



Universiteit
Leiden
The Netherlands

A standardised model for stool banking for faecal microbiota transplantation: a consensus report from a multidisciplinary UEG working group

Keller, J.J.; Ooijevaar, R.E.; Hvas, C.L.; Terveer, E.M.; Lieberknecht, S.C.; Hogenauer, C.; ...
; Vehreschild, M.J.G.T.

Citation

Keller, J. J., Ooijevaar, R. E., Hvas, C. L., Terveer, E. M., Lieberknecht, S. C., Hogenauer, C., ... Vehreschild, M. J. G. T. (2021). A standardised model for stool banking for faecal microbiota transplantation: a consensus report from a multidisciplinary UEG working group. *United European Gastroenterology Journal*, 9(2), 229-247.
doi:10.1177/2050640620967898

Version: Publisher's Version
License: [Creative Commons CC BY-NC-ND 4.0 license](https://creativecommons.org/licenses/by-nc-nd/4.0/)
Downloaded from: <https://hdl.handle.net/1887/3196113>

Note: To cite this publication please use the final published version (if applicable).

ORIGINAL ARTICLE

A standardised model for stool banking for faecal microbiota transplantation: a consensus report from a multidisciplinary UEG working group

Josbert J. Keller^{1,2,3} | Rogier E. Ooijsaar^{3,4} | Christian L. Hvas⁵ | Elisabeth M. Terveer^{3,6} | Simone C. Lieberknecht⁷ | Christoph Högenauer⁸ | Perttu Arkkila⁹ | Harry Sokol^{10,11,12}  | Oleksiy Gridnyev¹³ | Francis Mégraud¹⁴ | Patrizia K. Kump⁸ | Radislav Nakov¹⁵ | Simon D. Goldenberg¹⁶ | Reetta Satokari¹⁷  | Sergiy Tkatch¹⁸ | Maurizio Sanguinetti¹⁹ | Giovanni Cammarota²⁰ | Andrey Dorofeev²¹ | Olena Gubska²² | Gianluca Laniro²⁰ | Eero Mattila¹⁷ | Ramesh P. Arasaradnam²³  | Shiv K. Sarin²⁴ | Ajit Sood²⁵ | Lorenza Putignani²⁶ | Laurent Alric²⁷ | Simon M. D. Baunwall⁵  | Juozas Kupcinskas²⁸ | Alexander Link²⁹ | Abraham G. Goorhuis³⁰ | Hein W. Verspaget^{3,31} | Cyriel Ponsioen⁴ | Georgina L. Hold³² | Herbert Tilg³³ | Zain Kassam³⁴ | Ed J. Kuijper^{3,6,35,36} | Antonio Gasbarrini²⁰ | Chris J. J. Mulder^{3,4} | Horace R. T. Williams³⁷ | Maria J. G. T. Vehreschild^{7,36,38,39}

¹Department of Gastroenterology, Haaglanden Medical Center, The Hague, The Netherlands

²Department of Gastroenterology and Hepatology, Leiden University Medical Center, Leiden, The Netherlands

³Netherlands Donor Feces Bank, Leiden University Medical Center, Leiden, The Netherlands

⁴Department of Gastroenterology and Hepatology, Amsterdam University Medical Centers, Amsterdam, The Netherlands

⁵Department of Hepatology and Gastroenterology, Aarhus University Hospital, Aarhus, Denmark

⁶Department of Medical Microbiology, Leiden University Medical Center, Leiden, The Netherlands

⁷Department of Internal Medicine II, University Hospital Frankfurt, Frankfurt am Main, Germany

⁸Division of Gastroenterology and Hepatology, Medical University of Graz, Graz, Austria

⁹Department of Gastroenterology, Helsinki University Hospital and Helsinki University, Helsinki, Finland

¹⁰Gastroenterology Department, Saint Antoine Hospital, Paris, France

¹¹INRA, UMR1319 Micalis, AgroParisTech, Jouy-en-Josas, France

¹²French Group of Faecal Microbiota Transplantation (GFTF), Paris, France

¹³L.T. Malaya Therapy, National Institute of the National Academy of Medical Sciences of Ukraine, Kyiv, Ukraine

¹⁴INSERM U1053, University of Bordeaux and Bacteriology Laboratory, Pellegrin Hospital, Bordeaux, France

¹⁵Clinic of Gastroenterology, Tsaritsa Yoanna University Hospital, Sofia, Bulgaria

¹⁶Centre for Clinical Infection and Diagnostics Research (CIDR), King's College London and Guy's and St Thomas' NHS Foundation Trust, London, UK

¹⁷Human Microbiome Research Program, Faculty of Medicine, University of Helsinki, Helsinki, Finland

¹⁸Ukrainian Scientific and Practical Center of Endocrine Surgery, Transplantation of Endocrine Organs and Tissues, Kyiv, Ukraine

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

© 2020 The Authors. United European Gastroenterology Journal published by Wiley Periodicals LLC. on behalf of United European Gastroenterology.

- ¹⁹Dipartimento di Scienze di Laboratorio e Infettivologiche, Fondazione Policlinico Universitario A. Gemelli IRCCS, Rome, Italy
- ²⁰Gastroenterology Area, Fondazione Policlinico Universitario Gemelli, Rome, Italy
- ²¹Department of Internal Medicine, National Medical Academy of Postgraduate Education n.a. P.L. Shupic, Kiev, Ukraine
- ²²Department of Therapy, Infectious Diseases and Dermatology Postgraduate Education, Bogomolets National Medical University, Kyiv, Ukraine
- ²³Department of Academic Gastroenterology, University Hospital Coventry and Warwick, Coventry, UK
- ²⁴Department of Hepatology, Institute of Liver and Biliary Sciences, New Delhi, India
- ²⁵Department of Gastroenterology, Dayanand Medical College and Hospital, Ludhiana, India
- ²⁶Department of Laboratories, Bambino Gesù Children's Hospital, Rome, Italy
- ²⁷Department of Internal Medicine and Digestive Diseases, IRD Toulouse 3 University, Toulouse, France
- ²⁸Department of Gastroenterology and Institute for Digestive Research, Lithuanian University of Health Sciences, Kaunas, Lithuania
- ²⁹Department of Gastroenterology, Hepatology and Infectious Diseases, Otto-von-Guericke University, Magdeburg, Germany
- ³⁰Department of Internal Medicine, Amsterdam University Medical Centers, Amsterdam, The Netherlands
- ³¹LUMC Biobank, Leids Universitair Medisch Centrum, Leiden, The Netherlands
- ³²Microbiome Research Centre, St. George and Sutherland Clinical School, University of New South Wales, Sydney, New South Wales, Australia
- ³³Department of Gastroenterology, Hepatology, Endocrinology and Metabolism, Medical University Innsbruck, Innsbruck, Austria
- ³⁴Finch Therapeutics, Somerville, Massachusetts, USA
- ³⁵Centre for Microbiota Analysis and Therapeutics, Leiden University Medical Centre, Leiden, The Netherlands
- ³⁶ESCMID Study Group for Host and Microbiota Interaction (ESGHAMI), Basel, Switzerland
- ³⁷Faculty of Medicine, Imperial College London, London, UK
- ³⁸Department I of Internal Medicine, University of Cologne, Cologne, Germany
- ³⁹German Centre for Infection Research (DZIF), Partner site Bonn-Cologne, Germany

Correspondence

Josbert Keller, Haaglanden Medisch Centrum,
Lijnbaan 32 Den Haag, Den Haag, Zuid-
Holland 2596SR, The Netherlands.
Email: j.keller@haaglandenmc.nl

Funding information

UEG Standards and Guidelines Activity Grant

Abstract

Background: Faecal microbiota transplantation is an emerging therapeutic option, particularly for the treatment of recurrent *Clostridioides difficile* infection. Stool banks that organise recruitment and screening of faeces donors are being embedded within the regulatory frameworks described in the European Union Tissue and Cells Directive and the technical guide to the quality and safety of tissue and cells for human application, published by the European Council.

Objective: Several European and international consensus statements concerning faecal microbiota transplantation have been issued. While these documents provide overall guidance, we aim to provide a detailed description of all processes that relate to the collection, handling and clinical application of human donor stool in this document.

Methods: Collaborative subgroups of experts on stool banking drafted concepts for all domains pertaining to stool banking. During a working group meeting in the United European Gastroenterology Week 2019 in Barcelona, these concepts were discussed and finalised to be included in our overall guidance document about faecal microbiota transplantation.

Results: A guidance document for all domains pertaining to stool banking was created. This document includes standard operating manuals for several processes involved with stool banking, such as handling of donor material, storage and donor screening.

Conclusion: The implementation of faecal microbiota transplantation by stool banks in concordance with our guidance document will enable quality assurance and guarantee the availability of donor faeces preparations for patients.

KEYWORDS

Clostridioides difficile, faecal microbiota transplantation, legislation, stool bank

INTRODUCTION

Faecal microbiota transplantation (FMT) has become an established treatment for recurrent *Clostridioides difficile* infection (CDI),¹⁻⁴ and it appears promising as a treatment modality for other disorders.⁵ In order to ensure a safe, accessible and cost-effective implementation of FMT, stool banks that provide ready-to-use donor faeces preparations are required. Such stool banks may operate at an institutional, national or international level and are currently being set up in different European countries.^{6,7} To date, FMT and stool banking protocols vary significantly between institutions, mostly due to the novelty of this treatment approach and the scarcity of guidelines addressing FMT and stool banking. A recent international consensus meeting addressed FMT and stool banking,⁸ and a British guideline concerning the use of FMT was published in 2018.⁹ Based on: (a) available consensus reports; (b) previous experiences^{6,7,10,11}; and (c) lessons learned from blood banks,⁷ an attempt was made to define a standardised model for stool banks in Europe. In addition, the regulatory boundaries that are required for safe and cost-effective FMT are outlined. This resulted in a practice-oriented consensus report including templates for standard operational procedures and questionnaires (included in the Appendix in Supporting Information Material) that may help to standardise the working plans of stool banks, and facilitate further implementation and regulation of FMT. In addition, our report will support clinicians who want to offer this treatment to their patients.

The statements made throughout this paper are supported by all working group members and describe minimum requirements. In addition, country-specific regulations need to be taken into account to complement the statements.

METHODS

A multidisciplinary working group was formed with experts from mainly European countries. Authors of previously published consensus reports^{8,9} were invited to participate to prevent inconsistencies. Based on the working process of stool banks as previously described,^{5,6} and the clinical application of FMT, topics to be addressed were subdivided into five groups. Subgroups were formed based on the expertise of subgroup members. Subgroup-specific literature searches were performed prior to submission of concept documents addressing the previously defined issues/questions, and statements were phrased. During a working group meeting at the United European Gastroenterology Week in Barcelona, 23 October 2019, the concepts were discussed in depth by the entire working group. Although the aim of the working group was to provide a manual for stool banking in Europe, and not a guideline, an attempt was made to grade the evidence to support statements. The GRADE system (Grades of Recommendation Assessment, Development and Evaluation) was used to grade the strength of evidence (high/moderate/low/very low) and strength of recommendation (strong/weak).¹² Statements addressing organisational aspects of stool

banking were based on expert opinion or law governed. Of note, based on the lack of available evidence, most statements are based on expert opinion. An aggregate document was prepared based on the individual working group's concept documents and expert input during the meeting. After additional comments from experts, the document was finalised and is presented here as a summary. Additional information, containing questions for subgroups, statements discussed by the workgroup, practical standard operating procedure (SOP) templates and questionnaires are available as Supporting Information Material.

