



Universiteit
Leiden
The Netherlands

Functional fluorescent materials and migration dynamics of neural progenitor cells

Bossert, N.

Citation

Bossert, N. (2022, January 13). *Functional fluorescent materials and migration dynamics of neural progenitor cells*. *Casimir PhD Series*. Retrieved from <https://hdl.handle.net/1887/3249722>

Version: Publisher's Version

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/3249722>

Note: To cite this publication please use the final published version (if applicable).

2

FLUORESCENCE-TUNABLE AG-DNA BIOSENSOR WITH TAILORED CYTOTOXICITY FOR LIVE-CELL APPLICATIONS

DNA-stabilized silver clusters (Ag-DNA) show excellent promise as a multifunctional nanoagent for molecular investigations in living cells. The unique properties of these fluorescent nanomaterials allow for intracellular optical sensors with tunable cytotoxicity based on simple modifications of the DNA sequences. Three Ag-DNA nanoagent designs are investigated, exhibiting optical responses to the intracellular environments and sensing-capability of ions, functional inside living cells. Their sequence-dependent fluorescence responses inside living cells include (1) a strong splitting of the fluorescence peak for a DNA hairpin construct, (2) an excitation and emission shift of up to 120 nm for a single-stranded DNA construct, and (3) a sequence robust in fluorescence properties. Additionally, the cytotoxicity of these Ag-DNA constructs is tunable, ranging from highly cytotoxic to biocompatible Ag-DNA, independent of their optical sensing capability. Thus, Ag-DNA represents a versatile live-cell nanoagent addressable towards anti-cancer, patient-specific and anti-bacterial applications.

This chapter is based on the publication: Bossert N., de Bruin D., Gotz M., Bouwmeester D., Heinrich D.: Fluorescence-tunable Ag-DNA Biosensor with tailored Cytotoxicity for Live-Cell Applications, Scientific Reports (6.1) (2016) 1-8

2.1. INTRODUCTION

DNA-encapsulated silver nanoclusters (Ag-DNA) have recently attracted much attention, as they exhibit numerous unique fluorescence properties, like high photostability, fluorescence quantum yields of up to 90% and tunable excitation and emission spectra [1–6]. The high sensitivity to their surroundings is one of the most intriguing properties of Ag-DNA nanoclusters, allowing them to exhibit an optical response due to environmental changes: conformational changes of the DNA template lead to varying distances and interactions between DNA bases and silver clusters [7, 8]. In solution, Ag-DNA have been shown to even respond to single-base mutations [9, 10], and to detect molecule concentrations down to 0.37 nM [11]. Therefore, Ag-DNA are very promising for intracellular bio-sensing applications and to investigate intracellular dynamics, as so far is done mostly with fluorescent nanobead probes [12–15]. Most fluorescent agents show functional limitations, like photobleaching or low absorption coefficients and often fulfill one function only. This makes the search for alternative nanomaterials an ongoing quest.

The encapsulated silver in Ag-DNA nanoagents further allow for multifunctional applications, as these optical sensors exhibit tunable degrees of cytotoxicity [16]. In the past years, silver has been increasingly used in cosmetics, clothes, implant coatings, and for burn wound dressings, to obtain an antibacterial effect [17–20]. Its inherent toxicity makes the metal a particularly promising alternative to antibiotics, as bacterial resistances rise [21, 22], as well as an anti-cancer agent [23–25]. However, the toxicity mechanism of silver is not yet clearly understood [26, 27]. As nano-sized silver is highly bioactive, its broad utilization bears a potential risk for humans ([28–30], making a more controlled and tunable way of its application appealing.

Furthermore, these fluorescent Ag-DNA nanomaterials are easily compatible with DNA nanotechnology and DNA origami, and provide an easy attachment site for specific addition of desired targets and recognition elements. This has been implemented for specific labeling of cellular membrane and nuclei ([31–34], using the Ag-DNA emitters as a replacement of an organic dye. So far, however, the optical sensor functionality, particularly in combination with independently tunable cytotoxicity of Ag-DNA constructs, has not been investigated in live-cell interiors.

Fluorescent Ag-DNA are synthesized upon reduction of silver ions by sodium borohydride (NaBH_4) in the presence of DNA (Figure 2.1), resulting in rod-shaped silver cores comprised of 4–12 neutral Ag atoms ([35, 36]. Remarkably, these nanomaterials can exhibit peak emission wavelengths across the visible and near-IR spectrum ([37] determined by the variant sizes and aspect ratios of the noble metal nanoclusters ([36]. Similar to larger noble metal nanoparticles, larger silver rods correspond to a longer wavelength resonance than smaller particles ([38–40], with the DNA template also playing a role in the fluorescence variance of the emitters.

Using these dependencies, we selected and optimized Ag-DNA templates to function as an optical sensor in living cells' interior environments (Figure 2.1 b-d) (for DNA sequences please see section 2.4 Table 2.1). At the same time, by utilizing the endogenous silver core, we tuned Ag-DNA from cytotoxic to completely biocompatible (Figure 2.1b-d), creating an intracellular toxicity-tunable optical sensor. Here, we demonstrate the functionality of the designed nanoagents in a live-cell environment.

2.2. RESULTS AND DISCUSSION

2.2.1. AG-DNA NANOAGENT DESIGN

The choice of DNA sequences is the result of optimizing fluorescence and/or sensor properties of Ag-DNA within the cell by varying the base sequences for enhanced performance. The strong interaction of cytosine and guanine bases with Ag(I) ions allows effective stabilization of a silver cluster. Furthermore, the addition of adenine and thymine bases throughout the DNA strand increase the yields of fluorescent species and fluorescence quantum yields. By varying the base sequences, the resulting different sizes and shapes of Ag-DNA structures define their particular functionality as nanoagents.

First, we demonstrate a robust and bright Ag-DNA structure, applicable as a fluorescent marker inside living cells. The DNA template, consisting of 28 single bases, yields an Ag-DNA construct (Ag-28b in Figure 2.1 b) with strong and stable fluorescence properties [35], which we show are retained upon insertion into living cells. Thus, in this DNA-design, the encapsulated silver cluster preserves its fluorescence properties when inside living cells, not reacting to the complex intracellular environment.

While the relatively long single strand wrapping around the silver cluster seems to protect the cluster enough to maintain a strong fluorescence, any exposed parts of the silver surface could still interact with cellular molecules, altering cellular processes.

