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MRI for planning and characterization of uveal melanoma patients treated with proton beam therapy

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Eye specific quantitative dynamic contrast enhanced MRI analysis for patients with intraocular masses

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Dynamic contrast enhanced (DCE)-MRI is currently not generally used for intraocular masses as lesions are small, have an inhomogeneous T_1 and the eye is prone to motion. The aim of this paper is to address these eye specific challenges, enabling accurate ocular DCE-MRI. DCE-MRI of 19 uveal melanoma (UM) patients was acquired using a fat-suppressed 3D spoiled gradient echo sequence with TWIST (time-resolved angiography with stochastic trajectories sequence). The analysis consisted of a two-step registration method to correct for both head and eye motion. A T_1 -map was calculated to convert signal intensities to concentrations. Subsequently the Tofts model was fitted voxel wise to obtain K^{trans} and v_e . Registration significantly improved the concentration curve quality ($p < 0.001$). The T_1 of melanotic lesions was significantly lower than amelanotic lesions (888 ms vs 1350 ms, $p = 0.03$). The average achieved B_1^+ in the lesions was 91%. The average K^{trans} was 0.46 min^{-1} (range: 0.13-1.0) and the average v_e was 0.22 (range: 0.10-0.51). Using this eye-specific analysis, DCE of intraocular masses is possible which might aid in the diagnosis, prognosis and follow-up of UM.

6.1 Introduction

Most intraocular lesions are benign, such as choroidal nevi, haemangiomas and leiomyomas, but also various malignant intraocular masses exist. Although uveal melanoma (UM) is relatively rare, with an incidence of 14 per million in the Netherlands², it is the most common primary intraocular tumour. It mostly originates from the choroid (90%), but can also originate from the iris or ciliary body^{3,4}. Other malignant ocular lesions include mainly metastases from other tumour sites or even more rare lesions such as retinoblastoma^{5,6}. As the prognosis and treatment of benign lesions, UM and other malignant ocular lesions differ, it is important to have an accurate diagnosis^{3,6}. For the differentiation between these different lesions the ophthalmologist primarily relies on fundoscopic, fluorescent angiography and ultrasound imaging (figure 6.1a-d)⁷. However, in some patients this differentiation is quite challenging, especially for amelanotic melanomas, or lesions behind the iris.

In the last decade, advances in ocular MRI, such as dedicated receive coils⁸ and dedicated acquisition strategies⁹ have resulted in different new clinical applications of MRI for ocular conditions¹⁰. MRI offers a superior evaluation of the extend of eye lid tumours¹¹, can be instrumental in the diagnosis and assessment of disease progression in orbital disease involving extra-ocular muscles¹², provide insight into ocular complaints such as negative dysphotopsia¹³ and allows for a more accurate assessment of tumour dimensions for radiotherapy therapy planning^{14,15}. Furthermore, diffusion weighted imaging is emerging as a promising early marker of therapy response after ocular proton beam therapy¹⁶, while quantitative dynamic contrast enhanced MRI (DCE-MRI) could assist in the differential diagnosis of intraocular masses and monitoring of treatment response of UM¹⁷. The use of DCE-MRI of ocular masses is often limited to the evaluation of the time intensity curve (TIC, figure 6.1f-h)¹⁷⁻²⁰. However, Wei et al.¹⁸ and Kamrava et al.²¹ have shown contradicting results on the relation between tumour permeability and metastatic risk. Kamrava et al.²¹ found a higher K^{trans} in UM patients with monosomy 3, a subset of UM patients who have a strong increased risk of developing metastatic disease²². Conversely Wei et al.¹⁸ showed a decreased K^{trans} (a lower peak signal intensity) in patients with metastatic disease. Although these papers are an improvement compared to the current clinical practice and other research where some eye-related challenges were not addressed such as small lesion size, eye motion and difference in melanin content.

The limited use of quantitative DCE-MRI might be due to the eye-specific challenges of MR-imaging in general. One of the main challenges of DCE-MRI of intraocular masses is the small size of the eye, containing even smaller lesions, generally with a thickness of less than 5 mm. Furthermore, the eye is prone to

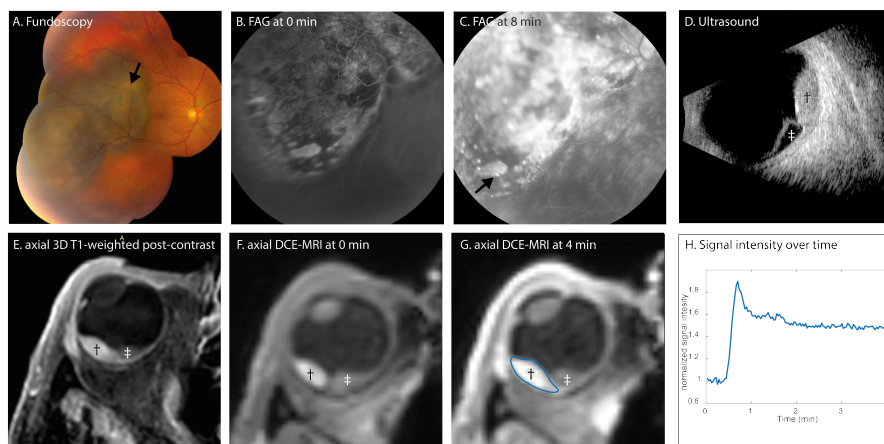


Figure 6.1: Conventional ophthalmic imaging and MRI of patient 13. (A–D) Conventional ophthalmic imaging of an UM. The fundus photo (A) shows a pigmented lesion with lipofuscin (arrow). The lesion is enhancing with pinpoints (C, arrow) on the fluorescein angiography (FAG, B, C). On ultrasound (US, D), the lesion (dagger) has an intermediate reflectivity, while the retinal detachment (double dagger) has a low reflectivity and the dimensions of the lesion are measured. On MRI, an enhancing lesion (dagger) with associated retinal detachment (double dagger) is visible (E–G). In contrast to the FAG, the change of the signal intensity after contrast administration can be visualized in DCE-MRI (H)

movement, which in combination with the small lesion size leads to a mismatch of the tumour location between timepoints¹⁹. Finally, intraocular lesions can be pigmented (large amount of melanin), unpigmented (no melanin) or partially pigmented²³. This varying degree of pigmentation results in a large variability in pre-contrast longitudinal relaxation time (T_1)²⁴, which directly affects the quantification of concentration of the, generally T_1 -based, contrast agent. However, recent improvements in ocular MR-imaging protocols such as the use of a surface coil for receiving the signal and implementation of time-resolved angiography with stochastic trajectories sequence²⁵ allow for the acquisition of DCE-MRI with sufficient temporal and spatial resolution to perform DCE-MRI¹⁹. The aim of this paper is to overcome eye specific challenges in the DCE-MRI analysis of intraocular masses.

