

Electrical and magnetic properties of ferritin: electron transport phenomena and electron paramagnetic resonance Labra Muñoz, J.A.

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EPR CHARACTERIZATION OF HUMAN-LIVER FERRITIN

In this chapter, we present a 9 GHz continuous wave EPR study that was used to elucidate the magnetic properties of the core of human-liver ferritin. The EPR description that is reported in this chapter is part of a larger study performed on ferritin, in collaboration with magnetometry experts. Since the article associated with the whole study has not been published yet [1], we include the full article in this chapter. It is important to clarify that only the measurements and analysis on EPR were performed by J. A. Labra-Muñoz. The magnetometry measurements, analysis, and theoretical model were performed by our collaborators. ¹

[•] Anton Lefering (RST-FAME, Delft University of Technology, Delft, The Netherlands): Magnetometry data acquisition.

[•] Ramon Egli (GeoSphere Austria, Department of Geophysics, Vienna, Austria): Magnetometry data processing and analysis. Also, R.E. proposed the idea and developed the theory behind the ferritin magnetic model.

Lucia Bossoni (Department of Applied Physics and Science Education, Eindhoven University of Technology, Eindhoven, the Netherlands): Magnetometry data acquisition, processing, and analysis.

5.1. ABSTRACT

Ferritin, the major iron storage protein in organisms, stores iron in the form of iron oxide, the mineral form of which is not well understood. Therefore, the question of how the ca. 2000 iron atoms in the ferritin core are magnetically coupled is still largely open. The ferritin core, with a diameter of 5–8 nm, is encapsulated in a protein shell that also catalyzes the uptake of iron and protects the core from outside interactions. Neurodegenerative disease is associated with iron imbalance, generating specific interest in the magnetic properties of ferritin. Here we present 9 GHz continuous wave EPR and a comprehensive set of magnetometry techniques including isothermal remanent magnetization (IRM) and AC susceptibility to elucidate the magnetic properties of the core of human liver ferritin. For the analysis of the magnetometry data, a new microscopic model of the ferritin-core spin structure is derived, showing that the magnetic moment is generated by surface-spin canting, rather than defects. The analysis explicitly includes the distribution of magnetic parameters, such as the distribution of the magnetic moment. This microscopic model explains some of the inconsistencies resulting from previous analysis approaches. The main findings are a mean magnetic moment of 337 $\mu_{\rm B}$ with a standard deviation of 0.947 $\mu_{\rm B}$. In contrast to previous reports, only a relatively small contribution of paramagnetic and ferrimagnetic phases is found, in the order of maximally 3%. For EPR, the over 30 mT wide signal of the ferritin core is analyzed using the model of the giant spin system [2]. Two components are needed minimally, and the broadening of these components suggests a broad distribution of the magnetic resonance parameters, the zero-field splitting, D, and the spin quantum number, S. We compare parameters from EPR and magnetometry and find that EPR is particularly sensitive to the surface spins of the core, revealing the potential to use EPR as a diagnostic for surface-spin disorder.

5.2. INTRODUCTION

Ferritin has fascinated scientists for decades. This ubiquitous iron-storage protein is made of a protein shell enclosing a core of bioavailable iron mineral [3]. The mammalian apoferritin shell contains two distinct polypeptide subunits: a heavy (21 kDa) and a light (19 kDa) chain [4]. These self-assemble into a 24-mer spherical structure which, depending on the organism and the specific organ it is found in, can have different heavyvs. light-chain ratios. While the heavy chain has a ferroxidase activity and protects cells from redox-active iron by the rapid uptake of Fe(II) and catalytic oxidation to Fe(III), the light chain promotes the nucleation and storage of iron as a biomineral [5]. The inner and outer dimensions of the protein shell are \sim 7–8 nm and \sim 12 nm, respectively. The biomineral inside the ferritin hollow cavity has received the attention of the biomedical community, because of a link between the protective function of ferritin against cellular iron toxicity[6] and altered core composition in the brain of patients with neurodegenerative diseases[7, 8]. From a physics standpoint, ferritin is also relevant in the fundamental study of nanoparticle properties. Through the biochemical machinery of iron incorporation and the protective protein shell, the composition, spin configuration, and size of the core are well-controlled and protected from post-assembly modification, factors that sometimes are difficult to control in man-made iron nanoparticles. Furthermore, the apoferritin shell prevents contact between the cores, eliminating magnetic exchange interactions.

Here, we focus on the magnetic properties of ferritin, which are an indicator of the spin structure and composition of the ferritin core. The magnetic properties of ferritin nano-particles, for example, their saturation magnetization, are directly linked to the relaxation rates of ferritin-rich tissue. As such, the magnetism of the protein influences the contrast of R2 and R2*-weighted MRI images [9–11].

In the past decades, 'bulk' magnetometry techniques have been used to characterize the magnetic and mineral state of ferritin [12, 13], along with spectroscopy techniques such as Mössbauer spectroscopy[13, 14], electron paramagnetic resonance (EPR) [15, 16], nuclear magnetic resonance (NMR) [17–19], as well as electron and X-ray microscopy techniques[20, 21], and diamond-based quantum spin relaxometry to study the ferritin room temperature magnetic properties[22]. Electron paramagnetic resonance (EPR), sometimes also referred to by the more general term electron magnetic resonance (EMR), has also been applied to ferritin [15, 16, 23–27], in spite of intrinsic challenges related to extreme spectral broadening.

It is generally agreed upon that the ferritin core is predominantly composed of a mineral resembling ferrihydrite[28], a poorly crystalline ferric oxyhydroxide. While some studies suggested a multiphase core composition [8, 29, 30], this hypothesis is not supported by NMR [18], magnetooptical measurements [19], Electron energy-loss spectroscopy [31], and only partially by muon spin rotation. On the other hand, there is a general consensus on the following properties: (1) iron-spins in the cores are antiferromagnetically (AF) coupled, (2) the cores possess a spontaneous magnetic moment of the order of ~ 300 $\mu_{\rm B}$, (3) the magnetic moments becomes blocked below $T_{\rm b} \approx 12$ K over the typical time span of magnetometric measurements, and (4) ferritin is superparamagnetic above $T_{\rm b}$ [15, 16, 23, 26]. However, several questions remain still unanswered: for instance, the origin of the spontaneous magnetic moment of the cores has been generically attributed to randomly distributed defects in the AF lattice [12, 32, 33], but the nature and location of these defects (e.g., in the bulk or at the surface) remain ambiguous [34–36]. The magnetic moment might also arise from core alteration, as it has been postulated in the case of ferrihydrite, which becomes partially ferrimagnetic during its transformation to hematite [37]. The maghemite-like ferrimagnetic phase resulting from this alteration process might explain the postulated low-coercivity phase in ferritin [29]. The apparent multiphase nature of ferritin might also originate from the interaction between different units within the core [28, 38], or between core and surface spins [15]. Such interactions can explain spin glass-like signatures such as shifted field-cooled hysteresis loops [39]. Finally, the absence of a spin-flop transition in fields up to 50 T [40], which is incompatible with the reported exchange and anisotropy fields of ferritin, questions the definition of anisotropy energy and energy barriers in AF nanoparticles [41].

Several limitations concur with our presently incomplete picture of the magnetic properties of ferritin. Interpretations of magnetometry and spectral techniques are intrinsically non-unique, therefore relying on models that require some prior knowledge of the spin structure of ferritin cores. Furthermore, most if not all magnetic parameters of ferritin are broadly distributed [42]. This can lead to erroneous conclusions if such broad distributions are replaced with mean values without considering possible correlations. Furthermore, broad parameter distributions make model fits very sensitive to initial assumptions and measurement noise, as seen with the multiple approaches used to model the superparamagnetic and linear contributions to in-field magnetization curves [12, 32, 34, 41, 43]. Finally, magnetometric and spectral techniques have been rarely combined [36], despite the intrinsic advantages of using complemental information to better constrain existing models.

In this work, we focus on the EPR and magnetometric properties of human-liver ferritin (HuLiFt) and address some of the issues mentioned above. Using these techniques, we explore the spin dynamics over a broad frequency range that includes DC (magnetization) to sub-kHz (AC-susceptibility) and microwave (9 GHz, EPR) measurements. The present work not only combines different methods, but also different fields of research, such as magnetism and, due to the mineral core of ferritin and the widespread occurrence of ferrihydrite in nature, geological aspects. We unified different naming conventions used for the characterization of magnetic materials using a single symbol for each quantity, except the magnetic moment, for which m is used in the case of magnetometric measurements and μ in the EPR context (see the list of symbols given in the Appendix).

Most EPR studies have been performed at 9 GHz on horse-spleen ferritin [15, 16, 23, 25, 26]. The broad superparamagnetic signal located at g' = 2 was associated with antiferromagnetically coupled Fe(III) ions in the ferritin cores, while a weak signal near g' = 4.3 was attributed to a small number of mononuclear Fe(III) centers showing typical paramagnetic behavior [15, 23, 25]. A few EPR studies have been performed on human ferritin, specifically from human spleens[16, 26], proposing that the broad EPR signal results from two overlapping broad contributions, a very anisotropic one at lower fields, and an isotropic one around g' = 2. EPR spectra have also been acquired at 35 GHz, facilitating the observation of these two broad components [26]. Interestingly, in Ref. [26] the lower field EPR component of horse-spleen ferritin has a much lower intensity than its counterpart in human-spleen ferritin, however, the article does not comment on this difference.

The EPR lineshape of superparamagnetic nanoparticles has been addressed using qualitative descriptions, such as the one by Noginova *et al.*[44] based on surface quantum effects, according to which the EPR intensity is proportional to $\exp(-\mu B/k_B T)$. Previously, Usselman *et al.*[45] used two different models to simulate temperature-dependent lineshape trends of iron oxide nanoparticles mineralized in *Listeria innocua* protein cages. The first model [46, 47] provides a qualitative description of the lineshape dependence on temperature [48] and frequency[49], assuming an ensemble of non-interacting single-domain particles, whose magnetization dynamics is described by the Landau-Lifshitz equation[47]. The second model [50] calculates the moment-distribution function by considering superparamagnetic fluctuations and ensemble broadening due to a distribution of anisotropy axes [50]. Other models include an improved static model to describe the decrease in the magnetic anisotropy as temperature increases[51]. However, these models do not reproduce all features of the measured EPR spectra.

Here, we use a quantum mechanical model [2, 52] to analyze 9 GHz EPR spectra of ferritin in solution, rather than in the freeze-dried state, to exclude ferritin-ferritin interactions. Our analysis reveals multiple EPR spectral components resulting from the distributed nature of the moment and anisotropy field distributions.

Magnetic simulations of equilibrium magnetization curves are used to understand the origin of the magnetic moment of ferritin cores. The theory behind these simulations is described in sections 5.3.6, 5.3.7, and 5.3.8, and its application in sections 4.2–4.5. Using these simulations and the measurement of isothermal remanent magnetization, we developed a new approach for the determination of the anisotropy field, magnetic moment, and blocking temperature distributions, as well as the relations existing between these parameters and between magnetometric and EPR measurements.

Our results can be explained by a simple model of AF nanoparticles whose magnetic moment is generated by surface-spin canting, rather than defects. This model explains the lack of a spin-flop transition below \sim 50 T. We also observe minor (<3%) contributions to the anisotropy field distribution that are compatible with a ferrimagnetic phase and a phase with very large anisotropy, respectively. Finally, we show that EPR is particularly sensitive to surface spins.

5.3. MATERIALS AND METHODS

5.3.1. PROPERTIES AND CHARACTERIZATION OF THE HUMAN-LIVER FER-RITIN

Commercial human-liver ferritin (HuLiFt) was obtained from LEE Biosolutions (Cat. No. 270-40, Lot 08E1805) and used without further purification. The protein concentration was 3.6 mg/ml with >95% purity as assessed by SDS-PAGE, Coomassie blue, and Prussian blue stains (see Appendix, for details). The protein loading factor was determined by inductively coupled plasma mass spectrometry (ICP-MS), yielding 1967±78 iron atoms per ferritin. The size distribution of ferritin cores was obtained from transmission electron microscopy (TEM) images (Fig. A5.19), using automatic circle detection based on the MATLAB function *regionprops*. The analysis of 2100 particles showed that core sizes strongly deviate from a unimodal lognormal-like distribution (Fig. 5.1). The main peak of the core size distribution is well approximated by a Weibull distribution with a mean of 6.5 nm and a median of 6.6 nm. An excess of small sizes with respect to the Weibull fit is observed below 4.5 nm and might represent incomplete fragmented cores.



Figure 5.1: Distribution of human ferritin core size and fit to an empirical function given by the sum (solid line) of a lognormal distribution with parameters $\mu = 1.128$ and $\sigma = 0.2704$, and a Weibull distribution with parameters $\beta = 6.797$ and $\eta = 11.08$ (dashed lines).

5.3.2. MAGNETOMETRY

The ferritin solution as purchased, i.e. without adding glycerol, see materials, was immersed in liquid nitrogen and subsequently freeze-dried over ~48 hours. For further details and sample handling see also Ref. [53]. The obtained powder sample was pressed into a gel capsule and loaded into a Quantum Design MPMS-XL SQUID magnetometer mounting the reciprocating sample option (RSO, noise floor: 1 pAm²). First, the magnetic moment in a 5 mT field was measured as a function of temperature after cooling to 5 K in zero field (zero-field-cooled, ZFC) and in 5 mT (field-cooled, FC), respectively. Then, the field-induced magnetization was measured at 5 K and 150 K (complete hysteresis loops) and in the 5–250 K range (initial magnetization curves). Low-field AC susceptibility was measured after ZFC to 20 K in a 5 mT DC field with superimposed longitudinal AC field of 0.38 mT amplitude and frequencies v = 0.113, 0.669, 4.481, 29.99, 59.9Hz. Only the in-phase AC susceptibility was processed, as the quadrature component was too noisy. ZFC and FC hysteresis loops were measured at 5 K and 25 K in order to detect the presence of an exchange field. Finally, high-resolution isothermal remanent magnetization (IRM) curves were acquired from 0 to 5 T in steps comprised between 1 and 200 mT, at 3, 5, 9, 11, 13, 17, and 20 K. Each point $M_r(B)$ of an IRM curve is obtained by ramping the field from 0 to B and then back to 0, with no overshoot. Repeated measurements were acquired to ensure reproducibility and enhance the signal-to-noise ratio (SNR): an SNR \geq 100 is ideally needed over the field range containing relevant coercivity contributions. All measurements are expressed as mass magnetization obtained by dividing the magnetic moment by the sample mass. All data analyses were carried out in Matlab2016a and Mathematica 12 using built-in non-linear minimization routines.

5.3.3. ELECTRON PARAMAGNETIC RESONANCE

In order to avoid ferritin-ferritin interactions that might occur in the freeze-dried samples [16, 23, 25] we used the buffered ferritin solution for EPR measurements. For this purpose, 100 μ L of the ferritin solution with 20% glycerol (vol/vol) solution were trans-

ferred into a 4 mm outer diameter EPR tube. The tube was then immediately frozen in liquid nitrogen. Continuous wave (CW) EPR measurements were performed with a 9 GHz ELEXSYS E680 EPR spectrometer (Bruker, Rheinstetten, Germany), equipped with a rectangular cavity. The spectra were recorded with 20 mW power, 90 kHz modulation frequency, and 29.46 $G_{\rm pp}$ field modulation amplitude at temperatures comprised between 5 and 210 K. The accumulation time was 11.2 min per spectrum. A helium flux cryostat was used to control the temperature. Temperatures were taken from the readout of the Oxford temperature unit connected to a thermocouple placed beneath the sample. Measurement of the buffered suspension instead of freeze-dried powders produces a lower EPR signal intensity; however, signal quality was still sufficient for further processing.

Simulations of measured spectra have been conducted with the EasySpin package (5.2.4) using a Matlab (R2019a) script. Spectra between 5 and 15 K were not analyzed because it was not possible to accurately discriminate the broad signal from background noise. Final simulations have been performed with two components. The parameters D and H_{strain} (Gaussian broadening) were adjusted independently for each component at each temperature, along with the relative contributions of the two components and the pepper routine parameters of the EasySin package, after choosing S = 10 and g = 2.01 as fixed parameters for both components. The sensitivity of the model to the parameters D and H_{strain} was tested by changing, for example, D of one component and leaving all other parameters unchanged until a visible lineshape alteration was detected (see section Sensitivity of EPR parameters in the Appendix).

5.3.4. Theoretical background of Magnetometry analysis

In the following, we discuss different models of the spin structure of ferritin cores and the implications they have on magnetometry and EPR results.

Starting from models proposed in the literature and our measurements we derive a new model for the spin structure of ferritin. First, we analyze the implications of polyphase ferritin cores for the interpretation of magnetometry results, showing that individual phases coexisting in the same core cannot be discriminated by isothermal magnetic measurements. Next, we use these findings to discuss possible spin structures that are compatible with equilibrium magnetization curves, showing that spin canting is needed to explain the ferritin-core susceptibility at lower fields and the lack of a spin-flop transition. The universal relation between mean magnetic moment and the number of Fe atoms in ferritin and ferrihydrite nanoparticles further confirms these findings. Finally, magnetometric parameters are discussed in relation to the energy barrier that needs to be overcome to switch the magnetic moment of ferritin cores, showing that the expression $E_b = KV$ is valid also in the case of AF nanoparticles and that a wide distribution of anisotropy fields can be a consequence of the fact that B_a , the anisotropy field, is inversely proportional to m, the magnetic moment.

5.3.5. MAGNETIC PHASES PROPOSED FOR FERRITIN IN THE LITERATURE

The use of TEM X-ray Adsorption Near Edge Spectroscopy (XANES) and Small-Angle X-ray Scattering (SAXS) for probing the composition of ferritin cores (i.e., the propor-

tion of Fe and O atoms), and of Electron Energy-Loss Spectroscopy (EELS) for probing the oxidation state of Fe ions suggests a polyphase structure[8, 29, 30] made mainly of ferrihydrite (Fh) or a phosphorous-rich phase whose structure is similar to that of ferrihydrite [19], with minor hematite (α -Fe₂O₃), magnetite (Fe₃O₄), and wüstite (FeO) contributions[30]. In the human brain, these secondary contributions tend to increase with age and in patients with neurological diseases [8]. Spatial EELS analyses suggest that magnetite is concentrated at the core surface [30]. On the other hand, other studies based on magneto-optical measurements suggest a single-phase core structure.

The hypothetical polyphase nature of ferritin cores might be the result of Fh alteration or variable iron storage mechanisms. Fh is known to form an ordered ferrimagnetic structure during aging [37], as an intermediate product on the pathway to full conversion to hematite. The conversion rate of synthetic Fh is very slow at room temperature, but it is greatly enhanced in the presence of ligands [54]. Heating during sample preparation might therefore be an issue for the assessment of ferritin core composition. On the other hand, a 3D morphology study suggests that ferritin cores are composed of up to eight regions with disordered surfaces, consistent with the eight channels in the protein shell that deliver iron to the central cavity [28]. Disordered surfaces are magnetically distinct from the bulk and can therefore be considered as an additional phase.

Ferritin-core phases identified so far are characterized by different forms of magnetic order, including AF (Fh, FeO), canted AF (hematite), ferrimagnetic (magnetite), and speromagnetic (surfaces). However, if these phases coexist within the same core units, exchange coupling is expected to produce a collective spin behavior that is not equivalent to the superposition of bulk-phase properties. Magnetometry data might still suggest a polyphase composition, for instance through a bimodal magnetic moment or energybarrier distribution [29, 38]. In our case, the existence of multiple magnetic phases is supported by IRM acquisition curves (see Sections 4 and 5).

