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Unraveling mucin type o-glycosylation signatures of colorectal cancer

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COLORECTAL CANCER CELL LINES SHOW STRIKING DIVERSITY OF THEIR O-GLYCOME REFLECTING THE CELLULAR DIFFERENTIATION PHENOTYPE

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Chapter 4



ABSTRACT

Alterations in protein glycosylation in colorectal cancer (CRC) have been extensively studied using cell lines as models. However, little is known about their *O*-glycome and the differences in glycan biosynthesis in different cell types. To provide a better understanding of the variation in *O*-glycosylation phenotypes and their association with other molecular features, an in-depth *O*-glycosylation analysis of 26 different CRC cell lines was performed. The released *O*-glycans were analyzed on porous graphitized carbon nano-liquid chromatography system coupled to a mass spectrometer via electrospray ionization (PGC-nano-LC-ESI-MS/MS) allowing isomeric separation as well as in-depth structural characterization. Associations between the observed glycan phenotypes with previously reported cell line transcriptome signatures were examined by canonical correlation analysis. Striking differences are observed between the *O*-glycomes of 26 CRC cell lines. Unsupervised principal component analysis reveals a separation between well differentiated colon-like and undifferentiated cell lines. Colon-like cell lines are characterized by a prevalence of I-branched and sialyl Lewis X/A epitope carrying glycans while most undifferentiated cell lines show absence of Lewis epitope expression resulting in dominance of truncated α 2-6-core sialylated glycans. Moreover, the expression of glycan signatures associates with the expression of glycosyltransferases that are involved in their biosynthesis, providing a deeper insight into the regulation of glycan biosynthesis in different cell types. This untargeted in-depth screening of cell line *O*-glycomes paves the way for future studies exploring the role of glycosylation in CRC development and drug response leading to discovery of novel targets for the development of anti-cancer antibodies.

INTRODUCTION

With over 18 million new cases worldwide in 2018, colorectal cancer (CRC) is the third most common cancer in the world¹. The disease is very heterogeneous with a high variability in patient prognosis and treatment response². There have been various attempts to classify CRC patients into clinically relevant groups^{2,3} using single genomic markers such as microsatellite instability (MSI), BRAF and KRAS mutations. While these markers can give insights into disease processes, they cannot provide a full understanding of the molecular pathology and prediction of patient outcome⁴. Therefore, the CRC subtyping consortium (CRCSC) has recently made a systematic comparison of gene mutation and gene expression of primary tumors from a large set of samples and defined four subtypes of CRC with different clinical and molecular markers⁴. Consensus molecular subtype (CMS)1 tumors show prevalence of MSI together with high immune infiltration in the tumor microenvironment, associated with a better patient prognosis³. In contrast, the mesenchymal CMS4 tumors are characterized by infiltration with cancer associated fibroblasts and upregulation of epithelial to mesenchymal transition (EMT) resulting in worse overall patient prognosis³. Both CMS2 and CMS3 tumors show strong epithelial differentiation signatures, with characteristic metabolic pathways dysregulation in the CMS3 group⁴. Although the proposed classification provides a deeper understanding of CRC and its differential molecular signatures, it is not yet clear which features will be relevant for accurate patient stratification. In order to design subtype specific therapeutic strategies, translation of the CMS classification to preclinical models is needed, to enable large-scale drug screenings.

Various genetic studies have confirmed that cancer cell lines recapitulate the molecular features of the tumors^{5,6} and the same has been confirmed for CMS⁷. Recently, profiling of 34 CRC cell lines revealed consistency at the gene, microRNA and protein levels, dominated by two distinct clusters. The colon-like cluster has high expression of gastrointestinal specific markers, while the second cluster contains undifferentiated cell lines showing upregulation of transforming growth factor (TGF)- β induced genes and EMT signatures⁸. These two groups significantly associate with CMS groups, where CMS1 and CMS4 cluster as undifferentiated, and CMS2 and CMS3 as colon-like⁸.

Next to genetic, metabolic and proteomic signatures, protein glycosylation is a major factor in colon differentiation and CRC development⁹. It has been shown that malignant transformation changes the glycosylation machinery of the cells, affecting the function of the oncogenic receptors that are involved in the control of cell proliferation and differentiation¹⁰. Moreover, glycan binding proteins, expressed by immune cells in the tumor microenvironment, respond to these changes, often resulting in an immunosuppressive response¹¹. Therefore, unravelling glycan-based interactions in cancer is instrumental for disclosure of molecular mechanisms underlying cancer biology.

Due to their continuous availability, cell lines are often used as models for studying glycosylation changes in cancer. Recently, the *N*-glycosylation of a set of CRC cell lines has been characterized, revealing association of antennary fucosylation with differentiation and Caudal Type Homeobox 1 (CDX1) expression^{12,13}. Another major class of colon glycans are mucin-type *O*-glycans, mainly carried by heavily glycosylated mucin proteins, which are the major components of mucus layer in the gastrointestinal tract. Mucin type *O*-linked glycosylation is initiated by the transfer of *N*-acetylgalactosamine (GalNAc) to Ser/Thr of both mucin and non-mucin glycoproteins which are shown to be altered in various cancers including CRC^{14,15}. Unfortunately, little is known about *O*-glycosylation of cell lines due to its complexity, the presence of multiple isomeric structures as well as the lack of enzymatic release methods, making it overall a challenging task¹⁴.

Here, we present an in depth structural analysis of *O*-glycosylation phenotypes of 26 CRC cell lines derived from both primary tumors and metastatic sites. We optimized a 96-well plate PVDF membrane based method¹² for preparation of released *O*-glycans from 500,000 cells via reductive β -elimination¹⁶. Released *O*-glycans were analyzed on a sensitive analytical platform, namely, porous graphitized carbon nano-liquid chromatography coupled to a tandem mass spectrometer (PGC nano-LC-ESI-MS/MS) using negative electrospray ionization. Major differences are observed between 26 analyzed CRC cell lines, revealing the diversity of the CRC cell line *O*-glycome. Moreover, associations are found between the observed glycome phenotypes and cell line gene expression as well as their differentiation.

Colorectal cancer cell lines show striking diversity of their *O*-glycome reflecting the cellular differentiation phenotype

MATERIALS AND METHODS

CELLS AND CELL CULTURE

Human CRC cell lines were obtained from the Department of Surgery of the Leiden University Medical Center (LUMC), Leiden, The Netherlands, as well as the Department of Pathology of the VU University Medical Center (VUmc), Amsterdam, The Netherlands. Further details are provided in **Supporting Information, S-1**.

O-GLYCAN RELEASE AND ANALYSIS

Lysed cell pellets containing 5×10^5 cells were loaded to the preconditioned PVDF membrane plate wells and denatured with guanidine hydrochloride and dithiothreitol (DTT) at 60 °C. After removing denaturation agent, *N*-glycans were released by PNGase F digestion overnight at 37 °C. Upon removal of *N*-glycans, 50 µL of 0.5 M sodium borohydride (NaBH₄) in 50 mM potassium hydroxide (KOH) was added to each well and incubated for 16 hrs at 50 °C for the release of *O*-glycans via reductive β-elimination. Desalting of the samples was performed using a cation exchange resin Dowex 50W X8 which was self-packed into 96-well filter plates. Desalted *O*-glycans were further purified via solid phase extraction by packing bulk sorbent carbograph slurry into 96-well filter plates. Analysis was performed using a PGC nano-LC-ESI-MS/MS platform. More details are provided in **Supporting Information, S-1**.

GLYCAN STRUCTURE ANALYSIS AND RELATIVE QUANTIFICATION

Identification of glycans was performed based on PGC retention time, known biosynthetic pathways, and manual inspection of fragmentation spectra following known MS/MS fragmentation pathways of *O*-glycan alditols in negative-ion mode^{17,18}. Glycan sequences and linkages were confirmed by the analysis of glycans upon α2-3 neuraminidase, α1-3/4 fucosidase, and β1-4 galactosidase digestion. Relative quantitation was performed on the total area of all *O*-glycans within one sample normalizing it to 100 %. MS/MS mass lists were exported from the DataAnalysis software for upload to Unicarb DR repository¹⁹. A more detailed description is provided in **Supporting Information, S-1**.