Small changes in the DNA template can have a significant effect on the functionality of the Ag-DNA nanomaterials. Here, we demonstrate that a simple modification of Ag-28b produces an Ag-DNA construct which shows optical sensor functionality inside living cells: shortening the 28b sequence by nine bases produces the Ag-19b construct [41] (Figure 2.1c), which exhibits an optical response to the intracellular environment. By using the shorter sequence, we reduce the shielding of cellular molecules from the silver surface as compared to Ag-28b, resulting in a higher level of cytotoxicity and yielding a stronger sensitivity to the change in conditions inside living cells. To decrease the toxicity level to a biocompatible Ag-DNA for an optical biosensor sensitive to intracellular ions, we designed an Ag-hairpin (Ag-HP) construct (Figure 2.1 c), as DNA hairpins with Poly-C loops have been shown to stabilize strongly fluorescent Ag-DNA species [37]. As the molecular dynamics of single- and double-stranded DNA heavily depend on the presence of salts [42], DNA-ions interactions lead to conformational changes of the hairpin loop resulting in optical emission shifts inside living cells, yielding an optical sensor. The amount of cytosine bases in the loop, and the length and sequence of the double stranded stem were optimized for sensitivity. The Poly-C-loop wrapping around the silver cluster through strong Cytosine-Ag(I) ions interactions prevents the silver surface from interacting with cellular molecules, yielding a biocompatible construct.

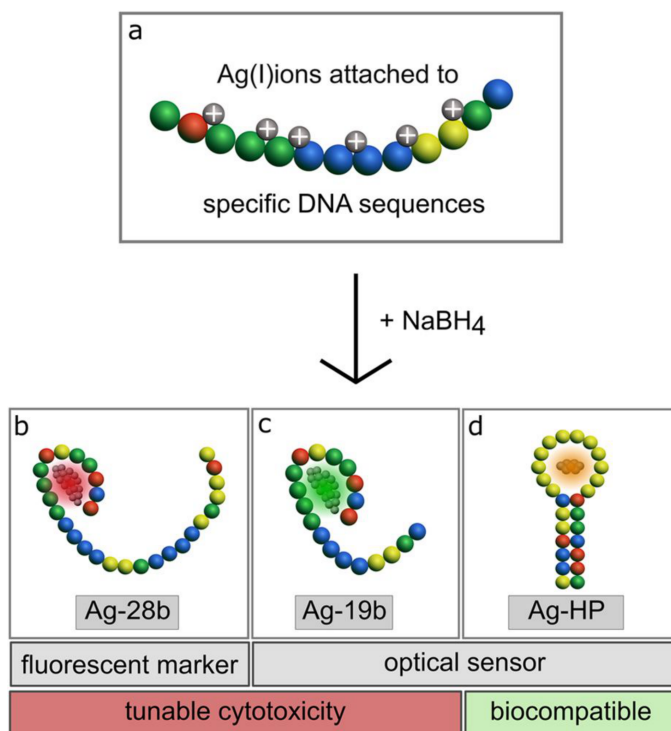


Figure 2.1: Schemes of general Ag-DNA synthesis (a) and specifically designed Ag-DNA constructs (b–d). Fluorescent Ag-DNA are synthesized upon reduction of silver ions by sodium borohydride (NaBH₄) in the presence of a specific DNA template. The designed Ag-DNA structures, utilized here, show different functionalities as a fluorescent marker or as an optical sensor, as well as exhibit tunable cytotoxicity or biocompatibility. DNA sequences color code are displayed for (b) Ag-28b, (c) Ag-19b and (d) Ag-HP by Cytosine=yellow, Guanine=green, Thymine=blue, Adenine=red.

2.2.2. FLUORESCENCE-TUNABLE PROPERTIES OF AG-DNA

OPTICAL SENSORS

By acquiring the excitation and emission spectra, the fluorescence properties of the Ag-DNA structures were investigated in solution and in comparison in real cell environments (Figure 2.2 and 2.3). All three types of Ag-DNA were quickly internalized into living cells and showed a homogeneous distribution throughout the cytoplasm and the cell nucleus (section 2.4 Figure 2.5, negative control section 2.4 Figure 2.6).

