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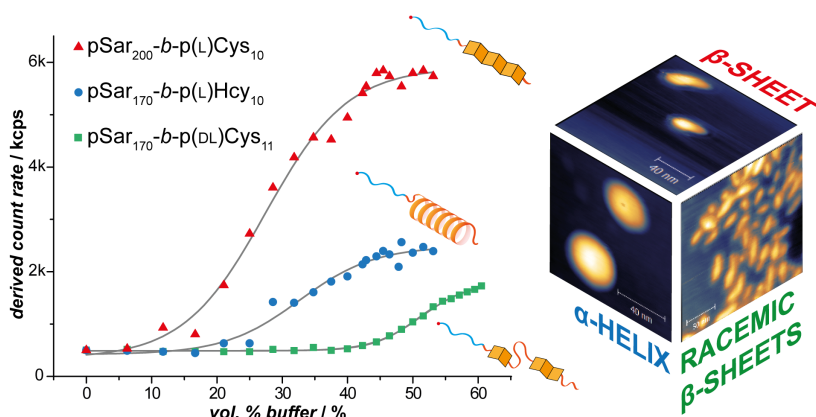
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Secondary Structure-Driven Self-Assembly of Thiol-Reactive Polypept(o)ides



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Secondary Structure-Driven Self-Assembly of Thiol-Reactive Polypept(o)ides

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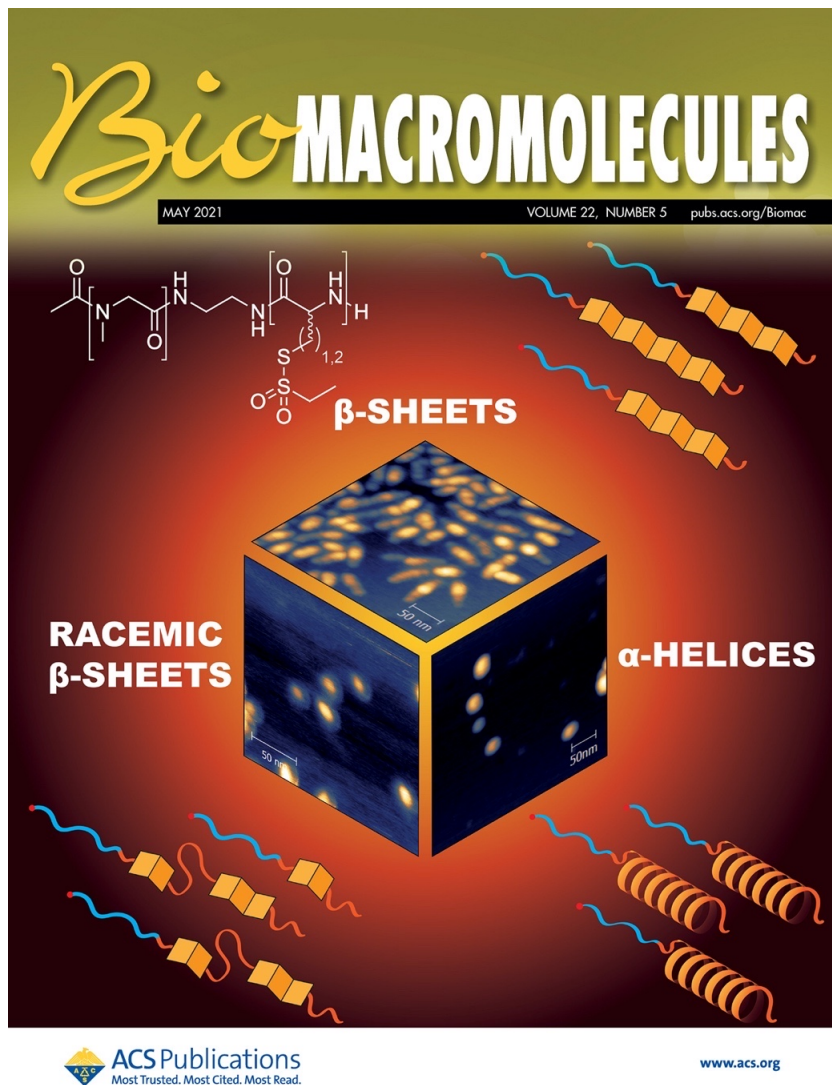
Abstract

Secondary structure formation differentiates polypeptides from most other synthetic polymers, and the transitions from random coil to rod-like α -helices or β -sheets represent an additional parameter to direct self-assembly and the morphology of nanostructures. We investigated the influence of distinct secondary structures on the self-assembly of reactive amphiphilic polypept(o)ides. The individual morphologies can be preserved by core cross-linking *via* chemoselective disulfide bond formation. A series of thiol-responsive copolymers of racemic polysarcosine-*block*-poly(*S*-ethylsulfonyl-DL-cysteine) (pSar-*b*-p(DL)Cys), enantiopure polysarcosine-*block*-poly(*S*-ethylsulfonyl-L-cysteine) (pSar-*b*-p(L)Cys), and polysarcosine-*block*-poly(*S*-ethylsulfonyl-L-homocysteine) (pSar-*b*-p(L)Hcy) was prepared by *N*-carboxyanhydride polymerization. The secondary structure of the peptide segment varies from α -helix (pSar-*b*-p(L)Hcy) to anti-parallel β -sheet (pSar-*b*-p(L)Cys), and disrupted β -sheet (pSar-*b*-p(DL)Cys). When subjected to nanoprecipitation, copolymers with anti-parallel β -sheets display the strongest tendency to self-assemble, whereas disrupted β -sheets hardly induce aggregation. This translates to worm-like micelles, solely spherical micelles or ellipsoidal structures, as analyzed by AFM and cryoTEM, which underlines the potential of secondary structure-driven self-assembly of synthetic polypeptides.

Keywords

secondary structure • polypept(o)ides • self-assembly • polymer • micelle • reactive polymer

Cover page



The secondary structure formation of reactive polypept(o)ides governs self-assembly, featuring control over the morphology of core cross-linked polymeric micelles.

Introduction

Secondary structure formation as a feature of polypeptides is the fundamental basis for the higher ordered structural diversity and specific function of proteins and other biomaterials.^{1–3} As a prominent example, the polyalanine-domains of spider silk undergo a transition from α -helix in solution to highly ordered β -sheets in the solid state, which, among other factors, accounts for feasible processing paired with exemptional elasticity and toughness.^{4–7} Consequently, not only from a polymer chemistry perspective, secondary structure displays an additional parameter influencing the preparation and application of peptide materials.^{8–11} For polypeptides derived from single amino acids, primarily α -helix or β -sheet are obtained, with exceptions for proline-type amino acids.^{1,12} Upon secondary structure formation, the induced coil-to-helix or coil-to-sheet transitions refer to a rigidification of the polymer segment (rod-like structures) and provide a driving-force for self-assembly.^{1,13} In particular for synthetic polypeptides of polyglutamic acid and polylysine, the stimuli which induce α -helix formation, such as pH or temperature, have been the subject of multiple investigations.^{14–20} Furthermore, Deming and co-workers intensively elucidated the thermo- and redox-responsiveness of α -helical polyhomocysteine derivatives.^{21,22} On the other hand, for polyserine or polycysteine, research has been more dedicated to resolving the strong induction of β -sheet formation, e.g., by side chain functionalization with oligo ethylene glycol moieties.^{23–25}

In contrast to polypeptides, polypeptoids, with the branching on the nitrogen atom instead of the α -position do not show classical secondary structure formation as no hydrogen bond donor is present if no specific modification was introduced.^{26–28} As such, polysarcosine (pSar, poly(*N*-methyl glycine)) adopts random coil conformation in aqueous solution, is highly water-soluble (A2 parameter of $3.50 \cdot 10^{-6} \text{ mol} \cdot \text{dm}^3 \cdot \text{g}^{-2}$ for pSar₁₈₅ at 20 °C), and can be considered a stealth-like material.^{29–33} For medical applications, polysarcosine is considered the most promising alternative to polyethylene glycol since substantial advantages like reduced proinflammatory cytokine secretion and reduced complement activation as well as evasion of the accelerated blood clearance (ABC) phenomenon have been reported.^{30,33–35} The combination of polypeptides and polysarcosine in polypept(o)ides thus represent a promising class of bio-based or even endogenous materials.^{26,36,37} While the concept of polypept(o)ides extents from linear block copolymers to cylindrical bottle-brush and star-shaped polymers including

miktoarm stars,^{36,38–40} polysarcosine itself has further been successfully combined with various other classes of synthetic polymers.^{37,41,42}

With its ability to form bio-reversible disulfide bonds, the non-essential amino acid cysteine plays a unique role within nature.^{43,44} In living cells, catalyzing proteins and small molecules orchestrate disulfide bond formation among specific cysteines generating functional proteins.⁴⁵ To transfer chemoselective disulfide bond formation to functional polymers, our group reported on the reactive *S*-alkylsulfonyl protecting group for cysteine and homocysteine.^{46,47} This reactive group differentiates hard and soft nucleophiles, and grants living amine-initiated ring-opening polymerization of *S*-alkylsulfonyl-protected *N*-carboxyanhydrides (NCAs). The *S*-alkylsulfonyl group can then be addressed by post polymerization modification reactions with soft nucleophiles, e.g., thiols, for disulfide bond formation. With respect to secondary structure, homopolypeptides of *S*-ethylsulfonyl-L-cysteine (p(L)Cys) were shown to adopt anti-parallel β -sheets, while β -sheets were even detected for the racemic mixture, poly(*S*-ethylsulfonyl-DL-cysteine) (p(DL)Cys), albeit to a much lesser extent.^{47,48} In contrast, poly(*S*-ethylsulfonyl-L-homocysteine) (p(L)Hcy), which has only one additional methylene group in the side chain as compared to the parent cysteine, was shown to adopt an α -helical structure in solution and solid state.⁴⁹

Since anti-parallel β -sheets of enantiopure polysarcosine-*block*-poly(*S*-ethylsulfonyl-L-cysteine) (pSar-*b*-p(L)Cys) were shown to direct self-assembly to worm-like micelles previously,⁵⁰ we now contribute a comprehensive study on the influence of the secondary structure on the self-assembly of reactive block copolypept(o)ides. Therefore, *S*-ethylsulfonyl-protected reactive copolymers with varying secondary structure (α -helix, anti-parallel β -sheet and a disrupted β -sheet) have been synthesized by ring-opening NCA polymerization and their aggregation behavior was analyzed by DLS, AFM and cryoTEM.

Materials and Methods

Unless stated otherwise, reagents and solvents were purchased from Sigma-Aldrich and used as received. *N,N*-Dimethyl formamide (DMF) was bought from Acros (99.8%, extra dry over molecular sieve) and purified by repetitive freeze-pump thaw cycles prior to use (water content < 50 ppm). *N*-*tert*-butoxycarbonyl (Boc)-1,2-diaminoethane was obtained from Sigma-Aldrich ($\geq 98\%$) and stored over activated molecular sieves before further use. MilliQ water was prepared using a MILLI-Q® Reference A+ System. Water was used at a resistivity of 18.2

$\text{M}\Omega\text{ cm}^{-1}$ and total organic carbon (TOC) of <5 ppm. Hexafluoro isopropanol (HFIP) and potassium trifluoro acetate was purchased from Fluorochem. Deuterated solvents were obtained from Deutero GmbH and were used as received.