STATISTICAL ANALYSIS

An imputation of the minimum positive number (0.0001) was performed to enable use of the statistical tools sensitive to the missing values such as principal component analysis (PCA). Regularized canonical correlation analysis was performed using rcc

function as it is implemented in the “mixOmics” package²⁰. Data analysis and visualization was performed in “R” software.

RESULTS

HIGH THROUGHPUT AND ROBUST O-GLYCAN RELEASE FROM CELLS

In order to analyze the *O*-glycosylation of 26 CRC cell lines, we established a high throughput sample preparation in 96-well format (**Supporting Information, Figure S1**). The combination and optimization of two previously established protocols^{12,16} allowed sequential release of both *N*- and *O*-glycans from cell lysates in a higher throughput manner using 96-well plates. *O*-glycans were analyzed on PGC nano-LC-ESI-MS/MS platform which revealed 178 different *O*-glycan structures. Of these, 153 passed the quality control criteria and were included in the analysis (**Supporting Information, Figure S3** and **Supporting Information, Table 1: S3- S31**). To assess the technical and biological variation of *O*-glycan profiles for each cell line, the complexity of each sample was reduced by compiling the relative peak areas for all glycans to single mass spectrometry average compositions (MSAC)²¹ (**Supporting Information, Table 1: S1**) representing the normalized number of sugar residues and modifications per glycan molecule. Low technical variability of our workflow is illustrated by the close clustering of scores in the PCA model from two technical replicates of each cell line as well as the triplicate of *O*-glycans released from bovine fetuin (standard) (**Supporting Information, Figure S2**). In addition, the close clustering scores from the cell lines which were cultured and analyzed in three biological replicates (HT29, HCT116, SW480, SW620, and HCT8) revealed a low biological variability (marked as A, B and C in the respective PCA plot). The highest variability in glycosylation profiles was observed in cell lines SW480, HT29 and HCT116, which may be attributed to the fact that the replicates for these cell lines were prepared at different sites (VUmc and LUMC) as well as using different media (detailed description in **Supporting Information, S-1**).

HIGH DIVERSITY OF CRC CELL LINES O-GLYCOSYLATION PROFILES

The comparison of the 26 CRC cell lines revealed striking differences between the *O*-glycomes (**Supporting Information, Figure S3**). Many, yet undescribed, *O*-GalNAc linked glycans were detected varying in size from two up to 14 monosaccharide residues. An illustration of the diversity between cell lines is shown in **Figure 1**. The

Colorectal cancer cell lines show striking diversity of their O-glycome reflecting the cellular differentiation phenotype

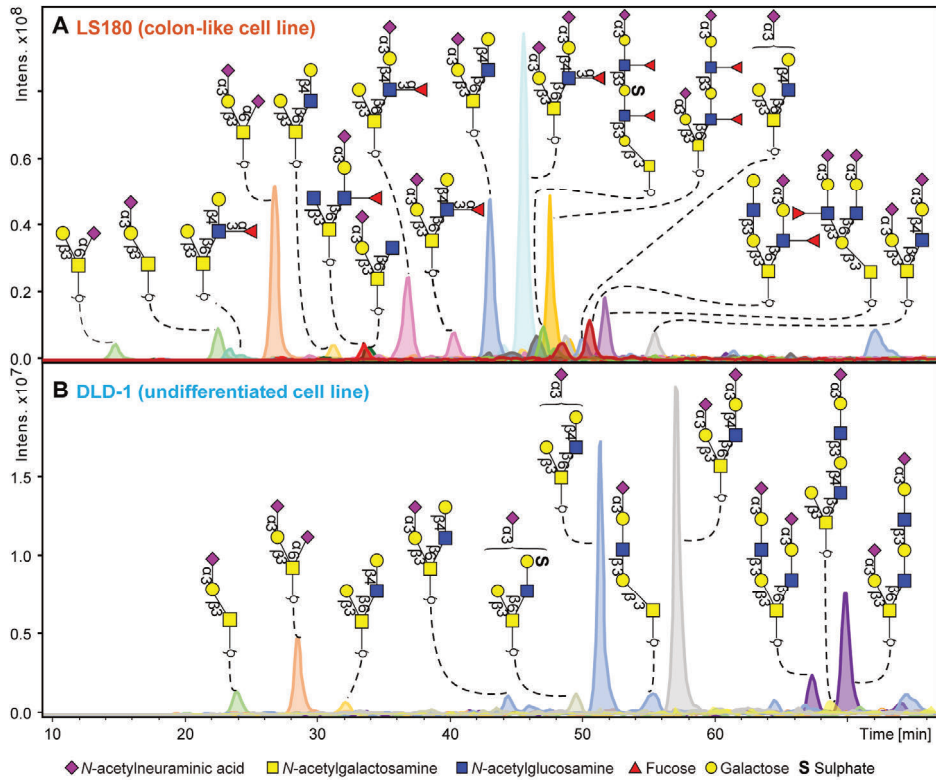


Figure 1. O-glycan profiles from two exemplary CRC cell lines. (A) O-glycan profile of the mucin secreting well differentiated (colon-like) cell line LS180 is characterised by high expression of sialyl Lewis X/A and Lewis X/A antigens. These antigens were found to be present on core 1, core 2 and core 4 glycan structures, both in linear and I-branched form. In contrast, panel (B) displays poorly differentiated human colon adenocarcinoma cell line DLD-1, showing an O-glycan profile which is dominated by core 2 sialylated glycans lacking any fucosylated antigens.

top panel shows the glycan profile of the mucin secreting cell line LS180 (human colon adenocarcinoma) which is characterized by high expression of sialyl Lewis X/A and Lewis X/A antigens. These antigens were found to be present on core 1, core 2 and core 4 glycan structures, both in linear and I-branched form. In contrast, DLD-1 (bottom panel), a poorly differentiated human colon adenocarcinoma cell line, showed an O-glycan profile which is dominated by core 2 sialylated glycans lacking any fucosylated antigens.

Only three glycans were present in all samples, albeit at highly varying abundances: disialyl-T antigen (NeuAc α 2-3Gal β 1-3(NeuAc α 2-6)GalNAcol), the α 2-3 sialyl-T antigen (NeuAc α 2-3Gal β 1-3GalNAcol), and the di-sialylated core 2 glycan (NeuAc α 2-

3Gal β 1-3(NeuAc α 2-3Gal β 1-4GlcNAc β 1-6)GalNAcol). The main core structures in most cell lines were core 2 followed by elongated structures of core 1. Core 3 structures were only detected in low amounts, while core 4 structures were expressed the most in HCT-15, LOVO and SW1116 cell lines (**Supporting Information, Figure S4**). Interestingly, all cell lines were dominated by sialylated glycan species (**Supporting Information, Figure S4**). *N*-acetylneuraminic acids (Neu5Ac) on *O*-glycans were mostly α 2-3-linked to a galactose residue. Moreover, α 2-6-linked sialylation was observed on the innermost GalNAc predominantly in the context of sialyl-T and disialyl-T antigens. Cell lines HT29 and WiDr showed the highest expression of α 2-3-sialylated *O*-glycans (**Supporting Information, Table 2: S2**). In addition, 42 *O*-glycans were found to contain sulphate modifications mainly expressed by SW948 and LS411N cell lines (**Supporting Information, Table 2: S2**).

