



Universiteit
Leiden
The Netherlands

Metabolic and functional evaluation of diabetic cardiomyopathy using MR Spectroscopy and MR Imaging

Bizino, M.B.

Citation

Bizino, M. B. (2022, November 16). *Metabolic and functional evaluation of diabetic cardiomyopathy using MR Spectroscopy and MR Imaging*. Retrieved from <https://hdl.handle.net/1887/3486006>

Version: Publisher's Version

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/3486006>

Note: To cite this publication please use the final published version (if applicable).

Part 1

Technical Advances in MRS and MRI to

Evaluate Diabetic Cardiomyopathy



Chapter 2

Metabolic Imaging of the Human Heart: Clinical Application of Magnetic Resonance Spectroscopy

Bizino MB, Hammer S, Lamb HJ
Heart. 2014 Jun;100(11):881-90.

Introduction

Cardiovascular MRI has earned its place in the field of clinical cardiac imaging. Regularly used techniques include anatomical imaging, functional imaging, perfusion and delayed enhancement (DE). Cardiac magnetic resonance spectroscopy (MRS) uses the same hardware, measuring the abundance of metabolites in the myocardium in vivo non-invasively without the use of radiation or external tracers. Its main application is currently scientific to gain insight into metabolic changes in cardiac pathologies.

The heart is a metabolically active organ using on average 6 kg of adenosine triphosphate (ATP) each day. As energy is crucial for both systole and diastole, derangements in energy metabolism may be the first step in failure of the heart¹. By using the gyromagnetic properties of ^1H , ^{31}P , ^{13}C and ^{23}Na MRS is a powerful tool to relate energy metabolism to (dys)function of the heart.

The aim of the present review is to provide an overview of the current use, opportunities and limitations of MRS in relation to common cardiac diseases: ischemic heart disease, heart failure, inherited cardiomyopathy, the metabolic syndrome, valvular heart disease and heart transplantation.

Cardiac energy metabolism

A simplified schematic representation of cardiac energy metabolism and opportunities to assess components of metabolism with MRS is depicted in figure 1. On average, the heart cycles 10 tons of blood each day in 100.000 heart beats. To meet the enormous ATP requirement, cardiomyocytes fuel themselves with free fatty acids (FFA) and glucose as the primary source of chemical energy. FFA and glucose contribute to ATP synthesis in terms of supply of chemical energy in a ratio of 3:1 in normal situations. Derangements in substrate utilization are associated with a wide variety of diseases which will be discussed below. The uptake of FFA by the fatty acid transporter is an energy-consuming process. Fatty acids enter the mitochondrion where beta-oxidation takes place after which the intermediate Acetyl-Coenzyme A (CoA) enters the Krebs cycle. The uptake of glucose by the glucose transporter type 4

(GLUT4) is insulin-dependent. Glucose is converted to pyruvate in the cytoplasm by glycolysis. Pyruvate enters the Krebs cycle in the mitochondrion (figure 1).

The process of oxidative phosphorylation is the basis for production of energy by the mitochondrial respiratory chain. This yields energy needed for the generation of ATP from adenosine diphosphate (ADP). Transfer of chemical energy (ATP) from mitochondrion to myofibril is provided by the creatine kinase energy shuttle. In the mitochondrion, creatine kinase catalyses conversion of ATP and creatine to phosphocreatine (PCr) and ADP (figure 1). PCr rapidly diffuses to the myofibrils. Once arrived there, creatine kinase catalyses the reaction back to ATP and free creatine. Creatine then diffuses back to the mitochondrion. Two thirds of the total creatine pool is phosphorylated to PCr; the other part remains in the cell as free creatine. The creatine kinase system acts as an important energy buffer, providing the heart of energy when the demand outweighs the supply. Such situation results in a decreased PCr level and increased ADP level whereas ATP levels are held constant. Overconsumption of PCr and subsequent increase in ADP lead to inhibition of many intracellular enzymes¹. Furthermore the process of energy production is regulated by calcium flux. For example calcium flux from the sarcoplasmic reticulum to the mitochondrion can directly activate oxidative metabolism². Both diastolic and systolic heart function are dependent upon energy synthesis and utilization. This is reflected by the fact that calcium-handling is an important determinant of ATP-consuming relaxation of cardiomyocytes as well as systolic function of the heart³.

Metabolic assessment of the myocardium with ¹H, ³¹P, ¹³C and ²³Na - MRS

MRS of the heart has been performed using 1.5 and 3.0 tesla (T) MR systems, and recently even using ultra-high 7T MR systems. Modern systems can be delivered with software to support MRS. Depending on the MR system (manufacturer, field strength) coils for ¹H and ³¹P spectroscopy are commercially available. However to initiate cardiac MRS technical development and support is needed to optimize i.e. shimming techniques, gradients and pulse sequences. Furthermore, each MR system

requires its own specific adjustments. For ^{13}C and ^{23}Na -MRS coils, pulse sequences and metabolic tracers are not routinely obtainable.

As the heart moves due to contraction and breathing, MRS is subject to motion artifacts. Therefore, ECG-triggering is necessary to correct for motion of the heart throughout the cardiac cycle. Respiratory motion compensation has been performed with respirometer-triggered acquisition, breath-hold scanning sequences and by using navigator echoes^{4,5}. Figure 2 shows a navigator-gated acquisition of a ^1H spectrum of the heart. The scanning time for ^1H -MRS varies from a single breath-hold to twenty minutes (excluding time required for ^1H imaging for planning of the voxel). The most commonly used pulse sequences are Stimulated Echo Acquisition Mode (STEAM) and Point Resolved Spectroscopy (PRESS). STEAM has the disadvantage of a lower signal-to-noise ratio (SNR) and increased susceptibility to motion, but enables acquisition with shorter echo time and hence better detection of fat. As SNR increases with field strength ^1H -MRS has recently been performed in one breath-hold at 3T with STEAM⁵. Based on the required variability in the measurement, mode of respiratory compensation and scanning time an appropriate method can be chosen. ^{31}P -MRS using the method depicted in figure 3 requires roundabout ten minutes. For postprocessing the spectra jMRUI is the most commonly used program. Table 1 shows an overview of the potential clinical applications of ^1H , ^{31}P , ^{13}C and ^{23}Na -MRS and challenges of the technique.

^1H is the most suitable nucleus for MRS because it has the highest MR sensitivity. ^1H -MRS can detect various metabolites in the myocardium that are associated with (dys)function (see figure 1). Assessment of triglyceride (TG) content of the myocardium is of major importance in view of the epidemic proportions of cardiovascular disease related to obesity in modern society. Using ^1H -MRS myocardial TG content has been shown to be an independent predictor of diastolic dysfunction in both healthy men⁶ and type 2 diabetes mellitus (T2DM) patients⁷. Furthermore, creatine content can be determined with ^1H -MRS. As part of the creatine kinase energy shuttle, creatine content reflects the energy status of the heart. In heart failure and non-viable myocardium creatine content is depleted^{8,9}. Other metabolites that can be detected are lactate, (de)oxymyoglobin and carnitine.

