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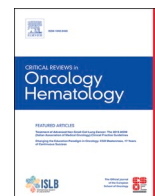
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Imaging response evaluation after neoadjuvant treatment in soft tissue sarcomas: Where do we stand?

Nicolò Gennaro^{a,b,c,*}, Sophie Reijers^d, Annemarie Bruining^c, Christina Messiou^{e,f}, Rick Haas^{g,h}, Piergiuseppe Colomboⁱ, Zuhir Bodalal^{c,j}, Regina Beets-Tan^{c,j,k}, Winan van Houdt^d, Winette T.A. van der Graaf^{l,m}

^a Humanitas Research and Cancer Center, Dept. of Radiology, Rozzano, Italy

^b Humanitas University, Dept. of Biomedical Sciences, Pieve Emanuele, Italy

^c The Netherlands Cancer Institute, Dept. of Radiology, Amsterdam, the Netherlands

^d The Netherlands Cancer Institute, Dept. of Surgical Oncology, Amsterdam, the Netherlands

^e The Royal Marsden NHS Foundation Trust, Dept. Of Radiology Sarcoma Unit, Sutton, United Kingdom

^f The Institute of Cancer Research, Sutton, United Kingdom

^g The Netherlands Cancer Institute, Dept. of Radiation Oncology, Amsterdam, the Netherlands

^h Leiden University Medical Center, Dept. of Radiation Oncology, the Netherlands

ⁱ Humanitas Research and Cancer Center, Dept. of Pathology, Rozzano, Italy

^j GROW School for Oncology and Developmental Biology, Maastricht University, Maastricht, the Netherlands

^k Danish Colorectal Cancer Center South, Vejle University Hospital, Institute of Regional Health Research, University of Southern Denmark, Denmark

^l The Netherlands Cancer Institute, Dept. of Medical Oncology, Amsterdam, the Netherlands

^m Erasmus MC Cancer Institute, Dept. of Medical Oncology, Erasmus University Medical Center, Rotterdam, the Netherlands

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ABSTRACT

Soft tissue sarcomas (STS) represent a broad family of rare tumours for which surgery with radiotherapy represents first-line treatment. Recently, neoadjuvant chemo-radiotherapy has been increasingly used in high-risk patients in an effort to reduce surgical morbidity and improve clinical outcomes. An adequate understanding of the efficacy of neoadjuvant therapies would optimise patient care, allowing a tailored approach. Although response evaluation criteria in solid tumours (RECIST) is the most common imaging method to assess tumour response, Choi criteria and functional and molecular imaging (DWI, DCE-MRI and ¹⁸F-FDG-PET) seem to outperform it in the discrimination between responders and non-responders. Moreover, the radiologic-pathology correlation of treatment-related changes remains poorly understood. In this review, we provide an overview of the imaging assessment of tumour response in STS undergoing neoadjuvant treatment, including conventional imaging (CT, MRI, PET) and advanced imaging analysis. Future directions will be presented to shed light on potential advances in pre-surgical imaging assessments that have clinical implications for sarcoma patients.

1. Introduction

Soft tissue sarcomas (STS) are a heterogeneous group of rare neoplastic diseases accounting for more than 70 different histologies (WHO, 2021). Locally-advanced sarcoma may benefit from neoadjuvant radiotherapy/chemotherapy to reduce surgical morbidity and improve clinical outcomes. Predicting the efficacy of neoadjuvant treatments is challenging; however, early identification of non-responders offers the potential to prevent unnecessary, potentially toxic treatment and select patients who truly benefit from neoadjuvant therapy. Therefore,

exploring robust methodologies to assess the predictive value of imaging for tumour response to neoadjuvant therapies is relevant and timely (Subbiah et al., 2017).

Although serial biopsies may, in theory, monitor the tumour response, they are invasive and prone to sampling errors which are significant in heterogeneous tumours. Moreover, a proper understanding of the correlation between imaging and chemotherapy- and/or radiation-induced changes in histology is crucial for the development of reliable imaging biomarkers (Schillaci et al., 2019; Hartman et al., 2016). Unfortunately, insights into changes at tissue level after systemic

* Corresponding author at: The Netherlands Cancer Institute (NKI), Dept. of Radiology, Plesmanlaan 121, 1066, Amsterdam, the Netherlands.

E-mail address: nicolo.gennaro@st.hunimed.eu (N. Gennaro).

or local therapy remain limited. Thus, considerable interest has recently risen in radiology-pathology correlation in STS before and after neoadjuvant therapies to identify predictive factors able to influence patient management.

This review gives an overview of the evidence for imaging of the STS in the neoadjuvant setting, identifying strengths and weaknesses and providing directions for future research.

2. Neoadjuvant treatment for soft tissue sarcomas

Neoadjuvant treatment is increasingly used in patients with STS. Specific nomograms can be used to better stratify patients and support clinical decisions (Callegaro et al., 2016; Donahue et al., 2010; Callegaro et al., 2017). Neoadjuvant radiotherapy is predominantly employed to reduce post-surgical local recurrence rates. In particular, radiation therapy may represent the only strategy for local control when surgery is declined, too morbid or not possible (Larrier et al., 2016). Preoperative radiotherapy is usually preferred because late morbidity (e.g. fibrosis, arthrosis, oedema and bone fractures) tends to be lower than in the adjuvant setting, resulting in improved long-term functional outcome and quality of life (Haas, 2018).

Moreover, high-risk STS may be treated upfront with systemic therapy to treat potential micro-metastases and thus prevent metastatic spread. For specific histotypes with locally advanced diseases (e.g. myxoid liposarcomas), chemo/radiotherapy can also be administered as an induction therapy to shrink the tumour volume and enable organ sparing resection (Pasquali and Gronchi, 2017; Saldaz et al., 2017). Recently, the European Organization for Research and Treatment of Cancer (EORTC) and Soft Tissue and Bone Sarcoma Group (STBSG) underwent a survey among their members that confirmed a substantial heterogeneity of treatment strategies and agents used both for pre- and postoperative chemo/radiotherapy in STS (Rothermundt et al., 2018). Multi-agent chemotherapy, like anthracycline plus ifosfamide, is the most common and accepted strategy, whereas doxorubicin and dacarbazine may be used for leiomyosarcoma (D'Ambrosio et al., 2020; Judson et al., 2014).

Some chemotherapy agents, like bevacizumab, sorafenib, sunitinib, pazopanib, and trabectedin, can also act as sensitizers prior to radiotherapy (Yoon et al., 2011; Gronchi et al., 2019; Jakob et al., 2016; Canter et al., 2014). Preoperative conventionally-fractionated RT is usually preferred, but hypo- and hyperfractionated schemes have also been proposed (Koseła-Paterczyk et al., 2014). Neoadjuvant chemo-radiation is a combined strategy for poorly resectable tumours with synergic effects and the potential to lower required RT doses (Look Hong et al., 2013; Spałek et al., 2020; Lehane et al., 2016). Hyperthermic isolated limb perfusion is a further neoadjuvant approach, and can be considered an alternative to amputation in specific circumstances (Song et al., 2019).

3. Response criteria for neoadjuvant treatments

Imaging assessment pursues different aims depending on which kind of therapy is delivered. After local therapy, radiology should investigate the persistence of viable tumour and possibly provide prognostic information about local recurrence. Conversely, imaging assessment after systemic therapy provides insights on histological response and envisages the possibility to provide prognostic information, similarly to what have been demonstrated in osteosarcoma and Ewing's sarcoma (Bielack et al., 2002; Picci et al., 1997). Traditionally, tumour response has been evaluated using RECIST 1.1 criteria, which uses unidimensional measurements (Eisenhauer et al., 2009). Apart from some reports on the effect of radiotherapy on tumour shrinkage in different histotypes, there is no information on a clear association between prognosis, the amount of tumour shrinkage or true histological response in STS treated by chemo/radiotherapy (Roberge et al., 2010; Tanaka et al., 2018; Canter et al., 2010; le Grange et al., 2014; Gui et al., 2019; Miki et al., 2010;

Betgen et al., 2013). The lack of relationship between percentage tumour shrinkage and survival was shown by Grünwald et al. in an EORTC analysis, suggesting that absence of progression rather than extent of tumour shrinkage defines prognosis in advanced STS patients treated with palliative chemotherapy (Grünwald et al., 2016). There is no evidence that the RECIST criterion of 30 %-unidimensional shrinkage of the target lesion always truly reflects a superior anticancer effect. The RECIST 30 %-cut-off is entirely arbitrary and dates back to the time when tumours were measured by physical examination. The main goal of RECIST was to standardise and harmonise endpoints of clinical trials in solid tumours. Furthermore, RECIST does not consider tumour changes like cystic transformation or haemorrhage, which can result in an increase in size even in case of a true histological response (Roberge et al., 2010; Tanaka et al., 2018; Canter et al., 2010; le Grange et al., 2014; Gui et al., 2019). To overcome this important limitation, Stacchiotti et al. demonstrated in 2009 how MR-based Choi criteria, which incorporate both signal intensity and size (Choi et al., 2007), were more accurate in predicting patient outcomes than RECIST (Stacchiotti et al., 2009). Further publications have confirmed such observations (Betgen et al., 2013; Marrari et al., 2020; Esser et al., 2018), and highlighted that RECIST responses are scarce and restricted to a few sarcoma types after neoadjuvant treatment (e.g. myxoid liposarcoma after radiation treatment) (Betgen et al., 2013; Chung et al., 2009; Lansu et al., 2020; Taieb et al., 2015; Stacchiotti et al., 2012). Nevertheless, in the absence of a validated alternative, most clinical sarcoma trials continue to use RECIST 1.1 as a response evaluation system (Schuetze et al., 2008). An alternative and standardised approach for imaging evaluation is needed to incorporate noninvasive imaging information into the decision-making algorithms of multidisciplinary sarcoma teams. This includes the definition of optimal acquisition and timing protocol along with a structured and objective image interpretation method. Timing of imaging assessment differs between radiation therapy and chemotherapy, as surgery is performed immediately following chemotherapy, whereas the interval between the end of radiation and surgery is usually 6–8 weeks (Subbiah et al., 2017).

Regarding the imaging evaluation of tumour response, there is no consensus on the choice of imaging modality. In 2016, a standardised approach for MR longitudinal evaluation of STS following preoperative radiotherapy (RT) was proposed by a task force endorsed by the EORTC-STBSG and Image Groups (Messiou et al., 2016). The paper advocated a standard MRI protocol to be performed at 4–6 weeks following neoadjuvant radiotherapy as close to the surgical date as possible to minimise the potential for pseudo-progression due to the acute effects of radiotherapy including vascular disruption, possibly causing an increase in size and enhancement. To ensure consistency in longitudinal evaluations, the same or comparable MRI equipment should be used. Furthermore, technical refinements regarding contrast medium administration have been recently proposed, highlighting how a 60s-acquisition delay optimises the response evaluation when adopting Choi criteria (Crombé et al., 2019a). Morphological pre- and post-contrast-enhanced sequences should be supplemented by diffusion-weighted MRI to explore changes in tissue cellularity (DWI) (Messiou et al., 2016; Costa et al., 2018). DWI is gaining interest as a widely available and quick quantitative MRI technique for oncology imaging. Predictive and prognostic data in STS are limited, but ongoing studies are shedding light on the role of multi-parametric MRI (which include contrast-enhanced imaging and DWI, but can also be complemented with additional DWI for estimation of intra-voxel incoherent motion (IVIM) and with multiple gradient-echo imaging, among others) for response assessment after neoadjuvant therapy (Costa et al., 2018; Dudeck et al., 2008; Winfield et al., 2019). Dynamic contrast-enhanced (DCE)-MRI has been investigated and shows some promise, but their widespread adoption in clinical practice is limited due to more extensive post-processing requirements and variability in acquisition (Crombé et al., 2019a; Soldatos et al., 2016). Despite a few preliminary reports, MRI-spectroscopy is not yet considered a feasible and reliable tool for

the response assessment of neoadjuvant treatment, since it is not available in most centres and the current evidence is debatable (Zhang et al., 2016).

