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Review

Cardiac and Vascular α_1 -Adrenoceptors in Congestive Heart Failure: A Systematic Review

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Abstract: As heart failure (HF) is a devastating health problem worldwide, a better understanding and the development of more effective therapeutic approaches are required. HF is characterized by sympathetic system activation which stimulates α - and β -adrenoceptors (ARs). The exposure of the cardiovascular system to the increased locally released and circulating levels of catecholamines leads to a well-described downregulation and desensitization of β -ARs. However, information on the role of α -AR is limited. We have performed a systematic literature review examining the role of both cardiac and vascular α_1 -ARs in HF using 5 databases for our search. All three α_1 -AR subtypes (α_{1A} , α_{1B} and α_{1D}) are expressed in human and animal hearts and blood vessels in a tissue-dependent manner. We summarize the changes observed in HF regarding the density, signaling and responses of α_1 -ARs. Conflicting findings arise from different studies concerning the influence that HF has on α_1 -AR expression and function; in contrast to β -ARs there is no consistent evidence for down-regulation or desensitization of cardiac or vascular α_1 -ARs. Whether α_1 -ARs are a therapeutic target in HF remains a matter of debate.

Keywords: α_1 -adrenoceptors; cardiomyocyte; blood vessel; heart failure

1. Introduction

The sympathetic nervous system primarily regulates cardiac function via β -adrenoceptors (β -ARs) [1,2], whereas α_1 -adrenoceptors (α_1 -ARs) play an important role in the regulation of vascular tone [3]. Yet, α_1 -ARs are also expressed in various cell types of the heart and can contribute to the regulation of cardiac contraction [4,5], rhythm [6,7] and hypertrophy [8]. However, the role of cardiac α_1 -ARs appears to be species-dependent with a considerable contribution in rodents but a much smaller (if any) contribution in the healthy human heart [1]. On the other hand, cardiac expression of other subtypes such as β_3 -ARs also appears to be lower in the human heart than in that of other species, but such receptors become up-regulated in congestive heart failure (CHF) [9].

The sympatho-adrenal system is activated in acute and chronic heart failure (HF). Accordingly, plasma levels of noradrenaline and its co-transmitters are increased in CHF [10,11]. This appears to correlate with a higher risk of mortality [12]. In line with this, patients with reduced left ventricular function appear to be more prone to post-operative vasoplegia that additionally increases the mortality and morbidity risk [13,14]. The regulation of β -ARs in CHF has been reviewed extensively and appears distinct for the three β -AR subtypes [1].

α_1 -ARs are also a subfamily with three members: α_{1A} , α_{1B} and α_{1D} [15]. Data outside the cardiovascular system suggest that they can undergo differential regulation upon extended agonist exposure [16]. Nevertheless, until now, the α_1 -AR regulation in HF has not been fully elucidated and results might vary a lot between different studies. Against this background we discuss the expression and function of α_1 -ARs and their subtypes in the heart and vasculature of patients with CHF and animal models thereof based upon a systematic review of the literature.

2. Materials and Methods

A broad literature search was performed in 5 databases (PubMed/MEDLINE, Embase, Web of Science, Cochrane Library, Emcare). The search was performed using the MeSH terms “Heart failure” and “Receptors, Adrenergic alpha 1” and by combining each term with different keyword variations and free text words. No language limits were applied. The final search was performed on the 15th of April 2020. Details of the search strategy are provided in the Supplementary Materials.

Animal and human studies were included that investigated changes of cardiac and/or vascular α_1 -AR expression or function in CHF. There were no limitations regarding the study design or the etiology/model of CHF. Two independent reviewers (GKM and OP) assessed all articles for eligibility that the search strategy had yielded. The first screening was performed by assessing only the titles and abstracts of the articles, while the final decision for inclusion was based upon reading the full text of the selected articles. The reference lists of all relevant articles and reviews were manually checked for inclusion of additional eligible studies that were not identified by the primary search strategy. Any disagreements that emerged during the study selection process were resolved by discussion. A Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA, www.prisma-statement.org) flowchart is shown as Figure 1.

The data from the eligible studies were retrieved and recorded in predefined Google Spreadsheet forms that offer the possibility of auditing. Data collection included study characteristics and study design information and changes that were identified regarding the expression and function of α_1 -ARs in cells, tissue and in vivo.

3. Considerations Related to Data Interpretation

Most investigators have reported mRNA levels after normalization to a reference gene, often glyceraldehyde-3-phosphate dehydrogenase (GADPH) [17]. However, they typically failed to document whether the expression of the reference gene was stable. A stable expression of reference genes cannot necessarily be expected [18]. Specifically, studies in multiple cell types and tissues [19] including the heart [2] have found that exposure to β -AR agonists can down-regulate GAPDH expression; based on the elevated plasma catecholamine levels in CHF [10,11] this is likely to be of relevance for the mRNA data discussed below.

Cardiac α_1 -AR expression at the protein level has primarily been studied by radioligand binding and antibody-based techniques. While radioligands such as [3 H]-prazosin or [125 I]-BE 2254 (also known as [125 I]-HEAT) have high specificity for α_1 -ARs, they do not discriminate between subtypes and, particularly in the case of [3 H]-prazosin, are not very sensitive. In contrast, antibody-based approaches such as immunoblotting or immunohistochemistry potentially can be subtype-selective and be used to examine distribution within a tissue. However, multiple studies have demonstrated that the vast majority of commercially available α_1 -AR subtype antibodies lack target specificity when tested under stringent conditions [20–22]. These limitations need to be considered in the interpretation of the subsequently discussed studies.

Another important factor that might influence the results of different studies is the exact location of α_1 -ARs in the peripheral vasculature. It has been demonstrated that α_1 -AR receptors are located not only on smooth muscle cells, but, also on endothelial, adventitial and nerve cells [23,24]. This is a rather important finding that should be taken into account when interpreting results from expression and functional studies, since the total α_1 -AR mRNA and the actual role of specific α_1 -subtypes mediating vascular responses might differ depending on the specific cell on which they are expressed.

CHF can be caused by different pathologies, for instance by chronic myocardial ischemia, valvular heart disease or viruses, or can be idiopathic. As CHF in humans can be caused by a variety of pathophysiologic mechanisms, it is important to realize that subjects within an animal model typically are more homogeneous than patient cohorts; on the other hand, each model reflects a different cause and, potentially, a different pathophysiology. Among the many reported animal models of CHF, post-myocardial infarction [25–27], cardiac pacing causing tachycardiomyopathy [28], drug-induced [29,30] and hereditary models [31] have been applied to studies of cardiovascular α_1 -ARs. Finally, pathophysiology and regulation of α_1 -ARs may not only differ between patients and experimental models but also between species. While most animal studies related to cardiovascular α_1 -ARs in CHF were done with rats [2,26,32], guinea pigs [33,34], mice [27,29], hamster [35], cats [36] and dogs [37,38] have also been used.

4. Results

4.1. Study Selection

The literature search resulted in 1461 unique, original articles. G.K.-M. and O.P. independently screened the titles and abstracts of all the articles and excluded 1312 articles because of lack of relevance. After assessing the full text of the remaining 149 potentially relevant articles, and resolving any disagreements, the two reviewers decided to include 65 articles (Figure 1). Additional relevant studies that were retrieved from other articles were also included. This study selection process led to an inclusion of 33 unique reports on cardiac and 31 unique reports on vascular α_1 -AR. One study was reviewed for both cardiac and vascular α_1 -ARs leading to a total number of included studies of 65.

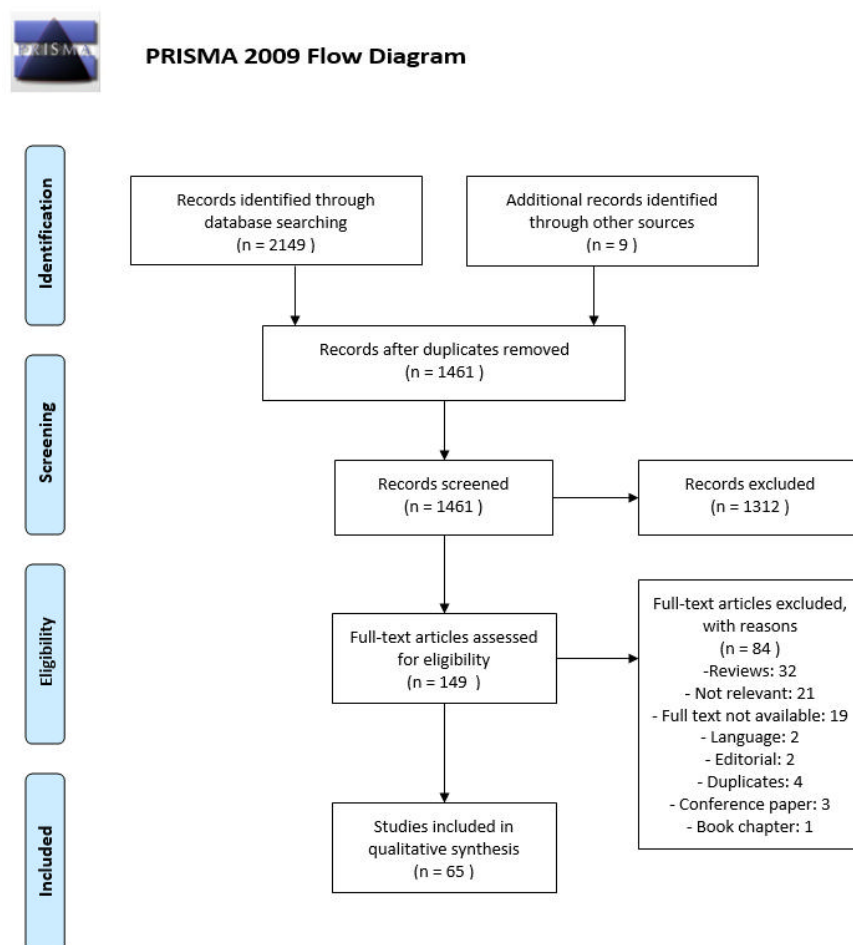


Figure 1. Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) study flow chart.

4.2. Cardiac α_1 -AR

4.2.1. mRNA Expression

There are three α_1 -AR subtypes: α_{1A} -, α_{1B} - and α_{1D} -ARs which have been identified in both animal and human hearts at the mRNA level [1,39,40]. All studies agree that α_{1A} -AR is the most abundant subtype in the heart but the presence of α_{1B} - and α_{1D} -AR mRNA were also confirmed [1]. Although animal hearts express more α_1 -AR in the heart, human and rodent hearts are reported to locate α_1 -AR subtypes similarly, as α_{1A} - and α_{1B} -subtypes are predominantly found in myocardium whereas the α_{1D} -subtype is expressed in coronary arteries [40,41]. Myagmar et al. claimed that α_{1A} -ARs are present in a myocyte subset while α_{1B} -ARs are in all cells [42]. Several studies reported changes in α_1 -AR subtype mRNA levels in CHF, but the results are heterogeneous (Table 1).

Table 1. Summary of the changes in cardiac mRNA studies for total α_1 -adrenoceptors (AR) and their subtypes in chronic heart failure.

mRNA	Species	Tissue	α_{1A} -AR	α_{1B} -AR	α_{1D} -AR	Total α_1 -AR
Fischer et al. [43]	human	LV, RV, LA, RA	↑	↑	-	n/a
Jensen et al. [41]	human	LV and RV	↑	X	X	X
Monto et al. [17]	human	LV and RV	X	X	X	n/a
Brattelid et al. [2]	rat	LV	X	X	↓	n/a

↑: increased; ↓: decreased; X: unchanged; n/a: no data available; LV, left ventricle; RV, right ventricle; LA, left atrium; RA, right atrium.

In human failing hearts, Fischer et al. demonstrated that mRNA of α_{1A} - and α_{1B} -ARs in the left ventricle was increased while no change in mRNA levels was observed in other cardiac regions [43]. Similarly, Jensen et al. reported that α_{1A} mRNA was increased in failing left ventricle LV and tended to increase in failing RV, but there was no change in α_{1B} and α_{1D} mRNA in any region [41]. On the other hand, Monto et al. did not confirm an altered expression of the three subtypes in human failing hearts; however, the relative abundance increased when considering the reduced β -AR expression [17]. They suggested that expression of α_{1A} -ARs in the LV correlates with the left ventricular ejection fraction (LVEF) suggesting that this subtype may contribute to maintain cardiac inotropy in the failing heart. Moreover, a study in a rat coronary ligation model of CHF reported α_{1D} -AR mRNA to be decreased; a numerical reduction of α_{1A} - and α_{1B} -AR mRNA did not reach statistical significance [2].

4.2.2. Protein Expression

Cardiac α_1 -ARs were first identified at the protein level in rats by Williams and Lefkowitz [44] and thereafter confirmed in many species. Their quantitative expression at the protein level ranges from a relatively high expression in rodents to a low expression in humans, where their role appears considerably lower than that of β -ARs [1,39,40]. Despite numerous studies, data on α_1 -AR binding properties in human HF have remained inconsistent (Table 2). The density of α_1 -ARs was reported as increased [45–48], unaltered [41,45,49–52], or decreased [43,53].

Table 2. Summary of cardiac α_1 -adrenoceptor (AR) protein expression in chronic heart failure.

