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Estradiol-driven metabolism in transwomen associates with reduced circulating extracellular vesicle microRNA-224/452

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Abstract

Objective: Sex steroid hormones like estrogens have a key role in the regulation of energy homeostasis and metabolism. In transwomen, gender-affirming hormone therapy like estradiol (in combination with antiandrogenic compounds) could affect metabolism as well. Given that the underlying pathophysiological mechanisms are not fully understood, this study assessed circulating estradiol-driven microRNAs (miRs) in transwomen and their regulation of genes involved in metabolism in mice.

Methods: Following plasma miR-sequencing (seq) in a transwomen discovery ($n = 20$) and validation cohort ($n = 30$), we identified miR-224 and miR-452. Subsequent systemic silencing of these miRs in male C57Bl/6 J mice ($n = 10$) was followed by RNA-seq-based gene expression analysis of brown and white adipose tissue in conjunction with mechanistic studies in cultured adipocytes.

Results: Estradiol in transwomen lowered plasma miR-224 and -452 carried in extracellular vesicles (EVs) while their systemic silencing in mice and cultured adipocytes increased lipogenesis (white adipose) but reduced glucose uptake and mitochondrial respiration (brown adipose). In white and brown adipose tissue, differentially expressed (miR target) genes are associated with lipogenesis (white adipose) and mitochondrial respiration and glucose uptake (brown adipose).

Conclusion: This study identified an estradiol-driven post-transcriptional network that could potentially offer a mechanistic understanding of metabolism following gender-affirming estradiol therapy.

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Introduction

Sex hormones like estrogens have a key role in the regulation of energy homeostasis and metabolism (1). Particularly upon binding the estrogen receptor α (ESR1) or ER β (ESR2) in adipose tissue, estrogen is known to affect adiposity and insulin sensitivity (2). Also, regular estradiol administration (in combination with antiandrogenic compounds) in transwomen may affect energy metabolism by increasing total body fat (3), fasting insulin, and HOMA of insulin resistance (HOMA-IR) (4), thereby reducing peripheral insulin sensitivity (5). This sex hormone-associated decline in metabolic health increases the risk for type 2 diabetes and future cardiovascular disease (CVD) in transwomen (6). However, the underlying pathophysiological mechanisms are not fully understood.

It has become increasingly clear that post-transcriptional networks through an intricate interplay of non-coding RNAs such as microRNAs (miRs) and RNA binding proteins coordinate the expression of multiple sets of functionally related genes, that together shape the functional response of cells to a change in metabolic demand (7). Also in human adipose tissue, studies have addressed this miR-mediated coordination of gene expression via the complementarity of base pairing at 3'-untranslated regions (3'-UTR) of target mRNAs (8). Interestingly, sex-specific expression of miRs may result via at least two mechanisms: (i) double dosage of X-chromosome located (X-linked) miRs due to incomplete silencing of the X-chromosome and (ii) estrogen regulation of miR transcription and processing (9). Given that increasing evidence is linking estrogen regulation in adipose tissue to whole-body metabolism (10), sex-specific, adipose tissue-derived miRs (11) could potentially alter metabolism at distal tissue sites (12).

Therefore, to understand how trans-hormones (estradiol) associate with post-transcriptional regulation of functionally related genes in metabolism, this study assessed circulating estradiol-driven miRs in transwomen and their regulation of genes involved in metabolism in mice.

Methods

A full description of the methods can be found in the Supplementary methods (see section on [supplementary materials](#) given at the end of this article). Below a summary can be found.

Patients

Transwomen received oral treatment with a daily dose of both 50 mg cyproterone acetate (CPA) (Androcur[®], Bayer) and 4 mg estradiol valerate (Progynova[®], Bayer) or 100 μ g/24 h transdermal estradiol (System[®], Janssen-Cilag) twice a week, as previously described (13). Paired plasma samples were obtained at the start of treatment and after 1 year of daily estradiol administration. Patient characteristics of the transwomen cohort 1 and 2 are displayed in [Tables 1](#) and [2](#), respectively.

Plasma RNA isolation

Plasma RNA from each patient sample was isolated from 200 μ L EDTA-plasma by using the RNeasy Micro Kit (Qiagen).

Library preparation and next-generation sequencing of plasma miRs

Plasma miR sequencing was performed by Exiqon according to protocol. Samples were sequenced on the Illumina NextSeq 500 system. Experiments were conducted at Exiqon Services, Denmark.

Plasma EV isolation

Plasma EVs (70 nm) were isolated by applying 125 μ L human plasma to a 3.64 mL Sepharose CL-2B size-exclusion chromatography (SEC) column, as previously described (14).

Table 1 Clinical characteristics of the pilot $n = 20$ transwomen cohort.

	Baseline	Estradiol	P-value
Age (years)	35 \pm 13	36 \pm 13	
BMI (kg/m ²)	25.2 \pm 4.1	24.5 \pm 6.8	0.623
Estradiol (pmol/L)	96 \pm 21	310 \pm 246	0.001
Testosterone (nmol/L)	19.9 \pm 8.7	0.8 \pm 0.2	0.001
SBP (mmHg)	133 \pm 10	128 \pm 12	0.051
DBP (mmHg)	86 \pm 11	82 \pm 9	0.167
Hemoglobin (mmol/L)	10.2 \pm 0.5	9.0 \pm 0.2	0.002
Hematocrit (L/L)	0.47 \pm 0.02	0.43 \pm 0.02	0.004
Glucose (mmol/L)	5.6 \pm 0.5	5.5 \pm 0.6	0.824
Insulin (pmol/L)	72 \pm 44	105 \pm 67	0.002
Creatinine (μ mol/L)	79 \pm 9	79 \pm 13	0.913
Cholesterol (mmol/L)	4.59 \pm 0.86	4.05 \pm 0.71	0.002
Triglyceriden (mmol/L)	1.06 \pm 0.42	0.97 \pm 0.36	0.303
HDL-cholesterol (mmol/L)	1.38 \pm 0.37	1.16 \pm 0.30	0.001

HDL, high density lipoprotein; SBP, systolic blood pressure; DBP, diastolic blood pressure.

Table 2 Clinical characteristics of the validation $n = 30$ transwomen cohort.

	Baseline	Estradiol	P-value
Age (years)	34 ± 12	35 ± 13	
BMI (kg/m ²)	23.5 ± 6.1	24.9 ± 4.3	0.074
Estradiol (pmol/L)	86 ± 22	275 ± 231	0.001
Testosterone (nmol/L)	20.4 ± 6.3	0.8 ± 0.4	0.001
SBP (mmHg)	127 ± 10	122 ± 9	0.003
DBP (mmHg)	80 ± 9	75 ± 8	0.003
Hemoglobin (mmol/L)	9.8 ± 0.5	8.8 ± 0.5	0.001
Hematocrit (L/L)	0.45 ± 0.03	0.42 ± 0.02	0.005
Glucose (mmol/L)	5.4 ± 0.7	5.2 ± 0.7	0.226
Insulin (pmol/L)	50.1 ± 30.9	71.8 ± 49.4	0.036
Creatinine (μmol/L)	78.2 ± 8.8	73.1 ± 9.0	0.001
Cholesterol (mmol/L)	4.7 ± 1.1	4 ± 0.8	0.001
Triglyceriden (mmol/L)	1.1 ± 0.5	0.9 ± 0.3	0.033
HDL-cholesterol (mmol/L)	1.4 ± 0.3	1.1 ± 0.3	0.001

HDL, high density lipoprotein; SBP, systolic blood pressure; DBP, diastolic blood pressure.

qPCR validation of plasma miR, EV miRs, and mRNA expression

Selected miRs were validated with quantitative PCR, using individual samples that comprised the pooled samples that were used for plasma miR sequencing. Taqman primers (Cat. 4427975, Thermo Fisher Scientific) were used according to the manufacturer's protocols. Target gene mRNA primer sequences are listed in Supplementary Table 2.

Animal experiments

Male C57Bl/6J mice ($n = 10$ per group, age = 8 weeks, Charles River Nederland) were randomized in three groups and received two s.c. injections of 25 mg/kg locked nucleic acid (LNA)-modified antisense miR-224 (antimiR-224), miR-452 (antimiR-452), or scrambled miR sequence (scramblemiR). Five days before injection, mice were individually housed in fully automated metabolic cages (LabMaster System, TSE Systems).

Plasma ELISA

Plasma insulin concentrations were measured by ELISA (Mercodia, 10-1247).

Clearance of radiolabeled glucose and lipoprotein-like particles

Glycerol tri[³H]oleate-labeled lipoprotein-like triglyceride (TG)-rich emulsion particles (80 nm) were prepared as

previously described (15) and [¹⁴C]deoxyglucose ([¹⁴C]DG) was added in a ³H:¹⁴C = 4:1 ratio. After 6 h of fasting, mice were injected with 200 μL of emulsion particles (1 mg TG per mouse) via the tail vein. After 15 min, organs were harvested and dissolved overnight at 56°C in Solvable (Perkin Elmer).

Tissue histology and immunohistochemistry

Formalin-fixed interscapular BAT (iBAT), s.c.WAT, and gonadal WAT (gWAT) were dehydrated in 70% EtOH, embedded in paraffin, and cut into 5-μm sections. Sections were stained with hematoxylin and eosin (H&E) using standard protocols.

