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IMMUNOPATHOLOGY AND INFECTIOUS DISEASES

Restoration of Regulatory T-Cell Function in Dry Eye Disease by Antagonizing Substance P/Neurokinin-1 Receptor



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Substance P (SP) is a tachykinin neuropeptide, implicated in the pathogenesis of various inflammatory conditions and a critical mediator in pain transmission. Recently, the role of SP was described in the pathogenesis of dry eye disease (DED) through its role in the maturation of antigen-presenting cells at the ocular surface after exposure to desiccating stress. However, the effect of SP on regulatory T cells (Tregs), which are functionally impaired in DED, remains unclear. This study examined the phenotypic and functional changes in Tregs in response to SP in DED. The *in vitro* cultures of normal Tregs in the presence of SP led to a significant reduction in both Treg frequencies and their suppressive function, which was prevented by the addition of an SP receptor (neurokinin-1 receptor) antagonist. Furthermore, *in vivo* treatment with the neurokinin-1 receptor antagonist in DED mice effectively restored Treg function, suppressed pathogenic T helper 17 response, and significantly ameliorated the disease. Our results show that a significant increase in SP levels promotes Treg dysfunction in DED, and blockade of SP effectively restores Treg function and suppresses DED severity. (*Am J Pathol* 2020, 190: 1859–1866; <https://doi.org/10.1016/j.ajpath.2020.05.011>)

Dry eye disease (DED) is a highly prevalent multifactorial ocular surface disorder that presents either as a stand-alone disease or as an ocular manifestation of a systemic disease such as graft-versus-host disease, Sjögren syndrome, and rheumatoid arthritis.^{1–5} Studies have calculated the prevalence of symptomatic DED in the United States to be 14.5%, and the total annual cost for the management of this disease is estimated to be near \$4 billion.^{6,7}

There is ample evidence indicating the role of autoimmune inflammation in the pathogenesis of DED.⁸ Multiple studies,^{9–12} have shown that inflammation in DED is principally mediated by the unrestrained effector T helper 17 (Th17) response resulting from dysfunction of the immunosuppressive regulatory T cells (Tregs).

Substance P (SP, also known as TAC1) is an 11 amino acid neuropeptide belonging to the tachykinin family.¹³ It is known to mediate neurotransmission of pain, as well as neuro-inflammation, by binding primarily to the neurokinin

1 receptor (NK-1R).^{14,15} It is secreted by sensory nerve endings, including corneal trigeminal nerves, and functions by binding to NK-1R.^{16–19} SP is also secreted by corneal epithelial cells and various immune cells, including macrophages, monocytes, and dendritic cells during inflammation.^{20–25} SP has known pro-inflammatory effects and plays a role in the pathogenesis of autoimmune disorders such as rheumatoid arthritis and inflammatory bowel disease, where dysfunction of Tregs is similarly involved in disease pathogenesis as in DED.^{12,26–28} Recent studies showed the role played by increased levels of SP on the ocular surface in promoting the maturation of corneal antigen-presenting

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cells, which subsequently migrate to the draining lymph nodes (DLNs) and activate effector Th17 cells in DED.²⁹ However, the precise function of SP on Tregs in autoimmunity or other inflammatory conditions remains unknown.

In the current study, we first evaluated the expression levels of SP in the DLNs and NK-1R by Tregs in DED mice. The results show that SP levels as well as the frequencies of NK-1R⁺ Tregs in DLNs are increased in DED. To test the hypothesis that SP directly influences Tregs through NK-1R, we conducted *in vitro* experiments of Tregs in the presence of SP. We also investigated the effects of inhibiting SP signaling by using NK-1R antagonists on Treg function and on recovery of regulatory function.

Materials and Methods

Animals

Eight-to-ten-week-old C57BL/6 female mice (Charles River Laboratories, Wilmington, MA) were used in all of the experiments. The protocol was approved by the Schepens Eye Research Institute Animal Care and Use Committee. All animals were treated in accordance with the Association for Research in Vision and Ophthalmology Statement for the use of animals in ophthalmic and vision research.³⁰

Induction of DED and Clinical Evaluation

DED was induced by continuously exposing the mice to desiccating stress (<20% relative humidity with an airflow of 15 L/min at a temperature of 21°C to 23°C) in a controlled-environment chamber for 14 days.³¹ Age- and sex-matched mice (8-to-10-week-old; C57BL/6 female) placed in the standard vivarium were used as control mice. Corneal fluorescein staining (CFS) was used to evaluate corneal epitheliopathy. Briefly, 1 μ L of 2% fluorescein was applied to the lateral conjunctival sac of the mice, and slit lamp examination was performed under cobalt blue light. Punctate staining was recorded in a masked fashion by using the standard National Eye Institute grading system with the score ranging from 0 to 3 for each of the five areas of the cornea.³²

