Universiteit
Leiden
The Netherlands

The genome sequence of hairy root Rhizobium rhizogenes strain LBA9402: bioinformatics analysis suggests the presence of a new opine system in the agropine Ri plasmid

Hooykaas, M.J.G.; Hooykaas, P.J.J.

Citation
Hooykaas, M. J. G., & Hooykaas, P. J. J. (2021). The genome sequence of hairy root Rhizobium rhizogenes strain LBA9402: bioinformatics analysis suggests the presence of a new opine system in the agropine Ri plasmid. *Microbiologyopen*, *10*(2). doi:10.1002/mbo3.1180

Version:	Publisher's Version
License:	Creative Commons CC BY-NC-ND 4.0 license
Downloaded from:	https://hdl.handle.net/1887/3254618

Note: To cite this publication please use the final published version (if applicable).

Check for updates

Received: 4 January 2021 | Revised: 24 February 2021 | Accepted: 26 February 2021

DOI: 10.1002/mbo3.1180

ORIGINAL ARTICLE

MicrobiologyOpen Open Access WILEY

The genome sequence of hairy root *Rhizobium rhizogenes* strain LBA9402: Bioinformatics analysis suggests the presence of a new opine system in the agropine Ri plasmid

Marjolein J. G. Hooykaas | **Paul J. J. Hooykaas**

Institute of Biology, Leiden University, Leiden, The Netherlands

Correspondence
Paul J. J. Hooykaas, Institute of Biology, Leiden University, Sylviusweg 72, Leiden, The Netherlands.
Email: p.j.j.hooykaas@biology.leidenuniv.nl

Abstract

We report here the complete genome sequence of the *Rhizobium rhizogenes* (formerly *Agrobacterium rhizogenes)* strain LBA9402 (NCPPB1855rifR), a pathogenic strain causing hairy root disease. To assemble a complete genome, we obtained short reads from Illumina sequencing and long reads from Oxford Nanopore Technology sequencing. The genome consists of a 3,958,212 bp chromosome, a 2,005,144 bp chromid (secondary chromosome) and a 252,168 bp Ri plasmid (pRi1855), respectively. The primary chromosome was very similar to that of the avirulent biocontrol strain K84, but the chromid showed a 724 kbp deletion accompanied by a large 1.8 Mbp inversion revealing the dynamic nature of these secondary chromosomes. The sequence of the agropine Ri plasmid was compared to other types of Ri and Ti plasmids. Thus, we identified the genes responsible for agropine catabolism, but also a unique segment adjacent to the TL region that has the signature of a new opine catabolic gene cluster including the three genes that encode the three subunits of an opine dehydrogenase. Our sequence analysis also revealed a novel gene at the very right end of the TL-DNA, which is unique for the agropine Ri plasmid. The protein encoded by this gene was most related to the succinamopine synthases of chrysopine and agropine Ti plasmids and thus may be involved in the synthesis of the unknown opine that can be degraded by the adjacent catabolic cluster. The available sequence will facilitate the use of *R. rhizogenes* and especially LBA9402 in both the laboratory and for biotechnological purposes.

KEYWORDS
Agrobacterium rhizogenes, chromid, opine, ornithine cyclodeaminase, Ri plasmid, succinamopine synthase

1 | INTRODUCTION

Hairy root, a neoplastic plant disease with a wide host range, is characterized by the formation of adventitious roots from infected wound sites (Riker, 1930). It was originally encountered as a problem in tree nurseries, but nowadays the disease is also increasingly causing problems in the greenhouse by inducing extensive root mats, thereby reducing the harvest of cucumbers and tomatoes (Weller et al., 2000). The causal agent, a bacterium that was for long called *Agrobacterium rhizogenes*, contains a large, about 200 kbp

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.
© 2021 The Authors. *MicrobiologyOpen* published by John Wiley & Sons Ltd.

root-inducing (Ri) plasmid, which contains the essential virulence determinants (White & Nester, 1980). The molecular mechanism by which hairy root is induced is similar to that which is used by the related bacterium *Agrobacterium tumefaciens* to induce crown gall tumors in plants. During infection, part of the Ri plasmid (the T region) is transferred to plant cells and integrated into the plant genome (Bevan & Chilton, 1982). Expression of *rol*-genes located in the transfer DNA (T-DNA) leads to the transformation of normal cells into tumor cells that develop into roots that can grow in in vitro culture in the absence of added plant growth regulators (Jouanin et al. 1987). In the cells of hairy roots, unusual compounds called opines are produced, which are specific condensates of amino acids and keto acids or sugars (Petit et al., 1983). These opines, which are formed by enzymes encoded by the T-DNA, support the growth of the pathogen, which contains the catabolic genes usually in a region adjacent to the T region on the Ri plasmid (Dessaux et al., 1993). Based on the specific opines formed and degraded, agropine, cucumopine, mannopine, and mikimopine Ri plasmids are nowadays distinguished.

Like *A. tumefaciens*, *A. rhizogenes* has been disarmed by deleting the T-DNA genes to convert this region into a vector useful for plant genome engineering (Collier et al., 2018). Also, the bacterium as such is used for biotechnological research and application; in research for instance for gene function and gene expression analysis in roots (Ron et al., 2014); and in the industry to obtain roots that can be grown in bioreactors for the production of secondary metabolites (Mehrotra et al., 2015).

It has become apparent over the years that in nature various bacteria of the *Rhizobiaceae* family may cause hairy root or crown gall, depending on whether they carry a Ri plasmid or a Ti plasmid. Based on their physiological properties, three different groups (biovars 1–3) were distinguished already long ago (Kerr & Panagopoulos, 1977). The species name *Rhizobium rhizogenes* is now commonly used for the bacteria belonging to biovar 2. *R. rhizogenes* strains have two megabase DNA circles, a chromosome and a plasmid-derived megacircle, sometimes called a chromid (Jumas-Bilak et al., 1998; Harrison et al. 2010). Draft genome sequences of several strains are available on NCBI, and one draft genome sequence has been published for *R. rhizogenes* strain ATCC15834 consisting of 43 scaffolds (Kajala et al., 2014). However, up to now, only one complete genome sequence is available for *R. rhizogenes*, that of the avirulent agrocin-producing biocontrol agent Kerr 84 (Slater et al., 2009). Here, we present a second complete genomic sequence of *R. rhizogenes*, that of the hairy root-inducing strain LBA9402. This strain is a rifampicin-resistant derivative of wild-type strain NCPPB1855, which is one of the most widely used laboratory strains (Desmet et al., 2020). By comparing the chromosome and chromid of LBA9402 with those of strain K84, we found that the chromosome was very similar, but that the chromid showed large differences due to a large 724 kb deletion accompanied by a large inversion, underscoring the dynamic nature of the chromid. Analysis of the sequence of the agropine Ri plasmid of LBA9402 revealed that this had a few unique areas including one that we predict encodes a new opine catabolic cluster, including the three genes characteristic for defining the three subunits of an opine dehydrogenase. A candidate gene for a novel opine synthase was identified at the very right end of the TL-DNA.

2 | MATERIALS AND METHODS

2.1 | Organism

Rhizobium rhizogenes strain LBA9402 is a rifampicin-resistant derivative of wild-type strain NCPPB1855, which was originally isolated from *Rosa* spp. (Hooykaas, 1979). The bacterium was grown on TY medium (Difco tryptone 5 g/l, Difco yeast extract 3 g/l, $CaCl_2.6H_2O$ 1.3 g/l). The bacterium was tested for virulence by puncturing the stems of *Kalanchoe daigremontiana* and *Kalanchoe tubiflora* with a sterile wooden toothpick that had been dipped into a colony of the bacterium.

2.2 | Sequencing methods

Rhizobium rhizogenes strain LBA9402 was cultured in TY medium (Beringer, 1974), followed by genomic DNA isolation using QIAGEN gravity-flow columns (QIAGEN Genomic-tip 100/G kit Cat No./ID: 10243). The genome of LBA9402 was sequenced using a combination of Illumina and Oxford Nanopore Technologies platforms. Nanopore sequencing was done in-house, but Illumina sequencing was performed at the Leiden Genome Technology Center (LGTC) of the Leiden University Medical Center (Leiden, The Netherlands), where TruSeq DNA Libraries were sequenced on an Illumina HiSeq 2000 machine. The Oxford Nanopore sequencing library was generated with 200 ng DNA using the SQK-RBK004 Rapid Barcoding Kit. The library was pooled with another library, followed by in-house sequencing on a MinION flow cell (version R9.4.1).

2.3 | Data processing methods

After base calling with Albacore (version 2.3.4), the MinION reads were demultiplexed (with Epi2me). The total yield for LBA9402 was 298,712 reads, totaling 1,027,720,149 bp, with a mean read length of 3,441 bp. Nanopore reads were end-trimmed and filtered on average quality (>Q10) and length (>5,000 bp) with NanoFilt (64-fold coverage after filtering). A total of 4,518,191 99-nucleotide paired-end Illumina reads were quality and adapter trimmed using Cutadapt (70-fold coverage). Hybrid assembly was performed using Unicycler version 0.4.7. Besides three contigs representing the two chromosomes and the Ri plasmid, the fourth contig of 5,386 bp was identified. This represented the bacteriophage ΦX174 genome sequence, which is spiked-in at low concentration during Illumina library preparation. This contig was therefore removed from the assembly. The assembly was annotated using NCBI Prokaryotic Genome Annotation Pipeline (PGAP). In addition, PHASTER was used to annotate prophage sequences (Arndt et al., 2016). For the functional characterization of

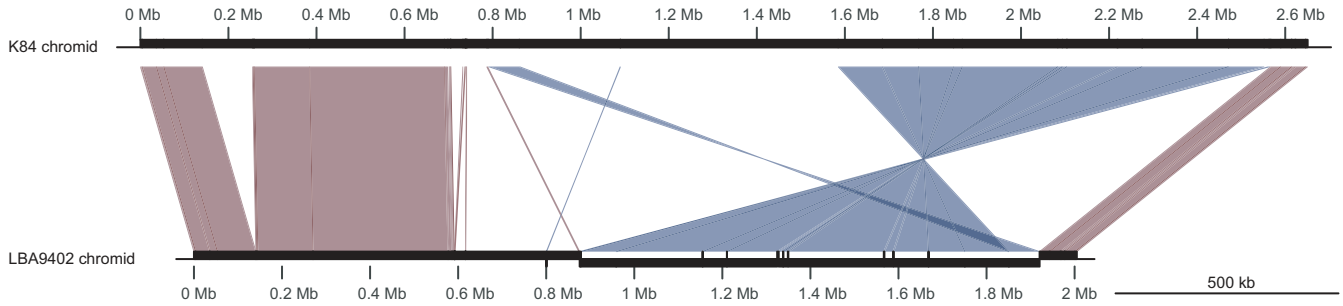


FIGURE 1 Large rearrangements in the chromid of *R. rhizogenes*. The chromid/secondary chromosome of LBA9402 was aligned to that of K84 using progressiveMauve. The aligned blocks were visualized with the R package genoplotR. Red areas are regions with the same orientation, whereas blue areas align in inverse orientation

