



Universiteit  
Leiden  
The Netherlands

## **Chronobiology and chronotherapy of osteoporosis**

Winter, E.M.; Kooijman, S.; Appelman-Dijkstra, N.M.; Meijer, O.C.; Rensen, P.C.; Schilperoort, M.

### **Citation**

Winter, E. M., Kooijman, S., Appelman-Dijkstra, N. M., Meijer, O. C., Rensen, P. C., & Schilperoort, M. (2021). Chronobiology and chronotherapy of osteoporosis. *Jbmr Plus*, 5(10). doi:10.1002/jbm4.10504

Version: Publisher's Version

License: [Creative Commons CC BY 4.0 license](#)

Downloaded from: <https://hdl.handle.net/1887/3237835>

**Note:** To cite this publication please use the final published version (if applicable).

# Chronobiology and Chronotherapy of Osteoporosis

Elizabeth M Winter,<sup>1,2,3</sup>  Sander Kooijman,<sup>1,2</sup> Natasha M Appelman-Dijkstra,<sup>1,2,3</sup>  Onno C Meijer,<sup>1,2</sup> Patrick CN Rensen,<sup>1,2</sup> and Maaïke Schilperoord<sup>1,2</sup> 

<sup>1</sup>Department of Medicine, Division of Endocrinology, Leiden University Medical Center, Leiden, The Netherlands

<sup>2</sup>Eindhoven Laboratory for Experimental Vascular Medicine, Leiden, The Netherlands

<sup>3</sup>Department of Medicine, Center for Bone Quality, Leiden University Medical Center, Leiden, The Netherlands

## ABSTRACT

Physiological circadian (ie, 24-hour) rhythms are critical for bone health. Animal studies have shown that genes involved in the intrinsic molecular clock demonstrate potent circadian expression patterns in bone and that genetic disruption of these clock genes results in a disturbed bone structure and quality. More importantly, circulating markers of bone remodeling show diurnal variation in mice as well as humans, and circadian disruption by, eg, working night shifts is associated with the bone remodeling disorder osteoporosis. In this review, we provide an overview of the current literature on rhythmic bone remodeling and its underlying mechanisms and identify critical knowledge gaps. In addition, we discuss novel (chrono)therapeutic strategies to reduce osteoporosis by utilizing our knowledge on circadian regulation of bone. © 2021 The Authors. *JBMR Plus* published by Wiley Periodicals LLC on behalf of American Society for Bone and Mineral Research.

**KEY WORDS:** BIOLOGICAL CLOCK; BONE REMODELING; CHRONOTHERAPY; CIRCADIAN RHYTHM; FRACTURES; OSTEOPOROSIS

## 1. Introduction

Healthy bone requires continuous remodeling to maintain its strength.<sup>(1)</sup> This delicate process is dependent on the well-coordinated activity of osteocytes, osteoclasts, and osteoblasts. After initiation of the bone remodeling cycle by osteocytes in response to mechanical loading or a microfracture, osteoclasts need to be recruited to the bone surface<sup>(2)</sup> to dissolve bone minerals and break down bone matrix. This is achieved by production of hydrochloric acid and lysosomal proteases (eg, cathepsin K),<sup>(3)</sup> which liberates growth factors that are trapped within the bone matrix (eg, bone morphogenetic proteins [BMPs], transforming growth factor beta [TGF- $\beta$ ], and insulin-like growth factor 1 [IGF-1]). Released bone matrix-derived factors promote migration and differentiation of osteoblast precursors.<sup>(4–6)</sup> In addition, osteoclasts can also directly interact with osteoblasts via recently identified “osteoclast-derived coupling factors,” which include cell surface regulatory proteins and secreted factors.<sup>(7)</sup> At the sites of bone resorption, osteoblasts produce new bone matrix predominantly composed of type I collagen. This newly formed osteoid is progressively mineralized by deposition of calcium (Ca<sup>2+</sup>) and phosphate (PO<sub>4</sub><sup>3-</sup>) in the form of hydroxyapatite crystals to increase mechanical strength of bone.<sup>(8,9)</sup> The mineralization process is facilitated by matrix vesicles produced from osteoblasts<sup>(10)</sup> and osteocyte-derived proteins such as dentin matrix protein 1 (DMP1),<sup>(11)</sup> after which

the bone remodeling cycle is concluded. During this process, some osteoblasts become trapped in the calcified matrix and differentiate into osteocytes, while others turn into bone lining cells (Fig. 1).

Osteoporosis develops if bone resorption exceeds bone formation, leading to bone fragility and increased fracture risk, which can result from either overactivity of osteoclasts<sup>(12)</sup> or hampered osteoblast activity or function.<sup>(13)</sup> The homeostatic activity of bone resorption and bone formation is regulated through the actions of various systemic hormones of which sex hormones are of particular importance. Although in men testosterone is the dominant circulating sex steroid hormone, estrogen is also formed in men from testosterone via aromatization. For women, estrogen is the dominant hormone, but they also have low levels of the androgens that are produced by the adrenal glands.<sup>(14,15)</sup> Androgens affect bone directly by preventing osteoblasts to undergo apoptosis.<sup>(14)</sup> (Androgen-derived) estrogens inhibit bone resorption by stimulating apoptosis of osteoclasts,<sup>(16)</sup> thus explaining the excessive bone resorption and increased fracture risk in postmenopausal women. Although androgens beneficially affect bone, the action of estrogen is thought to be stronger.<sup>(14)</sup> This is also demonstrated by the fact that blunted estrogen signaling due to mutations in the estrogen receptor gene<sup>(17)</sup> or aromatase deficiency<sup>(18–20)</sup> is associated with a lower bone mass in men.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

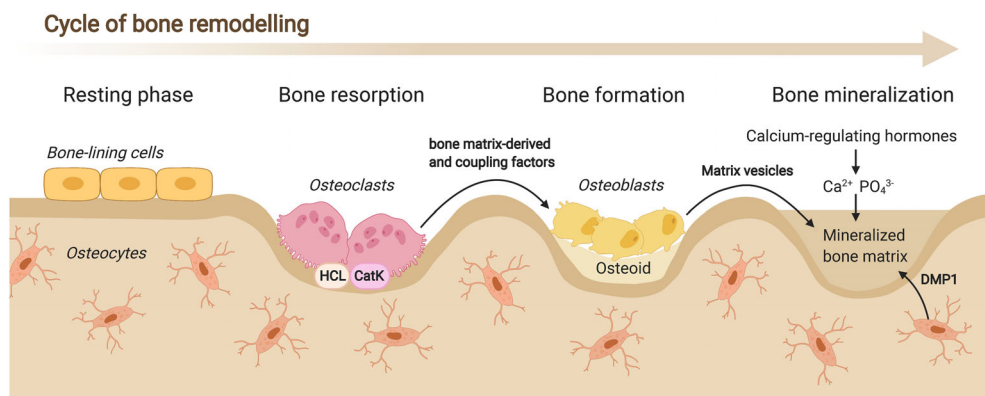
Received in original form December 28, 2020; revised form March 31, 2021; accepted April 10, 2021. Accepted manuscript online April 21, 2021.

Address correspondence to: Elizabeth M Winter, MD, PhD, Department of Medicine, Division of Endocrinology, and Center for Bone Quality, Postal zone C7-Q, Leiden University Medical Center, PO Box 9600, 2300 RC Leiden, The Netherlands. E-mail: e.m.winter@lumc.nl

JBMR<sup>®</sup> Plus (WOA), Vol. 5, No. 10, October 2021, e10504.

DOI: 10.1002/jbm4.10504

© 2021 The Authors. *JBMR Plus* published by Wiley Periodicals LLC on behalf of American Society for Bone and Mineral Research.



**Fig 1.** The bone remodeling cycle. See text for explanation.

In addition to sex hormones, the whole-body calcium regulators parathyroid hormone (PTH) and vitamin D are also important for bone health. PTH is produced by the parathyroid gland in response to low serum calcium levels and stimulates the expression of receptor activator of nuclear factor  $\kappa$ B ligand (RANKL) by osteoblasts.<sup>(21)</sup> RANKL binds to receptor activator of nuclear factor  $\kappa$ B (RANK) on osteoclasts, thereby promoting osteoclast proliferation and differentiation. Thus, by stimulating RANK-RANKL signaling, PTH enhances bone resorption and thereby the release of calcium and phosphate from bone.<sup>(22)</sup> PTH further increases serum calcium levels by promoting reabsorption of urinary calcium in the kidney, and indirectly, by stimulating intestinal calcium resorption. The latter is the result of 1-alpha-hydroxylase activity, induced by PTH. This enzyme converts 25-hydroxycholecalciferol into 1,25-dihydroxycholecalciferol, ie, the active form of vitamin D, which stimulates calcium and phosphate absorption from the gut.<sup>(23)</sup> The ensuing, immediate rise in serum calcium exerts negative feedback on PTH secretion to ensure that calcium levels are maintained within a narrow range. In addition, high serum calcium levels promote calcitonin production by the thyroid gland. Calcitonin counteracts the effects of PTH by inhibiting bone resorption and increasing renal calcium excretion, although its physiological importance in humans is debated.<sup>(24)</sup>