Isolation of Tregs and *in Vitro* Culture

The submandibular and cervical DLNs were harvested from normal and DED mice, and single-cell suspensions were prepared as previously described.¹² CD4⁺CD25⁺ Tregs were isolated by magnetic separation using a Treg isolation kit (Miltenyi Biotec, Somerville, MA). The purity of sorted cells (>95% Foxp3⁺ Tregs) was confirmed by using flow cytometry analysis. Tregs were resuspended in RPMI 1640-based complete medium at a concentration of 1×10^6 cells/mL. Subsequently, 100 μ L of cell suspension

was plated in each well of a 96-well plate (Corning Life Sciences Inc., Tewksbury, MA). SP (1 μ mol/L; Tocris Bioscience, Minneapolis, MN), Spantide I (10 μ mol/L; Tocris Bioscience), or both were added to the Treg cultures. After 48 hours of incubation, cells were collected for flow cytometry analysis.

Flow Cytometry Analysis

Single-cell suspensions were prepared from conjunctival tissues of mice, as previously described.³³ Briefly, conjunctivae were harvested and cut into small fragments, followed by digestion with 2 mg/mL collagenase type IV (MilliporeSigma, Burlington, MA) and 0.05 mg/mL DNase I (Roche Holding AG, Basel, Switzerland) for 1 hour at 37°C with agitation. The suspension was then filtered through a 70- μ m cell strainer (BD Biosciences Inc., San Jose, CA). Single-cell suspensions from DLNs were prepared by homogenizing tissues and subsequently filtering them through a 70- μ m cell strainer. Cells were stained with fluorescein isothiocyanate-conjugated anti-CD4, phycoerythrin (PE)/Cy7-conjugated Foxp3, APC-conjugated cytotoxic T lymphocyte associated antigen-4 (CTLA-4) or lymphocyte activation gene-3, and PE-conjugated programmed cell death 1 (PD-1) or glucocorticoid-induced tumor necrosis factor receptor (GITR) (BioLegend Inc., San Diego, CA) for flow cytometry analysis. NK-1R was stained with primary antibody (Santa Cruz Biotechnology Inc., Dallas, TX) and secondary antibody (fluorescein isothiocyanate; Jackson ImmunoResearch Laboratories, Inc., West Grove, PA). For intracellular IL-17A staining, the cells were stimulated with 50 ng/mL phorbol myristate acetate and 500 ng/mL ionomycin (MilliporeSigma) for 6 hours at 37°C and 5% carbon dioxide in the presence of GolgiStop (BD Biosciences Inc.) to inhibit cytokine secretion before staining and then stained with PE-Cy7-conjugated anti-IL-17A antibody (BioLegend Inc.). The stained cells were examined with an LSR II flow cytometer (BD Biosciences Inc.) and the results analyzed. Data were analyzed by using FlowJo software version X.5.3 (FlowJo LLC., Ashland, OR).

Treg Suppression Assay

The suppressive function of Tregs was examined by using the suppression assay, as previously described.¹² Briefly, Tregs and responder CD4⁺ T (Tresp) cells were isolated from the DLNs. Purified total CD4⁺ Tresp cells (1×10^5) were co-cultured with CD4⁺CD25⁺ Tregs (5×10^4), T cell-depleted syngeneic splenocytes (1×10^5), and 1 μ g/mL anti-CD3 antibody for 3 days. Proliferation of Tresp cells without adding Tregs was considered as control with 0% suppression. The proliferation was measured by using the BrdU proliferation kit (MilliporeSigma), and the percentage of suppression was calculated by using the following formula:

$$\% \text{suppression} = \frac{(\text{Tresp proliferation without Tregs} - \text{Tresp proliferation with Tregs})}{\text{Tresp proliferation without Tregs}} \times 100 \quad (1)$$

Real-Time PCR

DLNs were harvested from mice, and total RNA was isolated by using the RNeasy Micro kit (Qiagen Inc., Germantown, MD) as per the manufacturer's recommendations and reverse transcribed by using an RT transcription kit (Qiagen Inc.). Real-time PCR was performed by using the TaqMan Universal PCR Master mix and predesigned primers SP (*Tac1*, Mm01166996_m1) and *Gapdh* (Mm99999915_g1) (Thermo Fisher Scientific, Waltham, MA) in a Realplex2 system (Eppendorf GmbH, Hamburg, Germany). The *Gapdh* gene was used as an endogenous control for each reaction. The results of quantitative PCR were analyzed by using the comparative CT method in which the target change = $\Delta\Delta\text{CT}$. The results were normalized by the CT value of *Gapdh*, and the mean CT of the relative mRNA level in the control group was used as the calibrator.