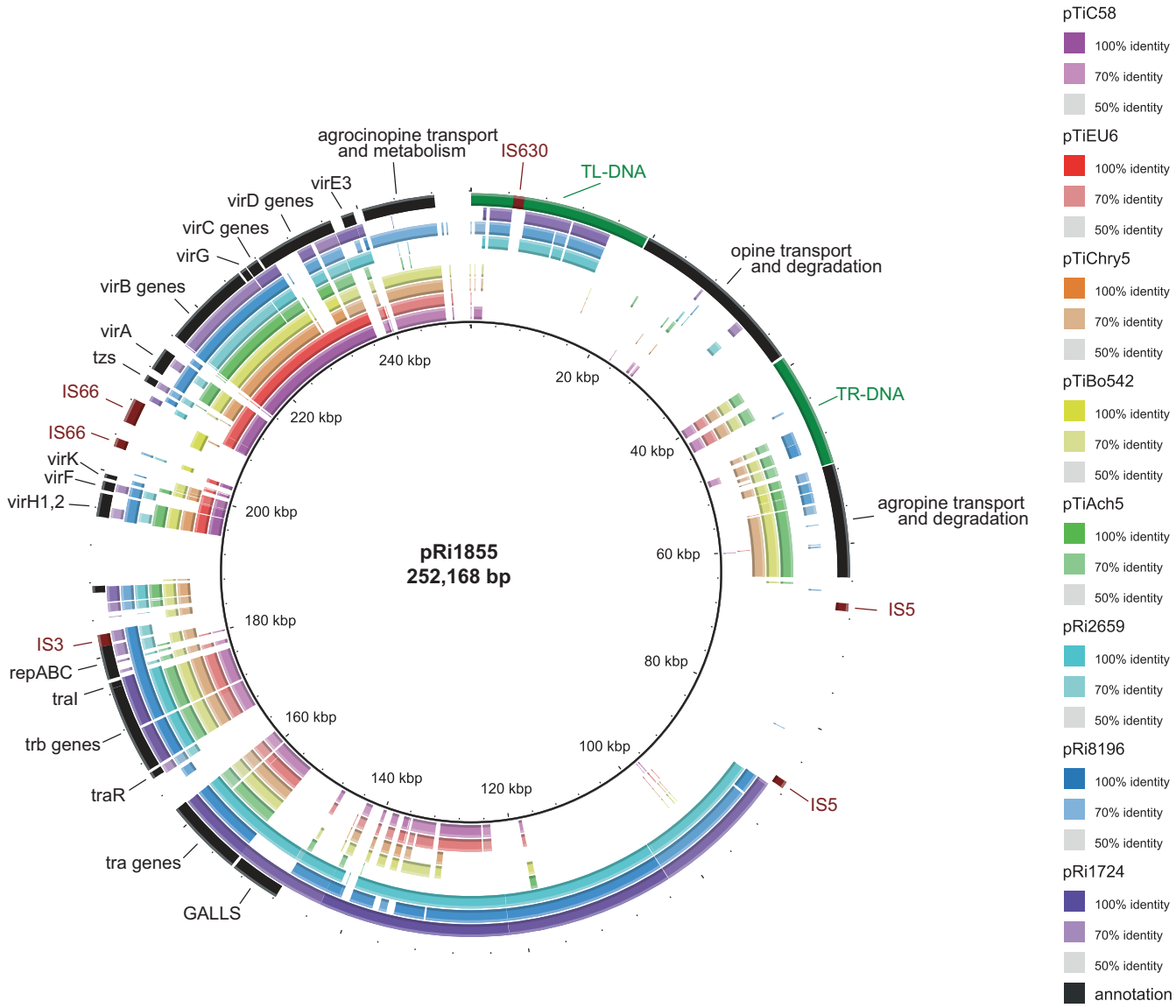


FIGURE 2 Circular representation of pRi1855 and comparative analysis to other Ri and Ti plasmids. The inner rings show BLASTn comparisons between Ri1855 and various Ri and Ti plasmids. Color intensity indicates the degree of sequence similarity, as shown in the legend. The number of regions and genes is indicated on the outermost ring

the encoded proteins, eggNOG-Mapper was employed (Huerta-Cepas et al., 2017). Insertion elements (IS elements) were identified using ISEScan (Xie & Tang, 2017). In Figure 2 and Appendix Figure A5, only complete insertion sequences, that is, including inverted repeats, are shown. IslandViewer was used to predict genomic islands (Dhillon et al., 2015), and CGView was used to generate a circular

map of pRi1855 (Stothard & Wishart, 2005). Mauve (progressiveMauve) (Darling et al., 2010) and MUMmer (NUCmer) (Kurtz et al., 2004) were used to align the LBA9402 and K84 genomes. Average nucleotide identity (ANI) values were calculated with fastANI (Jain et al., 2018), and Digital DNA–DNA hybridization (DDH) values were estimated with GGDC 2.1 (identities/HSP length) (Meier-Kolthoff et al., 2013). BRIG was used to compare pRi1855 with other Ri and Ti plasmids (with BLASTN, e-value cut-off 1e-10) and to visualize the hits in concentric rings (Alikhan et al., 2011). Percent identity values between specific regions of pRi1855 and pRiA4 were obtained with EMBOSS needle (Needleman-Wunsch pairwise global alignment). Single-nucleotide polymorphisms (SNPs) and small indels were detected with Snippy (https://github.com/tseemann/snippy, version 4.6.0, ran versus pRiA4 with the contigs option --ctgs). For the comparisons between erythritol catabolism regions and between pRi1855 and *Rhizobium lusitanum* strain 629, BLASTn was run locally with BLAST version 2.9.0+, and the similarities were visualized with the R package genoplotR (Guy et al., 2010). Protein alignments were performed with MAFFT version 7.471, L-INS-I method (Katoh, 2013), and visualized with Jalview version 2.11.1.2 (Waterhouse et al., 2009) and Adobe Illustrator. Percentage identities were calculated with the R package seqinr.

3 | RESULTS

3.1 | The genomic sequence of *Rhizobium rhizogenes* strain LBA9402

As Illumina sequencing data alone were not sufficient to obtain a high-quality and complete genome sequence of strain LBA9402, we additionally obtained long reads by Nanopore sequencing. Unicycler was used to obtain a hybrid assembly. This resulted in three circular contigs of 3,958,212 bp, 2,005,144 bp and 252,168 bp, respectively. More than 99.9% of both MinION and Illumina reads align to the assembly, indicating that the assembly is complete. The G + C content of the genome was 60%. The assembly was annotated using NCBI Prokaryotic Genome Annotation Pipeline (PGAP). In total, 5,822 coding sequences, 9 rRNA genes (3 operons), and 53 tRNA genes were annotated. COG categories were assigned to predicted coding sequences with eggNOG-mapper (Appendix Figure A1).

3.2 | Comparison to the sequence of *R. rhizogenes* strain K84

The LBA9402 genome sequence was uploaded to the Microbial Genome Atlas webserver for taxonomic classification by TypeMat, to verify that it is indeed a *R. rhizogenes* strain. Average nucleotide identity (ANI) to the type strain NBRC 13257 was very high (98.9%). The digital DNA–DNA hybridization (DDH) value as calculated with Genome-to-Genome Distance Calculator 2.1 (GGDC 2.1) was 91.6% versus the type strain. These high values (>95% ANI, >70% DDH) confirm that LBA9402 is a *R. rhizogenes* strain. The only other complete, high-quality genome of *R. rhizogenes* obtained thus far is that of the avirulent biocontrol strain K84 (Slater et al., 2009). This latter strain does not contain an Ri plasmid, but the two LBA9402 megacircles have high sequence similarity to those of K84 (both genomes have 99% ANI, 92.1% DDH). As can be seen in Appendix Figure A2, the largest contig of LBA9402 is largely collinear with the primary chromosome of K84 (96% of the sequence can be aligned to that of K84 at >95% sequence identity, 99% ANI). It contains the genes for replication such as for a DnaA replication protein, repair, and DNA recombination, for cell division, for transcription and translation including the 3 rRNA clusters of the bacterium and the 53 tRNA genes. We annotated an extra tRNA in one of the unique regions of the LBA9402 primary chromosome. The chromosome contains a putative genomic island of about 100 kbp with a set of genes for conjugative DNA transfer encoding not only a Type IV secretion system for mating pair formation (from position 3,324,071 to 3,314,501), but also the enzymes necessary for DNA transfer and replication (from position 3,302,269 to 3,307,564). The genomic island contains a gene for a putative integrase and is surrounded by a direct repeat of 15 bp, which may be (the remains of) two *att* sites. Larger differences between the chromosomes of LBA9402 and K84 are mainly due to the presence/absence of other mobile elements. For example, various proteins encoded in the unique segment of DNA from position 745,195 to 784,185 in LBA9402 have homology to phage proteins (Rhizobium phage vB_RleM_PPF1 and other tailed phages) as revealed by the phage search tool PHASTER (Arndt et al., 2016) and thus seems due to the insertion of a prophage (Appendix Figure A3).

The second-largest replicons are less similar (Appendix Figures A2, 1), but still, 85% of the LBA9402 sequence aligns to that of K84 (and 64% of K84 aligns to LBA9402, 99% ANI). The LBA9402 sequence is smaller, mainly due to a large approximately 724 kb deletion, which seems accompanied by a large 1.8 Mbp inversion (Figure 1). The large deletion did not affect any class of genes in particular as can be seen in Appendix Figure A1b, which shows a similar distribution of the predicted proteins in COG categories in both replicons.

This secondary megacircle has a plasmid-like RepABC replication system but has a similar GC content as the primary chromosome. Such secondary megacircles are considered (developing) secondary chromosomes that over evolutionary time exchange genes with the primary chromosome and have been coined “chromids” (Slater et al., 2009; Harrison et al. 2010). The chromid of LBA9402 contains many metabolic genes, but also genes for the production of cell wall polysaccharides and fimbriae/pili. We found in the chromid a set of genes homologous to the erythritol region in *Sinorhizobium meliloti* and *Rhizobium leguminosarum* (Appendix Figure A4) including a transport operon with genes *eryEFG*, a catabolic operon with genes *eryABCD* and a *deoR*-type regulator (also called *eryR*) followed by genes called *eryH* and *eryI* (Barbier et al., 2014; Geddes & Oresnik, 2012; Yost et al., 2006). The ability to catabolize erythritol is one of the key characteristics distinguishing biotype 1 and biotype 2 agrobacteria (Kerr & Panagopoulos, 1977). The presence of erythritol catabolic

genes was thus expected, but it was remarkable that they were present on the more dynamic chromid instead of the chromosome. Finally, the 252 kbp circle represents the agropine pRi1855 plasmid, which is very different from the large nopaline catabolic plasmid carried by strain K84 and will be described below.

3.3 | General properties of the Ri plasmids

The pRi1855 plasmid comprises 252,168 bp. It has an approximately 4% lower GC content than the rest of the genome. In total, 236 protein-coding sequences were identified with an average size of 898 bp (Appendix Figure A5). Recently, a draft of agropine Ri plasmid pRiA4 was published (Thompson et al., 2020). This plasmid is slightly smaller than pRi1855 with a size of 249,350 bp, but has a similar restriction profile (Jouanin, 1984) and is indeed very similar to pRi1855 (99% ANI). We compared the agropine Ri plasmid pRi1855 sequence to publicly available Ri and Ti plasmid sequences: octopine Ti plasmid pTiAch5 (CP007228; Henkel et al., 2014; Huang et al., 2015), nopaline Ti plasmid pTiC58 (AE007871; Goodner et al., 2001; Wood et al., 2001), succinamopine Ti plasmid pTiEU6 (KX388535; Shao et al., 2019), agropine Ti plasmid pTiBo542 (DQ058764; Oger et al., 2001), chrysopine Ti plasmid pTiChry5 (KX388536; Shao et al., 2018), mannopine Ri plasmid pRi8196 (Weisberg et al., 2020), cucumopine Ri plasmid pRi2659 (NZ_CP019703.3; Valdes Franco et al., 2016; Tong et al., 2018), and mikimopine Ri plasmid pRi pRi1724 (NC_002575; Moriguchi et al., 2001). The conservation of the different areas in these plasmids is visualized in Figure 2, and we shall discuss these in the following parts.

As can be seen in Figure 2, the replication (*repABC*) and conjugative transfer (*tra*, *trb*) genes are very similar to those of other Ti and Ri plasmids. This was previously already shown for the replication and conjugative transfer genes of the closely related agropine Ri plasmid pRiA4 (Nishiguchi et al., 1987; Wetzel et al., 2015). Indeed most of these genes are very similar in both agropine Ri plasmids. For example, pRi1855 *repABC* and *traI* are 100% identical to those of pRiA4. The *traAFBH* and *traCDG* genes are 95.3% identical, and the *trb* operon 88.8%. In contrast, *traR* and *trbK* are less conserved (62.7%, 47.5%), also compared to the other Ri plasmids.

