



Universiteit
Leiden
The Netherlands

Oxygenation of anandamide by lipoxygenases

Zadelhoff, G. van; Stelt, M. van der; Maccarrone, M.

Citation

Zadelhoff, G. van, & Stelt, M. van der. (2022). Oxygenation of anandamide by lipoxygenases. In M. Maccarrone (Ed.), *Methods in Molecular Biology* (pp. 307-316). New York: Humana. doi:10.1007/978-1-0716-2728-0_26

Version: Publisher's Version

License: [Licensed under Article 25fa Copyright Act/Law \(Amendment Taverne\)](#)

Downloaded from: <https://hdl.handle.net/1887/3571937>

Note: To cite this publication please use the final published version (if applicable).



Oxygenation of Anandamide by Lipoxygenases

Guus van Zadelhoff and Mario van der Stelt

Abstract

The endocannabinoids anandamide and 2-arachidonoylglycerol are not only metabolized by serine hydrolases, such as fatty acid amide hydrolase, monoacylglycerol lipase, and α,β -hydrolases 6 and 12, but they also serve as substrates for cyclooxygenases, cytochrome P450s, and lipoxygenases. These enzymes oxygenate the 1Z,4Z-pentadiene system of the arachidonic acid backbone of endocannabinoids, thereby giving rise to an entirely new array of bioactive lipids. Hereby, a protocol is provided for the enzymatic synthesis, purification, and characterization of various oxygenated metabolites of anandamide generated by lipoxygenases, which enables the biological study and detection of these metabolites.

Key words Lipoxygenase, Endocannabinoid, Anandamide, 2-Arachidonoylglycerol, Oxygenated metabolite

1 Introduction

The endocannabinoids anandamide (AEA, (5Z,8Z,11Z,14Z)-*N*-(2-hydroxyethyl)icosa-5,8,11,14-tetraenamide) and 2-arachidonoylglycerol (2-AG, 1,3-dihydroxypropan-2-yl (5Z,8Z,11Z,14Z)-icosa-5,8,11,14-tetraenoate) are signaling lipids that act on type 1 and type 2 (CB₁ and CB₂) cannabinoid receptors [1, 2]. They modulate a wide array of physiological functions. For example, in the brain they act as retrograde messengers that inhibit neurotransmitter release, whereas at the periphery they influence immune cell migration [3, 4]. The biological effect of endocannabinoids at CB₁ and CB₂ receptors depends on their life span in the extracellular space. Both endocannabinoids serve as substrates for metabolic serine hydrolases that hydrolyze the amide bond of AEA or the ester bond of 2-AG, thereby releasing arachidonic acid and ethanolamine or glycerol, respectively. Fatty acid amide hydrolase is the main enzyme responsible for anandamide hydrolysis, whereas 2-AG is converted by monoacylglycerol lipase, α,β -hydrolase 6 and 12 [5]. Hydrolysis of the amide or ester bond results in termination of CB₁ and CB₂ receptor signaling [6].

Both AEA and 2-AG contain an arachidonic acid backbone, which may serve as a substrate for enzymes such as cyclooxygenases and lipoxygenases [7–11]. Indeed, both enzyme classes convert endocannabinoids into oxygenated metabolites, such as prostamides, prostaglandin-glycerol esters, and hydro(pero)xy-derivatives. These oxygenated products do not necessarily serve to terminate endocannabinoid signaling but may exert biological actions of their own and/or modulate the pharmacology of endocannabinoids by interfering with fatty acid amide hydrolase activity. The action of cyclooxygenases on endocannabinoids has been studied in detail, and specific receptors have been found for the prostamides, whereas the characterization of the biological presence and action of the lipoxygenase products is lacking behind [12].

There are three types of lipoxygenases (5-, 12-, and 15-lipoxygenases) that insert O_2 at C5, C12, or C15 of arachidonic acid, respectively. The same enzymes oxygenate in a regio- and stereospecific manner also AEA and 2-AG, adding O_2 at different positions of their arachidonic acid backbone (*see* Fig. 1). The resulting hydroperoxide products are rapidly and non-enzymatically reduced in the cell to their corresponding hydroxyl derivatives.