Cytokine Expression Assay

For protein extraction, DLNs were harvested and homogenized in radioimmunoprecipitation assay buffer containing protease inhibitors (MilliporeSigma) on ice and centrifuged. The supernatant was assayed by using commercial enzyme-linked immunosorbent assay kits for SP (detection limit, 43.8 pg/mL; R&D Systems, Minneapolis, MN), IL-10, and transforming growth factor- β (TGF- β) (BioLegend Inc.), as well as total protein amount (Thermo Fisher Scientific).

In Vivo Treatment of Mice with Spantide I

DED mice were injected intraperitoneally with 36 μg synthetic Spantide I or vehicle control [phosphate-buffered saline (PBS)] once daily in 100 μL volume starting from one day before DED induction (ie, day -1), until day 14. CFS scoring was recorded on days 0, 3, 7, 10, and 14 to assess disease severity. On day 14, the mice were euthanized, and the tissues were harvested for further flow cytometry and enzyme-linked immunosorbent assay analyses.

Statistical Analysis

The *t*-test or *U*-test was used for comparison of means between the groups, and $P < 0.05$ was considered statistically significant. The data are presented as mean \pm SEM of at least three independent experiments. Prism for Windows software version 8.0 (GraphPad Software, La Jolla, CA) was used for statistical analysis.

Results

SP Expression Levels Increase in DLNs in DED

DLNs are the principal site where Tregs normally regulate effector T cells; however, Tregs have been shown to become dysfunctional in DED.¹² Thus, the association between Treg dysfunction and altered SP levels in the draining lymphoid tissues in DED was first investigated. After 14 days of DED induction, the mRNA expression levels of SP (*Tac1*) in DLNs increased more than fourfold (Figure 1A), associated with a 25% increase in SP protein levels (Figure 1B). There was no significant change in SP levels on day 7 after DED induction.

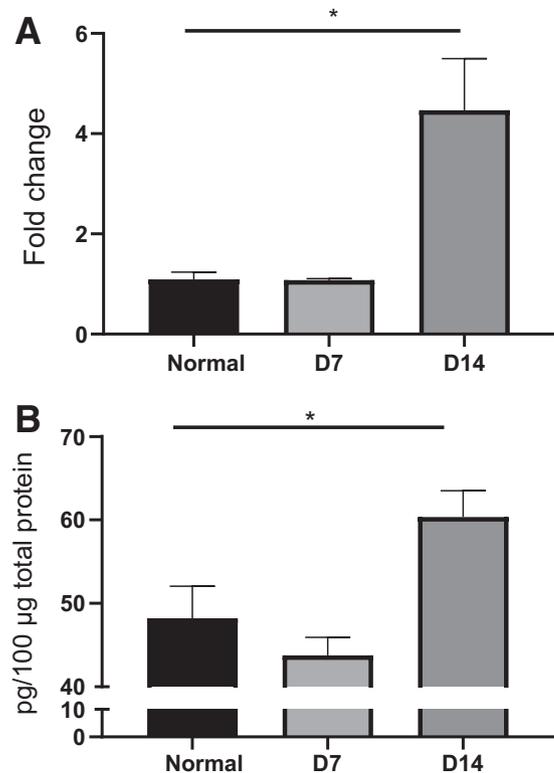


Figure 1 Expression of substance P (SP, also known as *Tac1*) at draining lymph nodes in dry eye disease (DED). The expression levels of SP in draining lymph nodes of normal and DED mice (after 7 and 14 days of induction) were quantified by real-time quantitative PCR data for mRNA [the y axis is fold change of SP expression compared with the normal group (normalized as 1.0)] (A) and by enzyme-linked immunosorbent assay for protein level (the y axis indicates the concentration of SP in lymph nodes per 100 μg of protein) (B). Both mRNA and protein level in DED mice at day 7 (D7) are the same as the normal group but increase significantly in DED mice at day 14 (D14) ($P = 0.04$ and 0.049 , respectively). Data are expressed as means \pm SEM of three independent experiments. $n = 5$ mice per group (A and B). $*P < 0.05$.

