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Rotational resonance NMR of ¹³C₂-labelled retinal: quantitative internuclear distance determination

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Abstract

Rotational resonance phenomena are investigated in the solid-state magic-angle spinning NMR of all-E-[11,20-¹³C₂]-retinal at a magnetic field of 4.7 T. We find good agreement between experiments and numerical simulations for the rotational resonance spectral peakshapes and for the rotor-driven magnetization exchange. The internuclear distance between the ¹³C-labelled C11 and C20 sites is determined to be 0.301 ± 0.008 nm (from rotational resonance spectra) and 0.300 ± 0.010 nm (from rotor-driven magnetization exchange), in agreement with the X-ray crystallographic distance of 0.296 nm. We show rotational resonance spectra which display perturbations from intermolecular homonuclear spin–spin interactions. © 1999 Elsevier Science B.V. All rights reserved.

Keywords: Rotational resonance NMR; Internuclear distance; ¹³C₂-labelled retinal

1. Introduction

Solid-state NMR may be used to determine molecular structure in systems which are difficult or

impossible to crystallize, such as many polymers, biological molecules, and surface species. Molecular structural information is obtained from the magnitudes of spin-spin interactions, such as through-space internuclear dipolar couplings [1–4], or chemical shift tensors [5–7]. It is also possible to extract molecular structural information from the relative or absolute orientations of spin interaction tensors [8–10].

The magnitude of the through-space dipole—dipole couplings between nuclear spins is a direct source of molecular structural information, because it only depends on fundamental constants and the inverse third power of the internuclear distance. However, these couplings are quite difficult to access in the context of high-resolution solid-state NMR. This

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is because magic-angle spinning (MAS) is normally used to enhance the sensitivity and the spectral resolution, by suppressing the effect of the chemical shift anisotropies. Unfortunately, MAS also tends to average out the structurally informative through-space dipole—dipole couplings.

A variety of methods have been proposed for retrieving the dipolar coupling information in the presence of magic-angle spinning. One of the simplest is rotational resonance (RR) [1.3.11.12]. Consider a sample containing dilute homonuclear spin pairs, for example, ¹³C pairs deliberately introduced by isotopic labelling. Rotational resonance occurs when the sample rotation frequency ω_r matches the condition $\omega_{\Lambda}^{\text{iso}} = n \omega_{\text{r}}$, where $\omega_{\Lambda}^{\text{iso}}$ is the difference in isotropic chemical shift frequencies of the paired spins, and n is a small integer denoting the order of the rotational resonance. There are two main consequences of the reintroduction of the through-space dipolar coupling. First, the normally narrow spectral peaks acquire splittings and broadenings; Second, there is rotor-driven exchange of longitudinal magnetization between the spin sites. These two effects provide alternative routes for the estimation of the dipolar spin-spin couplings, and hence internuclear distances [1,3].

So far, the exchange of longitudinal magnetization has been most widely used for biomolecular structural investigations [13–24]. This requires a two-dimensional experimental procedure, since longitudinal magnetization cannot be observed directly. The direct observation of spectral peakshapes at rotational resonance is potentially much faster [25], but has been relatively less explored for structural studies. Most quantitative treatments of rotational resonance spectra have been on compounds with short ¹³C-¹³C distances [1,3], although McDermott et al. reported the determination of an internuclear distance of about 0.27 nm between two ³¹P spins [13].

Although most of the reported spectral peakshapes conform to theoretical expectations, there are also several reports which are in conflict with current theory. For example, Peersen et al. showed rotational resonance spectral peakshapes for pairs of ¹³C nuclei separated by 0.37 nm in a small peptide, which do not conform to the present rotational resonance theory [22]. Rather similar observations were made by some of us on a set of ¹³C₂-labelled retinals at a magnetic field of 9.4 T [25]. In the latter work, the observed peaks typically had a very different form for the two sites, with one peak split and the other unsplit, whereas available theory always predicts a more-or-less symmetrical splitting at rotational resonance. Despite these striking discrepancies, it still proved possible to obtain quantitative distance estimations from the split peak by a semi-empirical calibration procedure [25]. In a forthcoming article [26] we consider the problem of the rotational resonance peakshapes of all-E-[11.20-¹³C₂]-retinal at the higher magnetic field of 9.4 T. We show that the anomalous peakshapes at high field may be explained in terms of the differential transverse relaxation of spins in the two sites. The transverse relaxation differential is due to imperfect heteronuclear decoupling, caused by an interference of the magicangle sample rotation and the proton decoupler field. A revised theory of rotational resonance peakshapes. capable of accommodating such effects, is presented [26].

