



Universiteit
Leiden
The Netherlands

Zebrafish atherosclerosis: experimental definitions and difficulties

Verwilligen, R.A.F.; Bussmann, J.; Eck, M. van

Citation

Verwilligen, R. A. F., Bussmann, J., & Eck, M. van. (2020). Zebrafish atherosclerosis: experimental definitions and difficulties. *Atherosclerosis*, 302, 52-54.

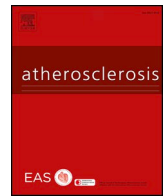
doi:10.1016/j.atherosclerosis.2020.04.007

Version: Publisher's Version

License: [Licensed under Article 25fa Copyright Act/Law \(Amendment Taverne\)](#)

Downloaded from: <https://hdl.handle.net/1887/3201342>

Note: To cite this publication please use the final published version (if applicable).



Correspondence

Zebrafish atherosclerosis: Experimental definitions and difficulties



ARTICLE INFO

Keywords:

Atherosclerosis
Macrophages
Zebrafish
Foam cell
Cholesterol

To the Editor,

Macrophages play an important role in atherogenesis. Early stages of the disease are characterized by the accumulation of foam cells, derived from macrophages that have taken up excessive amounts of cholesterol. Our lab studies the role of macrophages in atherosclerosis progression as well as atherosclerosis regression. Mice represent the most widely used model organism for studying atherosclerosis. However, a clear limitation of using mice is that only end-point atherosclerosis measurements are possible. In this context, zebrafish provides an attractive alternative atherosclerosis model because of its optical transparency and the associated ability for non-invasive cell, i.e. macrophage tracking. With great interest, we have therefore read the paper of Yan et al. (2018), who recently established a zebrafish atherosclerosis model showing high cholesterol diet (HCD)-induced vascular lipid accumulation and co-localization of myeloid cells with lipids in the caudal vein of zebrafish larvae [1].

The experimental setup of Yan and colleagues was based on a paper of Stoletov et al. (2009) who, more than 10 years ago, for the first time described the possibility to induce early atherogenesis in zebrafish [2]. Stoletov et al. demonstrated that wild-type adult (3-month old) zebrafish are susceptible to develop hypercholesterolemia, e.g. increased total cholesterol levels, and fatty streaks in the dorsal aorta upon 8–12 weeks of 4% (w/w) HCD feeding. Interestingly, larvae of wild-type zebrafish (15 days post fertilization (dpf)) also developed vascular lipid deposits in the caudal vein after only 10 days challenge with fluorescently labelled 4% HCD. In their studies, Stoletov et al. used the *Lyz:DsRed2* transgenic zebrafish line to examine the effect of HCD on myeloid cell recruitment. Upon HCD challenge, more myeloid cells were recruited to the caudal vein, but the possible co-localization of these cells with the vascular lipid deposits was not determined. In this light, it should be noted that under control of the lysozyme promoter, all myeloid cells are labelled and hence it is not macrophage-specific [3]. Similarly, in the recently published manuscript of Yan et al., the *Coronin1a:eGFP* line was used, which also labels all myeloid cells as well as T-lymphocytes [4].

To examine the specific role of macrophages in early atherogenesis in zebrafish larvae, we used a zebrafish line in which the *Mpeg1* promoter drives fluorescent transgene expression [5]. At five dpf, the 10 day-challenge with 4% HCD labelled with 10 µg/g cholesteryl ester Bodipy FLC12 was initiated as previously described [2] (Fig. 1A). Although adult zebrafish develop hypercholesterolemia upon long-term HCD feeding, we did not observe a difference in total body cholesterol levels in the zebrafish larvae after 10 days on HCD (Fig. 1B). However, we did note compensatory downregulation of genes involved in cholesterol metabolism, i.e. HMG-CoA reductase (*hmgcr*; –47%; $p = 0.05$) and the low-density lipoprotein receptor (*ldlr*; –85%; $p = 0.003$). No effect was found in the expression of ATP-binding cassette cholesterol efflux transporters *abca1* and *abcg1* (Fig. 1C). Co-localization of macrophages with labelled cholesteryl ester was used as readout for foam cell formation (early atherogenesis). We incidentally found foam cells in the caudal vein of the *Mpeg1:RFP* larvae. However, the numbers were very low and did not appear to be different between the control (CTRL) and HCD group (Fig. 1D).