Magnetic evidence used so far in support of significant contributions from phases other than Fh depends heavily on the way energy barrier and magnetic moment distributions are measured and modeled. For instance, the energy barrier distribution obtained from magnetic viscosity measurements is bimodal [38], while the same distribution derived from quadrature AC susceptibility data is strictly unimodal [55]. The case of the magnetic moment distribution is even more ambiguous, as the fit of equilibrium magnetization curves with two superparamagnetic components with distinct single-valued magnetic moments, as proposed by Brem *et al.* [29], is a valid alternative to the distributed moment model described in the Results section. Evidently, the additional degree of freedom of the two-component model enables better fits to the data. Nevertheless, as we demonstrate further below, this interpretation is incompatible with the IRM results obtained in the present study and therefore discarded.

5.3.6. THE MAGNETIC SIGNATURE OF IDEALIZED SPIN STRUCTURES

Consider a collinear two-sublattice AF particle with sublattice magnetization M_0 and exchange constants A_a , A_b , and A_{ab} . In the case of slightly uncompensated sublattices, the two sublattice magnetizations are given by $M_b = M_0$ and $M_a = (1 + \alpha)M_0$, respectively, where $\alpha > -1$ is the fraction of excess moment in one sublattice. This creates a

net spontaneous magnetic moment $m_{\rm uc} = \alpha M_0 V$ in a particle with volume V. In the following, it is assumed that $m_{\rm uc}$ is rigidly coupled to the sublattice magnetizations, due to the strong AF coupling [56, 57], so that any change of the magnetic moment is produced by the uniform rotation of all spins in both sublattices. Furthermore, particles possess a positive uniaxial magnetic anisotropy [58] with anisotropy constant K and easy axis parallel to the unit vector $\mathbf{e} = (\sin\phi, 0, \cos\phi)$, so that $E_a = -\frac{1}{2}KV[(1+\alpha)^2(\mathbf{e}\cdot\mathbf{u}_a)^2 + (\mathbf{e}\cdot\mathbf{u}_b)^2]$ is the anisotropy energy corresponding to sublattice magnetizations parallel to the unit vectors \mathbf{u}_a and \mathbf{u}_b , respectively [59]. In the absence of external fields, the total energy is minimized when the sublattice magnetizations are exactly antiparallel and oriented along the easy axis. The application of a field **B** rotates the lattice magnetizations away from the easy axis and introduces an induced spin canting. Following Bogdanov et al. [59], we define the spin canting angle $-\pi/2 \le \varepsilon \le \pi/2$, such that $\mathbf{u}_{a,b} = \pm \mathbf{p} \cos \varepsilon + \mathbf{n} \sin \varepsilon$, where **p** is the so-called Néel unit vector parallel to the staggered magnetization direction $\mathbf{u}_a - \mathbf{u}_b$, and $\mathbf{n} \perp \mathbf{p}$ is the unit vector parallel to the canting magnetization direction $\mathbf{u}_a + \mathbf{u}_b$. In a spherical coordinate system with $\mathbf{B} \parallel \hat{\mathbf{z}}, \mathbf{p} = (\sin\theta\cos\psi, \sin\theta\sin\psi, \cos\theta)$ and $\mathbf{n} = \mathbf{n}_1 \cos \lambda + \mathbf{n}_2 \sin \lambda$ with $\mathbf{n}_1 = (\hat{\mathbf{z}} \times \mathbf{p}) \times \mathbf{p}$ parallel to the plane spanned by **B** and **e**, $\mathbf{n}_2 = \hat{\mathbf{z}} \times \mathbf{p} \perp \mathbf{z}, \ 0 \le \theta \le \pi, \ \text{and} \ -\pi \le \psi, \lambda \le \pi.$ In this case, the total energy E per unit of volume of a particle with the above properties is given by

$$\frac{E}{M_0} = +2(1+\alpha)B_{\rm E}\sin^2\varepsilon + [(2+\alpha)\sin\varepsilon\cos\lambda\sin\theta - \alpha\cos\varepsilon\cos\theta]B -\frac{1}{2}[(\mathbf{e}\cdot\mathbf{p})^2\cos^2\varepsilon + (\mathbf{e}\cdot\mathbf{n})^2\sin^2\varepsilon]B_{\rm a} -\frac{1}{4}\alpha(2+\alpha)[(\mathbf{e}\cdot\mathbf{p})\cos\varepsilon + (\mathbf{e}\cdot\mathbf{n})\sin\varepsilon]^2B_{\rm a}$$
(5.1)

with the exchange field $B_E = A_{ab}M_0$ and the anisotropy field $B_a = 2K/M_0$. The equilibrium magnetization of an ensemble of non-interacting particles is given by [60]

$$M(B) = -\sum_{j} \frac{\sum_{i} \frac{\partial \mathcal{H}_{i}}{\partial B} e^{-\mathcal{H}_{i}(B)/k_{\rm B}T}}{\sum_{i} e^{-\mathcal{H}_{i}(B)/k_{\rm B}T}}$$
(5.2)

for all particles with easy axis orientations \mathbf{e}_j and states *i* of their Hamiltonian \mathcal{H} . In the classical case where the magnetic moments can take any orientation, $\mathcal{H} = E(\theta, \psi, \lambda, \varepsilon)$ and $\partial \mathcal{H}_i / \partial B = -M_0 V \zeta$, with

$$\zeta = \alpha \cos \varepsilon \cos \theta - (2 + \alpha) \sin \varepsilon \sin \theta \cos \lambda \tag{5.3}$$

being the ratio between the component of the magnetic moment parallel to **B**, and M_0V . In this case, summations in eq. (5.2) are replaced by integrals, obtaining

$$\frac{M(B)}{M_{\rm s}} = \int_{\phi=0}^{\pi/2} \sin\phi \,\mathrm{d}\phi \,\frac{\int_{\varepsilon,\lambda,\psi,\theta} \zeta(\phi) \, e^{-E(B,\phi)/k_{\rm B}T} \sin\theta \,\mathrm{d}\theta \,\mathrm{d}\psi \,\mathrm{d}\lambda \,\mathrm{d}\varepsilon}{\int_{\varepsilon,\lambda,\psi,\theta} e^{-E(B,\phi)/k_{\rm B}T} \sin\theta \,\mathrm{d}\theta \,\mathrm{d}\psi \,\mathrm{d}\lambda \,\mathrm{d}\varepsilon}.$$
(5.4)

This result cannot be further simplified, since the dependence of E on all five integration variables is not separable, except when spin canting and anisotropy are negligible: in this case eq. (5.4) converges to the well-known Langevin law of superparamagnetism [60].

Numerical evaluations of eq. (5.4) are extremely time-consuming, due to the five-fold integrals: on a PC, a single equilibrium magnetization calculation takes ~ 7 min using an optimized method (Appendix: Equilibrium magnetization models). Simulations of M(B) at 50 K using $B_{\rm E} \approx 320$ T for ferritin [57], show that the equilibrium magnetization is governed by two regimes (Fig. 5.2). In small fields, thermal fluctuations, which act on each degree of freedom, induce a small spin-canting angle that adds a random canting moment $m_{\rm c}$ perpendicular to the uncompensated moment $m_{\rm uc}$. If $m_{\rm uc} = 0$, the random canting moment generates a low-field susceptibility $\chi_{\rm lf} = \chi_{\perp}/3$, where $\chi_{\perp} = M_0/B_{\rm E}$ is the bulk perpendicular susceptibility of the AF lattice. In larger fields, the canting moment gets progressively aligned with the field, producing a transition to the highfield regime given by $M = \chi B$ (Fig. 5.2b). If $\alpha m_{\rm nc} B_{\rm E} \gg k_{\rm B} T$, the uncompensated moment is much larger than the canting moment, and the low-field magnetization converges to the Langevin model prediction, with $\chi_{\rm lf} = M_{\rm s} m_{\rm uc}/k_{\rm B} T$, regardless of single particle anisotropy. At larger fields, anisotropy decreases the equilibrium magnetization, because of the competition between field and easy axis alignment (dashed lines, Fig. 5.2a). The same result has been obtained previously [60, 61] for non-interacting ferromagnetic particles. In the simulations of Fig. 5.2, this ferromagnetic-like regime holds for $m_{\rm uc} \ge 150 \mu_{\rm B}$ and B < 5 T.

External fields increase the canting angle of all particles whose Néel vector **p** is not parallel to the field direction; however, at the same time, the uncompensated moment tends to align the Néel vector with the field, so that spin canting becomes less effective. As a result, the spin canting angle continues to be controlled only by thermal fluctuations until the alignment of the canting moment becomes energetically more favorable than that of $m_{\rm uc}$: at this point, a so-called spin-flop transition takes place through rotation of the Néel vector by 90°. In bulk antiferromagnets, this is a sudden event that occurs at the spin-flop field $B_{\rm sf} \approx (2B_{\rm E}B_{\rm a})^{1/2}$ predicted by mean-field theory [59]. This result does not hold for the equilibrium magnetization of AF nanoparticles, because thermally activated spin canting occurs in all fields. As shown by our simulations, the spin-flop field is defined, in this case, by the intersection of the Langevin law with the high-field regime $M = \chi_{\perp} B$ of perfect antiferromagnets (Fig. 5.2b). For sufficiently large values of α (e.g., $\alpha \approx 0.025$ for the 50 K simulations of Fig. 5.2), the spin-flop field for the equilibrium magnetization is then given by $B_{sfe} = \alpha B_E$. Contrary to bulk antiferromagnetism [41], magnetic anisotropy does not affect B_{sfe} ; instead, it increases the slope of M(B) in the Langevin saturation regime, until it becomes indistinguishable from the spin-flop regime (Fig. 5.3). This occurs because the additional Néel vector misalignment produced by randomly oriented anisotropy axes enhances the induced spin canting and its contribution to the equilibrium magnetization already in fields $< B_{sfe}$. Vice versa, the same anisotropy effect reduces the alignment of the canting moment above B_{sfe} .

The defect moment model illustrated above is our starting point for assessing the validity of the modified Langevin fit we used to estimate the distribution of m_{uc} from M(B) measurements, and for testing the origin of the uncompensated moment in ferritin cores.



Figure 5.2: Numerical simulations of the T = 50 K equilibrium magnetization of randomly oriented AF particles with no anisotropy (K = 0, solid lines), and with anisotropy (K = 18.3 kJ/m³, dashed lines), for selected values of m_{uc} . Other model parameters are $M_0 = 366.8$ kA/m, $B_E = 320$ T, and $M_0 V = 6000 \mu_B$. (a) and (b) represent the same simulations over different field ranges.

For this purpose, we used eq. (5.4) to calculate M(B) at 50 K for an ensemble of randomly oriented particles with the same lognormal distribution of $m_{\rm uc}$ obtained from the modified Langevin fit, together with model parameters representative for ferritin, that is, $K = 18.3 \text{ kJ/m}^3$, $B_{\rm E} = 320 \text{ T}$, and $M_0V = 6000 \,\mu_{\rm B}$ (Fig. 5.4b). According to this simulation, the spin-flop transition is expected to occur at ~10 T, instead of $(2B_{\rm E}B_{\rm a})^{1/2} \approx 36$ T. This is above the maximum field used in our M(B) measurements, but well below the ~50 T maximum field used in experiments that failed to detect such a transition [40, 41]. The lack of a spin-flop transition below 50 T has been attributed to larger-than-expected values of $B_{\rm E}$ and/or $B_{\rm a}$ [40, 41]. As shown by our simulations, $B_{\rm sfe}$ is not affected by single particle anisotropy, while the >5 times larger exchange field required to push $B_{\rm sfe}$ beyond the maximum field range of available measurements does not comply with $B_{\rm E}$ values obtained from high-field estimates of the AF susceptibility [41]. Therefore, the only plausible explanation for the discrepancy between the simulation of Fig. 5.4 and actual high-field measurements of M(B) is that the defect model of Néel [62] does not provide a correct description of the in-field magnetic moment of ferritin cores.

As far as the effect of anisotropy on the shape of M(B) is concerned, a relatively small but non-negligible reduction of the equilibrium magnetization occurs over the 0.5–3 T field range, before the onset of saturation (Fig. 5.4a). This alters the magnetic moment dis-



Figure 5.3: Numerical simulations of the T = 50 K equilibrium magnetization of randomly oriented AF particles with $m_{uc} = 300 \mu_B$ and selected values of the anisotropy constant. Other model parameters are the same as in Fig. 5.2.

tribution obtained by the modified Langevin fit, lowering the apparent mean moment by ~40% and increasing the apparent moment distribution width by ~30%. A-posteriori corrections of the Langevin fit according to these results, however, would not be meaningful, since the absence of a spin-flop transition in fields \leq 50 T requires a different model for the superparamagnetic behavior of ferritin. Therefore, we look for spin configurations that produce a net spontaneous moment that is not parallel to the Néel vector. A possible source for such spin configurations is surface anisotropy because it affects the orientation of surface spins with respect to the bulk.

Surface spins are often assumed to be in a disordered, spin-glass-like state created by a distribution of exchange field vectors pointing to different directions [63, 64]. The exchange interaction between surface and internal spins manifests itself through exchange bias, that is, the horizontal shift of FC hysteresis. The existence of this exchange bias in horse-spleen ferritin [12] and in our sample (see section 5.3.6), along with data from dynamic Mössbauer spectroscopy [36], testifies for the existence of a surface spin layer in ferritin cores. Due to the sensitivity of exchange interactions to the position of ions, surface spins can take multiple configurations that are not necessarily associated with a complete disorder, as seen for instance with the spike, throttled, and two-pole configurations obtained from simulations of ferrimagnetic nanoparticles [65]. These configurations decrease the net moment of particles with ferrimagnetic order, but represent a possible source of spontaneous moments in AF nanoparticles. Furthermore, the exchange coupling between surface and internal spins can alter the AF ordering of the whole particle [66]. Recent simulations of small AF nanoparticles demonstrate this effect, with spike, throttled, and disordered internal spin configurations, as well as spin canting (Fig. 11–12 in Laura-Ccahuana and De Biasi [67]). Most importantly, these simulations show that surface anisotropy increases the spin-flop field and/or limits spin-flopping to subregions of the particles or suppresses it completely, so that the bulk magnetization does no longer show the effects of a spin-flop transition [67]. Similar effects might also occur at interfaces between different phases in a polyphase model of ferritin cores, especially



Figure 5.4: Numerical simulations of the T = 50 K equilibrium magnetization of randomly oriented AF particles with the lognormal distribution of magnetic moments deduced from the modified Langevin fit of M(B) measurements (logarithmic mean: $215 \mu_B$, logarithmic standard deviation: 0.963), calculated using the Langevin model (dashed line), and the AF model of Fig. 5.2 (solid lines). (a) and (b) represent the same simulations over different field ranges.

if secondary phases consist of few surface atomic layers [30].

In principle, the equilibrium magnetization of AF particles with surface anisotropy can be calculated by evaluating eq. (5.2) with the Hamiltonian [65, 67]

$$\mathcal{H} = -2 \sum_{i,j \in V} J_{ij} \mathbf{S}_i \cdot \mathbf{S}_j - g \mu_{\rm B} \mathbf{B} \cdot \sum_{i \in V} \mathbf{S}_i - K_V \sum_{i \in V} (\mathbf{k}_V \cdot \mathbf{S}_i)^2 -K_S \sum_{i \in \partial V} (\mathbf{n}_i \cdot \mathbf{S}_i)^2,$$
(5.5)

where \mathbf{S}_i are the spin vectors, J the exchange constant, K_V the uniaxial volume anisotropy with easy axis \mathbf{k}_V , K_S the surface anisotropy, and \mathbf{n}_i the surface normal vector for the i-th surface spin. This requires sampling the whole parameter space spanned by the spin vectors. Some characteristics of the complete solution can be captured by simulations based on an equivalent homogeneous system where the spontaneous moment $m_c = 2M_0V\sin\varepsilon_s$ is produced by a zero-field canting angle ε_s . Spontaneous canting is generated by an additional energy term proportional to $\mathbf{c} \cdot (\mathbf{u}_a \times \mathbf{u}_b)$, with \mathbf{c} being the unit vector perpendicular to the canting plane [68]. In this case, the total magnetic energy per unit of volume becomes

$$\frac{E}{M_0} = B_{\rm E} \mathbf{u}_a \cdot \mathbf{u}_b + B_{\rm E} \tan(2\varepsilon_{\rm s}) \mathbf{c} \cdot (\mathbf{u}_a \times \mathbf{u}_b) - \mathbf{B}(\mathbf{u}_a + \mathbf{u}_b) + E_a(\mathbf{u}_a, \mathbf{u}_b, \mathbf{e}),$$
(5.6)

where the term E_a accounts for the combined effect of surface and volume anisotropy. In the isotropic case ($E_a = 0$), the total energy is insensitive to the direction **n** of canting, as long as it is perpendicular to **c**. Therefore, $E_a = 0$ describes an azimuthal dependence of E with respect to c, which requires an additional degree of freedom with respect to the case of a defect moment. Given the minor role played by realistic values of the magnetocrystalline anisotropy in the defect moment simulations of Fig. 5.4, numerical calculations have been performed with $E_a = 0$. As expected, the equilibrium magnetization of AF particles with a spontaneous canting moment is equivalent to the sum of a Langevin term that describes the superparamagnetism of m_c , and a linear term $M = \chi_{\perp} B$ that accounts for the induced spin canting (Fig. 5.5a). Because $m_{\rm c}$ is already perpendicular to the Néel vector, there is no spin-flop transition. The M(B) curve resulting from the same distribution of moments used to simulate uncompensated moment now contains a linear term $M = \chi B$ comparable with the non-paramagnetic component $\mathscr{L}(mB/k_{\rm B}T) + \chi B$ of the model used to fit M(B) data (Fig. 5.5b). The normalized slope χ/M_s of the linear term is ~40% smaller than the fitted value at 50 K. A good agreement, on the other hand, is obtained at 250 K. The temperature-dependent mismatch is due to the fact that this model does not account for the effects of single-particle anisotropy, and in particular surface anisotropy, on γ .

5.3.7. The role of distributed parameters

Magnetic properties of ferritin are usually expressed in terms of averaged quantities or treated as single-valued parameters (e.g., K, B_a , the blocking temperature T_b). This approach is correct only when it is applied to intrinsic properties of the material, such as the sublattice magnetizations and exchange constants, and the bulk magnetocrystalline anisotropy. Parameters that do not represent intrinsic properties of an AF crystal, such as the magnetic moment and the anisotropy field, must be treated as statistical distributions to avoid incorrect interpretations, as shown in the following example. The mean magnetic moment of $\sim 350 \,\mu_{\rm B}$ obtained from simple [12, 69] or distributed (this work) Langevin fits is usually attributed to an uncompensated moment matching one of the three models proposed by Néel [62]. The first Néel model associates $m_{\rm uc}$ with defects randomly distributed among the AF sublattices: in this case, $m_{\rm uc} \approx \mu_{\rm Fe} (cN)^{1/2}$, where $N \approx 2500$ is the total number of Fe ions in ferritin, $\mu_{\rm Fe} \sim 5\mu_{\rm B}$ their magnetic moment, and $c \ll 1$ the concentration of defects. The good match between $\langle m \rangle$ estimates obtained with c = 1 and with the Langevin fit has often been used as a validation of this model [12, 32, 33], even though a simple probabilistic analysis shows the largest possible mean moment $\langle m_{\rm uc} \rangle_{\rm max} \approx 0.56 \,\mu_{\rm Fe} N^{1/2}$ is obtained with c = 0.5.

