

#### **Immune checkpoint inhibitors in mesothelioma** Disselhorst, M.J.

#### **Citation**

Disselhorst, M. J. (2022, October 25). *Immune checkpoint inhibitors in mesothelioma*. Retrieved from https://hdl.handle.net/1887/3483978



**Note:** To cite this publication please use the final published version (if applicable).



# **Chapter 1**

General introduction and outline of thesis

Malignant pleural mesothelioma (MPM) is an aggressive tumor originating from the mesothelial cells of the pleural cavity. It has a causal relation with (occupational) asbestos exposure (1).

Asbestos is a group of 6 different mineral fibers naturally occurring throughout the world; all are composed of long and thin fibrous crystals. Two large subgroups are known as the serpentine and amphibole subgroup. Chrysotile (white asbestos) is a serpentine mineral, of which the fibers are relatively large and curly and it is the most commonly used type of asbestos. Amphibole minerals are needle-like and members of this class are amosite (brown asbestos), crocidolite (blue asbestos), tremolite, actinolite and anthophyllite. Asbestos is being used since prehistoric times due to its fire-resistant properties (2). In the last century it has been used extensively in buildings and ship-building, because of its strength, fire-resistance and isolating properties. Furthermore it is cheap. All types of asbestos fibers can cause mesothelioma.

Asbestos is banned from most countries in the world, but it is estimated that approximately 43,000 people will die from this disease worldwide (3). The survival is poor, with a 5 year survival rate in Europe of 7% (4). In the Netherlands, spray asbestos was banned in 1978 and complete use of asbestos in 1993. Unfortunately, exposure is still possible since it is incorporated in many buildings and sheds. With a latency time between asbestos exposure and diagnosis of mesothelioma of 20 to 50 years (1, 5) we are still confronted with 600 patients per year in the Netherlands.

The carcinogenic mechanism of how asbestos can cause MPM is not completely understood. Chronic inflammation may predispose individuals to develop this malignancy as is concluded from microscopic examinations. In the tumor microenvironment (TME) inflammation promotes proliferation and survival of malignant cells (6). Asbestos can cause an influx of mononuclear phagocytic cells into the tumor that internalize asbestos fibers. These phagocytic cells will release proinflammatory cytokines. In combination with chronic inflammation, oxygen radical release and DNA damage, these processes promote malignant transformation. In combination with the immunosuppressive environment, this promotes cancer growth. It has been shown that CXCR3 (the chemokine receptor on the surface of T helper cells) and the production of interferon gamma (IFN-γ) were reduced in peripheral CD4+ cells of asbestos-exposed patients, thereby showing the decreased antitumor immunity of asbestos (7).

Only a minority of asbestos exposed people develop mesothelioma. This might for some cases be explained by genetic susceptibility. Germline mutations in (BRCA1) associated protein-1 (BAP1) tumor suppressor gene cause the BAP1 tumor predisposition syndrome. Carriers have an increased risk of developing

mesothelioma, (uveal) melanoma, renal cell, basal cell and hepatocellular carcinoma. It is thought that loss of BAP1 may predispose to mesothelioma after asbestos exposure. Homozygous deletion of CDKN2A, loss of NF2 or germline PALB2 deletions may also favor the development of MPM (8, 9). Genetic susceptibility can predispose to MPM via chronic exposition.

MPM is classified in 3 histological subtypes, epithelioid, biphasic and sarcomatoid. The sarcomatoid subtype is composed of malignant spindle cells and occurs in 10-15% of MPM, is chemotherapy-resistant and has the worst survival. The epithelioid subtype is the most common variant. It accounts for 50-70% of all mesotheliomas, and is composed of epithelioid polygonal cells. The biphasic subtype has features of both epithelioid and sarcomatoid subtype, larger biopsies are needed to demonstrate both components. Examination of both tumor and surrounding stroma has revealed that features such as inflammation, cellular diversity and vacuolization within the stroma all have a prognostic effect, besides the histopathological findings (10).

#### **Diagnosis**

First step in diagnostic process is usually a contrast-enhanced computed tomography (CT) of chest or a positive-emission tomography (PET) with CT, showing pleural enlargement, pleural fluid and sometimes thoracic wall invasion.

A cytological diagnosis of mesothelioma is often difficult when thoracocentesis is used to obtain the pleural fluid. This material provides a diagnosis in 20-50% of patients and only in epithelioid subtype, but it can often exclude other diagnoses. Histological biopsies by thoracoscopy or ultrasound or CT-guided have a high diagnostic accuracy. Immunohistochemistry markers usually include calretinine, cytokeratin 5/6, Wilms Tumor 1 antigen (WT1), those should be positive. Markers for adenocarcinoma should be negative (TTF-1, CEA, Ber-EP4). The sensitivity for sarcomatoid subtype is poor. Absence of BAP1 expression could be an important extra tool, it is lost in up to 60% of cases, most often in epithelioid subtype (11, 12).

In the Netherlands, nearly all mesothelioma diagnoses (and possible diagnoses) are centrally reviewed by an expert pathology board, the "Nederlands Mesotheliomen Panel" because of the rareness of the disease and the difficulty of the diagnosis.

Comprehensive genomic and transcriptomic sequencing of MPM revealed large heterogeneity between patients. Most mutations found inactivation of tumor suppressor genes (f.e. BAP1, CDKN2A, NF2, TP53, SETD2) (13-15). Heterogeneity has been reported within the tumor location in the chest cavity. Kiyotani examined biopsies of patients at 3 different sites and showed intratumoral heterogeneity in somatic mutations and unique TCRβ clonotypes of TILs (16).