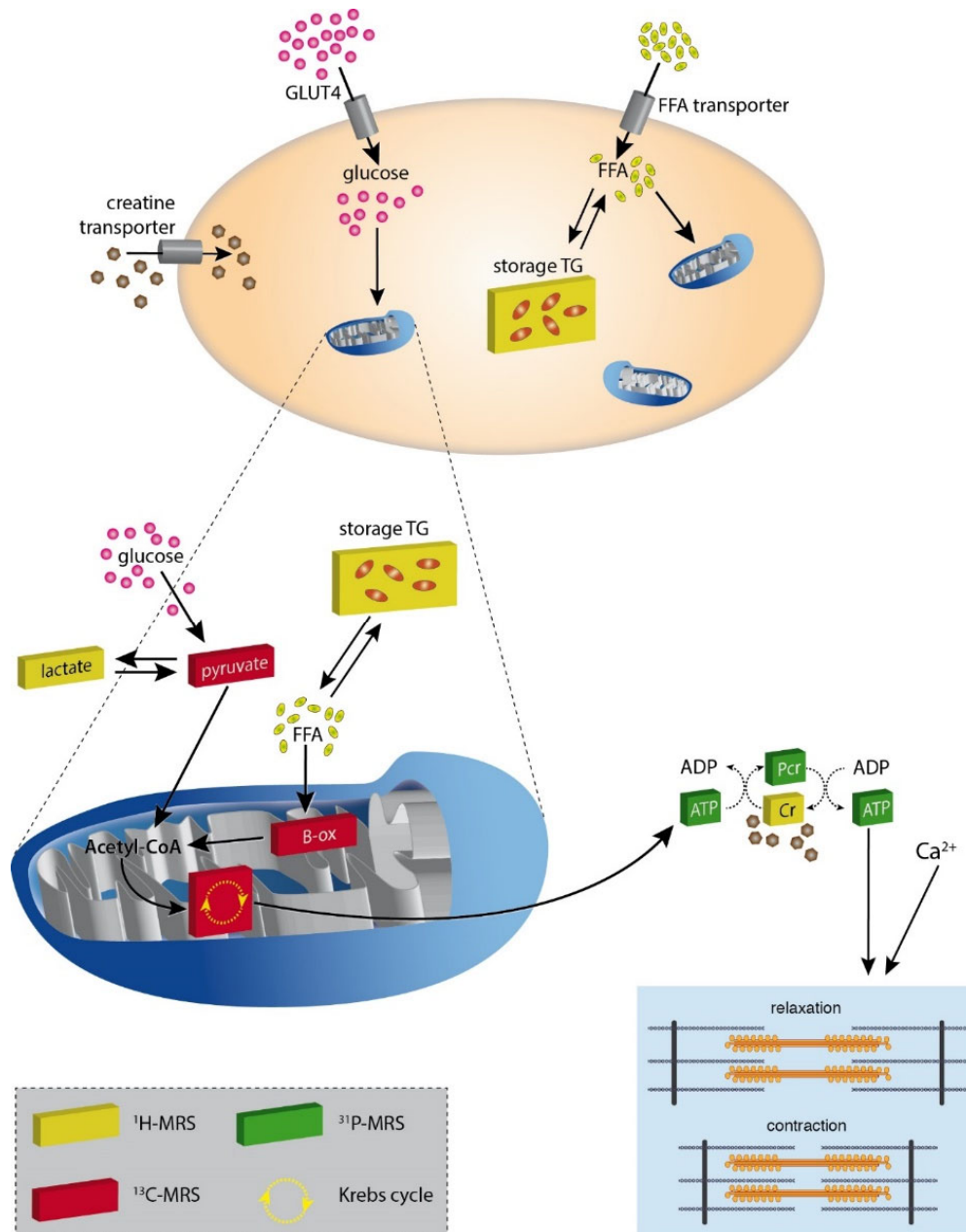


Figure 1 Magnetic resonance spectroscopy (MRS) assessment of cardiac energy metabolism. Upper figure depicts a schematic diagram of energy metabolism of the cardiomyocyte. In the mitochondrion (blue) ATP is generated from mainly glucose and FFAs. ATP and regulation by calcium flux are needed for contraction and relaxation of the myofibrils. The coloured boxes (yellow, green and red) indicate opportunities for measurement with MRS, respectively 1H , ^{31}P , and ^{13}C . Glucose and FFAs are the primary sources of chemical energy. Glucose uptake by the GLUT4 is insulin dependent; FFAs are transported into the cell via the FFA transporter. Glucose is subject to glycolysis and converted to pyruvate which enters the mitochondrion or is converted to lactate. FFAs are either stored as triglycerides or enter the mitochondrion. Both pyruvate and FFA (after β -oxidation) give rise to acetyl-CoA. Acetyl-CoA enters the Krebs cycle. In the mitochondrion the process of oxidative phosphorylation generates ATP. Outside the mitochondrion energy is transferred to PCr which releases its energy in the myofibrils giving rise to ATP and Cr. Acetyl-CoA, acetyl-coenzyme A; ADP, adenosine diphosphate; ATP, adenosine triphosphate; B-ox, β -oxidation; Ca^{2+} , ionised calcium; Cr, creatine; FFA, free fatty acid; GLUT4, glucose transporter type 4; PCr, phosphocreatine; TG, triglyceride content.