6.2 Methods

6.2.1 Study population

Nineteen patients diagnosed with UM were included for this study. Nine patients were scanned as part of a prospective study and were recruited randomly. This study has been approved by the local ethics committee and subjects were scanned after written informed consent. The data from the remaining 10 patients were selected from UM patients with a tumour prominence >3 mm who received an MRI as part of clinical care. This retrospective inclusion of data was approved by the local ethics committee. The patients were on average 63 years old (range 30 to 82 years), 68% ($n=13$) were male and had a BMI of 26.6 ± 4.5 . The lesions had an average prominence of 7.8 mm and an average largest basal diameter of 14.5 mm on ultrasound. The American Joint Committee on Cancer staging²⁶ ranged from T1 to T4 with most patients in the T3 ($n = 8$) and T4 ($n = 6$) stage. Most were primary tumours (18/19) but in one case (UM15) the patient had a large reoccurrence. The primary tumour was treated with ruthenium plaque therapy and was located at the other side of the eye. A detailed description of the cohort of patients can be found in table 6.1.

| Patient no. | Tumour stage | Age at diagnosis (years) | BMI | Prominence on US (mm) | LBD on US (mm) |
|-------------|--------------|--------------------------|---------|-----------------------|----------------|
| UM 1 | T3a | 71 | 20 | 3.3 | 14.9 |
| UM 2 | T3a | 61 | 30 | 12.3 | 14.4 |
| UM 3 | T2c | 82 | Missing | 4 | 10.5 |
| UM 4 | T2b | 68 | 22 | 5 | 11 |
| UM 5 | T4a | 71 | 29 | 7.3 | 15.7 |
| UM 6 | T2a | 73 | 32 | 2.8 | 9.5 |
| UM 7 | T3b | 30 | 22 | 6.1 | 14.9 |
| UM 8 | T3a | 37 | 35 | 6 | 14.6 |
| UM 9 | T2a | 50 | 29 | 2.5 | 10 |
| UM 10 | T4a | 80 | 27 | 12.1 | 15.5 |
| UM 11 | T1c | 73 | 28 | 5.8 | 6.6 |
| UM 12 | T4a | 62 | 19 | 11 | 22.6 |
| UM 13 | T4a | 59 | 31 | 5.7 | 17.4 |
| UM 14 | T4b | 75 | 26 | 13.9 | 18.8 |
| UM 15 | T3b | 83 | 26 | 9.5 | 15 |
| UM 16 | T3b | 64 | 21 | 8.4 | 15.6 |
| UM 17 | T4b | 45 | 27 | 13 | 18 |
| UM 18 | T3b | 65 | 29 | 9.1 | 14.8 |
| UM 19 | T3a | 53 | 29 | 9 | 15.1 |

Table 6.1: Patient characteristics
LBD: Largest basal diameter, US: Ultrasound

6.2.2 MRI protocol

All patients were scanned before treatment on a 3T MR scanner (Ingenia, Philips Healthcare, the Netherlands) using the protocol described by Ferreira et al¹⁹ and

a 47 mm diameter surface receive coil covering the affected eye (figure 6.2). The scan parameters of the relevant sequences are listed in table 6.2. Patients were instructed not to wear eye makeup and the affected eye was taped shut. In the last 6 patients a wet gauze was placed on top of the eye to minimize susceptibility artefacts. Patients were immobilized as much as possible using a radiotherapy head support (MaxSupportTM wide shaped, red variant, 117000 HSSETW, Medeo, Schöftland, Switzerland) (figure 6.2). The dynamic time series were acquired using a fat-suppressed 3D spoiled gradient echo sequence with a spatial resolution of 1.25x1.5x1.5 mm³. A bolus of 0.1 mmol/kg gadolinium (DOTAREM; Guerbet, Roissy CdG Cedex, France) was administered 6 s after the start of the scan followed by a 20 ml injection of isotonic saline, using a power injector with an injection rate of 2 ml/sec. The first eight patients were scanned with a flip angle of 5 degrees to match the then used 7 Tesla protocol, where the flip angle was limited to 5 degrees due to SAR restrictions. The flip angle was increased to 13 degrees for subsequent patients as this provided a more optimal contrast for the contrast agent concentrations in our patients as theoretically the optimal flip angle for our spoiled gradient echo sequence is between 13 and 16 degrees assuming a T₁ between 600 and 1500 ms²⁷. Time-resolved angiography with stochastic trajectories sequence (TWIST)²⁵, with a central size of 25% and a peripheral density of 20%, was implemented to increase the temporal resolution to 2 s per dynamic scan to reduce motion artefacts per dynamic image. To determine the baseline lesion T₁, a 3D spoiled gradient echo flip angle series, with flip angles of 2, 5, 9 and 15 degrees, was acquired before the dynamic scan with the same field of view (FOV). Additionally, a B₁⁺-map was acquired using the DREAM sequence²⁸. Finally a post-contrast 3DT₁-weighted scan (3DT₁gd) was acquired for anatomical reference.

| | T ₁ -mapping | B ₁ ⁺ -mapping | DCE-MRI | 3DT ₁ -weighted |
|-------------------------------|-------------------------|--------------------------------------|--------------------|------------------------------|
| Voxel size (mm ³) | 1.25x1.5x1.5 | 2.0x2.0x2.0 | 1.25x1.5x1.5 | 1.0x1.1x1.0 |
| FOV (mm ³) | 80x80x32 | 160x120x33 | 80x80x32 | 80x80x40 |
| TR (ms) | 7 | 7.1 | 4.5 | 350 |
| TE(1) (ms) / TE(2) (ms) | 3.1 | 4.6/6.9 | 2.3 | 9.4 |
| FA (deg) | 2/5/9/15 | 10 | 5 or 13 | 90 |
| Fat. sup. | Proset 11 | SPIR | Proset 11 | SPIR |
| Averages | 1 | 2 | 1 | 1 |
| Scantime (mm:ss) | 4 x 00:09 | 00:21 | 04:20 | 03:23 |
| Remarks | | 0.2 mm gap | 2 s/dynamic; TWIST | Post contrast administration |

Table 6.2: Scan parameters

FOV: Field of view, TR: repetition time, TE: echo time, FA: flip angle, Fat.Sup: fat suppression, SPIR: Spectral Presaturation with Inversion Recovery, TWIST²⁵: time-resolved angiography with stochastic trajectories sequence, a dynamic scan technique where the a semi-randomly part of the 20% of outer k-space is acquired per dynamic.

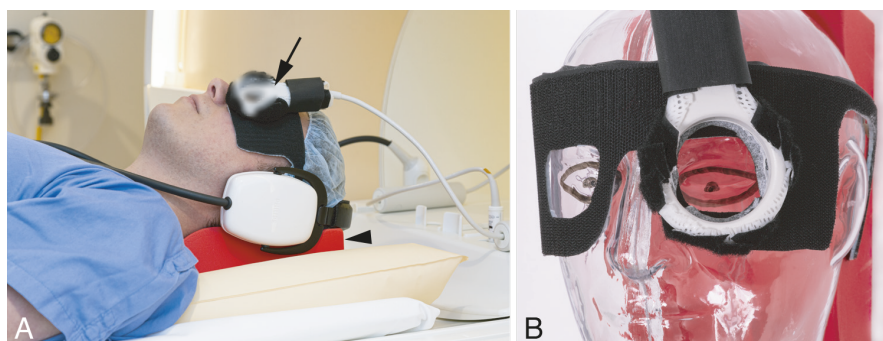


Figure 6.2: (A) The patient was scanned using a 47mm surface receive coil (arrow). This coil was positioned over the affected eye. The head was supported by a radiotherapy head support (arrowhead). (B) Clarification of the positioning of the eye coil on a phantom.

6.2.3 DCE analysis

The analysis of the DCE data consisted of four steps. First, the images of the dynamic scan, flip angle series, B_1^+ -map and 3DT₁gd scan were registered to the dynamic dataset and the lesion was segmented on the 3DT₁gd image. Second, a T_1 -map was calculated using the flip angle series and B_1^+ -map, which was subsequently used to calculate the gadolinium concentration for each lesion voxel in the dynamic scan. Finally, pharmacokinetic modelling was applied using the Tofts model²⁹.