Nutritional status and lifestyle factors fine-tune the hormonal feedback loops and are therefore also important determinants of bone health.<sup>(25)</sup> Malnutrition, smoking, and excessive alcohol consumption have a detrimental impact on bone, while physical activity promotes bone remodeling. Predominantly during high-impact and weight-bearing exercises, mechanical forces are exerted on the bone through ground reaction forces and by the contractile activity of muscles. These forces are sensed by an intricate network of osteocytes, which subsequently respond by shifting the balance in bone remodeling toward bone formation, thereby increasing bone strength.<sup>(26)</sup>

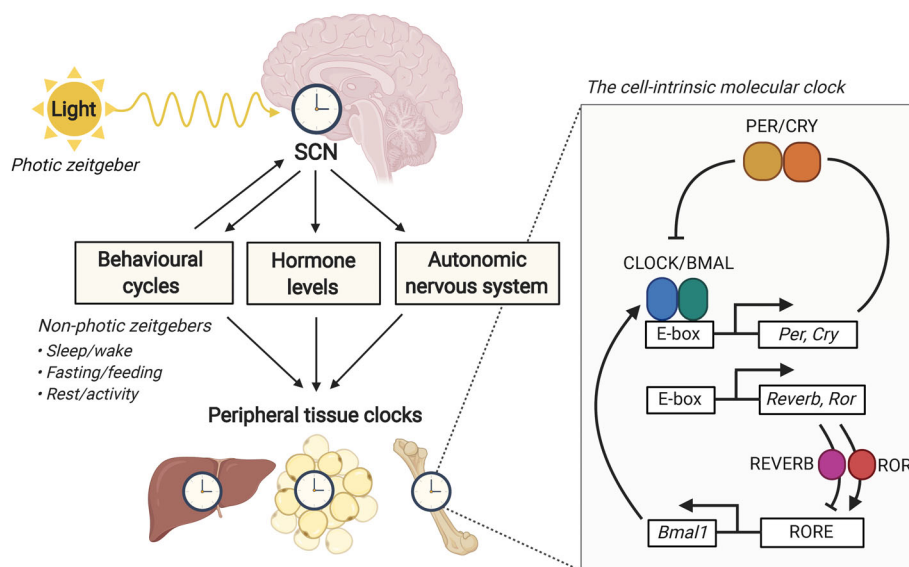
It is becoming increasingly clear that bone remodeling is under strict control of the biological clock and that disruption of circadian (ie, 24-hour) rhythms by night shift work is associated with osteoporosis and fractures.<sup>(27,28)</sup> The circadian timing system orchestrates daily rhythms in physiological processes through a small brain region in the hypothalamus, named the suprachiasmatic nucleus (SCN). The SCN connects the inner workings of the body to the outside environment, by receiving

photoc (ie, light) and nonphotoc input, and orchestrates coherent circadian rhythms in peripheral tissues including bone.<sup>(29)</sup> Transmission of external timing signals is mediated through regulation of autonomic nervous system activity, behavioral cycles (eg, sleep/wake, fasting/feeding, rest/activity), and circulating hormone levels, of which glucocorticoid (GC) hormones are especially important.<sup>(30)</sup> Behavior can also affect circadian clocks in peripheral tissues including bone in an SCN-independent manner, as described for time-restricted feeding<sup>(31–33)</sup> and scheduled exercise.<sup>(34)</sup> Through these SCN-dependent and -independent mechanisms, illustrated in Fig. 2, body rhythms are synchronized to the external 24-hour light/dark cycle.

Circadian rhythms within peripheral tissues are maintained through a cell-autonomous molecular clock (Fig. 2), generated by two interlocking transcriptional/translational feedback loops (TTFL).<sup>(35)</sup> The core TTFL consists of two activator proteins (CLOCK and BMAL1) and two repressor proteins (PER and CRY). The activator proteins CLOCK and BMAL1 heterodimerize and bind to a DNA cis-element E-box to initiate transcription of the repressor genes *PER* and *CRY*. *PER* and *CRY* proteins subsequently heterodimerize and translocate to the nucleus to inhibit the CLOCK and BMAL1 complex, thereby inhibiting their own transcription, which lasts until they are degraded after 24 hours. This thus results in a self-sustaining oscillation of core clock genes. The oscillations are fine-tuned via the induction of clock proteins REV-ERB $\alpha/\beta$  and ROR $\alpha/\beta$  by CLOCK/BMAL1 heterodimers, which inhibit and activate *BMAL1* transcription through retinoic acid-related orphan receptor response element (RORE) binding, respectively (Fig. 2). Together, these interlocking feedback loops produce robust 24-hour rhythms in the expression of genes and proteins comprising the molecular clock (collectively named “clock genes” and “clock proteins”).<sup>(36)</sup> Clock proteins not only regulate the expression of (other) clock genes but also can initiate transcription of tissue-specific target genes in a circadian oscillating pattern.<sup>(37)</sup> As a result, many important tissue-specific genes and proteins demonstrate a circadian rhythm.<sup>(38,39)</sup>

## 2. Chronobiology of Osteoporosis

We and others have demonstrated that the clock genes *Bmal1*, *Clock*, *Per1*, *Per2*, *Cry1*, and *Reverba* all exhibit diurnal expression



**Fig 2.** Regulation of the molecular clock in peripheral tissues by the suprachiasmatic nucleus and zeitgebers. The suprachiasmatic nucleus (SCN) receives input from different environmental factors called “zeitgebers” (German for “time-givers”). Behavioral signals such as sleep/wake, fasting/feeding, and rest/activity can feed back to the SCN and thereby act as “non-photic zeitgebers.” By receiving input from both photic and non-photic zeitgebers, the SCN can synchronize robust and coherent circadian rhythms throughout the body. Circadian rhythms within peripheral tissues are maintained by a cell-intrinsic molecular clock, consisting of clock proteins and clock genes.

patterns in murine calvaria and long bones,<sup>(40–42)</sup> indicating that circadian rhythms within bone are indeed maintained through the cell-autonomous molecular clock. Rhythmic clock gene expression has been detected in cultured osteoclasts<sup>(42,43)</sup> as well as osteoblasts,<sup>(44–46)</sup> but it is not yet known whether different cell types within bone demonstrate differential expression patterns of clock genes *in vivo*. It would be of interest to further explore this using novel single-cell sequencing-based approaches,<sup>(47)</sup> although cellular isolation of bone cells required for these techniques is challenging.<sup>(48)</sup> Various genes involved in osteoclast activity (eg, *Ctsk*, *Nfatc*, *Rankl*, *Opg*) show potent diurnal expression patterns in bone,<sup>(42,43,49)</sup> while the osteoblast markers *Runx2* and *Col1a1* were found not to be rhythmic.<sup>(40)</sup> However, as *Runx2* is mostly involved in osteoblast differentiation<sup>(50)</sup> and *Col1a1* lacks specificity for osteoblasts,<sup>(51)</sup> this does not necessarily preclude rhythmic osteoblast activity. Genetic disruption of clock genes<sup>(52–54)</sup> as well as environmental circadian disruption through shifting light/dark cycles<sup>(40)</sup> have been shown to affect bone mass in mice, stressing the importance of circadian rhythm in bone remodeling for maintaining skeletal integrity.

## 2.1 Rhythmic bone resorption

Bone remodeling can be assessed *in vivo* by measuring markers of bone resorption and bone formation. Many clinical studies reported a diurnal rhythm in the bone resorption marker carboxy-terminal collagen cross-links (CTX), which is a product of bone collagen degradation. In healthy men and women, CTX levels in serum peak at night or early morning,<sup>(55–59)</sup> indicating that bone resorption is particularly high during the resting phase. While mechanical unloading due to physical inactivity would be an obvious explanation for the increased bone resorption at night, CTX rhythm was found to be completely unaffected by timing of rest/activity.<sup>(57)</sup> Variation in CTX is also not explained

by diurnal variation in cortisol,<sup>(57,60)</sup> nor by light input,<sup>(57)</sup> but CTX is known to be significantly reduced upon fasting,<sup>(57)</sup> suggesting that rhythm in bone resorption is primarily mediated through cycles of fasting/feeding. This was confirmed in a randomized study in postmenopausal women showing that bone resorption is reduced upon food intake and increased upon prolonged fasting.<sup>(61)</sup> These results could partly be reproduced by exogenous and endogenous insulin stimulation.<sup>(61)</sup> The effect of food intake on bone remodeling is also mediated by gut-derived incretin hormones, including glucose-dependent insulinotropic polypeptide (GIP), glucagon-like peptide 1 (GLP-1), and glucagon-like peptide 2 (GLP-2). There are receptors for GIP and GLP-1 present on osteoblasts, and receptors for GLP-2 on osteoclasts, as nicely reviewed by Yavropoulou and colleagues.<sup>(62)</sup> In line with this notion, oral glucose loading results in an acute and more prominent decrease in bone resorption than intravenous glucose loading.<sup>(63)</sup> The response involves somatostatin, since the effect of oral glucose on bone can be abolished by octreotide.<sup>(64)</sup> Of note, rhythmic bone resorption remains present during fasting, although less pronounced.<sup>(65)</sup> Thus, fasting/feeding cycles likely have an effect on bone resorption independent of the SCN, but the exact underlying mechanisms remains uncharacterized.

