



Universiteit
Leiden
The Netherlands

Diversity of glucocorticoid receptor signaling: molecular mechanisms and therapeutic implications

Viho, E.M.G.

Citation

Viho, E. M. G. (2023, September 7). *Diversity of glucocorticoid receptor signaling: molecular mechanisms and therapeutic implications*. Retrieved from <https://hdl.handle.net/1887/3638839>

Version: Publisher's Version

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/3638839>

Note: To cite this publication please use the final published version (if applicable).



1

General introduction and outline

GENERAL INTRODUCTION

At the beginning of the last century, the physiologist Walter B. Cannon defined homeostasis as the internal “equilibrium”; a balance that is held by constant physiological adjustments [1]. Homeostasis can be challenged by “stressors”, as described in 1936 by Hans Selye who wrote: “*Experiments on rats show that if the organism is severely damaged by acute non-specific noxious agents [...], a typical syndrome appears, the symptoms of which are independent of the nature of the damaging agent [...] and represent rather a response to damage as such.*” His observations were the first steps in the theory of stress biology, where stress is defined as a threat to homeostasis [2]. Almost 15 years later, Selye published “Stress and the general adaptation syndrome” in the British Medical Journal, where he argued that any threat to life causes a stress response, and that adequate adaptability and resistance to stress are required for survival [3].

The physiological response to stress

The physiological stress response involves various neuronal and endocrine mediators, including neurotransmitters, neuropeptides, and steroid hormones. The onset of the stress response is associated with the release of catecholamines (e.g., noradrenaline) and the activation of the sympathetic nervous system [4]. The noradrenergic system primarily relies on the locus coeruleus (LC), located in the brainstem. Neurons from the LC project to most brain regions and are the dominant source of noradrenaline in the brain [5–7]. Noradrenaline acts through G protein-coupled receptors, which can cause rapid changes in neuronal electrical properties by altering the functionality of ion channels [8]. Noradrenergic neurotransmission triggered by the LC facilitates the behavioral, cognitive, and neuroendocrine adaptive mechanisms required in the response to stress, particularly those related to vigilance and arousal [9]. LC neurons also activate the sympathetic nervous system leading to the release of acetylcholine in the adrenal medulla which triggers catecholamine release [10, 11]. The sympathetic-adrenal-medullary system connects the rapid response to stress between the central nervous system and the periphery (**Fig. 1A**). The immediate stress response also relies on central release of corticotropin-releasing hormone (CRH) from the hypothalamic paraventricular nucleus (PVN) [12, 13]. The stress response induced by CRH is mediated by CRHR1 receptors that are expressed in the hippocampus, the amygdala and the LC [14–18]. In mice, the selective deletion of CRHR1 in forebrain glutamatergic neurons was shown to alter neuronal activity in the amygdala and the hippocampus, and alleviate anxiety-like symptoms [19]. The neuroendocrine response to stress via the hypothalamic pituitary adrenal (HPA) axis also depends on CRH release by hypothalamic PVN

neurons into the bloodstream, together with arginine vasopressin (AVP) [20]. In rats, it was previously shown that approximately half of the CRH neurons in the PVN are AVP-positive [21, 22]. CRH and AVP that are released from the hypothalamus bind to G protein-coupled receptors (CRHR1 and AVPR1B) in the anterior pituitary gland [23–26] (**Fig. 1B**).

Upon stimulation by CRH, corticotropic cells of the anterior pituitary release the adrenocorticotrophic hormone (ACTH) which results from the expression and cleavage of proopiomelanocortin (POMC) [27]. ACTH activates the melanocortin type 2 receptor (MC2R) in the cortex of the adrenal gland, which induces steroidogenesis and immediate secretion of glucocorticoid (GC) hormones [28–30].

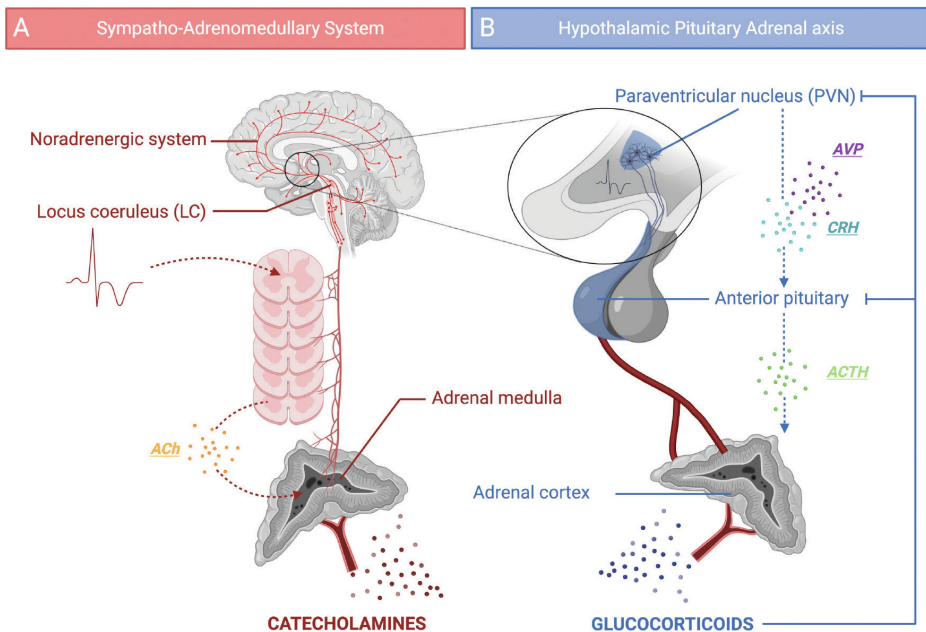


Figure 1. Neuroendocrine response to stress. The sympatho-adrenomedullary system (**A**) and the hypothalamic pituitary adrenal axis (**B**) coordinate the neuroendocrine response to stress. **Abbreviations:** ACh, acetylcholine; CRH, corticotropin-releasing hormone; AVP, arginine vasopressin; ACTH, adrenocorticotrophic hormone.

The zona fasciculata of the adrenal cortex produces and secretes the GC hormones, *i.e.*, corticosterone in rats and mice, and predominantly cortisol in humans [31–34] (**Fig. 1B**). To maintain homeostasis, GC hormones exert negative feedback at different levels of the HPA axis, particularly the hypothalamus, and the pituitary (**Fig. 1B**). Some studies have also reported intra-adrenal negative feedback by

GCs, although this phenomenon is less well characterized [35]. Regulation of the HPA axis by GCs also involves cortical and limbic brain regions, including the hippocampus, the amygdala, and the pre-frontal cortex. Indeed, GCs can alter the afferent projections from these regions to the PVN, thereby affecting the activation of CRH neurons [36]. The negative feedback on the HPA axis results in a circadian and ultradian rhythm in GC secretion unto the bloodstream [37].

Upon stress, the increased catecholamine release is rapidly followed by the secretion of GC hormones [38]. Even though both catecholamines and GC hormones have rapid effects to engage energy in response to stress, only GCs can regulate the long-term stress adaptation via genomic and epigenomic mechanisms. These latter effects form the focus of this thesis.

