



Universiteit
Leiden
The Netherlands

Oxidative stress in chronic diseases: causal inference from observational studies

Luo, J.

Citation

Luo, J. (2022, September 1). *Oxidative stress in chronic diseases: causal inference from observational studies*. Retrieved from <https://hdl.handle.net/1887/3454705>

Version: Publisher's Version

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/3454705>

Note: To cite this publication please use the final published version (if applicable).

CHAPTER 2

Ageing, age-related diseases, and oxidative stress: what to do next?

Jiao Luo, Kevin Mills, Saskia le Cessie, Raymond Noordam, Diana van Heemst

Ageing Res Rev. 2020 Jan; 57:100982.

Abstract

Among other mechanisms, oxidative stress has been postulated to play an important role in the rate of ageing. Oxidative damage contributes to the hallmarks of ageing and essential components in pathological pathways which are thought to drive multiple age-related diseases. Nonetheless, results from studies testing the hypothesis of oxidative stress in ageing and diseases showed controversial results. While observational studies mainly found detrimental effects of high oxidative stress levels on disease status, randomized clinical trials examining the effect of antioxidant supplementation on disease status generally showed null effects. However, re-evaluations of these counterintuitive observations are required considering the lack of reliability and specificity of traditionally used biomarkers for measuring oxidative stress. To facilitate these re-evaluations, this review summarizes the basic knowledge of oxidative stress and the present findings regarding oxidative damage and ageing and age-related diseases. Meanwhile, two approaches are highlighted, namely proper participants selection, together with the development of reliable biomarkers. We propose that oxidized vitamin E metabolites may be used to accurately monitor individual functional antioxidant levels, which might serve as promising key solutions for future elucidating the impact of oxidative stress on ageing and age-related diseases.

Introduction

It has been widely acknowledged that life expectancy has increased over the past centuries as a specific result of improved medical care, vaccination, and hygiene^{1,2}. The process of ageing is a dynamic, chronological process characterized by the gradual accumulation of damage to cells, progressive functional decline, and increased susceptibility and vulnerability to diseases. In addition, ageing is closely connected to the onset and progression of multiple age-related diseases, such as cancer, type 2 diabetes mellitus, and cardiovascular and neurodegenerative diseases³⁻⁵. The ageing process is postulated to originate from several basic molecular changes, better known as the hallmarks of ageing, which include four primary hallmarks, genomic instability, telomere attrition, epigenetic alterations, and loss of proteostasis, three antagonistic hallmarks, deregulated nutrient sensing, mitochondrial dysfunction, and cellular senescence, and two integrative hallmarks stem cell exhaustion, and altered intercellular communication⁶.

These hallmarks contributing to the ageing process could be caused by oxidative damage. For example, telomeres are highly sensitive to oxidative damage and their repair capacity is less well than other parts of the chromosome^{7,8}. Hence, oxidative damage may result in telomere attrition that accelerates ageing and increases the risk of age-related diseases⁹. The concept of oxidative stress was introduced in 1985 and updated later¹⁰⁻¹². Oxidative stress refers to “an imbalance between the generation of oxidants and their elimination systems, i.e. antioxidants, in favor of oxidants, leading to disruption of redox signaling and control and/or molecular damage”¹². Conceptually, the level of oxidative stress ranges from physiological levels for redox signaling to toxic levels of molecular or organelle damage (**Figure 1**). Redox signaling is essential for host defense as well as in a diverse array of signaling pathways^{13,14}. Other damages caused by non-physiological high oxidative stress lead to a wide range of phenotypic changes, including altered gene expression, arrested cell proliferation, cell growth, and cellular senescence¹⁵⁻¹⁷.

Antioxidants may act as scavengers of oxidants to maintain the biological redox steady states. Therefore, since the oxidative stress theory was proposed¹⁸, antioxidants were postulated to potentially play a protective role in ageing and age-related diseases. Considering the premise that adverse health consequences caused by oxidative stress can be counteracted by antioxidants, a comprehensive body of studies aiming to examine the beneficial effects of antioxidants on diseases have been carried out in the past three decades. However, results were often disappointing and counterintuitive. The most appealing and well-known example is vitamin E, a well elucidated chain-breaking antioxidant. Although lower disease risks in individuals with higher vitamin E concentration have been found in many observational studies¹⁹⁻²⁴, as well as protective properties of vitamin E in animal experiments^{25,26}, most clinical trials examining vitamin E supplementation failed to demonstrate any advantageous effects on the prevention or treatment of various age-related diseases²⁷⁻³².

Along with these conflicting evidence, it seems like the controversy about the oxidative stress theory in ageing and age-related diseases has never stopped (**Figure 2**). In addition, over the past 30 years, fluctuations in the use of antiox-

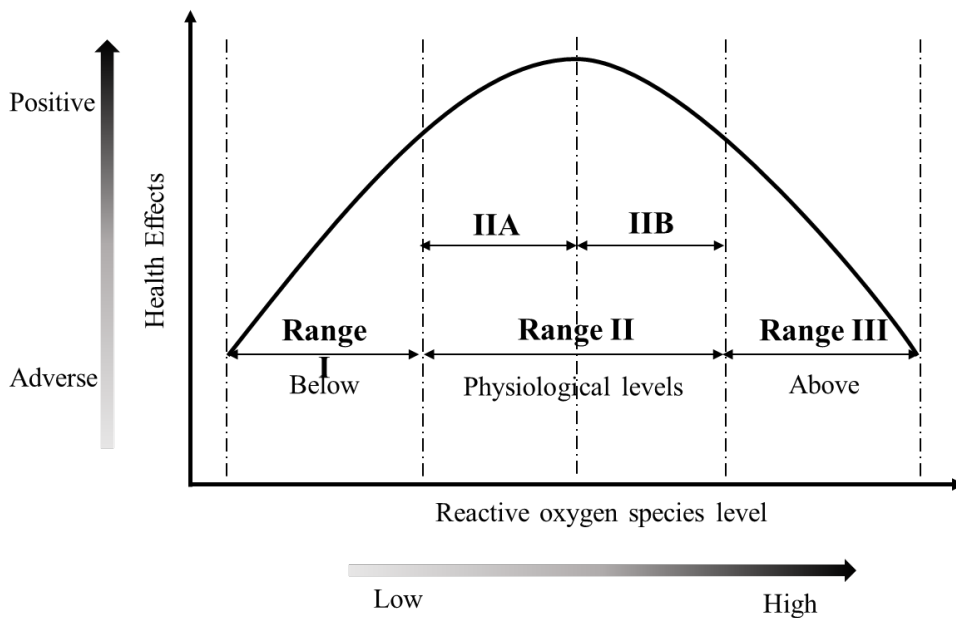


Figure 1 ROS levels and health effects: a dose-response model

Detailed discussion in the texts in part 2.2.

idant supplements were also observed, for example in the US (**Figure 3**)^{33,34}. The percentage of individuals using antioxidant supplements gradually increased from the 1980s and peaked in the 1990s. Of note, specifically, the use of vitamin E supplements steeply dropped in the early 21st century, where after decline turned to be stabilized. However, these observations neither imply that any consensus about the effect of antioxidants on diseases has been reached, nor that the oxidative stress theory has been refuted. Conversely, the annual publication count of antioxidant articles steadily increased since the 1990s, and more than 30,000 papers have been published in 2018 alone about this research topic³⁵.

So far, oxidative damages are thought to play a pivotal role in the pathological processes implicated in ageing and age-related diseases and the underlying biochemical mechanisms have been clarified in detail^{6,36}. However, there are still several questions unsettled such as the existing paradox regarding the preventive and therapeutic role of antioxidants (such as vitamin E), the lack of stable and representative biomarkers of oxidative stress, and whether oxidative stress is causally associated with ageing and age-related disease in the general population setting. Therefore, this review is organized as such to provide an overview of the chemical processes involved in oxidative stress and an update on the available evidence about associations with ageing and age-related diseases. In the last part of the review, antioxidants, especially the controversial role of vitamin E will be addressed, together with novel insights and directions for future research.

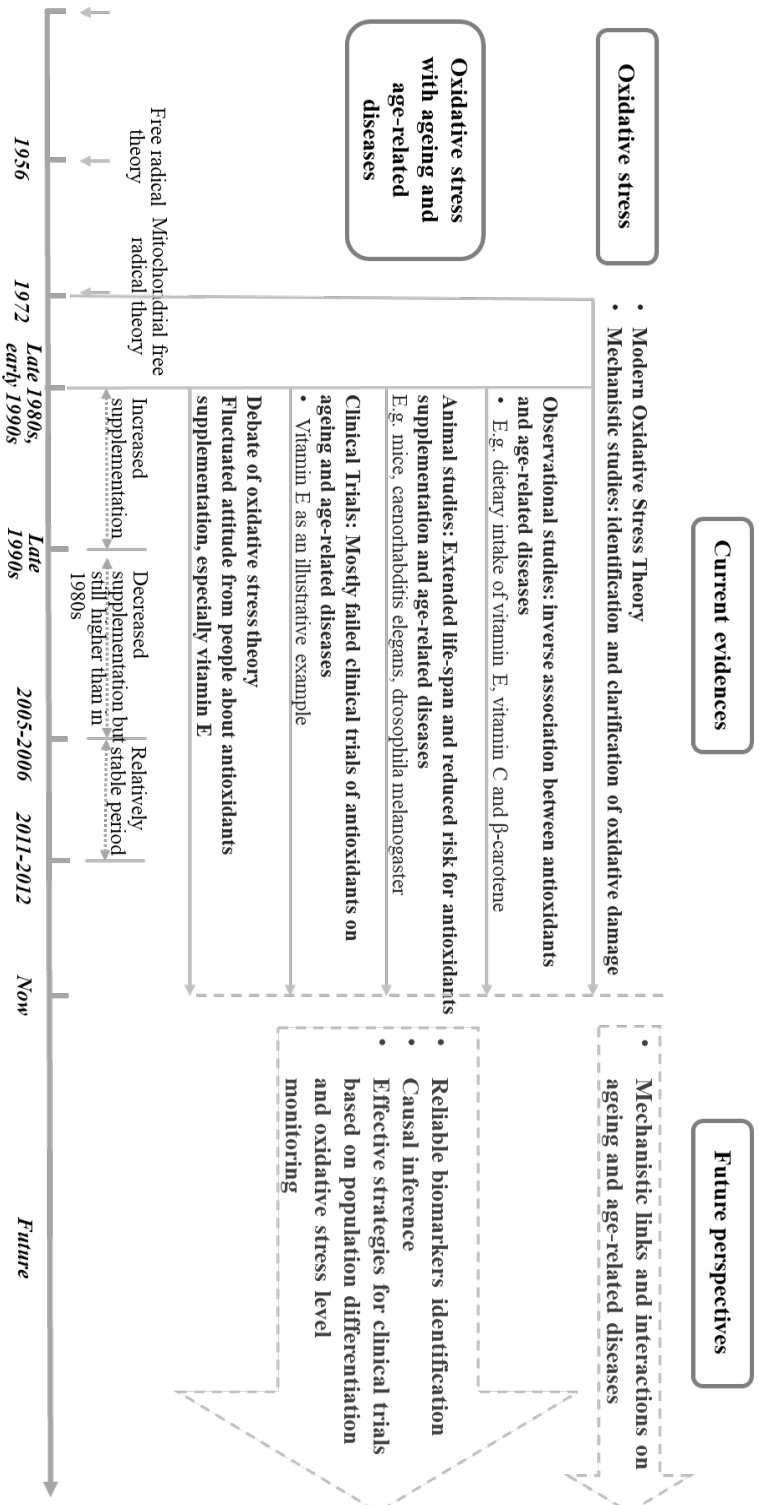


Figure 2 Schematic review of oxidative stress theory in ageing and age-related diseases

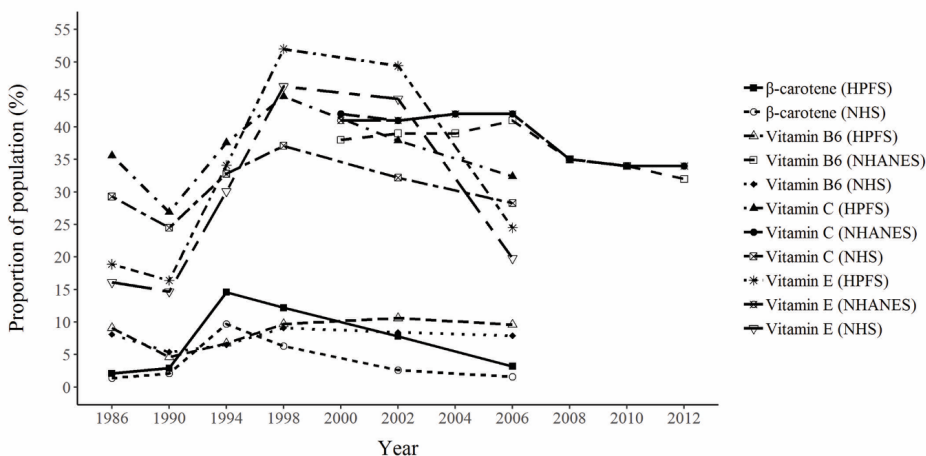


Figure 3 Trends in Antioxidants supplementation in the US

NHS: Nurses' Health Study, HPFS: Health Professionals Follow-up Study, NHANES: National Health and Nutrition Examination Survey. Data presented in the graph of NHS and HPFS are from 1986 to 2006, and data of NHANES are from 1999 to 2012.

The generation and health roles of Reactive Oxygen Species (ROS)

Endogenous generation of ROS

According to the "free radical theory" that was proposed in the 1950s and revised in the 1970s, damages induced by free radicals are the main cause of ageing and a shorter lifespan^{18,37,38}. Reactive oxygen species (ROS) are highly reactive molecules, primarily including typical free radicals that contain at least one unpaired electron (superoxide $O_2^{\cdot-}$, hydroxyl radical $\cdot OH$), and hydrogen peroxides (H_2O_2), and have been considered the main source of endogenous oxidative stress damage³⁹.