It has been proposed that PTH also contributes to rhythmic activity in bone resorption. PTH levels demonstrate a diurnal rhythm, with peak values at night and a nadir in the morning.<sup>(66)</sup> In turn, PTH rhythmicity may also be a reaction to rhythmic food intake, since PTH is the most important player in the negative feedback loop aiming for stable serum calcium levels within a narrow window. Serum PTH and calcium levels have a strong bidirectional and temporal relationship.<sup>(67)</sup> Theoretically, an absence of dietary calcium intake at night will result in a subsequent increase in PTH. Indeed, PTH rhythmicity is diminished by prolonged fasting, at least in healthy premenopausal women.<sup>(65)</sup> Fasting resulted in an increase in serum calcium

levels and bone resorption, which could explain a subsequent decrease in PTH as a result from the negative feedback loop. Human subjects in which PTH rhythm was blunted by continuous calcium infusion still showed a diurnal pattern in urinary excretion of amino-terminal collagen cross-links (NTX),<sup>(68)</sup> a bone resorption marker comparable to CTX. Intrinsic PTH rhythmicity may in itself thus contribute to rhythmic bone resorption, but this will be overruled in case of disturbances in the calcium homeostasis. Of note, long-term PTH exposure should be discriminated from intermittent PTH peaks. It is well known that administration of daily intermittent doses of PTH in humans promotes bone formation and increases bone mass by stimulating osteoblasts without inducing the osteoclastogenesis that occurs with continuous exposure to PTH.<sup>(22,69)</sup> Therefore, we anticipate that short-lived circadian peaks in PTH are more likely to affect osteoblast activity than osteoclast activity. Future studies are needed to identify the potential role of PTH and other calcium-regulating hormones in rhythmic bone formation. For example, osteocalcin and vitamin D demonstrate a diurnal rhythm in humans<sup>(70–72)</sup> and are known to modulate clock gene expression in bone of rats,<sup>(73,74)</sup> but they are mainly regulators of calcium homeostasis rather than major determinants of rhythmic osteoclast activity and bone resorption.

GCs directly affect the core clock in several ways, for example, by stimulation of *Per1/2* expression, to act as an extrinsic driving force on the intrinsic oscillators.<sup>(75)</sup> Bone rhythms are GC sensitive: Rhythmic bone resorption depends on rhythmic GCs. Circadian oscillations in gene expression in osteoclasts are affected by GC signaling. The synthetic GC dexamethasone has been shown to induce rhythmic gene expression in cultured osteoclasts,<sup>(42,43)</sup> and GC depletion by adrenalectomy in mice abolishes rhythm in clock genes as well as osteoclast-related genes in bone.<sup>(42)</sup> A single injection of GCs can restore circadian gene expression in adrenalectomized mice,<sup>(42)</sup> demonstrating the potency of GCs as a circadian timing signal for bone. Because GCs have been shown to stimulate the production of RANKL by osteoblasts, this thus promotes osteoclastic bone resorption through RANK-RANKL signaling,<sup>(76)</sup> which in itself has been shown to be rhythmic.<sup>(77)</sup> However, GC-dependent bone resorption is not rhythmic.<sup>(57,60)</sup> This is in contrast to osteocalcin for which rhythmicity depends on rhythmic cortisol expression<sup>(60)</sup> as described below.

Although osteocytes have been suggested to mediate rhythmic bone resorption by promoting osteoclast differentiation and function,<sup>(78)</sup> the osteocyte marker sclerostin does not show diurnal variation in human subjects, nor does it predict variation in CTX levels.<sup>(55)</sup> Also, while global and osteoblast-specific *Bmal1* deficiency in mice results in a low bone mass phenotype due to enhanced osteoclastogenesis and increased bone resorption,<sup>(54,77)</sup> this phenotype was not recapitulated by osteocyte-specific *Bmal1* deletion.<sup>(77)</sup>

## 2.2 Rhythmic bone formation

Albeit less pronounced than markers of bone resorption, markers of bone formation also demonstrate 24-hour serum profiles. One study found a minor rhythm in the commonly used bone formation marker procollagen type 1 N-terminal propeptide (P1NP) across different ethnic groups,<sup>(71)</sup> but this was not observed in many other studies.<sup>(55,58,79,80)</sup> The bone matrix protein osteocalcin, which is produced by active osteoblasts, shows a more robust diurnal rhythm in human serum.<sup>(58,59,81,82)</sup> As with CTX, osteocalcin levels are higher at nighttime compared with

daytime, suggesting that osteoblast activity and therefore bone formation is also highest during the resting phase. Osteocalcin levels are not affected by fasting,<sup>(65)</sup> but elimination of the morning peak in cortisol abolishes the expected morning decrease in osteocalcin in healthy individuals.<sup>(60,83)</sup> Thus, circadian GC rhythm is an important determinant of diurnal variation in osteoblast activity, which is supported by in vitro studies showing that dexamethasone can induce a rhythm in osteoblasts<sup>(46)</sup> and that the endogenous GC peak has an inhibitory effect on bone formation. In fact, endogenous GCs are required to maintain bone health, as demonstrated by diminished osteoblast differentiation and progressive bone loss upon strongly attenuated GC signaling in mice through adrenalectomy<sup>(84)</sup> and osteoblast-specific disruption of GC action.<sup>(85–87)</sup> However, excess GCs negatively impact bone formation by attenuating osteoblast differentiation.<sup>(85,88)</sup> GCs affect the Wnt signaling pathway, a critical regulator of osteoblastogenesis, in a dose-dependent manner, with upregulation of Wnt at lower doses and downregulation at higher doses.<sup>(89)</sup> It can be postulated that low GC concentrations observed at the natural trough of circadian GC rhythm could enable bone formation, while peak levels may have an inhibitory effect. However, this remains speculative and requires further investigation.

In contrast to GC signaling, which affects both osteoblasts and osteoclasts, signaling through adrenergic receptors modulates rhythm selectively in osteoblasts. The  $\beta$ -adrenergic receptor agonist isoprenaline was found to promote clock gene oscillations in cultured osteoblasts<sup>(45,46)</sup> but not osteoclasts.<sup>(42)</sup> Consistent with these observations, genetic ablation of  $\alpha$ 1-adrenergic receptor signaling in mice disrupts the expression of osteoblast-related and clock genes in bone,<sup>(44)</sup> and treatment of mice with the  $\alpha$ -adrenergic receptor antagonist prazosin lowers bone mass by reducing bone formation.<sup>(90)</sup> These findings collectively indicate that bone formation may be regulated through sympathetic nerve system activity, which is known to transmit signals from the SCN to peripheral tissue clocks. Nevertheless, additional studies are needed to confirm a direct relationship between adrenergic signaling, the circadian clock in bone, and rhythmic bone remodeling.

Bone formation is also regulated by melatonin, a hormone produced by the pineal gland in response to photic input from the SCN, at least in rodents. Melatonin release is inhibited by light and peaks during the dark phase when markers of bone formation are also high. Accordingly, melatonin has been found also to stimulate human osteoblast differentiation and proliferation in vitro<sup>(91,92)</sup> and to promote bone formation in animal models in vivo.<sup>(92,93)</sup> Melatonin depletion by pinealectomy as well as long-term melatonin administration affect circadian oscillations in bone formation markers in rats,<sup>(94)</sup> suggesting a role of melatonin in rhythmic bone formation. However, melatonin administration does not affect circulating osteocalcin levels or bone density in humans.<sup>(95)</sup> In addition, melatonin has been shown to suppress activation of osteoclasts in mice through downregulation of RANKL on osteoblasts,<sup>(96)</sup> but there is no significant relationship between melatonin rhythm and NTX rhythm in humans.<sup>(97)</sup> These studies demonstrate that at least in humans, melatonin rhythm may not be an important determinant of bone remodeling.

## 2.3 Rhythmic bone growth

Like bone formation and bone resorption, it can be understood that skeletal growth is time-of-day dependent.<sup>(98)</sup> Growth plates

and their chondrocytes exhibit a strong circadian expression pattern of clock genes,<sup>(99)</sup> including *Bmal1*.<sup>(100)</sup> BMAL1 appears critical to growth plate development, as *Bmal1*-deficient mice have significantly shorter femora and tibiae.<sup>(54)</sup> It is even suggested that a chondrocyte-specific peripheral clock might exist,<sup>(98)</sup> which could be an interesting topic for further research. PTH directly regulates circadian oscillation of the clock genes in chondrocytes<sup>(101)</sup> and is able to reset the robust circadian rhythm in chondroprogenitor cells.<sup>(102)</sup> This circadian oscillation is not only crucial for bone growth during aging but also important for endochondral bone formation during fracture healing.<sup>(103)</sup> Besides, FGF23 expression, which itself depends on the circadian clock, food intake, and sympathetic activity<sup>(104)</sup> and phosphate influence skeletal growth in a circadian fashion.<sup>(105)</sup> The circadian rhythmicity within growth plates seems thus an interesting field for further research.

