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ORIGINAL ARTICLE

Generation and reactivity analysis of human recombinant monoclonal antibodies directed against epitopes on HLA-DR

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In kidney transplantation, eplet mismatches between donor and recipient have been associated with de novo donor-specific antibody development. Eplets are theoretically defined configurations of polymorphic amino acids and require experimental verification to establish whether they can be bound by alloantibodies. Human HLA-specific monoclonal antibodies (mAbs) have been instrumental for this purpose but are largely lacking for HLA class II. In this study, we isolated single HLA-DR-specific memory B cells from peripheral blood of immunized individuals ($n = 3$) using HLA class II tetramers to generate recombinant human HLA-DR antigen-reactive mAbs ($n = 5$). Comparison of the amino acid composition of the reactive HLA alleles in relation to the antibody reactivity patterns led to identification of 3 configurations, 70Q 73A, 31F 32Y 37Y, and 14K 25Q recognized, respectively, by HLA-DRB1*01:01, HLA-DRB1*04:01, and HLA-DRB1*07:01 antigen-reactive mAbs. The first 2 correspond to eplets 70QA and 31FYY and can now be considered antibody verified. The latter indicates that eplet 25Q needs to be redefined before being considered as antibody verified. Generation and reactivity analysis of human HLA-DR mAbs allowed for identification of amino acid configurations corresponding to known eplets, whereas the other patterns may be used to redefine eplets with similar, but not identical predicted amino acid composition.

KEYWORDS

alloantibody, basic (laboratory) research / science, histocompatibility, immunogenetics, major histocompatibility complex (MHC), recipient selection, sensitization

Abbreviations: Å, angstrom; AA, amino acid; APC, allophycocyanin; BCM, background corrected mean fluorescence intensity; BCR, B cell receptors; BSA, bovine serum albumin; CDC, complement-dependent cytotoxicity; cDNA, complementary deoxyribonucleic acid; CDR, complementarity-determining region; DSA, donor-specific antibodies; EBV-LCL, Epstein-Barr virus-transformed lymphoblastoid cell line; EDTA, ethylenediaminetetraacetic acid; ELISA, enzyme-linked immunosorbent assay; FACS, fluorescence-activated cell sorting; FITC, fluorescein isothiocyanate; HLA, human leukocyte antigen; IgG, immunoglobulin G; IL, interleukin; IMDM, Iscove's modified Dulbecco's medium; mAbs, monoclonal antibodies; MFI, mean fluorescence intensity; OD, optical density; PBMC, peripheral blood mononuclear cells; PBS, phosphate-buffered saline; PCR, polymerase chain reaction; PE, phycoerythrin; RACE, rapid amplification cDNA ends; RNA, ribonucleic acid; SAB, single antigen beads; TerEp, Teresaki epitope; VH, heavy-chain variable domain; VL, light-chain variable domain.

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1 | INTRODUCTION

In kidney transplantation, mismatched donor human leucocyte antigens (HLA) can lead to the formation of de novo donor-specific antibodies (dnDSA), which are associated with inferior graft survival.¹ These dnDSA are induced by polymorphic amino acid (AA) configurations on mismatched HLA molecules that have been theoretically defined as eplets,^{2,3} which are listed in an online registry.^{4,5} Eplets are defined as configurations of surface exposed polymorphic AA within 3-3.5 Å radius.^{2,3} Various studies have shown an association between the number of eplet mismatches and dnDSA formation,⁶⁻⁸ especially for HLA-DR and HLA-DQ.^{9,10}

Eplets resemble functional epitopes but they are not necessarily identical. A functional epitope determines the specificity of an antibody through interaction, in most cases, with the complementarity-determining region 3 of the heavy chain (CDR-H3) of the antibody.¹¹⁻¹³ The complete surface area of an antigen that interacts with the paratope of an antibody is referred to as the structural epitope, which consists of additional AA configurations within 15 Å radius that are essential for binding and affinity.¹⁴⁻¹⁶

Because not every individual eplet is necessarily immunogenic due to the nature of the AA substitution, physiochemical properties, and the absence or presence of an accompanying T helper cell epitope, verification of the actual interaction of eplets with human antibodies is required to determine their clinical relevance.

For antibody verification of eplets human HLA-specific mAbs¹⁷⁻¹⁹ have been instrumental alongside HLA antibodies purified by absorption and elution from sera of alloimmunized individuals.²⁰⁻²² Several eplets have been listed as being verified based on mouse mAbs and/or polyclonal sera,^{4,5,23-26} which, in our opinion, is not sufficient to determine whether a single human antibody can interact with an eplet. The limited array of available HLA class II-specific mAbs hampers verification of many HLA class II eplets. Indeed, Sapir-Pichhadze et al recently observed a strong effect of HLA class I antibody-verified eplet mismatches on graft survival, with no residual effect of HLA class I nonverified eplets. For HLA class II, a similar effect was shown, albeit with a residual effect of HLA class II nonverified eplets.²⁷ These data indicate that for HLA class II verification of additional eplets will allow for better risk stratification for individual patients.

Human mAbs can be generated from isolated antigen-specific B cells using recombinant technology.²⁸⁻³⁰ Low frequency HLA-specific memory B cells in peripheral blood can be detected using flow cytometry and HLA-tetramers.³¹⁻³⁷ Here, we isolated HLA-DR-specific memory B cells from peripheral blood using HLA-DR tetramers for the subsequent generation of recombinant human HLA-DR mAbs. Subsequently, uniquely shared AA within 3-3.5 Å radius were deducted from SAB reactivity patterns and referred to as functional epitopes. These were also mapped to eplets, from which the reactive AAs are theoretically predefined. Overall, we present 5 recombinant human HLA-DR mAbs and antibody verification of 3 functional epitopes/eplets.

2 | MATERIALS AND METHODS

2.1 | Cells

Peripheral blood and serum samples were collected from healthy women (n = 3) who had developed HLA class II antibodies due to pregnancy, as detected with luminex single antigen bead (SAB) assays. All samples were collected with informed consent under guidelines issued by the medical ethics committee of Leiden University Medical Centre (Leiden, The Netherlands). Peripheral blood mononuclear cells (PBMC) were isolated by Ficoll-Paque (LUMC Pharmacy, Leiden, The Netherlands) density gradient centrifugation and kept frozen in liquid nitrogen until further use. HLA typed Epstein-Barr virus-transformed lymphoblastoid cell lines (EBV-LCLs) were cultured in Iscove's modified Dulbecco's medium (IMDM) (Gibco Invitrogen, Paisley, UK) containing 10% fetal bovine serum (FBS) (Sigma-Aldrich, Zwijndrecht, The Netherlands), 50 µM 2-mercaptoethanol (Sigma-Aldrich), 2 mM L-glutamine (Gibco Invitrogen), and 100 U/mL penicillin with 100 µg/mL streptomycin (Gibco Invitrogen) in T75 flasks (Greiner, Frickenhausen, Germany).

2.2 | HLA typing

All subjects were HLA typed for HLA-A, -B, -C, -DRB1, -DRB3/4/5, -DQB1, -DQA1, -DPB1, and -DPA1 loci by next-generation sequencing (NGS). Genomic DNA was automated bead-based isolated from PBMC (Chemagen, Perking Elmer, Baesweiler, Germany). NGSgo-AmpX kit (GenDx, Utrecht, The Netherlands) was used for the amplification of HLA genes. Next, library and sequence preparation were performed with NGSgo-LibrX/IndX kit (GenDx) and subsequent sequencing was carried on an Illumina MiniSeq (Illumina, San Diego, CA). NGS data were analyzed with NGSengine software version 2.11.0 (GenDx).

2.3 | HLA-DR-specific memory B cell isolation and expansion

After thawing, B cells were enriched from 40-60x10⁶ PBMC by negative selection using EasySep Human B cell enrichment kits (Stem Cell Technologies, Grenoble, France), according to the manufacturer's instructions (purity >95%). Enriched B cells were incubated for 45 minutes at 4°C with phycoerythrin (PE) and allophycocyanin (APC)-labeled HLA-DR tetramers (Table 1, ProImmune, Oxford, UK) and with the following mouse antihuman antibodies: CD3 (pacific blue, SP34-2), IgD (PE-Cy7, IA6-2) (both from BD Biosciences, Breda, The Netherlands), and CD27 (fluorescein isothiocyanate, FITC, CLB-CD27/1, 9F4) (Sanquin, Amsterdam, The Netherlands). A FACSAria III sorter (BD Biosciences) was used to sort CD3⁺CD27⁺IgD⁺ tetramer-APC⁺ and tetramer-PE⁺ cells at 1 cell per well in 96-well flat-bottom plates (Costar, Corning, NY), containing 100 000 irradiated (50Gy) CD40L-expressing EL4-B5 cells.³⁸ B cells were expanded for 13 days in IMDM containing 10% FBS,

