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Lipid model membrane systems as a tool for unraveling the underlying factors for skin barrier dysfunction

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Barrier capability of skin lipid models: Effect of ceramides and free fatty acid composition

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ABSTRACT

The skin is an effective barrier that prevents the influx of harmful substances from the environment and the efflux of body fluid. This barrier function is ascribed to the intercellular lipids present in the outermost layer of the skin referred to as the stratum corneum (SC). These lipids are composed mainly of ceramides (CERs), cholesterol, and free fatty acids (FFAs). Alterations in the SC lipid composition and barrier function impairment occur in several skin diseases including atopic dermatitis (AD). As the etiology of AD is multifactorial, establishing the relationship between the changes in SC lipid composition and barrier function impairment in the patients remains a challenge. Here, we employed model membrane systems to investigate the contribution of various anomalies in the SC CER and FFA composition observed in AD patients' skin to the barrier dysfunction. Using ethyl-p-aminobenzoate permeation and transepidermal water loss values as markers for barrier function, we determined that the alterations in SC lipid composition contribute to the impaired barrier function in AD patients. By the use of biophysical techniques, we established that the largest reduction in barrier capability was observed in the model with an increased fraction of short-chain FFAs, evident by the decrease in chain packing density. Modulations in the CER subclass composition impacted the lamellar organization while having a smaller effect on the barrier function. These findings provide evidence that AD therapies normalizing the FFA composition are at least as important as normalizing CER composition.

Keywords

Atopic dermatitis; Ceramide; Free fatty acids; Infrared spectroscopy; Permeability; Stratum corneum; X-ray scattering.

INTRODUCTION

Lipids can act as structural components for cell membranes but are also important for signaling and energy storage [1-6]. In most viable cell membranes the most important components are phospholipids, sphingolipids, and cholesterol (CHOL). A special group of sphingolipids is the ceramides (CERs). In the outermost layer of the epidermis, the stratum corneum (SC), these CERs together with cholesterol and free fatty acids (FFAs) are the most important components of the lipid matrix, a structure that is crucial in maintaining the skin barrier function [7]. A major difference between the SC lipid composition and other biological membranes is that the former is basically devoid of phospholipids. The SC lipid matrix surrounds the individual keratin-filled terminally differentiated cells, the corneocytes. The SC structure has been described as being analogous to bricks and mortar structure, the corneocyte being the bricks and the intercellular lipids the mortar [8]. X-ray diffraction studies have revealed that the intercellular lipids are assembled in two co-existing lamellar phases with periodicities of ~13 nm and ~6 nm known as the long and short periodicity phases (LPP and SPP) respectively [9, 10]. Within the lamellae, the majority of the SC lipids preferably adopt a dense orthorhombic lateral packing at skin temperature (30-32 °C), while a portion of the lipids adopts the less dense hexagonal packing [11, 12]. This is distinctive from most biological cell membranes, which often form a liquid (ordered) phase. The ordered lipid packing in SC is induced by; i) the FFAs with predominantly saturated chains, with carbon chain lengths ranging between 12 and 30, the majority of the FFA having a chain length of 22, 24, or 26 carbon atoms [13, 14] and ii) the CERs with a small head group and long hydrocarbon tails. The CERs can vary in the headgroup structure and chain length. Currently, at least 18 subclasses of CERs have been identified in the human SC [15-17]. The commonly adopted nomenclature for CER is based on Motta et al [18], in which the CERs are designated with 2-3 letters, defining the type of sphingoid base and linked fatty acid acyl chain (Figure 1).

Alterations in the SC lipid composition has been reported in one of the most prevalent, chronic inflammatory skin disease referred to as atopic dermatitis (AD). In industrialized nations, the incidence of AD has increased to about 15-30% in children and 2-10% in adults [19]. The etiology of AD is multifactorial, involving the interaction of immunological, genetic, and environmental factors [20]. A compromised skin barrier, indicated by increased water transport across the SC, is characteristic of patients with AD [21, 22]. Several studies have reported that the reduced skin barrier function is associated with changes in lipid composition. The following changes in lipid composition were described

in AD patients' skin compared to healthy skin: i) a reduction in the levels of CER NP, CER NH, CER EO classes, while CER NS and AS were increased [23-26], ii) a reduction in chain length of FFAs [27] and CERs, specifically increased level of CERs AS and NS (C16) with a total chain length of 34 carbon atoms [23, 26]. As these changes in lipid composition occur simultaneously [21, 28, 29], it is not yet established which of them is the underlying factor of the impaired skin barrier.

In the current study, we investigated in a systematic way, which changes in SC lipid composition reported in AD patients, resulted in the most dramatic changes in the lipid organization, thereby possibly increasing the lipid barrier dysfunctionality. Healthy skin model based on the SC lipid composition and diseased skin models that mimicked different aspects of the altered lipid composition in atopic dermatitis patients' skin was developed. We showed that an increased level of short-chain FFAs resulted in a more drastic reduction in barrier function than compositional changes of the CERs. Knowledge from this study could aid in design of rational therapy for the treatment of inflammatory skin diseases in which compositional changes in SC lipids are implicated.

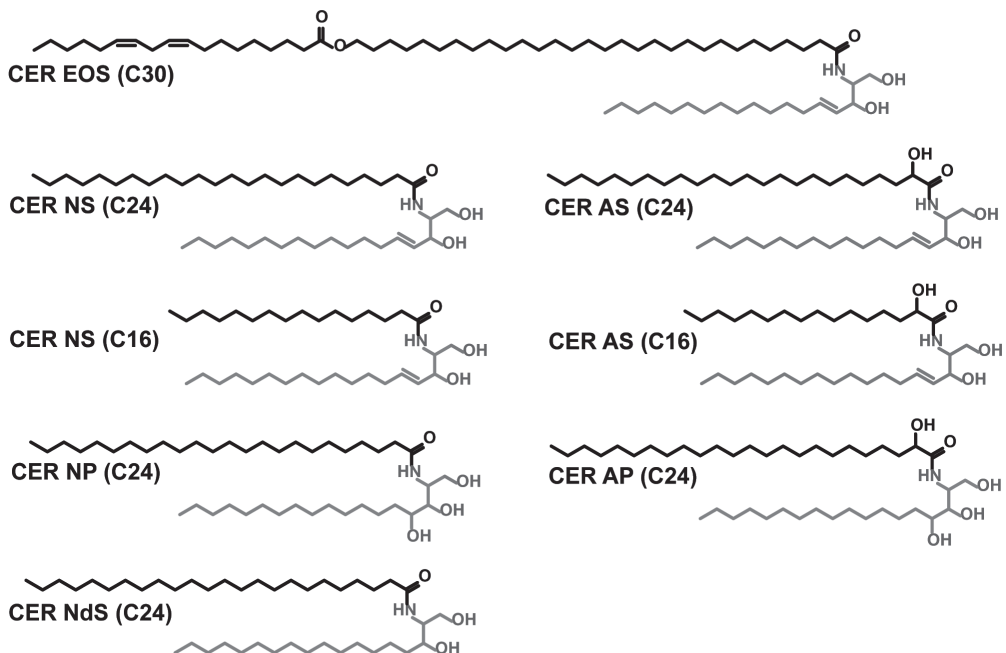


Figure 1: Molecular structure of the CERs used in the study.

CERs consist of a sphingoid base (marked in grey) linked to a fatty acid acyl chain (marked in black) via an amide bond. The fatty acid acyl chain is either non-hydroxylated [N], α-hydroxylated [A], or ω-esterified [EO], while the sphingoid base is either sphingosine [S] or phytosphingosine [P].

EXPERIMENTAL SECTION

Materials

The synthetic CERs: N-(tetracosanoyl)-sphingosine (CER NS), N-(tetracosanoyl)-phytosphingosine (CER NP), N-(2R-hydroxy-tetracosanoyl)-sphingosine (CER AS), N-(2R-hydroxy-tetracosanoyl)-phytosphingosine (CER AP) of acyl carbon chain length C24 and N-(30-Linoleoyloxy-triacontanoyl)-sphingosine (EOS C30) used in this study were generously donated by Evonik (Essen, Germany). CERs NS and AS C16 and N-(tetracosanoyl)-dihydrosphingosine (CER NdS C24) were purchased from Avanti. The CERs were of $\geq 90\%$ purity. The FFAs, ethyl-p-aminobenzoate (E-PABA), CHOL and acetate buffer salts were acquired from Sigma- Aldrich Chemie GmbH (Schnelldorf, Germany). The perdeuterated FFAs (DFFAs) with chain lengths of C18 and C20 were obtained from Cambridge Isotope Laboratories (Andover, Massachusetts). The DFFAs with chain lengths of C16 and C22 were purchased from Larodan (Malmö, Sweden). The DFFA with a chain length of C24 was obtained from Arc Laboratories B.V. (Apeldoorn, The Netherlands). All organic solvents were procured from Labscan Ltd. (Dublin, Ireland). All other reagents were of analytical grade and used without further purification. The water was of Millipore quality, obtained through a Milli-Q Integral Water Purification System (Millipore, Bedford, MA). Nucleopore polycarbonate filter disks (pore size 0.05 μm) were acquired from Whatman (Kent, UK).

Isolation of SC from native skin

SC was isolated from abdominal or mammary skin, which was obtained within 24 h after cosmetic surgery. The skin surface was first cleaned with 70% ethanol and Milli-Q water thereafter. The skin was dermatomed to a thickness of ~ 0.6 mm. The SC was separated from the epidermis by trypsin digestion; 0.1% in phosphate-buffered saline (PBS 0.1 M solution: NaCl, Na_2HPO_4 , KH_2PO_4 , and KCl in milli-Q water with a concentration of 8.13, 1.14, 0.20 and 0.19 g/l respectively at pH 7.4), after overnight incubation at 4 °C and subsequently at 37 °C for 1 h. The SC was then placed in a 0.1% solution of trypsin inhibitor, rinsed twice with Milli-Q water, and allowed to dry. The SC was stored in a silica-containing, opaque, sealed box, under nitrogen in order to prevent oxidation of the intercellular SC lipids. The isolated native SC was subsequently used in diffusion studies.