Definitions

The definitions used in this paper are mainly based on practice recommendations by the European Society of Clinical Microbiology and Infectious Diseases (ESCMID) and by the Infectious Diseases Society of America,^{1,2} and on the guide to the quality and safety of tissues and cells for human application (tissue guide) of the European Council.¹³

Diarrhoea

Diarrhoea is loose stools at a frequency of three or more per 24 or fewer consecutive hours.¹

CDI

CDI is defined as a combination of compatible signs and symptoms (diarrhoea, ileus, toxic megacolon) confirmed by microbiological evidence of *C. difficile* toxin and *C. difficile* in stools without reasonable evidence of another cause of diarrhoea or by findings demonstrating pseudomembranous colitis on endoscopy or histology.¹

Treatment response

Treatment response in CDI is present when stool frequency decreases to less than three per 24 h (or return to the pre-existent defecation frequency) over a period of at least 72 h in combination with improvement of parameters of disease severity (clinical, laboratory, radiological).

Recurrent CDI

Recurrence is present when CDI reoccurs after the resolution of symptoms of the previous episode. Recurrence within the first 8 weeks after the onset of the previous episode is mostly due to relapse, whereas renewed symptoms during long-term follow-up may be due to relapse or reinfection.¹

Refractory CDI

Lack of response to medication after 5 days or more of treatment.

Severe CDI

Severe CDI is defined by CDI in combination with either a white blood cell count of $15 \times 10^3 \mu\text{l}$ or greater or a creatinine level 1.5 times or greater of the baseline.²

Complicated CDI

CDI is complicated if at least one of the following signs or symptoms are present and related to CDI (2): hypotension requiring vasopressors, intensive care unit admission for a complication of CDI, ileus leading to placement of nasogastric tube, toxic megacolon, colonic perforation, colectomy or colostomy.

Adverse event

Any untoward occurrence associated with the procurement, testing, processing, storage or distribution of faecal preparations.

Adverse reaction

Any unintended response, including a communicable disease, in the donor or the recipient that is associated with the procurement or human application of faecal preparations.

REGULATORY FRAMEWORK AND ORGANISATIONAL PLAN

Classification of FMT

FMT is regarded as a medical treatment that is carried out by registered specialists.^{9,14,15} Although FMT is not yet uniformly regulated within Europe, it seems obvious that the voluntarily donated material (faeces) should be collected, handled and used according to the standards defined by the EU Commission in the EU Tissue and Cells Directive (EUTCD; 2004/23/ec).^{16,17} Faeces should be regarded as a substance of human origin and may thus be considered equal to tissue, as long as no additional modifications other than those necessary for conservation and/or administration are made.¹⁸ Unfortunately, some countries within the EU have currently classified donor faeces preparations as a drug, even though only modifications necessary for conservation and/or administration are applied. Although classification as a drug may enhance the

commercial environment for microbiome therapeutic drug product development, classification as a drug will inevitably result in time-consuming and costly registration processes, sharp and unjustified rise in costs and will negatively impact availability and innovation.¹⁷

Differentiation between stool banks and FMT services

In general, stool banks need to be differentiated from FMT services. While stool banks are responsible for the manufacture and distribution of FMT preparations, FMT services are responsible for patient treatment by the use of FMT preparations. While it is not mandatory, the organisation of an FMT service may be integrated into that of a stool bank. In relation to FMT administration, we suggest that the responsible person should be qualified to treat patients and have a specialist registration. Centres offering FMT should accommodate fully equipped treatment facilities to handle side effects and complications of FMT procedures.

Organisation of stool banks

A stool bank should comply with the following standard requirements.^{16,19}

Organisational description

The EUTCD requires a documented organisational plan for all intended activities and the institutional appointment of an overall responsible person.¹⁶ This responsible person should be a registered medical specialist. The stool bank coordinator should be in charge of the dedicated personnel, the overall quality control and the reporting and handling of adverse events and reactions. The organisational plan should feature a list or a chart of primary implicated partners, their assigned responsibilities, a description of required personnel, validation and qualification concepts, audit schedules, equipment and facilities for each of the three divisions; that is: donor recruitment, screening and manufacture.

Quality management

Stool banks should implement a quality system in accordance with the requirements described in detail in the guide to the quality and safety of tissues and cells for human application (tissue guide) of the European Council.¹³ Although the EU Guidelines for Good Manufacturing Practices (GMP) specifically guide the preparation of medicinal products, their content contains relevant aspects for tissue and stool banks as well. Therefore, the recently released fourth edition of the tissue guide contains good practice guidelines for tissue establishments including relevant elements of GMP.

For all activities, SOPs, guidelines, reference manuals, reporting forms and procedural protocols must be present, and should be updated continuously. Any substantial change to activities requires prior written approval from the responsible specialist. Any processes that may critically affect the quality or the safety of donated material must be identified and lead to implementation of precautionary measures. This includes describing responsible personnel, SOPs, logging and referencing manuals. All activities related to the preparation and distribution of donated material should be logged.

Incidents occurring during donor screening, the laboratory processing of donated faeces or distribution which may potentially impact patient outcome should be handled as adverse events and result in re-evaluation of (adherence to) protocols. Auditing by local or preferably (in the future) national authorities should be part of the quality management of a stool bank. Such audits should ensure that data are entered and maintained properly, and evaluate the working processes and quality assurance programme of the stool bank.

Personnel

All personnel should have clearly defined and documented work descriptions. To ensure competencies, the personnel should be trained in relevant hygiene measures, safety procedures, technical protocols and the organisation before conducting any activities.

Activity

A stool bank should maintain a registry of its activity and should be able to collect and report data on the patients treated with the preparations it distributes. Reporting should include the number of patients, the treatment modality used (colonoscopy, enema, gastric or duodenal tube, gastroscopy, capsules, etc.), as well as follow-up data including safety and outcome of FMT. In the near future, these data should also be entered into a national or international registry.

Distribution

The distribution of faecal preparations from a stool bank to an FMT service requires a (written) agreement between the stool bank and the FMT service in which each party's responsibilities are stated. Prior to distribution, the third party must confirm that it is in possession of sufficient facilities to conduct FMT safely. It is the duty of the stool bank to evaluate and select third parties. Clinical application and patient follow-up is the responsibility of the treating clinician at the FMT service. The FMT service has to report these data back to the stool bank as described above (see Section 3.3.4). SOPs are required for requests of FMT preparations by third-party

institutions, and clear instructions on how to handle the preparations should be available. Return policies of preparations from third-party institutions should be handled according to a shared SOP.

Adverse reaction documentation

Adverse events, adverse reactions (ARs) and serious ARs as defined by the International Council for Harmonisation of Technical Requirements for Pharmaceuticals for Human Use harmonised tripartite guidelines²⁰ should be documented. The stool bank should be able to communicate data related to performance, outcome and safety in an annual report for national health authorities or other auditing authorities. Quality auditing should be organised in collaboration with local or national authorities. Complications and ARs of FMT are generally underreported.^{21,22} Expected ARs following FMT include short-term (including among others abdominal pain, dyspepsia, diarrhoea, fever, constipation) and long-term (including postinfectious irritable bowel syndrome) reactions.²² In addition, unexpected ARs may occur and careful monitoring is required. A serious AR may be procedure related (such as aspiration, perforation), or not clearly related to the procedure. A distinction between procedure-related and unrelated complications should be made according to established guidelines.^{23,24} Each stool bank or FMT service should have a SOP to describe how to handle serious ARs. In the case of any serious AR that is deemed related to FMT, the unit is obligated to notify the appropriate authority immediately. The stool bank is responsible for ensuring notification to the health authorities and confirming that sufficient actions and adaptations are being made. All relevant personnel and divisions should be notified in the case of a serious AR to facilitate traceability and control of quality and safety. Effective and rapid procedures for recall and disposal must be in place in the case of a serious AR.

Data management

Anonymity

All data regarding donors and recipients should be anonymised to maintain a principle of voluntary altruism between donor and recipients without any revealing elements. This also applies to third-party data access. All steps of the donation should therefore be uniquely coded and linked using specific donor identifiers, donation identifiers and donation product identifiers.

Traceability

An efficient and well protected data storage system for logging should be in place, allowing accurate tracing of the entire process

from donor to recipient. Both donor and processing records should be kept. All data necessary to map the process and handling should be kept for a minimum of 30 years.

Storage

Paper-based documentation systems are acceptable, but secure electronic systems that are auditable, traceable and retrievable are preferred. Storage must be in accordance with local data protection regulations.

DONOR RECRUITMENT, SELECTION AND SCREENING

Donor recruitment

Donation of stool is a voluntary act and should not become a subject of trade. The use of unpaid donors reduces the risk that applicants withhold information during the screening process. Reimbursement of expenses for travel to the donation centre is reasonable. Universal donors are preferred over patient-selected donors. In individual cases, a patient-selected donor may be accepted, if (s)he fulfils all criteria determined in the screening process for universal donors. Healthcare workers may be approved as donors if they fulfil all necessary requirements, although some stool banks choose to exclude them because of a potential risk of multidrug resistant organism carriership.

Donor informed consent

All donors should be informed about the associated risks and benefits and provide written informed consent that covers the provision of personal information, the screening processes, the provision of multiple donations, the storage of donor data in a donor registry and future unscheduled contacts by the stool bank in case of adverse events or for research purposes. A proposed template for an informed consent form for healthy donors is included in the Appendix in Supporting Information Material.

Donor screening

Donor screening should be conducted in accordance with a locally approved set of SOPs and performed by dedicated staff. Meetings between the donor screening staff and the rest of the stool bank team need to be scheduled on a regular basis, in order to identify problems with respect to screening or the availability of donations well in advance. The collected data should be documented on pre-specified forms that facilitate the structured display of results obtained during the screening process (interviews/questionnaires and

microbiological testing) and include documentation of donor clearance.

Initial questionnaire

During the first step of the screening process, the medical history and risk behaviour of potential donors should be assessed by the use of a dedicated questionnaire. The results should be evaluated by a clinician. Objective evaluation should include a document clearly identifying the consequence of a specific response for the screening process. The topics and items that should be addressed in this questionnaire are listed in Table 1. A proposed questionnaire with consequences for positive answers is added to the Appendix (Supporting Information Material).

Microbiological testing

Once a potential donor has been found suitable for further evaluation based on the donor questionnaire and the physical examination, he or she should undergo blood and faeces screening for transmissible pathogens listed in Table 1.