Mapping and analysis of RNA-seq data

Mus musculus reference version GRCm38.p4 was used for the alignment of samples. The reads were mapped to the reference sequences using a short-read aligner based on the Burrows–Wheeler Transform. The read counts were loaded into the DESeq package v 1.10.1, a statistical package within the R platform v2.15.3. Additionally, RPKM/FPKM (reads/fragments per kilobase of exon per million reads mapped) values were calculated.

Pathway analysis

Pathway analysis was carried out using Ingenuity Pathway Analysis (IPA) software.

Cell culture of murine brown adipocytes and 3T3-L1 white adipocytes

Brown preadipocytes were isolated from interscapular BAT depots of 5-week-old male C57BL/6J mice as previously described (16), and upon confluence, cells were differentiated. Experiments were performed on days 12–14 of differentiation. 3T3-L1 preadipocytes (Zenbio) were differentiated in growth medium (DMEM/Ham's F-12 medium (1:1, v/v) supplemented with HEPES pH 7.4, 10% heat-inactivated FBS (Life Technologies Europe), human insulin, dexamethasone, penicillin and streptomycin, 3-isobutyl-1-methylxanthine (IBMX) and PPAR_γ agonist rosiglitazone.

Cell treatment

Differentiated 3T3-L1 adipocytes and differentiated brown adipocytes were incubated with 100 nM 17-β estradiol

(E2758, Sigma–Aldrich) for 48 h. After 48 h, cells were washed with PBS combined with Trizol to isolate RNA.

Oxygen consumption and extracellular acidification of murine brown adipocytes

Oxygen consumption rate (OCR) and extracellular acidification rate (ECAR) were measured using the Seahorse XF96 analyzer (Agilent Technologies).

Mitotracker experiment

Immortalized murine brown adipocytes were transfected with LNA-antimiR-224 and -452 and incubated for 30 min with MitoTracker Green FM (125 nM; Thermo Fisher) and MitoTracker RedCMXRos (250 nM; Thermo Fisher) in DMEM/F12 (Sigma) without FBS.

Glucose uptake experiments

Glucose uptake by immortalized murine brown adipocytes transfected with LNA-antimiR-224 and -452 was performed using a glucose uptake colorimetric assay kit (Sigma–Aldrich/Merck) according to the manufacturer's protocol.

Statistical analysis

Differential expression analysis of plasma miRs in the next generation sequencing experiment was done using the EdgeR statistical software package (Bioconductor). For normalization, the trimmed mean of the M-values method was used based on log-fold and absolute gene-wise changes in expression levels between samples (TMM normalization). All other data are expressed as mean \pm S.E.M. Variable distribution was tested using the Kolmogorov–Smirnov test for normal distribution. In addition, multivariable linear regression was used to adjust for possible confounders. Statistical analysis was performed with GraphPad software using a two-tailed paired or unpaired Student's *t*-test or ANOVA with Bonferroni's *post hoc* test.

Study approval

These studies were approved by the Institutional Review Boards of both the VU University Medical Center (Amsterdam, The Netherlands) and the Leiden University Medical Center (Leiden, The Netherlands) and complied with the ethical principles of the Declaration of Helsinki. All patients gave written informed consent. All animal

experiments and protocols were approved by the animal welfare committee of the veterinary authorities of the Leiden University Medical Center (Leiden, The Netherlands).

Data and resource availability

The datasets generated during and/or analyzed during the current study are available in the Gene Expression Omnibus (GEO) and are accessible under GSE147966 (reviewer token for access: ahmfqeqglzwpchf). No applicable resources were generated or analyzed during the current study.

Results

Identification of circulating estradiol-responsive miRs in transwomen

We performed a pilot study assessing plasma miR profiles in transwomen who received 1 year of estradiol supplementation prior to surgical transition (17). Next-generation sequencing of plasma miRs in four pooled EDTA-plasma samples (pooled based on age and estradiol concentration) from five transwomen, before ($n = 20$) and after ($n = 20$) 1 year of estradiol treatment, identified 667 miRs (Fig. 1A) of which 33 were differentially expressed (Supplementary Data 1). Specifically, miR-224, -122-5p, -539-5p, 23a-3p, -133b, -452-5p, 23b-3p, -3913-5p, -144-5p, -331-5p, -766-3p, -874-3p, 30b-5p, and miR-490-3p levels displayed a significant decrease, while circulating levels of miR-3138, -215-5p, -483-5p, -let-7b-5p, -6787-3p, -184, -3679-5p, -370-5p, -615-3p, -let-7d-3p, -432-5p, -139-3p, -6747-3p, -433b-3p, -584-5p, -3198, -3940-3p, -625-3p, and miR-6786-3p increased (Fig. 1B). The following miRs were selected for RT-qPCR validation: miR-224, -122, -23a, -452, -139, -133b, -215, -9, -874, -30b, -483, -539, and -652. These miRs were selected based on (i) a sufficient level of regulation across sample groups (>2-fold up- or down-regulation, corresponding to ± 1.0 log-FoldChange) and (ii) differential expression (based on *P*-value) as a result of 1-year estradiol treatment.

Validation of estrogen-responsive miRs in independent transgender cohorts

To validate these circulating miRs, we conducted a three-step validation process. First, we reassessed plasma miRs (Supplementary Fig. 1) by RT-qPCR in the individual samples of the same transwomen cohort that was used in the pilot study, before ($n = 20$) and after ($n = 20$) estradiol

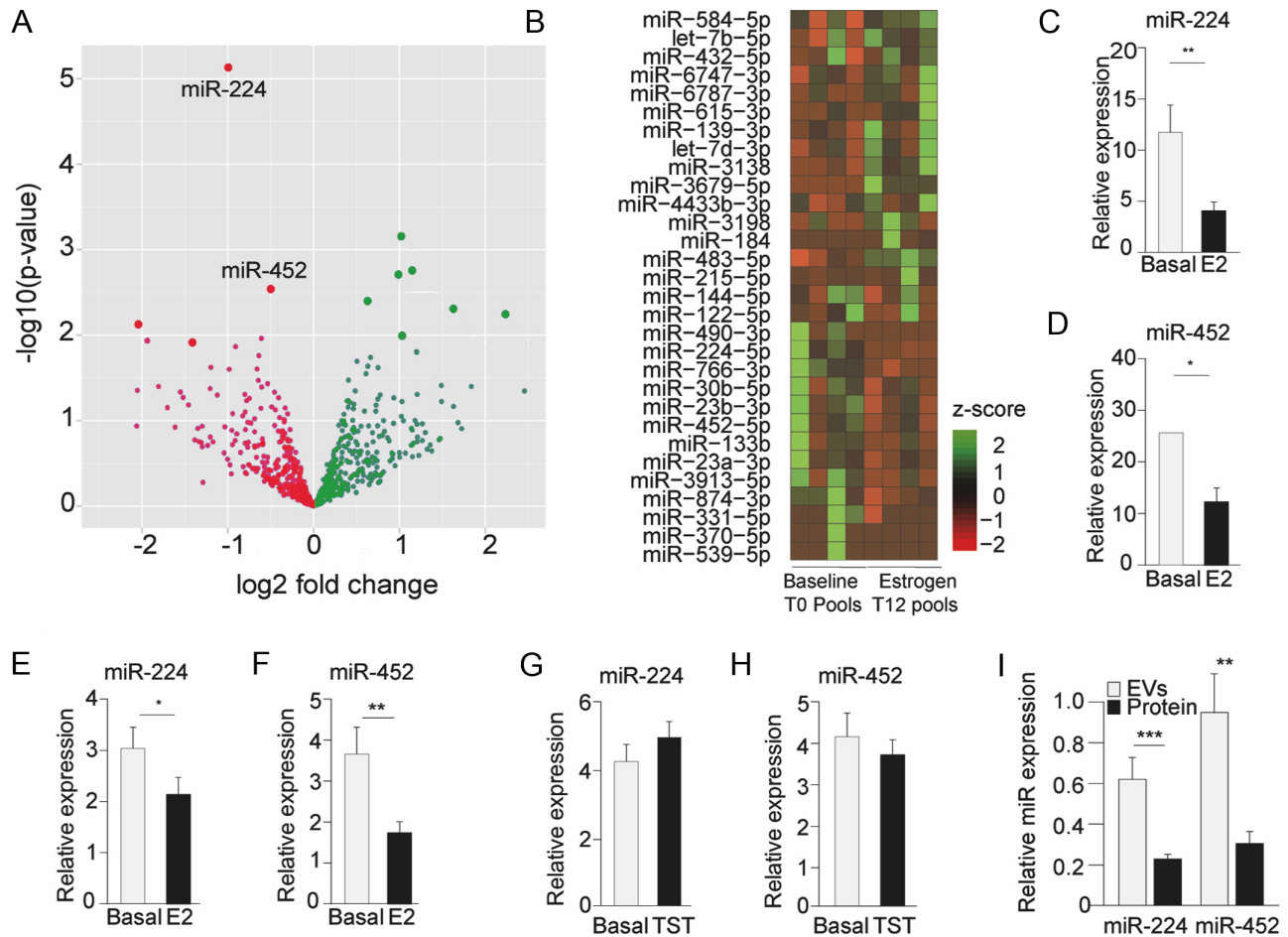


Figure 1

Identification of circulating miRNAs in human plasma of male-to-female transgender persons (transwomen). (A) Volcano plot depicting differentially expressed miRNAs after estrogen treatment. (B) Heatmap depicting differentially expressed estrogen-responsive miRNAs. (C and D) RT-qPCR validation of the differentially expressed miR-224 and miR-452 in the same cohort of 20 transwomen. (E and F) RT-qPCR validation of miR-224 and miR-452 in a second, independent transwomen cohort ($n = 39$). (G and H) Expression levels of circulating miR-224 and miR-452 in transmen after testosterone treatment ($n = 51$). (I) Higher expression levels of miR-224 and miR-452 in extracellular vesicles (EVs) compared to total plasma protein both at baseline and after estrogen supplementation in transwomen ($n = 20$). Basal, baseline state before hormone supplementation; E2, estradiol; TST, testosterone. Data are represented as means \pm s.e.m. * $P \leq 0.05$, ** $P \leq 0.01$, *** $P \leq 0.001$ according to the two-tailed paired and unpaired Student's t -test.