^1H NMR spectra were recorded on a Bruker Avance II 400 at room temperature at a frequency of 400 MHz. DOSY spectra were recorded on a Bruker Avance III HD 400 (400 MHz). Calibration of the spectra was achieved using the solvent signals. NMR spectra were analyzed with MestReNova version 12.0.4 from Mestrelab Research S.L. Degrees of polymerization (X_n) by ^1H NMR were calculated comparing the integral of the initiator peak and the integrals of the α -protons. For block copolymers containing polysarcosine, the chain lengths of the second block were calculated from the integral of the α -protons and the signals of the polysarcosine backbone, as derived from HFIP-GPC relative to polysarcosine standards.

Attenuated total reflectance Fourier-transformed infrared (ATR-FT-IR) spectroscopy was performed on a FT/IR-4100 (JASCO Corporation) with an ATR sampling accessory (MIRacle™, Pike Technologies). IR spectra were analyzed using Spectra Manager 2.0 (JASCO Corporation). NCA polymerization was monitored by FT-IR spectroscopy and the polymerization was judged to be completed if the carbonyl peaks at 1858 and 1788 cm^{-1} had vanished.

Analytical gel permeation chromatography (GPC) was performed on a Jasco GPC setup at a flow rate of 0.8 ml min^{-1} and a temperature of $40\text{ }^\circ\text{C}$. The eluent was HFIP equipped with 3 g L^{-1} potassium trifluoroacetate. The column material was modified silica gel (PFG columns, particle size: $7\text{ }\mu\text{m}$, porosity: $100\text{ }\text{\AA}$ and $4000\text{ }\text{\AA}$), purchased from PSS Polymer Standards Service GmbH. For polymer detection, a UV detector (Jasco UV-2075+) at a wavelength of $\lambda = 230\text{ nm}$ was employed. Molecular weights were determined by using a calibration with PMMA (PSS Polymer Standards Services GmbH) with toluene as internal standard. The elution diagram was evaluated with PSS WinGPC (PSS Polymer Standard Service GmbH).

Single-angle dynamic light scattering (DLS) measurements were performed with a ZetaSizer Nano ZS (Malvern Instruments Ltd., Worcestershire, UK) equipped with a He-Ne laser ($\lambda = 632.8\text{ nm}$). All measurements were performed at $25\text{ }^\circ\text{C}$ and a detection angle of 173° . Disposable polystyrene and organic solvent-resistant cuvettes (VWR, Darmstadt, Germany) were used for single-angle DLS

measurements. Cumulant size, polydispersity index (PDI), size distribution (intensity weighted) histograms, and the derived count rate were calculated based on the autocorrelation function of the samples, with automated position and attenuator adjustment at multiple scans.

Circular dichroism (CD) spectroscopy was performed on a Jasco J-815 spectrometer at a temperature of 20°C using Spectra Manager 1.53.04 (Jasco). The spectra were recorded in HFIP at a concentration of 0.1 g · L⁻¹ using a quartz cell with a path length of 1 mm. θ_{MR} was calculated with the equation below with $c_M = 0.1$ g · L⁻¹, $l = 0.1$ cm, and $M_{\text{Repeating Unit}} = 195.26$ g · mol⁻¹ for *S*-ethylsulfonyl-L-cysteine and *S*-ethylsulfonyl-DL-cysteine, $M_{\text{Repeating Unit}} = 209.26$ g · mol⁻¹ for *S*-ethylsulfonyl-L-homocysteine and $M_{\text{Repeating Unit}} = 71.08$ g · mol⁻¹ for sarcosine.

$$\theta_{MR} = \frac{\theta \cdot M_{\text{Repeating Unit}}}{10 \cdot c_M \cdot l} [\text{deg} \cdot \text{cm}^2 \cdot \text{mol}^{-1}]$$

Atomic force microscopy (AFM) was measured on mica using a Cypher™ AFM (Asylum Research) using tapping mode at a scan rate of 1 Hz. Samples were prepared by drop-casting of a particle solution ($V = 5$ μL, $\beta = 50$ mg · L⁻¹ in MilliQ water) onto freshly cleaned mica. The sample was dried overnight at room temperature. Images were evaluated using Gwyddion 2.49.

For cryogenic transmission electron microscopy (CryoTEM), 5 μL of CCPM solution (50 mg · L⁻¹, in MilliQ water) were applied to freshly glow-discharged carbon grids with a copper 200 mesh (Quantifoil Micro Tools GmbH). Excess fluid was removed by direct blotting (2.5 s) and the grids were individually plunge-frozen in liquid ethane. Grids were cryotransferred in liquid nitrogen using a Gatan cryoholder (model 626 DH) to a Technai T12 transmission electron microscope equipped with a field emission electron source and operating at 120 kV accelerating voltage. Images were recorded using a TemCam-F416 (TVIPS, Gauting, Germany). Software imageJ 1.52h (National Institutes of Health, USA) was used for image evaluation.

Polymer Synthesis. Polymers were prepared by amine-initiated ring-opening *N*-carboxyanhydride (NCA) polymerization in dry DMF under Schlenk-conditions. The respective monomers, sarcosine-NCA, *S*-ethylsulfonyl-DL-cysteine-NCA, *S*-ethylsulfonyl-L-cysteine-NCA, and *S*-ethylsulfonyl-L-homocysteine-NCA were synthesized and characterized as reported previously.^{46,48,49}

Polysarcosine (pSar_n) (P1, P2) Sarcosine-NCA (5.98 g, 52.0 mmol, 200 eq.) was transferred into a pre-dried Schlenk-tube, dissolved in 60 mL dry DMF and *N*-

(*tert*-butoxycarbonyl)-1,2-diaminoethane (41.6 mg, 0.26 mmol, 1.0 eq.) was added as a stock solution in dry DMF. The clear, colorless solution was stirred at 10 °C in the absence of light until the reaction was completed after six days, as monitored by IR spectroscopy. The sarcosine amino terminus was quenched by addition perfluorophenyl 4-azidobutanoate (153 mg, 0.52 mmol, 2.0 eq.) and *N,N*-diisopropylethylamine (221 μ L, 1.30 mmol, 5.0 eq.). The reaction mixture was stirred overnight, followed by addition of acetic anhydride (248 μ L, 2.60 mmol, 10 eq.) and *N,N*-diisopropylethylamine (884 μ L, 5.20 mmol, 20 eq.) to react residual end groups. The slightly yellow solution was stirred for one additional day at room temperature before the polymer was precipitated in 600 mL diethyl ether, collected on a Buchner funnel, washed with acetone and diethyl ether and dried *in vacuo*. Next, the Boc-protecting group was removed. The polymer (3.4 g) was dissolved in 50 mL of water, cooled to 0 °C and 50 mL trifluoro acetic acid were added in one portion. After 4 h at 0 °C, the solution was transferred into dialysis bags (MWCO 3.5 kDa) and dialyzed against MilliQ water, saturated sodium hydrogen carbonate solution, and MilliQ water. The aqueous solution was lyophilized, and the polysarcosine (P1) was obtained as a colorless powder (3.08g, 82%). ^1H NMR (400 MHz, DMSO-*d*₆) δ (ppm) = 4.50-3.76 (m, 2nH, -CH₂-), 3.06-2.76 (m, 3nH, -CH₃). The chain length was determined by HFIP GPC relative to polysarcosine standards.²⁹

Block copolymer syntheses. All block copolymers were prepared from pSar macroinitiators, following the procedure described below for pSar-*b*-p(DL)Cys using the respective NCAs.⁵⁰

Polysarcosine-*n*-block-poly(*S*-ethylsulfonyl-DL-cysteine)_m (pSar-*n*-*b*-p(DL)Cys_m) (P3-P6)

The polysarcosine macroinitiator (111.9 mg, 9.2 μ mol, 1.0 eq.) was weighed into a pre-dried Schlenk-tube and dried by azeotropic distillation with toluene *in vacuo* twice. Next, the macroinitiator was dissolved in freshly degassed dry DMF (1.45 mL), cooled to -10 °C, and *S*-ethylsulfonyl-L-cysteine NCA (33.0 mg, 138 μ mol, 15 eq.) was added as a stock solution in dry DMF. The polymerization was performed at an overall mass concentration of $\beta = 100 \text{ g} \cdot \text{L}^{-1}$ and monitored by IR spectroscopy. After 5 days, full conversion was observed, and the polymer was precipitated in THF. The suspension was centrifuged (4500 rpm, 15 min, 4°C) and decanted. This procedure was repeated twice concluding with pure diethyl ether. The product was dried *in vacuo* yielding pSar-*n*-*b*-p(DL)Cys_m (P3) as a colorless solid (130 mg, 93%). ^1H NMR (400 MHz, DMSO-*d*₆) δ (ppm) = 9.04 - 8.59 (b s, 1mH, NHCO), 4.90 - 4.60 (m, 1m H, α -CH_(DL-Cys)), 4.49 - 3.78 (m, 2n H, -CH₂-(Sar)),

3.69 - 3.41 (m, 4m H, $-\text{CH}_2\text{S}-$, $-\text{SO}_2\text{CH}_2-$), 3.06 - 2.61 (m, 3n H, $-\text{CH}_3(\text{Sar})$), 1.36 - 1.21 (t, 3m H, $-\text{CH}_3(\text{DL-Cys})$).

Polysarcosine_n-block-poly(S-ethylsulfonyl-L-cysteine)_p (pSar_n-b-p(L)Cys_p) (P7-P10)
 ^1H NMR (400 MHz, DMSO-*d*₆) δ (ppm) = 8.75 (b s, 1p H, NHCO), 4.69 (m, 1p H, $\alpha\text{-CH}_{(\text{L-Cys})}$), 4.49 - 3.78 (m, 2nH, $-\text{CH}_2-(\text{Sar})$), 3.69 - 3.41 (m, 4p H, $-\text{CH}_2\text{S}-$, $-\text{SO}_2\text{CH}_2-$), 3.06 - 2.61 (m, 3p H, $-\text{CH}_3(\text{Sar})$), 1.30 (t, 3p H, $-\text{CH}_3(\text{L-Cys})$).

Polysarcosine_n-block-poly(S-ethylsulfonyl-L-homocysteine)_q (pSar_n-b-p(L)Hcy_q) (P11-P14)
 ^1H NMR (400 MHz, DMSO-*d*₆) δ (ppm) = 8.34 (b s, 1q H, NHCO), 4.56 - 3.75 (m, 2n H + 1q H, $-\text{CH}_2-(\text{Sar})$, $\alpha\text{-CH}_{(\text{L-Hcy})}$), 3.64 - 3.45 (m, 2q H, $-\text{SO}_2\text{CH}_2-$), 3.19 - 3.08 (m, 2q H, $-\text{CH}_2\text{S}-$), 3.06 - 2.61 (m, 3n H, $-\text{CH}_3(\text{Sar})$), 2.18 - 1.85 (m, 2q H, $-\text{CH}_2\text{-CH}_2\text{S}-$), 1.30 (t, 3q H, $-\text{CH}_3(\text{L-Hcy})$).