### 3. Chronotherapy of Osteoporosis

#### 3.1 Prevention of osteoporosis associated with circadian disruption

Preventing disruption of the circadian timing system can be used to improve bone health of the general population. Shift work is likely the most widespread lifestyle associated with osteoporosis. Postmenopausal nurses who work in rotating shifts were reported to have a lower bone mineral density at the lumbar spine as well as femoral neck bones compared with daytime workers of the same age and sex.<sup>(28)</sup> Although more than 25% of shift workers had osteoporosis at the lumbar spine (defined by a *T*-score < -2.5), this was not observed for any of the daytime workers. In addition, women who worked in night shifts for more than 20 years demonstrated a significantly increased risk of wrist and hip fractures.<sup>(27)</sup> These negative effects of shift work on bone could be a direct consequence of disruption of the circadian timing system. However, shift workers have also been suggested to maintain an overall healthier lifestyle (eg, increased smoking, a higher alcohol intake, lower physical activity, and altered eating habits), which may be a confounding factor. Moreover, shift workers and day workers may not be comparable with respect to their income and the type of work they perform. Nevertheless, a recent intervention study showed that circadian disruption in combination with sleep restriction also decreases P1NP levels in healthy subjects,<sup>(106)</sup> pointing toward reduced bone formation. We have recently demonstrated that repeated shifts in light/dark cycle in mice, as a way to mimic human shift work, reduces markers of bone remodeling (ie, P1NP and CTX) and alters the material and structural properties of bone.<sup>(40)</sup> In addition, continuous light exposure has been shown to diminish bone volume and density in mice.<sup>(107)</sup> This could be the result of a disrupted circadian clock in combination with sleep deprivation, as sleep duration positively correlates to bone stiffness in humans,<sup>(108)</sup> and short sleep is associated with low bone mineral density.<sup>(109,110)</sup> These studies collectively support a causal relationship between environmental circadian disruption and osteoporosis. Although it is unclear whether the increased risk of osteoporosis in shift workers is a consequence of excessive or inadequate bone turnover, circadian rhythms are markedly dampened in older individuals,<sup>(111)</sup> indicating that osteoporosis associated with circadian disruption and aging could have a shared etiology.

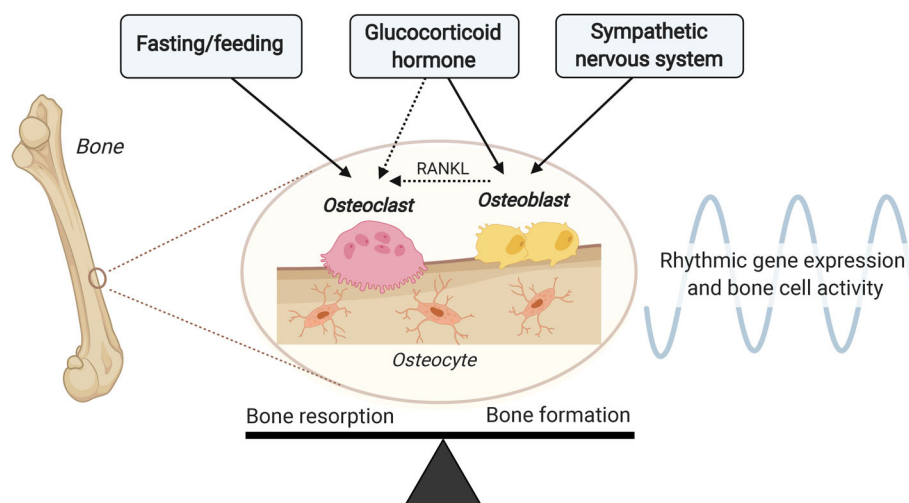
Considering the large number of individuals with irregular working schedules (ie, nowadays more than 20% of the

population in industrialized societies is involved in some form of shift work), strategies to preserve bone health in shift workers are highly warranted. Importantly, such strategies may also reduce the risk of age-related osteoporosis in these individuals, by strengthening intrinsic circadian rhythms. Prevention-focused strategies should preferably involve lifestyle interventions instead of pharmaceutical therapies that are costly and often accompanied by adverse effects. Two promising intervention strategies come to mind, ie, timed eating and timed exercise. In the previous sections, we discussed the importance of fasting/feeding cycles in regulating rhythmic bone remodeling. As bone resorption is increased by fasting at night and suppressed by feeding throughout the day, rhythmic patterns of food intake could strengthen rhythmic bone remodeling. Moreover, timing of food intake has been shown to entrain rhythm in a variety of peripheral tissues<sup>(32)</sup> as well as in the circadian master clock (ie, the SCN),<sup>(112)</sup> which was classically thought to be only affected by light. We have shown that time-restricted feeding improves adaptation to shifting light/dark cycles in mice,<sup>(49)</sup> indicating that timed eating could be an effective strategy to limit circadian disruption in shift workers.

In addition to food intake, physical activity has well-characterized effects on bone metabolism. Mechanical forces that are exerted on bone through exercise, depending on their direction and magnitude, reduce the production of sclerostin by osteocytes.<sup>(113)</sup> This relieves the inhibition of sclerostin on the Wnt signaling pathway, thereby promoting osteoblast differentiation.<sup>(114)</sup> In addition, Wnt signaling promotes the expression of osteoprotegerin (OPG) by osteoblasts,<sup>(115)</sup> which is a decoy receptor for RANKL, thus preventing RANK-RANKL interaction and osteoclastogenesis. These dual effects of Wnt on osteoblasts and osteoclasts result in a net increase in bone mass.<sup>(116)</sup> As such, high-force exercise such as progressive resistance strength training mildly increases bone density in postmenopausal women<sup>(117)</sup> and is proposed as a safe and effective way to prevent bone loss. Although it remains to be investigated whether timed exercise also entrains rhythm in bone, exercise during the night shift has been shown to adapt circadian temperature rhythms in humans,<sup>(118)</sup> stressing the importance of physical activity as a circadian timing signal. Moreover, exercise may be used to adjust a person's late acrophase into early acrophase, as the former is associated with increased fall risk.<sup>(119)</sup> Thus, both timed feeding and timed exercise may be implemented to prevent disruption of the biological clock and reduce the associated risk of osteoporosis.

#### 3.2 Prevention of osteoporosis by correct timing of bone-sparing agents

As bone metabolism exhibits potent circadian rhythms, it is not surprising that the response of bone to pharmacological treatment is time dependent. Various studies have shown that application of chronotherapy, ie, a method of treatment in which the administration of medication is coordinated with the biological clock, is an effective strategy to improve therapeutic efficacy of osteoporosis medication. Treatment of postmenopausal osteoporotic women with the recombinant PTH teriparatide differentially affects diurnal rhythm in bone turnover markers depending on the time of administration.<sup>(80)</sup> While subjects treated with teriparatide in the evening showed a pronounced peak in serum CTX at night, morning teriparatide administration more effectively diminished CTX levels, indicating that timing of teriparatide treatment could be important. Accordingly, 12-month



**Fig 3.** Circadian regulation of bone resorption by osteoclasts and bone formation by osteoblasts. Healthy bone depends on a delicate balance between bone resorption by osteoclasts and bone formation by osteoblasts. Studies indicate that these processes are strongly regulated by the circadian timing system, thereby producing diurnal variation in bone remodeling. While cycles of fasting/feeding primarily dictate osteoclast activity, sympathetic nervous system activity may selectively regulate rhythmic osteoblast activity. Glucocorticoid (GC) signaling is likely important for rhythm in both bone resorption and bone formation, as GCs can modulate the cellular clock of osteoclasts and osteoblasts, respectively. Of note, GCs could also affect rhythm in osteoclast activity indirectly through osteoblasts (as indicated by the dashed arrow), by regulating expression of the RANKL.

treatment of osteoporotic women with teriparatide in the morning was shown to be more effective in increasing bone mineral density compared with treatment in the evening.<sup>(120)</sup> Although as yet not clinically relevant, the effect of timed calcitonin treatment is illustrative: Healthy women who were treated with oral calcitonin just before dinner or in the evening showed a larger reduction in CTX compared with treatment in the morning,<sup>(121)</sup> in line with the opposing actions of PTH and calcitonin. However, differences in baseline CTX levels and fasting state are confounding factors in this study. For many novel drugs to treat osteoporosis, chronotherapy is likely relevant yet unexplored. These drugs include abaloparatide, a PTH-related protein that has recently been shown to have superior anabolic effects on bone compared with teriparatide,<sup>(122,123)</sup> and selective estrogen receptor modulators (SERMs) such as raloxifene<sup>(124,125)</sup> and bazedoxifene,<sup>(126–128)</sup> which are used to prevent and treat postmenopausal osteoporosis. Chronotherapy may be less relevant for the current standard treatment with bisphosphonates because of their very long half-life,<sup>(129)</sup> although chronotherapy might improve the poor oral bioavailability (<1% of the administered dose)<sup>(130)</sup> due to diurnal variation in intestinal absorption. In addition, monoclonal antibodies such as denosumab, which targets RANKL, and the more novel romosozumab, which targets sclerostin, have a half-life of a few weeks,<sup>(131,132)</sup> rendering chronotherapy to increase therapeutic efficacy of these compounds less likely. Nevertheless, 39% of clinical trials of drugs with a long half-life (more than 15 hours) still show dosing time dependence,<sup>(133)</sup> indicating that variability in pharmacokinetics across the 24-hour day could also modulate the effectiveness of longer-acting drugs. Moreover, time-of-day-dependent variation in pharmacokinetics is an important determinant of adverse drug effects, as explained below.