supplementation (Table 1 for clinical characteristics). Out of the 13 selected miRNAs, we confirmed a significant decrease in circulating levels of miR-224 (Fig. 1C), miR-452 (Fig. 1D), and miR-133b (Supplementary Fig. 1). Subsequently, in a second independent transwomen cohort (Patient characteristics in Table 2), we again assessed plasma levels of miR-224, miR-452, and miR-133b and demonstrated decreased plasma miR-224 and miR-452 (Fig. 1E and F) while plasma miR-133b was not affected (Supplementary Fig. 2). Because next to estradiol treatment, transwomen also received a daily dose of cyproterone acetate (CPA) to block testosterone secretion, we sought to exclude

testosterone effects on the observed differences in plasma miRNAs. Therefore, we assessed plasma miR-224, -452, and -133b in female-to-male transgender persons (transmen, $n = 50$, clinical characteristics in Supplementary Table 1) after 1 year of testosterone treatment. In transmen, these miRNAs were not affected (Supplementary Fig. 3). The supposedly coordinated regulation of miR-224 and miR-452 by estradiol could be consistent with the fact that both miRNAs are transcribed from a single transcript from the GABRE locus on the X-chromosome (Source: miRbase version 21). Following the above and given that both miRNAs are predominantly expressed in adipose tissue (18),

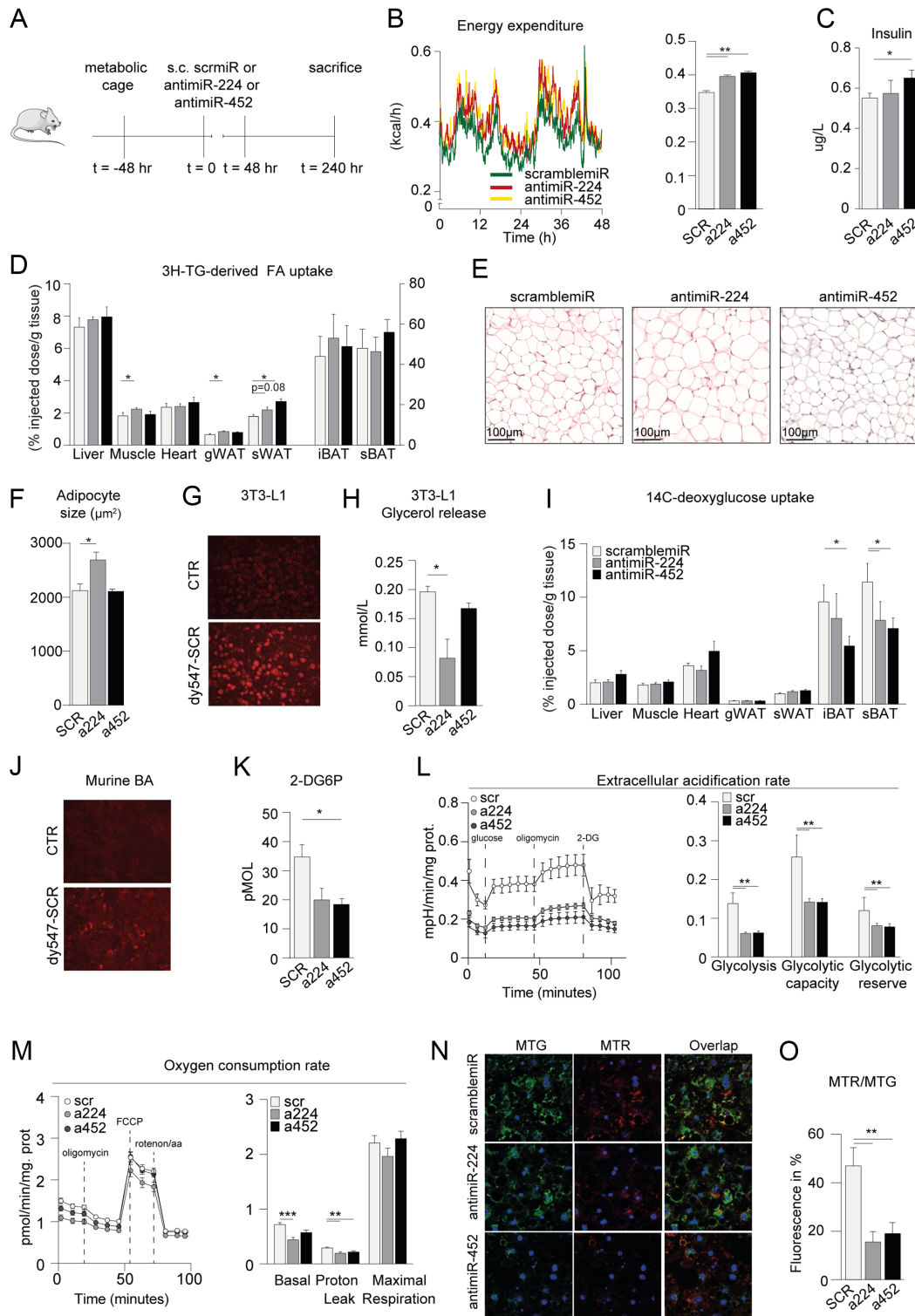


Figure 2

Systemic silencing of miR-224 and miR-452 affects adipocyte-specific nutrient uptake. (A) Experimental set up, mice were housed in calorimetric cages 48 h before i.p. injection of scramble miR ($n = 10$), anti-miR-224 ($n = 10$), and anti-miR-452 ($n = 10$). 240 h after injection mice were sacrificed. (B) Energy expenditure (EE) was assessed and quantified over 48 h. (C) miR-452 silencing increased plasma insulin levels. (D) miR-224 silencing increased triglyceride-derived fatty acid uptake in skeletal muscle, gonadal WAT

we subsequently focused our experiments on both miR-224 and miR-452. We next assessed whether miR-224 and miR-452 associate with EVs or plasma proteins and used size-exclusion chromatography (SEC, Supplementary Fig. 4) to isolate EVs from plasma of the transwomen pilot study cohort ($n=20$). This identified an approximately three-fold higher expression of both miRs in EVs compared to the plasma protein fraction (Fig. 11). Lastly, we examined the correlation of these miRs with the calculated homeostatic model assessment for insulin resistance (HOMA-IR) in transwomen to assess their association with metabolism. Particularly miR-224 displayed a negative correlation with HOMA-IR ($R = -0.38$, $P=0.03$, while miR-452 and HOMA-IR were not significantly correlated (data not shown).

Systemic silencing of miR-224 and miR-452 in mice affects adipocyte-specific nutrient uptake

To investigate the function of miR-224 and miR-452, we set up a mouse study allowing silencing these miRs to study its impact on lipid and glucose metabolism. First, male C57BL/6J mice ($n=10$) were housed in metabolic cages and injected with locked nucleic acid (LNA) antisense (anti-miR-224, LNA anti-miR-452, or control scramble miR (Fig. 2A). Within 48 h, anti-miR-224 and anti-miR-452 injection in mice elevated energy expenditure (Fig. 2B) although body weight, respiratory exchange ratio, carbohydrate oxidation, and fat oxidation were not significantly altered (data not shown). Given the altered energy expenditure we studied whether plasma insulin levels were affected and found that in anti-miR-452 treated mice, insulin levels were significantly increased after 48 h (Fig. 2C). Therefore, to

study the effect of both miRs on nutrient uptake, we next assessed triglyceride (TG)-derived fatty acid (FA) uptake in metabolic tissues such as skeletal muscle and adipose tissue, using glycerol tri- ^3H oleate packaged in TG-rich lipoprotein-like particles. In anti-miR-224 treated mice, ^3H oleate uptake was significantly increased in skeletal muscle and gonadal white adipose tissue (gWAT), and a trend toward increased ^3H oleate uptake was observed in s.c.WAT ($P=0.08$), while anti-miR-452 treatment of mice increased ^3H oleate uptake in s.c. white adipose tissue (Fig. 2D). To study whether the observed increase in ^3H oleate uptake led to an increase in white adipocyte size, we performed HE staining of s.c.WAT tissue, which demonstrated a marked increase in white adipocyte size in the anti-miR-224-treated mice (Fig. 2E and F). *In vitro*, in male murine 3T3-L1 adipocytes, miR-224 silencing (Fig. 2G indicates fluorescent signal detection upon dy547-labeled siRNA transfection indicating successful LNA transfer in these cells) similarly affected lipid uptake given the decrease in glycerol release into the culture medium (Fig. 2H) thereby confirming our findings *in vivo*. Subsequently, we studied *in vivo* whether miR silencing altered tissue-specific glucose uptake in mice by injecting ^{14}C deoxyglucose (DG). Anti-miR-224 treatment reduced ^{14}C DG uptake in subscapular brown adipose tissue (sBAT), while anti-miR-452 treatment reduced ^{14}C DG in both interscapular brown adipose tissue (iBAT) and sBAT (Fig. 2I). Next, in cultured brown adipocytes, we found less glucose uptake following miR-452 silencing only (Fig. 2K) (following confirmation of successful LNA transfer by fluorescent signal detection in brown adipocytes in Fig. 2J). In addition, we observed less glycolysis, glycolytic capacity, and glycolytic reserve in anti-miR-224/452 treated cells as determined with Seahorse