TABLE 1 HLA-DR tetramers used for cell sorting

HLA allele	Peptide	Peptide sequence	Fluorochrome
DRB1*07:01/ DRA1*01:01	CMV	PDDYSNTHSTRYVTV	PE & APC
DRB1*01:01/ DRA1*01:01	Negative control/ CLIP	PVSKMRMATPLLMQA	PE & APC
DRB1*04:01/ DRA1*01:01 ¹	Negative control/ CLIP	PVSKMRMATPLLMQA	PE & APC
DRB1*04:05/ DRA1*01:01 ¹	Negative control/ CLIP	PVSKMRMATPLLMQA	PE & APC

¹DRB1*04:01 and DRB1*04:05 were used together in one sort.

supplemented with 50 μM 2-mercaptoethanol, 2 mM L-glutamine, 100 U/mL penicillin with 100 μg/mL streptomycin, 20 μg/mL insulin-transferrin-sodium selenite (Sigma-Aldrich), 50 ng/mL IL-21 (Gibco), 1 ng/mL IL-1β (Miltenyi, Leiden, The Netherlands), 0.3 ng/mL TNFα (Miltenyi), and 0.5 μg/mL R848 (toll-like receptor 7/8 agonist, resiquimod) (Sigma-Aldrich).³⁹

2.4 | HLA-specific antibody detection

After expansion, supernatants were tested for the presence of IgG by enzyme-linked immunosorbent assay (ELISA), as previously described,⁴⁰ after which IgG-positive supernatants were screened for the presence of HLA antibodies with Lifecodes Lifescreen Deluxe screening kit (LMX, Immucor Transplant Diagnostics, Stamford, CT). The specificity of the HLA antibodies in positive supernatants was determined by Lifecodes HLA class II SAB assays (Immucor). Serum samples were treated with ethylenediaminetetraacetic acid (6% EDTA) prior to testing. Data were analyzed with Match It! Antibody software version 1.3.0 (Immucor). The screening data were analyzed using raw mean fluorescence intensity (MFI), and for the SAB data background corrected MFI (BCM) was used.

2.5 | Production of recombinant human monoclonal antibodies

RNA was isolated from HLA-antibody positive B cell clones using TRIzol (ThermoFisher Scientific, Waltham, MA). Next, the genes encoding the variable heavy chain (VH) and variable light chain (VL) were obtained and recombinant monoclonal antibodies (mAbs) were generated and purified as previously described.⁴¹ Briefly, SMART cDNA synthesis and 5'-RACE polymerase chain reaction (PCR) were performed to obtain the PCR products of VH and VL, which were cloned into pcDNA3.3 expression vectors containing the constant domains of human IgG1 (IGHG1*03), and kappa (κ) (IGKC) or lambda (λ) (IGLC2*01). Recombinant mAbs were expressed by transient co-transfection of heavy and light chain vectors of Expi293F cells with SV40-LT plasmid,⁴² ExpiFectamine, Opti-Mem, and Expi293 expression medium (ThermoFisher Scientific). Further purification was done using Amicon ProAffinity Concentration Kit

Protein G (Merck Millipore, Burlington, MA). Concentrations of the purified mAbs were determined using the protein A280 protocol of Nanodrop2000 (ThermoFisher Scientific), yielding the molecular concentration of each mAb based on AA sequence.

2.6 | Sequence analysis

Plasmids were sequenced by Sanger sequencing (Macrogen, Amsterdam, The Netherlands) to obtain nucleotide sequence data of VH and VL. The sequence data were analyzed with IgBLAST⁴³ to define the V(D)J genes of the VH and VL domains and clonality of B cell clones.

2.7 | Flow cytometric crossmatch assays

EBV-LCLs, 0.5 × 10⁶ were incubated with 25 μL mAb or PBS for 30 minutes at room temperature (RT). Cells were washed 3 times with phosphate-buffered saline (PBS) supplemented with 0.1% bovine serum albumin (BSA, Sigma). Next, cells were stained with mouse antihuman CD3 (PE, SK7), CD19 (APC, HIB19, both from BD Biosciences), and rabbit antihuman IgG F(ab')₂ (FITC, Dako, Leiden, The Netherlands) for 30 minutes at 4°C in the dark. After washing with 0.1%BSA/PBS, cells were fixed with 1% paraformaldehyde. Data were acquired using an Accuri C6 flow cytometer (BD Biosciences) and analyzed using FlowJo V10 software (Ashland, OR).

2.8 | Complement-dependent cytotoxicity (CDC) assay

Terasaki plates (Greiner) were oiled and filled with 1 μL of supernatant containing the mAb of interest in triplicate. Then, 3000 EBV-LCLs were added to each well and incubated for 60 minutes at RT. Next, 5 μL rabbit complement (Inno-train, Kronberg, Germany) was added and incubated for 60 minutes at RT. To visualize cytotoxicity, 5 μL propidium iodide ink was added to each well, and after 15 minutes incubation in the dark at RT the plates were analyzed using a Patimed (Leica Microsystems, Amsterdam, The Netherlands).

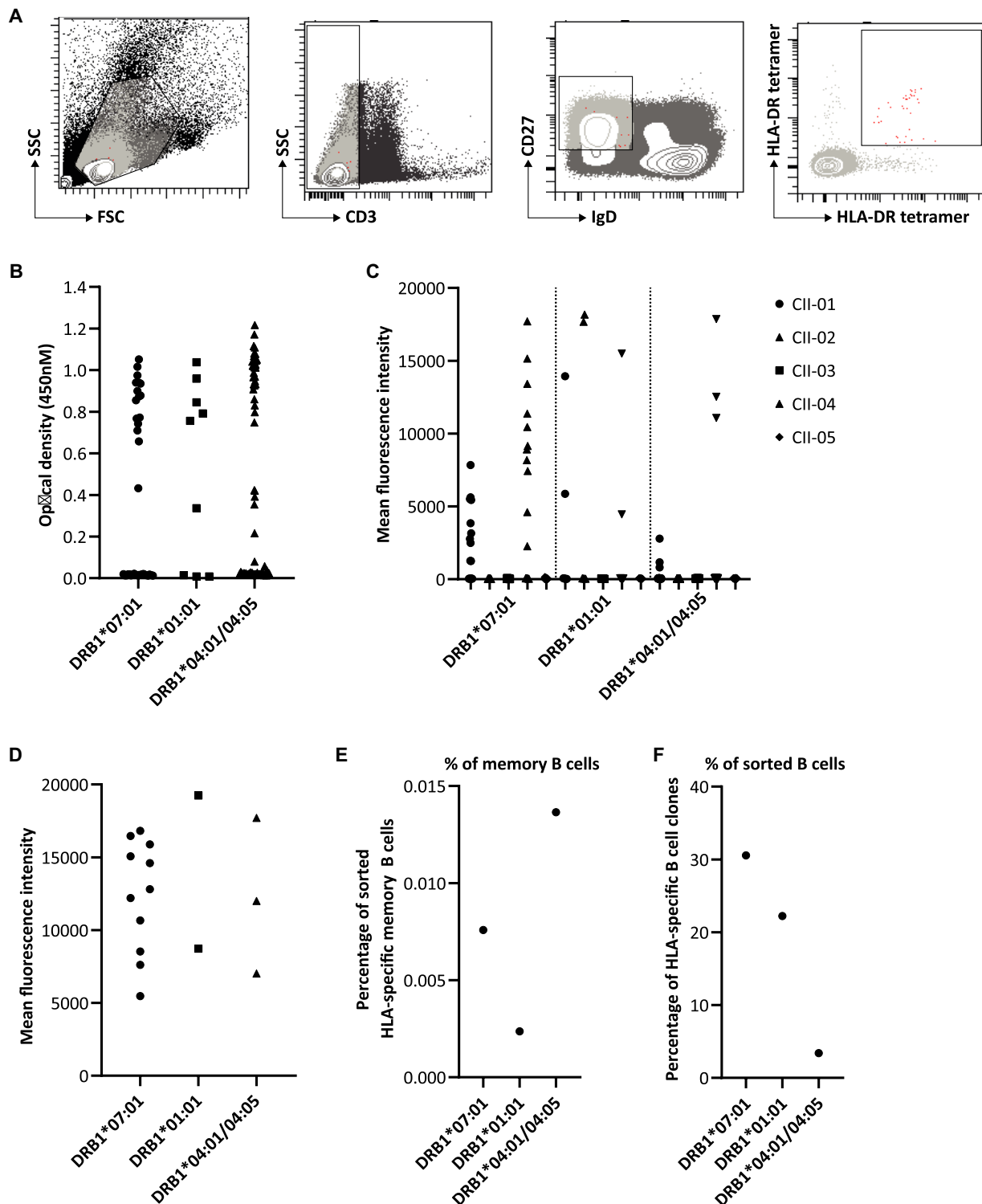


FIGURE 1 HLA-DR-specific memory B cell clones isolated from peripheral blood. **A**, Representative example of 3 independent experiments depicting the flow cytometry gating strategy to live single cell sort CD3⁺CD27⁺IgD⁺HLA-DR tetramer double positive B cells from PBMC. **B**, IgG antibody production by the clones was determined by ELISA. **C**, IgG positive clones were screened with HLA class II Lifecodes Lifescreen Deluxe kit to detect HLA antibody. The kit contains 5 groups of HLA class II beads and each data point represents a single bead group. **D**, HLA-specific B cell clones were tested with SAB assays to confirm tetramer specificity used for cell sorting. Each dot presents 1 clone and only MFI of bead with tetramer specificity is depicted. **E**, Percentage of sorted HLA-specific B cells from total memory B cells. **F**, Percentage of HLA antibody-producing B cell clones from sorted B cells. On the x-axis are the specificity of the tetramers used depicted. FSC, forward scatter; MFI, mean fluorescence intensity; PBMC, peripheral blood mononuclear cells; SAB, single antigen beads; SSC, side scatter [Color figure can be viewed at wileyonlinelibrary.com]