The bioavailability of a drug is dependent on circadian timing of the intestine, liver, and kidneys, which together importantly regulate the absorption, distribution, metabolism, and excretion

of a drug.<sup>(134)</sup> Accordingly, if biological timing is considered when administering a drug, this could not only optimize therapeutic outcomes but also reduce adverse effects. Indeed, a proof-of-principle study demonstrated that treatment of osteoporotic rats with vitamin D at the start of the dark phase instead of the light phase is not only more effective but also results into fewer side effects, such as hypercalcemia.<sup>(135)</sup> This thus indicates that chronotherapy is a promising strategy to reduce or prevent adverse effects associated with osteoporosis treatment in humans, such as hypercalcemia-induced dizziness and leg cramps in the case of PTH treatment.<sup>(69)</sup>

Another way in which chronotherapy could reduce negative effects on bone is by preventing GC-induced osteoporosis, which is the most common cause of secondary osteoporosis.<sup>(136)</sup> It was long thought that negative effects of GCs on bone are solely attributable to supraphysiological GC levels resulting from GC treatment, but we have recently shown that a disturbed GC rhythm could also contribute substantially.<sup>(137)</sup> This finding is in line with the pivotal role of GC rhythm in mediating healthy bone remodeling, as discussed above, and emphasizes the importance of timing of GC therapy to limit its adverse effects.

#### 4. Concluding remarks

It has become clear that circadian rhythm plays a pivotal role in maintaining bone health by regulating bone remodeling. Evident rhythms in osteocyte activity have not (yet) been observed, but based on pronounced rhythms in the bone resorption marker CTX and the bone formation marker osteocalcin, combined with the data on rhythmic gene expression and the effects that hormones such as GCs have on these rhythms, we may conclude that both osteoclasts and osteoblasts possess robust diurnal activity patterns in humans with a peak during the rest phase. While rhythm in osteoclastic bone resorption is primarily

regulated through fasting/feeding cycles and potentially fine-tuned by GC hormone levels, rhythm in osteoblastic bone resorption seems mainly mediated by GC hormone levels and sympathetic nervous system activity, as illustrated in Fig. 3. Calcium-regulating hormones and melatonin have also been implicated in rhythmic bone remodeling in cell cultures and rodents, but so far this is not supported by evidence in humans. Although a solid foundation has been laid, critical knowledge on the exact regulatory mechanisms of rhythmic bone remodeling is still scarce, and the importance of additional time cues, such as rhythm in physical activity and feeding, remains to be investigated. Future studies should also focus on investigating whether osteoclasts and osteoblasts are differentially regulated by different circadian timing signals and how this could be manipulated to promote bone strength and reduce bone remodeling disorders.

A significant part of the general population suffers from circadian disruption, thereby increasing the incidence of the bone remodeling disorder osteoporosis. Best established, osteoporosis risk is increased in the growing population of shift workers, which could be attenuated by promising novel interventions that prevent disruption of the circadian timing system, such as time-restricted eating and/or time-restricted exercise. In addition, osteoporosis risk may be increased in many individuals through mistimed administration of synthetic GCs, which disrupts the endogenous cortisol rhythm. Appropriate timing of GC medication could alleviate the risk of GC-induced osteoporosis.

A critical knowledge gap remains for optimally timed anabolic or antiresorptive treatment: Too little is known yet to advise at which time medication should be administered, not only for optimization of therapeutic efficacy but also for prevention of side effects. Although chronotherapy is a very intuitive and promising strategy to improve therapeutic efficacy and reduce adverse effects, it is not yet broadly applied in clinical practice. A recent evaluation of clinical trials reported that less than 0.2% of currently ongoing trials involve a form of circadian intervention, and of those that do involve a form of chronotherapy, only 1% is focused on diseases of muscle, bone, and cartilage.<sup>(138)</sup> These numbers emphasize the need for additional clinical studies that investigate circadian interventions to prevent and/or treat bone diseases such as osteoporosis. Thus, implementation of chronotherapy is crucial and should be further investigated, both for novel and existing therapies.

## Acknowledgments

This work was supported by a personal grant of the Leiden University Fund/Elise Mathilde Fund to EMW and by a personal grant from the Board of Directors of Leiden University Medical Center to MS. Figures were created with BioRender.com.

Authors' roles: Conceptualization, formal analysis, methodology, resources, validation, and writing original draft: EMW and MS. Writing—review and editing: EMW, SK, NMAD, OCM, PCNR, and MS.

## AUTHOR CONTRIBUTIONS

**Elizabeth Winter:** Conceptualization; formal analysis; methodology; resources; ; validation; writing-original draft; writing-review & editing. **Sander Kooijman:** Writing-review & editing. **Natasha**

**Appelman-Dijkstra:** Writing-review & editing. **Onno Meijer:** Writing-review & editing. **Patrick Rensen:** Writing-review & editing. **Maaïke Schilperoort:** Conceptualization; formal analysis; methodology; resources; validation; writing-original draft; writing-review & editing.

## References

- Zaidi M. Skeletal remodeling in health and disease. *Nat Med.* 2007;13(7):791-801.
- Heino TJ, Kurata K, Higaki H, Väänänen HK. Evidence for the role of osteocytes in the initiation of targeted remodeling. *Technol Health Care.* 2009;17(1):49-56.
- Teitelbaum SL. Bone resorption by osteoclasts. *Science.* 2000;289(5484):1504-1508.
- Xian L, Wu X, Pang L, et al. Matrix IGF-1 maintains bone mass by activation of mTOR in mesenchymal stem cells. *Nat Med.* 2012;18(7):1095-1101.
- Tang Y, Wu X, Lei W, et al. TGF-beta1-induced migration of bone mesenchymal stem cells couples bone resorption with formation. *Nat Med.* 2009;15(7):757-765.
- Fiedler J, Röderer G, Günther K-P, Brenner RE. BMP-2, BMP-4, and PDGF-bb stimulate chemotactic migration of primary human mesenchymal progenitor cells. *J Cell Biochem.* 2002;87(3):305-312.
- Sims NA, Martin TJ. Coupling signals between the osteoclast and osteoblast: how are messages transmitted between these temporary visitors to the bone surface? *Front Endocrinol.* 2015;6:41.
- Follet H, Boivin G, Rumelhart C, Meunier PJ. The degree of mineralization is a determinant of bone strength: a study on human calcanei. *Bone.* 2004;34(5):783-789.
- Boivin G, Bala Y, Doublier A, et al. The role of mineralization and organic matrix in the microhardness of bone tissue from controls and osteoporotic patients. *Bone.* 2008;43(3):532-538.
- Cui L, Houston DA, Farquharson C, MacRae VE. Characterisation of matrix vesicles in skeletal and soft tissue mineralisation. *Bone.* 2016;87:147-158.
- Feng JQ, Ward LM, Liu S, et al. Loss of DMP1 causes rickets and osteomalacia and identifies a role for osteocytes in mineral metabolism. *Nat Genet.* 2006;38(11):1310-1315.
- Eastell R, Szulc P. Use of bone turnover markers in postmenopausal osteoporosis. *Lancet Diabetes Endocrinol.* 2017;5(11):908-923.
- Cavalli L, Brandi ML. Age- and gender-related macro- and micro-architecture changes in bone structure and implications for treatment. *Int J Clin Rheumatol.* 2011;6(3):359-369.
- Clarke BL, Khosla S. Androgens and bone. *Steroids.* 2009;74(3):296-305.
- Mohamad NV, Soelaiman IN, Chin KY. A concise review of testosterone and bone health. *Clin Interv Aging.* 2016;11:1317-1324.
- Kameda T, Mano H, Yuasa T, et al. Estrogen inhibits bone resorption by directly inducing apoptosis of the bone-resorbing osteoclasts. *J Exp Med.* 1997;186(4):489-495.
- Smith EP, Boyd J, Frank GR, et al. Estrogen resistance caused by a mutation in the estrogen-receptor gene in a man. *N Engl J Med.* 1994;331(16):1056-1061.
- Bilezikian JP, Morishima A, Bell J, Grumbach MM. Increased bone mass as a result of estrogen therapy in a man with aromatase deficiency. *N Engl J Med.* 1998;339(9):599-603.
- Carani C, Qin K, Simoni M, et al. Effect of testosterone and estradiol in a man with aromatase deficiency. *N Engl J Med.* 1997;337(2):91-95.
- Morishima A, Grumbach MM, Simpson ER, Fisher C, Qin K. Aromatase deficiency in male and female siblings caused by a novel mutation and the physiological role of estrogens. *J Clin Endocrinol Metab.* 1995;80(12):3689-3698.
- Lee SK, Lorenzo JA. Parathyroid hormone stimulates TRANCE and inhibits osteoprotegerin messenger ribonucleic acid expression in murine bone marrow cultures: correlation with osteoclast-like cell formation. *Endocrinology.* 1999;140(8):3552-3561.