Figure 2 Continued

(gWAT), and s.c.WAT, while miR-452 silencing increased triglyceride-derived fatty acid uptake in s.c.WAT. (E and F) Hematoxylin and eosin (H&E) staining of white adipocyte size in anti-miR-224 and anti-miR-452 treated mice compared to scramble control mice. (G) Transfer of dy547-labeled siRNAs into male 3T3-L1 white adipocytes. (H) Decreased glycerol release in culture media of anti-miR-224 treated 3T3-L1 white adipocytes ($n=6$) only. (I) miR-224 silencing reduced deoxyglucose uptake in subscapular BAT (sBAT), while miR-452 silencing decreased deoxyglucose uptake in interscapular BAT (iBAT) and subscapular (sBAT). (J) Transfer of dy547-labeled siRNAs into murine male immortalized brown adipocytes. (K) Decreased glucose uptake in anti-miR-452 treated immortalized brown adipocytes ($n=4-6$). (L) Decreased extracellular acidification rate (ECAR) of anti-miR-224 and anti-miR-452 treated murine male immortalized brown adipocytes ($n=10$) followed by its quantification. (M) Oxygen consumption rate (OCR) of anti-miR-224 and anti-miR-452 treated murine male immortalized brown adipocytes ($n=10$) followed by its quantification. (N) Representative images of anti-miR-224 and anti-miR-452 treated, immortalized brown adipocytes ($n=4-6$) stained with MitoTracker Green FM (125 nM) and MitoTracker Red CMXRos (250 nM). Fluorescence of MitoTracker stained cells was imaged using a confocal laser scanning microscope (Leica TCS SP8, Leica Microsystems). (O) Quantification of MitoTracker Green (MTG) and MitoTrackers Red (MTR) using Imagej. SCR, scramble miR; a224, anti-miR-224; a452, anti-miR-452. Data are represented as means \pm S.E.M. * $P \leq 0.05$, ** $P \leq 0.01$ according to a one-way ANOVA, Bonferroni's *post-hoc* test.

respirometry (Fig. 2L). Because glucose consumption and glycolytic flux maintain mitochondrial respiration (19), we also measured mitochondrial respiration by proton flux in the culture media of anti-miR-224 and anti-miR-452 treated brown adipocytes. Although maximal respiration was not significantly affected, we observed a decrease in basal respiration in anti-miR-224 cells and a decrease in proton leak in anti-miR-224 and -452 treated cells (Fig. 2M). Next, we stained the mitochondria in anti-miR-224 and -452 treated murine brown adipocyte cell cultures using Mitotracker green (MTG; total) and Mitotracker red (MTR; active) (Fig. 2N), which stain mitochondria independent of- and dependent on membrane potential, respectively. After quantification of relative fluorescence intensity (MTR/MTG), which measures mitochondrial depolarization, a decrease in mitochondrial depolarization was seen in anti-miR-224 and -452 treated cells (Fig. 2O).

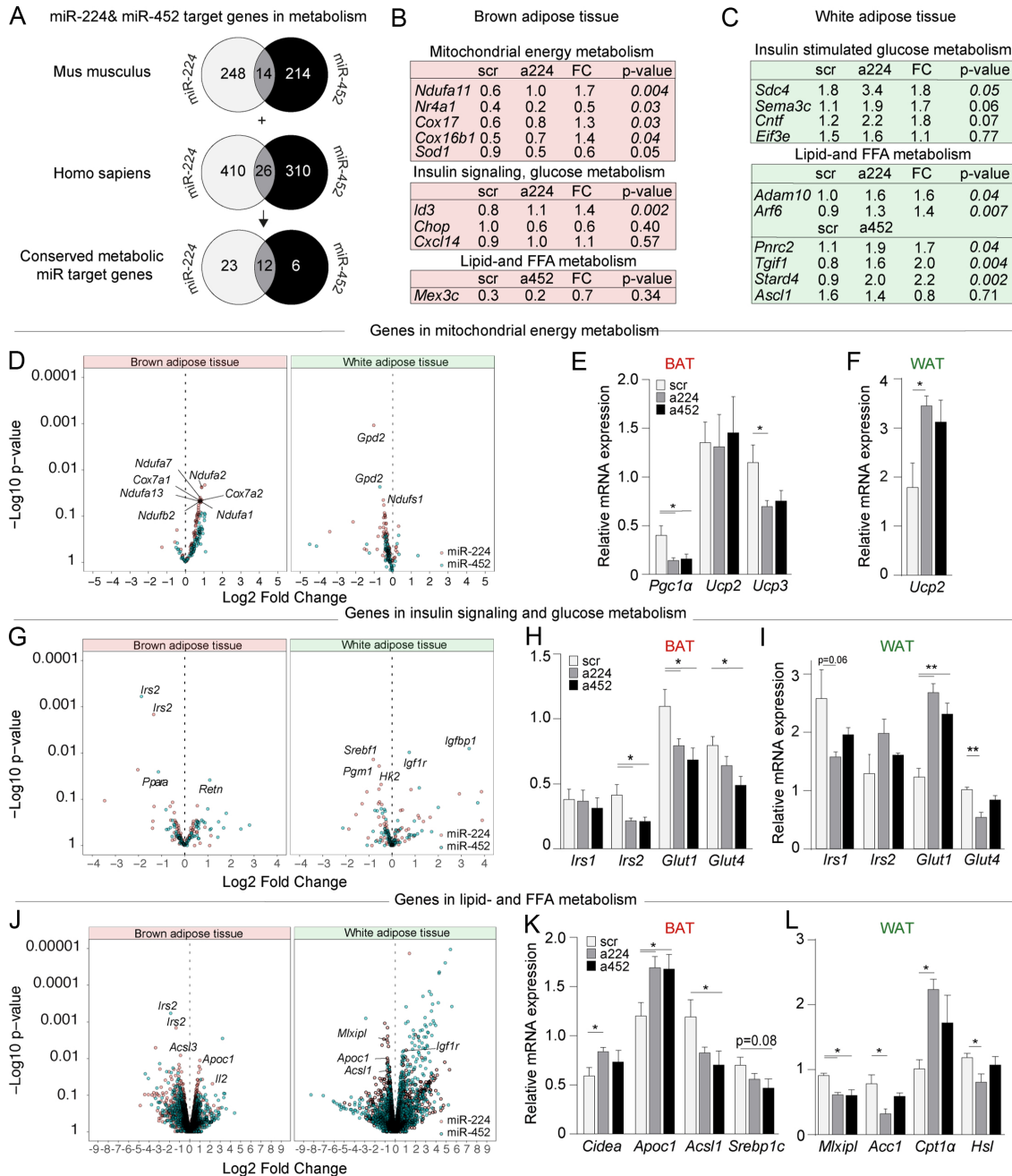
miR-224 and miR-452 target genes are involved in mitochondrial energy metabolism, insulin signaling, glucose metabolism, and lipogenesis

Given that silencing of these miRs stimulated fatty acid uptake in WAT and decreased glucose uptake in BAT, we next performed RNA sequencing (RNA-seq) of WAT and BAT to identify metabolism-associated gene expression changes. Interestingly, using Ingenuity Pathway Analysis (IPA), we found that both miR-224/452 target genes are strongly enriched for genes that associate with metabolism, potentially indicating a coordinated regulation of multiple metabolic pathways by these miRs (Fig. 3A). By further applying *in silico* analysis of the RNA-seq data with the IPA tool, we identified mitochondrial dysfunction and insulin receptor signaling in BAT as significant top canonical pathways affected (Supplementary Fig. 5). In contrast, in WAT tissue from anti-miR-224 treated mice, AMP-activated protein kinase (AMPK) signaling (regulates adipocyte metabolism) and signal transducer and activator of transcription 3 (STAT3) (controls lipogenesis and adipocyte hypertrophy) were affected, while anti-miR-452 treatment affected the HIPPO-pathway (adipocyte differentiation) and insulin receptor signaling. We subsequently used IPA to identify all potential miR-224/452 target genes that could serve as upstream regulators of differential top canonical pathways. When miR-224/452 target genes were plotted in heatmaps, many of them were upregulated in BAT and WAT (Supplementary Fig. 6, metabolic genes marked with an asterisk), and some key metabolism-

related target genes could be validated by qPCR (Fig. 3B and C). Specifically, we identified an increased expression of the miR-224 targets *Ndufa11*, *Cox17*, *Cox16b1*, and *Id3* in BAT (Fig. 3B), which associate with metabolic syndrome and obesity (20, 21). *Nr4a1* expression, an effector of BAT thermogenesis (22), was decreased with miR-224 silencing. Interestingly, in WAT (Fig. 3C), anti-miR-224 treatment coordinately upregulated genes that regulate lipid metabolisms such as *Arf6* (23, 24) and *Adam10* (25) or exacerbate insulin sensitivity like *Sema3C* (26) and *Sdc4* (27). Similarly, anti-miR-452 treated WAT tissue displayed higher expression of lipogenesis-associated genes, such as *Pnrc2* (28) and *Stard4* (29) as well as genes that promote insulin resistance like *Cntf* (30) or associate with weight gain and adiposity (*Tgfr-1*) (31, 32).