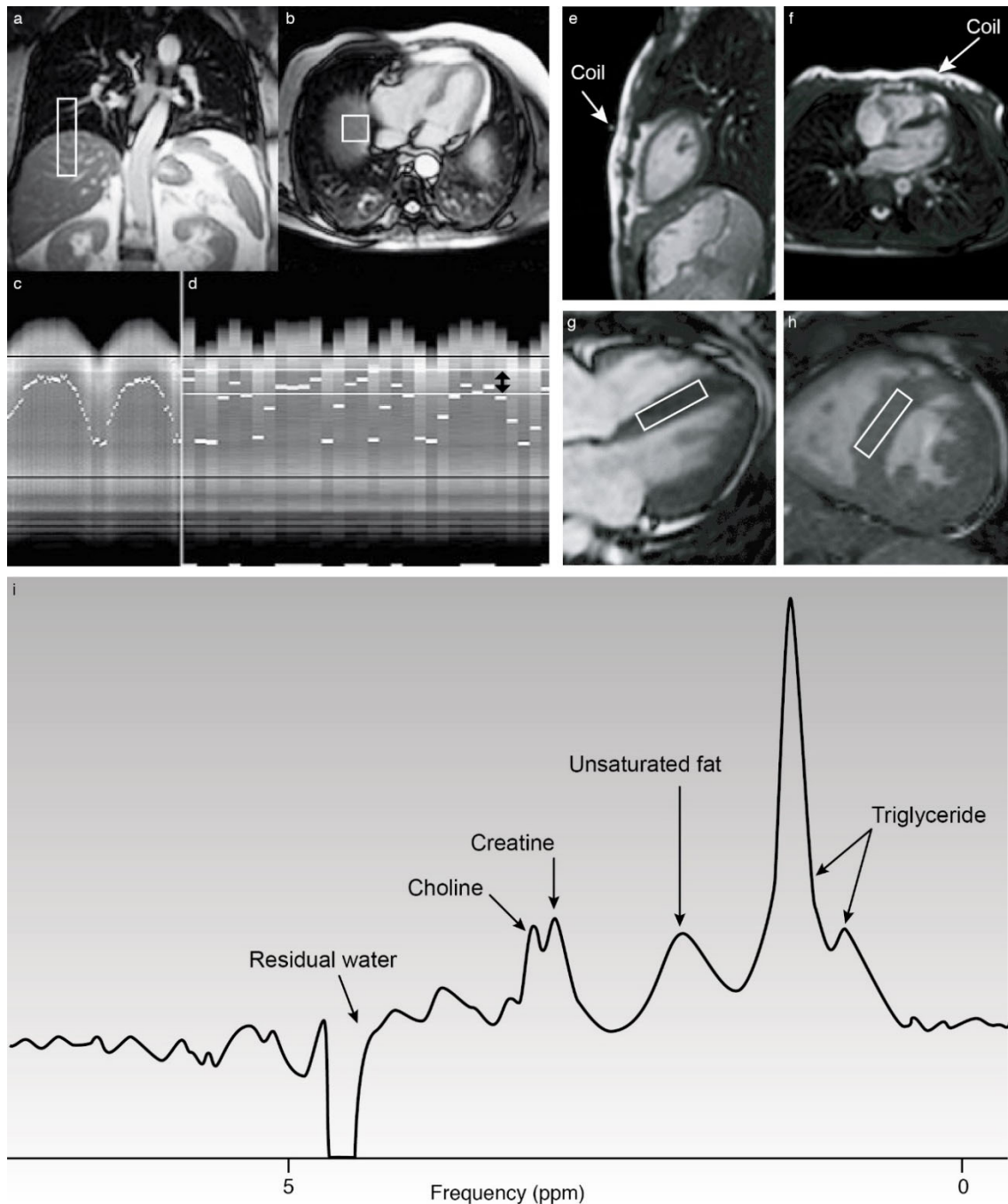


Figure 2 Navigator gated ^1H magnetic resonance (MR) spectroscopy of the heart. Upper left panel shows preparation and monitoring of the respiratory navigator gating technique. A pencil beam is placed on the dome of the right hemidiaphragm on coronal (A) and transverse (B) MR images. White dots (C) represent the automatically traced position of the diaphragm during calibration. In (D) horizontal lines indicate the acceptance window; whenever the detected position of the diaphragm is within the predefined window, the spectroscopic measurement is stored. The upper right panel shows the acquisition of the spectrum. The surface coil is positioned just below the anatomical level of the mitral valve on sagittal (E) and transverse (F) MR images. By using a four chamber (G) and short axis (H) view the spectroscopic volume is placed in the interventricular septum. As water is by far the most abundant substance, an ^1H spectrum as depicted in (I) can only be acquired after applying suppression of the water signal. Adapted from van der Meer et al.⁴

Table 1. Summary of potential clinical applications of cardiac MRS

Nucleus	Metabolites	Potential clinical application	Technical hurdles
¹ H	Creatine content	Heart Failure, determination of myocardial viability	- Need to suppress water signal - Need to compensate breathing artifacts - Improved spatial resolution required
	Triglyceride content	Obesity / Metabolic syndrome / DM2: myocardial triglyceride content	
³¹ P	PCr	Detection of ischemia, scar tissue, heart failure	- Very low spatial resolution
	ATP	Detection of scar tissue	
	PCr/ATP ratio	Decreased in: - ischemic heart disease - heart failure - valvular disease - DCM - HCM - Diabetic cardiomyopathy	
¹³ C	1- ¹³ C pyruvate: alanine, lactate, bicarbonate, pyruvate dehydrogenase flux	Ischemic heart disease, viability assessment, heart failure, diabetic and inherited cardiomyopathy	- Complex technique to generate hyperpolarized ¹³ C - Expensive
	2- ¹³ C-pyruvate: downstream metabolites of Acetyl-CoA (Krebs cycle and Acetyl carnitine)	Heart failure, inherited cardiomyopathy	
	¹³ C-bicarbonate: intracellular pH	Ischemic heart disease, heart failure, heritable metabolic cardiomyopathy	
²³ Na	Sodium content	Scar tissue	- Low spatial resolution - No absolute quantification of sodium possible
		Ischemia	
		Hibernating myocardium?	

³¹P-MRS has been used extensively to assess high energy phosphate metabolism which is crucial for maintaining normal function of the heart. ³¹P-MRS is therefore a useful tool to analyze the energy status of myocardium. The PCr peak and a combination of the three peaks of ATP are used to calculate the PCr/ATP-ratio (see

figure 1). This ratio is the most commonly used parameter in ^{31}P -MRS of the heart. In general there are two conditions associated with a decreased ratio: 1. ATP synthesis cannot meet the ATP requirement i.e. in ischemia; 2. Decreased total creatine supply is diminished. Absolute measurements of PCr and ATP can also be assessed albeit in a more complex way¹. Figure 3 shows an example of a technique to perform ^{31}P -MRS of the heart and an example of a phosphorus spectrum.

^{13}C -MRS is limited by the fact that it has too low MR sensitivity and abundance in myocardial tissue to be used in vivo. However, with the exciting new technique of hyperpolarization (dynamic nuclear polarization technique) the signal can be increased > 10.000 times. A hyperpolarized metabolic tracer is made by mixing the molecule with free radicals, placement in a magnetic field and then freezing it to a temperature close to 1K¹⁰. In cardiometabolic research, ^{1-13}C pyruvate, ^{2-13}C pyruvate and ^{13}C -bicarbonate tracers have been used in animal studies allowing analysis of pyruvate dehydrogenase flux and assessment of components of beta-oxidation and Krebs cycle (see figure 1). Figure 4 shows that ^{13}C -MRS could be used for detection of ischemia and / or infarction. Although ^{1-13}C pyruvate has been used in research setting in one human study, technical issues regarding safety have to be addressed before the technique of hyperpolarization gains regulatory approval for widespread use.

^{23}Na -MRS can be used to produce images based on the level of sodium content of the myocardium. The regional spectra can be converted into signals producing an MR image of the myocardium, based on sodium content. As the sodium content alters in ischemic and infarcted tissue, ^{23}Na MRI can be used to detect myocardial infarction and viability^{11,12}.