The agropine Ri plasmid has two T regions, one of which, the TL region, contains the *rol*-genes that are necessary and sufficient for the formation of hairy roots (Offringa et al., 1986; White et al., 1985). Other Ri plasmids have only one T region with very similar genes (Figure 3; Otten, 2018). In the agropine Ri plasmid, however, a copy of IS630 is inserted between *orf3* and *orf8*. At the very left end in the agropine Ri TL region and the mannopine Ri T region, a gene for agrocinopine synthase is present, but only remnants of these genes are still present in the cucumopine and mikimopine Ri plasmids. At the very right end of the T region one (in the cucumopine and mikimopine Ri plasmids) or two (in the mannopine Ri plasmids), non-conserved genes are present. These encode the cucumopine (Valdes Franco et al., 2016) and mikimopine synthases (Moriguchi et al., 2001), respectively, and the two genes necessary for mannopine synthesis in the mannopine Ri plasmid (Figure 3). The TL region of the agropine Ri plasmid pRiA4 was previously sequenced (Slightom et al., 1986), and this revealed at the right end the presence of *orf15/rolD* and three smaller *orfs*. In our pRi1855 sequence, we find besides *orf15/rolD* only one larger *orf*, hereinafter called *orf16*. The function of *orf16* is unknown but may encode an unknown opine synthase as will be discussed in the next paragraph. The agropine Ri plasmid has besides the conserved T region (TL region) an additional T region (TR region) containing *aux*-genes involved in the biosynthesis of the auxin indole acetic acid (Offringa et al., 1986) and the genes *mas1*, *mas2*, and *ags* for agropine biosynthesis (Bouchez and Tourneur 1991).

The virulence region of pRi1855 responsible for the transfer of the T-DNA into plant cells contains the essential virulence genes *virA*, *virB1-virB11*, *virG*, *virC1*, *virC2*, and *virD1-virD5* in the same order as in other Ri and Ti plasmids, but although *virE3* is present close to *virD5*, the *virE1* and *virE2* genes are missing and replaced by a new *orf* with some similarity to nopaline pTi *virF*. The sequence of the cucumopine and mikimopine Ri plasmids in this area is almost identical

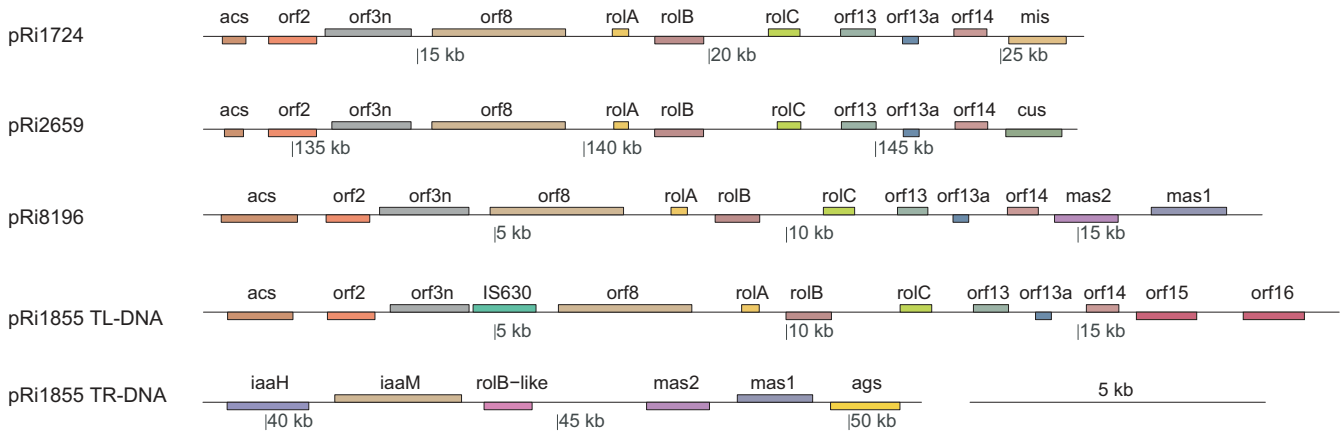


FIGURE 3 Unique genes located at the very right end of the T regions of Ri plasmids. Schematic overview of genes in the T regions of different Ri plasmids (pRi1724: mikimopine type, pRi2659: cucumopine type, pRi8196: mannopine type, pRi1855: agropine type). Homologous genes have the same color

suggesting that the deletion of *virE2* occurred once before the divergence of these Ri plasmids. The *virE2* gene is functionally replaced by a gene called *GALLS* as reported before for pRiA4 (Hodges et al., 2004). The *GALLS* gene was previously also identified in the cucumopine and mikimopine Ri plasmids by Southern analysis but is not present in the mannopine Ri plasmid, which still carries the *virE2* gene (Hodges et al., 2004). Together with a *tzs* gene, which encodes an enzyme that catalyzes the synthesis of the cytokinin zeatin riboside 5′-phosphate (Krall et al., 2002), *GALLS* is located outside the *vir* region, about 65 kbp clockwise from the *virE3* gene, near opine catabolic genes in the cucumopine and mikimopine Ri plasmids. We could now identify and locate the *GALLS* gene in the pRi1855 sequence at a completely different location next to the *traG* gene almost 90 kbp counterclockwise from the *virE3* gene. Besides the core set of *vir* genes mentioned above, the *vir* region of the agropine Ri plasmid contains next to the *virA* gene the *tzs* gene, *virK*, a second nopaline Ti-like *virF* gene, and finally *virH1*, *virH2*. It resembles in this respect the *vir* region of the nopaline Ti plasmid and like in the nopaline Ti plasmids *virJ* is absent. The *virD3* gene, which is very variable, also shows the highest similarity to that of nopaline Ti plasmids. Other types of Ri plasmids have similar *vir* regions, but a *virH2* gene is absent from the cucumopine and mikimopine Ri plasmids, and as mentioned above a *tzs* gene is present, but located in an entirely different area of the plasmid.

3.4 | Agropine and agrocinopine catabolism genes

Hairy roots formed by agropine strains contain agropine, agropinic acid, mannopinic acid, and mannopine (Petit et al., 1983). The agropine Ri plasmid enables host strains to degrade agropine. *R. rhizogenes* strains such as A4, but not NCPPB1855 contain a second, catabolic plasmid with genes for catabolism of the other three mannityl opines (Petit et al., 1983). We have now identified the genes for agropine transport and catabolism in pRi1855, which are located in a segment of the plasmid adjacent to the TR region (Figure 2). This region contains genes with high similarity to the genes described by Kim and Farrand (1996) on the octopine Ti plasmid involved in agropine uptake and degradation. These genes encode an agropine permease and also comprise *agcA* for the delactonase converting agropine into mannopine, *mocC* for oxidizing mannopine into deoxyfructosyl glutamine, and *mocDE* determining the deconjugase liberating an amino acid and a phosphorylated sugar. In the octopine Ti plasmid, the genes *mocA* and *mocB* encode enzymes with weak homology to glucose-6-phosphate dehydrogenase and 6-phosphogluconate dehydratase (Kim & Farrand, 1996). These are probably involved in further catabolism of the released phosphorylated sugar. However, while an intact homolog of *mocA* was present in pRi1855, as well as a homolog of *mocC*, in between only a truncated remnant of a gene homologous to *mocB* was present due to a deletion of more than 1 kbp. Regulators closely related to *mocS* and *mocR* are present in an identical position in front of *mocA* and between *mocC* and *mocD*. Our detection of a *mocD* gene in pRi1855 was remarkable as such gene was thought to be absent from the agropine Ri plasmid (Baek et al., 2005). The agropine Ri plasmid has adjacent to the left end of the TL region in pRi1855 a set of *acc* genes for agrocinopine catabolism (Figure 2), which matches the presence of an *acs* gene for the biosynthesis of agrocinopine in the TL region. These genes are also present in the mannopine Ri plasmid but absent from the cucumopine and mikimopine Ri plasmids.

3.5 | Genes for a novel opine system in pRi1855

The pRi1855 plasmid has several regions with genes of unknown function. It shares a large region of about 65 kbp (the area 88–153 kbp on the map of Figure 2) with the other Ri plasmids. This region contains genes putatively involved in sugar transport, glycerol metabolism and encodes several transcription regulators and two chemoreceptors (Moriguchi et al., 2001). Besides, the pRi1855 plasmid has several unique areas with genes that are found in none of the other types of Ri and Ti plasmids described. These include transposable elements (Figure 2) and two larger areas of about 20 kbp (the area 20–40 kbp on the map adjacent to the right border of the TL region in Figure 2) and about 24 kbp (the area 64–88 kbp on the map adjacent to the agropine catabolic genes). The latter area contains mainly metabolic genes and may have been introduced into pRi1855 by transposition as it is surrounded by IS5-like insertion sequences. It may have originated from the chromosome of another *Rhizobium* species, as a very similar stretch of DNA was detected in the recently sequenced chromosome of *Rhizobium lusitanum* strain 629 (Appendix Figure A6). The 20 kbp segment adjacent to the right border of the TL region may be involved in the transport and catabolism of a new opine. In this area, we identified all the three characteristic genes that together code for the three subunits of a putative flavin-containing opine dehydrogenase (Appendix Figures A7–A9). Flavin-containing opine dehydrogenases such as octopine, nopaline, and succinamopine dehydrogenase consist of three subunits OdhABC that are encoded by *noxABC*-like/*ooxABC*-like genes arranged in tandem in the genome (Watanabe et al., 2015). The three genes in pRi1855 (F3X89_28345, F3X89_28350, F3X89_28355) encode closely related proteins in which the characteristic binding sites for the FAD and FMN co-factors and the Fe-S cluster have fully been conserved (Appendix Figures A7–A9). These three genes are surrounded on both sides by genes for a transport system and a LysR-type regulator. In Ti and Ri plasmids, genes encoding an opine dehydrogenase are often accompanied by genes encoding the permease required for uptake of a specific opine into the bacterial cell. Also, genes encoding enzymes necessary for the further catabolism of the products liberated by the activity of the opine dehydrogenase on the opine substrates are often present in the vicinity. In this area of pRi1855 genes encoding such metabolic proteins are also present, including genes encoding a putative saccharopine dehydrogenase and a putative aminoadipate semialdehyde dehydrogenase, which may form part of a catabolic pathway of the amino acid lysine (de Mello Serrano et al., 2012). A gene for an AsnC/Lrp regulator is located at the end of this DNA segment. The Lrp family of

transcriptional regulators is known to control amino acid metabolism in bacteria (Brinkman et al., 2003).

If these genes are involved in the catabolism of an opine, a gene for an unknown opine synthase should be present in the T region of pRi1855. Genes for agrocinopine synthase are located at the extreme left end of the T region in Ti and Ri plasmids, while genes for nopaline synthase, octopine synthase, and succinamopine synthase are located at the extreme right end of the T region in Ti plasmids. In cucumopine, mikimopine, and mannopine Ri plasmids, the genes for cucumopine, mikimopine, and mannopine synthesis are likewise located immediately next to the right border repeat. We find at the very right end of the TL region of pRi1855 two related genes (Figure 3): *orf15*/*rolD* and *orf16*, which share 55% identity (Figure 4, Table 1). These genes are not present in the T regions of any of the other types of Ri plasmids (Figure 3). The *orf15* has been called *rolD*; the encoded RolD protein has weak sequence homology with ornithine cyclodeaminases and indeed can convert ornithine into proline (Trovato et al., 2001). The role of *rolD* in hairy root formation is marginal, but the gene can influence plant development by its metabolic activity (Trovato et al., 2018). Using BLASTP with the proteins encoded by *orf15* and *orf16* as a query, we picked up the succinamopine synthases encoded by the T region of chrysopine pTiChry5 (Shao et al., 2018) and the agropine pTiBo542 (Oger et al., 2001) as the most related proteins. The proteins encoded by *orf15* and *orf16* share 44–47% identity with the two succinamopine synthases, which themselves share 93% identity (Table 1). All these proteins (encoded by *orf15*/*rolD*, *orf16*, *susL*) are evolutionarily related to ornithine cyclodeaminases (encoded by *ocd* genes). They share, for instance, about 19–21% identity with the ornithine cyclodeaminase encoded by the nopaline Ti plasmid. That ornithine cyclodeaminase (*ocd*) genes can evolve novel biochemical functions during evolution is known for some time. For instance, its function has been reported to evolve into an alanine dehydrogenase in *Archaeoglobus fulgidus* and into a tauropine dehydrogenase in *Halichondria japonica* (Sharma et al., 2013; Watanabe et al., 2014). It can evolve also in an opine (succinamopine) synthase. Therefore, it would seem possible that either or both of the *ocd*-like genes at the right end of pRi1855 (*orf15*, *orf16*) similarly have evolved a novel opine synthase function, producing an unknown opine that can be degraded by the putative opine dehydrogenase encoded in the area with genes of unknown function located adjacent to the right border of the TL region.