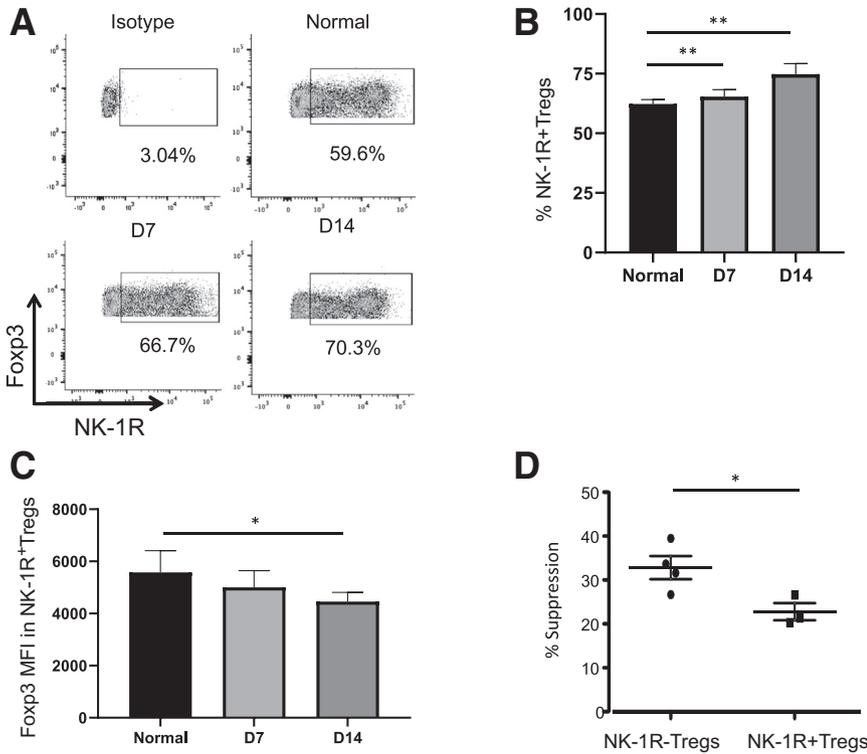


Figure 2 Neurokinin-1 receptor (NK-1R)—positive regulatory T cells (Tregs) increase in the course of dry eye disease (DED) and exhibit impaired suppressive function. **A** and **B**: Representative flow cytometry plots (**A**) and summary data (**B**) showing frequencies of NK-1R⁺ Tregs (CD4⁺CD25⁺FcγR3⁺) in draining lymph nodes of normal and DED mice after 7 and 14 days of DED induction. **C**: Expression levels of FcγR3 by NK-1R⁺ Tregs in different groups are assessed by using mean fluorescence intensity (MFI). **D**: Treg function is assessed by using the suppression assay. Data are expressed as means ± SEM of three independent experiments. *n* = 5 mice per group (**A–C**); *n* = 7 mice per group (**D**). **P* < 0.05, ***P* < 0.01. D7, day 7; D14, day 14; FcγR3, forkhead box P3.

Frequency of Functionally Impaired, SP-Binding NK-1R⁺ Treg Increases in DED

After detecting increased levels of SP in DLNs in DED, the frequencies of Tregs expressing the SP receptor (NK-1R) in DED were evaluated. As shown in Figure 2, A and B, the frequencies of NK-1R⁺ Tregs progressively increased in DED, with 65.4% NK-1R⁺ Tregs on day 7 and 74.8% NK-1R⁺ Tregs on day 14, compared with 50.4% in normal control. Because FcγR3 is the critical transcriptional factor determining the immunosuppressive functions of Tregs, its expression levels on NK-1R⁺ Tregs were examined.^{34,35}

The results showed that FcγR3 expression levels were significantly decreased in NK-1R⁺ Tregs derived from DED mice (Figure 2C). Furthermore, Treg suppression assay showed that the capacity to suppress effector T-cell proliferation was significantly reduced in NK-1R⁺ Tregs compared with NK-1R⁻ Tregs (Figure 2D).

SP Induces Treg Dysfunction *in Vitro* and Is Reversed by the NK-1R Antagonist Spantide I

After establishing elevated levels of SP expression and increased frequencies of NK-1R⁺ Tregs in DED, the direct