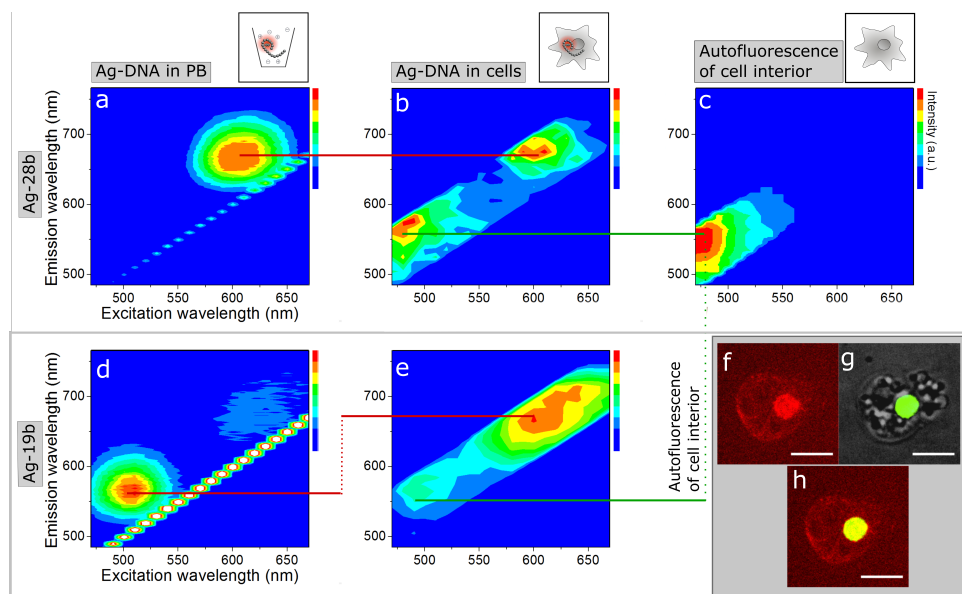


Figure 2.2: Fluorescence excitation/emission spectra of Ag-DNA nanoagents of type Ag-28b and Ag-19b, before (a,d) and after (b,e) internalization in *D.discoideum* wildtype cells. Fluorescence spectra are measured within the excitation range of 470–670 nm and the emission range of 490–760 nm, each in 10 nm steps. Ag-28b exhibits a fluorescence peak at $\lambda_{(ex)}=605$ nm / $\lambda_{(em)}=665$ nm in PB (a), which is preserved upon internalization in living cells (b) (illustrated by the red line). Without addition of Ag-DNA variants, cell interiors exhibit a peak at $\lambda_{(ex)}=470$ nm / $\lambda_{(em)}=550$ nm, corresponding to autofluorescence (c) (illustrated by the green lines). Ag-19b shows a dominant emitter at $\lambda_{(ex)}=495$ nm / $\lambda_{(em)}=560$ nm in PB (d). Upon internalization in living cells, the peak shifts to $\lambda_{(ex)}=625$ nm / $\lambda_{(em)}=680$ nm (e) yielding a shift of $\Delta\lambda_{(ex)}=130$ nm / $\Delta\lambda_{(em)}=120$ nm (illustrated by the red lines). The regions of interest (ROIs) for intracellular fluorescence spectra are displayed in section 2.4 Figure 2.7. Confocal fluorescence images display the accumulation of Ag-19b at cellular membranes and inside the cell nucleus (f). The nucleus is labeled by GFP-histone (green) (g) and nuclear localization is shown by the overlay (yellow) (h) of the Ag-19b signal (red) (f) with GFP-labeled histones. The scale bar is 10 μ m.

In phosphate buffer (PB), Ag-28b exhibits a single fluorescence peak at $\lambda_{(ex)}=605$ nm / $\lambda_{(em)}=665$ nm (Figure 2.3 a). When exposing these Ag-DNA structures to the interior of a living cell, the fluorescence properties of Ag-28b are not influenced, as indicated by the red line in Figure 2.3 a,b. The second peak at $\lambda_{(ex)}=470$ nm / $\lambda_{(em)}=550$ nm arises from the autofluorescence of *D.discoideum* wildtype cells (Figure 2.3b,c) (indicated by the green line), as observed in all intracellular fluorescence spectra.

Shortening the DNA template reveals a significant effect on the environmental sensing function of the Ag-DNA construct. Inside cells, Ag-19b shows a significant fluorescence shift of $\Delta\lambda_{(ex)}=130$ nm / $\Delta\lambda_{(em)}=120$ nm (Figure 2.2d,e) (indicated by the red lines). This relocates the dominant peak observed in PB at $\lambda_{(ex)}=495$ nm / $\lambda_{(em)}=560$ nm to $\lambda_{(ex)}=625$ nm / $\lambda_{(em)}=680$ nm in the living cell.

A fluorescence peak in this wavelength range is typically exhibited by Ag-DNA with DNA strands longer than 19 bases, such as in Ag-28b, which stabilize larger silver clusters. Hence, these larger clusters likely require higher molecular stability to reach the fluorescent state, which can be provided by the interactions of Ag-19b with the nuclear

DNA, as well as relatively dense cellular structures like membrane proteins and lipids. Accumulation of Ag-19b in these cellular regions was observed, as shown in Figure 2.2 f-h.

The hairpin structure (Ag-HP) was designed as a biosensor specific to intracellular ions. When in PB, Ag-HP emits at $\lambda_{(ex)}=540$ nm/ $\lambda_{(em)}=590$ nm (Figure 2.3 a). Upon internalization by cells, its fluorescence properties show a split into two peaks. The previously observed peak is largely retained. Additionally, a second peak is observed at $\lambda_{(ex)}=580$ nm/ $\lambda_{(em)}=650$ nm, thus providing a shift in emission of $\Delta\lambda_{(ex)}=60$ nm (Figure 2.3 a,b) (indicated by the red lines).

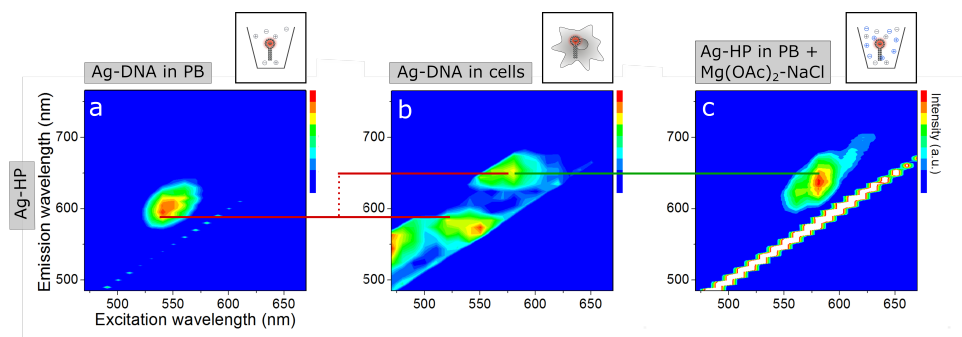


Figure 2.3: Fluorescence excitation/emission spectra of Ag-HP before (a) and after (b) internalization in *D.discoideum* wildtype cells, and in presence of specific ions (c). Fluorescence spectra were measured within the excitation range of 470–670 nm and the emission range of 490–760 nm, each in 10 nm steps. In PB, Ag-HP exhibits a peak at $\lambda_{(ex)}=540$ nm/ $\lambda_{(em)}=590$ nm (a). Upon internalization in cells, a split into a second peak at $\lambda_{(ex)}=580$ nm/ $\lambda_{(em)}=650$ nm occurs, yielding a shift of $\Delta\lambda_{(ex)}=40$ nm/ $\Delta\lambda_{(em)}=60$ nm, whereas the first peak is mainly preserved (illustrated by the red lines) (b). Addition of 40mM Mg(OAc)₂ and 30mM NaCl reproduces the red-shifted peak at $\lambda_{(ex)}=580$ nm/ $\lambda_{(em)}=640$ nm observed only inside cells (illustrated by the green line), yet not the peak in PB (c). The regions of interest (ROIs) for intracellular fluorescence spectra are displayed in Figure 2.7.

To confirm the role of ions in the observed red-shifted fluorescence peak, we exposed Ag-HP to various salt types and concentrations in PB. By the addition of 40 mM Mg(OAc)₂ in combination with 30 mM NaCl (Figure 2.3 c), the red-shifted intensity distribution near $\lambda_{(ex)}=580$ nm/ $\lambda_{(em)}=650$ nm, as observed inside living cells in Figure 2.3b, was reproduced (Figure 2.3 b,c) (indicated by the green line). Exposing Ag-HPs only to monovalent (Na⁺) or only to divalent (Mg²⁺) cations did not reproduce the spectrum in Figure 2.3 b. Cations in particular reduce the flexibility of double stranded DNA, and can induce a conformational change, depending on the type of cations bound [43]. In high salt conditions, even chloride anions have been suggested to localize on parts of the DNA surface, as well as in the grooves [44]. We observed that the addition of chloride anions was necessary for reproducing the shift of the peak in PB to higher wavelengths, that was measured in living cells, in Mg(OAc)₂-NaCl. Noble metal nanoparticles and clusters are known to exhibit sensitivity to their environment, like the dielectric constant of a medium, as is reflected by changes in their plasmonic resonance [39]. However, for small particles these effects typically lead to relatively small shifts in resonance [40],

which has been demonstrated recently for purified, individual Ag-DNA species, leading to an absorption shift of up to 8 nm [36].