Binding	Species	Tissue	α_{1A} -AR	α_{1B} -AR	α_{1D} -AR	Total α_1 -AR
Böhm et al. [49]	human	papillary	n/a	n/a	n/a	X
Bristow et al. [50]	human	LV and RV	n/a	n/a	n/a	X
Corr et al. [36]	cat	ventricle	n/a	n/a	n/a	↑
Dixon et al. [54]	rat	LV	n/a	n/a	n/a	↑
Erdmann and Böhm [51]	human	papillary	n/a	n/a	n/a	X
Fischer et al. [43]	human	LV, RV, LA, RA	↓	↓	↓	n/a
Gopalakrishnan et al. [32]	rat	LV and RV	n/a	n/a	n/a	X
Hwang et al. [45]	human	explanted hearts	n/a	n/a	n/a	↑ (in ICM) X (in DCM)
Jensen et al. [41]	human	LV and RV	X (↑ in RV)	X	undetected	X
Karliner et al. [33]	GP	whole heart	n/a	n/a	n/a	↑
Limas et al. [52]	human	LV and RV	n/a	n/a	n/a	↓ (in severe HF) X (in mild)
Litwin et al. [25]	rat	LV	n/a	n/a	n/a	X
Maisel et al. [34]	GP	LV	n/a	n/a	n/a	
Meggs et al. [26]	rat	LV and RV	n/a	n/a	n/a	X
Re et al. [37]	dog	LV, RV, LA, RA	n/a	n/a	n/a	↓
Shi et al. [53]	human	LV	↓	X	n/a	↓
Sjaastad et al. [55]	rat	LV and RV	n/a	n/a	n/a	X
Steinfath et al. [46]	human	LV	n/a	n/a	n/a	↑
Watanabe et al. [35]	hamster	whole heart	n/a	n/a	n/a	Slight↓
Zhao et al. [30]	rat	RA and LV	n/a	n/a	n/a	↓
Rowley et al. [56]	rat	atria	n/a	n/a	n/a	X
Vago et al. [47]	human	LV	n/a	n/a	n/a	↑

GP: guinea pig; ↑: increased; ↓: decreased; X: unchanged; n/a: no data available; LV, left ventricle; RV, right ventricle; LA, left atrium; RA, right atrium.

However, even a lack of change in α_1 -AR protein expression may be biologically relevant for two reasons: Firstly, it is interesting to note that expression does not consistently decrease despite exposure to elevated plasma catecholamines [10,11]. However, studies outside the cardiovascular system have found that prolonged agonist exposure does not always lead to down-regulation of α_1 -AR subtypes and in some cases can even be associated with an up-regulation [16]. Secondly, an unaltered expression of α_1 -ARs in face of a consistently found down-regulation of β -ARs [1,39] implies a change in the relative role of the two adrenoceptor sub-families, which in its own right may have implications for the regulation of cardiac function [50,55].

A possible explanation for the controversial results on the regulation of cardiac α_1 -AR mRNA and protein is heterogeneity based on various etiologies of CHF. For instance, it was reported that α_1 -AR protein was increased in ischemic human cardiomyopathy but remained constant in dilated cardiomyopathy within the same study [45]. Similarly, the degree of HF may contribute to heterogeneous findings. While α_1 -AR density was not changed in a mild-to-moderate heart failure group, it was decreased in severely failing human hearts [52]. Finally, the change in α_1 -AR density may depend on the cardiac region under investigation: α_{1A} - and α_{1B} -AR protein were not changed in failing left ventricle, but α_{1A} -AR binding was increased in right ventricle [41].

Studies in rats mainly found that α_1 -AR density remained constant after myocardial infarction [25,26,32,55,56], which is in line with the lack of a consistent change in human CHF (Table 2). However, an increase was also reported in a rat model of myocardial infarction [54]. Noradrenaline infusion, on the other hand, resulted in a decrease in α_1 -AR density in rats [30], demonstrating that cardiac α_1 -ARs are in principle susceptible to agonist-induced down-regulation.

There is also a substantial number of studies investigating the changes in cardiac α_1 -ARs using different animal models. Cardiomyopathic hamster [35] and dog [37] hearts were shown to have

decreased α_1 -AR concentrations. In contrast, an increase in cardiac α_1 -AR density was demonstrated in a guinea pig model of HF caused by aortic constriction [33]. In addition, increase α_1 -AR densities were reported in cat [36] and guinea pig [34] ischemia models following coronary ligation.

4.2.3. α_1 -AR Subtypes

For the interpretation of subtype expression data, there is a technical issue between α_1 -AR mRNA and protein levels. While mRNA of all α_1 -AR subtypes were detected in animal and human hearts, binding studies demonstrated the presence of only α_{1A} - and α_{1B} -subtypes, but not α_{1D} -AR [39,41,57]. Lack of α_{1D} -AR expression at the protein level has been shown in many tissues expressing corresponding mRNA [58].

Animal species do not only differ from humans by expressing more α_1 -AR in the heart but also by exhibiting a different subtype expression profile. For instance, rat [59] and mouse [58] hearts were reported to express α_{1A}/α_{1B} -AR in an approximate 20/80 ratio. Moreover, previous studies suggest that the three subtypes mediate different effects in the heart. α_{1A} -AR has been reported to mediate positive inotropy in right ventricles of human hearts and this response has been found to be reduced only slightly in failing hearts. Thus, α_{1A} -subtype-mediated inotropy is suggested to be significant source of inotropic support [60]. However, in nonfailing murine hearts, stimulation by the α_{1A} -AR selective agonist, A61603 or the non-subtype-selective α_1 -AR agonist, phenylephrine mediated negative inotropic effects in right ventricle which was switched to positive inotropy in failing hearts [29]. On the other hand, the inotropic response mediated by α_{1B} -subtype remained negative and unaltered. In the light of these findings, the authors suggested that α_{1A} -subtype may play a role in improving contraction in RV failure and might be a therapeutic target. Another study using transgenic mice with heart-specific overexpression of α_{1A} -AR displayed enhanced inotropy after myocardial infarction [61]. Moreover, α_{1D} -AR mRNA decreased in rats after coronary ligation; however, this study was inconclusive because there was a non-significant trend for reduced mRNA of α_{1A} - and α_{1B} -ARs [2]. Thus, the authors proposed that a decrease in α_{1D} -AR might be a protective response of the failing heart. However, the relevance of these data remains unclear because studies in several mouse tissues have demonstrated that many tissues express α_{1D} -AR at the mRNA but not at the protein level [58].

Patients who receive left ventricular assist device to unload the failing heart and improve cardiac function were shown to have a redistribution of α_{1A} -AR from a peri-myocytic to an intra-myocytic location along with a rise in the overall density of receptors [62]. On the other hand, α_{1A} -AR subtype was decreased in human dilated cardiomyopathy and this result was concluded to be consistent with the theory that α_{1A} -AR agonism may be cardioprotective [53]. Fischer et al. demonstrated that α_{1A} - and α_{1B} -AR mRNA levels in left ventricle of human failing hearts were increased, although α_{1A} -, α_{1B} - and α_{1D} -AR binding sites were decreased [43]. They suggested that a decrease of α_{1B} -AR can have cardioprotective effects while a decrease of α_{1A} -AR is more likely the result of impaired heart function.

4.2.4. Signaling Pathways Involved

A schematic representation of α_1 -AR signal transduction is shown in Figure 2. The prototypical signaling pathway of α_1 -ARs involves coupling to G_q with subsequent activation of phospholipase C (PLC), which releases Ca^{2+} from intracellular stores and activates protein kinase C (PKC) [63]. However, cardiac α_1 -ARs can also couple to G_i [5,7], $G_{12/13}$ [64], even G_s proteins [65] and G_h which is a GTP-binding protein with Transglutaminase activity [66] and to pathways such as inhibition of adenylyl cyclase [67], modulation of Ca^{2+} , K^+ and Cl^- channel activity [7,68,69], ras activity [70] or activation of various mitogen activated protein kinases [71]. Most of these pathways have also been studied in human CHF or CHF animal models [72].

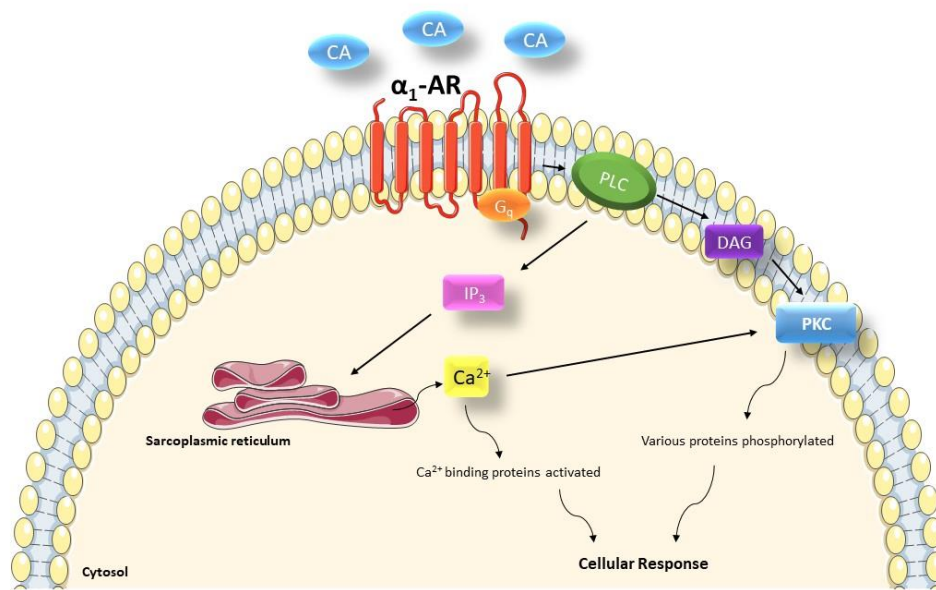


Figure 2. The prototypical signaling pathway of α_1 -ARs. CA: catecholamines, DAG: diacylglycerol, IP_3 : inositol triphosphate, PKC: protein kinase C, PLC: phospholipase C.

Activation of G_q -signaling pathways mediates the development of hypertrophy and the transition to HF [73]. Overexpression of G_q was demonstrated to result in marked hypertrophy [74] and HF [75]. Moreover, inhibition of G_q -mediated signaling was shown to inhibit cardiac dysfunction [76,77]. On the other hand, multiple studies in humans and several animal species have shown that HF is accompanied by an increased expression of G_i at the mRNA and protein level, whereas a reduced expression of G_s has been found less consistently [78]. Reduced GTP binding with Transglutaminase activity of $G\alpha_h$ were found in human HF [45]. α_1 -AR coupling to a GTP binding protein was detected only in pathological, but not in healthy human hearts [47].

α_1 -AR-mediated phosphoinositol turnover is not changed in both animal and human studies when accompanied by an unaltered receptor density in human [50] and rat [56] HF. On the other hand, in a rat model of myocardial infarction, an increase in phosphoinositol turnover was reported along with an increase in G-protein content and an upregulation of α -skeletal actin which suggests an involvement of α_1 -ARs in the reactive hypertrophic response [26].

α_1 -AR stimulation increases both the intracellular Ca^{2+} concentration, which activates muscle contraction, but also the Ca^{2+} sensitivity of cardiomyocytes. α_1 -AR-mediated Ca^{2+} -transients and phosphorylation of cardiac myosin regulatory light chain were increased in a mouse right ventricular HF model [29]. The increase in Ca^{2+} sensitivity expected from these observations was reported in a canine HF model, apparently occurring via the G_q -RhoA-ROCK signaling pathway [38]. Similarly, the switch from negative to positive inotropy observed in mouse failing right ventricles (see below) was claimed to be mediated through a pathway involving increased myofilament Ca^{2+} sensitivity [27].

4.2.5. Cell and Tissue Responses

α_1 -AR stimulation induces a positive inotropic effect in isolated cardiac preparations such as atria, left and right ventricles (Table 3) [25,55,56,60]. The positive inotropic effect of α_1 -AR stimulation involves multiple subtypes, but mostly occurs via α_{1B} -AR in rodents; when these become inactivated in vitro, the α_{1A} -component becomes stronger and compensates for the loss of α_{1B} response resulting in overall maintained contractile responses [79]. However, depending on experimental conditions α_1 -AR stimulation can also have negative inotropic effects (Table 3) [29]. Moreover, stimulation of cardiac α_1 -AR can increase heart rate [56], but systemic stimulation can reduce heart rate secondary to

an increase in total peripheral resistance and activation of the baroreflex [80], which can mask direct chronotropic effects of α_1 -AR stimulation.

Table 3. Summary of changes in cardiac α_1 -adrenoceptor (AR)-mediated responses in chronic heart failure.

	Species	Preparation	α_1 -AR-mediated Responses
Böhm et al. [49]	human	papillary	X
Erdmann and Böhm [51]	human	papillary	X
Litwin et al. [25]	rat	papillary	X
Sjaastad et al. [55]	rat	papillary	X
Cowley et al. [29]	mouse	trabecula	α_{1A} -med $\uparrow$ α_{1B} -med X
Janssen et al. [60]	human	trabecula	slight $\downarrow$
Steinfath et al. [46]	human	trabecula	$\downarrow$
Wang et al. [27]	mouse	trabecula	in RV switch from NIE to PIE in LV X
Skomedal et al. [81]	human	trabecula	PIE
Rowley et al. [56]	rat	atria	$\downarrow$

$\uparrow$: increased; $\downarrow$: decreased; X: unchanged; RV: right ventricle; LV: left ventricle; NIE: negative inotropic effect; PIE: positive inotropic effect.