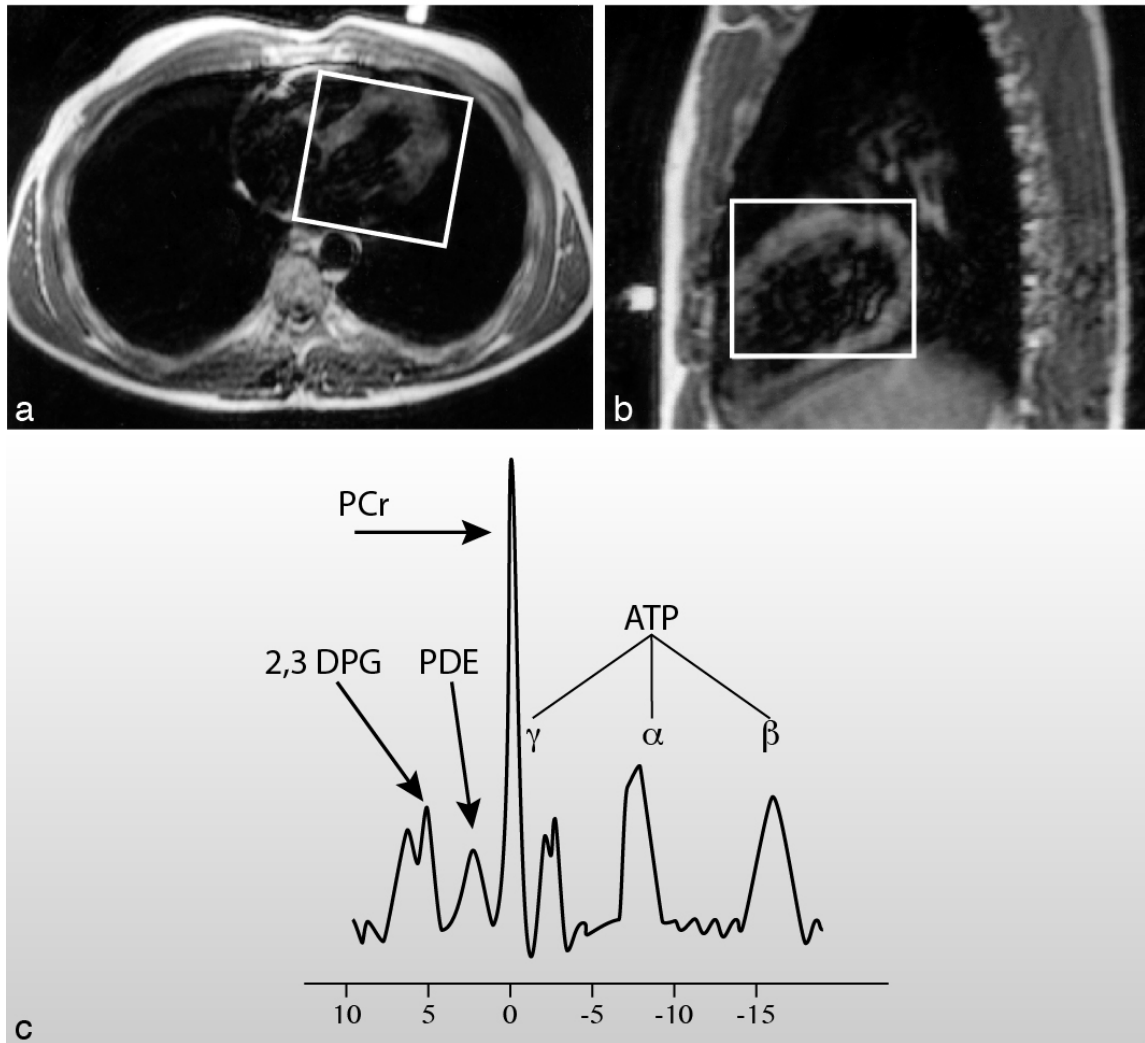


Figure 3 ^{31}P -magnetic resonance spectroscopy of the human heart. Volume of interest is positioned in an optimal orientation relative to the left ventricle based on scout images in the transverse (A) and sagittal imaging plane (B). The volume (square box) is placed perpendicular to the chest wall to prevent contamination with skeletal muscle. By adjusting the level of the volume selection in the caudo-cranial direction, contamination of the sensitive volumes by diaphragm muscle and liver tissue can be prevented (B). The white square on the chest wall originates from a reference sample in the centre of the surface coil. With this technique a spectrum as shown in (C) can be acquired. Note that with this technique no regional differences can be detected, but an average of the signals in the left ventricle is acquired. 2,3 DPG, 2,3-diphosphoglycerate; PDE, phosphodiester; PCr, phosphocreatine.

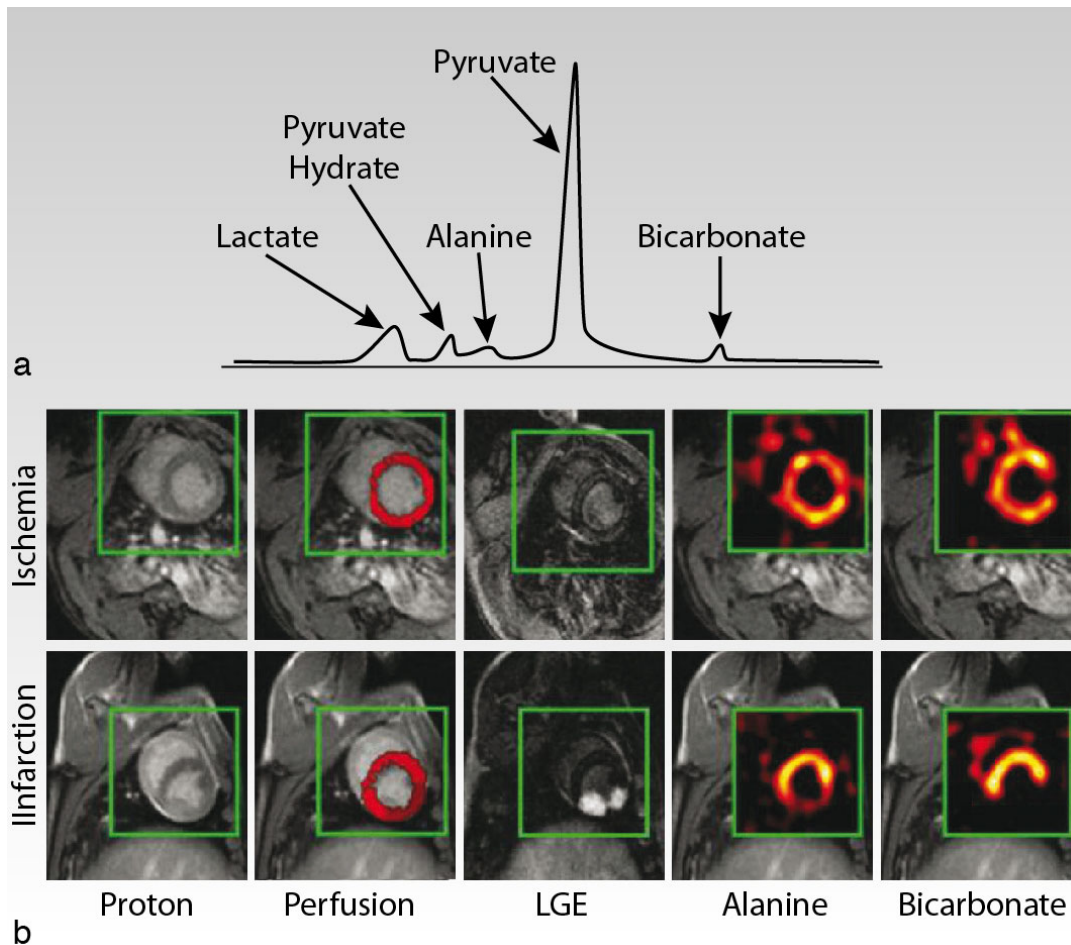


Figure 4 ^{13}C -magnetic resonance spectroscopy (MRS) combined with MRI of a pig heart. By combining proton MRI, perfusion and late gadolinium enhancement (LGE) with ^{13}C MRS with a ^{1-13}C pyruvate tracer, ischaemic and infarcted tissue can be characterised. These images were obtained in pig hearts. By superimposing the signals acquired in spectroscopy on proton MR images, maps can be made for different metabolites. Alanine reflects cytoplasmic cell metabolism whereas bicarbonate indicates mitochondrial activity. In the ischaemic model (15 min occlusion of left circumflex artery) only the bicarbonate changes, whereas in the infarction model (45 min of occlusion) the alanine signal drops and the bicarbonate signal almost disappears, spatially correlated to LGE area. Note that the spatial resolution is high enough to establish regional differences in intensity. Adapted from Golman et al.²³

Ischemic heart disease

Ischemic heart disease develops when oxygen demands outweigh the supply. In daily practice, coronary angiography (CAG) or coronary artery CT are the standard procedures to rule out significant obstructive coronary artery disease (CAD). However, ischemia can also occur without significant stenosis. The cause of ischemia in the absence of significant CAD may be microvascular disease and is associated with a worse prognosis in terms of survival and reinfarction¹³. Furthermore, the discrimination between non-viable (scarred) and viable (hibernating or stunned)

myocardium is of clinical importance to determine the potential benefit of revascularization.

