

# **Genetics of migraine aura: an update**

Boer, I. de; Terwindt, G.M.; Maagdenberg, A.M.J.M. van den

# **Citation**

Boer, I. de, Terwindt, G. M., & Maagdenberg, A. M. J. M. van den. (2020). Genetics of migraine aura: an update. *Journal Of Headache And Pain*, *21*(1). doi:10.1186/s10194-020-01125-2

Version: Publisher's Version License: [Creative Commons CC BY 4.0 license](https://creativecommons.org/licenses/by/4.0/) Downloaded from: <https://hdl.handle.net/1887/3182714>

**Note:** To cite this publication please use the final published version (if applicable).

# **REVIEW ARTICLE External intervention of the Community Community Community Community Community Community Community**

Check for updates

# Genetics of migraine aura: an update

Irene de Boer<sup>1</sup> $\odot$ [,](http://orcid.org/0000-0002-7261-762X) Gisela M. Terwindt<sup>1†</sup> and Arn M. J. M. van den Maagdenberg<sup>1,2\*†</sup>

## Abstract

Migraine is a common brain disorder with a large genetic component. Of the two main migraine types, migraine with aura and migraine without aura, the genetic underpinning in the former is least understood. Given the evidence from epidemiological studies in cohorts and families that the genetic contribution is highest in migraine with aura, this seems paradoxical. Various genetic approaches have been applied to identify genetic factors that confer risk for migraine. Initially, so-called candidate gene associations studies (CGAS) have been performed that test DNA variants in genes prioritized based on presumed a priori knowledge of migraine pathophysiology. More recently, genome-wide association studies (GWAS) tested variants in any gene in an hypothesis-free manner. Whereas GWAS in migraine without aura, or the more general diagnosis migraine have already identified dozens of gene variants, the specific hunt for gene variants in migraine with aura has been disappointing. The only GWAS specifically investigating migraine with aura yielded only one single associated single nucleotide polymorphism (SNP), near MTDH and PGCP, with genome-wide significance. However, interrogation of all genotyped SNPs, so beyond this one significant hit, was more successful and led to the notion that migraine with aura and migraine without aura are genetically more alike than different. Until now, most relevant genetic discoveries related to migraine with aura came from investigating monogenetic syndromes with migraine aura as a prominent phenotype (i.e. FHM, CADASIL and FASPS). This review will highlight the genetic findings relevant to migraine with aura.

Keywords: Migraine, Aura, Genetics, GWAS, Complex, Variant, Mutation, Monogenic

### Background

Migraine is a common brain disorder characterised by attacks of severe typically unilateral headache with nausea, vomiting, phono- and/or photophobia [[1](#page-8-0)]. In up to a third of patients, attacks are accompanied by an aura, consisting of transient focal neurological symptoms that are most often visual, but may also include sensory symptoms and/or speech disturbances. If the aura encompasses motor weakness the disease is diagnosed as hemiplegic migraine [[1\]](#page-8-0). The likely underlying mechanism of the aura is cortical spreading depolarisation (CSD), a brief wave of neuronal and glial depolarization

<sup>&</sup>lt;sup>2</sup> Department of Human Genetics, Leiden University Medical Center, Albinusdreef 2, PO Box 9600, 2300 RC Leiden, The Netherlands



that slowly self-propagates across the cerebral cortex, originating in the occipital cortex, that is followed by long-lasting suppression of brain activity [\[2](#page-8-0)–[4\]](#page-8-0).

Migraine has a strong genetic component. Epidemiological studies have shown that migraine runs in families and that migraine concordance rates are 1.5- to 2-fold higher in monozygotic compared to dizygotic twins [[5](#page-8-0)–[7](#page-8-0)]. A stronger family history of migraine is associated with migraine with aura, as well as a lower age-at-onset and a higher number of medication days [\[8\]](#page-8-0), which suggests a higher genetic susceptibility in this migraine type [\[8](#page-8-0)]. Several approaches, discussed below, have been used to identify genetic factors that confer migraine risk. The general assumption is that in common polygenic forms of migraine multiple DNA variants, each with a small effect size, together with environmental factors confer disease risk, whereas in rare monogenic migraine syndromes a single DNA mutation suffices to cause disease in a patient.

© The Author(s), 2020 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit [http://creativecommons.org/licenses/by/4.0/.](http://creativecommons.org/licenses/by/4.0/) The Creative Commons Public Domain Dedication waiver [\(http://creativecommons.org/publicdomain/zero/1.0/](http://creativecommons.org/publicdomain/zero/1.0/)) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

<sup>\*</sup> Correspondence: [A.M.J.M.van\\_den\\_Maagdenberg@lumc.nl](mailto:A.M.J.M.van_den_Maagdenberg@lumc.nl) †

<sup>&</sup>lt;sup>+</sup>Gisela M. Terwindt and Arn M. J. M. van den Maagdenberg contributed equally to this work.

<sup>&</sup>lt;sup>1</sup>Department of Neurology, Leiden University Medical Center, Albinusdreef 2, PO Box 9600, 2300 RC Leiden, The Netherlands

Candidate gene association studies in migraine with aura Initially, candidate gene association studies (CGAS) were performed that test whether specific genetic markers in genes that had been selected based on prior hypotheses that they are implicated in migraine disease pathways, show a different allele frequency in migraine cases vs. controls. CGAS have been performed for many candidate genes for both migraine types (for reviews see [\[9,](#page-8-0) [10](#page-8-0)]). Among the largest studies is one that included 841 migraine with aura patients and 884 controls in the discovery cohort and tested several thousand genetic markers in 155 ion transporter genes, but replication in an independent data set was essentially negative  $[11]$  $[11]$  $[11]$ . A large CGAS that systematically re-evaluated association findings of the 21 most promising genes from CGAS, but now using a much larger data set of several thousand cases and controls, did not lead to significant results [[12](#page-8-0)]. Hence one may draw the conclusion that many, if not all, of the initial CGAS results may be false-positives. Results are disappointing most likely because of methodological issues, foremost because, almost without exception, CGAS had too small sample sizes (in most cases less than a few hundred cases and an equal number of controls were tested). Another likely explanation is that the diagnosis of aura symptomatology is difficult to obtain through questionnaires and is often not validated through direct interviews by trained headache physicians [\[13,](#page-8-0) [14](#page-8-0)]. In addition, cases and controls may not have been adequately matched for genetic background, gender and age. Finally, without replication of the main results in an independent population in the same study, the risk of reporting false positive results is high. It remains to be tested whether rarer variants (instead of the more common variants usually tested in a CGAS) may in fact elevate risk in specific subgroups of migraine patients. There is also the distinct possibility that the selected genes are not those that confer genetic risk, even if they belong to molecular pathways clearly implicated in migraine pathophysiology (such as genes that play a role in serotonin and dopamine pathways). Perhaps genes regulating (in)directly these pathways would have been better candidates.

#### Genome-wide association study in migraine with aura

Due to improved technology and the development of cost-effective genotyping platforms genome-wide association studies (GWAS) became feasible in the last decade. Unlike CGAS, in a GWAS no prior hypothesis is formulated about the possible role of a DNA variant. In a GWAS, in one experiment, several hundred thousand to millions of single nucleotide polymorphisms (SNPs), close to equally distributed over the whole genome, are tested for association with a trait (e.g. a disease such as migraine) by assessing differences in allele frequencies between patients and controls. Findings are rigorously

corrected for the vast number of tests as an association is considered genome-wide significant only if the p-vale is  $< 5 \times 10^{-8}$ . In a GWAS typically only variants with a moderate to high minor allele frequency ( $\geq 0.05$ ) are genotyped; so alternative approaches, foremost next generation sequencing (NGS)-based whole-exome and whole-genome approaches, are used to assess the contribution of rarer variants.