The chronotropic and inotropic responses to the α_1 -AR agonist phenylephrine were decreased in isolated right and left atria of rats with myocardial infarction [56]. As this was not accompanied by alterations of receptor density or phosphoinositol turnover, this may reflect changes of post-receptor signaling (see above). In contrast, several studies in isolated papillary muscle from humans [49,51] or rats [25,55] reported unaltered inotropic responses in CHF, which was in line with unaltered receptor density reported in these studies. The positive inotropic response to α_{1A} -AR stimulation in human right ventricle also exhibited little change in HF in two studies [60,81]. In contrast, the phenylephrine-induced positive inotropic response in human CHF was abolished in another study [46]. In mouse models, negative inotropic effect observed in control animals was switched to positive inotropy in the failing right ventricles [29]. However PE-mediated positive inotropy in LV of nonfailing hearts remained unchanged in failing mouse hearts after myocardial infarction [27]. Cardiomyopathic hearts of Syrian hamsters were shown to have increased inotropy accompanied by increased Ca^{2+} levels and pertussis toxin-sensitive G-proteins [82]. Thus, with few exceptions, most studies reported little change in inotropic responses in the failing heart across species and models. This may nonetheless be biologically relevant because it would indicate an α_1/β -AR balance in light of the consistently found reduction in β -adrenergic function in CHF [50,55].

Moreover, there are opposing views on the role of α_1 -AR subtypes on hypertrophy. While some suggest that α_{1A} -AR were found to mediate hypertrophy development in neonatal rat cardiomyocytes in vitro [8], transgenic overexpression of α_{1B} -AR caused a hypertrophic response resulting in HF, while α_{1A} -AR overexpression did not affect hypertrophy and did not hasten development of HF [29,83].

4.3. Vascular α_1 -AR

4.3.1. mRNA Expression

Multiple studies have explored the expression of α_1 -AR subtype mRNA in arteries and veins of various species including rat [84–88], rabbit [89–91] and human [89,92–94]. It appears from these and many other studies, that all three α_1 -AR subtypes are expressed in the vasculature of these species in a species- and vascular bed-dependent manner. This conclusion is in line with studies on the

role of α_1 -AR subtypes in functional studies [3]. Our search did not identify any animal studies reporting on the regulation of vascular α_1 -AR subtype mRNA in CHF. However, it is likely that such regulation occurs because CHF is characterized by increased circulating catecholamine levels and the endogenous agonist noradrenaline has been shown to transiently down-regulate α_1 -AR mRNA in rabbit aortic smooth muscle cells whereas the down-regulation of α_1 -AR protein was longer lasting (at least 96 h) [95]. Moreover, CHF can be associated with hypoperfusion and hypoxia of tissues and vascular α_1 -AR mRNA expression can be regulated by hypoxia [96]. Vascular α_1 -AR mRNA expression is also regulated by age [97]. HF did not appear to affect the total levels of α_1 -AR mRNA in human coronary arteries [98]. On the contrary, the use of β -blockers was associated with a significant decrease in both α_{1D} and total α_1 -AR mRNA levels. α_{1D} -AR appeared to be the principal receptor in human epicardial coronary arteries, which resembles the mouse coronary arteries [99,100].

4.3.2. Protein Expression

Quantitative, ligand-binding studies have been performed in order to assess the α_1 -AR distribution in the various vascular beds of different species. Existing discrepancies might be due to interspecies differences, variant α_1 -AR subtype distribution in the peripheral vasculature, differences in models of HF, for example pacing-induced HF versus myocardial infarction, or even due to poorly reproducible results.

Such studies have been performed in animals and humans without coexisting diseases in order to assess the physiological α_1 -ARs at the protein level. In humans a considerable difference in α_1 -AR protein expression has been demonstrated between somatic (mammary artery) and splanchnic arteries [97]. However, this was not true for veins, in which the α_1 -AR density seemed to be independent from differences in vessel diameter. This might be explained by the importance of splanchnic circulation, especially during a situation of hypovolemic shock. Sympathetic stimulation of α_1 -ARs in those vascular territories leads to maintenance of blood flow and O₂ delivery to vital organs. On the other hand, constriction of conduit arteries and veins contribute to the preservation of blood pressure and cardiac afterload. In human epicardial coronary arteries 8.7 fmol/mg protein of total α_1 -ARs were found, of which 75% constituted of the α_{1D} subtype, almost twice as much as in the left ventricle [41,98].

Considering that progression of HF is associated with a decrease in β -AR protein in the heart, one could argue that similar findings would be expected for α_1 -ARs. On the contrary, α_1 -AR density has been reported to be increased, yet not statistically significant, in studies with myocardial infarction-induced CHF in rats [101,102]. However, the lack of statistical significance of this change leads to rather inconclusive results.

4.3.3. α_1 -AR Subtypes

All three subtypes have been identified in the peripheral vasculature of various species, however the subtype distribution varies between animals and humans, between different vascular beds and with age. For instance, α_{1B} and α_{1D} subtype mRNAs have been detected in human aorta [93] and are considered to mediate vascular constriction. A controversy exists in the rat aorta since some studies have identified only the subtype α_{1D} [103–105], while others report both α_{1B} and α_{1D} -ARs [101]. The α_{1B} subtype has been recognized to mediate vasoconstriction after sympathetic stimulation in the mouse, rabbit and dog aorta [106–108]. Similarly, mesenteric responses in humans have been found to be mediated by all three α_1 -AR subtypes [109], whereas in rats mesenteric vascular constriction is mediated by α_{1D} and α_{1B} subtypes in superior mesenteric artery and the mesenteric resistance arteries respectively [101,104]. In the small mesenteric arteries the subtype α_{1A} has also been identified [101,110]. The same receptor was found to mediate contractile responses in rat small renal arteries [111].

Similarities in vascular tone regulation between different species also exist. As it was mentioned before, α_{1D} is recognized as the main subtype that mediate coronary contraction in humans [98] and mice [99,100] and the same has also been reported in rats [105]. In this latter study, it was additionally

shown that CHF did not modify the function of α_{1D} -AR, neither in the carotid artery nor in the aorta, whereas the α_{1D} -AR-mediated vasoconstriction was enhanced in older animals.

Furthermore, blood pressure regulation in mice is considered to be achieved via the stimulation of all three subtypes. This is confirmed in knockout mice studies in which deletion of any subtype results in a decrease in resting blood pressure [106,112,113].

The heterogeneous α_1 -AR population in the various vascular beds might be modified depending on the disease state. In support of this, antagonist studies have explicitly displayed an alteration of the antagonism against α_1 -ARs before and after the onset of CHF [114–117] and showed changes from competitive to non-competitive antagonism, or the other way around, which, however, was also related to the specific agonist and vessel studied. Distinct patterns of antagonism have been identified in different disease states and in different vessels. The specific subtype populations, however, and their involvement in the vascular responses in different disease settings are still to be elucidated.

4.3.4. Signaling Pathways Involved

Changes in α_1 -AR responsiveness due to chronic exposure of the peripheral vasculature to the endogenously released catecholamines might also derive from alterations in signal transduction. These changes might be due to modification of Ca^{2+} sensitivity and/or other changes in the signaling pathways involved. For example, the continuous exposure of smooth muscle cells from rabbit aorta to α_1 -AR agonists led to a major loss in sensitivity, which did not seem to be due to down-regulation of α_1 -ARs, but rather to blunting of phosphatidylinositol turnover [118].

Disease progression might also modify the dependency of α_1 -AR-mediated vascular contraction on extracellular Ca^{2+} which seems to increase in end-stage HF [119,120]. More precisely, the existence of a high affinity component of the α_1 -AR-mediated responses was identified which is inhibitable by a PKC inhibitor, polymyxin B, and which becomes apparent with the progression of HF. Interestingly, in this study, the dorsal pedal artery presented lower dependency on PKC activation than the saphenous vein, which underlines the existing differences between separate vascular beds [120]. Likewise, after investigating the α_1 -AR in mesenteric resistance arteries from CHF rats, it was confirmed that Ca^{2+} plays a more crucial role in the vascular contractility as CHF develops [121]. Furthermore, it has been demonstrated that the diminished contractile response of rat mesenteric resistance arteries to phenylephrine and Ca^{2+} is not due to changes in Ca^{2+} sensitivity, nor in intracellular Ca^{2+} release, but rather to changes in the α_1 -AR-triggered Ca^{2+} influx [102].

Other studies, however, have identified a modification in the sensitivity of α_1 -AR vasoconstriction to Ca^{2+} . For instance, in CHF rats an increased responsiveness of CHF femoral resistance arteries was identified which appeared to be the result of first, a RhoA/Rho kinase-dependent increase in sensitivity to Ca^{2+} and second, of the IP3-induced increase in intracellular Ca^{2+} [122]. On the contrary, this arterial sensitivity to Ca^{2+} was reported to be decreased in femoral resistance and muscle resistance arteries of CHF rats, which presented as a dissociation between the intracellular Ca^{2+} and the generated wall tension [123].

4.3.5. Cell and Tissue Responses

α_1 -ARs constitute an important homeostatic mechanism that contributes to the maintenance of peripheral vascular resistance at normal conditions and also in response to shock. Nevertheless, α_1 -AR regulation and function might alter due to disease. Even though HF is characterized by enhanced vascular resistance [124], there is little, and conflicting evidence to date concerning the vascular contractile responses to α_1 -AR stimulation in HF vessels.

The chronic sympathetic stimulation during HF might result in the desensitization of vascular α_1 -AR and consequently, in the failure of the contractile apparatus to function properly. Attenuation of α_1 -AR responsiveness after continuous exposure to catecholamines has been validated in various animal studies [125,126] but also in vitro [127,128]. However, despite the fact that several studies have tried to assess the α_1 -AR mediated vascular responses in both animals and humans with HF,

the findings are not unanimous (Table 4). Multiple studies have shown an increase in vascular responsiveness to α_1 -AR stimulation in animals [114–117,119,120,122,129–131]. On the contrary, others have reported desensitization of α_1 -ARs in the peripheral vasculature in animals [102,121,132–136] and in humans [137–140], while numerous studies failed to demonstrate any alterations of the α_1 -AR responsiveness in vessels from animals [105,123,133,141–143] and humans [144].

Table 4. Summary of changes in vascular α_1 -adrenoceptor (AR)-mediated responses in chronic heart failure.

	Species	Preparation/Parameter Studied	α_1 -AR-Mediated Responses
Goldsmith et al. [137]	human	In vivo hemodynamic responses	↓
Wilson et al. [141]	dog	In vivo hindlimb vascular responses	X
Schwinn et al. [145]	human	In vivo BP response	X (awake)/ ↓ (on fentanyl anesthesia)
Borow et al. [138]	human	In vivo BP response	↓
Forster et al. [114]	dog	DPA and SV	↑
Forster et al. [115]	dog	DPA and SV	↑
Main et al. [132]	dog	LAD and LCX	↓
Forster et al. [119]	dog	DPA and SV	↑
Angus et al. [139]	human	Small arteries from gluteal skin biopsy	↓
Townsend et al. [129]	dog	Pulmonary arterial and venous pressure response	↑
Indolfi et al. [144]	human	In-vivo forearm blood flow response	X
Teerlink et al. [146]	rat	Thoracic aorta	↑ intact vessels/ ↓ denuded vessels
Forster et al. [120]	dog	DPA and SV	↑ significant only in SV
Bergdahl et al. [130]	rat	Basilar, femoral, and renal artery and iliac vein	↑ arteries (NS) ↓ vein
Forster et al. [116]	dog	DPA and SV	↑
Stassen et al. [102]	rat	MSA	↓
Stassen et al. [133]	rat	Thoracic aorta, coronary arteries, MSA	X thoracic aorta/ ↑ coronary arteries (NS)/↓ MSA
Mulder et al. [142]	rat	Abdominal aorta, Femoral artery, MSA	X
Le Tran et al. [117]	dog	DPA and SV	↑ (significant only for NA in DPA)
Martinez et al. [105]	rat	Aorta, Carotid artery	X
McMillon et al. [143]	dog	Intrapulmonary bronchial vessels	X
Feng et al. [134]	rat	In vivo BP response	↓
Bergdahl et al. [121]	rat	MSA	↓
Ahmadiasl et al. [135]	rabbit	Thoracic aorta, left renal artery and vein, lateral saphenous artery and vein, vena cava	↓ significant only in vena cava
Tamagawa et al. [131]	dog	LAD/ In vivo coronary pressure-flow relationship response	↑
Trautner et al. [123]	rat	Femoral artery (2nd order side branches) and muscle resistance arteries	X
Koida et al. [122]	rat	Femoral resistance artery (small branch)	↑
Van Tassel et al. [147]	human	In vivo BP response	↑ with up-titration of carvedilol
Ramchandra et al. [136]	sheep	Renal vessels/In vivo renal vasoconstrictor response	↓
Barrett-O’Keefe et al. [140]	human	CFA and CFV/In vivo leg blood flow response	↓ at rest

BP: blood pressure, CFA: common femoral artery, CFV: common femoral vein, DPA: dorsal pedal artery, LAD: left anterior descending coronary artery, LCX: left circumflex coronary artery, MSA: mesenteric small artery, NS: not significant, SV: saphenous vein, ↑: increase, ↓: decrease, X: similar.