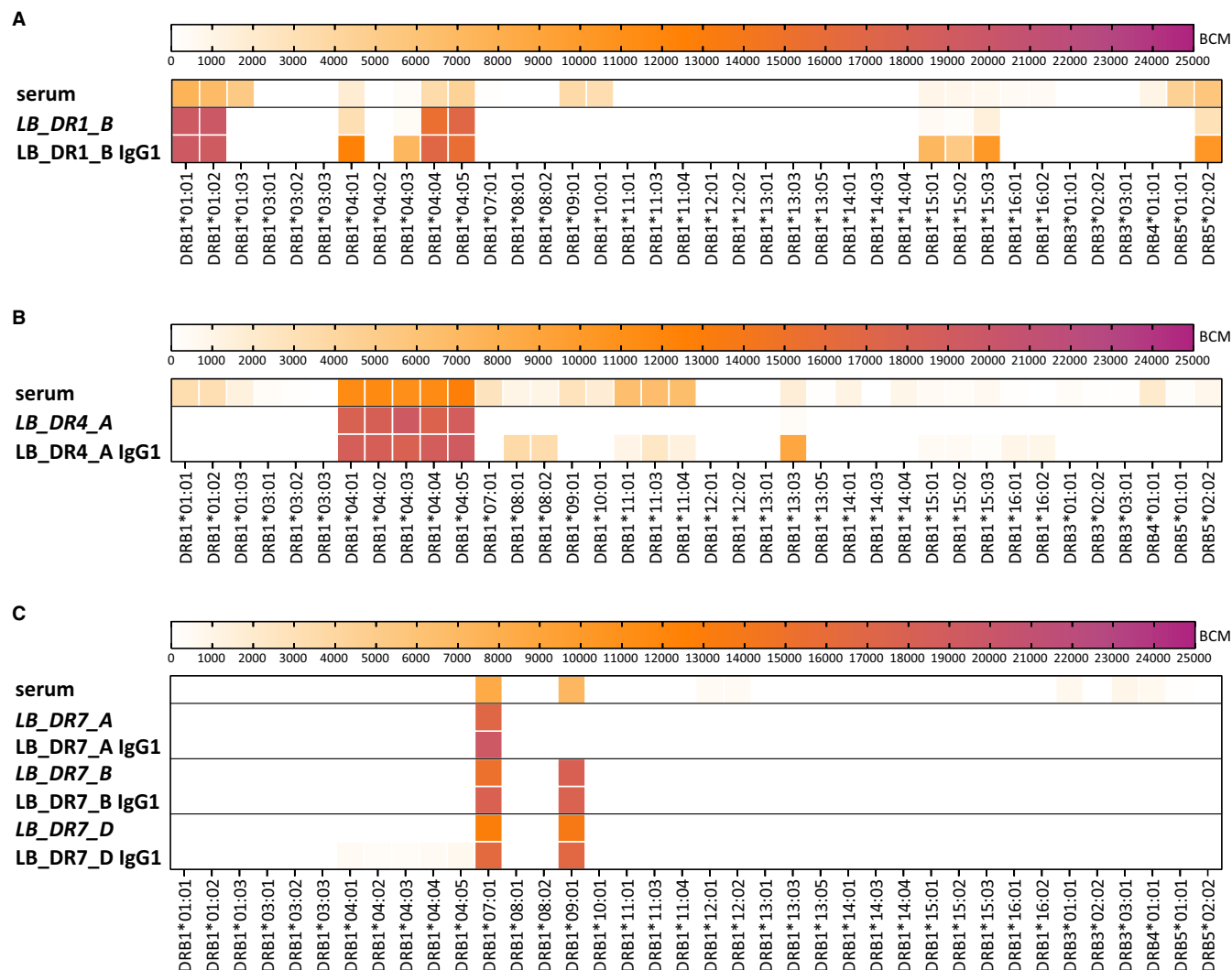


FIGURE 2 Recombinant human HLA-DR monoclonal antibodies have the same reactivity as the B cell clones. HLA-DR specificities in serum, supernatants of B cell clones (italic) and generated IgG1 mAbs. A, Serum of individual used for HLA-DRB1*01:01 sort and the generated LB_DR1_B mAb and with its respective B cell clone. B, Serum of individual used for HLA-DRB1*04:01/04:05 sort and the generated LB_DR4_A mAb and its respective B cell clone. C, Serum of individual used for HLA-DRB1*07:01 sort and the generated LB_DR7 mAbs with their respective B cell clones. Only DRB1/3/4/5 beads are shown as all other loci were negative for the B cell clones and mAbs. Purified recombinant monoclonal antibody concentration tested was 62.5 nM. BCM, background corrected mean fluorescence intensity; mAbs, monoclonal antibodies

2.9 | Antibody reactivity pattern analyses of mAbs

AA sequences of HLA alleles present in the Immucor SAB assay were obtained from IPD-IMGT/HLA (<https://www.ebi.ac.uk/ipd/imgt/hla/> accessed January 2019), in order to define shared AA of the reactive HLA alleles. To determine the eplets present on the reactive HLA alleles, reactivity patterns were analyzed with HLAMatchmaker (HLA DRDQDP Matching, version v2.0 and v3.0; <http://www.epitopes.net/>). Eplet antibody-verification status was extracted from <http://www.EpRegistry.com.br> (accessed on July 15, 2019 and February 12, 2020).

The positions of uniquely reactive AA were visualized with Swissviewer⁴⁴ using the following HLA-DR crystal structures: Protein Data Bank (PDB) 3PDO and 4MD4 (downloaded from

<https://www.rcsb.org/> on February 4, 2019). Swissviewer allows for the distance between 2 atoms to be estimated as well as for the display of atoms that are at a certain distance from a specific atom. These options were used to determine whether AAs were within 3-3.5 Å or 15 Å radius of each other.

3 | RESULTS

3.1 | HLA-DR-specific memory B cell clones isolated from peripheral blood

Flow cytometric cell sorting of HLA-specific memory B cells using HLA-DR-specific tetramers (Figure 1A) yielded an average of 44

(range 9-88) single memory B cells. After B cell expansion, IgG could be detected in 50.7% (range 40.9%-66.7%) of sorted wells with a wide concentration range (Figure 1B). HLA class II antibodies were present in 36.8% (range 8.3%-68.8%) of the IgG positive B cell clones with a wide MFI range (MFI 811-18168) (Figure 1C). Subsequent SAB assays confirmed that the HLA-specific B cell clones produced antibodies with the same specificity as the tetramers used for cell sorting (Figure 1D). Eventually, from the total pool of memory B cells an average of 0.008% (range 0.002%-0.014%) HLA-specific B cells were acquired (Figure 1E) and 18.7% (range 3.4%-30.6%) of the sorted cells produced HLA antibodies after expansion (Figure 1F). Overall, an average of 5 (range 2-11) HLA antibody-producing B cell clones were obtained, which is an average of 0.001% of memory B cells and 0.0002% of total B cells.

3.2 | Recombinant human HLA-DR antigen-reactive mAbs generated from HLA positive B cell clones

From several HLA-specific B cell clones HLA-DR antigen-reactive mAbs were generated, and in this proof of principle study we describe a DR1 mAb (LB_DR1_B), a DR4 mAb (LB_DR4_A) and 3 DR7 mAbs (LB_DR7_A, B and D). The specificity of these mAbs was confirmed by SAB analysis (Figure 2). As expected, the HLA-DR antigen-reactive mAbs showed almost identical reactivity to the supernatants of the B cell clones they were derived from. Flow cytometric crossmatches and CDC assays with EBV-LCL lines expressing

HLA alleles corresponding to the tetramers used for B cell isolation confirmed binding of the mAbs to their physiologically expressed HLA target (Figure S1A-C), as well as their cytotoxicity capacity (Figure S2A-C). Additionally, the mAbs also bound to other natively expressed HLA alleles that were reactive in SAB assays, whereas no binding was observed for nonreactive HLA alleles (Figure S1).

