci, M.R., et al., *Interobserver variability in the interpretation of tumor cell necrosis in uterine leiomyosarcoma*. Am J Surg Pathol, 2013;**37**(5):650-8.
21. Alexander-Sefre, F., Nibbs, R., Rafferty, T., et al., *Clinical value of immunohistochemically detected lymphatic and vascular invasions in clinically staged endometrioid endometrial cancer*. Int J Gynecol Cancer, 2009;**19**(6):1074-9.
22. Vandeput, I., Vanhove, T. Calster, B.V., et al., *The use of lymph vessel markers to predict endometrial cancer outcome*. Int J Gynecol Cancer, 2010;**20**(3):363-7.
23. Miyakuni, Y., Matsumoto, T., Arakawa, A., et al., *Lymphatic invasion according to D2-40 immunostaining is a predictor of nodal metastasis in endometrioid adenocarcinoma of the uterine corpus*. Pathol Int, 2008;**58**(8):471-6.
24. Alexander-Sefre, F., Singh, N., Ayhan, A. et al., *Detection of tumour lymphovascular space invasion using dual cytokeratin and CD31 immunohistochemistry*. J Clin Pathol, 2003;**56**(10):786-8.
25. Wei, S., Conner, M.G., Zhang, K., et al., *Juxtatumoral stromal reactions in uterine endometrioid adenocarcinoma and their prognostic significance*. Int J Gynecol Pathol, 2010;**29**(6):562-7.
26. Gadducci, A., Cosio, S., Fabrini, M.G., et al., *Patterns of failures in endometrial cancer: clinicopathological variables predictive of the risk of local, distant and retroperitoneal failure*. Anticancer Res, 2011;**31**(10):3483-8.
27. Chang, S.J., Kong, T.W., Kim, W.Y., et al., *Lymph-vascular space invasion as a significant risk factor for isolated para-aortic lymph node metastasis in endometrial cancer: a study of 203 consecutive patients*. Ann Surg Oncol, 2011;**18**(1):58-64.
28. Gao, Y., Liu, Z., Gao, F. et al., *High density of peritumoral lymphatic vessels is a potential prognostic marker of endometrial carcinoma: a clinical immunohistochemical method study*. BMC Cancer, 2010;**10**:131.
29. Narayan, K., Khaw, P., Bernshaw, D. et al., *Prognostic significance of lymphovascular space invasion and nodal involvement in intermediate- and high-risk endometrial cancer patients treated with curative intent using surgery and adjuvant radiotherapy*. Int J Gynecol Cancer, 2012;**22**(2):260-6.
30. Briet, J.M., Hollema, H., Reesink, N. et al., *Lymphovascular space involvement: an independent prognostic factor in endometrial cancer*. Gynecol Oncol, 2005;**96**(3):799-804.
31. Tsuruchi, N., Kaku, T., Kamura, T. et al., *The prognostic significance of lymphovascular space invasion in endometrial cancer when conventional hemotoxylin and eosin staining is compared to immunohistochemical staining*. Gynecol Oncol, 1995;**57**(3):307-12.
32. Thomas, S., Hussein, Y. Bandyopadhyay, S., et al., *Interobserver Variability in the Diagnosis of Uterine High-Grade Endometrioid Carcinoma*. Arch Pathol Lab Med, 2016;**140**(8):836-43.
33. Guan, H., Semaan, S., Bandyopadhyay, S., et al., *Prognosis and reproducibility of new and existing binary grading systems for endometrial carcinoma compared to FIGO grading in hysterectomy specimens*. Int J Gynecol Cancer, 2011;**21**(4):654-60.
34. van der Putten, L.J., van de Vijver, K., Bartosch, C. et al., *Reproducibility of measurement of myometrial invasion in endometrial carcinoma*. Virchows Arch, 2017;**470**(1):63-8.
35. McCluggage, W.G., Hirschowitz, L., Wilson, G.E., et al., *Significant variation in the assessment of cervical involvement in endometrial carcinoma: an interobserver variation study*. Am J Surg Pathol, 2011;**35**(2):289-94.
36. de Boer, S.M., Wortman, B.G., Bosse, T. et al., *Clinical consequences of upfront pathology review in the randomised PORTEC-3 trial for high-risk endometrial cancer*. Ann Oncol, 2018;**29**(2):424-430.
37. Weber, S.K., Sauerwald, A., Polcher, M. et al. *Detection of lymphovascular invasion by D2-40 (podoplanin) immunoexpression in endometrial cancer*. Int J Gynecol Cancer, 2012;**22**(8):1442-8.
38. Hoshino, A., Ishii, G., Ito, T. et al. *Podoplanin-positive fibroblasts enhance lung adenocarcinoma tumor formation: podoplanin in fibroblast functions for tumor progression*. Cancer Res, 2011;**71**(14):4769-79.

