



Universiteit
Leiden
The Netherlands

Large-scale neural recordings call for new insights to link brain and behavior

Urai, A.E.; Doiron, B.; Leifer, A.M.; Churchland, A.K.

Citation

Urai, A. E., Doiron, B., Leifer, A. M., & Churchland, A. K. (2022). Large-scale neural recordings call for new insights to link brain and behavior. *Nature Neuroscience*, 25(1), 11-19. doi:10.1038/s41593-021-00980-9

Version: Publisher's Version

License: [Licensed under Article 25fa Copyright Act/Law \(Amendment Taverne\)](#)

Downloaded from: <https://hdl.handle.net/1887/3443849>

Note: To cite this publication please use the final published version (if applicable).



Large-scale neural recordings call for new insights to link brain and behavior

Anne E. Urai^{1,2}, Brent Doiron³, Andrew M. Leifer⁴ and Anne K. Churchland^{1,5} ✉

Neuroscientists today can measure activity from more neurons than ever before, and are facing the challenge of connecting these brain-wide neural recordings to computation and behavior. In the present review, we first describe emerging tools and technologies being used to probe large-scale brain activity and new approaches to characterize behavior in the context of such measurements. We next highlight insights obtained from large-scale neural recordings in diverse model systems, and argue that some of these pose a challenge to traditional theoretical frameworks. Finally, we elaborate on existing modeling frameworks to interpret these data, and argue that the interpretation of brain-wide neural recordings calls for new theoretical approaches that may depend on the desired level of understanding. These advances in both neural recordings and theory development will pave the way for critical advances in our understanding of the brain.

Our understanding of how the nervous system controls behavior closely trails our ability to precisely measure its core components—the activity of groups of neurons. Recent years have seen an explosion in large-scale neural recordings during animal behavior, opening up new ways to measure and understand network-level neural codes for cognition in diverse species. In the present review, we highlight how advances in technology have enabled this progress, we reveal pitfalls and promises and we explain new analysis approaches and theoretical tools being developed to understand the vast quantities of data now being collected. We focus on technologies for recording the activity of individual neurons. Although measurements of neural mass signals (such as local field potentials, closely related scalp-level electrophysiological signals measured by electroencephalography and magnetoencephalography, widefield calcium imaging, fiber photometry, magnetic resonance imaging and functional ultrasound) have enabled advancements in studying large-scale brain networks, these techniques are beyond the scope of the present review. We also exclude measurement of non-neural brain cells (such as glia) and extracellular signaling molecules (for example, neuromodulators), as well as perturbation techniques such as optogenetics.

Each neuron does not act independently: the importance of not just studying neurons in isolation, but rather understanding simultaneous recordings of pairs of neurons, has been appreciated since the early days of neural recordings¹. Some theoretical frameworks argued that correlations among neurons limited the information that a neural population could encode^{2,3}, whereas others emphasized the importance of the neural cell assembly as a substrate for memory⁴ and stimulus processing⁵. Consequently, there has been a concerted effort to understand the mechanistic origin⁶ and computational role⁷ of correlated fluctuations in neuronal population activity. The importance of understanding these interactions grows with the increasing sizes of simultaneously measured neural populations.

For decades, however, technical constraints limited many experiments to simultaneous recordings from only a few cells. In practice, this meant that these neurons were usually hand-selected to respond strongly to experimenter-defined variables, such as visual

motion or contrast. Many influential frameworks to understand neural computation then relied heavily on the neuron as a single unit, aiming to extrapolate or infer its role in local and long-range circuits^{8–10}. In parallel, pioneering work in neuronal circuit models focused on capturing single-neuron statistics such as firing rates¹¹ or spiking variability^{12–14}.

With the increasing throughput of simultaneous recordings in the 1990s and 2000s (Box 1) came the (accurate) anticipation that large-scale recordings would speed up experiments and boost their statistical power. They also reduced the focus on hand-picked neurons and brain areas with well-characterized responses. What was less expected were the large changes in theoretical focus and a new depth of understanding. Big questions about how neural activity and behavior relate to each other are beginning to be within reach. For instance, how are neural representations distributed across brain areas and cell types? How do signals connected to task-related computations interact with signals related to other brain functions, such as movements and arousal? And how much of neural variability is truly stochastic ‘noise’, as opposed to a reflection of signals coming from other neurons, brain areas or behaviors that we couldn’t measure before? Some skepticism is warranted as well: what have we learned from these advances in larger-scale recordings and behavioral characterization, especially in small animals that allow for whole-brain recordings? Will large-scale recordings deliver the promise of new insights, when many neurons in such recordings are unresponsive? And, finally, where do current theoretical frameworks explain large-scale neural data, and where do they fall short?

In the present review, we aim to address these questions. We review how technological developments have brought increasing experimental throughput, allow large-scale surveys of neural responses across previously understudied brain areas and are prompting new developments in studying information flow within and across brain areas. We then discuss how large-scale recordings have offered four unexpected insights (Fig. 1):

- Neural representations of sensory and cognitive variables are distributed and sparse, and can be dwarfed by movement signals.

¹Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, USA. ²Cognitive Psychology Unit, Leiden University, Leiden, The Netherlands.

³University of Chicago, Chicago, IL, USA. ⁴Princeton University, Princeton, NJ, USA. ⁵University of California Los Angeles, Los Angeles, CA, USA.

✉e-mail: ACHurchland@mednet.ucla.edu

Box 1 | History and future of large-scale neural recordings

Can we ever expect to record all neurons in the brain simultaneously? The answer depends on the size and physical properties of the brain in question. Simultaneous whole-brain measurements of single-neuron activity have been acquired in small, transparent animals, notably *Caenorhabditis elegans*¹¹², larval zebrafish¹¹³, hydra¹¹⁴ and, perhaps soon, *Drosophila*^{115,116}. In mammals, electrophysiological recordings across all cortical neurons have not been achieved but may be possible in principle^{117,118}.

In 2011, Stevenson and Kording¹¹⁹ proposed a ‘Moore’s law’ for neural recordings, predicting a doubling of simultaneously recorded neurons every ± 7.4 years. This prediction has been borne out, and calcium imaging methods have seen even faster increases than predicted. Yield from imaging, however, comes with trade-offs in temporal resolution, signal-to-noise ratio or imaging depth.

Exponential increases in recording ability offer exciting prospects for whole-brain imaging in larger brains, but we remain orders of magnitudes away from recording a sizable fraction of the mammalian brain. Extrapolation suggests that whole-brain, single-neuron recordings in mice may become a reality between two decades and a century from now.

Are whole-brain recordings necessary to understand all aspects of nervous system function? Whole-brain recordings in small animals may hold valuable lessons: in systems such as *C. elegans*, hydra and zebrafish, whole-brain recordings could be subsampled to assess the value of measuring a more and more complete set of nervous system activity.

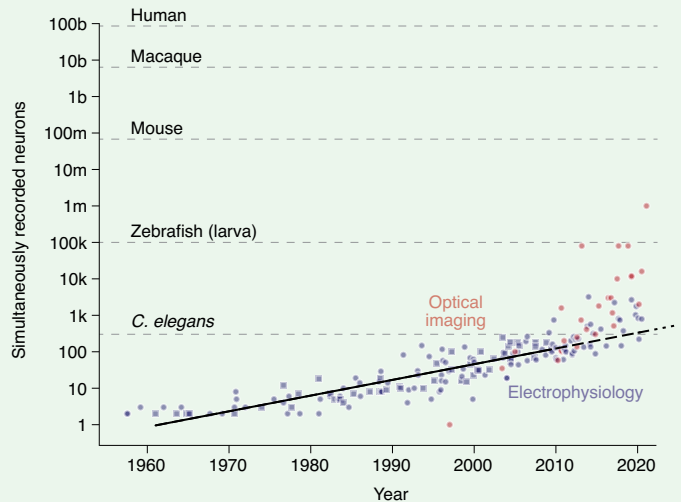
- Neural computations can be evident at the level of population dynamics but hidden at the level of single-neuron firing rates.
- Behaviorally relevant neural variance can often be explained by a small number of dimensions.
- Largely unstructured network architectures can drive highly structured responses.

We elaborate on existing theoretical frameworks for interpreting neural data, and how they are challenged by results from large-scale recordings. Finally, we speculate about future experimental and theoretical developments, and explore skepticism regarding the role of large-scale recording in helping us understand brain function.