3.3 | Reactivity analysis of LB_DR1_B mAb

Next, we analyzed the mAb reactivity patterns to determine if the reactive HLA alleles in SAB assays uniquely share AA within a 3-3.5 Å radius acting as the functional epitope, determining the antibody specificity.¹¹⁻¹³ Furthermore, we analyzed whether additional AA configurations within 15 Å radius of the functional epitope were an absolute requirement for the interaction between antibody and HLA alleles.⁴⁵

The HLA type of the immunizer of LB_DR1_B was unknown (Table 2). This mAb has a broad reactivity pattern including DRB1*01:01 and DRB1*01:02, but not DRB1*01:03 (Figure 2A). Interestingly, no individual AA at a specific position was uniquely shared between reactive HLA alleles and absent on nonreactive HLA alleles, but the combination of 70 glutamine (Q), and 73 alanine (A) was present only on the reactive alleles (Figure 3A). Indeed, HLAMatchmaker v3.0 also showed that the reactive alleles share eplet 70QA (70Q 73A). These AAs are located on top of the HLA molecule (Figure 3B) and within 3 Å radius of each other, suggesting

TABLE 2 Information of the 5 human HLA-DR monoclonal antibodies and description of the reactive HLA-DR alleles

Human mAb	HLA-DR antibody producer	HLA immunizer	HLA tetramer	Reactive HLA-DRB1/3/4/5 alleles	On reactive HLA-DRB1/3/4/5 alleles		
					Amino acids ¹	Eplet	TerEp
LB_DR1_B	DRB1*13:01 DRB1*13:02 DRB3*02:02 DRB3*03:01		DRB1*01:01	DRB1*01:01 DRB1*01:02 DRB1*04:01 DRB1*04:03 DRB1*04:04 DRB1*04:05 DRB1*15:01 DRB1*15:02 DRB1*15:03 DRB5*02:02	70Q 73A (71R 74A)	70QA	
LB_DR4_A	DRB1*03:01 DRB1*13:01 DRB3*01:01 DRB3*02:02	DRB1*04:04	DRB1*04:01/04:05	DRB1*04:01 DRB1*04:02 DRB1*04:03 DRB1*04:04 DRB1*04:05 DRB1*13:03 DRB1*08:01 DRB1*08:02 DRB1*11:03 DRB1*11:04 DRB1*11:01 DRB1*16:01 DRB1*16:02 DRB1*15:01 DRB1*15:02 DRB1*15:03	31F 32Y 37Y/S (13H 33H)	37YV	
LB_DR7_A	DRB1*11:01 DRB1*15:01 DRB3*02:02 DRB5*01:01		DRB1*07:01	DRB1*07:01	14K 25Q 11G 30L	25Q	#1008, #1405, #1602
LB_DR7_B & LB_DR7_D	DRB1*11:01 DRB1*15:01 DRB3*02:02 DRB5*01:01		DRB1*07:01	DRB1*07:01 DRB1*09:01	78V 96H 98E 120S	77TV 98ES	#1029

¹Amino acids in parentheses are present on the highly reactive HLA alleles, amino acids in italic are not exposed.

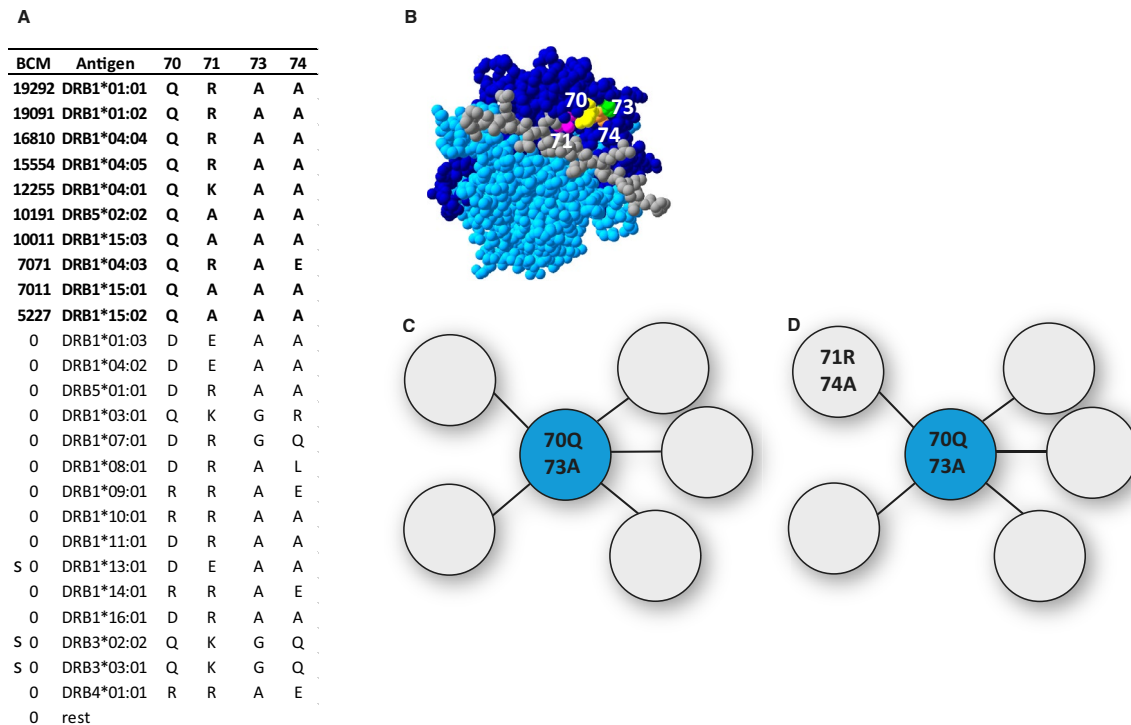


FIGURE 3 Reactivity analysis of LB_DR1_B monoclonal antibody. A, Comparison of the amino acid positions of interests of the reactive HLA-DR alleles of LB_DR1_B mAb and a selection of the nonreactive HLA-DR alleles. B, Locations of amino acid 70Q (yellow), 71R (magenta), 73A (green), and 74A (orange) are indicated on crystal structure of DRB1*01:01 (PDB: 3PDO). C, LB_DR1_B mAb interacts with HLA-DR alleles containing the functional epitope 70Q 73A. D, Schematic representation of the footprint of LB_DR1_B mAb that is highly reactive for HLA-DR alleles containing the functional epitope 70Q 73A (cyan) and additional amino acids 71R and 74A. In crystal structures the β chain is colored dark blue, α chain light blue, and peptide is gray. Purified recombinant monoclonal antibody concentration tested was 62.5nM. BCM, background corrected mean fluorescence intensity, negative values are presented as zero. mAbs, monoclonal antibodies; PDB, Protein Data Bank; s, self HLA alleles of antibody-producer

that 70Q and 73A are comprising the functional epitope (Figure 3C). This is in accordance with cellular assays, as LB_DR1_B binds only to cells expressing HLA alleles containing 70Q and 73A (Figure S1).

Some of the reactive alleles showed a lower MFI in SAB analysis, suggesting that additional AAs are involved in binding and affinity. Indeed, the alleles showing the highest MFI values share arginine (R) on position 71 and 74A, which are located near positions 70 and 73, within the area of the functional epitope. DRB1*04:03 also harbors a 71R but lacks 74A, which might explain the lower MFI values against this allele. Together, these data suggest that all 4 AAs are involved in binding of LB_DR1_B to HLA alleles with high MFI (Figure 3D). As the identified functional epitope corresponds to eplet 70QA, the latter can be considered as antibody verified by LB_DR1_B.

3.4 | Reactivity analysis of LB_DR4_A mAb

LB_DR4_A mAb showed a broad reactivity pattern in SAB assays with high reactivity observed for all included DR4 alleles, whereas 11 other alleles were reactive with low MFI values (Figures 2B and 4A). From the AA mismatches of the immunizing DRB1*04:04 with the HLA-DR constitution of antibody producer (DRB1*03:01

DRB1*13:01 DRB3*01:01 DRB3*02:02), only tyrosine (Y) on position 32 was shared by all reactive HLA alleles. However, 32Y is also present on nonreactive HLA alleles, suggesting that other AAs are involved in interaction with this mAb (Figure 4A). HLAMatchmaker v3.0 identified eplet 37YV (37Y 38V), which was present on 11 out of 16 reactive alleles. The other identified eplets were 96Y (96Y) present on all tested DR4 alleles, and 142M (142M) shared by 5 reactive HLA alleles with lowest MFI values in the positive range. These eplets are likely not involved in binding of this mAb since they are shared by a limited number of reactive alleles.