Genomic basis of stress adaptation

GC hormones bind to the glucocorticoid receptor (GR), and the mineralocorticoid receptor (MR) which is also a receptor for aldosterone. Endogenous glucocorticoid levels can be converted into inactive metabolites (*i.e.*, cortisone or 11-dehydrocorticosterone) by the 11 β hydroxysteroid dehydrogenase type 2 (11 β -HSD2) enzyme. In tissues that express high levels of 11 β -HSD2, MR is predominantly activated by aldosterone [39–41]. In the brain, MR activity is mostly regulated by GCs because 11 β -HSD2 is only expressed in a small number of brain nuclei that mediate the central effects of aldosterone on salt homeostasis [42–44]. High affinity of GCs for the MR results in its function as a sensor for basal GC levels, as the dynamic range of MR lies in small variations of hormone concentrations occurring during the circadian trough. On the other hand, GCs have a lower affinity for the GR, and this receptor therefore responds to elevated levels of GCs, during the endogenous circadian peak or upon stress [45, 46]. GR and MR belong to the nuclear steroid receptor family and are encoded by the nuclear receptor subfamily 3 group C member 1 and 2 gene, respectively (*Nr3c1/Nr3c2*). Nuclear steroid receptors are ligand-dependent transcription factors composed of an amino-terminal domain (NTD) which contains the activation domain 1 (AF1), a DNA-binding domain (DBD), a hinge region, and a carboxy-terminal ligand-binding domain that contains the activation domain 2 (AF2) [47]. This thesis focuses on the GR which consists of 777 amino acids in mice and diverges from other nuclear receptors mainly in the hinge region and NTD [48].

In the absence of a ligand, a multi-protein chaperone complex sequesters the GR in the cytosol. FK506-binding protein 5 (FKBP51), and heat shock proteins 70 (HSP70) and 90 (HSP90) are essential to this process [49]. Upon ligand binding,

the cytosolic GR changes its conformation, which exposes its nuclear localization signal [50]. FKBP51 is replaced by FKBP52, which, together with HSP90, induces GR transport into the nucleus using the microtubule network and dynein activity, resulting in a rapid nuclear translocation that is apparent within five minutes [51–55]. This process can also be slower and take up to 50 minutes, which allows GR targeting for proteasomal degradation [56, 57]. In the nucleus, GR associates with DNA at GR-binding sites (GBS). The most common GBS involve a glucocorticoid response element (GRE) with a palindromic sequence (AGAACA_nTTGTTCT). However, the GR can also bind to half sites, or inverted-repeat sites, that are associated with transcriptional inhibition and therefore referred to as negative GREs [47, 58]. Finally, the GR can indirectly interact with DNA by tethering to other transcription factors via protein-protein interactions [59–61].

The GR is thought to be able to bind to DNA as a monomer, although GR homodimers are the favored conformation [62, 63], and GRs could possibly form tetramers (*i.e.*, a dimer of dimers) [64, 65]. The nucleosome consists of histone protein octamers enveloped in DNA. The accessibility of GR to its target loci depends on the nucleosome status of the target region. Chromatin accessibility varies according to post-translational histone modifications and correlates with histone acetylation within genomic active regions. Conversely, histone methylation is often associated with inaccessible heterochromatin (**Fig. 2**).

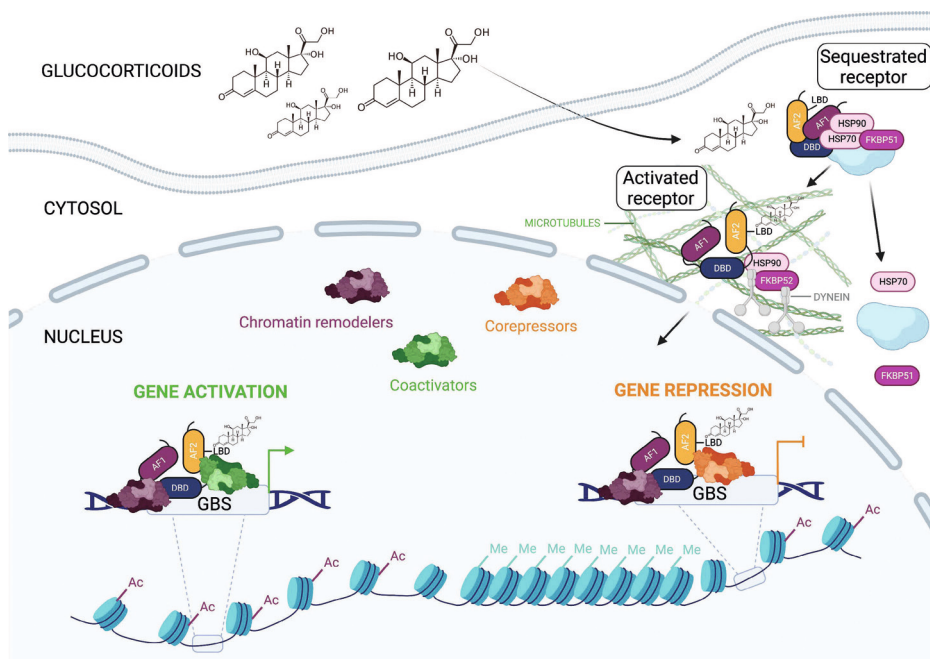


Figure 2. Glucocorticoid receptor transcriptional activity. Abbreviations: AF1, activation function 1; AF2, activation function 2; LBD, ligand-binding domain; DBD, DNA-binding domain; HSP70, heat shock protein 70; HSP90, heat shock protein 90; FKBP51, FK506-binding protein 5 isoform 1; FKBP52, FK506-binding protein 5 isoform 2; GBS, glucocorticoid receptor binding site; Ac, histone acetylation; Me, histone methylation.

At the genome, GR recruits coregulators that determine the outcome of GR signaling. These protein partners can be divided into two main categories: coactivators, that stimulate GR-driven gene transactivation, and corepressors, which lead to gene inhibition by the GR transcription complex. GR coactivators include the p160 steroid receptor coactivator (SRC) family, more specifically SRC-1, SRC-2, and SRC-3, which are also known as nuclear receptor coactivators (NCOA1, NCOA2, and NCOA3). The SRC family is a group of scaffolding proteins that can increase GR stability, but are mostly known to recruit other coregulators, histone acetyltransferases, or histone methyltransferases [47, 66, 67].

Another group of GR coactivators form the mediator complex, which regulates RNA polymerase II activity and the recruitment of transcription initiation factors [47, 68]. GR corepressors mostly include proteins from the nuclear receptor corepressor (NCOR) family and silencing mediators of the retinoic acid and thyroid hormone receptor (SMRT) family [69]. These corepressor complexes contain

histone deacetylases (HDACs) or methyltransferases (**Fig. 2**). A subset of nuclear coregulators can act as corepressors or coactivators depending on the context, *e.g.*, the tissue, cell type or the other proteins present in the transcriptional complex. Finally, the GR interacts with chromatin remodelers to actively change chromatin accessibility around target loci. Particularly, the GR recruits the Brahma (BRM) and Brahma-related-gene (BRG1) SWitch/Sucrose Non-Fermentable (SWI/SNF) complexes [70–72]. BRM and BRG1 are ATPases that catalyze changes in the nucleosome to facilitate chromatin accessibility to the GR transcription complexes (**Fig. 2**) [73–75]. The coactivators, corepressors, and chromatin remodelers compose the GR transcriptional complex and determine the outcome of GR signaling. The GR creates a link between the environment and the genome. The proteins required for the GR genomic activity are differentially expressed between tissues and between cell types, and the composition of the GR transcriptional complex can also be influenced by the context, *e.g.*, in different disease stages, upon stress, and during drug treatment. Consequently, GR signaling is tissue-, cell-, and context specific [76].