It is widely accepted that the bulk of ROS is generated by the mitochondrial electron transport chain during normal oxidative respiration in addition to numerous intracellular pathways (**Figure 4**). It is estimated that about 1-2% of the daily overall oxygen molecules consumed are reduced into $O_2^{\cdot-}$ with the leak of electrons⁴⁰. This process occurs mainly in two discrete complexes of mitochondrial electron transport chain in the matrix side of inner mitochondrial membrane, notably complex I (NADH-ubiquinone oxidoreductase) and complex III (ubiquinone-cytochrome c reductase)⁴¹. Iron-sulphur centers are thought as the most likely site of ROS production in complex I⁴². Complex III, also known as the Q cycle, which refers to a set of ubiquinone oxidation reactions, is responsible for the robust production of superoxide, the precursor of other ROS, by non-enzymatic transfer of electrons to molecular oxygen. Once generated, superoxide could be catalyzed by superoxide dismutase (SOD) into H_2O_2 , which is unstable and membrane-diffusible peroxide. Subsequently, in the presence of transition cation with reduced form (Fe^{2+} or Cu^+ , referred to as the Fenton reaction)

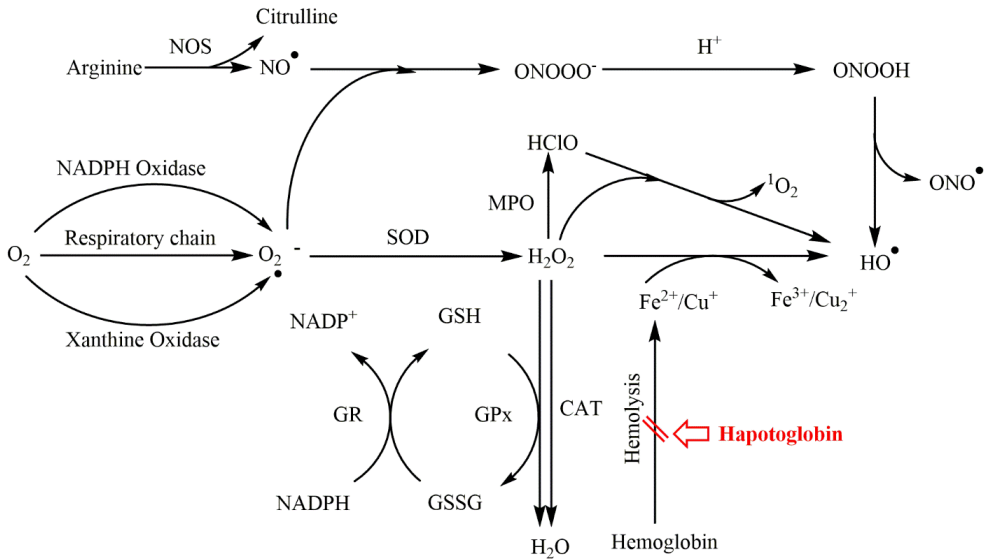


Figure 4 Reactive oxygen species generation

$O_2^{\bullet -}$: Super oxide anion, SOD: super oxidize dismutase, H_2O_2 : hydrogen peroxide, HO^{\bullet} hydroxyl radical, MPO: myeloid peroxide, $HClO$: hypochlorous acid, 1O_2 : singlet oxygen, NOS: nitric oxide synthase, NO^{\bullet} : nitric oxide radical, $ONOOO^-$: peroxyntirite, $ONOOH$: peroxyntirous acid, ONO^{\bullet} : nitrogen dioxide, GSH: glutathione, GSSG: glutathione disulfide, GPx: glutathione peroxidase, CAT: catalase, $NADP^+$: Nicotinamide adenine dinucleotide phosphate, $NADPH$: Nicotinamide adenine dinucleotide phosphate reduced form, GR: glutathione reductase.

The bulk of ROS are mainly generated by the mitochondrial electron transport chain during normal aerobic metabolism in addition to multiple ways encompassing cytosolic enzyme systems (NADPH oxidase), monoamine oxidase on the outer membrane of mitochondrial, xanthine oxidase, and uncoupled NOS. In the electron transport chain, oxygen molecules are univalent reduced into $O_2^{\bullet -}$ with the leak of electrons. The formation of superoxide is the initial step and the start of a cascade reaction of other ROS generation. Once generated, it could be catalyzed spontaneously by SOD into H_2O_2 . Subsequently in the presence of transition cation with reduced form (Fe^{2+} or Cu^+ , referred to as the Fenton reaction) or myeloid peroxide (MPO), H_2O_2 further dismutates to HO^{\bullet} . Meanwhile, H_2O_2 can also be reduced into the water by the enzymatic antioxidants such as CAT and GPx. Haptoglobin binds hemoglobin with high affinity and stability, preventing the release of heme iron from hemolysis into circulation, consequently terminating Fenton reaction and preventing the production of HO^{\bullet} .

or myeloid peroxide (MPO) ⁴³, H_2O_2 further dismutates to HO^{\bullet} , the extremely unstable and most reactive ion among all ROS. In summary, the main process of ROS generation in mitochondria could be schematically presented as $O_2 \rightarrow O_2^{\bullet -} \rightarrow H_2O_2 \rightarrow HO^{\bullet}$.

Hydroxyl radicals may lead to detrimental damages to macromolecules owing to its chemical properties ⁴⁴. Moreover, its formation relies on the presence of a reduced form transition cation, mainly iron generated from the hemoglobin heme group during hemolysis. Hence any component which can stabilize heme iron within hemoglobin could prevent oxidative damage caused by hydroxyl rad-

icals. In recent years, haptoglobin (Hp), an abundant, acute-phase inflammatory glycoprotein, which is predominantly synthesized in the liver and is regulated by cytokines, has been indicated to have an important role in the prevention of the generation of hydroxyl radicals by virtue of its ability to bind hemoglobin with high affinity and stability, thus preventing the release of heme iron from hemolysis into the circulation ⁴⁵, as shown in **Figure 4**. The Hp gene basically contains two common alleles, namely Hp1 and Hp2, with homozygous (1-1 or 2-2) and heterozygous (2-1) genotypes. In parallel with the theoretical evidence, both haptoglobin concentration and genotype, specifically Hp2-2, are associated with various age-related diseases, such as cancer, cardiovascular disease, etc. ⁴⁶. However, the underlying mechanisms related to the pathophysiology of these diseases still remain to be demonstrated.

The complex role of ROS in health maintenance and diseases

The role of ROS in the body is rather complex, and the influences on health vary largely along with changing ROS levels. ROS levels, as a reflection of oxidative stress, are modulated by oxidant generation and their elimination, and are linked to many pathophysiological processes. Within physiological levels, ROS are in a biological redox steady state ¹² and facilitate the maintenance of cellular homeostasis and function. However, ROS levels would go toward either side beyond dynamic balance (pathological states). Thus ROS levels have both beneficial and damaging aspects, as put forward in the concept of mitohormesis ⁴⁷. Consequently, both (too) low and (too) high levels of ROS will have adverse health effects, as illustrated in **Figure 1**.

Physiological levels: beneficial health effects

ROS may act as second messengers owing to the characteristics of having an intricate system for synthesis and removal as well as reversible signaling effects. Both superoxide and hydrogen peroxide could be potential messengers to regulate reduction-oxidation-dependent signaling mechanisms, while hydrogen peroxide has a higher advantage in signaling capacity given its stability and membrane permeability. The major mechanism underlying most redox-dependent signaling has been considered as the reversible modulation of cysteine thiol groups (thiolate anion to sulfenic form Cys-SOH) regulated by hydrogen peroxide ^{14,48}.

Within physiological levels (Range II in **Figure 1**), ROS can promote host defense mechanisms such as for the optimal activity for macrophages against bacteria, as well as in signaling pathways, such as toll-like receptors initiated pathways, Mitogen-Activated Protein Kinase (MAPK) signaling pathways, NF- κ B signaling pathway and Keap1-Nrf2-ARE signaling pathway ^{13,14,49,50}. Therefore, ROS levels within the physiological range are critical signaling molecules for many redox-dependent signaling processes including gene expression, metabolic regulation, inflammatory response, stem cell proliferation and differentiation, cancer pathogenesis as well as ageing. Intensive discussion regarding ROS signaling physiological consequences has been provided in previous reviews ^{13,14}.

Given the signaling effect of ROS, we can speculate that a certain increase of ROS in the physiological range would lead to better health effects, from health maintenance to health promotion, such as decreased risks of diseases, or even prolonged lifespan⁴⁷, as illustrated in IIA, **Figure 1**. However, with further increase of ROS levels and more damage events, this beneficial effect may decrease or disappear, but with no manifestation of pathological symptoms (IIB, **Figure 1**).

Elevated or Decreased Physiological levels (Pathological state): adverse health effects

When ROS levels are beyond the range of physiological levels, either (too) low or (too) high, adverse health effects can happen. For example, upon inflammatory stimulation, neutrophils are activated and generate large amounts of ROS for host defense. However, Ncf1 (neutrophil cytosolic factor 1) mutated mice, with low production of ROS, have higher type I interferon and develop an accelerated lupus-like disease⁵¹. Similarly, the Ncf1-339 T allele, related to reduced extracellular ROS production in neutrophils and increased type 1 interferon-regulated genes expression, was found enriched in systemic lupus erythematosus patients compared to healthy controls⁵². Moreover, patients with NADPH oxidase 2 (Nox2) mutations that are associated with lower ROS levels, were more susceptible to develop chronic granulomatous disease⁵³. These findings indicated an association between lower ROS generation that is not sufficient to maintain physiological processes which could lead to inferior health outcomes, as shown in Range I, **Figure 1**.

On the other hand, in the situation of high ROS levels (toxicity), excessive ROS might irreversibly react with intracellular macromolecules, including lipids, proteins, and DNA, both in the mitochondria and nucleus. Hazardous products are formed and accumulated, and the functions of macromolecules and organelles are altered. Once these exceed the repair capacity of the body, devastating damage will occur, and accelerated ageing or multiple diseases might manifest or progress (discussion in part 2.3 and part 3), as shown in Range III, **Figure 1**.

What is the physiological level?

Though **Figure 1** shows the possible ranges of ROS levels, several caveats should be noted. Firstly, how to define “physiological levels”. ROS generation happens all the time and signaling merits and damaging events occur simultaneously. For example, protein oxidation will produce many harmful byproducts, meanwhile, it is also involved in redox signaling and control, especially cysteine side chain peroxidation. When we are talking about the levels of either ROS or oxidative stress, we should be more cautious of using “high” and “low”, given that “normal” is undefined. Possibly, the so-called “high” or “low” ROS levels close to the physiological range should also be regarded as tolerable levels. For example, transgenic animal models with lower SOD production did not show a shortened lifespan in mice^{54,55}, and even an extended lifespan in *C.elegans*⁵⁶. This “lower SOD production” may lead to a relatively “higher ROS level”, but notably, this level could be tolerated by the experimented animals and they also function well for maintaining or promoting health, thus being potential physiological. In addition, antioxidants would have a similar dose-response manner since the change of

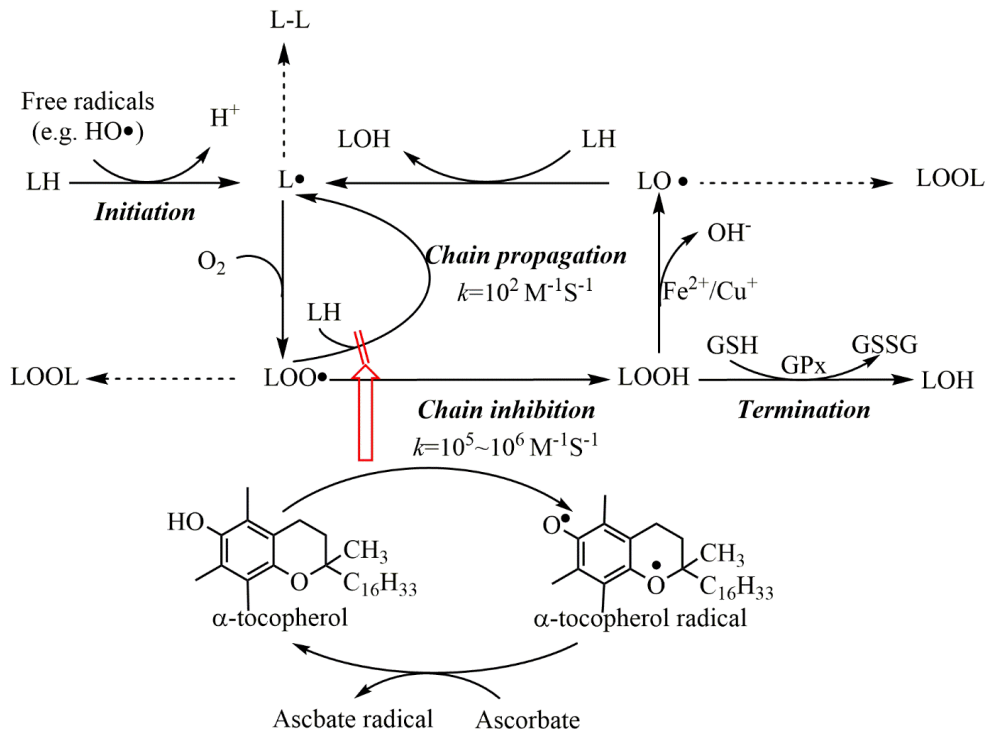


Figure 5 Lipid peroxidation chain reaction induced by reactive oxygen species

L: lipid radical, LOO: lipid peroxyl radical, LOOH: lipid hydroperoxides, LOOL: peroxide bridged dimer, L-L: fatty acid dimer, LOH: aldehydes (e.g., MDA, HNE), α -TocH: α -tocopherol, α -TocH \cdot : α -tocopherol radical, Asc: ascorbic, Asc \cdot : ascorbic radical

Three steps of lipid peroxide chain reaction:

Initiation: ROS (especially hydroxy radicals) initially trigger a reactive hydrogen atom extraction from the methylene group of polyunsaturated fatty acid forming carbon-centered radicals at the allylic position, and then react with oxygen molecule thus forming a peroxyl radical.

Propagation: Peroxyl radical, as an intermittent radical in the reaction chain, may further react with nearby lipids, yielding lipid hydroperoxides (LOOH), as well

antioxidant levels are mostly inversely associated with oxidants, for example, in *C. elegans*, an inverted U-shaped dose-response relationship between antioxidants and lifespan was observed⁵⁷. Consequently, the same situation might also be there for oxidative stress.

Damages caused by oxidative stress

Lipid peroxidation

Polyunsaturated fatty acid (PUFA), particularly with a high number of double bonds, such as arachidonic acid and linoleate, are highly susceptible to ROS and free radicals, known as autocatalytic chain reaction⁵⁸. Lipid peroxidation includes

three steps: initiation, propagation, and termination, as shown in **Figure 5**. Phospholipids peroxidation in lipid membranes may lead to a decrease in membrane fluidity and permeability, and inactivation of receptors, resulting in cell apoptosis. Furthermore, lipid radicals generated during oxidation processes can form a myriad of deleterious end products, including reactive aldehydes, alkanes, and alkenes⁵⁹. Among those products, malondialdehyde and 4-hydroxy-2-nonenal (4-HNE) are two of the most widely studied aldehydes. However, due to the high reactivity of these aldehydes, for example, they can react with proteins through the Michael addition reaction or with DNA to form adducts^{60,61}, it is difficult to accurately measure their free concentrations as valid oxidative damage levels. Moreover, urinary malondialdehyde levels are affected by dietary malondialdehyde content⁶², and glutathione S-transferases genetic polymorphisms could confound the metabolism of 4-HNE⁶¹. Therefore, both the properties of the products themselves, xenobiotic sources, and genetic predispositions present obstacles for their development and further use as reliable biomarkers.

