



Universiteit
Leiden
The Netherlands

Transient parvovirus B19 DNAemia after kidney transplantation: a 2-sided story

Russcher, A.; Backer, M.M.D.; Brouwer, C. de; Dijkstra, K.; Kers, J.; Vries, A. de; ... ; Kroes, A.

Citation

Russcher, A., Backer, M. M. D., Brouwer, C. de, Dijkstra, K., Kers, J., Vries, A. de, ... Kroes, A. (2023). Transient parvovirus B19 DNAemia after kidney transplantation: a 2-sided story. *Open Forum Infectious Diseases*, 10(3). doi:10.1093/ofid/ofad079

Version: Publisher's Version

License: [Creative Commons CC BY-NC-ND 4.0 license](https://creativecommons.org/licenses/by-nc-nd/4.0/)

Downloaded from: <https://hdl.handle.net/1887/3641584>

Note: To cite this publication please use the final published version (if applicable).

Transient Parvovirus B19 DNAemia After Kidney Transplantation: A 2-Sided Story

Anne Russcher,¹ Marijke Molenaar-de Backer,² Caroline de Brouwer,¹ Kyra Dijkstra,³ Jesper Kers,³ Aiko de Vries,⁴ Hans Zaaijer,² Ann Vossen,¹ and Aloysius Kroes¹

¹Department of Medical Microbiology, Leiden University Medical Center, Leiden, The Netherlands, ²Donor Medicine Research, Department of Blood-borne Infections, Sanquin Blood Supply Foundation, Amsterdam, The Netherlands, ³Department of Pathology and Leiden Transplant Center, Leiden University Medical Center, Leiden, The Netherlands, and ⁴Department of Nephrology and Leiden Transplant Center, Leiden University Medical Center, Leiden, The Netherlands

Parvovirus B19 (B19V) DNAemia appears to be a relatively common finding after kidney transplantation. However, not all DNAemia signifies active infection with replicating virus. This study screened 134 patients posttransplantation for B19V DNAemia and identified 2 cases in which viral DNA was present after transplantation, with the donor kidney as probable source of the DNA. In both cases intact viral particles could not be detected using an endonuclease method, indicating the presence of noninfectious DNA remnants.

Keywords. DNAemia; kidney transplantation; parvovirus B19; viremia; viral activity.

Parvovirus B19 (B19V) is a ubiquitous human pathogen with a unique predisposition to infect erythroid progenitor cells (EPCs), causing viremia with extremely high viral loads up to 10^{14} IU/mL during acute infection [1]. In immunocompromised patients such as transplant recipients, this may lead to severe, and often relapsing, anemia [2–4]. Apart from EPCs, B19V DNA appears to enter multiple cell types using mechanisms other than cell receptor-specific entry during acute infection [5, 6]. This results in lifelong detectability of B19V DNA in some tissues, including renal tissue, without apparent disease activity [7–9]. Also in blood, B19V DNA remains detectable for months after infection [10]. Of all transplant

patients, attention for B19V infection in kidney transplantation is particularly relevant as kidney recipients often suffer from anemia for various reasons, of which B19V infection may be one [11]. Detection of B19V DNA (DNAemia) in blood after kidney transplantation is not uncommon; shortly after transplantation, the incidence is 0.5% to 1.4%, but the overall incidence of B19V DNAemia is estimated at 7.6% in the year after kidney transplantation [12, 13]. However, due to the unique characteristic of B19V DNA to persist in tissue lifelong, the clinical relevance of DNAemia after kidney transplantation should be carefully considered. True B19V infection by kidney transplantation appears to be rare, as it supposes (semi)acute infection in the donor combined with a susceptible recipient. Due to an adult seroprevalence of 60%–70%, many recipients will not be susceptible [13]. Detectable DNA may originate from intact viral particles, causing true infection, or from DNA remnants released from previously infected cells [10, 14–16]. Application of an endonuclease-based assay can differentiate between these 2 scenarios [10]. Endonuclease treatment degrades DNA remnants but not DNA in intact viral particles, which is protected by the viral capsid. This study investigates DNAemia after kidney transplantation and presents 2 cases where DNAemia after kidney transplantation should not be mistaken for active B19V infection.