Insights from large-scale neural recordings

Recent studies have leveraged high-yield recording modalities (Box 2) in the hope of gaining new insights into behavior and brain activity. The ability to record many neurons at once has increased statistical power, and reduced the number of required research animals. It has also shifted the focus from hand-selected neurons and brain areas toward unbiased, global surveys of neural responses and cell types. Large-scale recording techniques also increase the probability of encountering neurons with rare responses, capturing small signals that are distributed over many cells, or recording rare cell types. For instance, matching high-density electrophysiological responses to anatomically identified cells makes it possible to define the responses of sparse cell types in the retina with distinctive morphology¹⁵. Furthermore, leveraging the simultaneous nature of large-scale neural recordings has brought new insights, by shifting focus from single units to larger neural populations and brain regions.

Large-scale recording efforts in rodents have begun to generate new insights into the distributed and sparse nature of neural responses to task-relevant information (Fig. 1a). In one recent study, Steinmetz et al.¹⁶ used Neuropixels probes to measure the responses



Scaling up of neural recordings. Dark blue points: number of simultaneously recorded neurons using electrophysiology (squares indicate the original data used for the fit¹¹⁹). Red points: number of simultaneously recorded neurons using optical imaging (two-photon or light-sheet). Line: exponential fit to the original data. Black dashed and dotted lines: extrapolation to the present and future, respectively. Gray lines: approximate number of neurons in the brains of species that are commonly used in neuroscience. Figure available at https://github.com/anne-urai/largescale_recordings under a CC-BY license.

of around 30,000 neurons in mice reporting spatial judgments about visual stimuli. Choice signals were sparse, widely distributed and plentiful in deep structures that had thus far been overlooked in studies of decision-making (for example, the midbrain reticular nucleus). It is interesting that only a small fraction (~18%) of neurons in V1 were responsive to visual grating stimuli. By showing mice a larger battery of visual stimuli (including drifting gratings, Gabors and flashes), another Neuropixels study¹⁷ could identify receptive fields in up to 70% of V1 neurons. The recording method matters here: since spike-sorted electrophysiology is biased toward cells with large or frequent spikes, it is more likely to overestimate the fraction of neurons responsive to visual stimulation^{18,19}. Indeed, when de Vries et al.²⁰ used calcium imaging to measure the activity of ~60,000 neurons in passively viewing mice (viewing the same set of stimuli as used in ref. 17), they found that few neurons were driven by static, abstract stimuli, and many neurons were not responsive to visual stimuli at all. Although the activity of some neurons could be well predicted by their response to visual stimuli, many more cells responded in ways that could not be captured by existing models of cortical function. Although the presence of so-called ‘dark neurons’ (neurons that don’t fire at all²¹ or don’t fire in responses to experimental variables¹⁹) has been long known, results from large-scale recordings emphasize the importance of understanding their prevalence and functional role¹⁹.

Large-scale recordings also showed that neural activity correlated with animal movements, including idiosyncratic, task-unrelated ‘fidgets’, is stronger and more widely distributed than previously thought^{22–24} (Fig. 1a). Although it has long been known that running modulates activity in the visual cortex^{25–28}, recent work extended these findings in important ways. First, the impact of movements on single neurons was not restricted to a single area^{22–24}. Second, movement-driven modulation was present even in animals who were not just passively viewing, but were instead engaged in expert cognitive behaviors^{22,24}. Although the existence of movement-related activity in untrained, passively viewing animals

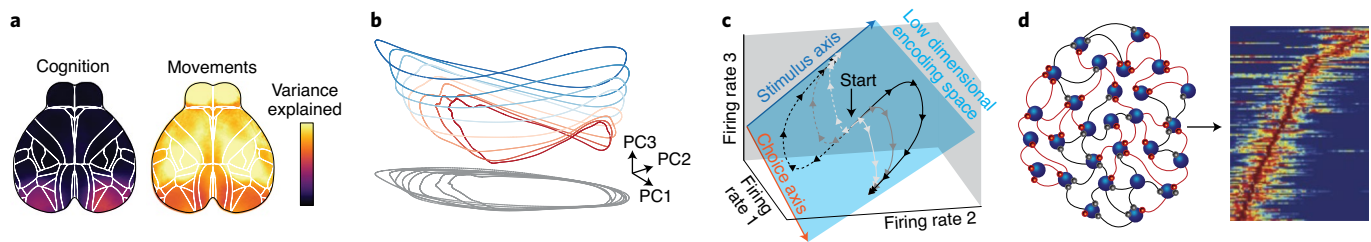


Fig. 1 | Insights from large-scale neural recordings. **a**, Neural representations of task/cognitive variables are distributed and sparse (left), and can be dwarfed by movement signals (right). Adapted from ref. ²². **b**, Neural computations can be evident in population dynamics but hidden at the level of single-neuron firing rates. Colors show neural responses in motor cortex during cycling at different speeds; each loop is once around a repeating cycle; blue is slowest. PC, principal component. Adapted from ref. ⁴⁶, CC BY. **c**, Behaviorally relevant neural variance is often explained by a small number of dimensions (blue, red axes). Adapted from ref. ³⁴, Springer Nature Ltd. **d**, Largely unstructured network architectures (left) drive highly structured neural responses (right). Adapted from ref. ⁵¹.

is perhaps unsurprising (after all, not much else is being asked of the brain in such circumstances), seeing movements dominate even in expert, engaged animals was unexpected: one would assume that task-related signals would dominate. Third, by using unsupervised video analysis, recent studies considered a far greater diversity of movements beyond running and pupil diameter^{22,23}, and evaluated these movements' roles in neural activity in a hypothesis-free way. This confirmed the importance of well-known movements such as pupil dilation, and also revealed previously ignored movements, such as hindlimb flexions and orofacial movements. The strong effect of facial movements on neural activity may indicate that these movements reflect animals' emotional states²⁹. An additional reason that orofacial and other movements are so widespread may be that many brain areas need to predict the sensory consequences of impending movements³⁰.

An anticipated challenge of large-scale recordings was the need to analyze them. What was not anticipated was that the increased neuron count would reveal that seemingly complex dynamics often reflect the sum of a small number of underlying motifs^{31,32}. Specifically, neural population recordings have uncovered that variance in neural activity, and specifically variance relevant for behavior, can often be accounted for by a small number of dimensions (Fig. 1c). For instance, a choice decoder built on the first principal component of neural activity in the higher visual cortex of monkeys performed almost as well as one built on the whole dataset³³. Low-dimensional activity is also apparent in the frontal cortex of monkeys performing complex tasks³⁴, in the premotor cortex of monkeys during reaching³⁵ and in *C. elegans* during fictive locomotion³⁶. In another example³⁷, researchers recorded the activity of about 150 neurons from the isolated nervous system of a medicinal leech. They used dimensionality reduction to identify an axis in neural state space along which the population's activity predicted behavioral responses to sensory stimulation (swimming or crawling) earlier in time than any single neuron. From the cells that strongly contributed to this population, they also identified one specific cell that could bias decisions toward crawling when electrically stimulated.

To what extent are neural codes truly low-dimensional (or merely appear that way)⁴⁰? Theoretical work suggests that higher-dimensional responses can be critical for computation and representation^{38,39}. In support of this, recent experimental work⁴¹ showed that the dimensional structure of neural responses in V1 allows for a code that balances efficiency with robustness to small perturbations in visual images.

Although the observations above benefited from large-scale recording technologies, some could in principle have been made with many neurons recorded sequentially. Simultaneous recordings additionally uncover the relationships among neurons: the way in

which their responses change together. For instance, internal states such as engagement tend to drive large fluctuations that are shared between many neurons: this variability would have looked like random trial-to-trial noise when recording one neuron at a time. Simultaneous large-scale neural recordings have enabled several important insights (below) that have changed the way we analyze and think about neural population activity.

Results from simultaneous recordings challenge classic population-coding approaches that focus on distributed input tuning over a set of neurons^{42,43}. This view treats the population as simply a collection of individual neurons, where decoders estimate inputs using a suitable weighting of static neuronal responses. Such population codes work well when most neurons show a simple and straightforward tuned response. However, large-scale recordings are revealing neural computations (such as movement planning and decision-making) that are evident at the level of population dynamics, even when single neurons do not show an obvious tuning to stimulus or task variables^{16,44–46}. Such heterogeneous activity has prompted new frameworks in which the representation is contained in the dynamics of the population response⁴⁷ (Fig. 1b).