F₂-Isoprostanes (F₂-IsoPs), another important class of lipid peroxidation end products, can be formed through the non-enzymatic oxidation of arachidonic acid. These are prostaglandin-like compounds initially formed in-situ esterified to phospholipids and subsequently released in free form by phospholipases⁶³. Isoprostanes have widely been considered as reliable biomarkers quantitating oxidative stress with their important merits of being (i) stable, both chemically and unaffected by diet or health status; (ii) sensitive to changes in oxidative stress; (iii) present in detectable quantities in many biological fluids, such as urine and plasma; and (iv) accessible to define a normal range in the population level which means the change of their formation can serve as the reflection of different oxidative stress levels and further relate to pathological status or diseases⁶⁴⁻⁶⁶. A meta-analysis identified different quantitative levels of 8-isoprostane-F_{2a} in patients across different pathological conditions, with relatively small increased levels observed in patients with, for example, hypertension and metabolic syndrome, while large increased levels were observed with patients such as kidney related pathologies and respiratory tract disorders⁶⁷.

Protein oxidation

Proteins are also important targets for ROS. Protein oxidation involves (i) oxidative modification of site-specific amino acid residues, including oxidation of sulphur-containing amino acid residues, particularly cysteine and methionine, as well as reactions with aromatic amino acid residues and peroxyxynitrite; (ii) protein fragmentation resulting from oxidative cleavage of the peptide backbone; (iii) generation of protein carbonyl derivatives; and (iv) generation of protein-protein cross-linkages⁶⁸⁻⁷⁰. Protein oxidation may lead to a change in its three-dimensional structures, modification of physiological properties such as enzyme activities and signal transduction networks which are related to cellular regulation and function, and further protein proteolytic degradation on one hand, or protein aggregation, partial unfolding, and altered conformation on the other hand⁷⁰⁻⁷². Carbonyl derivatives are a large group of end-products of protein oxidation that have been considered as the most widely used biomarkers of oxidative damage to protein. They can be formed through oxidative cleavage of backbones, direct oxidation of amino acid residues including lysine, arginine, proline, and threonine,

as well as the reaction of amino acid residues with aldehyde resulting from lipid peroxidation or carbohydrate⁷³. Among the protein carbonyls, advanced glycation end products (AGEs) are a heterogeneous group of additive derivatives produced by reactions between proteins and oxidation end products from PUFAs or carbohydrates, such as hazardous aldehyde⁷⁴. Other protein oxidation end-products include advanced oxidation protein products (AOPP) formed mainly by myeloperoxidase-derived chlorinated oxidants⁷⁵, oxidized low-density lipoprotein (oxLDL) derived from oxidative modification⁷⁶, and nitrotyrosine resulting from tyrosine oxidation⁷⁷. However, because of the structural heterogeneity caused by different biochemical pathways as well as the low specificity and sensitivity of antibodies in available detection assays, it is difficult to use them as stable and efficient biomarkers in large epidemiological studies.

Nuclear DNA oxidation

ROS, in particular the hydroxyl radicals, can cause oxidative damage to the DNA, which includes: (i) base mutation; (ii) strand breaking, both single and double; (iii) DNA-protein cross-links; and (iv) formation of DNA-adducts. In general, hydroxyl radicals could react with DNA bases and sugar-phosphate backbone, leading to erroneous base pairing and further common mutation of G to T and C to A bases⁷⁸. Meanwhile, hydroxyl radicals could also react with the deoxyribose moiety leading to the loss of DNA bases, generating base-free sites, thus single or double DNA strand(s) could break. DNA strand breaks are well-established risk factors of genome instability, cell cycle disruption, as well as cell death⁷⁹⁻⁸¹. DNA-protein cross-links involving thymine and tyrosine in the nucleoprotein complex of histones and DNA (nucleo-histone) can also be induced by the hydroxyl radicals⁸². In addition, adducts to DNA can be formed by the reaction of deoxyguanosine (M(1)G) and deoxyadenosine with other macromolecular modifications triggered by ROS, such as reactive aldehyde products generated during the lipid peroxidation⁸³.

Specific to the most common base mutations, there are mainly three ways: (i) adding double bonds to heterocyclic DNA bases; or (ii) abstracting a hydrogen atom from the methyl group of thymine; or (iii) forming allyl radical of thymine and carbon-centered sugar radicals^{84,85}. For pyrimidines, hydroxyl radicals react with the particularly sensitive C5- and C6- double bond of thymine and cytosine, generating a spectrum of adducts, including thymine glycol, cytosine glycol, 5-hydroxymethyl uracil and 5-formyluracil, 5-hydroxycytosine, uracil glycol, and 5-hydroxyuracil^{85,86}. For purines, similarly, C4-, C5, and C8- sites are more sensitive to radicals^{85,86}. Among all these DNA oxidative modifications, the mutagenic lesions of saturated imidazole ring 7,8-dihydroxy-8-oxo-7,8-dihydroguanine (8-oxodG), formed from hydroxylation of the C8 residue of guanine, has been the most abundant, best characterized, and widely considered as a potential biomarker of DNA oxidation^{85,87}. Theoretically, the concentration of 8-oxodG measured in urine, with its merits of long-term stability in urine and multiple measurement methods, reflects the accumulative DNA oxidation in the whole body and is thus considered being predictive for ageing and multiple diseases. However, unlike other molecules, oxidative damage to DNA might be repaired by the DNA repair systems⁸⁸. So the ROS-induced 8-oxodG concentration is not only related to oxidative damage but also depends on the DNA repair capacity.

Yet, due to the unknown inter-individual difference in DNA repair capacity, it is difficult to quantify the real 8-oxodG concentration induced by ROS accurately in individuals, therefore, the further use of this biomarker has been hampered.

Mitochondrial dysfunction

The mitochondrial respiratory chain is the main intracellular place for endogenous ROS generation, with steady higher concentrations of free radicals in the mitochondrial matrix, and this leads to close proximity of mitochondria to ROS. Evidence showed that an about 5- to 10-fold higher superoxide anion level is present in the mitochondrial matrix than in the cytosolic and nuclear spaces³⁸, making the mitochondria primary targets for ROS-induced damages.

Mitochondrial DNA (mtDNA) casts a critical role in energy generation during oxidative phosphorylation by the function of encoding important bioenergetic genes including 13 polypeptides, 22 transfer RNA, 2 ribosomal RNA, which are essential for the synthesis of multi-subunit enzymatic components of the electron transport chain⁸⁹. However, there are several factors ascribed that contributed to the higher vulnerability of mtDNA to oxidative damage: (i) close proximity to the oxidants forming site; (ii) the absence of histones in mtDNA; and (iii) limited mtDNA repair activity. This less active repair capacity is partly attributed to the multiple copies of mtDNA and a circular and compact coding arrangement in mtDNA⁹⁰. Therefore, damaged mtDNA would be degraded and replaced by newly produced ones copied from undamaged genomes, which gives a higher ability to mitochondria against detrimental mutation effects. Meanwhile, there are also fewer mtDNA repair mechanisms than that in nuclear DNA and mostly are poorly understood. Though base excision repair, mismatch repair, as well as recombination or nonhomologous end joining repair mechanisms, do exist in mtDNA, nucleotide excision repair is still absent, and the specific roles of each pathway, as well as proteins involved in repair processes, remain to be fully elucidated^{90,91}. It is estimated that oxidized bases are 10 times more frequent in mtDNA than nuclear DNA, for instance, the concentration of the common DNA lesion 8-oxodG is much higher in mtDNA than in nuclear DNA^{36,92}. When a mutation occurs, mutated and normal mtDNA co-exist in the same mitochondria, which is known as heteroplasmy^{93,94}. The percentage of an erroneous sequence of mtDNA changes during cell replication and division, and mitochondria could aggregate different mutations over time, reducing their bioenergetic capacity⁹⁵. When the increasing proportion of mutated mtDNA exceeds the critical threshold level of normal mitochondrial energy generation, bioenergetic defects in cells may occur. Besides, the sustaining existence and accumulation of damaged mtDNA in the mitochondria will inevitably lead to more ROS generation, which in turn causes further damage, making a vicious cycle⁹².

The mtDNA damages would lead to loss of redox homeostasis, perturbed Ca²⁺ homeostasis, damage to membrane proteins and lipids, as well as to abnormal mitochondrial energy transduction⁹⁵. All these modifications are the driving force for further mitochondrial dysfunction and loss of integrity, which will affect cell viability and cellular function^{96,97}. In addition, organs, especially the brain, heart, and muscles, being high-energy consuming and sensitive to bioenergetic defects, are strongly affected by mitochondrial dysfunctions, resulting in organ-specific

pathologies. Moreover, variations in mtDNA have also been shown to associate with several clinical phenotypes, including cardiovascular diseases, anthropometric and metabolic measures⁹⁸.

In addition to mtDNA, mitochondrial sirtuins have been found to be involved in the regulation of redox homeostasis. Sirtuins are a group of proteins (from SIRT1 to SIRT7), acting predominantly as nicotinamide adenine dinucleotide (NAD⁺) dependent deacetylases and ADP-ribosyltransferases. SIRT3, 4, and 5 are exclusively localized within mitochondrial, altering the NAD/NADPH ratio in mitochondria to respond to metabolic changes. SIRT3 is supposed as the major mitochondrial deacetylase and is the most thoroughly studied mitochondrial sirtuin. It plays an important role in mitochondrial bioenergetics⁹⁹⁻¹⁰², as well as promotes resistance to oxidative stress by reducing ROS via magnesium SOD deacetylation and activation^{99,100}. Though SIRT4 and SIRT5 are less well understood, SIRT5 has also been shown to be strongly associated with oxidative stress signaling and protect cells from ROS, suppressing oxidative stress levels¹⁰³. In contrast, SIRT4 might have dual roles, either to induce ROS production or to have antioxidative function¹⁰³. In addition, these mitochondrial sirtuins work in concert and present complicated interaction profiles¹⁰⁴, for example, SIRT3 and SIRT4 may function together to protect cells from stress and DNA damage¹⁰⁵. A series of reviews regarding sirtuins on mitochondrial and oxidative regulation has been published before^{99-102,106-108}. Nevertheless, additional studies are needed to in-depth ascertain and illustrate the mechanism of mitochondrial sirtuins in maintaining mitochondrial biological functions and redox homeostasis.

Causes and consequences of oxidative damage on ageing and age-related diseases

The progressive accumulation of oxidative damage to macromolecules and mitochondria will finally lead to pathophysiological alterations, functional decline, and accelerated ageing. Here we will discuss the harmful consequences of oxidative stress, mainly oxidative damage which are more closely related to accelerated ageing and multiple age-related diseases, with a special focus on ageing and lifespan, cardiovascular diseases (CVD), and neurodegenerative diseases (NNDs).

Ageing and lifespan

The best way to determine the effect of oxidative stress on ageing is to test whether changes in lifespan are dependent on alterations in oxidative stress levels. Numerous studies aimed to investigate this effect have been conducted on animals. Based on the free radical theory of ageing, attempts to extend lifespan have been carried out with a focus on two different targets: alterations of oxidant load (i.e. ROS generation) or alternations of mitochondrial antioxidant capacity. Alternations of oxidant load are usually investigated through calorie restriction, while alternation of mitochondrial antioxidant capacity is often examined through inducing genetic alterations, or through dietary supplementations.

Calorie reduction (CR) is defined as a 10-50% reduction of total energy intake without inducing malnutrition. A large body of evidence suggested that CR

might modulate the mitochondrial activity and lower ROS production resulting in the reduction of oxidative damage through sirtuins regulation, for example through primary activation of sirtuin 1 and further activation of PPAR γ coactivator-1 α (PGC-1 α), leading to a slower rate of aging-related decline and extended lifespan¹⁰⁹⁻¹¹¹. Several reviews indicated that, in lower model organisms ranging from yeast to mammals, CR is capable to extend both average and maximum lifespan and health span¹¹²⁻¹¹⁶. Interestingly, studies in humans also detected some favorable biomarkers of longevity induced by CR, mainly hormonal adaptations, such as lower insulin, and lower IGF-1 level, though the latter was mainly in response to protein restriction^{117,118}.

With respect to induced genetic alternations, antioxidants-related transgenic lower models have been frequently used, which have altered antioxidant capacity or a disrupted oxidative-related signaling pathway, leading to either a reduced or extended lifespan¹¹⁹⁻¹²³. Mice with antioxidant overexpression, for example, catalase¹²⁴, or with modifications in other signaling pathway components, such as p66^{sch-/-}¹²⁵, Igf1^{r+/-}¹²⁶ are generally considered to have increased longevity. However, other results are inconclusive about altered expression levels of, especially SOD^{54,55,127,128}, thioredoxin^{129,130}, or methionine sulfoxide reductase A (MsrA^{-/-}) that function to protect against protein oxidation^{131,132}. Moreover, Gpx1^{-/-} knocked-out mice with reduced glutathione peroxidase-1 surprisingly did not show accelerated ageing^{55,128}. Similarly, results on the longevity of transgenic invertebrate models, such as *C. elegans* or *drosophila*, with alternations of antioxidants capacity, are also quite divergent, and some studies even found a large extension of lifespan with elevated oxidative stress level^{56,133-140}. The inconsistency was also found in experiments with dietary antioxidants supplementation¹⁴¹.

However, oxidative stress may act as a double-edged sword to human health, as discussed in part 2.2. It is difficult to determine a lifespan-affecting ROS level and this further lead to the ambiguity on the relation of oxidative stress and ageing. Nevertheless, a recent review still concludes that mitochondria play a critical role in the ageing process, but it remains unclear whether it is a cause or consequence¹⁴².

Cardiovascular diseases (CVDs)

Cardiac myocytes have a high number of mitochondria which makes them more susceptible to oxidative damage. In addition to the mitochondrial pathway of ROS generation, crosstalk exists between mitochondria and NADPH oxidase, the major source of ROS in blood vessels, affecting vascular function by dysregulation or uncoupling eNOS, leading to endothelium dysfunction¹⁴³. Together with other ROS-induced damages involved in mitochondrial dysfunction, and ROS-induced macromolecule damages (e.g. ox-LDL), they all contribute to the pathogenesis and progress of CVD^{143,144}. To date, several lines of evidence have found an association between onset and progression of cardiovascular diseases (CVD), which include atherosclerosis¹⁴⁵, hypertension¹⁴⁶, heart failure¹⁴⁷, and peripheral artery disease¹⁴⁸.