Stool donation, quarantine and retesting of donors

Once a donor has been approved, (s)he should complete a second short questionnaire before each donation, assessing the occurrence of any event that may have occurred between donor approval and donation. Complete donor screening based on blood and faecal analyses should be repeated a minimum of every 3 months. A donation period should have a defined maximum duration, not exceeding 3 months and should start and end with a complete donor screening. Blood tests pertaining to the close-out screening should be performed a minimum of 4 weeks after the last donation to allow for a lag time of serology or polymerase chain reaction-based testing for certain pathogens. FMT products prepared during a donation period should be put under quarantine until the repeat donor screening results are available.

If donor screening is performed as recommended in this document, direct testing of each preparation is not mandatory. It may, however, be required by local regulations.

Donor screening for treatment of patients with severe immunosuppression

A distinction should be made between severely immunocompromised and moderately immunocompromised patients, as outlined in the section describing the clinical application of FMT below. FMT preparations for severely immunocompromised patients should be made with the following additional safety precautions:

TABLE 1 Items to be assessed during screening of donors before approval for FMT

Infectious diseases

- History or exposure to infectious diseases with chronic activity: HIV, hepatitis B virus (HBV) or hepatitis C virus (HCV), non
- successfully eradicated helicobacter pylori, syphilis, malaria, trypanosomiasis, tuberculosis, Chagas disease,
- strongyloidiasis
- Any currently active infection or those of relevance within the past 6 months
- Live attenuated vaccine within the past 8 weeks
- Country of birth

At-risk behaviour

- Current or previous intravenous drug use
- Ongoing high risk sexual behaviour within the past 6 months
- Travel to high-risk foreign countries within the past 6 months
- Current occupation in a setting facilitating acquisition of potential pathogens (e.g., veterinarian, animal attendant, gamekeeper, prison worker)
- Tattoo, piercing or acupuncture within the past 6 months
- Major surgery within the past 6 months
- Contact with human blood (e.g., accident, needle stick injury) within the past 6 months
- Previous prison term
- Previous tissue/organ transplantation
- Transfusion of blood products (e.g., packed red cells, plasma, platelets, immunoglobulins) within the past 6-month medical history

Medical history

- Chronic diseases
- (Risk of) Creutzfeldt–Jakob disease
- Allergies or atopy (e.g., food or drug allergies, asthma)
- hospitalisation within the past 4 months
- Ongoing pregnancy
- Antibiotic treatments, scheduled or received within the past 3 months
- Regular medication or nutritional supplements
- BMI (accepted if ≥ 20 and ≤ 25 kg/m²)^a
- Age (accepted if ≥ 18 and ≤ 60 years)

Intestinal health

- Previous or scheduled gastrointestinal surgery, except for appendectomy
- Gastrointestinal symptoms within the past 3 months (e.g., diarrhoea, constipation, haematochezia, vomiting, abdominal pain), or (removed) adenomatous polyps or sessile serrated lesions
- Any other relevant clinical sign or symptom within the past 3 months (e.g., fever or rash)

Note: Exclusion criteria related to each item are presented in Supporting Information Material 3.

Abbreviations: BMI, body mass index; FMT, faecal microbiota transplantation.

^aSome stool banks accept donors with a BMI more than >18 and less than <30 kg/m².

1. Additional screening tests are advised for the use of FMT preparations in severely immunocompromised patients (see Table 2 and Supporting Information Material 2).
2. FMT products should be put under quarantine until the donor has been found acceptable in a repeat screen. For severely immunocompromised patients (see paragraph ahead about the clinical

TABLE 2 Blood and faeces tests for donor screening before approval for FMT

Stool analyses

- Bacterial enteral pathogens: Shiga-like toxin-producing *E. coli* (STEC) stx1/stx2,^a *Shigella* spp., campylobacter jejuni and coli, *Salmonella* spp., *Yersinia enterocolitica* and *Clostridioides difficile* (all PCR), *Helicobacter pylori* (faecal antigen), *Vibrio* species (if visited or residing in tropical country within the past 6 months; culture)
- Antibiotic-resistant bacteria: Extended spectrum beta-lactamase-producing bacteria (ESBLs)/multidrug-resistant Gram-negative bacteria (MRGN) including carbapenemase-producing enterobacteriales, vancomycin-resistant enterococci (VRE), methicillin-resistant staphylococcus aureus (MRSA; all culture)
- Viruses: norovirus, rotavirus (all PCR), SARS-CoV-2^b
- Parasites: *Cryptosporidium* spp., *Giardia lamblia*, helminths, *Entamoeba histolytica*, *Dientamoeba fragilis*, *strongyloides stercoralis* (PCR), cyclospora and *Isospora*,^c *Blastocystis hominis*^d (PCR, antigen (if available) or microscopy)

Blood analyses

- General laboratory: CRP, creatinine, ALT, bilirubin, blood cell count
- Viruses: hepatitis A (IgM), hepatitis B (HbsAg), hepatitis C (anti HCV), HIV 1 and 2 (combined HIV antigen/antibody test), HEV serology, SARS-CoV-2^b
- Bacteria: *Treponema pallidum* (TPHA)

Additional screening tests advised for immunocompromised patients (see Supporting Information Material 8): Stool: *Plesiomonas shigelloides*, adenovirus, parechovirus, astrovirus, enterovirus, sapovirus, microsporidia. Blood: CMV, EBV, toxoplasmosis.

Abbreviations: ALT, alanine aminotransferase; CMV, cytomegalovirus; CRP, C-reactive protein; EBV, Epstein-Barr virus; FMT, faecal microbiota transplantation; PCR, polymerase chain reaction.

^aEnteropathogenic *Escherichia coli* (EPEC) testing may be considered in some countries.

^bCovid-19 (SARS-CoV-2) testing of asymptomatic donors combining serology and stool testing requires further validation.

^cIf residing in or foreign travel to tropical country within the past 6 months.

^dKolonisation with *Blastocystis hominis* is not considered an exclusion criterium. However, the working group advises to monitor for the effects of transmission.

^eSeveral stool banks also include HTLV 1 and 2 testing.

application of FMT), the prepared FMT preparation itself should undergo a quality control that includes (re)screening for potential pathogens, as listed in Table 2.

PROCESSING AND STORAGE OF DONOR FAECES PREPARATIONS

Donor faeces collection and preparation for FMT should follow a standard protocol to ensure that products are consistently produced and controlled with respect to the quality standards appropriate to their intended use. The process described below is based on experiences in different centres and has not been tested rigorously. There are no reported studies comparing different preparation protocols of faecal preparations. A possible protocol is presented in Table 3. Following a standard protocol enables the comparison of outcomes between institutions, facilitating quality assurance. The described protocol has proved effective for FMT treatment of recurrent CDI; for other potential indications, adjustment of the protocol may be required in the future.

Collection of donor stool

To promote standardised practice and a safe and effective product, clear written instructions should be provided to the donor for faeces

collection and delivery procedures. To prevent environmental/cross-contamination, faeces is collected by the donor in a faecal container for single use only (e.g., fecotainer).

Donors should hand in their faeces within 2 h after defecation; some stool banks prefer on-site donation. After defecation and until further processing, the stool sample can be stored at room temperature. If it takes more than 30 min to deliver the collected faeces to the stool bank, temporary storage in a cooler bag or refrigerator is preferred because faecal storage without stabilisation buffer significantly changes taxa abundances from 30 min onwards.²⁸⁻³¹

Processing of preparations

In most studies that performed FMT for recurrent CDI, donor faeces was processed within 6 h of defecation.^{4,11,25-27,32-48} Studies in which a longer interval between defecation and processing was allowed (24-48 h) report a somewhat lower cure rate of FMT (approximately 75%).⁴⁹⁻⁵² In addition, it is hypothesised that a high viability of bacteria in donor stool increases the efficacy of FMT. Subsequently, this could allow a reduced amount of donor faeces used per suspension. To minimise sample degradation and alteration, donor faeces should be processed as soon as possible as most faecal bacteria are anaerobic.

The preparation of faecal preparations under aerobic conditions is considered suitable for FMT treatment of recurrent CDI, as anaerobic processing does not increase cure rate.⁵³⁻⁵⁵ This

TABLE 3 Validated standardised conditions for processing of FMT suspensions and treatment of patients with recurrent *Clostridioides difficile* infection

Processing of FMT suspensions	
Amount of faeces	50 g ^a
Processing	Aerobic or anaerobic
Diluent	NaCl 0.9%
Cryoprotectant	Glycerol 10%
End volume	50–60 g: 200 cc 25–30 g: 100 cc
Storage	
Timeframe between collection and storage	<6 h (rapid processing preferred)
Treatment of patients	
FMT preparation	Frozen stool banked preparation
Pretreatment of patient	4–10 Days vancomycin 125–250 mg qid (or fidaxomicin 200 mg bid) Stop >24 h before FMT Bowel lavage with macrogol on day before FMT (Not necessary in case of capsule administration) ^b Colonoscopy Nasoduodenal tube (infusion 10–25 cc/minute) Capsules, if established protocol

Abbreviations: CDI, *Clostridioides difficile* infection; FMT, faecal microbiota transplantation.

^aSuspensions made of 25-g donor faeces may even suffice, but require validation.^{25–27}

^bFor upper gastrointestinal delivery for treatment of CDI, bowel lavage could be limited to 50% of the advised dose for colonoscopy cleansing.

observation may be explained by the fact that a considerable part of the bacterial genera can tolerate oxygen for a limited amount of time or produce resilient spores that allow transfer of at least a proportion of anaerobic intestinal bacteria.⁵⁶

Pooling (mixing) of multiple donor faeces during processing is not recommended because: (a) it hampers the traceability of the faecal preparation to the individual donor; (b) the risk of transmissible disease may be increased; and (c) the principle of transfusing a balanced microbiota preparation may be lost.

It is preferable to use 50 g or more of stool to prepare an FMT suspension for recurrent CDI treatment. Systematic reviews suggest that the use of 50 g or more of faeces is more effective than less than 50 g.^{57,58} However, the strength of this recommendation is impaired by the absence of comparative studies, limited numbers of recent studies and variation in delivery routes. Several experts report (mostly unpublished) positive results with less than 50 g of donor faeces.⁵⁹ Reducing the amount to 25 g (or even lower) could therefore be considered but should be validated.

In general, the faecal suspension is made with preservative-free sterile 0.9% saline as diluent.^{4,11,26,32,34–38,40–43,47–50,55,60–75} The suspension should not be too viscous for infusion through a nasoduodenal tube or biopsy channel of a gastroscope or colonoscope.

The suspension can be homogenised by a variety of methods such as blenders,^{32,35–37,42–44,46,50,53,55,60,61,65,67,69,73,74}

stomachers^{34,51,71} with mortar and pestle,⁶ or wooden spatulas.^{26,49,76} Of utmost importance is the use of clean consumables, which implies that materials should be decontaminated using a validated method if reused or should be disposable. To prevent clogging of the tube/biopsy channel during the administration procedure the faecal suspension should be filtered. Filtration can be performed by a gauze, filter paper, strainers or sieves. To prevent external contamination either a closed system or an open system in a biological safety cabinet is preferred. To combine homogenisation and filtration, a bag mixer appears easy to handle and is efficient.