Finally, ^{18}F -FDG-PET imaging is a well-established technique for assessing metabolic activity (Gennaro et al., 2020), and has been shown to be helpful in the early identification of chemotherapy responders (Grueisen et al., 2019; Lim et al., 2019). Alongside the EORTC-STBSG paper on imaging response evaluation guidelines (Messiou et al., 2016), the same group published in 2016 a standardised approach for pathology evaluation after neoadjuvant therapy, highlighting the importance of radiology-pathology correlation (Wardelmann et al., 2016). The proposed protocol provides specific instructions on the handling and photography of surgical specimens to enable correlation with radiologic findings. Moreover, such correlation may serve as a useful guide for pathologists to select the most representative pathology samples. The two publications were proposed as frameworks to build new evidence. Unfortunately, the first study to assess the prognostic value of the EORTC-STBSG response score failed to correlate the amount of viable tumour cells with the clinical outcome (Schaefer et al., 2017).

4. Radiology-pathology correlation in soft tissue sarcoma

Pathology is considered the gold standard for the definition of tumour changes after therapy. The most important factors for predicting tumour response and survival after chemo-radiotherapy are pre-treatment grading and histological subgroup. High-grade tumours and undifferentiated pleomorphic sarcomas (UPS) have been associated with improved response rates to the combination of doxorubicin and ifosfamide in the metastatic setting as compared to single agent doxorubicin (Young et al., 2017). Imaging findings have recently been shown to be complementary to tissue findings for the assessment of tumour grade (Fisher et al., 2016). A grade difference between biopsy and excision specimens of leiomyosarcomas was found in 68 % of cases, with all these cases showing an increase in grade from biopsy to excision specimen (Schneider et al., 2017). CT and MRI imaging features (e.g. necrotic areas) have been proposed to be complementary to improve the accuracy of histopathological grading (McCaddy et al., 2020; Crombé et al., 2019b; Zhao et al., 2014). CT and MRI texture analysis is an advanced image analysis referring to characterisation of regions in MR images, including DCE-MRI and DWI sequences, by their texture content. Both CT- and MRI-based texture analysis may also play a relevant role in this regard (Peeken et al., 2019; Hong et al., 2020; Zhang and Ren, 2020).

Dependent on histology and treatment, several changes in the specimen can be recognised after therapy, including the presence of ghost cells (necrotic cells with loss of nuclear and cytoplasmic detail), reduced cellularity (intended as reduction of stainable cells), granulation tissue (observed as foamy macrophages and lymphocytes), fibrosis, adipose tissue, hemosiderin deposition, changes in vessel micro density, development of calcifications, atrophic or regenerating non-neoplastic skeletal muscle (Lucas et al., 2008). A further complicating factor is the well-known substantial discordance in the evaluation of histological subtype, tumour grade and definitions of post-treatment changes existing between pathologists working at reference institutions and those who are not (up to 40 % of cases) (Young et al., 2017). To date, there is no agreement on measuring the effects of neoadjuvant chemotherapy/radiation therapy on STS. Expert pathologists recommend describing the morphological changes throughout the tumour (represented as a percentage), including the percentage of viable tumour, necrosis, haemorrhage, sclero-hyalinosis, reparative fibrosis, fibro-histiocytic reaction with deposits of hemosiderin, myxoid and cystic changes. In this setting, adequate sampling is of paramount importance (almost 1 sample per cm), including necrotic areas. The gross appearance of the tumour can be misleading, as areas that appear necrotic may represent myxoid or oedematous tissues (Shah et al., 2016; Rubin et al., 2010). A standardised system proposed in 2016 by the aforementioned EORTC-STBSG task force is summarised in Table 1

Table 1

Assessment of tumour response proposed by EORTC-STBSG in 2016 (Wardelmann et al., 2016).

Histological findings	Response grade
No stainable vital tumor cells	A
Single stainable tumor cells or small cluster (overall < 1%)	B
≥1% <10 % stainable vital tumour cells	C
10 % <50 % stainable vital tumour cells	D
≥50 % stainable vital tumour cells	E

(Wardelmann et al., 2016).

Different imaging modalities like MRI, CT and ^{18}F -FDG PET aim to macroscopically depict those changes, with a particular interest in detecting necrosis, as 95 % necrosis after neoadjuvant therapy was initially correlated to a halved risk of local recurrence and metastatic spread and pathology-related deaths at 5 and 10 years (Cousin et al., 2017; Eilber et al., 2001). These results are still controversial and advocate for larger studies to confirm that the extent of histopathological necrosis after neoadjuvant treatment may influence patient management, as several papers were unable to correlate extended tissue necrosis with improved survival (Canter et al., 2010; Vaynrub et al., 2015; Mullen et al., 2014; Menendez et al., 2007). This could be because necrosis does not reflect the biological behaviour of the tumour and its metastatic potential, including circulating tumour cells. Recently, the extent of hyalinization/fibrosis has also been suggested as a better predictor for recurrence-free survival and overall survival, supporting the hypothesis that histologic appearance can predict tumour biology and represent a future endpoint for neoadjuvant trials (Schaefer et al., 2017). Reliable imaging biomarkers would allow radiologists to complement pathology assessment and provide non-invasive, predictive and prognostic information (Chen et al., 2013).

Integration of multi-modality imaging with clinical data may support experienced radiologists to better interpret imaging changes in patients treated with neoadjuvant therapy. Beneath, we present the latest evidence for each imaging modality that has allowed a better understanding of radiologic-pathology correlation after neoadjuvant treatments in STS.

4.1. Magnetic resonance imaging (MRI)

MRI is the preferred imaging modality for STS, particularly for extremity STS. Its predominant role is to define the local extent of disease and to accurately depict anatomy (Raghavan, 2017). MRI imaging features include morphology, signal intensity on different sequences and enhancement after contrast medium. Features including peritumoral contrast enhancement and oedema, heterogeneous T2 signal intensity and necrotic areas have been associated with high histological grade and poorer outcomes (Crombé et al., 2019b; Zhao et al., 2014). Despite a preliminary study describing T1W hyper-intensity correlation with improved survival, relapse, and time to relapse (Sala et al., 2010), conventional morphologic MRI characteristics are subjective and generally inadequate in discriminating between patients with positive or negative clinical response (Favinger et al., 2018). Moreover, necrosis on imaging lacks an agreed/validated definition and is mostly defined as areas of low tumour perfusion, T2W hyper-intensity and high apparent diffusion coefficient (ADC) values at DWI (Soldatos et al., 2016; Crombé et al., 2019b; MacVicar et al., 1992) (Fig. 1). However, such correlations are not always straightforward, as pathological entities may show high ADC values (myxoid liposarcoma), as well as certain types of necrosis may manifest can also manifest with low ADC values (e.g. coagulative necrosis) (Figs. 2 and 3). Semi-automated volumetric segmentation techniques provide more objective measurements and have been shown to identify necrosis on MRI with similar accuracy compared to histopathology (Monksy et al., 2012). A study of multi-parametric MRI, entailing DCE and DWI MRI for assessing response to radiotherapy, has

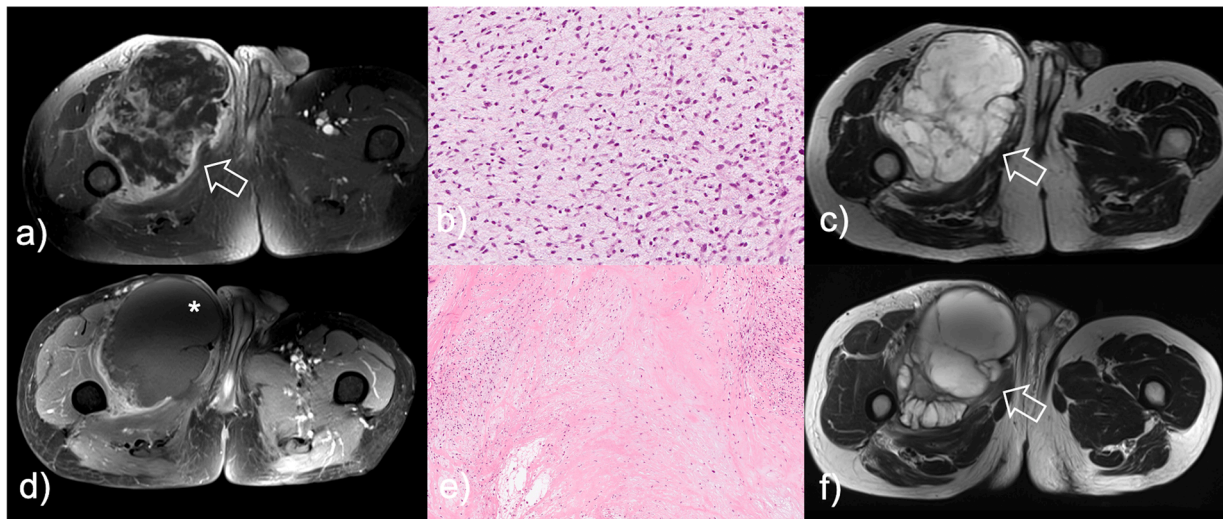


Fig. 1. a) Large lesion in the right thigh of a 52-year old man with heterogeneous contrast-enhancement on T1-weighted MRI due to internal areas of necrosis. b) Pre-treatment core needle biopsy shows high-grade spindle cell sarcoma with myogenic differentiation (Ki67 80 %). c) Liquid/myxoid content is shown on T2-weighted images and d) Contrast-enhanced T1-weighted MRI after chemoradiotherapy (3 cycles of adriamycin-dacarbazine and 50 Gy/25 fractions) shows no intralesional contrast-enhancement in keeping with response to therapy although the mass is stable by RECIST 1.1. e) Post-surgical pathology shows large areas of necrosis and reduction in neoplastic cellularity, partly visible on the top-right and accounting for 70 % of the whole specimen. f) Fibrohistiocytic changes and sclerosis are also present and are represented by hypointense septa on T2-weighted images.