Adipose tissue-specific differential expression of genes involved in metabolism following miR-224 and miR-452 silencing

Next, we aimed to identify differential expression of genes that function in the top altered canonical metabolic pathways. To that end, we first generated gene-distribution volcano plots confirming differential expression of genes in mitochondrial energy metabolism (Fig. 3D), insulin signaling and glucose metabolism (Fig. 3G), and lipid metabolism (Fig. 3J) (RNA-seq data can be found in Supplementary Data 2, while a selected subset of differentially expressed genes involved in these pathways is displayed in Table 3). Then we validated the differential expression of several genes whose functions are known to have a major impact on energy metabolism by RT-qPCR. With regard to mitochondrial energy metabolism in BAT, qPCR validation confirmed a reduced expression of *Pgc1 α* (Fig. 3E and F), which regulates mitochondrial biogenesis and whose loss predisposes to insulin resistance (33). Given the *in vivo* decrease in BAT-specific glucose uptake, we also validated the loss of BAT-specific glucose uptake genes namely *Glut4* and *Irs2* (34, 35) (Fig. 3H). Similar decreases in key insulin signaling genes like *Irs1* and *Glut4* were observed in anti-miR-224 treated WAT (Fig. 3I). Finally, in lipid metabolism in BAT, we validated a lower expression of *Cidea* (Fig. 3K), which associates with lipogenesis (36), a decreased expression of *Acs11*, a promoter of FA oxidation (37), and increased *Apoc1* expression. In WAT-specific lipid metabolism, the expression of *Mlxipl*, *Acc1*, and *Hsl* was decreased (Fig. 3L), which collective decrease is associated with more FA uptake (38, 39, 40).

**Figure 3**

Next-generation sequencing (NGS)-derived differential expression and qPCR validation of (miR target) genes involved in mitochondrial energy, glucose, and lipid metabolism. (A) Both miR-224 and miR-452 have a strong bias toward target genes that associate with metabolism in general. (B) Differentially expressed miR-224- and miR-452 target genes involved in mitochondrial energy metabolism, insulin signaling, glucose metabolism, and lipid metabolism in BAT and (C) WAT tissue. (D) Volcano plot of NGS-derived differential expression of genes ($n = 4$ per group) involved in mitochondrial energy metabolism. (E) RT-qPCR validation ($n = 8-10$ per group) of NGS derived, differential expression of genes involved in mitochondrial energy metabolism in BAT and (F) WAT. (G) Volcano plot of NGS-derived differential expression of genes involved in insulin signaling and glucose metabolism followed by RT-qPCR validation of several key genes in (H) BAT and (I) WAT. (J) Volcano plot of NGS-derived differential expression of genes involved in lipid metabolism followed by RT-qPCR validation in (K) BAT and (L) WAT. Data are represented as means \pm S.E.M. * $P \leq 0.05$, ** $P \leq 0.01$ according to a one-way ANOVA, Bonferroni's *post-hoc* test.

Table 3 Next-generation sequencing derived differential gene expression (FPKM, $P < 0.05$, $n = 4$) of a selected set of key-metabolism genes involved in glucose and lipid metabolism in brown and white adipose tissue after systemic silencing of miR-224 and miR-452 in mice.

	Glucose metabolism	Lipid metabolism	Fatty acid metabolism	Mitochondrial energy metabolism
BAT				
AntimiR-224 - BAT	<i>Irs1</i> ↓, <i>Irs2</i> ↓, <i>Eif4ebp1</i> ↑, <i>Frs2</i> ↓, <i>Jun</i> ↓, <i>Fbp1</i> ↑, <i>Pik3ca</i> ↑	<i>Cidea</i> ↑, <i>Fabp4</i> ↑, <i>Acaa2</i> ↑, <i>Apoc1</i> ↑, <i>Pdhb</i> ↑, <i>Pdk4</i> ↓, <i>Idh3b</i> ↑, <i>Suc1g1</i> ↑, <i>Acaa1a</i> ↑, <i>Acca2a</i> ↑, <i>Acat1</i> ↑, <i>Acadv1</i> ↑, <i>Acs13</i> ↓	<i>Fabp5</i> ↑, <i>Slc27a3</i> ↓	<i>Ppargc1a</i> ↓, <i>Cox5b</i> ↑, <i>Cox6b1</i> ↑, <i>Cox6c</i> ↑, <i>Cox7a2</i> ↑, <i>Cox7a2l</i> ↑, <i>Cox7b</i> ↑, <i>Cyc1</i> ↑, <i>Ndufa1</i> ↑, <i>Ndufa11</i> ↑, <i>Ndufa2</i> ↑, <i>Ndufa3</i> ↑, <i>Ndufa4</i> ↑, <i>Ndufa6</i> ↑, <i>Ndufa7</i> ↑, <i>Ndufa8</i> ↑, <i>Ppa2</i> ↑, <i>Sdhb</i> ↑, <i>Slc25a21</i> ↓, <i>Ucp3</i> ↓
AntimiR-452 - BAT	<i>Insr</i> ↓, <i>Irs2</i> ↓, <i>Pfkfb3</i> ↓, <i>Sorbs1</i> ↓, <i>Pdk1</i> ↓, <i>Pir3cb</i> ↓, <i>Prkc1</i> ↓, <i>Srebf1</i> ↓, <i>Sorbs1</i> ↓, <i>Frs2</i> ↓	<i>Scd2</i> ↓, <i>Srebp1c</i> ↓, <i>Acs11</i> ↓	<i>Prkaa1</i> ↓, <i>Prkacb</i> ↓	<i>Ppargc1a</i> ↓, <i>Atp6v0a2</i> ↓, <i>Cox6c</i> ↑, <i>Lhpp</i> ↑, <i>Slc25a15</i> ↓, <i>Ucp2</i> ↑
WAT				
AntimiR-224 - WAT	<i>Igf1r</i> ↑, <i>Grb2</i> ↑, <i>Ptpn1</i> ↑, <i>Ptprf</i> ↑, <i>Adra1d</i> ↓, <i>Srebf1</i> ↓, <i>Glut4</i> ↓, <i>PI3K</i> ↑, <i>mTOR</i> ↓	<i>Gpr120</i> ↓, <i>Mlxipl</i> ↓, <i>Acc1</i> ↓, <i>Acc2</i> ↓, <i>Srebp1c</i> ↓, <i>Hsl</i> ↓, <i>Acs11</i> ↓, <i>Apoc1</i> ↓	<i>Acads</i> ↓, <i>Acs15</i> ↑, <i>Cpt1a</i> ↑, <i>Crat</i> ↓, <i>Hadha</i> ↓, <i>Lipe</i> ↓, <i>Hmgcs2</i> ↑	<i>Ucp2</i> ↑, <i>Cox7a2l</i> ↑, <i>Gadd45b</i> ↑
AntimiR-452 - WAT	<i>Gys1</i> ↓, <i>Fbp2</i> ↑, <i>Igf1r</i> ↑, <i>Igf1r</i> ↑, <i>Ptprf</i> ↑, <i>Araft</i>	<i>Gpr120</i> ↓, <i>Mlxipl</i> ↓, <i>Acc1</i> ↓, <i>Apoc1</i> ↓	<i>Slc27a2</i> ↑, <i>Prkacb</i> ↑	<i>Slc25a15</i> ↑, <i>Slc25a21</i> ↑

Discussion

This study demonstrates that estradiol in transwomen lowers circulating miR-224 and -452 carried in extracellular vesicles. Following systemic silencing of these miRs in mice and cultured adipocytes, lipid uptake was increased in skeletal muscle and WAT, while glucose uptake and mitochondrial respiration were decreased in BAT. Differentially expressed genes in these tissues were involved in mitochondrial energy metabolism, glucose uptake, and lipogenesis. As such, this study identified novel estradiol-driven post-transcriptional networks that could potentially offer a novel mechanistic understanding of metabolism following gender-affirming therapy (Figure 4).

In transwomen, it remains challenging to separate estradiol administration effects from androgen withdrawal effects. Nonetheless, this estradiol-mediated repression of the miR-224/452 cluster was demonstrated before in ER-positive breast cancer patients (41) and is consistent with the idea that miR clusters tend to be regulated in a similar way (42). Moreover, the female genetic background makes transmen not the most suitable control group to exclude testosterone withdrawal effects on these miRs. Still, in women, we previously demonstrated increased circulating levels of miR-224/452 in contrast to no significant effects in the transmen cohort (Fig. 1G and H) (43).