The infused volume may be reduced using centrifugation,^{6,55,61,74} but the reported cure rate of 68% after multiple centrifugation and washing steps appears relatively low,⁴¹ and such an approach should be validated.

To prepare frozen preparations, a cryoprotective agent should be added prior to freezing. In general, the cryoprotectant glycerol is used in a final concentration of 10%–15%.^{26,27,34,41,53,55,61,66,69,71,73,74,77} Cryopreservation is a process of preservation of the biological and structural functions of tissues or cells when cooling to sub-zero temperatures. The viability of six representative groups of faecal bacteria after 6 months of storage at 80°C in normal saline with 10% glycerol did not differ from baseline, whereas viability was reduced in suspensions stored without glycerol.⁴¹ A possible side effect of large amounts of glycerol in the bowel

is a mild alteration in serum glucose. This is not observed with doses below 0.75 g of glycerol per kg body weight.⁷⁸ A faeces preparation of 200 cc, containing 10% glycerol (approximately 20 g) is far below this limit.

Encapsulation of glycerol preparations and cryopreservation of encapsulated donor faeces is feasible and could reduce the burden of FMT for patients in the future, but requires further validation.⁷⁹

Storage of FMT preparations

FMT preparations should be stored in sealed, clean plastic containers, with a unique code ensuring traceability of the sample (see data management). Storage of FMT preparations (intended for treatment) should be accommodated by the stool bank in a dedicated 80°C freezer with connected alarm notification to guarantee continuous registration of the storage conditions. Documentation requires a biobanking information and management system for coding, registrations and tracing of the samples.

All processed FMT preparations should be placed into quarantine and stored in clearly distinguishable sections until all donor screening results are available. The release of products must be documented, and following these quarantine measures, the FMT preparations should be cleared and transferred to another distinct storage section. Only cleared FMT preparations should be used for patient treatment.

Distribution of FMT preparations should be performed on dry ice (or 4°C or room temperature in the case of immediate use after transfer) shipment through a certified courier service. Registration in a “biobanking information and management system” related database for the shipped faecal preparations, including recipient institution and requestor information, should be undertaken so that biovigilance tracing can be performed in case of ARs. Long-term storage of faecal preparations at the recipient centre should be discouraged.

To ensure the maximum safety and quality of the FMT preparation, it is mandatory to specify a maximum storage time with an expiry date. The association between storage conditions and clinical efficacy has not been investigated. Storage at –80°C is preferred, although storage of faecal preparations at –20°C for up to one month (30 days) may not reduce efficacy.⁸⁰ In general, long-term storage should be at –80°C or lower to prevent sample degradation. Once preparations are transferred to a ward or an endoscopy unit for administration, temporary storage at –20°C is acceptable. High cure rates have been reported, with frozen FMT suspensions stored up to 10 months at –80°C^{6,26,27,34,53,55,73,74} but this could in theory be much longer. OpenBiome and The Netherlands Donor Feces Bank have good experiences with storage up to 1 and 2 years, respectively.^{6,81}

A sample of the original donor faeces and/or of the processed FMT preparation should be stored for a minimum of 10 years following the application for retrospective quality assessment in case of an AR.

Thawing of donor faeces preparations

Thaw times depend on the volume and type of FMT preparations. A 200-cc suspension can be thawed overnight in a 4°C refrigerator, or during 5 h at room temperature. Warm water baths (37°C) can be used to speed up thawing of FMT suspensions, but are not applicable to the thawing of capsules. However, only clean water baths with fresh water should be used to avoid cross-contamination (especially with water-dwelling bacteria such as *Pseudomonas*). Thawed faecal suspensions should be infused the same day, and should not be refrozen, because freeze thaw cycles adversely affect the viability of the microbial communities in the faecal suspension.⁸²

The faecal suspension should be at room temperature when infused into the recipient, to minimise abdominal discomfort.

CLINICAL APPLICATION OF FMT

The primary aim of a stool bank is to centralise the process of donor screening and processing and storage of donor faeces preparations. Given the novelty of this treatment modality, guidance for treating physicians may be required to optimise the safety and efficacy of large-scale FMT implementation. Stool banks can offer expert consultation at the request of the treating physician, or they can incorporate consultation as part of a standard procedure to verify the correct indication for FMT before delivering FMT preparations. Currently, stool preparations are indicated for recurrent CDI only. In a research or compassionate use setting, other indications could be considered. Stool should not be sold or delivered to patients upon their own request. Finally, the need for documentation and quality improvement requires a strong and well-defined infrastructure.

Consultation

To offer appropriate consultation, an expert panel is required. As indications for FMT are likely to become more numerous in the future, medical specialists in the field pertaining to the added indications should be included in the FMT expert panel of a stool bank. A standardised patient questionnaire with key information about the current episode of CDI, previous episodes of CDI, treatment effects and microbiological testing is required (see Supporting Information Material 8). The expert panel may assess whether the indication is appropriate and advise about additional diagnostics or an alternative therapy. Based on such a consultation process, The Netherlands Donor Feces Bank has rejected about a quarter of the requests for FMT preparations for treatment of recurrent CDI.⁶ An even higher rate of inappropriate FMT indications (around 50%) was noticed by a French reference centre (St Antoine Hospital). If regular consultation is not offered by a stool bank, treating clinicians should receive contact information for consultation in case of: (a) unexpected problems during the FMT treatment; and (b) unexpected ARs.

Patient data should be shared and stored appropriately, in accordance with European General Data Protection Regulation guidance and other local regulations.

Treatment protocol

A standardised treatment protocol for recurrent CDI will enable the comparison of outcomes among FMT centres, and subsequently increase safety and facilitate quality assurance. However, variation of almost all steps of the FMT treatment protocol is conceivable. Therefore, a validated treatment protocol is presented in Table 3.

Pretreatment of patients

In clinical practice, antibiotics are initiated directly after a positive *C. difficile* test. It is unknown if pretreatment with antibiotics increases the effectiveness of FMT, but most centres perform FMT following at least 4 days of oral vancomycin (125–250 mg qid) or fidaxomicin (200 mg bid). Antibiotics should be stopped the day before FMT (>24 h before FMT). Bowel lavage is required before a colonoscopy. In general, (modified) bowel cleansing is also performed one day before donor faeces infusion via the upper gastrointestinal route; this could for example include 2 instead of 4 l polyethylene glycol-electrolyte lavage solution.⁶ Whether bowel lavage increases clinical efficacy for the treatment of CDI is unknown.

Administration method

In a small study, FMT via a nasoenteric tube (upper gastrointestinal delivery) or colonoscopy (lower gastrointestinal delivery) appears equally effective ($p = 0.53$).⁷⁴ In addition, nasogastric administration is possible after pretreatment with proton pump inhibitors.³⁵ One study suggested that treatment with enemas more often requires repeated infusions,⁴⁹ but this study used low volume enemas (50 ml) and no bowel lavage. Capsules containing either lyophilised or glycerol-preserved faeces suspensions also appear effective.^{62,79} The choice for the route of delivery should be based on local preferences and patient characteristics, if applicable. A volume of 200 cc can be safely infused via the upper gastrointestinal route. Slow infusion of the donor faeces suspension (e.g., 10–25 cc/minute) may prevent nausea and regurgitation/aspiration. In patients with impaired gastrointestinal motility, infusion through a colonoscope is preferred to prevent regurgitation/aspiration. Moreover, upper gastrointestinal delivery (including capsules) should be avoided in patients with swallowing disorders, because of the increased risk of aspiration. Small intestinal bacterial overgrowth has not been recognised as an adverse event of upper gastrointestinal delivery of FMT. In frail patients,

ileocolonoscopy can be avoided by upper gastrointestinal delivery, rectosigmoidoscopy or enema. The infusion of the donor faeces suspension is straightforward and can be performed by any appropriately qualified healthcare professional.

Indications and contraindications for FMT

The development of national and international guidelines for the clinical applications of FMT is a dynamic process, driven by the emerging body of clinical evidence. The indications and contraindications outlined below represent the consensus of the working group.

Indications

FMT should be considered in patients with two or more CDI recurrences. A balance of risks and benefits should guide a joint decision of either FMT or (tapered) antibiotics with or without additional treatment with, for example, monoclonal antibody or nonabsorbable antibiotics. Few studies have addressed the effects of FMT in patients with refractory or severe CDI, although FMT as rescue treatment appears encouraging.^{40,63,83–86} We suggest FMT as an option for patients with refractory or severe CDI. The route of FMT application in severely ill patients should be guided by the complication risk of the individual patient. Toxic megacolon should mainly be treated surgically. In patients with fulminant disease, a diverting ileostomy or other surgical treatment options should be considered. In patients who refuse surgery, or in whom surgery is somehow contraindicated, FMT via the lower or upper gastrointestinal route can be cautiously considered and has resulted in cure.^{45,53} Because the efficacy of FMT in severe and complicated CDI appears lower, repeated infusions are often required.

Contraindications

Currently, no data from clinical studies or reviews indicate absolute contraindications for FMT. Based on expert opinion, the following contraindications for FMT apply: anatomical abnormalities that prevent safe application of FMT by endoscopy, enema or tube; signs of intestinal perforation; pregnancy and severe food (e.g., peanut) allergy. The latter may be solved by a patient-related donor with strict dietary restrictions before stool donation. Further limitations appear mainly related to the mode of application. Relative contraindications for upper gastrointestinal delivery are increased risk of regurgitation (e.g., known large hiatus hernia, severe gastro-oesophageal reflux disease), swallowing disorders and impaired small bowel motility or obstruction. An increased risk of sedation may be considered a contraindication for lower gastrointestinal delivery by colonoscopy.