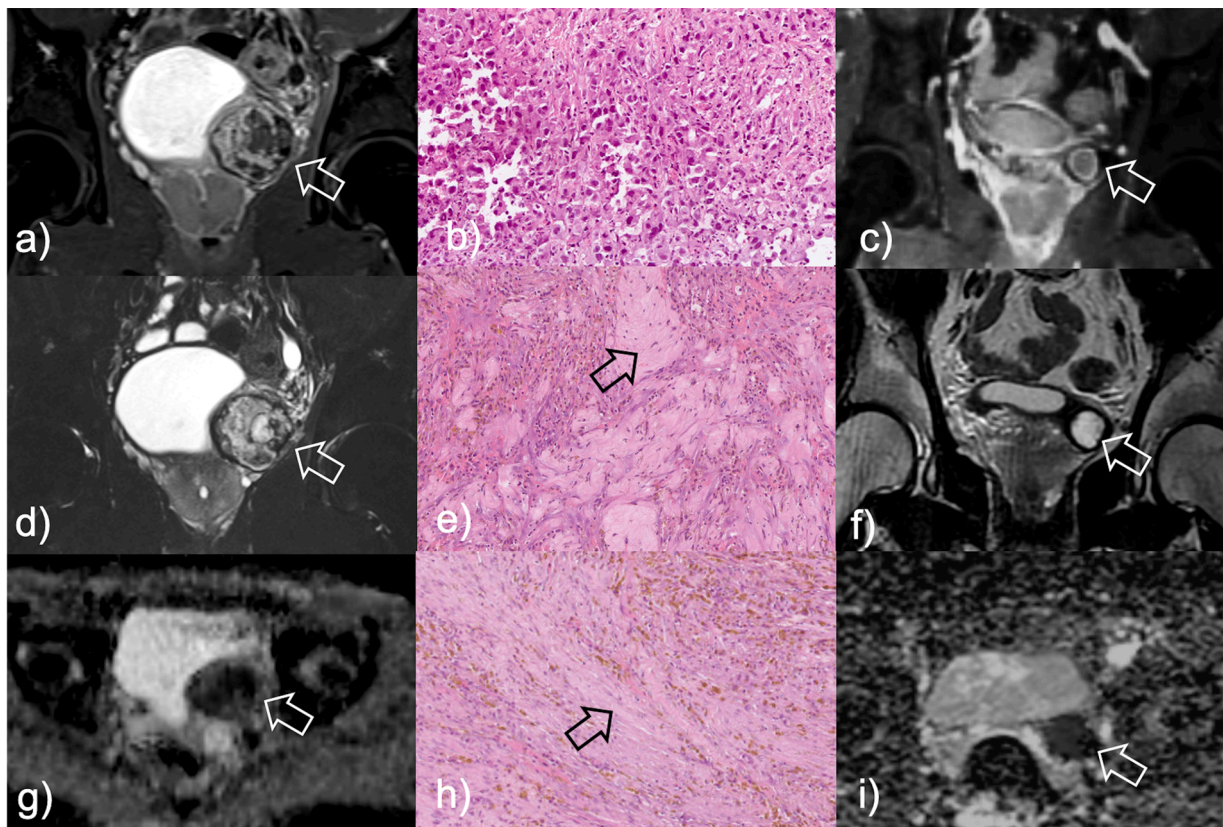


Fig. 2. a) Soft tissue mass arising from the left wall of the bladder of a 30-year old patient. a) Contrast enhanced T1-weighted MRI showing areas of non-enhancing internal necrosis (arrow). b) Pre-treatment core needle biopsy shows high grade mesenchymal neoplasm characterized by atypical vascular structures consistent with epithelioid angiosarcoma (Ki-67 80 %). c) Post-chemotherapy (3 cycles of adriamycin-ifosfamide) contrast-enhanced T1-weighted MRI shows reduction in tumor size and diminished contrast-enhancement (arrow) d) Pretreatment T2-weighted images showing heterogeneous signal with liquid/myxoid areas (arrow). e) Post-surgical specimen shows florid reparative fibrosis (arrow) and fibrohistiocytic reaction with hemosiderin deposition. f) There is a homogenous bright signal on post-treatment T2-weighted images (arrow) g) Pre-treatment ADC map showing low signal corresponding to high tumor cellularity, h) Higher magnification demonstrates foci of sclero jalinosis (arrow) and deposition of hemosiderin and foamy histiocytes. i) The post-treatment ADC map demonstrates low signal/ restricted diffusion possibly corresponding to coagulative necrosis and haemosiderin.

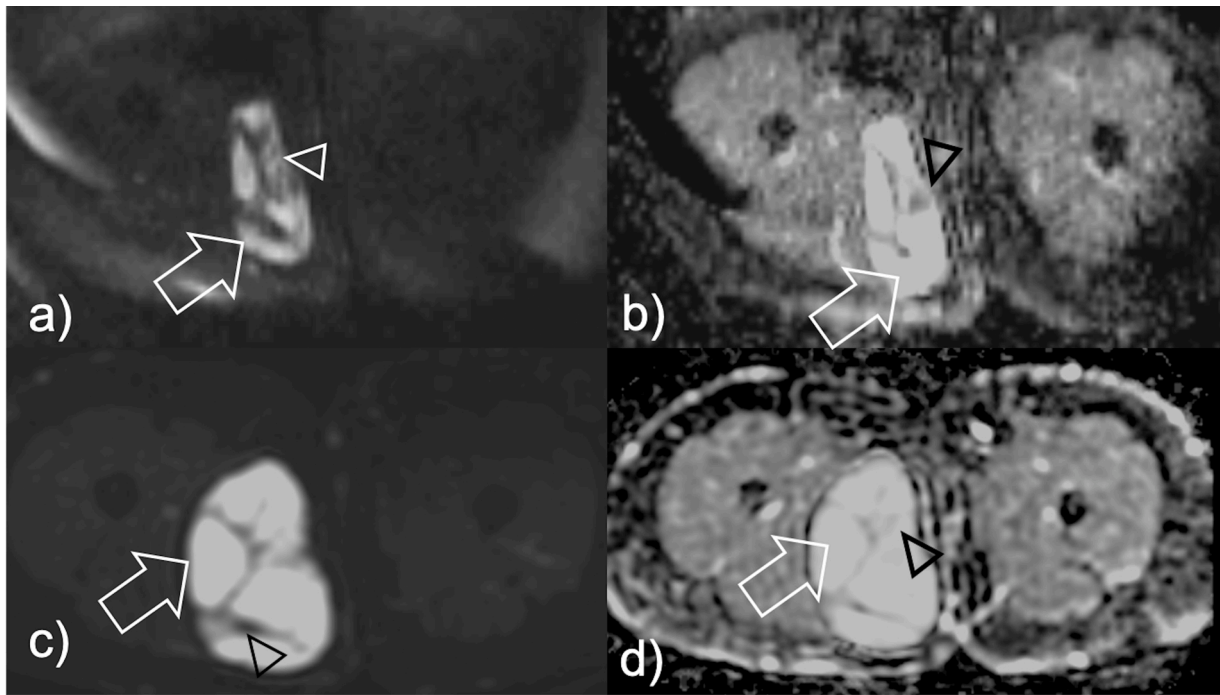


Fig. 3. a) Axial DWI scan shows a myxoid liposarcoma (<1/10 HPF) in the right thigh, composed of myxoid/cystic areas (arrow) and nodular septa (arrowhead); b) Despite the malignant nature, the mass is hyperintense at ADC map due to the high content in myxoid matrix c) After chemo-radiation, axial DWI and d) ADC map scans show a considerable increase in dimensions, especially of the myxoid areas. However, pathology described only 5% of viable cells (5/10 HPF) with 50 % fibrosis.

confirmed that ADC is not only a highly reproducible parameter but showed significant increase (suggesting response) post radiotherapy despite size stability. Furthermore, histopathological correlations confirmed correspondence between restriction and degree of cellularity

(Winfield et al., 2019). More recently, attention has shifted from the core of the tumour, where necrosis is easily found after neoadjuvant therapy, to the tumour periphery, where viable cells may persist (Baur et al., 2003). An infiltrative margin on post-contrast images has been

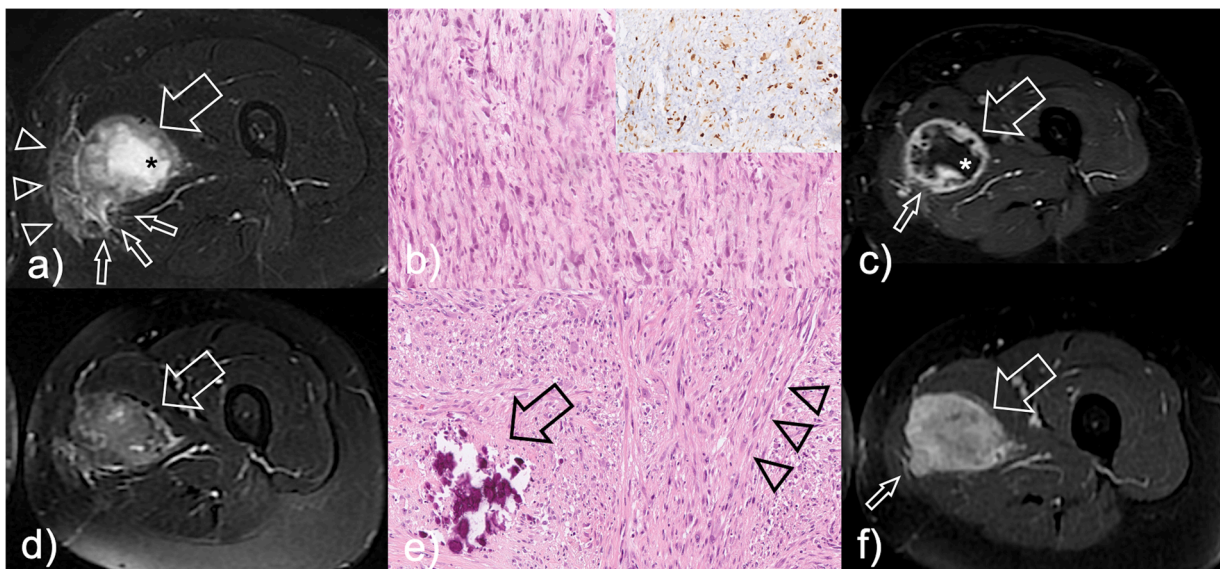


Fig. 4. a) Baseline axial fat-suppressed T2W showing a deep-seated mass in the right thigh (arrow) with perilesional oedema (arrowheads), internal cystic/myxoid areas (asterisk), with ill-defined margins in its posterior lateral surface (small arrows); b) Baseline core tissue biopsy reveals pleomorphic and spindle cells coherent with myxoid liposarcoma, grade III (7/10 HPF, Ki67(MIB1)60 %) with MDM2 amplification (right upper box); c) Baseline axial fat-suppressed contrast-enhanced T1W shows peripheral enhancement (arrow), internal necrotic areas (asterisks) and ill-defined margins (small arrow); d) After chemo-radiotherapy (doxorubicin, adriamycin 3 cycles and 50 Gy) axial fat-suppressed T2W shows no significant changes in tumour size (arrow), but cystic/myxoid areas are not assessable anymore (arrow); e) core tissue biopsy shows >95 % vital cells represented by dedifferentiated spindle cell (arrowheads) and pleomorphic liposarcoma cells (2/10 HPF) with small areas of low-grade, well-differentiated liposarcoma cells. Post-radiotherapy changes are clearly visible in the right lower corner (calcification, arrow); f) Axial fat-suppressed contrast-enhanced T1W shows homogeneous enhancement with no internal necrotic areas (arrow) and further invasion of nearby structures (small arrow), coherent with poor response to chemo-radiotherapy.

associated with a lower disease-specific and metastasis-free survival (MFS), highlighting the importance of peripheral growth pattern on MRI and histology (Fig. 4) (Nakamura et al., 2017; Liu et al., 2008).

Changes in peri-tumoral regions also appear to play a role in the assessment of response. In fact, the presence of satellite tumorous cells on surgical specimens is predictive of lower survival and disease-free survival (Kandel et al., 2013). Peritumoral oedema has been hypothesised to be proportional to peritumoral tumour satellites (Hanna et al., 1991; Beltran et al., 1987). Conventional MRI sequences accurately identify peritumoral oedema through fluid sensitive images, including T2W, T2W-FS or T2W-STIR. Stability or increase of peritumoral oedema has been recently associated with a poor histological response (>10 % viable cells), as well as stability or increase of pre-existing contrast-enhanced peritumoral areas (Crombé et al., 2018), but the clinical significance of peritumoral oedema is still controversial (White et al., 2005). Functional quantitative imaging refers to a collection of imaging parameters that aim to depict biological information and are showing encouraging results compared to morphological imaging in the definition of tumour response. Changes in DCE-MRI parameters evaluated after the second cycle of neoadjuvant chemotherapy have been shown to be highly predictive of histological response, whereas RECIST 1.1 was not (Cousin et al., 2017). A recent publication showed poor predictive value of conventional MRI (T1W, T2W, T2W with fat suppression images and T1W after contrast administration) for response when neoadjuvant therapies induce fibrosis and/or granulation tissue instead of necrosis (Soldatos et al., 2016). However, over 5% of early tumour enhancements and an average ADC value $> 2.2 \times 10^{-3} \text{ mm}^2/\text{sec}$ are considered markers of positive response, both reaching 100 % sensitivity in determining nonviable tumour areas (Soldatos et al., 2016). Further, studies confirmed the emerging role of quantitative DCE-MRI in early prediction using absolute parameters like K_{trans} , k_{ep} , AUC60 or time-to-peak (Huang et al., 2016; Meyer et al., 2013; O'Connor et al., 2011). DCE-MRI was validated in vivo models for the very early assessment (24 h) after isolated limb perfusion, once again with promising results (Alic et al., 2013; Preda et al., 2004).