In rodents, deletion of the NADPH oxidases gene (Nox1 and Nox2) or p47phx deficiency resulted in markedly lower ROS generation, and yielded a lower risk

of atherosclerosis, while Nox4 deletion gave an increased risk¹⁴⁹. Similarly, over/under-expression of different antioxidant enzymes modulated atherogenesis in different genetically-altered mouse models^{147,149}. In human studies, increased expression and activity of NADPH oxidase and xanthine oxidase subunits were associated with higher ROS production and an increased risk of coronary artery disease¹⁵⁰. Moreover, mtDNA copy number, the increase of which is indicative of early molecular changes compensating for oxidative stress-induced mitochondrial defects, has been shown to be associated with CVD. A study composed of 21870 participants from three large cohorts found that the CVD incidence increased 23% with a one-standard deviation decrease in mtDNA copy number after adjustment for traditional CVD risk factors¹⁵¹. In addition, epidemiological cohort studies consistently showed that higher dietary intake of antioxidants, either as diet components or supplements, including flavonoids, vitamin E, and C, are associated with lower CVD risk, such as coronary heart disease, ischemic stroke as well as CVD mortality^{152,153}. However, large randomized controlled clinical trials generally failed to show any beneficial effect of antioxidants supplementation, including vitamin E, A, C, B12, B6, and folic acid, on CVD risk, especially in atherosclerosis^{153,154}. Meta-analyses also concluded that there is insufficient evidence to support vitamins or other antioxidants supplementation to decrease CVD incidence^{30,155}. Interestingly, a very recent Mendelian Randomization study found there are causal associations between higher circulating vitamin E and the risk of coronary artery disease or myocardial infarction¹⁵⁶. However, the three vitamin E-associated instrumental SNPs used in this study are all associated with blood lipid levels or coronary artery disease itself. This violates the important condition for conducting Mendelian Randomization that no horizontal pleiotropy for instruments. Though sensitivity analysis was performed, this is only for one SNP but not for others, thus the results are of limited value. Therefore, despite the biological evidence from consistent results, the premise of interventions with the intention to decrease oxidative levels and consequently CVD risk is still limited.

Neurodegenerative diseases (NDDs)

NDDs are characterized by dynamic and progressive neuronal cell damage and loss of neurons, typically including Alzheimer's disease (AD) and Parkinson's disease (PD). Ageing is considered to be the predominant risk factor for NDDs, and accumulative oxidative damages during ageing are the main culprits of neurological deterioration^{96,157}. Neurological systems are extremely sensitive and vulnerable to ROS-induced damage due to their higher energy demand, lower rate of cellular renewal, membrane PUFA levels, as well as less active oxidative defense mechanisms¹⁵⁸. Notably, studies suggested that mitochondrial dysfunction plays a causal role in the pathogenesis of NDDs^{157,159}. Sirtuins, which provide a key role in mitochondrial function and oxidative stress regulation (part 2.3) are also associated with neuron fates, being either neuroprotective or neurotoxic^{102,160,161}. A large number of proteins that are implicated in NDDs are found to have mitochondrial involvement. AD is pathologically characterized by extracellular deposition of amyloid- β peptide (A β) and intracellular neurofibrillary tangles composed of hyper-phosphorylated tau. Oxidative stress may activate signaling pathways, such as Jun amino-terminal kinase and p38 mitogen-activated protein kinase, and give rise to amyloid precursor protein and β -secretases to form A β ,

as well as increase tau phosphorylation^{96,162}. In turn, A β proteins would directly induce ROS generation through activating NADPH oxidase (as shown in Figure 2), further exacerbating neuronal damage, amplifying neurotoxicity¹⁶³. Besides, mtDNA control-region adopted more mutations in 23 pathologically confirmed AD brain samples compared with 40 controls, and heteroplasmic mtDNA control-region mutations increased 63% on average in all AD brains, while markedly increased 130% in patients older than 80 years¹⁶⁴. 1-methyl-4-phenylpyridinium (MPP+), the oxidized product of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), and rotenone, both of which are mitochondrial complex I inhibitors, have been demonstrated to result in clinical PD phenotype by producing pathological degeneration of dopaminergic neurons of the substantia nigra^{165,166}. Other PD-related proteins including α -Synuclein, Parkin, DJ-1, PINK1, LRRK2, and HTRA2 were associated with mitochondrial dysfunction as well^{96,167}. In addition, ROS-induced macromolecule damage, for example, MDA and HNE, lipid peroxidation end products, may also play an important role in NDDs¹⁶⁸⁻¹⁷⁰. This can be evidenced by the fact that increased levels of oxidative stress biomarkers and decreased levels of antioxidative biomarkers were observed in pharmacologically induced or transgenic animal models with AD or PD¹⁶⁸. Several papers have speculated oxidative stress as the key component in the etiology of NDDs^{96,157,158,167,168,171-174}.

Many, but not all, observational studies indicated that a higher intake of antioxidants or supplements such as vitamin E could reduce the risk of NDDs¹⁷⁵⁻¹⁸¹. Plasma vitamin E levels are associated with better cognitive performance¹⁸², and patients with AD were found to have a lower nutrient status of vitamin E, C, B12 in blood and brain or cerebrospinal fluid^{183,184}. Animal experimental studies also showed a preventive effect of vitamin E supplementation on AD risk¹⁸⁵. However, most clinical trials have shown disappointing results in terms of the benefits on cognitive decline or dementia of antioxidants supplementation for NDDs, including vitamin E, C, B12, B6, and coenzyme Q10. Meta-analyses also suggested that there is no evidence of vitamin supplementation for prevention of cognitive decline or dementia in cognitively healthy adults¹⁸⁶, or for prevention of progression from mild cognitive impairment (MCI) to dementia, or for treatment of MCI, or for improvement of cognitive function in people with MCI or dementia due to AD^{32,187}. Similar results were partly observed for PD^{188,189}. A series of seminal papers on antioxidant therapy as well as the challenges in AD have been reviewed thoroughly¹⁹⁰⁻¹⁹².

In order to disentangle the causality of antioxidants and AD, two Mendelian Randomization studies were conducted recently. In European-ancestry individuals, there is no causal association between circulating vitamin E levels¹⁹³, as well as ascorbate, β -carotene, retinol¹⁹⁴, and AD risk. However, the SNPs selected as vitamin E genetic instruments are also associated with lipid metabolism, and only every single SNP for ascorbate and retinol was selected, limiting the power of the analyses. Meanwhile, these instruments only explained a small portion of the total variance thus may not fully represent the circulating vitamin levels. It remains unclear to what extent vitamin levels are associated with oxidative stress. Therefore, nevertheless the compelling biological plausibility and observational evidence, there is also a lack of convincing data for the antioxidant supplementation aiming to prevent or treat NDDs.

Future Perspectives and Concluding Remarks

Reasons for the failure of antioxidant supplements in clinical trials

Based on the free radical theory and positive results from both experimental and observational studies, numerous clinical trials examining antioxidants, particularly vitamin E, have been conducted for the prevention and/or treatment of age-related morbidity and mortality. However, results are apparently disappointing as partly discussed earlier in part 3. Here, we will have an in-depth summary of the critical points in clinical trials with a focus on the widely-studied vitamin E supplementation.

Vitamin E is a well-known chain-breaking antioxidant, as shown in **Figure 5**. Alpha-tocopherol competitively reacts with lipid peroxy radical with a higher reaction rate to inhibit chain propagation. Theoretically, abundant vitamin E will ameliorate oxidative damage by preventing lipid peroxidation. Not surprisingly, most observational studies specifically found detrimental/protective effects of high oxidative stress levels/antioxidants levels on disease status. However, oral vitamin E supplementation in middle-aged participants failed to demonstrate protective effects on neither primary and secondary prevention, nor on the treatment of various age-related diseases. Several potential reasons for this failure have been discussed extensively ¹⁹⁵⁻¹⁹⁸. Data reported that the plasma vitamin E levels of participants with normal baseline blood tocopherol levels could only increase less than 2-3 times no matter the amount or duration of supplementation ¹⁹⁹. Therefore, when individuals have a relatively low oxidative level and/or sufficient or high antioxidative level at enrollment in a study, it is highly unlikely to generate many benefits via extra supplementation, which might be explained as a potential “functional concentration limit”. This hypothesis is supported by studies in vitamin E supplementation stratified by haptoglobin genotype. Vitamin E supplementation provided cardiovascular-protective effects only in discriminatingly selected subgroups of individuals with both diabetes and Hp2-2 genotype ²⁰⁰⁻²⁰², who had high levels of oxidative stress and inferior antioxidant protection, in whom supplementation decreased cardiovascular events by 34% ²⁰³. Besides, a larger improvement was observed in nonalcoholic steatohepatitis patients carrying the Hp2 allele after vitamin E treatment compared with Hp1 allele carriers ²⁰⁴. Still, biological mechanisms underlying these observations should be explored in greater detail in future studies.

Another potential reason for the failure of vitamin E supplementation in clinical trials might lie in the difference of forms of tocopherols. In many studies, for example, in observational studies, vitamin E has been used, but this “vitamin E” was not specific to α -tocopherol and it is more about the combination of complicated tocopherol or tocotrienol isomers, while only α -tocopherol can be called “vitamin E” given its proved ability to avoid vitamin E deficiency ²⁰⁵. Natural vitamin E has the highest bioactivity in the human body compared to other tocopherols or tocotrienols. However, most of the oral supplementation of vitamin E is synthetic, with lower bioactivity than the natural one, characterized by preferentially non-oxidation metabolites in urine ²⁰⁶.

Besides, administration heterogeneity could result from various reasons and become critical in clinical trials, such as timing, monotherapy or not, dose and duration. Normally, it is hard to define a standardized strategy for supplement administration since chronic diseases often have an ambiguous onset time and the progression can take several years. Administration of the antioxidant supplements after irreversible ROS damage could have a negligible beneficial effect at that moment in time. Therefore, there might be a certain “critical window”, and antioxidants administrated at an optimal timing before disease onset or during the early stages of disease could be able to ameliorate ROS-induced damage. Similarly, the duration of intervention needs to be sufficiently long to observe these effects.

Moreover, there is still a lack of consensus about the proper dosage regimen for antioxidant supplements. The doses used in clinical trials are derived from observational studies and are relatively safe and low. However, the dose for vitamin E to suppress plasma F_2 -isoprostane concentration is 1600 IU or 3200 IU without observing any adverse effect after 16 weeks of supplementation, while the suppression of isoprostane were 35% and 49% respectively ²⁰⁷. Importantly, the authors suggested that a 49% reduction with the largest dose of 3200 IU/day was not so profound. Davies et al also concluded from two modeling techniques, that only a sufficiently high concentration of non-enzymatic antioxidants would have a chance for collision and interaction with free radicals ²⁰⁸. However, oral intake of vitamin E only modestly increases the plasma and tissue vitamin E levels, and this decreases the concentration of vitamin E at target cells or organelles and slows down the reaction rate with ROS, which was quite unlikely to affect health outcomes ²⁰⁹. And also, endogenously present defense systems will weaken the effects of non-enzymatic antioxidants when the reaction rate with free radicals is below a necessary threshold ²⁰⁸. Besides, some studies suggested vitamin E could also act as pro-oxidant after denoting a hydrogen atom to a lipid radical ²¹⁰, so regenerating the reduced form from the oxidized form guarantees the lower depletion rate as well as the antioxidant function of vitamin E. Theoretically, combined administration of vitamin E and other antioxidants, such as ascorbic acid, coenzyme Q10 which can reduce oxidized vitamin E, might be protective. However, trials regarding this point are also inconsistent ^{28,177}.

Ageing and age-related diseases are multifactorial, heterogeneous, and multidimensional. They do not respond to the organ-centered paradigm of “one cause - one mechanism - one disease/condition - one therapy”. Therefore, a single therapeutic component that targets only one aspect of the several factors contributing to aging and age-related diseases, even if highly weighted like oxidative stress, might be less effective as the solution by itself. Similarly, it is highly possible that only one component (vitamin E) might not be enough to show physiological significant effects on the complex, heterogeneous pathways of organ dysfunction with increasing age.

In addition to these potential explanations for the lack of significant findings of vitamin E supplementation, some opinions argued that oxidative damage could merely play a role in the pathophysiology instead of being a direct cause of diseases ^{171,198,211}. However, emerging evidence points to the possible etiological role of oxidative stress, especially ROS-induced mitochondrial dysfunction in

NDDs^{157,159,162}. To demonstrate the causality, the best way is to directly measure the association of ROS, or ROS-induced damage change with pathological disorders or diseases, or the improvement of clinical phenotypes led by prevention of oxidative damage^{84,198,212}. Yet, it is almost impossible to measure total ROS levels in the human body because of their chemical properties of short-existence and high reactivity. Similarly, it remains a challenge to accurately measure oxidative damage because of the lack of reliable biomarkers as well as technical difficulties, further leading to the uncertainty of causality.

Potential oxidative stress biomarkers in clinical trials

After oral supplementation, vitamin E is digested and absorbed into the circulation by the body. However, plasma vitamin E concentrations after supplementation in a healthy population with a normal range of baseline vitamin E could largely vary from one individual to another which may arise from variations in the regulation of vitamin E uptake and metabolism between subjects, such as genetic factors, particularly apolipoprotein polymorphism²¹³. Besides, though no causal association was shown in the previous Mendelian Randomization of circulating vitamin E and Alzheimer's disease¹⁹³, this may raise another argument that even circulating levels, especially induced by synthetic vitamin E, should not be considered the same as functional levels. First, vitamin E has two metabolic ways, to go either through non-enzymatic, namely free radical-dependent metabolism, opening the hydroxy-chromanol ring, or through enzymatic (mainly CYP4F2 of cytochrome P-450 family) metabolism with only successive shortening of phytol side chain²¹⁴ (**Figure 6**). As synthetic vitamin E preferentially goes through the non-oxidation pathway, it is essential to determine whether or not, it acts as an antioxidant in the body. Second, oxidative damage is often located and gathered in certain tissues, specific cell types, or organelles. Although plasma circulating vitamin E is incorporated into lipoproteins and non-specifically transported to tissues, the acquisition of vitamin E seems to be largely mediated via the delivery and selective uptake of vitamin E after lipoprotein particles binding to receptor²¹⁵. However, the mechanisms of the delivery and uptake of tissue vitamin E have been relatively poorly-characterized in other tissues except only for the hepatic tissue, and it appears to be essentially related to the expression and function of lipoprotein receptors. It is still hard to define the uptake level by many tissues precisely. Thus the accumulation of vitamin E could be different in various tissues, for instance, an only limited amount of vitamin E in the brain was observed after supplementation, or could be different for regions within the same organ, for example, the striatum contains the lowest amount of vitamin E compared with other areas in the brain²¹⁶. In addition, once taken up by cells, the intracellular distribution of vitamin E to organelles is regulated by different transport proteins, but these proteins and their activity could differ from one tissue to another²¹⁵. Hence the vitamin E concentration at different target tissues or organelles largely differ, and there might be insufficient concentration in certain target locations to counteract with overwhelmed oxidative stress damage, particularly in the high energy-consuming organs such as the brain. Obviously, the real functional vitamin E might be completely different with, to be more precisely, far less than the circulating vitamin E. Therefore, due to the differences in susceptibility to oxidative damage and response to vitamin E supplementation among individuals, even identical supplementation strategies

(e.g. vitamin E type, administration protocol) may possibly not lead to same health outcomes. What matters most should be the ability to take advantage of circulating vitamin E by the body, namely to what extent, it acts as antioxidants. In brief, these issues can be simplified as the detection and monitoring of the antioxidative function of vitamin E.