Cross-linker Synthesis. The synthesis of (*R*)-lipoic acid hydrazide (**3**) was performed in a two-step synthesis *via* (*R*)-methyl lipoate (**2**) starting from (*R*)-lipoic acid (**1**). The respective protocols were adapted and modified from Hassan and Maltman and Koufaki *et al.*^{51,52}

(*R*)-Lipoic acid (2.0 g, 9.69 mmol, 1.0 eq.) was dissolved in 20 mL of methanol, and a catalytic amount of sulfuric acid (5.2 μL , 0.10 mmol, 0.01 eq.) was added. The reaction mixture was stirred at room temperature for 18h and protected from light. The solvent was removed under reduced pressure and the crude solid was re-dissolved in dichloromethane. The organic phase was washed with saturated NaHCO_3 -solution and brine, dried over magnesium sulfate, filtered, and concentrated *in vacuo*. (*R*)-Methyl lipoate (**2**) was obtained as a yellow oil (1.90 g, 94%) and used without further purification. ^1H NMR (400 MHz, DMSO-*d*₆) δ (ppm) = 3.25 - 3.03 (s + m, 4H, $-\text{CH}-$, $-\text{OCH}_3$), 3.15 (m, 2H, $-\text{SCH}_2-$), 2.42 (m, 1H, $-\text{SCH}_2\text{CH}_2-$), 2.31 (t, $^3J = 7.3$ Hz, 2H, $-\text{COCH}_2-$), 1.87 (m, 1H, $-\text{SCH}_2\text{CH}_2-$), 1.73 - 1.48 (m, 4H, $-\beta\text{-CH}_2$, $-\delta\text{-CH}_2$), 1.42 - 1.30 (m, 2H, $-\gamma\text{-CH}_2-$).

(*R*)-Methyl lipoate (**2**) (1.90 g, 8.6 mmol, 1.0 eq.) was dissolved in 10 mL of methanol and hydrazine hydrate (1.3 mL, 25.8 mmol, 3.0 eq.) was added in one portion. The reaction mixture was stirred at room temperature for 72 h in the absence of light. The solution was concentrated *in vacuo*, re-dissolved in chloroform, and the organic layer was washed with brine. The organic phase was dried over magnesium sulfate, filtered, and concentrated under reduced pressure. (*R*)-lipoic acid hydrazide (**3**) was obtained as a yellow oil (1.22 g, 64%) and used without further purification. ^1H NMR (400 MHz, DMSO-*d*₆) δ (ppm) = 8.92 (bs,

1H, NHCO), 4.14 (bs, 2H, $-\text{NH}_2$), 3.60 (m, 1H, $-\text{CH}-$), 3.14 (m, 2H, $-\text{SCH}_2-$), 2.40 (m, 1H, $-\text{SCH}_2\text{CH}_2-$), 2.00 (t, $^3J = 7.3$ Hz, 2H, $-\text{COCH}_2$), 1.85 (m, 1H, $-\text{SCH}_2\text{CH}_2-$), 1.70 - 1.43 (m, 4H, $-\beta\text{-CH}_2$, $-\delta\text{-CH}_2$), 1.38 - 1.27 (m, 2H, $-\gamma\text{-CH}_2-$). ESI-MS $m/z = 243.20$ $[\text{M}+\text{Na}]^+$, (M (calc.) 243.06 g mol^{-1}).

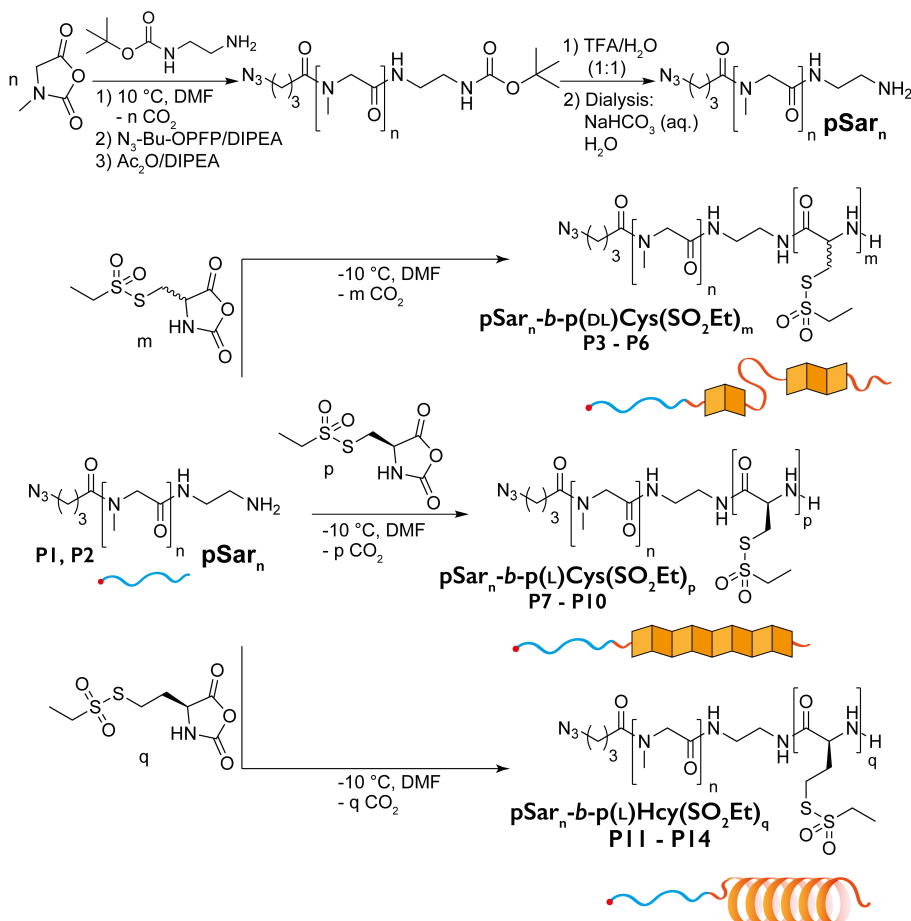
Aggregation Curves. The respective polypept(o)ide (P3-P14) was dissolved in DMAc at a concentration of 7.5 g L^{-1} and 300 μL were placed into a solvent-resistant plastic cuvette for DLS measurement. Aqueous sodium acetate buffer (1 mM, pH 4.75) was added in steps of 5 to 20 μL , and the derived count rate was obtained by DLS with automated positioning and attenuator adjustment at multiple scans. Data were analyzed using Origin 7.5 (OriginLab, USA) and a sigmoidal fit (Model: Dose Response).

Self-Assembly and Core Cross-Linking. The preparation of core cross-linked polymeric micelles was adapted and modified from Klinker *et al.*⁵⁰ Each polypept(o)ide was assembled at the respective turning point.

pSar_{170-b}-p(DL)Cys₁₁ (P3) was dissolved in DMAc at a concentration of 7.5 g L^{-1} for 1 h. Next, 51 vol.% of 1 mM acetate buffer (pH 4.75) was added and the solution was left to equilibrate at room temperature for 3 h, followed by dialysis against 1 mM acetate buffer (pH 4.75). The solution was filtered (GHP 450) and concentrated to 7 g L^{-1} by spin filtration (Amicon Ultra, MWCO 3 kDa), yielding the micelle solution. For cross-linking, in a separate flask, lipoic acid hydrazide was dissolved in ethanol at a concentration of $\beta = 20$ g L^{-1} and one equivalent of an aqueous solution of tris(2-carboxyethyl)phosphine hydrochloride (TCEP HCl) (50 g L^{-1}) was added per disulfide. After 18 h, the cross-linker solution was added to the micelle solution at equimolar amounts of thiols per cysteines. The reaction mixture was stirred at room temperature for 48 h. To remove residual cross-linker and free polymer, the solution was dialyzed against DMSO and MilliQ water (MWCO $6-8$ kDa), followed by repetitive spin filtration (Amicon Ultra, MWCO 100 kDa). Both, micelles and core cross-linked micelles were analyzed by DLS. For CCPMs, the absence of free polymer was verified by gel permeation chromatography in hexafluoro isopropanol.

Results and Discussion

To investigate the influence of secondary structure on the self-assembly of thiol-reactive polypept(o)ides, amphiphilic block copolymers of racemic polysarcosine-*block*-poly(*S*-ethylsulfonyl-DL-cysteine) (pSar-*b*-p(DL)Cys, P3-P6), enantiopure polysarcosine-*block*-poly(*S*-ethylsulfonyl-L-cysteine) (pSar-*b*-p(L)Cys, P7-P10), and polysarcosine-*block*-poly(*S*-ethylsulfonyl-L-homocysteine) (pSar-*b*-p(L)Hcy, P11-P14) have been prepared.



Scheme 1. Synthesis of block copolypept(o)ides with varying secondary structures.

As shown in Scheme 1, all polymers were synthesized by living ring-opening NCA polymerization. Respective secondary structures of the hydrophobic blocks, e.g.,

α -helix, β -sheet, or disrupted β -sheet, are illustrated as reported for the parent homopolymers.^{47–49} The polymerization of sarcosine NCA was initiated by *N*-Boc-1,2-diaminoethane, and pSar (P1, P2) was end-capped with perfluorophenyl-4-azidobutanoate before the Boc-group was removed and block copolypept(o)ides were obtained from pSar macroinitiators. This strategy allows for trifold copolymer modification by [3+2] cycloaddition reactions with alkyne-modified dyes or targeting moieties, reactions with reactive esters at the amine end-group as well as reactions addressing the *S*-ethylsulfonyl group.⁵³

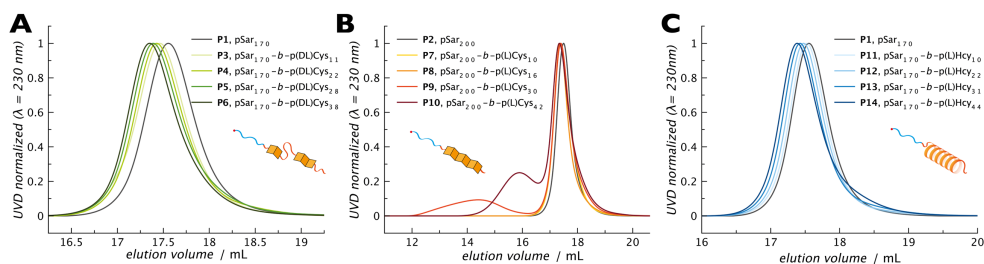


Figure 1. Analytical gel permeation chromatography in HFIP. (A) pSar-*b*-p(DL)Cys (P3-P6), (B) pSar-*b*-p(L)Cys (P7-P10), (C) pSar-*b*-p(L)Hcy (P11-P14), with the respective pSar macroinitiators (P1, P2).

For the preparation of block copolymers, when reacted at -10 °C in DMF, the reactive *S*-ethylsulfonyl protecting group remains intact,⁴⁶ and a shift compared to the pSar macroinitiator can be detected by GPC in hexafluoro isopropanol (HFIP) (Figure 1), which indicates successful chain extension. In detail, symmetric and monomodal molecular weight distributions with a distinct shift for increasing repetitive units were detected for pSar_{*n*}-*b*-p(DL)Cys_{*m*} with *X_m* from 11 to 38 (P3-P6) and pSar_{*n*}-*b*-p(L)Hcy_{*q*} *X_q* = 10 to 44 (P11-P14) (Figure 1A, 1C). Contrariwise, the strong anti-parallel β -sheets of enantiopure p(L)Cys_{*p*} could not be resolved by HFIP for higher chain lengths, accounting for the bimodal GPC in Figure 1B and the corresponding broad dispersity for block copolymers with *X_p* > 16. When analyzed by DOSY NMR in DMSO-*d*₆, however, only one diffusing species could be detected for all pSar_{*n*}-*b*-p(L)Cys_{*n*} polymers (P7-P10) verifying successful polymer synthesis and the absence of homopolymer (see SI, Figure S26-S29).