Here, we utilize the fact that synthesis of Ag-DNA yields a heterogeneous distribution of different Ag cluster sizes and DNA conformations [45]. After synthesis, the formed silver nanoclusters can occur in a fluorescent or a dark state [45]. Hence, we attribute the observed optical shifts to rearrangements in switching these optically different emitters 'on' or 'off', resulting from conformational changes of the DNA template. This way we facilitate much larger shifts of up to $\Delta\lambda_{(ex)}=120$ nm, which allow the varying responses of Ag-19b and Ag-HP to be easily separated and identified by conventional microscopy methods. In particular, DNA hairpin structures have been shown to typically stabilize a broader range of active fluorescent emitters than the single stranded variants [46], making them very suitable for biosensing applications.

2.2.3. TUNABLE CYTOTOXICITY OF AG-DNA

In addition to this biosensing functionality, we utilize the toxicity effect of endogenous silver clusters in Ag-DNA for live-cell applications. We show the tunability of cytotoxicity in Ag-DNA, which is controlled independently of the optical sensor ability of the Ag-DNA constructs.

In the cytotoxicity experiment, 100% cell viability corresponds to control experiments with cells not exposed to Ag-DNA or AgNO₃ agents under the same conditions. Figure 2.4 displays the concentration-dependent toxicity of Ag-28b, Ag-19b, and Ag-HP on *D.discoideum* wildtype cells, with exposures ranging between 75 nm and 36 μ M. Ag-HP exhibit biocompatibility, with cell viabilities within 80–100% at all concentrations. When incubated with Ag-19b and Ag-28b, cell survival was increasingly impaired with rising Ag-DNA concentrations. For Ag-28b a slight decrease in cell viability was observed, with over 60% viable cells at 36 μ M. Ag-19b had the strongest negative effect on cell survival, still yielding 30% of living cells at the highest concentration. As a reference measurement for cytotoxicity we used AgNO₃, which readily releases free Ag(I) ions in solution. This resulted in the highest cytotoxic effect for the same concentration of silver, as compared to the DNA-stabilized silver clusters.

To exclude other sources for cytotoxicity beside Ag-DNA, the following steps were taken. We removed potentially harmful byproducts or educts possibly causing toxicity by filtration of the synthesized Ag-DNA. Also we confirmed experimentally that a toxic effect caused by oligonucleotides is unlikely. Furthermore, for the cytotoxicity experiments, incubation steps and subsequent cell viability assays were performed in the dark, to exclude possible light-induced toxicity, like ROS generation. These points allow us to identify the silver in the Ag-DNA constructs as the only significant source of cytotoxicity, which is supported by the biocompatibility of Ag-HP.

We conclude from these results that the DNA templates can protect cells from silver and the selected DNA-sequence determines the toxic effect, ranging from non-toxic for Ag-HP to highly toxic for Ag-19b. This mechanism of protection results from distinct interactions of silver with DNA bases and the steric conditions of the Ag-DNA, varying the degree of silver exposure to cellular structures. Due to high binding energies of silver clusters to cytosine and guanine bases, which are several times stronger than Watson-

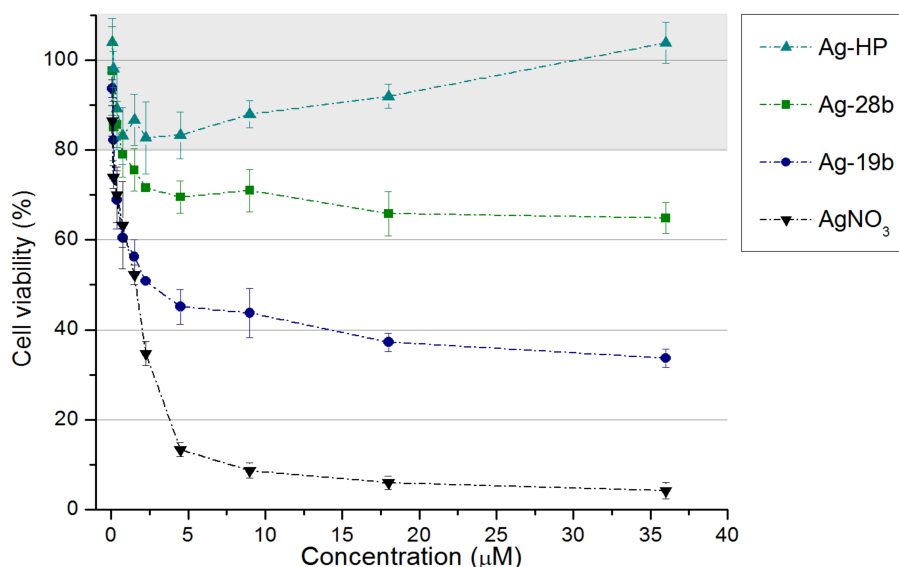


Figure 2.4: The number of living cells was quantified by incubation with Fluorescein diacetate and measuring the average fluorescence intensity. Cell viability of 100% was defined by a control incubated simultaneously without addition of any agents. Data shown represents the mean $\pm$ standard deviation ($n=3$).

Crick base pairs [46], we can assume that the silver clusters mainly remain contained in the Ag-DNA structures. Hence, the observed cytotoxicity effect of single stranded Ag-DNA nanomaterials most likely results from direct contact of cellular structures and molecules with exposed areas of the silver clusters as well as possibly released individual Ag(I) ions.

In Ag-HP, the silver cluster is surrounded by a Poly-C loop. As guanine and cytosine bases in particular show a high affinity to Ag(I) ions [45, 47], multiple Ag(I) ions-DNA base connections are formed which lead to a strongly shielded silver cluster, allowing for a decrease in Ag-DNA cytotoxicity. Both Ag-28b and Ag-19b constructs are single stranded DNA templates, which are expected to wrap around the silver cluster upon reduction with NaBH₄, due to Ag(I) ions-base bonding. The sequences in Ag-28b and Ag-19b lack the high density of cytosines of the Poly-C loop in Ag-HP, hence a larger area of the silver nanocluster is exposed in Ag-28b and Ag-19b, which can interact with surrounding molecules and impact cellular processes. Since Ag-28b provides a higher amount of cytosine and guanine bases, and additionally, its longer strand offers the possibility of a more stable Ag-DNA conformation, it yields a lower cytotoxicity than its shorter counterpart Ag-19b. In these three variants, we show the potential for cytotoxicity tuning by adapting the cytosine and guanine amount and location in the implemented DNA template.