6.2.4 Registration

All timepoints of the dynamic scan were rigidly registered to the 50th of the 125 timepoints in two steps using Elastix 4.9.0 [29]. The 50th timepoint was chosen as it had an intensity comparable to most timepoints, which was beneficial for automatic image registration. The first step consisted of registration of the full FOV to correct for head motion. Subsequently an eye-mask was created using an in-house build Mevislab network (3.0.2, MeVis Medical Solutions AG, Bremen, Germany³⁰). The eye-mask was used for a masked registration to reduce eye motion between timepoints. Additionally, the variable flip angle series, B_1^+ -map and 3DT₁gd scan were rigidly registered to the 50th timepoint using masked registration.

6.2.5 Segmentation

A lesion mask was created by manually segmenting the UM on the 3DT₁gd images using ITK-SNAP³¹. Elastix was used to translate this mask to the registered dynamic scan, using the earlier obtained transformation matrix. Subsequently the voxels within this mask were selected for the pharmacokinetic analysis.

6.2.6 T₁-mapping

The pre-contrast T₁ value of each voxel was obtained from the flip angle series in Matlab (version R2019b, MathWorks, Natick, Massachusetts, USA) as described by Fram et al.³² and Gupta et al.³³. The flip angles were corrected according to the median achieved B₁⁺ of the lesion. Subsequently, a masked 3D median filter with 26-connected components was applied to the T₁-map to remove potential outliers within the lesion. Voxels outside the lesion were excluded from the filter, as the vitreous has significantly higher T₁ values than the lesion³⁴. The UM were classified as melanotic, amelanotic or mixed based on description of the tumour in the medical status by an ophthalmologist.

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6.2.7 Pharmacokinetic modelling

The signal intensities from the DCE-images were converted to concentration time profiles, using the relations described by Tofts²⁹, assuming a gadolinium relaxivity of 3.4 L mmol⁻¹ s⁻¹³⁵. For each voxel the peak concentration was defined as the 95th percentile of the concentration over time. Voxel-by-voxel pharmacokinetic modelling (PKM) was performed using nonlinear least squares fitting of the standard Tofts model using in house build scripts in Matlab. First, the bolus arrival time (BAT) was determined for each patient by fitting the PKM for the first 40 time points of the median lesion concentration curve for 25 different BATs. The BAT with the lowest residuals was selected. The automatic determined BAT was visually correct in 50% of the patients. In the remaining patients the BAT was shifted with one timepoint in 8 cases and two timepoints in 3 cases. Subsequently the PKM was fitted to the full dynamic concentration curve (C(t)) for each voxel within the lesion to obtain the K^{trans} (vascular permeability [min⁻¹]) and v_e (extravascular extracellular space per volume of tissue [unitless]). As no major feeding arteries were in the field-of-view of the DCE scan, an earlier derived population arterial input function (AIF) was used, which was derived from the carotid arteries in ten brain cancer patients. The fit of the Parker AIF to the average AIF resulted in the following parameters: A1 = 46.7 mM·s, σ₁ = 3.5 s, T1 = 12.5 s, A2 = -4.7 mM·s, σ₂ = 5.2 s, T2 = 26.9 s, α = 1.4 mM, β = 0.003 s⁻¹, s = 8.3 s⁻¹, τ = 17.5 s.

6.2.8 Evaluation of the registration

The effect of the registration was evaluated by comparing the concentration curves before and after registration of fifteen randomly selected lesion voxels for all patients. The curves from all patients were randomized and presented unannotated to prevent a potential bias. The observer scored each curve as being sufficient or insufficient for automatic fitting and scored which of the two curves had the best quality or whether the quality was the same based on the amount of visual variance/spikes and motion artefacts in the concentration curve. Observer 1 (MJ) scored all 285 curves and observer 2 (JWB) scored a random subset of 50 to validate the scores. The scores were evaluated using a two-sided Wilcoxon signed rank test.

6.2.9 Evaluation of the error propagation

The effect of the precision error of the B_1^+ , T_1 and registration on the pharmacokinetic analysis were assessed. First, in two additional UM patients, the flip angle series and B_1^+ map were acquired twice to determine the repeatability of the B_1^+ and T_1 measurements. Secondly, the effect precision errors in B_1^+ , T_1 and registration on K^{trans} and v_e was assessed to determine the sensitivity of small inaccuracies of the different analysis steps on the final DCE parameters. To this end the results of different intermediate steps were manually modified in two different patients: a patient with a medium sized amelanotic tumour with a B_1^+ of 82% (patient 7) and a patient with a large melanotic tumour and a B_1^+ of 96% (patient 10). The effect of a precision error in B_1^+ measurement was assessed by increasing and decreasing the measured B_1^+ with 2 and 5 percent. The sensitivity to T_1 changes was determined by changing the measured T_1 for all voxels by 30 ms, the measured precision error for the T_1 mapping, or 60 ms, and 2 and 5 percent of the average tumour T_1 respectively. Finally, the effect of imprecise registration was estimated by artificially shifting the images of individual time-points. Regular eye-motion was simulated by a one voxel shift during 2 time-points after 175 seconds, while the most unfavourable case was simulated by the same shift but immediately after bolus arrival. For all cases, the median K^{trans} and v_e of the tumor were compared with the original analyses.

6.2.10 Statistical analysis

The impact of the different analysis steps on the final PKM was evaluated. The K^{trans} values with and without registration and with and without B_1^+ -correction were visualized. The effect of T_1 -mapping was evaluated by comparing the K^{trans} values based on concentration data with the average T_1 of all patients with the model in which data from the individual T_1 map was used. The difference

in K^{trans} between melanotic and amelanotic lesions for a population T_1 and individual T_1 was tested using unpaired t-tests. The reported PKM values in this paper are the median of the voxels within the lesion mask as the values are not normally distributed. The error bars shown in the figures are the 25th and 75th percentile (IQR). The reported unpaired t-tests were calculated using Matlab. A p-value of 0.05 or smaller was considered statistically significant.

6.3 Results

6.3.1 Registration

The centre of the eye moved up to 3.0 mm (average 1.3 mm) during the 4 minutes acquisition of the dynamic time series with rotations of up to 20 degrees (average 6 degrees) with respect to the first time point.

Registration resulted in a significant improvement in the quality of the concentration curves ($Z = 8.9$, $p < 0.001$). Figure 6.3 shows a representative curve before and after registration. Eye-motion can result in changes in the enhancement and concentration curves, as even small eye motion can result in mismatch between the ROI and actual lesion location, as can be seen in figure 6.3. Without registration, there were spikes (figure 6.3, asterisk) in the contrast agent concentration, which were caused by motion, most likely eye blinks or a different gaze angle. In this case, most of the artefacts were no longer apparent after registration of the eye, although still some residual motion artefacts can be seen in the concentration curve. Note that eye motion was not resolved with the unmasked registration.

Before registration 40% ($n = 113/285$) of the curves were scored as insufficient quality to perform a fit compared to 15% ($n = 43/285$) after registration. In 55% of the curves ($n = 157/285$) the curve was scored as improved, in 36% ($n = 102/285$) of the curves no clear effect of the registration was observed, while in 9% ($n = 26/285$) the curve was scored as deteriorated after registration although in 15/26 of these curves the curves received the same quality, indicating a minimal difference. No significant difference in scoring was found between the scoring of observer 1 and 2 ($p = 0.09$).