Frontline research in MRI evaluation of tumour response includes the innovative concept of “imaging habitats” (Napel et al., 2018). This mapping technique consists of a systematic approach of (1) intratumour segmentation into sub-regions based on pixel intensity (2) qualitative or quantitative analysis within each distinctive habitat to explore heterogeneity (3) estimation of percentage of necrosis (Sala et al., 2017). Pilot studies have already shown that necrosis can be identified with a 76 % accuracy through the analysis of radiologically-defined habitats on T2W and contrast-enhanced T1W images, possibly predicting the treatment outcome as well as the development of distant metastasis (Farhidzadeh et al., 2015a, b). A recently published method consists of plotting the voxel-based signal intensity distribution by histogram for analysis of the relative signal intensity compared to tumour volume. This approach allowed discrimination between viable (enhancing) tumour and necrosis (non-enhancing component) using a cut-off signal intensity threshold (Fields et al., 2020). Unsupervised clustering of such habitats can discriminate between responders and non-responders by analysing changes in k_{ep} , k_{trans} , and percent volume when DCE-MRI information is available (Yu-Cherng et al., 2019; Y-CC et al., 2017; Diwanji et al., 2019). Habitat imaging approaches are now also supported by machine learning (Blackledge et al., 2019). MRI texture analysis has already been shown to distinguish low from high-grade soft tissue sarcoma, and could be further explored in the assessment of tumour response (Corino et al., 2018). Recently, a T2-based Δ -radiomics approach has identified three features (Δ _Histogram_Entropy, Δ _Elongation, Δ _Surrounding_Edema) associated with good tumour response, defined as <10 % viable cell at pathology (Crombé et al., 2019c).

4.2. Computed tomography (CT)

CT is not frequently used for local staging due to poorer contrast

resolution compared to MRI. In general, areas that cease to enhance after treatment are considered necrotic and contribute to the definition of Choi criteria through the measurement of Hounsfield Units (Fig. 5). Unfortunately, contrast-enhanced CT cannot differentiate between hypo-densities caused by necrosis and or subacute-to-late haemorrhage or cystic components. Differentiating tumour enhancement from granulation or fibrosis is also very challenging with CT (Crombé et al., 2019b). Evaluation of neoadjuvant therapy using RECIST with MRI or CT has been used to predict radical resection and survival. Unfortunately, they did not correlate to other clinical or pathology treatment characteristics (Chen et al., 2013).

Recently, CT texture analysis has also been performed alongside with the evaluation of descriptive semantic tumour features, which include shape, size, location and attenuation (Esser et al., 2018; Tian et al., 2015). Fine to coarse texture parameters (e.g. entropy, mean value of positive pixels, skewness and kurtosis) were extracted from non-enhanced CT and compared with biologic markers of angiogenesis, showing that the mean value of positive pixels (defined as the average brightness of positive values of the image) showed a better clinical outcome and a definite correlation with micro-vessel density on immunohistochemistry. Interestingly, the entropy parameter also showed positive correlations with plasma VEGF and SVEGFR-1, which likely represent biomarkers for antiangiogenic therapy (Hayano et al., 2015). When analysing contrast-enhanced CT, mean perfusion peak (MPP) parameter correlated with necrosis on histology and allowed for differentiating (with 85 % sensitivity and 71 % specificity) responders from non-responders. Therefore, the authors suggested that MPP might be chosen as the best predictor for treatment response of STS, as shown in other non-sarcoma histologies (Goh et al., 2011; Ravanelli et al., 2013).

4.3. ^{18}F -FDG positron emission tomography (^{18}F -FDG-PET)

Recent research has shown that changes in standardised uptake value (SUV) are more accurate than changes in tumour size or density (measured on contrast-enhanced CT) to predict pathological response to neoadjuvant chemotherapy. Conversely, evidence pertaining to radiation therapy or chemo-radiotherapy is still limited (Favinger et al., 2018; Evilevitch et al., 2008; Tateishi et al., 2011; Benz et al., 2008, 2009). Both early and late metabolic response measured by means of changes in SUV peaks were the only two predictors of survival in a prospective study on 78 patients (Herrmann et al., 2012). Similarly, the metabolic response on ^{18}F -FDG PET using PET Response Criteria in Solid Tumors (PERCIST) predicted PFS and time to local and distant progression after 2–4 cycles of neoadjuvant chemotherapy with regional hyperthermia in sarcoma patients. When the predictive performance of PERCIST was compared with RECIST 1.1 and Choi, PERCIST was superior in identifying responders (Fendler et al., 2015).

A recent meta-analysis showed that a ≥ 60 % decrease in FDG-uptake resulted in a sensitivity and specificity of 100 % and 71 % respectively for assessment of histopathologic response (Lim et al., 2019). Despite the promising results, PET cannot replace MRI for surgical planning as surgeons require highly accurate anatomic visualisation for procedural planning. PET and MRI thus have a synergistic value in such assessment, and either the diffusion of hybrid PET/MRI equipment as well as post-processing software for fusion imaging might ease its application (Gennaro et al., 2020; Cassarino et al., 2020). It is partly with this ambition that efforts are being made in implementing PET and MRI standardised protocols across multi-vendor platforms to ensure reliable quantification of imaging biomarkers in multicentre trials (deSouza et al., 2018; Boellaard et al., 2015; Winfield et al., 2016).

5. Future research

Unlike bone sarcoma, where the prognostic value of tumour necrosis induced by neoadjuvant chemotherapy has been extensively validated

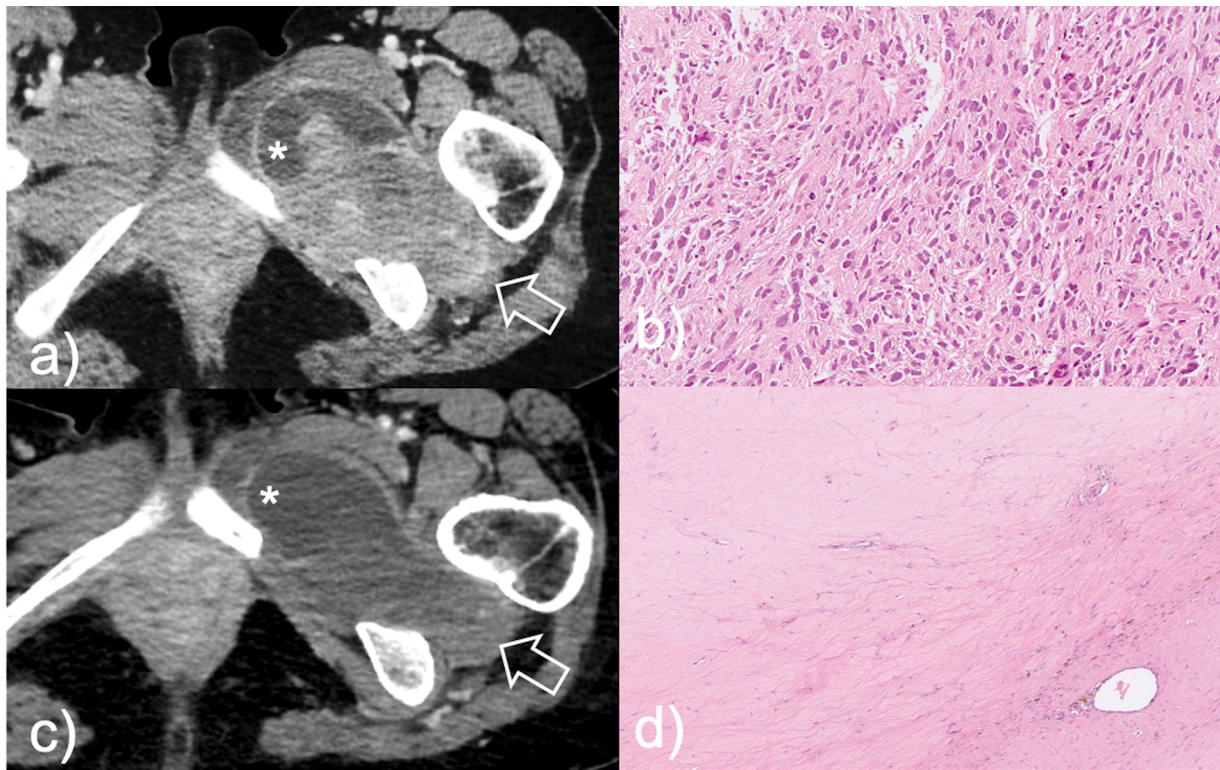


Fig. 5. a) Contrast-enhanced CT demonstrates a soft tissue mass occupying the left adductor compartment in a 37-year old female. The lesion is predominantly solid with some low density (arrow), likely representing cystic changes or necrosis (asterisk) b) Pre-treatment core-needle biopsy shows spindle and pleomorphic cells with 80 % Ki67 representing high-grade undifferentiated pleomorphic sarcoma c) Contrast enhanced CT post-chemo/radiotherapy (3 cycles of adriamycin-ifosfamide and 50 Gy/25 fractions with VMAT) shows no significant reduction in size and but there is a marked reduction in enhancement (arrow) and in the extent of the necrotic areas (asterisk); d) The surgical specimen reveals <1% viable tumor cells, isolated spindle and pleomorphic cells in a vast sclerotic stroma (<99 %); haemosiderin deposits are also present.

(Bielack et al., 2002; Picci et al., 1997), STS still lacks radiological and histological markers of tumour response and patient survival. In order to deliver personalised treatment strategies, clinicians demand from imaging a reproducible, objective assessment of the efficacy of local and systemic therapies, and they advocate imaging and histologic features capable of predicting the clinical behaviour of STS. Without a robust understanding of radiology- pathology correlations, developing solid imaging biomarkers is challenging. For this purpose, innovative techniques such as radiomics and radiogenomics complementing conventional imaging have arisen, where quantitative radiomic features derived from routine images are linked to tumour biology (Bodalal et al., 2019; Cromb e et al., 2020). While still in its infancy, radiogenomics promises to address many of the pitfalls of current biological profiling by analysing the full tumour burden of a patient (to account for intra-tumoural and intertumoural heterogeneity). Additionally, once radiogenomic models are better fine-tuned, longitudinal imaging will be the new frontier of AI imaging research. Both diagnostic disciplines, pathology and radiology, are likely to play complementary roles (Jha and Topol, 2016). In theory, the complementary data from these two diagnostic modalities can be integrated into a single AI algorithm with the potential for higher predictive performance (Bodalal et al., 2018; Chibon et al., 2019).

The recent introduction of the concept of “imaging habitats” to improve radiological mapping of different microenvironments has brought radiologists and pathologists closer in the characterisation of tumour heterogeneity (Blackledge et al., 2019; Gatenby et al., 2013). In-vivo models can also aid in the comprehension of radiologic-pathology treatment-related tissue changes, especially in the case of innovative imaging modalities (Shapiro et al., 2018; Sigal and Sebro, 2018). The huge potential of artificial intelligence models seems

promising in identifying features hidden in radiological images, possibly predicting or complementing histological and molecular information or even outcomes. Along with tissue and imaging biomarkers, correlations with serum/molecular biomarkers and circulating tumour cells/DNA might also be further explored (Aggerholm-Pedersen et al., 2019; Wang et al., 2019; Kambadakone et al., 2015).

6. Conclusion

Imaging can depict various changes induced by neoadjuvant therapies in the treatment of STS, but these changes are poorly correlated with histology and clinical outcomes. Evidence so far shows that Choi criteria, along with functional and molecular imaging information (DWI, DCE-MRI and ¹⁸F-FDG-PET), outperform commonly used response criteria like RECIST 1.1 in identifying good responders based on necrosis and viability of tumour cells after treatment. The introduction of advanced quantitative image analysis such as unsupervised clustering, texture analysis and dynamic post-contrast imaging appears promising in improving non-invasive evaluation of treatment response in soft tissue sarcoma, therefore complementing histological information. Histological and radiological response criteria advocated by the EORTC-STBSG and EORTC Imaging Group should shed light on the biological efficacy of neoadjuvant therapies through future studies. Such studies would not only help radiologists identify prognostic and early response predictive imaging biomarkers, but also help us understand how far imaging can interrogate cancer biology in the description of tumour response. This will allow for improved personalised treatment within the heterogeneous group of STS patients.