2.2.4. CONCLUSIONS

In conclusion, we show that the unique properties of Ag-DNA allow for nanomaterials with multiple functionalities, targeted at optical sensing in combination with tunable cytotoxicity, by simple modifications of Ag-DNA designs. Ag-28b exhibits stable fluorescence properties and is thus suitable for bright and persistent labeling of cells, as well as localization experiments. Its shorter counterpart, Ag-19b, is environmentally sensitive with an optical emission shift of $\Delta\lambda_{(em)}=120$ nm and exhibits higher cytotoxicity, making it interesting as an anti-bacterial or anti-cancer agent. The biocompatible, non-toxic Ag-HP optically responds to intracellular salt concentrations, visible as a large fluorescence shift of about $\Delta\lambda_{(em)}=60$ nm, which allows a convenient detection of signal changes for both, Ag-HP and Ag-19b, by commonly used fluorescence microscope systems.

These results will allow us to extend these Ag-DNA constructs to more complex DNA-origami structures, and the investigations from single cells to living organisms. Therefore, Ag-DNA represent a new nanomaterial with high potential for a multi-functional nanoagent, combining bio-sensing and effector functionality. Possible targeted DNA-sequence specific applications would open a large field of patient-specific treatment including cancer or rare and degenerative diseases, as well as specific antimicrobial targeting.

2.3. MATERIALS AND METHODS

2.3.1. CHEMICALS

DNA strands 5'-CAC CGC TTT TGC CTT TTG GGG ACG GAT A-3' (28b), 5'-TGC CTT TTG GGG ACG GAT A-3' (19b), and (b) 5'-CT TAC CTC CCC CCC CCC CCA GGT AAG-3' (HP) with standard desalting (Integrated DNA Technologies, USA), AgNO₃ (99.9999%, Sigma Aldrich, The Netherlands), NH₄OAc (99.99%, Sigma Aldrich, The Netherlands) NaBH₄ (99%, Sigma Aldrich, The Netherlands), KH₂PO₄ (>99%, Sigma Aldrich, The Netherlands), Na₂HPO₄ (>99%, Sigma Aldrich, The Netherlands), Maltose (Duchefa Biochemie B. V., Netherlands), Fluorescein diacetate (F7378, Sigma Aldrich Merck, Germany), HL5-C medium (Formedium™, United Kingdom), NaCl (Fisher Scientific Company LLC, USA), Mg(OAc)₂ (>99%, Sigma Aldrich, The Netherlands), Geneticin (Sigma Aldrich, The Netherlands), 3kDa and 50kDa centrifugal filter (Merck Millipore, Germany) were used as received without further purification. Bi-distilled water from a Millipore system (Milli-Q Academic A-10, Merck Millipore Germany) was used for all solutions.

2.3.2. AG-DNA PREPARATION

For synthesis of (a) Ag-28b and Ag19b, and (b) Ag-HP solutions, 50 nmol of the respective DNA molecules were mixed with AgNO₃ in a ratio of (a) 9.6 : 1 silver ions: DNA strands, or (b) 7 : 1 silver ions: DNA strands in 1ml 20 mM NH₄OAc. The solutions were incubated for 30min at 4° C and then reduced with NaBH₄ at 4° C over night. The reduction ratios were (a) 0.5 : 1 and (b) 0.3 : 1 NaBH₄ : Ag⁺. To exclude formed silver clumps and unbound Ag(I) ions as well as other salts, Ag-DNA constructs were filtered using

a 3kDa centrifugal filter and subsequently a 50kDa centrifugal filter, simultaneously exchanging the buffer with phosphate buffer (PB; 3.57mM KH_2PO_4 , 3.46mM Na^2HPO_4 , pH 6.0). The concentration of the DNA was calculated based on the absorption at 260 nm, measured with a Cary 50 UV-Vis spectrophotometer (Varian Medical Systems, Netherlands). For the reproduction of the observed peaks, various concentrations of $\text{Mg}(\text{OAc})_2$ and NaCl were added to a 15 μM solution of Ag-HP in PB and the fluorescence properties measured. The excitation and emission properties of the Ag-DNA in PB were determined by Cary Eclipse fluorimeter (Varian Medical Systems, Netherlands).

2.3.3. CELL CULTURE

Dictyostelium discoideum HG1729 histone-GFP in AX2 and *Dictyostelium discoideum* Wildtype in AX2 cells were cultured in HL5-C medium adjusted to pH 6.7 at 21° C; medium for HG1729 histone-GFP was supplemented with 10 $\mu\text{g}/\text{ml}$ Geneticin. For the experiments the cells were harvested below a confluency of 40% and washed three times with PB by centrifugation at 400 g for 5 min. Subsequently, cells dispersed in PB were seeded onto an observation chamber (composed of a cover glass and a custom-made Teflon® frame) and rested for 30 min, allowing the cells to settle down.

2.3.4. MICROSCOPY

Imaging of cells incubated with Ag-DNA was performed by confocal microscopy set-up equipped with a Yokogawa 10,000 rpm spinning disc unit (Andor Technology Ltd., UK), Eclipse TiE Nikon inverted microscope (Nikon Corporation, Japan) and an IXON Ultra EMCCD camera (Andor Technology Ltd., UK). Ag-28b and Ag-19b were added to *D. discoideum* HG1729 histone-GFP cells to an end concentration of 4.5 μM and 3 μM , respectively, in an observation chamber and immediately imaged with a 100x oil immersion objective with NA 1.45. Ag-28b and Ag-19b were excited at 640 nm, histone GFP at 480 nm. As negative controls, also cells without Ag-DNA were imaged, as well as cells incubated in 36 μM AgNO_3 and 4.5 μM 28b-DNA, separately and mixed. Local fluorescence spectra measurements were performed with a Leica TCS SP8 STED 3X Microscope and with a 63x objective lens (Leica Microsystems, Germany), additionally equipped with HyD detectors (Leica Microsystems, Germany). The excitation wavelengths ranged from 470 to 670 nm in 10 nm steps and the emission wavelengths ranged from 480 to 770 nm, with a step size of 10 nm and a detection window of 10 nm. For this assay, solutions of Ag-28b (4.5 μM), Ag-19b (3 μM) and Ag-HP (15 μM) were incubated with *D. discoideum* wildtype cells.