12(*S*)-Hydroxy-AEA has been shown to bind to the CB_1 receptor with comparable affinity as AEA, whereas the other metabolites showed reduced binding affinity [13]. Of note, all lipoxygenase products were found to be inhibitors of fatty acid amide hydrolase in the low to submicromolar range. In various physiological processes, the action of lipoxygenase metabolites of endocannabinoids has been suggested based on intervention studies with lipoxygenase inhibitors [14]. To interrogate the physiological relevance and function of lipoxygenase metabolites of endocannabinoids in more detail, it is important to have sufficient quantities of the oxygenated products. Here, we provide a protocol for the enzymatic synthesis, purification, and analysis of 5(*S*)-hydroxy-AEA, 11(*S*)-hydroxy-AEA, 12(*S*)-hydroxy-AEA, 15(*S*)-hydroxy-AEA, 5(*S*),15(*S*)-dihydroxy-AEA, and 8(*S*),15(*S*)-dihydroxy-AEA (*see* Note 1) [13–15].

2 Materials

2.1 Buffers and Solutions

- 0.1 M sodium borate buffer (pH 9.0): Weigh 6.18 g of boric acid and transfer to a 1-l conical flask. Add 900 mL of water and dissolve boric acid under magnetic stirring. Adjust the pH to 9.0 with 1 M NaOH and bring the volume to 1 L.
- 0.1 M sodium phosphate buffer (pH 7.0): Weigh 15.60 g of sodium phosphate monobasic dihydrate and transfer to a 1-l conical flask. Add 900 mL of water and dissolve sodium phosphate under magnetic stirring. Adjust the pH to 7.0 with 1 M NaOH and bring the volume to 1 L.