Since oxygen is crucial for the mitochondrial process of oxidative phosphorylation, ischemic myocardium is largely dependent on inefficient anaerobic glycolysis for the production of ATP. In ischemic myocardium ATP production, and thus formation of PCr decreases. A decrease in the PCr/ATP ratio reflects exhaustion of energy reserves of the myocardium.

³¹P-MRS has been used in extension to investigate cardiac energy metabolism in ischemic and infarcted tissue. Weiss et.al. performed a study in patients with > 70% stenosis of the left anterior descending coronary artery (LAD) before and after revascularization. Myocardial ischemia was provoked by handgrip exercise. Patients with stenosis of the LAD had a significantly decreased PCr/ATP-ratio during exercise whilst no difference between cases and controls could be established in rest. After revascularization the 'between group' and 'within group' difference was recovered¹⁴. An interesting finding was that in women admitted with chest pain with normal CAG, 7 out of 35 had an abnormal PCr/ATP ratio during handgrip exercise¹⁵. To evaluate the prognostic finding of an abnormal PCr/ATP ratio, a prospective follow up study was performed in women with chest pain, normal CAG with or without abnormal PCr/ATP ratio and a reference group of 352 women with known coronary artery disease. After adjusting for CAD and cardiac risk factors, a PCr/ATP ratio decrease of 1% increased the risk for cardiovascular events by 4%^{15,16}. The most likely explanation for these findings is that patients with normal CAG and abnormal ³¹P-MRS suffer from microvascular pathology.

Opposed to viable myocardial tissue, the PCr/ATP ratio is not useful in infarcted tissue since both PCr and ATP levels are reduced¹⁷. However, absolute levels of PCr and ATP do correlate with infarct size and viability respectively. In a study in humans with either a fixed or reversible defect on 201TI scan, myocardial PCr content decreased significantly in patients with reversible and fixed defects when compared to healthy controls. ATP content decreased significantly in subjects with fixed defects, but did not in those with reversible defects¹⁸. These data suggest that ³¹P-MRS could be used to determine myocardial viability. Moreover, in a rat model of myocardial

infarction, whole-heart PCr content was inversely correlated with infarct size, whereas ATP distribution provides a profile of viable myocardium around the infarction reflecting remodeling of the heart¹⁹.

In rats that underwent ligation of the LAD, ¹H-MRS proved lower creatine content as compared to controls²⁰. In line with this, in humans total creatine content was significantly lower in regions of infarction⁸.

In animal studies it has been shown that viable and nonviable myocardium have a different sodium content as detected by ²³Na-MRI. ²³Na signal intensity (SI) is elevated in acute ischemia and remains increased afterwards in humans²¹. To further investigate the role of ²³Na-MR imaging in ischemic heart disease, ²³Na-MRI was compared with cine MR imaging and delayed enhancement (DE) imaging in 15 patients with subacute infarction and 15 with chronic infarction. In subacute infarction all patients showed regional elevated ²³Na SI that correlated well with wall motion abnormality (WMA) and with DE ($r = 0.68$). In chronic infarction, the patients that had presented with WMA showed increased ²³Na SI even in cases without DE, indicating that ²³Na MRI could detect hibernating myocardial tissue²².

As shown in figure 4 ¹³C-MRS has the potential to distinct between viable and non-viable myocardial tissue in an experiment in pigs²³. However this technique has not been performed in cardiovascular research in humans as yet.

A major drawback of application of MRS in general, although significant in ischemic heart disease, is the low spatial resolution. Since ischemic heart disease does not involve the whole myocardium, information on localization of the ischemic or infarcted area is hard to establish with the current techniques. Overcoming these issues with for example higher field imaging or hyperpolarization could pave the way for MRS to be of diagnostic relevance, most notably in patients with suspected ischemic heart disease without coronary artery stenosis on CAG. Moreover, MRS could be of use in patients to discriminate between viable and non-viable myocardium.

Heart failure

Heart failure is the final common pathway of a diverse set of specific diseases which will be described in this section. Heart failure in general, its prognosis and the

effect of (non-) pharmacologic interventions have been studied with MRS. The most commonly studied causes of heart failure are dilated cardiomyopathy, hypertrophic cardiomyopathy and cardiomyopathy related to the metabolic syndrome and T2DM. Metabolic imaging in heart failure has generally focused on two aspects: 1) high energy phosphate metabolism (^{31}P) and 2) substrate utilization and myocardial triglyceride content (^1H).

Decreased ATP metabolism is the final common pathway in heart failure and can be measured by ^{31}P -MRS. Both the absolute level of ATP and ATP flux are diminished^{24,25}. The PCr/ATP ratio may underestimate the metabolic derangement because both PCr and ATP levels are decreased²⁶.

The causality between altered substrate preference (FFA vs glucose) and heart failure is controversial and beyond the scope of this review. To summarize clinically relevant findings:

- High levels of circulating FFA's are related to cardiac lipotoxicity and reduce cardiac glucose utilization²⁷.
- In advanced heart failure systemic and cardiac insulin resistance are related to myocardial lipotoxicity irrespective of the presence of obesity²⁸.
- Therapeutic interventions in substrate utilization can improve heart function. For example, inhibition of fatty acid oxidation and stimulation of glucose use with trimetazidine improved the PCr/ATP ratio and cardiac function in heart failure patients²⁹.

Dilated cardiomyopathy

Dilated cardiomyopathy is characterized by left ventricular dilatation, myocardial wall thinning and decreased systolic function. Underlying mechanisms are cardiomyocyte death and myocardial fibrosis caused by a heterogeneous group of mutations in different pathways³⁰. These mechanisms reflect global cardiac metabolic changes, allowing regional spectroscopy techniques to be representative of energy metabolism in the whole heart. MRS has provided insight into metabolism of cardiomyocytes in DCM. The severity of dilated cardiomyopathy in terms of LV end-diastolic wall thickness and ejection fraction showed a linear regression with the PCr/ATP ratio³¹. Whether metabolic derangements precede heart failure and play a

role in the etiology, or it merely is an innocent bystander, could not be extracted from this observational study. There are however indications that alterations in the high energy phosphate metabolism play an important role in the pathogenesis of dilated cardiomyopathy. First, in patients with dilated cardiomyopathy a decreased PCr/ATP-ratio has been shown to be an independent predictor of both total and cardiovascular mortality and offered significant independent prognostic information when compared to NYHA class³². Second, in patients and carriers with Duchenne or Becker muscular dystrophy (the Xp21 dystrophies) the reduced PCr/ATP ratio's were not associated with LV mass or ejection fraction. As patients and carriers of the Xp21 dystrophies frequently develop cardiac hypertrophy and dilated cardiomyopathy, this study suggests that the metabolic derangement precedes cardiac structural changes and failure³³. Moreover, diuretics and ACE-inhibitors induce an improvement in the PCr/ATP ratio parallel to clinical improvement, suggesting that MRS could be used as a quantitative marker of success of therapy³⁴. In the future ¹³C-MRS might be used to unravel the causality between substrate use and heart failure as shown in an experimental model of pigs with DCM³⁵.