4 | DISCUSSION

The second complete, high-quality genomic sequence of a *Rhizobium rhizogenes* strain and the first of a virulent strain enabled us to make a comparison with the sequence of the previously sequenced biocontrol strain K84. This revealed high conservation of the primary chromosome but showed large differences in the secondary megacircle, the chromid. It has been described that chromids have a plasmid-like RepABC replication system but have a similar GC content as the primary chromosome and this is also the case in LBA9402. It has been proposed that chromids are plasmids that evolve into secondary chromosomes and overtime exchange genes with the primary chromosome (Slater et al., 2009; Harrison et al. 2010). We found that the chromid of strain LBA9402 was much smaller than that of strain K84 due to the absence of a segment of 724 kbp that may have been deleted in LBA9402 or inserted in K84. Also, we found that this insertion/deletion was accompanied by a large inversion of a segment of 1.8 Mbp. The presence of such complex rearrangements is in line with their plasmid descent and the genes which they carry being mostly non-essential.

Our genomic sequence includes the complete sequence of the agropine pRi1855 plasmid. Over the years, sequences have already been published dealing with specific parts of the closely related agropine Ri plasmid pRiA4 and recently a draft of the completed pRiA4 sequence was published (Thompson et al., 2020). However, it still differs in numerous areas, both by base substitutions, small insertions/deletions (803 differences were detected with variant caller Snippy), and a few larger insertions/deletions encoding complete genes. Also, likely due to technical sequencing differences (the pRiA4-carrying bacterium was solely sequenced with Illumina "short read" technology whereas for LBA9402 we additionally obtained "long reads" with Nanopore sequencing), the repeat-containing GALLS gene sequence is shorter (presumably collapsed) in pRiA4.

Hairy roots formed by agropine strains contain agropine, agropinic acid, mannopinic acid, and mannopine (Petit et al., 1983). The agropine Ri plasmid, however, enables host strains to degrade agropine, but not the other mannityl opines. *R. rhizogenes* strains such as A4, but not NCPPB1855, contain a second, catabolic plasmid with genes for catabolism of the other three mannityl opines (Petit et al., 1983). We have now identified the genes for agropine catabolism in pRi1855 adjacent to the right border of the TR region. The region

TABLE 1 Percentage identities between the two rightmost proteins of the LBA9402 TL-DNA, succinamopine synthase (susL) of pTiBo542 and pTiChry5, and two bacterial ornithine cyclodeaminases.

	LBA9402 orf15	LBA9402 orf16	Bo542 susL	Chry5 susL	C58 ocd	Pseudomonas putida ocd
LBA9402 orf15	100%	55%	46%	47%	19%	19%
LBA9402 orf16	55%	100%	44%	44%	21%	21%
Bo542 susL	46%	44%	100%	93%	19%	18%
Chry5 susL	47%	44%	93%	100%	19%	16%
C58 ocd	19%	21%	19%	19%	100%	56%
Pseudomonas putida ocd	19%	21%	18%	16%	56%	100%

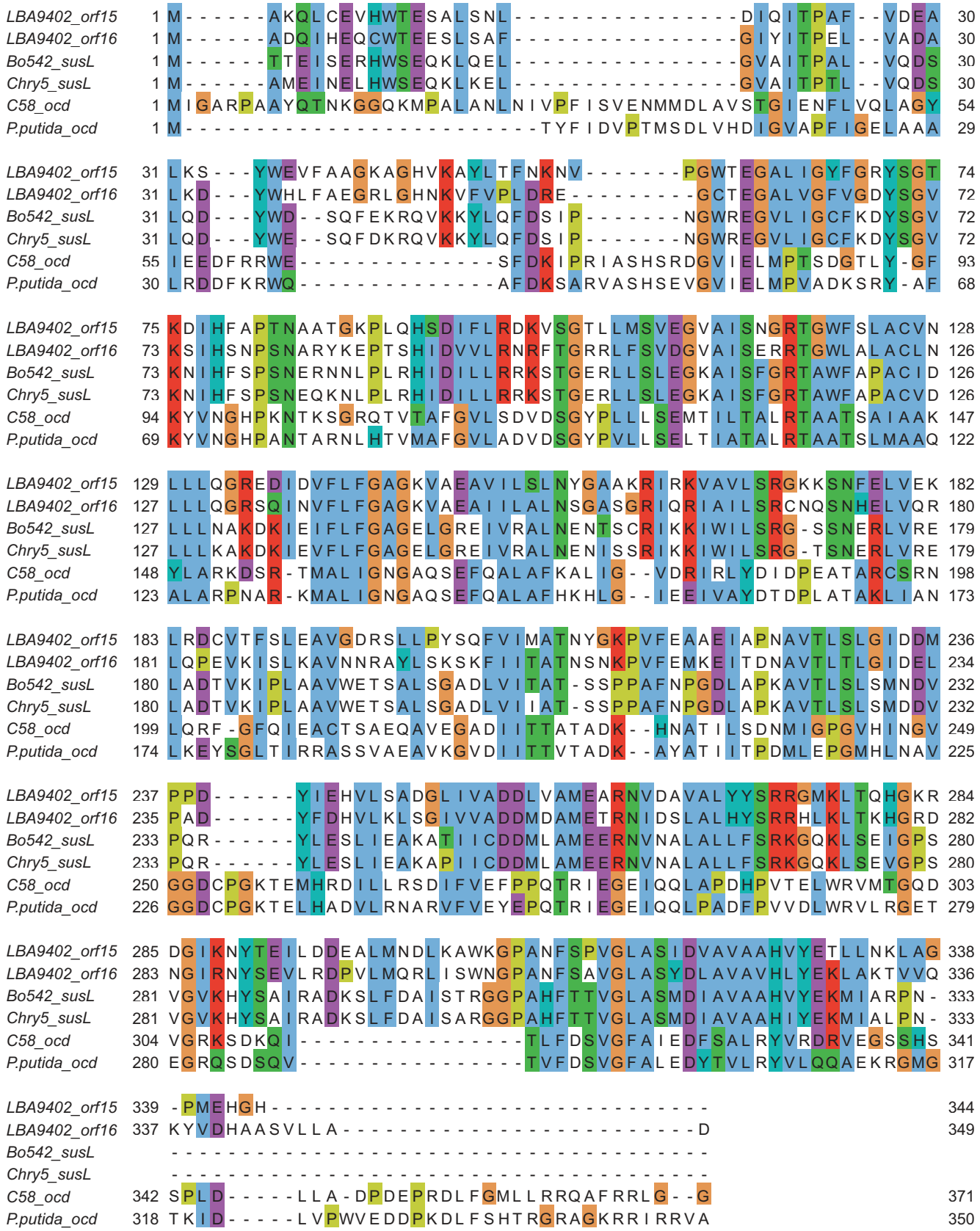


FIGURE 4 The proteins encoded by *orf15* and *orf16* are evolutionarily related to succinamopine synthase and more distantly to ornithine cyclodeaminase. Multiple sequence alignment of proteins encoded by *orf15* and *orf16* of pRi1855, *susL* from *A. tumefaciens* strains Chry5 and Bo542, and *ocd* from *A. tumefaciens* C58 and *Pseudomonasputida*. Colors are according to the Clustal X color scheme

embraces an *agcA* gene for the delactonase converting agropine into mannopine, *mocC* for oxidizing mannopine, and *mocD* and *mocE* together determining the enzymes that can release the amino acid and a phosphorylated sugar from the conjugate. This pathway would allow the bacterium to degrade both agropine and mannopine. However, it is known that the bacteria carrying pRi1855 cannot degrade mannopine. This may be because mannopine cannot induce the catabolic genes or because the bacterium cannot import mannopine into the cell. Indeed, the pRi1855 plasmid contains genes for an agropine permease, but not for a mannopine transport system.

Agrobacteria induce neoplasias in which opines are formed that serve as a nutritional source of the bacteria. All the different types of Ti and Ri plasmids described so far have a gene coding for an opine synthase at the very right end of the T region. Our sequence now shows that also the agropine Ri plasmid has one larger *orf* (*orf16*) at the very right end of the TL region, which shares 55% identity with the neighboring *rolD* gene. This gene encodes a protein that is evolutionary related to ornithine cyclodeaminase (ocd) and which still has ornithine cyclodeaminase activity (Trovato et al., 2001). Here, we discovered that both *rolD* and *orf16* have a significant identity of 44%–47% with the *susL* genes encoding succinamopine synthase in the agropine and chrysopine Ti plasmids. Ornithine cyclodeaminase encoding (*ocd*) genes have also been reported to have evolved into genes encoding new enzymatic activities such as alanine dehydrogenase activity in *Archaeoglobus fulgidus* and tauropine dehydrogenase activity in *Halichondria japonica* (Sharma et al., 2013; Watanabe et al., 2014). Previously, it was found that enzymes involved in the biosynthesis of mannopine are related to and thus may have been evolved from the enzymes required for degradation (Kim & Farrand, 1996). In this case, the enzymatic activity of the enzyme remained the same, but acted in the other direction, synthesis instead of degradation. We now find that an *ocd*-like gene may have evolved into a gene encoding a succinamopine synthase, which catalyzes a very different enzymatic step. Similarly, we hypothesize that an *ocd*-like gene may have evolved in the agropine Ri plasmid into a gene for a new opine synthase. Opine catabolic genes are often located close to the synthase gene, but on the other side of the right border, an arrangement seen in many different Ti and Ri plasmids. Indeed, three of the genes residing here together have the signature of the trios of genes such as *ooxABC* and *noxABC* (Appendix Figures A7–A9) that are known to encode the three subunits of octopine and nopaline dehydrogenase (Watanabe et al., 2015) and thus may encode the opine dehydrogenase needed for catabolism of the novel unknown opine.

Given its frequent application, the available sequence will facilitate the use of *R. rhizogenes* and especially LBA9402 in both the laboratory and for biotechnological purposes.

ACKNOWLEDGEMENTS

We are grateful to Amke den Dulk-Ras and Shuai Shao for help with DNA isolation, and Dr. Marcel van Verk and Dr. Chris Henkel for contributions to the initial analyses of the Illumina data. P.J.J.H. was supported by the fellowship associated with the appointment as Academy Professor by the Royal Netherlands Academy of Sciences.

CONFLICT OF INTEREST

None declared.

AUTHOR CONTRIBUTIONS

Marjolein J. G. Hooykaas: Conceptualization (equal); Data curation (equal); Formal analysis (lead); Funding acquisition (supporting); Investigation (lead); Methodology (lead); Project administration (equal); Resources (supporting); Software (lead); Supervision (supporting); Validation (lead); Visualization (lead); Writing-original draft (equal); Writing-review & editing (equal). **Paul J. J Hooykaas:** Conceptualization (equal); Data curation (equal); Formal analysis (supporting); Funding acquisition (lead); Investigation (supporting); Methodology (supporting); Project administration (equal); Resources (lead); Software (supporting); Supervision (lead); Validation (supporting); Visualization (supporting); Writing-original draft (equal); Writing-review & editing (equal).

ETHICS STATEMENT

None required.