METHODS

Patient Screening

One hundred ninety-two patients received a living kidney transplantation at Leiden University Medical Center in the period January 2013–June 2016. Of these, blood samples were available from 134 patients in the period 30–90 days after transplantation. For each patient, 1 blood sample in that period was screened with qualitative B19V polymerase chain reaction (PCR) for the presence of B19V DNA. If positive, additional donor and longitudinal patient blood samples were tested with quantitative B19V PCR. PCR was also performed on kidney biopsies from the time of transplantation. To determine viral activity, blood samples were also tested using endonuclease pretreatment.

B19V PCR and Sequencing

DNA extraction was performed by MagnaPure (Roche Diagnostics) with 200 μ L of plasma or serum as input for the PCR, according to the manufacturer's instructions. Subsequently, eluates were tested with an in-house B19V PCR targeting the VP1/VP2 region of all known 3 genotypes as described by Knoester et al [17]. For kidney biopsies, 5 sections of 10 μ m were deparaffinized and homogenized with a pretreatment

Received 17 November 2022; editorial decision 07 February 2023; accepted 07 February 2023; published online 14 February 2023

Correspondence: Anne Russcher MD, Leiden University Medical Center, Department of Medical Microbiology, Albinusdreef 2, 2333 ZA Leiden, The Netherlands (a.russcher@lumc.nl); Aloysius Kroes MD, PhD, Leiden University Medical Center, Department of Medical Microbiology, Albinusdreef 2, 2333 ZA Leiden, The Netherlands (a.c.m.kroes@lumc.nl).

Open Forum Infectious Diseases®

© The Author(s) 2023. Published by Oxford University Press on behalf of Infectious Diseases Society of America. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs licence (<https://creativecommons.org/licenses/by-nc-nd/4.0/>), which permits non-commercial reproduction and distribution of the work, in any medium, provided the original work is not altered or transformed in any way, and that the work is properly cited. For commercial re-use, please contact journals.permissions@oup.com

<https://doi.org/10.1093/ofid/ofad079>

as used for routine tissue diagnostic procedures in our laboratory. Subsequently, DNA extraction and PCR were performed as described above in duplicate. An in-house PCR for the housekeeping gene β -globin was performed in duplicate on the kidney biopsies to determine cell count, as described previously, to express DNA load in kidney biopsies as international units per cell [18]. Near whole genome sequencing on the kidney biopsy and plasma samples of patient 1 was performed as described previously [19].

B19V Serology

Immunoglobulin M (IgM) and immunoglobulin G (IgG) serology was performed on blood samples using the Liaison Biotrin Parvovirus IgM/IgG assay (Diasorin) according to the manufacturer's protocol.

Endonuclease (Benzonase) Assay

If B19V DNA positive, a blood sample was retested applying endonuclease treatment as described by Molenaar et al [10]. In short, each plasma sample was split into two 80–100 μ L aliquots, the exact amount depending on the availability of sufficient material. To one of the aliquots, 250 units of Benzonase (Sigma-Aldrich, the Netherlands) was added. Subsequently both aliquots were incubated for 1 hour at 37°C in a shaking incubator at 120 rpm. After cooling to room temperature, DNA extraction and quantitative dual-target NS-VP2 PCR were performed as described previously [20]. Negative, low-positive, and high-positive controls were included in the endonuclease assay.

Histochemical Staining of Kidney Biopsies

Staining of paraffin slides (10 μ m) was performed using monoclonal anti-B19V antibody, targeted against amino acid 328–344 of the VP2 capsid protein, clone R9276 (Sigma-Aldrich). A positive control stain was included in the analysis.