Ideally, there are two aspects to monitor the effect of vitamin E, one is to measure robust oxidative stress levels in a range of pathologies of specific targets, as well as its alterations corresponding to vitamin E supplementation, and the other is to measure the direct level change of vitamin E acting as antioxidants. However, the major limitation of the first approach is the identification of reliable biomarkers. An oxidative damage biomarker that relates to the pathology could be promising, such as MDA, HNE, carbonyls, and particularly isoprostanes. These biomarkers have been identified in experimental and population-based studies^{67,217-219}, together with the potential associations with ageing and age-related diseases. Nevertheless, the measured oxidative damages are often the result of the complex network of both endogenous and exogenous antioxidants systems. Moreover, these biomarkers are exclusive to certain macromolecule damage which cannot reflect the whole oxidative damage of the body. Although isoprostanes have been regarded as the most reliable biomarker with many merits mentioned in part 2.2.1 so far, several possible drawbacks impede its further use. In the pathway of isoprostane formation, direct oxygen addition would compete with the second 5-exo cyclization involving a carbon-centered reaction, so the formation relies on low oxygen tension which gives priority to the cyclization reaction^{63,220}. Besides, due to the ubiquitous existence of F₂-isoprostane in normal tissues, the products derived from the kidney may confound the urine isoprostane concentration^{63,66}. This could also be an explanation why a larger increase of isoprostane was found in the kidney pathologies than other disease conditions⁶⁷. Apart from the generation pathway, quantitation of isoprostanes remains a challenge due to multiple steps in the detection method, which is labor-intensive, as well as due to the impossibility to specify each isomer⁶¹. All these drawbacks hindered the possibility as a biomarker in large epidemiological studies or clinical trials to assess the specific oxidative damage change caused by vitamin E accurately.

In recent years, vitamin E metabolites have been proposed to help better elucidate vitamin E roles²²¹. Those metabolites reveal the biological process and reflect the authentic utility of vitamin E. Notably, identification of metabolites, especially free radical-dependent vitamin E metabolites, along with the determination of their levels may provide insights into oxidative stress-related physiological functions, as well as disease mechanisms and therapeutic strategies. When vitamin E acts as peroxy lipid radical scavenger, it forms vitamin E radicals, which further react with lipid peroxides into α -tocopherol quinone (α -TQ) (**Figure 6**)²²². In presence of NAD(P)H, α -TQ captures hydrogens converting into α -tocopherol hydroquinone (α -THQ), following the β -oxidation and cyclization of the phytol side chain, α -tocopheronic acid and α -tocopherol lactone (α -TL) are generated²²³. Finally, α -TL are excreted as polar glucuronidated and sulfated conjugates of α -TL hydroquinone (α -TLHQ) glucuronide and α -TLHQ sulphate²²⁴. Theoretically, disease states that are associated with increased oxidative stress are likely to have higher antioxidants requirements, which would result in depletion of circu-

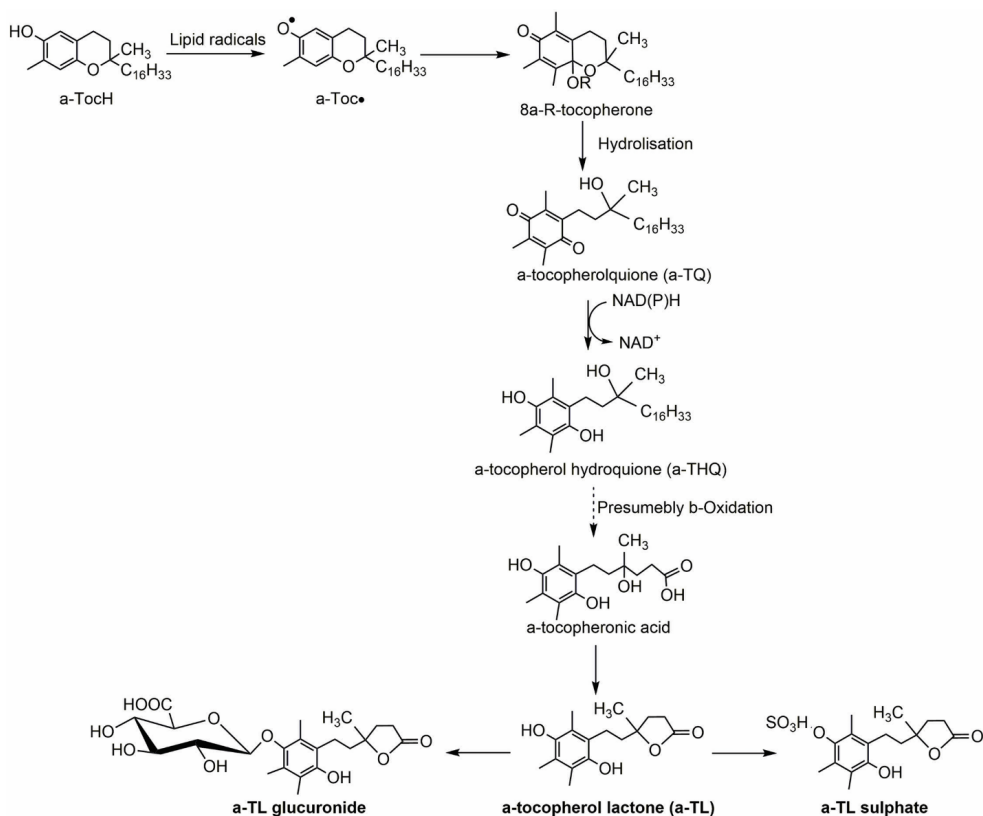


Figure 6 Alpha-tocopherol catabolism

The left panel shows the non-enzymatic metabolic pathway of α -tocopherol. Alpha-tocopherol reacts with lipid peroxy radicals, forming α -tocopherol radical, which further reacts with lipid peroxides, following hydrolyzation, generating α -tocopherol quinone (α -TQ). In presence of NAD(P)H, α -TQ captures hydrogens converting into α -tocopherol hydroquinone (α -THQ), following the β -oxidation and cyclisation of the phytol side chain, α -tocopheronic acid and α -tocopherol lactone (α -TL) are generated. Finally, α -TL are excreted as polar glucuronidate and sulfate conjugates of α -TL hydroquinone (α -TLHQ). The right panel presents the enzymatic metabolic pathway of α -tocopherol. This process is initiated in the liver, with the hydroxylation of the methyl group by hepatic CYP enzyme. Subsequently, the phytol side chain shortens successively with the removal of carbon units in β -oxidation. Finally, α -carboxyethyl-hydroxychroman (α -CEHC) are generated and excreted as their polar glucuronide or sulfate conjugates.

lating vitamin E, and consequently, a higher concentration of oxidized vitamin E metabolites in urine, as a reflection of real antioxidant capacity.

Few studies have shown that oxidized vitamin E has been associated with diseases. A level of about 3% to 11% of α -tocopherol as an oxidized form of α -TQ was found in all lipoprotein density fractions prepared from advanced human atherosclerotic plaque²²⁵. Older individuals without baseline cognitive impairment and with the highest tertile of α -TQ/cholesterol had a higher risk of prevalent

dementia²²⁶. A recent study also indicated that both plasma α -TQ and 4-HNE were higher in participants with non-alcoholic fatty liver diseases, with lipid peroxidation as one of the earliest pathogenic events, compared with healthy participants²²⁷. Though it has been claimed that the formation of α -tocopheronic acid and α -TL could be artificial products from the oxidation of α -carboxyethyl hydroxy chromans during the analytical process²²⁸, Sharma and colleagues developed a feasible way to measure these conjugates by avoiding the artifacts²²⁹, and found that the mean concentrations of α -TL conjugates were significantly higher in children with type 1 diabetes compared with age-matched controls²²⁴. Notably, all these studies were carried out in a small sample size, and it remains unclear whether the levels of the metabolites are consistent with the oxidative damage. Efforts are still required to validate the potential biomarkers in large population-based studies, and further use as clinical monitoring at the individual level after supplementation of vitamin E.

Remarks and Conclusions

There is an ongoing controversy in terms of antioxidant supplementation for the prevention and/or therapy of ageing and age-related diseases. However, large amounts of money are still spent, and tens of trials with different population sets and various administration strategies are in the plan, regarding single antioxidants clinical trials. Of note, “multifactor - multi treatments” might be more effective since there might be an additional or a synergistic benefit from two agents with acceptable safety and efficacy, for example, the combination of treatments (traditional treatment plus antioxidants treatment) to achieve better effects than antioxidants only. Interestingly, a recent trial showed that middle-aged type 2 diabetic patients who received metformin treatment plus vitamin E and/or vitamin C had significant improvement of glucose measures as well as lipid profiles compared to patients with metformin treatment alone, indicating that antioxidants use might be an adjuvant therapy in the management in type 2 diabetic patients²³⁰. But the combination use still requires additional efforts.

In summary, there are two critical points in clinical trials related to oxidative stress that need to be further discussed: (i) Target population. Participants should be selected by the stratification of certain features (e.g., genotype) which could induce significantly heterogeneous responses. So the selected participants subgroup are most likely to benefit from vitamin E supplementation. For example, vitamin E supplementation has been shown to be associated with an approximately 35% reduction in CVD specifically in individuals with both diabetes and Hp 2-2²⁰². Besides, a recent study also suggested that alpha-tocopherol supplementation was beneficial for cancer prevention only among carriers of homozygous low-activity alleles in the catechol-O-methyltransferase gene²³¹. Therefore, there is still a need for further studies aiming to identify potential geno-/pheno- types that could be a determinant of the possible benefits of supplementations, as well as the elucidation of underlying mechanisms. (ii) Reliable biomarkers. To date, there is still little consensus about the best systemic biomolecular measurements of oxidative stress. An optimal biomarker should be an authentic reflection of overall redox status, easily accessible, reasonably stable, easy to be detected accurately. Moreover, since ROS has properties of the second messenger which is necessary for the maintenance of cellular homeo-

stasis, as well as normal physiological function, it remains obscure to define an optimal range of an oxidative damage-induced biomarker from physiological to pathological disorders, i.e. which level is detrimental for health or promotes longevity and metabolic health outcomes.

There is rather convincing evidence that oxidative damage has an important role in aging and the pathogenesis of multiple age-related diseases. Yet, clinical trials did not demonstrate the preventive or therapeutic role of antioxidants supplementation. However, this does not mean the failure of the oxidative stress theory and the use of antioxidants. On the contrary, it compels us to rethink this issue with several open-ended questions: (i) Are these associations reliable in observational studies which could be biased by confounders and reverse causality. (ii) How do we infer the causality of oxidative stress and diseases at population levels. (iii) What should be the best biomarker of oxidative stress in clinical trials. (iv) How to develop proper preventive or therapeutic strategies taking all the critical points into account. Therefore, research in the coming years should be devoted to more in-depth studies examining the role of a more direct measure of oxidative stress in biofluids in the pathogenesis of age-related diseases and the ageing process, and to clarify the causality lying between oxidative stress and age-related diseases and the ageing process.

References

1. K. N. Eggleston, V. R. Fuchs. The New Demographic Transition: Most Gains in Life Expectancy Now Realized Late in Life. *J Econ Perspect* 2012; 26(3): 137-56.
2. R. Rappuoli, M. Pizza, G. Del Giudice, E. De Gregorio. Vaccines, new opportunities for a new society. *Proc Natl Acad Sci U S A* 2014; 111(34): 12288-93.
3. T. Wyss-Coray. Ageing, neurodegeneration and brain rejuvenation. *Nature* 2016; 539(7628): 180-6.
4. T. Finkel, M. Serrano, M. A. Blasco. The common biology of cancer and ageing. *Nature* 2007; 448(7155): 767-74.
5. N. J. Samani, P. van der Harst. Biological ageing and cardiovascular disease. *Heart* 2008; 94(5): 537-9.
6. C. López-Otin, M.A. Blasco, L. Partridge, et al. The hallmarks of aging. *Cell* 2013; 153(6): 1194-217.
7. T. von Zglinicki. Role of oxidative stress in telomere length regulation and replicative senescence. *Ann N Y Acad Sci* 2000; 908: 99-110.
8. T. von Zglinicki. Oxidative stress shortens telomeres. *Trends Biochem. Sci* 2002; 27(7): 339-44.
9. E. H. Blackburn, E. S. Epel, J. Lin. Human telomere biology: A contributory and interactive factor in aging, disease risks, and protection. *Science* 2015; 350(6265): 1193-8.
10. H. Sies. 1 - Oxidative Stress: Introductory Remarks. In: Sies H, ed. *Oxidative Stress*. London: Academic Press; 1985: 1-8.
11. H. Sies. Oxidative stress: a concept in redox biology and medicine. *Redox Biol* 2015; 4: 180-3.
12. H. Sies, C. Berndt, D. P. Jones. Oxidative Stress. *Annu Rev Biochem* 2017; 86: 715-48.
13. M. Schieber, N. S. Chandel. Chandel. ROS function in redox signaling and oxidative stress. *Curr Biol*; 24(10): R453-R62.
14. K.M. Holmström, T. Finkel. Cellular mechanisms and physiological consequences of redox-dependent signalling. *Nat Rev Mol Cell Biol* 2014; 15: 411.
15. R. H. Burdon. Superoxide and hydrogen peroxide in relation to mammalian cell proliferation. *Free Radic Biol Med* 1995; 18(4): 775-94.
16. K. Hensley, K. A. Robinson, S. P. Gabbita, et al. Floyd. Reactive oxygen species, cell signaling, and cell injury. *Free Radic Biol Med* 2000; 28(10): 1456-62.
17. S. Kreuz, W. Fischle. Oxidative stress signaling to chromatin in health and disease. *Epigenomics* 2016; 8(6): 843-62.
18. D. Harman. The biologic clock: the mitochondria? *J Am Geriatr Soc* 1972; 20(4): 145-7.
19. R. M. Bostick, J. D. Potter, D. R. McKenzie, et al. Reduced risk of colon cancer with high intake of vitamin E: the Iowa Women's Health Study. *Cancer Res* 1993; 53(18): 4230-7.
20. E. B. Rimm, M. J. Stampfer, A. Ascherio, et al. Willett. Vitamin E consumption and the risk of coronary heart disease in men. *N Engl J Med* 1993; 328(20): 1450-6.
21. M. J. Stampfer, C. H. Hennekens, J. E. Manson, et al. Vitamin E consumption and the risk of coronary disease in women. *N Engl J Med* 1993; 328(20): 1444-9.