FMT for other indications

Both patients and clinicians are currently exploring the opportunities for FMT to alter the course of a large variety of diseases. Although promising studies that confirm the efficacy of FMT for other disorders besides CDI are lacking, limited or contradicting and FMT is not mentioned in treatment guidelines for disorders beyond *C. difficile* infection. Although randomised trials suggest a beneficial effect in a subset of patients with hepatic encephalopathy and ulcerative colitis, many questions about the optimal use of FMT for those patients remain. For irritable bowel syndrome, the results of randomised studies are less consistent. Until further evidence becomes available, FMT for other indications should preferably be limited to the research setting or, in the absence of alternative therapeutic options, as compassionate use. The results of studies addressing FMT for other indications are summarised in Table 4.⁸⁷⁻¹⁰⁰

Special situations

Children

Although rare, the use of FMT in children suffering from CDI has been described.^{101,102} Antibiotics remain the cornerstone of initial CDI treatment, but recent European and American guidelines recommend FMT for children suffering from multiple recurrent CDI.^{2,103} FMT protocols for children and adults are similar. It is unknown whether age-matched donors should be considered for paediatric use. Donor preparations from adults appear safe, but long-term effects are unknown and careful follow-up is warranted. Particular attention is required because the microbiota rapidly develops during infancy, and maturation of the microbiota in children occurs parallel with the maturation of the immune system; processes which are likely to influence each other.¹⁰⁴ Endoscopy and the instillation of suspensions may be performed under general anaesthesia with intubation.^{101,102} As for adults, existing guidelines only recommend the use of FMT in children for recurrent CDI.²

Immunocompromised patients

FMT appears safe in immunocompromised patients.¹⁰⁵ The recent report of a severely immunocompromised patient's death following transfer of an MDRO calls for particular caution in this patient group.¹⁰⁶ The risk of developing complications after the transfer of potential pathogens appears increased in severely immunocompromised patients, defined as: (a) patients with current or foreseeable neutropenia within the next 14 days (defined as <500 neutrophils/ml); (b) patients scheduled for allogeneic stem cell transplantation (SCT) or having received allogeneic SCT within 100 days; and (c) patients with active graft versus host

disease requiring immunosuppressive treatment. For severely immunocompromised patients, we suggest: (a) additional screening tests of donors; and (b) testing of the FMT preparation or donated stool used for treatment of the individual patient, as described in the Appendix (Supporting Information Material). For less severely immunocompromised patients (<200 CD4 T cells/ml; or prolonged use of corticosteroids at a mean dose of more than 0.3 mg/kg/day of prednisone equivalent for over 3 weeks; or treatment with other recognised T-cell immunosuppressants, such as cyclosporin, TNF- α blockers, specific monoclonal antibodies (e.g., alemtuzumab), methotrexate or nucleoside analogues during the past 90 days); or inherited severe immunodeficiency (e.g., chronic granulomatous disease or severe combined immunodeficiency), normal donor screening appears sufficient. More stringent follow-up is justified in all immunocompromised patients, and careful reporting of outcomes and side effects is needed to confirm the safety of FMT in those patients.

COVID 19-pandemic

During the COVID-19 pandemic, stool banks have to re-evaluate their working processing to optimise safety for donors, patients and healthcare professionals, as outlined in a recent consensus report.¹⁰⁷ Screening for SARS-CoV-2b should be incorporated in donor screening protocols (Table 2).

Follow-up of patients after FMT

Patients who receive FMT should be registered by the stool bank with support of the treating FMT service, in order to document patient flow, ensure traceability and document treatment outcome and treatment-related complications. A structured patient follow-up should be conducted. This should consist of clinical information and relevant paraclinical tests, depending on the indication for FMT. Short term follow-up includes the documentation of side effects or complications that occur in relation to the treatment or within the first 24 h after any FMT-related procedure. Long-term follow-up includes the documentation of clinical details and relevant clinical results beyond the first 24 h. The duration of long-term follow-up depends on the indication, but should cover a minimum of 8 weeks and preferably 6 months. Relevant clinical variables to document during follow-up are general symptoms, disease-specific symptoms, standardised symptom or disease activity scores and patient weight.

FMT appears safe with limited side effects,^{21,108} although infectious complications have been described in severely immunocompromised recipients underlining the need for careful donor screening.¹⁰⁶ However, some concern about the long-term side effects of FMT has been raised. This may include weight gain, the development or worsening of inflammatory bowel diseases, cancer, autoimmune diseases, allergies or neurological diseases.¹⁰⁹

TABLE 4 Summary of studies addressing the efficacy of FMT for other disorders

Disease	Author	Study Design	Donor selection	Pretreatment	Route of administration	Number of FMTs	Outcome
Hepatic Encephalopathy	Bajaj et al. ⁸⁷	RCT	Single selected donor	Broad-spectrum antibiotics	Enema	3	Significant reduction in new HE episodes
Ulcerative colitis	Moayyedi et al. ⁸⁸	RCT	Multiple donors	None	Enema	6	Significant difference in achieving remission
	Rossen et al. ⁸⁹	RCT	Multiple donors	Bowel lavage	Duodenal tube	2	No significant difference in achieving remission
	Parasomthy et al. ⁹⁰	RCT	Pooled donors	Bowel lavage	Colonoscopy + enema	1 + 39	Significant difference in achieving remission
	Costello et al. ⁹¹	RCT	Pooled donors	Bowel lavage	Colonoscopy + enema	1 + 2	Significant difference in achieving remission
Crohn's disease	Sokol et al. ⁹²	RCT	Multiple donors	Bowel lavage	Colonoscopy	1	No difference in sustained engraftment between allogeneic FMT and sham
Irritable bowel syndrome	Holvoet et al. ⁹³	RCT	Multiple donors	NA	Jejunal tube	1	Significant difference in symptom relief
	Arionadis et al. ⁹⁴	RCT	Multiple donors	NA	Capsules	3	No difference in achieving symptom relief vs. placebo
	Halkjaer et al. ⁹⁵	RCT	Multiple donors	Bowel lavage	Capsules	12	Placebo treatment superior
	Johnsen et al. ⁹⁶	RCT	Multiple donors	Bowel lavage + loperamide 2 h prior to FMT	Colonoscopy	1	Significant difference in symptom relief
	Holster et al. ⁹⁷	RCT	Multiple donors/ autologous	Bowel lavage + loperamide prior to FMT	Colonoscopy	1	No difference in achieving symptom relief vs. autologous FMT
	El-Salhy et al. ⁹⁸	RCT	Single donor	Unknown	Duodenal delivery	1	Significant dose dependent difference in symptom severity reduction
Eradication of MDROs	Huttner et al. ¹⁰⁰	RCT (open-label)	Multiple donors	Nonabsorbable antibiotics	Capsule or nasogastric tube	1	Slightly decreased ESBL-E/CPE carriage

Abbreviations: CPE, *Carbapenemase-producing Enterobacteriaceae*; ESBL, extended spectrum beta-lactamase; FMT, faecal microbiota transplantation; MDROs, multidrug resistant organisms; NA, not available; RCT, randomised controlled trial.

Follow-up should include clinical and analytical data, also determined by the indication for FMT and comorbidity of the patient. National or institutional registries that are able to perform long-

term follow-up, and collect outcome data after FMT for various conditions are warranted to document the long-term safety of FMT.

TABLE 5 Summary of key recommendations regarding FMT and stool banking

Regulatory framework

- FMT should be classified as “tissue transplantation”, not as a drug
- Stool banks should work in compliance with the EU tissue and cells directive
- Stool banks should have an organisational plan describing the intended activities and involved personnel
- An efficient and well protected data storage system for logging should be in place, allowing accurate tracing of the entire process from donor to recipient

Donor recruitment, selection, screening

- Stool donation should remain a voluntary act
- In general, universal donors are preferred to patient-selected donors
- Extensive screening of donors before releasing material for patient care is mandatory
- FMT products should be placed under quarantine until the donor has been found acceptable in a repeat screen
- All information collected during donor screening, including identity and results, should be stored for at least 30 years

Processing and storage of preparations

- Pooling of donor material is not recommended
- Long-term storage of donor faeces preparations should be at -80°C
- A safety aliquot of each faecal suspension should be stored separately for up to 10 years after use

Clinical applications

- Currently, FMT is only considered standard of care in multiple recurrent CDI
- FMT can be used to treat multiple recurrent CDI in children or severely immunocompromised patients
- The choice for a route of delivery of FMT preparations should be based on local preferences and patient characteristics if applicable

Follow-up

- Stool banks should actively perform follow-up of all patients treated by FMT
- An FMT centre should maintain a register to document patient flow, performance, clinical outcome and safety measures

Abbreviations: CDI, *Clostridioides difficile* infection; FMT, faecal microbiota transplantation.

CONCLUSIONS

FMT is a new, safe and efficacious treatment strategy. The critical step of FMT is the process of donor selection and screening. Stool banks have enabled centralisation of this process, following the model of blood banks. We provide a model for the standardisation of stool banking; key recommendations are summarised in Table 5. Standardisation will result in further improvement of the quality and safety of FMT in Europe. Finally, stool banks will enable quality assurance of FMT and guarantee the availability of donor faeces preparations for patients.

ACKNOWLEDGEMENTS

The templates of standard operational procedures are based on SOPs provided by The Netherlands Donor Feces Bank and Aarhus University Hospital, Aarhus, Denmark. The presented informed consent form for healthy donors was provided by the Cologne Stool Bank, Germany. The working group was hosted by the European

Helicobacter and Microbiota Study Group (EHMSG), which is a member society of UEG. Delegates of the ESCMID study group for Host and Microbiota Interaction (ESGHAMI) group participated in the working group. The working group was supported by Romana Koönig. The working group was supported by an UEG standards and guidelines activity grant.

CONFLICT OF INTERESTS

Josbert J. Keller and Ed J. Kuijper have received research grants from Vedanta Biosciences. Cyriel Ponsioen has received grant support from Takeda, speaker's fees from Takeda, Tillotts and Pfizer and consultancy fees from Takeda, Shire and Pliant. Radislav Nakov is in the speaker's bureau of Takeda and Pfizer. Perttu Arkkila has received a research grant from Vifor Pharma, and speaker's fee from MSD. Maria JGT Vehreschild has received research grants from 3M, Astellas Pharma, DaVolterra, Gilead Sciences, Heel, Glycom, MaaT Pharma, Merck/MSD, Organobalance, Seres Therapeutics; speaker's fees from Astellas Pharma, Basilea, Falk Foundation, Gilead Sciences,

Merck/MSD, Organobalance, Pfizer and has been a consultant to Alb Fils Kliniken GmbH, Astellas Pharma, BioMerieux, DaVolterra, Ferring, MaaT Pharma, Merck/MSD and Roche. Simon D. Goldenberg reports consultancy fees from Astellas, Enterobiotix, Menarini, MSD, Pfizer and Shionogi. Oleksiy Gridnyev reports speaker's fees from Abbott and a research grant from Biocodex. Harry Sokol reports board membership, consultancy or lecture fees from Carenity, Abbvie, Astellas, Danone, Ferring, Biocodex, Mayoly Spindler, MSD, Novartis, Roche, Tillots, Enterome, Maat, BiomX, Biose, Novartis and Takeda; and is a co-founder of Exeliom Biosciences. Zain Kassam is an employee/shareholder of Finch Therapeutics. Alexander Link reports a grant from European Commission European funds for regional development (EFRE) related to micro-biome and FMT research. Georgina L. Hold reports consultancy fees from Ferring and Proctor and Gamble; and scientific board membership of Enterobiotix. Eero Mattila has been a scientific advisor and received lecture fees from MSD Finland. Christoph Hogenauer receives grant support from Seres Therapeutics and consultancy, or lecture fees from Abbvie, Astellas, Ferring, Falk, Janssen, Merck/MSD, Pfizer and Takeda. The other authors declare that there are no conflict of interests.

DATA AVAILABILITY STATEMENT

The data are available upon reasonable request to the corresponding author.