Preparation of the model lipid mixtures

Five SC lipid models (healthy and diseased skin) were prepared as equimolar mixtures of CER, CHOL, and FFAs. The lipid composition of the healthy skin

model (denoted as Healthy) was based on the native SC lipid composition [15, 30]. The most prevalent CERs in human skin were included in the model, except for CERs comprised of 6-hydroxy sphingoid bases that were not available. As CER EOS constitutes over 50% of the total CER EO class present in the human CER [15], it represented the CER EO in the healthy skin model. To prepare the diseased skin models, the lipid composition of the healthy skin model was adjusted, to mimic various aberrations reported in the skin of AD patients. The changes made are; i) replacing a fraction of CER NS (C24) and CER AS (C24) with by CER NS (C16) and CER AS (C16) respectively, denoted as Dis-CER_{ch}; ii) besides the presence of CERs NS and AS (C16), a reduction of the level of CER EOS and NP subclasses was included, counterbalanced by an increase in the levels of CER NS and AS subclasses, referred to as Dis-CER_{ch-SbC}; iii) increased fraction of short-chain FFAs referred to as Dis-FFA_{ch} and iv) Combination of all the changes in CERs & FFAs referred to as Dis-Combi. The CERs and FFAs composition of the models are presented in Tables 1 and 2 respectively. Additional models were prepared in which FFAs were replaced by their deuterated counterparts. The resulting models are denoted by D- before the model name (D-Healthy, D-Dis-CER_{ch}, D-Dis-CER_{ch-SbC}, D-Dis-FFA_{ch}, and D-Dis-Combi). Other models were prepared in which either CER NS (C24) or CER NS (C16) fatty acid acyl chain in the Dis-CER_{ch} model was substituted by their deuterated counterparts (D-CER NS (C24) or D-CER NS (C16)) resulting in Dis-CER_{ch-D-NS (C24)} and Dis-CER_{ch-D-NS (C16)} respectively.

Table 1: CER composition of the various models used in the study.

CER name and chain length	Model name & CER molar ratio (%)						
	Healthy	Dis-CER _{ch}	Dis-CER _{ch-D-NS (C24)}	Dis-CER _{ch-D-NS (C16)}	Dis-CER _{ch-SbC}	Dis-FFA _{ch}	Dis-Combi
CER EOS (C30)	15	15	15	15	7	15	7
CER NdS (C24)	13	13	13	13	13	13	13
CER NS (C24)	13	6.5	-	6.5	12	13	12
D-CER NS (C24)	-	-	6.5	-	-	-	-
CER NS (C16)	-	6.5	6.5	-	12	-	12
D-CER NS (C16)	-	-	-	6.5	-	-	-
CER NP (C24)	30	30	30	30	16	30	16
CER AS (C24)	13	6.5	6.5	6.5	12	13	12
CER AS (C16)	-	6.5	6.5	6.5	12	-	12
CER AP (C24)*	16	16	16	16	16	16	16
Total	100	100	100	100	100	100	100

* In our study, we used the native D-isomer of CER AP and CER AS which unlike the unnatural L-isomer has been shown to display native-like behavior[31]

Table 2: FFA composition of the various models used in the study.

FFA name and chain length	Model name & FFA molar ratio (%)		
	Healthy	Dis-CER _{ch} /Dis-CER _{ch-SbC}	Dis-FFA _{ch} /Dis-combi
Palmitic Acid (C16)	1.8	1.8	26
Stearic Acid (C18)	4.0	4.0	19
Arachidic Acid (C20)	7.6	7.6	10
Behenic Acid (C22)	47.8	47.8	21
Lignoceric Acid (C24)	38.8	38.8	24
Total	100	100	100

Preparation of SC model membranes for permeability studies and transepidermal water loss (TEWL) measurements

To prepare the SC model membranes, 0.9 mg of the appropriate lipid composition was dissolved in 200 μ l hexane:ethanol (2:1) solution. The solution was sprayed over an area of 10 x 10 mm², on a nucleopore polycarbonate filter disk with 0.05 μ m pore size (Whatman, Kent, UK), using a Linomat IV device with an extended y-axis arm (Camag, Muttenz, Switzerland) under a gentle stream of nitrogen. The solution was sprayed at 14 μ l/sec at an approximate distance of 1 mm between the nozzle and the spraying surface. The samples were equilibrated for 10 min at 85 °C, which was sufficient to ensure that the lipid mixtures had fully melted and then gradually cooled to room temperature. The heating and cooling cycle was repeated once more.

Permeability studies

The permeation studies were performed using PermeGear in-line diffusion cells (Bethlehem PA, USA) with a diffusion area of 0.28 cm². The sprayed membranes in various lipid compositions were mounted in the diffusion cells and hydrated for 1 h with the acceptor phase consisting of PBS at pH 7.4 before the experiment. Before use, the PBS was filtered and degassed for 30 min under vacuum to remove any dissolved air. The donor phase was a saturated E-PABA solution (0.65mg/ml) in pH 5 acetate buffer. The solution was stirred with a magnetic stirrer, perfused at a flow rate of about 2 ml/h, and degassed continuously by an online degasser. The experiment was performed under occlusive conditions, by closing the opening of the donor compartment with adhesive tape. The temperature of the membranes was maintained at 32 °C mimicking the skin temperature. The acceptor fluid was collected over 15 h at 1-h intervals. At the end of the diffusion study, the volume per collected fraction of PBS was determined by weight and the concentration of E-PABA was determined by UPLC. Permeation of multiple

samples of each model was analyzed $n > 6$. The steady-state flux values were calculated from the plots of E-PABA flux values versus time, at a time interval between the 5th and 15th hour.

Ultra performance liquid chromatography (UPLC)

The analysis of E-PABA was undertaken with Acquity UPLC systems (Waters Co., Milford, MA, USA). The UPLC systems consisted of a quaternary solvent manager (a high-pressure pump), a tunable ultraviolet/visible absorbance detector, and a sample manager. UPLC special analytical column packed with 1.7 μm , bridged, ethyl siloxane, hybrid particles, was used as the stationary phase. The column temperature was set at 40 °C. The mobile phase was composed of mixtures of 0.1% trifluoroacetic acid in acetonitrile: milli-Q water at 40:60 (v/v) ratio. The flow rate of the mobile phase was set at 1 ml/min. 10 μl of the sample was injected on the column. The detector wavelength was set at 286 nm. Data were collected and processed by MassLynx and TargetLynx software V4.1 SCN951 (Waters Co., Milford, USA).

0.5 mg/ml E-PABA stock solution in a 1:1 solution of methanol and milli-Q water was prepared. Eight different concentrations were prepared by serial dilutions of the stock solution with milli-Q water to plot the standard curve. The linearity of the relationship was evaluated in a concentration range of 0.1 -10 $\mu\text{g/ml}$ covering the range of concentrations obtained when analyzing the concentration of E-PABA permeating the model membranes. The calibration curves were obtained using least square linear regression and the linearity was confirmed with the R^2 values. The UPLC method was previously validated for E-PABA analysis as per ICH (International conference on harmonization) guidelines with respect to linearity, precision, limit of detection (LOD), and limit of quantification (LOQ) [32-34].

TEWL measurements

An AquaFlux (AF200, Biox Systems Ltd., London, UK) was used to measure the TEWL of the lipid models. The measurement has been previously described elsewhere [35]. Briefly, the sprayed membranes were mounted in a diffusion cell. The TEWL device was then coupled vertically with the donor compartment of the diffusion cell using a special measurement cap (Biox Systems Limited, UK) in order to seal the compartment and ensure vapour tight connectivity. The TEWL values for the models studied were recorded over a period of 30 min, after which an average reading during the final 10 min of the measurement was calculated, $n = 7$.

Sample preparation for FTIR and X-ray diffraction studies

The appropriate lipid composition was dissolved in chloroform:methanol (2:1) to a concentration of 5 mg/ml. Either 1.5 mg was sprayed over an area of 10 × 10 mm on an AgBr window (for FTIR studies) or 1 mg was sprayed over an area of 2 × 4 mm on mica (for X-ray diffraction studies) using a Linomat IV device (Camag, Muttenz, Switzerland) with an extended y-axis arm and equilibrated as described earlier (section 2.3). The samples were equilibrated for 10 min at 85 °C. The heating and cooling cycle was repeated once more. Finally, the samples were hydrated in deuterated acetate buffer (pH 5.0) and incubated at 37 °C for at least 15 h before measurements to ensure that the samples were fully hydrated.

FTIR measurements

FTIR measurements were implemented with a Varian 670-IR spectrometer (Agilent Technologies, Inc., Santa Clara, CA) equipped with a broad-band mercury cadmium telluride detector, cooled by liquid nitrogen. The samples were purged continuously under dry air, starting 30 min prior to data acquisition. The spectra were collected in transmission mode and acquired by the co-addition of 256 scans at 1 cm⁻¹ resolution, over 4 min. In order to examine the thermotropic phase behavior, the sample was measured between 0-100 °C at a heating rate of 0.25 °C/min, resulting in a 1 °C temperature rise per recorded spectrum. The spectra were deconvoluted using a half-width of 4 cm⁻¹, and an enhancement factor of 1.7. The software used was Agilent resolution pro (Agilent Technologies, Palo Alto CA, USA). Samples were measured over a range of 600–4000 cm⁻¹. The CH₂ symmetric stretching modes (2845–2855 cm⁻¹) and CD₂ symmetric stretching modes (2080–2100 cm⁻¹), referred to as $\nu_s\text{CH}_2$ and $\nu_s\text{CD}_2$ modes respectively, were chosen to examine the conformational ordering and phase transition of the lipid chains. The conformational ordering was determined at 10 °C, which was prior to the orthorhombic to hexagonal phase transition. The linear regression curve fitting method was used to determine the mid-transition temperature described previously [36]. The CH₂ rocking vibration mode (719–730 cm⁻¹) and scissoring mode (1462–1473 cm⁻¹) referred to as ρCH_2 and δCH_2 respectively were evaluated to monitor the lateral packing, while the CD₂ scissoring modes (1085–1095 cm⁻¹) was examined for determination of the mixing properties of the lipid chains. Multiple samples were measured for each condition, $n > 2$.

X-ray diffraction studies

The small- and wide-angle X-ray diffraction (SAXD and WAXD) diffraction measurements were performed at the European Synchrotron Radiation Facility (ESRF, Grenoble) at station BM26B. SAXD images were collected using a Pilatus 1M detector with 981 x 1043 pixels of 172 μm spatial resolution while the WAXD patterns were collected using a 300K Pilatus detector with 1475 x 195 pixels of 172 μm spatial resolution. The SAXD detector was calibrated using silver behenate while the WAXD detector was calibrated using the two strongest reflections of high-density polyethylene (HDPE). The X-ray wavelength was set at 0.1033 nm. The sample-to-detector distances were 1.98 m and 0.31 m for SAXD and WAXD respectively. The samples were hydrated for 24 h in milli-Q water prior to mounting in parallel to the primary beam in a sample holder with temperature controlled at 25 $^{\circ}\text{C}$. The diffraction patterns of the samples were acquired in two positions. At each position, the samples were measured for 90 sec. The scattering intensity (I) was measured as a function of the scattering vector (q). The latter is defined as $q = \frac{4\pi \sin \theta}{\lambda}$, where θ is the scattering angle and λ is the wavelength. From the positions of a series of equidistant peaks (q_h), the periodicity of a lamellar phase was calculated using the equation $d = \frac{2h\pi}{q_h}$ in which h is the order of the diffraction peak. The one-dimensional intensity profiles used in these calculations were obtained by the reduction of the 2D SAXD pattern from Cartesian (x,y) to polar (ρ,θ) coordinates and integrated over an θ range. WAXD was used to obtain the diffraction patterns related to the lateral packing of lipids. The diffraction pattern was subsequently collected between 25 and 85 $^{\circ}\text{C}$ in order to examine the temperature-induced phase changes in various models. Each sample condition was measured, $n > 2$.