It could be further argued that transmen do not distinguish causal effects of estradiol from CPA treatment nor the fact that a change in androgen/estrogen ratio could alter miR levels. Therefore, different patient cohorts like men receiving androgen withdrawal as prostate cancer treatment or hypogonadal men before and after initiation of testosterone replacement therapy should serve as control groups. However, such particular patients have other confounding characteristics that could affect these miRs. For instance, the *Gabre-miR-224/452* locus is downregulated and hypermethylated in prostate cancer patients (44) while low testosterone levels following hypogonadism almost always correlate with the presence of diabetes mellitus (DM) (45). It is also important to note that CPA could have slight glucocorticoid effects as well (46). As such it could potentially affect glucose homeostasis as well which could impact the results of this study. Instead of studying transwomen treated with a combination of CPA and estradiol, more studies are needed to assess the impact of miRs on metabolism. For instance, in transwomen who receive other gonadotropin-releasing hormone receptor (GnRHR) agonists like leuprolide acetate in combination with estradiol, which was found to have no effect on insulin and glucose levels compared to CPA and estradiol (47). However, these sex hormone-mediated effects on the metabolism have been described for estradiol as well (47) while our studies in mice demonstrate significant miR-

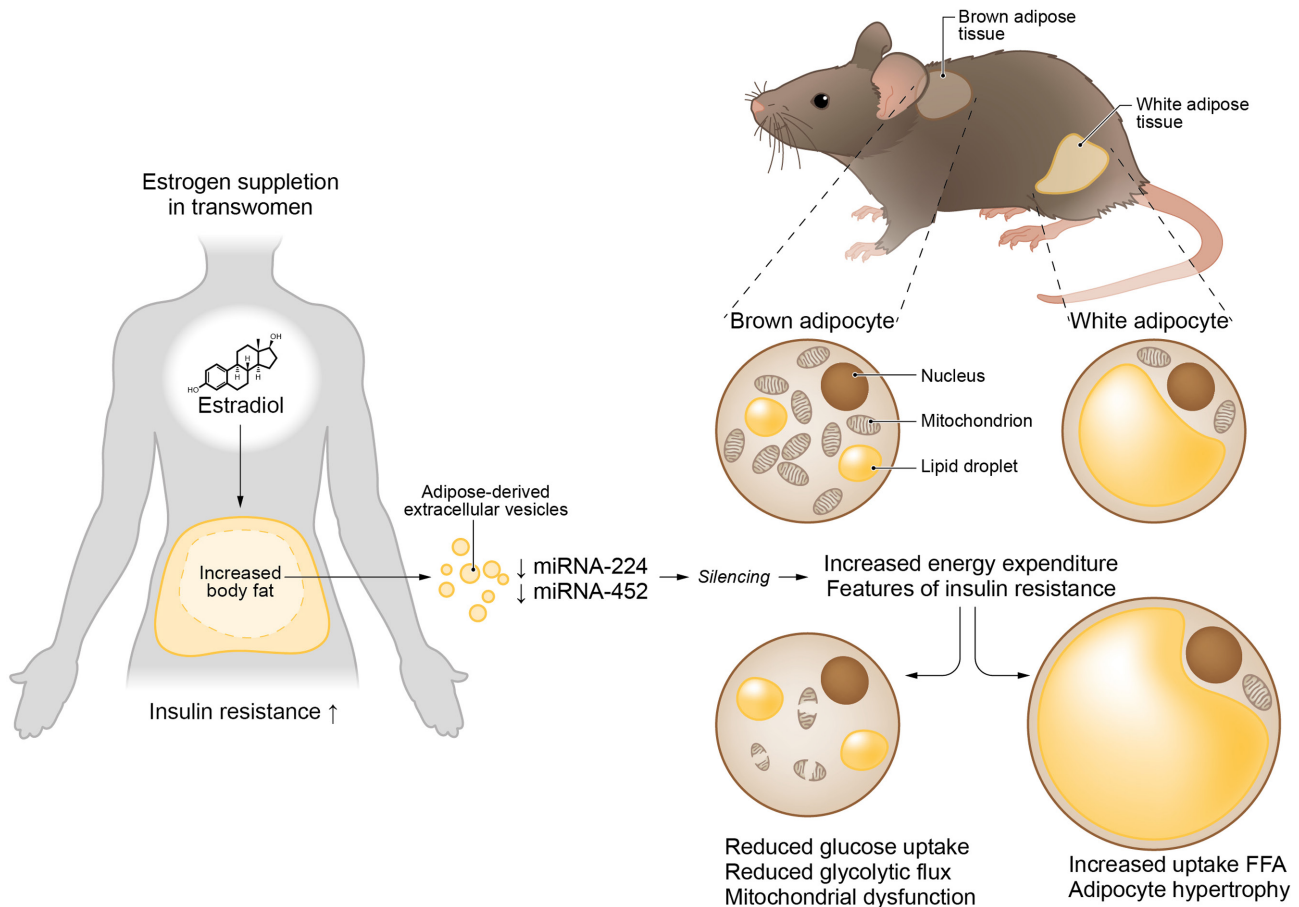


Figure 4

Proposed mechanism by which miR-224 and miR-452 affect metabolism in transwomen. Tightly balanced glucose uptake in brown adipose and lipid uptake in white adipose in lean metabolism is disrupted upon supraphysiological levels of estrogen that lower miR-224 and miR-452 in extracellular vesicles. The loss of both miRs lowers mitochondrial respiration and glucose uptake in brown adipose and results in more triglyceride-derived fatty acid uptake in white adipose and skeletal muscle.

mediated effects on glucose uptake and mitochondrial energy metabolism. Interestingly, when we performed IPA analyses of the differential proteome of omental adipose tissue from women with gestational diabetes mellitus (GDM) compared to adipose tissue from control subjects (48) we found that mitochondrial dysfunction topped the list of differentially expressed pathways (data not shown) further supporting the relationship between high levels of estradiol and metabolic changes. Lastly, body composition may affect metabolic health and as such may alter miR plasma levels as well (49), although BMI was not significantly affected in transwomen in this study (Tables 1 and 2). Still, future studies should investigate miR-224/452 in relation to body composition changes over time.

We conducted miR-224 and -452 silencing in male mice, particularly because we identified both miRs in male-female transgenders. It could be argued that testosterone

signaling in these mice may have confounded the *in vivo* data, while female mice, in which estrogen signaling is intact, would be more appropriate for direct testing of our hypothesis. However, it has been demonstrated that particularly female rats were previously found to become obese and showed impaired systemic insulin sensitivity following mitochondrial dysfunction in BAT, suggesting the potential of sex-specific miR effects in BAT tissue (50). Our observations in mice regarding increased energy expenditure and increased fatty acid uptake could also indicate beneficial effects to prevent lipotoxicity. Although we do not have a direct explanation for the elevated energy expenditure, we suspect that the onset of insulin resistance resulted in a shift in nutrient partitioning (change from glucose toward lipid consumption) and possibly futile cycling of those nutrients (energy expenditure due to a repeated elimination and composition of triglycerides).

Furthermore, the reduction of thermogenic genes in BAT and reduced BAT glucose uptake were shown to impact metabolic health negatively. Still, additional experiments are needed to confirm this miR-mediated phenotype, for example, by doing glucose tolerance testing and assessment of whether glucose uptake is indeed due to insulin resistance or due to the observed reduced expression of *Glut4*. The fact that miR-224 is known to control FA metabolism of 3T3-L1 adipocytes (51), prevents 3T3-L1 apoptosis upon inflammation (52) and controls low-density lipoprotein (LDL) metabolism (via its target PCSK9) (53) is consistent with the notion that miRs coordinately regulate functionally related genes in similar processes (54). A striking example in which the miR-224/452 cluster simultaneously controls cellular metabolism was found in malignant melanomas, in which both miRs targeted thioredoxin interacting protein (TXNIP), a key transcription factor involved in redox regulation and tumor suppression (55). However, more studies with (synergistic) silencing of both miRs are needed to investigate their regulation of (adipocyte) metabolism in mice.

Supplementary materials

This is linked to the online version of the paper at <https://doi.org/10.1530/EJ-21-0267>.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of this study.

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Author contributions statement

B W F conducted experiments, acquired and analyzed data, and wrote the manuscript. M K, E N K, R W A L L, and S K acquired and analyzed data. J D, J L, G D T, A B, Y Y, and W S researched the data. R N, T J R, P C N R, A J V Z, and R B contributed to the discussion and reviewed and edited the manuscript. B W F, A J V Z, and R B are guarantors of this work and, as such, had full access to all the data in the study and responsibility for the integrity of the data and the accuracy of the data analyses. J M G J D and M K contributed equally. R B and A J v Z shared senior authorship.