Declaration of Competing Interest

The authors report no declarations of interest

References

- Aggerholm-Pedersen, N., Maretty-Kongstad, K., Keller, J., Safwat, A., 2019. Serum biomarkers as prognostic factors for metastatic sarcoma. *Clin. Oncol. (R. Coll. Radiol.)* 31, 242–249. <https://doi.org/10.1016/j.clon.2019.01.011>.
- Alic, L., van Vliet, M., Wielopolski, P.A., ten Hagen, T.L.M., van Dijke, C.F., Niessen, W. J., et al., 2013. Regional heterogeneity changes in DCE-MRI as response to isolated limb perfusion in experimental soft-tissue sarcomas. *Contrast Media Mol. Imaging* 8, 340–349. <https://doi.org/10.1002/cmml.1528>.
- Baur, A., Stähler, A., Wendtner, C.M., Arbogast, S., Rahman, S.A., Santl, M., et al., 2003. MR-imaging changes of musculoskeletal soft-tissue sarcomas associated with neoadjuvant chemotherapy and hyperthermia. *Int. J. Hyperthermia* 19, 391–401. <https://doi.org/10.1080/0265673021000058366>.
- Beltran, J., Simon, D.C., Katz, W., Weis, L.D., 1987. Increased MR signal intensity in skeletal muscle adjacent to malignant tumors: pathologic correlation and clinical relevance. *Radiology* 162, 251–255. <https://doi.org/10.1148/radiology.162.1.3786772>.
- Benz, M.R., Allen-Auerbach, M.S., Eilber, F.C., Chen, H.J.J., Dry, S., Phelps, M.E., et al., 2008. Combined assessment of metabolic and volumetric changes for assessment of tumor response in patients with soft-tissue sarcomas. *J. Nucl. Med.* 49, 1579–1584. <https://doi.org/10.2967/jnumed.108.053694>.
- Benz, M.R., Czernin, J., Allen-Auerbach, M.S., Tap, W.D., Dry, S.M., Elashoff, D., et al., 2009. FDG-PET/CT imaging predicts histopathologic treatment responses after the initial cycle of neoadjuvant chemotherapy in high-grade soft-tissue sarcomas. *Clin. Cancer Res.* 15, 2856–2863. <https://doi.org/10.1158/1078-0432.CCR-08-2537>.
- Betgen, A., Haas, R.L.M., Sonke, J.-J., 2013. Volume changes in soft tissue sarcomas during preoperative radiotherapy of extremities evaluated using cone-beam CT. *J. Radiat. Oncol.* 2, 55–62. <https://doi.org/10.1007/s13566-012-0085-0>.
- Bielack, S.S., Kempf-Bielack, B., Delling, G., Exner, G.U., Flege, S., Helmke, K., et al., 2002. Prognostic factors in high-grade osteosarcoma of the extremities or trunk: an analysis of 1,702 patients treated on neoadjuvant cooperative osteosarcoma study group protocols. *J. Clin. Oncol.* 20, 776–790. <https://doi.org/10.1200/JCO.2002.20.3.776>.
- Blackledge, M.D., Winfield, J.M., Miah, A., Strauss, D., Thway, K., Morgan, V.A., et al., 2019. Supervised machine-learning enables segmentation and evaluation of heterogeneous post-treatment changes in multi-parametric MRI of soft-tissue sarcoma. *Front. Oncol.* 9, 941. <https://doi.org/10.3389/fonc.2019.00941>.
- Bodalal, Z., Trebeschi, S., Beets-Tan, R., 2018. Radiomics: a critical step towards integrated healthcare. *Insights Imaging* 9, 911–914. <https://doi.org/10.1007/s13244-018-0669-3>.
- Bodalal, Z., Trebeschi, S., Nguyen-Kim, T.D.L., Schats, W., Beets-Tan, R., 2019. Radiomics: bridging imaging and genomics. *Abdom. Radiol. (NY)* 44, 1960–1984. <https://doi.org/10.1007/s00261-019-02028-w>.
- Boellaard, R., Delgado-Bolton, R., Oyen, W.J.G., Giammarile, F., Tatsch, K., Eschner, W., et al., 2015. FDG PET/CT: EANM procedure guidelines for tumour imaging: version 2.0. *Eur. J. Nucl. Med. Mol. Imaging* 42, 328–354. <https://doi.org/10.1007/s00259-014-2961-x>.
- Callegaro, D., Miceli, R., Bonvalot, S., Ferguson, P., Strauss, D.C., Levy, A., et al., 2016. Development and external validation of two nomograms to predict overall survival and occurrence of distant metastases in adults after surgical resection of localised soft-tissue sarcomas of the extremities: a retrospective analysis. *Lancet Oncol.* 17, 671–680. [https://doi.org/10.1016/S1470-2045\(16\)00010-3](https://doi.org/10.1016/S1470-2045(16)00010-3).
- Callegaro, D., Miceli, R., Mariani, L., Raut, C.P., Gronchi, A., 2017. Soft tissue sarcoma nomograms and their incorporation into practice. *Cancer* 123, 2802–2820. <https://doi.org/10.1002/cncr.30721>.
- Canter, R.J., Martinez, S.R., Tamurian, R.M., Wilton, M., Li, C.-S., Ryu, J., et al., 2010. Radiographic and histologic response to neoadjuvant radiotherapy in patients with soft tissue sarcoma. *Ann. Surg. Oncol.* 17, 2578–2584. <https://doi.org/10.1245/s10434-010-1156-3>.
- Canter, R.J., Borys, D., Olusanya, A., Li, C.-S., Lee, L.-Y., Boutin, R.D., et al., 2014. Phase I trial of neoadjuvant conformal radiotherapy plus sorafenib for patients with locally advanced soft tissue sarcoma of the extremity. *Ann. Surg. Oncol.* 21, 1616–1623. <https://doi.org/10.1245/s10434-014-3543-7>.
- Cassarino, G., Evangelista, L., Giraudo, C., Capizzi, A., Carretta, G., Zucchetta, P., et al., 2020. 18F-FDG PET/MRI in adult sarcomas. *Clin. Transl. Imaging.* <https://doi.org/10.1007/s40336-020-00395-9>.
- Chen, Y., Yang, Y., Wang, C., Shi, Y., 2013. Radiographic response to neoadjuvant therapy and its impact on scope of surgery and prognosis in stage IIB/III soft tissue sarcomas. *BMC Cancer* 13, 591. <https://doi.org/10.1186/1471-2407-13-591>.
- Chibon, F., Lesluyes, T., Valentin, T., Le Guellec, S., 2019. CINSARC signature as a prognostic marker for clinical outcome in sarcomas and beyond. *Genes Chromosomes Cancer* 58, 124–129. <https://doi.org/10.1002/gcc.22703>.
- Choi, H., Charnsangavej, C., Faria, S.C., Macapinlac, H.A., Burgess, M.A., Patel, S.R., et al., 2007. Correlation of computed tomography and positron emission tomography in patients with metastatic gastrointestinal stromal tumor treated at a single institution with imatinib mesylate: proposal of new computed tomography response criteria. *J. Clin. Oncol.* 25, 1753–1759. <https://doi.org/10.1200/JCO.2006.07.3049>.
- Chung, P.W.M., Deheshi, B.M., Ferguson, P.C., Wunder, J.S., Griffin, A.M., Catton, C.N., et al., 2009. Radiosensitivity translates into excellent local control in extremity myxoid liposarcoma: a comparison with other soft tissue sarcomas. *Cancer* 115, 3254–3261. <https://doi.org/10.1002/cncr.24375>.
- Corino, V.D.A., Montini, E., Messina, A., Casali, P.G., Gronchi, A., Marchiano, A., et al., 2018. Radiomic analysis of soft tissues sarcomas can distinguish intermediate from high-grade lesions. *J. Magn. Reson. Imaging* 47, 829–840. <https://doi.org/10.1002/jmri.25791>.
- Costa, F.M., Martins, P.H., Canella, C., Lopes, F.P.P.L., 2018. Multiparametric MR imaging of soft tissue tumors and pseudotumors. *Magn. Reson. Imaging Clin. N. Am.* 26, 543–558. <https://doi.org/10.1016/j.mric.2018.06.009>.
- Cousin, S., Crombe, A., Stoeckle, E., Brouste, V., Le Loarer, F., Lucchesi, C., et al., 2017. Clinical, radiological and genetic features, associated with the histopathologic response to neoadjuvant chemotherapy (NAC) and outcomes in locally advanced soft tissue sarcoma (STS) patients (pts). *J. Clin. Oncol.* 35 https://doi.org/10.1200/JCO.2017.35.15_suppl.11014, 11014–11014.
- Crombé, A., Le Loarer, F., Stoeckle, E., Cousin, S., Michot, A., Italiano, A., et al., 2018. MRI assessment of surrounding tissues in soft-tissue sarcoma during neoadjuvant chemotherapy can help predicting response and prognosis. *Eur. J. Radiol.* 109, 178–187. <https://doi.org/10.1016/j.ejrad.2018.11.004>.
- Crombé, A., Le Loarer, F., Cornelis, F., Stoeckle, E., Buy, X., Cousin, S., et al., 2019a. High-grade soft-tissue sarcoma: optimizing injection improves MRI evaluation of tumor response. *Eur. Radiol.* 29, 545–555. <https://doi.org/10.1007/s00330-018-5635-4>.
- Crombé, A., Marcellin, P.-J., Buy, X., Stoeckle, E., Brouste, V., Italiano, A., et al., 2019b. Soft-tissue sarcomas: assessment of MRI features correlating with histologic grade and patient outcome. *Radiology* 291, 710–721. <https://doi.org/10.1148/radiol.2019181659>.
- Crombé, A., Périer, C., Kind, M., De Senneville, B.D., Le Loarer, F., Italiano, A., et al., 2019c. T2-based MRI Delta-radiomics improve response prediction in soft-tissue sarcomas treated by neoadjuvant chemotherapy. *J. Magn. Reson. Imaging* 50, 497–510. <https://doi.org/10.1002/jmri.26589>.
- Crombé, A., Fadli, D., Italiano, A., Saut, O., Buy, X., Kind, M., 2020. Systematic review of sarcomas radiomics studies: Bridging the gap between concepts and clinical applications? *Eur. J. Radiol.* 132, 109283 <https://doi.org/10.1016/j.ejrad.2020.109283>.
- D'Ambrosio, L., Touati, N., Blay, J.-Y., Grignani, G., Flippot, R., Czarnecka, A.M., et al., 2020. Doxorubicin plus dacarbazine, doxorubicin plus ifosfamide, or doxorubicin alone as a first-line treatment for advanced leiomyosarcoma: A propensity score matching analysis from the European Organization for Research and Treatment of Cancer Soft Tissue and Bone Sarcoma Group. *Cancer.* <https://doi.org/10.1002/cncr.32795>.
- deSouza, N.M., Winfield, J.M., Waterton, J.C., Weller, A., Papoutsaki, M.V., Doran, S.J., et al., 2018. Implementing diffusion-weighted MRI for body imaging in prospective multicentre trials: current considerations and future perspectives. *Eur. Radiol.* 28, 1118–1131. <https://doi.org/10.1007/s00330-017-4972-z>.
- Diwanji, T., Chang, Y.C., Subhawong, T.K., Wilky, B., Yechieli, R., Asher, D., et al., 2019. Prediction of treatment efficacy in Soft tissue sarcomas through analysis of habitat heterogeneity in Mpmri. *Int. J. Radiat. Oncol.* 105, E738. <https://doi.org/10.1016/j.ijrobp.2019.06.838>.
- Donahue, T.R., Kattan, M.W., Nelson, S.D., Tap, W.D., Eilber, F.R., Eilber, F.C., 2010. Evaluation of neoadjuvant therapy and histopathologic response in primary, high-grade retroperitoneal sarcomas using the sarcoma nomogram. *Cancer* 116, 3883–3891. <https://doi.org/10.1002/cncr.25271>.
- Dudeck, O., Zeile, M., Pink, D., Pech, M., Tunn, P.-U., Reichardt, P., et al., 2008. Diffusion-weighted magnetic resonance imaging allows monitoring of anticancer treatment effects in patients with soft-tissue sarcomas. *J. Magn. Reson. Imaging* 27, 1109–1113. <https://doi.org/10.1002/jmri.21358>.
- Eilber, F.C., Rosen, G., Eckardt, J., Forscher, C., Nelson, S.D., Selch, M., et al., 2001. Treatment-induced pathologic necrosis: a predictor of local recurrence and survival in patients receiving neoadjuvant therapy for high-grade extremity soft tissue sarcomas. *J. Clin. Oncol.* 19, 3203–3209. <https://doi.org/10.1200/JCO.2001.19.13.3203>.
- Eisenhauer, E.A., Therasse, P., Bogaerts, J., Schwartz, L.H., Sargent, D., Ford, R., et al., 2009. New response evaluation criteria in solid tumours: revised RECIST guideline (version 1.1). *Eur. J. Cancer* 45, 228–247. <https://doi.org/10.1016/j.ejca.2008.07.1026>.
- Esser, M., Kloth, C., Thaiss, W.M., Reinert, C.P., Fritz, J., Kopp, H.-G., et al., 2018. CT-response patterns and the role of CT-textural features in inoperable abdominal/retroperitoneal soft tissue sarcomas treated with trabectedin. *Eur. J. Radiol.* 107, 175–182. <https://doi.org/10.1016/j.ejrad.2018.09.006>.
- Evilevitch, V., Weber, W.A., Tap, W.D., Allen-Auerbach, M., Chow, K., Nelson, S.D., et al., 2008. Reduction of glucose metabolic activity is more accurate than change in size at predicting histopathologic response to neoadjuvant therapy in high-grade soft-tissue sarcomas. *Clin. Cancer Res.* 14, 715–720. <https://doi.org/10.1158/1078-0432.CCR-07-1762>.
- Farhizadeh, H., Chaudhury, B., Zhou, M., Goldof, D.B., Hall, L.O., Gatenby, R.A., et al., 2015a. Prediction of treatment outcome in soft tissue sarcoma based on radiologically defined habitats. In: Hadjiiski, L.M., Tourassi, G.D. (Eds.), *Medical Imaging 2015: Computer-Aided Diagnosis*, Vol. 9414. SPIE, p. 94141U. <https://doi.org/10.1117/12.2082324>.
- Farhizadeh, H., Goldof, D.B., Hall, L.O., Gatenby, R.A., Gillies, R.J., Raghavan, M., 2015b. Texture feature analysis to predict metastatic and necrotic soft tissue sarcomas. In: 2015 IEEE International Conference on Systems, Man, and Cybernetics. IEEE, pp. 2798–2802. <https://doi.org/10.1109/SMC.2015.488>.
- Favinger, J.L., Hippe, D.S., Davidson, D.J., Elojeimy, S., Roth, E.S., Lindberg, A.W., et al., 2018. Soft tissue sarcoma response to two cycles of neoadjuvant chemotherapy: a multireader analysis of MRI findings and agreement with RECIST criteria and change