Table 1. Analytical results of the polymer synthesis.

polymer	X_n (pSar) ^a	$X_{m,p,q}$ (react. block, calc.)	$X_{m,p,q}$ (react. block) ^b	M_n^c / kDa	M_w^c / kDa	$\bar{D}^c$	wt.% react. block
P1 , pSar ₁₇₀	170	-	-	37.7	45.7	1.21	-
P2 , pSar ₂₀₀	200	-	-	39.8	47.9	1.20	-
P3 , pSar ₁₇₀ - <i>b</i> -p(DL)Cys ₁₁	170	15	11	42.6	53.0	1.24	15
P4 , pSar ₁₇₀ - <i>b</i> -p(DL)Cys ₂₂	170	30	22	41.5	53.4	1.28	26
P5 , pSar ₁₇₀ - <i>b</i> -p(DL)Cys ₂₈	170	50	28	42.5	54.3	1.28	31
P6 , pSar ₁₇₀ - <i>b</i> -p(DL)Cys ₃₈	170	50	38	46.1	57.2	1.24	38
P7 , pSar ₂₀₀ - <i>b</i> -p(L)Cys ₁₀	200	20	10	42.3	53.4	1.26	12
P8 , pSar ₂₀₀ - <i>b</i> -p(L)Cys ₁₆	200	30	16	41.9	56.0	1.33	18
P9 , pSar ₂₀₀ - <i>b</i> -p(L)Cys ₃₀	200	50	30	47.7	215.5	4.52	29
P10 , pSar ₂₀₀ - <i>b</i> -p(L)Cys ₄₂	200	50	42	41.9	122.1	2.45	37
P11 , pSar ₁₇₀ - <i>b</i> -p(L)Hcy ₁₀	200	11	10	38.5	48.1	1.25	15
P12 , pSar ₁₇₀ - <i>b</i> -p(L)Hcy ₂₂	200	22	22	38.3	49.9	1.30	27
P13 , pSar ₁₇₀ - <i>b</i> -p(L)Hcy ₃₁	200	35	31	38.9	52.5	1.35	35
P14 , pSar ₁₇₀ - <i>b</i> -p(L)Hcy ₄₄	200	50	44	39.2	54.0	1.37	43

^a determined by HFIP GPC relative to pSar standards²⁹, ^b determined by ¹H NMR, ^c determined by HFIP GPC relative to PMMA standards.

Since α -helices promote NCA polymerization at pseudo first-order kinetics,^{1,12,20} full conversion was detected for the polymerization of L-Hcy(SO₂Et)-NCA, and thus, chain lengths determined for pSar-*b*-p(L)Hcy are in good agreement with the calculated chain lengths (Table 1). In contrast, the NCA polymerization of both, enantiopure and racemic L-Cys(SO₂Et)-NCA and DL-Cys(SO₂Et)-NCA, was governed by insoluble β -sheets referring to an Avrami-type of chain growth.^{47,48} Consequently, polymerizations were stopped when no further progress could be detected by FT-IR spectroscopy, accounting for the deviation between calculated and obtained chain lengths.⁵⁴ Of note, in our case, the addition of chaotropic thiourea, freshly purified by vacuum sublimation, did not help to overcome this limitation but solely resulted in homopolymer formation (Figure S1).⁵⁵ Nevertheless, as summarized in Table 1, three sets containing four reactive amphiphilic polypept(o)ides have been prepared successfully. These polymers are characterized by comparable chain lengths of 170 and 200 for hydrophilic pSar,

and 10 to 44 for the hydrophobic reactive *S*-ethylsulfonyl-protected blocks. The respective block ratios correspond to similar hydrophobic segments ranging from 12 to 43 weight%, well suitable for self-assembly to micellar structures.^{56–58}

To verify the secondary structure, block copolypept(o)ides were analyzed by FT-IR spectroscopy in the solid state.^{59,60} The characteristic signals for an α -helix can generally be found at 1650 cm^{-1} (Amide I) and 1546 cm^{-1} (Amide II), while for β -sheets these peaks can be observed at 1630 cm^{-1} (Amide I) and 1530 cm^{-1} (Amide II), as well as 1656 cm^{-1} (Amide I) and 1535 cm^{-1} (Amide II) for random coil structures.^{1,59–61} As shown in Figure 2A, predominant signals of the amide bands at 1546 and 1317 cm^{-1} (amide II, amide III; blue arrows) can be assigned to the α -helical p(L)Hcy block despite the co-existence with the random coil of polysarcosine which overlaps for the amide I peak. Furthermore, CD-spectroscopy in HFIP showed an increasing concision of the characteristic signals (minima at 212 nm and 226 nm, maximum at 194 nm) for growing p(L)Hcy chain lengths confirming the α -helical structure (Figure S2).⁶² For enantiopure pSar-*b*-p(L)Cys copolymers, the shoulder at 1703 cm^{-1} (red arrow) accounts for the anti-parallel orientation of the β -sheets, while no characteristic peaks can be distinguished for

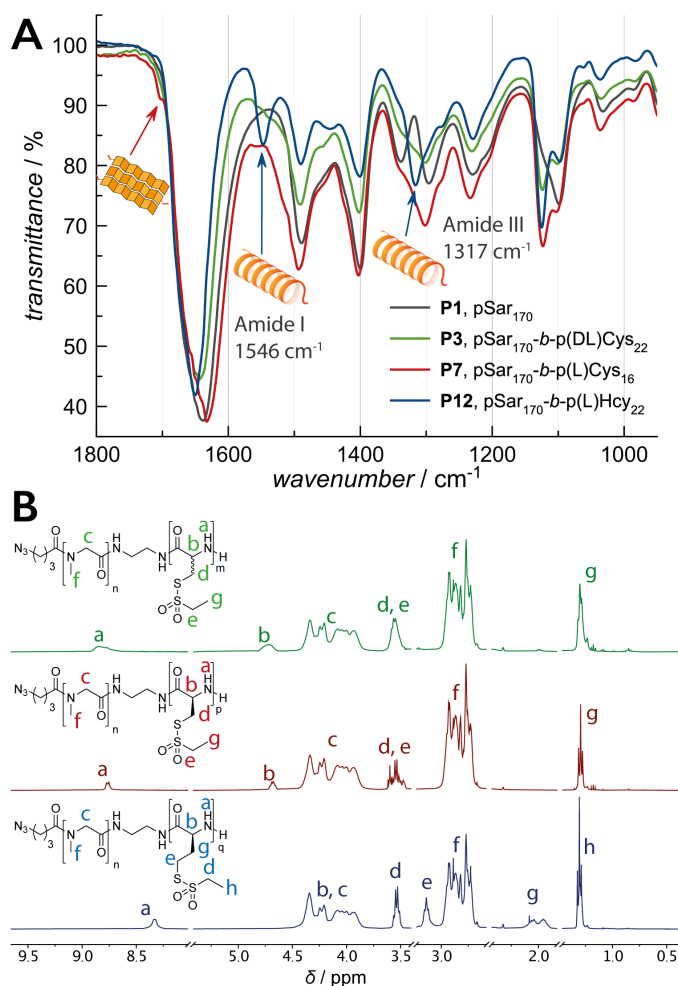


Figure 2. Secondary structure analysis of pSar-*b*-p(DL)Cys, pSar-*b*-p(L)Cys, and pSar-*b*-p(L)Hcy. (A) FT-IR spectroscopy, (B) ^1H NMR spectroscopy.

racemic pSar-*b*-p(DL)Cys copolymers. When analyzed by NMR spectroscopy (Figure 2B), block copolymers of pSar-*b*-p(DL)Cys show much broader peaks compared to enantiopure pSar-*b*-p(L)Cys, suggesting a rather atactic polymer structure similar to observations for the racemic p(DL)Cys homopolymers.⁴⁸ Taken together and referring to the secondary structures detected for the respective homopeptides in solution by circular dichroism spectroscopy,^{47–49} GPC, IR and NMR spectroscopy confirm the successful preparation of reactive block

copolypept(o)ides with secondary structures varying from α -helix to disrupted or anti-parallel β -sheets.^{63,64}

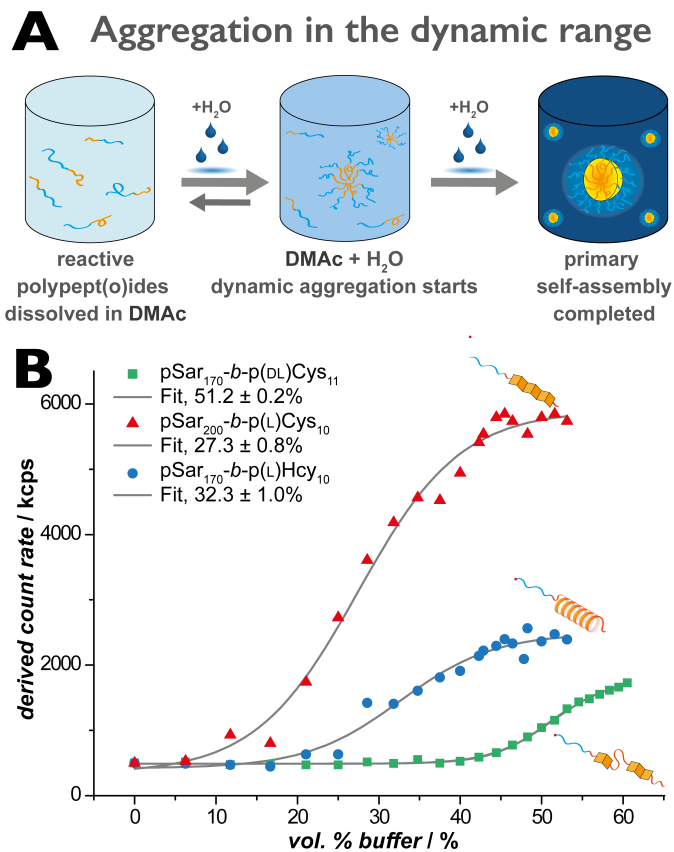


Figure 3. Influence of the secondary structure on the aggregation behavior. (A) Schematic illustration of the dynamic range. (B) Aggregation curve of polypept(o)ides (P3, P7, P11) with sigmoidal fit.