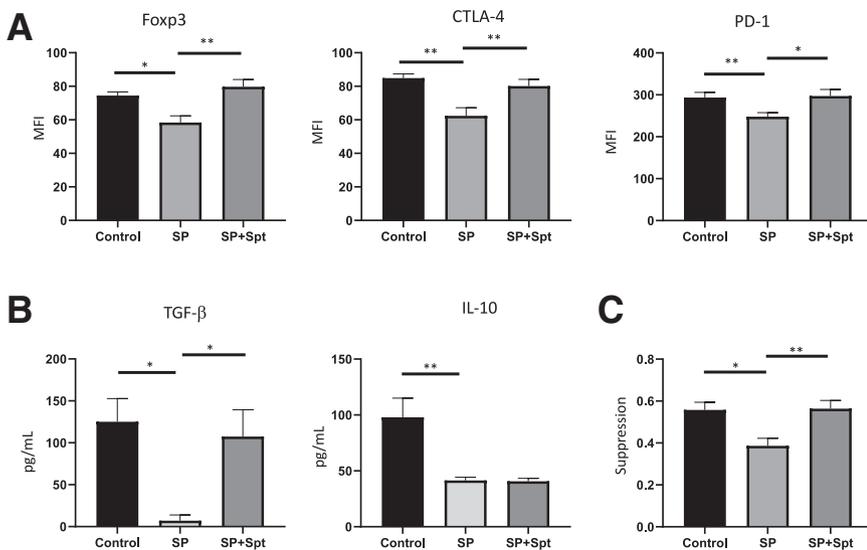


Figure 3 Substance P directly impairs regulatory T cell (Treg) function *in vitro*. Tregs were isolated from lymph nodes of normal mice and incubated in the presence of Substance P (SP; 1 μmol/L), with or without Spantide I (Spt; 10 μmol/L) for 48 hours. Phosphate-buffered saline was used as a control. **A**: Flow cytometry analysis of cultured Tregs showing mean fluorescence intensities (MFIs) of forkhead box P3 (FcγR3), cytotoxic T lymphocyte associated antigen-4 (CTLA-4), and programmed cell death 1 (PD-1) expression by Tregs. **B**: Protein levels of transforming growth factor-β (TGF-β) and IL-10 in culture supernatants are quantified by using enzyme-linked immunosorbent assay. **C**: Suppressive functions of cultured Tregs are assessed by using Treg suppression assay. Data are expressed as means ± SEM of three independent experiments., *n* = 5 mice per group (**A**); *n* = 4 mice per group (**B**); *n* = 9 mice per group (**C**). **P* < 0.05, ***P* < 0.01.