The reported increase in quality of the concentration curves after registration resulted in a change of the PKM parameter values. The maximum absolute change in K^{trans} was 0.25 min^{-1} with an average absolute change of 0.06 min^{-1} (figure 6.3c & table 6.3).

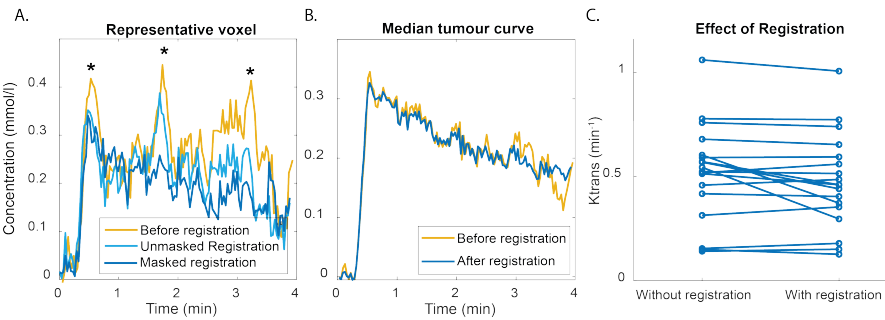


Figure 6.3: (A,B) Concentration time curves of a representative patient (18) before and after registration. Before registration motion artefacts were present (asterisk) furthermore, unmasked registration was not able to fully resolve the motion. A movie of the dynamic scan of patient 18 visualizing the motion in the scan can be found in the supplementary materials. (C) Registration changes the K^{trans} of the UM patients with an average of 0.06 min^{-1}

| Patient | Ktrans | Without registration | % change | Without B1+ | % change | Average T1 | % change |
|---------|--------|----------------------|----------|-------------|----------|------------|----------|
| UM 1 | 0.20 | 0.18 | -10 | 0.27 | 36 | 0.26 | 31 |
| UM 2 | 0.46 | 0.57 | 24 | 0.77 | 68 | 0.70 | 51 |
| UM 3 | 0.37 | 0.60 | 62 | 0.50 | 33 | 0.19 | -50 |
| UM 4 | 0.13 | 0.15 | 18 | 0.17 | 35 | 0.16 | 23 |
| UM 5 | 0.46 | 0.51 | 11 | 0.55 | 20 | 0.52 | 13 |
| UM 6 | 0.15 | 0.14 | -6 | 0.18 | 20 | 0.11 | -28 |
| UM 7 | 0.30 | 0.54 | 84 | 0.44 | 49 | 0.53 | 80 |
| UM 8 | 0.40 | 0.42 | 3 | 0.50 | 24 | 0.55 | 35 |
| UM 9 | 0.35 | 0.32 | -11 | 0.44 | 24 | 0.52 | 47 |
| UM 10 | 0.77 | 0.78 | 1 | 0.85 | 9 | 0.94 | 22 |
| UM 11 | 0.18 | 0.15 | -14 | 0.29 | 64 | 0.19 | 7 |
| UM 12 | 0.44 | 0.57 | 29 | 0.44 | 0 | 0.43 | -2 |
| UM 13 | 0.65 | 0.68 | 4 | 0.76 | 16 | 0.46 | -30 |
| UM 14 | 0.51 | 0.52 | 2 | 0.72 | 39 | 0.29 | -44 |
| UM 15 | 0.59 | 0.59 | -1 | 0.80 | 35 | 0.27 | -55 |
| UM 16 | 0.49 | 0.46 | -6 | 0.61 | 26 | 0.20 | -59 |
| UM 17 | 1.01 | 1.06 | 5 | 1.03 | 3 | 1.17 | 17 |
| UM 18 | 0.56 | 0.52 | -7 | 0.68 | 21 | 0.25 | -55 |
| UM 19 | 0.74 | 0.76 | 3 | 0.70 | -6 | 0.82 | 10 |

Table 6.3: K^{trans}

6.3.2 B_1^+ and T_1 -mapping

The median lesion T_1 per patient showed a wide range from 522 ms to 1509 ms as is shown in figure 6.4b. The average T_1 of all patients was 1122 ms, this value was used as population T_1 for subsequent comparisons on the effect of T_1 on the PKM. Amelanotic lesions had an average T_1 of 1350 ms while the average T_1 of 888 ms for the melanotic lesions was significantly lower ($p = 0.03$). The average T_1 of the mixed lesions was 1193 ms. The average achieved B_1^+ in the

lesion was 91% (range 77% to 104%). Correction of the flip angles resulted in an average absolute change in T_1 of 208 ms and an absolute change in K^{trans} of 0.11 min^{-1} , figure 6.4c & table 6.3.

When a population average T_1 was used the K^{trans} appeared to mainly resemble the amount of pigmentation, with the melanotic showing a significant lower K^{trans} compared to amelanotic lesions ($p < 0.01$, figure 6.4d). However, when the actual T_1 was included in the analysis, the K^{trans} changed with 0.15 min^{-1} on average (figure 6.4d & table 6.3) and the bias was resolved as no systematic difference was found between the K^{trans} of melanotic and amelanotic lesions ($p = 0.37$).

The strong effect of the melanin concentration on the perfusion quantification can be seen in a mixed lesion with both a melanotic and amelanotic lobe, Figure 6.5. On the TIC, the amelanotic part of the lesion appeared to be enhancing stronger than the melanotic part, 225% vs 150%. The melanotic lobe has, however, an almost 1000 ms shorter T_1 than the amelanotic lobe, on average 494 ms vs 1464 ms respectively. When the T_1 was included in the conversion to concentration, a very similar concentration was found in both lesions, although still a higher peak concentration was measured in the early timepoints of the amelanotic lobe.

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6.3.3 Pharmacokinetic parameters

When all corrections were applied, a wide range of K^{trans} values was observed (figure 6.6c). The median K^{trans} per lesion ranged from 0.13 to 1.0 min^{-1} with a mean of 0.46 min^{-1} . The median v_e was 0.22 on average with a range from 0.10 to 0.51. Within lesions a wide distribution of K^{trans} and v_e was observed. The maximum IQR (75th -25th percentile) of the K^{trans} was 0.99 min^{-1} and the average IQR was 0.40 min^{-1} . For the v_e the maximum IQR was 0.83 and the average IQR was 0.18.