Another major insight that has been gained from simultaneous recordings is that transient or fluctuating responses at the single-neuron level can give way to stability at the population level (Fig. 1d). For example, persistent activity in single neurons was once thought to be the sole substrate of slow-timescale cognitive processes, such as working memory^{48,49} and were traditionally modeled by fixed point attractors¹³. Recent work extends this idea by demonstrating that dynamic single-neuron responses can coexist with a stable, lower-dimensional subspace coding that offers comparable benefits⁵⁰. Moreover, populations of transiently responding neurons can be generated by networks with minimal structure, and offer benefits over persistent activity in single neurons in terms of robustness and flexibility⁵¹.

Large-scale simultaneous recordings have also challenged the role of sequentially firing neurons (Fig. 1d). The predominant theory was that sequences of neurons reflect highly structured neural circuits, such as synfire chains⁵². However, large-scale recordings uncovered that, instead, sequential firing can emerge through cooperation between recurrent synaptic interactions and external inputs⁵¹, which argues that neural sequences can emerge gradually from largely unstructured network architectures. Hippocampal replay, thought to be involved in memory consolidation, offers another example of sequential activation that can be best studied when populations of many neurons are recorded simultaneously⁵³. Observing a hippocampal replay event requires precisely noting the relative timing of a population of place cells on the tens of milliseconds timescale. The insight that hippocampal activity

Box 2 | Tools and technologies to observe brain and behavior

Recent decades have seen dramatic progress in the ability to record and process neural activity, and to connect it to behavior. The number of simultaneously, electrically recorded cells has been increasing, and the development of Neuropixels probes accelerated this considerably. These linear probes with up to ~10,000 recording sites allow simultaneous recording of large neural populations spanning multiple areas^{17,120,121} (see the figure, panel c). Furthermore, the thin shank ($70 \times 20 \mu\text{m}^2$) causes minimal tissue displacement relative to previously used braided wires, and the probe's active circuits for amplifying, digitizing and multiplexing lower noise levels. Improvements in flexible electrodes, either part of an injectable mesh or inserted by a stiff guide, may further help to reduce tissue damage, and can enable stable, long-term interrogation of neural circuits during behavior¹²².

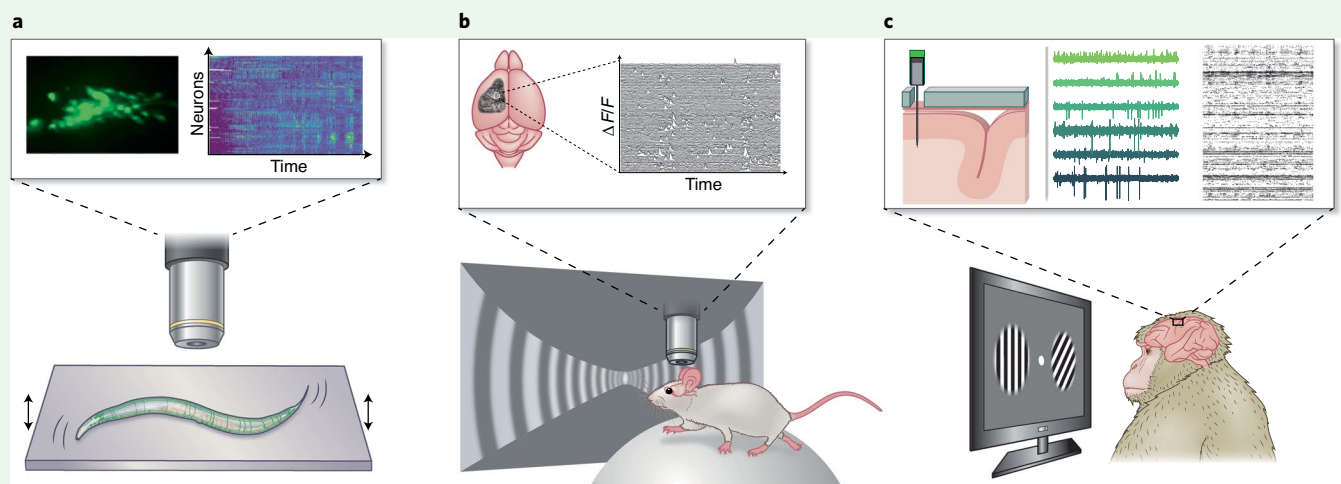
A major challenge for large-scale electrophysiology is a lack of consensus on spike sorting. Although recording from densely spaced sites can facilitate automated spike sorting, the problem still requires significant manual curation¹²⁵. Certain experiments may tolerate imperfect spike sorting¹²⁶, but many demand confidence in knowing which neuron was recorded. Simultaneous juxtacellular and extracellular recordings can provide ground-truth benchmark data on spike sorting, against which different algorithms have been systematically evaluated¹²⁷. These approaches are moving the field forward from traditional laboratory-specific and manually curated spike sorting toward standardized, less subjective practices¹²⁸.

Optical imaging has recently overtaken electrophysiology in its neural yield (Box 1). Recent progress has pushed the number of simultaneously recorded cells in the mouse brain to one million, about a tenth of its cortex¹²⁹. Imaging has high spatial resolution and coverage, allows labeling of specific cell types or projection targets¹³⁰, and can reveal the spatial organization of activity

patterns⁶². Imaging through thinned skulls, cranial windows or in transparent animals is minimally invasive, and allows long-term monitoring of the same structures (see the figure, panel b). Head-mounted microscopes additionally allow imaging in freely moving animals, and targeting of deep structures^{131,132}. However, as both calcium kinetics and the dynamics of calcium indicators are slow compared with neural firing, calcium imaging can be used only as a coarse proxy for spike timing and rate^{18,133,134}.

In smaller, transparent animals such as larval zebrafish, *C. elegans* and hydra, the activity of most neurons can be imaged simultaneously at cellular resolution in the fully intact animal^{112–114}. *C. elegans* recordings, for example, have provided a sandbox to test new theories of nonlinear dynamic system models applied to whole-brain dynamics^{135–138}. Fast-tracking microscopes allow for large-scale population imaging even as the animal swims or crawls^{123,139–142} (see the figure, panel a) and have revealed the importance of population codes for representing locomotion even in relatively simple animals¹⁰⁴. Multicolor labeling strategies to register entire brains on to an atlas with single-neuron accuracy¹⁴³ now allow whole-brain activity to be compared across individuals at cellular resolution and to further be linked to gene expression¹⁰⁸, opening up new ways to study individual variability in neural coding.

A final technological advance is in new ways of quantifying behavior more fully, largely driven by progress in video tracking and processing^{144–146}. Such data-driven approaches in parsing spontaneous behavior^{97,147–149} can allow us to interpret neural activity in the context of the behavior it produces, with or without experimenter-imposed task structure. This has benefited the study of both traditional, well-controlled behaviors^{150,151} and more ethological ones^{99,152,153}.



Large-scale neural recordings in behaving animals. **a**, Confocal microscopy of all neurons in *C. elegans* as it freely moves on an adjustable platform. Adapted from ref. ¹²³ under a CC-BY license. **b**, Mesoscope two-photon imaging of the cortical surface while a mouse moves through a virtual reality by running on a ball. Adapted from ref. ¹²⁴ under a CC-BY license. **c**, High-density electrophysiological recordings using Neuropixels probes, while a monkey performs a psychophysical decision-making task.

replays an animal's previous experience, forward or backward in time, thus relies crucially on the ability to observe relative timing among simultaneously recorded neurons.