The first migraine GWAS consisted of 2731 patients with migraine with aura, obtained from three European specialized headache clinics (from Finland, Germany and the Netherlands), and 10,747 population-matched controls in the discovery cohort [\[15](#page-8-0)]. Surprisingly, only one single SNP, rs835740, reached genome-wide significance in this sample  $(p = 5.38 \times 10^{-9})$ , odds ratio = 1.23). The finding was successfully replicated in an independent set finding was successfully replicated in an independent set of samples from Iceland, Denmark, Germany and the Netherlands encompassing 3202 cases and 40,062 controls and was highly significant in an overall metaanalysis ( $p = 1.69 \times 10^{-11}$ , odds ratio = 1.18). The associated SNP is located between MTDH (better known as astrocyte elevated gene 1 (AEG-1)) and PGCP (encoding plasma glutamate carboxypeptidase); both gene products have been implicated in glutamate homeostasis. A subsequent analysis of gene expression in lymphoblastoid cell lines demonstrated that the risk allele correlated with a higher transcript level of the *MTDH* gene [[15\]](#page-8-0). However, the relation between MTDH and migraine remains controversial, also because the SNP did not show significance in subsequent meta-analyses of GWAS data, neither in studies that investigated association with migraine  $(p = 5 \times 10^{-3}$  [\[16](#page-8-0)],  $p = 0.12$  [[17\]](#page-8-0)) nor migraine with aura (both when the cohorts used to identify the locus were included  $(p = 1.49 \times 10^{-4})$  or excluded  $(p = 0.59)$  [17]) 0.59) [[17\]](#page-8-0)).

The most recent migraine GWAS included 59,674 cases and 316,078 controls and identified up to 38 distinct genomic regions associated with migraine [\[16](#page-8-0)]. When considering the genes closest to the associated SNPs, which is the typical way SNPs are being linked to implicated genes, it was concluded that current GWAS findings seem to link to neuronal, vascular, metalloproteinase, pain and metal-iron-related pathways [[16](#page-8-0)–[18](#page-8-0)]. Only in sub-analyses the contribution of associated loci to specific migraine types was given further consideration. Whereas seven loci (i.e. near TSPAN2, TRPM8, PHACTR1, FHL5, ASTN2, near FGF6, and LRP1) were associated with migraine without aura (in a sample of 8348 cases and 139,622 controls), none were associated with migraine with aura (6332 cases vs. 144,883 controls) [\[16](#page-8-0)]. Of note, the seven loci had already been identified in the initial analysis with migraine in general, not subdivided in its subtypes, as phenotype. Given that two-third of migraineurs suffers from migraine without

aura, this finding is perhaps not that surprising as it merely shows that the most abundant migraine type likely seems responsible for the association signals.

#### Is the genetic component in migraine with and without aura different?

A possible explanation for why GWAS appears more successful in migraine without aura might be that both migraine types are distinct disorders with a different genetic architecture. The suggestion fuels the longstanding debate whether or not both types are different disease entities. It should be noted that the migraine types are diagnosed differently according to the ICHD-III classification as migraine with aura, unlike migraine without aura, does not have to fulfil two out of four headache characteristics (moderate/severe pain intensity, unilateral location, pulsating quality, aggravation by routine physical activity) and accompanying symptoms (photophobia and phonophobia and/or nausea or vomiting). Moreover, to be labeled as patient the experienced number of attacks is less in migraine with aura (at least two attacks) compared with migraine without aura (at least five attacks) [[1](#page-8-0)]. Notwithstanding, there are long-recognized clinical observations suggestive for a shared etiology of both migraine types, not only because attacks of both types can co occur in a patient (or family) or that the migraine type can change over lifetime (aura attacks may develop in the elderly patient) [\[19](#page-8-0)].

It was the hope that GWAS findings could settle the debate. One argument for the distinction of migraine types is the previously mentioned observation that GWAS particularly identified loci for migraine without aura. Another genetic argument is that a polygenetic risk score analysis of 21 migraine-associated SNPs from GWAS showed association only with migraine without aura [\[20](#page-8-0)]. Importantly, it should be noted that in that study participants with migraine with aura (152 cases) were fewer than participants with migraine without aura (295 cases), so the study likely lacks statistical power to pick up a signal for migraine with aura. A recent study that took into account all available genetic information, so not just the migraine-associated SNPs ('top hits'), in a much larger data set, however, found statistical evidence for a significant overlap of genes associated with migraine with aura (4505 cases) and migraine without aura (4038 cases) and concluded that the genetic architecture of both types is more alike than dissimilar [\[21](#page-8-0)]. Likewise, a heterogeneity analysis of SNP effects in migraine with aura and migraine without aura showed that for 12 migraine-associated SNPs the risk-increasing allele was the same [[22\]](#page-8-0). In fact, when more than 23,000 SNPs were analyzed the majority of SNP effects were in the same direction in migraine with aura (5118 cases) and

migraine without aura (7101 cases) [[22](#page-8-0)]. Finally, the international Brainstorm consortium that compared GWAS data of over 20 brain disorders (and investigated genetic information of 265,218 patients and 784,643 controls) found substantial genetic correlation between migraine with aura and migraine without aura [\[23](#page-8-0)]. In conclusion, although migraine with aura and migraine without aura seem genetically more alike than dissimilar, the challenge remains to identify more of the underlying causal variants as only a fraction of them have been discovered.

#### Where is the missing heritability of migraine with aura?

Like what has been proposed for other complex diseases, one can postulate that migraine with aura may be determined by multiple rarer medium- to high-risk alleles that are not captured by a GWAS approach  $[24]$  $[24]$  (Fig. [1](#page-4-0)). Finding an association for rarer alleles is notoriously difficult as it requires the analysis of even larger welldefined migraine with aura cohorts and representation of rarer variants in SNP arrays. For rarer variants a more logical approach is to genotype them by NGS, which has not been done yet.

Part of the missing heritability could also be explained by genetic variants in mitochondrial DNA (mtDNA). Mitochondrial DNA is separated from nuclear DNA and inherited exclusively along the maternal lineage. Each cell contains hundreds to thousands of copies of mtDNA, and there may be genetic variation between mtDNA molecules [[25\]](#page-8-0). In fact, migraine-like headache has been reported in several mitochondrial disorders, such as mitochondrial encephalopathy, lactic acidosis and stroke-like episodes (MELAS) [[26\]](#page-8-0). Additionally, structurally abnormal mitochondria and findings indicative of impaired energy metabolism have been demonstrated in migraine [[27\]](#page-8-0). While small CGAS focusing on mtDNA have reported associations [\[28\]](#page-8-0), these could not be replicated in a recent mitochondrial GWAS (with 4021 migraine cases included) [[29\]](#page-9-0). Furthermore, this mitochondrial GWAS found no additional associations with migraine [[29\]](#page-9-0). It seems likely that a larger sample size is required to find variants robustly associated with migraine.