Magnetic moment estimates obtained from ferritin nanoparticles with different iron loadings support a power law of the form $m \propto N^p$ with *p* comprised between 1/2 and 2/3, the latter being the exponent expected from the uncompensated moments arising



Figure 5.5: (a) Numerical simulations of the T = 50 K equilibrium magnetization of randomly oriented AF particles with no anisotropy for selected values of the canting moment m_c . Other model parameters are $M_0 = 366.8$ kA/m, $B_E = 320$ T, and $M_0V = 6000 \mu_B$. (b) Same as (a) for a lognormal distribution of magnetic moments with logarithmic mean of $215 \mu_B$ and logarithmic standard deviation of 0.963. The Langevin model and the sum of the Langevin and linear terms of the measurement fits are shown for comparison.

from surface spins belonging to one sublattice only [34]. The same empirical power law applies to a large compilation of available data on ferritin and ferrihydrite, which yields $p \approx 0.59 \pm 0.06$, with no systematic differences related to particle composition (Fig. 5.6). The maximum possible defect moment $\langle m_{\rm uc} \rangle_{\rm max}$ associated with p = 1/2 is compatible only with two measurements out of a total of 28 so that the Néel defect model must be discarded. The empirical trend $\langle m \rangle \approx 0.4 \mu_{\rm Fe} N^{2/3}$ fits the data almost optimally. Its compatibility with a surface-spin-canting origin of the magnetic moment is discussed below.

A better insight into the origin of the peculiar magnetic properties of ferritin is provided by the joint analysis of the magnetic moment and anisotropy distributions, through the relation between m, B_a , K, and T_b imposed by the Néel-Arrhenius model

$$E_{\rm b} = \frac{1}{2}mB_{\rm a} = KV = k_{\rm B}T_{\rm b}\ln\frac{t_{\rm m}}{\tau_0}$$
(5.7)

for the energy barrier of uniaxial single-domain particles, with t_m being the measurement time and τ_0 the attempt-time for spin reversal.



Figure 5.6: Mean magnetic moment vs. number of Fe atoms in ferritin and ferrihydrite particles (circles), obtained from a compilation of literature data [12, 32–34, 70–79]. Lines show best fits with different power laws described in the text.

We also tried the alternative model of fitting measured equilibrium magnetization curves with two distinct magnetic moments, as proposed by Brem et al. [29]. Replication of this approach with our data yields superparamagnetic and linear contributions that are almost identical to our original model, with a slightly smaller misfit (Appendix, Figure A5.37). The magnetic moments $m_1 = 97 \,\mu_{\rm B}$ and $m_2 = 540 \,\mu_{\rm B}$ and the relative superparamagnetic contributions (57% and 43%, respectively) are similar to those obtained by Brem *et al.* for horse-spleen ferritin, where m_2 was attributed to magnetite. Using a magnetic moment of 4.1 $\mu_{\rm B}$ per formula unit of Fe₃O₄, magnetite must contain ~20% of the ~2000 Fe atoms in our ferritin cores in order to explain the magnetic moment of 540 $\mu_{\rm B}$ attributed to this phase. The model of Brem *et al.* requires the two moments to be fully uncoupled in order to be modeled by the linear combination of two independent Langevin functions, in which case their contributions to the remanent magnetization would add linearly in the same proportions as the superparamagnetic contributions. However, the magnetite-like component deduced from our IRM measurements contributes to $\sim 0.6\%$ of the total blocked magnetization, much less than deduced from the two-moment Langevin model[29]. This discrepancy is too large to be explained by uncertainties in the Langevin fit or in fits of the IRM acquisition curves. Therefore, we assume that ferritin cores are made either by a single phase or by different phases with rigidly coupled spins, justifying the representation of the core magnetization by a fixed magnetic moment.

5.3.8. SURFACE-SPIN MODEL FOR FERRITIN

As discussed above, surface spin canting in ferritin cores appears to be the only source of a spontaneous magnetic moment that is compatible with all magnetic characterizations reported so far. This model represents the basis of our approach to fit our equilibrium magnetization curves and obtain the magnetic moment distribution (see section 5.4.4).

Along with the anisotropy field distribution obtained from the analysis of IRM acquisition curves, these results allow to verify the consistency of all magnetometry measurements, as explained in the following.

If the magnetic moment of ferritin cores is controlled by surface effects, we can expect $m = N_{\rm s} \mu_{\rm Fe} \eta_{\rm s}$ to be proportional to the number $N_{\rm s}$ of surface spins, their magnetic moment $\mu_{\rm Fe} \approx 5\mu_{\rm B}$, and the degree $\eta_{\rm s} = \sin\varepsilon_{\rm s}$ of canting, regardless of the detailed spin configuration. Internal spins might also experience some canting through coupling with the surface [67]. In all cases, the source of spin canting is related to the surface, and therefore, we assume $N_{\rm s} = \kappa_{\rm s} V^{2/3}$ with $\kappa_{\rm s} = (36\pi)^{1/3} (\rho_{\rm f} N_{\rm A}/u_{\rm f})^{2/3}$, where $\rho_{\rm f} \approx 3.9$ g/cm³ is the density of ferritin cores[80], $N_{\rm A}$ the Avogadro constant, and $u_{\rm f} \approx 96$ g/mol the molar mass per Fe atom obtained with the chemical formula 5Fe₂O₃ · 9H₂O of six-line ferrihydrite. The resulting expression $m = \kappa_{\rm s} \mu_{\rm Fe} \eta_{\rm s} V^{2/3}$ explains the data compilation in Fig. 5.6, yielding $\eta_{\rm s} \approx 0.4$ if only surface spins are canted. Much lower canting angles are required if spin canting extends to internal spins [67]. Alternate compositions have been proposed for the ferritin mineral core [19]: if this composition is indeed significantly different from that of ferrihydrite, it does not affect the magnetic moment, as seen in Fig. 5.6.

According to this model, the distribution of $\ln(m)$, which is usually assumed to be Gaussian when fitting M(B) curves, is given by $g_m = g_{\kappa\mu\eta} * g_{V^{2/3}}$, where $g_{\kappa\mu\eta}$ and $g_{V^{2/3}}$ are the distributions of $\ln(\kappa_s\mu_{Fe}\eta_s)$ and $\ln V^{2/3}$, respectively, and "*" is the convolution operator. Deconvolution of g_m obtained from fitting M(B) curves with $g_{V^{2/3}}$ obtained from TEM observations thus yields an estimate of $g_{\kappa\mu\eta}$, from which the distribution of η_s easily derived. The maximum range of this distribution should not exceed $\eta_s = 1$ for a physically reasonable spin canting model.

The surface spin model must also satisfy the Néel-Arrhenius model of eq. (5.7) when the distributions of m, B_a and T_b are considered. The normalized temperature dependence $M_r(T)/M_r(0)$ of the saturation remanent magnetization M_r yields, by definition, the integral of the blocking temperature distribution, $f_b(T)$. The function $f_b(T)$ can also be reconstructed from $E_b = mB_a/2$ using the distributions of m and B_a obtained from M(B) and from IRM acquisition curves, respectively. These distributions, however, are extremely broad, so that the product mB depends critically on the type of relation existing between m and B, and not just on the respective mean values. In the case of ferrimagnetic SD particles with spontaneous magnetization M_s , $m = M_s V$ and $B_a = 2K/M_s$ are independent variables, because M_s is a fixed material property. Accordingly, the distribution of $\ln(mB_a)$ is given by the convolution of the distributions of $\ln(m)$ and $\ln(B_a)$, respectively. This approach, however, does not hold for AF particles, where $M_s = m/V$ is itself distributed. Accordingly, in the case of ferritin, convolution of the distributions of $\ln(m)$ and $\ln(B_a)$ yields an extremely broad distribution of energy barriers, which does not match $f_b(T)$ (Appendix, Figure A5.38).

The correct expression for the anisotropy field of AF particles obtained from eq. (5.7) is $B_a = 2K/m$: in this case, B_a is inversely proportional to the magnetic moment, and the two factors in $E_b = mB_a/2$ are no longer independent variables. The inverse relation between *m* and B_a can be understood by considering that the work required to reverse all spins of the AF lattice must be provided by the Zeeman energy of the magnetic moment

in the switching field B_{sw} , so that a smaller magnetic moment must be compensated by a larger $B_{sw} \propto B_a$. The above model for *m* yields $B_a = 2KV^{1/3}/\kappa_s\mu_{Fe}\eta_s$. If *V* is a narrow distribution, as in the case of ferritin cores, any size dependence of *K* and η_s is negligible, and B_a is the product of almost completely independent satistical variables. The resulting anisotropy field distribution is then given by $g_a \approx g_{2K/\kappa\mu} * g_{V^{1/3}} * g_{\eta^{-1}}$, where $g_{2K/\kappa\mu}, g_{V^{1/3}}$, and $g_{\eta^{-1}}$ are the distributions of $\ln(2K/\kappa_s\mu_{Fe})$, $\ln V^{1/3}$, and $\ln \eta^{-1}$, respectively so that g_a can be reconstructed from estimates of *K*, *V*, and *m* obtained from magnetometry measurements. On the other hand, g_a can also be obtained directly from IRM acquisition curves, using the well-known relation $B_a = 2.083B_{sw}$ for randomly oriented, uniaxial SD particles [81]. The two reconstructions of the anisotropy field distribution must coincide if the surface spin model described above correctly describes the spin configuration of ferritin cores.

5.4. RESULTS

5.4.1. ELECTRON PARAMAGNETIC RESONANCE

Figure 5.7a shows the continuous wave 9 GHz EPR spectra between 5 and 210 K. The most prominent feature is a broad signal with a linewidth of ~100 mT at 190 K, which is centered at g' = 2.0. This broad signal is due to the mineral core in the protein shell of ferritin, in agreement with what has been reported in literature [15, 16, 23, 24, 27]. Three narrow signals overlap with the broad spectrum at g' = 2.0, 4.3, and 5.8, respectively (arrows). The g' = 4.3 and g' = 5.8 signals are usually attributed to mononuclear rhombic Fe(III) sites [15, 23, 24, 53, 82] and high-spin Fe(III) in methemoglobin [83], respectively. Multiple lines at g' = 2.0 [53, 83] might be ascribed to radical impurities, Cu(II), and possibly a small indication of Mn(II) [24], but the origin is not further investigated. The amplitude of the g' = 4.3 signal is inversely proportional to the logarithm of temperature, as expected from a paramagnetic contribution (data not shown). The other two narrow signals at g' = 2.0 and 5.8 follow qualitatively the same trend.

The lineshape of the broad signal, which is due to the magnetic moment of ferritin cores (Fig. 5.7a), is nearly Lorentzian at higher temperatures. Below 100 K, the shape becomes more asymmetric and is better fitted to a Gaussian shape. At 5–10 K, the amplitude of the broad signal has decreased to the point of being barely identifiable. The double integral of the broad EPR component, which reflects the total number of ferritin-core spins in the sample, increases with temperature, reaching a plateau at 100 K, followed by a slight decrease above 180 K (Fig. 5.7b). The increase in EPR signal amplitude with temperature is typical for superparamagnetic particles with an antiferromagnetic ground state [16, 23, 25, 45], and can be explained by the fact that only unblocked particles, whose fraction increases with temperature, contribute to the signal. Once all particles are unblocked, a paramagnetic behavior, characterized by a decrease of the EPR spectral amplitude with increasing temperature is observed. Therefore, the maximum value of the signal intensities (Fig. 5.7b), at $T \sim 100$ K, marks the transition from a regime of progressive unblocking of the magnetic moments to a regime where all moments are unblocked, thus representing the maximum blocking temperature of ferritin cores.



Figure 5.7: (a) EPR spectra acquired between 5 and 210 K (in order of ascending amplitude from lowest to highest temperature). Arrows point to the narrow signals centered at g' = 2, 4.3, and 5.8, respectively. (b) The double integral of the broad signal component in (a), as a function of measurement temperature.

EPR simulations provide further insights into the nature of the broad signals. For this purpose, the ferritin core is considered as a single large spin *S* resulting from the coupling of individual iron ions in the core [2, 52]. The corresponding spin Hamiltonian, used to describe the spectra, is given by

$$\mathcal{H} = \mu_{\rm B} \,\mathbf{S} \cdot \mathbf{g} \cdot \mathbf{B} + \mathbf{S} \cdot \mathbf{D} \cdot \mathbf{S},\tag{5.8}$$

with $\mu_{\rm B}$ being the Bohr magneton, **S** the electron spin operator associated with the spin number *S*, **g** the g-factor tensor, **D** the traceless zero-field splitting tensor, and **B** the applied magnetic field. In relation to magnetometry, the total spin of a particle can be expressed as [52]

$$S = \frac{\mu}{g\mu_{\rm B}},\tag{5.9}$$

with μ being the effective magnetic moment of the particle [52]. The zero-field splitting

is then expressed by

$$D = -\frac{g\mu_{\rm B}B_{\rm a}}{2S},\tag{5.10}$$

with B_a being the effective anisotropy field [44]. In the case of particles with uniaxial anisotropy, $B_a = 2KV/m$, with *K* being the anisotropy factor and *V* the particle volume.

Simulations based on a single component produce a poor fit to the data (Fig. A5.22), suggesting that at least two components are needed in order to capture the relevant spectral features. Automated two-component fitting approaches, however, yield unphysical results even in the case of limited parameter sets (see Appendix: "EPR alternative fitting approach"). This is because the line shape of high-spin systems such as ferritin does not depend in a simple manner on *D* and *S*. Furthermore, very different component combinations can fit broad line shapes equally well, so that meaningful fitting results depend critically on the initial parameters.



Figure 5.8: Selected EPR spectra (black lines) and their simulations (red dashed lines), at 30 K (a), 80 K (b), 130 K (c), and 190 K (d).

Physically meaningful initial parameters have been obtained at selected temperatures by visually matching the measured spectra with a set of components covering a wide range of values for *D*, *S* and the Gaussian broadening parameter H_{strain} (see Appendix: "EPR simulations of individual components"). Because of the excessive computation time required for simulating realistic values of *S* in excess of ~100, the scaling procedure of Fittipaldi *et al.* [2, 52]

$$S_{\text{real}} = S \cdot n, \ D_{\text{real}} = D/n, \ T_{\text{real}} = T \cdot n \tag{5.11}$$

has been used to relate realistic parameters to those used for fitting, through a scaling factor *n*. For instance, a simulated spectrum with S = 100 is obtained by rescaling a corresponding calculation performed for S = 10, using n = 10. The combination of two model spectra with g' = 2.01 and S = 10, which most closely reproduced a chosen experimental spectrum, served as an initial guess for the final optimization of the componentspecific parameters D and H_{strain} . Simulated spectra obtained with this procedure are in excellent agreement with experimental data at each measurement temperature (Fig. 5.8, see Fig. A5.23 for all temperatures). Details of the optimization procedure are explained in Material and Methods, and the optimized model parameters are listed in Table A5.6. Both components are centered at $B_0 = 336$ mT, but the first component (E1) is significantly narrower than the second one (E2) (Fig. 5.9a, see Fig. A5.24 for other temperatures). The temperature dependence of E1 and E2 is small compared with the respective error margins (Appendix, Fig. A5.25). A systematic variation of D, H_{strain} , and the relative contributions of E1 and E2 cannot be excluded, but the trend is not sufficiently well defined to support further interpretations.



Figure 5.9: Total EPR spectrum (solid line) and components E1, E2 (dashed lines) at 80 K.

5.4.2. DC SUSCEPTIBILITY AND HYSTERESIS

The superparamagnetic behavior of human-liver ferritin is well captured by FC-ZFC measurements (Fig. 5.10a). The curves bifurcate at $T_{b,max} \approx 24$ K, which corresponds

to the largest unblocking temperature of the particles. The ZFC data display a peak at $\hat{T}_{\rm b} = 10.5 \pm 0.5$ K, in agreement with an earlier characterization of ferritin [12]. The relation $\hat{T}_{\rm b} < T_{\rm b,max}$ is indicative of a distribution of blocking temperatures. The opening of the hysteresis loop below $\hat{T}_{\rm b}$ confirms the blocking process and the occurrence of magnetic irreversibility (Fig. 5.10b). The slight horizontal offset of the FC hysteresis loops highlights the presence of an exchange coupling field $B_{\rm ex} \approx 25$ mT, persistent at 5 and 25 K (Fig. 5.10d). This exchange field is similar to $B_{\rm ex} \approx 32$ mT reported for horse-spleen ferritin [12].



Figure 5.10: (a) ZFC-FC magnetization curves, measured at 5 mT. (b) Isothermal-induced magnetization measured at 150 K (full squares) and 5 K (empty circles). Both temperatures were reached in ZFC conditions. (c) Close-up of (b) centered on the origin, showing hysteresis opening at low temperature. (d) Detail of ZFC (black diamonds) and FC (orange stars) hysteresis around the origin, measured at 5 K in a maximum field of 0.3 T.

5.4.3. AC SUSCEPTIBILITY

The in-phase AC susceptibility $\chi'(T)$ (Fig. 5.11) shows a broad peak that shifts towards higher temperatures upon increasing the frequency of the AC field [84]. In-phase measurements have been fitted to a model derived from Gittleman *et al.*[85], while the imaginary part was ignored, due to its low SNR (see Appendix, Fig. A5.35). Full $\chi'(T)$ curves were calculated by integrating the analytical expression [85] for χ' over a distribution $\Gamma(E_b)$ of anisotropy energy barriers E_b , thereby relaxing any assumption about the analytical dependence of E_b on the particles volume distribution:

$$\frac{\chi'(T;\omega)}{\chi_1} = 1 + \frac{1}{\beta \langle E_b \rangle} \int_0^{k_{\rm B} T \ln(r_0 \omega)} \left(\frac{E_b}{3aT} - 1\right) \Gamma(E_b, \mu_E, \sigma_E) dE_b$$
(5.12)

where $\omega = 2\pi v$ is the AC frequency in rad/s, τ_0 is the inverse attempt frequency of thermal activations, χ_1 is the susceptibility in the blocked state, $a = \langle \sin^2 \phi \rangle / 2$, where the average is over all angles ϕ between easy axis and field, with a = 1/3 in case of random orientations, and β is a scaling factor with a theoretical value of 1 for blocked particles described by the model of Stoner and Wohlfarth [81]. We note that the single-domain susceptibility of blocked particles is not temperature-independent as assumed in the fitting equation, since it depends on the temperature-dependent anisotropy constant and spontaneous magnetization. Therefore, this remains a coarse approach to determine $E_{\rm b}$. Equation 5.12 was used to fit the AC susceptibility measurements using the Gamma function $\Gamma(E_{\rm b}, \mu_E, \sigma_E)$ with mean μ_E and width parameter σ_E as energy barrier distribution [55]. The attempt time was fixed to 9 ps, based on reported AC susceptibility ferritin data [32].

The mean and standard deviation of the energy-barrier distribution used to fit the data (eq. 5.12) are 144.74 K and 57.02 K, respectively. Note that the scaling parameter, β , decreased by 30%, upon decreasing the frequency.

5.4.4. EQUILIBRIUM MAGNETIZATION

Isothermal magnetization curves acquired in fields up to 7 T at temperatures ≥ 15 K (Fig. 5.12a) are close to equilibrium: the residual hysteresis opening at 15 K is ~3% of M_s , and drops to ~0.5% at 27 K. A modified Langevin model of the form $M(B) = \mathcal{S}(B, f) + \chi B$ was used to fit the equilibrium magnetization, where \mathcal{S} is a superparamagnetic term resulting from the partial alignment of unblocked magnetic moments with distribution f in the applied field, and χB is a linear term that includes superantiferromagnetic and paramagnetic contributions. In case of isotropic particles with identical magnetic term is proportional to the Langevin function $\mathcal{L}(\xi) = \operatorname{coth} \xi - \xi^{-1}$ with $\xi = mB/k_BT$, where k_B is the Boltzmann constant, and T the absolute temperature [32, 34]. This model reproduces the equilibrium magnetization calculations for canted spins discussed in section 5.3.6 (Fig. 5.5).