DATA AVAILABILITY STATEMENT

The complete genome sequence of *R. rhizogenes* LBA9402 is available in GenBank under accession numbers CP044122, CP044123, and CP044124. The raw reads are deposited in the Sequence Read Archive under accession numbers SRR10177303 and SRR10177304. BioProject PRJNA566100: https://www.ncbi.nlm.nih.gov/bioproject/PRJNA566100

ORCID

Paul J. J. Hooykaas https://orcid.org/0000-0002-9736-6927

REFERENCES

Alikhan, N. F., Petty, N. K., Zakour, N. L. B., & Beatson, S. A. (2011). BLAST Ring Image Generator (BRIG): simple prokaryote genome comparisons. *BMC Genomics*, *12*, 402.

Arndt, D., Grant, J. R., Marcu, A., Sajed, T., Pon, A., Liang, Y., & Wishart, D. S. (2016). PHASTER: a better, faster version of the PHAST phage search tool. *Nucleic Acids Research*, *44*(W1), W16–W21.

Baek, C., Farrand, S. K., Park, D.-K., Lee, K.-E., Hwang, W., & Kim, K.-S. (2005). Genes for utilization of deoxyfructosyl glutamine (DFG), an amadori compound, are widely dispersed in the family Rhizobiaceae. *FEMS Microbiology Ecology*, *53*, 221–233.

Barbier, T., Collard, F., Zúñiga-Ripa, A., Moriyón, I., Godard, T., Becker, J., Wittmann, C., Schaftingen, E. V., & Letesson, J.-J. (2014). Erythritol feeds the pentose phosphate pathway via three new isomerases leading to D-erythrose-4-phosphate in Brucella. *Proceedings of the National Academy of Sciences of the United States of America*, *111*, 17815–17820.

Beringer, J. E. (1974). R factor transfer in *Rhizobium leguminosarum*. *Journal of General Microbiology*, *84*, 188–198.

Bevan, M. W., & Chilton, M. D. (1982). T-DNA of the *Agrobacterium* Ti and Ri plasmids. *Annual Review of Genetics*, *16*, 357–384.

Bouchez, D., & Tourneur, J. (1991). Organization of the agropine synthesis region of the T-DNA of the Ri plasmid from Agrobacterium rhizogenes. *Plasmid*, *25*, 27–39.

Brinkman, A. B., Ettema, T. J., de Vos, W. M., & van der Oost, J. (2003). The Lrp family of transcriptional regulators. *Molecular Microbiology*, *48*, 287–294.

Collier, R., Thomson, J., & Thilmony, R. (2018). A versatile and robust Agrobacterium-based gene stacking system generates high quality transgenic Arabidopsis plants. *The Plant Journal*, *95*, 573–583.

Darling, A. E., Mau, B., & Perna, N. T. (2010). progressiveMauve: Multiple genome alignment with gene gain, loss and rearrangement. *PLoS One*, *5*, e11147.

de Mello Serrano, G. C., e Silva Figueira, T. R., Kiyota, E., Zanata, N., & Arruda, P. (2012). Lysine degradation through the saccharopine pathway in bacteria: LKR and SDH in bacteria and its relationship to the plant and animal enzymes. *FEBS Letters*, *586*, 905–911.

Desmet, S., Dhooghe, E., De Keyser, E., Van Huylenbroeck, J., Müller, R., Geelen, D., & Lütken, H. (2020). Rhizogenic agrobacteria as an innovative tool for plant breeding: current achievements and limitations. *Applied Microbiology and Biotechnology*, *104*, 2435–2451.

Dessaux, Y., Petit, A., & Tempe, J. (1993). Chemistry and biochemistry of opines, chemical mediators of parasitism. *Phytochemistry*, *34*, 31–38.

Dhillon, B. K., Laird, M. R., Shay, J. A., Winsor, G. L., Lo, R., Nizam, F., Pereira, S. K., Waglechner, N., McArthur, A. G., Langille, M. G. I., & Brinkman, F. S. L. (2015). IslandViewer 3: more flexible, interactive genomic island discovery, visualization and analysis. *Nucleic Acids Research*, *43*, W104–W108.

Franco, J. A. V., Collier, R., Wang, Y., Huo, N., Gu, Y., Thilmony, R., & Thomson, J. G. (2016). Draft genome sequence of Agrobacterium rhizogenes strain NCPPB2659. *Genome Announcements*, *4*, e00746-16.

Geddes, B. A., & Oresnik, I. J. (2012). Genetic characterization of a complex locus necessary for the transport and catabolism of erythritol, adonitol and l-arabitol in *S.meliloti*. *Microbiology*, *158*, 2180–2191.

Goodner, B., Hinkle, G., Gattung, S., Miller, N., Blanchard, M., Qurollo, B., Goldman, B. S., Cao, Y., Askenazi, M., Halling, C., Mullin, L., Houmiel, K., Gordon, J., Vaudin, M., Iartchouk, O., Epp, A., Liu, F., Wollam, C., Allinger, M., … Flanagan, C. (2001). Genome sequence of the plant pathogen and biotechnology agent *Agrobacterium tumefaciens* C58. *Science*, *294*, 2323–2328.

Guy, L., Kultima, J. R., & Andersson, S. G. (2010). genoPlotR: comparative gene and genome visualization in R. *Bioinformatics 26*: 2334–2335.

Harrison, P. W., Lower, R. P. J., Kim, N. K. D., & Young, J. P. W. (2010). Introducing the bacterial 'chromid': not a chromosome, not a plasmid. *Trends Microbiology*, *18*, 141–148.

Henkel, C. V., den Dulk-Ras, A., Zhang, X., & Hooykaas, P. J. (2014). Genome sequence of the octopine-type *Agrobacterium tumefaciens* strain Ach5. *Genome Announcements*, *2*, e00225-14.

Hodges, L. D., Cuperus, J., & Ream, W. (2004). Agrobacterium rhizogenes GALLS protein substitutes for Agrobacterium tumefaciens single-stranded DNA-binding protein VirE2. *Journal of Bacteriology*, *186*, 3065–3077.

Hooykaas, P. J. J. (1979). *The role of plasmid determined functions in the interactions of Rhizobiaceae with plant cells. A genetic approach*. PhD Thesis. Leiden University, The Netherlands.

Huang, Y. Y., Cho, S.-T., Lo, W.-S., Wang, Y.-C., Lai, E.-M., & Kuo, C.-H. (2015). Complete genome sequence of *Agrobacterium tumefaciens* Ach5. *Genome Announcements*, *3*, e00570-15.

Huerta-Cepas, J., Forslund, K., Coelho, L. P., Szklarczyk, D., Jensen, L. J., von Mering, C., & Bork, P. (2017). Fast genome-wide functional annotation through orthology assignment by eggNOG-mapper. *Molecular Biology and Evolution*, *34*, 2115–2122.

Jain, C., Rodriguez-R, L. M., Phillippy, A. M., Konstantinidis, K. T., & Aluru, S. (2018). High throughput ANI analysis of 90K prokaryotic genomes reveals clear species boundaries. *Nature Communications*, *9*, 5114.

Jouanin, L. (1984). Restriction map of an agropine-type Ri plasmid and its homologies with Ti plasmids. *Plasmid*, *12*, 91–102.

Jouanin, L., Vilaine, F., Tourneur, J., Tourneur, C., Pautot, V., Muller, J. F., & Caboche, M. (1987). Transfer of a 4.3 kb fragment of the TL-DNA of *Agrobacterium rhizogenes* strain A4 confers the pRi transformed phenotype to regenerated tobacco. *Plant Science*, *53*, 53–63.

Jumas-Bilak, E., Michaux-Charachon, S., Bourg, G., Ramuz, M., & Allardet-Servent, A. (1998). Unconventional genomic organization in the alpha subgroup of the Proteobacteria. *Journal of Bacteriology*, *180*, 2749–2755.

Kajala, K., Coil, D. A., & Brady, S. M. (2014). Draft genome sequence of *Rhizobium rhizogenes* strain ATCC15834. *Genome Announcements*, *2*, e01108-14.

Katoh, S. (2013). MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Molecular Biology and Evolution*, *30*, 772–780.

Kerr, A., & Panagopoulos, C. G. (1977). Biotypes of *Agrobacterium radiobacter* var. *tumefaciens* and their biological control. *Journal of Phytopathology*, *90*, 172–179.

Kim, K. S., & Farrand, S. K. (1996). Ti plasmid-encoded genes responsible for catabolism of the crown gall opine mannopine by Agrobacterium tumefaciens are homologs of the T-region genes responsible for synthesis of this opine by the plant tumor. *Journal of Bacteriology*, *178*, 3275–3284.

Krall, L., Raschke, M., Zenk, M. H., & Baron, C. (2002). The Tzs protein from Agrobacterium tumefaciens C58 produces zeatin riboside 5′-phosphate from 4-hydroxy-3-methyl-2-(E)-butenyl diphosphate and AMP. *FEBS Letters*, *527*, 315–318.

Kurtz, S., Phillippy, A., Delcher, A. L., Smoot, M., Shumway, M., Antonescu, C., & Salzberg, S. L. (2004). Versatile and open software for comparing large genomes. *Genome Biology*, *5*, R12.

Mehrotra, S., Srivastav, V., Rahman, L. U., & Kukreja, A. K. (2015). Hairy root biotechnology-indicative timeline to understand missing links and future outlook. *Protoplasma*, *252*, 1189–1201.

Meier-Kolthoff, J. P., Auch, A. F., Klenk, H. P., & Göker, M. (2013). Genome sequence-based species delimitation with confidence intervals and improved distance functions. *BMC Bioinformatics*, *14*, 60.

Moriguchi, K., Maeda, Y., Satou, M., Hardayani, N. S., Kataoka, M., Tanaka, N., & Yoshida, K. (2001). The complete nucleotide sequence of a plant root-inducing (Ri) plasmid indicates its chimeric structure and evolutionary relationship between tumor-inducing (Ti) and symbiotic (Sym) plasmids in Rhizobiaceae. *Journal of Molecular Biology*, *307*, 771–784.

Nishiguchi, R., Takanami, M., & Oka, A. (1987). Characterization and sequence determination of the replicator region in the hairy-root-inducing plasmid pRiA4b. *Molecular and General Genetics MGG*, *206*, 1–8.

Offringa, I. A., Melchers, L. S., Regensburg-Tuink, A. J. G., Costantino, P., Schilperoort, R. A., & Hooykaas, P. J. J. (1986). Complementation of Agrobacterium tumefaciens tumor-inducing aux mutants by genes from the TR-region of the Ri plasmid of Agrobacterium rhizogenes. *Proceedings of the National Academy of Sciences of the United States of America*, *83*, 6935–6939.

Oger, P. M., Reich, C., Olsen, G. J., & Farrand, S. K. (2001). Complete nucleotide sequence and analysis of pTiBo542: what genomics tells us about structure and evolution of plasmids in the family *Rhizobiaceae*. *Plasmid*, *45*, 169–170.

Otten, L. (2018). The *Agrobacterium* phenotypic plasticity (*Plast*) genes. *Current Top Microbiology Immunology*, *418*, 375–420.

Petit, A., David, C., Dahl, G. A., Ellis, J. G., Guyon, P., Casse-Delbart, F., & Tempé, J. (1983). Further extension of the opine concept: Plasmids in *Agrobacterium rhizogenes* cooperate for opine degradation. *Molecular and General Genetics MGG*, *190*, 204–214.

Riker, A. J. (1930). Studies on infectious hairy root of nursery apple trees. *Journal of Agricultural Research*, *41*, 507–540.

Ron, M., Kajala, K., Pauluzzi, G., Wang, D., Reynoso, M. A., Zumstein, K., Garcha, J., Winte, S., Masson, H., Inagaki, S., Federici, F., Sinha, N., Deal, R. B., Bailey-Serres, J., & Brady, S. M. (2014). Hairy root transformation using *Agrobacterium rhizogenes* as a tool for exploring cell type-specific gene expression and function using tomato as a model. *Plant Physiology*, *166*, 455–469.