Ethics

The investigation was done in accordance with the principles outlined in the declaration of Helsinki. The ethical committee of the Leiden University Medical Center approved the research protocol. As the study was not subject to the Medical Research Involving Human Subjects Act and patient data cannot be traced back to the individual patient, informed consent was waived.

RESULTS

A total of 134 kidney transplant patients were screened for B19V DNAemia 30–90 days after kidney transplantation. Clinical characteristics are shown in Table 1. Two of the 134 patients tested positive for B19V DNA in the screening, after which additional donor kidney and recipient blood samples were tested. The longitudinal B19V DNAemia profiles of

Table 1. Patient Characteristics

Characteristic	n = 134
Age, years, mean (SD)	51 (14)
Male sex, No. (%)	83 (61.9)
B19V IgG positive at baseline ^a , No. (%)	93 (69.4)
Time to screening sample, days after transplantation, mean (SD)	65 (14)
Positive B19V PCR result, No.	2

Abbreviations: B19V, parvovirus B19; IgG, immunoglobulin G; PCR, polymerase chain reaction; SD, standard deviation.

^aOne patient missing IgG status.

both patients are shown in Figure 1, accompanied by clinical information and the course of hemoglobin (Hb) levels.

Patient 1, a 49-year-old man, was B19V IgG seropositive before transplantation and DNA negative in blood. He received a living-related kidney transplant. The kidney biopsy at the time of transplantation tested positive for B19V DNA (200 IU/cell). After transplantation, patient 1 tested consistently positive for B19V DNA in blood for >10 months, with a gradual decrease in DNA load. Endonuclease testing of all DNA-positive blood samples showed 100% degradability of the detected DNA. Patient 1 showed no obvious symptoms of active B19V infection after transplantation. The patient suffered from anemia 4 months posttransplantation with a nadir Hb level of 8.0 g/dL (Figure 1), which recovered in the following months. The presence of B19V DNAemia was unknown to clinicians in the years after transplantation and he received no specific B19V treatment such as intravenous immunoglobulin. His immunosuppressive regimen was moderated by a temporary stop of tacrolimus due to a concomitant BK nephropathy (Figure 1), but this did not lead to clearance of B19V DNA, whereas his Hb levels did improve.

Patient 2, a 54-year-old man, was B19V IgG seronegative and B19V DNA negative before transplantation. The patient received a living-related kidney transplant from a B19V-seropositive donor. The kidney biopsy at the time of transplantation tested weak-positive in 1 of 2 duplicates for B19V DNA, indicating a DNA load around the limit of detection. After transplantation, patient 2 tested positive at 1 time-point with a low B19V DNA load in blood of 79 IU/mL. A false-positive finding was ruled out by confirmation with a different dual-target PCR method in a different laboratory. The sample was 100% degradable in the endonuclease assay. Clinically, the patient showed no signs of a B19V infection, with a stable Hb level of around 13 g/dL in the months posttransplantation. His immunosuppressive regimen was also moderated due to a concomitant BK nephropathy, but in the months following the positive sample, the patient remained B19V IgM and IgG negative.

DISCUSSION

This study describes 2 patients testing positive for B19V DNA in blood after kidney transplantation, which was found to be