Another explanation for the missing heritability could be epigenetic processes. Epigenetics encompasses changes to the DNA structure without changing the genetic code and affects gene expression and has also been implicated in neurological disorders [[30](#page-9-0)]. Epigenetic changes can be dynamic, be passed on through cell divisions or transgenerational inheritance. The main epigenetic modifications are DNA methylation and post-translational modification of histone proteins. So far, small steps have been taken to dis-sect the epigenetic contribution towards migraine [\[31\]](#page-9-0). The first genome-wide analysis of DNA methylation in migraine

<span id="page-4-0"></span>

identified 62 independent differentially methylated regions in blood samples without distinguishing between migraine without and with aura [\[32\]](#page-9-0). While epigenetic factors might contribute towards migraine pathophysiology, for instance towards migraine chronification, this field is still in its infancy and therefore it is too soon to speculate towards its relevance for migraine with aura.

#### Genes in monogenetic migraine with aura syndromes

Until now successful identification of genetic factors relevant to migraine with aura almost exclusively came from investigating rare monogenetic syndromes. The identification of causal genes for such syndromes is more straightforward than for polygenic disorders, already because the hunt is for a single genetic factor that is sufficient to cause disease in a patient (Fig. 1). Sequencing of the coding regions of the DNA of a patient (and family members) by direct sequencing of exons of possible candidate genes, in the past, and nowadays by sequencing all coding regions

in a single experiment by whole exome sequencing has led to the identifications of thousands of disease genes for many disorders [\[33\]](#page-9-0); among which a number for migraine with aura syndromes that will be discussed below. Whole exome sequencing is more and more ready to be used in the diagnostic process of migraine with aura syndromes, as shown in a large Finnish sample [\[34\]](#page-9-0).

#### Familial hemiplegic migraine

The most investigated monogenetic migraine with aura syndrome is hemiplegic migraine [[1\]](#page-8-0). In patients with hemiplegic migraine, the auras include motor weakness in addition to other focal neurological features commonly seen in migraine with aura patients. Additional clinical symptoms in hemiplegic migraine may include cerebellar ataxia, seizures, cognitive impairment, and decreased consciousness [\[35\]](#page-9-0). Of note, many patients also suffer from attacks of migraine with aura and/or migraine without aura [[36\]](#page-9-0). This finding, but also the

observation that factors that trigger attacks are the same as for migraine with aura  $[37]$  $[37]$ , supports the hypothesis that hemiplegic migraine is part of the spectrum of migraine disorders. DNA sequencing of candidate genes in genomic regions shared by affected family members, have identified causal mutations in three hemiplegic migraine genes: CACNA1A (FHM1), ATP1A2 (FHM2), SCN1A (FHM3) [[38\]](#page-9-0). In recent years, whole genome/ whole exon NGS was used to identify additional causal genes in those patients in which no mutation in one of the three genes had been found. Surprisingly, this has not led to undisputed additional genes [[38](#page-9-0)]. One important explanation may be that there is clinical heterogeneity among patients with a hemiplegic migraine diagnosis [[38](#page-9-0)]. Chances of finding a causal mutation in hemiplegic migraine patients in one of the known genes is higher in patients with a lower age of disease onset, when there are more affected family members, and when attacks are (1) triggered by mild head trauma, (2) include confusion, (3) have extensive motor weakness, (4) have symptoms of brainstem pathology, or (5) are associated with brain edema; mental retardation and progressive ataxia are only found in hemiplegic migraine patients with a causal mutation in one of the three known genes.

#### Other monogenetic migraine with aura syndromes

In patients with monogenic cerebral small vessel diseases a high prevalence of migraine with aura can be observed. Almost half of the patients with cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy (CADASIL) suffer from migraine with aura, which is often the first symptom of disease [\[39](#page-9-0)]. Other symptoms of CADASIL include mid-adult onset of recurrent ischemic stroke, cognitive decline progressing to dementia, mood disturbances and apathy. CADASIL is caused by mutations, typically affecting the number of cysteines, in Notch3 that plays an important role in vascular smooth muscle cell functioning [[40](#page-9-0), [41\]](#page-9-0). Another small vessel disease in which migraine with (and without) aura is often present is retinal vasculopathy with cerebral leukoencephalopathy and systemic manifestations (RVCL-S) [[42](#page-9-0), [43\]](#page-9-0). RVCL-S is caused by carboxyl terminal truncating mutations in TREX1 [\[44\]](#page-9-0). In addition to migraine (with and without aura), mutation carriers typically suffer from retinal vasculopathy, focal and global neurological symptoms and systemic manifestations including impaired liver and renal function [\[42](#page-9-0)–[44\]](#page-9-0). In RVCL-S migraine onset appears later in life  $(>40 \text{ years})$  [\[43](#page-9-0), [45\]](#page-9-0). This might indicate that migraine is a secondary phenomenon due to the progressive cerebral vasculopathy seen in patients with RVCL-S. Of note, in another cerebral small vessel disease, Dutch-type hereditary cerebral amyloid angiopathy (D-CAA) that is caused by a single point mutation (E693Q) in the amyloid precursor protein (APP) gene, there also is an increased prevalence of migraine with aura [\[46](#page-9-0)]. D-CAA is characterized by intracerebral hemorrhages and vascular dementia [\[47\]](#page-9-0). Migraine with aura is the presenting symptom in close to 80% of patients [\[46\]](#page-9-0). Patients with specific mutations in COL4A1 also appear to have an increased prevalence of migraine with aura, in addition to clinical features such as porencephaly type 1, brain small vessel disease with or without ocular anomalies, nonsyndromic autosomal dominant congenital cataract, hereditary angiopathy with nephropathy, aneurysms, muscle cramps syndrome, and/or tortuosity of retinal arteries [[48](#page-9-0)–[50\]](#page-9-0). At present it is not known what causes the higher prevalence of migraine with aura in these genetic vasculopathies. Possible suggested mechanisms are neurovascular changes, including endothelial and vascular smooth muscle dysfunction and amyloid depositions [[40](#page-9-0), [51](#page-9-0)–[56](#page-9-0)].

Also in patients with non-small vessel disease familial advanced sleep phase syndrome (FASPS), which is caused by specific missense mutations in the CSNK1D gene, migraine with aura is prevalent [\[57](#page-9-0)]. Although little over a handful of mutation carriers in two FASPS families have been reported, almost without exception they also suffer from migraine with aura [[57](#page-9-0)]. Given the fact that FASPS is characterized by a profound phase advancement of the sleep-wake, melatonin and temperature rhythms [\[57](#page-9-0)–[59](#page-9-0)], other genes involved in sleep regulation might be implicated in familial migraine. A good candidate is PER2, which is already a known cause of FASPS [[60](#page-9-0)]; it is however unknown whether *PER2* mutation carriers also have an increased risk for migraine.

Other genes implicated in migraine with aura syndromes? In recent years several genes have been put forward as possible hemiplegic migraine genes (i.e. PRRT2, SLC1A3 and SLC4A4 [\[61](#page-9-0)–[65\]](#page-9-0)), but evaluation of available data should cast doubt on such claims.

PRRT2 - Whereas heterozygous PRRT2 mutations (typically involving similar deletions in the same cytosine stretch) have been identified in a few patients with hemiplegic migraine [\[63,](#page-9-0) [64,](#page-9-0) [66,](#page-9-0) [67](#page-9-0)], it must be taken into account that many hundreds of patients with such mutation who suffer from paroxysmal kinesigenic dyskinesia, benign familial infantile seizures, or infantile convulsion chor-eoathetosis syndrome [[66](#page-9-0)] do *not* have hemiplegic migraine. So the presence of such PRRT2 mutation does not suffice to cause hemiplegic migraine in the Mendelian fashion seen with the three hemiplegic migraine genes [[64](#page-9-0)]. Thus, a PRRT2 mutation may increase the risk of hemiplegic migraine as "add-on" mutation but it seems unlikely that PRRT2 is the FHM4 gene. Still PRRT2 itself is relevant given functional evidence that mutant PRRT2, which affects the neuronal neurotransmitter release machinery, leads to abnormal neuronal hyperexcitability, as was shown in a knockout rat model [[68\]](#page-9-0).