2.3.5. CYTOTOXICITY ASSAY

D. discoideum wildtype cells were harvested and washed three times with a maltose-loaded phosphate buffer (3.57 mM KH_2PO_4 , 3.46 mM Na^2HPO_4 , 52.58 mM Maltose, pH 6.70) by centrifugation at 400 g for 4 min. Cells were incubated for 2 h with Ag-28b, Ag-19b and Ag-HP, with concentrations ranging between 75 nm and 36 μM , as triplicates with 25000 cells/condition in the dark. As a positive control, incubation with AgNO_3 was

performed under the same conditions, as well as a negative control without agent incubation. To determine cell viability, cells were incubated with 50 $\mu\text{g}/\text{ml}$ Fluorescein diacetate for 15 min, and the average fluorescence intensity was measured using a TECAN Infinite M200 well plate reader (Tecan Trading AG, Switzerland). To avoid background fluorescence from the Ag-DNA, 50 mM NaCl was introduced to all samples prior to measurements.

2.4. ADDITIONAL INFORMATION

In this section additional information and figures are displayed that support the data presented in this chapter.

Table 2.1: DNA sequences used for Ag-DNA synthesis.

DNA sequence	Abbreviation
5'-CAC CGC TTT TGC CTT TTG GGG ACG GAT A-3'	28b
5'-TGC CTT TTG GGG ACG GAT A-3'	19b
5'-CT TAC CTC CCC CCC CCC CCA GGT AAG-3'	HP

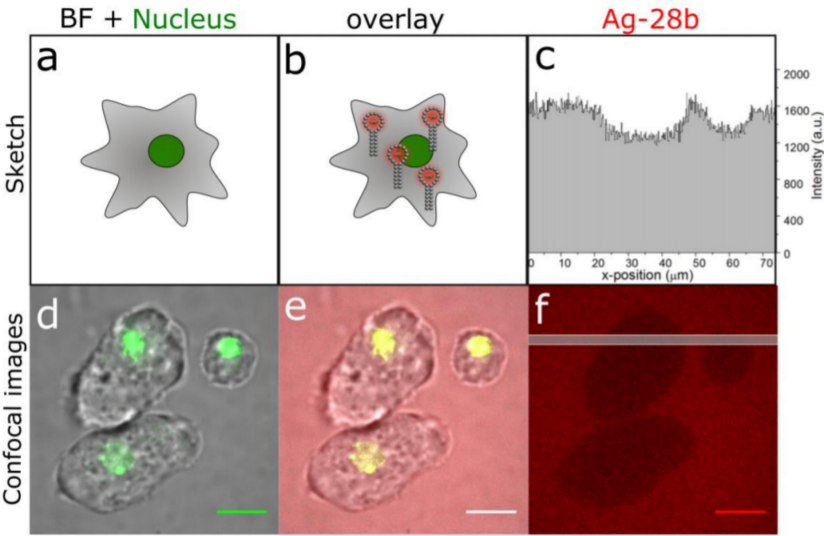


Figure 2.5: Internalization and distribution of Ag-28b inside *D. discoideum* cells. Overlay of brightfield images of cells with labeled GFP histones (green), showing the cell nucleus (a, d) (excitation at 488 nm), and including Ag-28b fluorescence (b, e) (excitation at 640 nm). Profile of the mean fluorescence intensity of Ag-28b (c) for positions along the grey line in the image of red Ag-28b fluorescence (f), showing about 80% of the intensity from the surrounding medium at positions corresponding to the cell interior. The scale bars are 10 μm .

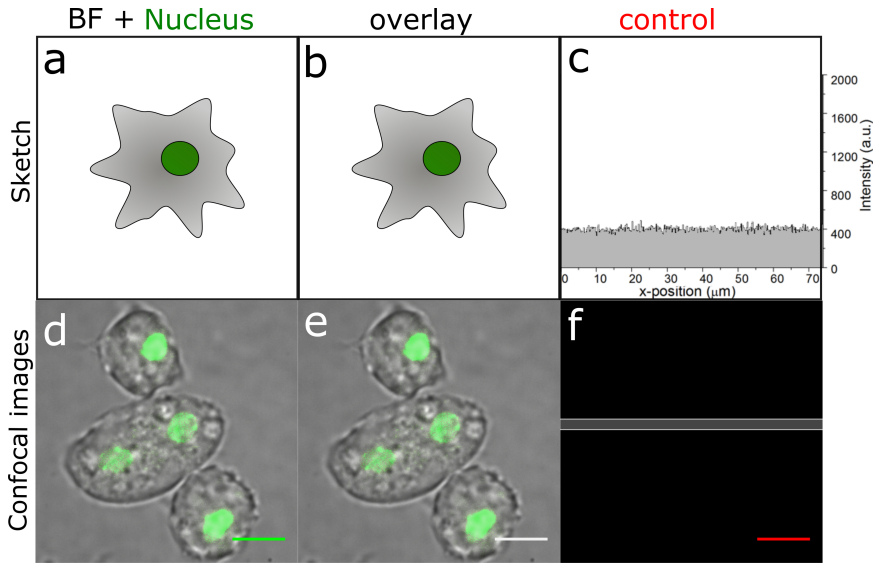


Figure 2.6: Negative control for the internalization and distribution of Ag-28b inside *D. discoideum* cells in Figure 2.5. Overlay of brightfield images of cells with labeled GFP histones, showing the cell nucleus (a, d) (excitation at 488 nm), and including any red autofluorescence (b, e) (excitation at 488 nm and 640 nm). Profile of the mean fluorescence intensity at 640 nm excitation (c) for positions along the grey line in the image of only the red channel (f). This control shows that there is only slight autofluorescence (e) when excited at 640 nm compared to the strong signal when employing Ag-28b as in 2.5 e. The scale bars are 10 μm .