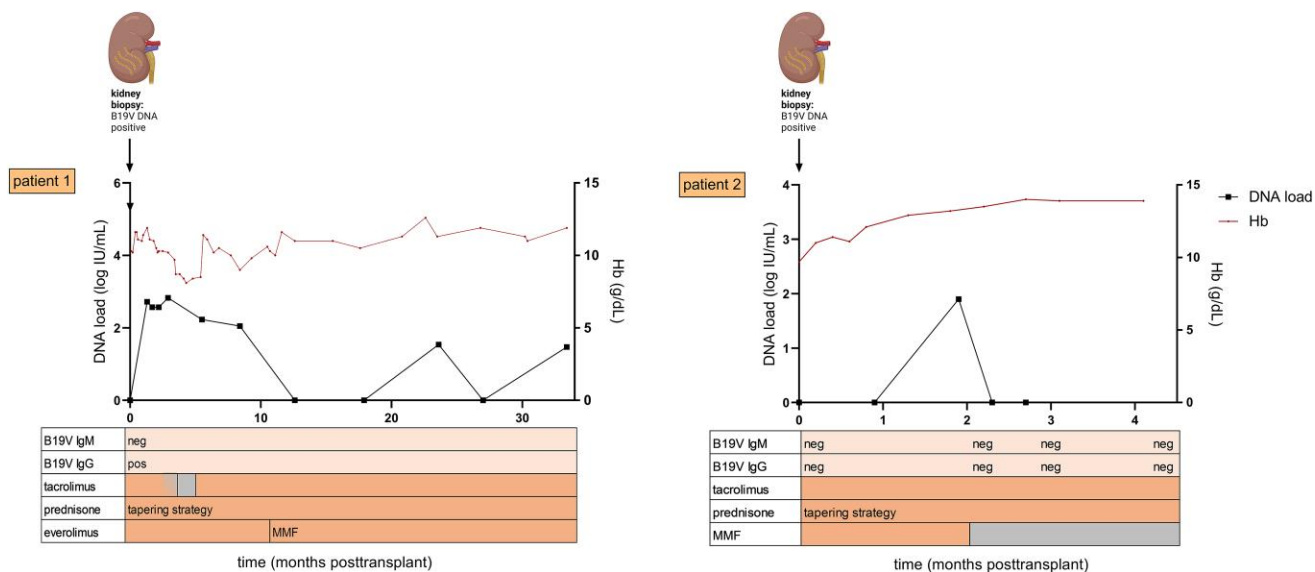


Figure 1. Longitudinal course of B19V DNAemia in 2 kidney transplant recipients, in relation to hemoglobin levels and information regarding seroconversion and immunosuppressive treatment. Dark orange bars represent active treatment; gray bars represent no treatment. Abbreviations: MMF, mycophenolate mofetil. Figure partly created with biorender.com.

unrelated to B19V replication and could be attributed to “passive” B19V DNA remnants, most likely originating from the donor kidney. This interpretation was supported by the degradation of the B19V DNA in an endonuclease test; by the fact that the B19V load did not decrease with tapering of immunosuppression; by the relatively low DNA loads in blood as opposed to the high viral loads commonly seen in active infection; by the absence of seroconversion; and by the lack of symptoms consistent with B19V infection. Patient 1 did suffer from anemia, but anemia occurs in 20%–51% of kidney transplant recipients at various time points after transplantation and the improving course of anemia does not correspond to the persistent detection of B19V DNA [11].

In previous studies, it was already observed that low-level B19V DNAemia after transplantation does not correspond to symptomatic infection. Barzon et al described a pediatric cohort of 75 transplantations in which 24 kidney biopsies tested positive for B19V DNA. Twenty recipients become DNA positive in the year after transplantation, of which 10 were B19V seronegative before transplantation. However, only 4 patients were reported to develop symptoms of B19V infection after transplantation. Data regarding seroconversion were not mentioned [21]. A similar observation was made recently by Reza Hosseini et al [22]. In their study, 6 previously B19V IgG-negative SOT recipients were identified with B19V DNAemia after transplantation. Seroconversion was documented in 4 patients but in the other 2 patients, DNAemia was low-level and transient and IgG was negative 1 year after transplantation. This illustrates that posttransplant B19V

DNAemia can be caused by either of 2 mechanisms: true B19V infection with viral replication, or clinically irrelevant presence of inactive B19V DNA fragments (ie, remnants from earlier infection). Although the above-mentioned studies by Barzon et al and Reza Hosseini et al report on patients in which passive transfer of clinically irrelevant DNA seems probable, they do not distinguish between these 2 manifestations by additional laboratory testing and do not comment on the possible different interpretations of DNAemia. The relative proportions of true infection and clinically irrelevant DNAemia can still be considered unknown. The endonuclease assay used in this study appears to be a useful and reliable tool to distinguish between active B19V infection and presence of inactive B19V DNA remnants, which can be applied in research settings, for example, to determine proportions of true infection versus irrelevant DNAemia, and in clinical practice to guide patient management.