The influence of secondary structure on solution self-assembly was investigated by a titration- experiment. As illustrated in Figure 3A, the reactive polypept(o)ides were dissolved in a good solvent, *N,N*-dimethylacetamide (DMAc), and water, equipped with 1 mM sodium acetate (pH 4.75), was added stepwise. When a certain buffer concentration is reached, polymers start to assemble, and the scattering intensity increases, which correlates with the derived count rate detected by DLS. The count rate continues to increase as long as new micelles form or larger aggregates are growing ($I \sim r^6$).⁶⁵ Finally, a plateau gets reached when the concentration of the block-selective solvent exceeds a certain limit. Due to the low solubility of free polymer, micelles remain stable.^{56,66,67} Besides additional parameters such as solvents and polymer concentration, the dynamic range of self-assembly is specific for each polymer. For head-to-head comparison, racemic pSar-*b*-p(DL)Cys, enantiopure pSar-*b*-p(L)Cys, and pSar-*b*-p(L)Hcy were dissolved in DMAc at equal concentrations, and the aqueous buffer was added at comparable time intervals. Interestingly, the three copolymers of comparable chain lengths showed significant variations in their aggregation behavior (Figure 3B). Sigmoidal shapes were obtained for all aggregation curves, however, the sigmoidal fit revealed a turning point at $27.3 \pm 0.8\%$ for the enantiopure pSar₂₀₀-*b*-p(L)Cys₁₀, while turning points of $51.2 \pm 0.2\%$ and $32.3 \pm 1.0\%$ were derived for racemic pSar₁₇₀-*b*-p(DL)Cys₁₁ and pSar₁₇₀-*b*-p(L)Hcy₁₀, respectively. Moreover, the final plateau level of pSar₂₀₀-*b*-p(L)Cys₁₀ was found ~ 3 -fold higher compared to racemic pSar₁₇₀-*b*-p(DL)Cys₁₁ and ~ 2 -fold higher compared to pSar₁₇₀-*b*-p(L)Hcy₁₀. As the primary structure does not differ significantly among the selected copolymers, the observed effects can be attributed to the variation of the secondary structure, which testifies anti-parallel β -sheets an assembly-promoting character stronger than α -helices, likely caused by attractive interactions between individual β -sheets.

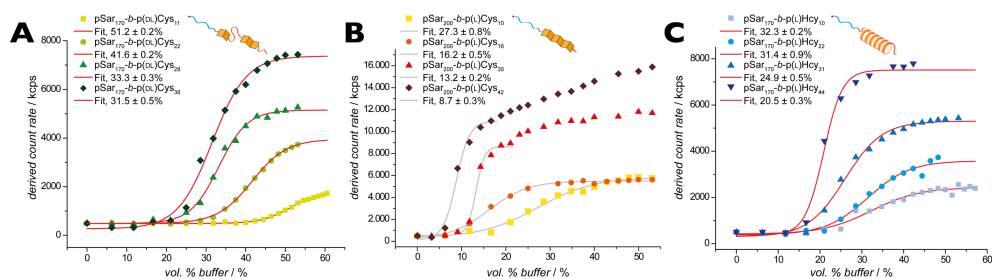


Figure 4. Aggregation curves for copolymers P3 - P14. (A) pSar-*b*-p(DL)Cys, (B) pSar-*b*-p(L)Cys, (C) pSar-*b*-p(L)Hcy.

When the chain lengths of the hydrophobic polypeptide blocks are increased, turning points generally shift to aggregation at lower buffer concentrations for all copolymers (Figure 4). In addition, the dynamic range, thus the area between the start of the aggregation and the primary plateau, becomes narrower by increasing the hydrophobic segment. As such, the dynamic range extents from 20 to 50 volume% buffer for pSar₁₇₀-*b*-p(L)Hcy₁₀, but from 15 to 30 vol.% buffer for pSar₁₇₀-*b*-p(L)Hcy₄₄. Furthermore, the trend of anti-parallel β -sheets as being the strongest promotor of self-assembly can be confirmed by the shift of the turning point from 27.3 ± 0.8 vol.% for pSar₂₀₀-*b*-p(L)Cys₁₀ to concentrations as low as 8.7 ± 0.3 vol.% buffer for pSar₂₀₀-*b*-p(L)Cys₄₂. Interestingly, for pSar₂₀₀-*b*-p(L)Cys₃₀ and pSar₂₀₀-*b*-p(L)Cys₄₂, the primary aggregation seems accompanied by a secondary aggregation, as the sigmoidal fit cannot be applied over the entire range, indicating further re-arrangement of the initially formed micelles.^{68,69}

A Self-Assembly and Core Cross-Linking

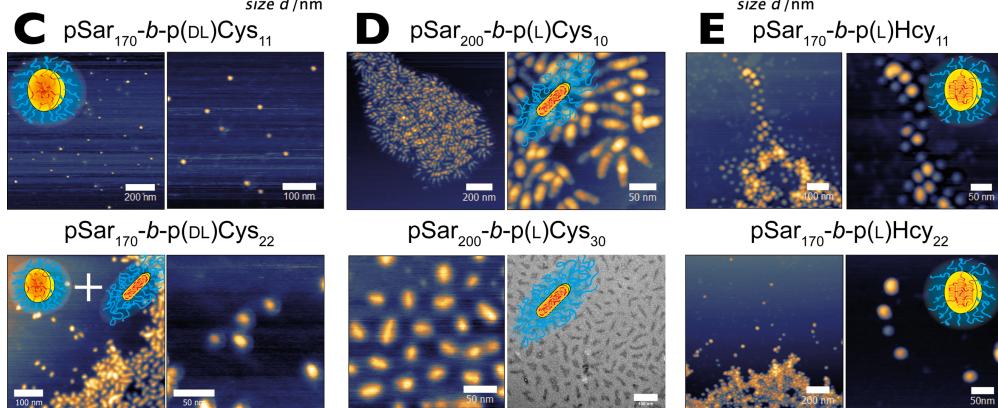
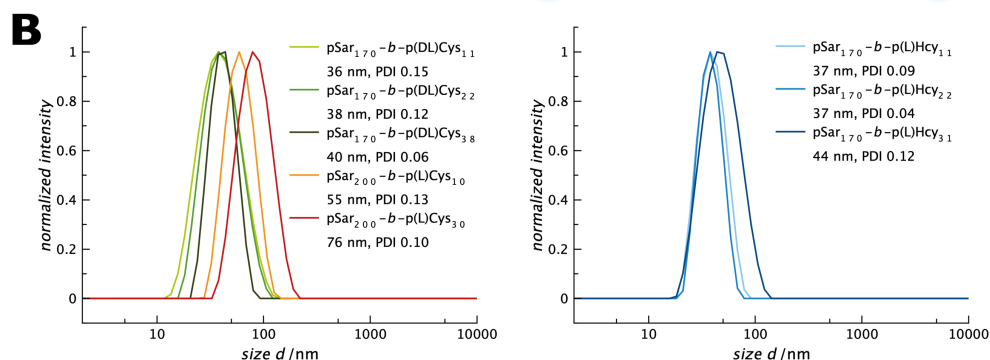
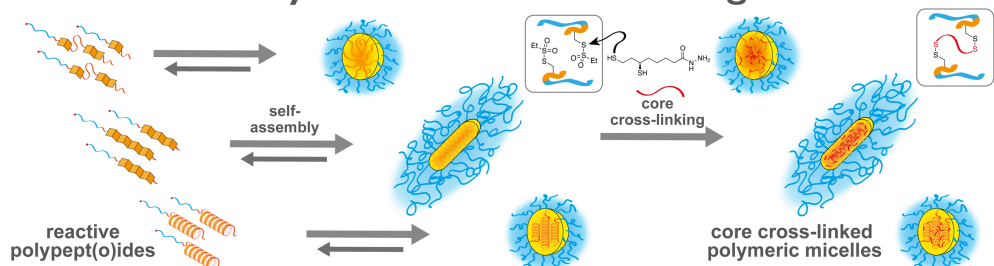


Figure 5. Self-assembly and core cross-linking. (A) Schematic illustration of secondary structure-driven self-assembly. (B) Single-angle DLS analysis of polymeric micelles (left: $\text{pSar}\text{-}b\text{-p(DL)Cys}$ and $\text{pSar}\text{-}b\text{-p(L)Cys}$, right: $\text{pSar}\text{-}b\text{-p(L)Hcy}$). Morphological Analysis of core cross-linked polymeric micelles by AFM and cryoTEM: (C) $\text{pSar}\text{-}b\text{-p(DL)Cys}$, (D) $\text{pSar}\text{-}b\text{-p(L)Cys}$, (E) $\text{pSar}\text{-}b\text{-p(L)Hcy}$.

Finally, the findings on the aggregation behavior were transferred to the preparation of core cross-linked polymeric micelles (CCPMs) from the reactive polypept(o)ides, as illustrated in Figure 5A. The copolymers were dissolved in DMAc and equilibrated by addition of buffer at their respective turning points, before solvent exchange was completed by dialysis. When analyzed by DLS (Figure 5B), both, α -helical pSar-*b*-p(L)Hcy and racemic pSar-*b*-p(DL)Cys yielded small polymeric micelles of similar size (D_h 36-44 nm) and narrow PDI (0.04 - 0.15), despite increasing chain lengths of the respective hydrophobic blocks. In contrast, an increased size was detected for enantiopure pSar-*b*-p(L)Cys (D_h 55-76 nm) directing to larger structures for polymers with longer hydrophobic blocks. For further analysis, micelles were cross-linked by reaction of the *S*-ethylsulfonyl-group with dihydro lipoic acid hydrazide (3), and the morphology of the obtained disulfide cross-linked micelles was investigated by AFM (Figure 5C-E, Figure S5-S8). Of note, the cross-linking reaction did not influence the overall size distribution (Figure S4). Analysis by AFM revealed spherical structures with diameters well below 50 nm for CCPMs from racemic pSar-*b*-p(DL)Cys containing 15 wt.% p(DL)Cys, while spherical to slightly elongated particles could be observed for 26 wt.% p(DL)Cys (Figure 5C, Figure S5). As shown by AFM and cryoTEM (Figure 5D, Figure S6 - S7), solely worm-like micelles were obtained for pSar-*b*-p(L)Cys containing both, 12 or 29 wt.% of enantiopure p(L)Cys, which corresponds well with previous findings as well as with the larger sized determined by DLS.⁵⁰ Spherical CCPMs only were obtained from α -helical pSar-*b*-p(L)Hcy of 15 or 27 wt.% p(L)Hcy, as shown in Figure 5E and Figure S8.⁷⁰

Taken together, secondary structure formation is a major parameter affecting the self-assembly of polypept(o)ides. In direct comparison, the reactive amphiphilic block copolymer with an α -helical segment, pSar-*b*-p(L)Hcy, only yielded spherical structures, whereas β -sheets in enantiopure pSar-*b*-p(L)Cys strongly direct self-assembly to worm-like particles and even affect racemic pSar-*b*-p(DL)Cys with longer hydrophobic segments.

Conclusion

In conclusion, our head-to-head comparison underlines the influence of secondary structure formation on the aggregation and self-assembly of block copolypept(o)ides. Herein, we reported the successful synthesis of reactive, *S*-ethylsulfonyl-protected pSar-*b*-p(DL)Cys, pSar-*b*-p(L)Cys, and pSar-*b*-p(L)Hcy. Despite comparable primary structures, the secondary structure of these block

copolymers varies from disrupted β -sheet to anti-parallel β -sheet and α -helix, which impacts their tendency to self-assemble during the solvent switch. The anti-parallel β -sheets of enantiopure pSar-*b*-p(L)Cys strongly induce early aggregation compared to the racemic pSar-*b*-p(DL)Cys counterpart and narrow the dynamic range. Moreover, the α -helix in pSar-*b*-p(L)Hcy, favors the formation of spherical structures, whereas β -sheets direct self-assembly into worm-like micelles for enantiopure pSar-*b*-p(L)Cys and even influence the self-assembly of pSar-*b*-p(DL)Cys in which the anti-parallel orientation is disrupted by the racemic hydrophobic block. A detailed understanding of the secondary structure enables powerful control over the morphology of micellar assemblies and fine-tuning of particle properties beyond the hydrophilic-hydrophobic ratio. This implies a major significance for polypeptides as drug delivery systems and in materials science.