## **Clinical**

Patients with MPM are typically men and older than 65. Symptoms are gradually worsening and include dyspnea, chest pain, cough, night sweats, fatigue and weight loss. Tumor spreads throughout the pleural cavity, and can result in pleural effusions. Metastases are rare, but can involve the lungs, bone, liver and CNS. Most patients present with advanced disease, which is incurable.

## **Treatment**

Surgical treatment for MPM remains controversial in many parts of the world, since it is always incomplete. Whether cytoreductive surgery prolongs overall survival is unclear, studies did not provide a clear positive outcome that outweighs the risk, with high morbidity for surgery. This is beyond the scope of this thesis, which is focused on systemic treatment.

For almost 20 years, platinum containing chemotherapy combined with an antifolate has been the standard of care for patients. Leading to a median overall survival of about 12 to 16 months. Unfortunately, the mean progression free survival (PFS) is only 6 months (17, 18).

The MAPS trial (Mesothelioma Avastin plus Pemetrexed-Cisplatin) showed that standard of care chemotherapy combined with bevacizumab (a monoclonal antibody targeting vascular endothelial growth factor), improved survival over chemotherapy alone (18.8 vs 16.1 months). Although there is a survival improvement, there is also an increased adverse event profile for bevacizumab. So it failed to be approved as standard treatment (19). Other anti-angiogenetic drugs also failed to show benefit (20).

In the past it has been observed that installation of BCG (Bacillus Calmette-Guérin) vaccine immunotherapy could have an improved survival rate for MPM (21).

This led to the idea that the immune system could play an important role in the biology and treatment of MPM. Cancer immunotherapy makes use of the host system to induce

or enhance an effective immune response against cancer cells. Different types of immunotherapy use different parts of the immune system to evoke effect on tumor cells.

Immune checkpoint proteins are crucial for maintenance of self-tolerance. Expression of these proteins is dysregulated in tumor cells, thereby making the tumor cell immune resistant. Immune checkpoint inhibitors (ICI) can block inhibitory checkpoints, thereby restoring immune system function and evoking an anti-cancer immune response.

Anti-Cytotoxic T-Lymphocyte-associated protein 4 (CTLA-4) antibodies impact the lymphoid compartment; increasing the number and broadening the tumor antigen reactive T cells; stimulating priming of naive T cells and enhancing antigen presentation. PD-L1 checkpoints are mainly expressed in activated lymphocytes and exhausted T cells. Anti-PD-(L)1 antibodies can promote T cell activation during the effector phase and can restore exhausted T cell functionality, mainly in the tumor microenvironment.

These immune checkpoint inhibitors are the most widely used agents of cancer immunotherapy and completely changed treatment of many cancer types over the last decade. In 2011, ipilimumab was the first checkpoint inhibitor approved by the FDA for treatment of melanoma (22). Ipilimumab blocks immune checkpoint molecule CTLA-4. After that PD-1 (nivolumab, pembrolizumab, cemiplimab), and PD-L1 (atezolizumab, durvalumab, avelumab) checkpoint inhibitors are approved for many cancer types.

For mesothelioma some promising data on ICI treatment have been reported in the second or later lines, mostly in single arm trials. Single agent PD-1 ICI have consistent objective response rates of about 20%, and disease control rates (DCR) between 48 and 72% in mainly phase II trials (23-28). The single agent CTLA-4 checkpoint inhibitor tremelimumab however, did not show any benefit compared to placebo (29).

The first randomized trial of pembrolizumab (PD-1 antibody) failed to improve PFS or OS over single agent chemotherapy (vinorelbine or gemcitabine) in later lines. Although pembrolizumab did have a higher overall response rate (ORR), 22% versus 6% (P=0.004) (30).

The second phase III trial of monotherapy of anti-PD-1 (nivolumab)showed a survival benefit of nivolumab over best supportive care in relapsed MPM, mOS was 10.2 months (95% CI 8.5-12.1) in the nivolumab group versus 6.9 months (5.0-8.0) in the placebo group (adjusted HR 0.69 [95% CI 0.52-0.91]; p=0.0090). Placebo was used for the comparator arm since no approved second line therapy exists (31).

Combining aPD-(L)1 and aCTLA-4 therapy has been shown to induce synergistic effects in preclinical and clinical trials (32, 33). Combining them can induce a more potent antitumor immune response (34).

This led to setting up a clinical trial in MPM with combination therapy, the INITIATE trial, which is described in chapter in this thesis (35).

For combination treatment with anti-PD-1 plus anti-CTLA-4, the ORR is around 27% and mPFS 6 months in single arm phase II trials, in recurrent disease (27, 35, 36).

In 2021, the Checkmate 743 trial was published. This international randomized phase III trial compared standard of care chemotherapy with combined nivolumab plus ipilimumab. ICI treatment significantly increased overall survival compared to chemotherapy by 4 months (mOS 18.1 months [95% CI 16.8 – 21.4] versus 14.1 months [95% CI 12.4-16.2], HR 0.74 [p=0.0020]). This lead to approval of nivolumab plus ipilimumab as first line therapy for MPM by the FDA and EMA. The benefit is most prominent in the non-epithelioid subgroup, as revealed by a post-hoc subgroup analysis, epithelioid subgroup HR 0.86 (95% CI 0.69–1.08) and non-epithelioid subgroup HR 0.46 (95% CI 0.31–0.68) (37).

## **Tumor microenvironment**

The mesothelioma tumor microenvironment (TME) is composed of heterogeneous stromal, endothelial and immune cells.

The TME in MPM is known to be highly immunosuppressive, with large numbers of tumor associated macrophages (TAMs), myeloid-derived suppressor cells (MDSC) and regulatory T cells (Tregs) (38-41).

