



Universiteit  
Leiden  
The Netherlands

## **Cross-reactivity of anti-modified protein antibodies is also present in predisease and individuals without rheumatoid arthritis**

Reijm, S.; Brehler, A.S.; Rantapaa-Dahlqvist, S.; Kawakami, A.; Maeda, T.; Kawashiri, S.Y.; ... ; Toes, R.E.M.

### **Citation**

Reijm, S., Brehler, A. S., Rantapaa-Dahlqvist, S., Kawakami, A., Maeda, T., Kawashiri, S. Y., ... Toes, R. E. M. (2022). Cross-reactivity of anti-modified protein antibodies is also present in predisease and individuals without rheumatoid arthritis. *Annals Of The Rheumatic Diseases*, 81(9), 1332-1334. doi:10.1136/annrheumdis-2022-222326

Version: Publisher's Version

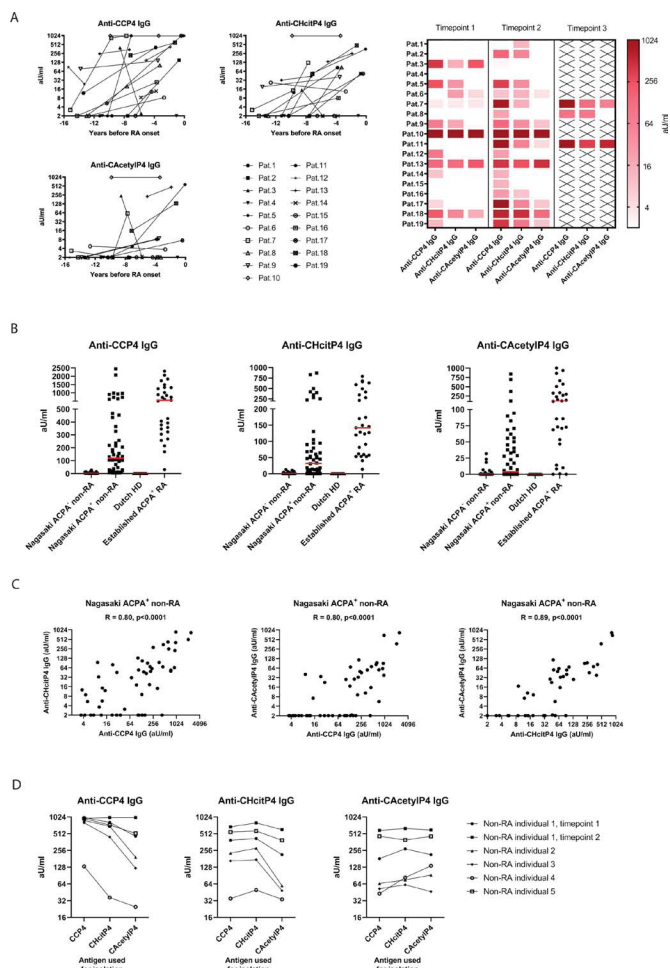
License: [Licensed under Article 25fa Copyright Act/Law \(Amendment Taverne\)](#)

Downloaded from: <https://hdl.handle.net/1887/3513117>

**Note:** To cite this publication please use the final published version (if applicable).

## Cross-reactivity of anti-modified protein antibodies is also present in predisease and individuals without rheumatoid arthritis

The presence of anti-citrullinated protein antibodies (ACPAs), anti-carbamylated protein antibodies (anti-CarPAs) and anti-acetylated protein antibodies (AAPAs) is a hallmark of rheumatoid arthritis (RA). ACPA and anti-CarPA can already be detected years before RA onset.<sup>1</sup> Moreover, it has been shown that the citrullinated epitope recognition profile of ACPA expands before RA develops. Recently, it has become clear that ACPA can display cross-reactivity to other post-translational modifications (PTMs), more specifically homocitrulline and



**Figure 1** (A) ACPA, anti-CarP and AAPA IgG levels, using the CCP4, CHcitP4 and CAcetylP4 peptides as antigen, in arbitrary units per ml (aU/mL) over time of 19 patients with RA before disease onset. Left graphs show the data in years before onset. The heatmap on the right shows a summary of the AMPA IgG levels per time point. (B) ACPA, anti-CarP and AAPA IgG levels in aU/ml of Japanese ACPA-non-RA samples (n=197), Japanese ACPA+non-RA samples (n=54), Dutch healthy donors (n=30) and established patients with RA (n=29). (C) Correlations of ACPA, anti-CarP and AAPA levels in aU/ml in Japanese ACPA+non-RA samples. R=correlation coefficient. (D) ACPA, anti-CarP and AAPA IgG levels in aU/mL of six samples from Japanese ACPA+non-RA samples after antibody isolation using CCP4, CHcitP4 or CAcetylP4 peptides. ACPA, anticitrullinated protein antibody; AAPA, antiacetylated protein antibody; anti-CarPA, anti-carbamylated protein antibodies; RA, rheumatoid arthritis.