Although both Stoletov et al. [2] as well as Yan et al. [1] have shown that early atherogenesis can be modulated in zebrafish larvae, we have not been able to reproduce their findings. As we followed the same protocol as published (Table 1), it is hard to pinpoint the underlying reason for the differential effects. In agreement with our current findings, Yoon et al. (2013) also did not find accumulation of leukocytes in the caudal vein of zebrafish larvae after 10 days on HCD [6]. The differential outcomes amongst the different laboratories might be due to small differences in general zebrafish husbandry, affecting the zebrafish microbiome and thereby immunity of the host [7]. Alternatively, the definition of early atherogenesis applied might influence the outcome.

In mammalian atherosclerosis, macrophages generally make up the majority of myeloid cells present in the early atherosclerotic lesion. Our apparently negative results whilst using a macrophage-specific transgenic line as compared to more general myeloid transgenics could indicate that other myeloid cells drive early atherosclerosis in zebrafish larvae. In support, Bandaru et al. (2019) demonstrated that 5 days of

DOI of original article: <https://doi.org/10.1016/j.atherosclerosis.2018.11.016>

<https://doi.org/10.1016/j.atherosclerosis.2020.04.007>

Received 31 March 2020; Accepted 3 April 2020

Available online 19 April 2020

0021-9150/© 2020 Elsevier B.V. All rights reserved.