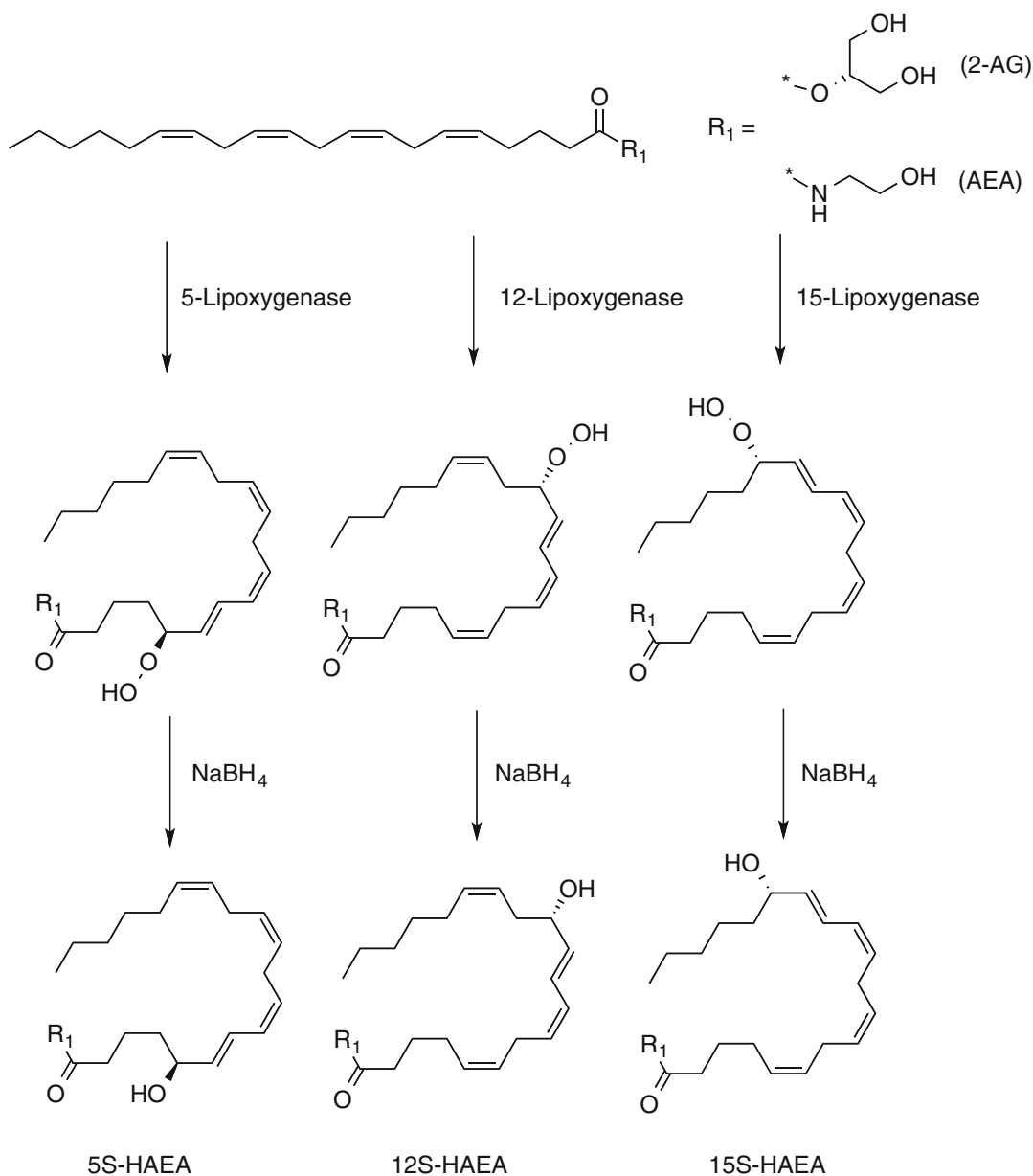


Fig. 1 Structures of oxygenated metabolites of anandamide produced by distinct lipoxygenases

- PBS (pH 7.4): Weigh 8 g of sodium chloride, 0.2 g of potassium chloride, 1.80 g of sodium phosphate dibasic dehydrate, and 0.24 g of potassium phosphate monobasic anhydrous and transfer to a 1-L conical flask. Add 900 mL of water and dissolve the salts under magnetic stirring. Adjust the pH to 7.0 with 0.1 M HCl and bring the volume to 1 l.
- Hypertonic PBS: Dissolve 2.7 g of NaCl in 100 mL of PBS.

- 0.05 M sodium acetate buffer (pH 5.5), containing 134 g/L ammonium sulfate: Add 2.90 mL of glacial acetic acid to 700 mL of H₂O, and bring to pH 5.5 with 3 M NaOH. Add 134 g of ammonium sulfate, dissolve, and bring the volume to 1 L with H₂O.
- 0.15 M sodium citrate: Dissolve 22 g of sodium citrate tribasic dehydrate in 500 mL of water.
- 0.15 M potassium phosphate (pH 6.0), containing 2 M ammonium sulfate: Dissolve 2 g of potassium dihydrogen phosphate in 50 mL of water, and bring pH to 6.0 with 3 M KOH. Add 26.8 g of ammonium sulfate, dissolve, and bring volume to 100 mL with H₂O.
- 100 mM AEA: Weigh 35 mg of AEA and dissolve in 1 mL of methanol in an autosampler vial.
- Tetrahydrofuran/methanol/water/acetic acid mixture (25/30/45/0.1, vol/vol/vol/vol): Mix 250 mL of tetrahydrofuran, 300 mL of methanol, 450 mL of water, and 1 mL of glacial acetic acid. Degas the HPLC solvent just before use by purging helium through the solution for 15 min.
- Methanol/water/acetic acid (75/25/0.1, vol/vol/vol): Mix 750 mL of methanol, 250 mL of water, and 1 mL of glacial acetic acid. Degas the HPLC solvent just before use by purging helium through the solution for 15 min.
- Silylation reagent solution (pyridine/1,1,1,3,3,3-hexamethyldisilazane/trimethylchlorosilane, 5/1/1, vol/vol/vol): Mix 500 μ L of anhydrous 99.8% pyridine, 100 μ L of 99.9% 1,1,1,3,3,3-hexamethyldisilazane, and 100 μ L of 99.0% trimethylchlorosilane just before use in a 1-mL glass vial.