Single-particle magnetic anisotropy decreases the magnetic moment alignment of mechanically blocked, randomly oriented particles as soon as the linear regime of \mathscr{L} is left [60, 86, 87]. In the limit case of infinite uniaxial anisotropy, particles possess only



Figure 5.11: AC-magnetic moment and susceptibility of human-liver ferritin, probed in the subkHz frequency range. Top Panel: normalized in-phase susceptibility data (circles) and fit (solid line) to equation 5.12. Bottom Panel: Residuals of the fit.

two magnetic states with m parallel or antiparallel to the easy axis, in which case \mathcal{L} is replaced by $\mathscr{G}(\xi) = \langle \cos\phi \tanh(\xi \cos\phi) \rangle$, where $\langle . \rangle$ denotes the average of individual particle contributions over all angles ϕ between easy axes and field. This function was originally proposed by Néel [62], and used by Gilles [43] as a model for ferritin superparamagnetism. \mathscr{L} and \mathscr{G} have the same slope at B = 0, but their $B \to \infty$ limits are 1 and 1/2, respectively. Particles of volume V and finite anisotropy constant K are characterized by intermediate equilibrium magnetization functions comprised between $\mathcal L$ and \mathscr{G} , whose shape is controlled by the anisotropy parameter $\kappa = KV/k_{\rm B}T$. Unfortunately, these functions cannot be expressed analytically, so \mathscr{L} or \mathscr{G} are used instead, regardless of the effective particle anisotropy [12, 32, 57, 69, 87]. The Langevin function is a good approximation of the superparamagnetic behavior for $\kappa < 2$ (maximum error: 2%), and still considerably better than \mathscr{G} for $\kappa < 10$ (maximum error: 20%). Using $K = 18.3 \text{ kJ/m}^3$ for our ferritin sample (see section 5.5.3) and the volume of spherical cores with a diameter of 7 nm (Fig. 5.1), the $\kappa < 2$ and $\kappa < 10$ conditions are fulfilled for T > 100 K and T > 24 K, respectively, which means that \mathscr{L} is a valid model for all measurements shown in Fig. 5.12a, except the 15 K one.



Figure 5.12: (a) Measured isothermal magnetization curves (dots) and corresponding best fits with eq. (5.13) (lines), at temperatures indicated by numbers. Residuals, defined as the difference between measurements and model, normalized by the measured magnetization at 7 T, are plotted below. (b) Same as (a), after subtracting the modeled linear term from each curve. The 15 and 20 K magnetizations have been multiplied by 1.13 and 1.05, respectively, for better visualization. (c) Logarithmic mean of the magnetic moment distribution at measurement (dots, with 2σ error bars). The dashed line is the best-fitting antiferromagnetic magnon law obtained from > 50 K moment estimates, with $\mu_m(0) = 215.4 \ \mu_B$ and $\alpha = 0.0043$. (d) Non-paramagnetic susceptibility χ from the χB term in eq. (5.13) (black dots with 2σ error bars, left axis), and $1/\chi$ (red dots with 2σ error bars, right axis). The dashed line represents the best-fitting Curie-Weiss law above 150 K, with $\Theta = -194$ K.

Implementation of the superparamagnetic term for a single-valued magnetic moment yields large model residuals with systematic trends (Appendix, Fig. A5.34), indicating that the real moment distribution is a broad function. Therefore, \mathcal{L} needs to be integrated over the moment distribution f(m), usually assumed to be a lognormal function with unknown logarithmic mean μ_m and logarithmic standard deviation σ_m [43, 69]. Furthermore, the presence of paramagnetic spins, suggested by EPR measurements, requires to split the linear term of the fitting function into a term associated with the ferritin cores, and another term for the paramagnetic contributions. The resulting model is given by

$$M(B,T) = \alpha \mu_m \int_0^\infty \mathscr{L}\left(\frac{\mu_B m}{k_B T}\right) f(m,\mu_m,\sigma_m) \,\mathrm{d}m + \chi(T)B + \eta \mathscr{B}_J\left(J\frac{g\mu_B B}{k_B T}\right),$$
(5.13)

where α is a scaling factor, such that $M_s = \alpha \mu$ is the saturation magnetization of the superparamagnetic particles, χ is the high-field magnetic susceptibility of the cores, and \mathscr{B}_J is the Brillouin function describing the magnetization of paramagnetic spins with total spin quantum number J and magnetic moment $g\mu_B$. The scaling factors α and η account for the unknown concentration of ferritin cores and paramagnetic spins, respectively. The use of $M_s = \alpha \mu$ ensures that the saturation magnetization has the same temperature dependence as the magnetic moment, as expected for a superparamagnetic term is justified by the identification of a corresponding EPR component with g' = 4.3, originating from mononuclear Fe³⁺ ions with J = S = 5/2. This EPR component covers a large field range and is therefore dominant over other paramagnetic contributions with g' = 2 and 5.8. In order to avoid model instabilities caused by the similar shapes of the Langevin and Brillouin functions, the shape of the latter has been fixed using J = 5/2 while maintaining g unconstrained.

All M(B) curves in the 15–250 K range have been fitted globally, that is, with common temperature-independent parameters α , η , σ_m , g, and one set of temperature-dependent parameters μ_m and χ for each curve. The assumption that σ_m does not depend on temperature is justified by the fact that the width of a broad moment distribution is relatively insensitive to possible differences between the temperature dependencies of small and large magnetic moments. Parameter confidence intervals have been calculated using a Monte Carlo error estimation, which consisted in adding random errors to the data, based on the standard deviation of the random component of model residuals. Residuals are comprised between $\pm 0.5\%$ of the maximum magnetization at 7 T (Fig. 5.12a), with a common field-dependent trend limited to $\pm 0.2\%$ and a random component associated with measurement errors. The small systematic misfit might be caused by a nonlognormal distribution of magnetic moments, by deviations from the Langevin model due to single particle anisotropy, or by a small field dependence of χ . An almost complete saturation of the superparamagnetic contribution in the 7 T maximum field is attained at the lowest temperatures (Fig. 5.12b), meaning that the moment distribution can be recovered from the data, up to a small fraction of the smallest moments, whose magnetization saturates in larger fields.

The temperature dependencies of μ_m and χ (Figure 5.12c,d) are qualitatively similar to those obtained by Makhlouf et al. [12] and Gilles et al. [57]. The maximum mean moment $\langle m \rangle = \exp(\mu_m + \sigma_m^2/2) \approx 333 \,\mu_B$ is slightly smaller than the single-valued estimate of ~ $350\,\mu_{\rm B}$ obtained from horse-spleen ferritin using a simple Langevin fit with linear term [12]. The temperature dependence of μ_m is characterized by two opposed trends: a $\sim 10\%$ increase over the 15–27 K range, followed by a quadratic decrease compatible with the bulk antiferromagnetic magnon law, $\mu_m(T) = \mu_m(0)(1 - aT^2)$ (Fig. 5.12c) [69]. The initial increase $\mu_m(T)$ is likely an artifact of the Langevin model. As previously mentioned, the equilibrium magnetization of particles with finite anisotropy becomes proportional to $\mathscr{G}(\xi)$ at $T \to 0$. In a Langevin fit, this function is approximated by ~0.5 $\mathscr{L}(2\xi)$. Because $\xi \propto m$, the apparent moment obtained from the Langevin fit decreases as the appropriated model function changes from $\mathscr{L}(\xi)$ to $\mathscr{G}(\xi)$. The non-paramagnetic susceptibility γ decreases monotonically with temperature, approaching a Curie-Weiss law with $\Theta_{CW} \approx -194$ K above ~100 K (Fig. 5.12d). Deviations from this trend are expected in the case of AF nanoparticles, because of superantiferromagnetic contributions arising for instance from uncompensated spin planes [41]. Spin frustration might also contribute to χ , as seen by similar temperature dependencies encountered in systems dominated by this effect [88].

5.4.5. COERCIVITY DISTRIBUTIONS

IRM acquisition curves $M_r(B)$ describe the acquisition of a remanent magnetization from an initially demagnetized state after the application of increasingly large fields, until the so-called saturation remanent magnetization M_{rs} is reached. Only particles that are blocked over the time needed to zero the magnetic field and measure the magnetic moment (about a minute in the case of MPMS measurements) contribute to the IRM. Accordingly, the amplitude decrease of $M_r(B)$ with increasing temperature is caused by the progressive unblocking of magnetic moments (Fig. 5.13), and $M_{rs}(T)$ is the integral of a blocking temperature distribution probed by remanent magnetization measurements.

IRM curves become non-monotonic around 20 K, with an inflection around 0.15 T. The negative slope section denotes a low-coercivity phase that acquires a negative remanent magnetization. Because of the nature of the IRM acquisition protocol, which is formally equivalent to a partial hysteresis between B = 0 and B_{irm} , negative $M_r(B)$ slopes must be associated with inverted hysteresis [89], a phenomenon that arises from the exchange coupling between phases with different coercivities. These phases can be detected by fitting $M_r(B)$ with a linear combination of model curves representing their individual contributions:

$$M_{\mathbf{r}}(B) = \sum_{i} M_{i} F(B, \mathbf{p}_{i}), \qquad (5.14)$$

where M_i is the saturation remanent magnetization of the *i*-th component, and *F* the primitive of a model function *f* used to represent the corresponding coercivity distribution [90, 91], whose shape is controlled by a set of parameters **p**. Assemblages of magnetic particles with uniform composition, size, and shape are characterized by unimodal coercivity distributions, which, on a logarithmic field scale, are well approximated by a normal distribution $\mathcal{N}(\log B, \log \mu_B, \sigma_B)$ with logarithmic mean μ_B and standard devia-



Figure 5.13: IRM acquisition curves at selected temperatures (dots), model components used to fit the data (solid curves labeled by component number), and modeled total magnetization (unlabelled gray curve). The shaded band around each component corresponds to the 1-standard deviation uncertainty obtained from bootstrap simulations of measurement errors. Model residuals, expressed in percent of the maximum magnetization at 5 T, are plotted below.

ion σ_B [90, 92], or, more often, by slightly left-skewed generalizations of the normal distribution [91, 93]. Depending on the distribution skewness, μ_B is more or less close to the median acquisition field $B_{1/2}$, which is the field required to acquire half of the saturation remanent magnetization.

Coercivity distributions do not have necessarily an intrinsic physical meaning, being just defined as the first derivative of magnetization curves. A notable exception is represented by uniaxial single-domain particles described by the Stoner-Wohlfarth model [81]. In this case, f(B) represents the relative contribution of particles with switching field $B_{sw} = B$ to M_{rs} . Ferritin behaves as an assemblage of non-interacting Stoner-Wohlfarth particles, as seen from the identity between the shape of IRM acquisition curves and curves obtained by applying the same protocol to samples with a previously imparted negative saturation remanent magnetization [94, 95]. Well below the blocking temperature, the switching field is related to the anisotropy field $B_a = 2KV/m$ by $B_{sw} = \zeta B_a$ with $\zeta \approx 0.524$, if particles are randomly oriented [96].

A linear combination of four coercivity components of the form

$$F(B) = \Theta_1(T) \mathscr{S}(\frac{B}{B_{1/2}\Theta_2(T)})$$
(5.15)

has been used to model IRM acquisition curves at selected measurement temperatures. In eq. (5.15), $\mathscr{S}(x)$ is a sigmoidal function with $\mathscr{S}(0) = 0$ and $\mathscr{S}(\infty) = 1$, based either on the Langevin or the tanh function, $\Theta_1(T)$ is a function describing the cumulative blocking temperature distribution of the corresponding component, with $\Theta_1(0) = 1$, $B_{1/2}$ is the median acquisition field at 0 K, and $\Theta_2(T)$ is a monotonically decreasing function describing the decline of $B_{1/2}$ with temperature, caused by thermally activated moment switching[96]. The effects of exchange coupling between a higher- and a lower-coercivity component are modelled by multiplying F(B) of the lower-coercivity component with a smoothed sign function, centered at the mean value of the exchange field, which roughly coincides with the inflection point of the IRM curve (Appendix: "Equilibrium magnetization models").

Below ~15 K, the IRM is dominated by two coercivity components, C1 and C2, which contribute to 96% of the total $M_{\rm rs}$. The blocking temperatures of the other two components, C3 and C4, are much larger than those of the bulk sample, contributing mainly to the IRM curves acquired at 20 and 25 K. In the case of C4, the maximum $T_{\rm b}$ is close to that of magnetoferritin [97]. C1 and C2 are characterized by slightly different temperature and field dependencies, with $B_{1/2} \approx 1$ T at 3 K. C3 and C4 are characterized by strongly contrasting median acquisition fields, with $B_{1/2} \approx 3.2$ T and ~0.12 T, respectively, at 20 K. C3 is heavily unsaturated at 5 T. Extrapolation of the model function used to fit this component to higher fields suggests that saturation occurs above 50 T (Fig. 5.14b), similarly to what has been reported for goethite [98]. The field dependence of C4 is within the range that can be expected from ferrimagnetic minerals: equidimensional magnetite and maghemite nanoparticles with sizes above 4 nm are characterized by coercivities of 40–50 mT at 5 K [99, 100]. The larger median field of C4 might be explained by additional contributions from shape and surface anisotropy, in case of smaller, irregular crystals



Figure 5.14: (a) Logarithmic switching field distribution of Components 1+2 at 3, 5, 9, 13, and 17 K (solid lines in order of decreasing amplitude), and the predicted temperature dependence of the peak for particles with randomly oriented uniaxial anisotropy axes (dashed line). (b) Same as (a) for Components 3 and 4. (c) Fraction of blocked magnetic moments for Components 1+2, 3, and 4, as a function of temperature.

partially replacing the Fh core.

C3 and C4 are coupled by an exchange field $B_{ex} \approx 82$ mT. At fields $\ll B_{ex}$, C4 acquires a significant fraction of its saturation remanent magnetization, while C3 is still close to its initial demagnetized state, owing to its much larger coercivity. As C3 becomes progressively magnetized in the positive direction, negative exchange coupling causes C4 to be switched to the opposite direction, leading to a decrease of the total IRM. When C4 is negatively saturated, around 200 mT, the total IRM starts to increase again, due to the continuing IRM acquisition of C3. The non-monotonic IRM acquisition characteristics of these two negatively coupled components become clearly visible above 20 K, when C1 and C2 are almost completely superparamagnetic, no longer contributing to the remanent magnetization.

While the identification of C3 and C4 with independent entities is justified by the exchange coupling signature, the existence of C1 and C2 as independent components, instead of a single component, might just reflect the need to use two model functions to describe the complex shape of a single coercivity distribution [91]. Because of their similar field and temperature dependencies, C1 and C2 are merged into a single component, labeled as C1+2. C1+2 is characterized by a very broad coercivity distribution, which extends over ~ 4 orders of magnitude (Fig. 5.14a), and a maximum blocking temperature of ~20 K (Fig. 5.14c), which is close to the merging point of FC-ZFC low-field magnetization curves (Fig. 5.10a). The temperature dependence of $B_{1/2}$, which can be identified with the coercivity distribution peaks in Fig. 5.14a, is well described by the thermal activation model of Egli and Lowrie [101] for the switching field of randomly oriented single-domain particles with uniaxial anisotropy energy $mB_a/2$, when $B_a = 2B_{1/2}(T = 0)$ is taken from the extrapolation of the IRM fitting model to 0 K, and $m = 325 \,\mu_{\rm B}$ is assumed (dashed line in Fig. 5.14a). The required magnetic moment is close to the mean value of ~ 333 $\mu_{\rm B}$ derived from the Langevin model of isothermal magnetization curves. Consideration of the random particle orientation is very important, as simpler thermal activation models based on aligned anisotropy axes [102] require unrealistically large moments of the order of ~1000 $\mu_{\rm B}$ to fit the distribution maxima in Fig. 5.14a.

5.5. DISCUSSION

In this work, we present a comprehensive investigation of human-liver ferritin by indepth electron paramagnetic resonance and an extensive set of magnetometry techniques. The goal is to determine the spin structure of ferritin in order to elucidate the composition of the ferritin core in terms of magnetic phases. Figure 5.15 shows the combination of techniques used to determine the properties of ferritin cores and produce a model for their spin configuration. Magnetometry measurements yield the magnetic moment, blocking temperature, and energy barrier distributions, while EPR provides an important constraint on the paramagnetic contribution to M(B) curves, as well as independent estimates of the anisotropy field and the blocking temperature over a much shorter time range of the order of 0.1 ns. Comparison with magnetometric blocking temperatures permits to verify the Néel-Arrhenius law and estimate the attempt time of thermal activations. The volume distribution of ferritin cores is obtained from TEM observations and yields, in combination with the anisotropy field distribution, an estimate of the magnetocrystalline anisotropy constant. The volume and magnetic moment distributions provide also important constraints on the spin configuration of ferritin cores.

The combination of this unusually broad set of experimental data shows that several of the previous approaches to interpret ferritin data give inconsistent results, requiring a new theoretical model to fit the data and derive the magnetic properties of this ironoxide nanoparticle. One of the crucial findings is that the distribution of magnetic parameters that do not reflect intrinsic properties of the core material, such as μ (*m*) and B_a , need to be taken into account explicitly.

The following discussion starts with the properties of ferritin derived from the EPR analysis and continues with key parts of the analysis of magnetometry results. The latter leans heavily on sections 5.3.6, 5.3.7, and 5.3.8, which describes the framework on which the interpretation is based. The discussion is concluded by the joint interpretation of EPR and magnetometric models, leading to the description of the spin structure of the core of human-liver ferritin.



Figure 5.15: Schematic representation of different techniques used for ferritin characterization and their use for the determination of relevant properties. Abbreviations: PM — paramagnetic contribution, exchange — exchange coupling between magnetic components, spin conf. — spin configuration. For the rest of the symbols, the reader is referred to the main text.

5.5.1. EPR SIMULATIONS

In this section, we describe the EPR spectra of ferritin, which are characterized by broad signal attributed to the core material (Fig. 5.7). Compared to standard EPR signals, ferritin spectra are extremely broad and the changes with temperature are small, thus lacking clearly resolved features. The several hundred mT width and the overall Gaussian shape at temperatures smaller than 100 K suggest an ensemble of ferritin cores with slightly different properties resulting in a distribution of EPR parameters. To analyze the EPR spectra, we use a quantum-mechanical description of the magnetic properties of ferritin cores, and in particular, we focus on the lineshape and its temperature dependence. This approach allows us to directly obtain the spin structure of ferritin from the simulated EPR spectra. For other approaches see the Appendix.

There are several challenges associated with analyzing the EPR spectra. For high-spin systems such as ferritin, variations in the lineshape due to the spin state *S* and the zero-field splitting *D* are not systematic, making it challenging to predict which parameters should be used in the simulations of the EPR spectra. Therefore, equivalent simulated spectra can be obtained by many different sets of EPR parameters, such as *S* and *D*, defying standard optimization methods. As an illustration of this problem, an attempt to perform an automatic fit by varying just a few parameters did not lead to a global minimum (see Appendix: "EPR alternative fitting approach"). Therefore, a different approach based on the use of carefully chosen model spectra (see section 5.4.1) was used, as discussed in the following.

A minimum of two components are required to simulate the experimental spectra in a satisfactory manner (Fig. A5.22 vs. Fig. 5.8b). Simulations with more than two components were not attempted, to avoid an excessive number of free parameters. No assump-

tions were made about the temperature behavior, such as, for example, that the relative weight of the components or the D values have to be constant for all temperatures, even if these assumptions might be justified by certain models.[2] The model parameters for the two components E1 and E2 were selected from simulated spectral lineshapes obtained for a range of D and S values (see Appendix: EPR Model Simulations, Fig. A5.30). A summary of the parameters used in the final simulations is given in the Appendix, Table A5.3 and Fig. A5.25. The temperature dependence of the parameters does not exclude a systematic variation of D with temperature, but the trend is not sufficiently well defined to support further interpretations.