1

Glucocorticoid receptor signaling in metabolic and neuropsychiatric diseases

The GR is widely expressed throughout the body and has pleiotropic functions that both support circadian alignment of tissues as well as adaptation to stress. However, excessive exposure to stressors or GCs, as defined by high intensity, repetition, or prolonged exposure, can lead to maladaptation. Chronic elevations in GC levels can be triggered by chronic stress such as psychological trauma but can also result from repeated and prolonged corticosteroid medication or pathological hypercortisolism, as occurs in patients with Cushing's syndrome. The immune system is one target of GR signaling. The acute stress response in part via GCs enhances adaptive immunity, but GCs are best known for their anti-inflammatory effects to counteract an excessive initial response to infections [77]. As a deleterious consequence, chronic GC exposure disturbs immune responses by decreasing the number and activity of immune cells (leukopenia) [78, 79]. In addition, excessive exposure to GCs can trigger and exacerbate metabolic diseases. Cushing's syndrome is an extreme example of the extent of metabolic consequences of uncontrolled GR signaling. Hypercortisolism in patients with Cushing's syndrome leads to metabolic abnormalities, such as hyperlipidemia, hyperglycemia, and insulin resistance [80, 81]. GC medication for inflammatory diseases is associated with the same cluster of metabolic disturbances [82–85]. As an example, excessive GC exposure or chronic stress in conjunction with high-fat diet can lead to non-alcoholic fatty liver disease (NAFLD), which can progress to non-alcoholic steatohepatitis (NASH) [86, 87]. The prevalence of NAFLD in patients with Cushing's syndrome is approximately 20% [88].

GC-induced liver lipid accumulation is attributed to increases in food intake, gluconeogenesis, lipogenesis, fatty acid uptake, and inhibition of lipid β -oxidation [86, 89]. Yet another side effect of synthetic GCs is the suppression of the HPA axis, which can result in adrenal insufficiency [90] (**Fig. 3**).

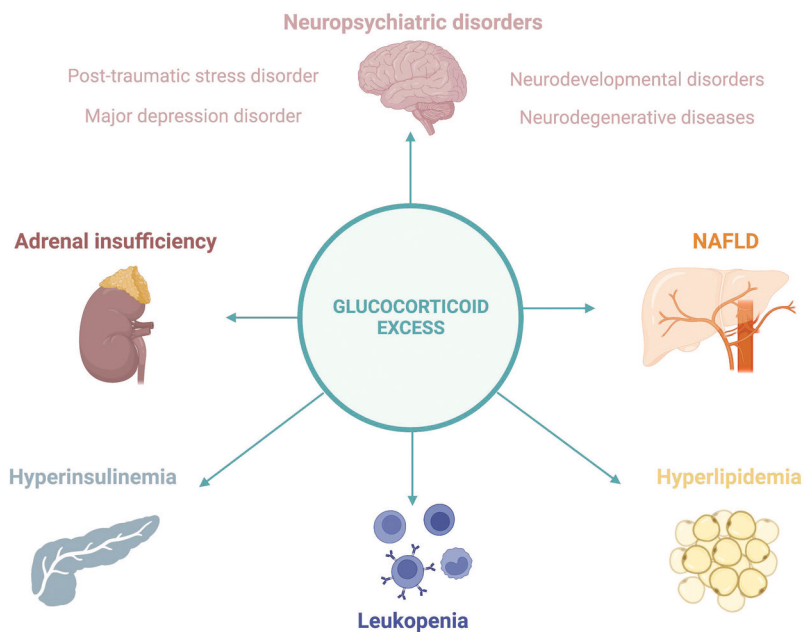


Figure 3. Metabolic and neuropsychiatric disturbances involving the glucocorticoid excess. Abbreviations: NAFLD, non-alcoholic fatty liver disease.

In the brain, disruption in GR signaling as a result of chronic stress or excessive GC exposure, can disrupt neurodevelopment [91, 92], lead to neuropsychiatric disorders, and aggravate neurodegenerative diseases such as Alzheimer's disease [93, 94] (**Fig. 3**). GCs are thought to play an important role in the pathogenesis of post-traumatic stress disorder (PTSD) which can develop in trauma-exposed individuals, with an estimated lifetime prevalence of 6.8% in the United States and 7.4% in the Netherlands before 2010 [95, 96]. More recently, the cross-national prevalence was determined for 24 countries, and the global lifetime PTSD prevalence was 3.9% in the total population and increased to 5.6% in the subset of trauma-exposed individuals [97]. Not all individuals exposed to trauma develop PTSD, suggesting that this pathology is associated with biological vulnerability factors. PTSD is linked to the dysregulation of GR signaling [98, 99], and systematic studies have shown a correlation between PTSD susceptibility and low morning cortisol

levels [100]. Furthermore, a risk allele of *Nr3c1* has recently been associated with lower hair cortisol concentrations and PTSD susceptibility in war veterans (**Fig. 3**) [101].

Depression is the most characterized neuropsychiatric comorbidity in Cushing's syndrome, and studies of major depressive disorder in patients with Cushing's syndrome have reported a prevalence of 50 – 80% [102, 103]. In major depressive disorder, GR-driven negative feedback on the HPA axis is impaired and patients show increased levels of circulating GCs [104, 105]. Depression may be caused by prolonged exposure to stress and GCs. For instance, systemic and inhaled GC medications are correlated with a reduction in white matter integrity, which may lead to adverse neuropsychiatric side effects [106].

The glucocorticoid receptor as a therapeutic target: selectivity and specificity

Considering the variety of metabolic and psychiatric diseases related to alterations in GR signaling, the development of novel therapeutics to target GR is of clinical interest. Mifepristone (RU486) is the predominant GR antagonist used in research and is approved for clinical use. Although surgery remains the first-line treatment for Cushing's syndrome, surgery is not always feasible and is unsuccessful in 20 – 50% of cases [107]. Mifepristone is used to treat patients with Cushing's syndrome in the U.S.A.; but unfortunately, its cross-reactivity with the progesterone receptor makes it less suitable for female patients and the lack of GR selectivity generally increases the risk of adverse side effects [108]. Therefore, a first goal in the development of new GR antagonists is to achieve receptor selectivity, with ligands that have no significant affinity for the MR, or the receptors for progesterone, androgen and estrogens [109]. Such ligands would also be of great benefit to the research community, considering sex dimorphism in stress and GC effects [110–115]. GR agonists have numerous applications mainly due to their anti-inflammatory and immunosuppressive properties. Commonly used GR agonists in clinical practice are prednisone/prednisolone/methylprednisolone, dexamethasone, and betamethasone [116]. Unfortunately, as described in the previous section, excessive activation of the GR causes severe metabolic and neuropsychiatric adverse effects [116–118]. In the case of dexamethasone, the side effects can also be in part attributed to endogenous GC depletion [119–121].

Over the past decade, efforts have been made to develop GR ligands that would retain the beneficial effects of GR activation or inhibition while minimizing adverse side effects. This new category of GR ligands includes “dissociating compounds” that preserve GC anti-inflammatory properties but limit GR-driven transactivation

to only a subset of genes, “soft” steroids with very short half-lives, gene-selective compounds, and selective glucocorticoid receptor modulators [122, 123]. Dissociating ligands typically are aimed to separate transcriptional activation from transcriptional repression, under the assumption that the latter effects are responsible for the therapeutic efficacy [124]. Selective GR modulators are specific in the sense that they target only a subset of gene networks and pathways by inducing a GR conformation that allows distinct GR interactions with downstream signaling factors [109, 125–127].

The ability to modulate the GR interactome results in tissue-, cell-, gene-, and context-specific GR signaling. Selective GR antagonists and selective GR modulators both bear substantial promise in the therapeutic targeting of stress- and GC-related diseases. Beyond their therapeutic value, selective GR antagonists and modulators allow the identification of the molecular mechanisms responsible for the adaptive and maladaptive effects of GCs on metabolic health and brain function.