2.2 Preparation of Enzymes

Soybean lipoxygenase-1 (soyLOX, from Maple Glen cultivar) is a 15-lipoxygenase that can be obtained from Sigma-Aldrich. The protein concentration is estimated from the absorbance at 280 nm ($\epsilon_{280} = 1.6 \times 10^5 \text{ L}\cdot\text{mol}^{-1}\cdot\text{cm}^{-1}$). The enzyme is stored at 4 °C at a concentration of 1.9 mg/mL (corresponding to 20 mM) in 0.05 M sodium acetate buffer (pH 5.5) containing 134 g/L ammonium sulfate. 5-Lipoxygenase, mostly isolated from potatoes, can be obtained from Cayman Chemicals. The enzyme is stored at 4 °C at a concentration of 1 mg/mL in 0.15 M potassium phosphate (pH 6.0) containing 2 M ammonium sulfate. Bovine lipoxygenase (bovLOX) is a 12-lipoxygenase isolated from bovine leukocytes, performing the entire purification at room temperature as follows. Bovine blood is collected in a vessel containing 0.15 M sodium (100 mL/L) as an anticoagulant. The anticoagulated blood is centrifugated at 470 g for 20 min in a swing-out rotor to separate blood cells from plasma: the lower

part of the tube contains the red and white blood cells, and the upper part contains the platelet-rich plasma that is discarded. The leukocyte fraction is separated from erythrocytes by lysis. To this end, 2 volumes of distilled water are added to the cell pellet and, after gently shaking for 30 s, 1 volume of hypertonic PBS is also added to make the final suspension hypotonic. The mixture is centrifuged for 10 min at 470 g, and the supernatant containing the lysed erythrocytes is removed. The pellet is resuspended in 10 mL of PBS, and the lysis procedure is repeated once more. The erythrocyte-free leukocyte pellet is suspended in 15 mL of PBS, carefully layered on 14 mL of Ficoll-Paque Plus (GE healthcare) in a 50-mL tube, and centrifuged at 400 g for 40 min in a swing-out rotor. Leukocytes are found at the bottom of the tube, whereas the lymphocytes and monocytes are found on top of the Ficoll-Paque plus. Leukocytes are washed with PBS and centrifuged at 2000 g, and the final cell pellet is resuspended in 10 mL of PBS.

3 Methods

3.1 Production of Hydroperoxy-AEA

1. Incubate for 45 min at room temperature 1 U enzyme (soyLOX, barLOX, or bovLOX) with 5 μ mol AEA (50 μ L from a 100 mM stock) in 50 mL of rigorously stirred, air-saturated appropriate buffer: 0.1 M sodium borate buffer, pH 9.0 (for soyLOX), 0.1 M sodium phosphate buffer, pH 7.0 (for barLOX), or PBS, pH 7.4 (for bovLOX) (*see* **Notes 2–4**).
2. Stop reactions by acidifying the reaction mixtures to pH 4.0 with 3 M HCl.
3. Extract metabolites using 60-mg OASIS HLB solid-phase extraction columns (or equivalent, like Bakerbond C18).
Condition cartridge with 5 mL of methanol and 5 mL of H₂O, respectively. Do not allow the cartridge to run dry. Load reaction mixture into a 100cc syringe connected to the SPE column via an SPE tube adaptor. Positive pressure is achieved by the plunger of the 100cc syringe. Wash the column with 5 mL of H₂O until dryness, and elute the concentrated and purified reaction products with 2.5 mL of methanol.
4. Azeotropically evaporate residual water by co-evaporation with methanol under a gentle stream of nitrogen gas.
5. Dissolve products in 200 μ L of methanol, and store at -25°C until use.

3.2 Reduction of Hydroperoxy-AEA to Hydroxy-AEA

1. Reduce lipoxygenase products with a molar excess of NaBH₄ in 3 mL of methanol.
2. Stir the reaction mixture for 30 min at 0°C under a N₂ atmosphere.

3. Terminate the reaction by adding 15 mL of water and acidify the mixture to pH 4.0 with 3 M HCl. Stir until gas production ends.
4. Extract and concentrate the reduced products with an SPE column, as described above.
5. Dissolve the metabolites in 100 μ L of methanol.