SLC1A3 - The observation of a missense mutation P290R in SLC1A3 in a single patient with alternating hemiplegia, episodic ataxia, and seizures, does not qualify SLC1A3 as a bona fide hemiplegic migraine gene either [[62\]](#page-9-0). SLC1A3 encodes the excitatory amino acid transporter 1 (EAAT1) that is involved in glutamatergic neurotransmission. More recently, SLC1A3 missense mutation T387P, shown to diminish potassium binding to EAAT1, was identified in a patient with hemiplegic migraine, strengthening the believe that EAAT1 might be involved in hemiplegic migraine. Still, doubts concerning causality remains as the mutation did not segregate with the phenotype as the patient's father was a mutation carrier but did not suffer from hemiplegic migraine [\[65](#page-9-0)]. At this moment proof for causality based on isolated cases in which the clinical diagnosis is not firmly established or in which the mutation does not segregate in the family seems not sufficient  $[62, 65]$  $[62, 65]$  $[62, 65]$  $[62, 65]$ .

SLC4A4 - In two patients with proximal renal tubular acidosis and a homozygous frameshift mutation S982NfsX4 in SLC4A4, which encodes for a sodium bicarbonate cotransporter NBCe1, hemiplegic migraine was reported, while some heterozygous carriers reported common forms of migraine [[61](#page-9-0)]. Given the notion that NBCe1 dysfunction affects pH and the functioning of astrocytes, the possible involvement of SLC4A4 in migraine pathophysiology deserves further investigation.

Of note, none of the mutations described in SLC1A3 and SLC4A4 above are found in gnomAD, a sequence variant reference database containing the information of over 100,000 exomes [\[69](#page-9-0)]. Absence in gnomAD indicates that the mutations must be rare and underscores that the mutations may indeed cause disease albeit that the association with migraine needs more proof.

In addition to the potentially (hemiplegic) migraine genes mentioned above, there is also an interesting report on a gene, KCNK18, that was identified in a large multigenerational pedigree suffering from migraine with aura [[70\]](#page-9-0). Thus far, genetic evidence that specific frameshift mutations (such as F139WfsX24) in the KCNK18 gene, encoding the TRESK channel, cause migraine came from only one single family in which the TRESK mutation seemed to segregate perfectly with the phenotype. It was also not reported whether there were other mutations, in the area of linkage, that could equally be disease gene candidates. This scenario is not that unlikely as only a set of 150 ion transporter genes (in the entire genome) were screened for mutations in a set of 110 unrelated migraine patients in the original study [[71\]](#page-9-0). Moreover, loss of one functional copy of TRESK per se does not seem to lead to disease as another variant, C110R, was found in both controls and patients [[70](#page-9-0)]. Where functional analyses of cells expressing the frameshift mutation indicated effect on plasma membrane localization leading to hyperexcitability of trigeminal ganglion neurons [[72](#page-9-0)], this second mutation (C110R) did not increase the excitability of trigeminal ganglion neurons [[73](#page-9-0), [74\]](#page-9-0). Recent intriguing work sheds some light on the complex mechanism on how TRESK mutations could lead to different functional effects, as frameshift mutations F139WfsX24 and Y121LfsX44, but not C110R, lead to an alternative start codon and consequently production of a second TRESK protein, which co-assembles with TREK1 and TREK2 to downregulate their function leading to increased trigeminal ganglion neuron excitability [\[75\]](#page-9-0). This might be the reason why only specific frameshift mutations induce hyperexcitability in trigeminal ganglion neurons. Using CRISPR-Cas9 engineering to correct the F139WfsX24 mutation in induced pluripotent stem cellbased models, a reversal of the neuronal excitability phenotype was demonstrated [\[76](#page-10-0)].

Why shed doubt whether TRESK1 mutations are indeed causal for migraine? Firstly, proof of causality based on one single family, even though segregation is seemingly perfect, is not sufficient, especially when realizing that the linkage region was not thoroughly scanned for other possible causal mutations. Secondly, whereas another frameshift mutation Y121LfsX44 was reported in ClinVar [\[77\]](#page-10-0) (RCV000490385.1) to be associated with migraine (notably without any record clinical details) [[75](#page-9-0)], for a second case no association with disease was reported. Thirdly, and foremost, frameshift mutations in KCNK18 in gnomAD are among the most common variants in KCNK18  $[69]$  $[69]$ . The Y121LfsX44 and the F139WfsX24 mutations were found no less than 665 and 123 times, respectively. Although in theory all individuals may have familial migraine, this sheds reasonable doubt on whether TRESK is indeed a migraine gene. The more likely scenario is that TRESK mutations that lead to inactive protein function are well tolerated, which is also suggested by the observation that there are 12 homozygous carriers reported in gnomAD. While it thus seems unlikely that a single mutation in TRESK is sufficient to cause migraine with aura, this does not exclude a role for TRESK in migraine pathophysiology. Based on their mode of action TREK1 and TREK2 might qualify as potentially interesting therapeutic targets for migraine.

#### Functional consequences of gene mutations causing migraine with aura syndromes

To unravel the pathophysiological mechanisms causing hemiplegic migraine, functional consequences of gene mutations have been studied in cellular and animal models (for detailed reviews see [[78](#page-10-0)–[80](#page-10-0)]). First clues came from studies in cellular models in which mutant cDNAs were overexpressed. FHM1 mutations in the  $\alpha$ 1 subunit of voltage-gated  $Ca<sub>V</sub>2.1$  (P/Q-type) calcium channels result in a gain of channel function [[81\]](#page-10-0). FHM2

mutations in the  $\alpha$ 2 subunit of sodium-potassium ATPases result in a loss of function in glial cells [\[82](#page-10-0)]. The functional consequences of FHM3 mutations on  $Na<sub>V</sub>1.1$  channels were at first difficult to assess but recent evidence suggest that they are associated with a gain of channel functioning [[83](#page-10-0)]. Together these findings predict increased neurotransmitter and potassium ion levels in vivo at the synaptic cleft, which facilitates CSD and thereby can explain the aura features in patients with the mutations [[82](#page-10-0)–[87](#page-10-0)].

For all three FHM types knock-in mouse models have been generated by expressing pathogenic human gene mutations in the respective endogenous mouse gene. FHM1 mutant mice with either the R192Q or the S218L mutation reveal increased neuronal calcium influx and enhanced (cortical) excitatory, but not inhibitory, neurotransmitter release [[88](#page-10-0)–[91\]](#page-10-0). FHM2 mutant mice with either the W887R or the G301R mutation showed abnormal glutamate uptake through glial cells [[92](#page-10-0)–[94](#page-10-0)]. For the W887R mutant a reduced rate of glutamate and  $K^+$ clearance, and a reduced density of GLT-1a glutamate transporters in astrocytic processes surrounding glutamatergic synapses, was shown which facilitated the ignition of experimentally induced CSD [\[92](#page-10-0), [93\]](#page-10-0). In line with these findings, both the FHM1 and FHM2 mutant mice also demonstrate an increased susceptibility for experimentally induced CSD [\[90](#page-10-0), [92,](#page-10-0) [95](#page-10-0), [96\]](#page-10-0). Recently, the first FHM3 mouse model (expressing the L263V mutation) was generated that demonstrated, for the first time, spontaneous CSD events [\[97](#page-10-0)]. Of note, the waves of SD consistently propagated from visual to motor cortex, in line with clinical observations in humans.