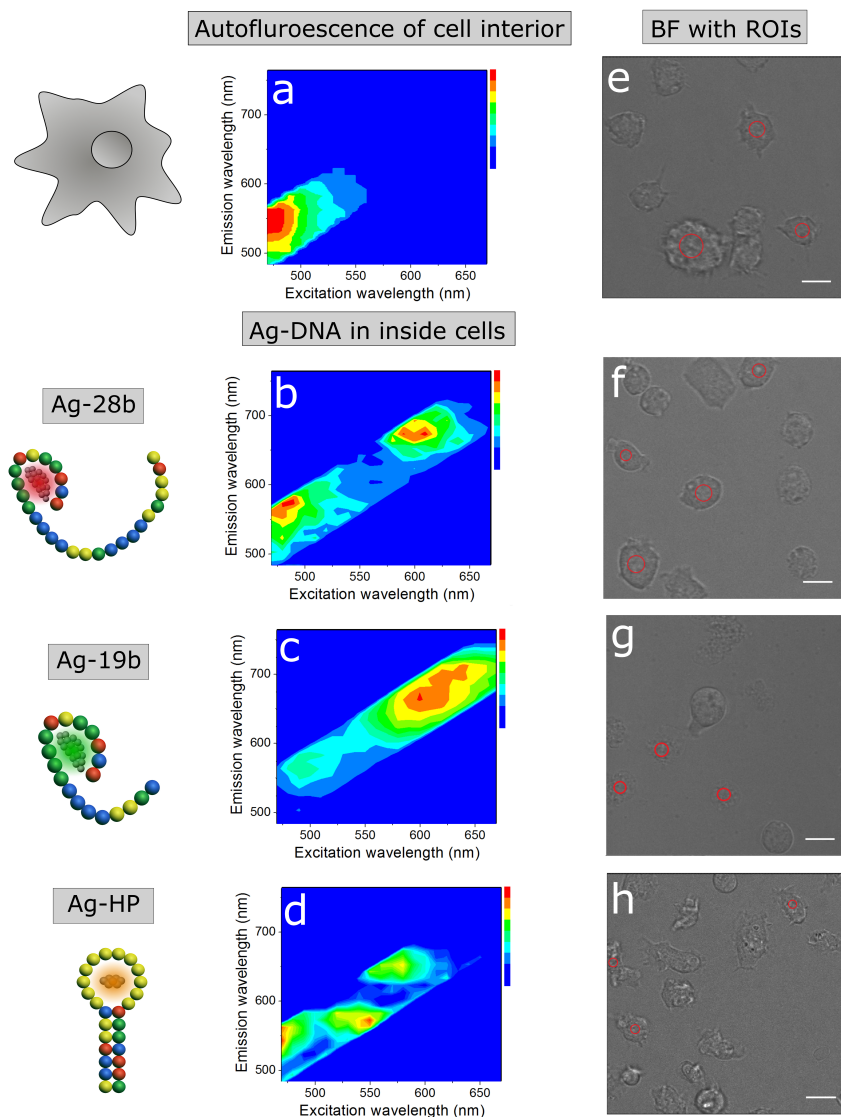


Figure 2.7: Region of interest (ROI) selection (e-h) for the fluorescence spectra analysis of Ag-DNA variants after internalization in *D. discoideum* wildtype cells (a-d). The 1st column shows sketches of used Ag-DNAs. DNA sequence color code: Cytosine = yellow, Guanine = green, Thymine = blue, Adenine = red. The spectra correspond to the fluorescence collected from the ROIs designated by red circles in the microscopy images. Fluorescence spectra were measured within excitation range of 470-670 nm and emission range of 490-760 nm, each in 10 nm steps. ROIs were chosen to remain entirely within the center of the cell throughout the measurement to avoid collection of light from the surrounding medium. The scale bars are 10 μ m.

2.5. REFERENCES

- [1] Jaswinder Sharma et al. “Silver nanocluster aptamers: in situ generation of intrinsically fluorescent recognition ligands for protein detection”. In: *Chemical Communications* 47.8 (2011), pp. 2294–2296.
- [2] Bidisha Sengupta et al. “DNA templates for fluorescent silver clusters and i-motif folding”. In: *The Journal of Physical Chemistry C* 113.45 (2009), pp. 19518–19524.
- [3] Chris I Richards et al. “Oligonucleotide-stabilized Ag nanocluster fluorophores”. In: *Journal of the American Chemical Society* 130.15 (2008), pp. 5038–5039.
- [4] Tom Vosch et al. “Strongly emissive individual DNA-encapsulated Ag nanoclusters as single-molecule fluorophores”. In: *Proceedings of the National Academy of Sciences* 104.31 (2007), pp. 12616–12621.
- [5] Danielle Schultz et al. “Dual-color nanoscale assemblies of structurally stable, few-atom silver clusters, as reported by fluorescence resonance energy transfer”. In: *ACS nano* 7.11 (2013), pp. 9798–9807.
- [6] Bidisha Sengupta et al. “Base-directed formation of fluorescent silver clusters”. In: *The Journal of Physical Chemistry C* 112.48 (2008), pp. 18776–18782.
- [7] Til Driehorst et al. “Distinct conformations of DNA-stabilized fluorescent silver nanoclusters revealed by electrophoretic mobility and diffusivity measurements”. In: *Langmuir* 27.14 (2011), pp. 8923–8933.
- [8] Hsin-Chih Yeh et al. “A DNA- silver nanocluster probe that fluoresces upon hybridization”. In: *Nano letters* 10.8 (2010), pp. 3106–3110.
- [9] Hsin-Chih Yeh et al. “A fluorescence light-up Ag nanocluster probe that discriminates single-nucleotide variants by emission color”. In: *Journal of the American Chemical Society* 134.28 (2012), pp. 11550–11558.
- [10] Weiwei Guo et al. “Highly sequence-dependent formation of fluorescent silver nanoclusters in hybridized DNA duplexes for single nucleotide mutation identification”. In: *Journal of the American Chemical Society* 132.3 (2010), pp. 932–934.
- [11] Jing-Jing Liu et al. “Label-free and fluorescence turn-on aptasensor for protein detection via target-induced silver nanoclusters formation”. In: *Analytica chimica acta* 749 (2012), pp. 70–74.
- [12] Delphine Arcizet et al. “Temporal analysis of active and passive transport in living cells”. In: *Physical review letters* 101.24 (2008), p. 248103.
- [13] M Götz et al. “Probing cytoskeleton dynamics by intracellular particle transport analysis”. In: *The European Physical Journal Special Topics* 224.7 (2015), pp. 1169–1183.
- [14] Marcus Otten et al. “Local motion analysis reveals impact of the dynamic cytoskeleton on intracellular subdiffusion”. In: *Biophysical journal* 102.4 (2012), pp. 758–767.
- [15] Carina Pelzl et al. “Axonal guidance by surface microstructuring for intracellular transport investigations”. In: *Chemphyschem* 10.16 (2009), pp. 2884–2890.