3.3 Purification of Hydroxy-AEA

- Purify the lipoxygenase metabolites with preparative HPLC on a Cosmosil 5C18-ARII (5 μ m, 250 $\times$ 10 mm i.d., or equivalent) column, using a tetrahydrofuran/methanol/water/acetic acid mixture (25/30/45/0.1, vol/vol/vol/vol) as the eluent, at a flow rate of 3 mL/min.
 1. Dilute the collected fractions with a 10-fold excess of water.
 2. Concentrate the diluted fractions with an SPE column, as described above.
 3. Dissolve the purified compounds in 200 μ L of methanol.
 4. Store compounds under N₂ at -25°C .
 5. Chiral separations of the purified sodium borohydride-reduced AEA metabolites can be carried out on a Chiralcel OD-R column (5 μ m, 250 $\times$ 4.6 mm, or equivalent) with methanol/water/acetic acid (75/25/0.1 vol/vol/vol) as the eluent, at a flow rate of 0.5 mL/min (*see Note 5*). Chirally pure AEA metabolites can be concentrated again with an SPE column as described above and are stored under N₂ at -25°C .

3.4 Analytical Characterization

1. Dry the hydroxyl-AEA derivatives under a stream of N₂ gas.
2. Dissolve them in CHCl₃ or CDCl₃ in order to perform FTIR- or ¹H-NMR measurements, respectively. Record IR spectra on an FTIR spectrometer from 4000 cm⁻¹ to 400 cm⁻¹ with a resolution of 4 cm⁻¹ and a co-addition of 60 scans in a N₂ atmosphere at 20 $^{\circ}\text{C}$. Correct the spectra with a co-addition of 60 blank scans and a manual baseline correction.
3. ¹H-NMR spectra are recorded in CDCl₃ with an AC 300 MHz or 500 MHz spectrometer at 27 $^{\circ}\text{C}$.
4. For GC-MS analysis of the hydroxyl position, dry aliquots of the purified hydroxy-AEA metabolites under a stream of N₂ gas.
 - 4.1 Redissolve in 1 mL of hexane and hydrogenate the double bonds with a catalytic amount (5%) of palladium on calcium carbonate under a gentle stream of H₂ gas.
 - 4.2 After 30 min, remove the catalyst by filtration over a pre-washed (hexane) piece of cotton wool using an empty Pasteur pipette.

- 4.3 Evaporate hexane under N₂.
- 4.4 Add 50 μ L of freshly prepared silylation reagent solution.
- 4.5 After 30 min at room temperature, evaporate the silylation reagent under a stream of N₂ and redissolve the residue in 10 μ L of hexane.
- 4.6 Analyze aliquots by GC/MS, equipped with a CP-Sil 5 CB-MS (or equivalent) column (25 m $\times$ 0.25 mm $\times$ 0.25 μ m). The column temperature is held at 200 °C for 1 min, increased in 13 min to 330 °C, and held at this temperature for 2 min. Mass spectra are recorded under electron impact with an ionization energy of 70 eV.
5. Record IR spectra on an FTIR spectrometer from 4000 cm⁻¹ to 400 cm⁻¹ with a resolution of 4 cm⁻¹ and a co-addition of 60 scans in a N₂ atmosphere at 20 °C. Correct the spectra with a co-addition of 60 blank scans and a manual baseline correction.
6. Record UV absorption spectra from 200 to 300 nm on 40 μ M hydroxyl-AEA in methanol with a diode array spectrophotometer (*see Note 6*).
7. Record CD-spectra (resolution of 1 nm and 20 scans, 10 nm/min) in a 1.0-cm cuvette at 20 °C on a CD-spectrophotometer from 210 to 270 nm, using metabolites at a concentration of 40 μ M in methanol.