Macrophages are plentiful present in MPM, with large heterogeneity, in both the epithelial and non-epithelial subtype. Mesothelial cells produce cytokines, which give chemotactic and stimulatory signals to immune cells of the myeloid lineage and recruit monocytes. In the tumor mass the monocytes differentiate into macrophages. Interleukins such as IL-1, IL-4, and IL-10 produced by tumor infiltrating lymphocytes (TILs) promote differentiation of macrophages towards a certain phenotype. This phenotype is pro-tumorgenic and promotes tumor growth by production of multiple cytokines. Higher percentages of macrophages are negatively correlated with overall survival and are positively correlated to the number of Tregs in tumor microenvironment (42-45).

MDSCs are immature myeloid cells and have immunosuppressive properties. They induce Tregs and produce nitric oxide and arginase, which leads to loss of function of CD4+ and CD8+ T cells (46).

T-Lymphocytes play an important role in the immune defense in cancer. These immune cells may influence tumor growth, but also mediate response to therapy. Twenty to 42% of the cells in the immune infiltrate consist of CD3+ T-lymphocytes. Besides the CD8+ T-lymphocytes, regulatory CD4+ FoxP3+ T-cells are frequently observed (39, 47, 48). Some studies suggest that higher levels of CD8+ T-cells have a favorable prognostic impact, while others found that high CD4+ and CD20+ and low FoxP3+ cells are linked to a better outcome (47-49).

The composition of the TME is different between subtypes, between individuals and within individuals (16, 40).

#### **Biomarkers**

Although a number of patients with cancer benefit from ICI treatment, many patients do not. Different mechanisms are proposed to explain these (non)responses to ICI treatment in cancers in general and in mesothelioma specifically.

The number of non-synonymous single nucleotide variants, referred to as the tumor mutation burden (TMB) may affect the odds of generating immunogenic peptides and thereby influence ICI response. In different tumor types the response to ICI treatment is positively correlated to TMB; a higher TMB resulting in a higher overall response rate (50). However, some tumor types respond better (51) and some worse (52) than would be expected based on TMB alone. And even within a specific tumor type some patients respond better than others. So although the association between TMB and ICI response is pretty robust, other factors are involved. MPM shows a rather low mutation rate, so TMB alone does not explain the response rates (13, 14, 16, 53).

It is hypothesized by Mansfield et al. that the number of alterations actually targeted by T cells, may have a stronger association with ICI response than does TMB (54). This includes immunogenic translocations or insertions/deletions, called chromoplexy and chromothrypsis.

A strong expression of the immune-checkpoint gene VISTA was found on tumor cells in epithelioid subtype. VISTA is a negative checkpoint regulator, possibly it avoids

an antitumor immune response (15, 55). The immunoregulatory impact needs to be elucidated.

In MPM, PD-L1 expression on tumor cells is observed in about 40% and is frequently associated with non-epithelioid subtype. PD-L1 expression is a prognostic marker and associated with worse outcome, when used in patients that are not treated with IO agents (56-61).

In NSCLC, PD-L1 expression is predictive of response to PD-1 checkpoint inhibitors. Tumors with higher PD-L1 expression usually respond better to IO treatment. But responses occur even in PD-L1 negative tumors and not all patient with high PD-L1 expression respond to treatment. In different other tumor types PD-L1 expression on tumor cells is not associated with response, whereas PD-L1 expression on tumor-infiltrating immune cells is (62-64).

In some phase II trials with ICI treatment for MPM, a (poor) correlation of PD-L1 expression with objective response rate and/or survival is shown. But in most other trials no correlation was found. Data are inconsistent (24-27, 31).

The predictive role of PD-L1 expression for dual agent ICI treatment has not been established either.

In the Checkmate 743 trial a relatively large amount of patients had PD-L1 positive tumors (77%), PD-L1 expression did not correlate with outcome. However survival with chemotherapy was better in patients with PD-L1 expression of less than 1% than in those with expression higher than 1%, this is probably more prognostic than predictive (35, 37).

In several tumor-types it is shown that density of tumor-infiltrating lymphocytes (TIL) is a positive prognostic indicator (regardless of ICI treatment) (65). In melanoma it is shown that pre-treatment TIL-density at the invasive margin is associated with response to anti-PD-1 treatment (62). Standardization is difficult, especially in MPM, which does not even have a distinct invasive margin.

Not only density of TILs impacts ICI outcomes, but also the type of immune cells. In melanoma, response to ICI treatment relies on pre-treatment infiltration of activated CD8 T-effector cells (62). In many more different cancers the association of infiltrating CD8+ cytotoxic T cells with longer disease free survival and/or overall survival has been demonstrated (66). In NSCLC, CD8 cell infiltration was positively correlated with ORR and PFS in patients treated with PD-1 blockade (67). A positive correlation of CD8+ T cells with overall survival has been reported, but not in all studies. One study even described opposite negative correlation (47-49, 68). This might be caused by sampling bias due to a

heterogeneous distribution in tumors, more advanced stage or from functional variability. The CD8+ cells could be exhausted cytotoxic T cells, with relatively high expression of multiple inhibitory receptors.

One study reported more CD8+ cells in PD-L1+ tumors versus PD-L1- tumors (69). Another study showed a higher ratio of cytotoxic T cells to malignant cells in the sarcomatoid subtype (70). Furthermore, CD8+ cells increased after administration of platinum plus pemetrexed, examined in paired biopsies (71).

For further analyses of mechanism of effect of ICI treatment, longitudinal tumor biopsies are needed. However these are not always possible to obtain, since in patients having a complete or partial response it is no longer possible to biopsy.