Data analysis

Unpaired t-test was performed for analysis of the permeability studies data for the healthy skin model versus native skin. One-way ANOVA with Dunnett's multiple comparison test was performed to analyze the permeability studies' and FTIR data for the healthy against the diseased skin models. The significance level for rejection of null hypothesis was set at $P < 0.05$ (*), $P < 0.01$ (**), $P < 0.001$ (***) and $P < 0.0001$ (****).

RESULTS

The healthy skin model membrane properties mimic that of human SC

First, we determined whether the lipid properties of the healthy skin model membrane mimic those of human SC. The lamellar and lateral lipid organization of the healthy skin model were characterized by SAXD and FTIR respectively. The SAXD profile exhibits two lamellar phases with repeat distances of 12.8 and 5.4 nm corresponding to the LPP and SPP in the native human SC, respectively [9, 10] (figure 2A). With respect to the lateral lipid organization, the thermotropic response of the νCH_2 mode is provided in figure 2B. The νCH_2 contour is split into two peaks positioned at ~ 720 and 730 cm^{-1} . The splitting is a result of the short-range coupling of adjacent νCH_2 modes and occurs only in a phase possessing orthorhombic packed chains, as observed in human SC [37]. As the temperature increases, the 730 cm^{-1} peak, which is attributed exclusively to the orthorhombic phase reduces in intensity, implying a loss in orthorhombic packing within the lipid population. At $32\text{ }^\circ\text{C}$, the peak appears as a weak shoulder, indicating a significant reduction in the proportion of lipids adopting an orthorhombic packing and finally disappears at $36\text{ }^\circ\text{C}$, signaling a complete orthorhombic to hexagonal phase transition of the lipids. The permeation of E-PABA through the isolated native human SC and healthy skin model membrane were compared. The E-PABA steady-state flux is presented in (figure 2C). There is no significant difference in

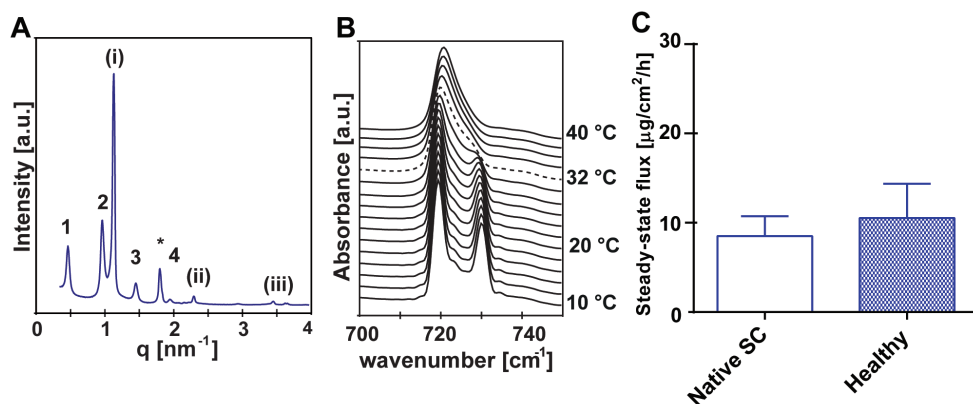


Figure 2: Characterization of the healthy skin model.

A) SAXD profile at $25\text{ }^\circ\text{C}$. The arabic numbers (1, 2, 3,4) indicate the diffraction orders of the LPP, $d = 12.8$ positioned at $q = 0.49, 0.98, 1.46$ and 1.96 nm^{-1} . The roman numbers (i, ii, iii) indicate the diffraction orders of the SPP, $d = 5.4$, positioned at $q = 1.16, 2.3$ and 3.48 nm^{-1} . B) The thermotropic response of the νCH_2 modes. The presence of two peaks positioned at ~ 720 and 730 cm^{-1} indicate orthorhombic lateral packing while a single peak at $\sim 720\text{ cm}^{-1}$ indicates hexagonal lateral packing. C) The average steady-state flux of E-PABA across the native SC and healthy skin model (5-15 h). Data presented as the mean \pm SD, $n \geq 6$, significance level ($P < 0.05$).

permeability between the native SC and the healthy skin model membrane. As the healthy skin model membrane properties are sufficiently similar to that of the lipid matrix in human SC, the composition of the membrane was modulated to mimic several aspects of the lipid composition in SC of AD.

Changes in lipid composition resulted in increased permeability in AD patients' skin models

In order to examine whether the changes in the lipid composition affect the lipid barrier function, the permeability of the healthy and diseased model membranes was monitored. The average fluxes of E-PABA across the models are displayed in figure 3A. The E-PABA's average steady-state flux values are plotted in figure 3B. The highest E-PABA steady-state flux was observed in Dis-combi, where the alterations in the CER and FFA composition were combined.

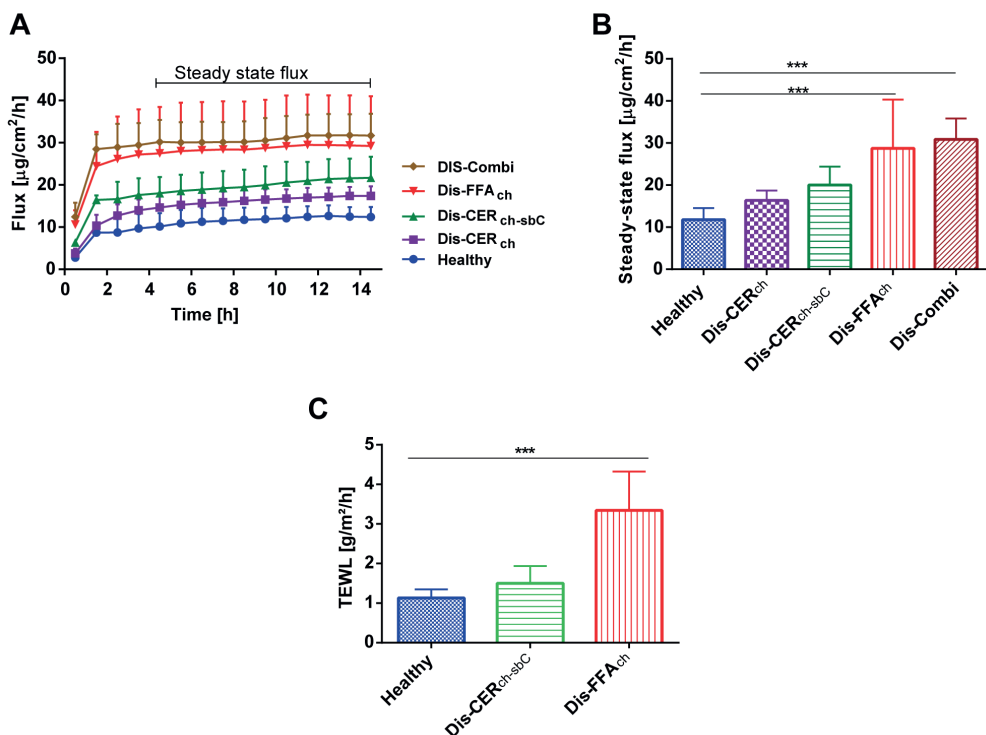


Figure 3: Permeability of the model membranes.

A) The flux of E-PABA across the model membranes over 15 h. B) The average steady-state flux of E-PABA across the model membranes (5-15 h). Data presented as the means \pm SD, $n \geq 6$. E-PABA steady-state flux was significantly higher in the models containing an increased level of short-chain FFAs compared to the healthy skin model, *** ($P < 0.001$). C) TEWL across the model membranes, Dis FFA_{ch} showing significantly higher permeability compared to the healthy skin model, $n = 7$, *** $P < 0.001$.

The flux was significantly higher compared to that of the healthy skin model. To determine which change in composition contributed the most to the reduced SC lipid barrier, the contribution of each of the anomalies in the CER and FFA composition on permeability was examined. First, the CER composition was examined. Replacing 50% of the CER NS and CER AS (C24) by CER NS and CER AS (C16), respectively (Model Dis-CER_{ch}) resulted in a slight but not significant increase in E-PABA flux compared to the healthy skin model. In the next step, the total CERs NS and AS (C24) fraction were increased, of which 50% were the C16 counterparts, while CERs NP and EOS were reduced. This resulted in a further increase (not statistically significant) in permeability. In contrast, when focusing on the FFA composition, the E-PABA steady-state flux across Dis-FFA_{ch}, which contains increased levels of short-chain FFAs, was significantly higher than that across the healthy skin model.

Table 3: The E-PABA steady-state flux across the model membranes with their SD, the wavenumber at 10 °C and midpoint temperatures of the orthorhombic-hexagonal phase transition ($T_{M,OR-HEX}$) in mixtures with protiated CERs and FFA chains and the ordered to disordered phase transition ($T_{M,HEX-LIQ}$) of the v_sCH_2 mode and v_sCD_2 mode in mixtures with protiated CER chains and deuterated FFA chains.

Lipid model	Steady-state flux ($\mu\text{g}/\text{cm}^2/\text{h}$)	wavenumber (cm^{-1})	$T_{M,OR-HEX}$ ($^{\circ}\text{C}$)	v_sCH_2 mode $T_{M,HEX-LIQ}$ ($^{\circ}\text{C}$)	v_sCD_2 mode $T_{M,HEX-LIQ}$ ($^{\circ}\text{C}$)
(D-)Healthy	11.8 ± 2.7	2848.7 ± 0.2	28.0 ± 1.7	67.1 ± 1.5	66.9 ± 2.5
(D-)Dis-CER _{ch}	16.4 ± 2.3	2848.8 ± 0.2	27.1 ± 0.8	65.9 ± 1.4	64.9 ± 2.5
(D-)Dis-CER _{ch-sbc}	20.0 ± 3.6	2848.6 ± 0.2	27.6 ± 0.9	63.1 ± 2.4	62.2 ± 1.7
(D-)Dis-FFA _{ch}	28.7 ± 11.6	2849.0 ± 0.4	24.9 ± 1.3	62.8 ± 0.7	63.7 ± 0.2
(D-)Dis-Combi	30.8 ± 5.0	2849.3 ± 0.3	Undefined	62.9 ± 1.1	63.2 ± 0.1

Changes in lipid composition resulted in increased TEWL in AD patients' skin models

TEWL is often monitored to measure the inside-out skin barrier function in clinical studies. For this reason, TEWL was also used to detect whether selected changes in CERs and FFAs composition resulted in an increased TEWL indicating a reduced lipid barrier. The TEWL values of Healthy, DIS-CER_{ch-sbc}, and Dis FFA_{ch} are plotted in figure 3C. The diseased model membranes show higher TEWL values than the healthy skin model indicating reduced barrier capability. The TEWL value of DIS-CER_{ch-sbc} was 1.2 times higher than that of the healthy skin model and not statistically significant. In contrast, DIS-FFA_{ch} resulted in significantly higher TEWL values, exceeding that of the healthy skin model by 2.8 times. This finding is consistent with the results of the E-PABA permeation study.