Inconsistency in findings might originate from differences in the anatomy and function of distinct vascular beds. Additionally, differences in CHF model and disease state might also influence the results of different studies. For example, in some studies it was demonstrated that the saphenous vein of CHF dogs was at all times more sensitive than the dorsal pedal artery [114–116,120]. This might be associated to a heterogeneous smooth muscle function between the artery and the vein, but, also, to differences in receptor population in the canine vasculature [148]. In addition, differences in vascular responsiveness between different vascular beds were also identified in rat models of HF [123,130]. Moreover, differences have also been identified between the various arteries studied. For example, in similar models of pacing-induced CHF in dogs, some studies have demonstrated an increased responsiveness to α_1 -AR stimulation in the dorsal pedal artery and the contralateral saphenous vein [114–116,119], while others have identified a desensitization of α_1 -ARs in coronary arteries of CHF dogs [132].

Other important parameters that influence the peripheral vascular responses are the integrity and physiological function of endothelium. More specifically, the endothelium-induced vasodilation counteracts the contraction via α_1 -ARs. Therefore, the malfunction of the endothelium might lead to the absence of this inhibitory function in CHF vessels [143] and result in an enhancement of vascular contraction. Increased vascular responses to noradrenaline were reported in the intact thoracic aorta from HF rats compared to controls, however the actual vascular α_1 -AR responsiveness was diminished when the endothelium was removed [146]. On the other hand, an increase in endothelial function as was observed in other studies might oppose the α_1 -AR-mediated vascular contractions [131,132]. Others, however, did not demonstrate a difference in vascular responses between intact and denuded vessels [123].

Even though the vascular responsiveness in patients with HF has not been widely investigated, many studies have yielded data that might have important clinical implications. More precisely, Schwinn et al. [145] reported on the vascular responsiveness of HF patients that underwent coronary artery revascularization. The results from this study support that left ventricular dysfunction and more precisely, left ventricular ejection fraction equal to or lower than 40% is associated with a diminished vascular responsiveness during fentanyl anesthesia in this patient population. In contrast to more recent reports that identify cardiac surgery and cardiopulmonary bypass as predisposing factors for an attenuated vascular responsiveness [13,14,149], Schwinn et al. [145] reported that phenylephrine-induced changes in systemic vascular resistance were increased during cardiopulmonary bypass and aortic cross-clamp in both HF and control subjects. This vascular responsiveness is of interest in HF patients that undergo cardiac surgery, since this patient population exhibits an increased risk for post-operative vasoplegia [150]. This complication is characterized by low systemic vascular resistance, severe hypotension, normal or increased cardiac output and a blunted or absent response to the administration of vasopressors [151]. The diminished sensitivity of HF patients is thought to originate from the continuous, chronic exposure of these patients' vascular α_1 -ARs to catecholamines which leads to desensitization and/or downregulation of those receptors. If vasoplegia is not promptly treated, the resulted tissue malperfusion might lead to end-organ dysfunction and multiple organ failure. Therefore, this post-operative complication significantly increases the risk of morbidity and mortality [152,153]. Moreover, another study identified a maladapted sympathetic response of the skeletal muscle vasculature to exercise that was characterized by sustained vascular constriction [140]. This can have serious implications on normal blood flow during physical movement and might hamper the patient's daily life. Furthermore, the apparent up-regulation of α_1 -AR responsiveness after chronic carvedilol treatment in HF patients [147] and the lack of change after chronic angiotensin-converting enzyme inhibitors [142] might yield important consequences regarding HF therapy.

5. Discussion

Understanding the role of α_1 -ARs is particularly important in HF because this condition is associated with high adrenergic activation leading to down-regulation and desensitization of cardiac

β -ARs. α_1 -ARs have been identified in both animal and human hearts and vasculature with a high expression in rodents but a much lower abundance in humans. Thus, caution should be applied while extrapolating data from animals to humans.

The α_{1A} -AR is the most abundant subtype in the human heart and α_{1B} -AR in the rodent heart, while α_{1D} -ARs are mainly found in coronary arteries (Figure 3). However, studies on a change in the expression of the subtypes in failing hearts are inconsistent, which may indicate a relative α_1/β -AR imbalance in light of the well-known reduction in β -adrenergic function in CHF.

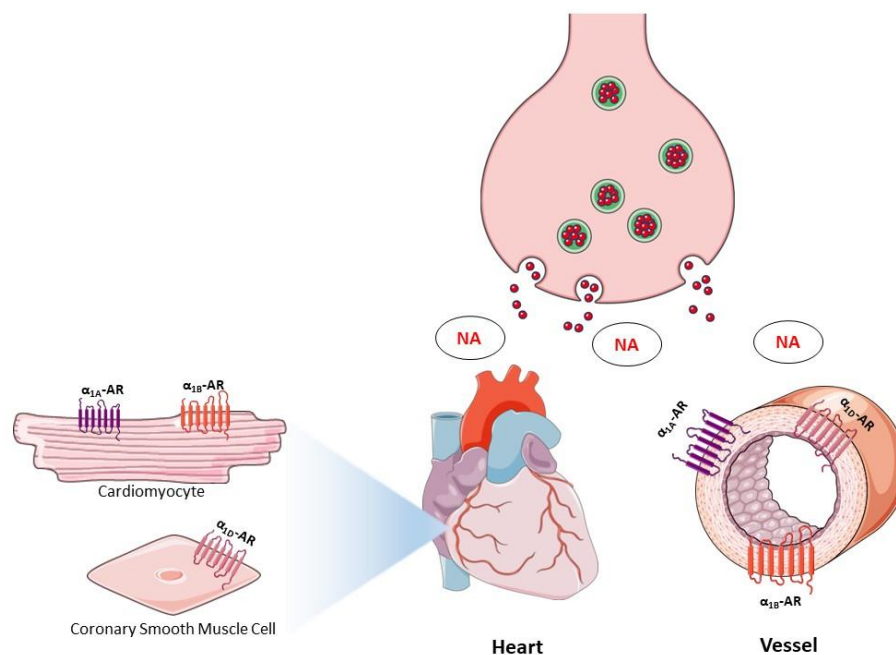


Figure 3. Differential expressions of α_1 -ARs in the cardiovascular system. NA: noradrenaline.

Another important point is that, while in heart α_{1A} - and α_{1B} -ARs dominate, all three subtypes are expressed in the vasculature and their relative expression and physiological role appear to differ between vascular beds and species (Figure 3) [3]. This varied expression of α_1 -AR has been reported to correlate with the adrenergic innervation in peripheral vessels [154]. Moreover, studies with transfected receptors suggest that the three α_1 -AR subtypes undergo differential regulation upon exposure to agonists [16]. Certainly, those discrepancies also affect the way in which blood pressure is regulated depending on which α_1 -AR subtypes play the predominant role. For that reason, the differences in subtype distribution might also have influenced studies that have investigated the function of α_1 -ARs in vivo or in vitro, leading to conflicting results.

α_1 -ARs can couple to multiple intracellular signal transduction pathways in the heart. Under most conditions α_1 -AR signal transduction results in enhanced inotropy. Although data on their role in HF is limited, there is evidence indicating that they mediate enhanced positive inotropy, as well as physiological hypertrophy and decreased apoptosis [46]. α_1 -AR occupancy was estimated as 10% in failing human heart [155]. Thus, activation of unoccupied receptors with an exogenous ligand is suggested to be cardioprotective [46]. Clinical trials such as ALLHAT in hypertension and V-HeFT in CHF reported that treatment with α_1 -AR antagonists was associated with greater mortality and apoptosis than with other treatments [156]. Moreover, double knockout of α_{1A} - and α_{1B} -AR was shown to augment apoptosis, worsen cardiomyopathy and reduce survival [157]. Taken together, cardiac α_1 -ARs appear to mediate cardioprotective effects. Data on the regulation of vascular α_1 -ARs in HF are also conflicting. This is unfortunate because of their role in controlling total peripheral resistance and the role of afterload reductions in the treatment of patients with HF. However, clinical studies in hypertensive patients have shown that α_1 -AR antagonism provides less protection from HF than

other classes of anti-hypertensive drugs [156]. More specifically, the non-selective α -blocker doxazosin was associated with significantly higher risk of HF compared with the other treatment arms of this trial. In the absence of a placebo arm, it remained unclear whether this reflects an adverse effect of doxazosin or less protection against hypertension-induced HF. The results from the V-HeFT trial [158] further support the detrimental effects of α -blocking therapy-, since prazosin use gave a signal for possibly increased mortality that failed to reach statistical significance. The reason for those results could possibly be explained by the findings from knock-out mice studies which confirm the importance of α_1 -AR signaling for the adaptational mechanisms of cardiomyocytes in disease settings [157,159].

In contrast to those studies that investigated the effect of selective α -blockade and discouraged their widespread use, the mixed β - and α_1 -AR antagonist carvedilol was demonstrated to be more beneficial than the β_1 -selective antagonist metoprolol in diminishing the risk of mortality [155]. It could, therefore, be postulated that the favorable effects of carvedilol are to some extent related to its α -blocking properties. Nevertheless, the combination of an α - with a β -blocker for long-term treatment in chronic CHF patients, did not yield better clinical outcomes than β -blocker therapy alone [160], which highlights the fact that such a combined therapy does not generate satisfactory outcomes similar to that of carvedilol.

The exact reason for those discrepancies in effectiveness still remains unclear. One explanation could be that carvedilol's α -blocking properties are less potent compared to both its innate β -blocking feature and to the α -blockade produced by a specific α -AR antagonist like prazosin. This is in fact supported by the results of a small trial in which carvedilol did not antagonize phenylephrine induced responses [161]. Another hypothesis is that the existence of distinct pharmacologic properties and mechanisms of action between the various α - and β -blockers could also lead to the occurrence of different side effects. Based on this assumption, one could possibly explain the contradictory findings from studies with carvedilol versus studies investigating the combination of an α - with a β -blocker. Last but not least, carvedilol use leads to pharmacologic tolerance after long term use [162]. This implies that its beneficial effects at long-term use derive mostly from its β -blocking properties rather than antagonizing α_1 -ARs.

In brief, one could state that α_1 -AR agonism could potentially be a valid treatment option for HF patients that would stimulate innate adaptive signaling procedures and would eventually illustrate its cardioprotective features. However, caution is required, since α_1 agonism could also have detrimental effects by increasing peripheral vascular resistance leading to an increased cardiac afterload and by inducing coronary vasoconstriction in those patients. Therefore, future research is necessary which should focus on compartment and/or subtype selective treatment in HF patients and which will investigate in more detail the treatment modalities of α_1 agonism.

On the other hand, carvedilol seems promising for the treatment of HF patients and might be an option for the pretreatment of HF patients that will undergo cardiac surgery. Short term α_1 antagonism with carvedilol treatment could shield α_1 -ARs against continuous catecholamine exposure in the preoperative period. This could perhaps increase the sensitivity of those receptors to the administration of catecholamines for the treatment of vasoplegia during the postoperative course [147].

In conclusion, more research is required to further understand the complex mechanisms of α_1 adrenergic function and adaptation in HF. Taking into account the existing differences between animals and humans regarding the expression of α_1 -ARs, we should carefully consider our next steps and we should promote additional research which would focus more on human tissue (heart and vessels). Moreover, a cardioselective approach in the case of α_1 agonism appears to be crucial for the exploitation of only its advantageous characteristics by avoiding potential vascular side effects. A subtype approach could also be beneficial but further studies are necessary in order to gain more insight in such treatment techniques. Finally, in view of the positive outcomes from clinical studies with carvedilol, more extensive investigation is to be expected so as to better comprehend the pharmacologic properties and mechanism of action of this agent. It is, therefore, essential that we expand our knowledge, and we apprehend current inconsistencies in order to pave the way to safer and more effective therapies.