To lesser extent, it has been investigated how gene mutations of the non-hemiplegic migraine genes (NOTCH3, TREX1, APP, COL4A1, CSNK1D) may cause migraine pathophysiology. Notch3 protein plays an important role in vascular smooth muscle cells of the small blood vessels of the brain [\[41\]](#page-9-0). Notch3 knockout mice and mice that overexpress the R90C mutation specifically in vascular smooth muscle cells were found to have an enhanced susceptibility to experimentally induced SD [[51](#page-9-0)]. In mutant mice that express the FASPS CKIδ-T44A mutation in CSNK1D, which encodes CK1D a well-known regulator of circadian rhythms, the susceptibility is also enhanced [[57](#page-9-0)]. It was hypothesized that migraine with aura in FASPS is caused by vascular dysfunction through abnormal astrocytic signaling. There have been no studies yet that investigated the susceptibility of CSD in mutant mice with TREX1, APP or COL4A1 mutations. Regardless, there are indirect clues how mutations in these genes may cause a migraine phenotype. RVCL-S mutations in TREX1, the major 3′-5'exonuclease in mammals, were shown to affect the regulation of oligosaccharyltransferase activity [[98](#page-10-0)], but not the protein's exonuclease function [\[44](#page-9-0), [98\]](#page-10-0). Recently, homozygous knock-in mice that express the RVCL-S V235fs mutation were shown to demonstrate signs of abnormal vascular function and increased sensitivity to experimentally induced stroke [\[52](#page-9-0)]. Of note, vasculopathy and endothelial dysfunction have also been demonstrated in vivo in patients with RVCL-S [\[53\]](#page-9-0). Amyloid precursor protein (APP) also is implicated in regulating endothelial function [\[54\]](#page-9-0), in addition to synapse formation [\[55](#page-9-0)] and neuroplasticity [\[56\]](#page-9-0). In APPDutch mice (overexpressing the E693Q APP mutation) neuronal expression of APP-Dutch leads to cerebrovascular amyloidosis, hemorrhage and smooth muscle cell degeneration [\[54\]](#page-9-0), supportive of the idea that vasculopathy may be implicated in migraine. Finally, COL4A1, which encodes the procollagen type IV α1, a basement-membrane protein, causes a pleiotropy of vascular disorders. Mutant mice (expressing the *Col4a1*<sup>Δex40</sup> mutation) develop a compromised cerebral vasculature (intracellular accumulation of mutant collagen in vascular endothelial cells and pericytes triggers small vessel disease) leading to recurrent hemorrhagic strokes and agerelated macroangiopathy [\[50,](#page-9-0) [99](#page-10-0)]. Based on the investigations in the mutant mice, it is well possible that vascular changes, such as endothelial dysfunction, may directly or via neuronal pathways increase migraine susceptibility, although this needs to be further investigated.

#### Are hemiplegic migraine and the common forms of migraine part of one genetic spectrum?

Recently, the close link between hemiplegic migraine (in those that do not have a causal mutation in the known genes) and migraine with aura was further demonstrated in a large Finnish study [\[100](#page-10-0)] that used polygenic risk scoring, based on genetic information of common variants from the most recent migraine GWAS [\[16](#page-8-0)]. It was shown that both familial migraine with aura and hemiplegic migraine cases had higher polygenic risk scores, so to larger extent are explained by the presence of common variants, compared to familial migraine without aura cases. The group of patients with hemiplegic migraine had a higher genetic load than in migraine with (typical) aura but the difference did not reach statistical significance  $(p = 0.09)$  [[100](#page-10-0)], which could be due to a smaller sample size or because sensory symptoms in some patients with migraine with aura erroneously were mistaken for motor symptoms so these patients ended up in the hemiplegic group. Notwithstanding, the study provides an independent argument that hemiplegic migraine and migraine with aura are part of the migraine spectrum.

#### Conclusion

Great effort went into unraveling the genetic architecture of migraine, both of migraine with aura and migraine without aura. Although the CGAS approach has

<span id="page-8-0"></span>been used in many (often very small) studies, findings were not replicated in a much larger GWAS data set. Of note, when taking all genotype information (not just the SNPs that surpass the threshold for genome-wide significance) into account, the genetic architecture of migraine with aura and migraine without aura is more similar than dissimilar. Until now most knowledge of the genetics of migraine with aura came from studies of monogenic syndromes that pointed to roles for neurotransmission and vasculature in migraine pathophysiology. Finally, the genetic load, i.e. the contribution of common genetic variants in families, was particularly high in hemiplegic migraine and migraine with aura compared to migraine without aura re-emphasizing the existence of a migraine spectrum. By increasing patient cohorts, by testing the contribution or rare(r) variants using recently developed SNP arrays and next generation sequencing approaches and by studying possible epigenetic contributors, it is expected that we will get a better insight of the genetic architecture of migraine, including migraine with aura.

#### Abbreviations

CADASIL: Cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy; CGAS: Candidate gene associations studies; CSD: Cortical spreading depolarization; D-CAA: Dutch-type hereditary cerebral amyloid angiopathy; GWAS: Genome-wide association studies; FHM: Familial hemiplegic migraine; FASPS: Familial advanced sleep phase syndrome; MELAS: Mitochondrial encephalopathy, lactic acidosis and strokelike episodes; mtDNA: mitochondrial DNA; NGS: Next generation sequencing; RVCL-S: Retinal vasculopathy with cerebral leukoencephalopathy and systemic manifestations; SHM: Sporadic hemiplegic migraine; SNP: Singlenucleotide polymorphisms

#### Acknowledgements

Not applicable.

#### Authors' contributions

All authors were involved in conceptualization, writing and revising the manuscript. All authors read and approved the final manuscript.

#### Funding

The article-processing charges for the article has been sponsored by the European Headache Federation.

#### Availability of data and materials

Not applicable.

Ethics approval and consent to participate

Not applicable.

#### Consent for publication

Not applicable.

#### Competing interests

The authors declare that they have no competing interests.