Increased levels of short-chain FFAs resulted in lower conformational ordering and significantly lower OR-HEX phase transition temperature

The lateral organization and conformational ordering of the lipids in the healthy and diseased skin models were examined using FTIR. Information about the conformational ordering and packing of lipid chains are provided by the $\nu_s\text{CH}_2$ and δCH_2 frequencies respectively. Low $\nu_s\text{CH}_2$ frequency below 2850 cm^{-1} is characteristic of highly ordered hydrocarbon chains typically packed either in the orthorhombic or hexagonal lateral phase. The thermotropic response of the $\nu_s\text{CH}_2$ mode in the healthy and diseased skin models are presented in figure 4. The average $\nu_s\text{CH}_2$ frequencies for the various models at $10\text{ }^\circ\text{C}$ are presented in Table 3. Dis-Combi exhibits the highest wavenumber indicating a lower conformational ordering compared to the other models. However, the difference was not significant. As the temperature increases, an approximate 1 cm^{-1} rise in the wavenumber occurs between 20 and $40\text{ }^\circ\text{C}$ indicative of an OR-HEX phase transition. The midpoint temperatures of the OR-HEX phase transition ($T_{M,OR-HEX}$) are presented in Table 3. The $T_{M,OR-HEX}$ of Dis-FFA_{ch} is significantly lower than that of the healthy skin model. The $T_{M,OR-HEX}$ of Dis-Combi was not defined indicating that the lipid chains were primarily in the hexagonal phase at the initial temperature.

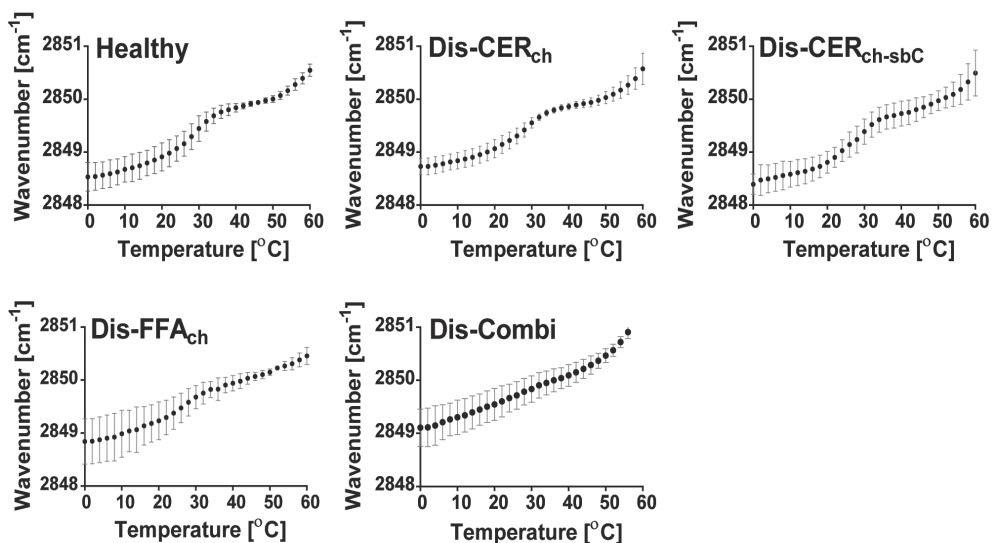


Figure 4: The thermotropic response of the $\nu_s\text{CH}_2$ modes of the lipid models.

The $\nu_s\text{CH}_2$ modes of all models showing frequencies between 0 and $60\text{ }^\circ\text{C}$. The wavenumber at $10\text{ }^\circ\text{C}$ is higher in the Dis-FFA_{ch} and Dis-Combi compared to the healthy skin model indicating the highest conformational ordering in the latter. The midpoint temperature of the OR-HEX phase transition is significantly lower in Dis-FFA_{ch} compared to the healthy skin model while the transition is undefined in Dis-Combi. Data presented as the means \pm SD, $n \geq 3$.

Increased level of short-chain FFAs induced hexagonal lateral packing at physiological temperature

In addition to the ρCH_2 mode, the δCH_2 and δCD_2 modes are affected by the packing arrangements in ordered phases. Broadening or splitting of the scissoring contour signifies an orthorhombic packing. In contrast, hexagonal packing is characterized by a single peak located at $\sim 1467\text{ cm}^{-1}$. The thermotropic response of the δCH_2 frequencies is provided in figure 5. The contour of the δCH_2 modes of the healthy skin model at 10°C is split into a strong doublet located at ~ 1462 and 1473 cm^{-1} indicating orthorhombic lateral packing. At 32°C , the δCH_2 mode of the healthy skin model displayed a weaker doublet with a peak shift in the lower frequency component to $\sim 1467\text{ cm}^{-1}$ indicating a co-existence

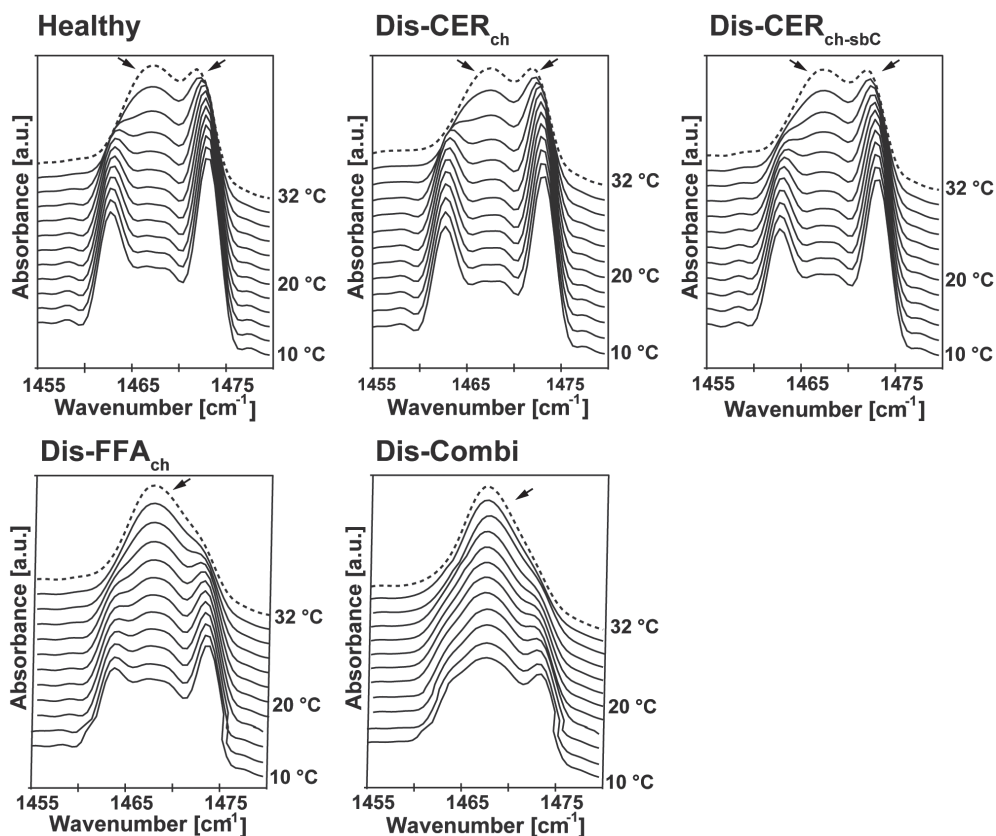


Figure 5: The thermotropic response of the δCH_2 modes of the healthy and diseased skin models.

At 32°C , the modes of Healthy, Dis-CER_{ch} and Dis-CER_{ch-sbC} display two peaks at $\sim 1467\text{ cm}^{-1}$ and 1473 cm^{-1} indicated by two arrows, showing co-existence of orthorhombic and hexagonal lateral packing while those of Dis-FFA_{ch} and Dis-combi display a single peak located at $\sim 1467\text{ cm}^{-1}$ indicated by single arrows representing hexagonal phase.

with a hexagonal phase. A similar profile was observed in the δCH_2 modes of Dis-CER_{ch} and Dis-CER_{ch-sbC}. In contrast, the δCH_2 mode of Dis-FFA_{ch} displayed a weak doublet at 10 °C indicating a substantially lower proportion of lipids in an orthorhombic phase already at low temperatures. The δCH_2 mode of Dis-combi at 10 °C displays a strong central peak located at $\sim 1467\text{ cm}^{-1}$ and a weak peak at $\sim 1473\text{ cm}^{-1}$ showing that the lipids form predominantly a hexagonal phase. At 32 °C, the δCH_2 mode of Dis-FFA_{ch} and Dis-combi display a single peak at $\sim 1467\text{ cm}^{-1}$ indicating the lipids exist predominantly in the hexagonal phase.