ORCID

Harry Sokol  <https://orcid.org/0000-0002-2914-1822>

Reetta Satokari  <https://orcid.org/0000-0002-4258-3414>

Ramesh P. Arasaradnam  <https://orcid.org/0000-0002-2231-3062>

Simon M. D. Baunwall  <https://orcid.org/0000-0002-5135-7435>

REFERENCES

1. Debast SB, Bauer MP, Kuijper EJ, et al. European Society of Clinical Microbiology and Infectious Diseases: update of the treatment guidance document for *Clostridium difficile* infection. *Clin Microbiol Infect.* 2014;20(Suppl 2):1–26. <https://doi.org/10.1111/1469-0691.12418>.
2. McDonald LC, Gerding DN, Johnson S, et al. Clinical practice guidelines for *Clostridium difficile* infection in adults and children: 2017 update by the Infectious diseases Society of America (IDSA) and Society for healthcare epidemiology of America (SHEA). *Clin Infect Dis.* 2018;66:987–94. <https://doi.org/10.1093/cid/ciy149>.
3. Ooijsaar RE, van Beurden YH, Terveer EM, et al. Update of treatment algorithms for *Clostridium difficile* infection. *Clin Microbiol Infect.* 2018;24:452–62. <https://doi.org/10.1016/j.cmi.2017.12.022>.
4. van Nood E, Vrieze A, Nieuwdorp M, et al. Duodenal infusion of donor feces for recurrent *Clostridium difficile*. *N Engl J Med.* 2013;368:407–15. <https://doi.org/10.1056/NEJMoa1205037>.
5. Ooijsaar RE, Terveer EM, Verspaget HW, et al. Clinical application and potential of fecal microbiota transplantation. *Annu Rev Med.* 2019;70:335–51. <https://doi.org/10.1146/annurev-med-111717-122956>.
6. Terveer EM, Vendrik KE, Ooijsaar RE, et al. Faecal microbiota transplantation for *Clostridioides difficile* infection: four years' experience of The Netherlands Donor Feces Bank. *United European Gastroenterol J.* 2020.
7. Jorgensen SMD, Hvas CL, Dahlerup JF, et al. Banking feces: a new frontier for public blood banks?. *Transfusion.* 2019;59:2776–82. <https://doi.org/10.1111/trf.15422>.
8. Cammarota G, Ianiro G, Kelly CR, et al. International consensus conference on stool banking for faecal micro- biota transplantation in clinical practice. *Gut.* 2019;68:2111–21. <https://doi.org/10.1136/gutjnl-2019-319548>.
9. Mullish BH, Quraishi MN, Segal JP, et al. The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications: joint British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS) guidelines. *Gut.* 2018;67:1920–41. <https://doi.org/10.1136/gutjnl-2018-316818>.
10. Peri R, Aguilar RC, Tuffers K, et al. The impact of technical and clinical factors on fecal microbiota transfer outcomes for the treatment of recurrent *Clostridioides difficile* infections in Germany. *United Eur Gastroenterol J.* 2019;7:716–22. <https://doi.org/10.1177/2050640619839918>.
11. van Beurden YH, de Groot PF, van Nood E, et al. Complications, effectiveness, and long term follow-up of fecal microbiota transfer by nasoduodenal tube for treatment of recurrent *Clostridium difficile* infection. *United Eur Gastroenterol J.* 2017;5:868–79. <https://doi.org/10.1177/2050640616678099>.
12. Guyatt GH, Oxman AD, Vist GE, et al. GRADE: an emerging consensus on rating quality of evidence and strength of recommendations. *BMJ.* 2008;336:924–6. <https://doi.org/10.1136/bmj.39489.470347>.
13. HealthCare EDftQoM. *Guide to the Quality and safety of tissues and cells for human application.* 4th ed. Strasbourg, France: European Directorate for the Quality of Healthcare & Medicines; 2019. <https://www.edqm.eu/en/organs-tissues-and-cells-technical-guides>. Accessed December 10 2019.
14. Konig J, Siebenhaar A, Hogenauer C, et al. Consensus report: faecal microbiota transfer - clinical applications and procedures. *Aliment Pharmacol Therapeut.* 2017;45:222–39. <https://doi.org/10.1111/apt.13868>.
15. Verbeke F, Janssens Y, Wynendaele E, et al. Faecal microbiota transplantation: a regulatory hurdle?. *BMC Gastroenterol.* 2017;17. <https://doi.org/10.1186/s12876-017-0687-5>.
16. European Union Journal. *Directive 2004/23/EC of the European Parliament and of the Council of 31 March 2004 on setting standards of quality and safety for the donation, procurement, testing, processing, preservation, storage and distribution of human tissues and cells.* 2004. <https://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2004:102:0048:0058>. Accessed March 24 2019.
17. Keller JJ, Vehreschild MJ, Hvas CL, et al. Stool for fecal microbiota transplantation should be classified as a transplant product and not as a drug. *United Eur Gastroenterol J.* 2019;7:1408–10. <https://doi.org/10.1177/2050640619887579>.
18. European Commission. *Competent Authorities on Substances of human origin expert group (CASoHO E01718), Meeting of the competent Authorities for Tissues and cells, 3-4 December 2014 Summary Report;* 2015. https://ec.europa.eu/health/sites/health/files/blood_tissues_organs/docs/ev_20141203_sr_en.pdf. Accessed March 24 2019.
19. Jorgensen SMD, Hansen MM, Erikstrup C, et al. Faecal microbiota transplantation: establishment of a clinical application framework. *Eur J Gastroenterol Hepatol.* 2017;29:e36–45. <https://doi.org/10.1097/MEG.0000000000000958>.
20. International Council for Harmonisation of Technical Requirements for Pharmaceuticals for Human Use (ICH). *Clinical safety data management: definitions and standards for expedited reporting* E2A; 1994. <https://www.ich.org/products/guidelines/>

- [efficacy/article/efficacy-guidelines.html](#). Accessed October 9 2020.
21. Wang S, Xu M, Wang W, et al. Systematic review: adverse events of fecal microbiota transplantation. *PLoS One*. 2016;11:e0161174. <https://doi.org/10.1371/journal.pone.0161174>.
 22. Baxter M, Colville A. Adverse events in faecal microbiota transplant: a review of the literature. *J Hosp Infect*. 2016;92:117–27. <https://doi.org/10.1016/j.jhin.2015.10.024>.
 23. *Detailed guidance on the collection, verification and presentation of adverse event/reaction reports arising from clinical trials on medicinal products for human use ('CT-3')*. 2011. [https://eur-lex.europa.eu/legal-content/EN/TXT/?uri=CELEX:52011XC0611\(01\)](https://eur-lex.europa.eu/legal-content/EN/TXT/?uri=CELEX:52011XC0611(01)). Accessed December 10 2019.
 24. Edwards IR, Biriell C. Harmonisation in pharmacovigilance. *Drug Saf*. 1994;10:93–102. <https://doi.org/10.2165/00002018-199410020-00001>.
 25. Mattila E, Uusitalo-Seppala R, Wuorela M, et al. Fecal transplantation, through colonoscopy, is effective therapy for recurrent *Clostridium difficile* infection. *Gastroenterology*. 2012;142:490–6. <https://doi.org/10.1053/j.gastro.2011.11.037>.
 26. Satokari R, Mattila E, Kainulainen V, et al. Simple faecal preparation and efficacy of frozen inoculum in faecal microbiota transplantation for recurrent *Clostridium difficile* infection - an observational cohort study. *Aliment Pharmacol Therapeut*. 2015;41:46–53. <https://doi.org/10.1111/apt.13009>.
 27. Lahtinen P, Mattila E, Anttila VJ, et al. Faecal micro-biota transplantation in patients with *Clostridium difficile* and significant comorbidities as well as in patients with new indications: a case series. *World J Gastroenterol*. 2017;23:7174–84. <https://doi.org/10.3748/wjg.v23.i39.7174>.
 28. Gorzelak MA, Gill SK, Tasnim N, et al. Methods for improving human gut microbiome data by reducing variability through sample processing and storage of stool. *PLoS One*. 2015;10:e0134802. <https://doi.org/10.1371/journal.pone.0134802>.
 29. Sinha R, Chen J, Amir A, et al. Collecting fecal samples for microbiome analyses in epidemiology studies. *Cancer Epidemiol Biomarkers Prev*. 2016;25:407–16. <https://doi.org/10.1158/1055-9965.EPI-15-0951>.
 30. Guo Y, Li SH, Kuang YS, et al. Effect of short-term room temperature storage on the microbial community in infant fecal samples. *Sci Rep*. 2016;6:26648. <https://doi.org/10.1038/srep26648>.
 31. Shaw AG, Sim K, Powell E, et al. Latitude in sample handling and storage for infant faecal microbiota studies: the elephant in the room?. *Microbiome*. 2016;4. <https://doi.org/10.1186/s40168-016-0186-x>.
 32. Cammarota G, Masucci L, Ianaro G, et al. Randomised clinical trial: faecal microbiota transplantation by colonoscopy vs. vancomycin for the treatment of recurrent *Clostridium difficile* infection. *Aliment Pharmacol Therapeut*. 2015;41:835–43. <https://doi.org/10.1111/apt.13144>.
 33. Bakken JS, Borody T, Brandt LJ, et al. Treating *Clostridium difficile* infection with fecal microbiota transplantation. *Clin Gastroenterol Hepatol*. 2011;9:1044–9. <https://doi.org/10.1016/j.cgh.2011.08.014>.
 34. Goldenberg SD, Batra R, Beales I, et al. Comparison of different strategies for providing fecal microbiota transplantation to treat patients with recurrent *Clostridium difficile* infection in two English hospitals: a review. *Infect Dis Therapy*. 2018;7:71–86. <https://doi.org/10.1007/s40121-018-0189-y>.
 35. Aas J, Gessert CE, Bakken JS. Recurrent *Clostridium difficile* colitis: case series involving 18 patients treated with donor stool administered via a nasogastric tube. *Clin Infect Dis*. 2003;36:580–5. <https://doi.org/10.1086/367657>.
 36. Lagier JC, Delord M, Million M, et al. Dramatic reduction in *Clostridium difficile* ribotype 027-associated mortality with early fecal transplantation by the nasogastric route: a preliminary report. *Eur J Clinical Microbiol Infect Dis: official publication of the European Society of Clinical Microbiology*. 2015;34:1597–601. <https://doi.org/10.1007/s10096-015-2394-x>.
 37. Girotra M, Garg S, Anand R, et al. Fecal microbiota transplantation for recurrent *Clostridium difficile* infection in the elderly: long-term outcomes and microbiota changes. *Dig Dis Sci*. 2016;61:3007–15. <https://doi.org/10.1007/s10620-016-4229-8>.
 38. Fischer M, Sipe BW, Rogers NA, et al. Faecal microbiota transplantation plus selected use of vancomycin for severe-complicated *Clostridium difficile* infection: description of a protocol with high success rate. *Aliment Pharmacol Therapeut*. 2015;42:470–6. <https://doi.org/10.1111/apt.13290>.
 39. Kelly CR, de Leon L, Jasutkar N. Fecal microbiota transplantation for relapsing *Clostridium difficile* infection in 26 patients: methodology and results. *J Clin Gastroenterol*. 2012;46:145–9. <https://doi.org/10.1097/MCG.0b013e318234570b>.
 40. Fischer M, Sipe B, Cheng YW, et al. Fecal microbiota transplant in severe and severe-complicated *Clostridium difficile*: a promising treatment approach. *Gut Microb*. 2017;8:289–302. <https://doi.org/10.1080/19490976.2016.1273998>.
 41. Hirsch BE, Saraiya N, Poeth K, et al. Effectiveness of fecal-derived microbiota transfer using orally administered capsules for recurrent *Clostridium difficile* infection. *BMC Infect Dis*. 2015;15:191. <https://doi.org/10.1186/s12879-015-0930-z>.
 42. MacConnachie AA, Fox R, Kennedy DR, et al. Faecal transplant for recurrent *Clostridium difficile*-associated diarrhoea: a UK case series. *QJM*. 2009;102:781–4. <https://doi.org/10.1093/qjmed/hcp118>.
 43. Rubin TA, Gessert CE, Aas J, et al. Fecal microbiome transplantation for recurrent *Clostridium difficile* infection: report on a case series. *Anaerobe*. 2013;19:22–6. <https://doi.org/10.1016/j.anaerobe.2012.11.004>.
 44. Walton J, Burns D, Gaehle KE. Process and outcome of fecal microbiota transplants in patients with recurrent *Clostridium difficile* infection: a prospective study. *Gastroenterol Nursing: the official journal of the Society of Gastroenterology Nurses and Associates*. 2017;40:411–9. <https://doi.org/10.1097/SGA.0000000000000233>.
 45. Kelly CR, Khoruts A, Staley C, et al. Effect of fecal microbiota transplantation on recurrence in multiply recurrent *Clostridium difficile* infection: a randomized trial. *Ann Intern Med*. 2016;165:609–16. <https://doi.org/10.7326/M16-0271>.
 46. Patel NC, Griesbach CL, DiBaise JK, et al. Fecal microbiota transplant for recurrent *Clostridium difficile* infection: Mayo Clinic in Arizona experience. *Mayo Clin Proc*. 2013;88:799–805. <https://doi.org/10.1016/j.mayocp.2013.04.022>.
 47. Bamba S, Nishida A, Imaeda H, et al. Successful treatment by fecal microbiota transplantation for Japanese patients with refractory *Clostridium difficile* infection: a prospective case series. *J Microbiol, Immunol Infect*. 2019;52:663–6. <https://doi.org/10.1016/j.jmii.2017.08.027>.
 48. Pathak R, Enuh HA, Patel A, et al. Treatment of relapsing *Clostridium difficile* infection using fecal microbiota transplantation. *Clin Exp Gastroenterol*. 2013;7:1–6. <https://doi.org/10.2147/CEG.S53410>.
 49. Lee CH, Steiner T, Petrof EO, et al. Frozen vs fresh fecal microbiota transplantation and clinical resolution of diarrhea in patients with recurrent *Clostridium difficile* infection: a randomized clinical trial. *J Am Med Assoc*. 2016;315:142–9. <https://doi.org/10.1001/jama.2015.18098>.