A major advantage of PGC chromatography is the high separation power that enables to discriminate between glycan linkages and positional isomers^{17,18}. Identification of *O*-glycans was performed based on PGC retention time, described biosynthetic pathways and manual inspection of fragmentation spectra following known MS/MS fragmentation patterns of *O*-glycan alditols in negative-ion mode^{17,22}. All annotated

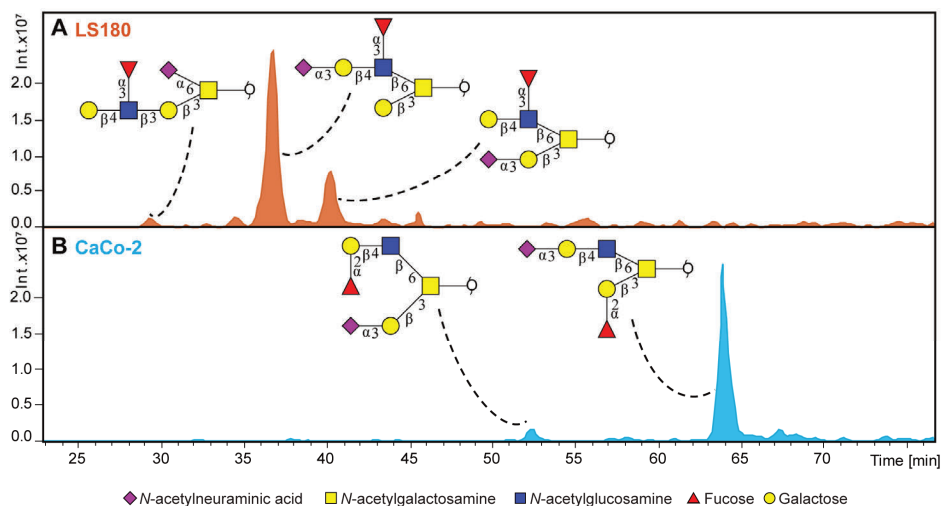


Figure 2. Chromatographic separation of *O*-glycan isomers at m/z 1186.40 [M-H] carrying different antigens. The extracted ion chromatograms show a different retention behaviour of Lewis antigen isomers in the (A) LS180 cell line. Namely, core 1 carrying Lewis X antigen with α 2-6-linked NeuAc linked to the core GalNAc (RT= 30.2 min), core 2 glycan carrying sialyl-Lewis X antigen on the 6 arm (RT= 37.2 min), and core 2 glycan carrying Lewis x antigen on the 6 arm (RT = 40.7 min). Additionally, the (B) CaCo-2 cell line illustrates the separation of core 2 blood group antigen H type 2 (RT = 52.4 min), and a core 2 mucin blood group antigen H type 3 (RT = 63.8 min).

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structures are listed in **Supporting Information, Table 2: S1**. MS/MS peak lists with glycan annotations per cell line are available via an online repository Unicarb DR¹⁹ (<http://unicarb-dr.biomedicine.gu.se/>). **Figure 2** shows the powerful chromatographic separation of five glycan isomers with the same composition H2N2F1S1 in both LS180 and CaCo-2 cell lines. Glycan sequences and linkages were confirmed by the analysis of glycans upon α 2-3 neuraminidase digestion, as well as additional combined α 1-3/4 fucosidase and β 1-4 galactosidase digestion as demonstrated by **Supporting Information, Figure S5-7**. With this approach we were able to identify the most abundant glycans representing more than 95% of the relative intensity for 13 cell lines, and over 90 % of the relative intensity for the remaining 13 cell lines.

GLYCAN TRAITS ARE ASSOCIATED WITH CELL LINE DIFFERENTIATION

To explore the specific *O*-glycan phenotypes in CRC more in-depth, the assigned glycans were relatively quantified and grouped based on glycan structural features such as core (1, 2, 3, or 4), I-branch (GlcNAc β 1-6Gal-R), α 2-3- or α 2-6-sialylation, Lewis X/A (Gal β 1-4/3(Fuca α 1-3/4)GlcNAc-R, sialyl Lewis x/a (NeuAca2-3Gal β 1-4/3(Fuca α 1-3/4)GlcNAc-R, (sialyl) dimeric Lewis X/A (NeuAca2-3Gal β 1-4/3(Fuca α 1-3/4)GlcNAc β 1-3Gal β 1-4/3(Fuca α 1-3/4)GalNAc-R, blood group A GalNAca1-3(Fuca α 1-2)Gal β 1-3/4-R, blood group B (Gal α 1-3(Fuca α 1-2)Gal β 1-3/4-R, blood group H type 3 (Fuca α 1-2Gal β 1-3(R-)GalNAcol) and blood group H type 2 (Fuca α 1-2Gal β 1-4GlcNAc-R) (**Supporting Information, Table 2: S2**). The structures that could not be unambiguously assigned were not included in the calculations of the structural features. The obtained CRC *O*-glycosylation signatures were further explored by PCA (**Figure 3**). Analysis of the score and loadings plot (**Figure 3A** and **B**, respectively) of the model shows that HT29 and its derivative WiDr cell line are positioned close to the center of the Hotelling circle and as such could be viewed as the examples of the average glycosylation profile. The cell lines derived from the same patient (HCT15, HCT8 and DLD-1) could also be considered as the examples of the average profile. Yet, a distance between them indicates the differences in their *O*-glycomes. This is supported by the **Figure 4** where DLD-1 cell line shows higher expression of I-branched glycans, and no expression of blood group antigen H, while the HCT8 and HCT15 express glycans carrying blood group H-antigens. Moreover, another two cell lines derived from the same patient, SW480 from primary tumor and SW620 from lymph node metastasis, do not cluster together on the score plot. Here, differences

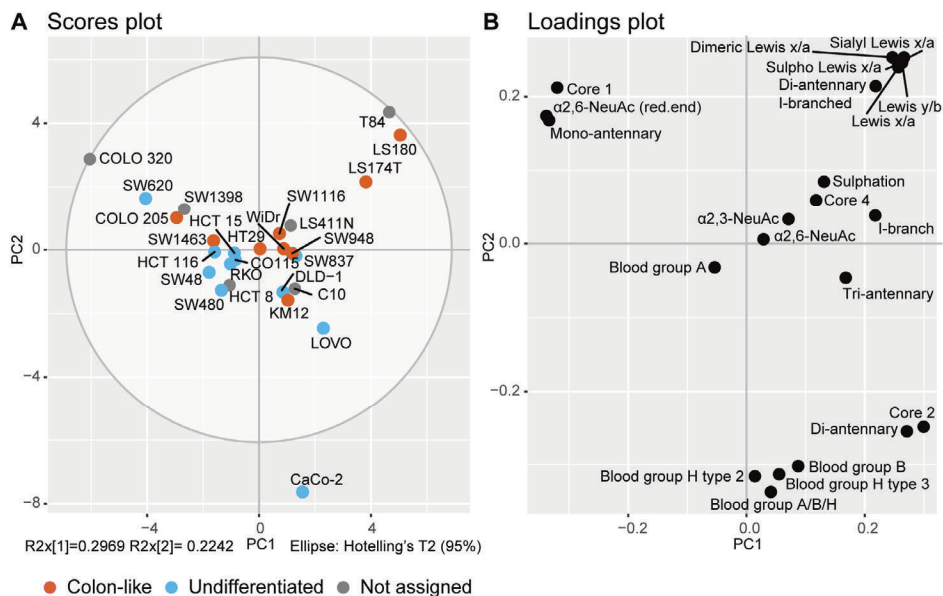


Figure 3. PCA based on relative abundance (%) of calculated structural glycan features. A separation between colon-like and undifferentiated cell lines is illustrated in the (A) PCA score plot of PC1 against PC2. (B) The PCA loading plot displays the variables that drive the separation in the PCA model. The top three of principal components explain 68.58 % of the variation within the data. Biological and technical replicates were averaged per cell line.

are found in the expression of blood group antigen H carrying glycans, which are present in higher levels in SW480 cell line, while SW620 does not express any fucosylated epitopes. Finally, the model reveals a similarity in the glycosylation profiles of the closely positioned cell line variants LS180 and LS174T on the outskirts of the score plot, as a result of very high expression of Lewis x/a antigens.