6.3.4 Error propagation

The average precision error of the achieved B_1^+ and T_{11} was 0.2% and 30 ms respectively. Major inaccuracies (5%) in the measurement of the achieved B_1^+ can result in a difference up to 12% in the determined K^{trans} and 19% in the determined v_e (table 4). Differences in the T_1 measurement (30 ms) can lead to changes in K^{trans} and v_e up to 5% (table 4). Imprecisions in the registration during the second half of the acquisition had a minimal effect on the outcomes of K^{trans} and v_e ($< 1\%$), while a similar imprecisions directly after contrast uptake

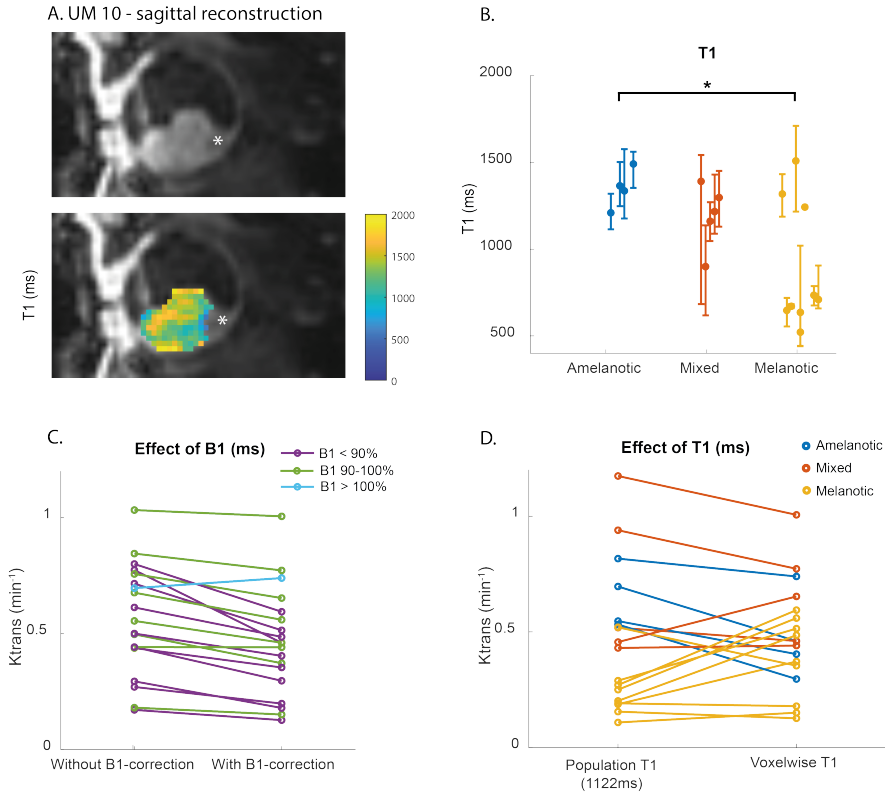


Figure 6.4: T1-mapping
(A) Slice of UM patient 10 showing an inhomogeneous T_1 of the lesion. The retinal detachment (Asterix) is excluded from the analysis. (B) T_1 values of amelanotic, mixed, melanotic and non-UM lesions (median and IQR). The three melanotic lesions with a higher T_1 are small lesions most likely suffering from partial volume effects. The T_1 between amelanotic and melanotic lesions is significantly different ($p = 0.03$). No significant difference was found between mixed lesions and either amelanotic or melanotic lesions. (B,C) B_1^+ -correction change the K^{trans} with an average of 0.11 min^{-1} . (D) When a population T_1 was used the K^{trans} mainly resembled the amount of pigmentation with a low K^{trans} for melanotic lesions. When the actual T_1 was used the K^{trans} changed with an average of 0.15 min^{-1} .

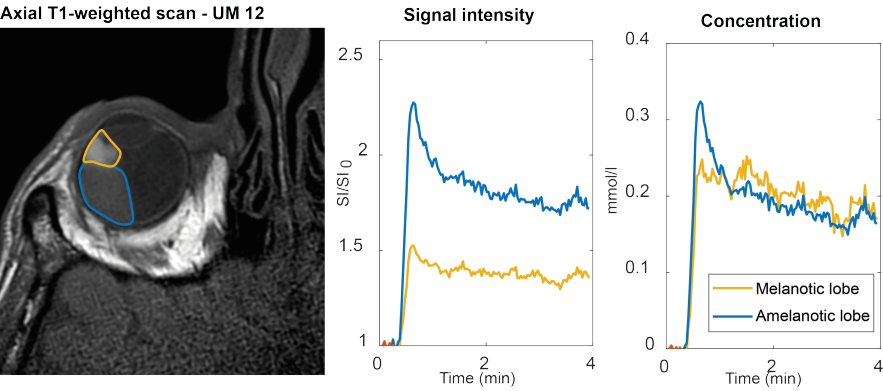


Figure 6.5: The effect of T_1 on the signal intensity
Bilobar lesion with both an amelanotic and a melanotic lobe. The amelanotic part of the lesion appears to be enhancing stronger than the melanotic part. When, however, the actual T_1 was included in the calculation of the contrast agent concentration, a very similar concentration was found in both lesions, although still a distinct difference was present in the early timepoints.

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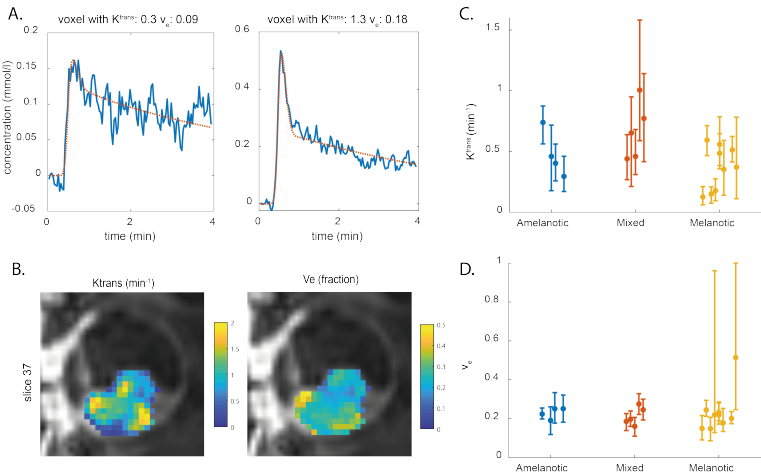


Figure 6.6: Pharmacokinetic modelling results
(A) Concentration curve and fit of two voxels of patient 10 showing the inhomogeneity of the lesion. (B) A sagittal slice of UM patient 10 showing K^{trans} and v_e maps showing an inhomogeneous values of K^{trans} and v_e in the lesion. (C&D). The median and IQR of the K^{trans} and v_e in the lesion. All patients except one UM had a v_e lower than 0.3.

| | | | | | | | | | |
|--|--|------------------------------------|-------|------------------------------------|-------|-----------|-------|-----------|-------|
| <i>Sensitivity to inaccuracies in the registration</i> | | 2-3 minutes after contrast arrival | | Immediately after contrast arrival | | | | | |
| Relative change in | | Ktrans | ve | Ktrans | ve | | | | |
| Plateau curve / Medium sized UM | | -0.5% | -0.6% | 3.0% | -0.5% | | | | |
| Washout curve / Large UM | | -0.2% | 0.1% | 2.6% | 0.1% | | | | |
| <i>Sensitivity to inaccuracies in B1+</i> | | B1+ - 5% | | B1+ - 2% | | B1++ 2% | | B1+ + 5% | |
| Relative change in | | Ktrans | ve | Ktrans | ve | Ktrans | ve | Ktrans | ve |
| Low achieved B1 (82%) | | -10.7% | -7.2% | -3.6% | 0.3% | 6.4% | 10.6% | 14.1% | 18.7% |
| Normal achieved B1 (96%) | | -11.8% | -7.4% | -5.8% | -1.2% | 2.5% | 7.5% | 8.9% | 14.2% |
| <i>Sensitivity to inaccuracies in T1</i> | | T1 - 60ms | | T1 - 30ms | | T1 + 30ms | | T1 + 60ms | |
| Relative change in | | Ktrans | ve | Ktrans | ve | Ktrans | ve | Ktrans | ve |
| Amelanotic UM | | 10.7% | 9.4% | 5.4% | 4.8% | -5.0% | -4.7% | -8.9% | -9.2% |
| Melanotic UM | | 6.7% | 6.0% | 3.3% | 3.1% | -3.4% | -2.8% | -6.8% | -5.8% |

Table 6.4: Error propagation

resulted in 3% change in Ktrans.