- in SUVmax. *Acad. Radiol.* 25, 470–475. <https://doi.org/10.1016/j.acra.2017.10.013>.
- Fendler, W.P., Lehmann, M., Todica, A., Herrmann, K., Knösel, T., Angele, M.K., et al., 2015. PET response criteria in solid tumors predicts progression-free survival and time to local or distant progression after chemotherapy with regional hyperthermia for soft-tissue sarcoma. *J. Nucl. Med.* 56, 530–537. <https://doi.org/10.2967/jnumed.114.152462>.
- Fields, B.K.K., Hwang, D., Cen, S., Desai, B., Gulati, M., Hu, J., et al., 2020. Quantitative magnetic resonance imaging (q-MRI) for the assessment of soft-tissue sarcoma treatment response: a narrative case review of technique development. *Clin. Imaging* 63, 83–93. <https://doi.org/10.1016/j.clinimag.2020.02.016>.
- Fisher, S.M., Joodi, R., Madhuranthakam, A.J., Öz, O.K., Sharma, R., Chhabra, A., 2016. Current utilities of imaging in grading musculoskeletal soft tissue sarcomas. *Eur. J. Radiol.* 85, 1336–1344. <https://doi.org/10.1016/j.ejrad.2016.05.003>.
- Gatenby, R.A., Grove, O., Gillies, R.J., 2013. Quantitative imaging in cancer evolution and ecology. *Radiology* 269, 8–15. <https://doi.org/10.1148/radiol.13122697>.
- Gennaro, N., Marrari, A., Renne, S.L., Cananzi, F.C.M., Quagliuolo, V.L., Di Brina, L., et al., 2020. Multimodality imaging of adult rhabdomyosarcoma: the added value of hybrid imaging. *Br. J. Radiol.* 93 <https://doi.org/10.1259/bjr.20200250>, 20200250.
- Goh, V., Ganeshan, B., Nathan, P., Juttla, J.K., Vinayan, A., Miles, K.A., 2011. Assessment of response to tyrosine kinase inhibitors in metastatic renal cell cancer: CT texture as a predictive biomarker. *Radiology* 261, 165–171. <https://doi.org/10.1148/radiol.11110264>.
- Gronchi, A., Hindi, N., Cruz, J., Blay, J.Y., Lopez-Pousa, A., Italiano, A., et al., 2019. Trabectedin and RAdiotherapy in Soft tissue sarcoma (TRASTS): results of a phase I study in myxoid liposarcoma from spanish (GEIS), italian (ISG), french (FSG) sarcoma groups. *EclinicalMedicine* 9, 35–43. <https://doi.org/10.1016/j.eclinm.2019.03.007>.
- Grueneisen, J., Schaarschmidt, B., Demircioglu, A., Chodyla, M., Martin, O., Bertram, S., et al., 2019. 18F-FDG PET/MRI for therapy response assessment of isolated limb perfusion in patients with soft-tissue sarcomas. *J. Nucl. Med.* 60, 1537–1542. <https://doi.org/10.2967/jnumed.119.226761>.
- Grünwald, V., Litière, S., Young, R., Messiou, C., Lia, M., Wardelmann, E., et al., 2016. Absence of progression, not extent of tumour shrinkage, defines prognosis in soft-tissue sarcoma - an analysis of the EORTC 62012 study of the EORTC STBSG. *Eur. J. Cancer* 64, 44–51. <https://doi.org/10.1016/j.ejca.2016.05.023>.
- Gui, C., Morris, C.D., Meyer, C.F., Levin, A.S., Fraccia, D.A., Deville, C., et al., 2019. Characterization and predictive value of volume changes of extremity and pelvis soft tissue sarcomas during radiation therapy prior to definitive wide excision. *Radiat. Oncol. J.* 37, 117–126. <https://doi.org/10.3857/roj.2018.00549>.
- Haas, Rick L., 2018. Preoperative radiotherapy in soft tissue sarcoma: from general guidelines to personalized medicine. *Chin. Clin. Oncol.*
- Hanna, S.L., Fletcher, B.D., Parham, D.M., Bugg, M.F., 1991. Muscle edema in musculoskeletal tumors: MR imaging characteristics and clinical significance. *J. Magn. Reson. Imaging* 1, 441–449. <https://doi.org/10.1002/jmri.1880010408>.
- Hartman, M., Silverman, J., Spruill, L., Hill, J., 2016. Radiologic-pathologic correlation - an advanced fourth-year elective: how we do it. *Acad. Radiol.* 23, 889–893. <https://doi.org/10.1016/j.acra.2016.01.019>.
- Hayano, K., Tian, F., Kambadakone, A.R., Yoon, S.S., Duda, D.G., Ganeshan, B., et al., 2015. Texture analysis of non-contrast-enhanced computed tomography for assessing angiogenesis and survival of Soft tissue sarcoma. *J. Comput. Assist. Tomogr.* 39, 607–612. <https://doi.org/10.1097/RCT.0000000000000239>.
- Herrmann, K., Benz, M.R., Czernin, J., Allen-Auerbach, M.S., Tap, W.D., Dry, S.M., et al., 2012. 18F-FDG-PET/CT Imaging as an early survival predictor in patients with primary high-grade soft tissue sarcomas undergoing neoadjuvant therapy. *Clin. Cancer Res.* 18, 2024–2031. <https://doi.org/10.1158/1078-0432.CCR-11-2139>.
- Hong, J.H., Jee, W.-H., Jung, C.-K., Chung, Y.-G., 2020. Tumor grade in soft-tissue sarcoma: prediction with magnetic resonance imaging texture analysis. *Medicine (Baltimore)* 99, e20880. <https://doi.org/10.1097/MD.00000000000020880>.
- Huang, W., Beckett, B.R., Tudorica, A., Meyer, J.M., Afzal, A., Chen, Y., et al., 2016. Evaluation of Soft tissue sarcoma response to preoperative chemoradiotherapy using dynamic contrast-enhanced magnetic resonance imaging. *Tomography* 2, 308–316. <https://doi.org/10.18383/j.tom.2016.00202>.
- Jakob, J., Simeonova, A., Kasper, B., Ronellenfitsch, U., Rauch, G., Wenz, F., et al., 2016. Combined sunitinib and radiation therapy for preoperative treatment of soft tissue sarcoma: results of a phase I trial of the German interdisciplinary sarcoma group (GISG-03). *Radiat. Oncol.* 11, 77. <https://doi.org/10.1186/s13014-016-0654-2>.
- Jha, S., Topol, E.J., 2016. Adapting to artificial intelligence: radiologists and pathologists as information specialists. *JAMA* 316, 2353–2354. <https://doi.org/10.1001/jama.2016.17438>.
- Judson, I., Verweij, J., Gelderblom, H., Hartmann, J.T., Schöffski, P., Blay, J.-Y., et al., 2014. Doxorubicin alone versus intensified doxorubicin plus ifosfamide for first-line treatment of advanced or metastatic soft-tissue sarcoma: a randomised controlled phase 3 trial. *Lancet Oncol.* 15, 415–423. [https://doi.org/10.1016/S1470-2045\(14\)70063-4](https://doi.org/10.1016/S1470-2045(14)70063-4).
- Kambadakone, A., Yoon, S.S., Kim, T.-M., Karl, D.L., Duda, D.G., DeLaney, T.F., et al., 2015. CT perfusion as an imaging biomarker in monitoring response to neoadjuvant bevacizumab and radiation in soft-tissue sarcomas: comparison with tumor morphology, circulating and tumor biomarkers, and gene expression. *AJR Am. J. Roentgenol.* 204, W11–8. <https://doi.org/10.2214/AJR.13.12412>.
- Kandel, R., Coakley, N., Werier, J., Engel, J., Ghert, M., Verma, S., et al., 2013. Surgical margins and handling of soft-tissue sarcoma in extremities: a clinical practice guideline. *Curr. Oncol.* 20, e247–54. <https://doi.org/10.3747/co.20.1308>.
- Kosela-Paterczyk, H., Szacht, M., Morysiński, T., Ługowska, I., Dziejewski, W., Falkowski, S., et al., 2014. Preoperative hypofractionated radiotherapy in the treatment of localized soft tissue sarcomas. *Eur. J. Surg. Oncol.* 40, 1641–1647. <https://doi.org/10.1016/j.ejso.2014.05.016>.
- Lansu, J., Bovée, J., Braam, P., van Boven, H., Flucke, U., Bonenkamp, J.J., et al., 2020. Dose reduction of preoperative radiotherapy in myxoid liposarcoma: a nonrandomized controlled trial. *JAMA Oncol.* <https://doi.org/10.1001/jamaoncol.2020.5865>.
- Larrier, N.A., Czito, B.G., Kirsch, D.G., 2016. Radiation therapy for soft tissue sarcoma: indications and controversies for neoadjuvant therapy, adjuvant therapy, intraoperative radiation therapy, and brachytherapy. *Surg. Oncol. Clin. N. Am.* 25, 841–860. <https://doi.org/10.1016/j.soc.2016.05.012>.
- le Grange, F., Cassoni, A.M., Seddon, B.M., 2014. Tumour volume changes following preoperative radiotherapy in borderline resectable limb and trunk soft tissue sarcoma. *Eur. J. Surg. Oncol.* 40, 394–401. <https://doi.org/10.1016/j.ejso.2014.01.011>.
- Lehane, C., Ho, F., Thompson, S.R., Links, D., Lewis, C., Smees, R., et al., 2016. Neoadjuvant chemoradiation (modified Eilber protocol) versus adjuvant radiotherapy in the treatment of extremity soft tissue sarcoma. *J. Med. Imaging Radiat. Oncol.* 60, 539–544. <https://doi.org/10.1111/1754-9485.12442>.
- Lim, H.-J., Johnny Ong, C.-A., Tan, J.W.-S., Ching Teo, M.C., 2019. Utility of positron emission tomography/computed tomography (PET/CT) imaging in the evaluation of sarcomas: a systematic review. *Crit. Rev. Oncol. Hematol.* 143, 1–13. <https://doi.org/10.1016/j.critrevonc.2019.07.002>.
- Liu, Q.-Y., Li, H.-G., Chen, J.-Y., Liang, B.-L., 2008. [Correlation of MRI features to histopathologic grade of soft tissue sarcoma]. *Ai Zheng* 27, 856–860.
- Look Hong, N.J., Hornicek, F.J., Harmon, D.C., Choy, E., Chen, Y.-L., Yoon, S.S., et al., 2013. Neoadjuvant chemoradiotherapy for patients with high-risk extremity and truncal sarcomas: a 10-year single institution retrospective study. *Eur. J. Cancer* 49, 875–883. <https://doi.org/10.1016/j.ejca.2012.10.002>.
- Lucas, D.R., Kshirsagar, M.P., Biermann, J.S., Hamre, M.R., Thomas, D.G., Schuetz, S. M., et al., 2008. Histologic alterations from neoadjuvant chemotherapy in high-grade extremity soft tissue sarcoma: clinicopathological correlation. *Oncologist* 13, 451–458. <https://doi.org/10.1634/theoncologist.2007-0220>.
- MacVicar, A.D., Olliff, J.F., Pringle, J., Pinkerton, C.R., Husband, J.E., 1992. Ewing sarcoma: MR imaging of chemotherapy-induced changes with histologic correlation. *Radiology* 184, 859–864. <https://doi.org/10.1148/radiology.184.3.1509080>.
- Marrari, A., Bertuzzi, A., Bozzarelli, S., Gennaro, N., Giordano, L., Quagliuolo, V., et al., 2020. Activity of regorafenib in advanced pretreated soft tissue sarcoma: results of a single-center phase II study. *Medicine (Baltimore)* 99, e20719. <https://doi.org/10.1097/MD.00000000000020719>.
- Mcaddy, N.C., Hallin, M., Strauss, D., Smith, M., Hayes, A., Yusuf, S., et al., 2020. CT imaging improves histopathological grading of retroperitoneal leiomyosarcomas. *Eur. J. Surg. Oncol.* 46, 288–292. <https://doi.org/10.1016/j.ejso.2019.10.007>.
- Menendez, L.R., Ahlmann, E.R., Savage, K., Cluck, M., Fedenko, A.N., 2007. Tumor necrosis has no prognostic value in neoadjuvant chemotherapy for soft tissue sarcoma. *Clin. Orthop. Relat. Res.* 455, 219–224. <https://doi.org/10.1097/01.blo.0000238864.69486.59>.
- Messiou, C., Bonvalot, S., Gronchi, A., Vanel, D., Meyer, M., Robinson, P., et al., 2016. Evaluation of response after pre-operative radiotherapy in soft tissue sarcomas; the European Organisation for Research and Treatment of Cancer-Soft Tissue and Bone Sarcoma Group (EORTC-STBSG) and Imaging Group recommendations for radiological examination and reporting with an emphasis on magnetic resonance imaging. *Eur. J. Cancer* 56, 37–44. <https://doi.org/10.1016/j.ejca.2015.12.008>.
- Meyer, J.M., Perlewitz, K.S., Hayden, J.B., Doung, Y.-C., Hung, A.Y., Vetto, J.T., et al., 2013. Phase I trial of preoperative chemoradiation plus sorafenib for high-risk extremity soft tissue sarcomas with dynamic contrast-enhanced MRI correlates. *Clin. Cancer Res.* 19, 6902–6911. <https://doi.org/10.1158/1078-0432.CCR-13-1594>.
- Miki, Y., Ngan, S., Clark, J.C.M., Akiyama, T., Choong, P.F.M., 2010. The significance of size change of soft tissue sarcoma during preoperative radiotherapy. *Eur. J. Surg. Oncol.* 36, 678–683. <https://doi.org/10.1016/j.ejso.2010.05.021>.
- Monksy, W.L., Jin, B., Molloy, C., Canter, R.J., Li, C.S., Lin, T.C., et al., 2012. Semi-automated volumetric quantification of tumor necrosis in soft tissue sarcoma using contrast-enhanced MRI. *Anticancer Res.* 32, 4951–4961.
- Mullen, J.T., Hornicek, F.J., Harmon, D.C., Raskin, K.A., Chen, Y.-L., Szymonifka, J., et al., 2014. Prognostic significance of treatment-induced pathologic necrosis in extremity and truncal soft tissue sarcoma after neoadjuvant chemoradiotherapy. *Cancer* 120, 3676–3682. <https://doi.org/10.1002/cncr.28945>.
- Nakamura, T., Matsumine, A., Matsubara, T., Asanuma, K., Yada, Y., Hagi, T., et al., 2017. Infiltrative tumor growth patterns on magnetic resonance imaging associated with systemic inflammation and oncological outcome in patients with high-grade soft-tissue sarcoma. *PLoS One* 12, e0181787. <https://doi.org/10.1371/journal.pone.0181787>.
- Napel, S., Mu, W., Jardim-Perassi, B.V., Aerts, H.J.W.L., Gillies, R.J., 2018. Quantitative imaging of cancer in the postgenomic era: Radio(geno)omics, deep learning, and habitats. *Cancer* 124, 4633–4649. <https://doi.org/10.1002/cncr.31630>.
- O'Connor, J.P.B., Tofts, P.S., Miles, K.A., Parkes, L.M., Thompson, G., Jackson, A., 2011. Dynamic contrast-enhanced imaging techniques: CT and MRI. *Br. J. Radiol.* 84 (2), S112–20. <https://doi.org/10.1259/bjr/55166688>.
- Pasquali, S., Gronchi, A., 2017. Neoadjuvant chemotherapy in soft tissue sarcomas: latest evidence and clinical implications. *Ther. Adv. Med. Oncol.* 9, 415–429. <https://doi.org/10.1177/1758834017705588>.
- Peeken, J.C., Bernhofer, M., Spraker, M.B., Pfeiffer, D., Devecka, M., Thamer, A., et al., 2019. CT-based radiomic features predict tumor grading and have prognostic value in patients with soft tissue sarcomas treated with neoadjuvant radiation therapy. *Radiother. Oncol.* 135, 187–196. <https://doi.org/10.1016/j.radonc.2019.01.004>.
- Picci, P., Böhlring, T., Bacci, G., Ferrari, S., Sangiorgi, L., Mercuri, M., et al., 1997. Chemotherapy-induced tumor necrosis as a prognostic factor in localized Ewing's