50. Fischer M, Kao D, Kelly C, et al. Fecal microbiota transplantation is safe and efficacious for recurrent or refractory *Clostridium difficile* infection in patients with inflammatory bowel disease. *Inflamm Bowel Dis*. 2016;22:2402–9. <https://doi.org/10.1097/MIB.0000000000000908>.
51. Hota SS, Sales V, Tomlinson G, et al. Oral vancomycin followed by fecal transplantation versus tapering oral vancomycin treatment for recurrent *Clostridium difficile* infection: an open-label, randomized controlled trial. *Clin Infect Dis*. 2017;64:265–71. <https://doi.org/10.1093/cid/ciw731>.
52. Hota SS, Surangiwalwa S, Paterson AS, et al. Regional variability in fecal microbiota transplantation practices: a survey of the Southern Ontario fecal microbiota transplantation Movement. *CMAJ open*. 2018;6:E184–90. <https://doi.org/10.9778/cmajo.20170109>.
53. Costello SP, Conlon MA, Vuaran MS, et al. Faecal microbiota transplant for recurrent *Clostridium difficile* infection using long-term frozen stool is effective: clinical efficacy and bacterial viability data. *Aliment Pharmacol Therapeut*. 2015;42:1011–8. <https://doi.org/10.1111/apt.13366>.
54. Emanuelsson F, Claesson BE, Ljungstrom L, et al. Faecal microbiota transplantation and bacteriotherapy for recurrent *Clostridium difficile* infection: a retrospective evaluation of 31 patients. *Scand J Infect Dis*. 2014;46:89–97. <https://doi.org/10.3109/00365548.2013.858181>.
55. Hamilton MJ, Weingarden AR, Sadowsky MJ, et al. Standardized frozen preparation for transplantation of fecal microbiota for recurrent *Clostridium difficile* infection. *Am J Gastroenterol*. 2012;107:761–7. <https://doi.org/10.1038/ajg.2011.482>.
56. Browne HP, Forster SC, Anonye BO, et al. Culturing of 'unculturable' human microbiota reveals novel taxa and extensive sporulation. *Nature*. 2016;533:543–6. <https://doi.org/10.1038/nature17645>.
57. Gough E, Shaikh H, Manges AR. Systematic review of intestinal microbiota transplantation (fecal bacteriotherapy) for recurrent *Clostridium difficile* infection. *Clin Infect Dis*. 2011;53:994–1002. <https://doi.org/10.1093/cid/cir632>.
58. Ianiro G, Maida M, Burisch J, et al. Efficacy of different faecal microbiota transplantation protocols for *Clostridium difficile* infection: a systematic review and meta-analysis. *United Eur Gastroenterol J*. 2018;6:1232–44. <https://doi.org/10.1177/2050640618780762>.
59. O'Brien K, Osman M, Eysenbach L, et al. Su1737 Clinical efficacy of fecal microbiota transplantation for recurrent *Clostridium difficile* infection from an international public stool bank: results from a 1,406 patient multi-center cohort. *Gastroenterology*. 2016;150:S539–40. [https://doi.org/10.1016/s0016-5085\(16\)31854-6](https://doi.org/10.1016/s0016-5085(16)31854-6).
60. Ianiro G, Valerio L, Masucci L, et al. Predictors of failure after single faecal microbiota transplantation in patients with recurrent *Clostridium difficile* infection: results from a 3-year, single-centre cohort study. *Clin Microbiol Infect*. 2017;23:e337. <https://doi.org/10.1016/j.cmi.2016.12.025>.
61. Khoruts A, Rank KM, Newman KM, et al. Inflammatory bowel disease affects the outcome of fecal microbiota transplantation for recurrent *Clostridium difficile* infection. *Clin Gastroenterol Hepatol*. 2016;14:1433–8. <https://doi.org/10.1016/j.cgh.2016.02.018>.
62. Youngster I, Russell GH, Pindar C, et al. Oral, capsulized, frozen fecal microbiota transplantation for relapsing *Clostridium difficile* infection. *J Am Med Assoc*. 2014;312:1772–8. <https://doi.org/10.1001/jama.2014.13875>.
63. Agrawal M, Aroniadis OC, Brandt LJ, et al. The longterm efficacy and safety of fecal microbiota transplant for recurrent, severe, and complicated *Clostridium difficile* infection in 146 elderly individuals. *J Clin Gastroenterol*. 2016;50:403–7. <https://doi.org/10.1097/MCG.0000000000000410>.
64. Alrabaa S, Jariwala R, Zeitler K, et al. Fecal microbiota transplantation outcomes in immunocompetent and immunocompromised patients: a single-center experience. *Transplant Infect Dis*. 2017. <https://doi.org/10.1111/tid.12726>.
65. Brandt LJ, Aroniadis OC, Mellow M, et al. Long-term follow-up of colonoscopic fecal microbiota transplant for recurrent *Clostridium difficile* infection. *Am J Gastroenterol*. 2012;107:1079–87. <https://doi.org/10.1038/ajg.2012.60>.
66. Chin SM, Sauk J, Mahabamunuge J, et al. Fecal microbiota transplantation for recurrent *Clostridium difficile* infection in patients with inflammatory bowel disease: a single-center experience. *Clin Gastroenterol Hepatol*. 2017;15:597–9. <https://doi.org/10.1016/j.cgh.2016.11.028>.
67. Friedman-Korn T, Livovsky DM, Maharshak N, et al. Fecal transplantation for treatment of *Clostridium difficile* infection in elderly and debilitated patients. *Dig Dis Sci*. 2018;63:198–203. <https://doi.org/10.1007/s10620-017-4833-2>.
68. Garborg K, Waagsbo B, Stallemo A, et al. Results of faecal donor instillation therapy for recurrent *Clostridium difficile*-associated diarrhoea. *Scand J Infect Dis*. 2010;42:857–61. <https://doi.org/10.3109/00365548.2010.499541>.
69. Costello SP, Tucker EC, La Brooy J, et al. Establishing a Fecal microbiota transplant service for the treatment of *Clostridium difficile* infection. *Clin Infect Dis*. 2016;62:908–14. <https://doi.org/10.1093/cid/civ994>.
70. Jiang ZD, Ajami NJ, Petrosino JF, et al. Randomised clinical trial: faecal microbiota transplantation for recurrent *Clostridium difficile* infection fresh, or frozen, or lyophilised microbiota from a small pool of healthy donors delivered by colonoscopy. *Aliment Pharmacol Therapeut*. 2017;45:899–908. <https://doi.org/10.1111/apt.13969>.
71. Kao D, Roach B, Silva M, et al. Effect of oral capsule- vs colonoscopy-delivered fecal microbiota transplantation on recurrent *Clostridium difficile* infection: a randomized clinical trial. *J Am Med Assoc*. 2017;318:1985–93. <https://doi.org/10.1001/jama.2017.17077>.
72. Rohlke F, Surawicz CM, Stollman N. Fecal flora reconstitution for recurrent *Clostridium difficile* infection: results and methodology. *J Clin Gastroenterol*. 2010;44:567–70. <https://doi.org/10.1097/MCG.0b013e3181dadab10>.
73. Youngster I, Mahabamunuge J, Systrom HK, et al. Oral, frozen fecal microbiota transplant (FMT) capsules for recurrent *Clostridium difficile* infection. *BMC Med*. 2016;14. <https://doi.org/10.1186/s12916-016-0680-9>.
74. Youngster I, Sauk J, Pindar C, et al. Fecal microbiota transplant for relapsing *Clostridium difficile* infection using a frozen inoculum from unrelated donors: a randomized, open-label, controlled pilot study. *Clin Infect Dis*. 2014;58:1515–22. <https://doi.org/10.1093/cid/ciu135>.
75. Cohen NA, Livovsky DM, Yaakovovitch S, et al. A retrospective comparison of fecal microbial transplantation methods for recurrent *Clostridium difficile* infection. *Israel Med Assoc J*. 2016;18:594–9.
76. Lee CH, Belanger JE, Kassam Z, et al. The outcome and long-term follow-up of 94 patients with recurrent and refractory *Clostridium difficile* infection using single to multiple fecal microbiota transplantation via retention enema. *Eur J Clin Microbiol Infect Dis*. 2014;33:1425–8. <https://doi.org/10.1007/s10096-014-2088-9>.
77. Hamilton MJ, Weingarden AR, Unno T, et al. High-throughput DNA sequence analysis reveals stable engraftment of gut microbiota