#### Received: 15 April 2020 Accepted: 19 May 2020 Published online: 05 June 2020

#### References

- 1. Classification Committee of The International Headache Society (IHS) (2018) The international classification of headache disorders, 3rd edition. Cephalalgia 38(1):1–211
- Lauritzen M (1994) Pathophysiology of the migraine aura. The spreading depression theory. Brain 117(Pt 1):199–210
- 3. Hadjikhani N, Sanchez Del Rio M, Wu O, Schwartz D, Bakker D, Fischl B et al (2001) Mechanisms of migraine aura revealed by functional MRI in human visual cortex. Proc Natl Acad Sci U S A 98(8):4687–4692
- 4. Gorji A, Scheller D, Straub H, Tegtmeier F, Kohling R, Hohling JM et al (2001) Spreading depression in human neocortical slices. Brain Res 906(1–2):74–83
- 5. Russell MB, Iselius L, Olesen J (1996) Migraine without aura and migraine with aura are inherited disorders. Cephalalgia 16(5):305–309
- 6. Ulrich V, Gervil M, Kyvik KO, Olesen J, Russell MB (1999) Evidence of a genetic factor in migraine with aura: a population-based Danish twin study. Ann Neurol 45(2):242–246
- 7. Gervil M, Ulrich V, Kyvik KO, Olesen J, Russell MB (1999) Migraine without aura: a population-based twin study. Ann Neurol 46(4):606–611
- 8. Pelzer N, Louter MA, van Zwet EW, Nyholt DR, Ferrari MD, van den Maagdenberg AM et al (2019) Linking migraine frequency with family history of migraine. Cephalalgia. 39(2):229–236
- 9. Gasparini CF, Sutherland HG, Griffiths LR (2013) Studies on the pathophysiology and genetic basis of migraine. Curr Genomics 14(5):300–315
- 10. Sutherland HG, Albury CL, Griffiths LR (2019) Advances in genetics of migraine. J Headache Pain. 20(1):72
- 11. Nyholt DR, LaForge KS, Kallela M, Alakurtti K, Anttila V, Farkkila M et al (2008) A high-density association screen of 155 ion transport genes for involvement with common migraine. Hum Mol Genet 17(21):3318–3331
- 12. de Vries B, Anttila V, Freilinger T, Wessman M, Kaunisto MA, Kallela M et al (2016) Systematic re-evaluation of genes from candidate gene association studies in migraine using a large genome-wide association data set. Cephalalgia 36(7):604–614
- 13. van Oosterhout WP, Weller CM, Stam AH, Bakels F, Stijnen T, Ferrari MD et al (2011) Validation of the web-based LUMINA questionnaire for recruiting large cohorts of migraineurs. Cephalalgia. 31(13):1359–1367
- 14. Hagen K, Stovner LJ, Zwart JA (2007) Potentials and pitfalls in analytical headache epidemiological studies--lessons to be learned from the head-HUNT study. Cephalalgia. 27(5):403–413
- 15. Anttila V, Stefansson H, Kallela M, Todt U, Terwindt GM, Calafato MS et al (2010) Genome-wide association study of migraine implicates a common susceptibility variant on 8q22.1. Nat Genet 42(10):869–873
- 16. Gormley P, Anttila V, Winsvold BS, Palta P, Esko T, Pers TH et al (2016) Metaanalysis of 375,000 individuals identifies 38 susceptibility loci for migraine. Nat Genet 48(8):856–866
- 17. Anttila V, Winsvold BS, Gormley P, Kurth T, Bettella F, McMahon G et al (2013) Genome-wide meta-analysis identifies new susceptibility loci for migraine. Nat Genet 45(8):912–917
- 18. Nyholt DR, Borsook D, Griffiths LR (2017) Migrainomics identifying brain and genetic markers of migraine. Nat Rev Neurol 13(12):725–741
- 19. Launer LJ, Terwindt GM, Ferrari MD (1999) The prevalence and characteristics of migraine in a population-based cohort: the GEM study. Neurology. 53(3):537–542
- 20. Pisanu C, Preisig M, Castelao E, Glaus J, Pistis G, Squassina A et al (2017) A genetic risk score is differentially associated with migraine with and without aura. Hum Genet 136(8):999–1008
- 21. Zhao H, Eising E, de Vries B, Vijfhuizen LS, Anttila V, Winsvold BS et al (2016) Gene-based pleiotropy across migraine with aura and migraine without aura patient groups. Cephalalgia 36(7):648–657
- 22. Nyholt DR, Anttila V, Winsvold BS, Kurth T, Stefansson H, Kallela M et al (2015) Concordance of genetic risk across migraine subgroups: impact on current and future genetic association studies. Cephalalgia 35(6):489–499
- 23. Anttila V, Bulik-Sullivan B, Finucane HK, Walters RK, Bras J, Duncan L et al (2018) Analysis of shared heritability in common disorders of the brain. Science (New York, NY) 360(6395):eaap8757
- 24. Tam V, Patel N, Turcotte M, Bosse Y, Pare G, Meyre D (2019) Benefits and limitations of genome-wide association studies. Nat Rev Genet 20(8):467–484
- 25. Wallace DC (2018) Mitochondrial genetic medicine. Nat Genet 50(12):1642–1649
- 26. Vollono C, Primiano G, Della Marca G, Losurdo A, Servidei S (2018) Migraine in mitochondrial disorders: prevalence and characteristics. Cephalalgia. 38(6): 1093–1106
- 27. Stuart S, Griffiths LR (2012) A possible role for mitochondrial dysfunction in migraine. Mol Gen Genomics 287(11–12):837–844
- 28. Zaki EA, Freilinger T, Klopstock T, Baldwin EE, Heisner KR, Adams K et al (2009) Two common mitochondrial DNA polymorphisms are highly associated with migraine headache and cyclic vomiting syndrome. Cephalalgia. 29(7):719–728
- <span id="page-9-0"></span>29. Borte S, Zwart JA, Skogholt AH, Gabrielsen ME, Thomas LF, Fritsche LG et al (2020) Mitochondrial genome-wide association study of migraine - the HUNT study. Cephalalgia. 40(6):625–634
- 30. Qureshi IA, Mehler MF (2018) Epigenetic mechanisms underlying nervous system diseases. Handb Clin Neurol 147:43–58
- 31. Eising E, Datson NA, van den Maagdenberg AM, Ferrari MD (2013) Epigenetic mechanisms in migraine: a promising avenue? BMC Med 11:26
- 32. Gerring ZF, McRae AF, Montgomery GW, Nyholt DR (2018) Genome-wide DNA methylation profiling in whole blood reveals epigenetic signatures associated with migraine. BMC Genomics 19(1):69
- 33. Fogel BL (2018) Genetic and genomic testing for neurologic disease in clinical practice. Handb Clin Neurol 147:11–22
- 34. Hiekkala ME, Vuola P, Artto V, Happola P, Happola E, Vepsalainen S et al (2018) The contribution of CACNA1A, ATP1A2 and SCN1A mutations in hemiplegic migraine: a clinical and genetic study in Finnish migraine families. Cephalalgia. 38(12):1849–1863
- 35. Pelzer N, Stam AH, Haan J, Ferrari MD, Terwindt GM (2013) Familial and sporadic hemiplegic migraine: diagnosis and treatment. Curr Treat Options Neurol 15(1):13–27
- 36. Thomsen LL, Olesen J, Russell MB (2003) Increased risk of migraine with typical aura in probands with familial hemiplegic migraine and their relatives. Eur J Neurol 10(4):421–427
- 37. Hansen JM, Hauge AW, Ashina M, Olesen J (2011) Trigger factors for familial hemiplegic migraine. Cephalalgia. 31(12):1274–1281