- [16] Jaswinder Sharma et al. "A DNA-templated fluorescent silver nanocluster with enhanced stability". In: *Nanoscale* 4.14 (2012), pp. 4107–4110.
- [17] Richard Kuehl et al. "Preventing implant-associated infections by silver coating". In: *Antimicrobial agents and chemotherapy* 60.4 (2016), pp. 2467–2475.
- [18] Sogol Asghari, Sarvesh Logsetty, and Song Liu. "Imparting commercial antimicrobial dressings with low-adherence to burn wounds". In: *Burns* 42.4 (2016), pp. 877–883.
- [19] Fenella D Halstead et al. "Antimicrobial dressings: Comparison of the ability of a panel of dressings to prevent biofilm formation by key burn wound pathogens". In: *Burns* 41.8 (2015), pp. 1683–1694.
- [20] Svitlana Chernousova and Matthias Epple. "Silver as antibacterial agent: ion, nanoparticle, and metal". In: *Angewandte Chemie International Edition* 52.6 (2013), pp. 1636–1653.
- [21] A Kedziora et al. "Silver nanoforms as a therapeutic agent for killing *Escherichia coli* and certain ESKAPE pathogens". In: *Current microbiology* 73.1 (2016), pp. 139–147.
- [22] MA Radzig et al. "Antibacterial effects of silver nanoparticles on gram-negative bacteria: influence on the growth and biofilms formation, mechanisms of action". In: *Colloids and Surfaces B: Biointerfaces* 102 (2013), pp. 300–306.
- [23] Samad Mussa Farkhani et al. "Enhancing antitumor activity of silver nanoparticles by modification with cell-penetrating peptides". In: *Artificial cells, nanomedicine, and biotechnology* 45.5 (2017), pp. 1029–1035.
- [24] Pallab Sanpui, Arun Chattopadhyay, and Siddhartha Sankar Ghosh. "Induction of apoptosis in cancer cells at low silver nanoparticle concentrations using chitosan nanocarrier". In: *ACS applied materials & interfaces* 3.2 (2011), pp. 218–228.
- [25] Muthuraman Pandurangan et al. "Time and concentration-dependent therapeutic potential of silver nanoparticles in cervical carcinoma cells". In: *Biological trace element research* 170.2 (2016), pp. 309–319.
- [26] Bin-Hsu Mao et al. "Mechanisms of silver nanoparticle-induced toxicity and important role of autophagy". In: *Nanotoxicology* 10.8 (2016), pp. 1021–1040.
- [27] Yan Li et al. "Differential genotoxicity mechanisms of silver nanoparticles and silver ions". In: *Archives of toxicology* 91.1 (2017), pp. 509–519.
- [28] Esther E Fröhlich and Eleonore Fröhlich. "Cytotoxicity of nanoparticles contained in food on intestinal cells and the gut microbiota". In: *International journal of molecular sciences* 17.4 (2016), p. 509.
- [29] Anastasia Georgantzopoulou et al. "Effects of silver nanoparticles and ions on a co-culture model for the gastrointestinal epithelium". In: *Particle and fibre toxicology* 13.1 (2015), pp. 1–17.
- [30] Ivana Vinković Vršek et al. "Comparison of in vitro toxicity of silver ions and silver nanoparticles on human hepatoma cells". In: *Environmental toxicology* 31.6 (2016), pp. 679–692.

- [31] Jinjin Yin et al. "One-step engineering of silver nanoclusters–aptamer assemblies as luminescent labels to target tumor cells". In: *Nanoscale* 4.1 (2012), pp. 110–112.
- [32] Zhipeng Sun et al. "Ag cluster–aptamer hybrid: specifically marking the nucleus of live cells". In: *Chemical Communications* 47.43 (2011), pp. 11960–11962.
- [33] Junhua Yu et al. "Live cell surface labeling with fluorescent Ag nanocluster conjugates". In: *Photochemistry and photobiology* 84.6 (2008), pp. 1435–1439.
- [34] Jingjing Li et al. "Binding-induced fluorescence turn-on assay using aptamer-functionalized silver nanocluster DNA probes". In: *Analytical chemistry* 84.12 (2012), pp. 5170–5174.
- [35] Danielle Schultz et al. "Evidence for rod-shaped DNA-stabilized silver nanocluster emitters". In: *Advanced materials* 25.20 (2013), pp. 2797–2803.
- [36] Stacy M Copp et al. "Cluster plasmonics: dielectric and shape effects on DNA-stabilized silver clusters". In: *Nano letters* 16.6 (2016), pp. 3594–3599.
- [37] Patrick R O'Neill et al. "Hairpins with poly-C loops stabilize four types of fluorescent Ag n: DNA". In: *The Journal of Physical Chemistry C* 113.11 (2009), pp. 4229–4233.
- [38] K Lance Kelly et al. *The optical properties of metal nanoparticles: the influence of size, shape, and dielectric environment*. 2003.
- [39] Huanjun Chen et al. "Shape- and size-dependent refractive index sensitivity of gold nanoparticles". In: *Langmuir* 24.10 (2008), pp. 5233–5237.
- [40] Danielle Schultz and Elisabeth Gwinn. "Stabilization of fluorescent silver clusters by RNA homopolymers and their DNA analogs: C, G versus A, T (U) dichotomy". In: *Chemical communications* 47.16 (2011), pp. 4715–4717.
- [41] Danielle Schultz and Elisabeth G Gwinn. "Silver atom and strand numbers in fluorescent and dark Ag: DNAs". In: *Chemical Communications* 48.46 (2012), pp. 5748–5750.
- [42] Vladimir P Denisov and Bertil Halle. "Sequence-specific binding of counterions to B-DNA". In: *Proceedings of the National Academy of Sciences* 97.2 (2000), pp. 629–633.
- [43] Péter Várnai and Krystyna Zakrzewska. "DNA and its counterions: a molecular dynamics study". In: *Nucleic acids research* 32.14 (2004), pp. 4269–4280.
- [44] OO Liubysh, AV Vlasiuk, and SM Perepelytsya. "Structuring of counterions around DNA double helix: a molecular dynamics study". In: *arXiv preprint arXiv:1503.05334* (2015).
- [45] Elisabeth G Gwinn et al. "Sequence-Dependent fluorescence of DNA-Hosted silver nanoclusters". In: *Advanced Materials* 20.2 (2008), pp. 279–283.
- [46] Steven M Swasey et al. "Silver (I) as DNA glue: Ag⁺-mediated guanine pairing revealed by removing Watson-Crick constraints". In: *Scientific reports* 5.1 (2015), pp. 1–9.

- [47] Hidetaka Torigoe et al. "Thermodynamic and structural properties of the specific binding between Ag⁺ ion and C: C mismatched base pair in duplex DNA to form C–Ag–C metal-mediated base pair". In: *Biochimie* 94.11 (2012), pp. 2431–2440.