3.5 Chemical Data of the Oxygenated Derivatives

5R/S-HAEA [5(*R/S*)-Hydroxy-eicosa-6*E*,8*Z*,11*Z*,14*Z*-tetraenoyl-*N*-(2-hydroxyethyl) amine] Produced by barLOX: ¹H NMR (CDCl₃): δ 6.55 (dd, J = 13.6, 10.6 Hz 1H), 6.02 (t J = 10.5 Hz 1H), 5.71 (dd, J = 13.6 Hz, 1H), 5.39 (m, 5H), 4.16 (m, 1H), 3.72 (t, 2H), 3.42 (q, 2H), 2.97 (m, 2H), 2.81 (m, 2H), 2.22 (t, J = 7.8 Hz, 2H), 2.11 (m, 2H), 1.72 (q, J = 6.2 Hz 2H), 1.34 (m, 6H), 0.89 (t, J = 6.2 Hz, 3H). NaBH₄ and H₂ reduced, hydrogenated trimethyl silyl ether GC/MS m/z 515 [M⁺], 500 [M⁺-CH₃], 313 [C₁₆H₃₂OTMS⁺], 304 [M⁺-C₁₅H₃₁], 214 [304-TMSOH], 116 [C₂H₅OTMS⁺], 73 [TMS⁺].

11S-HAEA [11(*S*)-Hydroxy-eicosa-5*Z*,8*Z*,12*E*,14*Z*-tetraenoyl-*N*-(2-hydroxyethyl)amine] Produced by barLOX: ¹H NMR (CDCl₃): δ 6.52 (dd, J = 11.0; 10.1 Hz 1H), 5.98 (t, J = 11.0 Hz 1H), 5.69 (dd, J = 15.1; 6.5 Hz 1H), 5.40 (m, 5H), 4.23 (q, 2H), 3.72 (t, J = 4.6 Hz, 2H), 3.42 (q J = 5.5 Hz; 2H), 2.81 (m, 2H), 2.32 (m, 2H), 2.22 (t, J = 7.4 Hz; 2H), 2.12 (m, 4H), 1.73 (q, 2H), 1.25–1.38 (m, 6H), 0.88 (t, J = 6.9 Hz 3H). NaBH₄ and H₂ reduced, hydrogenated trimethyl silyl ether GC/MS m/z 515 [M⁺], 500 [M⁺-CH₃], 388 [M⁺-C₉H₁₉], 229 [C₁₀H₂₀OTMS⁺], 116 [C₂H₅OTMS⁺], 73 [TMS⁺].

12S-HAEA [12(*S*)-Hydroxy-eicosa-5*Z*,8*Z*,10*E*,14*Z*-tetraenoyl-*N*-(2-hydroxyethyl)amine] Produced by bovLOX: ^1H NMR (CDCl_3): δ 6.58 (dd, $J = 14.2$ Hz 1H), 5.99 (t, $J = 10.5$ Hz 1H), 5.74 (dd, $J = 6.2$ Hz 1H), 5.40 (m, 5H), 4.25 (q, 1H), 3.72 (t, $J = 4.6$ Hz 2H), 3.42 (q, $J = 4.6$ Hz 2H), 2.95 (m, 2H), 2.33 (m, 2H), 2.21 (t, $J = 7.5$ Hz 2H), 2.10 (m, 4H), 1.74 (q, $J = 7.3$ Hz, 2H), 1.28 (m, 6H), 0.89 (t, $J = 6.9$ Hz 3H). NaBH_4 and H_2 reduced, hydrogenated trimethyl silyl ether GC/MS m/z 515 [M^+], 500 [$\text{M}^+ - \text{CH}_3$], 402 [$\text{M}^+ - \text{C}_8\text{H}_{17}^+$], 215 [$\text{C}_9\text{H}_{18}\text{OTMS}^+$], 116 [$\text{C}_2\text{H}_3\text{OTMS}^+$], 73 [TMS^+].

15S-HAEA [15(*S*)-Hydroxy-eicosa-5*Z*,8*Z*,11*Z*,13*E*-tetraenoyl-*N*-(2-hydroxyethyl)amine] Produced by soyLOX: ^1H NMR (CDCl_3): δ 6.55 (dd, $J = 15.4; 12.2$ Hz 1H), 6.00 (t, $J = 10.7$ Hz, 1H), 5.72 (dd, $J = 7.2$ Hz, 1H), 5.40 (m, 5H), 4.12 (q, 1H), 3.72 (t, 2H), 3.42 (q, 2H), 2.97 (m, 2H), 2.82 (m, 2H), 2.22 (t, $J = 7.5$ Hz, 2H), 2.11 (m, 2H), 1.72 (q, $J = 7.3$ Hz, 2H), 1.56 (m, 2H), 1.31 (m, 6H), 0.89 (t, $J = 6.9$ Hz, 3H). NaBH_4 and H_2 reduced, hydrogenated trimethyl silyl ether GC/MS m/z 515 [M^+], 500 [$\text{M}^+ - \text{CH}_3$], 444 [$\text{M}^+ - \text{C}_5\text{H}_{11}$], 173 [$\text{C}_6\text{H}_{11}\text{OTMS}^+$], 116 [$\text{C}_2\text{H}_3\text{OTMS}^+$], 73 [TMS^+].