acetyllysine, as shown at both the monoclonal and polyclonal antibody level.<sup>2,3</sup> B cell receptor analysis of ACPA-expressing B cells from patients with RA has shown that ACPAs have undergone extensive somatic hypermutation and that this can facilitate epitope spreading to multiple citrullinated epitopes.<sup>4</sup> Given the association of ACPA epitope spreading with progression to disease, it is relevant to obtain more insights when cross-reactivity to other PTMs is introduced. Furthermore, insights in whether cross-reactivity is also present in ACPA-positive subjects without RA or confined to subjects that will—or have developed RA will also help to better understand the evolution of anti-modified protein antibody (AMPA) responses. Therefore, we analysed cross-reactivity of the ACPA response in pre-disease samples and ACPA-positive individuals

without RA. To this end, ACPA, anti-CarPA and AAPA in different cohorts were measured using modified peptides as described in online supplemental materials. First, we analysed the AMPA-IgG response in samples from 19 different Swedish subjects who later developed RA. As expected, ACPA could be detected years before disease onset with a rise in antibody level over time (figure 1A). We detected a similar pattern for anti-CarPA and AAPA. Interestingly, for most patients with detectable ACPA, anti-CarPA and/or AAPA, these antibodies could be detected at the same timepoint, indicating their simultaneous appearance years before disease onset. Next, we analysed AMPA levels in samples from ACPA-positive and ACPA-negative Japanese individuals without RA, derived from the community-based Nagasaki Island study (figure 1B, online supplemental figure S1).<sup>5</sup> Intriguingly, a strong correlation between levels of the different individual AMPA-reactivities was observed, pointing to cross-reactivity of the antibodies (figure 1C). To experimentally confirm cross-reactivity, we selected six samples from ACPA-positive non-RA individuals with high AMPA values, isolated ACPA, anti-CarPA and AAPA and determined the reactivity of the isolated antibodies to the three different PTMs. Isolated ACPAs were highly reactive to the homocitrullinated and acetylated antigen and vice versa, showing that AMPA in individuals without RA are also cross-reactive towards different PTMs (figure 1D). These results were confirmed on post-translationally modified fibrinogen and FCS (online supplemental figure S2). Interestingly, the reactivity to citrullinated/homocitrullinated peptides was higher when AMPA were isolated with a citrullinated or homocitrullinated antigen than with an acetylated antigen. This suggests cross-reactivity between ACPA and anti-CarPA is stronger than between either of them and AAPA. Together, our data show that ACPA, anti-CarPA and AAPA already coexist before disease onset. Moreover, ACPA can be cross-reactive towards homocitrulline and acetyllysine in ACPA-positive individuals without RA. These results indicate that cross-reactivity towards different PTMs emerges when AMPA responses become detectable and provide evidence that cross-reactivity towards different PTMs is an intrinsic characteristic of AMPA responses. This finding is in line with the observation that (germline) ACPA-IgM can be cross-reactive towards other PTMs as well.<sup>6</sup> Although cross-reactivity seems to be an intrinsic feature of AMPA, it is tempting to speculate that the most cross-reactive B cells are selected during progression towards RA, explaining the increase of the ACPA epitope recognition profile in time towards disease onset. Although cross-reactivity is already present before disease onset, the further increase in AMPA cross-reactivity and level could be a valuable biomarker in predicting transition towards disease.

Sanne Reijm<sup>1</sup>, Astrid S Brehler,<sup>1</sup> Solbritt Rantapää-Dahlqvist<sup>2</sup>, Atsushi Kawakami,<sup>3</sup> Takahiro Maeda,<sup>4</sup> Shin-ya Kawashiri,<sup>5</sup> Mami Tamai,<sup>5</sup> Diane van der Woude,<sup>1</sup> René E M Toes<sup>1</sup>

<sup>1</sup>Department of Rheumatology, Leiden University Medical Center, Leiden, The Netherlands

<sup>2</sup>Department of Public Health and Clinical Medicine/Rheumatology, Umeå Universitet, Umeå, Sweden

<sup>3</sup>Department of Immunology and Rheumatology, Nagasaki University Graduate School of Biomedical Sciences, Nagasaki, Japan

<sup>4</sup>Department of Community Medicine, Nagasaki University Graduate School of Biomedical Sciences, Nagasaki, Japan

<sup>5</sup>Department of Immunology and Rheumatology Graduate School of Biomedical Sciences, Nagasaki University, Nagasaki, Japan

**Correspondence to** Sanne Reijm, Department of Rheumatology, Leiden University Medical Center, Leiden 2333 ZA, The Netherlands; s.reijm@lumc.nl

**Handling editor** Josef S Smolen

**Acknowledgements** We thank the LUMC Biobank for collecting the healthy donor samples and the Biobank Research Unit at Umeå University for providing the samples. We thank Jan Wouter Drijfhout for the synthesis of the peptides.