A final advantage of simultaneously recorded neurons is that co-fluctuations among neurons offer insights into multi-region communication⁵⁴. For instance, some slow drifts in neural responses

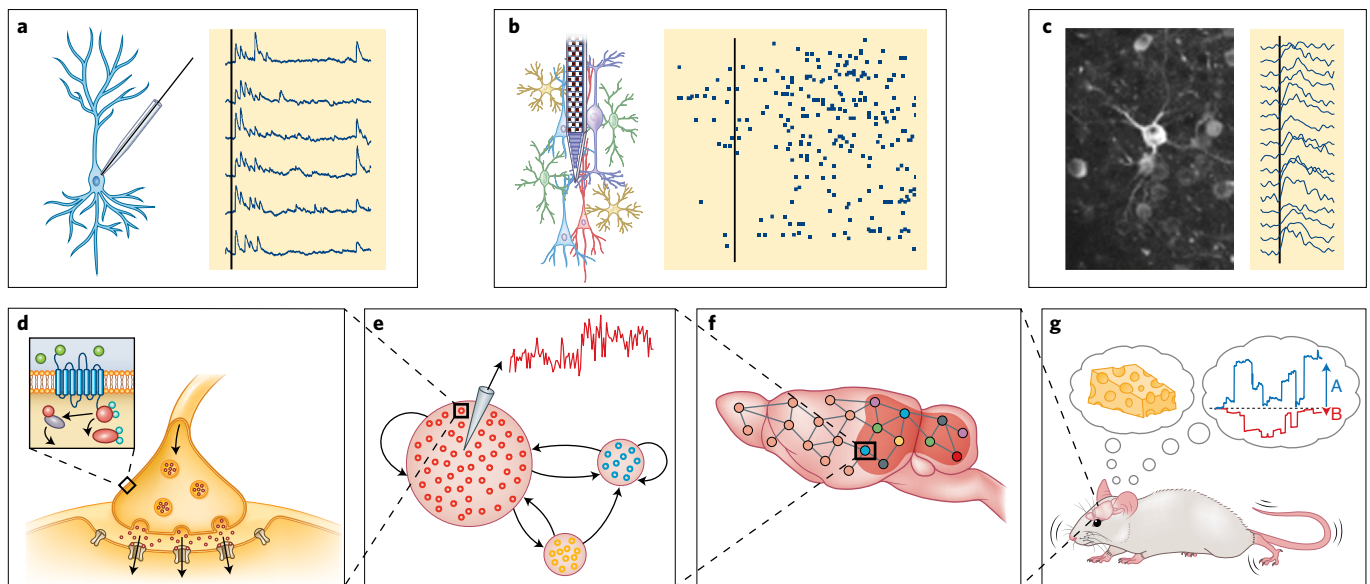


Fig. 2 | Understanding trial-to-trial neural variability across scales. a–c, Trial-to-trial variability of single-neuron responses in different recording modalities. **a,** Variability in responses to presynaptic stimulation, measured using patch clamp in vitro. Adapted from ref. ⁷², Springer Nature Ltd. **b,** Variability in spike timing and rate to visual stimulus presentation, measured using extracellular silicon probes in mouse visual cortex. **c,** Variability in calcium responses to whisker stimulation in mouse somatosensory cortex. Adapted from ref. ¹¹⁰ with permission from the author. **d–g,** Understanding neural variability across spatial scales. **d,** Cellular noise at the level of synapses and membrane dynamics. **e,** Circuit noise, arising from the dynamics of local populations of excitatory and inhibitory neurons. Adapted from ref. ¹¹¹ under a CC-BY license. **f,** Whole-brain noise, arising from the interactions between brain areas that may propagate or damp variability. **g,** Interpreting neural variability in the context of animal behavior, quantified from, for example, computational models of task-related cognitive processes, body movements and pupil-linked arousal, and task-unrelated physiological states.

are shared between V4 and the prefrontal cortex and can predict a monkey's fluctuating impulsivity in a decision-making task over the course of an experimental session⁵⁵. As the direction of the drifting signal was diverse across neurons, it could be uncovered only by analyzing large-scale simultaneous recordings. Other analyses of communication across cortical areas have revealed that some activity fluctuations are communicated to downstream structures, whereas others remain private⁵⁶. Such approaches are crucial for understanding ever-larger, multi-region neural recordings and promise a shift toward understanding neural activity in brain-wide models.

Analyses of multi-region communication often infer connectivity via correlations⁵⁷, in part because the connections of each neuron within an area are not known. In animals for which full^{58,59} or partial^{60,61} connectomes are known, interpreting multi-region recordings is becoming more concrete. In *Drosophila*, neural recordings had previously identified cells that track the animal's heading direction, putatively functioning as a ring-attractor network^{62,63}. The connectome then made it possible to extend these physiological results to arrive at a circuit model for how these neurons, and their connection weights, can compute a transformation from egocentric to allocentric coordinates. The critical observation from the connectome was the precise offset of synaptic weights between two types of cell (PFN and h Δ B)^{64–66}. This approach will probably benefit researchers likewise seeking to understand how allocentric traveling direction is computed in rodents⁶⁷. New experiments can evaluate whether the concrete model predictions garnered from *Drosophila*⁶⁶ are realized in the rodent. The challenge of connecting insights from invertebrates to mammals remains, and may be helped in the future by expanded knowledge of the mammalian connectome.

Theoretical frameworks: more is different

A potential criticism of large-scale recording studies is that they are observational rather than hypothesis-driven, and lack the ability to

distinguish concrete mechanistic models. In some sense, this is a fair criticism. A full understanding of brain function will require more than simply a list of all neurons and the extent to which each is modulated by one variable or another. On the other hand, some current studies deploy descriptive models partly out of necessity, as the complexity of the measured activity can make it difficult to relate it to existing theoretical frameworks. In the past, optimism suggested that, if only we could record more of the right neurons, our elegant assumed models would be readily confirmed. But when one records neural activity, the diversity of variables that modulate neural activity makes it challenging to argue that a signal at hand truly reflects a hypothesized computation. For instance, models of evidence accumulation offer appealing explanations for decisions made in the face of noisy evidence^{68,69}. Neural activity that 'ramps' during decision formation is certainly reminiscent of evidence accumulation⁷⁰, but such ramps can also reflect idiosyncratic combinations of stimuli and movements²² or the average of multiple, disparate sensory and decision-related motifs⁷¹. Thus, large-scale recordings are currently uncovering such unexpected neural responses that the ability to connect them to theoretical mechanisms may seem, at least momentarily, out of reach. A probable way forward is that the current focus on detailed characterization of neurons across brain structures will give way to more hypothesis-driven experiments in the near future. In this section, we discuss how new datasets can start to inform physiological models of large-scale brain networks, and point to the need for new (and different) theoretical frameworks to integrate brain and behavior.

What is needed is a blueprint for how large-scale datasets can first inform and then be a test for mechanistic models of cortical circuits. One promising approach to creating such models is to better understand the causes and consequences of variability in neural activity: trial-to-trial neuronal responses and within-trial spiking dynamics in diverse brain areas are famously variable⁷² (Fig. 2a–c).

Physicists have long used variability as a window into the dynamic interactions between components of a larger complex system⁷³. In the brain, any response variability reflects the underlying biology of the nervous system; however, the vast spatial and temporal scales over which this biology operates make it challenging to unravel the underlying neuronal mechanics. Indeed, past modeling efforts have been constrained by the experimental techniques used to record from local, small-scale cortical circuits^{12,14,74–76}. A severe limitation of this approach is that these models must make assumptions about any variability inherited from outside the circuit⁷⁷.

We here review how large-scale recordings can alleviate this shortcoming and contribute to understanding neural variability across spatial scales. Rather than aim for a single model of the brain (perhaps a large-scale simulation that gives rise to the sorts of computations observed in real brains), we include models of varying complexity; this is a hybrid (rather than hierarchical) approach that allows high-level and fine-grained models to coexist, each explaining different features of the data⁷⁸.

Perhaps best categorized is variability at the smallest scales of membrane and synaptic dynamics (Fig. 2d). Seminal work has shown that synaptic vesicle release and recovery are very unreliable^{79,80} and voltage-gated ion channels in the cellular membrane open and close randomly^{81,82}. At a larger spatial scale, neuronal recordings have provided compelling evidence of the spiking variability of single neurons, and populations of neurons in a local circuit (Fig. 2e). This variability is so pervasive that successful statistical modeling frameworks often take neurons to behave like assumed Poisson processes^{83,84}. One often-cited mechanism underlying such spiking variability is the emergent population dynamics in networks with strong and balanced excitatory and inhibitory recurrent interactions^{12,13}. Balanced excitatory–inhibitory networks with structured connectivity account for correlated pairwise variability^{76,85,86}, and even low dimensional, population-wide shared variability^{77,87,88}. Extending such mechanistic understanding to how variability is distributed over multiple brain regions remains a significant challenge in the new era of large datasets.

Compared with cellular or local network scales, much less is known about how the trial-to-trial neural variability in one brain region depends on the variability distributed over the rest of the brain (Fig. 2f). Large-scale recordings are starting to show how spiking variability in one brain region is inherited or filtered by another, prominently along the visual pathway^{54,89}, and in the songbird system⁹⁰. Yet it is not clear if all the variability in one region should be attributed to outside sources⁹¹, or that some component is internally generated from interactions within the circuit itself^{74,77,92}. Promising new analysis methods have started to identify specific activity patterns in one brain area, the variability of which is inherited from activity in an upstream region: a ‘communication subspace’⁵⁶. In the macaque visual system, only a small subset of the scope of V1 population variability drives population variability in V2, and this V1–V2 predictive dimension is largely nonoverlapping with the feedback V2–V1 subspace⁵⁶. This may allow V1 to route selective activity to different downstream areas and reduce unwanted co-fluctuations in downstream areas⁹³. Such dimensionality reduction approaches to link variability across brain regions will be needed to keep any ensuing mechanistic models tractable in the era of large-scale neural recordings⁹⁴.