Peripheral blood T cells can provide insight of understanding immunological responses induced by ICI treatment. It also can provide biomarkers to monitor or predict response to ICI treatment. In lung cancer, it is shown that an increase in Ki-67+ PD-1+ CD8+ T cells is seen after ICI treatment in most patients. This may indicate activation of tumor-specific CD8+ T cells. These cells co-expressed CTLA-4 after PD-1 antibody treatment (72). In melanoma, presence of neoantigen specific T cells in peripheral blood is shown. Mainly in CD8+ PD-1+ T cells, which account for < 5% of all peripheral blood lymphocytes, patient specific neoantigens that target mutant and/or shared tumor neoantigens in all the melanoma patients (73).

Other blood biomarkers have been a focus in biomarker research, since they are easily accessible, are independent from intra- and inter-tumor heterogeneity, and reflect multiple factors (e.g. tumor cells, tumor-microenvironment and patient's immune system). Inflammation is a mechanism of immune-resistance in patients with cancer, promoting cancer growth and dissemination, based on activating oncogenic signaling pathways. Proposed inflammatory biomarkers that might be prognostic or predictive include LDH, CRP, white blood cells, absolute neutrophil count, neutrophil to lymphocyte ratio (NLR) derived neutrophil to lymphocyte ratio (dNLR; absolute neutrophil count/(white blood cell concentration – absolute neutrophil)). In melanoma a pro-inflammatory status is correlated with poor outcomes in patients treated with ICIs (74, 75). In NSCLC pretreatment Lung Immune Prognostic Index (LIPI), combining dNLR greater than 3 and LDH greater than ULN was correlated with worse outcome for ICI, but not for chemotherapy (76).

Exhaled breath analysis has shown potential as a non-invasive and easy-to-use technology for diagnosis and phenotyping of a wide range of diseases including mesothelioma and lung cancer. Electronic nose (eNose) technology can be used for this breath analysis (77- 81). This eNose could be used for immunotherapy response in lung cancer. In lung cancer

it has been shown that exhaled breath analysis before start of treatment could identify patients that show progressive disease to anti-PD-1 therapy, thereby ICI treatment could possibly be with-held (82). In addition, it can identify patients with an objective response to anti-PD-1 therapy early during treatment (83).

### **Outline of thesis**

This thesis aims to contribute to a better treatment of malignant pleural mesothelioma, and is specifically evaluating dual checkpoint inhibitor treatment. Besides the clinical effect of ICI treatment also the search for an explanation for the effect of this treatment, thereby aiming to predict response to treatment.

**Part I** summarizes what is known about treatment of mesothelioma

Chapter 2 is a review that discusses optimal systemic therapy for patients with advanced MPM. Including first-line, maintenance and second-line therapy, as well as antibody drug conjugates and targeted agents.

Chapter 3 is a review that focusses more in detail on novel treatment options in MPM, including immune checkpoint inhibitors.

Chapter 4 is a chapter from the ESMO handbook Immuno-Oncology on mesothelioma, describing what is known about immune checkpoint inhibition in MPM.

**Part II** is the clinical part of this thesis.

Chapter 5 describes the single center, single arm, phase 2 clinical INITIATE trial of nivolumab plus ipilimumab. This combination of checkpoint inhibitors shows marked efficacy in MPM, with no new safety concerns.

**Part III** is the translational research part of the thesis.

In chapter 6 immune cell profiling was performed on screening and on treatment peripheral blood samples of MPM patients treated with nivolumab (anti PD-1 antibody) monotherapy or a combination of nivolumab and ipilimumab (anti-CTLA-4 antibody). High proportions of effector memory CD8 T cells that re-expressed RA (TEMRA) and cytokine production by TEMRAs before treatment was associated with a better clinical outcome.

In chapter 7 exhaled breath analysis of volatile organic compounds by electronic technology (eNose) is performed in patients treated with nivolumab plus ipilimumab. An eNose is able to discriminate between responders and non-responders to treatment at baseline.

In chapter 8 immunohistochemistry analysis was performed on baseline and on-treatment biopsies from INITIATE trial and from a clinical trial using nivolumab in MPM. Cell density of CD4+, CD8+, and FoxP3+ cells is higher in patients having a response to nivolumab plus ipilimumab compared to patients having progressive disease at 24 weeks.

In chapter 9 RNA and whole genome sequencing was performed on the same biopsies as described above. A particular gene set demonstrated an interaction with tumor junction burden and was predictive of overall survival. Thus, analysis of structural variants and gene expression may facilitate patient selection for immune checkpoint inhibitors.