**Contributors** SR: concept design, acquisition, analysis and interpretation of data, drafting the manuscript. ASB: acquisition of data. SRD: providing samples and data analysis. MT, TM, S-yK and AK: providing samples. DvdW: interpretation of data. REMT: concept design and interpretation of data. All authors critically revised the manuscript and approved the final version to be published.

**Funding** The project leading to this application has received funding from the European Research Council (ERC) under the European Union's Horizon 2020 research and innovation programme (grant agreement No 884796), Target2B1 (LSHM18055-SGF), Reuma Nederland (NR 17-1-402) and from the Swedish Research Council (Dnr:2018-02551).

**Competing interests** None declared.

**Patient and public involvement** Patients and/or the public were not involved in the design, or conduct, or reporting, or dissemination plans of this research.

**Patient consent for publication** Not applicable.

**Ethics approval** All participants have given their written informed consent and the Regional Ethical Review Board Committees approved the studies.

**Provenance and peer review** Not commissioned; externally peer reviewed.

**Supplemental material** This content has been supplied by the author(s). It has not been vetted by BMJ Publishing Group Limited (BMJ) and may not have been peer-reviewed. Any opinions or recommendations discussed are solely those of the author(s) and are not endorsed by BMJ. BMJ disclaims all liability and responsibility arising from any reliance placed on the content. Where the content includes any translated material, BMJ does not warrant the accuracy and reliability of the translations (including but not limited to local regulations, clinical guidelines, terminology, drug names and drug dosages), and is not responsible for any error and/or omissions arising from translation and adaptation or otherwise.

© Author(s) (or their employer(s)) 2022. No commercial re-use. See rights and permissions. Published by BMJ.

► Additional supplemental material is published online only. To view, please visit the journal online (<http://dx.doi.org/10.1136/annrheumdis-2022-222326>).



**To cite** Reijm S, Brehler AS, Rantapää-Dahlqvist S, *et al.* *Ann Rheum Dis* 2022;**81**:1332–1334.

Received 9 February 2022

Accepted 12 April 2022

Published Online First 22 April 2022

*Ann Rheum Dis* 2022;**81**:1332–1334. doi:10.1136/annrheumdis-2022-222326

#### ORCID iDs

Sanne Reijm <http://orcid.org/0000-0002-4424-4256>

Solbritt Rantapää-Dahlqvist <http://orcid.org/0000-0001-8259-3863>

René E M Toes <http://orcid.org/0000-0002-9618-6414>

#### REFERENCES

- Shi J, van de Stadt LA, Levarht EWN, *et al.* Anti-carbamylated protein (anti-CarP) antibodies precede the onset of rheumatoid arthritis. *Ann Rheum Dis* 2014;**73**:780–3.
- Kissel T, Reijm S, Slot LM, *et al.* Antibodies and B cells recognising citrullinated proteins display a broad cross-reactivity towards other post-translational modifications. *Ann Rheum Dis* 2020;**79**:472–80.
- Sahlström P, Hansson M, Steen J, *et al.* Different hierarchies of Anti-Modified protein autoantibody reactivities in rheumatoid arthritis. *Arthritis Rheumatol* 2020;**72**:1643–57.
- Kongpachith S, Lingampalli N, Ju C-H, *et al.* Affinity maturation of the Anti-Citrullinated protein antibody paratope drives epitope spreading and polyreactivity in rheumatoid arthritis. *Arthritis Rheumatol* 2019;**71**:507–17.
- Kawashiri S-Y, Tsuji Y, Tamai M, *et al.* Effects of cigarette smoking and human T-cell leukaemia virus type 1 infection on anti-citrullinated peptide antibody production in Japanese community-dwelling adults: the Nagasaki islands study. *Scand J Rheumatol* 2021;**50**:295–8.
- Reijm S, Kissel T, Stoeken-Rijsbergen G, *et al.* Cross-Reactivity of IgM anti-modified protein antibodies in rheumatoid arthritis despite limited mutational load. *Arthritis Res Ther* 2021;**23**:230.