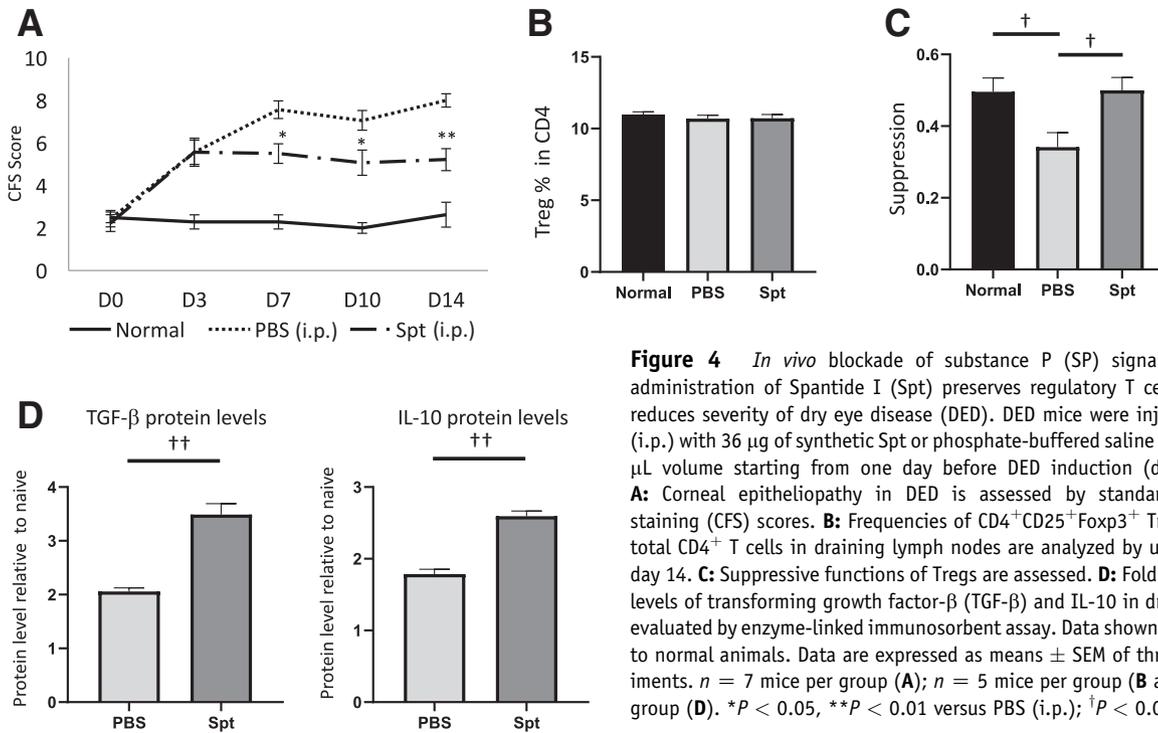


Figure 4 *In vivo* blockade of substance P (SP) signaling through systemic administration of Spantide I (Spt) preserves regulatory T cell (Treg) function and reduces severity of dry eye disease (DED). DED mice were injected intraperitoneally (i.p.) with 36 μg of synthetic Spt or phosphate-buffered saline (PBS) once daily in 100 μL volume starting from one day before DED induction (day -1) until day 14. **A:** Corneal epitheliopathy in DED is assessed by standard corneal fluorescein staining (CFS) scores. **B:** Frequencies of $\text{CD4}^+\text{CD25}^+\text{Foxp3}^+$ Tregs as a proportion of total CD4^+ T cells in draining lymph nodes are analyzed by using flow cytometry at day 14. **C:** Suppressive functions of Tregs are assessed. **D:** Fold increase in the protein levels of transforming growth factor- β (TGF- β) and IL-10 in draining lymph nodes as evaluated by enzyme-linked immunosorbent assay. Data shown are the relative change to normal animals. Data are expressed as means \pm SEM of three independent experiments. $n = 7$ mice per group (**A**); $n = 5$ mice per group (**B** and **C**); $n = 4$ mice per group (**D**). * $P < 0.05$, ** $P < 0.01$ versus PBS (i.p.); $\dagger P < 0.05$, $\dagger\dagger P < 0.01$. D, day.

effect of SP on Treg function was examined. Tregs isolated from normal mice were cultured in the presence of SP with or without Spantide I, an NK-1R antagonist. After 48 hours of culture, Tregs were analyzed for their inhibitory phenotypes and suppressive function. The data show that SP-treated Tregs expressed significantly lower levels of Foxp3 ($P = 0.0101$) and the inhibitory molecules CTLA-4 ($P \leq 0.001$) and PD-1 ($P = 0.039$) compared with control. However, these phenotypic changes in Tregs were reversed with the addition of the NK-1R antagonist (Spantide I) to the culture. Tregs treated with both SP and Spantide I had higher expression levels of Foxp3 ($P = 0.005$), CTLA-4 ($P = 0.0097$), and PD-1 ($P = 0.043$) compared with SP-treated Tregs (Figure 3A). No significant changes were observed in the expression of GITR or lymphocyte activation gene-3 by Tregs after SP treatment (data not shown). The function of cultured Tregs was further evaluated by examining the secreted anti-inflammatory cytokines TGF- β and IL-10 in the supernatants. Both TGF- β ($P = 0.026$) and IL-10 ($P = 0.0026$) protein levels were significantly reduced in the presence of SP compared with controls. Blockade of SP signaling by Spantide I inhibited the SP-induced decrease in TGF- β production by Tregs ($P = 0.038$) compared with the SP-treated group (Figure 3B).

Finally, the suppressive function of cultured Tregs was assessed. Tregs cultured in the presence of SP exhibited significantly decreased capability in suppressing effector T-cell proliferation compared with the Tregs cultured alone ($P = 0.013$), and the addition of Spantide I along with SP prevented the functional impairment of cultured Tregs ($P = 0.0047$) compared with the SP-treated group (Figure 3C). To ensure that Spantide I did not have a direct

effect on Treg function, Tregs were cultured with Spantide I. The data show that Spantide I alone does not significantly affect Treg phenotypes or functions (data not shown).

In Vivo Treatment with an NK-1R Antagonist Reduces Corneal Epitheliopathy in DED

Next, it was investigated whether blockade of SP by Spantide I can reduce DED severity by modulating the underlying immune responses. Spantide I was administered to mice daily by intraperitoneal injection from one day before DED induction until day 14. PBS-injected mice were used as controls. CFS scores were used to evaluate corneal epitheliopathy. The data show significantly lower CFS scores in Spantide I-treated mice compared with the control mice on days 7 ($P = 0.03$), 10 ($P = 0.015$), and 14 ($P < 0.001$) (Figure 4A).

NK-1R Antagonist Prevents Treg Dysfunction in DED

To understand the underlying mechanisms by which Spantide I ameliorates DED *in vivo*, Treg frequencies, inhibitory phenotypes, and suppressive functions in treated DED mice were further analyzed. No significant differences were observed in the frequencies of Tregs as a proportion of total CD4^+ T cells in DLNs of normal (non-DED) mice, PBS-treated DED mice, and Spantide I-treated DED mice (Figure 4B). However, consistent with a previous report on Treg dysfunction in DED, it was observed that Tregs derived from the PBS-treated group had significantly compromised suppressive function compared with normal