22. E. J. Mayer-Davis, T. Costacou, I. King, D. et al. Plasma and dietary vitamin E in relation to incidence of type 2 diabetes: The Insulin Resistance and Atherosclerosis Study (IRAS). *Diabetes Care* 2002; 25(12): 2172-7.
23. M. E. Wright, K. A. Lawson, S. J. Weinstein, et al. Higher baseline serum concentrations of vitamin E are associated with lower total and cause-specific mortality in the Alpha-Tocopherol, Beta-Carotene Cancer Prevention Study. *Am J Clin Nutr* 2006; 84(5): 1200-7.
24. F. J. Li, L. Shen, H. F. Ji. Dietary intakes of vitamin E, vitamin C, and beta-carotene and risk of Alzheimer's disease: a meta-analysis. *J Alzheimers Dis* 2012; 31(2): 253-8.
25. R. Banks, J. R. Speakman, C. Selman. Vitamin E supplementation and mammalian lifespan. *Mol Nutr Food Res* 2010; 54(5): 719-25.
26. F. L. Muller, M. S. Lustgarten, Y. Jang, et al. Trends in oxidative aging theories. *Free Radic Biol Med* 2007; 43(4): 477-503.
27. E. R. Miller 3rd, R. Pastor-Barriuso, D. Dalal, et al. Meta-analysis: high-dosage vitamin E supplementation may increase all-cause mortality. *Ann Intern Med* 2005; 142(1): 37-46
28. G. Bjelakovic, D. Nikolova, L. L. Gluud, et al. Mortality in randomized trials of antioxidant supplements for primary and secondary prevention: systematic review and meta-analysis. *JAMA* 2007; 297(8): 842-57.
29. E. A. Klein, I. M. Thompson, Jr., C. M. Tangen, et al. Vitamin E and the risk of prostate cancer: the Selenium and Vitamin E Cancer Prevention Trial (SELECT). *JAMA* 2011; 306(14): 1549-56.
30. S. K. Myung, W. Ju, B. Cho, et al. Efficacy of vitamin and antioxidant supplements in prevention of cardiovascular disease: systematic review and meta-analysis of randomised controlled trials. *BMJ* 2013; 346: f10.
31. L. Wang, H. D. Sesso, R. J. Glynn, et al. Vitamin E and C supplementation and risk of cancer in men: posttrial follow-up in the Physicians' Health Study II randomized trial. *Am J Clin Nutr* 2014; 100(3): 915-23.
32. N. Farina, D. Llewellyn, Mgekn Isaac, et al. Vitamin E for Alzheimer's dementia and mild cognitive impairment. *Cochrane Database of Syst Rev* 2017; (4).
33. H. J. Kim, E. Giovannucci, B. Rosner, et al. Longitudinal and secular trends in dietary supplement use: Nurses' Health Study and Health Professionals Follow-Up Study, 1986-2006. *J Acad Nutr Diet* 2014; 114(3): 436-43.
34. E. D. Kantor, C. D. Rehm, M. Du, et al. Giovannucci. Trends in Dietary Supplement Use Among US Adults From 1999-2012. *JAMA* 2016; 316(14): 1464-74.
35. A. W. K. Yeung, N. T. Tzvetkov, Osama S. El-Tawil, et al. Abdel-Daim, Atanas G. Atanasov. Antioxidants: Scientific Literature Landscape Analysis. *Oxid Med Cell Longev* 2019; 2019: 11.
36. H. Cui, Y. Kong, H. Zhang. Oxidative stress, mitochondrial dysfunction, and aging. *J Signal Transduct* 2012; 2012: 646354.
37. D. Harman. Aging: a theory based on free radical and radiation chemistry. *J Gerontol* 1956; 11(3): 298-300.
38. E. Cadenas, K. J. Davies. Mitochondrial free radical generation, oxidative stress, and aging. *Free Radic Biol Med* 2000; 29(3-4): 222-30.
39. S. I. Liochev. Reactive oxygen species and the free radical theory of aging. *Free Radic Biol Med* 2013; 60: 1-4.

40. J. F. Turrens. Mitochondrial formation of reactive oxygen species. *J Physiol* 2003; 552(Pt 2): 335-44.
41. T. Finkel, N. J. Holbrook. Oxidants, oxidative stress and the biology of ageing. *Nature* 2000; 408(6809): 239-47.
42. A. Y. Andreyev, Y. E. Kushnareva, A. A. Starkov. Mitochondrial metabolism of reactive oxygen species. *Biochemistry (Mosc)* 2005; 70(2): 200-14.
43. R. Castagna, J. P. Eiserich, M. S. Budamagunta, et al. Hydroxyl radical from the reaction between hypochlorite and hydrogen peroxide. *Atmos Environ* 2008; 42(26): 6551-4.
44. B. Halliwell. Free radicals, reactive oxygen species and human disease: a critical evaluation with special reference to atherosclerosis. *Br J Exp Pathol* 1989; 70(6): 737-57.
45. C. B. F. Andersen, K. Stodkilde, K. L. Saederup, et al. Haptoglobin. *Antioxid Redox Signal* 2017; 26(14): 814-31.
46. A. P. Levy, R. Asleh, S. Blum, et al. Haptoglobin: basic and clinical aspects. *Antioxid Redox Signal* 2010; 12(2): 293-304.
47. M. Ristow, K. Schmeisser. Mitohormesis: Promoting Health and Lifespan by Increased Levels of Reactive Oxygen Species (ROS). *Dose Response* 2014; 12(2): 288-341.
48. C. R. Reczek, N. S. Chandel. ROS-dependent signal transduction. *Curr Opin Cell Biol* 2015; 33: 8-13.
49. J. Zhang, X. Wang, V. Vikash, et al. ROS and ROS-Mediated Cellular Signaling. *Oxid Med Cell Longev* 2016; 2016: 4350965.
50. S. Hekimi, J. Lapointe, Y. Wen. Taking a "good" look at free radicals in the aging process. *Trends Cell Biol* 2011; 21(10): 569-76.
51. V. Urbanaviciute, H. Luo, C. Sjowall, et al. Low Production of Reactive Oxygen Species Drives Systemic Lupus Erythematosus. *Trends Mol Med* 2019 Oct;25(10):826-835
52. L. M. Olsson, Å. C. Johansson, B. Gullstrand, et al. A single nucleotide polymorphism in the *NCF1* gene leading to reduced oxidative burst is associated with systemic lupus erythematosus. *Ann Rheum Dis* 2017; 76(9): 1607.
53. T. Kelkka, D. Kienhöfer, M. Hoffmann, et al. Reactive Oxygen Species Deficiency Induces Autoimmunity with Type 1 Interferon Signature. *Antioxid Redox Signal* 2014; 21(16): 2231-45.
54. Y. C. Jang, V. I. Perez, W. Song, et al. Overexpression of Mn superoxide dismutase does not increase life span in mice. *J Gerontol A Biol Sci Med Sci* 2009; 64(11): 1114-25.
55. Y. Zhang, Y. Ikeno, W. Qi, et al. Mice deficient in both Mn superoxide dismutase and glutathione peroxidase-1 have increased oxidative damage and a greater incidence of pathology but no reduction in longevity. *J Gerontol A Biol Sci Med Sci* 2009; 64(12): 1212-20.
56. J. M. Van Raamsdonk, S. Hekimi. Deletion of the mitochondrial superoxide dismutase *sod-2* extends lifespan in *Caenorhabditis elegans*. *PLoS Genet* 2009; 5(2): e1000361.
57. D. Desjardins, B. Cacho-Valadez, J. L. Liu, et al. Antioxidants reveal an inverted U-shaped dose-response relationship between reactive oxygen species levels and the rate of aging in *Caenorhabditis elegans*. *Aging Cell* 2017; 16(1): 104-12.
58. H. Yin, L. Xu, N. A. Porter. Free radical lipid peroxidation: mechanisms and analysis. *Chem Rev* 2011; 111(10): 5944-72.
59. B. C. Sousa, A. R. Pitt, C. M. Spickett. Chemistry and analysis of HNE and other prominent carbonyl-containing lipid oxidation compounds. *Free Radic Biol Med* 2017; 111: 294-308.

60. C. M. Spickett. The lipid peroxidation product 4-hydroxy-2-nonenal: Advances in chemistry and analysis. *Redox Biol* 2013; 1(1): 145-52.
61. D. Il'yasova, P. Scarbrough, I. Spasojevic. Urinary biomarkers of oxidative status. *Clin. Chim. Acta* 2012; 413(19-20): 1446-53.
62. E. D. Brown, V. C. Morris, D. G. Rhodes, et al. Urinary malondialdehyde-equivalents during ingestion of meat cooked at high or low temperatures. *Lipids* 1995; 30(11): 1053-6.
63. G. L. Milne, E. S. Musiek, J. D. Morrow. F2-isoprostanes as markers of oxidative stress in vivo: an overview. *Biomarkers* 2005; 10 Suppl 1: S10-23.
64. P. Montuschi, P. J. Barnes, L. J. Roberts, 2nd. Isoprostanes: markers and mediators of oxidative stress. *Faseb j* 2004; 18(15): 1791-800.
65. N. K. Gopaul, B. Halliwell, E. E. Anggard. Measurement of plasma F2-isoprostanes as an index of lipid peroxidation does not appear to be confounded by diet. *Free Radic Res* 2000; 33(2): 115-27.
66. L. J. Roberts, J. D. Morrow. Measurement of F(2)-isoprostanes as an index of oxidative stress in vivo. *Free Radic Biol Med* 2000; 28(4): 505-13.
67. T. J. van't Erve, M. B. Kadiiska, S. J. London, et al. Classifying oxidative stress by F2-isoprostane levels across human diseases: A meta-analysis. *Redox Biol* 2017; 12: 582-99..
68. E. R. Stadtman. Protein oxidation and aging. *Free Radic Res* 2006; 40(12): 1250-8.
69. B. S. Berlett, E. R. Stadtman. Protein oxidation in aging, disease, and oxidative stress. *J Biol Chem* 1997; 272(33): 20313-6.
70. M. J. Davies. Protein oxidation and peroxidation. *Biochem J* 2016; 473(7): 805-25.
71. A. Hohn, T. Jung, T. Grune. Pathophysiological importance of aggregated damaged proteins. *Free Radic Biol Med* 2014; 71: 70-89.
72. K. J. Davies. Protein oxidation and proteolytic degradation. General aspects and relationship to cataract formation. *Adv Exp Med Biol* 1990; 264: 503-11.
73. E. R. Stadtman, R. L. Levine. Free radical-mediated oxidation of free amino acids and amino acid residues in proteins. *Amino Acids* 2003; 25(3-4): 207-18.
74. R. Singh, A. Barden, T. Mori, et al. Advanced glycation end-products: a review. *Diabetologia* 2001; 44(2): 129-46.
75. V. Witko-Sarsat, M. Friedlander, T. Nguyen Khoa, et al. Advanced oxidation protein products as novel mediators of inflammation and monocyte activation in chronic renal failure. *J Immunol* 1998; 161(5): 2524-32.
76. A. Trpkovic, I. Resanovic, J. Stanimirovic, et al. Oxidized low-density lipoprotein as a biomarker of cardiovascular diseases. *Crit Rev Clin Lab Sci* 2015; 52(2): 70-85.
77. S. Bartesaghi, G. Ferrer-Sueta, G. Peluffo, et al. Protein tyrosine nitration in hydrophilic and hydrophobic environments. *Amino Acids* 2007; 32(4): 501-15.
78. A. P. Grollman, M. Moriya. Mutagenesis by 8-oxoguanine: an enemy within. *Trends Genet* 1993; 9(7): 246-9.
79. T. Yan, S. E. Berry, A. B. Desai, et al. DNA mismatch repair (MMR) mediates 6-thioguanine genotoxicity by introducing single-strand breaks to signal a G2-M arrest in MMR-proficient RKO cells. *Clin Cancer Res* 2003; 9(6): 2327-34.
80. V. Dehennaut, I. Loison, M. Dubuissez, et al. DNA double-strand breaks lead to activation of hypermethylated in cancer 1 (HIC1) by SUMOylation to regulate DNA repair. *J Biol Chem* 2013; 288(15): 10254-64.

81. P. A. Johnson, P. Clements, K. Hudson, et al. The mitotic spindle and DNA damage-induced apoptosis. *Toxicol Lett* 2000; 112-113: 59-67.
82. M. Dizdaroglu, E. Gajewski, P. Reddy, et al. Structure of a hydroxyl radical induced DNA-protein cross-link involving thymine and tyrosine in nucleohistone. *Biochemistry* 1989; 28(8): 3625-8.
83. L. J. Marnett. Oxy radicals, lipid peroxidation and DNA damage. *Toxicology* 2002; 181-182: 219-22.
84. C. C. Winterbourn. Reconciling the chemistry and biology of reactive oxygen species. *Nat Chem Biol* 2008; 4(5): 278-86.
85. M. Dizdaroglu, P. Jaruga, M. Birincioglu, et al. Free radical-induced damage to DNA: mechanisms and measurement. *Free Radic Biol Med* 2002; 32(11): 1102-15.
86. A. P. Breen, J. A. Murphy. Reactions of oxyl radicals with DNA. *Free Radic Biol Med* 1995; 18(6): 1033-77.
87. J. Cadet, T. Douki, D. Gasparutto, et al. Oxidative damage to DNA: formation, measurement and biochemical features. *Mutat Res* 2003; 531(1-2): 5-23.
88. N. Chatterjee, G. C. Walker. Mechanisms of DNA damage, repair, and mutagenesis. *Environ Mol Mutagen* 2017; 58(5): 235-63.
89. S. Anderson, A. T. Bankier, B. G. Barrell, et al. Sequence and organization of the human mitochondrial genome. *Nature* 1981; 290: 457.
90. N. M. Druzhyna, G. L. Wilson, S. P. LeDoux. Mitochondrial DNA repair in aging and disease. *Mech Ageing Dev* 2008; 129(7): 383-90.
91. D. L. Croteau, R. H. Stierum, V. A. Bohr. Mitochondrial DNA repair pathways. *Mutat Res* 1999; 434(3): 137-48.
92. B. N. Ames, M. K. Shigenaga, Tory M. Hagen. Mitochondrial decay in aging. *Biochim Biophys* 1995; 1271(1): 165-70.
93. T. Ozawa, M. Tanaka, W. Sato, et al. Mitochondrial DNA Mutations as an Etiology of Human Degenerative Diseases. In: Kim CH, Ozawa T, eds. *Bioenergetics: Molecular Biology, Biochemistry, and Pathology*. Boston, MA: Springer US; 1990: 413-27.
94. J. B. Stewart, P. F. Chinnery. The dynamics of mitochondrial DNA heteroplasmy: implications for human health and disease. *Nat Rev Genet* 2015; 16(9): 530-42.
95. D. C. Wallace, D. Chalkia. Mitochondrial DNA genetics and the heteroplasmy conundrum in evolution and disease. *Cold Spring Harb Perspect Biol* 2013; 5(11): a021220.
96. M. T. Lin, M. F. Beal. Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature* 2006; 443(7113): 787-95.
97. D. C. Wallace. A mitochondrial bioenergetic etiology of disease. *J Clin Invest* 2013; 123(4): 1405-12.
98. A. T. Kraja, C. Liu, J. L. Fetterman, et al. Associations of Mitochondrial and Nuclear Mitochondrial Variants and Genes with Seven Metabolic Traits. *Am J Hum Genet* 2019; 104(1): 112-38.
99. L. Guarente. Sirtuins, Aging, and Medicine. *N Engl J Med* 2011; 364(23): 2235-44.
100. M. N. Sack, T. Finkel. Mitochondrial metabolism, sirtuins, and aging. *Cold Spring Harb Perspect Biol* 2012; 4(12), pii: a013102.
101. S. I. Imai, L. Guarente. It takes two to tango: NAD(+) and sirtuins in aging/longevity control. *NPJ Aging Mech Dis* 2016; 2: 16017.