How different are the two components E1 and E2 chosen for fitting EPR measurements? For all temperatures, D_1 is 2.5 times smaller than D_2 , on average. However, D and S are not independent but inversely related (eq. 5.10), so that a change in D can be compensated by the inverse change in S without changing the overall shape (see Appendix: EPR S-D inverse compensation). For instance, in the case of equal D values for both components, S_1 must be ~2.5 times smaller than S_2 to obtain the same total EPR spectrum. The two components do not necessarily represent two distinct families of ferritin, rather they should be considered as a mathematical construct to represent an overall broad distribution. The hypothesis of an underlying broad distribution is supported by the fact that the Gaussian broadening used in the simulations exceeds the D parameters by at least an order of magnitude, which can only be explained by a distribution of ferritincore spin configurations within the ferritin population. The overall Gaussian lineshape of the EPR signal below 100 K reveals an inhomogeneous broadening typical of a resonance that consists of centers with a distribution of anisotropic magnetic parameters. The resonance narrows at higher temperatures, yielding an overall Lorentzian lineshape above 100 K, revealing that a dynamic process averages the differences in the anisotropy of the different centers. Such a line narrowing at higher temperatures was attributed to anisotropy averaging [48] or, alternatively, anisotropic melting [49, 50].

SCALING MODEL OF EPR PARAMETERS

The real value of *S* for ferritin cores must be much larger than S = 10 used in our simulations. The scaling approach proposed by Fittipaldi *et al.* [2] (eq. 5.11) permits to find equivalent parameters S_{real} and D_{real} corresponding to more realistic magnetic moment estimates, while maintaining the same lineshape. Two examples with scaling factors n = 10 and n = 30 are given in Table 5.1.

EPR results can be compared with ferritin properties reported in the literature by deriving μ and B_a from S_{real} and D_{real} through eq. (5.9) and eq. (5.10), respectively. Our range of estimates for μ is within the range published by Brem *et al.* [29] (128-556 μ_B) and by Koralewski *et al.* [103] (133-239) μ_B . On the other hand, our estimated interval of 0.1–0.3 T for B_a is independent of scaling, and agrees well with the $0.08 \le B_a \le 0.27$ T range reported in the literature [15, 104] for 5.4–7.0 nm ferritin cores. Previous EPR studies on horse-spleen and human-spleen ferritin focused on describing qualitative aspects of the EPR lineshape (EPR intensity and width variations vs. temperature and different ferritin samples), all agreeing on the broad signal located at g' = 2.0 and its observed temperature dependence[15, 16, 23, 25, 26, 105]. However, a few EPR spectra present a

more asymmetric lineshape, especially at lower fields[25, 26], the shape of which does not agree well with our spectra.

Table 5.1:	EPR parameter scaling and c	orresponding magnetic	: moments μ for 2	T = 20 K. n is the
scaling fac	tor and B_a the scale-independent	lent anisotropy field.		

	E1 ($B_a = 0.1 \text{ T}$)		E2 ($B_a = 0.3 \text{ T}$)		E1 & E2
n	S_{real}	D_{real} (MHz)	S_{real}	D_{real} (MHz)	μ ($\mu_{ m B}$)
1	10	-180	10	-450	20
10	100	-18	100	-45	200
30*	300	-6	300	-15	601

*maximum scaling factor, see text.

5.5.2. FERRITIN PROPERTIES DERIVED FROM MAGNETOMETRY

Combined analysis of M(B) curves, AC susceptibility, and IRM curves highlights the inconsistency of models used in the literature to interpret the magnetic properties of ferritin (see sections 5.3.6, 5.3.7, and 5.3.8). IRM acquisition curves (Fig. 5.13) support the existence of multiple magnetic phases. The main phase, which we attribute to Fh cores, has a median blocking temperature of ~8 K and contributes to ~96% of the remanent magnetization at 3 K. The remaining magnetization is carried by a magnetite-like low-coercivity phase (~0.6%), exchange-coupled with a high-coercivity phase, which become dominant above ~ 17 K. The very large saturation field of the high-coercivity phase is compatible with goethite nanoparticles [98], and possibly also wüstite [106]. The contribution of these two secondary phases in our sample is negligibly small, compared to estimates obtained with diffractometric techniques (e.g., 30% magnetite in horse-spleen and human-liver ferritin).[8]. While these secondary phases must coexist within the same protein shells due to their exchange coupling, it is not clear whether they represent fully altered ferritin cores, or if they coexist with Fh as intermediate alteration products. In any case, non-Fh phases detected with the analysis of IRM curves are negligible. The main IRM component exhibits a magnetic behavior explainable with a fixed total magnetic moment, whose distribution is given by the Langevin model described in section 5.4.4. This does not exclude that this component consists of two or more phases with a sufficiently strong magnetic coupling, such that the vector sum of their magnetic moments does not change during isothermal magnetization measurements.

Additional proof for the above conclusion is provided through the analysis of the relations existing between blocking temperature, magnetic moment, and anisotropy field distributions. In the case of single-domain particles with a fixed spin configuration, these relations are established by the Néel-Arrhenius model (eq. 5.7). We start with the normalized temperature dependence of the saturation remanent magnetization $M_{\rm rs}$ corresponding to the sum of the IRM components C1 and C2, which we attributed to Fh (Fig. 5.16). The normalized saturation remanent magnetization $M_{\rm rs}(T)/M_{\rm rs}(0)$ of C1+2, coincides, by definition, with the integral of the blocking temperature distribution $f_{\rm b}(T)$ of the Fh cores. Assuming the anisotropy constant *K* to be the same for all ferritin cores, one can expect from eq. (5.7) that *V* is proportional to $T_{\rm b}$, so that a rescaled version $cf_V(cV)$ of the volume distribution $f_V(V)$ obtained from TEM statistics (Fig. 5.1) will match $f_b(T)$ for a certain value of the proportionality constant $c = V/T_b$. Indeed, a good match with the IRM component C1 is obtained using $c \approx 20 \text{ nm}^3/\text{K}$ (Fig. 5.16). The match is less good if one considers C1+2, because of the wider composite blocking temperature distribution. It is therefore possible that C1 and C2 represent two distinct groups of ferritin cores with a narrower and a wider anisotropy distribution, respectively, so that the narrow anisotropy distribution of C1 is well approximated by a mean *K* value. In this case, $K = c^{-1}k_B \ln t/t_0 \approx 19 \text{ kJ/m}^3$ is obtained for C1 using $\tau_0 = 10^{-11} \text{ s}$ [84] and t = 5 s for the IRM measurement time. Comparable values have been reported in the literature, e.g., $K \sim 25 \text{ kJ/m}^3$ for horse spleen ferritin [40], and $K \sim 17 \text{ kJ/m}^3$ for bacterial ferrihydrite [70]. A slightly worse match is obtained when a similar rescaling procedure is applied to the distribution of E_b derived from susceptibility measurements (Fig. 5.16). In this case, the lack of contributions at temperatures smaller than 4 K reflects the shape of the gamma distribution used to fit the measurements. The relatively well-constrained value of the bulk anisotropy *K* suggests that most Fe ions are embedded in a crystalline structure.



Figure 5.16: Cumulative blocking temperature distributions reconstructed from IRM measurements (Comp. 1 and Comp. 1+2), from low-field susceptibility measurements (Susceptibility), and from $E_{\rm h} = KV$ with $K \approx 19$ kJ/m³ (Volume).

Our surface-spin model of ferritin (section 5.3.8) predicts that the distribution of $\ln(m)$, which we assumed to be Gaussian when fitting M(B) curves, is given by $g_m = g_{\kappa\mu\eta} * g_{V^{2/3}}$, where $g_{\kappa\mu\eta}$ and $g_{V^{2/3}}$ are the distributions of $\ln(\kappa_s\mu_{Fe}\eta_s)$ and $\ln V^{2/3}$, respectively, and "*" is the convolution operator. Because κ_s and μ_{Fe} are fixed quantities, and the standard deviation $\sigma \approx 0.08$ of $\ln V^{2/3}$ is much smaller than $\sigma \approx 0.96$ for $\ln(m)$, the moment distribution is controlled mainly by the degree of spin canting $\eta_s = \sin(\epsilon_s)$, and thus, by the surface anisotropy. The distributions (Appendix, Figure A5.39) is almost perfectly Gaussian, owing to the fact that g_m used in the Langevin model was a normal distribution. About 96% of the distribution is comprised between $\eta_s \approx 1\%$ and ~50%, which is a reasonable upper limit for the alignment of surface spins.

Finally, we compare the anisotropy-field distributions obtained from IRM acquisition
curves and from the magnetic moment and volume distributions, respectively. In the first case, we identify the distribution of B_a with the IRM coercivity component C1+2, using the relation $B_{sw} = \zeta B_a$ between B_a and the switching field of single-domain particles well below the blocking temperatures. In the case of randomly oriented particles with uniaxial anisotropy [96], $\zeta \approx 0.524$. In the second case, the distribution of $\ln(B_a)$ is given by $g_a = g_{2K/\kappa\mu} * g_{V^{1/3}} * g_{\eta^{-1}}$ (section 5.3.8). A good match between the two estimates is obtained using $B_a = 1.8B_{sw}$ (Fig. 5.17). The corresponding $\zeta \approx 0.56$ is slightly larger than the value expected for randomly oriented uniaxial single-domain particles and might be explained by a single-particle anisotropy with a small degree of non-uniaxiality.



Figure 5.17: Comparison between a direct estimate of the anisotropy field distribution obtained from $B_a = 1.8B_{sw}$, with B_{sw} being the switching field distribution of the IRM Component 1+2 at 3 K, and a reconstruction based on the magnetic moment model explained in the text.

In summary, the modified Langevin model used to fit M(B) curves (section 5.4.4), the lack of a spin-flop transition (section 5.3.6), the magnetic moment distribution (Sections 5.3.7 and 5.3.8), the coercivity distribution (section 5.4.5) and the compatibility of volume, magnetic moment, blocking temperature, and anisotropy field distributions with the Néel-Arrhenius model (section 5.3.8, Figure 5.17) point, altogether, to a model for the spin structure of ferritin cores where the magnetic moment arises from surfacespin canting, rather than randomly distributed defects in the AF sublattices. The ferritin cores behave as single-domain particles with uniaxial magnetocrystalline anisotropy. The wide distribution of m is explained by the variability of spin configurations deviating from the two-sublattice AF model, rather than the volume distribution. Despite the strong heterogeneity of magnetic configurations, all ferritin cores appear to share a well-defined magnetic anisotropy constant K. This is consistent with a model where spins are rigidly coupled and switching occurs by uniform rotation, so that most of the work necessary to overcome the energy barrier originates from the intrinsic anisotropy of the AF sublattices. Finally, the wide distribution of switching fields, from ~ 0.03 to > 30T, is also a direct consequence of this model, which predicts $B_{sw} \propto m^{-1}$ for AF particles with a spontaneous moment *m*. In particular, $m \rightarrow 0$ yields extremely large switching fields, which are limited only by the sublattice exchange field $B_{\rm E}$.

The above model explains magnetometry measurements almost completely. As discussed in section 5.4.5 only ~4% of the total remanent magnetization is carried by phases that cannot be associated with a Fh core composition. M(B) curves contain a paramagnetic contribution that is carried by a small fraction (~6.5%, Table 5.2) of mononuclear Fe³⁺ atoms. This fraction of ~6.5% is obtained from the ratio $\alpha \mu_m / \eta$ between the saturation magnetization of these atoms and the Fh cores in eq. 5.13. The last magnetic parameter that describes the ferritin cores is the non-paramagnetic susceptibility χ that contributes with a linear term χB to the M(B) curves. This term originates from the field-induced spin canting of surface spins and of the inner spins with AF order. The estimate of χ at 50 K obtained from M(B) curves fits with eq. 5.13 is ~1.4 times larger than the value expected from the simple model of spin-canted AF nanoparticles discussed in section 5.5.3 (Table 5.2). Measurements of horse-spleen ferritin up to 50 T show that the slope of M(B) curves measured at 4.2 K continues to decrease up to ~35 T, before merging with the superantiferromagnetic trend [41]. It is therefore possible that the excess

Table 5.2: Magnetic properties of ferritin cores derived from magnetometry (M) and EPR (E) measurements. μ — spontaneous magnetic moment; σ_{μ} — standard deviation of the distribution of $\ln(\mu)$; B_a — anisotropy field; σ_B — standard deviation of the distribution of $\ln(B_a)$; B_c — coercive field of hysteresis; E_b — energy barrier; σ_E — standard deviation of the distribution of $\ln(E_b)$; K uniaxial anisotropy constant; σ_K — standard deviation of the distribution of $\ln(K)$; M_p/M_{Fh} — paramagnetic saturation magnetization, normalized by the saturation magnetization of the superparamagnetic contribution; N_p/N_{Fh} — number of paramagnetic Fe atoms, normalized by number of superparamagnetic atoms of ferritin core; χ/χ_c — non-paramagnetic susceptibility in M(B) fits with eq. (5.13), normalized by the value expected from superparamagnetic particles with spin canting moments (section 5.3.6).

Droporty	From	From	
rioperty	magnetometry	EPR	
$\mu/\mu_{\rm B}$	$\mu/\mu_{\rm B}$ 337 ^{a)}		
σ_{μ}	0.947 ^{b)}	—	
B_{a} (T)	1.9 ^{c)}	0.1–0.3 ⁿ⁾	
σ_B	1.44 ^{d)}	—	
$B_{\rm c}$ (T)	0.08 ^{e)}	—	
$E_{\rm b}/k_{\rm B}$	$227^{\text{ f}}$	_	
σ_E	0.679 ^{g)}	—	
<i>K</i> (kJ/m ³)	19 ^{h)}	_	
σ_K	0.417 ⁱ⁾	_	
$M_{\rm p}/M_{\rm Fh}$	2.39 ^{j)}		
$\dot{N_{\rm p}}/N_{\rm Fh}$	0.0645 ^{k)}	0.0036 ^{o)}	
$\dot{\chi}/\chi_{\rm c}$	$\sim 1.4^{1)}$	_	

a) Mean, Fig. 5.12c, 0 K b) Langevin fit c) Median of C1+2, 3 K d) C1+2, 3 K e) Hysteresis f) T_b of C1+2 g) T_b of C1+2 h) eq. 5.7 i) eq. 5.7 j) $\eta/\alpha\mu_m$ in eq. 5.13 k) $\eta \langle m \rangle / \alpha \mu_m N \mu_{\text{Fe}}$ in eq. 5.13 l) 50 K, Fig. 5.5b m) for *n* =10, 30, i.e., *S* = 100 and S = 300, respectively n) E1 and E2, 20 K o) for S = 300; for $S = 100 N_p / N_{Fh} = 4 \cdot 10^{-4}$ From Appendix, section "Ferritin core and mononuclear Fe(III) EPR intensities"

susceptibility in our fits of M(B) curves limited to a maximum field of 7 T is caused by the linear approximation of a small residual curvature due to superparamagnetic contributions that saturate in much larger fields. These contributions might require a magnetic moment distribution with a heavier left tail associated with much smaller Fh cores or isolated core fragments, as suggested by the measured volume distribution (Fig. 5.1).

5.5.3. Comparison between EPR and magnetometry

BLOCKING TEMPERATURE FROM EPR AND MAGNETOMETRY

According to the Results section (Fig. 5.7b), the maximum EPR blocking temperature, $T_b \sim 100$ K, is in agreement with values reported for horse-spleen ferritin [15, 45]. The blocking temperature values obtained from magnetometry, e.g., from the ZFC susceptibility maximum at ~10.5 K, are much lower than those obtained from EPR, owing to the drastically different characteristic timescales $t_{ZFC} \sim 100$ s and $t_{EPR} \sim 0.1$ ns. From the Néel-Arrhenius law

$$\alpha = \frac{T_{b,EPR}}{T_{b,ZFC}} = \frac{\ln(t_{ZFC}/\tau_0)}{\ln(t_{EPR}/\tau_0)}$$
(5.16)

the attempt time estimate $\tau_0 = (t_{EPR}^{\alpha}/t_{ZFC})^{1/(\alpha-1)} = 3.9$ ps is obtained, in good agreement with the range of values reported for ferritin [107] and other AF nanoparticles of similar size [108].

MAGNETIC MOMENT AND ANISOTROPY FIELD FROM EPR AND MAGNETOMETRY

As discussed above, it is not possible to perform a full inversion of EPR spectra to resolve the magnetic moment and anisotropy-field distributions needed for a quantitative comparison with magnetometry results. The amount of paramagnetic vs. ferrihydrite-like phases by EPR is obtained from the ratio of the intensities of the g = 4.3 signal and the broad EPR signal, resulting in 0.4% of mononuclear Fe(III) atoms (Table 5.2). The difference with respect to the magnetometry results (6.5 %) is dominated, from the EPR side, by the uncertainty in the spin quantum number S of the ferritin core (see Appendix, section "Ferritin core and mononuclear Fe(III) EPR intensities"). Also, only the mononuclear Fe(III) signal is taken into account, leading to a possible underestimation of the paramagnetic contribution from EPR. From the magnetometry side, small clusters of iron ions with a superparamagnetic contribution similar to the Brillouin function used to model the paramagnetic phase might lead to an overestimation of the paramagnetic contribution, whereas in EPR such clusters may escape detection due to broadening or unfavorable relaxation properties. The existence of small iron clusters or incomplete ferritin cores with a much smaller magnetic moment is supported by the < 4.5 nm tail of the core size distribution obtained from TEM (Fig. 5.1). In view of the above mentioned uncertainties in the determination of para- and superparamagnetic contributions, the agreement between EPR and magnetometry can be considered satisfactory. Nevertheless, it is still possible to verify the compatibility of the two EPR components E1 and E2 with magnetometric parameters. For this purpose, we plot the estimated μ and B_a ranges obtained from both techniques (Fig. 5.18). As discussed in section 5.5.3, an inverse relationship of the form $B_a = 2E_b/m$ holds between the anisotropy fields and magnetic moments of individual ferritin cores. Because the distribution of E_b is much narrower than those of B_a and μ , possible magnetic moment and anisotropy field combinations of individual cores lie close to a line with slope -1 in the bilogarithmic plot of Fig. 5.18. On the other hand, simulated EPR spectra sharing the same value of *SD* are characterized by almost identical shapes (Appendix, Fig. A5.26), owing to the scaling rules of Fittipaldi *et al.* [2, 52]. Because $SD = g\mu_B B_a$ (eq. 5.9–5.10), EPR spectra with same B_a are indistinguishable, which means that the magnetic moment is totally unconstrained. Accordingly, the EPR components E1 and E2 define two horizontal lines in Fig. 5.18. If these components are interpreted as the discrete representation of a broad distribution of EPR parameters, the corresponding B_a values (Table 5.1) can be interpreted as two discrete samples of a broad anisotropy field distribution.



Figure 5.18: Anisotropy field (B_a) vs. magnetic moment (μ) from EPR (components E1 and E2 at 20 K) and from magnetometry. Shading around the magnetometry and EPR trends correspond to the quartiles of T_B distribution (Fig. 5.16) and to the range of EPR parameters yielding similar simulated spectra, respectively. The dot represents the averages of B_a and μ obtained from magnetometry. The distributions of μ and B_a obtained from magnetometry are plotted on the corresponding axes.

The intersection between the constraints plotted in Fig. 5.18 gives a rough estimate of the anisotropy fields and magnetic moments probed by EPR, which are centered around $B_a \approx 0.2$ T and $\mu \approx 4000 \,\mu_B$, respectively. This combination of parameters is off by a factor

~10 with respect to mean values obtained from magnetometry ($B_a \approx 2 \text{ T}$ and $\mu \approx 337 \mu_B$). This discrepancy originates from the ~10 times smaller anisotropy fields of E1 and E2. A possible explanation for this difference is discussed in the next section.