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References

1. Brodde, O.E.; Michel, M.C. Adrenergic and muscarinic receptors in the human heart. *Pharmacol. Rev.* **1999**, *51*, 651–689.
2. Brattellid, T.; Tveit, K.; Birkeland, J.A.; Sjaastad, I.; Qvigstad, E.; Krobert, K.A.; Hussain, R.I.; Skomedal, T.; Osnes, J.B.; Levy, F.O. Expression of mRNA encoding G protein-coupled receptors involved in congestive heart failure; A quantitative RT-PCR study and the question of normalisation. *Basic Res. Cardiol.* **2007**, *102*, 198–208.
3. Guimaraes, S.; Moura, D. Vascular adrenoceptors: An update. *Pharmacol. Rev.* **2001**, *53*, 319–356.
4. Puceat, M.; Terzic, A.; Clement, O.; Scamps, F.; Vogel, S.M.; Vassort, G. Cardiac α_1 -adrenoceptors mediate positive inotropy via myofibrillar sensitization. *Trends Pharmacol. Sci.* **1992**, *13*, 263–265. [[PubMed](#)]
5. Steinberg, S.F.; Drugge, E.D.; Bilezikian, J.P.; Robinson, R.B. Acquisition by innervated cardiac myocytes of a pertussis toxin-specific regulatory protein linked to the alpha 1-receptor. *Science* **1985**, *230*, 186–188.
6. Li, K.; He, H.; Li, C.; Sirois, P.; Rouleau, J.L. Myocardial α_1 -adrenoceptor: Inotropic effect and physiologic and pathologic implications. *Life Sci.* **1997**, *60*, 1305–1318. [[PubMed](#)]
7. Anyukhovskiy, E.P.; Rybin, V.O.; Nikashin, A.V.; Budanova, O.P.; Rosen, M.R. Positive chronotropic responses induced by alpha 1-adrenergic stimulation of normal and “ischemic” Purkinje fibers have different receptor-effector coupling mechanisms. *Circ. Res.* **1992**, *71*, 526–534.
8. Knowlton, K.U.; Michel, M.C.; Itani, M.; Shubeita, H.E.; Ishihara, K.; Brown, J.H.; Chien, K.R. The α_{1A} -adrenergic receptor subtype mediates biochemical, molecular, and morphologic features of cultured myocardial cell hypertrophy. *J. Biol. Chem.* **1993**, *268*, 15374–15380. [[PubMed](#)]
9. Dal Monte, M.; Evans, B.A.; Arioglu-Inan, E.; Michel, M.C. Upregulation of β_3 -adrenoceptors—A general marker of and protective mechanism against hypoxia? *Naunyn-Schmiedeberg's Arch. Pharmacol.* **2020**, *393*, 141–146.
10. Cohn, J.N. Abnormalities of peripheral sympathetic nervous system control in congestive heart failure. *Circulation* **1990**, *82*, 59–67.
11. Maisel, A.S.; Scott, N.A.; Motulsky, H.J.; Michel, M.C.; Boublik, J.H.; Rivier, J.E.; Ziegler, M.; Allen, R.S.; Brown, M.R. Elevation of plasma neuropeptide Y levels in congestive heart failure. *Am. J. Med.* **1989**, *86*, 43–48. [[CrossRef](#)]
12. Cohn, J.N.; Levine, T.B.; Olivari, M.T.; Garberg, V.; Lura, D.; Francis, G.S.; Simon, A.B.; Rector, T. Plasma Norepinephrine as a Guide to Prognosis in Patients with Chronic Congestive Heart Failure. *N. Engl. J. Med.* **1984**, *311*, 819–823. [[CrossRef](#)]
13. Carrel, T.; Englberger, L.; Mohacsi, P.; Neidhart, P.; Schmidli, J. Low systemic vascular resistance after cardiopulmonary bypass: Incidence, etiology, and clinical importance. *J. Card. Surg.* **2000**, *15*, 347–353. [[CrossRef](#)]
14. Sun, X.; Zhang, L.; Hill, P.C.; Lowery, R.; Lee, A.T.; Molyneaux, R.E.; Corso, P.J.; Boyce, S.W. Is incidence of postoperative vasoplegic syndrome different between off-pump and on-pump coronary artery bypass grafting surgery? *Eur. J. Cardiothorac. Surg.* **2008**, *34*, 820–825. [[CrossRef](#)]
15. Michel, M.C.; Kenny, B.; Schwinn, D.A. Classification of alpha 1-adrenoceptor subtypes. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1995**, *352*, 1–10. [[CrossRef](#)] [[PubMed](#)]

16. Yang, M.; Ruan, J.; Voller, M.; Schalken, J.; Michel, M.C. Differential regulation of human α_1 -adrenoceptor subtypes. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1999**, *359*, 439–446. [\[CrossRef\]](#)
17. Monto, F.; Oliver, E.; Vicente, D.; Rueda, J.; Aguero, J.; Almenar, L.; Ivorra, M.D.; Baretino, D.; D'Ocon, P. Different expression of adrenoceptors and GRKs in the human myocardium depends on heart failure etiology and correlates to clinical variables. *Am. J. Physiol. Heart Circ. Physiol.* **2012**, *303*, 368–376. [\[CrossRef\]](#)
18. Hendriks-Balk, M.C.; Michel, M.C.; Alewijnse, A.E. Pitfalls in the normalization of real-time polymerase chain reaction data. *Basic Res. Cardiol.* **2007**, *102*, 195–197. [\[CrossRef\]](#)
19. Michel-Reher, M.B.; Michel, M.C. Regulation of GAPDH expression by treatment with the β -adrenoceptor agonist isoprenaline—Is GAPDH a suitable loading control in immunoblot experiments? *Naunyn-Schmiedeberg's Arch. Pharmacol.* **2015**, *388*, 1119–1120. [\[CrossRef\]](#)
20. Pradidarcheep, W.; Stallen, J.; Labruyere, W.T.; Dabhoiwala, N.F.; Michel, M.C.; Lamers, W.H. Lack of specificity of commercially available antisera against muscarinic and adrenergic receptors. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **2009**, *379*, 397–402. [\[CrossRef\]](#) [\[PubMed\]](#)
21. Jensen, B.C.; Swigart, P.M.; Simpson, P.C. Ten commercial antibodies for α_1 -adrenergic receptor subtypes are nonspecific. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **2009**, *379*, 409–412. [\[CrossRef\]](#)
22. Böhmer, T.; Pfeiffer, N.; Gericke, A. Three commercial antibodies against α_1 -adrenergic receptor subtypes lack specificity in paraffin-embedded sections of murine tissues. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **2014**, *387*, 703–706. [\[CrossRef\]](#)
23. McGrath, J.C. Localization of α -adrenoceptors: JR Vane Medal Lecture. *Br. J. Pharmacol.* **2015**, *172*, 1179–1194. [\[CrossRef\]](#)
24. Faber, J.E.; Yang, N.; Xin, X. Expression of α -Adrenoceptor Subtypes by Smooth Muscle Cells and Adventitial Fibroblasts in Rat Aorta and in Cell Culture. *J. Pharmacol. Exp. Ther.* **2001**, *298*, 441.
25. Litwin, S.E.; Vatner, D.E.; Morgan, J.P. Inotropic effects of α_1 -adrenergic agonists in myocardium from rats with postinfarction heart failure. *Am. J. Physiol.* **1995**, *269*, H1553–H1563. [\[CrossRef\]](#)
26. Meggs, L.G.; Tillotson, J.; Huang, H.; Sonnenblick, E.H.; Capasso, J.M.; Anversa, P. Noncoordinate regulation of α_1 -adrenoceptor coupling and reexpression of α skeletal actin in myocardial infarction-induced left ventricular failure in rats. *J. Clin. Investig.* **1990**, *86*, 1451–1458. [\[CrossRef\]](#)
27. Wang, G.Y.; Yeh, C.C.; Jensen, B.C.; Mann, M.J.; Simpson, P.C.; Baker, A.J. Heart failure switches the RV α_1 -adrenergic inotropic response from negative to positive. *Am. J. Physiol. Heart Circ. Physiol.* **2010**, *298*, H913–H920. [\[CrossRef\]](#)
28. Calderone, A.; Bouvier, M.; Li, K.; Juneau, C.; de Champlain, J.; Rouleau, J.L. Dysfunction of the beta- and α -adrenergic systems in a model of congestive heart failure. The pacing-overdrive dog. *Circ. Res.* **1991**, *69*, 332–343. [\[CrossRef\]](#)
29. Cowley, P.M.; Wang, G.; Chang, A.N.; Makwana, O.; Swigart, P.M.; Lovett, D.H.; Stull, J.T.; Simpson, P.C.; Baker, A.J. The α_{1A} -adrenergic receptor subtype mediates increased contraction of failing right ventricular myocardium. *Am. J. Physiol. Heart Circ. Physiol.* **2015**, *309*, H888–H896. [\[CrossRef\]](#)
30. Zhao, M.; Hagler, H.K.; Muntz, K.H. Regulation of α_1 -, β_1 -, and β_2 -adrenergic receptors in rat heart by norepinephrine. *Am. J. Physiol.* **1996**, *271*, H1762–H1768. [\[CrossRef\]](#) [\[PubMed\]](#)
31. Eschenhagen, T.; Diederich, M.; Kluge, S.H.; Magnussen, O.; Mene, U.; Muller, F.; Schmitz, W.; Scholz, H.; Weil, J.; Sent, U.; et al. Bovine hereditary cardiomyopathy: An animal model of human dilated cardiomyopathy. *J. Mol. Cell Cardiol.* **1995**, *27*, 357–370. [\[CrossRef\]](#)
32. Gopalakrishnan, M.; Triggle, D.J.; Rutledge, A.; Kwon, Y.W.; Bauer, J.A.; Fung, H.L. Regulation of K^+ and Ca^{2+} channels in experimental cardiac failure. *Am. J. Physiol.* **1991**, *261*, H1979–H1987. [\[CrossRef\]](#)
33. Karliner, J.S.; Barnes, P.; Brown, M.; Dollery, C. Chronic heart failure in the guinea pig increases cardiac α_1 - and β -adrenoceptors. *Eur. J. Pharmacol.* **1980**, *67*, 115–118. [\[CrossRef\]](#)
34. Maisel, A.S.; Motulsky, H.J.; Ziegler, M.G.; Insel, P.A. Ischemia- and agonist-induced changes in α - and β -adrenergic receptor traffic in guinea pig hearts. *Am. J. Physiol. Heart Circ. Physiol.* **1987**, *253*, H1159–H1166. [\[CrossRef\]](#)
35. Watanabe, K.; Shibata, A.; Wakabayashi, H.; Shimada, K.; Sasaki, K.; Nagatomo, T. Cardiomyopathic hamster hearts: Long-term effects of drugs on catecholamine contents and binding characteristics of α_1 - and β_1 -adrenergic receptors. *Biol. Pharm. Bull.* **1993**, *16*, 660–663. [\[CrossRef\]](#)