Differentiation phenotype of the cells (colon-like and undifferentiated)⁸ appears as a visible trend separating the first two components of the score plot (**Figure 3A**), while no association could be found with other cancer cell characteristics (e.g. MSI, oncogene mutations, tumor stages; data not shown). In order to decipher which structural glycan features drove the separation in the PCA model, we explored the variables in the PCA loadings plot (**Figure 3B**). Cell lines clustering in the left part of the score plot in **Figure 3A** (Colo320, SW620, SW1398, SW48, SW480, HCT116, RKO, Co115, and HCT15) have higher expression of core 1 and α2-6-linked Neu5Ac linked to the core (**Figure 3B**) which is also displayed in **Supporting Information, Figure S4**. Most of the cell lines in this cluster have been previously classified as

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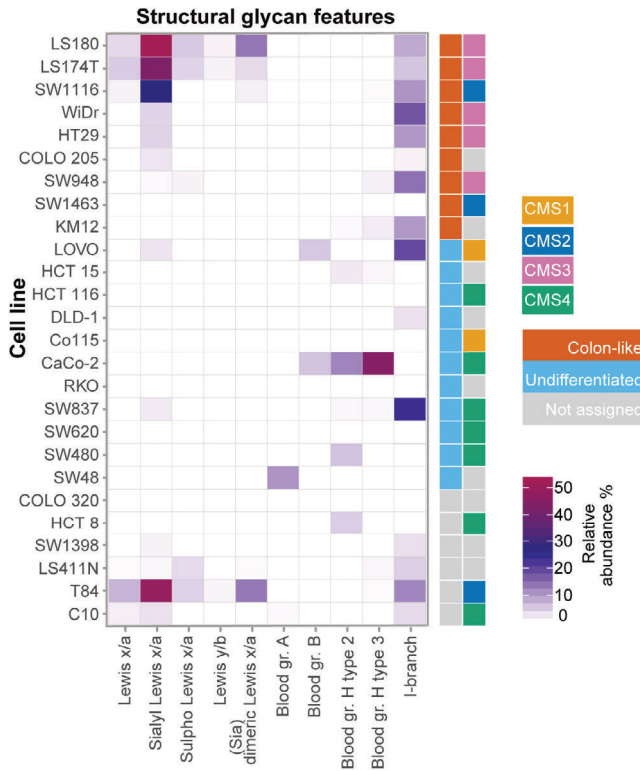


Figure 4. Relative abundance of structural glycan features per cell type. Geometrical tile of the relative abundance (%) of the calculated structural glycan features (x-axis) and cell line type (y-axis). Different classifications of the cell lines are displayed with colour codes based on gene expression (colon-like in red and undifferentiated in light blue) and consensus molecular subtypes (CMS1 in yellow, CMS2 in dark blue, CMS3 in pink, and CMS4 in green). Non-assigned cell lines were marked in grey for the gene expression as well as for the CMS status.

undifferentiated based on low expression of gastrointestinal specific genes⁸. On the other hand, mucin secreting cell lines LS180, LS174T²³ and T84²⁴ are characterized by abundant and diverse glycosylation with very high expression of Lewis antigens clustering in the upper right part of the PCA score plot. In contrast, CaCo-2 cell line displays a different phenotype, rich in blood group H type 2 and type 3 antigens, as well as Sd^a (GalNAc β 1-4(NeuAca2-3)Gal β -) and Cad (GalNAc β 1-4(NeuAca2-3)Gal β 1-3(NeuAca2-6)-) antigens. Colon-like cell lines show higher expression of Lewis-like antigens, predominantly sialyl Lewis X/A epitopes (**Figure 4**) which could be a trait of the CMS3 metabolic subtype (represented by LS180, LS174T, HT29, WiDr and SW948). The I-antigen branching is found more often in the colon-like cell lines

(**Figure 5**) such as WiDr, SW948, SW1116 and HT29. However, one of the undifferentiated cell lines, SW837, shows the highest expression of I-branched glycans. Most of CMS4 cell lines are characterized by a high level of overall α 2-3-sialylation, no expression of Lewis antigens (with exception of SW837 and C10 cell lines), together with relatively higher expression of blood group antigen type 2.

GLYCOMIC SIGNATURES ASSOCIATE WITH CELL LINE TRANSCRIPTOMICS

In order to understand which genes are underlying the glycosylation phenotypes observed in this study, we performed a canonical correlation analysis using mRNA expression data from an online available dataset⁸. Primarily, we selected genes involved in the biosynthesis of *O*-glycans which showed the largest fold change in expression ($\log_e > 0.5$) when comparing colon-like and undifferentiated cell lines with high statistical significance (Bonferroni corrected p -value < 0.05), displayed in the volcano plot (**Supporting Information, Figure S8**). Colon-like cells show higher expression of the following genes encoding for glycosyltransferases (GTs): *B3GNT3*, *FUT2*, *FUT3*, *FUT6*, *GALNT3*, *GCNT3*, and *ST6GALNAC1*. In contrast, the undifferentiated cell lines show a slightly higher expression of *ST3GAL3*. The correlations of glycan structural features with expression of the selected genes are illustrated in the clustered image map of the canonical model (**Figure 5**). A moderate correlation between the *FUT3* and Lewis x/a ($r = 0.40$) as well as between *FUT6* and sialyl Lewis x/a epitopes ($r = 0.36$) is observed. *ST6GALNAC1* gene expression also correlates with the expression of sialyl Lewis X/A ($r = 0.38$) and Lewis x/a epitopes ($r = 0.40$). Additionally, *GCNT3* shows a strong correlation with α 2-3-sialylation ($r = 0.70$) and I-branching ($r = 0.66$) reflecting the expression of sialylated I-branched diantennary and triantennary *O*-glycans. *GALNT3* encoding for *O*-glycosylation initiating enzyme shows positive correlation with core 2 ($r = 0.43$), I-branching ($r = 0.44$), and α 2-3-sialylation ($r = 0.38$), as well as a negative correlation with α 2-6-core sialylation ($r = -0.50$), and core 1 glycans ($r = -0.45$). *B3GNT3*, a gene encoding for β -1-3-*N*-acetylglucosaminyltransferase 3 and involved in the biosynthesis of poly-*N*-acetyllactosamine chains, is found to be negatively correlated with the expression of blood group A, B or H (blood group A/B/H) antigens ($r = -0.57$), core 2 glycans ($r = -0.45$) and shows a positive correlation with core 1 glycans ($r = 0.40$). *ST3GAL3* is the only gene involved in the *O*-glycan biosynthesis which shows higher expression in the

Colorectal cancer cell lines show striking diversity of their *O*-glycome reflecting the cellular differentiation phenotype undifferentiated cell lines, displaying correlation with core 1 ($r = 0.45$) and $\alpha 2$ -6-sialylated glycan expression ($r = 0.46$).

TRANSCRIPTIONAL REGULATION OF GLYCOSYLTRANSFERASE EXPRESSION

To gain more insight into the regulation of *O*-glycosylation, we also examined the associations of glycan epitopes with transcription factors which showed the highest fold change in expression ($\log_e > 0.5$) when comparing colon-like and undifferentiated cell lines⁸ with high statistical significance (Bonferroni corrected p -value < 0.05) (**Supplementary Figure S8**). The colon-like cells show significantly higher expression of the following transcription factors: *CDX1*, *ETS2*, *HNF1A*, *HNF4A*, *MECOM* and *MYB*. In contrast, the undifferentiated cell lines showed elevated expression levels of the following transcription factors: *MLLT10*, *MSX1*, *SIX4*, *ZNF286A* and *ZNF286B*. The clustered image map of the canonical model (**Figure 5**) displays the moderate positive correlation between the expression of Lewis antigens and transcription factors *CDX1* and *ETS2*. Relatively strong correlation is seen for *ETS2* with $\alpha 2$ -3-sialylation ($r = 0.57$) and I-branching ($r = 0.51$). The *MYB* gene, which is highly expressed in colon-like CRC cells, showed a strong positive correlation with the expression of core 1 ($r = 0.59$) and $\alpha 2$ -6-sialylated glycans ($r = 0.57$) and a strong negative correlation with blood group A, B or H (Blood group A/B/H) carrying structures ($r = -0.62$). On the other hand, two transcription factors *ZNF286A* and *ZNF286B*, expressed in undifferentiated cell lines, show high negative correlation with $\alpha 2$ -3-sialylation ($r = -0.63$, and $r = -0.60$ respectively) and I-branching ($r = -0.55$ and $r = -0.53$ respectively), together with a positive correlation with blood group antigen A, B or H carrying glycans ($r = 0.58$ and $r = 0.53$, respectively). To identify the possible transcriptional regulation of specific GTs, we tested the associations between transcription factor expression and GT expression (**Supplementary Figure S9**). In contrast with the rather weak correlations of *MYB* expression and Lewis antigen expression, *MYB* expression does show correlation with *FUT3* and *B3GNT3* genes. We also observe strong negative correlations between expression of *ZNF286A* and *ZNF286B* genes with the expression of *GCNT3* and *B3GNT3* genes involved in the elongation and branching of *O*-glycan molecules next to a positive correlation with the expression of *ST3GAL3*.