Hypertrophic cardiomyopathy

Cardiac hypertrophy has different etiologies. It can be caused by genetic factors (hypertrophic cardiomyopathy), may be the result of chronic hypertension and is also seen physiologically in athletes. Metabolism in these different hypertrophic hearts has been studied predominantly with ³¹P-MRS. In hypertrophic cardiomyopathy a reduced PCr/ATP ratio has been a consistent finding, in the context of hypertension³⁶, as well as in young asymptomatic HCM patients³⁷. On the contrary, there is a normal PCr/ATP ratio in physiologic hypertrophy³⁸. The fact that a reduced PCr/ATP ratio precedes the symptomatology of HCM patients, suggests a causative role for metabolic derangement in HCM. Further evidence of this is that patients with a genetic hypertrophic cardiomyopathy due to sarcomeric gene mutations, the metabolic derangement was not correlated with the degree of hypertrophy³⁹. This study therefore suggests that 30% decrease in the PCr/ATP ratio is not a consequence of hypertrophy / heart failure / strain but may be at the beginning of the pathophysiological process. The fact that energy deficiency is at least in part

responsible for heart failure was generated by a study by Abozguia et.al. They proved in a placebo-controlled RCT that the PCr/ATP ratio increased with perhexiline, an agent thought to improve cardiac energetics by providing a shift from fatty acid oxidation to glucose utilization. The increased PCr/ATP ratio was associated with an increase in exercise capacity and improvement in diastolic function^{39,40}. ³¹P-MRS has the potential to play a role in determining prognosis of patients with HCM and support intervention trials focused on the potential of improving energy metabolism to ameliorate cardiac function.

Metabolic syndrome

Apart from an increased risk of coronary events, patients with the metabolic syndrome (including T2DM) have an increased risk of developing heart failure without evidence of ischemia / infarction, the so called diabetic cardiomyopathy. The pathophysiology of cardiac dysfunction in the metabolic syndrome is rather complex and reviewed by van der Meer et.al.⁴¹. It appears that cardiac energy metabolism plays a crucial role in diastolic and systolic dysfunction in these patients. Cardiac energy metabolism in the metabolic syndrome is characterized by an increase of fatty acid uptake and oxidation becomes less efficient²⁷. Consistent with this hypothesis, the PCr/ATP ratio was inversely correlated to fasting FFA concentrations in T2DM patients whilst the PCr/ATP ratio was lower compared to healthy controls, despite the fact that cardiac mass and function appeared to be normal⁴². Accumulation of triglycerides (as detected by ¹H-MRS) in the heart could play a role in the pathogenesis of heart failure in patients with obesity and / or diabetes. There is profound evidence that myocardial triglycerides reflect cardiomyocyte function by a process called lipotoxicity: the accumulation of toxic lipids such as diacylglycerol and ceramide^{28,43}. Indeed myocardial triglyceride content has been shown to be an independent predictor of diastolic dysfunction³. Interestingly diastolic dysfunction can be reversed, in parallel with decreased myocardial steatosis, by prolonged caloric restriction in T2DM patients⁴⁴. Another study with T2DM patients has proven that a short term increment of FFA flux induced by fasting, is associated with myocardial steatosis and diastolic dysfunction⁴⁵. However, in morbidly obese patients undergoing bariatric surgery, the improvement of diastolic function after six months was not associated with a change

in myocardial triglyceride content⁴⁶. ¹H-MRS and ³¹P-MRS were also used to investigate the mechanism of improvement of cardiac function by pioglitazone. Here, the functional improvement was not associated with metabolic changes⁴⁷. In the future MRS could be helpful to unravel pathophysiology of cardiovascular disease in the metabolic syndrome, and aid in the evaluation of the effect of life style and / or pharmacologic interventions, especially when combined with cardiovascular magnetic resonance imaging. ¹³C-MRS could help to study the complex interplay between substrate use and heart failure in patients with the metabolic syndrome / diabetic cardiomyopathy⁴⁸.

Valvular heart disease

In valvular heart disease the timing of valve replacement is an important issue: a balanced decision must be taken on the basis of expected benefits versus the risk related to the replacement procedure. Studies involving ³¹P-MRS of the heart have been performed in patients with aortic stenosis (AoS), aortic incompetence (AoI) and mitral regurgitation (MR). In line with observations in patients with heart failure, the PCr/ATP ratio was reduced in AoS and AoI patients with symptoms of heart failure when compared to asymptomatic patients⁴⁹. A similar study performed by Neubauer et. al. concordantly showed a decreased ratio only in patients with NYHA class 3 or 4. The ratio was more disturbed in AoS patients with the highest wall stress⁵⁰. In 22 patients with MR the PCr/ATP ratio was higher in those with symptoms and more severe disease. Again, there was a correlation between the PCr/ATP ratio and various parameters of ventricular dysfunction. In this study there was no correlation with wall stress however⁵¹. Breyerbacht et.al. performed ³¹P-MRS before and 40 weeks after aortic valve replacement in 9 AoS patients. PCr/ATP ratio's improved significantly from 1,28 to 1,47⁵². The fact that the PCr/ATP ratio is a marker of disease severity in valvular heart disease and is an independent predictor of mortality in DCM, suggests that serial ³¹P- MRS assessments in patients with valvular heart disease could prove to play a role in the timing of a surgical procedure. However further studies are needed in that respect. In the research setting, MRS could aid to unravel the causal relationship between impaired cardiac energy metabolism and mechanical changes: how do they interact to ultimately result in a failing heart?

Heart transplantation

Cardiac allograft patients are at risk for rejection. The golden standard to diagnose rejection is based on histology of the myocardium which warrants repeated biopsy procedures in the post-transplant period⁵³. Therefore a non-invasive tool for this diagnosis is desirable. To investigate the sensitivity of MRS in predicting rejection, ³¹P-MRS at rest was performed in 14 patients and 17 healthy controls late after transplantation (months to years). The PCr/ATP ratio was significantly lower in transplant recipients than healthy controls. However ³¹P-MRS could not reliably predict in which patients augmented immunosuppressive therapy was warranted⁵⁴. Van Dobbenburgh et al. performed a similar study but assessed ³¹P-MRS repeatedly in the early post-transplant period to investigate whether ³¹P-MRS could detect early acute rejection. Again PCr/ATP-ratio's were lower in heart transplant recipients than controls, with the ratio recovering in time. However, no correlation was found between PCr/ATP ratio's and biopsy scores of rejection: ³¹P-MRS was unable to predict early acute rejection⁵⁵. Evanochko et.al. performed a ³¹P-MRS stress test in heart transplant recipients with normal CAG. Interestingly 10 out of 25 had a positive stress test with a mean decrease of the PCr/ATP ratio of $25,6 \pm 3.6\%$. Further research is needed to determine the cause of this finding. A possible explanation could be that the reduced ratio is an early sign of chronic allograft vasculopathy⁵⁶. These findings are hopeful with respect to implementing MRS techniques in the follow-up of heart transplant patients, although the exact role of MRS remains to be established.

Future direction

The fact that function of the heart depends on myocardial energy metabolism makes MRS an excellent technique to evaluate diseased myocardium. However, the low spectral and temporal resolution combined with the lack of availability of MRS to study the heart in most clinics hampers widespread use clinical cardiology. Moreover, the currently used techniques remain time consuming.