6.4 Discussion

Recent developments in ocular MR-imaging allow for the acquisition of DCE-images with sufficient temporal and spatial resolution to perform DCE-MRI in the eye, as the currently achievable isotropic spatial resolution of 1 mm is sufficient to assess the smaller intraocular lesions, while the 2 s temporal resolution yields more than sufficient time points to determine the inflow characteristics of the lesion¹⁹. With this improved protocol, DCE-MRI was performed in 19 intraocular lesions and the effect of eye specific challenges on the quantification was investigated.

As the eye can rotate within the head, registering the complete Field-of-View(FOV) of all time points is not sufficient to correct for eye motion. Therefore a dedicated registration method was developed to mitigate the effect of gazing variations on the measured concentration curve. These corrections resulted in absolute changes in K^{trans} up to 0.25 min⁻¹. Although rigid registration of the complete FOV has been proposed to correct for motion of intraocular lesions²¹, we proved with our experiments that it is not sufficient, since the eye moves within the head. We showed that masked-registration improved the quality of the curves significantly, however, still the quality of 15% of the concentration curves was scored as insufficient to perform an automatic fit on. Smaller lesions had a higher percentage of insufficient quality voxels indicating that our registration cannot fully resolve effects of motion at the edge of the lesion. In 4% (n = 10/285) of the curves, across different subjects and tumour sizes, registration seemed to have deteriorated the quality at some timepoints although the deterioration was minor compared to the improvement by the registration in the other voxels. This mostly occurred at the edge of the tumour and might be attributed to partial volume effects, although inhomogeneities in the tumour might play a role as well. A MRI-protocol with an increased resolution might therefore also be favourable

to decrease the effect of these residual errors in the registration. Although cued blinking might be implemented to reduce motion during acquisition, the resulting twofold reduction in temporal resolution will likely not be beneficial for the pharmacokinetic modelling^{8,9}. It might be beneficial to register all scans to the higher resolution 3DT₁gd instead of one of the dynamic timepoints.

Only one paper²¹ was found that used T₁ -mapping for the quantitative DCE analysis of ocular lesions. We found a significant difference in T₁ between melanotic and amelanotic lesions and a strong effect on the quantification indicating that T₁ -mapping is a crucial step in DCE quantification. We therefore recommend to include the actual T₁ in the analysis of DCE of ocular lesions to prevent the bias introduced by the amount of melanin in the tumour. The effect of melanin can be seen clearly in patients with a bilobar lesion, as shown in figure 5. A similar effect was observed between lesions, where amelanotic lesions appears to enhance more than melanotic lesions. This was, however, primarily the result of its longer T₁ and not of an increased contrast agent concentration.

Finally, our results indicate that registration and B₁⁺-correction are important steps in the quantitative analysis of ocular DCE, but these steps affect the pharmacokinetic parameters to a lesser extent than the differences in T₁, and are independent of the type of lesion. DCE scans are clinically often evaluated by assessing the Time Intensity Curves (TIC), instead of the actual gadolinium concentration which can be calculated from these intensities^{17–20}. However, by comparing these TICs between lesions, the potential differences in T₁ between these lesions are ignored, which can lead to erroneous interpretation of the data. A higher (maximum) relative enhancement or area under the curve might, for example, be interpreted as an increased perfusion in the lesion, but might actually be the result of a less pigmented lesion which has therefore a higher T₁. As the overall shape of the TIC is not affected by the scaling effect of the T₁, a classification based on the curve pattern can still be useful when only the signal intensity is available.^{36,37}

In line with earlier genetic and pathology studies,^{38–41} our results indicate that uveal melanomas are inhomogeneous, making a voxel wise volumetric assessment of ocular lesions preferred over a single 2D ROI analysis. The relatively thick slices of 3 mm used in the study of Kamrava²¹, likely still resulted in representative sample of the lesion, due to averaging in the slice-direction, resulting in the observed correlation between monosomy 3 and Ktrans. However, a lower resolution limits the possibility for motion correction and analysis of small intraocular lesions, making a higher resolution with a voxel wise analysis preferred. Overall, ROI based (semi-)quantitative DCE-MRI analysis without T₁ -mapping for ocular lesions is not recommended as this would result in a significant bias as most DCE measures are dependent on the baseline T₁.

There are areas in which ocular DCE can be further improved. First of all, the flip angle of the DCE-MRI acquisition was 5 degrees for the first eight patients and 13 degrees for subsequent patients. For the determination of reference PKM values of lesions a standardized protocol should be used for all patients. On the hardware side, a multichannel eye-coil^{42,43} might result in multiple advances in ocular DCE. An increased channel count might not only enable an increased temporal or spatial resolution, but also a potential larger FOV which can be used to determine the AIF on a patient-specific level. As with the single loop-coil approach no major blood vessels were available within the FOV, we relied on a population based AIF. Population based AIFs are a widely used approach but result a less accurate estimation of especially K^{trans} as the AIF is influenced by body mass and cardiac output^{44,45}. Eye muscles were investigated as reference tissue, but the perfusion of the eye muscles appeared not to be consistent within a single subject and might be also influenced by lesion location and therefore unreliable.

Although the spatial and temporal resolution achieved with our protocol are high compared to other ocular DCE studies, the spatial resolution of $1.25 \times 1.5 \times 1.5 \text{ mm}^3$ is still a limiting factor for small ocular lesions. The smallest lesion included in this study was 32 voxels. A higher resolution would be preferred as this allows for the edge voxels to be removed from the analysis, as the results from these voxels are less reliable due to partial volume effects, not only of the dynamic scan, but also in the T_1 -mapping. Therefore, for the current resolution, DCE-MRI of small lesions (i.e. thickness $< 2 \text{ mm}$) is likely less accurate. In situations where the conditions are less optimal (e.g. no orbital coil is or only a 1.5 T MRI is present) DCE-MRI of intraocular lesions could be performed, although the decrease in image resolution would increase the lesion size required for reliable results. Moreover, although the TWIST-sequence successfully reduces the motion artefacts by reducing the acquisition time per dynamic, the effective spatial resolution for rapidly changing concentrations, in particular the inflow of the contrast agent, might be lower. Nevertheless, the effect of precision errors in analysis steps, such as T_1 determination or registration, on the obtained pharmacokinetic parameters generally showed to be less than 5%. Furthermore, some choices in the proposed analysis pipeline might benefit from a more thorough evaluation to further improve the analysis, such as the optimal reference image for the registration, the potential benefit of applying the median filter to the source FA-series images instead of the resulting T_1 -map and other PK-models, such as the extended Tofts model to include a vascular component⁴⁶. Finally, it is important to assess the reproducibility of DCE-MRI for ocular lesions between visits and centers.