22. Silva BC, Bilezikian JP. Parathyroid hormone: anabolic and catabolic actions on the skeleton. *Curr Opin Pharmacol*. 2015;22:41-50.
23. Khundmiri SJ, Murray RD, Lederer E. PTH and vitamin D. *Compr Physiol*. 2016;6(2):561-601.
24. Talmage RV, Grubb SA, Norimatsu H, Vanderwiell CJ. Evidence for an important physiological role for calcitonin. *Proc Natl Acad Sci U S A*. 1980;77(1):609-613.
25. Pouresmaeili F, Kamalidehghan B, Kamarehei M, Goh YM. A comprehensive overview on osteoporosis and its risk factors. *Ther Clin Risk Manag*. 2018;14:2029-2049.
26. Marędziaek M, Śmieszek A, Chrzęstek K, Basinska K, Marycz K. Physical activity increases the total number of bone-marrow-derived mesenchymal stem cells, enhances their osteogenic potential, and inhibits their adipogenic properties. *Stem Cells Int*. 2015;2015:379093.
27. Feskanich D, Hankinson SE, Schernhammer ES. Nightshift work and fracture risk: the Nurses' Health Study. *Osteoporos Int*. 2009;20(4):537-542.
28. Quevedo I, Zuniga AM. Low bone mineral density in rotating-shift workers. *J Clin Densitom*. 2010;13(4):467-469.
29. Dibner C, Schibler U, Albrecht U. The mammalian circadian timing system: organization and coordination of central and peripheral clocks. *Annu Rev Physiol*. 2010;72:517-549.
30. Oster H, Challet E, Ott V, et al. The functional and clinical significance of the 24-hour rhythm of circulating glucocorticoids. *Endocr Rev*. 2017;38(1):3-45.
31. Damiola F, Le Minh N, Preitner N, Kornmann B, Fleury-Olela F, Schibler U. Restricted feeding uncouples circadian oscillators in peripheral tissues from the central pacemaker in the suprachiasmatic nucleus. *Genes Dev*. 2000;14(23):2950-2961.
32. Stokkan KA, Yamazaki S, Tei H, Sakaki Y, Menaker M. Entrainment of the circadian clock in the liver by feeding. *Science*. 2001;291(5503):490-493.
33. Wakamatsu H, Yoshinobu Y, Aida R, Moriya T, Akiyama M, Shibata S. Restricted-feeding-induced anticipatory activity rhythm is associated with a phase-shift of the expression of mPer1 and mPer2 mRNA in the cerebral cortex and hippocampus but not in the suprachiasmatic nucleus of mice. *Eur J Neurosci*. 2001;13(6):1190-1196.
34. Aoyama S, Shibata S. The role of circadian rhythms in muscular and osseous physiology and their regulation by nutrition and exercise. *Front Neurosci*. 2017;11:63.
35. Partch CL, Green CB, Takahashi JS. Molecular architecture of the mammalian circadian clock. *Trends Cell Biol*. 2014;24(2):90-99.
36. Yang X, Downes M, Yu RT, et al. Nuclear receptor expression links the circadian clock to metabolism. *Cell*. 2006;126(4):801-810.
37. Eckel-Mahan KL, Patel VR, Mohney RP, et al. Coordination of the transcriptome and metabolome by the circadian clock. *Proc Natl Acad Sci U S A*. 2012;109(14):5541-5546.
38. Panda S, Antoch MP, Miller BH, et al. Coordinated transcription of key pathways in the mouse by the circadian clock. *Cell*. 2002;109(3):307-320.
39. Zhang R, Lahens NF, Balance HI, Hughes ME, Hogenesch JB. A circadian gene expression atlas in mammals: implications for biology and medicine. *Proc Natl Acad Sci U S A*. 2014;111(45):16219-16224.
40. Schilperoort M, Bravenboer N, Lim J, et al. Circadian disruption by shifting the light-dark cycle negatively affects bone health in mice. *FASEB J*. 2020;34(1):1052-1064.
41. Zvonic S, Ptitsyn AA, Kilroy G, et al. Circadian oscillation of gene expression in murine calvarial bone. *J Bone Miner Res*. 2007;22(3):357-365.
42. Fujihara Y, Kondo H, Noguchi T, Togari A. Glucocorticoids mediate circadian timing in peripheral osteoclasts resulting in the circadian expression rhythm of osteoclast-related genes. *Bone*. 2014;61:1-9.
43. Xu C, Ochi H, Fukuda T, et al. Circadian clock regulates bone resorption in mice. *J Bone Miner Res*. 2016;31(7):1344-1355.
44. Hirai T, Tanaka K, Togari A.  $\alpha$ 1B-adrenergic receptor signaling controls circadian expression of Tnfrsf11b by regulating clock genes in osteoblasts. *Biol Open*. 2015;4(11):1400-1409.
45. Hirai T, Tanaka K, Togari A.  $\beta$ -Adrenergic receptor signaling regulates Ptg2 by driving circadian gene expression in osteoblasts. *J Cell Sci*. 2014;127(Pt 17):3711-3719.
46. Komoto S, Kondo H, Fukuta O, Togari A. Comparison of  $\beta$ -adrenergic and glucocorticoid signaling on clock gene and osteoblast-related gene expressions in human osteoblast. *Chronobiol Int*. 2012;29(1):66-74.
47. Shapiro E, Biezuner T, Linnarsson S. Single-cell sequencing-based technologies will revolutionize whole-organism science. *Nat Rev Genet*. 2013;14(9):618-630.
48. Greenblatt MB, Ono N, Ayturk UM, Debnath S, Lalani S. The unmixing problem: a guide to applying single-cell RNA sequencing to bone. *J Bone Miner Res*. 2019;34(7):1207-1219.
49. Schilperoort M, van den Berg R, Dollé MET, et al. Time-restricted feeding improves adaptation to chronically alternating light-dark cycles. *Sci Rep*. 2019;9(1):7874.
50. Ducy P, Zhang R, Geoffroy V, Ridall AL, Karsenty G. Osf2/Cbfa1: a transcriptional activator of osteoblast differentiation. *Cell*. 1997;89(5):747-754.
51. Dacic S, Kalajzic I, Visnjic D, Lichtler AC, Rowe DW. Col1a1-driven transgenic markers of osteoblast lineage progression. *J Bone Miner Res*. 2001;16(7):1228-1236.
52. Fu L, Patel MS, Bradley A, Wagner EF, Karsenty G. The molecular clock mediates leptin-regulated bone formation. *Cell*. 2005;122(5):803-815.
53. Yuan G, Hua B, Yang Y, et al. The circadian gene clock regulates bone formation via PDIA3. *J Bone Miner Res*. 2017;32(4):861-871.
54. Samsa WE, Vasanji A, Midura RJ, Kondratov RV. Deficiency of circadian clock protein BMAL1 in mice results in a low bone mass phenotype. *Bone*. 2016;84:194-203.
55. Swanson C, Shea SA, Wolfe P, et al. 24-hour profile of serum sclerostin and its association with bone biomarkers in men. *Osteoporos Int*. 2017;28(11):3205-3213.
56. Pellegrini GG, Gonzales Chaves MMS, Fajardo MA, et al. Salivary bone turnover markers in healthy pre- and postmenopausal women: daily and seasonal rhythm. *Clin Oral Investig*. 2012;16(2):651-657.
57. Qvist P, Christgau S, Pedersen BJ, Schlemmer A, Christiansen C. Circadian variation in the serum concentration of C-terminal telopeptide of type I collagen (serum CTx): effects of gender, age, menopausal status, posture, daylight, serum cortisol, and fasting. *Bone*. 2002;31(1):57-61.
58. van der Spoel E, Oei N, Cachucho R, et al. The 24-hour serum profiles of bone markers in healthy older men and women. *Bone*. 2019;120:61-69.
59. Dovio A, Generali D, Tampellini M, et al. Variations along the 24-hour cycle of circulating osteoprotegerin and soluble RANKL: a rhythmometric analysis. *Osteoporos Int*. 2009;19(1):113-117.
60. Heshmati HM, Riggs BL, Burritt MF, McAlister CA, Wollan PC, Khosla S. Effects of the circadian variation in serum cortisol on markers of bone turnover and calcium homeostasis in normal postmenopausal women. *J Clin Endocrinol Metab*. 1998;83(3):751-756.
61. Bjarnason NH, Henriksen EEG, Alexandersen P, Christgau S, Henriksen DB, Christiansen C. Mechanism of circadian variation in bone resorption. *Bone*. 2002;30(1):307-313.
62. Yavropoulou MP, Yovos JG. Incretins and bone: evolving concepts in nutrient-dependent regulation of bone turnover. *Hormones (Athens)*. 2013;12(2):214-223.
63. Clowes JA, Robinson RT, Heller SR, Eastell R, Blumsohn A. Acute changes of bone turnover and PTH induced by insulin and glucose: euglycemic and hypoglycemic hyperinsulinemic clamp studies. *J Clin Endocrinol Metab*. 2002;87(7):3324-3329.
64. Clowes JA, Allen HC, Prentis DM, Eastell R, Blumsohn A. Octreotide abolishes the acute decrease in bone turnover in response to oral glucose. *J Clin Endocrinol Metab*. 2003;88(10):4867-4873.
65. Schlemmer A, Hassager C. Acute fasting diminishes the circadian rhythm of biochemical markers of bone resorption. *Eur J Endocrinol*. 1999;140(4):332-337.