Changes in CER composition reduced the mixing of CERs and FFAs at low temperature

By substituting the FFAs with their deuterated counterparts, the phase transition of the CER and DFFA chains can be monitored simultaneously since the frequencies of vibrational modes of protonated and deuterated acyl chains appear at different positions in the IR spectrum. The $T_{M, \text{HEX-LIQ}}$ of the $\nu_5\text{CH}_2$ and $\nu_5\text{CD}_2$ modes are presented in table 3. In all the models, the CERs and DFFA chains melted in the same temperature range (not shown) indicating the mixing of the CER and DFFA chains. To obtain detailed information on the mixing of protonated and deuterated chains, we examined the δCH_2 and δCD_2 modes. When CER and DFFA chains mix in the same orthorhombic lattice and are thus adjacent to each other, the δCH_2 and δCD_2 vibrational frequencies are sufficiently different in energy to preclude the short-range vibrational coupling between the chains thereby precluding the mode splitting characteristic of orthorhombic packing. Consequently, only a single peak is observed in the δCH_2 and δCD_2 modes. The δCD_2 modes of the healthy and diseased skin models are displayed in figure 6. At 10 °C, the δCD_2 mode from the DFFAs in D-Healthy, D-Dis-CER_{ch'} and D-Dis-CER_{ch-sbC} are split into weak doublets indicating coupling of adjacent DFFA chains. A fraction of the DFFA chains were still able to interact as they are located at neighbouring positions in the orthorhombic lattice [38]. The magnitude of the band splitting is indicative of the degree of inter-chain interactions and the size of the domains with an orthorhombic organization [12, 37, 39]. The splitting values in the δCD_2 mode of the samples were 3.1 ± 0.2 , 3.5 ± 0.2 , and $4.2 \pm 0.5\text{ cm}^{-1}$ for D-Healthy, D-Dis-CER_{ch'} and D-Dis-CER_{ch-sbC} respectively. The mixing of the CERs and DFFA chains was more enhanced in the healthy compared with the diseased skin models as indicated by the lower splitting value and shallower minima between the δCD_2 peaks in the spectrum. D-Dis-CER_{ch-sbC} exhibited the highest splitting value and deepest minima between the δCD_2 peaks indicating the most CD_2 - CD_2 interaction. Upon further increase in temperature, the splitting collapse into single peaks in all the models at $\sim 25\text{ °C}$ resulting from

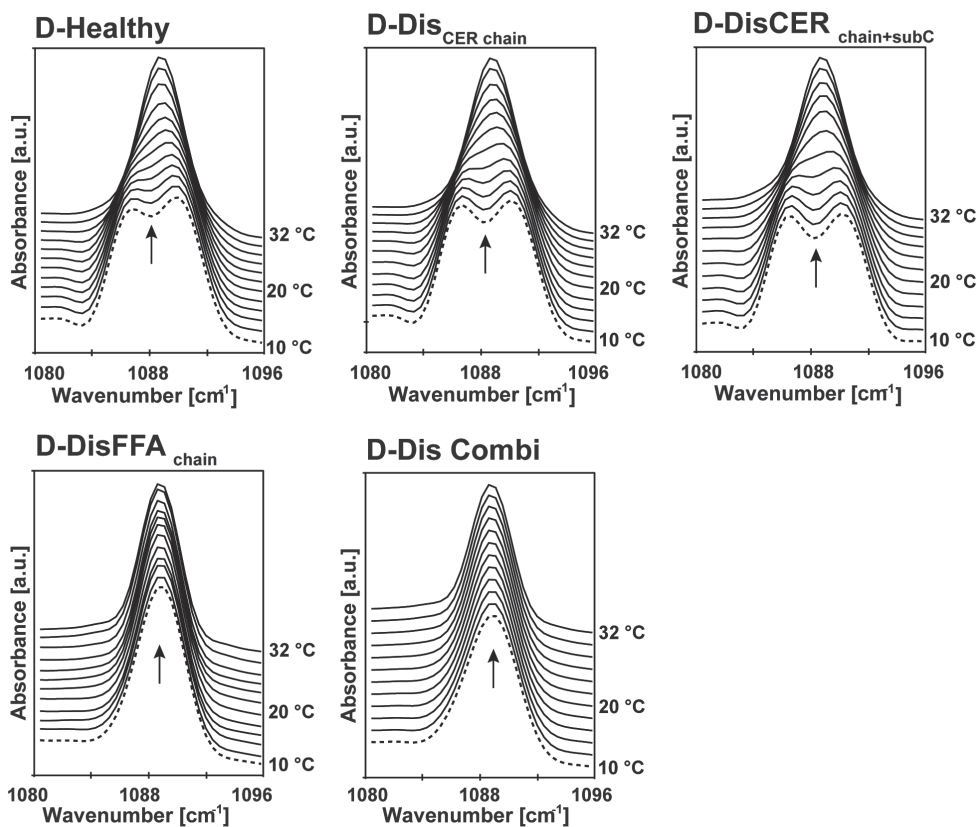


Figure 6: δCD_2 modes of the lipid models.

D-Dis-CER_{ch-sbC} δCD_2 mode doublets displayed the deepest minima at 10 °C when compared with D-Dis-CER_{ch} and D-Healthy indicating the greatest interaction between DFFA chains in the former, while D-Dis-FFA_{ch} and D-Dis-combi displayed single peaks since the chains pack predominantly in a hexagonal phase.

a combined effect of mixing of the CERs and DFFA chains and transition to the hexagonal phase. The δCD_2 modes of D-Dis-FFA_{ch} and Dis-combi at 10 °C exhibit a single central peak since the chains pack predominantly in a hexagonal phase as shown in the corresponding protonated mixtures.

Short-chain CERs present in diseased models are less ordered than the physiological chain length CERs

Another application of selective lipid deuteration is monitoring the individual acyl chain of CER NS conformational ordering in the $\nu_s\text{CD}_2$ mode of mixtures. For these measurements, the deuterated acyl chain in either CER NS (C16) or CER NS (C24) in the Dis-CER_{ch} model was selected. We compared the conformational ordering of the long and short chains in Dis CER_{ch}. The compositions of both

models are presented in table 1 and their $\nu_5\text{CD}_2$ wavenumbers are shown in figure 7. The $\nu_5\text{CD}_2$ wavenumber of Dis-CER_{ch-D-NS (C16)} at 10 °C is $2089.9 \pm 0.4 \text{ cm}^{-1}$ being significantly higher than $2088.4 \pm 0.04 \text{ cm}^{-1}$ in Dis-CER_{ch-D-NS (C24)} ($n = 3, p < 0.01$) indicating lower conformational ordering in the former.

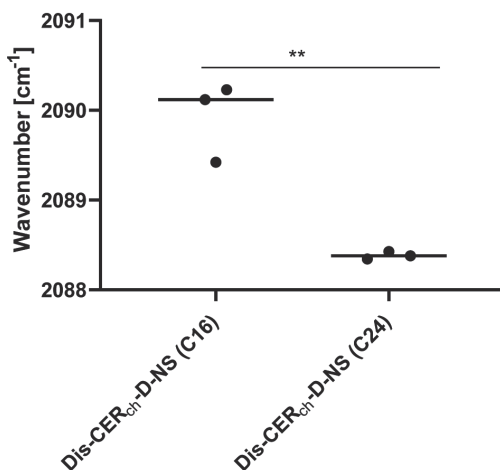


Figure 7: $\nu_5\text{CD}_2$ wavenumber at 10 °C.

The average peak position of the $\nu_5\text{CD}_2$ curve for Dis-CER_{ch-D-NS (C16)} and Dis-CER_{ch-D-NS (C24)}. Dis-CER_{ch-D-NS (C16)} wavenumber is significantly higher than that of Dis-CER_{ch-D-NS (C24)} indicating more disordering of the lipid chains in the former $n = 3, ** p < 0.01$

Changes in CER composition affects the lamellar organization in skin lipid models

The influence of the changes in lipid composition in the diseased model on the lamellar organization was examined by SAXD. The SAXD profile of the healthy skin model is shown earlier in figure 2A while those of the diseased models are provided in figure 8. Both the healthy and the diseased skin models revealed a number of diffraction peaks attributed to the LPP and the SPP with repeat distances of $\sim 12.8 \text{ nm}$ and 5.4 nm respectively. The diffraction pattern of Dis-CER_{ch} displayed an extra peak at $q = 1.5 \text{ nm}^{-1}$ that did not match the spacing of either the LPP or SPP, suggesting the presence of an additional phase. When focusing on Dis-CER_{ch-sbC} the LPP formation was drastically reduced compared to the healthy skin model. Meanwhile, the diffraction pattern of Dis-FFA_{ch} did not markedly differ from that of Dis-CER_{ch}. The diffraction pattern of Dis-combi was similar to the Dis-CER_{ch-sbC} model aside from an extra peak at $q = 1.3 \text{ nm}^{-1}$ in the diffraction profile of the former. All the SAXD profiles studied revealed the presence of a peak related to a spacing of 3.4 nm attributed to crystalline domains of phase-separated CHOL.

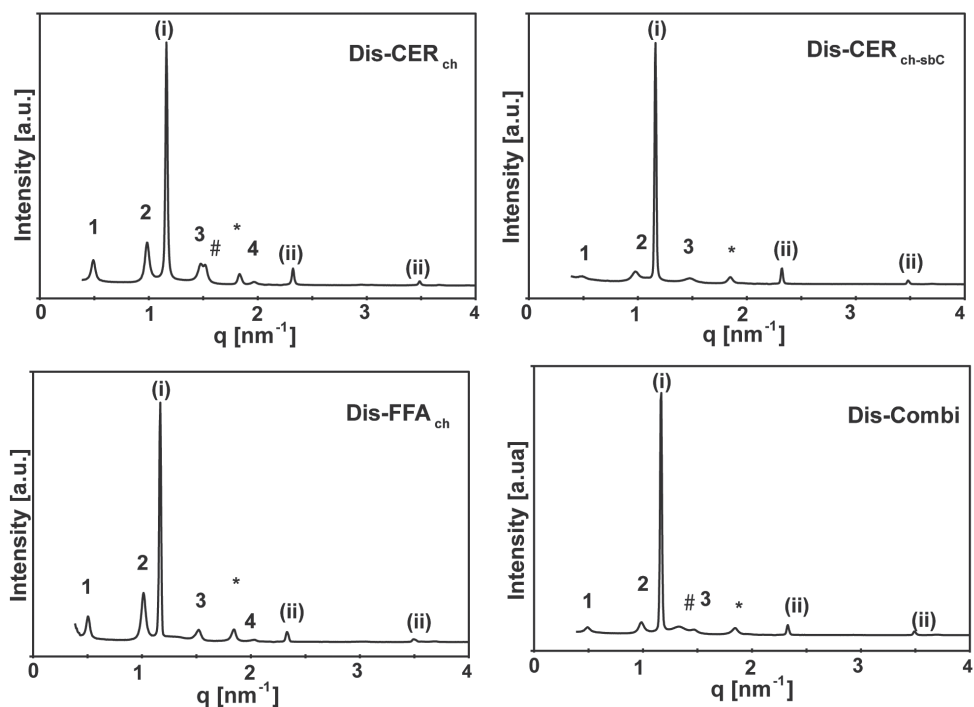


Figure 8: SAXD profile of the diseased skin models at 25 °C.