Some of the proposed steps to enable reliably ocular DCE-MRI, such as the

eye-specific registration, might be less easily incorporated into clinical practice. However, the inclusion of differences in T_1 between lesions in the analysis, which has the strongest effect on the PK-measures, is available in various clinical software packages. Although the elementary TIC-classification, eg. a distinction between a washout and plateau curve, is not affected by the T_1 , other elementary measures, in particular the relative enhancement, are significantly affected. Clinically, this conversion to concentration is particularly important as conversely to UM, the majority of other intraocular lesions are non-pigmented, resulting a biased evaluation. When the pharmacokinetic parameters have been determined in a larger cohort of patients with intraocular lesions, they could aid to differentiate between benign and different malignant intraocular lesions. Although the eye specific motion correction is currently not available clinically, head motion can still be corrected with regular registration methods. When only this form of registration is available, a careful evaluation of the individual data is needed to screen for motion and to potentially remove motion-corrupted time points. We anticipate that this approach will be clinically sufficient to aid in the differential diagnosis, especially as other information, such as DWI, can be included in the considerations. This research furthermore showed that some UM are inhomogeneous in composition. For subsequent studies to assess these inhomogeneities, and assess their potential relation to genetic factors and the patients' prognosis²¹, full motion correction will be needed, as these inhomogeneities can amplify the effect of eye motion on the final parameters.

6.5 Conclusion

Although MRI of eyes is challenging in many aspects, we showed that quantitative DCE-MRI analysis can be performed for intraocular lesions by increasing the temporal and spatial resolution of the dynamic scan and using dedicated registration and T_1 -mapping with B_1^+ -correction in the analysis. In the clinic DCE-MRI analysis might aid in the diagnosis, prognosis and follow-up of intraocular masses.

6.6 References

- [1] M. G. Jaarsma-Coes, T. A. Ferreira, P. J. van Houdt, U. A. van der Heide, G. P. Luyten, and J.-W. M. Beenakker, "Eye-specific quantitative dynamic contrast-enhanced mri analysis for patients with intraocular masses," *Magnetic Resonance Materials in Physics, Biology and Medicine*, pp. 1–13, 2021.
- [2] IKNL, *NKR cijfers*. [Online]. Available: <https://iknl.nl/nkr-cijfers>.

- [3] S. Kaliki and C. L. Shields, *Uveal melanoma: Relatively rare but deadly cancer*, 2017. DOI: 10.1038/eye.2016.275.
- [4] C. L. Shields *et al.*, "Metastasis of uveal melanoma millimeter-by-millimeter in 8033 consecutive eyes," *Archives of Ophthalmology*, vol. 127, no. 8, pp. 989–998, 2009, ISSN: 00039950. DOI: 10.1001/archophthamol.2009.208.
- [5] I. D. Fabian and M. S. Sagoo, "Understanding retinoblastoma: epidemiology and genetics," *Community eye health*, vol. 31, no. 101, p. 7, 2018, ISSN: 0953-6833.
- [6] V. M. Cohen, "Ocular metastases," in *Eye (Basingstoke)*, 2013. DOI: 10.1038/eye.2012.252.
- [7] C. L. Shields *et al.*, "Choroidal nevus transformation into melanoma: Analysis of 2514 consecutive cases," *Archives of Ophthalmology*, 2009, ISSN: 00039950. DOI: 10.1001/archophthamol.2009.151.
- [8] J. W. Beenakker, G. A. van Rijn, G. P. Luyten, and A. G. Webb, "High-resolution MRI of uveal melanoma using a microcoil phased array at 7 T," *NMR in Biomedicine*, 2013, ISSN: 09523480. DOI: 10.1002/nbm.3041.
- [9] B. A. Berkowitz, C. McDonald, Y. Ito, P. S. Tofts, Z. Latif, and J. Gross, "Measuring the human retinal oxygenation response to a hyperoxic challenge using MRI: Eliminating blinking artifacts and demonstrating proof of concept," *Magnetic Resonance in Medicine*, vol. 46, no. 2, pp. 412–416, 2001, ISSN: 07403194. DOI: 10.1002/mrm.1206.
- [10] T. Niendorf *et al.*, *Ophthalmic Magnetic Resonance Imaging: Where Are We (Heading To)?* 2021. DOI: 10.1080/02713683.2021.1874021.
- [11] T. Ferreira *et al.*, "MR and CT Imaging of the Normal Eyelid and its Application in Eyelid Tumors," *Cancers*, vol. 12, p. 658, Mar. 2020. DOI: 10.3390/cancers12030658.
- [12] K. R. Keene *et al.*, "The feasibility of quantitative MRI of extra-ocular muscles in myasthenia gravis and Graves' orbitopathy," *eng, NMR in biomedicine*, vol. 34, no. 1, e4407–e4407, Jan. 2021, ISSN: 1099-1492. DOI: 10.1002/nbm.4407. [Online]. Available: <https://pubmed.ncbi.nlm.nih.gov/32893386/><https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7757175/>.
- [13] L. van Vught, D. P. Shamonin, G. P. Luyten, B. C. Stoel, and J.-W. M. Beenakker, "MRI-based 3D retinal shape determination," *[submitted]*, 2021.

- [14] J. W. M. Beenakker *et al.*, "Clinical evaluation of ultra-high-field MRI for three-dimensional visualisation of tumour size in uveal melanoma patients, with direct relevance to treatment planning," *Magnetic Resonance Materials in Physics, Biology and Medicine*, vol. 29, no. 3, pp. 571–577, 2016, ISSN: 13528661. DOI: 10.1007/s10334-016-0529-4.
- [15] E. Fleury *et al.*, "3D MRI-based treatment planning approach for non-invasive ocular proton therapy," *Medical Physics*, 2020, ISSN: 0094-2405.
- [16] P. V. Foti *et al.*, "Uveal melanoma: quantitative evaluation of diffusion-weighted MR imaging in the response assessment after proton-beam therapy, long-term follow-up," *Radiologia Medica*, 2017, ISSN: 18266983. DOI: 10.1007/s11547-016-0697-3.
- [17] X. Jiang *et al.*, "Dynamic contrast-enhanced MRI of ocular melanoma," *Melanoma Research*, 2015, ISSN: 14735636. DOI: 10.1097/CMR.0000000000000142.
- [18] W. Wei *et al.*, "Dynamic Contrast-Enhanced Magnetic Resonance Imaging of Ocular Melanoma as a Tool to Predict Metastatic Potential," *Journal of Computer Assisted Tomography*, 2017, ISSN: 15323145. DOI: 10.1097/RCT.0000000000000598.
- [19] T. A. Ferreira, L. G. Fonk, M. G. Jaarsma-Coes, G. G. van Haren, M. Marinkovic, and J. W. M. Beenakker, "MRI of uveal melanoma," *Cancers*, vol. 11, no. 3, pp. 1–20, 2019, ISSN: 20726694. DOI: 10.3390/cancers11030377.
- [20] F. Rodjan *et al.*, "Retinoblastoma: Value of dynamic contrast-enhanced MR imaging and correlation with tumor angiogenesis," *American Journal of Neuroradiology*, 2012, ISSN: 01956108. DOI: 10.3174/ajnr.A3119.
- [21] M. Kamrava *et al.*, "Quantitative multiparametric MRI in uveal melanoma: increased tumor permeability may predict monosomy 3," *Neuroradiology*, vol. 57, no. 8, pp. 833–840, 2015, ISSN: 14321920. DOI: 10.1007/s00234-015-1546-0.
- [22] M. Y. Chang, N. P. Rao, B. L. Burgess, L. Johnson, and T. A. Mccannel, "Heterogeneity of monosomy 3 in fine needle aspiration biopsy of choroidal melanoma," *Molecular Vision*, 2013, ISSN: 10900535.
- [23] J. L. Chien, K. Sioufi, T. Surakiatchanukul, J. A. Shields, and C. L. Shields, *Choroidal nevus: A review of prevalence, features, genetics, risks, and outcomes*, 2017. DOI: 10.1097/ICU.0000000000000361.
- [24] W. S. Enochs, P. Petherick, A. Bogdanova, U. Mohr, and R. Weissleder, "Paramagnetic metal scavenging by melanin: MR imaging," *Radiology*, 1997, ISSN: 00338419. DOI: 10.1148/radiology.204.2.9240529.