To identify the AA configuration involved, we analyzed the DRB1*04:01 crystal structure and observed that 37Y is located within 4 Å radius of 32Y, whereas 38 valine (V) is 6 Å away from 32Y and not surface exposed (Figure 4B). In addition, 31 phenylalanine (F) is located within 3.5 Å radius of 32Y, and also 37Y is located within 3.5 Å radius of 31F. Interestingly, the previous version of HLAMatchmaker (v2.0) identified eplet 31FYY (31F 32Y 37Y) present on the same 11 reactive alleles as defined for eplet 37YV. Therefore, we deduced that positions 31F, 32Y, and 37Y together form the functional epitope, and indeed this configuration is unique for the reactive HLA alleles, but only for the highly reactive HLA alleles. Five of the lower reactive HLA alleles have a serine (S) instead of 37Y, and the combination of 31F, 32Y, and 37S is absent on the

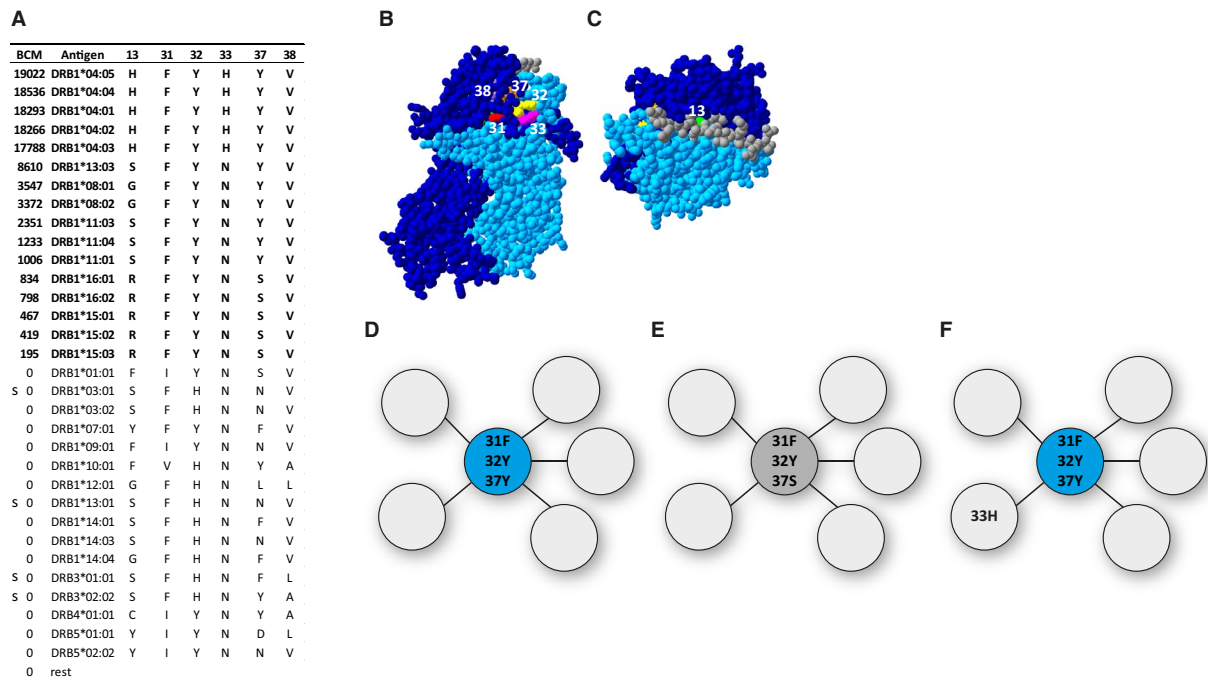


FIGURE 4 Reactivity analysis of LB_DR4_A monoclonal antibody. A, Comparison of the amino acid positions of interests of the reactive HLA-DR alleles of LB_DR4_A mAb and a selection of the nonreactive HLA-DR alleles. B, Locations of amino acid 31F (red), 32Y (yellow), 33H (magenta), 37Y (orange), 38V (lilac), and (C) 13H (green) are indicated on crystal structure of DRB1*04:01 (PDB: 4MD4). D, Schematic representation of the footprint of LB_DR4_A mAb interacting with the functional epitope 31F 32Y 37Y (cyan) (E) LB_DR4_A mAb weakly binds to HLA alleles containing 31F 32Y 37S. F, Schematic representation of the footprint of LB_DR4_A mAb interacting with the highly reactive HLA-DR4 alleles. In crystal structures the β chain is colored dark blue, α chain light blue, and peptide is gray. Purified recombinant monoclonal antibody concentration tested was 62.5nM. BCM, background corrected mean fluorescence intensity, negative values are presented as zero. mAbs, monoclonal antibodies; PDB, Protein Data Bank; s, self HLA alleles of antibody-producer

nonreactive HLA alleles. CDC assays showed that LB_DR4_A mAb can lyse cells expressing HLA alleles carrying 31F 32Y 37Y (Figure S2C,D), whereas no specific lysis was observed for cells expressing HLA alleles with 31F 32Y 37S (Figure S2F-H).

Because 32Y was the AA shared by all reactive HLA alleles and mismatched with the antibody producer, we deduce that the functional epitope of LB_DR4_A consists of 31F 32Y 37Y (Figure 4D) with the mAb weakly binding to HLA alleles containing 37S instead of 37Y (Figure 4E). The stronger interaction observed for the DR4 alleles suggests involvement of AAs solely present on DR4 alleles, which are 96Y, 180 leucine (L), and histidine (H) on position 13 and 33. The latter 2 AAs are within 15 Å radius of 32Y, but only 33 is exposed and is therefore most likely involved in the interaction with the antibody (Figure 4F). The functional epitope, 31F 32Y 37Y, corresponds to eplet 31FYY and thereby this eplet can be considered as antibody verified by LB_DR4_A.

3.5 | Reactivity analysis of LB_DR7 mAbs

LB_DR7 mAbs were obtained from an individual of which the immunizing event was unknown. We analyzed 3 LB_DR7 mAbs from which the variable domains were acquired by sequencing (Table 3), showing different V(D)J usage, and unique VH and VL clonotypes.

This indicates that memory B cells with B cell receptors recognizing different AA configurations can be isolated with a single tetramer specificity.

LB_DR7_A is only reactive with HLA-DRB1*07:01 (Figure 2C), strongly suggesting that this was the immunizing allele. Upon comparing the AA sequence of DRB1*07:01 with the nonreactive HLA-DR alleles present in SAB assay, glycine (G) on position 11, lysine (K) on position 14, Q on position 25, and L on position 30 were identified as unique AAs for DRB1*07:01 (Figure 5A). Three of these AAs are present within the eplet 25Q (25Q 30L 14K), which is also the eplet determined upon analysis with HLAMatchmaker v3.0. The 4 unique AAs correspond to TerEp #1602^{22,46} and have been previously described for mouse mAbs.⁴⁷⁻⁵⁰ Positions 11 and 30 are located at the bottom of the peptide-binding groove (Figure 5B), while 14K and 25Q are surface exposed and within 3.5 Å radius of each other (Figure 5C). Due to location of 11G and 30L and as neither are within 3.5 Å radius of 14K and/or 25Q, it is unlikely that those form the functional epitope. Additionally, mutation assays with mouse mAbs showed that only mutation of 14K and 25Q affected binding.⁴⁷⁻⁵⁰ Altogether, we suggest that 14K and 25Q comprise the functional epitope without 30L being involved (Figure 5D).

LB_DR7_B and LB_DR7_D bind to DRB1*07:01 and DRB1*09:01 in both SAB assays (Figure 2C) and flow crossmatch,

TABLE 3 V(D)J usage of the different HLA-DR monoclonal antibodies

Clone	HLA	Heavy chain			Light chain		
		V gene	D gene	J gene	κ or λ	V gene	J gene
LB_DR1_B	DRB1*01:01	4-59	3-10	6	λ	3-1	2 or 3 ¹
LB_DR4_A	DRB1*04:01/04:05	3-48	3-16 or 6-6 ¹	4	λ	1-40	1
LB_DR7_A	DRB1*07:01	4-61	6-13 or 6-25 or 6-6 ¹	4	λ	2-14	2 or 3 ¹
LB_DR7_B	DRB1*07:01	3-30	3-10	4	κ	1-5	1
LB_DR7_D	DRB1*07:01	3-30	6-19	6	κ	1-16	2

¹Multiple equivalent top matches.

whereas HLA alleles with low reactivity to LB_DR7_D in SAB did not react with natively expressed alleles in flow (Figure S1). DRB1*07:01 and DRB1*09:01 share a valine (V) on position 78, which is absent on all nonreactive HLA alleles (Figure 5E) and located on top of the molecule (Figure 5F) and correspond to TerEp #1029. Concomitantly, eplets 77TV (77T 78V) and 98ES (98E 120S) were identified as unique for the reactive alleles. Analysis with HLA-Matchmaker v2.0 suggested that 96H could potentially be involved, based on previously listed eplet 78V₂, which was the predecessor of 77TV and 98ES. Interestingly, although the individual AAs 96H, 98E, and 120S are present on nonreactive HLA alleles, including self, the configuration of the 3 is only present on DRB1*07:01 and DRB1*09:01. Because this configuration is exposed but not within 15 Å radius of 78V (Figure 5G), either 78V (Figure 5H) or 96H 98E 120S (Figure 5I) act as contact site for the CDR-H3 of LB_DR7_B and/or LB_DR7_D.

4 | DISCUSSION

Increasing numbers of HLA class II eplet mismatches are associated with the development of *dn*DSA,⁶⁻⁸ which led to the hypothesis that eplet mismatch loads can be used as predictor of DSA occurrence.^{9,10} However, eplets have been theoretically defined and for several eplets it remains to be established whether they are indeed reactive with antibodies and thus clinically relevant. Therefore, eplets require experimental verification, either by human mAbs or absorption and elution studies,^{4,5,19-21} to establish if interaction with an antibody can occur. This is of importance for the implementation of eplet matching in allocation systems aiming at prevention of *dn*DSA development. By performing eplet matching solely on relevant functional eplets, patients will not be denied an organ offer based on irrelevant eplet disparities with the donor.