Shao, S., van Heusden, G. P. H., & Hooykaas, P. J. J. (2019). Complete sequence of succinamopine Ti-plasmid pTiEU6 reveals its evolutionary relatedness with nopaline-type Ti-plasmids. *Genome Biology and Evolution*, *11*, 2480–2491.

Shao, S., Zhang, X., van Heusden, G. P. H., & Hooykaas, P. J. J. (2018). Complete sequence of the tumor-inducing plasmid pTiChry5 from the hypervirulent Agrobacterium tumefaciens strain Chry5. *Plasmid*, *96–97*, 1–6.

Sharma, S., Shinde, S., & Verslues, P. E. (2013). Functional characterization of an ornithine cyclodeaminase-like protein of *Arabidopsis thaliana*. *BMC Plant Biology*, *13*, 182.

Slater, S. C., Goldman, B. S., Goodner, B., Setubal, J. C., Farrand, S. K., Nester, E. W., Burr, T. J., Banta, L., Dickerman, A. W., Paulsen, I., Otten, L., Suen, G., Welch, R., Almeida, N. F., Arnold, F., Burton, O. T., & Du, Z. (2009). Genome sequences of three *Agrobacterium* biovars help elucidate the evolution of multichromosome genomes in bacteria. *Journal of Bacteriology*, *191*, 2501–2511.

Slightom, J. L., Durand-Tardif, M., Jouanin, L., & Tepfer, D. (1986). Nucleotide sequence analysis of TL-DNA of *Agrobacterium rhizogenes* agropine type plasmid. Identification of open reading frames. *Journal of Biological Chemistry*, *261*(1), 108–121.

Stothard, P., & Wishart, D. S. (2005). Circular genome visualization and exploration using CGView. *Bioinformatics*, *21*, 537–539.

Thompson, M. G., Cruz-Morales, P., Moore, W. M., Pearson, A. N., Keasling, J. D., Scheller, H. V., & Shih, P. M. (2020). Draft genome sequence of *Agrobacterium fabrum* ARqua1. *Microbiology Resource Announcements*, *9*, e00506-20.

Tong, X., Li, Y., Xiang, T., Chen, Z., Wang, L., Zhang, C., Feng, M., & Wu, L. (2018). The complete genome sequence of cucumopine-type *Agrobacterium rhizogenes* strain K599 (NCPPB2659), a nature's genetic engineer inducing hairy roots. *International Journal of Agriculture and Biology*, *20*, 1167–1174.

Trovato, M., Maras, B., Linhares, F., & Costantino, P. (2001). The plant oncogene *rolD* encodes a functional ornithine cyclodeaminase. *Proceedings of the National Academy of Sciences of the United States of America*, *98*, 13449–13453.

Trovato, M., Mattioli, R., & Costantino, P. (2018). From *A. rhizogenes* RolD to plant P5CS: exploiting proline to control plant development. *Plants*, *7*, 108.

Watanabe, S., Sueda, R., Fukumori, F., & Watanabe, Y. (2015). Characterization of flavin-containing opine dehydrogenase from bacteria. *PLoS One*, *10*, e0138434.

Watanabe, S., Tozawa, Y., & Watanabe, Y. (2014). Ornithine cyclodeaminase/μ-crystallin homolog from the hyperthermophilic archaeon *Thermococcus litoralis* functions as a novel Δ1-pyrroline-2-carboxylate reductase involved in putative *trans*-3-hydroxy-l-proline metabolism. *FEBS Open Bio*, *4*, 2211–5463.

Waterhouse, A. M., Procter, J. B., Martin, D. M. A., Clamp, M., & Barton, G. J. (2009). Jalview Version 2-a multiple sequence alignment editor and analysis workbench. *Bioinformatics*, *25*, 1189–1191.

Weisberg, A. J., Davis, E. W., Tabima, J., Belcher, M. S., Miller, M., Kuo, C.-H., Loper, J. E., Grünwald, N. J., Putnam, M. L., & Chang, J. H. (2020). Unexpected conservation and global transmission of agrobacterial virulence plasmids. *Science*, *368*, eaba5256.

Weller, S. A., Stead, D. E., O'Neill, T. M., Hargreaves, D., & McPherson, G. M. (2000). Rhizogenic *Agrobacterium* biovar 1 and cucumber root mat in the UK. *Plant Pathology*, *49*, 43–50.

Wetzel, M. E., Olsen, G. J., Chakravartty, V., & Farrand, S. K. (2015). The *repABC* plasmids with quorum-regulated transfer systems in members of the Rhizobiales divide into two structurally and separately evolving groups. *Genome Biology and Evolution*, *7*, 3337–3357.

White, F. F., & Nester, E. W. (1980). Hairy root: plasmid encodes virulence traits in *Agrobacterium rhizogenes*. *Journal of Bacteriology*, *141*, 1134–1141.

White, F. F., Taylor, B. H., Huffman, G. A., Gordon, M. P., & Nester, E. W. (1985). Molecular and genetic analysis of the transferred DNA regions of the root-inducing plasmid of *Agrobacterium rhizogenes*. *Journal of Bacteriology*, *164*, 33–44.

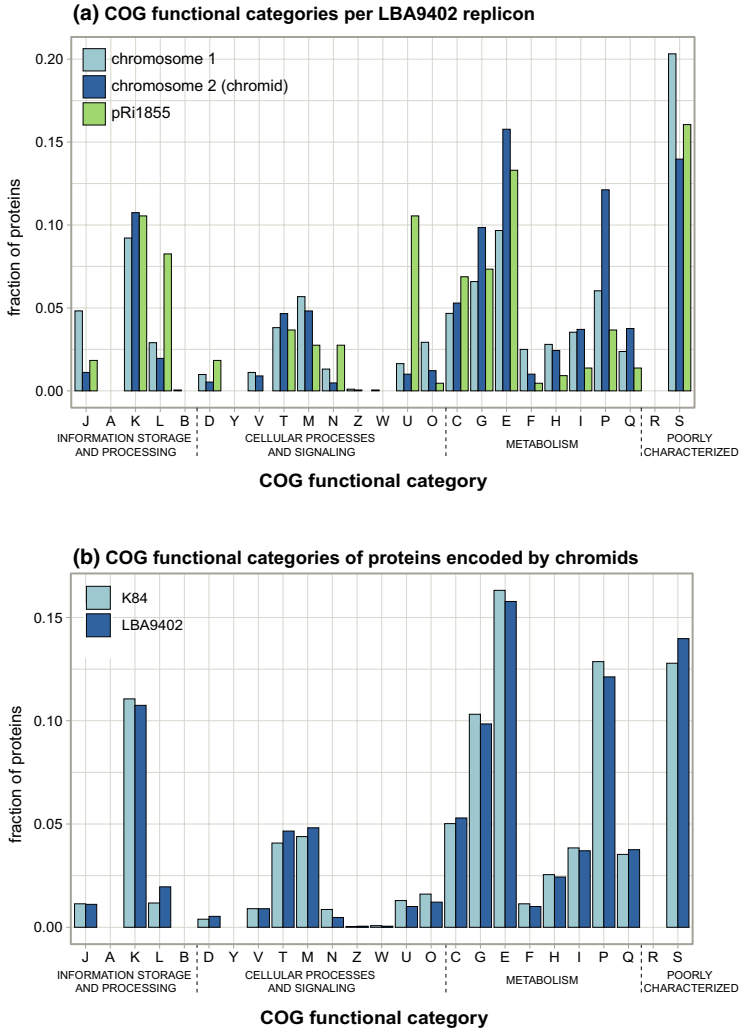
Wood, D. W., Setubal, J. C., Kaul, R., Monks, D. E., Kitajima, J. P., Okura, V. K., Zhou, Y., Chen, L., Wood, G. E., Almeida, N. F. Jr, Woo, L., Chen, Y., Paulsen, I. T., Eisen, J. A., Karp, P. D., Bovee, D. Sr, Chapman, P., Clendenning, J., Deatherage, G., … Nester, E. W. (2001). The genome of the natural genetic engineer *Agrobacterium tumefaciens* C58. *Science*, *294*, 2317–2323.

Xie, Z., & Tang, H. (2017). ISEScan: automated identification of insertion sequence elements in prokaryotic genomes. *Bioinformatics*, *33*, 3340–3347.

Yost, C. K., Rath, A. M., Noel, T. C., & Hynes, M. F. (2006). Characterization of genes involved in erythritol catabolism in *Rhizobium leguminosarum* bv. *Viciae*. *Microbiology*, *152*, 2061–2074.

How to cite this article: Hooykaas MJ, Hooykaas PJ. The genome sequence of hairy root *Rhizobium rhizogenes* strain LBA9402: Bioinformatics analysis suggests the presence of a new opine system in the agropine Ri plasmid. *MicrobiologyOpen*. 2021;10:e1180. https://doi.org/10.1002/mbo3.1180

APPENDIX



COG functional categories

Information storage and processing	J	Translation, ribosomal structure and biogenesis
	A	RNA processing and modification
	K	Transcription
	L	Replication, recombination and repair
	B	Chromatin structure and dynamics
Cellular processes and signaling	D	Cell cycle control, cell division, chromosome partitioning
	Y	Nuclear structure
	V	Defense mechanisms
	T	Signal transduction mechanisms
	M	Cell wall/membrane/envelope biogenesis
	N	Cell motility
	Z	Cytoskeleton
	W	Extracellular structures
	U	Intracellular trafficking, secretion, and vesicular transport
	O	Posttranslational modification, protein turnover, chaperones
Metabolism	C	Energy production and conversion
	G	Carbohydrate transport and metabolism
	E	Amino acid transport and metabolism
	F	Nucleotide transport and metabolism
	H	Coenzyme transport and metabolism
	I	Lipid transport and metabolism
	P	Inorganic ion transport and metabolism
	Q	Secondary metabolites biosynthesis, transport and catabolism
Poorly characterized	R	General function prediction only
	S	Function unknown

FIGURE A1 LBA9402 and K84 proteins were annotated with eggNOG (evolutionary genealogy of genes: Non-supervised Orthologous Groups (OGs)) mapper. Shown in the figure are the frequencies at which the COG functional categories were assigned to proteins (normalized to the total number of proteins). In cases where multiple categories were assigned to a single protein, each category was counted once. On the right, a description of each COG category is shown. (a) The relative frequencies are shown per LBA9402 replicon. (b) Functional categories are shown for the chromids (secondary chromosomes) of K84 and LBA9402

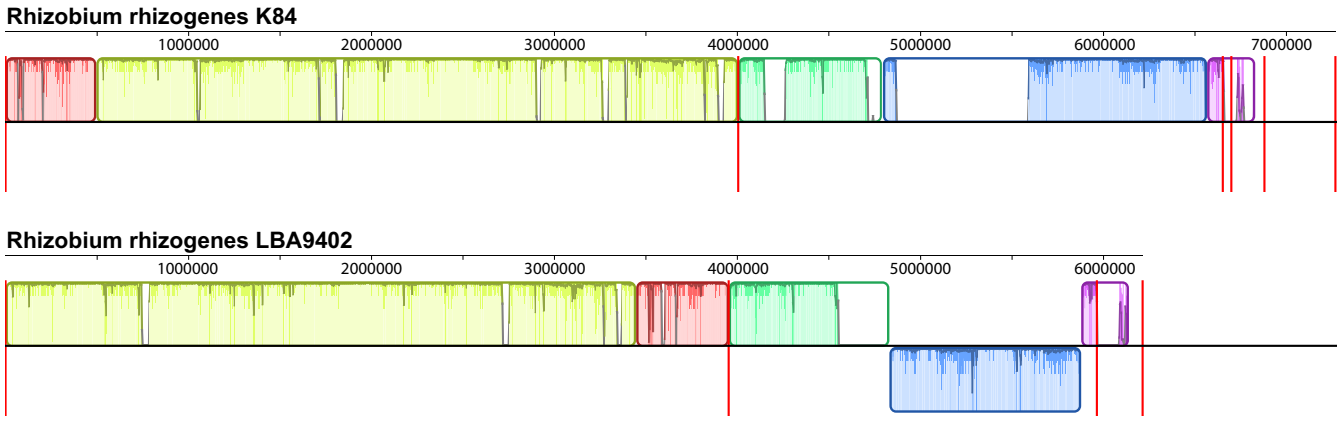


FIGURE A2 The LBA9402 and K84 genomes were aligned using progressiveMauve. Colored boxes are local collinear blocks. Blocks below the center line are aligned in reverse complementary orientation compared to the reference (top) sequence, and blocks above the center line are in the forward orientation. Similarity profiles are shown inside the boxes