ORIGIN OF THE LOWER EPR ANISOTROPY FIELD

As discussed in section 5.5.3, Ba estimates obtained from EPR spectra are about one order of magnitude smaller than the median B_a obtained from IRM acquisition curves, meaning that the two measurements probe distinct processes related to particle anisotropy. In the case of IRM acquisition curves, B_a is probed through the switching field $B_{\rm sw}$, defined as the field required to reverse the remanent magnetic moment of a singledomain particle. In the case of a sufficiently rigid coupling between spins, particles with uniaxial anisotropy possess only two antiparallel magnetic states, and B_{sw} is the field in which a transition occurs between these two states. Coherent rotation [81], fanning [109], and curling [110], are just some examples of such two-state models of singledomain particles. The type of reversal mechanism determines the dependence of B_{sw} on the angle between anisotropy axis and applied field, and thus the relation between the bulk B_{sw} of IRM acquisition curves and B_a , e.g. $B_{sw} \approx 0.524B_a$ in case of coherent rotation. Regardless of the reversal mechanism, transitions between the two magnetic states in IRM and EPR measurements are governed by the same equation for the particle energy: $E = -B_a \mu(\mathbf{e} \cdot \mathbf{n})^2 / 2 - B \mu(\hat{\mathbf{z}} \cdot \mathbf{n})$, where **e** and **n** are the unit vectors parallel to the easy axis and the magnetic moment respectively, and the applied field is parallel to \hat{z} . Accordingly, the same B_a is sensed by the two methods.

Surface anisotropy complicates the description of the particle magnetization by introducing intermediate magnetic states along the path that produces a field-induced reversal of the bulk magnetic moment [111]. In practice, the switching process begins by reversing discrete groups of surface spins in increasingly large fields, until the reversal of the internal core spins completes the process. As a result, single-particle hysteresis contains several discrete magnetization jumps corresponding to partial reversals [111], instead of the single jump at B_{sw} of two-state particles. The way multistep magnetic moment reversals are recorded depends on the measurement protocol. In the case of IRM acquisition curves, partial reversals of surface spins will not be recorded, because the strong exchange coupling with the internal core spins recovers the initial spontaneous moment as soon as the field is removed. Accordingly, switching occurs only in a field that is sufficiently strong to reverse the internal core spins. As shown in section 5.5.2, this field is given by $B_a = 2KV/m$, where K is the anisotropy constant of the AF-coupled core spins. On the other hand, the entire sequence of partial reversals is recorded by the hysteresis loop (Fig. 5.10). As a result, the coercive field of hysteresis, $B_c \approx 0.1$ T, is ~10 times smaller than the median switching field $B_{1/2} \approx 1$ T obtained from IRM curves. For comparison, randomly oriented Stoner-Wohlfarth particles [112] are characterized by $B_{1/2}/B_c \approx 1.2$. Like hysteresis, EPR spectra are expected to record all transitions between magnetic states, and therefore also partial reversals occurring at lower fields. The apparent anisotropy field of EPR spectra is therefore more similar to the coercive field of hysteresis than the field required for a complete reversal. This explains why a direct comparison of EPR and magnetometry data, as in Fig. 5.18, does not work.

The existence of multiple magnetic states for the ferritin cores questions the applicability of the Langevin model for fitting M(B) curves, since particles in the superparamagnetic state can undergo thermally activated transitions between any pair of states. Intermediate states with partially reversed surface spins have higher energies than the ground states, and the whole sequence of reversal steps is represented by a path in a multidimensional energy landscape, which connects a series of local minima distributed along a "valley" running from a ground state to its opposite [113]. The presence of local minima along this valley raises the probability of finding particles with these discrete intermediate states, compared to the continuous transition between ground states of particles with no intermediate states. Close to the blocking temperature, the probability of magnetic configurations different from ground states is negligibly small, and intermediate states do not play any role. At higher temperatures, the existence of multiple states with different energy levels has a similar effect as multiaxial anisotropy, and deviations from the Langevin law can be expected to be smaller than those produced by uniaxial anisotropy (section 5.3.6), owing to the smaller switching fields associated with transitions between intermediate states. Because the effect of uniaxial anisotropy is small in the temperature range of our M(B) measurements (Fig. 5.4), the Langevin model is ex-

pected to be a good representation of the equilibrium magnetization even in the case of particles whose magnetic moment is not reversed in a single step. Consequently, the Langevin model, as applied in the present context is a good approximation for the magnetic properties of ferritin.

5.6. CONCLUSIONS

In this work, we have probed the magnetic response of a sample of human-liver ferritin with a broad set of available experimental techniques and at different excitation frequencies, sparsely covering the DC-to-9 GHz interval. While magnetic analyses of ferritin are not new, our synergetic comparison of magnetometry and electron paramagnetic resonance offers, for the first time, an in-depth description of ferritin's spin behavior. Our methods reconcile recurring discrepancies on the origin of the magnetic moment in ferritin on the one hand, while offering a quantitative approach to the challenging determination of spin Hamiltonian components emerging from EPR data fitting on the other hand, over a temperature range extending close to the DC-blocking temperature.

With regard to EPR data, the broad and seemingly featureless nature of the spectra can be well captured by at least two components, with *S* and *D* parameters returning an anisotropy field in the 0.1-0.6 T range, and an *S* parameter above 50. These two components most likely represent a broad distribution of magnetic properties. While the magnetic moment can only indirectly be derived from the EPR data, the technique identifies also paramagnetic contributions, providing information complementary to magnetometry.

Moreover, a more complete understanding of the magnetometry data of ferritin, which has been the subject of extensive debate, can be achieved with IRM acquisition data. IRM acquisition curves reveal (i) the mineral composition of the protein core, which complements the information from other methods, such as energy-resolved electron microscopy; (ii) the blocking temperature distribution, in reasonable agreement with the particle volume distribution and the assumption of a single anisotropy constant; and (iii) the existence of a negative exchange field between the dominant antiferromagnetic phase and a minor ferrimagnetic phase within the same core. This exchange interaction prevents the magnetic moments of the two phases to display independent superparamagnetic behaviors.

From a new theoretical description of the induced magnetization data, we conclude that the magnetic moment of ferritin is controlled mainly by spin canting caused by surface anisotropy. The wide distribution of magnetic moments can be explained by the strong heterogeneity of surface-spin configurations, to which the EPR spectrum seems to be particularly sensitive. Because of the inverse proportionality relation between magnetic moment and anisotropy field imposed by the Néel-Arrhenius law, the coercivity of ferritin is also represented by a broad distribution.

5.7. APPENDIX**5.7.1.** TEM description



Figure A5.19: Transmission electron microscopy (TEM) characterization of human-liver ferritin. (a) TEM image of human-liver ferritin cores. (b) Stained-SEM image of human-liver ferritin particles allowing the protein shell visualization.

5.7.2. FERRITIN PURITY ASSESSMENT

NativeMark Unstained Protein Standard (Cat. No. LC0725) was obtained from Life Technologies Corporation. The purity and the homogeneity of the protein were assessed by a 7.5% non-denaturing polyacrylamide gel electrophoresis. Gel electrophoresis was run at 4°C and 90 V until samples reached the running gel, then the voltage was increased to 160 V for 3 hours. After electrophoresis, Coomassie blue staining was used to stain proteins. For iron staining, the gel was immersed in Prussian blue staining solution, which was freshly prepared by mixing 2% K₄Fe(CN)₆ and 2% HCl (1g Potassium ferrocyanide + 47,3 ml H₂O + 2,7 ml HCl) for 1h at RT, then 1h in water, changing water every 15-20 min. Finally, the gel was incubated in a solution containing 0.025% DAB (3,3'-Diaminobenzidine-Sigma) and 0.05% H₂O₂ in 1X TBS [12,5mg DAB resuspended in 500 µl DMSO, added to 50 ml TBS 1x, 75-180 µl H₂O₂ added just before incubation] for 15-30 minutes at RT to enhance the signal.

5



Figure A5.20: Purity assessment of human-liver ferritin. (a,b) Dynamic light scattering showing the particle size distribution: one main Gaussian peak with μ =12.2 nm and σ =4.9 nm. (c) Non-denaturing polyacrylamide gel electrophoresis of human-liver ferritin stained with Coomassie. The strong band of 500 kDa agrees with ferritin monomers (24-mer protein) while the weaker bands of 1.048 and 1.236 kDa correspond to ferritin multimers (dimers/trimers) or aggregates of ferritin protein. Markers refers to NativeMark Unstained Protein Standard (Cat. No. LC0725).

5.7.3. EPR BASELINE CORRECTION

The baseline correction of the spectra was performed by a home-made script implemented in Matlab 2019a, which automatically looks for a linear transformation (Fig. A5.21a), such that the slope of the baseline correction is the optimal value by which the high magnetic field values of the second integral of the spectra reach a plateau-like region (Fig. A5.21c), which is equivalent to having a zero first integral value at the highest field values (Fig. A5.21b).



Figure A5.21: Automatic baseline correction algorithm. (a) The experimental spectrum (in blue), the optimal line (red dotted line), and the transformed spectrum (yellow curve). (b) The first integral of the baseline-corrected spectrum. Its high field values reach zero. (c) The second integral of the baseline-corrected spectrum. The high field values reach a plateau-like region.

5.7.4. EPR SIMULATION WITH ONE COMPONENT

Figure A5.22 shows an example of one of the best fits obtained by considering a simulation with only one EPR component. It shows the discrepancy between the data and the model, indicating the need of two or more EPR components.



Figure A5.22: Simulation (light-blue line) of EPR spectrum (black line) with single component only (80 K). The simulation is performed with one spin component of *S* =15, *g* = 2.01, *D* = -335 MHz, and $H_{\text{strain}} = 6500$ MHz.



5.7.5. EPR SIMULATIONS, FROM 20 K TO 210 K

Figure A5.23: Simulations (red dotted lines) of EPR spectra (black lines) for all data recorded between 20 and 210 K. The simulations are performed with two spin systems, as described in the main text. The system parameters are shown in Table A5.3.



5.7.6. EPR SIMULATIONS OF INDIVIDUAL COMPONENTS

Figure A5.24: Total EPR simulations (red lines) performed at 30 K, 80 K, 130 K, and 190 K. The simulations are performed with two spin systems, as described in the main text. The first system component is depicted in green. The second component is shown in blue.

5.7.7. EPR SIMULATIONS PARAMETERS

Table A5.3: System parameters used for simulations of the EPR spectra. Simulation parameters: S = 10, *D*: axial zero-field splitting, H_{strain} : Gaussian broadening parameter. The parameters at 30 K, 80 K, 130 K, and 190 K were determined first.

	Component 1				Component 2	
T(K)	D (MHz)	H _{strain} (MHz)	Weight (%)	D (MHz)	H _{strain} (MHz)	Weight (%)
20	180	6000	7.7	450	11000	92.3
30	170	6000	9.1	400	10500	90.9
40	155	5500	9.5	370	10500	90.5
50	155	4600	10.0	370	10400	90.0
60	175	4500	10.5	300	11000	89.5
80	150	3900	11.1	370	8900	88.9
100	150	3900	15.4	370	8700	84.6
130	150	3900	22.2	350	8500	77.8
160	130	3900	24.4	350	8500	75.6
190	100	3600	26.3	250	8500	73.7
210	100	3500	26.3	250	8300	73.7

The simulation parameters used for all experimental spectra are shown in Table A5.3. The total spin for component 1 (E1) and component 2 (E2) is 10. Figure A5.25 shows the system-parameters temperature dependence for E1 and E2. A gentle slope is observed for the *D* values of both components, with a total decrease of 44% for both of them (Fig. A5.25a) as temperature increases. While the weight of E1 increases with the increase of the temperature, the weight of E2 decreases, although it remains preponderant over all the temperature range (see Fig. A5.25b). While decreasing the temperature, a gentle slope is observed for the *H*_{strain} (Gaussian broadening) of E1 and E2. However, around 50 K a sudden jump is observed in both cases.

The relative weight of E2 with respect to E1 decreases with temperature, from ~93% at 20 K to ~75% at 210 K (Fig. A5.25a). Finally, both components display similar temperature dependencies on *D* and H_{strain} (Fig. A5.25). The real temperature dependencies of E1 and E2 depend on the scaling factor given by the ratio between the unknown S_{real} , and S = 10 used for the model. For instance, $S_{\text{real}} = 100$ yields n = 10, according to the scaling laws of eq. 5.11.



Figure A5.25: Simulation parameters as a function of temperature, for component 1 (green dots) and component 2 (blue dots). (a) *D* values. (b) Weights. (c) Broadening (H_{strain}). The parameters are given in Table A5.3.



5.7.8. EPR SIMULATIONS WITH DIFFERENT SCALING FACTORS

Figure A5.26: Simulations (colored lines) of EPR spectra (black lines) recorded at 20 K, 30 K, 80 K, and 130 K. The simulations are performed with two spin systems using different scaling factors (n). The remaining parameters are shown in Table A5.3. For the entire temperature range, the line shape presents negligible changes, confirming the scaling method as a suitable candidate to reproduce and study the EPR line shape of high-spin systems.

5.7.9. EPR S-D INVERSE COMPENSATION



Figure A5.27: Simulations of EPR spectra performed at 80 K. The original simulation is shown in red. The simulations indicated by the green and yellow curves were obtained by considering both components with equal *D*.

The parameters *S* and *D* are inversely related. That means, an increase (decrease) of *S* of one of the components followed by a decrease (increase) of *D* of the same component by the same amount, results in a simulation with the same lineshape as the original simulation. It's important to mention that a correction in the weight of the component needs to be applied to preserve the same simulation amplitude. The correcting factor for components 1 (n1) and component 2 (n2) is defined by

n1 =
$$\frac{S1_{\text{orig}} \cdot (S1_{\text{orig}} + 1)}{S1_{\text{new}} \cdot (S1_{\text{new}} + 1)} = \frac{10 \cdot 11}{4 \cdot 5} = 5.5,$$
 (5.17)

$$n2 = \frac{S2_{\text{orig}} \cdot (S2_{\text{orig}} + 1)}{S2_{\text{new}} \cdot (S2_{\text{new}} + 1)} = \frac{10 \cdot 11}{25 \cdot 26} = 0.1692,$$
(5.18)

where SX_{orig} refers to the *S* used in the original simulations and SX_{new} to the change in *S* that is being applied and will result in the opposite change in *D*, by the same factor. X = 1,2 refers to EPR components 1 and 2, respectively.



5.7.10. SENSITIVITY OF EPR PARAMETERS

Figure A5.28: Simulations (colored lines) of EPR spectra (black lines) at 80 K performed by varying *D*2. An example of what is considered to be an acceptable simulation is depicted in red. Examples of rejected simulations are shown in light blue and yellow.

5.7.11. FERRITIN CORE AND MONONUCLEAR FE(III) EPR INTENSITIES

Mononuclear Fe(III) gives a characteristic EPR signal, with a sharp resonance at g' = 4.3, the so-called g' = 4.3 signal. This signal is also observed in the ferritin EPR spectra and the temperature dependence of its intensity is that of a paramagnetic species. Here we compare the amount of these species with respect to the ferritin core signal, the broad g' = 2 signal. The EPR active spin concentration ratio of Fe(III) (g' = 4.3) and the ferritin core spin system can be determined from[114]

$$\frac{[\text{Fe}_{m}]}{[\text{core}]} = \frac{A_{\text{Fe}_{m}}R_{\text{Fe}_{m}}(\text{Scan}_{\text{Fe}_{m}})^{2}G_{\text{core}} \cdot (B_{m})_{\text{core}} \cdot (g_{\text{core}})^{2}S_{\text{core}}(S_{\text{core}}+1)}{A_{\text{core}}R_{\text{core}} \cdot (\text{Scan}_{\text{core}})^{2}G_{\text{Fe}_{m}} \cdot (B_{m})_{\text{Fe}_{m}} \cdot (g_{\text{Fe}_{m}})^{2} \cdot S_{\text{Fe}_{m}}(S_{\text{Fe}_{m}}+1)},$$
(5.19)

where *g* is the g tensor, *S* electron spin number of each species and *A* is the enclosed area (second integral) that is calculated from the blue (mononuclear Fe(III)) and red (for the core) curves shown in Fig. A5.29. 'Scan' is the sweep range, *G* is the relative gain of the signal amplifier, B_m is the modulation amplitude, *R* is a factor related to the hyperfine interactions, but for both, Fe_m and core, the hyperfine interactions are neglected. Scan, *G*, B_m , and *R* are the same for both Fe_m and core, so they cancel out. Thus, equation 6.1 becomes

$$\frac{[\text{Fe}_{\text{m}}]}{[\text{core}]} = \frac{A_{\text{Fe}_{\text{m}}} \cdot (g_{\text{core}})^2 \cdot S_{\text{core}}(S_{\text{core}} + 1)}{A_{\text{core}} \cdot (g_{\text{Fe}_{\text{m}}})^2 \cdot S_{\text{Fe}_{\text{m}}}(S_{\text{Fe}_{\text{m}}} + 1)},$$
(5.20)

and follows

$$\frac{[\text{Fe}_{\text{m}}]}{[\text{core}]} = \frac{4135 \cdot (2.01539)^2 \cdot S_{\text{core}}(S_{\text{core}}+1)}{6463828 \cdot (1.9825)^2 \cdot \frac{5}{2}(\frac{5}{2}+1)} = 7.56 \cdot 10^{-5} \cdot S_{\text{core}}(S_{\text{core}}+1),$$
(5.21)

this expression is solely dependent on the total spin of the ferritin core signal. As an example, this ratio is 0.8 and 6.8 if the spin number *S* of the core is 100 and 300, respectively.

In order to compare the calculated EPR active spin concentration ratio $\frac{|\text{Fe}_{m}|}{|\text{core}|}$ to the number of paramagnetic Fe atoms, normalized by the number of superparamagnetic atoms, obtained by magnetometry ($N_{\rm p}/N_{\rm Fh}$) we need to multiply [core] with the number of iron atoms contributing to the ferrihydrite-like ferritin-core-EPR signal ($N_{\rm fh}$). The amount of iron determined by ICP-MS ($N_{\rm ICP}$) includes both mononuclear ($N_{\rm Fe}_{\rm m}$) and ferritin-like iron ions ($N_{\rm fh}$). However, the number of mononuclear Fe-ions that contribute to their EPR signal is small (2nd integral: 4575) compared to the ones contributing to the ferritin EPR signal (6778579). Therefore, we approximate the value obtained by ICP-MS to the number of iron atoms that contribute to the ferritin EPR signal, i.e., $N_{\rm fh} = 1967\pm78$.

The error of $\frac{[Fe_m]}{[core] \cdot N_{fh}}$ was calculated using the following error propagation expression

$$\delta(F) = \frac{\Delta(F)}{|F|} \approx \frac{\Delta(M_1)}{|M_1|} + \frac{\Delta(M_2)}{|M_2|} + \frac{\Delta(M_3)}{|M_3|},$$
(5.22)

where *F* is $\frac{[Fe_m]}{[core] \cdot N_{fh}}$, M_1 is the second integral of the ferritin-core EPR signal, M_2 is the second integral of the mononuclear Fe EPR signal, M_3 is the number of iron atoms in



Figure A5.29: Simulations (colored lines) of EPR spectra (black line) at 40 K. In red, the simulation of the broad signal. In blue, the simulation of the rhombic mononuclear Fe(III) ion signal located at g'=4.3, using S = 2.5, D = 20.96 GHz, E = 6.97 GHz, g = [1.9852,2.0139,1.9484], g_{strain} = [0.1328,0.0242,0.5009]. In green, the superposition of both simulations.

the ferritin core $N_{\rm fh}$. The errors of M_1 and M_2 were estimated by considering mainly the associated error of the simulations, using the Montecarlo error propagation with a small sampling number. Thus, δM_1 is 310 and δM_2 is 41. Finally, the error of M_3 is 78.