- sarcoma of the extremities. *J. Clin. Oncol.* 15, 1553–1559. <https://doi.org/10.1200/JCO.1997.15.4.1553>.
- Preda, A., Wielopolski, P.A., Ten Hagen, T.L.M., van Vliet, M., Veerland, J.F., Ambagtsheer, G., et al., 2004. Dynamic contrast-enhanced MRI using macromolecular contrast media for monitoring the response to isolated limb perfusion in experimental soft-tissue sarcomas. *MAGMA* 17, 296–302. <https://doi.org/10.1007/s10334-004-0050-z>.
- Raghavan, M., 2017. Conventional modalities and novel, emerging imaging techniques for musculoskeletal tumors. *Cancer Control* 24, 161–171. <https://doi.org/10.1177/107327481702400208>.
- Ravanelli, M., Farina, D., Morassi, M., Roca, E., Cavalleri, G., Tassi, G., et al., 2013. Texture analysis of advanced non-small cell lung cancer (NSCLC) on contrast-enhanced computed tomography: prediction of the response to the first-line chemotherapy. *Eur. Radiol.* 23, 3450–3455. <https://doi.org/10.1007/s00330-013-2965-0>.
- Roberge, D., Skamene, T., Nahal, A., Turcotte, R.E., Powell, T., Freeman, C., 2010. Radiological and pathological response following pre-operative radiotherapy for soft-tissue sarcoma. *Radiother. Oncol.* 97, 404–407. <https://doi.org/10.1016/j.radonc.2010.10.007>.
- Rothermundt, C., Fischer, G.F., Bauer, S., Blay, J.-Y., Grünwald, V., Italiano, A., et al., 2018. Pre- and postoperative chemotherapy in localized extremity Soft tissue sarcoma: a european organization for research and treatment of Cancer expert survey. *Oncologist* 23, 461–467. <https://doi.org/10.1634/theoncologist.2017-0391>.
- Rubin, B.P., Cooper, K., Fletcher, C.D.M., Folpe, A.L., Gannon, F.H., Hunt, J.L., et al., 2010. Protocol for the examination of specimens from patients with tumors of soft tissue. *Arch. Pathol. Lab. Med.* 134, e31–9. <https://doi.org/10.1043/1543-2165-134.4.e31>.
- Sala, N., Robert, L., Lopez-Pousa, A., Quintana, M.J., Llauger, J., Bagaña, S., et al., 2010. Neoadjuvant treatment evaluation in soft tissue sarcomas (STS): looking for news in radiologic and pathologic response. *J. Clin. Oncol.* 28 https://doi.org/10.1200/jco.2010.28.15_suppl.e20509 e20509–e20509.
- Sala, E., Mema, E., Himoto, Y., Veeraraghavan, H., Brenton, J.D., Snyder, A., et al., 2017. Unravelling tumour heterogeneity using next-generation imaging: radiomics, radiogenomics, and habitat imaging. *Clin. Radiol.* 72, 3–10. <https://doi.org/10.1016/j.crad.2016.09.013>.
- Salduz, A., Alban, B., Valiyev, N., Özmen, E., İribaş, A., Ağaoğlu, F., et al., 2017. Neoadjuvant radiotherapy for myxoid liposarcomas: oncologic outcomes and histopathologic correlations. *Acta Orthop. Traumatol. Turc.* 51, 355–361. <https://doi.org/10.1016/j.aott.2017.03.009>.
- Schaefer, I.-M., Hornick, J.L., Barysaukas, C.M., Raut, C.P., Patel, S.A., Royce, T.J., et al., 2017. Histologic appearance after preoperative radiation therapy for Soft tissue sarcoma: assessment of the european organization for research and treatment of cancer-soft tissue and bone sarcoma group response score. *Int. J. Radiat. Oncol. Biol. Phys.* 98, 375–383. <https://doi.org/10.1016/j.ijrobp.2017.02.087>.
- Schillaci, O., Scimeca, M., Toschi, N., Bonfiglio, R., Urbano, N., Bonanno, E., 2019. Combining diagnostic imaging and pathology for improving diagnosis and prognosis of cancer. *Contrast Media Mol. Imaging* 2019, 9429761. <https://doi.org/10.1155/2019/9429761>.
- Schneider, N., Strauss, D.C., Smith, M.J., Miah, A.B., Zaidi, S., Benson, C., et al., 2017. The adequacy of core biopsy in the assessment of smooth muscle neoplasms of soft tissues: implications for treatment and prognosis. *Am. J. Surg. Pathol.* 41, 923–931. <https://doi.org/10.1097/PAS.0000000000000867>.
- Schuetze, S.M., Baker, L.H., Benjamin, R.S., Canetta, R., 2008. Selection of response criteria for clinical trials of sarcoma treatment. *Oncologist* 13 (Suppl 2), 32–40. <https://doi.org/10.1634/theoncologist.13-S2-32>.
- Shah, C., Wang, J., Mubako, T., Fisher, C., Thway, K., 2016. Gross examination and reporting of soft tissue tumours: evaluation of compliance with the UK Royal College of Pathologists soft tissue sarcoma dataset. *J. Clin. Pathol.* 69, 761–766. <https://doi.org/10.1136/jclinpath-2015-203495>.
- Shapiro, M.C., Tang, T., Dasgupta, A., Kurenbekova, L., Shuck, R., Gaber, M.W., et al., 2018. In vitro and in vivo characterization of a preclinical irradiation-adapted model for ewing sarcoma. *Int. J. Radiat. Oncol. Biol. Phys.* 101, 118–127. <https://doi.org/10.1016/j.ijrobp.2018.01.095>.
- Sigal, I.R., Sebros, R., 2018. Preclinical PET tracers for the evaluation of sarcomas: understanding tumor biology. *Am. J. Nucl. Med. Mol. Imaging* 8, 428–440.
- Soldatos, T., Ahlawat, S., Montgomery, E., Chalian, M., Jacobs, M.A., Fayad, L.M., 2016. Multiparametric assessment of treatment response in high-grade soft-tissue sarcomas with anatomic and functional MR imaging sequences. *Radiology* 278, 831–840. <https://doi.org/10.1148/radiol.2015142463>.
- Song, Y., Bruce, A.N., Fraker, D.L., Karakousis, G.C., 2019. Isolated limb perfusion and infusion in the treatment of melanoma and soft tissue sarcoma in the era of modern systemic therapies. *J. Surg. Oncol.* 120, 540–549. <https://doi.org/10.1002/jso.25600>.
- Spalek, M.J., Kozak, K., Czarnecka, A.M., Bartnik, E., Borkowska, A., Rutkowski, P., 2020. Neoadjuvant treatment options in soft tissue sarcomas. *Cancers (Basel)* 12. <https://doi.org/10.3390/cancers12082061>.
- Stacchiotti, S., Collini, P., Messina, A., Morosi, C., Barisella, M., Bertulli, R., et al., 2009. High-grade soft-tissue sarcomas: tumor response assessment-pilot study to assess the correlation between radiologic and pathologic response by using RECIST and Choi criteria. *Radiology* 251, 447–456. <https://doi.org/10.1148/radiol.2512081403>.
- Stacchiotti, S., Verderio, P., Messina, A., Morosi, C., Collini, P., Lombart-Bosch, A., et al., 2012. Tumor response assessment by modified Choi criteria in localized high-risk soft tissue sarcoma treated with chemotherapy. *Cancer* 118, 5857–5866. <https://doi.org/10.1002/cncr.27624>.
- Subbiah, V., Chuang, H.H., Gambhire, D., Kairemo, K., 2017. Defining clinical response criteria and early response criteria for precision oncology: current state-of-the-Art and future perspectives. *Diagnostics (Basel)* 7. <https://doi.org/10.3390/diagnostics7010010>.
- Taieb, S., Saada-Bouزيد, E., Tresch, E., Rycckewaert, T., Bompas, E., Italiano, A., et al., 2015. Comparison of response evaluation criteria in solid tumours and Choi criteria for response evaluation in patients with advanced soft tissue sarcoma treated with trabectedin: a retrospective analysis. *Eur. J. Cancer* 51, 202–209. <https://doi.org/10.1016/j.ejca.2014.11.008>.
- Tanaka, K., Ogawa, G., Mizusawa, J., Naka, N., Kawai, A., Takahashi, M., et al., 2018. Prospective comparison of various radiological response criteria and pathological response to preoperative chemotherapy and survival in operable high-grade soft tissue sarcomas in the Japan Clinical Oncology Group study JCOG0304. *World J. Surg. Oncol.* 16, 162. <https://doi.org/10.1186/s12957-018-1462-y>.
- Tateishi, U., Kawai, A., Chuman, H., Nakatani, F., Bepptu, Y., Seki, K., et al., 2011. PET/CT allows stratification of responders to neoadjuvant chemotherapy for high-grade sarcoma: a prospective study. *Clin. Nucl. Med.* 36, 526–532. <https://doi.org/10.1097/RLU.0b013e3182175856>.
- Tian, F., Hayano, K., Kambadakone, A.R., Sahani, D.V., 2015. Response assessment to neoadjuvant therapy in soft tissue sarcomas: using CT texture analysis in comparison to tumor size, density, and perfusion. *Abdom. Imaging* 40, 1705–1712. <https://doi.org/10.1007/s00261-014-0318-3>.
- Vayntrub, M., Taheri, N., Ahlmann, E.R., Yao, C., Fedenko, A.N., Allison, D.C., et al., 2015. Prognostic value of necrosis after neoadjuvant therapy for soft tissue sarcoma. *J. Surg. Oncol.* 111, 152–157. <https://doi.org/10.1002/jso.23775>.
- Wang, X., Liu, S., Zhao, X., Fang, E., Zhao, X., 2019. The value of C-reactive protein as an independent prognostic indicator for disease-specific survival in patients with soft tissue sarcoma: a meta-analysis. *PLoS One* 14, e0219215. <https://doi.org/10.1371/journal.pone.0219215>.
- Wardelmann, E., Haas, R.L., Bovée, J.V.M.G., Terrier, P., Lazar, A., Messiou, C., et al., 2016. Evaluation of response after neoadjuvant treatment in soft tissue sarcomas; the European Organization for Research and Treatment of Cancer-Soft Tissue and Bone Sarcoma Group (EORTC-STBSG) recommendations for pathological examination and reporting. *Eur. J. Cancer* 53, 84–95. <https://doi.org/10.1016/j.ejca.2015.09.021>.
- White, L.M., Wunder, J.S., Bell, R.S., O'Sullivan, B., Catton, C., Ferguson, P., et al., 2005. Histologic assessment of peritumoral edema in soft tissue sarcoma. *Int. J. Radiat. Oncol. Biol. Phys.* 61, 1439–1445. <https://doi.org/10.1016/j.ijrobp.2004.08.036>.
- WHO Classification of Tumours of Soft Tissue and Bone. Fourth Edition - WHO - OMS - n. d. <http://apps.who.int/bookorders/anglais/detart1.jsp?codlan=1&codcol=70&codcch=4005> (accessed February 3, 2019).
- Winfield, J.M., Collins, D.J., Priest, A.N., Quest, R.A., Glover, A., Hunter, S., et al., 2016. A framework for optimization of diffusion-weighted MRI protocols for large field-of-view abdominal-pelvic imaging in multicenter studies. *Med. Phys.* 43, 95. <https://doi.org/10.1118/1.4937789>.
- Winfield, J.M., Miah, A.B., Strauss, D., Thway, K., Collins, D.J., deSouza, N.M., et al., 2019. Utility of multi-parametric quantitative magnetic resonance imaging for characterization and radiotherapy response assessment in soft-tissue sarcomas and correlation with histopathology. *Front. Oncol.* 9, 280. <https://doi.org/10.3389/fonc.2019.00280>.
- Y-CC, Chang, Ackerstaff, E., Tschudi, Y., Jimenez, B., Foltz, W., Fisher, C., et al., 2017. Delineation of tumor habitats based on dynamic contrast enhanced MRI. *Sci. Rep.* 7, 9746. <https://doi.org/10.1038/s41598-017-09932-5>.
- Yoon, S.S., Duda, D.G., Karl, D.L., Kim, T.-M., Kambadakone, A.R., Chen, Y.-L., et al., 2011. Phase II study of neoadjuvant bevacizumab and radiotherapy for resectable soft tissue sarcomas. *Int. J. Radiat. Oncol. Biol. Phys.* 81, 1081–1090. <https://doi.org/10.1016/j.ijrobp.2010.07.024>.
- Young, R.J., Litière, S., Lia, M., Hogendoorn, P.C.W., Fisher, C., Mechttersheimer, G., et al., 2017. Predictive and prognostic factors associated with soft tissue sarcoma response to chemotherapy: a subgroup analysis of the European Organisation for Research and Treatment of Cancer 62012 study. *Acta Oncol.* 56, 1013–1020. <https://doi.org/10.1080/0284186X.2017.1315173>.
- Yu-Cherng, C.C., Mahan, J., Subhawong Ty, K., Wilky, B.A., Breto, A.L., et al., 2019. Quantitative imaging features of habitats in Soft tissue sarcomas are associated with treatment response: a pilot study. *Cancer Stud. Ther. J.* 4, 1–6. <https://doi.org/10.31038/CST.2019433>.
- Zhang, L., Ren, Z., 2020. Comparison of CT and MRI images for the prediction of soft-tissue sarcoma grading and lung metastasis via a convolutional neural networks model. *Clin. Radiol.* 75, 64–69. <https://doi.org/10.1016/j.crad.2019.08.008>.
- Zhang, X., Y-LE, Chen, Lim, R., Huang, C., Chebib, I.A., El Fakhri, G., 2016. Synergistic role of simultaneous PET/MRI-MRS in soft tissue sarcoma metabolism imaging. *Magn. Reson. Imaging* 34, 276–279. <https://doi.org/10.1016/j.mri.2015.10.027>.
- Zhao, F., Ahlawat, S., Farahani, S.J., Weber, K.L., Montgomery, E.A., Carrino, J.A., et al., 2014. Can MR imaging be used to predict tumor grade in soft-tissue sarcoma? *Radiology* 272, 192–201. <https://doi.org/10.1148/radiol.14131871>.