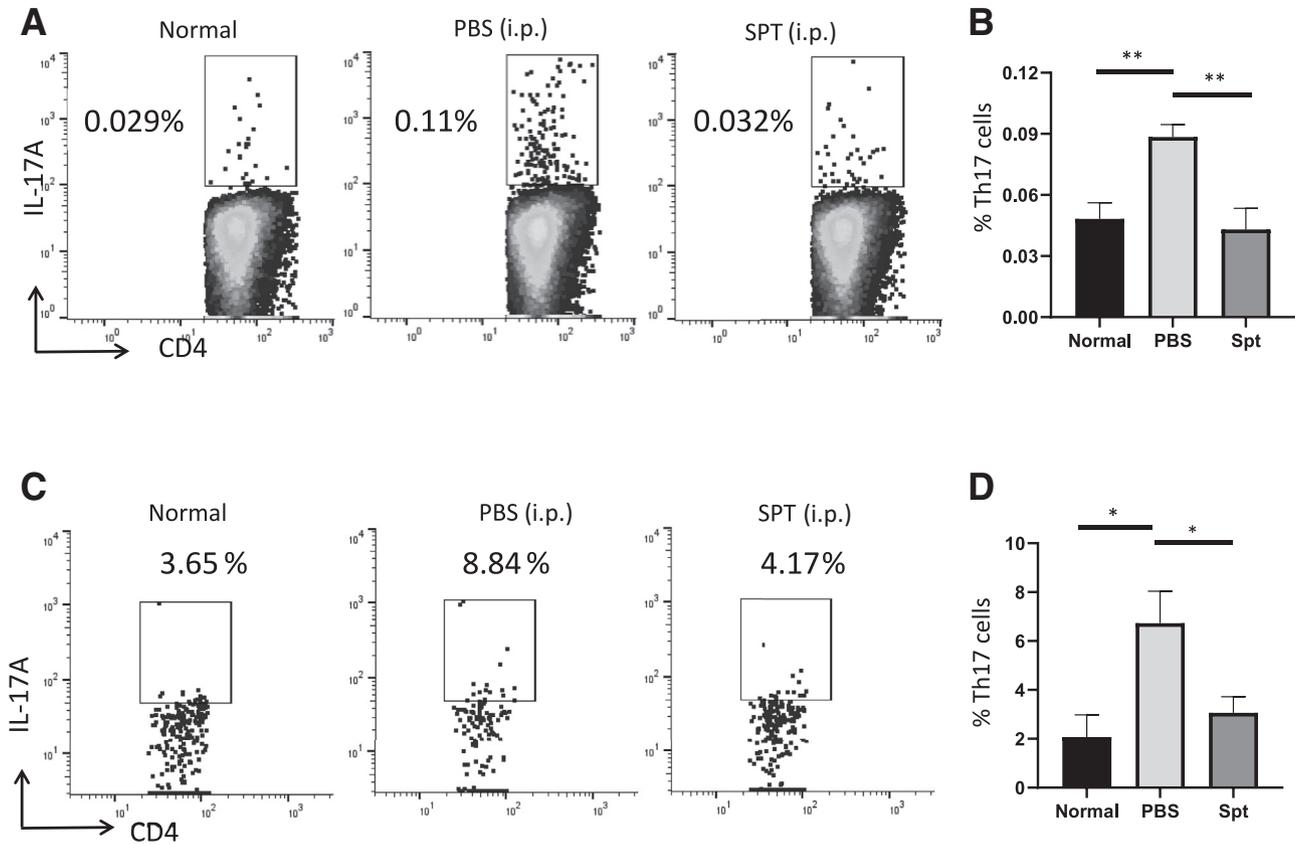


Figure 5 *In vivo* blockade of substance P (SP) signaling through systemic administration of Spantide I (SPT) suppresses T helper 17 (Th17) cell response. At day 14, frequencies of IL-17–producing CD4⁺ T cells (Th17 cells) in draining lymph nodes (**A** and **B**) and conjunctivae (**C** and **D**) were analyzed by using flow cytometry. Phosphate-buffered saline (PBS) (i.p.) indicates intraperitoneal injection of PBS; SPT (i.p.), intraperitoneal injection of Spantide I (36 μ g per mouse daily). Representative flow cytometry dot plots are shown on the left (**A** and **C**), and bar graph summaries are shown on the right (**B** and **D**). Data are expressed as means \pm SEM of three independent experiments. $n = 4$ draining lymph nodes and conjunctiva, with four tissues pooled together as one sample (**B** and **D**). * $P < 0.05$, ** $P < 0.01$.

mice.¹² In addition, treatment of DED mice with Spantide I was efficacious in preserving the function of Tregs in suppressing effector T-cell proliferation (percent suppression, $50.0 \pm 3.2\%$ versus $34.1 \pm 3.7\%$; $P = 0.027$) (Figure 4C). The protein levels of TGF- β and IL-10 in DLNs were also evaluated. The levels of TGF- β and IL-10 were significantly higher in Spantide I–treated DED mice ($P < 0.001$ for both cytokines) compared with the PBS-treated group (Figure 4D).

NK-1R Antagonist Suppresses Th17 Response in DED

The frequencies of Th17 cells, the principal effector T-cell population implicated in the pathogenesis of DED, were analyzed.¹² Systemic treatment with Spantide I significantly reduced Th17 frequencies in DLNs compared with the PBS-treated group ($0.048 \pm 0.008\%$ versus $0.088 \pm 0.005\%$; $P = 0.005$) (Figure 5, A and B). Similar results were observed in the conjunctiva, with the control-treated group having significantly higher Th17 cell frequencies ($6.7 \pm 1.1\%$) compared with the Spantide I–treated mice ($3.1 \pm 0.6\%$; $P = 0.039$) (Figure 5, C and D).