36. Corr, P.B.; Shayman, J.A.; Kramer, J.B.; Kipnis, R.J. Increased alpha-adrenergic receptors in ischemic cat myocardium. A potential mediator of electrophysiological derangements. *J. Clin. Investig.* **1981**, *67*, 1232–1236. [\[CrossRef\]](#) [\[PubMed\]](#)
37. Re, G.; Bergamasco, L.; Badino, P.; Borgarelli, M.; Odore, R.; Tarducci, A.; Zanatta, R.; Girardi, C. Canine dilated cardiomyopathy: Lymphocyte and cardiac α_1 - and β -adrenoceptor concentrations in normal and affected great danes. *Vet. J.* **1999**, *158*, 120–127. [\[CrossRef\]](#)
38. Suematsu, N.; Satoh, S.; Kinugawa, S.; Tsutsui, H.; Hayashidani, S.; Nakamura, R.; Egashira, K.; Makino, N.; Takeshita, A. Alpha₁-adrenoceptor-Gq-RhoA signaling is upregulated to increase myofibrillar Ca²⁺ sensitivity in failing hearts. *Am. J. Physiol. Heart Circ. Physiol.* **2001**, *281*, H637–H646. [\[CrossRef\]](#)
39. Brodde, O.E.; Leineweber, K. Autonomic receptor systems in the failing and aging human heart: Similarities and differences. *Eur. J. Pharmacol.* **2004**, *500*, 167–176. [\[CrossRef\]](#)
40. Jensen, B.C.; O'Connell, T.D.; Simpson, P.C. Alpha-1-adrenergic receptors in heart failure: The adaptive arm of the cardiac response to chronic catecholamine stimulation. *J. Cardiovasc. Pharmacol.* **2014**, *63*, 291–301. [\[CrossRef\]](#) [\[PubMed\]](#)
41. Jensen, B.C.; Swigart, P.M.; de Marco, T.; Hoopes, C.; Simpson, P.C. α_1 -Adrenergic receptor subtypes in nonfailing and failing human myocardium. *Circ. Heart Fail.* **2009**, *2*, 654–663. [\[CrossRef\]](#)
42. Myagmar, B.E.; Flynn, J.M.; Cowley, P.M.; Swigart, P.M.; Montgomery, M.D.; Thai, K.; Nair, D.; Gupta, R.; Deng, D.X.; Hosoda, C.; et al. Adrenergic Receptors in Individual Ventricular Myocytes: The Beta-1 and Alpha-1B are in All Cells, the Alpha-1A is in a Subpopulation, and the Beta-2 and Beta-3 are Mostly Absent. *Circ. Res.* **2017**, *120*, 1103–1115. [\[CrossRef\]](#)
43. Fischer, V.; Gabauer, I.; Tillinger, A.; Novakova, M.; Pechan, I.; Krizanov, O.; Kvetnansky, R.; Myslivecek, J. Heart adrenoceptor gene expression and binding sites in the human failing heart. *Ann. N. Y. Acad. Sci.* **2008**, *1148*, 400–408. [\[CrossRef\]](#)
44. Williams, R.S.; Lefkowitz, R.J. Alpha-adrenergic receptors in rat myocardium. Identification by binding of [³H]dihydroergocryptine. *Circ. Res.* **1978**, *43*, 721–727. [\[CrossRef\]](#)
45. Hwang, K.C.; Gray, C.D.; Sweet, W.E.; Moravec, C.S.; Im, M.J. Alpha 1-adrenergic receptor coupling with Gh in the failing human heart. *Circulation* **1996**, *94*, 718–726. [\[CrossRef\]](#)
46. Steinfath, M.; Danielsen, W.; von der Leyen, H.; Mende, U.; Meyer, W.; Neumann, J.; Nose, M.; Reich, T.; Schmitz, W.; Scholz, H.; et al. Reduced alpha 1- and beta 2-adrenoceptor-mediated positive inotropic effects in human end-stage heart failure. *Br. J. Pharmacol.* **1992**, *105*, 463–469. [\[CrossRef\]](#)
47. Vago, T.; Bevilacqua, M.; Norbiato, G.; Baldi, G.; Chebat, E.; Bertora, P.; Baroldi, G.; Accinni, R. Identification of alpha 1-adrenergic receptors on sarcolemma from normal subjects and patients with idiopathic dilated cardiomyopathy: Characteristics and linkage to GTP-binding protein. *Circ. Res.* **1989**, *64*, 474–481. [\[CrossRef\]](#)
48. Grigore, A.; Poindexter, B.; Vaughn, W.K.; Nussmeier, N.; Frazier, O.H.; Cooper, J.R.; Gregoric, I.D.; Buja, L.M.; Bick, R.J. Alterations in alpha adrenoceptor density and localization after mechanical left ventricular unloading with the Jarvik flowmaker left ventricular assist device. *J. Heart Lung Transplant.* **2005**, *24*, 609–613. [\[CrossRef\]](#)
49. Bohm, M.; Diet, F.; Feiler, G.; Kemkes, B.; Erdmann, E. Alpha-adrenoceptors and alpha-adrenoceptor-mediated positive inotropic effects in failing human myocardium. *J. Cardiovasc. Pharmacol.* **1988**, *12*, 357–364. [\[CrossRef\]](#)
50. Bristow, M.R.; Minobe, W.; Rasmussen, R.; Hersherberger, R.E.; Hoffman, B.B. Alpha-1 adrenergic receptors in the nonfailing and failing human heart. *J. Pharmacol. Exp. Ther.* **1988**, *247*, 1039–1045.
51. Erdmann, E.; Bohm, M. Positive inotropic stimulation in the normal and insufficient human myocardium. *Basic Res. Cardiol.* **1989**, *84*, 125–133. [\[CrossRef\]](#)
52. Limas, C.J.; Limas, C.; Goldenberg, I.F. Intracellular distribution of adrenoceptors in the failing human myocardium. *Am. Heart J.* **1989**, *117*, 1310–1316. [\[CrossRef\]](#)
53. Shi, T.; Moravec, C.S.; Perez, D.M. Novel proteins associated with human dilated cardiomyopathy: Selective reduction in α_{1A} -adrenergic receptors and increased desensitization proteins. *J. Recept. Signal Transduct.* **2013**, *33*, 96–106. [\[CrossRef\]](#)
54. Dixon, I.M.C.; Dhalla, N.S. Alterations in cardiac adrenoceptors in congestive heart failure secondary to myocardial infarction. *Coron. Artery Dis.* **1991**, *2*, 805–814.
55. Sjaastad, I.; Schiander, I.; Sjetnan, A.; Qvigstad, E.; Bokenes, J.; Sandnes, D.; Osnes, J.B.; Sejersted, O.M.; Skomedal, T. Increased contribution of α_1 - vs. β -adrenoceptor-mediated inotropic response in rats with congestive heart failure. *Acta Physiol. Scand.* **2003**, *177*, 449–458. [\[CrossRef\]](#)

56. Rowley, K.G.; Tung, L.H.; Hodsman, G.P.; Howes, L.G.; Jarrott, B.; Beart, P.M.; Louis, W.J. Altered α_1 -Adrenoceptor-Mediated Responses in Atria of Rats with Chronic Left Ventricular Infarction. *J. Cardiovasc. Pharmacol.* **1991**, *17*, 474–479. [[CrossRef](#)]
57. O'Connell, T.D.; Jensen, B.C.; Baker, A.J.; Simpson, P.C. Cardiac α_1 -adrenergic receptors: Novel aspects of expression, signaling mechanisms, physiologic function, and clinical importance. *Pharmacol. Rev.* **2014**, *66*, 308–333. [[CrossRef](#)]
58. Yang, M.; Reese, J.; Cotecchia, S.; Michel, M.C. Murine α_1 -adrenoceptor subtypes. I. Radioligand binding studies. *J. Pharmacol. Exp. Ther.* **1998**, *286*, 841–847.
59. Michel, M.C.; Hanft, G.; Gross, G. Radioligand binding studies of α_1 -adrenoceptor subtypes in rat heart. *Br. J. Pharmacol.* **1994**, *111*, 533–538. [[CrossRef](#)] [[PubMed](#)]
60. Janssen, P.M.L.; Canan, B.D.; Kilic, A.; Whitson, B.A.; Baker, A.J. Human Myocardium Has a Robust α_{1A} -Subtype Adrenergic Receptor Inotropic Response. *J. Cardiovasc. Pharmacol.* **2018**, *72*, 136–142. [[CrossRef](#)]
61. Du, X.J.; Gao, X.M.; Kiriazis, H.; Moore, X.L.; Ming, Z.; Su, Y.; Finch, A.M.; Hannan, R.A.; Dart, A.M.; Graham, R.M. Transgenic α_{1A} -adrenergic activation limits post-infarct ventricular remodeling and dysfunction and improves survival. *Cardiovasc. Res.* **2006**, *71*, 735–743. [[CrossRef](#)]
62. Schnee, P.M.; Shah, N.; Bergheim, M.; Poindexter, B.J.; Buja, L.M.; Gemmato, C.; Radovancevic, B.; Letsou, G.V.; Frazier, O.H.; Bick, R.J. Location and density of α - and β -adrenoreceptor sub-types in myocardium after mechanical left ventricular unloading. *J. Heart Lung Transplant.* **2008**, *27*, 710–717. [[CrossRef](#)]
63. Hieble, J.P.; Bylund, D.B.; Clarke, D.E.; Eikenburg, D.C.; Langer, S.Z.; Lefkowitz, R.J.; Minneman, K.P.; Ruffolo, R.R., Jr. International Union of Pharmacology. X. Recommendation for nomenclature of α_1 -adrenoceptors: Consensus update. *Pharmacol. Rev.* **1995**, *47*, 267–270.
64. Maruyama, Y.; Nishida, M.; Sugimoto, Y.; Tanabe, S.; Turner, J.H.; Kozasa, T.; Wada, T.; Nagao, T.; Kurose, H. $G_{\alpha_{12/13}}$ Mediates α_1 -Adrenergic receptor-induced cardiac hypertrophy. *Circ. Res.* **2002**, *91*, 961–969. [[CrossRef](#)]
65. Pönicke, K.; Heinroth-Hoffmann, I.; Brodde, O.E. Role of β_1 - and β_2 -adrenoceptors in hypertrophic and apoptotic effects of noradrenaline and adrenaline in adult rat ventricular cardiomyocytes. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **2003**, *367*, 592–599. [[CrossRef](#)] [[PubMed](#)]
66. Baek, K.J.; Das, T.; Gray, C.; Antar, S.; Murugesan, G.; Im, M.J. Evidence that the Gh protein is a signal mediator from α_1 -adrenoceptor to a phospholipase C. I. Identification of α_1 -adrenoceptor-coupled Gh family and purification of Gh7 from bovine heart. *J. Biol. Chem.* **1993**, *268*, 27390–27397. [[PubMed](#)]
67. Barrett, S.; Honbo, N.; Karliner, J.S. α_1 -Adrenoceptor-mediated inhibition of cellular cAMP accumulation in neonatal rat ventricular myocytes. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1993**, *347*, 384–393. [[CrossRef](#)]
68. Hool, L.C.; Oleksa, L.M.; Harvey, R.D. Role of G proteins in α_1 -adrenergic inhibition of the β -adrenergically activated chloride current in cardiac myocytes. *Mol. Pharmacol.* **1997**, *51*, 853–860. [[CrossRef](#)] [[PubMed](#)]
69. Liu, Q.Y.; Karpinski, E.; Pang, P.K. L-channel modulation by α_1 -adrenoceptor activation in neonatal rat ventricular cells: Intracellular mechanisms. *J. Pharmacol. Exp. Ther.* **1994**, *271*, 944–951.
70. LaMorte, V.J.; Thorburn, J.; Absher, D.; Spiegel, A.; Brown, J.H.; Chien, K.R.; Feramisco, J.R.; Knowlton, K.U. Gq- and ras-dependent pathways mediate hypertrophy of neonatal rat ventricular myocytes following α_1 -adrenergic stimulation. *J. Biol. Chem.* **1994**, *269*, 13490–13496.
71. Bogoyevitch, M.A.; Clerk, A.; Sugden, P.H. Activation of the mitogen-activated protein kinase cascade by pertussis toxin-sensitive and -insensitive pathways in cultured ventricular cardiomyocytes. *Biochem. J.* **1995**, *309*, 437–443. [[CrossRef](#)]
72. Jensen, B.C.; O'Connell, T.D.; Simpson, P.C. α_1 -Adrenergic receptors: Targets for agonist drugs to treat heart failure. *J. Mol. Cell Cardiol.* **2011**, *51*, 518–528. [[CrossRef](#)]
73. Salazar, N.C.; Chen, J.; Rockman, H.A. Cardiac GPCRs: GPCR signaling in healthy and failing hearts. *Biochim. Biophys. Acta* **2007**, *1768*, 1006–1018. [[CrossRef](#)]
74. D'Angelo, D.D.; Sakata, Y.; Lorenz, J.N.; Boivin, G.P.; Walsh, R.A.; Liggett, S.B.; Dorn, G.W., II. Transgenic α_1 -adrenoceptor overexpression induces cardiac contractile failure in mice. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 8121–8126. [[CrossRef](#)] [[PubMed](#)]

75. Adams, J.W.; Sakata, Y.; Davis, M.G.; Sah, V.P.; Wang, Y.; Liggett, S.B.; Chien, K.R.; Brown, J.H.; Dorn, G.W., II. Enhanced Galphaq signaling: A common pathway mediates cardiac hypertrophy and apoptotic heart failure. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 10140–10145. [[CrossRef](#)]
76. Akhter, S.A.; Luttrell, L.M.; Rockman, H.A.; Iaccarino, G.; Lefkowitz, R.J.; Koch, W.J. Targeting the receptor-Gq interface to inhibit in vivo pressure overload myocardial hypertrophy. *Science* **1998**, *280*, 574–577. [[CrossRef](#)]
77. Esposito, G.; Rapacciuolo, A.; Naga Prasad, S.V.; Takaoka, H.; Thomas, S.A.; Koch, W.J.; Rockman, H.A. Genetic alterations that inhibit in vivo pressure-overload hypertrophy prevent cardiac dysfunction despite increased wall stress. *Circulation* **2002**, *105*, 85–92. [[CrossRef](#)]
78. Lohse, M.J.; Engelhardt, S.; Eschenhagen, T. What is the role of beta-adrenergic signaling in heart failure? *Circ. Res.* **2003**, *93*, 896–906. [[CrossRef](#)] [[PubMed](#)]
79. Michel, M.C.; Hanft, G.; Gross, G. Functional studies on α_1 -adrenoceptor subtypes mediating inotropic effects in rat right ventricle. *Br. J. Pharmacol.* **1994**, *111*, 539–546. [[CrossRef](#)]
80. Barcroft, H.; Konzett, H. On the actions of noradrenaline, adrenaline and isopropyl noradrenaline on the arterial blood pressure, heart rate and muscle blood flow in man. *J. Physiol.* **1949**, *110*, 194–204. [[CrossRef](#)]
81. Skomedal, T.; Borthne, K.; Aass, H.; Geiran, O.; Osnes, J.B. Comparison between alpha-1 adrenoceptor-mediated and beta adrenoceptor-mediated inotropic components elicited by norepinephrine in failing human ventricular muscle. *J. Pharmacol. Exp. Ther.* **1997**, *280*, 721–729.
82. Sen, L.; Liang, B.T.; Colucci, W.S.; Smith, T.W. Enhanced alpha 1-adrenergic responsiveness in cardiomyopathic hamster cardiac myocytes. Relation to the expression of pertussis toxin-sensitive G protein and alpha 1-adrenergic receptors. *Circ. Res.* **1990**, *67*, 1182–1192. [[CrossRef](#)]
83. Woodcock, E.A.; Du, X.J.; Reichelt, M.E.; Graham, R.M. Cardiac alpha 1-adrenergic drive in pathological remodelling. *Cardiovasc. Res.* **2008**, *77*, 452–462. [[CrossRef](#)]
84. Perez, D.M.; Piascik, M.T.; Malik, N.; Gaivin, R.; Graham, R.M. Cloning, expression, and tissue distribution of the rat homolog of the bovine alpha 1C-adrenergic receptor provide evidence for its classification as the alpha 1A subtype. *Mol. Pharmacol.* **1994**, *46*, 823.
85. Piascik, M.T.; Smith, M.S.; Soltis, E.E.; Perez, D.M. Identification of the mRNA for the novel alpha 1D-adrenoceptor and two other alpha 1-adrenoceptors in vascular smooth muscle. *Mol. Pharmacol.* **1994**, *46*, 30.
86. Ping, P.; Faber, J.E. Characterization of alpha-adrenoceptor gene expression in arterial and venous smooth muscle. *Am. J. Physiol. Heart Circ. Physiol.* **1993**, *265*, H1501–H1509. [[CrossRef](#)]
87. Price, D.T.; Chari, R.S.; Berkowitz, D.E.; Meyers, W.C.; Schwinn, D.A. Expression of alpha 1-adrenergic receptor subtype mRNA in rat tissues and human SK-N-MC neuronal cells: Implications for alpha 1-adrenergic receptor subtype classification. *Mol. Pharmacol.* **1994**, *46*, 221. [[PubMed](#)]
88. Kaiming, X.; Chide, H. Quantification of mRNAs for three α_1 -adrenoceptor subtypes in rat aorta by solution hybridization. *Life Sci.* **1996**, *59*, PL343–PL347. [[CrossRef](#)]
89. Diehl, N.L.; Martin Shreeve, S. Identification of the α_{1C} -adrenoceptor in rabbit arteries and the human saphenous vein using the polymerase chain reaction. *Eur. J. Pharmacol.* **1994**, *268*, 393–398. [[CrossRef](#)]
90. Miyamoto, S.; Taniguchi, T.; Suzuki, F.; Takita, M.; Kosaka, N.; Negoro, E.; Okuda, T.; Kosaka, H.; Murata, S.; Nakamura, S.; et al. Cloning, functional expression and tissue distribution of rabbit α_{1a} -adrenoceptor. *Life Sci.* **1997**, *60*, 2069–2074. [[CrossRef](#)]
91. Suzuki, F.; Miyamoto, S.; Takita, M.; Oshita, M.; Watanabe, Y.; Kakizuka, A.; Narumiya, S.; Taniguchi, T.; Muramatsu, I. Cloning, functional expression and tissue distribution of rabbit α_{1d} -adrenoceptor1The nucleotide sequence reported in this paper has been submitted to the GenBank under the accession number U64032.1. *Biochim. Biophys. Acta Biomembr.* **1997**, *1323*, 6–11. [[CrossRef](#)]
92. Faure, C.; Gouhier, C.; Langer, S.Z.; Graham, D. Quantification of α_1 -Adrenoceptor Subtypes in Human Tissues by Competitive RT-PCR Analysis. *Biochem. Biophys. Res. Commun.* **1995**, *213*, 935–943. [[CrossRef](#)]
93. Price, D.T.; Lefkowitz, R.J.; Caron, M.G.; Berkowitz, D.; Schwinn, D.A. Localization of mRNA for three distinct alpha 1-adrenergic receptor subtypes in human tissues: Implications for human alpha-adrenergic physiology. *Mol. Pharmacol.* **1994**, *45*, 171.
94. Uhlén, M.; Fagerberg, L.; Hallström, B.M.; Lindskog, C.; Oksvold, P.; Mardinoglu, A.; Sivertsson, Å.; Kampf, C.; Sjöstedt, E.; Asplund, A.; et al. Tissue-based map of the human proteome. *Science* **2015**, *347*, 1260419. [[CrossRef](#)]