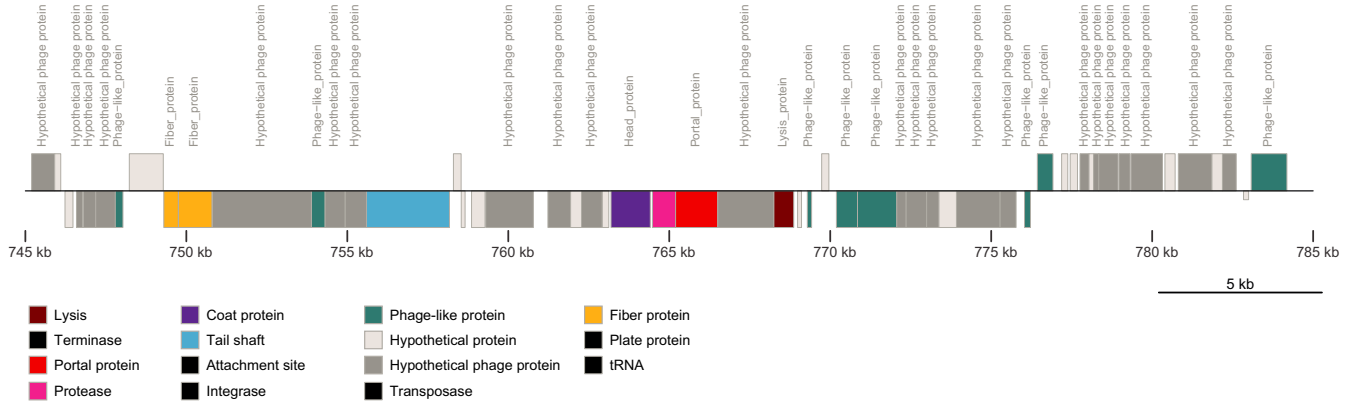


FIGURE A3 Multiple bacteriophage sequences were detected in LBA9402 chromosome 1. Phage search tool PHASTER was used to find prophage sequences. The schematic of the particular genomic region was generated by visualizing the raw output from PHASTER with genoplotR. The default PHASTER legend was slightly adapted by changing the color of protein categories not present in the displayed region to black. Genes annotated as "Hypothetical phage proteins" had a hit in the PHASTER phages database, but no gene function was known, whereas "Hypothetical proteins" did not have a hit in the PHASTER database

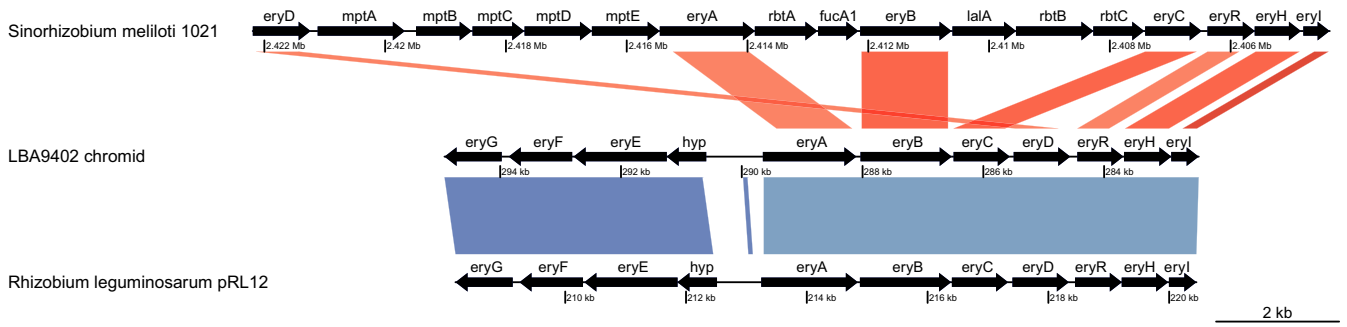


FIGURE A4 Comparison between the erythritol transport and catabolism regions of *Sinorhizobiummeliloti* 1021, *Rhizobiumleguminosarum* plasmid pRL12, and LBA9402. The red (same orientation) and blue (reverse orientation) ribbons show BLASTn hits, with darker shading indicating higher similarity. The data were visualized with genoplotR

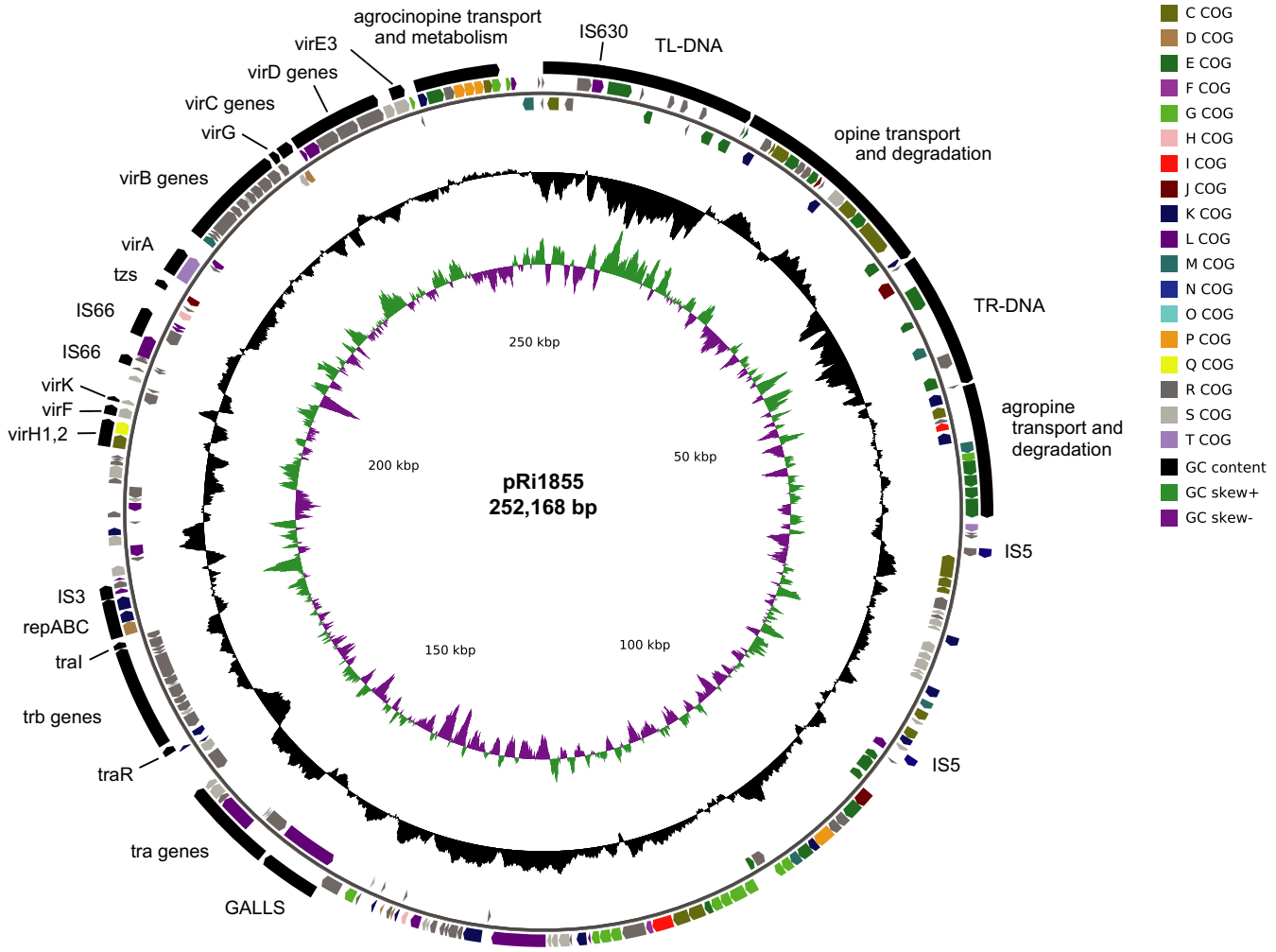


FIGURE A5 Map of pRi1855. From center to border, the rings represent GC skew, GC content, locations of genes predicted by PGAP (the colors of the arrows represent the COG functional categories predicted for the proteins; a description of the COG letters is provided in Appendix Figure A1), and finally the location of the number of features with their annotation

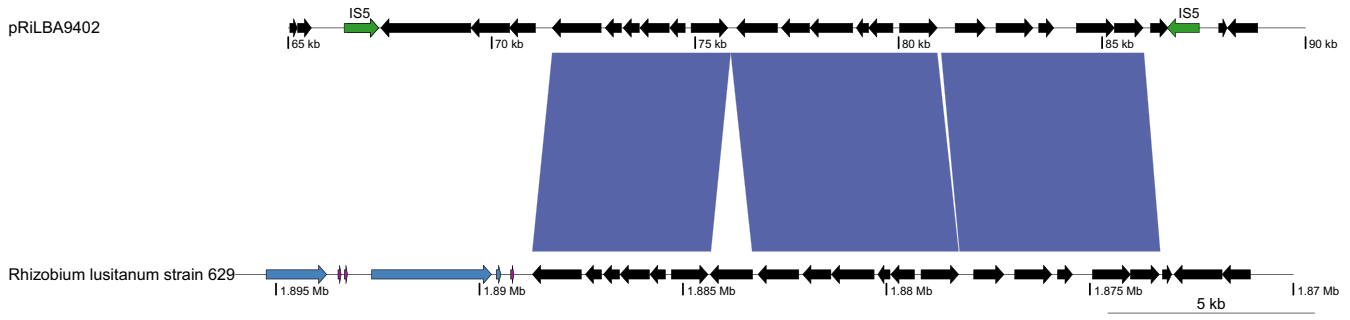


FIGURE A6 The region between two IS5 family insertion sequences shows similarity to a region in the *Rhizobiumlusitanum* strain 629 chromosome. A megablast search, with the sequence between both insertions sequences as a query, against the NCBI RefSeq genomes database (restricted to Rhizobiales sequences), and excluding Ri plasmid sequences, yielded the chromosome of *Rhizobiumlusitanum* strain 629 as best hit (67% query coverage, 94.50% sequence identity). Depicted are the areas with a high similarity between both sequences (plotted with genoplotR). Black arrows represent coding sequences, green arrows insertion sequences, blue arrows rRNAs, and pink arrows tRNAs