#### **References**

- 1. Wolff H, Vehmas T, Oksa P, Rantanen J, Vainio H. Asbestos, asbestosis, and cancer, the Helsinki criteria for diagnosis and attribution 2014: recommendations. Scandinavian journal of work, environment & health. 2015;41(1):5-15.
- 2. Dilek Y, Newcomb S. Ophiolite concept and the evolution of geological thought: Geological Society of America; 2003.
- 3. Stayner L, Welch LS, Lemen R. The worldwide pandemic of asbestos-related diseases. Annual review of public health. 2013;34:205-16.
- 4. Francisci S, Minicozzi P, Pierannunzio D, Ardanaz E, Eberle A, Grimsrud TK, et al. Survival patterns in lung and pleural cancer in Europe 1999-2007: Results from the EUROCARE-5 study. European journal of cancer (Oxford, England : 1990). 2015;51(15):2242-53.
- 5. McDonald JC. Epidemiology of malignant mesothelioma--an outline. The Annals of occupational hygiene. 2010;54(8):851-7.
- 6. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011;144(5):646-74.
- 7. Maeda M, Chen Y, Lee S, Kumagai-Takei N, Yoshitome K, Matsuzaki H, et al. Induction of IL-17 production from human peripheral blood CD4+ cells by asbestos exposure. Int J Oncol. 2017;50(6):2024-32.
- 8. Désage AL, Karpathiou G, Peoc'h M, Froudarakis ME. The Immune Microenvironment of Malignant Pleural Mesothelioma: A Literature Review. Cancers (Basel). 2021;13(13).
- 9. Panou V, Røe OD. Inherited Genetic Mutations and Polymorphisms in Malignant Mesothelioma: A Comprehensive Review. International journal of molecular sciences. 2020;21(12).
- 10. Courtiol P, Maussion C, Moarii M, Pronier E, Pilcer S, Sefta M, et al. Deep learning-based classification of mesothelioma improves prediction of patient outcome. Nature medicine. 2019;25(10):1519-25.
- 11. Carbone M, Yang H, Pass HI, Krausz T, Testa JR, Gaudino G. BAP1 and cancer. Nature reviews Cancer. 2013;13(3):153-9.
- 12. Cheung M, Testa JR. BAP1, a tumor suppressor gene driving malignant mesothelioma. Translational lung cancer research. 2017;6(3):270-8.
- 13. Guo G, Chmielecki J, Goparaju C, Heguy A, Dolgalev I, Carbone M, et al. Whole-Exome Sequencing Reveals Frequent Genetic Alterations in <em>BAP1</em>, <em>NF2</em>, <em>CDKN2A</ em>, and <em>CUL1</em> in Malignant Pleural Mesothelioma. Cancer Res. 2015;75(2):264-9.
- 14. Bueno R, Stawiski EW, Goldstein LD, Durinck S, De Rienzo A, Modrusan Z, et al. Comprehensive genomic analysis of malignant pleural mesothelioma identifies recurrent mutations, gene fusions and splicing alterations. Nature genetics. 2016;48(4):407-16.
- 15. Hmeljak J, Sanchez-Vega F, Hoadley KA, Shih J, Stewart C, Heiman D, et al. Integrative Molecular Characterization of Malignant Pleural Mesothelioma. Cancer discovery. 2018;8(12):1548-65.
- 16. Kiyotani K, Park JH, Inoue H, Husain A, Olugbile S, Zewde M, et al. Integrated analysis of somatic mutations and immune microenvironment in malignant pleural mesothelioma. Oncoimmunology. 2017;6(2):e1278330.
- 17. Vogelzang NJ, Rusthoven JJ, Symanowski J, Denham C, Kaukel E, Ruffie P, et al. Phase III study of pemetrexed in combination with cisplatin versus cisplatin alone in patients with malignant pleural mesothelioma. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2003;21(14):2636-44.
- 18. van Meerbeeck JP, Gaafar R, Manegold C, Van Klaveren RJ, Van Marck EA, Vincent M, et al. Randomized phase III study of cisplatin with or without raltitrexed in patients with malignant pleural mesothelioma: an intergroup study of the European Organisation for Research and Treatment of Cancer Lung Cancer Group and the National Cancer Institute of Canada. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2005;23(28):6881-9.
- 19. Zalcman G, Mazieres J, Margery J, Greillier L, Audigier-Valette C, Moro-Sibilot D, et al. Bevacizumab for newly diagnosed pleural mesothelioma in the Mesothelioma Avastin Cisplatin Pemetrexed Study (MAPS): a randomised, controlled, open-label, phase 3 trial. Lancet. 2016;387(10026):1405-14.
- 20. Scagliotti GV, Gaafar R, Nowak AK, Nakano T, van Meerbeeck J, Popat S, et al. Nintedanib in combination with pemetrexed and cisplatin for chemotherapy-naive patients with advanced malignant pleural mesothelioma (LUME-Meso): a double-blind, randomised, placebocontrolled phase 3 trial. The Lancet Respiratory medicine. 2019;7(7):569-80.
- 21. Webster I, Cochrane JW, Burkhardt KR. Immunotherapy with BCG vaccine in 30 cases of mesothelioma. South African medical journal = Suid-Afrikaanse tydskrif vir geneeskunde. 1982;61(8):277-8.
- 22. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, et al. Improved survival with ipilimumab in patients with metastatic melanoma. The New England journal of medicine. 2010;363(8):711-23.
- 23. Alley EW, Lopez J, Santoro A, Morosky A, Saraf S, Piperdi B, et al. Clinical safety and activity of pembrolizumab in patients with malignant pleural mesothelioma (KEYNOTE-028): preliminary results from a non-randomised, open-label, phase 1b trial. The Lancet Oncology. 2017;18(5):623- 30.
- 24. Metaxas Y, Rivalland G, Mauti LA, Klingbiel D, Kao S, Schmid S, et al. Pembrolizumab as Palliative Immunotherapy in Malignant Pleural Mesothelioma. Journal of thoracic oncology : official publication of the International Association for the Study of Lung Cancer. 2018;13(11):1784-91.
- 25. Quispel-Janssen J, van der Noort V, de Vries JF, Zimmerman M, Lalezari F, Thunnissen E, et al. Programmed Death 1 Blockade With Nivolumab in Patients With Recurrent Malignant Pleural Mesothelioma. Journal of thoracic oncology : official publication of the International Association for the Study of Lung Cancer. 2018;13(10):1569-76.
- 26. Okada M, Kijima T, Aoe K, Kato T, Fujimoto N, Nakagawa K, et al. Clinical Efficacy and Safety of Nivolumab: Results of a Multicenter, Open-label, Single-arm, Japanese Phase II study in Malignant Pleural Mesothelioma (MERIT). Clinical cancer research : an official journal of the American Association for Cancer Research. 2019;25(18):5485-92.