95. Izzo, N.J., Jr.; Seidman, C.E.; Collins, S.; Colucci, W.S. Alpha 1-adrenergic receptor mRNA level is regulated by norepinephrine in rabbit aortic smooth muscle cells. *Proc. Natl. Acad. Sci. USA* **1990**, *87*, 6268–6271. [\[CrossRef\]](#)
96. Eckhart, A.D.; Zhu, Z.; Arendshorst, W.J.; Faber, J.E. Oxygen modulates alpha 1B-adrenergic receptor gene expression by arterial but not venous vascular smooth muscle. *Am. J. Physiol. Heart Circ. Physiol.* **1996**, *271*, H1599–H1608. [\[CrossRef\]](#)
97. Rudner, X.L.; Berkowitz, D.E.; Booth, J.V.; Funk, B.L.; Cozart, K.L.; D’Amico, E.B.; El-Moalem, H.; Page, S.O.; Richardson, C.D.; Winters, B.; et al. Subtype Specific Regulation of Human Vascular α_1 -Adrenergic Receptors by Vessel Bed and Age. *Circulation* **1999**, *100*, 2336–2343. [\[CrossRef\]](#) [\[PubMed\]](#)
98. Jensen, B.C.; Swigart, P.M.; Laden, M.E.; DeMarco, T.; Hoopes, C.; Simpson, P.C. The alpha-1D is the predominant alpha-1-adrenergic receptor subtype in human epicardial coronary arteries. *J. Am. Coll. Cardiol.* **2009**, *54*, 1137–1145. [\[CrossRef\]](#)
99. Chalothorn, D.; McCune, D.F.; Edelmann, S.E.; Tobita, K.; Keller, B.B.; Lasley, R.D.; Perez, D.M.; Tanoue, A.; Tsujimoto, G.; Post, G.R.; et al. Differential Cardiovascular Regulatory Activities of the α_{1B} - and α_{1D} -Adrenoceptor Subtypes. *J. Pharmacol. Exp. Ther.* **2003**, *305*, 1045. [\[CrossRef\]](#) [\[PubMed\]](#)
100. Turnbull, L.; McCloskey, D.T.; O’Connell, T.D.; Simpson, P.C.; Baker, A.J. α_1 -Adrenergic receptor responses in α_1 AB-AR knockout mouse hearts suggest the presence of α_1 D-AR. *Am. J. Physiol. Heart Circ. Physiol.* **2003**, *284*, H1104–H1109. [\[CrossRef\]](#)
101. Stassen, F.R.; Willemsen, M.J.; Janssen, G.M.; DeMey, J.G. α_1 -adrenoceptor subtypes in rat aorta and mesenteric small arteries are preserved during left ventricular dysfunction post-myocardial infarction. *Cardiovasc. Res.* **1997**, *33*, 706–713. [\[CrossRef\]](#)
102. Stassen, F.R.; Willemsen, M.J.; Janssen, G.M.; Fazzi, G.E.; Schiffers, P.M.; Smits, J.F.; de Mey, J.G. Reduced responsiveness of rat mesenteric resistance artery smooth muscle to phenylephrine and calcium following myocardial infarction. *Br. J. Pharmacol.* **1997**, *120*, 1505–1512. [\[CrossRef\]](#)
103. Piascik, M.T.; Guarino, R.D.; Smith, M.S.; Soltis, E.E.; Saussy, D.L.; Perez, D.M. The specific contribution of the novel alpha-1D adrenoceptor to the contraction of vascular smooth muscle. *J. Pharmacol. Exp. Ther.* **1995**, *275*, 1583. [\[PubMed\]](#)
104. Piascik, M.T.; Hrometz, S.L.; Edelmann, S.E.; Guarino, R.D.; Hadley, R.W.; Brown, R.D. Immunocytochemical Localization of the Alpha-1B Adrenergic Receptor and the Contribution of This and the Other Subtypes to Vascular Smooth Muscle Contraction: Analysis with Selective Ligands and Antisense Oligonucleotides. *J. Pharmacol. Exp. Ther.* **1997**, *283*, 854.
105. Martinez, L.; Carmona, L.; Villalobos-Molina, R. Vascular alpha 1D-adrenoceptor function is maintained during congestive heart failure after myocardial infarction in the rat. *Arch. Med. Res.* **1999**, *30*, 290–297. [\[CrossRef\]](#)
106. Cavalli, A.; Lattion, A.L.; Hummler, E.; Nenniger, M.; Pedrazzini, T.; Aubert, J.F.; Michel, M.C.; Yang, M.; Lembo, G.; Vecchione, C.; et al. Decreased blood pressure response in mice deficient of the α_{1B} -adrenergic receptor. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 11589–11594. [\[CrossRef\]](#)
107. Muramatsu, I.; Kigoshi, S.; Ohmura, T. Subtypes of α_1 -Adrenoceptors Involved in Noradrenaline-Induced Contractions of Rat Thoracic Aorta and Dog Carotid Artery. *Jpn. J. Pharmacol.* **1991**, *57*, 535–544. [\[CrossRef\]](#)
108. Suzuki, E.; Tsujimoto, G.; Tamura, K.; Hashimoto, K. Two pharmacologically distinct alpha 1-adrenoceptor subtypes in the contraction of rabbit aorta: Each subtype couples with a different Ca^{2+} signalling mechanism and plays a different physiological role. *Mol. Pharmacol.* **1990**, *38*, 725.
109. Shibata, K.; Hirasawa, A.; Foglar, R.; Ogawa, S.; Tsujimoto, G. Effects of Quinidine and Verapamil on Human Cardiovascular α_1 -Adrenoceptors. *Circulation* **1998**, *97*, 1227–1230. [\[CrossRef\]](#)
110. Fetscher, C.; Schäfers, R.F.; Philipp, T.; Michel, M.C.; Chen, H.; Wambach, G. Effects of noradrenaline and neuropeptide Y on rat mesenteric microvessel contraction. *Naunyn-Schmiedeberg’s Arch. Pharmacol.* **1996**, *353*, 314–323. [\[CrossRef\]](#)
111. Chen, H.; Bischoff, A.; Schäfers, R.F.; Wambach, G.; Philipp, T.; Michel, M.C. Vasoconstriction of rat renal interlobar arteries by noradrenaline and neuropeptide Y. *J. Auton. Pharmacol.* **1997**, *17*, 137–146. [\[CrossRef\]](#) [\[PubMed\]](#)
112. Rokosh, D.G.; Simpson, P.C. Knockout of the α_1 A/C-adrenergic receptor subtype: The α_1 A/C is expressed in resistance arteries and is required to maintain arterial blood pressure. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 9474–9479. [\[CrossRef\]](#) [\[PubMed\]](#)

113. Tanoue, A.; Nasa, Y.; Koshimizu, T.; Shinoura, H.; Oshikawa, S.; Kawai, T.; Sunada, S.; Takeo, S.; Tsujimoto, G. The α_{1D} -adrenergic receptor directly regulates arterial blood pressure via vasoconstriction. *J. Clin. Investig.* **2002**, *109*, 765–775. [[CrossRef](#)]
114. Forster, C.; Carter, S.; Armstrong, P. Vascular smooth muscle responsiveness to noradrenaline and phenylephrine following experimental heart failure in dogs. *Cardiovasc. Res.* **1989**, *23*, 489–497. [[CrossRef](#)]
115. Forster, C.; Armstrong, P.W. Pacing-induced heart failure in the dog: Evaluation of peripheral vascular alpha-adrenoceptor subtypes. *J. Cardiovasc. Pharmacol.* **1990**, *16*, 708–718. [[CrossRef](#)]
116. Forster, C. Interaction of 5-methyl-urapidil with α_1 -adrenoceptors in canine blood vessels: Impact of pacing-induced heart failure. *Eur. J. Pharmacol.* **1996**, *318*, 55–63. [[CrossRef](#)]
117. Le Tran, Y.; Forster, C. Chloroethylclonidine and alpha-adrenoceptor agonist interaction in blood vessels following heart failure. *Eur. J. Pharmacol.* **1997**, *336*, 177–185. [[CrossRef](#)]
118. Lurie, K.G.; Tsujimoto, G.; Hoffman, B.B. Desensitization of alpha-1 adrenergic receptor-mediated vascular smooth muscle contraction. *J. Pharmacol. Exp. Ther.* **1985**, *234*, 147–152.
119. Forster, C.; Campbell, P.M. Nifedipine inhibits responses to alpha-adrenoceptor stimulation in canine blood vessels: Impact of heart failure. *Eur. J. Pharmacol.* **1993**, *242*, 119–128. [[CrossRef](#)]
120. Forster, C. Inhibition of vascular contractions to alpha-adrenoceptor agonists by polymyxin B: Impact of heart failure state. *Eur. J. Pharmacol.* **1995**, *283*, 241–250. [[CrossRef](#)]
121. Bergdahl, A.; Valdemarsson, S.; Sun, X.Y.; Hedner, T.; Edvinsson, L. Enhanced Ca^{2+} -induced contractions and attenuated alpha-adrenoceptor responses in resistance arteries from rats with congestive heart failure. *Eur. J. Heart Fail.* **2001**, *3*, 7–13. [[CrossRef](#)]
122. Koida, S.; Ohyanagi, M.; Ueda, A.; Mori, Y.; Iwasaka, T. Mechanism of increased alpha-adrenoceptor-mediated contraction in small resistance arteries of rats with heart failure. *Clin. Exp. Pharmacol. Physiol.* **2006**, *33*, 47–52. [[CrossRef](#)]
123. Trautner, S.; Amtorp, O.; Boesgaard, S.; Andersen, C.B.; Galbo, H.; Haunsøe, S.; Sheykhzade, M. Noradrenaline-induced increases in calcium and tension in skeletal muscle conductance and resistance arteries from rats with post-infarction heart failure. *Eur. J. Pharmacol.* **2006**, *537*, 143–154. [[CrossRef](#)]
124. Schrier, R.W.; Abraham, W.T. Hormones and Hemodynamics in Heart Failure. *N. Engl. J. Med.* **1999**, *341*, 577–585. [[CrossRef](#)]
125. Maze, M.; Spiss, C.K.; Tsujimoto, G.; Hoffman, B.B. Epinephrine infusion induces hyporesponsiveness of vascular smooth muscle. *Life Sci.* **1985**, *37*, 1571–1578. [[CrossRef](#)]
126. Snively, M.D.; Ziegler, M.G.; Insel, P.A. Subtype-Selective Down-Regulation of Rat Renal Cortical α - and β -Adrenergic Receptors by Catecholamines. *Endocrinology* **1985**, *117*, 2182–2189. [[CrossRef](#)]
127. Wikberg, J.E.S.; Akers, M.; Caron, M.G.; Hagen, P.O. Norepinephrine-induced down regulation of alpha₁ adrenergic receptors in cultured rabbit aorta smooth muscle cells. *Life Sci.* **1983**, *33*, 1409–1417. [[CrossRef](#)]
128. Leeb-Lundberg, L.M.; Cotecchia, S.; DeBlasi, A.; Caron, M.G.; Lefkowitz, R.J. Regulation of adrenergic receptor function by phosphorylation. I. Agonist-promoted desensitization and phosphorylation of alpha₁-adrenergic receptors coupled to inositol phospholipid metabolism in DDT1 MF-2 smooth muscle cells. *J. Biol. Chem.* **1987**, *262*, 3098–3105.
129. Townsley, M.I.; Pitts, V.H.; Ardell, J.L.; Zhao, Z.; Johnson, W.H., Jr. Altered pulmonary microvascular reactivity to norepinephrine in canine pacing-induced heart failure. *Circ. Res.* **1994**, *75*, 347–356. [[CrossRef](#)]
130. Bergdahl, A.; Valdemarsson, S.; Pantev, E.; Ottosson, A.; Feng, Q.P.; Sun, X.Y.; Hedner, T.; Edvinsson, L. Modulation of vascular contractile responses to α_1 - and α_2 -adrenergic and neuropeptide Y receptor stimulation in rats with ischaemic heart failure. *Acta Physiol. Scand.* **1995**, *154*, 429–437. [[CrossRef](#)]
131. Tamagawa, K.; Saito, T.; Oikawa, Y.; Maehara, K.; Yaoita, H.; Maruyama, Y. Alterations of alpha-adrenergic modulations of coronary microvascular tone in dogs with heart failure. *J. Card. Fail.* **2005**, *11*, 388–395. [[CrossRef](#)]
132. Main, J.S.; Forster, C.; Armstrong, P.W. Inhibitory role of the coronary arterial endothelium to alpha-adrenergic stimulation in experimental heart failure. *Circ. Res.* **1991**, *68*, 940–946. [[CrossRef](#)]
133. Stassen, F.R.M.; Fazzi, G.E.; Leenders, P.J.A.; Smits, J.F.M.; de Mey, J.G.R. Coronary Arterial Hyperreactivity and Mesenteric Arterial Hyporeactivity After Myocardial Infarction in the Rat. *J. Cardiovasc. Pharmacol.* **1997**, *29*, 780–788. [[CrossRef](#)]