5,15-diHAEA [5,15-Dihydroxy-eicosa-6*E*,8*Z*,11*Z*,13*E*-tetraenoyl-*N*-(2-hydroxyethyl) amine] Produced by soyLOX: ^1H NMR (CDCl_3): δ 6.58 (m, 2H), 6.01 (m, 2H), 5.72 (m, 2H), 5.43 (m, 2H), 4.21 (m, 2H), 3.72 (t, $J = 4.6$ Hz, 2H), 3.42 (q, $J = 4.6$ Hz, 2H), 2.97 (m, 2H), 2.28 (t, $J = 6.9$ Hz, 2H), 1.76 (m, 2H), 1.57 (m, 4H), 1.30 (m, 6H), 0.89 (t, 3H). NaBH_4 and H_2 reduced, hydrogenated trimethyl silyl ether GC/MS m/z 603 [M^+], 588 [$\text{M}^+ - \text{CH}_3$], 532 [$\text{M}^+ - \text{C}_5\text{H}_{11}$], 304 [$\text{M}^+ - \text{C}_{15}\text{H}_{31}$], 214 [304-TMSOH], 116 [$\text{C}_2\text{H}_3\text{OTMS}^+$], 73 [TMS^+].

8,15-diHAEA [8,15-Dihydroxy-eicosa-5*Z*,9*E*,11*Z*,13*E*-tetraenoyl-*N*-(2-hydroxyethyl) amine] Produced by soyLOX: ^1H NMR (CDCl_3): δ 6.70 (m, 2H), 5.97 (m, 2H), 5.74 (m, 2H), 5.46 (m, 2H), 4.27 (m, 1H), 4.18 (m, 1H), 3.72 (t, $J = 4.6$ Hz, 2H), 3.42 (q, $J = 4.6$ Hz, 2H), 2.32 (m, 2H), 2.19 (t, $J = 6.9$ Hz, 2H), 2.09 (q, 2H), 1.70 (m, 2H), 1.50 (m, 2H), 1.32 (m, 6H), 0.88 (t, 3H). NaBH_4 and H_2 reduced, hydrogenated trimethyl silyl ether GC/MS m/z 603 [M^+], 588 [$\text{M}^+ - \text{CH}_3$], 532 [$\text{M}^+ - \text{C}_5\text{H}_{11}$], 346 [$\text{M}^+ - \text{C}_{12}\text{H}_{24}\text{OTMS}$], 173 [$\text{C}_6\text{H}_{11}\text{OTMS}^+$], 116 [$\text{C}_2\text{H}_3\text{OTMS}^+$], 73 [TMS^+].

4 Notes

1. This procedure is not suitable for the generation of lipoxygenase products of 2-AG because of the chemical instability of this endocannabinoid that rapidly isomerizes to 1-AG during the incubation.
2. All solutions where organic solvents or reagents are involved should be in glass. Small beaker glasses (20–100 mL) or auto-sampler vials (1–20 mL) with PTFE septa are recommended.
3. Use 5 U soyLOX to produce 8(*S*),15(*S*)-dihydroxy AEA and 5(*S*),15(*S*)-dihydroxy-AEA.
4. Incubate 2 U barLOX with 5 μmol AEA in 50 mL of 0.1 M phosphate buffer (pH 7.4) for 60–90 min at 20 °C to produce 5(*S*)- and 11(*S*)-hydroxy AEA.
5. Incubate 2 U bovLOX with 5 μmol AEA in 50 mL of 0.1 M phosphate buffer (pH 7.4) for 60–90 min at 20 °C to produce 12*S*-hydroxy-AEA. Skip **step 2** because the products are already reduced.
6. Alternatively, hydroxyl-AEAs can be separated with a 20-min linear gradient of methanol/water/acetic acid, from 60/40/0.1 (vol/vol/vol) to 95/5/0.1 (vol/vol/vol), at 1 mL/min on an analytical 5C18 ARII (5 μm , 250 $\times$ 4.6 mm) column, or equivalent.
7. A molar absorbance of 23,000 $\text{M}^{-1} \text{cm}^{-1}$ at 236 nm is used to quantify hydroxy-AEAs. For 8,15- and 5,15-dihydroxy-AEAs, molar absorbances of 40,000 $\text{M}^{-1} \text{cm}^{-1}$ at 269 nm and of 33,500 $\text{M}^{-1} \text{cm}^{-1}$ at 243 nm are used, respectively.