- 27. Scherpereel A, Mazieres J, Greillier L, Lantuejoul S, Do P, Bylicki O, et al. Nivolumab or nivolumab plus ipilimumab in patients with relapsed malignant pleural mesothelioma (IFCT-1501 MAPS2): a multicentre, open-label, randomised, non-comparative, phase 2 trial. The Lancet Oncology. 2019;20(2):239-53.
- 28. Hassan R, Thomas A, Nemunaitis JJ, Patel MR, Bennouna J, Chen FL, et al. Efficacy and Safety of Avelumab Treatment in Patients With Advanced Unresectable Mesothelioma: Phase 1b Results From the JAVELIN Solid Tumor Trial. JAMA oncology. 2019.
- 29. Maio M, Scherpereel A, Calabro L, Aerts J, Cedres Perez S, Bearz A, et al. Tremelimumab as second-line or third-line treatment in relapsed malignant mesothelioma (DETERMINE): a multicentre, international, randomised, double-blind, placebo-controlled phase 2b trial. The Lancet Oncology. 2017;18(9):1261-73.
- 30. Popat S, Curioni-Fontecedro A, Dafni U, Shah R, O'Brien M, Pope A, et al. A multicentre randomised phase III trial comparing pembrolizumab versus single-agent chemotherapy for advanced pre-treated malignant pleural mesothelioma: the European Thoracic Oncology Platform (ETOP 9-15) PROMISE-meso trial. Annals of oncology : official journal of the European Society for Medical Oncology. 2020;31(12):1734-45.
- 31. Fennell DA, Ewings S, Ottensmeier C, Califano R, Hanna GG, Hill K, et al. Nivolumab versus placebo in patients with relapsed malignant mesothelioma (CONFIRM): a multicentre, doubleblind, randomised, phase 3 trial. The Lancet Oncology. 2021.
- 32. Selby MJ, Engelhardt JJ, Johnston RJ, Lu LS, Han M, Thudium K, et al. Preclinical Development of Ipilimumab and Nivolumab Combination Immunotherapy: Mouse Tumor Models, In Vitro Functional Studies, and Cynomolgus Macaque Toxicology. PLoS One. 2016;11(9):e0161779.
- 33. Das R, Verma R, Sznol M, Boddupalli CS, Gettinger SN, Kluger H, et al. Combination therapy with anti-CTLA-4 and anti-PD-1 leads to distinct immunologic changes in vivo. J Immunol. 2015;194(3):950-9.
- 34. Bluthgen MV, Basté N, Recondo G. Immunotherapy combinations for the treatment of patients with solid tumors. Future Oncol. 2020;16(23):1715-36.
- 35. Disselhorst MJ, Quispel-Janssen J, Lalezari F, Monkhorst K, de Vries JF, van der Noort V, et al. Ipilimumab and nivolumab in the treatment of recurrent malignant pleural mesothelioma (INITIATE): results of a prospective, single-arm, phase 2 trial. The Lancet Respiratory medicine. 2019;7(3):260-70.
- 36. Calabro L, Morra A, Giannarelli D, Amato G, D'Incecco A, Covre A, et al. Tremelimumab combined with durvalumab in patients with mesothelioma (NIBIT-MESO-1): an open-label, non-randomised, phase 2 study. The Lancet Respiratory medicine. 2018;6(6):451-60.
- 37. Baas P, Scherpereel A, Nowak AK, Fujimoto N, Peters S, Tsao AS, et al. First-line nivolumab plus ipilimumab in unresectable malignant pleural mesothelioma (CheckMate 743): a multicentre, randomised, open-label, phase 3 trial. Lancet. 2021;397(10272):375-86.
- 38. Hegmans JP, Hemmes A, Hammad H, Boon L, Hoogsteden HC, Lambrecht BN. Mesothelioma environment comprises cytokines and T-regulatory cells that suppress immune responses. The European respiratory journal. 2006;27(6):1086-95.
- 39. Awad MM, Jones RE, Liu H, Lizotte PH, Ivanova EV, Kulkarni M, et al. Cytotoxic T Cells in PD-L1- Positive Malignant Pleural Mesotheliomas Are Counterbalanced by Distinct Immunosuppressive Factors. Cancer immunology research. 2016;4(12):1038-48.
- 40. Minnema-Luiting J, Vroman H, Aerts J, Cornelissen R. Heterogeneity in Immune Cell Content in Malignant Pleural Mesothelioma. International journal of molecular sciences. 2018;19(4).
- 41. Chu GJ, van Zandwijk N, Rasko JEJ. The Immune Microenvironment in Mesothelioma: Mechanisms of Resistance to Immunotherapy. Front Oncol. 2019;9:1366.
- 42. Mantovani A, Sozzani S, Locati M, Allavena P, Sica A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. Trends Immunol. 2002;23(11):549-55.
- 43. Burt BM, Rodig SJ, Tilleman TR, Elbardissi AW, Bueno R, Sugarbaker DJ. Circulating and tumor-infiltrating myeloid cells predict survival in human pleural mesothelioma. Cancer. 2011;117(22):5234-44.
- 44. Cornelissen R, Lievense LA, Maat AP, Hendriks RW, Hoogsteden HC, Bogers AJ, et al. Ratio of intratumoral macrophage phenotypes is a prognostic factor in epithelioid malignant pleural mesothelioma. PLoS One. 2014;9(9):e106742.
- 45. Marcq E, Siozopoulou V, De Waele J, van Audenaerde J, Zwaenepoel K, Santermans E, et al. Prognostic and predictive aspects of the tumor immune microenvironment and immune checkpoints in malignant pleural mesothelioma. Oncoimmunology. 2017;6(1):e1261241.
- 46. Yap TA, Aerts JG, Popat S, Fennell DA. Novel insights into mesothelioma biology and implications for therapy. Nature reviews Cancer. 2017;17(8):475-88.
- 47. Anraku M, Cunningham KS, Yun Z, Tsao MS, Zhang L, Keshavjee S, et al. Impact of tumorinfiltrating T cells on survival in patients with malignant pleural mesothelioma. The Journal of thoracic and cardiovascular surgery. 2008;135(4):823-9.
- 48. Chee SJ, Lopez M, Mellows T, Gankande S, Moutasim KA, Harris S, et al. Evaluating the effect of immune cells on the outcome of patients with mesothelioma. British journal of cancer. 2017;117(9):1341-8.
- 49. Yamada N, Oizumi S, Kikuchi E, Shinagawa N, Konishi-Sakakibara J, Ishimine A, et al. CD8+ tumor-infiltrating lymphocytes predict favorable prognosis in malignant pleural mesothelioma after resection. Cancer immunology, immunotherapy : CII. 2010;59(10):1543-9.
- 50. Yarchoan M, Hopkins A, Jaffee EM. Tumor Mutational Burden and Response Rate to PD-1 Inhibition. The New England journal of medicine. 2017;377(25):2500-1.
- 51. Nghiem P, Bhatia S, Lipson EJ, Sharfman WH, Kudchadkar RR, Brohl AS, et al. Three-year survival, correlates and salvage therapies in patients receiving first-line pembrolizumab for advanced Merkel cell carcinoma. Journal for immunotherapy of cancer. 2021;9(4).
- 52. Galbraith NJ, Wood C, Steele CW. Targeting Metastatic Colorectal Cancer with Immune Oncological Therapies. Cancers (Basel). 2021;13(14).