- [25] L. A. Tudorica *et al.*, "A feasible high spatiotemporal resolution breast DCE-MRI protocol for clinical settings," *Magnetic Resonance Imaging*, 2012, ISSN: 0730725X. DOI: 10.1016/j.mri.2012.04.009.
- [26] E. D. Baron, M. Di Nicola, and C. L. Shields, *Updated ajcc classification for posterior uveal melanoma: A case example of a patient with choroidal melanoma is discussed in light of the latest edition of this cancer staging manual*, 2018.
- [27] D. De Naeyer, J. Verhulst, W. Ceelen, P. Segers, Y. De Deene, and P. Verdonck, "Flip angle optimization for dynamic contrast-enhanced mri-studies with spoiled gradient echo pulse sequences," *Physics in Medicine & Biology*, vol. 56, no. 16, p. 5373, 2011.
- [28] K. Nehrke and P. Börnert, "DREAM-a novel approach for robust, ultrafast, multislice B1 mapping," *Magnetic Resonance in Medicine*, 2012, ISSN: 07403194. DOI: 10.1002/mrm.24158.
- [29] P. S. Tofts, "T1-weighted DCE imaging concepts: modelling, acquisition and analysis," *signal*, vol. 500, no. 450, p. 400, 2010.
- [30] F. Ritter *et al.*, "Medical Image Analysis: A Visual Approach," *IEEE Pulse*, vol. 2, no. 6, pp. 60–70, 2011, ISSN: 2154-2287. DOI: 10.1109/MPUL.2011.942929. [Online]. Available: http://ieeexplore.ieee.org/xpls/abs_all.jsp?arnumber=6088923&tag=1.
- [31] P. A. Yushkevich *et al.*, "User-guided 3D active contour segmentation of anatomical structures: significantly improved efficiency and reliability," *Neuroimage*, vol. 31, no. 3, pp. 1116–1128, 2006, ISSN: 1053-8119.
- [32] E. K. Fram *et al.*, "Rapid calculation of T1 using variable flip angle gradient refocused imaging," *Magnetic Resonance Imaging*, 1987, ISSN: 0730725X. DOI: 10.1016/0730-725X(87)90021-X.
- [33] R. K. Gupta, "A new look at the method of variable nutation angle for the measurement of spin-lattice relaxation times using fourier transform NMR," *Journal of Magnetic Resonance (1969)*, 1977, ISSN: 00222364. DOI: 10.1016/0022-2364(77)90138-X.
- [34] K. Koolstra, J. W. M. Beenakker, P. Koken, A. Webb, and P. Börnert, "Cartesian MR fingerprinting in the eye at 7T using compressed sensing and matrix completion-based reconstructions," *Magnetic Resonance in Medicine*, 2019, ISSN: 15222594. DOI: 10.1002/mrm.27594.
- [35] Geneesmiddeleninformatiebank. "Samenvatting van de productkenmerken: Dotarem 0,5 mmol/ml." (), [Online]. Available: https://www.geneesmiddeleninformatiebank.nl/smpc/h27039_smpc.pdf (visited on 09/30/2019).

- [36] S. R. Ro, P. Asbach, E. Siebert, E. Bertelmann, B. Hamm, and K. Erb-Eigner, "Characterization of orbital masses by multiparametric MRI," *European Journal of Radiology*, 2016, ISSN: 18727727. DOI: 10.1016/j.ejrad.2015.11.041.
- [37] Y. Yuan, X. P. Kuai, X. S. Chen, and X. F. Tao, "Assessment of dynamic contrast-enhanced magnetic resonance imaging in the differentiation of malignant from benign orbital masses," *European Journal of Radiology*, 2013, ISSN: 0720048X. DOI: 10.1016/j.ejrad.2013.03.001.
- [38] K. G. Griewank and R. Murali, "Pathology and genetics of uveal melanoma," *Pathology - Journal of the RCPA*, vol. 45, no. 1, 2013, ISSN: 0031-3025. [Online]. Available: https://journals.lww.com/pathologyrcpa/Fulltext/2013/01000/Pathology_and_genetics_of_uveal_melanoma.3.aspx.
- [39] W. Maat *et al.*, "The Heterogeneous Distribution of Monosomy 3 in Uveal Melanomas: Implications for Prognostication Based on Fine-Needle Aspiration Biopsies," *Archives of Pathology & Laboratory Medicine*, vol. 131, no. 1, pp. 91–96, Jan. 2007, ISSN: 0003-9985. DOI: 10.5858/2007-131-91-THDOMI. [Online]. Available: <https://doi.org/10.5858/2007-131-91-THDOMI>.
- [40] A. K. Miller, M. J. Benage, D. J. Wilson, and A. H. Skalet, "Uveal Melanoma with Histopathologic Intratumoral Heterogeneity Associated with Gene Expression Profile Discordance," *Ocular oncology and pathology*, vol. 3, no. 2, pp. 156–160, Jul. 2017, ISSN: 2296-4681. DOI: 10.1159/000453616. [Online]. Available: <https://pubmed.ncbi.nlm.nih.gov/28868288/><https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5566757/>.
- [41] I. W. McLean, W. D. Foster, L. E. Zimmerman, and J. W. Gamel, "Modifications of Callender's classification of uveal melanoma at the Armed Forces Institute of Pathology," *American journal of ophthalmology*, vol. 96, no. 4, pp. 502–509, 1983, ISSN: 0002-9394.
- [42] A. Graessl *et al.*, "Ophthalmic magnetic resonance imaging at 7 T using a 6-channel transceiver radiofrequency coil array in healthy subjects and patients with intraocular masses," *Investigative radiology*, vol. 49, no. 5, pp. 260–270, 2014, ISSN: 0020-9996.
- [43] R. K. Glarin *et al.*, "MR-EYE: High-Resolution MRI of the Human Eye and Orbit at Ultrahigh Field (7T)," *Magnetic Resonance Imaging Clinics of North America*, vol. 29, no. 1, pp. 103–116, 2021, ISSN: 1064-9689. DOI: <https://doi.org/10.1016/j.mric.2020.09.004>. [Online]. Available:

- <https://www.sciencedirect.com/science/article/pii/S1064968920300696>.
- [44] A. Fedorov *et al.*, "A comparison of two methods for estimating DCE-MRI parameters via individual and cohort based AIFs in prostate cancer: A step towards practical implementation," *Magnetic Resonance Imaging*, 2014, ISSN: 18735894. DOI: 10.1016/j.mri.2014.01.004.
- [45] G. J. Parker *et al.*, "Experimentally-derived functional form for a population-averaged high-temporal-resolution arterial input function for dynamic contrast-enhanced MRI," *Magnetic Resonance in Medicine*, 2006, ISSN: 07403194. DOI: 10.1002/mrm.21066.
- [46] S. P. Sourbron and D. L. Buckley, "On the scope and interpretation of the Tofts models for DCE-MRI," *Magnetic Resonance in Medicine*, 2011, ISSN: 07403194. DOI: 10.1002/mrm.22861.