Table A5.4 shows the $\frac{[\text{Fe}_{m}]}{[\text{core}]\cdot N_{\text{fh}}}$ estimated for both S_{core} of 100 and 300. The largest number (3.6 \cdot 10⁻³) is obtained when considering $S_{\text{core}} = 300$. This value is one order of magnitude smaller than the fraction $N_{\text{p}}/N_{\text{Fh}}$, found by magnetometry (0.0645). The relative errors in Table A5.4 round up to 10%. The difference in $\frac{[\text{Fe}_{m}]}{[\text{core}]\cdot N_{\text{fh}}}$ for the two different *S* of a factor of 10 suggests that the uncertainty of $\frac{[\text{Fe}_{m}]}{[\text{core}]\cdot N_{\text{fh}}}$ is the dominating source of uncertainty and could be in the range of an order of magnitude.

Table A5.4: EPR active spin concentration ratio per ferritin particle $\frac{[Fe_m]}{[core] \cdot N_{PR}}$

Score	$\frac{[Fe_m]}{[core] \cdot N_{fh}}$	Abs. error	Rel. error (%)
100	$4 \cdot 10^{-4}$	$2 \cdot 10^{-5}$	5
300	$3.6 \cdot 10^{-3}$	$2 \cdot 10^{-4}$	6

Table A5.5: Data used in the error propagation calculation of $\frac{ \mathbf{r} ^2}{ \mathbf{c} ^2}$
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	M1 (a.u.)	M2 (a.u.)	M3 (iron ions)
Abs. value	6463828	4135	1967
Δ	310	41	78

5.7.12. EPR MODEL SIMULATIONS

In this section, some examples of the large set of spectra that constitute the EPR Model Simulations (80 K), are provided. The Model Simulations are used to constrain certain EPR parameters based on the trends seen. Figure A5.30 shows simulations of one EPR component with S = 10 by varying the D parameter from 100 to 5000 MHz and considering two different Gaussian broadening parameters (3000 and 8000 MHz). For the smallest broadening, D values larger than 500 MHz can not be used to account for the features observed in our experimental spectrum, this is shown in Fig. A5.30a where a sharp feature at low field is visible for the green curve (1000 MHz) and in Fig. A5.30c where the spectra become distorted. For the largest broadening, D cannot be greater than 1000 MHz (see Fig. A5.30d) since the spectrum starts to get progressively sharp at the lowest fields and flat at higher fields. In this way, to simulate the experimental spectrum measured at 80 K, figures A5.30a, b are used as starting points to inspect all parameters for D, H_{strain} and weights considering these two base components



Figure A5.30: Model simulations (colored lines) performed at 80 K. The simulation parameters are S = 10, (a,c) $H_{\text{strain}} = 3000$ MHz and (b,d) $H_{\text{strain}} = 8000$ MHz. In blue, D = 100 MHz. In red, D = 500 MHz. In green, D = 1000 MHz. In purple, D = 2000 MHz. In yellow, D = 5000 MHz.

5.7.13. EPR ALTERNATIVE FITTING APPROACH

Following a different approach, a systematical fit of the spectra recorded at different temperatures was performed. First, We determined the *S*, *D*, and weight values of each component by fitting the EPR spectra recorded at 80 K (close to the middle-temperature range). The fitting was performed with a Matlab (R2019a) script, using the EasySpin package (5.2.4), using the scaling approach described in the main text.

The optimized scaled *S* and *D* values found for both EPR components, *g* and their respective weights are given in Table A5.6. Then, to fit the spectra from 20 to 190 K we used the scaled *S*, *D*, and weight of each component determined at 80 K and only the Gaussian broadening given by an H_{strain} parameter was used as the fitting parameter for both components (see Fig. A5.33). The spectra between 5 and 15 K were omitted from the analysis because it was not possible to accurately discriminate the broad signal from the background noise.

The two individual components are shown in figure A5.32 at 20 K, 80 K, and 190 K. Component 2 (blue curve) is narrower and highly contributes to the total intensity of the spectra. Comparing the lineshape evolution at the different temperatures, it can be noticed that component 2 (green curve) is responsible for the overall lineshape change. Whereas, component 1, being quite broad and lower in intensity, appears almost temperature independent. This is a sign of a poor fitting result since component 1 can be hiding many features. *D* parameter does not change the simulation shape. The H_{strain} changes the lineshape, but in an unsystematic way. In spite of the good agreement of simulations with the experiment, simulation parameters do not give physical insight. Therefore, a different approach was used (see main text).



Figure A5.31: Simulations (red dash lines) of EPR spectra (black lines) for all data recorded between 20 and 190 K. The simulations are performed with two spin systems, as described above. The system parameters are shown in Table A5.6. The scaled H_{strain} of both systems (fitting parameters) are depicted in figure A5.33.



Figure A5.32: Total simulations (red lines) of EPR spectra (black lines) performed at 20 K, 80 K, and 190 K. The simulations are performed with two spin systems, as described above. The first system component is depicted in green. The second component is shown in blue.

Table A5.6: System parameters used for simulations of the EPR spectra. *S* is the Spin number and *D* the Zero-field axial component. The scaling factor is n = 14.6. The real parameters can be derived as follows: $D_{real} = D/n$, $S_{real} = S \cdot n$.

	S	g	D (MHz)	weight (%)
System 1	20	2.01	-167.2	36
System 2	7	2.01	-489.5	64

5.7.14. EQUILIBRIUM MAGNETIZATION MODELS

The evaluation of 5.4 requires numerical integration. Standard numerical integration routines require excessive computation time to evaluate the five-fold integral, while Monte Carlo integration does not work well, due to the different evaluation points at the numerator and denominator. Therefore, integrals have been replaced with sums over discrete states for a finite number of particles, that is

$$M(B) = \frac{\sum_{\phi=j\delta\phi} \sin\phi \frac{\sum_{\varepsilon=i\delta\varepsilon} \sum_{\lambda=l\delta\lambda} \sum_{\psi=k\delta\psi} \sum_{\theta=n\delta\theta} \zeta e^{-E/k_{\rm B}T} \sin\theta}{\sum_{\varepsilon=i\delta\varepsilon} \sum_{\lambda=l\delta\lambda} \sum_{\psi=k\delta\psi} \sum_{\theta=n\delta\theta} e^{-E/k_{\rm B}T}} \sum_{\phi=j\delta\phi} (5.23)$$

where $\delta \phi = \delta \lambda = \delta \psi = \delta \theta = \pi/2N$ are exact dividers of $\pi/2$ and $\delta \varepsilon$ an exact divider of the maximum canting angle ε_{max} . The maximum canting angle is defined as the mean root square of ε weighted by $e^{-E/k_{\rm B}T}$ and avoids the evaluation of strongly canted states with negligible occurrence. As ε increases with the applied field, ε_{max} is evaluated iteratively starting for a series of field values starting from $\varepsilon_{max} = \pi B/2B_E$. The number N + 1 of samples in each dimension is chosen to be the largest possible for given memory and time constraints. The numerical procedure has been implemented in Wolfram Mathematica and compiled in C using the built-in compiler. A PC with 64 GB RAM supports



Figure A5.33: Broadening fitting parameters (H_{strain}) as a function of temperature, for component 1 (green dots) and component 2 (blue dots).

calculations up to N = 27 for a total of 10^9 combinations of particle orientations and magnetic states, for an angular resolution of ~ 3.3° in ϕ , λ , ψ , and θ .



Figure A5.34: (a) Initial magnetization curves measured at temperatures comprised between 3 K (blue circles) and 200 K (yellow circles), and global fit to the modified Langevin model with one superparamagnetic component without moment distribution. (b) Fit residuals as a function of the predicted magnetization.



Figure A5.35: Real μ' and Imaginary μ'' parts of the magnetic moment of the human-liver ferritin (raw data). The legend refers to the frequency of the oscillating B_1 field.



Figure A5.36: Full hysteresis acquired at 5 K and 25 K both in ZFC and FC conditions, to determine the exchange bias of human-liver ferritin.



Figure A5.37: Measured isothermal magnetization curves (dots) and corresponding best fits with a two-component Langevin law obtained from eq. (5.13) after replacing the magnetic moment distribution with two single-valued moments (lines), at temperatures indicated by numbers. Residuals, defined as the difference between measurements and model, normalized by the measured magnetization at 7 T, are plotted below.



Figure A5.38: Comparison between a direct estimate of the energy barrier distribution, obtained from the blocking temperature distribution of the IRM Component 1, and the energy barrier distribution expected from $E_{\rm b} = mB_{\rm a}/2$ with the incorrect assumption that *m* and $B_{\rm a}$ are independent variables.



Figure A5.39: Reconstructed distribution of η_s obtained from the deconvolution of the magnetic moment and volume distributions (solid line), and best-fit normal distribution $N \log \eta_s$, -1,0.392).

LIST OF SYMBOLS AND ACRONYMS

В	Applied magnetic field. Also B .
B_0	Resonance field in EPR spectra.
$B_{\rm EX}$	Exchange field.
Ba	Anisotropy field.
$B_{\rm esf}$	Equilibrium spin-flop field.
$B_{\rm ex}$	Exchange coupling field.
$B_{ m sf}$	Spin-flop field.
D	Zero-field splitting.
$E_{\rm b}$	Anisotropy energy.
G _{pp}	Amplitude peak to peak of field modulation.
H _{strain}	Gaussian broadening.
J	Exchange constant.
Κ	Anisotropy constant.
K _S	Surface anisotropy.
$K_{\rm V}$	Uniaxial volume anisotropy.
M_0	Sublattice magnetization.
$M_{ au}$	Remanent magnetization.
$M_{\rm s}$	Saturation magnetization.
$N_{\rm A}$	Avogadro constant.
$N_{ m s}$	Surface spins.
S	Total spin of the particle.
Т	Temperature.
$T_{\rm b}$	Blocking temperature.
V	Particle volume.
χ_{\perp}	Bulk perpendicular susceptibility.
χnc	low-field susceptibility.
D	Traceless zero-field splitting tensor.
S	Electron spin operator.
g	Landé g-factor tensor.
μ	(1) Magnetic moment in EPR. (2) Mean value of distributions.
$\mu_{ m B}$	Bohr magneton.
ε_{s}	Zero-field-canting angle.

с	Concentration of defects.
g'	Effective <i>g</i> .
$k_{\rm B}$	Boltzmann constant.
т	Magnetic moment in magnetometry.
m _c	Canting moment.
m _{uc}	Uncompensated moment in magnetometry.
AF	Antiferromagnetic.
E1	EPR simulation component 1.
E2	EPR simulation component 2.
EELS	Electron Energy-Loss Spectroscopy.
EMR	Electron magnetic resonance.
EPR	Electron paramagnetic resonance.
FC	Field-cooled.
Fh	Ferrihydrite.
HuLiFt	Human-liver ferritin.
ICP-MS	Inductively coupled plasma mass spectrometry.
IRM	Isothermal remanent magnetization.
MRI	Magnetic resonance imaging.
NMR	Nuclear magnetic resonance.
SAXS	Small-Angle X-ray Scattering.
SDS-PAGE	Sodium dodecyl sulfate-polyacrylamide gel electrophoresis.
SNR	Signal-to-noise ratio.
TEM	Transmission electron microscopy.
XANES	X-ray adsorption near edge spectroscopy.
ZFC	Zero-Field-cooled.

REFERENCES

- L. Bossoni, J. A. Labra-Muñoz, H. S. van der Zant, V. Čaluković, A. Lefering, R. Egli, and M. Huber, *In-depth magnetometry and EPR analysis of the spin structure of human-liver ferritin: from DC to 9 GHz*, Phys. Chem. Chem. Phys (2023), accepted for publication.
- [2] M. Fittipaldi, R. Mercatelli, S. Sottini, P. Ceci, E. Falvo, and D. Gatteschi, Sensing the quantum behaviour of magnetic nanoparticles by electron magnetic resonance, Phys. Chem. Chem. Phys. 18, 3591 (2016).
- [3] N. Chasteen and P. M. Harrison, *Mineralization in ferritin: An efficient means of iron storage*, Journal of Structural Biology **126**, 182 (1999).
- [4] D. Finazzi and P. Arosio, *Biology of ferritin in mammals: an update on iron storage, oxidative damage and neurodegeneration, Archives of toxicology* **88**, 1787 (2014).
- [5] P. Arosio, L. Elia, and M. Poli, *Ferritin, cellular iron storage and regulation*, IUBMB Life 69, 414 (2017).
- [6] G. Bartzokis, T. a. Tishler, P. H. Lu, P. Villablanca, L. L. Altshuler, M. Carter, D. Huang, N. Edwards, and J. Mintz, *Brain ferritin iron may influence age- and gender-related risks of neurodegeneration*, Neurobiology of Aging 28, 414 (2007).
- [7] C. Quintana, S. Bellefqih, J. Laval, J.-L. Guerquin-Kern, W. Ting-Di, J. Avila, I. Ferrer, R. Arranz, and C. Patiño, *Study of the localization of iron, ferritin, and hemosiderin in alzheimer's disease hippocampus by analytical microscopy at the subcellular level*, Journal of structural biology 153, 42 (2006).
- [8] C. Quintana and L. Gutiérrez, Could a dysfunction of ferritin be a determinant factor in the aetiology of some neurodegenerative diseases? Biochimica et Biophysica Acta (BBA) - General Subjects 1800, 770 (2010).
- [9] O. Strbak, L. Balejcikova, M. Kmetova, J. Gombos, J. Kovac, D. Dobrota, and P. Kopcansky, Longitudinal and transverse relaxivity analysis of native ferritin and magnetoferritin at 7 t mri, International Journal of Molecular Sciences 22, 8487 (2021).
- [10] Y. Gossuin, R. N. Muller, and P. Gillis, *Relaxation induced by ferritin: a better understanding for an improved mri iron quantification*, NMR in Biomedicine **17**, 427.
- [11] L. Bossoni, I. Hegeman-Kleinn, S. G. van Duinen, M. Bulk, L. H. P. Vroegindeweij, J. G. Langendonk, L. Hirschler, A. Webb, and L. van der Weerd, Off-resonance saturation as an mri method to quantify mineral- iron in the post-mortem brain, Magnetic Resonance in Medicine 87, 1276.
- [12] S. A. Makhlouf, F. T. Parker, and A. E. Berkowitz, *Magnetic hysteresis anomalies in ferritin*, Phys. Rev. B 55, R14717 (1997).

- [13] I. Alenkina, M. Oshtrakh, Z. Klencsár, E. Kuzmann, A. Chukin, and V. Semionkin, 57fe mössbauer spectroscopy and electron paramagnetic resonance studies of human liver ferritin, ferrum lek and maltofer[®], Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy 130, 24 (2014).
- [14] D. E. Madsen, M. F. Hansen, J. Bendix, and S. Mørup, *On the analysis of magnetization and mössbauer data for ferritin*, Nanotechnology **19**, 315712 (2008).
- [15] E. Wajnberg, L. J. El-Jaick, M. P. Linhares, and D. M. Esquivel, *Ferromagnetic resonance of horse spleen ferritin: Core blocking and surface ordering temperatures,* Journal of Magnetic Resonance **153**, 69 (2001).
- [16] M. P. Weir, T. J. Peters, and J. F. Gibson, *Electron spin resonance studies of splenic ferritin and haemosiderin*, Biochimica et Biophysica Acta (BBA) Protein Structure and Molecular Enzymology 828, 298 (1985).
- [17] Z. Jang, B. Suh, A. Lascialfari, R. Sessoli, and F. Borsa, *Proton nmr and susceptibility measurements on the magnetic core of ferritin*, Applied Magnetic Resonance 19, 557 (2012).
- [18] Y. Gossuin, D. Hautot, R. Muller, Q. Pankhurst, J. Dobson, C. Morris, P. Gillis, and J. Collingwood, *Looking for biogenic magnetite in brain ferritin using nmr relaxometry*, NMR in biomedicine 18, 469 (2005).
- [19] M. Koralewski, L. Balejčíková, Z. Mitróová, M. Pochylski, M. Baranowski, and P. Kopčanský, Morphology and magnetic structure of the ferritin core during iron loading and release by magnetooptical and nmr methods, ACS Applied Materials & Interfaces 10, 7777 (2018).
- [20] E. Schäfer-Nolte, L. Schlipf, M. Ternes, F. Reinhard, K. Kern, and J. Wrachtrup, *Tracking temperature-dependent relaxation times of ferritin nanomagnets with a wideband quantum spectrometer*, Phys. Rev. Lett. **113**, 217204 (2014).
- [21] E. Lee, D. H. Kim, J. Hwang, K. Lee, S. Yoon, B. J. Suh, K. Hyun Kim, J.-Y. Kim, Z. H. Jang, B. Kim, B. I. Min, and J.-S. Kang, *Size-dependent structural evolution* of the biomineralized iron-core nanoparticles in ferritins, Applied Physics Letters 102, 133703 (2013).
- [22] E. S. Grant, L. T. Hall, L. C. L. Hollenberg, G. McColl, and D. A. Simpson, Nonmonotonic superparamagnetic behavior of the ferritin iron core revealed via quantum spin relaxometry, ACS Nano 17, 372 (2023).
- [23] S. Aime, B. Bergamasco, D. Biglino, G. Digilio, M. Fasano, E. Giamello, and L. Lopiano, *Epr investigations of the iron domain in neuromelanin*, Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease 1361, 49 (1997).
- [24] V. Nandwana, S.-R. Ryoo, S. Kanthala, A. Kumar, A. Sharma, F. Castro, Y. Li, B. Hoffman, S. Lim, and V. Dravid, *Engineered ferritin nanocages as natural contrast agents in magnetic resonance imaging*, RSC Adv. 7, 34892 (2017).

- [25] J. Boas and T. G.J., *Electron paramagnetic resonance studies of a range of ferritins and haemosiderins*, Biochim Biophys Acta **229**, 68 (1971).
- [26] N. Deighton, A. Abu-Raqabah, I. J. Rowland, M. C. R. Symons, T. J. Peters, and R. J. Ward, *Electron paramagnetic resonance studies of a range of ferritins and haemosiderins*, J. Chem. Soc., Faraday Trans. 87, 3193 (1991).
- [27] F. S. Otsuka, M. C. G. Otaduy, O. R. Nascimento, and C. E. G. Salmon, *Quantifica*tion of Paramagnetic Ions in Human Brain Tissue Using EPR, Brazilian Journal of Physics 52, 94 (2022).
- [28] Y.-H. Pan, K. Sader, J. J. Powell, A. Bleloch, M. Gass, J. Trinick, A. Warley, A. Li, R. Brydson, and A. Brown, 3d morphology of the human hepatic ferritin mineral core: New evidence for a subunit structure revealed by single particle analysis of haadf-stem images, Journal of Structural Biology 166, 22 (2009).
- [29] F. Brem, G. Stamm, and A. M. Hirt, *Modeling the magnetic behavior of horse spleen ferritin with a two-phase core structure*, Journal of Applied Physics **99**, 123906 (2006).
- [30] N. Gálvez, B. Fernández, P. Sánchez, R. Cuesta, M. Ceoláin, M. Clemente-Leáon, S. Trasobares, M. Lýez-Haro, J. J. Calvino, O. Stéphan, and J. M. Domínguez-Vera, *Comparative structural and chemical studies of ferritin with gradual removal of their iron contents*, Journal of the American Chemical Society 130, 8062 (2008).
- [31] G. Jutz, P. van Rijn, B. Santos Miranda, and A. Böker, *Ferritin: A versatile building block for bionanotechnology*, Chemical Reviews **115**, 1653 (2015).
- [32] S. H. Kilcoyne and R. Cywinski, *Ferritin: a model superparamagnet*, Journal of Magnetism and Magnetic Materials 140–144, 1466 (1995).
- [33] M. S. Seehra, V. S. Babu, and A. Manivannan, *Neutron scattering and magnetic studies of ferrihydrite nanoparticles*, Physical Review B **61**, 3513 (2000).
- [34] J. C. E. Harris, J. E. Grimaldi, D. D. Awschalom, A. Chiolero, and D. Loss, *Excess spin* and the dynamics of antiferromagnetic ferritin, Physical Review B **60**, 3453 (1999).
- [35] R. Brooks, J. Vymazal, R. Goldfarb, J. Bulte, and P. Aisen, *Relaxometry and magne-tometry of ferritin*, Magnetic resonance in medicine 40, 227 (1998).
- [36] G. C. Papaefthymiou, *The mössbauer and magnetic properties of ferritin cores*, Biochimica et Biophysica Acta (BBA) - General Subjects 1800, 886 (2010), ferritin: Structures, Properties and Applications.
- [37] F. M. Michel, V. Barrón, J. Torrent, M. P. Morales, C. J. Serna, J.-F. Boily, Q. Liu, A. C. Ambrosini, A.and Cismasu, and G. E. Brown, Ordered ferrimagnetic form of ferrihydrite reveals links among structure, composition, and magnetism, Proceedings of the National Academy of Sciences of the United States 107, 2787 (2010).