39. Pula, B., Jethon, A., Piotrowska, A. et al. *Podoplanin expression by cancer-associated fibroblasts predicts poor outcome in invasive ductal breast carcinoma*. *Histopathology*, 2011;**59**(6):1249-60.
40. Harris, E.I., Lewin, D.N., Wang, H.L., et al. *Lymphovascular invasion in colorectal cancer: an interobserver variability study*. *Am J Surg Pathol*, 2008;**32**(12):1816-21.
41. Fan, L., Mac, M.T., Frishberg, D.P., et al. *Interobserver and intraobserver variability in evaluating vascular invasion in hepatocellular carcinoma*. *J Gastroenterol Hepatol*, 2010;**25**(9):1556-61.
42. Beggan, C., Fives, C., O'Leary, G., et al. *Pattern of invasion and lymphovascular invasion in squamous cell carcinoma of the floor of the mouth: an interobserver variability study*. *Histopathology*, 2016;**69**(6):914-920.

Supplementary table 1A. Questions and response options in phase 1.

Take a look at the indicated focus. Do you think this is a focus of LVSI?

Yes

No, this is a shrinkage artifact

No, there is no endothelial lining

No, there are no tumour cells

No, this is a focus with MELF pattern invasion (MELF = micro cystic, elongated and fragmented)

No, this is tumour spill

No, because of other reasons

Unsure (explain below)

We would like to learn from your answer. You can elucidate your answer in the box below. (optional)

Supplementary table 1B. Questions and response options in phase 2.

Take a look at the indicated focus and its surroundings. Do you think this case shows LVSI?

No, there is no LVSI

Yes, this case shows mild LVSI

Definition: mild LVSI = a single focus of LVSI around a tumour.

Yes, this case shows substantial LVSI

Definition: diffuse or multifocal LVSI around a tumour.

We would like to learn from your answer. You can elucidate your answer in the box below. (optional)

Supplementary table 2. Raw data phase 1.

CASE	OBSERVER					
	A	B	C	D	E	F
1	1	1	1	1	1	1
2	6	1	6	9	1	6
3	7	3	1	1	1	1
4	6	1	6	1	7	6
5	6	1	6	6	1	1
6	1	1	1	1	1	1
7	6	1	1	1	1	1
8	1	4	4	1	1	3
9	1	1	1	1	1	1
10	1	1	3	1	1	1
11	1	6	9	9	1	1
12	1	1	1	1	1	9
13	1	1	1	1	1	1
14	3	3	9	2	3	2
15	6	1	1	1	1	1
16	1	3	5	1	1	1
17	2	1	1	1	1	3
18	6	6	7	6	6	6
19	6	1	9	1	1	1
20	6	2	1	1	1	1
21	1	1	1	1	1	1
22	1	1	9	1	1	1
23	9	1	1	2	1	2
24	6	5	1	6	1	1
25	5	1	1	5	1	5
26	6	1	1	1	1	9
27	1	1	1	1	1	1
28	5	3	5	5	5	5
29	6	2	6	1	1	1
30	1	6	6	1	1	7
31	7	4	1	4	1	1
32	1	1	1	1	1	1
33	2	2	1	2	1	2
34	5	3	1	5	1	5
35	1	1	5	5	1	5
36	2	2	1	1	1	2
37	1	1	6	1	1	1
38	1	1	9	1	1	1
39	1	1	1	5	1	2
40	6	1	1	9	1	9
41	1	1	1	1	1	2
42	1	2	1	1	1	1
43	9	5	1	6	1	2
44	1	2	1	1	1	1
45	4	4	4	4	1	6
46	3	3	1	3	1	2
47	4	4	9	1	1	1
48	1	1	1	1	1	1

Key: A to F: observers. 1: LVSI positive; 2: No LVSI – shrinkage; 3: No LVSI – no endothelial lining; 4: No LVSI – no tumour cells; 5: No LVSI – MELF; 6: No LVSI – spill; 7: No LVSI – other reasons; 9: Unsure.

Supplementary table 3. Raw data phase 2.

CASE	OBSERVER					
	A	B	C	D	E	F
1	2	2	2	2	2	2
2	1	1	2	0	2	2
3	1	2	1	2	2	2
4	1	2	2	2	2	2
5	0	1	2	1	2	2
6	2	1	2	2	1	2
7	2	1	2	2	1	2
8	1	0	2	1	1	0
9	0	2	2	2	2	2
10	2	2	2	2	2	2
11	2	2	0	2	2	2
12	0	0	2	2	1	1
13	1	1	1	1	1	1
14	0	1	2	1	2	2
15	0	1	1	0	1	1
16	0	1	2	1	2	2
17	1	1	2	2	2	2
18	2	2	2	1	2	2
19	1	1	2	2	2	2
20	1	1	0	0	1	1
21	2	2	2	2	2	2
22	0	2	2	2	2	2
23	0	0	2	1	1	1
24	2	1	2	2	2	2
25	2	2	1	2	2	2
26	2	0	2	1	2	2
27	1	1	2	2	1	2
28	2	1	2	2	2	2
29	0	1	2	2	2	2
30	0	2	2	2	2	2
31	2	1	2	0	1	2
32	1	2	2	2	2	2
33	1	1	2	1	2	2
34	0	2	2	2	1	2
35	2	2	2	2	2	2
36	2	2	2	2	2	2
37	2	1	2	1	1	2
38	1	0	2	0	1	2
39	1	1	2	0	2	2
40	0	1	2	1	1	2
41	0	2	2	2	2	2
42	0	2	2	0	2	2

Key: A to F: observers. 0: no LVSI; 1: focal LVSI; 2: substantial LVSI.