This study also showed that B19V DNAemia can arise from the graft after kidney transplantation. Patient 1 was B19V IgG seropositive before transplantation, so B19V DNA may have been derived from a previous infection in the recipient. However, the course and properties of DNAemia in patient 1 strongly suggested that DNAemia originated from the donor kidney, which was retrospectively found to be positive in the B19V PCR. In patient 2, it is also likely that the B19V DNA-positive kidney was the source of the B19V DNA transmission, as the patient had no evidence of a previous B19V infection. Unfortunately, it was not possible to investigate the genetic relationship between donor and recipient B19V DNA, because the viral loads proved too low for reliable sequencing (data not shown).

It is remarkable that B19V DNAemia is reported in many patients with evident kidney damage, for example, kidneys from deceased donors or concomitant BK reactivation [23]. Our 2 patients also showed BK reactivation. It can be hypothesized that B19V DNA present in tissue will be released into the bloodstream during tissue damage, for example, induced by BK reactivation. This mechanism has been described for donor-derived cell-free DNA, which is used as a biomarker for tissue injury [24]. This mechanism could also explain clinically irrelevant B19V DNAemia after transplantation. Immunohistochemical staining for B19V capsid antigen in kidney biopsies was negative (data not shown), which is in line with persistence of DNA in kidney tissue but not persistence of intact viral particles.

In summary, this study showed that B19V DNAemia after kidney transplantation is not necessarily associated with active infection. The nature and origin of B19V DNAemia should be carefully considered before it is assumed to have clinical relevance, to avoid erroneous treatment decisions.

Notes

Acknowledgments. We thank Dr I. Bajema of the Department of Pathology for providing the kidney biopsies for PCR testing and Dr E. Wessels for facilitating PCR testing on the biopsies.

Potential conflicts of interest. All authors: No reported conflicts.