References

1. Chiurchiù V, van der Stelt M, Centonze D, Maccarrone M (2018) The endocannabinoid system and its therapeutic exploitation in multiple sclerosis: clues for other neuroinflammatory diseases. *Prog Neurobiol* 160:82–100
2. Baggeelaar MP, Maccarrone M, van der Stelt M (2018) 2-Arachidonoylglycerol: a signaling lipid with manifold actions in the brain. *Prog Lipid Res* 71:1–17
3. Di Marzo V (2011) Endocannabinoid signaling in the brain: biosynthetic mechanisms in the limelight. *Nat Neurosci* 14:9–15
4. Piomelli D (2014) More surprises lying ahead. The endocannabinoids keep us guessing. *Neuropharmacology* 76:228–234
5. Cravatt BF, Giang DK, Mayfield SP et al (1996) Molecular characterization of an enzyme that degrades neuromodulatory fatty-acid amides. *Nature* 384:83–87
6. van Egmond N, Straub VM, van der Stelt M (2021) Targeting endocannabinoid signaling: FAAH and MAG lipase inhibitors. *Annu Rev Pharmacol Toxicol.* 61:441–463
7. Hampson AJ, Hill WA, Zan-Phillips M et al (1995) Anandamide hydroxylation by brain lipoxygenase: metabolite structures and potencies at the cannabinoid receptor. *Biochim Biophys Acta* 1259:173–179
8. Ueda N, Yamamoto K, Kurahashi Y et al (1995) Oxygenation of arachidonylethanolamide (anandamide) by lipoxygenases. *Adv Prostaglandin Thromboxane Leukot Res* 23: 163–165

9. Edgemon WS, Hillard CJ, Falck JR et al (1998) Human platelets and polymorphonuclear leukocytes synthesize oxygenated derivatives of arachidonylethanolamide (anandamide): their affinities for cannabinoid receptors and pathways of inactivation. *Mol Pharmacol* 54:180–188
10. Yu M, Ives D, Ramesha CS (1997) Synthesis of prostaglandin E2 ethanolamide from anandamide by cyclooxygenase-2. *J Biol Chem* 272: 21181–21186
11. Kozak KR, Rowlinson SW, Marnett LJ (2000) Oxygenation of the endocannabinoid, 2-arachidonylglycerol, to glyceryl prostaglandins by cyclooxygenase-2. *J Biol Chem* 275: 33744–33749
12. Urquhart P, Nicolaou A, Woodward DF (2015) Endocannabinoids and their oxygenation by cyclo-oxygenases, lipoxygenases and other oxygenases. *Biochim Biophys Acta* 1851:366–376
13. Van der Stelt M, van Kuik JA, Bari M et al (2002) Oxygenated metabolites of anandamide and 2-arachidonoylglycerol: conformational analysis and interaction with cannabinoid receptors, membrane transporter, and fatty acid amide hydrolase. *J Med Chem* 45:3709–3720
14. Veldhuis W, van der Stelt M, Wadman MW et al (2003) Neuroprotection by the endogenous cannabinoid anandamide and arvanil against in vivo excitotoxicity in the rat: role of vanilloid receptors and lipoxygenases. *J Neurosci* 23: 4127–4133
15. Van Zadelhoff G, Veldink GA, Vliegthart JF (1998) With anandamide as substrate plant 5-lipoxygenases behave like 11-lipoxygenases. *Biochem Biophys Res Commun* 248:33–38