Author Contribution

Experiment design, polymer synthesis, nanoparticle preparation, AFM, and CD measurements were performed by T.A.B., J.I. contributed to polymer synthesis and nanoparticle formation, and C.M. contributed to polymer synthesis. B.W. performed cryoTEM analysis. The manuscript was written by T.A.B. and M.B. The project was supervised by M.B. All authors have given approval to the final version of the manuscript.

Notes

The authors declare the following competing financial interest(s): Matthias Barz holds the patent Thiol-protected amino acid derivatives and uses thereof WO2015169908A1.

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Abbreviations

AFM, atomic force microscopy; Boc, *tert*-butoxycarbonyl; CCPM, core cross-linked polymeric micelle; CD, circular dichroism; DLS, dynamic light scattering;

DMAc, *N,N*-dimethylacetamide; DMF, *N,N*-dimethylformamide; DMSO, dimethylsulfoxide; GPC, gel permeation chromatography; FT-IR spectroscopy, Fourier-transformed infrared spectroscopy; HFIP, 1,1,1,-3,3,3-hexafluoropropan-2-ol; MWCO, molecular weight cut-off; NCA, *N*-carboxyanhydride; pSar, polysarcosine; p(L)Cys, poly(*S*-ethylsulfonyl-L-cysteine); p(DL)Cys, poly(*S*-ethylsulfonyl-DL-cysteine); p(L)Hcy, poly(*S*-ethylsulfonyl-L-homocysteine); cryoTEM, cryogenic transmission electron microscopy.

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Supporting Information

Results and Discussion

NCA-Polymerization with thiourea

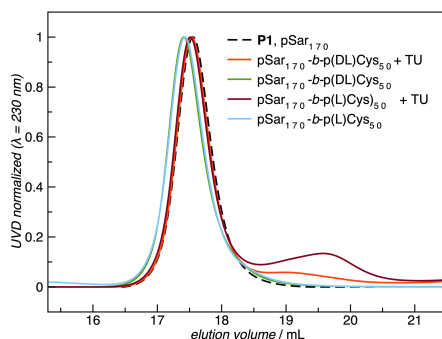


Figure S1. HPLC-GPC analysis of NCA polymerizations in the presence or absence of 1M thiourea in DMF. All polymerizations were performed with 50 eq. of DL-Cys(SO₂Et)- or L-Cys(SO₂Et)-NCA. No significant shift but only homopolymer formation was detected for polymerizations performed in the presence of 1M thiourea.

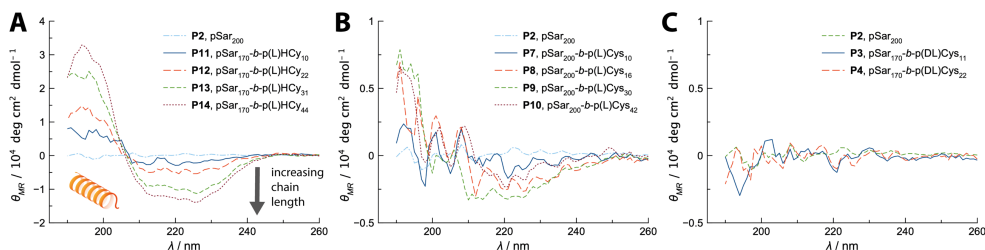


Figure S2. CD spectroscopy of polypept(o)ides P2 - P14. (A) pSar-*b*-p(L)Hcy. The α -helical secondary structure becomes more pronounced for growing chain lengths of p(L)Hcy. (B) pSar-*b*-p(L)Cys. No significant secondary structure detectable. (C) pSar-*b*-p(DL)Cys. No significant secondary structure detectable. Note that pSar significantly contributes to the overall absorbance limiting the resolution of this method.

Core Cross-Linked Polymeric Micelles

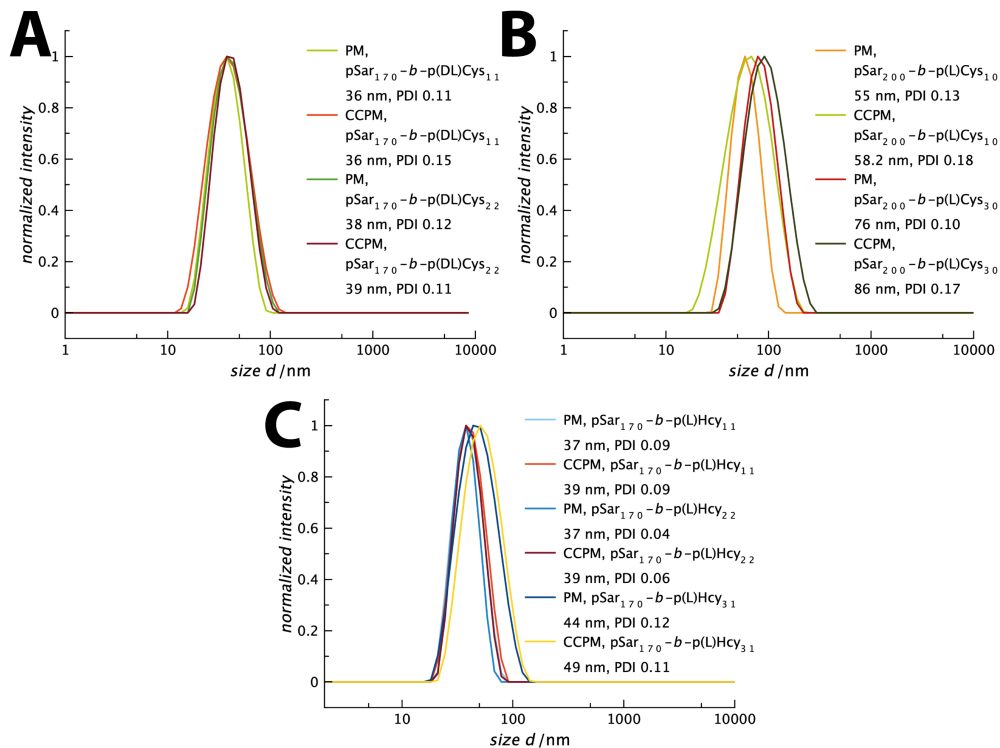


Figure S3. Single-Angle DLS analysis of polymeric micelles before (PM) and after cross-linking (CCPM). Nanoparticles were formed from (A) pSar-*b*-p(DL)Cys, (B) pSar-*b*-p(L)Cys, (C) pSar-*b*-p(L)Hcy.

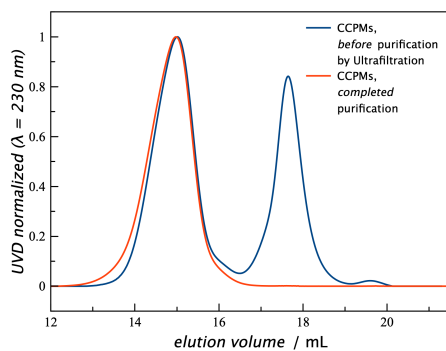


Figure S4. HFIP GPC analysis of CCPMs before and after purification by ultrafiltration (Amicon Ultra, MWCO 100kDa).

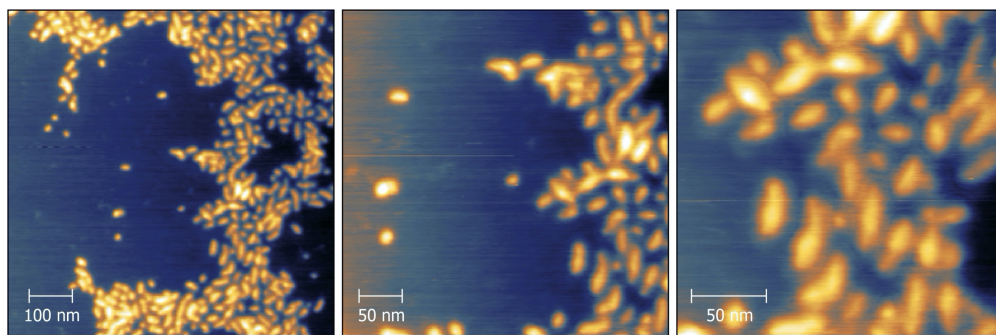


Figure S5. Additional AFM images of pSar₁₇₀-b-p(DL)Cys₂₂ (P4).

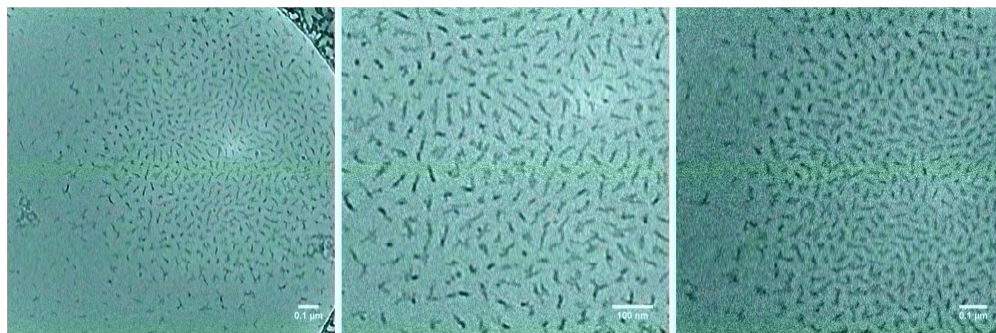


Figure S6. Additional CryoTEM images of pSar₂₀₀-b-p(L)Cys₃₀ (P9).

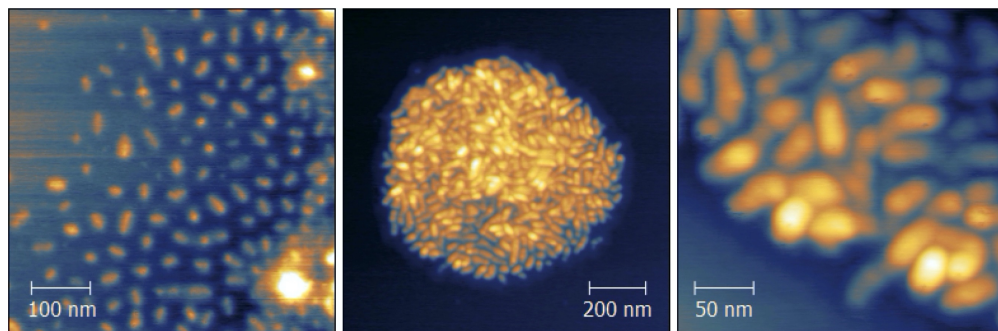


Figure S7. Additional AFM images of pSar₂₀₀-*b*-p(L)Cys₃₀ (P9).