- following transplantation of previously frozen fecal bacteria. *Gut Microb*. 2013;4:125–35. <https://doi.org/10.4161/gmic.23571>.
78. Thornitt DN, Sander B, la Cour M, et al. The effects of peroral glycerol on plasma osmolality in diabetic patients and healthy individuals. *Basic Clin Pharmacol Toxicol*. 2009;105:289–93. <https://doi.org/10.1111/j.1742-7843.2009.00462.x>.
 79. Staley C, Hamilton MJ, Vaughn BP, et al. Successful resolution of recurrent *Clostridium difficile* infection using freeze-dried, encapsulated fecal microbiota; pragmatic cohort study. *Am J Gastroenterol*. 2017;112:940–7. <https://doi.org/10.1038/ajg.2017.6>.
 80. Takahashi M, Ishikawa D, Sasaki T, et al. Faecal freezing preservation period influences colonization ability for faecal microbiota transplantation. *J Appl Microbiol*. 2019;126:973–84. <https://doi.org/10.1111/jam.14167>.
 81. Elliott RJ, Njenga M, Ladha A, et al. Stool processing speed and storage duration do not impact clinical effectiveness of fecal microbiota transplantation across 1,924 *Clostridium difficile* infection patients: 119. *Am J Gastroenterol*. 2016;111:S57.
 82. Sleight SC, Wigginton NS, Lenski RE. Increased susceptibility to repeated freeze thaw cycles in *Escherichia coli* following long-term evolution in a benign environment. *BMC Evol Biol*. 2006;6(104). <https://doi.org/10.1186/1471-2148-6-104>.
 83. Alukal J, Dutta SK, Surapaneni BK, et al. Safety and efficacy of fecal microbiota transplant in 9 critically ill patients with severe and complicated *Clostridium difficile* infection with impending colectomy. *J Dig Dis*. 2019;20:301–7. <https://doi.org/10.1111/1751-2980.12750>.
 84. Hocquart M, Lagier JC, Cassir N, et al. Early fecal microbiota transplantation improves survival in severe *Clostridium difficile* infections. *Clin Infect Dis*. 2018;66:645–50. <https://doi.org/10.1093/cid/cix762>.
 85. Ianiro G, Masucci L, Quaranta G, et al. Randomized clinical trial: single-infusion FMT versus multiple infusion FMT for the treatment of severe *C. difficile* infection. *Dig Liver Dis*. 2018;50: E66–7.
 86. Cheng YW, Phelps E, Nemes S, et al. Fecal microbiota transplant decreases mortality in patients with refractory severe or fulminant *Clostridioides difficile* infection. *Clin Gastroenterol Hepatol*. 2020;18:2234–43. <https://doi.org/10.1016/j.cgh.2019.12.029>.
 87. Bajaj JS, Kassam Z, Fagan A, et al. Fecal microbiota transplant from a rational stool donor improves hepatic encephalopathy: a randomized clinical trial. *Hepatology*. 2017;66:1727–38. <https://doi.org/10.1002/hep.29306>.
 88. Moayyedi P, Surette MG, Kim PT, et al. Fecal microbiota transplantation induces remission in patients with active ulcerative colitis in a randomized controlled trial. *Gastroenterology*. 2015;149:102–9. <https://doi.org/10.1053/j.gastro.2015.04.001>.
 89. Rossen NG, Fuentes S, van der Spek MJ, et al. Findings from a randomized controlled trial of fecal transplantation for patients with ulcerative colitis. *Gastroenterology*. 2015;149:110–8. <https://doi.org/10.1053/j.gastro.2015.03.045>.
 90. Paramsothy S, Kamm MA, Kaakoush NO, et al. Multidonor intensive faecal microbiota transplantation for active ulcerative colitis: a randomised placebo-controlled trial. *Lancet*. 2017;389:1218–28. [https://doi.org/10.1016/S0140-6736\(17\)30182-4](https://doi.org/10.1016/S0140-6736(17)30182-4).
 91. Costello SP, Hughes PA, Waters O, et al. Effect of fecal microbiota transplantation on 8-week remission in patients with ulcerative colitis: a randomized clinical trial. *J Am Med Assoc*. 2019;321:156–64. <https://doi.org/10.1001/jama.2018.20046>.
 92. Sokol H, Landman C, Seksik P, et al. Fecal microbiota transplantation to maintain remission in Crohn's disease: a pilot randomized controlled study. *Microbiome*. 2020;8. <https://doi.org/10.1186/s40168-020-0792-5>.
 93. Holvoet T, Joossens M, Jerina B, et al. Fecal microbiota transplantation in irritable bowel syndrome with predominant abdominal bloating: results from a double blind, placebo-controlled clinical trial. *Gastroenterology*. 2018;154:S130. [https://doi.org/10.1016/S0016-5085\(18\)30860-6](https://doi.org/10.1016/S0016-5085(18)30860-6).
 94. Aroniadis OC, Brandt LJ, Oneto C, et al. A double-blind, randomized, placebo-controlled trial of fecal microbiota transplantation capsules (FMT) for the treatment of diarrhea-predominant irritable bowel syndrome (IBS-D). *Gastroenterology*. 2018;154:S154–S155. [https://doi.org/10.1016/S2468-1253\(19\)30198-0](https://doi.org/10.1016/S2468-1253(19)30198-0).
 95. Halkjaer S, Christensen A, Lo B, et al. Fecal microbiota transplantation alters gut microbiota in patients with irritable bowel syndrome: results from a randomized, double-blind placebo controlled study. *Gastroenterology*. 2018;154:S181. <https://doi.org/10.1136/gutjnl-2018-316434>.
 96. Johnsen PH, Hilpusch F, Cavanagh JP, et al. Faecal microbiota transplantation versus placebo for moderate-to-severe irritable bowel syndrome: a double-blind, randomised, placebo-controlled, parallel group, single-centre trial. *Lancet Gastroenterol Hepatol*. 2018;3:17–24. [https://doi.org/10.1016/S2468-1253\(17\)30338-2](https://doi.org/10.1016/S2468-1253(17)30338-2).
 97. Holster S, Lindqvist CM, Repsilber D, et al. The effect of allogenic versus autologous fecal microbiota transfer on symptoms, visceral perception and fecal and mucosal microbiota in irritable bowel syndrome: a randomized controlled study. *Clin Transl Gastroenterol*. 2019;10:e00034. <https://doi.org/10.14309/ctg.0000000000000034>.
 98. El-Salhy M, Hatlebakk JG, Gilja OH, et al. Efficacy of faecal microbiota transplantation for patients with irritable bowel syndrome in a randomised, double-blind, placebo-controlled study. *Gut*. 2020;69:859–67. <https://doi.org/10.1136/gutjnl-2019-319630>.
 99. Lahtinen P, Jalanka J, Hartikainen A, et al. Randomised clinical trial: faecal microbiota transplantation versus autologous placebo administered via colonoscopy in irritable bowel syndrome. *Aliment Pharmacol Therapeut*. 2020;51:1321–31. <https://doi.org/10.1111/apt.15740>.
 100. Huttner BD, de Lastours V, Wassenberg M, et al. A 5-day course of oral antibiotics followed by faecal transplantation to eradicate carriage of multidrug-resistant *Enterobacteriaceae*: a randomized clinical trial. *Clin Microbiol Infect*. 2019;25:830–8. <https://doi.org/10.1016/j.cmi.2018.12.009>.
 101. Nicholson MR, Mitchell PD, Alexander E, et al. Efficacy of fecal microbiota transplantation for *Clostridium difficile* infection in children. *Clin Gastroenterol Hepatol*. 2020;18:612–9. <https://doi.org/10.1016/j.cgh.2019.04.037>.
 102. Ooijsaar RE, van Rossen TM, Vandenbroucke-Grauls C, et al. Faecal transplants for children with recurrent infections. *Ned Tijdschr Geneeskd*. 2019;163:D3739.
 103. Davidovics ZH, Michail S, Nicholson MR, et al. Fecal microbiota transplantation for recurrent *Clostridium difficile* infection and other conditions in children: a joint position paper from the North American Society for Pediatric Gastroenterology, Hepatology, and Nutrition and the European Society for Pediatric Gastroenterology, Hepatology, and Nutrition. *J Pediatr Gastroenterol Nutr*. 2019;68:130–43. <https://doi.org/10.1097/MPG.0000000000002205>.
 104. Lynch SV. Fecal microbiota transplantation for recurrent *Clostridium difficile* infection in pediatric patients: encouragement wrapped in caution. *J Pediatr Gastroenterol Nutr*. 2015;60:1–3. <https://doi.org/10.1097/MPG.0000000000000612>.
 105. Tariq R, Saha S, Wahab A, et al. Efficacy of fecal microbiota transplantation for *Clostridium difficile* infection in immunocompromised patients: a systematic review and meta-analysis. *Am J Gastroenterol*. 2018;113:S121.

106. DeFilipp Z, Bloom PP, Torres Soto M, et al. Drug-resistant *E. coli* bacteremia transmitted by fecal microbiota transplant. *N Engl J Med*. 2019;381:2043–50. <https://doi.org/10.1056/NEJMoa1910437>.
107. Ianiro G, Mullish BH, Kelly CR, et al. Reorganisation of faecal microbiota transplant services during the COVID-19 pandemic. *Gut*. 2020;69:1555–63. <https://doi.org/10.1136/gutjnl-2020-321829>.
108. Dailey FE, Turse EP, Daglilar E, et al. The dirty aspects of fecal microbiota transplantation: a review of its adverse effects and complications. *Curr Opin Pharmacol*. 2019;49:29–33. <https://doi.org/10.1016/j.coph.2019.04.008>.
109. Perler BK, Chen B, Phelps E, et al. Long-term efficacy and safety of fecal microbiota transplantation for treatment of recurrent *Clostridioides difficile* infection. *J Clin Gastroenterol*. 2020;54:701–6. <https://doi.org/10.1097/MCG.0000000000001281>.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of this article.

How to cite this article: Keller JJ, Ooijevaar RE, Hvas CL, et al. A standardised model for stool banking for faecal microbiota transplantation: A consensus report from a multidisciplinary UEG working group. *United European Gastroenterol J*. 2021;9:229–247. <https://doi.org/10.1177/2050640620967898>