In this study, we isolated HLA-DR-specific memory B cells from peripheral blood of immunized individuals using HLA-DR-specific tetramers. Although tetramers have been used to detect and isolate HLA class I-specific B cells,^{35,37,51} to our knowledge this study is the first to use tetramers for the isolation of HLA class II-specific memory B cells. Here, we describe generation of 5 HLA-DR mAbs with 4 different specificities: DR7, DR7/DR9, DR1/DR9/DR10/DR51, and DR4/DR1303/DR8/DR11/DR15/DR16.

Overall, the specificity of the generated mAbs resembled the antibody repertoire observed in the serum. For LB_DR7_D additional reactive HLA alleles were observed, albeit with very low MFI. A possible explanation is that the memory B cell compartment may contain a broader repertoire than that of circulating antibodies.⁵²⁻⁵⁴ However, the additional reactive HLA alleles could not be confirmed with flow cytometric crossmatch assays using natively expressed HLA alleles (Figure S1). Therefore, the additional reactivity for the mAbs appears to be due to nonspecific binding in SAB assays or due to the mAb concentration used.

Based on both SAB and cellular data presented here, the eplets 70QA and 31FYY, corresponding to AA configurations

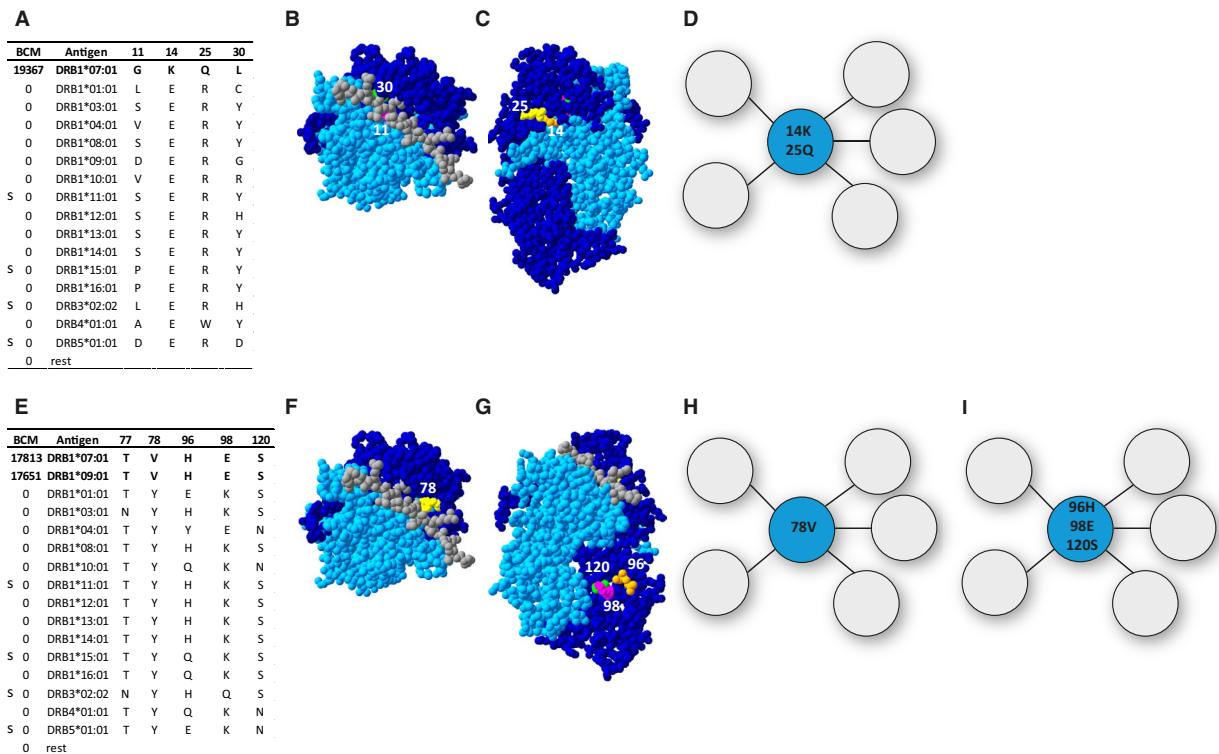


FIGURE 5 Reactivity analyses of LB_DR7_A, and LB_DR7_B and LB_DR7_D monoclonal antibodies. A, Comparison of the amino acids of the reactive HLA-DR alleles of LB_DR7_A mAb with nonreactive HLA-DR alleles. B, Positions 30 (green), and 11 (magenta) and (C) 25 (yellow), and 14 (orange) are indicated on the crystal structure of DRB1*01:01 (PDB: 3PDO). D, A schematic representation of the footprint of LB_DR7_A mAb with 14K 25Q as functional epitope (cyan). E, Comparison of the amino acids of the reactive HLA-DR alleles of LB_DR7_B and LB_DR7_D with the nonreactive HLA-DR alleles, of which only a selection is shown. Only BCM of LB_DR7_B are depicted. F, Location of position 78 (yellow) and (G) of 96 (orange), 98 (magenta), and 120 (green) on the DRB1*01:01 crystal structure. H, A schematic representation of LB_DR7_B and LB_DR7_D footprint with 78V or (I) with 96H 98E 120S as the functional epitope (cyan) of the mAb. In crystal structures the β chain is colored dark blue, α chain light blue, and peptide is gray. Purified recombinant monoclonal antibody concentration tested was 62.5nM. BCM, background corrected mean fluorescence intensity, negative values are presented as zero. mAbs, monoclonal antibodies; PDB, Protein Data Bank; s, self HLA-DR alleles of antibody-producer

70Q 73A and 31F 32Y 37Y, have been antibody verified by the human mAbs LB_DR1_B and LB_DR4_A, respectively, despite the limitation of missing HLA typing of the immunizer for LB_DR1_B. Eplet 31FFY was previously registered as antibody verified based on reactivity analysis of mouse mAbs^{5,55} and pregnancy sera.²⁵ Peculiarly, this eplet is no longer present on the HLA Epitope Registry as it was deemed redundant. The data presented here suggest that this eplet does actually exist and should be relisted on the Registry.

For the narrow reactivity patterns of LB_DR7_A, LB_DR7_B, and LB_DR7_D HLAMatchmaker defined eplets 25Q, and 77TV and 98ES, respectively, to be uniquely shared. For eplet 25Q the current description of the AAs involved exceeds the original definition of an eplet since the suggested residues are not located within 3.5 Å radius of each other. The same applies to eplet 37YV, defined by HLAMatchmaker v3.0 for LB_DR4_A. Provided that eplet 25Q (25Q 14K 30L) is to be redefined as 14K and 25Q, mAb LB-DR7_A allows for antibody verification of this eplet.

Eplet 98ES is currently listed as antibody verified based on pregnancy sera, but our reactivity analysis shows that either 78V or 96H 98E 120S can act as contact site for mAb

reactive for HLA-DRB1*07:01 and HLA-DRB1*09:01. In contrast to HLAMatchmaker v2.0, the newly defined 98ES no longer includes residue 96H in v3.0. Reactivity analysis of the HLA-DR mAbs showed that AAs on reactive HLA alleles not always correspond to predefined eplets identified by HLAMatchmaker. For some of these mAbs we were able to identify different eplets on basis of version 2.0 of HLAMatchmaker, indicating that the list of eplets in this program and on the HLA Epitope Registry is subject to change without broadly accepted and validated arguments showing the need to install an international nomenclature committee for the definition of antibody-verified eplets and/or epitopes. Overall, the data presented herein indicate that the current, widely used list of eplets contains inaccuracies. Furthermore, our results show that performing reactivity analysis of human HLA mAbs based on AA rather than on predefined eplets may be more useful in defining the relevant AA configurations, and this will require several mAbs.^{18,56}

Antibody reactivity analyses based solely on SAB assay can be complex and can benefit from additional functional assays to determine true reactivity. AA substitutions within the functional epitope that do not affect binding of mAbs to an HLA allele in SAB assay may affect the ability to induce complement-dependent cytotoxicity,¹⁷

as was the case for LB_DR4_A. In addition, MFI values can reflect differential affinity of the mAbs for specific HLA alleles⁵⁷ and AA substations within the structural epitope can lead to lower affinity, which can be reflected in the MFI values.⁵⁸ Mutation studies have been informative on determining the involvement of single AAs in the interaction between the HLA molecule and antibody.^{48,49,59} However, it is important to realize that AA substitutions can affect the tertiary structure and surface electrostatic potential of the HLA molecule.^{60,61}

In the present study we obtained multiple B cell clones from one individual with subtle differences in specificity reflecting the polyclonal reactivity of serum. These observations substantiate the notion that antibody verification of eplets should be done only by using human mAbs or absorption and elution studies^{22,62-64} and not based on sera of women after first or second pregnancy, as is currently done for various eplets.²⁴⁻²⁶ Although in this proof of principle study, we present HLA-DR mAbs obtained from 3 subjects, the inventory of HLA-DR mAbs will expand soon, which will result in identification and antibody verification of additional relevant AA configurations. In addition, we are developing methods to utilize HLA-DQ monomers³³ to isolate HLA-DQ-specific memory B cells for the generation of recombinant HLA-DQ mAbs and subsequently reactivity analysis to verify HLA-DQ eplets, because HLA-DQ DSA are most prevalent after transplantation and associated with rejection.^{8,9,65-67}

These human HLA class II mAbs can be used in functional studies to provide more insight in the respective roles of HLA-specific IgG antibodies in causing graft damage,⁶⁸⁻⁷² with the possibility of all IgG subclasses to be generated.⁴¹ In addition, as shown for LB_DR7, distinct B cell clones with different levels of affinity maturation, as suggested by the different binding strengths and efficacy in cell lysis, can be obtained from a single individual. Thus, the method described herein can contribute to understanding the development of the HLA-specific memory B cell compartment⁷³ besides their use in eplet verification.