102. C. H. Westphal, M. A. Dipp, L. Guarente. A therapeutic role for sirtuins in diseases of aging? *Trends Biochem Sci* 2007; 32(12): 555-60.
103. C. K. Singh, G. Chhabra, M. A. Ndiaye, et al. The Role of Sirtuins in Antioxidant and Redox Signaling. *Antioxid Redox Signal* 2018; 28(8): 643-61.
104. W. Yang, K. Nagasawa, C. Münch, et al. Mitochondrial Sirtuin Network Reveals Dynamic SIRT3-Dependent Deacetylation in Response to Membrane Depolarization. *Cell* 2016; 167(4): 985-1000.e21.
105. S. M. Jeong, C. Xiao, L. W. S. Finley, et al. SIRT4 Has Tumor-Suppressive Activity and Regulates the Cellular Metabolic Response to DNA Damage by Inhibiting Mitochondrial Glutamine Metabolism. *Cancer Cell* 2013; 23(4): 450-63.
106. L. Guarente. Mitochondria--a nexus for aging, calorie restriction, and sirtuins? *Cell* 2008; 132(2): 171-6.
107. W. Grabowska, E. Sikora, A. B. Zmijewska. Sirtuins, a promising target in slowing down the ageing process. *Biogerontology* 2017; 18(4): 447-76.
108. L. Guarente. Sirtuins, aging, and metabolism. *Cold Spring Harb Symp Quant Biol* 2011; 76: 81-90.
109. G. López-Lluch, P. Navas. Calorie restriction as an intervention in ageing. *The J Physiol* 2016; 594(8): 2043-60.
110. A. Zullo, E. Simone, M. Grimaldi, et al. Sirtuins as Mediator of the Anti-Ageing Effects of Calorie Restriction in Skeletal and Cardiac Muscle. *Int J Mol Sci* 2018; 19(4): 928.
111. L. Guarente. Calorie restriction and sirtuins revisited. *Genes Dev* 2013; 27(19): 2072-85.
112. G. Testa, F. Biasi, G. Poli, et al. Calorie restriction and dietary restriction mimetics: a strategy for improving healthy aging and longevity. *Curr Pharm Des* 2014; 20(18): 2950-77.
113. L. Fontana, L. Partridge, V. D. Longo. Extending healthy life span--from yeast to humans. *Science* 2010; 328(5976): 321-6.
114. D. K. Ingram, R. de Cabo. Calorie restriction in rodents: Caveats to consider. *Ageing Res Rev* 2017; 39: 15-28.
115. J. Most, V. Tosti, L. M. Redman, et al. Calorie restriction in humans: An update. *Ageing Res Rev* 2017; 39: 36-45.
116. L. K. Heilbronn, E. Ravussin. Calorie restriction and aging: review of the literature and implications for studies in humans. *Am J Clin Nutr* 2003; 78(3): 361-9.
117. L. K. Heilbronn, L. de Jonge, M. I. Frisard, et al. Effect of 6-month calorie restriction on biomarkers of longevity, metabolic adaptation, and oxidative stress in overweight individuals: a randomized controlled trial. *JAMA* 2006; 295(13): 1539-48.
118. D. Lettieri-Barbato, E. Giovannetti, K. Aquilano. Effects of dietary restriction on adipose mass and biomarkers of healthy aging in human. *Ageing (Albany NY)* 2016; 8(12): 3341-55.
119. C. Y. Liao, B. K. Kennedy. Mouse models and aging: longevity and progeria. *Curr Top Dev Biol* 2014; 109: 249-85.
120. H. Liang, E. J. Masoro, J. F. Nelson, et al. Genetic mouse models of extended lifespan. *Exp Gerontol* 2003; 38(11-12): 1353-64.
121. A. B. Salmon, A. Richardson, V. I. Perez. Update on the oxidative stress theory of aging: does oxidative stress play a role in aging or healthy aging? *Free Radic Biol Med* 2010; 48(5): 642-55.

122. Y. Chen, C. Wu, C. Kao, et al. Longevity and lifespan control in mammals: Lessons from the mouse. *Ageing Res Rev* 2010; 9: S28-S35.
123. C. Selman, D. J. Withers. Mammalian models of extended healthy lifespan. *Philos Trans R Soc Lond B Biol Sci* 2011; 366(1561): 99-107.
124. D. F. Dai, Y. A. Chiao, G. M. Martin, et al. Mitochondrial-Targeted Catalase: Extended Longevity and the Roles in Various Disease Models. *Prog Mol Biol Transl Sci* 2017; 146: 203-41.
125. E. Migliaccio, M. Giorgio, S. Mele, et al. The p66shc adaptor protein controls oxidative stress response and life span in mammals. *Nature* 1999; 402(6759): 309-13.
126. R. K. Junnila, E. O. List, D. E. Berryman, et al. The GH/IGF-1 axis in ageing and longevity. *Nat Rev Endocrinol* 2013; 9(6): 366-76.
127. V. I. Perez, H. Van Remmen, A. Bokov, et al. The overexpression of major antioxidant enzymes does not extend the lifespan of mice. *Aging Cell* 2009; 8(1): 73-5.
128. H. Van Remmen, Y. Ikeno, M. Hamilton, et al. Life-long reduction in MnSOD activity results in increased DNA damage and higher incidence of cancer but does not accelerate aging. *Physiol Genomics* 2003; 16(1): 29-37.
129. G. M. Cunningham, L. C. Flores, M. G. Roman, et al. Thioredoxin overexpression in both the cytosol and mitochondria accelerates age-related disease and shortens lifespan in male C57BL/6 mice. *Geroscience* 2018; 40(5-6): 453-68.
130. V. I. Perez, L. A. Cortez, C. M. Lew, et al. Thioredoxin 1 overexpression extends mainly the earlier part of life span in mice. *J Gerontol A Biol Sci Med Sci* 2011; 66(12): 1286-99.
131. J. Moskovitz, S. Bar-Noy, W. M. Williams, et al. Methionine sulfoxide reductase (MsrA) is a regulator of antioxidant defense and lifespan in mammals. *Proc Natl Acad Sci U S A* 2001; 98(23): 12920-5.
132. A. B. Salmon, V. I. Perez, A. Bokov, et al. Lack of methionine sulfoxide reductase A in mice increases sensitivity to oxidative stress but does not diminish life span. *Faseb j* 2009; 23(10): 3601-8.
133. R. Doonan, J. J. McElwee, F. Matthijssens, et al. Against the oxidative damage theory of aging: superoxide dismutases protect against oxidative stress but have little or no effect on life span in *Caenorhabditis elegans*. *Genes Dev* 2008; 22(23): 3236-41.
134. W. Yang, J. Li, S. Hekimi. A Measurable increase in oxidative damage due to reduction in superoxide detoxification fails to shorten the life span of long-lived mitochondrial mutants of *Caenorhabditis elegans*. *Genetics* 2007; 177(4): 2063-74.
135. C. Yee, W. Yang, S. Hekimi. The Intrinsic Apoptosis Pathway Mediates the Pro-Longevity Response to Mitochondrial ROS in *C.elegans*. *Cell* 2014; 157(4): 897-909.
136. M. Uno, E. Nishida. Lifespan-regulating genes in *C. elegans*. *NPJ Aging Mech Dis* 2016; 2: 16010.
137. T. Oberacker, J. Bajorat, S. Ziola, et al. Enhanced expression of thioredoxin-interacting-protein regulates oxidative DNA damage and aging. *FEBS Letters* 2018; 592(13): 2297-307.
138. I. Reveillaud, J. Phillips, B. Duyf, et al. Phenotypic rescue by a bovine transgene in a Cu/Zn superoxide dismutase-null mutant of *Drosophila melanogaster*. *Mol Cell Biol* 1994; 14(2): 1302-7.
139. W. Yang, S. Hekimi. A mitochondrial superoxide signal triggers increased longevity in *Caenorhabditis elegans*. *PLoS Biol* 2010; 8(12): e1000556.

140. A. B. Hwang, E. A. Ryu, M. Artan, et al. Feedback regulation via AMPK and HIF-1 mediates ROS-dependent longevity in *Caenorhabditis elegans*. *Proc Natl Acad Sci U S A* 2014; 111(42): E4458.
141. I. Sadowska-Bartosz, G. Bartosz. Effect of antioxidants supplementation on aging and longevity. *Biomed Res Int* 2014; 2014: 404680.
142. A. Sanz. Mitochondrial reactive oxygen species: Do they extend or shorten animal lifespan? *Biochim Biophys Acta* 2016; 1857(8): 1116-26.
143. A. Daiber, F. Di Lisa, M. Oelze, et al. Crosstalk of mitochondria with NADPH oxidase via reactive oxygen and nitrogen species signalling and its role for vascular function. *Br J Pharmacol* 2017; 174(12): 1670-89.
144. A. Daiber. Redox signaling (cross-talk) from and to mitochondria involves mitochondrial pores and reactive oxygen species. *Biochim Biophys Acta* 2010; 1797(6-7): 897-906.
145. A. J. Kattoor, N. V. K. Pothineni, D. Palagiri, et al. Oxidative Stress in Atherosclerosis. *Curr Atheroscler Rep* 2017; 19(11): 42.
146. S. Chrissobolis, Q. N. Dinh, G. R. Drummond, et al. Role of Oxidative Stress in Hypertension. In: Rodriguez-Porcel M, Chade AR, Miller JD, eds. *Studies on Atherosclerosis*. Boston, MA: Springer US; 2017: 59-78.
147. T. Munzel, G. G. Camici, C. Maack, et al. Impact of Oxidative Stress on the Heart and Vasculature: Part 2 of a 3-Part Series. *J Am Coll Cardiol* 2017; 70(2): 212-29.
148. S. Steven, A. Daiber, J. F. Doppeide, et al. Peripheral artery disease, redox signaling, oxidative stress - Basic and clinical aspects. *Redox Biol* 2017; 12: 787-97.
149. U. Forstermann, N. Xia, H. Li. Roles of Vascular Oxidative Stress and Nitric Oxide in the Pathogenesis of Atherosclerosis. *Circ Res* 2017; 120(4): 713-35.
150. T. J. Guzik, J. Sadowski, B. Guzik, et al. Coronary artery superoxide production and nox isoform expression in human coronary artery disease. *Arterioscler Thromb Vasc Biol* 2006; 26(2): 333-9.
151. F. N. Ashar, Y. Zhang, R. J. Longchamps, et al. Association of Mitochondrial DNA Copy Number With Cardiovascular Disease. *JAMA Cardiol* 2017; 2(11): 1247-55.
152. Z. Ye, H. Song. Antioxidant vitamins intake and the risk of coronary heart disease: meta-analysis of cohort studies. *Eur J Cardiovasc Prev Rehabil* 2008; 15(1): 26-34.
153. H. N. Siti, Y. Kamisah, J. Kamsiah. The role of oxidative stress, antioxidants and vascular inflammation in cardiovascular disease (a review). *Vascul Pharmacol* 2015; 71: 40-56.
154. T. Gori, T. Munzel. Oxidative stress and endothelial dysfunction: therapeutic implications. *Ann Med* 2011; 43(4): 259-72.
155. H. Boeing, M. Gottschald, S. Dietrich, et al. Dietary Supplements and Risk of Cause-Specific Death, Cardiovascular Disease, and Cancer: A Systematic Review and Meta-Analysis of Primary Prevention Trials. *Adv Nutr* 2017; 8(1): 27-39.
156. T. Wang, L. Xu. Circulating Vitamin E Levels and Risk of Coronary Artery Disease and Myocardial Infarction: A Mendelian Randomization Study. *Nutrients* 2019; 11(9), 2153(9).
157. M. T. Islam. Oxidative stress and mitochondrial dysfunction-linked neurodegenerative disorders. *Neurol Res* 2017; 39(1): 73-82.
158. A. C. Rego, C. R. Oliveira. Mitochondrial dysfunction and reactive oxygen species in excitotoxicity and apoptosis: implications for the pathogenesis of neurodegenerative diseases. *Neurochem Res* 2003; 28(10): 1563-74.