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References

- Morselli E, Santos RS, Criollo A, Nelson MD, Palmer BF & Clegg DJ. The effects of oestrogens and their receptors on cardiometabolic health. *Nature Reviews: Endocrinology* 2017 **13** 352–364. (<https://doi.org/10.1038/nrendo.2017.12>)
- Zhou Z, Moore TM, Drew BG, Ribas V, Wanagat J, Civelek M, Segawa M, Wolf DM, Norheim F, Seldin MM *et al*. Estrogen receptor α controls metabolism in white and brown adipocytes by regulating *polg1* and mitochondrial remodeling. *Science Translational Medicine* 2020 **12** eaax8096. (<https://doi.org/10.1126/scitranslmed.aax8096>)
- Klaver M, de Blok CJM, Wiepjes CM, Nota NM, Dekker MJHJ, de Mutser R, Schreiner T, Fisher AD, T'Sjoen G & den Heijer M. Changes in regional body fat, lean body mass and body shape in trans persons using cross-sex hormonal therapy: results from a multicenter prospective study. *European Journal of Endocrinology* 2018 **178** 163–171. (<https://doi.org/10.1530/EJE-17-0496>)
- Shadid S, Abosi-Appadu K, De Maertelaere AS, Defreyne J, Veldeman L, Holst JJ, Lapauw B, Vilsboll T & T'Sjoen G. Effects of gender-affirming hormone therapy on insulin sensitivity and incretin responses in transgender people. *Diabetes Care* 2020 **43** 411–417. (<https://doi.org/10.2337/dc19-1061>)
- Auer MK, Ebert T, Pietzner M, Defreyne J, Fuss J, Stalla GK & T'Sjoen G. Effects of sex hormone treatment on the metabolic syndrome in transgender individuals: focus on metabolic cytokines. *Journal of Clinical Endocrinology and Metabolism* 2018 **103** 790–802. (<https://doi.org/10.1210/jc.2017-01559>)
- Wierckx K, Elaut E, Declercq E, Heylens G, De Cuypere G, Taes Y, Kaufman JM & T'Sjoen G. Prevalence of cardiovascular disease and cancer during cross-sex hormone therapy in a large cohort of trans persons: a case-control study. *European Journal of Endocrinology* 2013 **169** 471–478. (<https://doi.org/10.1530/EJE-13-0493>)
- Herranz H & Cohen SM. MicroRNAs and gene regulatory networks: managing the impact of noise in biological systems. *Genes and Development* 2010 **24** 1339–1344. (<https://doi.org/10.1101/gad.1937010>)
- Arner P & Kulyte A. MicroRNA regulatory networks in human adipose tissue and obesity. *Nature Reviews: Endocrinology* 2015 **11** 276–288. (<https://doi.org/10.1038/nrendo.2015.25>)
- Florijn BW, Bijkerk R, van der Veer EP & van Zonneveld AJ. Gender and cardiovascular disease: are sex-biased microRNA networks a driving force behind heart failure with preserved ejection fraction in women? *Cardiovascular Research* 2018 **114** 210–225. (<https://doi.org/10.1093/cvr/cvx223>)
- Kim JH, Cho HT & Kim YJ. The role of estrogen in adipose tissue metabolism: insights into glucose homeostasis regulation. *Endocrine Journal* 2014 **61** 1055–1067. (<https://doi.org/10.1507/endocr.je14-0262>)
- Shi Z, Zhao C, Guo X, Ding H, Cui Y, Shen R & Liu J. Differential expression of microRNAs in omental adipose tissue from gestational diabetes mellitus subjects reveals miR-222 as a regulator of ER α expression in estrogen-induced insulin resistance. *Endocrinology* 2014 **155** 1982–1990. (<https://doi.org/10.1210/en.2013-2046>)
- Thomou T, Mori MA, Dreyfuss JM, Konishi M, Sakaguchi M, Wolfrum C, Rao TN, Winnay JN, Garcia-Martin R, Grinspoon SK *et al*. Adipose-derived circulating miRNAs regulate gene expression in other tissues. *Nature* 2017 **542** 450–455. (<https://doi.org/10.1038/nature21365>)
- van Velzen DM, Paldino A, Klaver M, Nota NM, Defreyne J, Hovingh GK, Thijs A, Simsek S, T'Sjoen G & den Heijer M. Cardiometabolic effects of testosterone in transmen and estrogen plus cyproterone acetate in transwomen. *Journal of Clinical Endocrinology and Metabolism* 2019 **104** 1937–1947. (<https://doi.org/10.1210/jc.2018-02138>)
- Florijn BW, Duijs JMGJ, Levels JH, Dallinga-Thie GM, Wang Y, Boing AN, Yuana Y, Stam W, Limpens RWAL, Au YW *et al*. Diabetic nephropathy alters the distribution of circulating angiogenic microRNAs Among extracellular vesicles, HDL, and Ago-2. *Diabetes* 2019 **68** 2287–2300. (<https://doi.org/10.2337/db18-1360>)