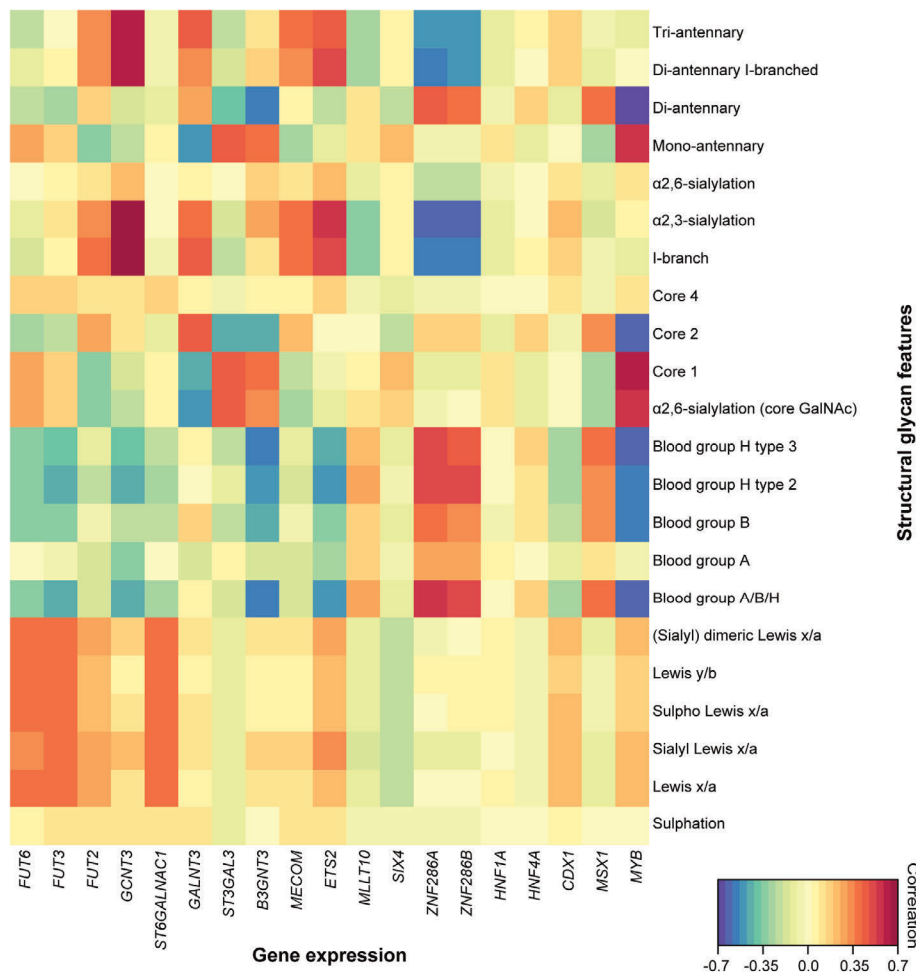


Figure 5. Associations of structural glycan features with gene expression. The clustered image map of the canonical model displaying associations of glycan structural features with gene expression of relevant GTs and transcription factors differentially expressed in colon-like vs undifferentiated cell lines

DISCUSSION

In the present study, we investigated the *O*-glycosylation phenotypes of 26 CRC cell lines derived from both primary tumors and metastatic sites, revealing pronounced differences between the cell lines. The PGC nano-LC-MS/MS platform allowed separation of isomeric *O*-glycan species which were structurally elucidated relying on negative mode tandem mass spectra and exoglycosidase treatment. Using this approach, we were able to create a detailed CRC cell line *O*-glycan MS/MS spectral

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library, which will be available via <https://unicarb-dr.biomedicine.gu.se/>, serving as an important resource leading towards automated *O*-glycan identification via spectral matching tools.

An exploratory, qualitative analysis of the entire pool of our data clearly shows a difference in *O*-glycome profiles between the colon-like well differentiated cell lines and undifferentiated ones. A proposed model explaining the differences in glycosylation and biosynthesis in different cell types is displayed in **Figure 6**. For instance, the well-differentiated cell lines show an overall higher expression of Lewis antigens and I-branched glycans, while the undifferentiated cell lines show a higher abundance of glycans carrying an α 2-6-linked NeuAc to the core GalNAc. Few exceptions could also be observed. For instance, Colo205 a colon-like but also a metastatic cell line²⁵, clusters together with the undifferentiated cell lines expressing higher relative amounts of core 1 α 2-6-sialylated glycan species. Similarly, the rectal adenocarcinoma SW1463 cell line²³ does not show expression of Lewis antigens characteristic for other mucus producing cell lines. Such observations can be valuable on their own, but offer only a limited space for a functional interpretation of the data. Taking advantage of the existing pool of published transcriptomics data on the selected cell lines, we attempted to generate regulational hypotheses using the correlations between the *O*-glycan profiles and the transcripts most strongly associated with the differentiation status of the studied cell lines. Associations were found between gene expression and cell line differentiation based on the *O*-glycome profiles of 26 CRC cell lines. Here, Lewis antigen expression was found to be the most abundant in the mucin secreting cell lines LS174T²⁴, LS180 and T84²⁶ correlating with the expression of genes *FUT3* and *FUT6* encoding for the GTs involved in the biosynthesis of the Lewis antigens. Expression of Lewis antigens was also associated with *ST6GALNAC1* gene expression encoding for an α 2-6-sialyltransferase which acts on the core GalNAc residue. Although we have detected sialyl-Tn antigen in the mucin secreting cell lines (LS180 and LS174T), we could not quantify mono- and disaccharide *O*-glycan alditols in a reliable manner due to limitations of the method. Previously, sialyl-Tn antigen was found to be highly expressed in LS174T cell line, and also showed correlation with upregulation of *ST6GALNAC1*²⁷. In concordance with previous studies, demonstrating an association of CDX-1 transcription factor with the expression of GTs *FUT3* and *FUT6* and expression of multifucosylated *N*-glycans in