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DISCLOSURE

The authors of this manuscript have no conflicts of interest to disclose as described by the *American Journal of Transplantation*.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

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REFERENCES

1. Loupy A, Lefaucheur C, Vernerey D, et al. Complement-binding anti-HLA antibodies and kidney-allograft survival. *N Engl J Med*. 2013;369(13):1215-1226.
2. Duquesnoy RJ, Askar M. HLA-Matchmaker: a molecularly based algorithm for histocompatibility determination. V. Eplet matching for HLA-DR, HLA-DQ, and HLA-DP. *Hum Immunol*. 2007;68(1):12-25.
3. Duquesnoy RJ. A structurally based approach to determine HLA compatibility at the humoral immune level. *Hum Immunol*. 2006;67(11):847-862.
4. Duquesnoy RJ, Marrari M, Mulder A, da Mata Sousa LCD, da Silva AS, do Monte SJH. First report on the antibody verification of HLA-ABC epitopes recorded in the website-based HLA Epitope Registry. *Tissue Antigens*. 2014;83(6):391-400.
5. Duquesnoy RJ, Marrari M, Tambur AR, et al. First report on the antibody verification of HLA-DR, HLA-DQ and HLA-DP epitopes recorded in the HLA Epitope Registry. *Hum Immunol*. 2014;75(11):1097-1103.
6. Kosmoliaptis V, Mallon DH, Chen Y, Bolton EM, Bradley JA, Taylor CJ. Alloantibody Responses After Renal Transplant Failure Can Be Better Predicted by Donor-Recipient HLA Amino Acid Sequence and Physicochemical Disparities Than Conventional HLA Matching. *Am J Transplant*. 2016;16(7):2139-2147.
7. Lachmann N, Niemann M, Reinke P, et al. Donor-Recipient Matching Based on Predicted Indirectly Recognizable HLA Epitopes Independently Predicts the Incidence of De Novo Donor-Specific HLA Antibodies Following Renal Transplantation. *Am J Transplant*. 2017;17(12):3076-3086.
8. Wiebe C, Pochinco D, Blydt-Hansen TD, et al. Class II HLA epitope matching-A strategy to minimize de novo donor-specific antibody development and improve outcomes. *Am J Transplant*. 2013;13(12):3114-3122.
9. Snanoudj R, Kamar N, Cassuto E, et al. Epitope load identifies kidney transplant recipients at risk of allosensitization following minimization of immunosuppression. *Kidney Int*. 2019;95(6):1471-1485.
10. Wiebe C, Kosmoliaptis V, Pochinco D, et al. HLA-DR/DQ molecular mismatch: A prognostic biomarker for primary alloimmunity. *Am J Transplant*. 2019;19(6):1708-1719.
11. Amit AG, Mariuzza RA, Phillips SE, Poljak RJ. Three-dimensional structure of an antigen-antibody complex at 2.8 Å resolution. *Science (New York, NY)*. 1986;233(4765):747-753.
12. Ippolito GC, Schelonka RL, Zemlin M, et al. Forced usage of positively charged amino acids in immunoglobulin CDR-H3 impairs B cell development and antibody production. *J Exp Med*. 2006;203(6):1567-1578.
13. Xu JL, Davis MM. Diversity in the CDR3 region of V(H) is sufficient for most antibody specificities. *Immunity*. 2000;13(1):37-45.
14. MacCallum RM, Martin AC, Thornton JM. Antibody-antigen interactions: contact analysis and binding site topography. *J Mol Biol*. 1996;262(5):732-745.
15. Padlan EA. Anatomy of the antibody molecule. *Mol Immunol*. 1994;31(3):169-217.
16. Sela-Culang I, Kunik V, Ofra Y. The structural basis of antibody-antigen recognition. *Front Immunol*. 2013;4:302-302.
17. Duquesnoy RJ, Marrari M, Jelenik L, Zeevi A, Claas FH, Mulder A. Structural aspects of HLA class I epitopes reacting with human monoclonal antibodies in Ig-binding, C1q-binding and lymphocytotoxicity assays. *Hum Immunol*. 2013;74(10):1271-1279.
18. Duquesnoy RJ, Marrari M, Mulder A, Claas FH, Mosteck J, Balazs I. Structural aspects of human leukocyte antigen class I epitopes detected by human monoclonal antibodies. *Hum Immunol*. 2012;73(3):267-277.
19. Duquesnoy RJ, Mulder A, Askar M, Fernandez-Vina M, Claas FH. HLA-Matchmaker-based analysis of human monoclonal antibody reactivity demonstrates the importance of an additional contact site