159. C. Cheignon, M. Tomas, D. Bonnefont-Rousselot, et al. Oxidative stress and the amyloid beta peptide in Alzheimer's disease. *Redox Biol* 2018; 14: 450-64.
160. L. Sansone, V. Reali, L. Pellegrini, et al. SIRT1 silencing confers neuroprotection through IGF-1 pathway activation. *J Cell Physiol* 2013; 228(8): 1754-61..
161. A. Satoh, S. I. Imai, L. Guarente. The brain, sirtuins, and ageing. *Nat Rev Neurosci* 2017; 18: 362.
162. Y. Zhao, B. Zhao. Oxidative stress and the pathogenesis of Alzheimer's disease. *Oxid Med Cell Longev* 2013; 2013: 316523.
163. Z. Liu, T. Zhou, A. C. Ziegler, et al. Oxidative Stress in Neurodegenerative Diseases: From Molecular Mechanisms to Clinical Applications. *Oxid Med Cell Longev* 2017; 2017: 2525967.
164. P. E. Coskun, M. F. Beal, D. C. Wallace. Alzheimer's brains harbor somatic mtDNA control-region mutations that suppress mitochondrial transcription and replication. *Proc Natl Acad Sci U S A* 2004; 101(29): 10726-31.
165. R. Betarbet, T. B. Sherer, G. MacKenzie, et al. Chronic systemic pesticide exposure reproduces features of Parkinson's disease. *Nat Neurosci* 2000; 3(12): 1301-6.
166. F. Fornai, O. M. Schluter, P. Lenzi, et al. Parkinson-like syndrome induced by continuous MPTP infusion: convergent roles of the ubiquitin-proteasome system and alpha-synuclein. *Proc Natl Acad Sci U S A* 2005; 102(9): 3413-8.
167. T. Jiang, Q. Sun, S. Chen. Oxidative stress: A major pathogenesis and potential therapeutic target of antioxidative agents in Parkinson's disease and Alzheimer's disease. *Prog Neurobiol* 2016; 147: 1-19.
168. E. Niedzielska, I. Smaga, M. Gawlik, et al. Oxidative Stress in Neurodegenerative Diseases. *Mol Neurobiol* 2016; 53(6): 4094-125.
169. D. A. Butterfield, M. L. Bader Lange, R. Sultana. Involvements of the lipid peroxidation product, HNE, in the pathogenesis and progression of Alzheimer's disease. *Biochim Biophys Acta* 2010; 1801(8): 924-9.
170. F. Coppede, L. Migliore. DNA damage in neurodegenerative diseases. *Mutat Res* 2015; 776: 84-97.
171. G. H. Kim, J. E. Kim, S. J. Rhie, S. Yoon. The Role of Oxidative Stress in Neurodegenerative Diseases. *Exp Neurobiol* 2015; 24(4): 325-40.
172. E. Khusnutdinova, I. Gilyazova, E. Ruiz-Pesini, et al. A Mitochondrial Etiology of Neurodegenerative Diseases: Evidence from Parkinson's Disease. *Ann N Y Acad Sci* 2008; 1147(1): 1-20.
173. E. Mariani, M. C. Polidori, A. Cherubini, et al. Oxidative stress in brain aging, neurodegenerative and vascular diseases: an overview. *J Chromatogr B Analyt Technol Biomed Life Sci* 2005; 827(1): 65-75.
174. J. Blesa, I. Trigo-Damas, A. Quiroga-Varela, et al. Oxidative stress and Parkinson's disease. *Front Neuroanat* 2015 Jul 8; 9(91).
175. M. C. Morris, L. A. Beckett, P. A. Scherr, et al. Vitamin E and vitamin C supplement use and risk of incident Alzheimer disease. *Alzheimer Dis Assoc Disord* 1998; 12(3): 121-6.
176. M. C. Morris, D. A. Evans, J. L. Bienias, et al. Dietary intake of antioxidant nutrients and the risk of incident Alzheimer disease in a biracial community study. *JAMA* 2002; 287(24): 3230-7.

177. P. P. Zandi, J. C. Anthony, A. S. Khachaturian, et al. Reduced risk of Alzheimer disease in users of antioxidant vitamin supplements: the Cache County Study. *Arch Neurol* 2004; 61(1): 82-8.
178. S. M. Zhang, M. A. Hernan, H. Chen, et al. Intakes of vitamins E and C, carotenoids, vitamin supplements, and PD risk. *Neurology* 2002; 59(8): 1161-9.
179. M. J. Engelhart, M. I. Geerlings, A. Ruitenberg, et al. Dietary intake of antioxidants and risk of Alzheimer disease. *JAMA* 2002; 287(24): 3223-9.
180. J. A. Luchsinger, M. X. Tang, S. Shea, et al. Antioxidant vitamin intake and risk of Alzheimer disease. *Arch Neurol* 2003; 60(2): 203-8.
181. K. Yaffe, T. E. Clemons, W. L. McBee, et al. Impact of antioxidants, zinc, and copper on cognition in the elderly: a randomized, controlled trial. *Neurology* 2004; 63(9): 1705-7.
182. V. Boccardi, M. Baroni, F. Mangialasche, et al. Vitamin E family: Role in the pathogenesis and treatment of Alzheimer's disease. *Alzheimers Dement* 2016; 2(3): 182-91.
183. M. C. de Wilde, B. Vellas, E. Girault, et al. Lower brain and blood nutrient status in Alzheimer's disease: Results from meta-analyses. *Alzheimers Dement* 2017; 3(3): 416-31.
184. S. Lopes da Silva, B. Vellas, S. Elemans, et al. Plasma nutrient status of patients with Alzheimer's disease: Systematic review and meta-analysis. *Alzheimers Dement* 2014; 10(4): 485-502.
185. A. Gugliandolo, P. Bramanti, E. Mazzon. Role of Vitamin E in the Treatment of Alzheimer's Disease: Evidence from Animal Models. *Int J Mol Sci* 2017; 18(12).
186. A. W. S. Rutjes, D. A. Denton, M. Di Nisio, et al. Vitamin and mineral supplementation for maintaining cognitive function in cognitively healthy people in mid and late life. *Cochrane Database Syst Rev* 2018; (12).
187. J. McCleery, R. P. Abraham, D. A. Denton, et al. Vitamin and mineral supplementation for preventing dementia or delaying cognitive decline in people with mild cognitive impairment. *Cochrane Database Syst Rev* 2018; (11).
188. Z. Zhu, M. Sun, W. Zhang, et al. The efficacy and safety of coenzyme Q10 in Parkinson's disease: a meta-analysis of randomized controlled trials. *Neurol Sci* 2017; 38(2): 215-24.
189. A. Negida, A. Menshawy, G. El Ashal, et al. Coenzyme Q10 for Patients with Parkinson's Disease: A Systematic Review and Meta-Analysis. *CNS Neurol Disord Drug Targets* 2016; 15(1): 45-53.
190. P. Mecocci, V. Boccardi, R. Cecchetti, et al. A Long Journey into Aging, Brain Aging, and Alzheimer's Disease Following the Oxidative Stress Tracks. *J Alzheimers Dis* 2018; 62(3): 1319-35.
191. P. Mecocci, M. C. Polidori. Antioxidant clinical trials in mild cognitive impairment and Alzheimer's disease. *Biochim Biophys Acta* 2012; 1822(5): 631-8.
192. M. C. Polidori, G. Nelles. Antioxidant clinical trials in mild cognitive impairment and Alzheimer's disease - challenges and perspectives. *Curr Pharm Des* 2014; 20(18): 3083-92.
193. G. Liu, Y. Zhao, S. Jin, et al. Circulating vitamin E levels and Alzheimer's disease: a Mendelian randomization study. *Neurobiol Aging* 2018; 72: 189.e1-e9.
194. D. M. Williams, S. Hägg, N. L. Pedersen. Circulating antioxidants and Alzheimer disease prevention: a Mendelian randomization study. *Am J Clin Nutr* 2018; 109(1): 90-8.
195. G. J. Brewer. Why vitamin E therapy fails for treatment of Alzheimer's disease. *J Alzheimers Dis* 2010; 19(1): 27-30.

196. S. R. Steinhubl. Why have antioxidants failed in clinical trials? *Am J Cardiol* 2008; 101(10a): 14d-9d.
197. I. Robinson, D. G. de Serna, A. Gutierrez, et al. Vitamin E in humans: an explanation of clinical trial failure. *Endocr Pract* 2006; 12(5): 576-82.
198. M. P. Murphy. Antioxidants as therapies: can we improve on nature? *Free Radic Biol Med* 2014; 66: 20-3.
199. A. Munteanu, J. M. Zingg, A. Azzi. Anti-atherosclerotic effects of vitamin E--myth or reality? *J Cell Mol Med* 2004; 8(1): 59-76.
200. M. Vardi, N. S. Levy, A. P. Levy. Vitamin E in the prevention of cardiovascular disease: the importance of proper patient selection. *J Lipid Res* 2013; 54(9): 2307-14.
201. J. P. Seferovic, M. Asanin, P. M. Seferovic. Haptoglobin and haptoglobin genotypes in diabetes: A silver bullet to identify the responders to antioxidant therapy? *Eur J Prev Cardiol* 2018; 25(14): 1498-501.
202. I. Hochberg, E. M. Berinstein, U. Milman, et al. Interaction Between the Haptoglobin Genotype and Vitamin E on Cardiovascular Disease in Diabetes. *Curr Diab Rep* 2017; 17(6): 42.
203. R. Asleh, A. Briasoulis, E. M. Berinstein, et al. Meta-analysis of the association of the haptoglobin genotype with cardiovascular outcomes and the pharmacogenomic interactions with vitamin E supplementation. *Pharmgenomics Pers Med* 2018; 11: 71-82.
204. B. A. Banini, S. C. Cazanave, K. P. Yates, et al. Haptoglobin 2 Allele is Associated With Histologic Response to Vitamin E in Subjects With Nonalcoholic Steatohepatitis. *J Clin Gastroenterol* 2018; 53(10):750-758.
205. A. Azzi. Many tocopherols, one vitamin E. *Mol Aspects Med* 2018; 61: 92-103.
206. M. G. Traber, A. Elsner, R. Brigelius-Flohé. Synthetic as compared with natural vitamin E is preferentially excreted as α -CEHC in human urine: studies using deuterated α -tocopheryl acetates. *FEBS Letters* 1998; 437(1): 145-8.
207. L. J. Roberts 2nd, J. A. Oates, M. F. Linton, et al. The relationship between dose of vitamin E and suppression of oxidative stress in humans. *Free Radic Biol Med* 2007; 43(10): 1388-93.
208. A. M Davies, A. Holt. Why antioxidant therapies have failed in clinical trials. *J Theor Biol* 2018; 457: 1-5.
209. N. G. Stephens, A. Parsons, P. M. Schofield, et al. Randomised controlled trial of vitamin E in patients with coronary disease: Cambridge Heart Antioxidant Study (CHAOS). *Lancet* 1996; 347(9004): 781-6.
210. A. Kontush, B. Finckh, B. Karten, et al. Antioxidant and prooxidant activity of alpha-tocopherol in human plasma and low density lipoprotein. *J Lipid Res* 1996; 37(7): 1436-48.
211. E. Niki. Evidence for beneficial effects of vitamin E. *Korean J Intern Med* 2015; 30(5): 571-9.
212. M. P. Murphy, A. Holmgren, N. G. Larsson, et al. Unraveling the biological roles of reactive oxygen species. *Cell Metab* 2011; 13(4): 361-6.
213. H. E. Roxborough, G. W. Burton, F. J. Kelly. Inter- and intra-individual variation in plasma and red blood cell vitamin E after supplementation. *Free Radic Res* 2000; 33(4): 437-45.
214. P. Torquato, O. Ripa, D. Giusepponi, et al. Analytical strategies to assess the functional metabolome of vitamin E. *J Pharm Biomed Anal* 2016; 124: 399-412.
215. M. Hacquebard, Y. A. Carpentier. Vitamin E: absorption, plasma transport and cell uptake. *Curr Opin Clin Nutr Metab Care* 2005; 8(2): 133-8.

216. B. Shukitt-Hale, G. Cao, J. A. Joseph, et al. Effect of Fruits, Vegetables, or Vitamin E-Rich Diet on Vitamins E and C Distribution in Peripheral and Brain Tissues: Implications for Brain Function. *J Gerontol A Biol Sci Med Sci* 2000; 55(3): B144-B51.
217. M. L. Bartoli, F. Novelli, F. Costa, et al. Malondialdehyde in exhaled breath condensate as a marker of oxidative stress in different pulmonary diseases. *Mediators Inflamm* 2011; 2011: 891752.
218. S. Dalleau, M. Baradat, F. Guéraud, et al. Cell death and diseases related to oxidative stress: 4-hydroxynonenal (HNE) in the balance. *Cell Death Differ* 2013; 20: 1615.
219. N. A. Strobel, R. G. Fassett, S. A. Marsh, et al. Oxidative stress biomarkers as predictors of cardiovascular disease. *Int J Cardiol* 2011; 147(2): 191-201.
220. J. P. Fessel, N. A. Porter, K. P. Moore, et al. Discovery of lipid peroxidation products formed in vivo with a substituted tetrahydrofuran ring (isofurans) that are favored by increased oxygen tension. *Proc Natl Acad Sci U S A* 2002; 99(26): 16713-8.
221. F. Galli, A. Azzi, M. Birringer, et al. Vitamin E: Emerging aspects and new directions. *Free Radic Biol Med* 2017; 102: 16-36.
222. E. Herrera, C. Barbas. Vitamin E: action, metabolism and perspectives. *J Physiol Biochem* 2001; 57(1): 43-56.
223. R. Brigelius-Flohe, M. G. Traber. Vitamin E: function and metabolism. *Faseb j* 1999; 13(10): 1145-55.
224. G. Sharma, D. P. Muller, S. M. O'Riordan, et al. Urinary conjugated α -tocopheronolactone—a biomarker of oxidative stress in children with type 1 diabetes. *Free Radic Biol Med* 2013; 55: 54-62.
225. X. Niu, V. Zammit, J. M. Upston, et al. Coexistence of oxidized lipids and alpha-tocopherol in all lipoprotein density fractions isolated from advanced human atherosclerotic plaques. *Arterioscler Thromb Vasc Biol* 1999; 19(7): 1708-18.
226. G. Ravaglia, P. Forti, A. Lucicesare, et al. Plasma tocopherols and risk of cognitive impairment in an elderly Italian cohort. *Am J Clin Nutr* 2008; 87(5): 1306-13.
227. P. Torquato, D. Bartolini, D. Giusepponi, et al. Increased plasma levels of the lipoperoxyl radical-derived vitamin E metabolite alpha-tocopheryl quinone are an early indicator of lipotoxicity in fatty liver subjects. *Free Radic Biol Med* 2019; 131: 115-25.
228. S. A. Pope, P. T. Clayton, D. P. Muller. A new method for the analysis of urinary vitamin E metabolites and the tentative identification of a novel group of compounds. *Arch Biochem Biophys* 2000; 381(1): 8-15.
229. S. Gayatri, D. Muller, S. O'Riordan, et al. A novel method for the direct measurement of urinary conjugated metabolites of α -tocopherol and its use in diabetes. *Mol Nutr Food Res* 2010; 54(5): 599-600.
230. A. A. El-Aal, E. A. A. El-Ghffar, A. A. Ghali, M. R. Zughbur, M. M. Sirdah. The effect of vitamin C and/or E supplementations on type 2 diabetic adult males under metformin treatment: A single-blinded randomized controlled clinical trial. *Diabetes Metab Syndr* 2018; 12(4): 483-9.
231. K. T. Hall, J. E. Buring, K. J. Mukamal, et al. COMT and Alpha-Tocopherol Effects in Cancer Prevention: Gene-Supplement Interactions in Two Randomized Clinical Trials. *J Natl Cancer Inst* 2019.