Recent work has increasingly appreciated the large role of internal states^{95,96} and rich, spontaneous behaviors^{22,23,97} as crucial predictors of trial-to-trial neural variability (Fig. 2g). As these studies have remained largely descriptive, future theoretical approaches are needed to integrate the presence of such varied signals with the core computations carried out by neural circuits. As neural recordings increasingly capture most or all of the brain (Figure, Box 1), distinct implications for theoretical models may arise. In one extreme, ever-larger neural population, recordings may reduce the need for

detailed behavioral quantification for capturing neuronal variability. In this view, behavior is simply a proxy for (yet to be measured) neuronal activity. Alternatively, behavior may be a complex expression of distributed neuronal activity and must be an equal partner in any comprehensive model of brain activity and its variability. Ultimately, a resolution to this issue may lead us to fully understand the brain as part of the whole animal, with an appreciation for its evolutionary past and ethological niche^{98,99}.

The possibility of whole-brain recordings in small animals shows us both promise and warning: larger observations bring more nuance, but also lay bare gaps in our tools for interpreting brain-wide neural dynamics. Large-scale recordings in *C. elegans*, zebrafish and *Drosophila* spp. have revealed that many neurons are tuned to diverse aspects of behavior, often in subtle ways^{100–104}. These small systems may be ideal models in which there is a semblance of ‘ground truth’ for testing new theoretical frameworks, before applying them to larger brains¹⁰⁵. Although they provide striking opportunities for demonstrating the predictive power of such recordings, for example, by ‘mindreading’ animal behavior, they also remind us of how much of neural activity (measured by variance explained or otherwise) we still don’t understand.

Conclusions and outlook

Over the last few decades, our ability to perform large-scale neural recordings during behavior has grown by orders of magnitude: a postdoc can now record more neurons in a day than their principal investigator could collect over the course of an entire postdoctoral fellowship. We have here reviewed the technical progress and major insights gained from such experiments. We have highlighted how these advances have answered key questions in the field, and raised new ones for which theoretical frameworks are only starting to be developed.

With the progression of large-scale recording technology and computational capacity, what can we expect in the years and decades to come? Accurately predicting and decoding behavior from neural activity will probably become feasible for species with larger brains, building on the successes of small invertebrate models. Especially for brain–computer interfaces, this may have a significant impact on translational neuroscience. More simultaneously recorded neurons across connected brain regions will give a tighter handle on the sources of neural variability, and the distributed nature of neural circuit computations. We also expect to see an increasing appreciation for how neural computations depend on internal states, individual animals’ individual life history and a diversity in behavioral strategies. We hope that, in future work, deep understanding of animal behavior (from psychophysics to body movements and ethology) will be central to interpreting neural data. Improvements in dimensionality reduction, going beyond linear techniques such as principal component or factor analysis, will be needed to answer important questions about the size and complexity of neural circuits required for specific computations. More direct cross-species comparison will facilitate the transfer of insights from smaller, more tractable brains to larger organisms such as ourselves.

Ultimately, just recording many neurons will be insufficient to fully understand the brain—even when accompanied by well-quantified behavior and a connectome. Instead, a multi-level network description with all synaptic weights and additional molecular details may be required to perform causal inference and make behavioral predictions. The completeness of whole-brain recordings also stands in contrast to the inaccessibility of other signals in the brain, including neuromodulators¹⁰⁶, glia and glia-like cells¹⁰⁷. We also don’t yet fully understand how neural recordings that we observe relate to neural wiring¹⁰⁵ or gene expression¹⁰⁸. Some argue that, even in a world with perfect and complete data, we may not be able to understand the brain to our satisfaction. This question has recently led to lively debates, often centering around the

fundamental question of what constitutes an explanation in neuroscience. Given the immense progress of the past few decades, we optimistically predict that the future will continue to bring in-depth, unexpected and multifaceted understanding of brain function in all its complexities.

Citation diversity statement. Recent work in several fields of science has identified a bias in citation practices such that papers from women and other minority scholars are under-cited relative to the number of such papers in the field¹⁰⁹. In the present review, we sought to proactively consider choosing references that reflect the diversity of the field. Expected proportions estimated from five top neuroscience journals since 1997 are 6.7% woman/woman, 9.4% man/woman, 25.5% woman/man and 58.4% man/man¹⁰⁹. By this measure, our references (excluding those before 1997) contain 9.5% woman/woman, 10.9% man/woman, 21.9% woman/man and 57.7% man/man.

Data availability

All data used to generate Fig. 2 are available at https://github.com/anne-urai/largescale_recordings under a CC-BY 4.0 license.

Code availability

All code used to generate Fig. 2 are available at https://github.com/anne-urai/largescale_recordings under a CC-BY 4.0 license.

Received: 31 July 2020; Accepted: 8 November 2021;