THESIS OUTLINE

The aim of this thesis is to dissect the underlying genomic and epigenomic mechanisms of GR signaling in metabolic diseases and brain function, and to further characterize the treatment properties of current selective GR antagonists and modulators. In **Chapter 2**, we describe the development of a preclinical pipeline to identify novel selective GR antagonists with beneficial properties in metabolic diseases, from which we identified CORT125329 as the most promising candidate for further clinical evaluation. In **Chapter 3**, we compared the properties of the selective GR antagonist, relacorilant, with those of the clinically used antagonist mifepristone. Relacorilant induced modest disinhibition of the HPA axis compared to mifepristone, which represents a considerable advantage over mifepristone in the treatment of Cushing's syndrome. This effect was associated with lack of GR antagonism in the brain and the absence of classic corepressors in the GR interactome. In **Chapter 4**, we investigated the molecular mechanisms underlying the beneficial effects of the GR modulator CORT118335 in NAFLD and NASH. Our results suggest that the beneficial properties of CORT118335 rely on cell- and gene-specific transcriptional effects and a unique GR interactome that lacks important chromatin remodelers. In **Chapter 5**, we provide an overview of GR and MR actions in the brain and highlight the potential of selective GR targeting in stress-related psychiatric disorders. In **Chapter 6**, we created a gene expression atlas in the mouse hippocampus that recapitulated the cell-specific expression of GR and MR, their target genes, transcriptional coregulators, and the neuropeptides and neurotransmitter receptor repertoire. This study shows the complex cellular heterogeneity of corticosteroid receptor signaling networks in the brain. Finally, in **Chapter 7**, we explored GR signaling in a mouse model of the neurodevelopmental disorder Angelman syndrome, which is characterized by the neuronal absence of the GR interacting protein UBE3A. Our results showed that mice with Angelman syndrome are more sensitive to acute elevations in GC levels and, therefore, are more likely to develop stress maladaptation.

REFERENCES

1. Cannon WB (1929). Organization for physiological homeostasis. *Physiological Reviews*. 9(3): 399–431. doi: 10.1152/physrev.1929.9.3.399.
2. Selye H (1936). A Syndrome produced by Diverse Nocuous Agents. *Nature*. 138(3479): 32–32. doi: 10.1038/138032a0.
3. Selye H (1950). Stress and the General Adaptation Syndrome. *Br Med J*. 1(4667): 1383–1392. doi: 10.1136/bmj.1.4667.1383.
4. Joëls M, and Baram TZ (2009). The neuro-symphony of stress. *Nat Rev Neurosci*. 10(6): 459–466. doi: 10.1038/nrn2632.
5. Sara SJ (2009). The locus coeruleus and noradrenergic modulation of cognition. *Nat Rev Neurosci*. 10(3): 211–223. doi: 10.1038/nrn2573.
6. van Dongen PAM (1981). The central noradrenergic transmission and the locus coeruleus: A review of the data, and their implications for neurotransmission and neuromodulation. *Progress in Neurobiology*. 16(2): 117–143. doi: 10.1016/0301-0082(81)90009-5.
7. Amaral DG, and Sinnamon HM (1977). The locus coeruleus: neurobiology of a central noradrenergic nucleus. *Progress in Neurobiology*. 9(3): 147–196. doi: 10.1016/0301-0082(77)90016-8.
8. Krugers H, Karst H, and Joels M (2012). Interactions between noradrenaline and corticosteroids in the brain: from electrical activity to cognitive performance. *Frontiers in Cellular Neuroscience*. 6.
9. Morilak DA, Barrera G, Echevarria DJ, Garcia AS, Hernandez A, Ma S, and Petre CO (2005). Role of brain norepinephrine in the behavioral response to stress. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*. 29(8): 1214–1224. doi: 10.1016/j.pnpbp.2005.08.007.
10. Ulrich-Lai YM, and Herman JP (2009). Neural regulation of endocrine and autonomic stress responses. *Nat Rev Neurosci*. 10(6): 397–409. doi: 10.1038/nrn2647.
11. Cortez V, Santana M, Marques AP, Mota A, Rosmaninho-Salgado J, and Cavadas C (2012). Regulation of catecholamine release in human adrenal chromaffin cells by β -adrenoceptors. *Neurochemistry International*. 60(4): 387–393. doi: 10.1016/j.neuint.2011.12.018.
12. Cook CJ (2004). Stress induces CRF release in the paraventricular nucleus, and both CRF and GABA release in the amygdala. *Physiology & Behavior*. 82(4): 751–762. doi: 10.1016/j.physbeh.2004.06.013.
13. Kim JS, Han SY, and Iremonger KJ (2019). Stress experience and hormone feedback tune distinct components of hypothalamic CRH neuron activity. *Nat Commun*. 10(1): 5696. doi: 10.1038/s41467-019-13639-8.
14. Hauger RL, Risbrough V, Brauns O, and Dautzenberg FM Corticotropin Releasing Factor (CRF) Receptor Signaling in the Central Nervous System: New Molecular Targets. *CNS & Neurological Disorders - Drug Targets*. 5(4): 453–479.
15. Sommer WH, Rimondini R, Hansson AC, Hipskind PA, Gehlert DR, Barr CS, and Heilig MA (2008). Upregulation of Voluntary Alcohol Intake, Behavioral Sensitivity to Stress, and Amygdala Crhr1 Expression Following a History of Dependence. *Biological Psychiatry*. 63(2): 139–145. doi: 10.1016/j.biopsych.2007.01.010.
16. Chen Y, Bender RA, Frotscher M, and Baram TZ (2001). Novel and Transient Populations of Corticotropin-Releasing Hormone-Expressing Neurons in Developing Hippocampus Suggest Unique Functional Roles: A Quantitative Spatiotemporal Analysis. *J Neurosci*. 21(18): 7171–7181. doi: 10.1523/JNEUROSCI.21-18-07171.2001.