- [38] T. G. St. Pierre, N. T. Gorham, P. D. Allen, J. L. Costa-Krämer, and K. V. Rao, Apparent magnetic energy-barrier distribution in horse-spleen ferritin: Evidence for multiple interacting magnetic entities per ferrihydrite nanoparticle, Physical Review B 65, 024436 (2001).
- [39] N. J. O. Silva, V. S. Amaral, A. Uritzberea, R. Bustamante, A. Millán, F. Palacio, E. Kampert, U. Zeitler, S. de Brion, O. Iglesias, and A. Labarta, *Shifted loops* and coercivity from field-imprinted high-energy barriers in ferritin and ferrihydrite nanoparticles, Physical Review B 84, 104427 (2011).
- [40] R. P. Guertin, N. Harrison, Z. X. Zhou, S. McCall, and F. Drymiotis, *Very high field magnetization and AC susceptibility of native horse spleen ferritin*, Journal of Magnetism and Magnetic Materials **308**, 97 (2007).
- [41] N. J. O. Silva, A. Millán, F. Palacio, E. Kampert, U. Zeitler, H. Rakoto, and V. S. Amaral, *Temperature dependence of antiferromagnetic susceptibility in ferritin*, Physical Review B 79, 104405 (2009).
- [42] N. J. O. Silva, V. S. Amaral, L. D. Carlos, B. Rodríguez-González, L. M. Liz-Marzán, T. S. Berquó, S. K. Banerjee, V. de Zea Bermudez, A. Millán, and F. Palacio, *Evidence* of random magnetic anisotropy in ferrihydrite nanoparticles based on analysis of statistical distributions, Physical Review B 77, 134426 (2008).
- [43] C. Gilles, P. Bonville, K. K. W. Wong, and S. Mann, Non-Langevin behavior of the uncompensated magnetization in nanoparticles of artificial ferritin, The European Physical Journal B – Condensed Matter and Complex Systems 17, 417 (2000).
- [44] N. Noginova, F. Chen, T. Weaver, E. P. Giannelis, A. B. Bourlinos, and V. A. Atsarkin, Magnetic resonance in nanoparticles: between ferro- and paramagnetism, Journal of Physics: Condensed Matter 19, 246208 (2007).
- [45] R. J. Usselman, S. E. Russek, M. T. Klem, M. A. Allen, T. Douglas, M. Young, Y. U. Idzerda, and D. J. Singel, *Temperature dependence of electron magnetic resonance spectra of iron oxide nanoparticles mineralized in listeria innocua protein cages*, Journal of Applied Physics **112**, 084701 (2012).
- [46] R. Berger, J. Kliava, J.-C. Bissey, and V. Baïetto, *Magnetic resonance of superparamagnetic iron-containing nanoparticles in annealed glass*, Journal of Applied Physics **87**, 7389 (2000).
- [47] R. Berger, J.-C. Bissey, J. Kliava, R. Berger, and J.-C. Bissey, *Lineshapes in magnetic resonance spectra*, Journal of Physics: Condensed Matter **12**, 9347 (2000).
- [48] R. Berger, J.-C. Bissey, J. Kliava, H. Daubric, and C. Estournès, *Temperature dependence of superparamagnetic resonance of iron oxide nanoparticles*, Journal of Magnetism and Magnetic Materials 234, 535 (2001).
- [49] R. J. Usselman, M. T. Klem, M. Allen, E. D. Walter, K. Gilmore, T. Douglas, M. Young, Y. Idzerda, and D. J. Singel, *Electron magnetic resonance of iron oxide nanoparticles mineralized in protein cages*, Journal of Applied Physics **97**, 10M523 (2005).

- [50] Y. L. Raikher and V. I. Stepanov, Ferromagnetic resonance in a suspension of singledomain particles, Phys. Rev. B 50, 6250 (1994).
- [51] M. M. Yulikov and P. A. Purtov, *Fmr study of superparamagnetic ni particles with weak and strong magnetic anisotropy*, Applied Magnetic Resonance **29**, 1613 (2005).
- [52] M. Fittipaldi, L. Sorace, A.-L. Barra, C. Sangregorio, R. Sessoli, and D. Gatteschi, Molecular nanomagnets and magnetic nanoparticles: The emr contribution to a common approach, Phys. Chem. Chem. Phys. 11, 6555 (2009).
- [53] K. S. Kumar, R. R. Pasula, S. Lim, and C. A. Nijhuis, *Long-range tunneling processes across ferritin-based junctions*, Advanced Materials 28, 1824 (2016).
- [54] V. Barrón and T. J., *Evidence for a simple pathway to maghemite in earth and mars soils*, Geochimica et Cosmochimica Acta **66**, 2801 (2002).
- [55] F. Luis, E. del Barco, J. M. Hernández, E. Remiro, J. Bartolomé, and T. J., *Resonant spin tunneling in small antiferromagnetic particles*, Physical Review B 59, 11837 (1999).
- [56] S. Mørup, *Spin-flop in antiferromagnetic microcrystals*, Surface Science **156**, 888 (1985).
- [57] C. Gilles, P. Bonville, H. Rakoto, J. Broto, K. Wong, and S. Mann, *Magnetic hysteresis and superantiferromagnetism in ferritin nanoparticles*, Journal of Magnetism and Magnetic Materials 241, 430 (2002).
- [58] S. V. Komogortsev, D. A. Balaev, A. A. Krasikov, S. V. Stolyar, R. N. Yaroslavtsev, V. P. Ladygina, and R. S. Iskhakov, *Magnetic hysteresis of blocked ferrihydrite nanopar-ticles*, AIP Advances 11, 015329 (2021).
- [59] A. N. Bogdanov, A. V. Zhuravlev, and U. K. Rössler, Spin-flop transition in uniaxial antiferromagnets: Magnetic phases, reorientation effects, and multidomain states, Physical Review B 75, 094425 (2007).
- [60] P. J. Cregg and L. Bessais, Series expansion for the magnetisation of a solid superparamagnetic system of non-interacting particles with anisotropy, Journal of Magnetism and Magnetic Materials 202, 554 (1999).
- [61] M. Hanson, C. Johansson, and S. Mørup, *The influence of magnetic anisotropy on the magnetization of small ferromagnetic particles*, Journal of Physics: Condensed Matter 5, 725 (1993).
- [62] L. Néel, *Superparamagnétisme des grains très fins antiferromagnétiques*, Comptes Rendus Hebdomadaires des Seances de l'Academie des Sciences **252**, 4075 (1961).
- [63] R. H. Kodama, A. E. Berkowitz, E. J. McNiff, and S. Foner, Surface spin disorder in ferrite nanoparticles (invited), Journal of Applied Physics 81, 5552 (1997).
- [64] R. H. Kodama, *Magnetic nanoparticles*, Journal of Magnetism and Magnetic Materials 200, 359 (1999).
- [65] J. Mazo-Zuluaga, J. Restrepo, and J. Mejía-López, Effect of surface anisotropy on the magnetic properties of magnetite nanoparticles: A Heisenberg-Monte Carlo Study, Journal of Applied Physics 103, 113906 (2008).
- [66] R. H. Kodama and A. E. Berkowitz, Surface-driven effects on the magnetic behavior of oxide nanoparticles, in Surface Effects in Magnetic Nanoparticles, edited by D. Fiorani (Springer, New York, 2005) 1st ed., Chap. 6, pp. 189–216.
- [67] D. Laura-Ccahuana and E. De Biasi, *Spin-flop transition in small nanoparticles: Internal magnetic study of surface effects*, Journal of Magnetism and Magnetic Materials **549**, 169018 (2022).
- [68] A. H. Morrish, Canted Antiferromagnetism: Hematite (World Scientific, 1994).
- [69] N. J. Silva, V. S. Amaral, and L. D. Carlos, *Relevance of magnetic moment distribution and scaling law methods to study the magnetic behavior of antiferromagnetic nanoparticles: Application to ferritin*, Physical Review B - Condensed Matter and Materials Physics **71**, 1 (2005).
- [70] D. Balaev, A. A. Krasikov, A. A. Dubrovskiy, S. I. Popkov, S. V. Stolyar, O. A. Bayukov, R. S. Iskhakov, V. P. Ladygina, and R. N. Yaroslavtsev, *Magnetic properties of heat treated bacterial ferrihydrite particles*, Journal of Magnetism and Magnetic Materials **410**, 171 (2016).
- [71] C. Rani and S. D. Tiwari, *Estimation of particle magnetic moment for antiferromagnetic ferrihydrite particles*, Journal of Magnetism an Magnetic Materials 385, 272 (2015).
- [72] D. Balaev, S. S. V., Y. V. Knyazev, R. N. Yaroslavtsev, A. I. Pankrats, M. Vorotynov, A. A. Krasikov, D. A. Veikanov, O. A. Bayukov, V. P. Ladygina, and R. S. Iskhakov, *Role of the surface effects and interparticle magnetic interactions in the temperature evolution of magnetic resonance spectra of ferrihydrite nanoparticle ensembles*, Results in Physics 35, 105340 (2022).
- [73] M.-E. Y. Mohie-Eldin, R. B. Frankel, and L. Gunther, *A comparison of the magnetic properties of polysaccharide iron complex (pic) and ferritin, Journal of Magnetism and Magnetic Materials* **135**, 65 (1994).
- [74] Z. Yuan, P. Atanassov, A. M. Alsmadi, S. G. E. te Velthuis, U. Welp, C. I. Hammetter, R. Hjelm, and H. Nakotte, *Magnetic properties of self-assembled ferritin-core arrays*, Journal of Applied Physics **99**, 08Q509 (2006).
- [75] M. B. Madsen and S. Mørup, *Magnetic properties of ferrihydrite*, Hyperfine Interactions 27, 329 (1986).

5

- [76] Y. Guyodo, R. Banerjee, S. K. an Lee Penn, D. Burleson, T. S. Berquó, T. Seda, and P. Solheid, *Magnetic properties of six-line ferrihydrite nanoparticles*, Physics of the Earth and Planetary Interiors 154, 222 (2006).
- [77] D. Balaev, S. I. Popkov, A. A. Krasikov, A. D. Balaev, A. A. Dubrovskiy, S. S. V., R. N. Yaroslavtsev, V. P. Ladygina, and R. S. Iskhakov, *Temperature behavior of the anti-ferromagnetic susceptibility of nanoferrihydrite from the measurements of the magnetization curves in fields up to 250 koe*, Physics of the Solid State 59, 1940 (2017).
- [78] A. Punnoose, T. Phanthavady, M. S. Seehra, N. Shah, and G. P. Huffman, *Magnetic properties of ferrihydrite nanoparticles doped with ni, mo, and ir, Physics Review B* 69, 054425 (2004).
- [79] C.-W. Hsieh, B. Zheng, and S. Hsieh, *Ferritin protein imaging and detection by magnetic force microscopy*, Chemical Communications **46**, 1655 (2010).
- [80] T. Hiemstra and W. H. Van Riemsdijk, A surface structural model for ferrihydrite i: Sites related to primary charge, molar mass, and mass density, Geochimica et Cosmochimica Acta 73, 4423 (2009).
- [81] E. C. Stoner and E. P. Wohlfarth, Apparent magnetic energy-barrier distribution in horse-spleen ferritin: Evidence for multiple interacting magnetic entities per ferrihydrite nanoparticle, Philosophical Transactions of the Royal Society of London, Series A 240, 599 (1948).
- [82] E. Vroegindeweij, L. Bossoni, A. Boon, J. Wilson, M. Bulk, J. Labra-Muñoz, M. Huber, A. Webb, L. van der Weerd, and J. Langendonk, *Quantification of different iron forms in the aceruloplasminemia brain to explore iron-related neurodegeneration*, NeuroImage: Clinical **30**, 102657 (2021).
- [83] R. S. Silva, P. C. Morais, F. Qu, A. M. Alcalde, N. O. Dantas, and H. S. L. Sullasi, Synthesis process controlled magnetic properties of pb1xmnxs nanocrystals, Applied Physics Letters 90, 253114 (2007).
- [84] P. D. Allen, T. G. St Pierre, W. Chua-anusorn, V. Ström, and K. V. Rao, *Low-frequency low-field magnetic susceptibility of ferritin and hemosiderin*, Biochimica et Bio-physica Acta 1500, 186 (2000).
- [85] J. I. Gittleman, B. Abeles, and S. Bozowski, *Superparamagnetism and relaxation effects in granular Ni-SiO2 and Ni-Al2O3 films, Physical Review B* **9**, 3891 (1974).
- [86] M. Respaud, Magnetization process of noninteracting ferromagnetic cobalt nanoparticles in the superparamagnetic regime: Deviation from Langevin law, Journal of Applied Physics 86, 556 (1999).
- [87] D. E. Madsen, S. Mørup, and M. F. Hansen, On the interpretation of magnetization data for antiferromagnetic particles, Journal of Magnetism and Magnetic Materials 305, 95 (2006).

- [88] J. Khatua, M. Gomilšek, J. C. Orain, A. M. Strydom, Z. Jagličić, C. V. Colin, S. Petit, A. Ozarowski, L. Magnin-Thro, M. S. Ramachandra Rao, A. Zorko, and P. Khuntia, *Signature of a randomness-driven spin-liquid state in a frustrated magnet*, Communications Physics 5, 99 (2022).
- [89] M. J. O'Shea and A. A. L., Inverted hysteresis in magnetic systems with interface exchange, Journal of Applied Physics 75, 6673 (1994).
- [90] D. J. Robertson and D. E. France, *Discrimination of remanence-carrying minerals in mixtures, using isothermal remanent magnetisation acquisition curves, Physics* of the Earth and Planetary Interiors **82**, 223 (1994).
- [91] R. Egli, *Analysis of the field dependence of remanent magnetization curves*, Journal of Geophysical Research **108**, 2081 (2003).
- [92] P. P. Kruiver, M. J. Dekkers, and D. Heslop, *Quantification of magnetic coercivity components by the analysis of acquisition curves of isothermal remanent magnetisation*, Earth and Planetary Science Letters **189**, 269 (2001).
- [93] X. Zhao, M. Fujii, Y. Suganuma, X. Zhao, and Z. Jiang, *Applying the Burr Type XII distribution to decompose remanent magnetization curves*, Journal of Geophysical Research: Solid Earth **123**, 8298 (2018).
- [94] L. X. Tian, C. Q. Cao, Q. S. Liu, and P. Pan, Y. *Low-temperature magnetic properties of horse spleen ferritin*, Chinese Science Bulletin **55**, 3174 (2010).
- [95] R. Egli, Magnetic characterization of geologic materials with first-order reversal curves, in Magnetic Measurement Techniques for Materials Characterization, edited by V. Franco and B. Dodrill (Springer Nature Publishing Group, New York, 2021) pp. 455–604.
- [96] H. Pfeiffer, *Determination of anisotropy field distribution in particle assemblies taking into account thermal fluctuations*, Physica Status Solidi (a) **118**, 295 (1990).
- [97] B. M. Moskowitz, R. B. Frankel, S. A. Walton, D. P. E. Dickson, K. K. W. Wong, T. Douglas, and M. S., *Determination of the preexponential frequency factor for superparamagnetic maghemite particles in magnetoferritin*, Journal of Geophysical Research 102, 22671 (1997).
- [98] P. Rochette, P.-E. Mathé, L. Esteban, H. Rakoto, J.-L. Bouchez, Q. Liu, and J. Torrent, Non-saturation of the defect moment of goethite and fine-grained hematite up to 57 Teslas, Geophysical Research Letters 32, L22309 (2005).
- [99] D. Carunto, G. Carunto, and C. O'Connor, *Magnetic properties of variable-sized Fe*₃O₄ *nanoparticles synthesized from non-aqueous homogeneous solutions of polyols*, J. Phys. D: Appl. Phys. **40**, 5801 (2007).
- [100] T. N. shendruk, R. D. Desautels, B. W. Southern, and J. van Lierop, *The effect of surface spin disorder on the magnetism of* γ *-fe*₂ o_3 *nanoparticle dispersions*, Nanotechnology **18**, 455704 (2007).

- [101] R. Egli and W. Lowrie, *Anhysteretic remanent magnetization of fine magnetic particles*, Journal of Geophysical Research **107**, 2209 (2002).
- [102] L. Breth, D. Suess, C. Vogler, B. Bergmair, M. Fuger, R. Heer, and H. Brueckl, *Ther-mal switching field distribution of a single domain particle for field-dependent at-tempt frequency*, Journal of Applied Physics **112**, 023903 (2012).
- [103] M. Koralewski, M. Pochylski, and J. Gierszewski, Magnetic properties of ferritin and akaganeite nanoparticles in aqueous suspension, Journal of Nanoparticle Research 15, 1902 (2013).
- [104] S. H. Bell, M. P. Weir, D. P. Dickson, J. F. Gibson, G. A. Sharp, and T. J. Peters, *Mössbauer spectroscopic studies of human haemosiderin and ferritin*, Biochimica et Biophysica Acta (BBA) - Protein Structure and Molecular Enzymology **787**, 227 (1984).
- [105] R. Krzyminiewski, Z. Kruczyński, B. Dobosz, A. Zając, A. Mackiewicz, E. Leporowska, and S. Folwaczna, *Epr study of iron ion complexes in human blood*, Applied Magnetic Resonance 40, 321 (2011).
- [106] M. Gheisari, M. Mozaffari, M. Acet, and J. Amighian, *Preparation and investigation of magnetic properties of wüstite nanoparticles*, Journal of Magnetism and Magnetic Materials **320**, 2618 (2008).
- [107] S. Ueno and M. Sekino, *Biomagnetics: Principles and Applications of Biomagnetic Stimulation and Imaging* (CRC Press, 2018) p. 234.
- [108] S. Mørup, D. E. Madsen, C. Frandsen, C. R. H. Bahl, and M. F. Hansen, *Experimen*tal and theoretical studies of nanoparticles of antiferromagnetic materials, Journal of Physics: Condensed Matter 19, 213202 (2007).
- [109] I. S. Jacobs and C. P. Bean, An approach to elongated fine-particle magnets, Physical Review 100, 1060 (1955).
- [110] A. Aharoni, Curling reversal mode in nonellipsoidal ferromagnetic particles, Journal of Applied Physics 86, 1041 (1999).
- [111] R. Skomski, J. P. Liu, C. B. Rong, and D. J. Sellmyer, *Hysteresis of ultrasmall fe-pt particles*, Journal of Applied Physics 103, 07E139 (2008).
- [112] D. J. Dunlop, *Theory and application of the day plot (mrs/ms versus hcr/hc) 1. theoretical curves and tests using titanomagnetite data, Journal of Geophysical Research: Solid Earth* **107**, EPM 4 (2002).
- [113] M. Winklhofer, K. Fabian, and F. Heider, *Magnetic blocking temperatures of magnetite calculated with a three-dimensional micromagnetic model*, Journal of Geophysical Research **102**, 22695 (1997).
- [114] J. A. Weil and J. R. Bolton, *Electron Paramagnetic Resonance. Elementary Theory and Practical Applications* (John Wiley and Sons, Inc., Hoboken, New Jersey, 2007) p. 546.