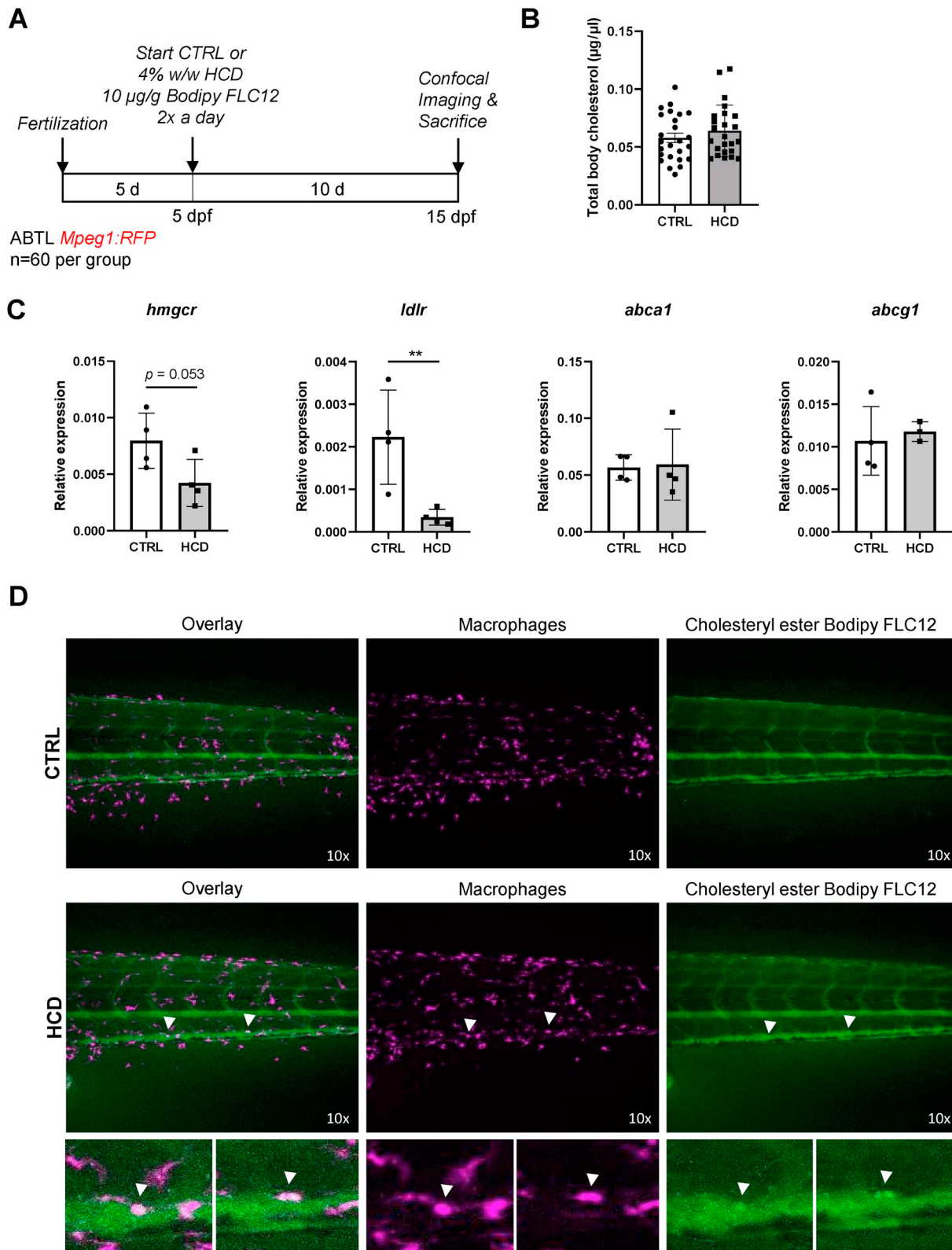


Fig. 1. No evidence for enhanced macrophage foam cell formation in ABTL wild-type zebrafish larvae upon 10 days of HCD feeding. (A) Experimental setup (B); total body cholesterol levels (n = 25); (C) relative expression levels of the key genes involved in cholesterol metabolism (n = 4 pools of 7 fish each); (D) representative images of the caudal vein showing cholesteryl esters in green and macrophages in purple. Incidentally found foam cells, as identified by purple and green co-localization, are indicated by the white arrowheads. All data represent means \pm SEM; ** $p < 0.01$. (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

Table 1
Overview of the similarities and differences in setup between zebrafish larvae atherosclerosis studies.

	Current study	Stoletov et al. [2]	Yan et al. [1]	Yoon et al. [6]	Bandaru et al. [8]
Tg zebrafish line	Tg(Mpeg1:RFP)	Tg(Lyz:DSred2)	Tg(Coronin1a:eGFP)	Tg(FH1:GFP)	Tg(Mpo:eGFP)
Feeding	2 times a day	- n/d	- n/d	- n/d	2 times a day
Label	4% (w/w) HCD Standard dry food	4% (w/w) HCD Artificial Artemia	4% (w/w) HCD - n/d	4% (w/w) HCD Artificial Artemia	4% (w/w) HCD Standard dry food
Days of HCD	Cholesteryl ester analog	Cholesteryl ester analog	Cholesteryl ester analog	Cholesteryl ester analog	Lipid-droplet marker Monodansylpentane
	10 µg/g Bodipy FLC12	10 µg/g Bodipy 576/589 C11	10 µg/g Bodipy FLC12	10 µg/g Bodipy 576/589 C11	Monodansylpentane
	10 days	10 days	10 days	10 days	5 days

n/d = not described.

HCD feeding induces co-localization of lipids with neutrophils in the caudal vein [8].

In conclusion, our data show that wild-type zebrafish larvae are not as susceptible to HCD-induced accumulation of macrophage foam cells in caudal vein, as previously suggested. With this letter we aim to underscore the importance of clearly defining atherosclerosis when studying a new animal experimental model, as the application of different definitions can affect the interpretation of the results.

Declaration of competing interest

The authors declared they do not have anything to disclose regarding conflict of interest with respect to this manuscript.

References

- [1] Y. Yan, et al., The important role of apolipoprotein A-II in ezetimibe driven reduction of high cholesterol diet-induced atherosclerosis, *Atherosclerosis* 280 (2019) 99–108.
- [2] K. Stoletov, et al., Vascular lipid accumulation, lipoprotein oxidation, and macrophage lipid uptake in hypercholesterolemic zebrafish, *Circ. Res.* 104 (2009) 952–960.
- [3] E.A. Harvie, A. Huttenlocher, Neutrophils in host defense: new insights from zebrafish, *J. Leukoc. Biol.* 98 (2015) 523–537.
- [4] H.-D. Song, et al., Hematopoietic gene expression profile in zebrafish kidney marrow, *Proc. Natl. Acad. Sci. U. S. A* 101 (2004) 16240–16245.
- [5] F. Ellett, L. Pase, J.W. Hayman, A. Andrianopoulos, G.J. Lieschke, mpeg1 promoter transgenes direct macrophage-lineage expression in zebrafish, *Blood* 117 (2011) e49–e56.
- [6] Y. Yoon, et al., High cholesterol diet induces IL-1 β expression in adult but not larval zebrafish, *PLoS One* 8 (2013).
- [7] A. López Nadal, et al., Feed, microbiota, and gut immunity: using the zebrafish model to understand fish health, *Front. Immunol.* 11 (2020).
- [8] M.K. Bandaru, et al., Zebrafish larvae as a model system for systematic characterization of drugs and genes in dyslipidemia and atherosclerosis, *bioRxiv* (2019) 502674, <https://doi.org/10.1101/502674>.

R.A.F. Verwilligen*, J. Bussmann, M. Van Eck
 Division BioTherapeutics, Leiden Academic Centre for Drug Research
 (LACDR), Leiden University, the Netherlands
 E-mail address:
r.a.f.verwilligen@lacdr.leidenuniv.nl (R.A.F. Verwilligen).

* Corresponding author.