17. Valentino RJ, and Van Bockstaele E (2008). Convergent regulation of locus coeruleus activity as an adaptive response to stress. *European Journal of Pharmacology*. 583(2): 194–203. doi: 10.1016/j.ejphar.2007.11.062.
18. Van Pett K, Viau V, Bittencourt JC, Chan R KW, Li H-Y, Arias C, Prins GS, Perrin M, Vale W, and Sawchenko PE (2000). Distribution of mRNAs encoding CRF receptors in brain and pituitary of rat and mouse. *Journal of Comparative Neurology*. 428(2): 191–212. doi: 10.1002/1096-9861(20001211)428:2<191::AID-CNE1>3.0.CO;2-U.
19. Refojo D, Schweizer M, Kuehne C, Ehrenberg S, Thoeringer C, Vogl AM, Dedic N, Schumacher M, von Wolff G, Avrabos C, Touma C, Engblom D, Schütz G, Nave K-A, Eder M, Wotjak CT, Sillaber I, Holsboer F, Wurst W, and Deussing JM (2011). Glutamatergic and Dopaminergic Neurons Mediate Anxiogenic and Anxiolytic Effects of CRHR1. *Science*. 333(6051): 1903–1907. doi: 10.1126/science.1202107.
20. Swanson LW, Sawchenko PE, Rivier J, and Vale WW (1983). Organization of ovine corticotropin-releasing factor immunoreactive cells and fibers in the rat brain: an immunohistochemical study. *Neuroendocrinology*. 36(3): 165–186. doi: 10.1159/000123454.
21. Whitnall MH, Smyth D, and Gainer H (1987). Vasopressin Coexists in Half of the Corticotropin-Releasing Factor Axons Present in the External Zone of the Median Eminence in Normal Rats. *NEN*. 45(5): 420–424. doi: 10.1159/000124768.
22. Scott LV, and Dinan TG (1998). Vasopressin and the regulation of hypothalamic-pituitary-adrenal axis function: Implications for the pathophysiology of depression. *Life Sciences*. 62(22): 1985–1998. doi: 10.1016/S0024-3205(98)00027-7.
23. Jard S, Gaillard RC, Guillon G, Marie J, Schoenenberg P, Muller AF, Manning M, and Sawyer WH (1986). Vasopressin antagonists allow demonstration of a novel type of vasopressin receptor in the rat adenohypophysis. *Mol Pharmacol*. 30(2): 171–177. 3016500.
24. Lolait SJ, O'Carroll AM, Mahan LC, Felder CC, Button DC, Young WS, Mezey E, and Brownstein MJ (1995). Extrahypothalamic expression of the rat V1b vasopressin receptor gene. *Proceedings of the National Academy of Sciences*. 92(15): 6783–6787. doi: 10.1073/pnas.92.15.6783.
25. De Souza EB (1995). Corticotropin-releasing factor receptors: Physiology, pharmacology, biochemistry and role in central nervous system and immune disorders. *Psychoneuroendocrinology*. 20(8): 789–819. doi: 10.1016/0306-4530(95)00011-9.
26. Raftoyianni A, Roth LC, García-González D, Bus T, Kühne C, Monyer H, Spergel DJ, Deussing JM, and Grinevich V (2018). Deciphering the Contributions of CRH Receptors in the Brain and Pituitary to Stress-Induced Inhibition of the Reproductive Axis. *Frontiers in Molecular Neuroscience*. 11.
27. Miller WL (2018). The Hypothalamic-Pituitary-Adrenal Axis: A Brief History. *HRP*. 89(4): 212–223. doi: 10.1159/000487755.
28. Mitani F (2014). Functional zonation of the rat adrenal cortex: the development and maintenance. *Proceedings of the Japan Academy, Series B*. 90(5): 163–183. doi: 10.2183/pjab.90.163.
29. Sheng JA, Bales NJ, Myers SA, Bautista AI, Roueifar M, Hale TM, and Handa RJ (2021). The Hypothalamic-Pituitary-Adrenal Axis: Development, Programming Actions of Hormones, and Maternal-Fetal Interactions. *Frontiers in Behavioral Neuroscience*. 14.
30. Smith SM, and Vale WW (2006). The role of the hypothalamic-pituitary-adrenal axis in neuroendocrine responses to stress. *Dialogues in Clinical Neuroscience*. 8(4): 383–395. doi: 10.31887/DCNS.2006.8.4/ssmith.
31. Gallo-Payet N, and Battista M-C (2014). Steroidogenesis—Adrenal Cell Signal Transduction. In: *Comprehensive Physiology*. John Wiley & Sons, Ltd; pp 889–964.
32. Hawley JM, and Keevil BG (2016). Endogenous glucocorticoid analysis by liquid chromatography–tandem mass spectrometry in routine clinical laboratories. *The Journal of Steroid Biochemistry and Molecular Biology*. 162: 27–40. doi: 10.1016/j.jsbmb.2016.05.014.

33. Travers S, Martinerie L, Bouvattier C, Boileau P, Lombès M, and Pussard E (2017). Multiplexed steroid profiling of gluco- and mineralocorticoids pathways using a liquid chromatography tandem mass spectrometry method. *The Journal of Steroid Biochemistry and Molecular Biology*. 165: 202–211. doi: 10.1016/j.jsbmb.2016.06.005.
34. Kulle AE, Welzel M, Holterhus P-M, and Riepe FG (2013). Implementation of a Liquid Chromatography Tandem Mass Spectrometry Assay for Eight Adrenal C-21 Steroids and Pediatric Reference Data. *HRP*. 79(1): 22–31. doi: 10.1159/000346406.
35. Walker JJ, Spiga F, Gupta R, Zhao Z, Lightman SL, and Terry JR (2015). Rapid intra-adrenal feedback regulation of glucocorticoid synthesis. *Journal of The Royal Society Interface*. 12(102): 20140875. doi: 10.1098/rsif.2014.0875.
36. Gjerstad JK, Lightman SL, and Spiga F (2018). Role of glucocorticoid negative feedback in the regulation of HPA axis pulsatility. *Stress*. 21(5): 403–416. doi: 10.1080/10253890.2018.1470238.
37. Lightman SL, Birnie MT, and Conway-Campbell BL (2020). Dynamics of ACTH and Cortisol Secretion and Implications for Disease. *Endocrine Reviews*. 41(3): bnaa002. doi: 10.1210/endo/bnaa002.
38. Joëls M (2018). Corticosteroids and the brain. *Journal of Endocrinology*. 238(3): R121–R130. doi: 10.1530/JOE-18-0226.
39. Funder JW, Pearce PT, Smith R, and Smith AI (1988). Mineralocorticoid Action: Target Tissue Specificity Is Enzyme, Not Receptor, Mediated. *Science*. 242(4878): 583–585. doi: 10.1126/science.2845584.
40. Funder JW (2017). Aldosterone and Mineralocorticoid Receptors—Physiology and Pathophysiology. *International Journal of Molecular Sciences*. 18(5): 1032. doi: 10.3390/ijms18051032.
41. Edwards CRW, Burt D, McIntyre MA, De Kloet ER, Stewart PM, Brett L, Sutanto WS, and Monder C (1988). LOCALISATION OF 11 β -HYDROXYSTEROID DEHYDROGENASE—TISSUE SPECIFIC PROTECTOR OF THE MINERALOCORTICOID RECEPTOR. *The Lancet*. 332(8618): 986–989. doi: 10.1016/S0140-6736(88)90742-8.
42. Wyrwoll CS, Holmes MC, and Seckl JR (2011). 11 β -Hydroxysteroid dehydrogenases and the brain: From zero to hero, a decade of progress. *Frontiers in Neuroendocrinology*. 32(3): 265–286. doi: 10.1016/j.yfrne.2010.12.001.
43. Leenen FHH (2010). The central role of the brain aldosterone–“ouabain” pathway in salt-sensitive hypertension. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*. 1802(12): 1132–1139. doi: 10.1016/j.bbadis.2010.03.004.
44. Geerling JC, and Loewy AD (2009). Aldosterone in the brain. *American Journal of Physiology-Renal Physiology*. 297(3): F559–F576. doi: 10.1152/ajprenal.90399.2008.
45. de Kloet ER, Joëls M, and Holsboer F (2005). Stress and the brain: from adaptation to disease. *Nature Reviews Neuroscience*. 6(6): 463–475. doi: 10.1038/nrn1683.
46. REUL JMHM, and KLOET ERD (1985). Two Receptor Systems for Corticosterone in Rat Brain: Microdistribution and Differential Occupation. *Endocrinology*. 117(6): 2505–2511. doi: 10.1210/endo-117-6-2505.
47. Weikum ER, Knuesel MT, Ortlund EA, and Yamamoto KR (2017). Glucocorticoid receptor control of transcription: precision and plasticity via allosterity. *Nat Rev Mol Cell Biol*. 18(3): 159–174. doi: 10.1038/nrm.2016.152.
48. Weikum ER, Liu X, and Ortlund EA (2018). The nuclear receptor superfamily: A structural perspective. *Protein Sci*. 27(11): 1876–1892. doi: 10.1002/pro.3496.
49. Pratt WB, Morishima Y, Murphy M, and Harrell M (2006). Chaperoning of Glucocorticoid Receptors. In: Starke K, Gaestel M, editors *Molecular Chaperones in Health and Disease*. Springer, Berlin, Heidelberg; pp 111–138.
50. Scheschowitsch K, Leite J, and Assreuy J (2017). New insights in glucocorticoid receptor signaling – more than just a ligand binding receptor. *Frontiers in Endocrinology*. 8.