Whether cardiac spectroscopy will aid in diagnostic and therapeutic decision making depends on the achievement of better spatial and temporal resolution as well

as reproducibility. There are various technical opportunities to achieve this goal. First of all, by using higher field MR systems (> 3 Tesla), the advantage of increased SNR could result in better resolution and reproducibility⁵⁷. Furthermore, development of improved shimming algorithms, pulse sequences and receiver coils could aid in achieving higher spatial resolution MRS, i.e. to be able to perform an MRS-based analysis in line with the 17-segment model for detection of regional metabolic alterations specific for (non) viable myocardium. One of the most promising new techniques is hyperpolarization. Hyperpolarized ¹³C with various tracers can assess substrate flux and thereby gain a position in clinical practice^{48,58}.

Conclusion

Magnetic resonance spectroscopy is a unique non-invasive, non-irradiating tool to assess cardiac metabolism in humans. In research setting, MRS has provided insight into pathophysiology of multiple cardiac diseases. The tight relation between energy metabolism and cardiac function potentiates spectroscopy to be of use in diagnostics, prognosis and treatment guidance in cardiology practice. Continuous improvements regarding technical optimization, and exciting new techniques such as hyperpolarization, may help to implement cardiac MRS in selected patients in daily practice.

Acknowledgement

We thank G. Kracht for support on figures and table.

References

1. Neubauer S. The failing heart--an engine out of fuel. *N Engl J Med* 2007; **356**(11): 1140-51.
2. Tarasov AI, Griffiths EJ, Rutter GA. Regulation of ATP production by mitochondrial Ca(2+). *Cell Calcium* 2012; **52**(1): 28-35.
3. Louch WE, Stokke MK, Sjaastad I, Christensen G, Sejersted OM. No rest for the weary: diastolic calcium homeostasis in the normal and failing myocardium. *Physiology (Bethesda)* 2012; **27**(5): 308-23.
4. van der Meer RW, Doornbos J, Kozerke S, et al. Metabolic imaging of myocardial triglyceride content: reproducibility of 1H MR spectroscopy with respiratory navigator gating in volunteers. *Radiology* 2007; **245**(1): 251-7.
5. Rial B, Robson MD, Neubauer S, Schneider JE. Rapid quantification of myocardial lipid content in humans using single breath-hold 1H MRS at 3 Tesla. *Magn Reson Med* 2011; **66**(3): 619-24.
6. van der Meer RW, Rijzewijk LJ, Diamant M, et al. The ageing male heart: myocardial triglyceride content as independent predictor of diastolic function. *Eur Heart J* 2008; **29**(12): 1516-22.
7. Rijzewijk LJ, van der Meer RW, Smit JW, et al. Myocardial steatosis is an independent predictor of diastolic dysfunction in type 2 diabetes mellitus. *J Am Coll Cardiol* 2008; **52**(22): 1793-9.
8. Bottomley PA, Weiss RG. Non-invasive magnetic-resonance detection of creatine depletion in non-viable infarcted myocardium. *Lancet* 1998; **351**(9104): 714-8.
9. Nakae I, Mitsunami K, Omura T, et al. Proton magnetic resonance spectroscopy can detect creatine depletion associated with the progression of heart failure in cardiomyopathy. *J Am Coll Cardiol* 2003; **42**(9): 1587-93.
10. Tyler DJ. Cardiovascular Applications of Hyperpolarized MRI. *Curr Cardiovasc Imaging Rep* 2011; **4**(2): 108-15.
11. Horn M, Weidensteiner C, Scheffer H, Przyklenk K, von KM, Neubauer S. Use of 23Na MRS to discriminate viable from non viable tissue: experimental studies. *MAGMA* 2000; **11**(1-2): 42-3.
12. Kim RJ, Lima JA, Chen EL, et al. Fast 23Na magnetic resonance imaging of acute reperfused myocardial infarction. Potential to assess myocardial viability. *Circulation* 1997; **95**(7): 1877-85.
13. Yilmaz A, Sechtem U. Angina pectoris in patients with normal coronary angiograms: current pathophysiological concepts and therapeutic options. *Heart* 2012; **98**(13): 1020-9.
14. Weiss RG, Bottomley PA, Hardy CJ, Gerstenblith G. Regional myocardial metabolism of high-energy phosphates during isometric exercise in patients with coronary artery disease. *N Engl J Med* 1990; **323**(23): 1593-600.
15. Buchthal SD, den Hollander JA, Merz CN, et al. Abnormal myocardial phosphorus-31 nuclear magnetic resonance spectroscopy in women with chest pain but normal coronary angiograms. *N Engl J Med* 2000; **342**(12): 829-35.
16. Johnson BD, Shaw LJ, Buchthal SD, et al. Prognosis in women with myocardial ischemia in the absence of obstructive coronary disease: results from the National Institutes of Health-National Heart, Lung, and Blood Institute-Sponsored Women's Ischemia Syndrome Evaluation (WISE). *Circulation* 2004; **109**(24): 2993-9.
17. von KM, Rosch C, Le FY, et al. Three-dimensional 31P magnetic resonance spectroscopic imaging of regional high-energy phosphate metabolism in injured rat heart. *Magn Reson Med* 1998; **39**(5): 731-41.
18. Yabe T, Mitsunami K, Inubushi T, Kinoshita M. Quantitative measurements of cardiac phosphorus metabolites in coronary artery disease by 31P magnetic resonance spectroscopy. *Circulation* 1995; **92**(1): 15-23.
19. Friedrich J, Apstein CS, Ingwall JS. 31P nuclear magnetic resonance spectroscopic imaging of regions of remodeled myocardium in the infarcted rat heart. *Circulation* 1995; **92**(12): 3527-38.
20. Bottomley PA, Weiss RG. Noninvasive localized MR quantification of creatine kinase metabolites in normal and infarcted canine myocardium. *Radiology* 2001; **219**(2): 411-8.
21. Sandstede JJ, Hillenbrand H, Beer M, et al. Time course of 23Na signal intensity after myocardial infarction in humans. *Magn Reson Med* 2004; **52**(3): 545-51.