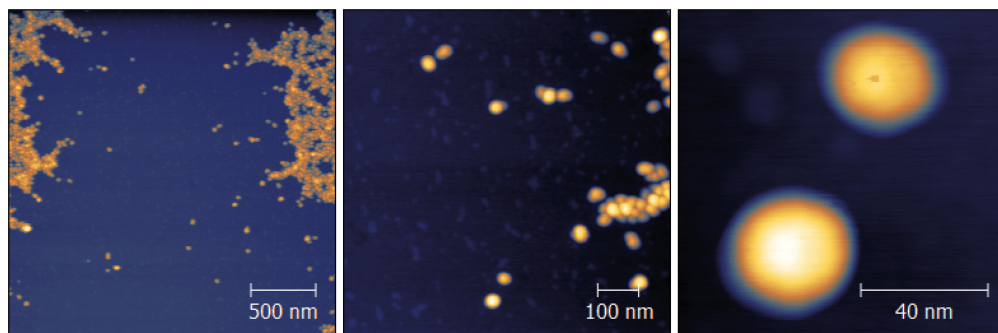
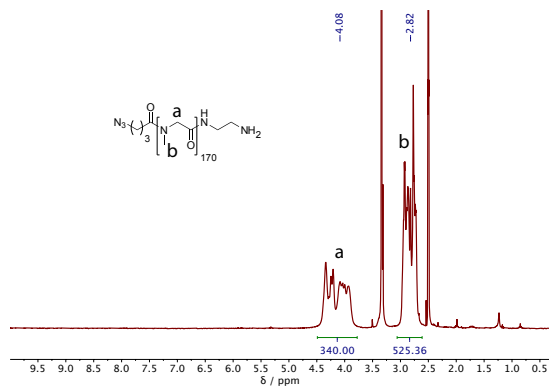
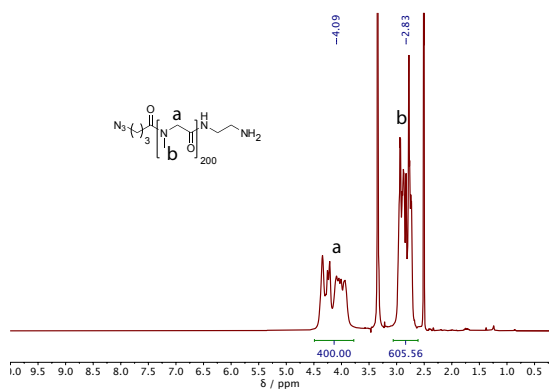


Figure S8. Additional AFM images of pSar₁₇₀-*b*-p(L)Hcy₂₂ (P12).

NMR-Data

**Figure S9.** ¹H NMR analysis of **P1**, pSar₁₇₀, in DMSO-*d*₆.**Figure S10.** ¹H NMR analysis of **P2**, pSar₂₀₀, in DMSO-*d*₆.

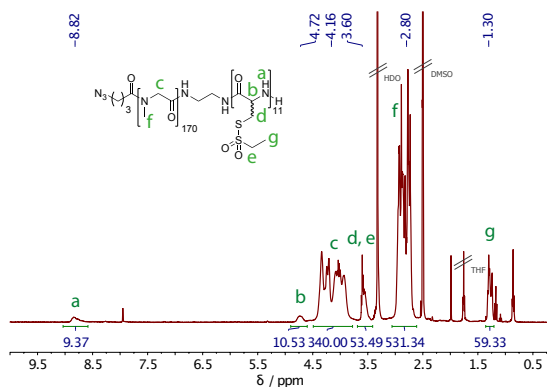


Figure S11. ¹H NMR analysis of **P3**, pSar₁₇₀-b-p(DL)Cys₁₁, in DMSO-*d*₆.

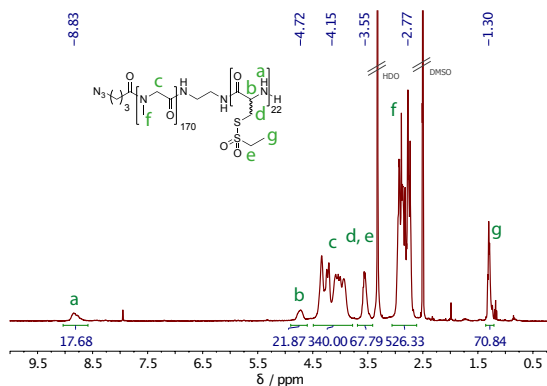


Figure S12. ¹H NMR analysis of **P4**, pSar₁₇₀-b-p(DL)Cys₂₂, in DMSO-*d*₆.

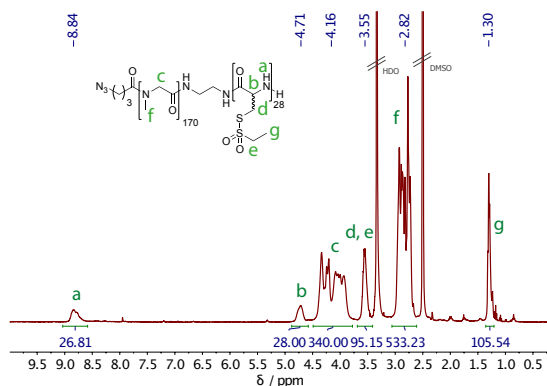


Figure S13. ¹H NMR Analysis of **P5**, pSar₁₇₀-b-p(DL)Cys₂₈, in DMSO-*d*₆.

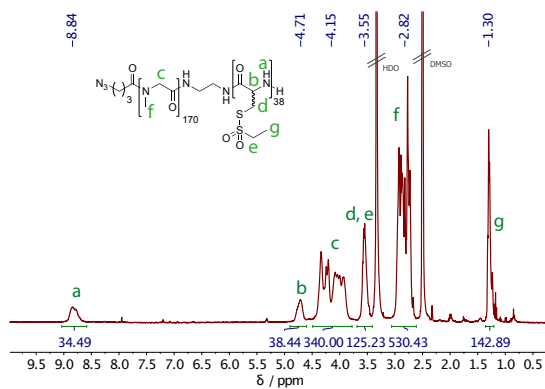


Figure S14. ¹H NMR analysis of **P6**, pSar₁₇₀-b-p(DL)Cys₃₈, in DMSO-*d*₆.

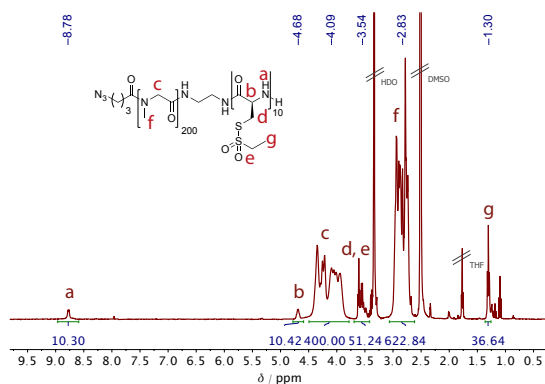


Figure S15. ¹H NMR analysis of **P7**, pSar₂₀₀-b-p(L)Cys₁₀, in DMSO-*d*₆.

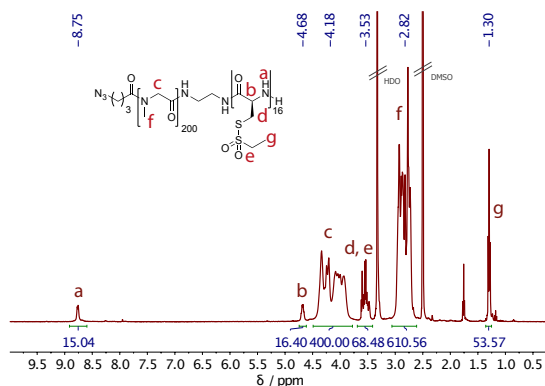


Figure S16. ¹H NMR Analysis of **P8**, pSar₂₀₀-b-p(L)Cys₁₆, in DMSO-*d*₆.

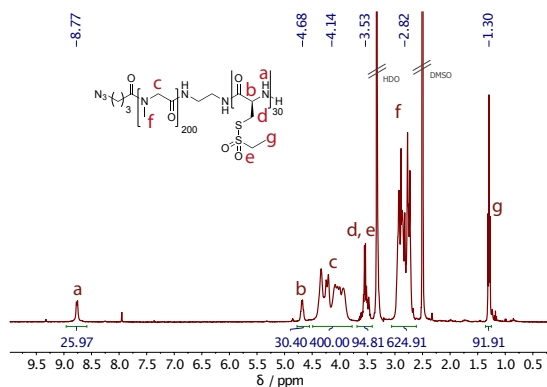


Figure S17. ¹H NMR analysis of **P9**, pSar₂₀₀-b-p(L)Cys₃₀, in DMSO-*d*₆.

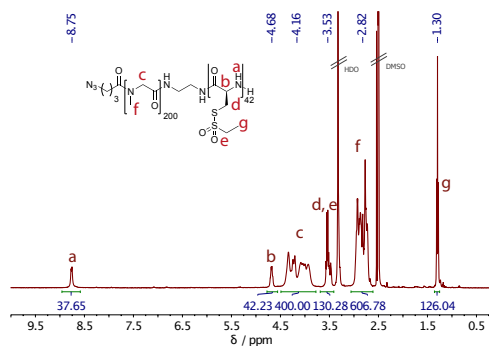


Figure S18. ¹H NMR analysis of **P10**, pSar₂₀₀-b-p(L)Cys₄₂, in DMSO-*d*₆.

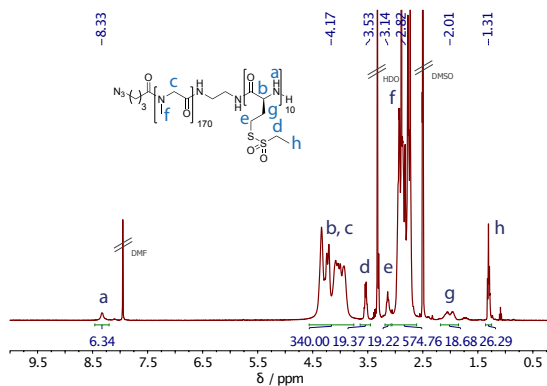


Figure S19. ¹H NMR analysis of **P11**, pSar₁₇₀-b-p(L)Hcy₁₀, in DMSO-*d*₆.

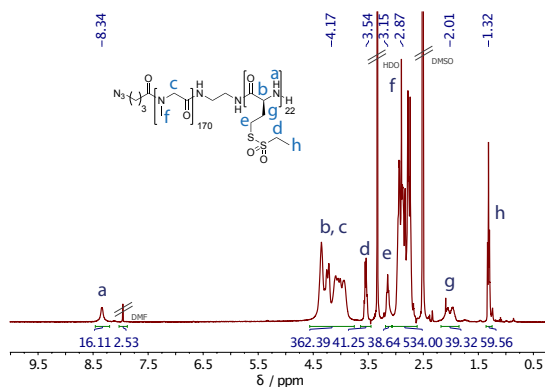


Figure S20. ¹H NMR analysis of **P12**, pSar_{170-b}-p(L)Hcy₂₂, in DMSO-*d*₆.

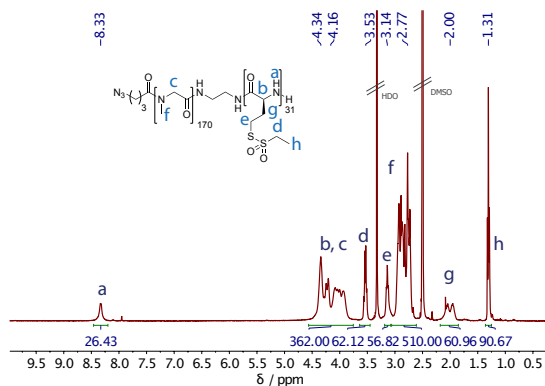


Figure S21. ¹H NMR Analysis of **P13**, pSar_{170-b}-p(L)Hcy₃₁, in DMSO-*d*₆.