**1**

- 53. Lo Iacono M, Monica V, Righi L, Grosso F, Libener R, Vatrano S, et al. Targeted next-generation sequencing of cancer genes in advanced stage malignant pleural mesothelioma: a retrospective study. Journal of thoracic oncology : official publication of the International Association for the Study of Lung Cancer. 2015;10(3):492-9.
- 54. Mansfield AS, Peikert T, Smadbeck JB, Udell JBM, Garcia-Rivera E, Elsbernd L, et al. Neoantigenic Potential of Complex Chromosomal Rearrangements in Mesothelioma. Journal of thoracic oncology : official publication of the International Association for the Study of Lung Cancer. 2019;14(2):276-87.
- 55. Muller S, Victoria Lai W, Adusumilli PS, Desmeules P, Frosina D, Jungbluth A, et al. V-domain Igcontaining suppressor of T-cell activation (VISTA), a potentially targetable immune checkpoint molecule, is highly expressed in epithelioid malignant pleural mesothelioma. Mod Pathol. 2020;33(2):303-11.
- 56. Mansfield AS, Roden AC, Peikert T, Sheinin YM, Harrington SM, Krco CJ, et al. B7-H1 expression in malignant pleural mesothelioma is associated with sarcomatoid histology and poor prognosis. Journal of thoracic oncology : official publication of the International Association for the Study of Lung Cancer. 2014;9(7):1036-40.
- 57. Cedres S, Ponce-Aix S, Zugazagoitia J, Sansano I, Enguita A, Navarro-Mendivil A, et al. Analysis of expression of programmed cell death 1 ligand 1 (PD-L1) in malignant pleural mesothelioma (MPM). PLoS One. 2015;10(3):e0121071.
- 58. Nguyen BH, Montgomery R, Fadia M, Wang J, Ali S. PD-L1 expression associated with worse survival outcome in malignant pleural mesothelioma. Asia Pac J Clin Oncol. 2018;14(1):69-73.
- 59. Chapel DB, Stewart R, Furtado LV, Husain AN, Krausz T, Deftereos G. Tumor PD-L1 expression in malignant pleural and peritoneal mesothelioma by Dako PD-L1 22C3 pharmDx and Dako PD-L1 28-8 pharmDx assays. Human pathology. 2019;87:11-7.
- 60. Brosseau S, Danel C, Scherpereel A, Mazieres J, Lantuejoul S, Margery J, et al. Shorter Survival in Malignant Pleural Mesothelioma Patients With High PD-L1 Expression Associated With Sarcomatoid or Biphasic Histology Subtype: A Series of 214 Cases From the Bio-MAPS Cohort. Clin Lung Cancer. 2019;20(5):e564-e75.
- 61. Brcic L, Klikovits T, Megyesfalvi Z, Mosleh B, Sinn K, Hritcu R, et al. Prognostic impact of PD-1 and PD-L1 expression in malignant pleural mesothelioma: an international multicenter study. Translational lung cancer research. 2021;10(4):1594-607.
- 62. Tumeh PC, Harview CL, Yearley JH, Shintaku IP, Taylor EJ, Robert L, et al. PD-1 blockade induces responses by inhibiting adaptive immune resistance. Nature. 2014;515(7528):568-71.
- 63. Herbst RS, Soria JC, Kowanetz M, Fine GD, Hamid O, Gordon MS, et al. Predictive correlates of response to the anti-PD-L1 antibody MPDL3280A in cancer patients. Nature. 2014;515(7528):563- 7.
- 64. Kluger HM, Zito CR, Turcu G, Baine MK, Zhang H, Adeniran A, et al. PD-L1 Studies Across Tumor Types, Its Differential Expression and Predictive Value in Patients Treated with Immune Checkpoint Inhibitors. Clinical cancer research : an official journal of the American Association for Cancer Research. 2017;23(15):4270-9.
- 65. Fridman WH, Pages F, Sautes-Fridman C, Galon J. The immune contexture in human tumours: impact on clinical outcome. Nature reviews Cancer. 2012;12(4):298-306.
- 66. Bruni D, Angell HK, Galon J. The immune contexture and Immunoscore in cancer prognosis and therapeutic efficacy. Nature reviews Cancer. 2020;20(11):662-80.
- 67. Hu-Lieskovan, Lisberg S, Zaretsky A, Grogan JM, Rizvi TR, Wells H, et al. Tumor Characteristics Associated with Benefit from Pembrolizumab in Advanced Non–Small Cell Lung Cancer. Clinical Cancer Research. 2019;25(16):5061 - 8.
- 68. Ujiie H, Kadota K, Nitadori JI, Aerts JG, Woo KM, Sima CS, et al. The tumoral and stromal immune microenvironment in malignant pleural mesothelioma: A comprehensive analysis reveals prognostic immune markers. Oncoimmunology. 2015;4(6):e1009285.