51. Galigniana MD, Radanyi C, Renoir J-M, Housley PR, and Pratt WB (2001). Evidence That the Peptidylprolyl Isomerase Domain of the hsp90-binding Immunophilin FKBP52 Is Involved in Both Dynein Interaction and Glucocorticoid Receptor Movement to the Nucleus*. *Journal of Biological Chemistry*. 276(18): 14884–14889. doi: 10.1074/jbc.M010809200.
52. Elbi C, Walker DA, Romero G, Sullivan WP, Toft DO, Hager GL, and DeFranco DB (2004). Molecular chaperones function as steroid receptor nuclear mobility factors. *Proceedings of the National Academy of Sciences*. 101(9): 2876–2881. doi: 10.1073/pnas.0400116101.
53. Davies TH, Ning Y-M, and Sánchez ER (2002). A New First Step in Activation of Steroid Receptors: HORMONE-INDUCED SWITCHING OF FKBP51 AND FKBP52 IMMUNOPHILINS*. *Journal of Biological Chemistry*. 277(7): 4597–4600. doi: 10.1074/jbc.C100531200.
54. Akner G, Wikström A-C, and Gustafsson J-Å (1995). Subcellular distribution of the glucocorticoid receptor and evidence for its association with microtubules. *The Journal of Steroid Biochemistry and Molecular Biology*. 52(1): 1–16. doi: 10.1016/0960-0760(94)00155-F.
55. Harrell JM, Murphy PJM, Morishima Y, Chen H, Mansfield JF, Galigniana MD, and Pratt WB (2004). Evidence for Glucocorticoid Receptor Transport on Microtubules by Dynein*. *Journal of Biological Chemistry*. 279(52): 54647–54654. doi: 10.1074/jbc.M406863200.
56. Galigniana MD, Harrell JM, Housley PR, Patterson C, Fisher SK, and Pratt WB (2004). Retrograde transport of the glucocorticoid receptor in neurites requires dynamic assembly of complexes with the protein chaperone hsp90 and is linked to the CHIP component of the machinery for proteasomal degradation. *Molecular Brain Research*. 123(1): 27–36. doi: 10.1016/j.molbrainres.2003.12.015.
57. Echeverría PC, Mazaira G, Erlejman A, Gomez-Sanchez C, Pilipuk GP, and Galigniana MD (2009). Nuclear Import of the Glucocorticoid Receptor-hsp90 Complex through the Nuclear Pore Complex Is Mediated by Its Interaction with Nup62 and Importin β . *Molecular and Cellular Biology*. 29(17): 4788–4797. doi: 10.1128/MCB.00649-09.
58. Surjit M, Ganti KP, Mukherji A, Ye T, Hua G, Metzger D, Li M, and Chambon P (2011). Widespread Negative Response Elements Mediate Direct Repression by Agonist-Liganded Glucocorticoid Receptor. *Cell*. 145(2): 224–241. doi: 10.1016/j.cell.2011.03.027.
59. Hua G, Ganti KP, and Chambon P (2016). Glucocorticoid-induced tethered transrepression requires SUMOylation of GR and formation of a SUMO-SMRT/NCoR1-HDAC3 repressing complex. *Proceedings of the National Academy of Sciences*. 113(5): E635–E643. doi: 10.1073/pnas.1522826113.
60. Præsthholm SM, Correia CM, and Grøntved L (2020). Multifaceted Control of GR Signaling and Its Impact on Hepatic Transcriptional Networks and Metabolism. *Frontiers in Endocrinology*. 11.
61. Desmet SJ, and Bosscher KD (2017). Glucocorticoid receptors: finding the middle ground. *J Clin Invest*. 127(4): 1136–1145. doi: 10.1172/JCI88886.
62. Lim H-W, Uhlenhaut NH, Rauch A, Weiner J, Hübner S, Hübner N, Won K-J, Lazar MA, Tuckermann J, and Steger DJ (2015). Genomic redistribution of GR monomers and dimers mediates transcriptional response to exogenous glucocorticoid in vivo. *Genome Res*. 25(6): 836–844. doi: 10.1101/gr.188581.114.
63. Louw A (2019). GR Dimerization and the Impact of GR Dimerization on GR Protein Stability and Half-Life. *Frontiers in Immunology*. 10.
64. Presman DM, and Hager GL (2017). More than meets the dimer: What is the quaternary structure of the glucocorticoid receptor? *Transcription*. 8(1): 32–39. doi: 10.1080/21541264.2016.1249045.
65. Presman DM, Ganguly S, Schiltz RL, Johnson TA, Karpova TS, and Hager GL (2016). DNA binding triggers tetramerization of the glucocorticoid receptor in live cells. *Proceedings of the National Academy of Sciences*. 113(29): 8236–8241. doi: 10.1073/pnas.1606774113.

66. Trousson A, Grenier J, Fonte C, Massaad-Massade L, Schumacher M, and Massaad C (2007). Recruitment of the p160 coactivators by the glucocorticoid receptor: Dependence on the promoter context and cell type but not hypoxic conditions. *The Journal of Steroid Biochemistry and Molecular Biology*. 104(3): 305–311. doi: 10.1016/j.jsbmb.2007.03.018.
67. Szapary D, Huang Y, and Simons SS Jr (1999). Opposing Effects of Corepressor and Coactivators in Determining the Dose-Response Curve of Agonists, and Residual Agonist Activity of Antagonists, for Glucocorticoid Receptor-Regulated Gene Expression. *Molecular Endocrinology*. 13(12): 2108–2121. doi: 10.1210/mend.13.12.0384.
68. Allen BL, and Taatjes DJ (2015). The Mediator complex: a central integrator of transcription. *Nat Rev Mol Cell Biol*. 16(3): 155–166. doi: 10.1038/nrm3951.
69. Mottis A, Mouchiroud L, and Auwerx J (2013). Emerging roles of the corepressors NCoR1 and SMRT in homeostasis. *Genes Dev*. 27(8): 819–835. doi: 10.1101/gad.214023.113.
70. Narlikar GJ, Sundaramoorthy R, and Owen-Hughes T (2013). Mechanisms and Functions of ATP-Dependent Chromatin-Remodeling Enzymes. *Cell*. 154(3): 490–503. doi: 10.1016/j.cell.2013.07.011.
71. Engel KB, and Yamamoto KR (2011). The Glucocorticoid Receptor and the Coregulator Brm Selectively Modulate Each Other's Occupancy and Activity in a Gene-Specific Manner. *Molecular and Cellular Biology*. 31(16): 3267–3276. doi: 10.1128/MCB.05351-11.
72. Fryer CJ, and Archer TK (1998). Chromatin remodelling by the glucocorticoid receptor requires the BRG1 complex. *Nature*. 393(6680): 88–91. doi: 10.1038/30032.
73. Wallberg AE, Neely KE, Hassan AH, Gustafsson J-Å, Workman JL, and Wright APH (2000). Recruitment of the SWI-SNF Chromatin Remodeling Complex as a Mechanism of Gene Activation by the Glucocorticoid Receptor τ 1 Activation Domain. *Molecular and Cellular Biology*. 20(6): 2004–2013. doi: 10.1128/MCB.20.6.2004-2013.2000.
74. King HA, Trotter KW, and Archer TK (2012). Chromatin remodeling during glucocorticoid receptor regulated transactivation. *Biochim Biophys Acta*. 1819(7): 716–726. doi: 10.1016/j.bbagr.2012.02.019.
75. Collingwood TN, Urnov FD, and Wolffe AP (1999). Nuclear receptors: coactivators, corepressors and chromatin remodeling in the control of transcription. *Journal of Molecular Endocrinology*. 23(3): 255–275. doi: 10.1677/jme.0.0230255.
76. Quatrini L, and Ugolini S (2021). New insights into the cell- and tissue-specificity of glucocorticoid actions. *Cell Mol Immunol*. 18(2): 269–278. doi: 10.1038/s41423-020-00526-2.
77. Munck A, Guyre PM, and Holbrook NJ (1984). Physiological Functions of Glucocorticoids in Stress and Their Relation to Pharmacological Actions*. *Endocrine Reviews*. 5(1): 25–44. doi: 10.1210/edrv-5-1-25.
78. Dhabhar FS (2009). Enhancing versus Suppressive Effects of Stress on Immune Function: Implications for Immunoprotection and Immunopathology. *NIM*. 16(5): 300–317. doi: 10.1159/000216188.
79. McEwen BS (2017). Neurobiological and Systemic Effects of Chronic Stress. *Chronic Stress*. 1: 2470547017692328. doi: 10.1177/2470547017692328.
80. Arnaldi G, Angeli A, Atkinson AB, Bertagna X, Cavagnini F, Chrousos GP, Fava GA, Findling JW, Gaillard RC, Grossman AB, Kola B, Lacroix A, Mancini T, Mantero F, Newell-Price J, Nieman LK, Sonino N, Vance ML, Giustina A, and Boscaro M (2003). Diagnosis and Complications of Cushing's Syndrome: A Consensus Statement. *The Journal of Clinical Endocrinology & Metabolism*. 88(12): 5593–5602. doi: 10.1210/jc.2003-030871.
81. Ferrau F, and Korbonits M (2015). Metabolic comorbidities in Cushing's syndrome. *European Journal of Endocrinology*. 173(4): M133–M157. doi: 10.1530/EJE-15-0354.
82. Davis GF (1986). Adverse effects of corticosteroids: II. systemic. *Clinics in Dermatology*. 4(1): 161–169. doi: 10.1016/0738-081X(86)90020-9.