- Pelzer N, Haan J, Stam AH, Vijfhuizen LS, Koelewijn SC, Smagge A et al (2018) Clinical spectrum of hemiplegic migraine and chances of finding a pathogenic mutation. Neurology. 90(7):e575–ee82
- 39. Guey S, Mawet J, Herve D, Duering M, Godin O, Jouvent E et al (2016) Prevalence and characteristics of migraine in CADASIL. Cephalalgia. 36(11): 1038–1047
- 40. Domenga V, Fardoux P, Lacombe P, Monet M, Maciazek J, Krebs LT et al (2004) Notch3 is required for arterial identity and maturation of vascular smooth muscle cells. Genes Dev 18(22):2730–2735
- 41. Joutel A, Corpechot C, Ducros A, Vahedi K, Chabriat H, Mouton P et al (1996) Notch3 mutations in CADASIL, a hereditary adult-onset condition causing stroke and dementia. Nature. 383(6602):707–710
- 42. Stam AH, Kothari PH, Shaikh A, Gschwendter A, Jen JC, Hodgkinson S et al (2016) Retinal vasculopathy with cerebral leukoencephalopathy and systemic manifestations. Brain. 139(11):2909–2922
- 43. Pelzer N, Hoogeveen ES, Haan J, Bunnik R, Poot CC, van Zwet EW et al (2019) Systemic features of retinal vasculopathy with cerebral leukoencephalopathy and systemic manifestations: a monogenic small vessel disease. J Intern Med 285(3):317–332
- 44. Richards A, van den Maagdenberg AM, Jen JC, Kavanagh D, Bertram P, Spitzer D et al (2007) C-terminal truncations in human 3′-5′ DNA exonuclease TREX1 cause autosomal dominant retinal vasculopathy with cerebral leukodystrophy. Nat Genet 39(9):1068–1070
- 45. Stewart WF, Wood C, Reed ML, Roy J, Lipton RB (2008) Cumulative lifetime migraine incidence in women and men. Cephalalgia. 28(11):1170–1178
- 46. Koemans EA, Voigt S, Rasing I, van Etten ES, van Zwet EW, van Walderveen MAA et al (2020) Migraine With Aura as Early Disease Marker in Hereditary Dutch-Type Cerebral Amyloid Angiopathy. Stroke 51(4):1094–1099
- 47. Wattendorff AR, Frangione B, Luyendijk W, Bots GT (1995) Hereditary cerebral haemorrhage with amyloidosis, Dutch type (HCHWA-D): clinicopathological studies. J Neurol Neurosurg Psychiatry 58(6):699–705
- 48. Breedveld G, de Coo IF, Lequin MH, Arts WF, Heutink P, Gould DB et al (2006) Novel mutations in three families confirm a major role of COL4A1 in hereditary porencephaly. J Med Genet 43(6):490–495
- 49. Lanfranconi S, Markus HS (2010) COL4A1 mutations as a monogenic cause of cerebral small vessel disease: a systematic review. Stroke. 41(8):e513–e518
- 50. Gould DB, Phalan FC, van Mil SE, Sundberg JP, Vahedi K, Massin P et al (2006) Role of COL4A1 in small-vessel disease and hemorrhagic stroke. N Engl J Med 354(14):1489–1496
- 51. Eikermann-Haerter K, Yuzawa I, Dilekoz E, Joutel A, Moskowitz MA, Ayata C (2011) Cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy syndrome mutations increase susceptibility to spreading depression. Ann Neurol 69(2):413–418
- 52. Mulder IA, Rubio-Beltran E, Ibrahimi K, Dzyubachyk O, Khmelinskii A, Hoehn M et al (2020) Increased mortality and vascular phenotype in a Knock-in mouse model of retinal vasculopathy with cerebral Leukoencephalopathy and systemic manifestations. Stroke. 51(1):300–307
- 53. de Boer I, Stam AH, Buntinx L, Zielman R, van dS I, van den Maagdenberg AMJM et al (2018) RVCL-S and CADASIL display distinct impaired vascular function. Neurology. 91(10):e956–ee63
- 54. Herzig MC, Winkler DT, Burgermeister P, Pfeifer M, Kohler E, Schmidt SD et al (2004) Abeta is targeted to the vasculature in a mouse model of hereditary cerebral hemorrhage with amyloidosis. Nat Neurosci 7(9): 954–960
- 55. Priller C, Bauer T, Mitteregger G, Krebs B, Kretzschmar HA, Herms J (2006) Synapse formation and function is modulated by the amyloid precursor protein. J Neurosci 26(27):7212–7221
- 56. Turner PR, O'Connor K, Tate WP, Abraham WC (2003) Roles of amyloid precursor protein and its fragments in regulating neural activity, plasticity and memory. Prog Neurobiol 70(1):1–32
- 57. Brennan KC, Bates EA, Shapiro RE, Zyuzin J, Hallows WC, Huang Y et al (2013) Casein kinase idelta mutations in familial migraine and advanced sleep phase. Sci Transl Med 5(183):183ra56 1–11
- 58. Xu Y, Padiath QS, Shapiro RE, Jones CR, Wu SC, Saigoh N et al (2005) Functional consequences of a CKIdelta mutation causing familial advanced sleep phase syndrome. Nature. 434(7033):640–644
- 59. Jones CR, Campbell SS, Zone SE, Cooper F, DeSano A, Murphy PJ et al (1999) Familial advanced sleep-phase syndrome: a short-period circadian rhythm variant in humans. Nat Med 5(9):1062–1065
- 60. Toh KL, Jones CR, He Y, Eide EJ, Hinz WA, Virshup DM et al (2001) An hPer2 phosphorylation site mutation in familial advanced sleep phase syndrome. Science (New York, NY) 291(5506):1040–1043
- 61. Suzuki M, Van Paesschen W, Stalmans I, Horita S, Yamada H, Bergmans BA et al (2010) Defective membrane expression of the Na(+)-HCO (3)(−) cotransporter NBCe1 is associated with familial migraine. Proc Natl Acad Sci U S A 107(36):15963–15968
- 62. Jen JC, Wan J, Palos TP, Howard BD, Baloh RW (2005) Mutation in the glutamate transporter EAAT1 causes episodic ataxia, hemiplegia, and seizures. Neurology. 65(4):529–534
- 63. Marini C, Conti V, Mei D, Battaglia D, Lettori D, Losito E et al (2012) PRRT2 mutations in familial infantile seizures, paroxysmal dyskinesia, and hemiplegic migraine. Neurology. 79(21):2109–2114
- 64. Pelzer N, de Vries B, Kamphorst JT, Vijfhuizen LS, Ferrari MD, Haan J et al (2014) PRRT2 and hemiplegic migraine: a complex association. Neurology. 83(3):288–290
- 65. Kovermann P, Hessel M, Kortzak D, Jen JC, Koch J, Fahlke C et al (2017) Impaired K(+) binding to glial glutamate transporter EAAT1 in migraine. Sci Rep 7(1):13913
- 66. Meneret A, Gaudebout C, Riant F, Vidailhet M, Depienne C, Roze E (2013) PRRT2 mutations and paroxysmal disorders. Eur J Neurol 20(6):872–878
- 67. Heron SE, Dibbens LM (2013) Role of PRRT2 in common paroxysmal neurological disorders: a gene with remarkable pleiotropy. J Med Genet 50(3):133–139
- 68. Mo J, Wang B, Zhu X, Wu X, Liu Y (2019) PRRT2 deficiency induces paroxysmal kinesigenic dyskinesia by influencing synaptic function in the primary motor cortex of rats. Neurobiol Dis 121:274–285
- 69. Karczewski KJ, Francioli LC, Tiao G, Cummings BB, Alföldi J, Wang Q et al (2019) Variation across 141,456 human exomes and genomes reveals the spectrum of loss-of-function intolerance across human protein-coding genes. bioRxiv 531210
- 70. Lafreniere RG, Cader MZ, Poulin JF, Andres-Enguix I, Simoneau M, Gupta N et al (2010) A dominant-negative mutation in the TRESK potassium channel is linked to familial migraine with aura. Nat Med 16(10):1157–1160
- 71. Lafreniere RG, Rouleau GA (2011) Migraine: role of the TRESK two-pore potassium channel. Int J Biochem Cell Biol 43(11):1533–1536