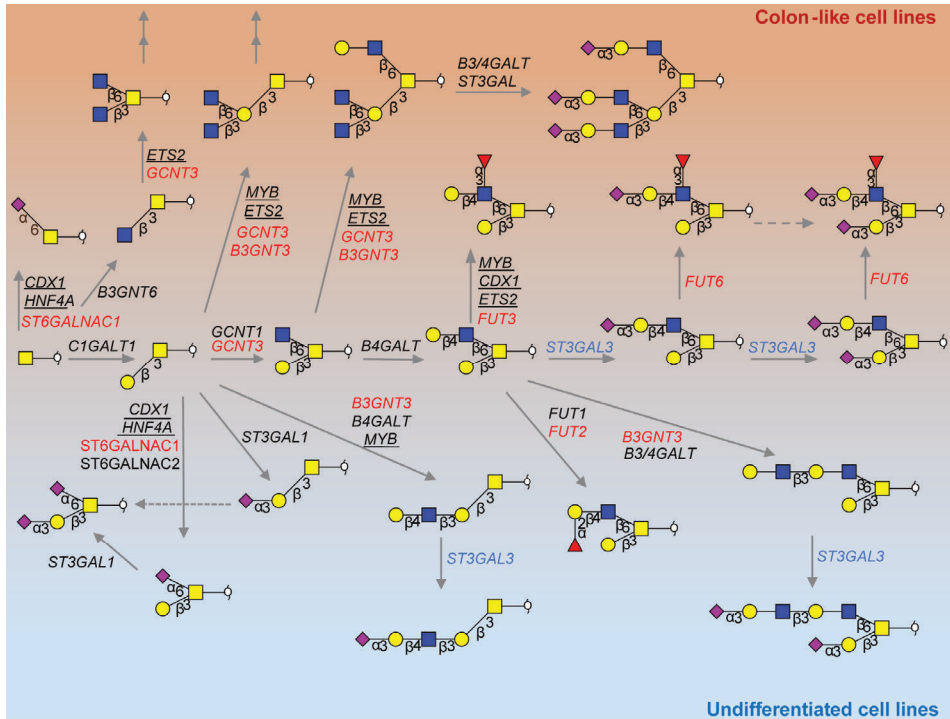


Figure 6. Proposed model explaining the differences in glycosylation between colon-like and undifferentiated cell lines. The most abundant structures in colon-like and undifferentiated cell lines and their biosynthesis pathways with genes encoding for the GTs involved in the biosynthesis. GTs up-regulated in colon-like cell lines are marked in red, whereas genes upregulated in undifferentiated cell lines are marked in blue. Doubled arrows indicate structures that can be elongated further by action of different GTs. Transcription factors showing correlation with the expression of GTs are underlined. Please note that other GTs could also be involved in the biosynthesis of the displayed glycans

cell lines^{12,13}, a correlation between Lewis antigen expressing cell lines and *CDX1* expression was observed. Moreover, some cell lines with high expression of α 2-3-sialylated and multifucosylated *N*-glycans¹² (HT29, WiDr, T84, and LOVO) also revealed a high expression of *O*-glycans carrying sialyl Lewis X/A antigens. Strikingly, the cell lines with the highest expression of sialylated Lewis X/A type antigens on *O*-glycans (LS180, LS174T and SW1116) express mainly non-sialylated Lewis antigens on *N*-glycans¹². These findings emphasize the importance of studying glycosylation in a glycan type-specific manner, as they can exhibit different glycan motifs and may convey different functions.

In contrast to high Lewis antigen and I-branching signatures of colon-like cell lines, the undifferentiated cell lines show a high abundance of core 1 glycans carrying an α 2-6-

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linked Neu5Ac to the core GalNAc. This signature is mainly reflecting the high relative abundance of the ubiquitous disialyl-T antigen compared to a lower abundance of elongated and branched *O*-glycans. The addition of Neu5Ac in the α 2-6-position of the Tn or T antigen will prevent formation of different core structures and further elongation of cores with different structural epitopes²⁸. This may explain the absence of core 3 structures in cell lines, which similarly to inflamed colon tissue²¹, show low abundance of core 3 structures compared to normal colon tissue. Some reports have shown that upregulation of *ST3GAL3*, *ST6GALNAC1* and *ST6GALNAC2* could be associated with invasion to lymph nodes and poor patient prognosis²⁹. Furthermore, enhanced sialylation of tumour cells can engage inhibitory sialic acid binding lectin-Siglec receptors on natural killer (NK) cells, providing an immune evasion mechanism¹¹. The competition between different GTs for the core GalNAc residue can explain the strong inverse correlations between core 2 and α 2-6-sialylation observed in our data (**Figure 6**). Competition involved in core 2 glycan biosynthesis such as *GCNT1*, *GCNT3* and *ST6GALNAC1* have been described extensively in the literature^{30,31}. *GCNT3* gene encodes a mucin type β 1,6-*N*-acetylglucosamine transferase, which adds a *N*-acetylglucosamine (GlcNAc) to the core GalNAc forming core 2 or core 4 structures. Importantly, *GCNT3* also has I-antigen biosynthetic activity, by adding the residue to the terminal galactose forming another branching point. *GCNT3* expression has been associated with reduction of proliferation and invasion as well as with higher sensitivity to the chemotherapeutic 5-Fluorouracil³². Low *GCNT3* expression has been proposed as prognostic marker that could be used to identify early-stage colon cancer patients at high risk of relapse³³. It has been demonstrated that the expression of *GCNT3* is higher in the colon-like cell lines⁸, showing a strong correlation with the expression of I-branched glycans as well as with α 2-3 sialylation in our data. However, one of the undifferentiated cell lines SW837, derived from the rectum³⁴, shows the highest expression of I-branched glycans.

Our results also show expression of histo-blood group ABH antigens carried by *O*-glycans mainly from undifferentiated cell lines. Apart from the erythrocytes, the ABH antigens can be expressed also in the gastrointestinal tract and secretions, where *Se-FUT* enzyme (*FUT2*) is responsible for their biosynthesis³⁵. However, several reports have demonstrated the expression of blood group ABH antigens in colon cancers from non-secretors with inactive enzyme^{36,37}. Cell lines showing expression of blood group

H antigens such as CaCo-2, HCT15, HCT8, SW948, SW1116, and SW837 were derived from tumors of blood group O secretor positive individuals³⁸. We have also observed expression of blood group B antigens by CaCo-2 cell line, and this incompatible expression in the gut of blood group O individuals has been previously observed in colon cancer³⁹. LOVO cell line, derived from a B blood type individual, expressed the blood group B carrying glycans, however the SW48 (AB blood group individual) shows expression of only blood group A glycans. Cell lines SW480, SW620 and HT29 derived from blood group A individuals do not show expression of the A antigen carried by O-glycans although these cell lines have shown a substantial A transferase activity⁴⁰. HCT8 cell line derived from the ileocecal colon shows higher expression of blood group antigens H type 2, compared to HCT15 derived from sigmoid colon of the same individual, which is in concordance with the reports demonstrating a decreasing gradient of ABH antigen expression towards the distal part of the colon⁴¹. However, more research is needed to see if CRC cell line glycosylation is comparable with blood group antigen distribution in the healthy colon. We have also observed that blood group antigen H types 2 and 3 were more abundant than H type 1 antigen in CRC cell lines. Similarly, antibody staining studies showed region specific expression of mainly H type 1 blood group antigen H in healthy colon tissue, in contrast to colon cancer tissues aberrantly expressing blood group antigens H type 2 and 3 associated with tumor progression in the distal colon and rectum³⁶.

The majority of the cell line O-glycans had α 2-3-linked Neu5Ac at the terminal end of the glycan. Overall sulphation was found to be much less abundant than sialylation, showing highest expression in SW948 and LS411N cell lines. As compared with normal colon tissue⁴¹, cancer associated O-glycans often show increased sialylation and less sulphation⁴². Furthermore, we detected structures carrying Sd^a/Cad epitopes only in one cell line, namely, CaCo-2, supporting other reports from the literature⁴³. Structures terminated by Sd^a/Cad epitopes have been described before as characteristic for normal colon mucin tissue^{44,45}. Our results also show a dominance of core 2 and core 1 O-linked glycans, and a very low abundance of core 3 and core 4 structures. Core 3 and core 4 are known to be characteristic for the healthy colon mucins⁴⁶ and most of the cell lines did not show high expression of those structures. Moreover, core 3 synthase has been reported to be downregulated in colon cancer⁴⁷, with a loss of activity in many CRC cell lines⁴⁸. Additionally, a decrease in core 3

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structures together with a concomitant increase in core 1 structures has been observed when moving from healthy tissue to tumour regions¹⁵. Previously, no core 3 structures could be detected in the glycan profiles of five CRC cell lines, in contrast to substantial expression in CRC tissues²⁷. We can hypothesize that due to the lack of precursor core 3 structures, *GCNT3*, which is highly expressed in many cell lines, exhibits the core 2 and I-branching activity, resulting in predominance of core 2 and I-branched structures in the cell lines.