The arabic numbers (1, 2, 3,4) indicate the 1st, 2nd, 3rd, and 4th order diffraction peaks of the LPP respectively, while the roman numbers (i, ii, iii) indicate the diffraction orders of the SPP. In the diffraction profile of Dis-CER_{ch}, the peak positions at $q = 0.48, 0.97, 1.45,$ and 1.92 nm^{-1} are attributed to the LPP with a repeat distance of 12.9 nm. While in Dis-CER_{ch-sbC} diffraction profile, the peak positions at $q = 0.49, 0.98,$ and 1.47 nm^{-1} are attributed to the LPP with a repeat distance of 12.8 nm. In the diffraction pattern of Dis-FFA_{ch}, the reflections at $q = 0.49, 0.99, 1.48,$ and 1.96 nm^{-1} are attributed to the LPP with a repeat distance of 12.7 nm. Finally, the Dis-Combi diffraction profile is characterized by a series of sequential peaks positioned at $q = 0.48, 0.97,$ and 1.47 nm^{-1} attributed to the LPP with a repeat distance of 12.9 nm. In addition, an extra peak indicated by # is located at $q=1.3$. In all the models, the series of diffraction peaks positioned at $q = 1.16, 2.3,$ and 3.48 nm^{-1} indicate the 1st, 2nd, and 3rd diffraction orders attributed to the SPP with a repeat distance of 5.4 nm and the peaks attributed to crystalline CHOL located at $q = 1.83 \text{ nm}^{-1}$ are indicated by an asterisk (*).

In addition to the lamellar organization, X-ray diffraction provides information about lateral packing of lipid chains. Hexagonal lateral packing is characterized by a single $\sim 0.41 \text{ nm}$ peak, while orthorhombic packing is characterized by two peaks at ~ 0.41 and $\sim 0.37 \text{ nm}$. We examined the changes in lamellar and lateral lipid organization simultaneously. A temperature range from 25 to 85 °C was chosen. We selected three models; Healthy, along with Dis-CER_{ch-sbC} and Dis-FFA_{ch}, which exhibited pronounced alterations in the lamellar and lateral lipid organization respectively. Sequential SAXD and WAXD plots as a function of

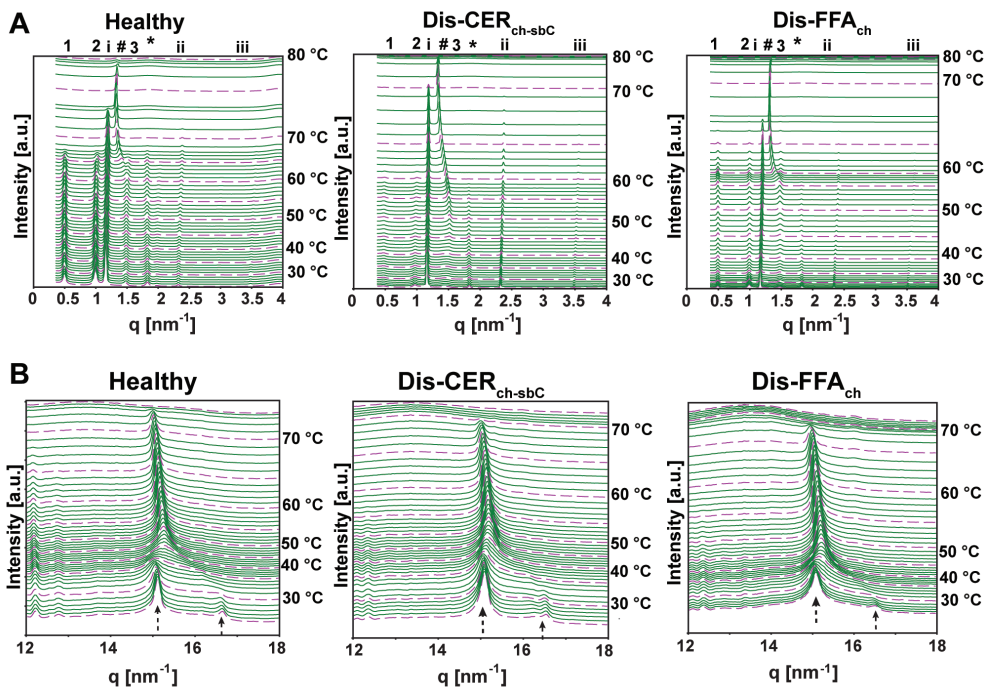


Figure 9: X-ray diffraction profile of the healthy and diseased skin lipid models 25 to 85 °C.

A) SAXD profile showing the changes in the lamellar organization with increasing temperature. The arabic numbers (1, 2, 3) indicate the diffraction orders of the LPP. The roman numbers (i, ii, iii) indicate the diffraction orders of the SPP. * and # indicate phase-separated CHOL and additional phase respectively. B) WAXD showing the changes in the lateral packing with increasing temperature. The arrows show the 0.41 and 0.37 nm peaks located at $q = \sim 15.20$ and 16.7 nm^{-1} respectively.

temperature are presented in figures 9A and B respectively, while a summary of the phase changes in the various models is provided in Table 4. In all three models, the SPP is generally more thermostable than the LPP and the collapse of the SPP coincides with the disappearance of the hexagonal phase. Focusing on the WAXD profile, the intensity of the 0.41 nm peak in the models decreases with increasing temperature, shifts slightly to the right before finally disappearing marking the collapse of the hexagonal phase. Dis-CER_{ch-sbC} LPP was the least thermostable, while Dis-FFA_{ch} exhibited the least thermostable orthorhombic and hexagonal phases. In the SAXD profile, the 3.4 nm reflections attributed to CHOL also disappear at 70 °C, 60 °C, and 57 °C in Healthy, Dis-CER_{ch-sbC}, and Dis-FFA_{ch} respectively. After the disappearance of the CHOL peaks, a broad 3.4 nm peak indicated by # appear in the models, shifting to lower q-values and increasing in intensity with increasing temperature, remaining present till 85 °C.

Table 4: Summary of phase changes on temperature increase.

Model	Temperature at phase disappearance (°C)			
	LPP	SPP	OR phase	HEX phase
Healthy	70	74	38	74
Dis-CER _{ch-sbc}	63	72	37	71
Dis-FFA _{ch}	68	68	33	68

DISCUSSION

Though several studies have reported the association of filaggrin deficiency with AD and ichthyosis vulgaris [40-44], the role of filaggrin gene mutation in barrier function impairment of AD patients remains contentious [16, 22, 45], suggesting other factors also play a role. Some studies have reported a strong correlation between altered lipid composition and TEWL in the skin of AD patients [23, 26, 46]. As CERs and FFAs have a common synthetic pathway [47, 48], changes in their composition were observed simultaneously in AD patients' skin including reduction in chain length of CERs and FFAs and changes in CER subclass composition. However, it is not yet known which changes in the lipid composition impact most on the skin barrier in AD, as multiple compositional changes occur simultaneously. Therefore, we have investigated the contribution of various abnormalities common to the lipid matrix of AD patients' skin using model membrane systems. This allows us to systematically study the impact of each of these changes on the lipid barrier [49].

For a model membrane to be used as a substitute for the native skin, preferably the lipid composition and organization should be similar. In the present study, we developed a model membrane mimicking the CER composition of human SC as closely as possible taking into account, the commercially available synthetic CER subclasses. A previous study from our group employed a model based on pig CER composition, as when the model was developed most of the CER subclasses in pigskin were available [35]. The pig CER model differs from the human CER model by the presence of short-chain CERs (particularly CER NP (C16)) and the presence of an abundant amount of CER NS at the expense of the more hydrophilic subclasses. When comparing the human lipid model with that of the previous pig CER model, there was hardly any difference in the lipid organization in the models. In both models, the lipids assembled in the two characteristic lamellar phases and adopted orthorhombic lateral packing. However, the LPP repeat distance was slightly longer in the human CER model, thereby more closely resembling that of the native human skin.

Changes in the CER profile altered the lamellar organization in model SC

Changes in the CER subclass distribution, as well as a reduction in the chain length, occur in the skin of AD patients. In a previous report, the level of CER NS (C16) was shown to correlate strongly with the TEWL values in AD patients' skin [23]. Subsequently, an excellent correlation was demonstrated between CER chain length reduction and barrier function impairment in AD patients [26]. In the current study, we show that the inclusion of CERs NS(C16) and AS (C16) in the SC model membrane (Dis-CER_{ch}) results in a slightly higher permeability compared to the healthy skin model. By selective deuteration of either the C16 fatty acid acyl chain of CER NS (C16) or the C24 fatty acid acyl chain of CER NS (C24) in Dis-CER_{ch}, we demonstrated that shorter chain length CERs are less conformationally ordered than the long physiological acyl chain length CERs. This is a plausible explanation for the slight increase in the permeability of Dis-CER_{ch} compared to the healthy skin model. Previous studies reported significantly higher permeability in model membranes with the CER fraction made up of short-chain length CERs with fatty acid acyl chain length C4-C8 as opposed to physiological chain length CERs [50, 51]. While the substitution of CER NS/AS (C24) in the healthy skin model by their C16 CER counterparts in Dis-CER_{ch} is substantial, the permeability difference between the models was not significant. This indicates that although a strong correlation was observed between the skin barrier and the presence of CER NS (C16) in the clinical studies, this might not be the prime underlying factor for an impaired skin barrier.

For a comprehensive representation of the CER profile of AD patients, we made additional changes in CER composition, thereby mimicking the CER subclass composition more closely as found in SC of AD patients. The levels of sphingosine-based CERs AS and NS were increased at the expense of CER NP and CER EOS fractions. These simultaneous changes in the CER chain length and subclass (Dis-CER_{ch-sbC}) resulted in a further increase in permeability, but not yet significant. The compositional change resulted in a drastic reduction of the lipid fraction forming the LPP, mainly attributed to the reduction in CER EOS level as reported in the literature. For instance, in a lipid mixture based on human isolated CERs, predominantly the SPP was formed at the expense of the LPP when CER EOS was excluded from the mixture [52]. In addition, model mixtures based on synthetic lipids formed exclusively the short phase when CER EOS was excluded and showed a reduced lipid barrier [53-56]. Furthermore, a reduced level of CER NP is also expected to contribute to a reduced lipid barrier, as recently it was reported that a higher permeability was observed in sphingosine-based CERs membranes compared to their phytosphingosine counterparts [34]. Although

a further increase in permeability was observed, our results indicate that a change in ceramide subclass composition is not a major underlying factor for the increased permeability of the skin in AD patients.

Increased level of short-chain length FFAs reduced the packing density and conformational ordering in model SC

While maintaining the CER composition of the healthy skin, an increased fraction of short-chain FFAs (Dis-FFA_{ch}), as reported in lesional skin of AD patients [27], resulted in a significant increase in permeability compared with the healthy skin model and the changes in permeability were more pronounced than induced by the changes in CER profile. By calculating the average carbon chain length in the diseased skin models relative to the healthy skin model, we deduced that increased levels of short-chain FFAs (mostly C16 and C18) decreased the average FFA chain length by 2.4 carbon atoms in Dis-FFA_{ch}. While increased levels of CERs NS and AS (C16) at the expense of their C24 counterparts resulted in a 1.0 carbon atom reduction in Dis-CER_{ch}. Interestingly, a greater reduction in carbon atoms (3.8) was obtained in Dis-CER_{ch-sbC}, attributed to the reduced concentration of the exceptionally long CER EOS. The compositional changes in CERs composition had more consequences on the carbon chain length than compositional changes in the FFAs and therefore the FFAs did not have a stronger reduction in chain length that could account for the higher impact on the barrier function.