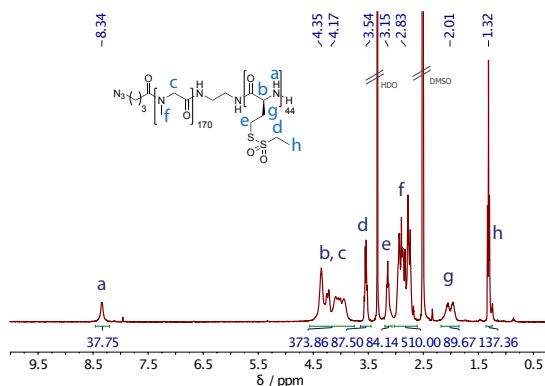


Figure S22. ¹H NMR analysis of **P14**, pSar_{170-b}-p(L)Hcy₄₂, in DMSO-*d*₆.

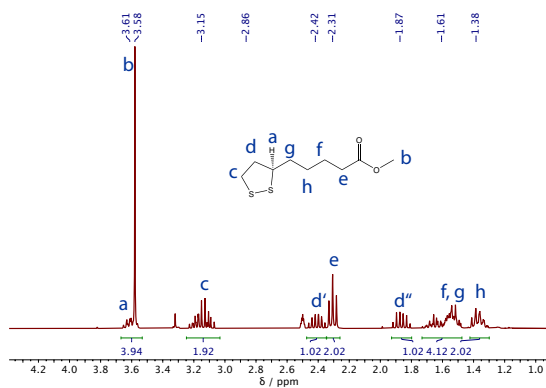


Figure S23. ¹H NMR analysis of *R*-methyl lipoate (**2**) in DMSO-*d*₆.

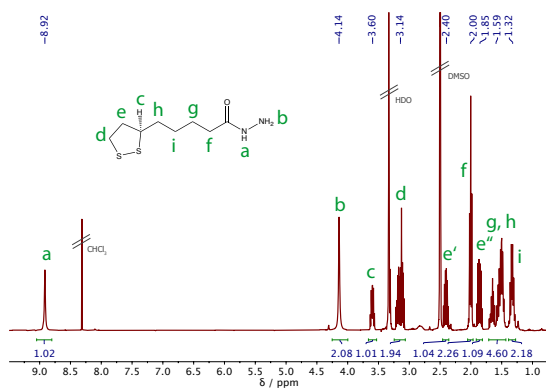


Figure S24. ¹H NMR Analysis of *R*-lipoic acid hydrazide (**3**) in DMSO-*d*₆.

DOSY-NMRs

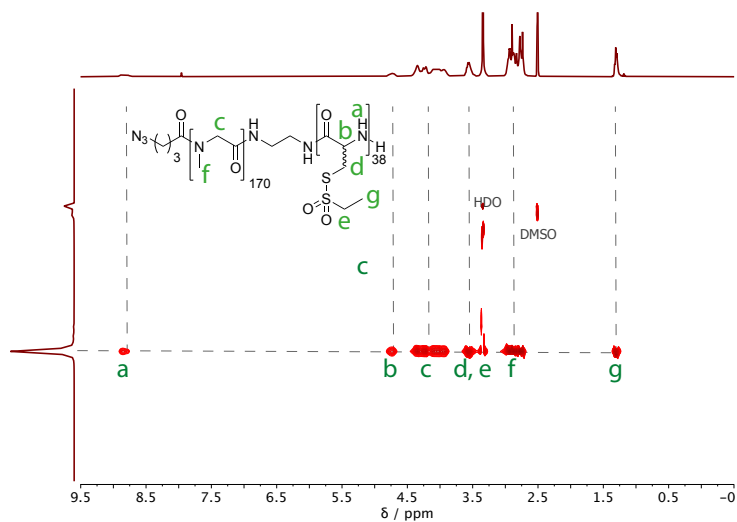


Figure S25. DOSY NMR analysis of **P6**, pSar_{170-b}-p(DL)Cys₃₈, in DMSO-*d*₆.

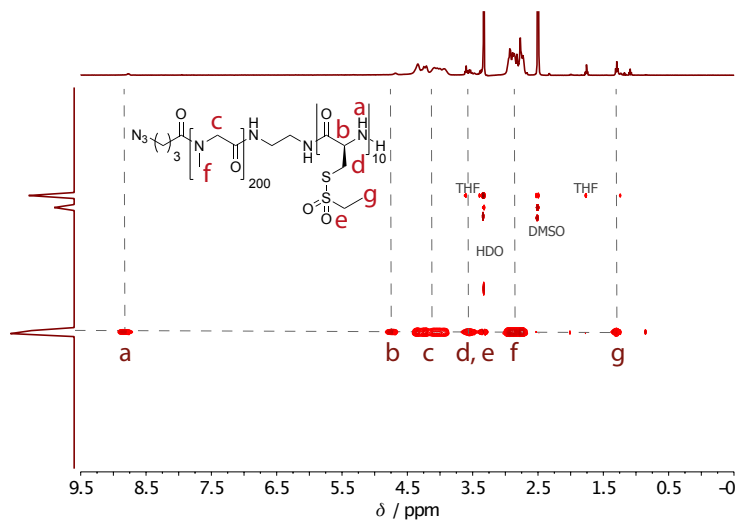


Figure S26. DOSY NMR analysis of **P7**, pSar_{200-b}-p(L)Cys₁₀, in DMSO-*d*₆.

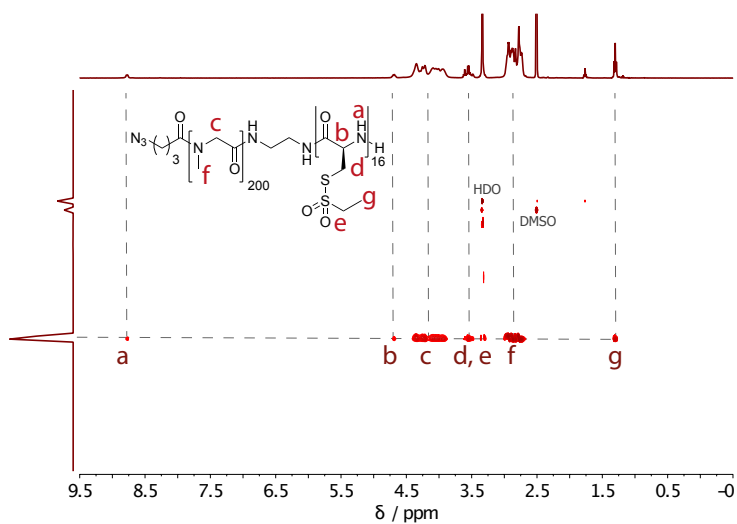


Figure S27. DOSY NMR analysis of **P8**, pSar₂₀₀-b-p(L)Cys₁₆, in DMSO-*d*₆.

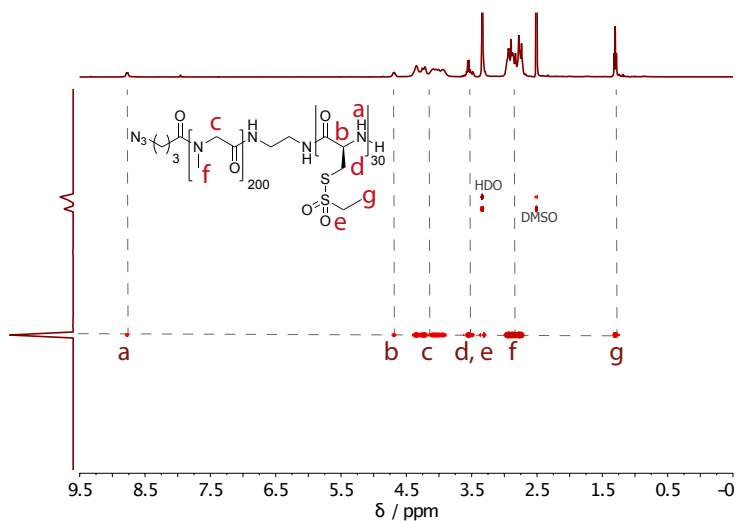


Figure S28. DOSY NMR analysis of **P9**, pSar₂₀₀-b-p(L)Cys₃₀, in DMSO-*d*₆.

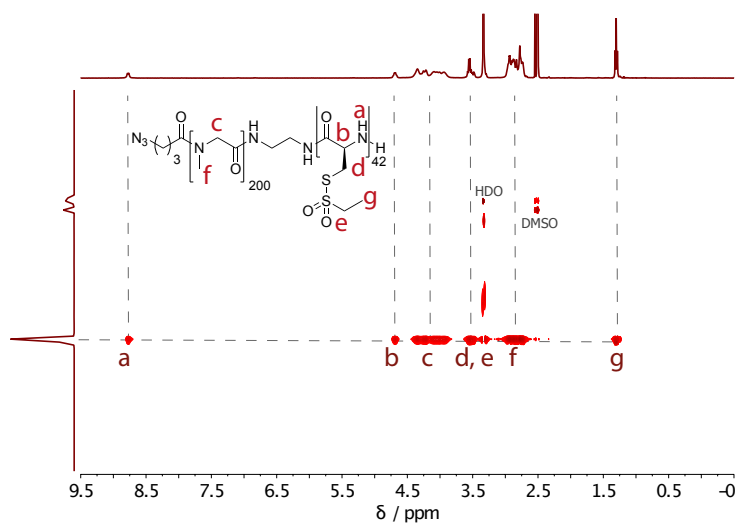


Figure S29. DOSY NMR analysis of **P10**, pSar₂₀₀-b-p(L)Cys₄₂, in DMSO-*d*₆.

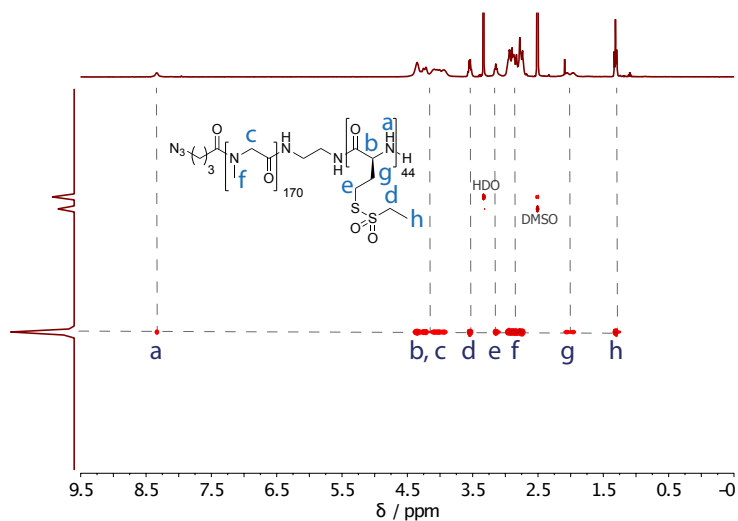


Figure S30. DOSY NMR analysis of **P14**, pSar₁₇₀-b-p(L)Hcy₄₄, in DMSO-*d*₆.