References

1. Heegaard ED, Brown KE. Human parvovirus B19. *Clin Microbiol Rev* **2002**; 15: 485–505.
2. Beckhoff A, Steffen I, Sandoz P, Hirsch HH, Schaub S. Relapsing severe anaemia due to primary parvovirus B19 infection after renal transplantation: a case report and review of the literature. *Nephrol Dial Transplant* **2007**; 22:3660–3.
3. Eid AJ, Brown RA, Patel R, Razonable RR. Parvovirus B19 infection after transplantation: a review of 98 cases. *Clin Infect Dis* **2006**; 43:40–8.
4. Plentz A, Würdinger M, Kudlich M, Modrow S. Low-level DNAemia of parvovirus B19 (genotypes 1–3) in adult transplant recipients is not associated with anaemia. *J Clin Virol* **2013**; 58:443–8.
5. Pyöriä L, Toppinen M, Mäntylä E, et al. Extinct type of human parvovirus B19 persists in tonsillar B cells. *Nat Commun* **2017**; 8:14930.
6. Bieri J, Ros C. Globoside is dispensable for parvovirus B19 entry but essential at a postentry step for productive infection. *J Virol* **2019**; 93:e00972–19.
7. Soderlund-Venermo M, Hokynar K, Nieminen J, Rautakorpi H, Hedman K. Persistence of human parvovirus B19 in human tissues. *Pathol Biol* **2002**; 50: 307–16.
8. Norja P, Hokynar K, Aaltonen LM, et al. Bioportfolio: lifelong persistence of variant and prototypic erythrovirus DNA genomes in human tissue. *Proc Natl Acad Sci U S A* **2006**; 103:7450–3.
9. Adamson-Small LA, Ignatovich IV, Laemmerhirt MG, Hobbs JA. Persistent parvovirus B19 infection in non-erythroid tissues: possible role in the inflammatory and disease process. *Virus Res* **2014**; 190:8–16.
10. Molenaar-de Backer MW, Russcher A, Kroes AC, Koppelman MH, Lanfermeijer M, Zaaier HL. Detection of parvovirus B19 DNA in blood: viruses or DNA remnants? *J Clin Virol* **2016**; 84:19–23.
11. Gafer-Gvili A, Gafer U. Posttransplantation anemia in kidney transplant recipients. *Acta Haematol* **2019**; 142:37–43.
12. Yu Y, Wei C, Lyu J, et al. Donor-derived human parvovirus B19 infection in kidney transplantation. *Front Cell Infect Microbiol* **2021**; 11:753970.
13. Thongprayoon C, Khoury NJ, Bathini T, et al. Epidemiology of parvovirus B19 and anemia among kidney transplant recipients: a meta-analysis. *Urol Ann* **2020**; 12:241–7.
14. Russcher A, Verdonshot J, Molenaar-de Backer MWA, Heymans SRB, Kroes ACM, Zaaier HL. Parvovirus B19 DNA detectable in hearts of patients with dilated cardiomyopathy, but absent or inactive in blood. *ESC Heart Fail* **2021**; 8: 2723–30.
15. Reber U, Moser O, Dilloo D, Eis-Hubinger AM. On the utility of the Benzonase treatment for correct laboratory diagnosis of parvovirus B19 infection. *J Clin Virol* **2017**; 95:10–1.
16. Alves ADR, Langella BB, Lima MMS, et al. Evaluation of molecular test for the discrimination of “naked” DNA from infectious parvovirus B19 particles in serum and bone marrow samples. *Viruses* **2022**; 14:843.
17. Knoester M, von dem Borne PA, Vossen AC, Kroes AC, Claas EC. Human parvovirus B19 genotype 3 associated with chronic anemia after stem cell transplantation, missed by routine PCR testing. *J Clin Virol* **2012**; 54:368–70.
18. de Vries JJ, Vesseur A, Rotteveel LJ, et al. Cytomegalovirus DNA detection in dried blood spots and perilymphatic fluids from pediatric and adult cochlear implant recipients with prelingual deafness. *J Clin Virol* **2013**; 56:113–7.
19. Molenaar-de Backer MW, Lukashov VV, van Binnendijk RS, Boot HJ, Zaaier HL. Global co-existence of 2 evolutionary lineages of parvovirus B19 1a, different in genome-wide synonymous positions. *PLoS One* **2012**; 7:e43206.
20. Molenaar-de Backer MW, de Waal M, Sjerps MC, Koppelman MH. Validation of new real-time polymerase chain reaction assays for detection of hepatitis A virus RNA and parvovirus B19 DNA. *Transfusion* **2016**; 56:440–8.
21. Barzon L, Murer L, Pacenti M, et al. Detection of viral DNA in kidney graft preservation and washing solutions is predictive of posttransplant infections in pediatric recipients. *J Infect Dis* **2009**; 200:1425–33.
22. Rezahosseini O, Ekenberg C, Möller DL, et al. Incidence and impact of parvovirus B19 infection in seronegative solid organ transplant recipients. *J Infect Dis* **2021**; 224:865–9.
23. Baek CH, Kim H, Yang WS, Han DJ, Park SK. Risk factors and long-term outcomes of parvovirus B19 infection in kidney transplant patients. *Transpl Infect Dis* **2017**; 19:e12754.
24. Oellerich M, Sherwood K, Keown P, et al. Liquid biopsies: donor-derived cell-free DNA for the detection of kidney allograft injury. *Nat Rev Nephrol* **2021**; 17: 591–603.