In a previous study, it was shown that shorter FFA chain lengths resulted in an increased conformational disordering and a reduced barrier [57], but in addition to this change in conformational ordering, the reduction in FFA chain length in the present study resulted also in a reduced lipid fraction assembled in the orthorhombic phase. This change in lateral packing was not observed by reducing the average chain length of the CERs and may therefore explain the higher permeability in the Dis-FFA_{ch} model. Our results agree with those of previous studies that reported an increased permeability of model membranes as a consequence of reduced packing density [35, 58]. A reduced lateral packing density was also observed in AD skin in comparison with healthy skin model and may be caused by the shorter FFA rather than the shorter CER chains in AD [25, 26, 59]. Thus, the reduction in FFA chain length can result in the reduced conformational ordering of lipid chains observed in AD patients' skin, compared to healthy skin model [26].

There was extensive mixing of the CER and FFA chains in the models studied since no domains of phase-separated FFA were identified in the δCD_2 modes of the models. As previously reported, the δCD_2 of pure DFFA mixture is

characterized by large splitting, measuring 7.2 cm^{-1} and remaining present till $72\text{ }^{\circ}\text{C}$ [34]. The introduction of C16 acyl chain length CERs NS and AS in D-DIS CER_{ch} resulted in slightly less miscibility of the CER and FFA chains compared to the healthy skin model at the low temperature indicated by the slight increase in splitting value of the δCD_2 mode. Further increase in splitting value was recorded after modulations in the CER subclass fractions. Our data suggest that a lower level of CER EOS may reduce the miscibility of CERs and FFA as we observed that the absence of CER EOS reduced the miscibility of CERs and FFAs (unpublished work).

Previous FTIR studies of ternary SC model mixtures (CER NS (C24)/CHOL/FFA (C16) or (C24)) reported phase separation when FFA (C16) was incorporated into the SC model mixture and was homogeneously mixed on the incorporation of C24 chain length FFA [36, 60, 61]. Separate domains of FFA occurred in CER NS (C24)/CHOL/FFA (C16) as the short-chain length FFA did not fit with the long-chain CER. The reason for the extensive mixing of the CERs and FFA chains in D-DIS CER_{ch} irrespective of the introduction of short-chain length CERs may be the variety in FFA chain lengths in the present mixture. Despite the simultaneous reduction of CER EOS in Dis-CER_{ch-sbC}, CER C16 still fitted in the lamellar structure. This is evident by the absence of phase-separated CER in the CH_2 mode of D-Dis-CER_{ch-sbC} (not shown). The absence of phase separation was also reported in complex SC models containing CER (C24)/CHOL/FFAs (C16-C24) and our pig SC models [36, 62]. A previous study reported that such FFA chain length distribution in the SC may be required for the coexistence of a proportion of solid lipids and fluid domains, which is suggested to be important for normal skin barrier function [63]. Complex SC model mixtures with a variety of CER subclasses and FFAs chain length as employed in the current study, appear to be resilient to phase separation even when the level of short acyl chain lengths are increased. CER headgroup diversity and FFA chain length distribution may provide robustness to the barrier necessary for withstanding challenges in the environment.

SUMMARY AND CONCLUSIONS

The changes in the lipid composition of model membranes that mimic the changes observed in AD patients resulted in a reduced barrier capability. Modulations in CER composition altered the lamellar organization, while changes in FFAs composition induced the phase transition from the dense orthorhombic to the less dense hexagonal packing. The present study shows

that FFA compositional changes in AD skin has a greater impact on the barrier function than changes in the CER profile most likely due to a change in packing density, which does not occur when shortening the CER chain length. These findings are highly suggestive that AD therapies that target the normalization of the FFA composition are at least as important as the normalization of CER composition.

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REFERENCES

1. Hannun, Y. A., Functions of ceramide in coordinating cellular responses to stress. *Science* **1996**, 274 (5294), 1855-9.
2. Brasaemle, D. L., Thematic review series: adipocyte biology. The perilipin family of structural lipid droplet proteins: stabilization of lipid droplets and control of lipolysis. *J. Lipid Res.* **2007**, 48 (12), 2547-59.
3. van Blitterswijk, W. J.; van der Luit, A. H.; Veldman, R. J.; Verheij, M.; Borst, J., Ceramide: second messenger or modulator of membrane structure and dynamics? *Biochem. J.* **2003**, 369 (Pt 2), 199-211.
4. Wang, X., Lipid signaling. *Curr. Opin. Plant Biol.* **2004**, 7 (3), 329-336.
5. Quinn, P. J., Structure of sphingomyelin bilayers and complexes with cholesterol forming membrane rafts. *Langmuir* **2013**, 29 (30), 9447-56.
6. Gonzalez-Ramirez, E. J.; Artetxe, I.; Garcia-Arribas, A. B.; Goni, F. M.; Alonso, A., Homogeneous and heterogeneous bilayers of ternary lipid compositions containing equimolar ceramide and cholesterol. *Langmuir* **2019**, 35 (15), 5305-5315.
7. Weerheim, A.; Ponc, M., Determination of stratum corneum lipid profile by tape stripping in combination with high-performance thin-layer chromatography. *Arch. Dermatol. Res.* **2001**, 293 (4), 191-199.
8. Mendelsohn, R.; Flach, C. R.; Moore, D. J., Determination of molecular conformation and permeation in skin via IR spectroscopy, microscopy, and imaging. *Biochim. Biophys. Acta* **2006**, 1758 (7), 923-933.
9. Bouwstra, J. A.; Gooris, G. S.; van der Spek, J. A.; Bras, W., Structural investigations of human stratum corneum by small-angle X-ray scattering. *J. Invest. Dermatol.* **1991**, 97 (6), 1005-12.
10. White, S. H.; Mirejovsky, D.; King, G. I., Structure of lamellar lipid domains and corneocyte envelopes of murine stratum corneum. An X-ray diffraction study. *Biochemistry* **1988**, 27 (10), 3725-32.
11. Bouwstra, J.; Gooris, G.; Ponc, M., The lipid organisation of the skin barrier: Liquid and crystalline domains coexist in lamellar phases. *Journal of biological physics* **2002**, 28 (2), 211-223.
12. Boncheva, M.; Damien, F.; Normand, V., Molecular organization of the lipid matrix in intact stratum corneum using ATR-FTIR spectroscopy. *Biochim. Biophys. Acta* **2008**, 1778 (5), 1344-55.
13. Norlen, L.; Nicander, I.; Lundsjo, A.; Cronholm, T.; Forslind, B., A new HPLC-based method for the quantitative analysis of inner stratum corneum lipids with special reference to the free fatty acid fraction. *Arch. Dermatol. Res.* **1998**, 290 (9), 508-16.
14. van Smeden, J.; Boiten, W. A.; Hankemeier, T.; Rissmann, R.; Bouwstra, J. A.; Vreeken, R. J., Combined LC/MS-platform for analysis of all major stratum corneum lipids, and the profiling of skin substitutes. *Biochim. Biophys. Acta* **2014**, 1841 (1), 70-79.
15. t'Kindt, R.; Jorge, L.; Dumont, E.; Couturon, P.; David, F.; Sandra, P.; Sandra, K., Profiling and characterizing skin ceramides using reversed-phase liquid chromatography-quadrupole time-of-flight mass spectrometry. *Anal. Chem.* **2012**, 84 (1), 403-411.
16. van Smeden, J.; Bouwstra, J. A., Stratum corneum lipids: Their role for the skin barrier function in healthy subjects and atopic dermatitis patients. *Curr. Probl. Dermatol.* **2016**, 49, 8-26.
17. Wertz, P. W.; Miethke, M. C.; Long, S. A.; Strauss, J. S.; Downing, D. T., The composition of the ceramides from human stratum corneum and from comedones. *J. Invest. Dermatol.* **1985**, 84 (5), 410-2.

18. Motta, S.; Monti, M.; Sesana, S.; Caputo, R.; Carelli, S.; Ghidoni, R., Ceramide composition of the psoriatic scale. *Biochim. Biophys. Acta* **1993**, *1182* (2), 147-51.
19. Alanne, S.; Nermes, M.; Soderlund, R.; Laitinen, K., Quality of life in infants with atopic dermatitis and healthy infants: a follow-up from birth to 24 months. *Acta Paediatr.* **2011**, *100* (8), e65-70.
20. Proksch, E.; Folster-Holst, R.; Jensen, J. M., Skin barrier function, epidermal proliferation and differentiation in eczema. *J. Dermatol. Sci.* **2006**, *43* (3), 159-69.
21. Jensen, J. M.; Folster-Holst, R.; Baranowsky, A.; Schunck, M.; Winoto-Morbach, S.; Neumann, C.; Schutze, S.; Proksch, E., Impaired sphingomyelinase activity and epidermal differentiation in atopic dermatitis. *J. Invest. Dermatol.* **2004**, *122* (6), 1423-31.
22. Jakasa, I.; Koster, E. S.; Calkoen, F.; Irwin McLean, W. H.; Campbell, L. E.; Bos, J. D.; Verberk, M. M.; Kezic, S., Skin Barrier Function in Healthy Subjects and Patients with Atopic Dermatitis in Relation to Filaggrin Loss-of-Function Mutations. *J. Invest. Dermatol.* **2011**, *131* (2), 540-542.
23. Ishikawa, J.; Narita, H.; Kondo, N.; Hotta, M.; Takagi, Y.; Masukawa, Y.; Kitahara, T.; Takema, Y.; Koyano, S.; Yamazaki, S.; Hatamochi, A., Changes in the ceramide profile of atopic dermatitis patients. *J. Invest. Dermatol.* **2010**, *130* (10), 2511-4.
24. Di Nardo, A.; Wertz, P.; Giannetti, A.; Seidenari, S., Ceramide and cholesterol composition of the skin of patients with atopic dermatitis. *Acta Derm-Venereol.* **1998**, *78* (1), 27-30.
25. Imokawa, G.; Abe, A.; Jin, K.; Higaki, Y.; Kawashima, M.; Hidano, A., Decreased level of ceramides in stratum corneum of atopic dermatitis: an etiologic factor in atopic dry skin? *J. Invest. Dermatol.* **1991**, *96* (4), 523-6.
26. Janssens, M.; van Smeden, J.; Gooris, G. S.; Bras, W.; Portale, G.; Caspers, P. J.; Vreeken, R. J.; Hankemeier, T.; Kezic, S.; Wolterbeek, R.; Lavrijsen, A. P.; Bouwstra, J. A., Increase in short-chain ceramides correlates with an altered lipid organization and decreased barrier function in atopic eczema patients. *J. Lipid Res.* **2012**, *53* (12), 2755-66.
27. van Smeden, J.; Janssens, M.; Kaye, E. C.; Caspers, P. J.; Lavrijsen, A. P.; Vreeken, R. J.; Bouwstra, J. A., The importance of free fatty acid chain length for the skin barrier function in atopic eczema patients. *Exp. Dermatol.* **2014**, *23* (1), 45-52.
28. Elias, P. M., Primary role of barrier dysfunction in the pathogenesis of atopic dermatitis. *Exp. Dermatol.* **2018**, *27* (8), 847-851.
29. Cork, M. J.; Danby, S. G.; Vasilopoulos, Y.; Hadgraft, J.; Lane, M. E.; Moustafa, M.; Guy, R. H.; MacGowan, A. L.; Tazi-Ahnini, R.; Ward, S. J., Epidermal Barrier Dysfunction in Atopic Dermatitis. *J. Invest. Dermatol.* **2009**, *129* (8), 1892-1908.
30. Wertz, P.W.; Dowing, D.T., *Physiology, biochemistry and molecular biology of the skin*. second ed.; Oxford University Press: New York, 1991; Vol. 1, p 205-236.
31. Schmitt, T.; Lange, S.; Sonnenberger, S.; Dobner, B.; Demé, B.; Neubert, R. H. H.; Gooris, G.; Bouwstra, J. A., Determination of the influence of C24 D/(2R)- and L/(2S)-isomers of the CER[AP] on the lamellar structure of stratum corneum model systems using neutron diffraction. *Chemistry and Physics of Lipids* **2017**, *209*, 29-36.
32. International Conference on Harmonization (ICH) Q2 (R1), Validation of analytical procedures: Text and Methodology. 2005.
33. European Pharmacopoeia. 8 ed.; European Directorate for the quality of medicines & healthcare. Strasbourg, 2014; Vol. 1.
34. Uche, L. E.; Gooris, G. S.; Beddoes, C. M.; Bouwstra, J. A., New insight into phase behavior and permeability of skin lipid models based on sphingosine and phytosphingosine ceramides. *Biochim. Biophys. Acta* **2019**, *1861* (7), 1317-1328.
35. Mojumdar, E. H.; Helder, R. W.; Gooris, G. S.; Bouwstra, J. A., Monounsaturated fatty acids reduce the barrier of stratum corneum lipid membranes by enhancing the formation of a hexagonal lateral packing. *Langmuir* **2014**, *30* (22), 6534-43.