- 72. Liu P, Xiao Z, Ren F, Guo Z, Chen Z, Zhao H et al (2013) Functional analysis of a migraine-associated TRESK K+ channel mutation. J Neurosci 33(31): 12810–12824
- 73. Guo Z, Liu P, Ren F, Cao YQ (2014) Nonmigraine-associated TRESK K+ channel variant C110R does not increase the excitability of trigeminal ganglion neurons. J Neurophysiol 112(3):568–579
- Andres-Enguix I, Shang L, Stansfeld PJ, Morahan JM, Sansom MS, Lafreniere RG et al (2012) Functional analysis of missense variants in the TRESK (KCNK18) K channel. Sci Rep 2:237
- 75. Royal P, Andres-Bilbe A, Avalos Prado P, Verkest C, Wdziekonski B, Schaub S et al (2019) Migraine-Associated TRESK Mutations Increase Neuronal Excitability through Alternative Translation Initiation and Inhibition of TREK. Neuron 101(2):232–45.e6
- <span id="page-10-0"></span>76. Pettingill P, Weir GA, Wei T, Wu Y, Flower G, Lalic T et al (2019) A causal role for TRESK loss of function in migraine mechanisms. Brain. 142(12):3852–3867
- 77. Landrum MJ, Lee JM, Benson M, Brown G, Chao C, Chitipiralla S et al (2016) ClinVar: public archive of interpretations of clinically relevant variants. Nucleic Acids Res 44(D1):D862–D868
- 78. Pietrobon D, Brennan KC (2019) Genetic mouse models of migraine. J Headache pain 20(1):79
- 79. Chen SP, Tolner EA, Eikermann-Haerter K (2016) Animal models of monogenic migraine. Cephalalgia. 36(7):704–721
- 80. Ferrari MD, Klever RR, Terwindt GM, Ayata C, van den Maagdenberg AM (2015) Migraine pathophysiology: lessons from mouse models and human genetics. Lancet Neurol 14(1):65–80
- 81. Pietrobon D (2013) Calcium channels and migraine. Biochim Biophys Acta 1828(7):1655–1665
- 82. Tavraz NN, Friedrich T, Durr KL, Koenderink JB, Bamberg E, Freilinger T et al (2008) Diverse functional consequences of mutations in the Na+/K+-ATPase alpha2-subunit causing familial hemiplegic migraine type 2. J Biol Chem 283(45):31097–31106
- 83. Cestele S, Schiavon E, Rusconi R, Franceschetti S, Mantegazza M (2013) Nonfunctional NaV1.1 familial hemiplegic migraine mutant transformed into gain of function by partial rescue of folding defects. Proc Natl Acad Sci U S A 110(43):17546–17551
- 84. Garza-Lopez E, Sandoval A, Gonzalez-Ramirez R, Gandini MA, Van den Maagdenberg A, De Waard M et al (2012) Familial hemiplegic migraine type 1 mutations W1684R and V1696I alter G protein-mediated regulation of Ca(V)2.1 voltage-gated calcium channels. Biochim Biophys Acta 1822(8): 1238–1246
- 85. Serra SA, Fernandez-Castillo N, Macaya A, Cormand B, Valverde MA, Fernandez-Fernandez JM (2009) The hemiplegic migraine-associated Y1245C mutation in CACNA1A results in a gain of channel function due to its effect on the voltage sensor and G-protein-mediated inhibition. Pflugers Archiv 458(3):489–502
- 86. Weiss N, Sandoval A, Felix R, Van den Maagdenberg A, De Waard M (2008) The S218L familial hemiplegic migraine mutation promotes deinhibition of Ca(v)2.1 calcium channels during direct G-protein regulation. Pflugers Archiv 457(2):315–326
- 87. Cestele S, Scalmani P, Rusconi R, Terragni B, Franceschetti S, Mantegazza M (2008) Self-limited hyperexcitability: functional effect of a familial hemiplegic migraine mutation of the Nav1.1 (SCN1A) Na+ channel. J Neurosci 28(29): 7273–7283
- 88. Eikermann-Haerter K, Dilekoz E, Kudo C, Savitz SI, Waeber C, Baum MJ et al (2009) Genetic and hormonal factors modulate spreading depression and transient hemiparesis in mouse models of familial hemiplegic migraine type 1. J Clin Invest 119(1):99–109
- 89. Tottene A, Conti R, Fabbro A, Vecchia D, Shapovalova M, Santello M et al (2009) Enhanced excitatory transmission at cortical synapses as the basis for facilitated spreading depression in Ca(v)2.1 knockin migraine mice. Neuron. 61(5):762–773
- 90. van den Maagdenberg AM, Pietrobon D, Pizzorusso T, Kaja S, Broos LA, Cesetti T et al (2004) A Cacna1a knockin migraine mouse model with increased susceptibility to cortical spreading depression. Neuron. 41(5):701–710
- 91. Vecchia D, Tottene A, van den Maagdenberg AM, Pietrobon D (2014) Mechanism underlying unaltered cortical inhibitory synaptic transmission in contrast with enhanced excitatory transmission in CaV2.1 knockin migraine mice. Neurobiol Dis 69:225–234
- 92. Leo L, Gherardini L, Barone V, De Fusco M, Pietrobon D, Pizzorusso T et al (2011) Increased susceptibility to cortical spreading depression in the mouse model of familial hemiplegic migraine type 2. PLoS Genet 7(6): e1002129
- 93. Capuani C, Melone M, Tottene A, Bragina L, Crivellaro G, Santello M et al (2016) Defective glutamate and K+ clearance by cortical astrocytes in familial hemiplegic migraine type 2. EMBO Mol Med 8(8):967–986
- 94. Bottger P, Glerup S, Gesslein B, Illarionova NB, Isaksen TJ, Heuck A et al (2016) Glutamate-system defects behind psychiatric manifestations in a familial hemiplegic migraine type 2 disease-mutation mouse model. Sci Rep 6:22047
- 95. Unekawa M, Ikeda K, Tomita Y, Kawakami K, Suzuki N (2018) Enhanced susceptibility to cortical spreading depression in two types of Na+,K+- ATPase α2 subunit-deficient mice as a model of familial hemiplegic migraine 2. Cephalalgia 38(9):1515–1524
- 96. van den Maagdenberg AM, Pizzorusso T, Kaja S, Terpolilli N, Shapovalova M, Hoebeek FE et al (2010) High cortical spreading depression susceptibility and migraine-associated symptoms in Ca(v)2.1 S218L mice. Ann Neurol 67(1):85–98
- 97. Jansen NA, Dehghani A, Linssen MML, Breukel C, Tolner EA, van den Maagdenberg A (2020) First FHM3 mouse model shows spontaneous cortical spreading depolarizations. Ann Clin Transl Neurol 7(1):132–138
- 98. Hasan M, Fermaintt CS, Gao N, Sakai T, Miyazaki T, Jiang S et al (2015) Cytosolic nuclease TREX1 regulates Oligosaccharyltransferase activity independent of nuclease activity to suppress immune activation. Immunity. 43(3):463–474
- 99. Jeanne M, Labelle-Dumais C, Jorgensen J, Kauffman WB, Mancini GM, Favor J et al (2012) COL4A2 mutations impair COL4A1 and COL4A2 secretion and cause hemorrhagic stroke. Am J Hum Genet 90(1):91–101
- 100. Gormley P, Kurki MI, Hiekkala ME, Veerapen K, Happola P, Mitchell AA et al (2018) Common Variant Burden Contributes to the Familial Aggregation of Migraine in 1,589 Families. Neuron 98(4):743–53.e4

#### Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

#### Ready to submit your research? Choose BMC and benefit from:

- · fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- · gold Open Access which fosters wider collaboration and increased citations
- · maximum visibility for your research: over 100M website views per year

#### At BMC, research is always in progress.

Learn more biomedcentral com/submissions