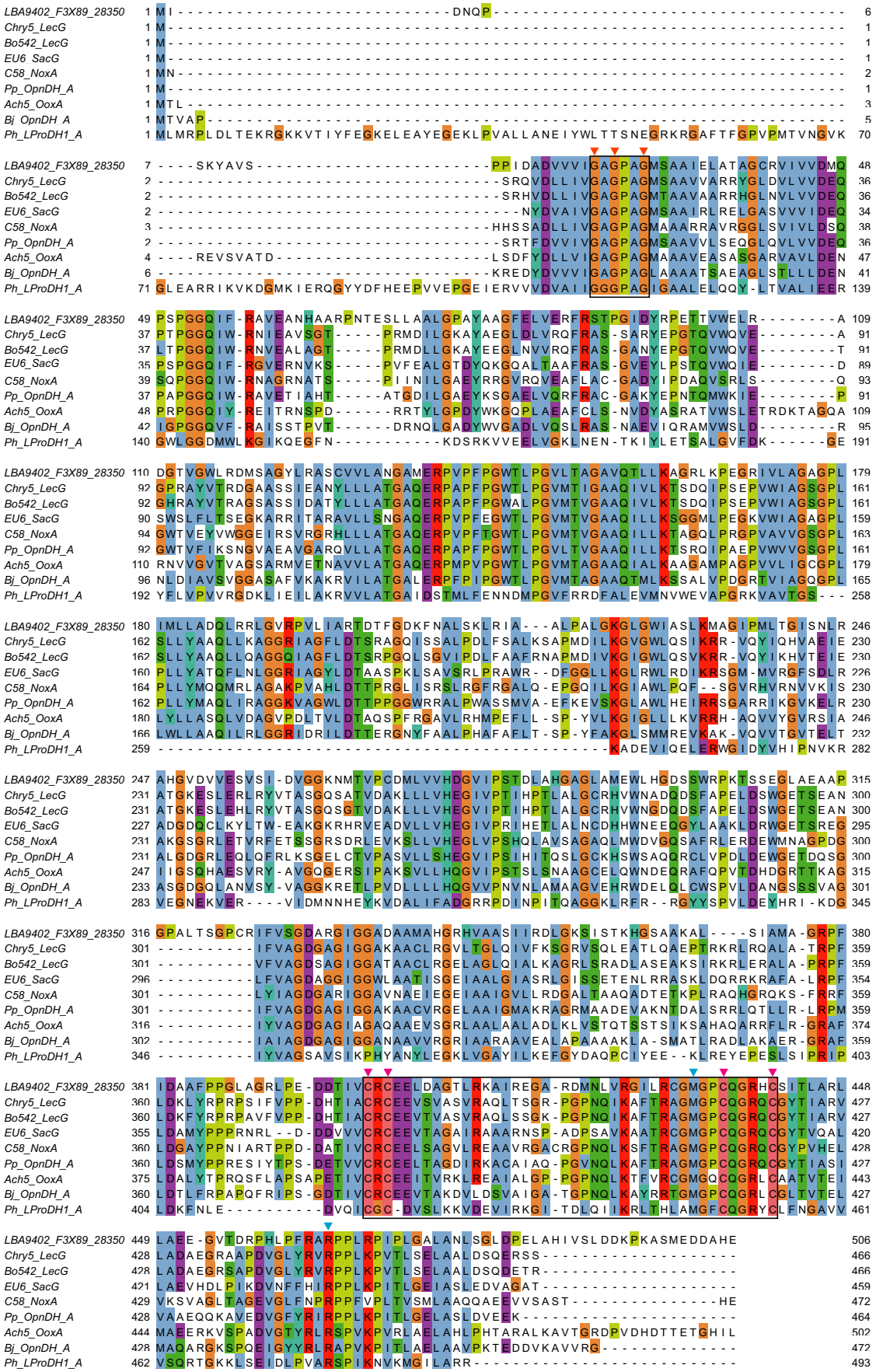

LBA9402_F3X89_28350
Chry5_LecG
Bo542_LecG
EU6_SacG
C58_NoxA
Pp_OpnDH_A
Ach5_OoxA
Bj_OpnDH_A
Ph_LProDH1_A

FIGURE A7 Multiple sequence alignment of α subunits of various opine dehydrogenases. Indicated with orange triangles is the ADP binding motif Gly-X-Gly-X-X-Gly and with pink triangles the [2Fe-2S] iron–sulfur cluster binding motif (Cys-X-Cys-$X_{31\text{-}33}$-Cys-X_4-Cys). The residues putatively interacting with FMN are indicated with blue triangles. This analysis was based on the crystal structure of *Pyrococcushorikoshi* (Ph) L-Proline dehydrogenase (Ph_L-ProDH) (Tsuge et al. 2005. Crystal structure of a novel FAD-, FMN-, and ATP-containing L-proline dehydrogenase complex from *Pyrococcushorikoshii*. J Biol Chem 280: 31045–31049). Shown are the amino acid sequences of opine dehydrogenase subunits encoded by the *Agrobacterium* Ti plasmids carried by strains Chry5, Bo542, EU6, C58, and Ach5, which function as succinamopine, nopaline, and octopine dehydrogenases, respectively. In addition, sequences from *Pseudomonasputida* (Pp) and *Bradyrhizobiumjaponicum* (diazoefficiens) USDA110 opine dehydrogenases (which function as nopaline and octopine dehydrogenases, respectively). Finally, the sequence of the L-Proline dehydrogenase of *Pyrococcushorikoshi* (Ph) is shown. The background color of the amino acids is according to the Clustal X color scheme.

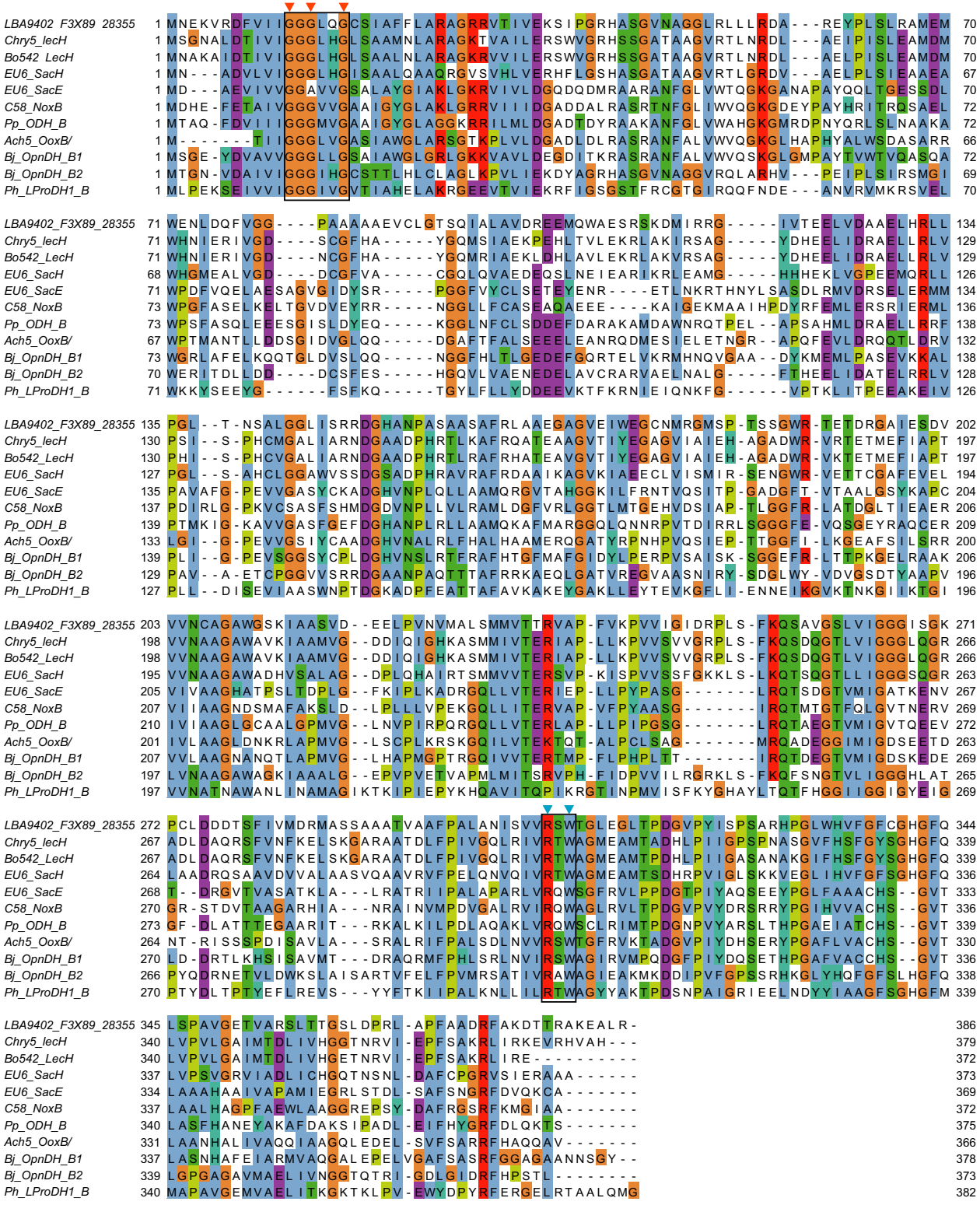


FIGURE A8 Multiple sequence alignment of β subunits of various opine dehydrogenases. Indicated with orange triangles is the ADP binding motif Gly-X-Gly-X-X-Gly. The arginine and tryptophan residues (putatively) interacting with FMN (as based on *Pyrococcushorikoshi* (Ph) L-Proline dehydrogenase (Ph_L-ProDH) crystal structure data) are indicated with blue triangles. Abbreviations of species are described in the legend of Appendix Figure A7.

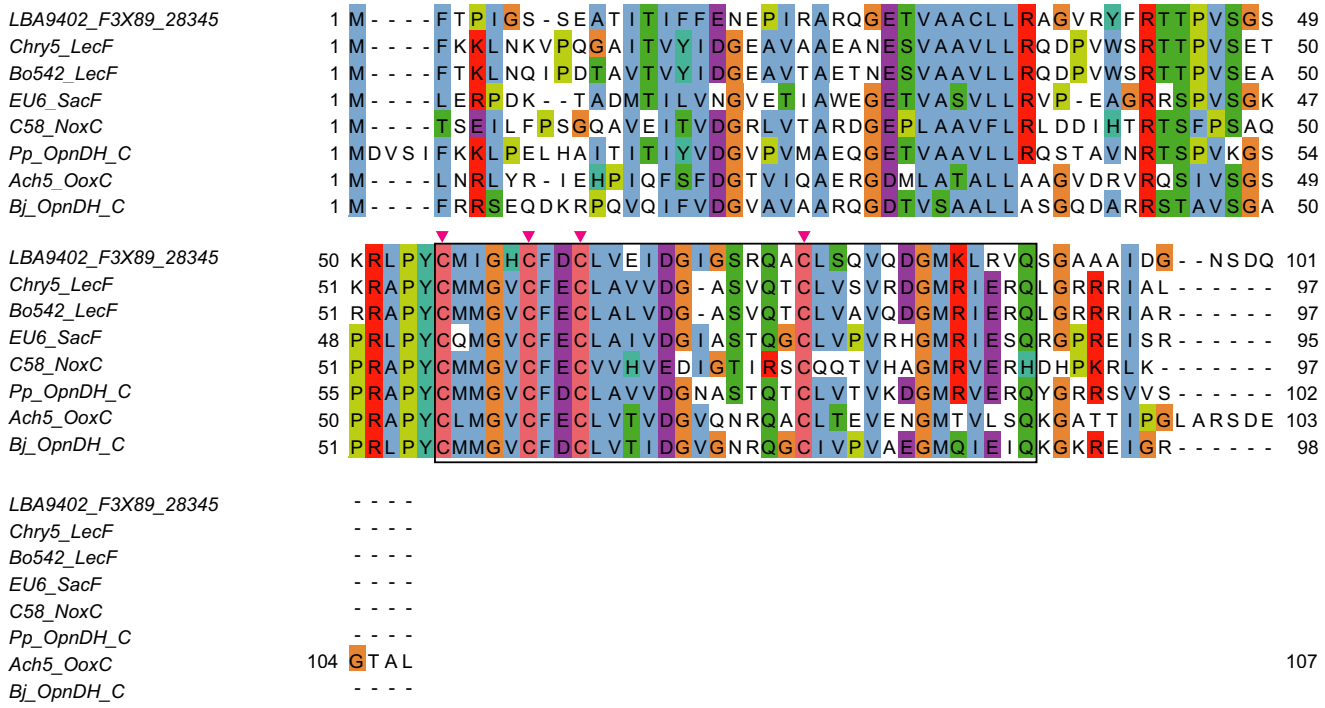


FIGURE A9 Multiple sequence alignment of ϒ subunits of various opine dehydrogenases. Indicated with pink triangles is the [2Fe-2S] iron–sulfur cluster binding motif (Cys-X_4-Cys-X_2-Cys-$X_{11\text{-}12}$-Cys). Abbreviations of species are described in the legend of Appendix Figure A7.