83. Gallant C, and Kenny P (1986). Oral glucocorticoids and their complications: A review. *Journal of the American Academy of Dermatology*. 14(2, Part 1): 161-177. doi: 10.1016/S0190-9622(86)70018-2.
84. Covar RA, Leung DYM, McCormick D, Steelman J, Zeitler P, and Spahn JD (2000). Risk factors associated with glucocorticoid-induced adverse effects in children with severe asthma. *Journal of Allergy and Clinical Immunology*. 106(4): 651-659. doi: 10.1067/mai.2000.109830.
85. Oray M, Abu Samra K, Ebrahimiadib N, Meese H, and Foster CS (2016). Long-term side effects of glucocorticoids. *Expert Opinion on Drug Safety*. 15(4): 457-465. doi: 10.1517/14740338.2016.1140743.
86. Rahimi L, Rajpal A, and Ismail-Beigi F (2020). Glucocorticoid-Induced Fatty Liver Disease. *Diabetes Metab Syndr Obes*. 13: 1133-1145. doi: 10.2147/DMSO.S247379.
87. Liu Y-Z, Chen J-K, Zhang Y, Wang X, Qu S, and Jiang C-L (2014). Chronic stress induces steatohepatitis while decreases visceral fat mass in mice. *BMC Gastroenterology*. 14(1): 106. doi: 10.1186/1471-230X-14-106.
88. Rockall AG, Sohaib SA, Evans D, Kaltsas G, Isidori AM, Monson JP, Besser GM, Grossman AB, and Reznick RH (2003). Hepatic steatosis in Cushing's syndrome: a radiological assessment using computed tomography. *European Journal of Endocrinology*. 149(6): 543-548. doi: 10.1530/eje.0.1490543.
89. Woods CP, Hazlehurst JM, and Tomlinson JW (2015). Glucocorticoids and non-alcoholic fatty liver disease. *The Journal of Steroid Biochemistry and Molecular Biology*. 154: 94-103. doi: 10.1016/j.jsbmb.2015.07.020.
90. Prete A, and Bancos I (2021). Glucocorticoid induced adrenal insufficiency. *BMJ*. 374: n1380. doi: 10.1136/bmj.n1380.
91. Fukumoto K, Morita T, Mayanagi T, Tanokashira D, Yoshida T, Sakai A, and Sobue K (2009). Detrimental effects of glucocorticoids on neuronal migration during brain development. *Mol Psychiatry*. 14(12): 1119-1131. doi: 10.1038/mp.2009.60.
92. Miller JG, Chahal R, and Gotlib IH (2022). Early Life Stress and Neurodevelopment in Adolescence: Implications for Risk and Adaptation. In: Miczek KA, Sinha R, editors *Neuroscience of Social Stress*. Springer International Publishing, Cham; pp 313-339.
93. Sharma VK, and Singh TG Navigating Alzheimer's Disease via Chronic Stress: The Role of Glucocorticoids. *Current Drug Targets*. 21(5): 433-444.
94. Saeedi M, and Rashidy-Pour A (2021). Association between chronic stress and Alzheimer's disease: Therapeutic effects of Saffron. *Biomedicine & Pharmacotherapy*. 133: 110995. doi: 10.1016/j.biopha.2020.110995.
95. Kessler RC, Berglund P, Demler O, Jin R, Merikangas KR, and Walters EE (2005). Lifetime Prevalence and Age-of-Onset Distributions of DSM-IV Disorders in the National Comorbidity Survey Replication. *Archives of General Psychiatry*. 62(6): 593-602. doi: 10.1001/archpsyc.62.6.593.
96. de Vries G-J, and Olff M (2009). The lifetime prevalence of traumatic events and posttraumatic stress disorder in the Netherlands. *Journal of Traumatic Stress*. 22(4): 259-267. doi: 10.1002/jts.20429.
97. Koenen KC et al. (2017). Posttraumatic stress disorder in the World Mental Health Surveys. *Psychological Medicine*. 47(13): 2260-2274. doi: 10.1017/S0033291717000708.
98. Sarapas C, Cai G, Bierer LM, Golier JA, Galea S, Ising M, Rein T, Schmeidler J, Müller-Myhsok B, Uhr M, Holsboer F, Buxbaum JD, and Yehuda R (2011). Genetic markers for PTSD risk and resilience among survivors of the World Trade Center attacks. *Disease Markers*. 30(2-3): 101-110. doi: 10.3233/DMA-2011-0764.
99. Zuiden M van, Geuze E, Willemsen HLDM, Vermetten E, Maas M, Amarouchi K, Kavelaars A, and Heijnen CJ (2012). Glucocorticoid Receptor Pathway Components Predict Posttraumatic Stress Disorder Symptom Development: A Prospective Study. *Biological Psychiatry*. 71(4): 309-316. doi: 10.1016/j.biopsych.2011.10.026.