134. Feng, Q.; Sun, X.; Lu, X.; Edvinsson, L.; Hedner, T. Decreased responsiveness of vascular postjunctional α_1 -, α_2 -adrenoceptors and neuropeptide Y1 receptors in rats with heart failure. *Acta Physiol. Scand.* **1999**, *166*, 285–291. [[CrossRef](#)]
135. Ahmadiasl, N.; Rostami, A.; Mohammadi, N.M.; Rajabi, F. Effects of noradrenaline and KCl on peripheral vessels in doxorubicin induced model of heart failure. *Pathophysiology* **2002**, *8*, 259–262. [[CrossRef](#)]
136. Ramchandra, R.; Xing, D.T.; Matear, M.; Lambert, G.; Allen, A.M.; May, C.N. Neurohumoral interactions contributing to renal vasoconstriction and decreased renal blood flow in heart failure. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2019**, *317*, R386–R396. [[CrossRef](#)]
137. Goldsmith, S.R.; Francis, G.S.; Cohn, J.N. Norepinephrine infusions in congestive heart failure. *Am. J. Cardiol.* **1985**, *56*, 802–804. [[CrossRef](#)]
138. Borow, K.M.; Neumann, A.; Arensman, F.W.; Yacoub, M.H. Cardiac and peripheral vascular responses to adrenoceptor stimulation and blockade after cardiac transplantation. *J. Am. Coll. Cardiol.* **1989**, *14*, 1229–1238. [[CrossRef](#)]
139. Angus, J.A.; Ferrier, C.P.; Sudhir, K.; Kaye, D.M.; Jennings, G.L. Impaired contraction and relaxation in skin resistance arteries from patients with congestive heart failure. *Cardiovasc. Res.* **1993**, *27*, 204–210. [[CrossRef](#)]
140. Barrett-O’Keefe, Z.; Lee, J.F.; Ives, S.J.; Trinity, J.D.; Witman, M.A.H.; Rossman, M.J.; Groot, H.J.; Sorensen, J.R.; Morgan, D.E.; Nelson, A.D.; et al. α -Adrenergic receptor regulation of skeletal muscle blood flow during exercise in heart failure patients with reduced ejection fraction. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2019**, *316*, R512–R524. [[CrossRef](#)]
141. Wilson, J.R.; Matthai, W.; Lanoce, V.; Frey, M.; Ferraro, N. Effect of experimental heart failure on peripheral sympathetic vasoconstriction. *Am. J. Physiol.* **1988**, *254*, H727–H733. [[CrossRef](#)]
142. Mulder, P.; Compagnon, P.; Devaux, B.; Richard, V.; Henry, J.P.; Elfertak, L.; Wimart, M.C.; Thibout, E.; Comoy, E.; Mace, B.; et al. Response of large and small vessels to alpha and beta adrenoceptor stimulation in heart failure: Effect of angiotensin converting enzyme inhibition. *Fundam. Clin. Pharmacol.* **1997**, *11*, 221–230. [[CrossRef](#)]
143. McMillon, R.K.; Townsley, M.I. Alpha-adrenergic vasoreactivity of canine intrapulmonary bronchial arteries in pacing-induced heart failure. *Am. J. Physiol.* **1999**, *277*, H1392–H1402.
144. Indolfi, C.; Maione, A.; Volpe, M.; Rapacciuolo, A.; Esposito, G.; Ceravolo, R.; Rendina, V.; Condorelli, M.; Chiariello, M. Forearm vascular responsiveness to alpha 1- and alpha 2-adrenoceptor stimulation in patients with congestive heart failure. *Circulation* **1994**, *90*, 17–22. [[CrossRef](#)]
145. Schwinn, D.A.; McIntyre, R.W.; Hawkins, E.D.; Kates, R.A.; Reves, J.G. alpha 1-Adrenergic responsiveness during coronary artery bypass surgery: Effect of preoperative ejection fraction. *Anesthesiology* **1988**, *69*, 206–217. [[CrossRef](#)] [[PubMed](#)]
146. Teerlink, J.R.; Gray, G.A.; Clozel, M.; Clozel, J.P. Increased vascular responsiveness to norepinephrine in rats with heart failure is endothelium dependent. Dissociation of basal and stimulated nitric oxide release. *Circulation* **1994**, *89*, 393–401. [[CrossRef](#)]
147. Van Tassel, B.W.; Rondina, M.T.; Huggins, F.; Gilbert, E.M.; Munger, M.A. Carvedilol increases blood pressure response to phenylephrine infusion in heart failure subjects with systolic dysfunction: Evidence of improved vascular α_1 -adrenoreceptor signal transduction. *Am. Heart J.* **2008**, *156*, 315–321. [[CrossRef](#)]
148. Mey, J.D.; Vanhoutte, P.M. Uneven distribution of postjunctional alpha 1-and alpha 2-like adrenoceptors in canine arterial and venous smooth muscle. *Circ. Res.* **1981**, *48*, 875–884. [[CrossRef](#)]
149. Van Vessel, M.E.; Petrus, A.H.J.; Palmen, M.; Braun, J.; Schalij, M.J.; Klautz, R.J.M.; Beeres, S.L.M.A. Vasoplegia after Restrictive Mitral Annuloplasty for Functional Mitral Regurgitation in Patients with Heart Failure. *J. Cardiothorac. Vasc. Anesth.* **2019**, *33*, 3273–3280. [[CrossRef](#)] [[PubMed](#)]
150. Argenziano, M.; Chen, J.M.; Choudhri, A.F.; Cullinane, S.; Garfein, E.; Weinberg, A.D.; Smith, C.R., Jr.; Rose, E.A.; Landry, D.W.; Oz, M.C. Management of vasodilatory shock after cardiac surgery: Identification of predisposing factors and use of a novel pressor agent. *J. Thorac. Cardiovasc. Surg.* **1998**, *116*, 973–980. [[CrossRef](#)]
151. Orozco Vinasco, D.M.; Triana Schoonewolff, C.A.; Orozco Vinasco, A.C. Vasoplegic syndrome in cardiac surgery: Definitions, pathophysiology, diagnostic approach and management. *Rev. Esp. Anesthesiol. Reanim.* **2019**, *66*, 277–287. [[CrossRef](#)]

152. Van Vessel, M.E.; Palmen, M.; Couperus, L.E.; Stijnen, T.; Berendsen, R.R.; Aarts, L.; de Jonge, E.; Klautz, R.J.M.; Schalij, M.J.; Beeres, S. Vasoplegia after Surgical Left Ventricular Restoration: 2-Year Follow-Up. *Ann. Thorac. Surg.* **2018**, *106*, 1371–1378. [[CrossRef](#)] [[PubMed](#)]
153. Tecson, K.M.; Lima, B.; Lee, A.Y.; Raza, F.S.; Ching, G.; Lee, C.H.; Felius, J.; Baxter, R.D.; Still, S.; Collier, J.D.G.; et al. Determinants and Outcomes of Vasoplegia Following Left Ventricular Assist Device Implantation. *J. Am. Heart Assoc.* **2018**, *7*, e008377. [[CrossRef](#)]
154. Stassen, F.R.; Maas, R.G.; Schiffrers, P.M.; Janssen, G.M.; de Mey, J.G. A positive and reversible relationship between adrenergic nerves and alpha-1A adrenoceptors in rat arteries. *J. Pharmacol. Exp. Ther.* **1998**, *284*, 399–405.
155. Bristow, M.R.; Feldman, A.M.; Adams, K.F., Jr.; Goldstein, S. Selective versus nonselective beta-blockade for heart failure therapy: Are there lessons to be learned from the COMET trial? *J. Card. Fail.* **2003**, *9*, 444–453. [[CrossRef](#)]
156. Furberg, C.D.; Wright, J.T.; Davis, B.R.; Cutler, J.A.; Alderman, M.; Black, H.; Cushman, W.; Grimm, R.; Haywood, J.; Leenen, F.; et al. Major Cardiovascular Events in Hypertensive Patients Randomized to Doxazosin vs Chlorthalidone; The Antihypertensive and Lipid-Lowering Treatment to Prevent Heart Attack Trial (ALLHAT). *JAMA* **2000**, *283*, 1967–1975.
157. O’Connell, T.D.; Swigart, P.M.; Rodrigo, M.C.; Ishizaka, S.; Joho, S.; Turnbull, L.; Tecott, L.H.; Baker, A.J.; Foster, E.; Grossman, W.; et al. Alpha₁-adrenergic receptors prevent a maladaptive cardiac response to pressure overload. *J. Clin. Investig.* **2006**, *116*, 1005–1015. [[CrossRef](#)]
158. Cohn, J.N. The Vasodilator-Heart Failure Trials (V-HeFT). Mechanistic data from the VA Cooperative Studies. Introduction. *Circulation* **1993**, *87*, VII–VI4.
159. O’Connell, T.D.; Ishizaka, S.; Nakamura, A.; Swigart, P.M.; Rodrigo, M.C.; Simpson, G.L.; Cotecchia, S.; Rokosh, D.G.; Grossman, W.; Foster, E.; et al. The α_1 A/C- and α_1 B-adrenergic receptors are required for physiological cardiac hypertrophy in the double-knockout mouse. *J. Clin. Investig.* **2003**, *111*, 1783–1791. [[CrossRef](#)]
160. Kukin, M.L.; Kalman, J.; Mannino, M.; Freudenberger, R.; Buchholz, C.; Ocampo, O. Combined alpha-beta blockade (doxazosin plus metoprolol) compared with beta blockade alone in chronic congestive heart failure. *Am. J. Cardiol.* **1996**, *77*, 486–491. [[CrossRef](#)]
161. Tham, T.C.; Guy, S.; McDermott, B.J.; Shanks, R.G.; Riddell, J.G. The dose dependency of the alpha- and beta-adrenoceptor antagonist activity of carvedilol in man. *Br. J. Clin. Pharmacol.* **1995**, *40*, 19–23. [[CrossRef](#)]
162. Kubo, T.; Azevedo, E.R.; Newton, G.E.; Parker, J.D.; Floras, J.S. Lack of evidence for peripheral alpha₁-adrenoceptor blockade during long-term treatment of heart failure with carvedilol. *J. Am. Coll. Cardiol.* **2001**, *38*, 1463. [[CrossRef](#)]

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