Discussion

It is known that the adoptive transfer of CD4⁺ T cells from DED to normal mice induces severe inflammation at the ocular surface, and Treg suppressive function in DED is impaired even though there is no significant change in their frequencies.^{12,36} These data suggest that the immune homeostasis breakdown occurs as a consequence of impaired Treg function, leading to unchecked effector CD4⁺ T-cell function in DED. The novelty of the current study lies in showing the increased expression of SP in the lymphoid tissues in DED, which directly impairs Treg suppressive function. Furthermore, blockade of SP signaling prevents SP-induced Treg dysfunction both *in vitro* and *in vivo* and significantly suppresses the severity of DED.

Changes in expression of SP in human tears have been reported in various conditions, such as allergic conjunctivitis and corneal hypoesthesia, and following laser-assisted *in situ* keratomileusis.^{37–39} Evidence suggests that SP plays a critical role in augmenting inflammatory responses principally by high-affinity binding to the NK-1R present on dendritic cells and macrophages.⁴⁰ In addition, Marriott

et al²¹ reported that SP and various other pro-inflammatory cytokines increase NK-1R⁺ macrophages. Blockade of NK-1R leads to decreases in the influx of neutrophils and CD4⁺ T cells in herpetic stromal keratitis, suppressing the expression of interferon- γ , IL-18, tumor necrosis factor- α , and IL-6 in corneal infection, and enhancing IL-10.^{41–43} Recently, it was reported that topical blockade of NK-1R suppresses maturation and activation of corneal CD11b⁺ cells in DED.²⁶ In aggregate, all of the aforementioned studies suggest an intimate link between SP and innate immune amplification.

SP is expressed by various cell types, regulating innate immune cell migration, activation of neutrophils, and promoting increases of IL-1 β , IL-6, tumor necrosis factor- α , and IL-23 levels by monocytes.^{19,44,45} However, very little is known about the direct effect of SP on Treg phenotype and function. Herein, we investigated the phenotypic and functional changes of Tregs in DLNs in a validated murine model of DED and correlated these findings to SP expression levels and Treg functions. This study found that SP impairs Treg function by reducing the expression of Foxp3 and other inhibitory membrane-bound molecules such as CTLA-4 and PD-1, as well as secreted cytokines, TGF- β , and IL-10. *In vitro*, antagonizing NK-1R prevents SP-induced Treg dysfunction as well as recovered TGF- β levels in the culture, although the SP-induced decrease of IL-10 secretion by Tregs is not prevented by blocking NK-1R. In the *in vitro* cultures, TGF- β may play a dominant role for Treg functions, as evidenced by its 30-fold higher levels compared with IL-10 in the supernatant. Interestingly, antagonizing NK-1R *in vivo* prevents Treg dysfunction by restoring both TGF- β and IL-10 expressions in DED mice, which is consistent with previous reports.^{42,43} A possible explanation for this differential effect of NK-1R antagonism on IL-10 expression between *in vitro* and *in vivo* may be a consequence of the effect of NK-1R blockade on other immune cells producing IL-10, such as dendritic cells, T helper 1 and 2 cells, natural killer cells, and macrophages.⁴⁶

SP is known to promote IL-17 secretion by human memory-phenotype T cells.⁴⁵ Our results show that treatment with an NK-1R antagonist suppresses Th17 responses on the ocular surface, as well as in the lymphoid tissue in DED, and consequently reduces disease severity. This scenario suggests that SP/NK-1R antagonism may be a viable therapeutic strategy for suppressing DED severity by rescuing Treg dysfunction and suppressing the Th17 response, which plays an essential function in DED pathogenesis.^{12,47}

Conclusions

The current findings provide new insights into the effects of SP on the phenotype and function of Tregs. We show for the first time that SP directly induces Treg dysfunction, and blockade of SP-NK-1R signaling effectively abolishes such an effect. Using the DED model, the benefits of NK-1R blockade in improving Treg functions and reducing corneal

epitheliopathy are further validated, indicating a broad translational potential of blocking SP-NK-1R signaling to treat autoimmune diseases.

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Author Contributions

Y.T., S.K.C., Y.C., and R.D. designed the study; Y.T., A.M., T.H.D., and Y.C. acquired data; Y.T. and Y.C. analyzed the data; Y.T. prepared the manuscript; A.A., R.B.S., Y.C., and R.D. revised the manuscript; and S.K.C., Y.C., and R.D. supervised the study; all authors read and approved the final manuscript.

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