66. Jubiz W, Canterbury JM, Reiss E, Tyler FH. Circadian rhythm in serum parathyroid hormone concentration in human subjects: correlation with serum calcium, phosphate, albumin, and growth hormone levels. *J Clin Invest.* 1972;51(8):2040-2046.
67. Markowitz ME, Arnaud S, Rosen JF, Thorpy M, Laximinarayan S. Temporal interrelationships between the circadian rhythms of serum parathyroid hormone and calcium concentrations. *J Clin Endocrinol Metab.* 1988;67(5):1068-1073.
68. Ledger GA, Burritt MF, Kao PC, O'Fallon WM, Riggs BL, Khosla S. Role of parathyroid hormone in mediating nocturnal and age-related increases in bone resorption. *J Clin Endocrinol Metab.* 1995;80(11):3304-3310.
69. Neer RM, Arnaud CD, Zanchetta JR, et al. Effect of parathyroid hormone (1-34) on fractures and bone mineral density in postmenopausal women with osteoporosis. *N Engl J Med.* 2001;344(19):1434-1441.
70. Radjaipour M, Kindtner E, Rösler H, Eggstein M. Circadian rhythm of parathyrin and calcitonin concentrations in the serum. *J Clin Chem Clin Biochem.* 1986;24(3):175-178.
71. Redmond J, Fulford AJ, Jarjou L, Zhou B, Prentice A, Schoenmakers I. Diurnal rhythms of bone turnover markers in three ethnic groups. *J Clin Endocrinol Metab.* 2016;101(8):3222-3230.
72. Rejnmark L, Lauridsen AL, Vestergaard P, Heickendorff L, Andreasen F, Mosekilde L. Diurnal rhythm of plasma 1,25-dihydroxyvitamin D and vitamin D-binding protein in postmenopausal women: relationship to plasma parathyroid hormone and calcium and phosphate metabolism. *Eur J Endocrinol.* 2002;146(5):635-642.
73. Mengatto CM, Mussano F, Honda Y, Colwell CS, Nishimura I. Circadian rhythm and cartilage extracellular matrix genes in osseointegration: a genome-wide screening of implant failure by vitamin D deficiency. *PLoS One.* 2011;6(1):e15848.
74. Shao P, Shinoda H. Effect of calcitonin on the expression of clock genes in rat bone and kidney. *Int Congr Ser.* 2005;1284:342-343.
75. So AY, Bernal TU, Pillsbury ML, Yamamoto KR, Feldman BJ. Glucocorticoid regulation of the circadian clock modulates glucose homeostasis. *Proc Natl Acad Sci U S A.* 2009;106(41):17582-17587.
76. Xiong J, Onal M, Jilka RL, et al. Matrix-embedded cells control osteoclast formation. *Nat Med.* 2011;17(10):1235-1241.
77. Takarada T, Xu C, Ochi H, et al. Bone resorption is regulated by circadian clock in osteoblasts. *J Bone Miner Res.* 2017;32(4):872-881.
78. O'Brien CA, Nakashima T, Takayanagi H. Osteocyte control of osteoclastogenesis. *Bone.* 2013;54(2):258-263.
79. Wolthers OD, Heuck C, Heickendorff L. Diurnal variations in serum and urine markers of type I and type III collagen turnover in children. *Clin Chem.* 2001;47(9):1721-1722.
80. Luchavova M, Zikan V, Michalska D, Raska I Jr, Kubena AA, Stepan JJ. The effect of timing of teriparatide treatment on the circadian rhythm of bone turnover in postmenopausal osteoporosis. *Eur J Endocrinol.* 2011;164(4):643-648.
81. Pietschmann P, Resch H, Woloszczuk W, Willvonseder R. A circadian rhythm of serum osteocalcin levels in postmenopausal osteoporosis. *Eur J Clin Invest.* 1990;20(3):310-312.
82. Greenspan SL, Dresner-Pollak R, Parker RA, London D, Ferguson L. Diurnal variation of bone mineral turnover in elderly men and women. *Calcif Tissue Int.* 1997;60(5):419-423.
83. Nielsen HK, Brixen K, Kassem M, Charles P, Mosekilde L. Inhibition of the morning cortisol peak abolishes the expected morning decrease in serum osteocalcin in normal males: evidence of a controlling effect of serum cortisol on the circadian rhythm in serum osteocalcin. *J Clin Endocrinol Metab.* 1992;74(6):1410-1414.
84. Durbridge TC, Morris HA, Parsons AM, et al. Progressive cancellous bone loss in rats after adrenalectomy and oophorectomy. *Calcif Tissue Int.* 1990;47(6):383-387.
85. Rauch A, Seitz S, Baschant U, et al. Glucocorticoids suppress bone formation by attenuating osteoblast differentiation via the monomeric glucocorticoid receptor. *Cell Metab.* 2010;11(6):517-531.
86. Sher LB, Harrison JR, Adams DJ, Kream BE. Impaired cortical bone acquisition and osteoblast differentiation in mice with osteoblast-targeted disruption of glucocorticoid signaling. *Calcif Tissue Int.* 2006;79(2):118-125.
87. Sher LB, Woitge HW, Adams DJ, et al. Transgenic expression of 11beta-hydroxysteroid dehydrogenase type 2 in osteoblasts reveals an anabolic role for endogenous glucocorticoids in bone. *Endocrinology.* 2004;145(2):922-929.
88. Han L, Wang B, Wang R, et al. The shift in the balance between osteoblastogenesis and adipogenesis of mesenchymal stem cells mediated by glucocorticoid receptor. *Stem Cell Res Ther.* 2019;10(1):377.
89. Mak W, Shao X, Dunstan CR, et al. Biphasic glucocorticoid-dependent regulation of Wnt expression and its inhibitors in mature osteoblastic cells. *Calcif Tissue Int.* 2009;85(6):538-545.
90. Tanaka K, Rudelius M, Bierie B.  $\alpha$ 1B-Adrenoceptor signalling regulates bone formation through the up-regulation of CCAAT/enhancer-binding protein  $\delta$  expression in osteoblasts. *Br J Pharmacol.* 2016;173(6):1058-1069.
91. Nakade O, Koyama H, Arijji H, Yajima A, Kaku T. Melatonin stimulates proliferation and type I collagen synthesis in human bone cells in vitro. *J Pineal Res.* 1999;27(2):106-110.
92. Satomura K, Tobiume S, Tokuyama R, et al. Melatonin at pharmacological doses enhances human osteoblastic differentiation in vitro and promotes mouse cortical bone formation in vivo. *J Pineal Res.* 2007;42(3):231-239.
93. Ladizesky MG, Cutrera RA, Boggio V, et al. Effect of melatonin on bone metabolism in ovariectomized rats. *Life Sci.* 2001;70(5):557-565.
94. Ostrowska Z, Kos-Kudla B, Nowak M, et al. The relationship between bone metabolism, melatonin and other hormones in sham-operated and pinealectomized rats. *Endocr Regul.* 2003;37(4):211-224.
95. Kotlarczyk MP, Lassila HC, O'Neil CK, et al. Melatonin Osteoporosis Prevention Study (MOPS): a randomized, double-blind, placebo-controlled study examining the effects of melatonin on bone health and quality of life in perimenopausal women. *J Pineal Res.* 2012;52(4):414-426.
96. Koyama H, Nakade O, Takada Y, Kaku T, Lau KHW. Melatonin at pharmacologic doses increases bone mass by suppressing resorption through down-regulation of the RANKL-mediated osteoclast formation and activation. *J Bone Miner Res.* 2002;17(7):1219-1229.
97. St Hilaire MA, Rahman SA, Gooley JJ, Witt-Enderby PA, Lockley SW. Relationship between melatonin and bone resorption rhythms in premenopausal women. *J Bone Miner Metab.* 2019;37(1):60-71.
98. Samsa WE, Zhou X, Zhou G. Signaling pathways regulating cartilage growth plate formation and activity. *Semin Cell Dev Biol.* 2017;62:3-15.
99. Okubo N, Minami Y, Fujiwara H, et al. Prolonged bioluminescence monitoring in mouse ex vivo bone culture revealed persistent circadian rhythms in articular cartilages and growth plates. *PLoS One.* 2013;8(11):e78306.
100. Takarada T, Kodama A, Hotta S, et al. Clock genes influence gene expression in growth plate and endochondral ossification in mice. *J Biol Chem.* 2012;287(43):36081-36095.
101. Hinoi E, Ueshima T, Hojo H, et al. Up-regulation of per mRNA expression by parathyroid hormone through a protein kinase A-CREB-dependent mechanism in chondrocytes. *J Biol Chem.* 2006;281(33):23632-23642.
102. Hosokawa T, Tsuchiya Y, Okubo N, Kunimoto T, Minami Y. Robust circadian rhythm and parathyroid hormone-induced resetting during hypertrophic differentiation in ATDC5 chondroprogenitor cells. *Acta Histochem Cytochem.* 2015;48(6):165-171.
103. Kunimoto T, Okubo N, Minami Y, et al. A PTH-responsive circadian clock operates in ex vivo mouse femur fracture healing site. *Sci Rep.* 2016;6:22409.
104. Kawai M, Kinoshita S, Shimba S, Ozono K, Michigami T. Sympathetic activation induces skeletal Fgf23 expression in a circadian rhythm-dependent manner. *J Biol Chem.* 2014;289(3):1457-1466.
105. Noguchi T, Hussein AI, Horowitz N, et al. Hypophosphatemia regulates molecular mechanisms of circadian rhythm. *Sci Rep.* 2018;8(1):13756.