Nicolò Gennaro, MD Board-certified radiologist from Humanitas Clinical and Research Center in Milan, Italy. He is actively involved in the institutional Sarcoma Multidisciplinary Board and since 2019 he joined the Netherlands Cancer Institute as research fellow in oncologic imaging, focusing mainly on soft tissue sarcoma. His research areas mainly include oncologic imaging of the abdomen and musculo-skeletal system, including interventional procedures.

Sophie Reijers, MD PhD candidate at the Surgical Oncology Department of the Netherlands Cancer Institute, Amsterdam. Her work is mainly focused on neoadjuvant treatments for soft tissue sarcoma.

Annemarie Bruining, MD Board-certified radiologist at the Netherlands Cancer Institute, Amsterdam. Her areas of interest are breast, abdominal and pelvic imaging. She is actively involved in the Sarcoma Multidisciplinary Board of the Netherland Cancer Institute.

Christina Messiou, MD, PhD Board-certified radiologist at the Royal Marsden Hospital, London. Currently, she is Reader at The Institute of Cancer Research, London. Her areas of interest are melanoma, myeloma and sarcoma imaging. She is actively involved in the NCRI Expert Advisory Panel and works as Steering Committee of the European Organisation for the Research and Treatment of Cancer (Soft Tissue and Bone Sarcoma Group).

Rick Haas, MD, PhD Board-certified radiation oncologist with focus on bone and soft tissue sarcoma at the Netherlands Cancer Institute. He is professor of Radiotherapy at Leiden University Medical Center and is current president of the Connective Tissue Oncology Society. He is also Board Member of the European Musculo-Skeletal Oncology Society.

Piergiuseppe Colombo, MD Head of the Surgical Pathology Unit at Humanitas Clinical and Research Center, Milan. He developed extensive experience in sarcoma along with bladder, prostate and renal cancer, that also represents his research focus.

Zuhir Bodalal, MD PhD candidate at the Netherlands Cancer Institute. Amsterdam in the Department of Radiology. His works is focused on Imaging in Immunotherapy and Radiomics and AI applications in radiology.

Regina Beets-Tan, MD, PhD Head of the Department of Radiology at the Netherlands Cancer Institute in Amsterdam. She is full Professor of radiology at the University of Maastricht and adjunct professor of abdominal and oncological radiology at the University of Southern Denmark. She is a member of the European Society of Radiology (ESR) Executive Council, past president of the European Society of Oncologic Imaging (ESOI) and the president elect of the European Society of Gastrointestinal and Abdominal Radiology (ESGAR).

Winan van Houdt, MD, PhD Sarcoma surgeon and surgical oncologist at The Netherland Cancer Institute. Amsterdam, the Netherlands. He has a special interest in sarcoma, melanoma and translational research. He is secretary of EORTC Soft Tissue and Bone Sarcoma Group

Winette T.A. van der Graaf, MD, PhD Professor at Erasmus MC Rotterdam, The Netherlands and works as Medical Oncologist at the Netherland Cancer Institute, with a focus on sarcoma. She is Member of the EORTC board, chair of the Dutch Sarcoma Study Group and the Dutch AYA 'Young and Cancer' Network.