22. Sandstede JJ, Pabst T, Beer M, et al. Assessment of myocardial infarction in humans with (23)Na MR imaging: comparison with cine MR imaging and delayed contrast enhancement. *Radiology* 2001; **221**(1): 222-8.
23. Golman K, Petersson JS, Magnusson P, et al. Cardiac metabolism measured noninvasively by hyperpolarized 13C MRI. *Magn Reson Med* 2008; **59**(5): 1005-13.
24. Smith CS, Bottomley PA, Schulman SP, Gerstenblith G, Weiss RG. Altered creatine kinase adenosine triphosphate kinetics in failing hypertrophied human myocardium. *Circulation* 2006; **114**(11): 1151-8.
25. Weiss RG, Gerstenblith G, Bottomley PA. ATP flux through creatine kinase in the normal, stressed, and failing human heart. *Proc Natl Acad Sci U S A* 2005; **102**(3): 808-13.
26. Beer M, Seyfarth T, Sandstede J, et al. Absolute concentrations of high-energy phosphate metabolites in normal, hypertrophied, and failing human myocardium measured noninvasively with (31)P-SLOOP magnetic resonance spectroscopy. *J Am Coll Cardiol* 2002; **40**(7): 1267-74.
27. Lopaschuk GD, Folmes CD, Stanley WC. Cardiac energy metabolism in obesity. *Circ Res* 2007; **101**(4): 335-47.
28. Chokshi A, Drosatos K, Cheema FH, et al. Ventricular assist device implantation corrects myocardial lipotoxicity, reverses insulin resistance, and normalizes cardiac metabolism in patients with advanced heart failure. *Circulation* 2012; **125**(23): 2844-53.
29. Fragasso G, Perseghin G, De CF, et al. Effects of metabolic modulation by trimetazidine on left ventricular function and phosphocreatine/adenosine triphosphate ratio in patients with heart failure. *Eur Heart J* 2006; **27**(8): 942-8.
30. Watkins H, Ashrafian H, Redwood C. Inherited cardiomyopathies. *N Engl J Med* 2011; **364**(17): 1643-56.
31. Neubauer S, Horn M, Pabst T, et al. Contributions of 31P-magnetic resonance spectroscopy to the understanding of dilated heart muscle disease. *Eur Heart J* 1995; **16 Suppl O**: 115-8.
32. Neubauer S, Horn M, Cramer M, et al. Myocardial phosphocreatine-to-ATP ratio is a predictor of mortality in patients with dilated cardiomyopathy. *Circulation* 1997; **96**(7): 2190-6.
33. Crilley JG, Boehm EA, Rajagopalan B, et al. Magnetic resonance spectroscopy evidence of abnormal cardiac energetics in Xp21 muscular dystrophy. *J Am Coll Cardiol* 2000; **36**(6): 1953-8.
34. Neubauer S, Krahe T, Schindler R, et al. 31P magnetic resonance spectroscopy in dilated cardiomyopathy and coronary artery disease. Altered cardiac high-energy phosphate metabolism in heart failure. *Circulation* 1992; **86**(6): 1810-8.
35. Schroeder MA, Lau AZ, Chen AP, et al. Hyperpolarized (13)C magnetic resonance reveals early- and late-onset changes to in vivo pyruvate metabolism in the failing heart. *Eur J Heart Fail* 2013; **15**(2): 130-40.
36. Lamb HJ, Beyerbach HP, van der Laarse A, et al. Diastolic dysfunction in hypertensive heart disease is associated with altered myocardial metabolism. *Circulation* 1999; **99**(17): 2261-7.
37. Jung WI, Sieverding L, Breuer J, et al. 31P NMR spectroscopy detects metabolic abnormalities in asymptomatic patients with hypertrophic cardiomyopathy. *Circulation* 1998; **97**(25): 2536-42.
38. Pluim BM, Lamb HJ, Kayser HW, et al. Functional and metabolic evaluation of the athlete's heart by magnetic resonance imaging and dobutamine stress magnetic resonance spectroscopy. *Circulation* 1998; **97**(7): 666-72.
39. Crilley JG, Boehm EA, Blair E, et al. Hypertrophic cardiomyopathy due to sarcomeric gene mutations is characterized by impaired energy metabolism irrespective of the degree of hypertrophy. *J Am Coll Cardiol* 2003; **41**(10): 1776-82.
40. Abozguia K, Elliott P, McKenna W, et al. Metabolic modulator perhexiline corrects energy deficiency and improves exercise capacity in symptomatic hypertrophic cardiomyopathy. *Circulation* 2010; **122**(16): 1562-9.
41. van der Meer RW, Lamb HJ, Smit JW, de Roos A. MR imaging evaluation of cardiovascular risk in metabolic syndrome. *Radiology* 2012; **264**(1): 21-37.
42. Scheuermann-Freestone M, Madsen PL, Manners D, et al. Abnormal cardiac and skeletal muscle energy metabolism in patients with type 2 diabetes. *Circulation* 2003; **107**(24): 3040-6.
43. Schaffer JE. Lipotoxicity: when tissues overeat. *Curr Opin Lipidol* 2003; **14**(3): 281-7.

44. Hammer S, Snel M, Lamb HJ, et al. Prolonged caloric restriction in obese patients with type 2 diabetes mellitus decreases myocardial triglyceride content and improves myocardial function. *J Am Coll Cardiol* 2008; **52**(12): 1006-12.
45. Hammer S, van der Meer RW, Lamb HJ, et al. Short-term flexibility of myocardial triglycerides and diastolic function in patients with type 2 diabetes mellitus. *Am J Physiol Endocrinol Metab* 2008; **295**(3): E714-8.
46. Gaborit B, Jacquier A, Kober F, et al. Effects of bariatric surgery on cardiac ectopic fat: lesser decrease in epicardial fat compared to visceral fat loss and no change in myocardial triglyceride content. *J Am Coll Cardiol* 2012; **60**(15): 1381-9.
47. van der Meer RW, Rijzewijk LJ, de Jong HW, et al. Pioglitazone improves cardiac function and alters myocardial substrate metabolism without affecting cardiac triglyceride accumulation and high-energy phosphate metabolism in patients with well-controlled type 2 diabetes mellitus. *Circulation* 2009; **119**(15): 2069-77.
48. Schroeder MA, Clarke K, Neubauer S, Tyler DJ. Hyperpolarized magnetic resonance: a novel technique for the in vivo assessment of cardiovascular disease. *Circulation* 2011; **124**(14): 1580-94.
49. Conway MA, Allis J, Ouwerkerk R, Niioka T, Rajagopalan B, Radda GK. Detection of low phosphocreatine to ATP ratio in failing hypertrophied human myocardium by ³¹P magnetic resonance spectroscopy. *Lancet* 1991; **338**(8773): 973-6.
50. Neubauer S, Horn M, Pabst T, et al. Cardiac high-energy phosphate metabolism in patients with aortic valve disease assessed by ³¹P-magnetic resonance spectroscopy. *J Investig Med* 1997; **45**(8): 453-62.
51. Conway MA, Bottomley PA, Ouwerkerk R, Radda GK, Rajagopalan B. Mitral regurgitation: impaired systolic function, eccentric hypertrophy, and increased severity are linked to lower phosphocreatine/ATP ratios in humans. *Circulation* 1998; **97**(17): 1716-23.
52. Beyerbach HP, Lamb HJ, van Der LA, et al. Aortic valve replacement in patients with aortic valve stenosis improves myocardial metabolism and diastolic function. *Radiology* 2001; **219**(3): 637-43.
53. Patel JK, Kittleson M, Kobashigawa JA. Cardiac allograft rejection. *Surgeon* 2011; **9**(3): 160-7.
54. Bottomley PA, Weiss RG, Hardy CJ, Baumgartner WA. Myocardial high-energy phosphate metabolism and allograft rejection in patients with heart transplants. *Radiology* 1991; **181**(1): 67-75.
55. Van Dobbenburgh JO, De Groot MC, de JN, et al. Myocardial high-energy phosphate metabolism in heart transplant patients is temporarily altered irrespective of rejection. *NMR Biomed* 1999; **12**(8): 515-24.
56. Evanochko WT, Buchthal SD, den Hollander JA, et al. Cardiac transplant patients response to the (³¹P) MRS stress test. *J Heart Lung Transplant* 2002; **21**(5): 522-9.
57. Stephenson MC, Gunner F, Napolitano A, et al. Applications of multi-nuclear magnetic resonance spectroscopy at 7T. *World J Radiol* 2011; **3**(4): 105-13.
58. Malloy CR, Merritt ME, Sherry AD. Could ¹³C MRI assist clinical decision-making for patients with heart disease? *NMR Biomed* 2011; **24**(8): 973-9.