36. Oguri, M.; Gooris, G. S.; Bito, K.; Bouwstra, J. A., The effect of the chain length distribution of free fatty acids on the mixing properties of stratum corneum model membranes. *Biochim. Biophys. Acta* **2014**, *1838* (7), 1851-1861.
37. Mendelsohn, R.; Liang, G. L.; Strauss, H. L.; Snyder, R. G., IR spectroscopic determination of gel state miscibility in long-chain phosphatidylcholine mixtures. *Biophys. J.* **1995**, *69* (5), 1987-1998.
38. Groen, D.; Gooris, G. S.; Bouwstra, J. A., Model membranes prepared with ceramide EOS, cholesterol, and free fatty acids form a unique lamellar phase. *Langmuir* **2010**, *26* (6), 4168-75.
39. Snyder, R. G.; Goh, M. C.; Srivatsavoy, V. J. P.; Strauss, H. L.; Dorset, D. L., Measurement of the growth-kinetics of microdomains in binary N-alkane solid-solutions by infrared-spectroscopy. *J. Phys. Chem-US* **1992**, *96* (24), 10008-10019.
40. Rodriguez, E.; Illig, T.; Weidinger, S., Filaggrin loss-of-function mutations and association with allergic diseases. *Pharmacogenomics* **2008**, *9* (4), 399-413.
41. Gruber, R.; Elias, P. M.; Crumrine, D.; Lin, T. K.; Brandner, J. M.; Hachem, J. P.; Presland, R. B.; Fleckman, P.; Janecke, A. R.; Sandilands, A.; McLean, W. H.; Fritsch, P. O.; Mildner, M.; Tschachler, E.; Schmuth, M., Filaggrin genotype in ichthyosis vulgaris predicts abnormalities in epidermal structure and function. *Am. J. Pathol.* **2011**, *178* (5), 2252-63.
42. Irvine, A. D.; McLean, W. H.; Leung, D. Y., Filaggrin mutations associated with skin and allergic diseases. *N. Engl. J. Med.* **2011**, *365* (14), 1315-27.
43. Hudson, T. J., Skin barrier function and allergic risk. *Nat. Genet.* **2006**, *38* (4), 399-400.
44. McGrath, J. A., Filaggrin and the great epidermal barrier grief. *Australas J. Dermatol.* **2008**, *49* (2), 67-73; quiz 73-4.
45. Danso, M.; Boiten, W.; van Drongelen, V.; Gmelig Meijling, K.; Gooris, G.; El Ghalbzouri, A.; Absalah, S.; Vreeken, R.; Kezic, S.; van Smeden, J.; Lavrijsen, S.; Bouwstra, J., Altered expression of epidermal lipid bio-synthesis enzymes in atopic dermatitis skin is accompanied by changes in stratum corneum lipid composition. *J. Dermatol. Sci.* **2017**, *88* (1), 57-66.
46. Janssens, M.; van Smeden, J.; Gooris, G. S.; Bras, W.; Portale, G.; Caspers, P. J.; Vreeken, R. J.; Kezic, S.; Lavrijsen, A. P.; Bouwstra, J. A., Lamellar lipid organization and ceramide composition in the stratum corneum of patients with atopic eczema. *J. Invest. Dermatol.* **2011**, *131* (10), 2136-8.
47. van Smeden, J.; Janssens, M.; Boiten, W. A.; van Drongelen, V.; Furio, L.; Vreeken, R. J.; Hovnanian, A.; Bouwstra, J. A., Intercellular skin barrier lipid composition, and organization in Netherton syndrome patients. *J. Invest. Dermatol.* **2014**, *134* (5), 1238-1245.
48. Kihara, A., Synthesis and degradation pathways, functions, and pathology of ceramides and epidermal acylceramides. *Prog. Lipid Res.* **2016**, *63*, 50-69.
49. Kováčik, A.; Pullmannová, P.; Maixner, J.; Vávrová, K., Effects of ceramide and dihydroceramide stereochemistry at C-3 on the phase behavior and permeability of skin lipid membranes. *Langmuir* **2018**, *34* (1), 521-529.
50. Skolova, B.; Janusova, B.; Zbytovska, J.; Gooris, G.; Bouwstra, J.; Slepicka, P.; Berka, P.; Roh, J.; Palat, K.; Hrabalek, A.; Vavrova, K., Ceramides in the skin lipid membranes: length matters. *Langmuir* **2013**, *29* (50), 15624-33.
51. Janusova, B.; Zbytovska, J.; Lorenc, P.; Vavrysova, H.; Palat, K.; Hrabalek, A.; Vavrova, K., Effect of ceramide acyl chain length on skin permeability and thermotropic phase behavior of model stratum corneum lipid membranes. *Biochim. Biophys. Acta* **2011**, *1811* (3), 129-37.
52. Bouwstra, J. A.; Gooris, G. S.; Dubbelaar, F. E.; Ponec, M., Phase behavior of lipid mixtures based on human ceramides: coexistence of crystalline and liquid phases. *J. Lipid Res.* **2001**, *42* (11), 1759-70.

53. Groen, D.; Gooris, G. S.; Barlow, D. J.; Lawrence, M. J.; van Mechelen, J. B.; Deme, B.; Bouwstra, J. A., Disposition of ceramide in model lipid membranes determined by neutron diffraction. *Biophys. J.* **2011**, *100* (6), 1481-1489.
54. Mojumdar, E. H.; Groen, D.; Gooris, G. S.; Barlow, D. J.; Lawrence, M. J.; Deme, B.; Bouwstra, J. A., Localization of cholesterol and fatty acid in a model lipid membrane: a neutron diffraction approach. *Biophys. J.* **2013**, *105* (4), 911-8.
55. de Jager, M.; Groenink, W.; Bielsa i Guivernau, R.; Andersson, E.; Angelova, N.; Ponec, M.; Bouwstra, J., A novel in vitro percutaneous penetration model: evaluation of barrier properties with p-aminobenzoic acid and two of its derivatives. *Pharm. Res.* **2006**, *23* (5), 951-60.
56. Opalka, L.; Kovacik, A.; Maixner, J.; Vavrova, K., Omega-O-Acylceramides in skin lipid membranes: effects of concentration, sphingoid base, and model complexity on microstructure and permeability. *Langmuir* **2016**, *32* (48), 12894-12904.
57. Uchiyama, M.; Oguri, M.; Mojumdar, E. H.; Gooris, G. S.; Bouwstra, J. A., Free fatty acids chain length distribution affects the permeability of skin lipid model membranes. *Biochim. Biophys. Acta* **2016**, *1858* (9), 2050-9.
58. Skolova, B.; Jandovska, K.; Pullmannova, P.; Tesar, O.; Roh, J.; Hrabalek, A.; Vavrova, K., The role of the trans double bond in skin barrier sphingolipids: permeability and infrared spectroscopic study of model ceramide and dihydroceramide membranes. *Langmuir* **2014**, *30* (19), 5527-35.
59. Pilgram, G. S. K.; Vissers, D. C. J.; van der Meulen, H.; Koerten, H. K.; Pavel, S.; Lavrijsen, S. P. M.; Bouwstra, J. A., Aberrant lipid organization in stratum corneum of patients with atopic dermatitis and lamellar ichthyosis. *J. Invest. Dermatol.* **2001**, *117* (3), 710-717.
60. Paz Ramos, A.; Lafleur, M., Chain length of free fatty acids influence the phase behavior of stratum corneum model membranes. *Langmuir* **2015**, *31* (42), 11621-11629.
61. Skolova, B.; Hudska, K.; Pullmannova, P.; Kovacik, A.; Palat, K.; Roh, J.; Fleddermann, J.; Estrela-Lopis, I.; Vavrova, K., Different phase behavior and packing of ceramides with long (C16) and very long (C24) acyls in model membranes: infrared spectroscopy using deuterated lipids. *J Phys Chem-US* **2014**, *118* (35), 10460-70.
62. de Jager, M. W.; Gooris, G. S.; Ponec, M.; Bouwstra, J. A., Lipid mixtures prepared with well-defined synthetic ceramides closely mimic the unique stratum corneum lipid phase behavior. *J. Lipid Res.* **2005**, *46* (12), 2649-2656.
63. Chen, X.; Kwak, S.; Lafleur, M.; Bloom, M.; Kitson, N.; Thewalt, J., Fatty acids influence "solid" phase formation in models of stratum corneum intercellular membranes. *Langmuir* **2007**, *23* (10), 5548-56.