Published online: 3 January 2022

References

- Perkel, D. H., Gerstein, G. L. & Moore, G. P. Neuronal spike trains and stochastic point processes. *Biophys. J.* **7**, 391–418 (1967).
- Abbott, L. F. & Dayan, P. The effect of correlated variability on the accuracy of a population code. *Neural Comput.* **11**, 91–101 (1999).
- Sompolinsky, H., Yoon, H., Kang, K. & Shamir, M. Population coding in neuronal systems with correlated noise. *Phys. Rev. E* **64**, 051904 (2001).
- Hebb, D. O. *The Organization of Behavior: A Neuropsychological Theory*. (Wiley, 1949).
- Abeles, M. *Corticonics: Neural Circuits of the Cerebral Cortex*. (Cambridge Univ. Press, 1991).
- Doiron, B., Litwin-Kumar, A., Rosenbaum, R., Ocker, G. K. & Josić, K. The mechanics of state-dependent neural correlations. *Nat. Neurosci.* **19**, 383–393 (2016).
- Kohn, A., Coen-Cagli, R., Kanitscheider, I. & Pouget, A. Correlations and neuronal population information. *Annu. Rev. Neurosci.* **39**, 237–256 (2016).
- Barlow, H. in *Sensory Communication* (ed. Rosenblith, W. A.) 217–234 (MIT Press, 1961).
- Britten, K. H., Shadlen, M. N., Newsome, W. T. & Movshon, A. Responses of neurons in macaque MT to stochastic motion signals. *Vis. Neurosci.* **10**, 1157–1169 (1993).
- Parker, A. J. & Newsome, W. T. Sense and the single neuron: probing the physiology of perception. *Annu. Rev. Neurosci.* **21**, 227–277 (1998).
- Wilson, H. R. & Cowan, J. D. Excitatory and inhibitory interactions in localized populations of model neurons. *Biophys. J.* **12**, 1–24 (1972).
- van Vreeswijk, C. & Sompolinsky, H. Chaos in neuronal networks with balanced excitatory and inhibitory activity. *Science* **274**, 1724–1726 (1996).
- Amit, D. J. & Brunel, N. Model of global spontaneous activity and local structured activity during delay periods in the cerebral cortex. *Cereb. Cortex* **7**, 237–252 (1997).
- Shadlen, M. N. & Newsome, W. T. The variable discharge of cortical neurons: implications for connectivity, computation, and information coding. *J. Neurosci.* **18**, 3870–3896 (1998).
- Li, P. H. et al. Anatomical identification of extracellularly recorded cells in large-scale multielectrode recordings. *J. Neurosci.* **35**, 4663–4675 (2015).
- Steinmetz, N. A., Zatka-Haas, P., Carandini, M. & Harris, K. D. Distributed coding of choice, action and engagement across the mouse brain. *Nature* **576**, 266–273 (2019).
- Siegle, J. H. et al. Survey of spiking in the mouse visual system reveals functional hierarchy. *Nature* **592**, 86–92 (2021).
- Siegle, J. H. et al. Reconciling functional differences in populations of neurons recorded with two-photon imaging and electrophysiology. *eLife* **10**, e69068 (2021).
- Humphries, M. *The Spike: An Epic Journey through the Brain in 2.1 Seconds* (Princeton Univ. Press, 2021).
- de Vries, S. E. J. et al. A large-scale standardized physiological survey reveals functional organization of the mouse visual cortex. *Nat. Neurosci.* **23**, 138–151 (2020).
- Shoham, S., O'Connor, D. H. & Segev, R. How silent is the brain: is there a 'dark matter' problem in neuroscience? *J. Comp. Physiol. A* **192**, 777–784 (2006).
- Musall, S., Kaufman, M. T., Juavinett, A. L., Gluf, S. & Churchland, A. K. Single-trial neural dynamics are dominated by richly varied movements. *Nat. Neurosci.* **22**, 1677–1686 (2019).
- Stringer, C. et al. Spontaneous behaviors drive multidimensional, brainwide activity. *Science* **364**, eaav7893 (2019).
- Salkoff, D. B., Zagha, E., McCarthy, E. & McCormick, D. A. Movement and performance explain widespread cortical activity in a visual detection task. *Cereb. Cortex* **30**, 421–437 (2020).
- Niell, C. M. & Stryker, M. P. Modulation of visual responses by behavioral state in mouse visual cortex. *Neuron* **65**, 472–479 (2010).
- Eriskens, S. et al. Effects of locomotion extend throughout the mouse early visual system. *Curr. Biol.* **24**, 2899–2907 (2014).
- Vinck, M., Batista-Brito, R., Knoblich, U. & Cardin, J. A. Arousal and locomotion make distinct contributions to cortical activity patterns and visual encoding. *Neuron* **86**, 740–754 (2015).
- Roth, M. M. et al. Thalamic nuclei convey diverse contextual information to layer 1 of visual cortex. *Nat. Neurosci.* **19**, 299–307 (2016).
- Dolensek, N., Gehrlach, D. A., Klein, A. S. & Gogolla, N. Facial expressions of emotion states and their neuronal correlates in mice. *Science* **368**, 89–94 (2020).
- Keller, G. B. & Mrsic-Flogel, T. D. Predictive processing: a canonical cortical computation. *Neuron* **100**, 424–435 (2018).
- Cunningham, J. P. & Yu, B. M. Dimensionality reduction for large-scale neural recordings. *Nat. Neurosci.* **17**, 1500–1509 (2014).
- Pang, R., Lansdell, B. J. & Fairhall, A. L. Dimensionality reduction in neuroscience. *Curr. Biol.* **26**, R656–R660 (2016).
- Ni, A., Ruff, D. A., Alberts, J. J., Symmonds, J. & Cohen, M. R. Learning and attention reveal a general relationship between population activity and behavior. *Science* **359**, 463–465 (2018).
- Aoi, M. C., Mante, V. & Pillow, J. W. Prefrontal cortex exhibits multidimensional dynamic encoding during decision-making. *Nat. Neurosci.* **23**, 1410–1420 (2020).
- Churchland, M. M. et al. Neural population dynamics during reaching. *Nature* **487**, 51–56 (2012).
- Kato, S. et al. Global brain dynamics embed the motor command sequence of *Caenorhabditis elegans*. *Cell* **163**, 656–669 (2015).
- Briggman, K. L., Abarbanel, H. D. I. & Kristan, W. B. Optical imaging of neuronal populations during decision-making. *Science* **307**, 896–901 (2005).
- Fusi, S., Miller, E. K. & Rigotti, M. Why neurons mix: high dimensionality for higher cognition. *Curr. Opin. Neurobiol.* **37**, 66–74 (2016).
- Gao, P. et al. A theory of multineuronal dimensionality, dynamics and measurement. Preprint at *bioRxiv* <https://doi.org/10.1101/214262> (2017).
- Humphries, M. D. Strong and weak principles of neural dimension reduction. *Neurons Behav. Data Anal. Theory* **5**, 1–28 (2021).
- Stringer, C., Pachitariu, M., Steinmetz, N., Carandini, M. & Harris, K. D. High-dimensional geometry of population responses in visual cortex. *Nature* **1**, 197 (2019).
- Georgopoulos, A., Schwartz, A. B. & Kettner, R. E. Neuronal population coding of movement direction. *Science* **233**, 1416–1419 (1986).
- Averbeck, B. B., Latham, P. E. & Pouget, A. Neural correlations, population coding and computation. *Nat. Rev. Neurosci.* **7**, 358–366 (2006).
- Shenoy, K. V., Sahani, M. & Churchland, M. M. Cortical control of arm movements: a dynamical systems perspective. *Annu. Rev. Neurosci.* **36**, 337–359 (2013).
- Russo, A. A. et al. Neural trajectories in the supplementary motor area and motor cortex exhibit distinct geometries, compatible with different classes of computation. *Neuron* **107**, 745–758.e6 (2020).
- Saxena, S., Russo, A. A., Cunningham, J. P. & Churchland, M. M. Motor cortex activity across movement speeds is predicted by network-level strategies for generating muscle activity. Preprint at *bioRxiv* <https://doi.org/10.1101/2021.02.01.429168> (2021).
- Saxena, S. & Cunningham, J. P. Towards the neural population doctrine. *Curr. Opin. Neurobiol.* **55**, 103–111 (2019).
- Fuster, J. M. & Alexander, G. E. Neuron activity related to short-term memory. *Science* **173**, 652–654 (1971).
- Constantinidis, C. et al. Persistent spiking activity underlies working memory. *J. Neurosci.* **38**, 7020–7028 (2018).
- Murray, J. D. et al. Stable population coding for working memory coexists with heterogeneous neural dynamics in prefrontal cortex. *Proc. Natl Acad. Sci. USA* **114**, 394–399 (2017).