106. Swanson CM, Shea SA, Kohrt WM, Wright KP. Sleep restriction with circadian disruption negatively alter bone turnover markers in women. *J Clin Endocrinol Metab.* 2020;105(7):2456-2463.
107. Lucassen EA, Coomans CP, van Putten M, de Kreijl SR. Environmental 24-hr cycles are essential for health. *Curr Biol.* 2016;26(14):1843-1853.
108. Cheng L, Pohlabeln H, Ahrens W, et al. Cross-sectional and longitudinal associations between sleep duration, sleep quality, and bone stiffness in European children and adolescents. *Osteoporos Int.* 2021;32(5):853-863.
109. Albayrak I, Aydogmus M, Ozerbil OM, Levendoglu F. The association between bone mineral density, quality of life, quality of sleep and fatigue. *Acta Clin Belg.* 2016;71(2):92-98.
110. Cunningham TD, Di Pace BS. Is self-reported sleep duration associated with osteoporosis? Data from a 4-year aggregated analysis from the National Health and Nutrition Examination Survey. *J Am Geriatr Soc.* 2015;63(7):1401-1406.
111. Hood S, Amir S. The aging clock: circadian rhythms and later life. *J Clin Invest.* 2017;127(2):437-446.
112. Castillo MR, Hochstetler KJ, Tavernier RJ Jr, Greene DM, Bult-Ito A. Entrainment of the master circadian clock by scheduled feeding. *Am J Physiol Regul Integr Comp Physiol.* 2004;287(3):R551-R555.
113. Robling AG, Niziolek PJ, Baldrige LA, et al. Mechanical stimulation of bone in vivo reduces osteocyte expression of Sost/sclerostin. *J Biol Chem.* 2008;283(9):5866-5875.
114. Krishnan V, Bryant HU, Macdougald OA. Regulation of bone mass by Wnt signaling. *J Clin Invest.* 2006;116(5):1202-1209.
115. Glass DA 2nd, Bialek P, Ahn JD, et al. Canonical Wnt signaling in differentiated osteoblasts controls osteoclast differentiation. *Dev Cell.* 2005;8(5):751-764.
116. Canalis E. Wnt signalling in osteoporosis: mechanisms and novel therapeutic approaches. *Nat Rev Endocrinol.* 2013;9(10):575-583.
117. Howe TE, Shea B, Dawson LJ, et al. Exercise for preventing and treating osteoporosis in postmenopausal women. *Cochrane Database Syst Rev.* 2011;(7):Cd000333. PMID: 21735380.
118. Eastman CI, Hoese EK, Youngstedt SD, Liu L. Phase-shifting human circadian rhythms with exercise during the night shift. *Physiol Behav.* 1995;58(6):1287-1291.
119. Rogers TS, Blackwell TL, Lane NE, et al. Rest-activity patterns and falls and fractures in older men. *Osteoporos Int.* 2017;28(4):1313-1322.
120. Michalska D, Luchavova M, Zikan V, Raska I Jr, Kubena AA, Stepan JJ. Effects of morning vs. evening teriparatide injection on bone mineral density and bone turnover markers in postmenopausal osteoporosis. *Osteoporos Int.* 2012;23(12):2885-2891.
121. Karsdal MA, Byrjalsen I, Riis BJ, Christiansen C. Investigation of the diurnal variation in bone resorption for optimal drug delivery and efficacy in osteoporosis with oral calcitonin. *BMC Clin Pharmacol.* 2008;8:12.
122. Miller PD, Hattersley G, Riis BJ, et al. Effect of abaloparatide vs placebo on new vertebral fractures in postmenopausal women with osteoporosis: a randomized clinical trial. *JAMA.* 2016;316(7):722-733.
123. Leder BZ, O'Dea LSL, Zanchetta JR, et al. Effects of abaloparatide, a human parathyroid hormone-related peptide analog, on bone mineral density in postmenopausal women with osteoporosis. *J Clin Endocrinol Metab.* 2015;100(2):697-706.
124. Ettinger B, Black DM, Mitlak BH, et al. Reduction of vertebral fracture risk in postmenopausal women with osteoporosis treated with raloxifene: results from a 3-year randomized clinical trial. Multiple Outcomes of Raloxifene Evaluation (MORE) investigators. *JAMA.* 1999;282(7):637-645.
125. Delmas PD, Ensrud KE, Adachi JD, et al. Efficacy of raloxifene on vertebral fracture risk reduction in postmenopausal women with osteoporosis: four-year results from a randomized clinical trial. *J Clin Endocrinol Metab.* 2002;87(8):3609-3617.
126. Silverman SL, Chines AA, Kendler DL, et al. Sustained efficacy and safety of bazedoxifene in preventing fractures in postmenopausal women with osteoporosis: results of a 5-year, randomized, placebo-controlled study. *Osteoporos Int.* 2012;23(1):351-363.
127. Silverman SL, Christiansen C, Genant HK, et al. Efficacy of bazedoxifene in reducing new vertebral fracture risk in postmenopausal women with osteoporosis: results from a 3-year, randomized, placebo-, and active-controlled clinical trial. *J Bone Miner Res.* 2008;23(12):1923-1934.
128. Miller PD, Chines AA, Christiansen C, et al. Effects of bazedoxifene on BMD and bone turnover in postmenopausal women: 2-yr results of a randomized, double-blind, placebo-, and active-controlled study. *J Bone Miner Res.* 2008;23(4):525-535.
129. Khan SA, Kanis JA, Vasikaran S, et al. Elimination and biochemical responses to intravenous alendronate in postmenopausal osteoporosis. *J Bone Miner Res.* 1997;12(10):1700-1707.
130. Mitchell DY, Barr WH, Eusebio RA, et al. Risedronate pharmacokinetics and intra- and inter-subject variability upon single-dose intravenous and oral administration. *Pharm Res.* 2001;18(2):166-170.
131. Yonemori K, Fujiwara Y, Minami H, et al. Phase 1 trial of denosumab safety, pharmacokinetics, and pharmacodynamics in Japanese women with breast cancer-related bone metastases. *Cancer Sci.* 2008;99(6):1237-1242.
132. Padhi D, Jang G, Stouch B, Fang L, Posvar E. Single-dose, placebo-controlled, randomized study of AMG 785, a sclerostin monoclonal antibody. *J Bone Miner Res.* 2011;26(1):19-26.
133. Ruben MD, Smith DF, FitzGerald GA, Hogenesch JB. Dosing time matters. *Science.* 2019;365(6453):547-549.
134. Dallmann R, Brown SA, Gachon F. Chronopharmacology: new insights and therapeutic implications. *Annu Rev Pharmacol Toxicol.* 2014;54:339-361.
135. Tsuruoka S, Nishiki K, Sugimoto K, Fujimura A. Chronotherapy with active vitamin D3 in aged stroke-prone spontaneously hypertensive rats, a model of osteoporosis. *Eur J Pharmacol.* 2001;428(2):287-293.
136. Chotiyanwong P, McCloskey EV. Pathogenesis of glucocorticoid-induced osteoporosis and options for treatment. *Nat Rev Endocrinol.* 2020;16(8):437-447.
137. Winter EM, Schilperoort M, Kroon J, et al. Loss of glucocorticoid rhythm induces an osteoporotic phenotype in mice. Presented at: the 22nd European Congress of Endocrinology. *Endocrine Abstracts.* 2020;70:OC2.4.
138. Selfridge JM, Gotoh T, Schiffhauer S, et al. Chronotherapy: intuitive, sound, founded...but not broadly applied. *Drugs.* 2016;76(16):1507-1521.