- 15 Rensen PC, van Dijk MC, Havenaar EC, Bijsterbosch MK, Kruijt JK & van Berkel TJ. Selective liver targeting of antivirals by recombinant chylomicrons: a new therapeutic approach to hepatitis B. *Nature Medicine* 1995 **1** 221–225. (<https://doi.org/10.1038/nm0395-221>)
- 16 Liu J, Kuipers EN, Sips HCM, Dorleijn JC, van Dam AD, Christodoulides C, Karpe F, Zhou G, Boon MR, Rensen PCN *et al*. Conditionally immortalized brown preadipocytes can switch between proliferative and differentiated states. *Biochimica et Biophysica Acta: Molecular and Cell Biology of Lipids* 2019 **1864** 158511. (<https://doi.org/10.1016/j.bbalip.2019.08.007>)
- 17 Dekker MJ, Wierckx K, Van Caenegem E, Klaver M, Kreukels BP, Elaut E, Fisher AD, van Trotsenburg MA, Schreiner T, den Heijer M *et al*. A European network for the investigation of gender incongruence: endocrine part. *Journal of Sexual Medicine* 2016 **13** 994–999. (<https://doi.org/10.1016/j.jsxm.2016.03.371>)
- 18 Ludwig N, Leidinger P, Becker K, Backes C, Fehlmann T, Pallasch C, Rheinheimer S, Meder B, Stahler C, Meese E *et al*. Distribution of miRNA expression across human tissues. *Nucleic Acids Research* 2016 **44** 3865–3877. (<https://doi.org/10.1093/nar/gkw116>)
- 19 Winther S, Isidor MS, Basse AL, Skjoldborg N, Cheung A, Quistorff B & Hansen JB. Restricting glycolysis impairs brown adipocyte glucose and oxygen consumption. *American Journal of Physiology: Endocrinology and Metabolism* 2018 **314** E214–E223. (<https://doi.org/10.1152/ajpendo.00218.2017>)
- 20 Henegar C, Tordjman J, Achard V, Lacasa D, Cremer I, Guerre-Millo M, Poitou C, Basdevant A, Stich V, Viguier N *et al*. Adipose tissue transcriptomic signature highlights the pathological relevance of extracellular matrix in human obesity. *Genome Biology* 2008 **9** R14. (<https://doi.org/10.1186/gb-2008-9-1-r14>)
- 21 Cutchins A, Harmon DB, Kirby JL, Doran AC, Oldham SN, Skafien M, Klibanov AL, Meller N, Keller SR, Garmey J *et al*. Inhibitor of differentiation-3 mediates high fat diet-induced visceral fat expansion. *Arteriosclerosis, Thrombosis, and Vascular Biology* 2012 **32** 317–324. (<https://doi.org/10.1161/ATVBAHA.111.234856>)
- 22 Kanzleiter T, Schneider T, Walter I, Bolze F, Eickhorst C, Heldmaier G, Klaus S & Klingenspor M. Evidence for Nr4a1 as a cold-induced effector of brown fat thermogenesis. *Physiological Genomics* 2005 **24** 37–44. (<https://doi.org/10.1152/physiolgenomics.00204.2005>)
- 23 Liu Y, Zhou D, Abumrad NA & Su X. ADP-ribosylation factor 6 modulates adrenergic stimulated lipolysis in adipocytes. *American Journal of Physiology: Cell Physiology* 2010 **298** C921–C928. (<https://doi.org/10.1152/ajpcell.00541.2009>)
- 24 Davies JC, Bain SC & Kanamarlapudi V. ADP-ribosylation factor 6 regulates endothelin-1-induced lipolysis in adipocytes. *Biochemical Pharmacology* 2014 **90** 406–413. (<https://doi.org/10.1016/j.bcp.2014.06.012>)
- 25 Moest H, Frei AP, Bhattacharya I, Geiger M, Wollscheid B & Wolfrum C. Malfunctioning of adipocytes in obesity is linked to quantitative surfaceome changes. *Biochimica et Biophysica Acta* 2013 **1831** 1208–1216. (<https://doi.org/10.1016/j.bbalip.2013.04.001>)
- 26 Mejhert N, Wilfling F, Esteve D, Galitzky J, Pellegrinelli V, Kolditz CI, Viguier N, Tordjman J, Naslund E, Trayhurn P *et al*. Semaphorin 3C is a novel adipokine linked to extracellular matrix composition. *Diabetologia* 2013 **56** 1792–1801. (<https://doi.org/10.1007/s00125-013-2931-z>)
- 27 De Luca M, Klimentidis YC, Casazza K, Chambers MM, Cho R, Harbison ST, Jumbo-Lucioni P, Zhang S, Leips J & Fernandez JR. A conserved role for syndecan family members in the regulation of whole-body energy metabolism. *PLoS ONE* 2010 **5** e11286. (<https://doi.org/10.1371/journal.pone.0011286>)
- 28 Zhou D, Shen R, Ye JJ, Li Y, Tsark W, Isbell D, Tso P & Chen S. Nuclear receptor coactivator PNRC2 regulates energy expenditure and adiposity. *Journal of Biological Chemistry* 2008 **283** 541–553. (<https://doi.org/10.1074/jbc.M703234200>)
- 29 Bazuine M, Stenkula KG, Cam M, Arroyo M & Cushman SW. Guardian of corpulence: a hypothesis on p53 signaling in the fat cell. *Clinical Lipidology* 2009 **4** 231–243. (<https://doi.org/10.2217/clp.09.2>)
- 30 White UA, Stewart WC, Mynatt RL & Stephens JM. Neurotrophin attenuates adipogenesis and induces insulin resistance in adipocytes. *Journal of Biological Chemistry* 2008 **283** 22505–22512. (<https://doi.org/10.1074/jbc.M710462200>)
- 31 Pramfalk C, Eriksson M & Parini P. Role of TG-interacting factor (Tgif) in lipid metabolism. *Biochimica et Biophysica Acta* 2015 **1851** 9–12. (<https://doi.org/10.1016/j.bbalip.2014.07.019>)
- 32 Dib L, Bugge A & Collins S. LXRalpha fuels fatty acid-stimulated oxygen consumption in white adipocytes. *Journal of Lipid Research* 2014 **55** 247–257. (<https://doi.org/10.1194/jlr.M043422>)
- 33 Kleiner S, Mepani RJ, Laznik D, Ye L, Jurczak MJ, Jornayvay FR, Estall JL, Chatterjee Bhowmick D, Shulman GI & Spiegelman BM. Development of insulin resistance in mice lacking PGC-1alpha in adipose tissues. *PNAS* 2012 **109** 9635–9640. (<https://doi.org/10.1073/pnas.1207287109>)
- 34 Olsen JM, Sato M, Dallner OS, Sandstrom AL, Pisani DE, Chambard JC, Amri EZ, Hutchinson DS & Bengtsson T. Glucose uptake in brown fat cells is dependent on mTOR complex 2-promoted GLUT1 translocation. *Journal of Cell Biology* 2014 **207** 365–374. (<https://doi.org/10.1083/jcb.201403080>)
- 35 Fasshauer M, Klein J, Ueki K, Kriaucianus KM, Benito M, White MF & Kahn CR. Essential role of insulin receptor substrate-2 in insulin stimulation of glut4 translocation and glucose uptake in brown adipocytes. *Journal of Biological Chemistry* 2000 **275** 25494–25501. (<https://doi.org/10.1074/jbc.M004046200>)
- 36 Puri V, Ranjit S, Konda S, Nicoloso SM, Straubhaar J, Chawla A, Chouinard M, Lin C, Burkart A, Corvera S *et al*. Cidea is associated with lipid droplets and insulin sensitivity in humans. *PNAS* 2008 **105** 7833–7838. (<https://doi.org/10.1073/pnas.0802063105>)
- 37 Ellis JM, Li LO, Wu PC, Koves TR, Ilkayeva O, Stevens RD, Watkins SM, Muoio DM & Coleman RA. Adipose acyl-CoA synthetase-1 directs fatty acids toward beta-oxidation and is required for cold thermogenesis. *Cell Metabolism* 2010 **12** 53–64. (<https://doi.org/10.1016/j.cmet.2010.05.012>)
- 38 Eissing L, Scherer T, Todter K, Knippschild U, Greve JW, Buurman WA, Pinnuschmidt HO, Rensen SS, Wolf AM, Bartelt A *et al*. De novo lipogenesis in human fat and liver is linked to ChREBP-beta and metabolic health. *Nature Communications* 2013 **4** 1528. (<https://doi.org/10.1038/ncomms2537>)
- 39 Osuga J, Ishibashi S, Oka T, Yagyu H, Tozawa R, Fujimoto A, Shionoiri F, Yahagi N, Kraemer FB, Tsutsumi O *et al*. Targeted disruption of hormone-sensitive lipase results in male sterility and adipocyte hypertrophy, but not in obesity. *PNAS* 2000 **97** 787–792. (<https://doi.org/10.1073/pnas.97.2.787>)
- 40 Harada N, Oda Z, Hara Y, Fujinami K, Okawa M, Ohbuchi K, Yonemoto M, Ikeda Y, Ohwaki K, Aragane K *et al*. Hepatic de novo lipogenesis is present in liver-specific ACC1-deficient mice. *Molecular and Cellular Biology* 2007 **27** 1881–1888. (<https://doi.org/10.1128/MCB.01122-06>)
- 41 Zhou X, Wang X, Huang Z, Xu L, Zhu W & Liu P. An ER-associated miRNA signature predicts prognosis in ER-positive breast cancer. *Journal of Experimental and Clinical Cancer Research* 2014 **33** 94. (<https://doi.org/10.1186/s13046-014-0094-5>)
- 42 Schmeier S, MacPherson CR, Essack M, Kaur M, Schaefer U, Suzuki H, Hayashizaki Y & Bajic VB. Deciphering the transcriptional circuitry of microRNA genes expressed during human monocytic differentiation. *BMC Genomics* 2009 **10** 595. (<https://doi.org/10.1186/1471-2164-10-595>)
- 43 Florijn BW, Valstar GB, Duijs J, Menken R, Cramer MJ, Teske AJ, Ghossein-Doha C, Rutten FH, Spaanderman MEA, den Ruijter HM *et al*. Sex-specific microRNAs in women with diabetes and left ventricular diastolic dysfunction or HFpEF associate with microvascular injury. *Scientific Reports* 2020 **10** 13945. (<https://doi.org/10.1038/s41598-020-70848-8>)
- 44 Kristensen H, Haldrup C, Strand S, Mundbjerg K, Mortensen MM, Thorsen K, Ostenfeld MS, Wild PJ, Arsov C, Goering W *et al*. Hypermethylation of the GABRE-miR-452-miR-224 promoter in prostate cancer predicts biochemical recurrence after radical prostatectomy. *Clinical Cancer Research* 2014 **20** 2169–2181. (<https://doi.org/10.1158/1078-0432.CCR-13-2642>)

- 45 Zarotsky V, Huang MY, Carman W, Morgentaler A, Singhal PK, Coffin D & Jones TH. Systematic literature review of the risk factors, comorbidities, and consequences of hypogonadism in men. *Andrology* 2014 **2** 819–834. (<https://doi.org/10.1111/andr.274>)
- 46 Gava G, Mancini I, Alvisi S, Seracchioli R & Meriggiola MC. A comparison of 5-year administration of cyproterone acetate or leuprolide acetate in combination with estradiol in transwomen. *European Journal of Endocrinology* 2020 **183** 561–569. (<https://doi.org/10.1530/EJE-20-0370>)
- 47 Mauvais-Jarvis F, Clegg DJ & Hevener AL. The role of estrogens in control of energy balance and glucose homeostasis. *Endocrine Reviews* 2013 **34** 309–338. (<https://doi.org/10.1210/er.2012-1055>)
- 48 Ma Y, Gao J, Yin J, Gu L, Liu X, Chen S, Huang Q, Lu H, Yang Y, Zhou H *et al.* Identification of a novel function of adipocyte plasma membrane-associated protein (APMAP) in gestational diabetes mellitus by proteomic analysis of omental adipose tissue. *Journal of Proteome Research* 2016 **15** 628–637. (<https://doi.org/10.1021/acs.jproteome.5b01030>)
- 49 Assmann TS, Riezu-Boj JI, Milagro FI & Martínez JA. Circulating adiposity-related microRNAs as predictors of the response to a low-fat diet in subjects with obesity. *Journal of Cellular and Molecular Medicine* 2020 **24** 2956–2967. (<https://doi.org/10.1111/jcmm.14920>)
- 50 Nadal-Casellas A, Bauza-Thorbrugge M, Proenza AM, Gianotti M & Llado I. Sex-dependent differences in rat brown adipose tissue mitochondrial biogenesis and insulin signaling parameters in response to an obesogenic diet. *Molecular and Cellular Biochemistry* 2013 **373** 125–135. (<https://doi.org/10.1007/s11010-012-1481-x>)
- 51 Peng Y, Xiang H, Chen C, Zheng R, Chai J, Peng J & Jiang S. MiR-224 impairs adipocyte early differentiation and regulates fatty acid metabolism. *International Journal of Biochemistry and Cell Biology* 2013 **45** 1585–1593. (<https://doi.org/10.1016/j.biocel.2013.04.029>)
- 52 Qi R, Huang J, Wang Q, Liu H, Wang R, Wang J & Yang F. MicroRNA-224-5p regulates adipocyte apoptosis induced by TNFalpha via controlling NF-kappaB activation. *Journal of Cellular Physiology* 2018 **233** 1236–1246. (<https://doi.org/10.1002/jcp.25992>)
- 53 Naeli P, Mirzadeh Azad F, Malakootian M, Seidah NG & Mowla SJ. Post-transcriptional regulation of PCSK9 by miR-191, miR-222, and miR-224. *Frontiers in Genetics* 2017 **8** 189. (<https://doi.org/10.3389/fgene.2017.00189>)
- 54 Tsang JS, Ebert MS & van Oudenaarden A. Genome-wide dissection of microRNA functions and cotargeting networks using gene set signatures. *Molecular Cell* 2010 **38** 140–153. (<https://doi.org/10.1016/j.molcel.2010.03.007>)
- 55 Knoll S, Furst K, Kowtharapu B, Schmitz U, Marquardt S, Wolkenhauer O, Martin H & Putzer BM. E2F1 induces miR-224/452 expression to drive EMT through TXNIP downregulation. *EMBO Reports* 2014 **15** 1315–1329. (<https://doi.org/10.15252/embr.201439392>)

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