100. Morris MC, Compas BE, and Garber J (2012). Relations among posttraumatic stress disorder, comorbid major depression, and HPA function: A systematic review and meta-analysis. *Clinical Psychology Review*. 32(4): 301–315. doi: 10.1016/j.cpr.2012.02.002.
101. Castro-Vale I, Durães C, van Rossum EFC, Staufenbiel SM, Severo M, Lemos MC, and Carvalho D (2021). The Glucocorticoid Receptor Gene (NR3C1) 9β SNP Is Associated with Posttraumatic Stress Disorder. *Healthcare*. 9(2): 173. doi: 10.3390/healthcare9020173.
102. Pivonello R, Simeoli C, De Martino MC, Cozzolino A, De Leo M, Iacuanillo D, Pivonello C, Negri M, Pellecchia MT, Iasevoli F, and Colao A (2015). Neuropsychiatric disorders in Cushing's syndrome. *Frontiers in Neuroscience*. 9.
103. Starkman MN (2013). Neuropsychiatric Findings in Cushing Syndrome and Exogenous Glucocorticoid Administration. *Endocrinology and Metabolism Clinics of North America*. 42(3): 477–488. doi: 10.1016/j.ecl.2013.05.010.
104. Pariante CM (2009). Risk Factors for Development of Depression and Psychosis. *Annals of the New York Academy of Sciences*. 1179(1): 144–152. doi: 10.1111/j.1749-6632.2009.04978.x.
105. Anacker C, Zunszain PA, Carvalho LA, and Pariante CM (2011). The glucocorticoid receptor: Pivot of depression and of antidepressant treatment? *Psychoneuroendocrinology*. 36(3): 415–425. doi: 10.1016/j.psyneuen.2010.03.007.
106. Meulen M van der, Amaya JM, Dekkers OM, and Meijer OC (2022). Association between use of systemic and inhaled glucocorticoids and changes in brain volume and white matter microstructure: a cross-sectional study using data from the UK Biobank. *BMJ Open*. 12(8): e062446. doi: 10.1136/bmjopen-2022-062446.
107. Castinetti F, Brue T, and Conte-Devolx B (2012). The use of the glucocorticoid receptor antagonist mifepristone in Cushing's syndrome. *Current Opinion in Endocrinology, Diabetes and Obesity*. 19(4): 295–299. doi: 10.1097/MED.0b013e32835430bf.
108. Robbins A, and Spitz IM (1996). Mifepristone: Clinical Pharmacology. *Clinical Obstetrics and Gynecology*. 39(2): 436–450.
109. Meijer OC, Koorneef LL, and Kroon J (2018). Glucocorticoid receptor modulators. *Annales d'Endocrinologie*. 79(3): 107–111. doi: 10.1016/j.ando.2018.03.004.
110. McEwen BS (2014). Sex, stress and the brain: interactive actions of hormones on the developing and adult brain. *Climacteric*. 17(sup2): 18–25. doi: 10.3109/13697137.2014.949662.
111. Wolf DC, Desgent S, Sanon NT, Chen J-S, Elkaim LM, Bosoi CM, Awad PN, Simard A, Salam MT, Bilodeau G-A, Duss S, Sawan M, Lewis EC, and Weil AG (2021). Sex differences in the developing brain impact stress-induced epileptogenicity following hyperthermia-induced seizures. *Neurobiology of Disease*. 161: 105546. doi: 10.1016/j.nbd.2021.105546.
112. Bangasser DA (2013). Sex differences in stress-related receptors: "micro" differences with "macro" implications for mood and anxiety disorders. *Biology of Sex Differences*. 4(1): 2. doi: 10.1186/2042-6410-4-2.
113. Buurstede JC, Paul SN, De Bosscher K, Meijer OC, and Kroon J (2022). Hepatic glucocorticoid-induced transcriptional regulation is androgen-dependent after chronic but not acute glucocorticoid exposure. *The FASEB Journal*. 36(4): e22251. doi: 10.1096/fj.202101313R.
114. Spaanderman DCE, Nixon M, Buurstede JC, Sips HHCM, Schilperoord M, Kuipers EN, Backer EA, Kooijman S, Rensen PCN, Homer NZM, Walker BR, Meijer OC, and Kroon J (2019). Androgens modulate glucocorticoid receptor activity in adipose tissue and liver. *Journal of Endocrinology*. 240(1): 51–63. doi: 10.1530/JOE-18-0503.
115. Kroon J, Pereira AM, and Meijer OC (2020). Glucocorticoid Sexual Dimorphism in Metabolism: Dissecting the Role of Sex Hormones. *Trends in Endocrinology & Metabolism*. 31(5): 357–367. doi: 10.1016/j.tem.2020.01.010.

116. Williams DM (2018). Clinical Pharmacology of Corticosteroids. *Respiratory Care*. 63(6): 655–670. doi: 10.4187/respcare.06314.
117. Ellero-Simatos S, Szymańska E, Rullmann T, Dokter WH, Ramaker R, Berger R, van Iersel TM, Smilde AK, Hankemeier T, and Alkema W (2012). Assessing the metabolic effects of prednisolone in healthy volunteers using urine metabolic profiling. *Genome Medicine*. 4(11): 94. doi: 10.1186/gm395.
118. Ciriaco M, Ventrice P, Russo G, Scicchitano M, Mazzitello G, Scicchitano F, and Russo E (2013). Corticosteroid-related central nervous system side effects. *Journal of Pharmacology and Pharmacotherapeutics*. 4(1_suppl): S94–S98. doi: 10.4103/0976-500X.120975.
119. Koning A-SCAM, Habets PC, Bogaards M, Kroon J, Santen HM van, Bont JM de, and Meijer OC (2022). Mineralocorticoid receptor status in the human brain after dexamethasone treatment: a single case study. *Endocrine Connections*. 11(3). doi: 10.1530/EC-21-0425.
120. Meijer OC, and de Kloet ER (2017). A Refill for the Brain Mineralocorticoid Receptor: The Benefit of Cortisol Add-On to Dexamethasone Therapy. *Endocrinology*. 158(3): 448–454. doi: 10.1210/en.2016-1495.
121. Koorneef LL, van der Meulen M, Kooijman S, Sánchez-López E, Scheerstra JF, Voorhoeve MC, Ramesh ANN, Rensen PCN, Giera M, Kroon J, and Meijer OC (2022). Dexamethasone-associated metabolic effects in male mice are partially caused by depletion of endogenous corticosterone. *Frontiers in Endocrinology*. 13.
122. McMaster A, and Ray DW (2008). Drug Insight: selective agonists and antagonists of the glucocorticoid receptor. *Nat Rev Endocrinol*. 4(2): 91–101. doi: 10.1038/ncpendmet0745.
123. Lesovaya E, Yemelyanov A, Swart AC, Swart P, Haegeman G, and Budunova I (2015). Discovery of Compound A – a selective activator of the glucocorticoid receptor with anti-inflammatory and anti-cancer activity. *Oncotarget*. 6(31): 30730–30744. doi: 10.18632/oncotarget.5078.
124. De Bosscher K, Berghe WV, Beck IME, Van Molle W, Hennuyer N, Hapgood J, Libert C, Staels B, Louw A, and Haegeman G (2005). A fully dissociated compound of plant origin for inflammatory gene repression. *Proceedings of the National Academy of Sciences*. 102(44): 15827–15832. doi: 10.1073/pnas.0505554102.
125. Monczor F, Chatzopoulou A, Zappia CD, Houtman R, Meijer OC, and Fitzsimons CP (2019). A Model of Glucocorticoid Receptor Interaction With Coregulators Predicts Transcriptional Regulation of Target Genes. *Frontiers in Pharmacology*. 10.
126. van der Laan S, Lachize SB, Vreugdenhil E, de Kloet ER, and Meijer OC (2008). Nuclear Receptor Coregulators Differentially Modulate Induction and Glucocorticoid Receptor-Mediated Repression of the Corticotropin-Releasing Hormone Gene. *Endocrinology*. 149(2): 725–732. doi: 10.1210/en.2007-1234.
127. Viho EMG, Buurstede JC, Mahfouz A, Koorneef LL, van Weert LTCM, Houtman R, Hunt HJ, Kroon J, and Meijer OC (2019). Corticosteroid Action in the Brain: The Potential of Selective Receptor Modulation. *NEN*. 109(3): 266–276. doi: 10.1159/000499659.