- 69. Thapa B, Salcedo A, Lin X, Walkiewicz M, Murone C, Ameratunga M, et al. The Immune Microenvironment, Genome-wide Copy Number Aberrations, and Survival in Mesothelioma. Journal of thoracic oncology : official publication of the International Association for the Study of Lung Cancer. 2017;12(5):850-9.
- 70. Brockwell NK, Alamgeer M, Kumar B, Rivalland G, John T, Parker BS. Preliminary study highlights the potential of immune checkpoint inhibitors in sarcomatoid mesothelioma. Translational lung cancer research. 2020;9(3):639-45.
- 71. Pasello G, Zago G, Lunardi F, Urso L, Kern I, Vlacic G, et al. Malignant pleural mesothelioma immune microenvironment and checkpoint expression: correlation with clinical-pathological features and intratumor heterogeneity over time. Annals of oncology : official journal of the European Society for Medical Oncology. 2018;29(5):1258-65.
- 72. Kamphorst AO, Pillai RN, Yang S, Nasti TH, Akondy RS, Wieland A, et al. Proliferation of PD-1+ CD8 T cells in peripheral blood after PD-1-targeted therapy in lung cancer patients. Proc Natl Acad Sci U S A. 2017;114(19):4993-8.
- 73. Gros A, Parkhurst MR, Tran E, Pasetto A, Robbins PF, Ilyas S, et al. Prospective identification of neoantigen-specific lymphocytes in the peripheral blood of melanoma patients. Nature medicine. 2016;22(4):433-8.
- 74. Ferrucci PF, Ascierto PA, Pigozzo J, Del Vecchio M, Maio M, Antonini Cappellini GC, et al. Baseline neutrophils and derived neutrophil-to-lymphocyte ratio: prognostic relevance in metastatic melanoma patients receiving ipilimumab. Annals of oncology : official journal of the European Society for Medical Oncology. 2016;27(4):732-8.
- 75. Weide B, Martens A, Hassel JC, Berking C, Postow MA, Bisschop K, et al. Baseline Biomarkers for Outcome of Melanoma Patients Treated with Pembrolizumab. Clinical cancer research : an official journal of the American Association for Cancer Research. 2016;22(22):5487-96.
- 76. Mezquita L, Auclin E, Ferrara R, Charrier M, Remon J, Planchard D, et al. Association of the Lung Immune Prognostic Index With Immune Checkpoint Inhibitor Outcomes in Patients With Advanced Non-Small Cell Lung Cancer. JAMA oncology. 2018;4(3):351-7.
- 77. Chapman EA, Thomas PS, Stone E, Lewis C, Yates DH. A breath test for malignant mesothelioma using an electronic nose. The European respiratory journal. 2012;40(2):448-54.

**1**

- 78. Dragonieri S, van der Schee MP, Massaro T, Schiavulli N, Brinkman P, Pinca A, et al. An electronic nose distinguishes exhaled breath of patients with Malignant Pleural Mesothelioma from controls. Lung Cancer. 2012;75(3):326-31.
- 79. Lamote K, Brinkman P, Vandermeersch L, Vynck M, Sterk PJ, Van Langenhove H, et al. Breath analysis by gas chromatography-mass spectrometry and electronic nose to screen for pleural mesothelioma: a cross-sectional case-control study. Oncotarget. 2017;8(53):91593-602.
- 80. Brusselmans L, Arnouts L, Millevert C, Vandersnickt J, van Meerbeeck JP, Lamote K. Breath analysis as a diagnostic and screening tool for malignant pleural mesothelioma: a systematic review. Translational lung cancer research. 2018;7(5):520-36.
- 81. Behera B, Joshi R, Anil Vishnu GK, Bhalerao S, Pandya HJ. Electronic nose: a non-invasive technology for breath analysis of diabetes and lung cancer patients. Journal of breath research. 2019;13(2):024001.
- 82. de Vries R, Muller M, van der Noort V, Theelen W, Schouten RD, Hummelink K, et al. Prediction of response to anti-PD-1 therapy in patients with non-small-cell lung cancer by electronic nose analysis of exhaled breath. Annals of oncology : official journal of the European Society for Medical Oncology. 2019;30(10):1660-6.
- 83. Buma AIG, Muller M, de Vries R, Sterk PJ, van der Noort V, Wolf-Lansdorf M, et al. eNose analysis for early immunotherapy response monitoring in non-small cell lung cancer. Lung Cancer. 2021;160:36-43.