51. Rajan, K., Harvey, C. D. & Tank, D. W. Recurrent network models of sequence generation and memory. *Neuron* **90**, 128–142 (2016).
52. Herrmann, M. J., Hertz, J. A. & Prügel-Bennett, A. Analysis of synfire chains. *Netw. Comput. Neural Syst.* **6**, 403–414 (1995).
53. Ólafsdóttir, F., Bush, D. & Barry, C. The role of hippocampal replay in memory and planning. *Curr. Biol.* **28**, R37–R50 (2018).
54. Ruff, D. A. & Cohen, M. R. Stimulus dependence of correlated variability across cortical areas. *J. Neurosci.* **36**, 7546–7556 (2016).
55. Cowley, B. R. et al. Slow drift of neural activity as a signature of impulsivity in macaque visual and prefrontal cortex. *Neuron* **108**, 551–567.e8 (2020).
56. Semedo, J. D., Zandvakili, A., Machens, C. K., Yu, B. M. & Kohn, A. Cortical areas interact through a communication subspace. *Neuron* **102**, 249–259.e4 (2019).
57. Das, A. & Fiete, I. R. Systematic errors in connectivity inferred from activity in strongly recurrent networks. *Nat. Neurosci.* **23**, 1286–1296 (2020).
58. White, J. G., Southgate, E., Thomson, J. N. & Brenner, S. The structure of the nervous system of the nematode *Caenorhabditis elegans*. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **314**, 1–340 (1986).
59. Witvliet, D. et al. Connectomes across development reveal principles of brain maturation. *Nature* **596**, 257–261 (2021).
60. Eichler, K. et al. The complete connectome of a learning and memory centre in an insect brain. *Nature* **548**, 175–182 (2017).
61. Scheffer, L. K. et al. A connectome and analysis of the adult *Drosophila* central brain. *eLife* **9**, e57443 (2020).
62. Seelig, J. D. & Jayaraman, V. Neural dynamics for landmark orientation and angular path integration. *Nature* **521**, 186–191 (2015).
63. Kim, S. S., Rouault, H., Druckmann, S. & Jayaraman, V. Ring attractor dynamics in the *Drosophila* central brain. *Science* **356**, 849–853 (2017).
64. Hulse, B. K. et al. A connectome of the *Drosophila* central complex reveals network motifs suitable for flexible navigation and context-dependent action selection. *eLife* <https://elifesciences.org/articles/66039> (2021).
65. Lu, J. et al. Transforming representations of movement from body- to world-centric space. *Nature* (in the press).
66. Lyu, C., Abbott, L. F. & Maimon, G. A neuronal circuit for vector computation builds an allocentric traveling-direction signal in the *Drosophila* fan-shaped body. *Nature* (in the press).
67. Gardner, R. J. et al. Toroidal topology of population activity in grid cells. Preprint at *bioRxiv* <https://doi.org/10.1101/2021.02.25.432776> (2021).
68. Ratcliff, R. & McKoon, G. The diffusion decision model: theory and data for two-choice decision tasks. *Neural Comput.* **20**, 873–922 (2008).
69. Brunton, B. W., Botvinick, M. M. & Brody, C. D. Rats and humans can optimally accumulate evidence for decision-making. *Science* **340**, 95–98 (2013).
70. Churchland, A. K. & Kiani, R. Three challenges for connecting model to mechanism in decision-making. *Curr. Opin. Behav. Sci.* **11**, 74–80 (2016).
71. Meister, M. L. R., Hennig, J. A. & Huk, A. C. Signal multiplexing and single-neuron computations in lateral intraparietal area during decision-making. *J. Neurosci.* **33**, 2254–2267 (2013).
72. Faisal, A. A., Selen, L. P. J. & Wolpert, D. M. Noise in the nervous system. *Nat. Rev. Neurosci.* **9**, 292–303 (2008).
73. Kubo, R., Toda, M. & Hashitsume, N. *Statistical Physics II: Nonequilibrium Statistical Mechanics* (Springer, 1991).
74. Rajan, K., Abbott, L. F. & Sompolsky, H. Stimulus-dependent suppression of chaos in recurrent neural networks. *Phys. Rev. E* **82**, 011903 (2010).
75. Churchland, A. K. et al. Variance as a signature of neural computations during decision making. *Neuron* **69**, 818–831 (2011).
76. Litwin-Kumar, A. & Doiron, B. Slow dynamics and high variability in balanced cortical networks with clustered connections. *Nat. Neurosci.* **15**, 1498–1505 (2012).
77. Huang, C. et al. Circuit models of low-dimensional shared variability in cortical networks. *Neuron* **101**, 337–348.e4 (2019).
78. Churchland, A. K. & Abbott, L. F. Conceptual and technical advances define a key moment for theoretical neuroscience. *Nat. Neurosci.* **19**, 348–349 (2016).
79. Calvin, W. H. & Stevens, C. F. Synaptic noise and other sources of randomness in motoneuron interspike intervals. *J. Neurophysiol.* **31**, 574–587 (1968).
80. Branco, T. & Staras, K. The probability of neurotransmitter release: variability and feedback control at single synapses. *Nat. Rev. Neurosci.* **10**, 373–383 (2009).
81. Neher, E. & Sakmann, B. Single-channel currents recorded from membrane of denervated frog muscle fibres. *Nature* **260**, 799–802 (1976).
82. White, J. A., Rubinstein, J. T. & Kay, A. R. Channel noise in neurons. *Trends Neurosci.* **23**, 131–137 (2000).
83. Pillow, J. W. et al. Spatio-temporal correlations and visual signalling in a complete neuronal population. *Nature* **454**, 995–999 (2008).
84. Goris, R. L. T., Movshon, J. A. & Simoncelli, E. P. Partitioning neuronal variability. *Nat. Neurosci.* **17**, 858–865 (2014).
85. Rosenbaum, R., Smith, M. A., Kohn, A., Rubin, J. E. & Doiron, B. The spatial structure of correlated neuronal variability. *Nat. Neurosci.* **20**, 107–114 (2017).
86. Darshan, R., van Vreeswijk, C. & Hansel, D. Strength of correlations in strongly recurrent neuronal networks. *Phys. Rev. X* **8**, 031072 (2018).
87. Landau, I. D. & Sompolsky, H. Coherent chaos in a recurrent neural network with structured connectivity. *PLoS Comput. Biol.* **14**, e1006309 (2018).
88. Mastrogiuseppe, F. & Ostojic, S. Linking connectivity, dynamics, and computations in low-rank recurrent neural networks. *Neuron* **99**, 609–623.e29 (2018).
89. Gómez-Laberge, C., Smolyanskaya, A., Nassi, J. J., Kreiman, G. & Born, R. T. Bottom-up and top-down input augment the variability of cortical neurons. *Neuron* **91**, 540–547 (2016).
90. Woolley, S. C., Rajan, R., Joshua, M. & Doupe, A. J. Emergence of context-dependent variability across a basal ganglia network. *Neuron* **82**, 208–223 (2014).
91. Hires, A., Gutnisky, D. A., Yu, J., O'Connor, D. H. & Svoboda, K. Low-noise encoding of active touch by layer 4 in the somatosensory cortex. *eLife* **4**, e06619 (2015).
92. Meshulam, L., Gauthier, J. L., Brody, C. D., Tank, D. W. & Bialek, W. Collective behavior of place and non-place neurons in the hippocampal network. *Neuron* **96**, 1178–1191.e4 (2017).
93. Kaufman, M. T., Churchland, M. M., Ryu, S. I. & Shenoy, K. V. Cortical activity in the null space: permitting preparation without movement. *Nat. Neurosci.* **17**, 440–448 (2014).
94. Williamson, R. C., Doiron, B., Smith, M. A. & Yu, B. M. Bridging large-scale neuronal recordings and large-scale network models using dimensionality reduction. *Curr. Opin. Neurobiol.* **55**, 40–47 (2019).
95. McGinley, M. J. et al. Waking state: rapid variations modulate neural and behavioral responses. *Neuron* **87**, 1143–1161 (2015).
96. Musall, S., Urai, A. E., Sussillo, D. & Churchland, A. K. Harnessing behavioral diversity to understand neural computations for cognition. *Curr. Opin. Neurobiol.* **58**, 229–238 (2019).
97. Wilschko, A. B. et al. Mapping sub-second structure in mouse behavior. *Neuron* **88**, 1121–1135 (2015).
98. Krakauer, J. W., Ghazanfar, A. A., Gomez-Marin, A., MacIver, M. A. & Poeppel, D. Neuroscience needs behavior: correcting a reductionist bias. *Neuron* **93**, 480–490 (2017).
99. Juavinett, A. L., Erlich, J. C. & Churchland, A. K. Decision-making behaviors: weighing ethology, complexity, and sensorimotor compatibility. *Curr. Opin. Neurobiol.* **49**, 42–50 (2018).
100. Chen, X. et al. Brain-wide organization of neuronal activity and convergent sensorimotor transformations in larval zebrafish. *Neuron* **100**, 876–890.e5 (2018).
101. Pacheco, D. A., Thiberge, S. Y., Pnevmatikakis, E. & Murthy, M. Auditory activity is diverse and widespread throughout the central brain of *Drosophila*. *Nat. Neurosci.* **24**, 93–104 (2021).
102. Ji, N. et al. A neural circuit for flexible control of persistent behavioral states. *eLife* **10**, e62889 (2021).
103. Kaplan, H. S., Salazar Thula, O., Khoss, N. & Zimmer, M. Nested neuronal dynamics orchestrate a behavioral hierarchy across timescales. *Neuron* **105**, 562–576.e9 (2020).
104. Hallinen, K. M. et al. Decoding locomotion from population neural activity in moving *C. elegans*. *eLife* **10**, e66135 (2021).
105. Bargmann, C. I. & Marder, E. From the connectome to brain function. *Nat. Methods* **10**, 483–490 (2013).
106. Marder, E. Neuromodulation of neuronal circuits: back to the future. *Neuron* **76**, 1–11 (2012).
107. Barres, B. A. The mystery and magic of glia: a perspective on their roles in health and disease. *Neuron* **60**, 430–440 (2008).
108. Taylor, S. R. et al. Molecular topography of an entire nervous system. *Cell* **184**, 4329–4347.e23 (2021).
109. Dworkin, J. D. et al. The extent and drivers of gender imbalance in neuroscience reference lists. *Nat. Neurosci.* **23**, 918–926 (2020).
110. Couto, J. et al. Spatially segregated responses to visuo-tactile stimuli in mouse neocortex during active sensation. Preprint at *bioRxiv* <https://doi.org/10.1101/199364> (2019).
111. Bos, H., Oswald, A.-M. & Doiron, B. Untangling stability and gain modulation in cortical circuits with multiple interneuron classes. Preprint at *bioRxiv* <https://doi.org/10.1101/2020.06.15.148114> (2020).
112. Schrödel, T., Prevedel, R., Aumayr, K., Zimmer, M. & Vaziri, A. Brain-wide 3D imaging of neuronal activity in *Caenorhabditis elegans* with sculpted light. *Nat. Methods* **10**, 1013–1020 (2013).
113. Ahrens, M. B., Orger, M. B., Robson, D. N., Li, J. M. & Keller, P. J. Whole-brain functional imaging at cellular resolution using light-sheet microscopy. *Nat. Methods* **10**, 413–420 (2013).
114. Yamamoto, W. & Yuste, R. Whole-body imaging of neural and muscle activity during behavior in *Hydra vulgaris*: effect of osmolarity on contraction bursts. *eNeuro* <https://doi.org/10.1101/2019.12.20.883835> (2020).