To provide a better understanding of regulation of *O*-glycosylation, we also examined the associations of glycan epitopes with transcription factor expression, and generated hypotheses by association analysis. *ETS2* transcription factor expression shows associations with I-branched and α 2-3-sialylated glycan expression (**Figure 5**) supported by its correlation with the expression of the *GCNT3* gene (**Supplementary Figure S9**). *ETS2* is a Wnt pathway target gene, whose inactivation leads to increased crypt cell proliferation⁴⁹. It has previously been associated with *MGAT2* promoter activation for the biosynthesis of complex type *N*-glycans, however its potential regulation of *O*-glycan biosynthesis is still unknown. Oncogenic transcription factor c-Myb, important for both cell proliferation and cell differentiation⁵⁰, is also involved in the Wnt pathway activation, and shows a positive correlation with the expression of core 1 and α 2-6-core sialylated glycans (**Figure 5**). Activated β -catenin and *MYB* induce upregulation of *MYC* promoter, and c-Myc transcription factor expression⁵¹. It has been found that *ST3GAL1, 2 and 4* are transcriptionally upregulated by c-Myc⁵², which might contribute to the higher expression of disialyl and sialyl-T antigens, together with a relatively lower expression of core 2 structures (**Figure 5**). The very strong negative correlation with blood group antigen H expression in the samples could be explained by the strong correlation of *MYB* gene with *B3GNT3* gene and is in direct competition with the biosynthesis of blood group antigens³¹ (**Figure 6**). Another strong positive correlation was found between the expression of *ZNF286A* and *ZNF286B* genes and the expression of blood group antigens. This might be a consequence of the strong negative correlations with both the expression of *B3GNT3* and *GCNT3* genes, which are involved in the elongation and branching of *O*-glycan structures. Additionally, a positive correlation with the expression of *ST3GAL3*, encoding the α 2-3-sialyltransferase, terminating the elongation of glycan structures

with Neu5Ac residues was found. Further studies need to be performed to confirm these hypotheses.

Although it has been demonstrated before that cell lines can be good representatives of the tumors based on gene mutations, gene expression, and protein expression⁶⁻⁸, a large scale study comparing glycosylation of cell lines and tumor tissues is necessary to evaluate whether the cell lines can be used as glycobiological model systems. A small scale study, which compared *N*- and *O*-glycomes derived from CRC cell lines and tumors, revealed that some cell lines are not representing the *O*-glycosylation of the tumor tissues, showing lack of expression of sialyl- Tn and core 3 disialyl Lewis X hexasaccharide²⁷, previously associated with malignant transformation⁵³. Additionally, we found notable differences between the *O*-glycomes of cell lines cultured at different sites in different culturing conditions, therefore the impact of these variables also needs to be considered.

4 The presented mass spectrometric approach revealing the complete repertoire of glycans expressed by a specific cell type is a valuable resource for understanding glycosylation related changes in cancer. While the CRC cell lines have shown enormous diversity of individual glycan structures, their structural features could be associated with the cell differentiation. Moreover, glyco-phenotypes were found to be associated with the expression of specific glycosyltransferases involved in their biosynthesis, providing more insight into the regulation of glycan biosynthesis in different cell types. Finally, this untargeted in-depth screening of cell line glycome phenotypes will provide an important resource for future studies exploring the role of cell glycosylation in CRC progression and drug response leading to discovery of novel targets for the development of anti-cancer antibodies.

SUPPORTING INFORMATION

Supporting information is available free of charge via doi.org/10.1007/s00018-020-03504-z

DATA AVAILABILITY

The raw mass spectrometric data files that support the findings of this study are available in GlycoPOST in mzXML format, with the identifier GPST000035, accessible via the following link:

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<https://glycopost.glycosmos.org/preview/863090025d7bddcbd8bcf>. The MS/MS spectra of glycan structures are available in UnicarbDR repository, accessible via the following link <https://unicarb-dr.biomedicine.gu.se/references>.

AUTHOR CONTRIBUTIONS

K.M. performed the experiments. K.M., S.H., T.Z., K.S. and M.W. conceptually designed the work. K.M., G.L., O.M. and M.W. wrote the manuscript. N.C. and C.J. assisted with data interpretation. All authors read and approved the final manuscript.

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REFERENCES

1. Ferlay, J. *et al.* Cancer incidence and mortality worldwide: Sources, methods and major patterns in GLOBOCAN 2012. *Int. J. Cancer* **136**, E359–E386 (2015).
2. De Sousa E Melo, F. *et al.* Poor-prognosis colon cancer is defined by a molecularly distinct subtype and develops from serrated precursor lesions. *Nat. Med.* **19**, 614–618 (2013).
3. Becht, E. *et al.* Immune and stromal classification of Colorectal cancer is associated with molecular subtypes and relevant for precision immunotherapy. *Clin. Cancer Res.* **22**, 4057–4066 (2016).
4. Guinney, J. *et al.* The consensus molecular subtypes of colorectal cancer. *Nat. Med.* **21**, 1350–1356 (2015).
5. Linnekamp, J. F. *et al.* Consensus molecular subtypes of colorectal cancer are recapitulated in in vitro and in vivo models. *Cell Death Differ.* **25**, 616–633 (2018).
6. Mouradov, D. *et al.* Colorectal cancer cell lines are representative models of the main molecular subtypes of primary cancer. *Cancer Res.* **74**, 3238–3247 (2014).
7. Sveen, A. *et al.* Colorectal cancer consensus molecular subtypes translated to preclinical models uncover potentially targetable cancer cell dependencies. *Clin. Cancer Res.* **24**, 794–806 (2018).
8. Berg, K. C. G. *et al.* Multi-omics of 34 colorectal cancer cell lines - a resource for biomedical studies. *Mol. Cancer* **16**, 116 (2017).
9. Holst, S., Wuhler, M. & Rombouts, Y. *Glycosylation characteristics of colorectal cancer. Advances in Cancer Research* **126**, (Elsevier Inc., 2015).

10. Pinho, S. S. & Reis, C. A. Glycosylation in cancer: mechanisms and clinical implications. *Nat. Rev. Cancer* **15**, 540–555 (2015).
11. Rodríguez, E., Schetters, S. T. T. & Van Kooyk, Y. The tumour glyco-code as a novel immune checkpoint for immunotherapy. *Nature Reviews Immunology* **18**, 204–211 (2018).
12. Holst, S. *et al.* N-glycosylation Profiling of Colorectal Cancer Cell Lines Reveals Association of Fucosylation with Differentiation and Caudal Type Homeobox 1 (CDX1)/Villin mRNA Expression. *Mol. Cell. Proteomics* **15**, 124–140 (2016).
13. Holst, S. *et al.* N-Glycomic and Transcriptomic Changes Associated with CDX1 mRNA Expression in Colorectal Cancer Cell Lines. *Cells* **8**, 273 (2019).
14. Ju, T., Aryal, R. P., Kudelka, M. R., Wang, Y. & Cummings, R. D. The Cosmc connection to the Tn antigen in cancer. *Cancer Biomarkers* **14**, 63–81 (2014).
15. Mihalache, A. *et al.* Structural Characterization of Mucin O-Glycosylation May Provide Important Information to Help Prevent Colorectal Tumor Recurrence. *Front. Oncol.* **5**, 217 (2015).
16. Jensen, P. H., Karlsson, N. G., Kolarich, D. & Packer, N. H. Structural analysis of N- and O-glycans released from glycoproteins. *Nat. Protoc.* **7**, 1299–1310 (2012).
17. Karlsson, N. G., Schulz, B. L. & Packer, N. H. Structural determination of neutral O-linked oligosaccharide alditols by negative ion LC-electrospray-MSn. *J. Am. Soc. Mass Spectrom.* **15**, 659–672 (2004).
18. Karlsson, N. G. *et al.* Negative ion graphitised carbon nano-liquid chromatography/mass spectrometry increases sensitivity for glycoprotein oligosaccharide analysis. *Rapid Commun. Mass Spectrom.* **18**, 2282–2292 (2004).
19. Rojas-Macias, M. A. *et al.* Towards a standardized bioinformatics infrastructure for N- and O-glycomics. *Nat. Commun.* **10**, 3275 (2019).
20. Rohart, F., Gautier, B., Singh, A. & Lê Cao, K.-A. mixOmics: An R package for 'omics feature selection and multiple data integration. *PLoS Comput. Biol.* **13**, e1005752 (2017).
21. Hayes, C. A., Nemes, S. & Karlsson, N. G. Statistical analysis of glycosylation profiles to compare tissue type and inflammatory disease state. *Bioinformatics* **28**, 1669–1676 (2012).
22. Seymour, J. L., Costello, C. E. & Zaia, J. The Influence of Sialylation on Glycan Negative Ion Dissociation and Energetics. *J. Am. Soc. Mass Spectrom.* **17**, 844–854 (2006).
23. Kerbel, R. S., Pross, H. F. & Leibovitz, A. Analysis of established human carcinoma cell lines for lymphoreticular-associated membrane receptors. *Int. J. Cancer* **20**, 673–679 (1977).
24. Van Klinken, B. J. W. *et al.* The human intestinal cell lines Caco-2 and LS174T as models to study cel-type specific mucin expression. in *Glycoconjugate Journal* **13**, 757–768 (1996).
25. Semple, T. U., Quinn, L. A., Woods, L. K. & Moore, G. E. Tumor and lymphoid cell lines from a patient with carcinoma of the colon for a cytotoxicity model. *Cancer Res.* **38**, 1345–55 (1978).
26. McCool, D. J., Marcon, M. A., Forstner, J. F. & Forstner, G. G. The T84 human colonic adenocarcinoma cell line produces mucin in culture and releases it in response to various secretagogues. *Biochem. J.* **267**, 491–500 (1990).