- for specific recognition of triplet-defined epitopes. *Hum Immunol.* 2005;66(7):749-761.
20. El-Awar N, Lee JH, Tarsitani C, Terasaki PI. HLA class I epitopes: recognition of binding sites by mAbs or eluted alloantibody confirmed with single recombinant antigens. *Hum Immunol.* 2007;68(3):170-180.
 21. El-Awar N, Nguyen A, Almeshari K, et al. HLA class II DQA and DQB epitopes: recognition of the likely binding sites of HLA-DQ alloantibodies eluted from recombinant HLA-DQ single antigen cell lines. *Hum Immunol.* 2013;74(9):1141-1152.
 22. El-Awar N, Terasaki PI, Cai J, et al. Epitopes of HLA-A, B, C, DR, DQ, DP and MICA antigens. *Clin Transpl.* 2009;295-321.
 23. Duquesnoy RJ, Marrari M, Marroquim MS, et al. Second update of the International Registry of HLA Epitopes. I. The HLA-ABC Epitope Database. *Hum Immunol.* 2019;80(2):103-106.
 24. Duquesnoy RJ, Honger G, Hosli I, Marrari M, Schaub S. Antibody-defined epitopes on HLA-DQ alleles reacting with antibodies induced during pregnancy and the design of a DQ eplet map. *Hum Immunol.* 2016;77(10):824-831.
 25. Duquesnoy RJ, Honger G, Hosli I, Marrari M, Schaub S. Identification of epitopes on HLA-DRB alleles reacting with antibodies in sera from women sensitized during pregnancy. *Hum Immunol.* 2016;77(2):214-222.
 26. Duquesnoy RJ, Honger G, Hosli I, Marrari M, Schaub S. Detection of newly antibody-defined epitopes on HLA class I alleles reacting with antibodies induced during pregnancy. *Int J Immunogenet.* 2016;43(4):200-208.
 27. Sapir-Pichhadze R, Zhang X, Ferradji A, et al. Epitopes as characterized by antibody-verified eplet mismatches determine risk of kidney transplant loss. *Kidney Int.* 2019;97(4):778-785.
 28. de Wildt RM, Steenbakkers PG, Pennings AH, van den Hoogen FH, van Venrooij WJ, Hoet RM. A new method for the analysis and production of monoclonal antibody fragments originating from single human B cells. *J Immunol Methods.* 1997;207(1):61-67.
 29. Liao H-X, Levesque MC, Nagel A, et al. High-throughput isolation of immunoglobulin genes from single human B cells and expression as monoclonal antibodies. *J Virol Methods.* 2009;158(1-2):171-179.
 30. Wrammert J, Smith K, Miller J, et al. Rapid cloning of high-affinity human monoclonal antibodies against influenza virus. *Nature.* 2008;453(7195):667-671.
 31. Han M, Rogers JA, Lavingia B, Stastny P. Peripheral blood B cells producing donor-specific HLA antibodies in vitro. *Hum Immunol.* 2009;70(1):29-34.
 32. Heidt S, Roelen DL, de Vaal YJH, et al. A NOVEL ELISPOT assay to quantify HLA-specific B cells in HLA-immunized individuals. *Am J Transplant.* 2012;12(6):1469-1478.
 33. Karahan GE, de Vaal YJ, Roelen DL, Buchli R, Claas FH, Heidt S. Quantification of HLA class II-specific memory B cells in HLA-sensitized individuals. *Hum Immunol.* 2015;76(2-3):129-136.
 34. Karahan GE, de Vaal YJH, Krop J, et al. A Memory B Cell Crossmatch Assay for Quantification of Donor-Specific Memory B Cells in the Peripheral Blood of HLA-Immunized Individuals. *Am J Transplant.* 2017;17(10):2617-2626.
 35. Mulder A, Eijssink C, Kardol MJ, et al. Identification, isolation, and culture of HLA-A2-specific B lymphocytes using MHC class I tetramers. *J Immunol.* 2003;171(12):6599-6603.
 36. Mulder A, Kardol MJ, Kamp J, et al. Determination of the frequency of HLA antibody secreting B-lymphocytes in alloantigen sensitized individuals. *Clin Exp Immunol.* 2001;124(1):9-15.
 37. Zachary AA, Kopchaliiska D, Montgomery RA, Leffell MS. HLA-specific B cells: I. A method for their detection, quantification, and isolation using HLA tetramers. *Transplantation.* 2007;83(7):982-988.
 38. Wen L, Hanvanich M, Werner-Favre C, Brouwers N, Perrin LH, Zubler RH. Limiting dilution assay for human B cells based on their activation by mutant EL4 thymoma cells: total and anti-malaria responder B cell frequencies. *Eur J Immunol.* 1987;17(6):887-892.
 39. Lighaam LC, Vermeulen E, den Bleker T, et al. Phenotypic differences between IgG4+ and IgG1+ B cells point to distinct regulation of the IgG4 response. *J Allergy Clin Immunol.* 2014;133(1): 267-270 e261-266.
 40. Heidt S, Roelen DL, Eijssink C, et al. Calcineurin inhibitors affect B cell antibody responses indirectly by interfering with T cell help. *Clin Exp Immunol.* 2010;159(2):199-207.
 41. Kramer CSM, Franke-van Dijk MEI, Priddey AJ, et al. Recombinant human monoclonal HLA antibodies of different IgG subclasses recognising the same epitope: Excellent tools to study differential effects of donor-specific antibodies. *HLA.* 2019;94(5):415-424.
 42. Vink T, Oudshoorn-Dickmann M, Roza M, Reitsma JJ, de Jong RN. A simple, robust and highly efficient transient expression system for producing antibodies. *Methods.* 2014;65(1):5-10.
 43. Ye J, Ma N, Madden TL, Ostell JM. IgBLAST: an immunoglobulin variable domain sequence analysis tool. *Nucleic Acids Res.* 2013;41(Web Server issue):W34-W40.
 44. Guex N, Peitsch MC. SWISS-MODEL and the Swiss-PdbViewer: an environment for comparative protein modeling. *Electrophoresis.* 1997;18(15):2714-2723.
 45. Kramer CSM, Roelen DL, Heidt S, Claas FHJ. Defining the immunogenicity and antigenicity of HLA epitopes is crucial for optimal epitope matching in clinical renal transplantation. *HLA.* 2017;90(1):5-16.
 46. Cai J, Kohanof S, Terasaki PI. HLA-DR antibody epitopes. *Clin Transpl.* 2006;103-114.
 47. Alber CA, Watts R, Klohe EP, et al. Multiple regions of HLA-DR beta 1 chains determine polymorphic epitopes recognized by monoclonal antibodies. *J Immunol.* 1989;143(7):2248-2255.
 48. Fu XT, Drover S, Marshall WH, Karr RW. HLA-DR residues accessible under the peptide-binding groove contribute to polymorphic antibody epitopes. *Hum Immunol.* 1995;43(4):243-250.
 49. Fu XT, Yu WY, Alber C, et al. Identification of residues involved in polymorphic antibody binding epitopes on HLA-DR molecules. *Hum Immunol.* 1992;33(1):47-56.
 50. Marsh SG, Bodmer JG. HLA-DR and -DQ epitopes and monoclonal antibody specificity. *Immunol Today.* 1989;10(9):305-312.
 51. Ouisse L-H, Gautreau-Rolland L, Devilder M-C, et al. Antigen-specific single B cell sorting and expression-cloning from immunoglobulin humanized rats: a rapid and versatile method for the generation of high affinity and discriminative human monoclonal antibodies. *BMC Biotechnol.* 2017;17(1):3.
 52. Karahan GE, Krop J, Wehmeier C, et al. An Easy and Sensitive Method to Profile the Antibody Specificities of HLA-specific Memory B Cells. *Transplantation.* 2019;103(4):716-723.
 53. Lavinder JJ, Horton AP, Georgiou G, Ippolito GC. Next-generation sequencing and protein mass spectrometry for the comprehensive analysis of human cellular and serum antibody repertoires. *Curr Opin Chem Biol.* 2015;24:112-120.
 54. Snanoudj R, Claas FH, Heidt S, Legendre C, Chatenoud L, Candon S. Restricted specificity of peripheral alloreactive memory B cells in HLA-sensitized patients awaiting a kidney transplant. *Kidney Int.* 2015;87(6):1230-1240.
 55. Drover S, Karr RW, Fu XT, Marshall WH. Analysis of monoclonal antibodies specific for unique and shared determinants on HLA-DR4 molecules. *Hum Immunol.* 1994;40(1):51-60.
 56. Mulder A, Kardol M, Regan J, Buelow R, Claas F. Reactivity of twenty-two cytotoxic human monoclonal HLA antibodies towards soluble HLA class I in an enzyme-linked immunosorbent assay (PRA-STAT). *Hum Immunol.* 1997;56(1-2):106-113.
 57. Daga S, Moyse H, Briggs D, et al. Direct quantitative measurement of the kinetics of HLA-specific antibody interactions with isolated HLA proteins. *Hum Immunol.* 2018;79(2):122-128.

58. Visentin J, Leu DL, Mulder A, et al. Measuring anti-HLA antibody active concentration and affinity by surface plasmon resonance: Comparison with the luminex single antigen flow beads and T-cell flow cytometry crossmatch results. *Mol Immunol*. 2019;108:34-44.
59. Maurer D, Gorski J. Transfer of polymorphic monoclonal antibody epitopes to the first and second domains of HLA-DR beta-chains by site-directed mutagenesis. *J Immunol*. 1991;146(2):621-626.
60. Kosmoliaptsis V, Dafforn TR, Chaudhry AN, Halsall DJ, Bradley JA, Taylor CJ. High-resolution, three-dimensional modeling of human leukocyte antigen class I structure and surface electrostatic potential reveals the molecular basis for alloantibody binding epitopes. *Hum Immunol*. 2011;72(11):1049-1059.
61. Mallon DH, Bradley JA, Winn PJ, Taylor CJ, Kosmoliaptsis V. Three-Dimensional Structural Modelling and Calculation of Electrostatic Potentials of HLA Bw4 and Bw6 Epitopes to Explain the Molecular Basis for Alloantibody Binding: Toward Predicting HLA Antigenicity and Immunogenicity. *Transplantation*. 2015;99(2):385-390.
62. Cai J, Terasaki PI, Mao Q, et al. Development of nondonor-specific HLA-DR antibodies in allograft recipients is associated with shared epitopes with mismatched donor DR antigens. *Am J Transplant*. 2006;6(12):2947-2954.
63. Mao Q, Terasaki PI, Cai J, El-Awar N, Rebellato L. Analysis of HLA class I specific antibodies in patients with failed allografts. *Transplantation*. 2007;83(1):54-61.
64. Everly MJ, Rebellato LM, Haisch CE, et al. Incidence and impact of de novo donor-specific alloantibody in primary renal allografts. *Transplantation*. 2013;95(3):410-417.
65. DeVos JM, Gaber AO, Knight RJ, et al. Donor-specific HLA-DQ antibodies may contribute to poor graft outcome after renal transplantation. *Kidney Int*. 2012;82(5):598-604.
66. Willicombe M, Brookes P, Sergeant R, et al. De novo DQ donor-specific antibodies are associated with a significant risk of antibody-mediated rejection and transplant glomerulopathy. *Transplantation*. 2012;94(2):172-177.
67. Ntokou I-S, Iniotaki AG, Kontou EN, et al. Long-term follow up for anti-HLA donor specific antibodies postrenal transplantation: high immunogenicity of HLA class II graft molecules. *Transpl Int*. 2011;24(11):1084-1093.
68. Gu Y, Wong YH, Liew CW, et al. Defining the structural basis for human alloantibody binding to human leukocyte antigen allele HLA-A*11:01. *Nat Commun*. 2019;10(1):893.
69. Kushihata F, Watanabe J, Mulder A, Claas F, Scornik JC. Human leukocyte antigen antibodies and human complement activation: role of IgG subclass, specificity, and cytotoxic potential. *Transplantation*. 2004;78(7):995-1001.
70. Honger G, Amico P, Arnold ML, Spriewald BM, Schaub S. Effects of weak/non-complement-binding HLA antibodies on C1q-binding. *HLA*. 2017;90(2):88-94.
71. Li F, Zhang X, Jin YP, Mulder A, Reed EF. Antibody ligation of human leukocyte antigen class I molecules stimulates migration and proliferation of smooth muscle cells in a focal adhesion kinase-dependent manner. *Hum Immunol*. 2011;72(12):1150-1159.
72. Valenzuela NM, Mulder A, Reed EF. HLA class I antibodies trigger increased adherence of monocytes to endothelial cells by eliciting an increase in endothelial P-selectin and depending on subclass, by engaging FcγRs. *J Immunol*. 2013;190(12):6635-6650.
73. Chong AS, Rothstein DM, Safa K, Riella LV. Outstanding questions in transplantation: B cells, alloantibodies, and humoral rejection. *Am J Transplant*. 2019;19(8):2155-2163.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

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