115. Mann, K., Gallen, C. L. & Clandinin, T. R. Whole-brain calcium imaging reveals an intrinsic functional network in *Drosophila*. *Curr. Biol.* **27**, 2389–2396.e4 (2017).
116. Aimon, S. et al. Fast near-whole-brain imaging in adult *Drosophila* during responses to stimuli and behavior. *PLoS Biol.* **17**, e2006732 (2019).
117. Marblestone, A. H. et al. Physical principles for scalable neural recording. *Front. Comput. Neurosci.* **7**, 134 (2013).
118. Kleinfeld, D. et al. Can one concurrently record electrical spikes from every neuron in a mammalian brain? *Neuron* **103**, 1005–1015 (2019).
119. Stevenson, I. H. & Kording, K. P. How advances in neural recording affect data analysis. *Nat. Neurosci.* **14**, 139–142 (2011).
120. Steinmetz, N. A. et al. Neuropixels 2.0: A miniaturized high-density probe for stable, long-term brain recordings. *Science* **372**, eabf4588 (2021).
121. Juavinett, A. L., Bekheet, G. & Churchland, A. K. Chronically implanted Neuropixels probes enable high-yield recordings in freely moving mice. *eLife* **8**, e47188 (2019).
122. Hong, G. & Lieber, C. M. Novel electrode technologies for neural recordings. *Nat. Rev. Neurosci.* **20**, 330–345 (2019).
123. Nguyen, J. P. et al. Whole-brain calcium imaging with cellular resolution in freely behaving *Caenorhabditis elegans*. *Proc. Natl Acad. Sci. USA* **113**, E1074–E1081 (2016).
124. Sofroniew, N. J. Q&A: the brain under a mesoscope: the forest and the trees. *BMC Biol.* **15**, 82 (2017).
125. Harris, K. D., Quiroga, R. Q., Freeman, J. & Smith, S. L. Improving data quality in neuronal population recordings. *Nat. Neurosci.* **19**, 1165–1174 (2016).
126. Trautmann, E. M. et al. Accurate estimation of neural population dynamics without spike sorting. *Neuron* **103**, 292–308.e4 (2019).
127. Magland, J. et al. SpikeForest, reproducible web-facing ground-truth validation of automated neural spike sorters. *eLife* **9**, e55167 (2020).
128. Buccino, A. P. et al. SpikeInterface, a unified framework for spike sorting. *eLife* **9**, e61834 (2020).
129. Demas, J. et al. High-speed, cortex-wide volumetric recording of neuroactivity at cellular resolution using light beads microscopy. *Nat. Methods* **18**, 1103–1111 (2021).
130. Grienberger, C. & Konnerth, A. Imaging calcium in neurons. *Neuron* **73**, 862–885 (2012).
131. Ghosh, K. K. et al. Miniaturized integration of a fluorescence microscope. *Nat. Methods* **8**, 871–878 (2011).
132. Aharoni, D., Khakh, B. S., Silva, A. J. & Golshani, P. All the light that we can see: a new era in miniaturized microscopy. *Nat. Methods* **16**, 11–13 (2019).
133. Najafi, F. et al. Excitatory and inhibitory subnetworks are equally selective during decision-making and emerge simultaneously during learning. *Neuron* **105**, 165–179.e8 (2020).
134. Huang, L. et al. Relationship between simultaneously recorded spiking activity and fluorescence signal in GCaMP6 transgenic mice. *eLife* **10**, e51675 (2021).
135. Brennan, C. & Proekt, A. A quantitative model of conserved macroscopic dynamics predicts future motor commands. *eLife* **8**, e46814 (2019).
136. Costa, A. C., Ahamed, T. & Stephens, G. J. Adaptive, locally linear models of complex dynamics. *Proc. Natl Acad. Sci. USA* **116**, 1501–1510 (2019).
137. Linderman, S., Nichols, A., Blei, D., Zimmer, M. & Paninski, L. Hierarchical recurrent state space models reveal discrete and continuous dynamics of neural activity in *C. elegans*. Preprint at *bioRxiv* <https://doi.org/10.1101/621540> (2019).
138. Fieseler, C., Zimmer, M. & Kutz, N. Unsupervised learning of control signals and their encodings in *Caenorhabditis elegans* whole-brain recordings. *J. R. Soc. Interface* **17**, 20200459 (2020).
139. Venkatachalam, V. et al. Pan-neuronal imaging in roaming *Caenorhabditis elegans*. *Proc. Natl Acad. Sci. USA* **113**, E1082–E1088 (2016).
140. Cong, L. et al. Rapid whole brain imaging of neural activity in freely behaving larval zebrafish (*Danio rerio*). *eLife* **6**, e28158 (2017).
141. Kim, D. H. et al. Pan-neuronal calcium imaging with cellular resolution in freely swimming zebrafish. *Nat. Methods* **14**, 1107–1114 (2017).
142. Symvoulidis, P. et al. NeuTracker—imaging neurobehavioral dynamics in freely behaving fish. *Nat. Methods* **14**, 1079–1082 (2017).
143. Yemini, E. et al. NeuroPAL: a multicolor atlas for whole-brain neuronal identification in *C. elegans*. *Cell* **184**, 272–288.e11 (2021).
144. Machado, A. S., Darmohray, D. M., Fayad, J., Marques, H. G. & Carey, M. R. A quantitative framework for whole-body coordination reveals specific deficits in freely walking ataxic mice. *eLife* **4**, e07892 (2015).
145. Mathis, M. W. & Mathis, A. Deep learning tools for the measurement of animal behavior in neuroscience. *Curr. Opin. Neurobiol.* **60**, 1–11 (2020).
146. Pereira, T. D. et al. Fast animal pose estimation using deep neural networks. *Nat. Methods* **16**, 117–125 (2019).
147. Berman, G. J., Choi, D. M., Bialek, W. & Shaevitz, J. W. Mapping the stereotyped behaviour of freely moving fruit flies. *J. R. Soc. Interface* **11**, 20140672 (2014).
148. Batty, E. et al. BehaveNet: nonlinear embedding and Bayesian neural decoding of behavioral videos. in *Advances in Neural Information Processing Systems* 32 (eds. Wallach, H. et al.) 15680–15691 (Curran, 2019).
149. Calhoun, A. J., Pillow, J. W. & Murthy, M. Unsupervised identification of the internal states that shape natural behavior. *Nat. Neurosci.* **22**, 2040–2049 (2019).
150. Carandini, M. & Churchland, A. K. Probing perceptual decisions in rodents. *Nat. Neurosci.* **16**, 824–831 (2013).
151. The International Brain Laboratory et al. Standardized and reproducible measurement of decision-making in mice. *eLife* **10**, e63711 (2021).
152. Dulac, C., O'Connell, L. A. & Wu, Z. Neural control of maternal and paternal behaviors. *Science* **345**, 765–770 (2014).
153. Karigo, T. et al. Distinct hypothalamic control of same- and opposite-sex mounting behaviour in mice. *Nature* **589**, 258–263 (2021).

Acknowledgements

A.E.U. is supported by the German National Academy of Sciences Leopoldina and the International Brain Research Organization. B.D. is supported by National Institutes of Health (NIH) (grant no. 1U19NS107613-01, R01EB026953), Vannevar Bush faculty fellowship (no. N00014-18-1-2002) and the Simons Foundation Collaboration on the Global Brain. A.M.L. is supported by the National Institute of Neurological Disorders and Stroke of the NIH (under New Innovator award no. DP2NS116768), and Simons Foundation Award (no. SCGB 543003). A.K.C. is supported by the NIH (nos. R01EY022979 and R01EB026949) and the Simons Collaboration on the Global Brain. We thank N. Sofroniew for sharing the mesoscope image panel shown in the figure in Box 2, panel b, E. Trautman and K. Shenoy for the primate electrophysiology (Neuropixels) data in the figure in Box 2, panel c, and D. Maizels for graphic design. I. Stevenson, K. Svoboda, P. Rupprecht, A. Charles and G. Meijer suggested data points shown in Box 1, and J. Couto provided helpful comments on an earlier version of the manuscript. J. Tuthill provided insights on interpreting data from the fly connectome.

Competing interests

The authors declare no competing interests.

Additional information

Correspondence should be addressed to Anne K. Churchland.

Peer review information *Nature Neuroscience* thanks the anonymous reviewers for their contribution to the peer review of this work.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

© Springer Nature America, Inc. 2022