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27. Chik, J. H. L. *et al.* Comprehensive glycomics comparison between colon cancer cell cultures and tumours: Implications for biomarker studies. *J. Proteomics* **108**, 146–162 (2014).
28. Brockhausen, I., Yang, J., Lehotay, M., Ogata, S. & Itzkowitz, S. Pathways of mucin O-glycosylation in normal and malignant rat colonic epithelial cells reveal a mechanism for cancer-associated Sialyl-Tn antigen expression. *Biol. Chem.* **382**, 219–232 (2001).
29. Schneider, F. *et al.* Overexpression of sialyltransferase CMP-sialic acid:Galbeta1,3GalNAc-R alpha6-Sialyltransferase is related to poor patient survival in human colorectal carcinomas. *Cancer Res.* **61**, 4605–11 (2001).
30. Lo, C. Y. *et al.* Competition between core-2 GlcNAc-transferase and ST6GalNAc-transferase regulates the synthesis of the leukocyte selectin ligand on human P-selectin glycoprotein ligand-1. *J. Biol. Chem.* **288**, 13974–13987 (2013).
31. Brockhausen, I. Pathways of O-glycan biosynthesis in cancer cells. *Biochim. Biophys. Acta - Gen. Subj.* **1473**, 67–95 (1999).
32. Fernández, L. P. *et al.* The role of glycosyltransferase enzyme GCNT3 in colon and ovarian cancer prognosis and chemoresistance. *Sci. Rep.* **8**, 8485 (2018).
33. González-Vallinas, M. *et al.* Clinical relevance of the differential expression of the glycosyltransferase gene GCNT3 in colon cancer. *Eur. J. Cancer* **51**, 1–8 (2015).
34. Leibovitz, A. *et al.* Human Colorectal Adenocarcinoma Cell Lines. *Cancer Res.* **36**, 4562–4569 (1976).
35. Dotz, V. & Wuhrer, M. Histo-blood group glycans in the context of personalized medicine. *Biochim. Biophys. Acta - Gen. Subj.* **1860**, 1596–1607 (2016).
36. Fujitani, N. *et al.* Expression of H type 1 antigen of ABO histo-blood group in normal colon and aberrant expressions of H type 2 and H type 3/4 antigens in colon cancer. *Glycoconj. J.* **17**, 331–338 (2000).
37. Orntoft, T. F., Greenwell, P., Clausen, H. & Watkins, W. M. Regulation of the oncodevelopmental expression of type 1 chain ABH and Lewisb blood group antigens in human colon by α -2-L-fucosylation. *Gut* **32**, 287–293 (1991).
38. Dahiya, R., Itzkowitz, S. H., Byrd, J. C. & Kim, Y. S. ABH Blood Group Antigen Expression, Synthesis, and Degradation in Human Colonic Adenocarcinoma Cell Lines. *Cancer Res.* **49**, 4550–4556 (1989).
39. Clausen, H., Hakomori, S., Graem, N. & Dabelsteen, E. Incompatible A antigen expressed in tumors of blood group O individuals: immunochemical, immunohistologic, and enzymatic characterization. *J. Immunol.* **136**, 326–30 (1986).
40. Dahiya, R., Itzkowitz, S. H., Byrd, J. C. & Kim, Y. S. ABH blood group antigen expression, synthesis, and degradation in human colonic adenocarcinoma cell lines. *Cancer Res.* **49**, 4550–6 (1989).
41. Robbe, C. *et al.* Evidence of regio-specific glycosylation in human intestinal mucins: Presence of an acidic gradient along the intestinal tract. *J. Biol. Chem.* **278**, 46337–46348 (2003).
42. Brockhausen, I. Mucin-type O-glycans in human colon and breast cancer: Glycodynamics and functions. *EMBO Rep.* **7**, 599–604 (2006).

43. Malagolini, N., Dall'Olio, F. & Serafini-Cessi, F. UDP-Ga1NAc:NeuAc α 2,3Gal β -R (GalNAc to Gal) β 1,4-N-acetyl-galactosaminyltransferase responsible for the Sda specificity in human colon carcinoma CaCo-2 cell line. *Biochem. Biophys. Res. Commun.* **180**, 681–686 (1991).
44. Capon, C., Maes, E., Michalski, J. C., Leffler, H. & Kim, Y. S. Sd(a)-antigen-like structures carried on core 3 are prominent features of glycans from the mucin of normal human descending colon. *Biochem J* **358**, 657–664 (2001).
45. Groux-Degroote, S. *et al.* B4GALNT2 gene expression controls the biosynthesis of Sda and sialyl Lewis X antigens in healthy and cancer human gastrointestinal tract. *Int. J. Biochem. Cell Biol.* **53**, 442–449 (2014).
46. Robbe, C., Capon, C., Coddeville, B., & Michalski, J. C. Structural diversity and specific distribution of O-glycans in normal human mucins along the intestinal tract. *Biochem. J.* **384**, 307–316 (2004).
47. Iwai, T. *et al.* Core 3 synthase is down-regulated in colon carcinoma and profoundly suppresses the metastatic potential of carcinoma cells. *Proc. Natl. Acad. Sci.* **102**, 4572–4577 (2005).
48. Vavasseur, F., Yang, J. M., Dole, K., Paulsen, H. & Brockhausen, I. Synthesis of O-glycan core 3: Characterization of UDP-GlcNAc: GalNAc-R β 3-N-acetyl-glucosaminyltransferase activity from colonic mucosal tissues and lack of the activity in human cancer cell lines. *Glycobiology* **5**, 351–357 (1995).
49. Múnera, J., Ceceña, G., Jedlicka, P., Wankell, M. & Oshima, R. G. Ets2 regulates colonic stem cells and sensitivity to tumorigenesis. *Stem Cells* **29**, 430–439 (2011).
50. Alitalo, K. *et al.* Aberrant expression of an amplified c-myc oncogene in two cell lines from a colon carcinoma. *Proc. Natl. Acad. Sci. U. S. A.* **81**, 4534–4538 (1984).
51. Ciznadija, D. *et al.* Intestinal adenoma formation and MYC activation are regulated by cooperation between MYB and Wnt signaling. *Cell Death Differ.* **16**, 1530–1538 (2009).
52. Sakuma, K., Aoki, M. & Kannagi, R. Transcription factors c-Myc and CDX2 mediate E-selectin ligand expression in colon cancer cells undergoing EGF/bFGF-induced epithelial-mesenchymal transition. *Proc. Natl. Acad. Sci. U. S. A.* **109**, 7776–7781 (2012).
53. Robbe-Masselot, C. *et al.* Expression of a core 3 disialyl-Lex hexasaccharide in human colorectal cancers: A potential marker of malignant transformation in colon. *J. Proteome Res.* **8**, 702–711 (2009).