The same labelled compounds which give discrepancies between theory and experiments at a field of 9.4 T provide "normal" rotational resonance spectra at a lower magnetic field of 4.7 T. In this article, we present a study of the rotational resonance spectrum of one of these compounds, polycrystalline all-*E*-[11,20-¹³C₂]-retinal, at the lower field of 4.7 T. As shown below, the rotational resonance peakshapes and magnetization exchange curves may be simulated quite well at this lower field, using the "conventional" theory, as presented in Ref. [3].

2. Materials

This study employed three different samples of polycrystalline all-*E*-retinal (Fig. 1).

- (1) Natural abundance all-*E*-retinal, purchased commercially (Fluka, Switzerland) and used without further purification. We refer to this sample as nat-RET.
- (2) All-*E*-retinal labelled to a level of larger than 99% with 13 C at both positions C11 and C20. This was synthesized according to published procedures [27] and crystallized at -20° C from *n*-pentane. We refer to this sample as 99%- 13 C₂-RET.

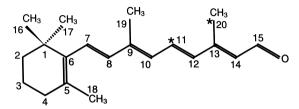


Fig. 1. Numbering scheme of the all-*E*-retinal molecule. Asterisks denote the ¹³C labelled sites

(3) A solid solution of $99\%^{-13}C_2$ -RET in nat-RET, at a molar ratio of 15:100. This sample was prepared by dissolving the appropriate amounts of $99\%^{-13}C_2$ -RET and nat-RET in *n*-pentane and crystallizing. We refer to this sample as $15\%^{-13}C_2$ -RET.

Neighboring all-*E*-retinal molecules in the crystalline solid have an antiparallel orientation, as determined by X-ray diffraction [28]. The internuclear distance between the C11 and C20 sites within the same molecule is 0.296 ± 0.001 nm. The shortest distances between 13 C labels on neighboring molecules are as follows: $r_{11,11'} = 0.384$ nm; $r_{11,20'} = 0.458$ nm; $r_{20,20'} = 0.573$ nm. In most of the spectra discussed below, we employed the 15%- 13 C₂-RET sample in order to reduce the probability of the intermolecular spin–spin interactions.

3. Methods

3.1. MAS spectra

¹³C MAS spectra were recorded at a field strength $B_0 = 4.7$ T (¹³C Larmor frequency = -50.323 MHz) using the standard CP-MAS pulse scheme shown in Fig. 2a. The ¹³C transverse magnetization was enhanced by ramped cross-polarization (CP) from protons [29–31]. The proton $\pi/2$ pulse duration was 4.7 μs. The proton decoupling field strength during data acquisition corresponded to a nutation frequency of $|\omega_{\text{nut}}^{\text{I}}/2\pi| \approx 96$ kHz.

The rotational resonance spectra were taken for a variety of orders n, chosen by adjusting the spinning frequency to the appropriate condition. The relevant spinning frequencies in the field of $B_0=4.7$ T were $|\omega_{\rm r}/2\pi|=6.098$ kHz for n=1 RR, $|\omega_{\rm r}/2\pi|=3.049$ kHz for n=2 RR, and $|\omega_{\rm r}/2\pi|=2.033$ kHz for n=3 RR.

3.2. Zeeman magnetization exchange

The Zeeman magnetization exchange between the labelled ¹³C sites was followed using the pulse scheme shown in Fig. 2b. The first part of this pulse scheme (up to and including the first $\pi/2^{-13}$ C pulse) creates a spin density operator ρ containing a term proportional to the difference angular momentum operator $S_{iz} - S_{kz}$, where j and k are the two labelled spin sites. This is done by allowing the crosspolarized ¹³C magnetization to evolve under the isotropic shift difference of the two sites. Four π pulses are applied at carefully timed intervals so as to suppress the effect of chemical shift anisotropy, as described in Ref. [32]. A $\pi/2$ pulse of suitable phase converts the out-of-phase transverse magnetization into differential longitudinal magnetization. The longitudinal difference magnetization created by the first $\pi/2$ pulse is allowed to evolve under the interval τ . Another strong $\pi/2$ pulse reconverts the Zeeman magnetization into transverse magnetization, which generates an NMR signal in the following interval. A standard phase cycling scheme is used to select signal components passing through longitudi-

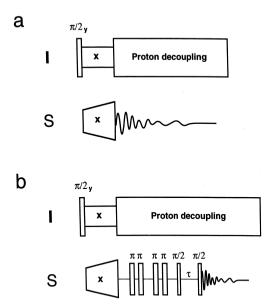


Fig. 2. The rf pulse scheme for rotational resonance spectral peakshape measurements (a), and for the Zeeman magnetization exchange measurements (b). The ramp of the cross polarization field and switching of the decoupler irradiation amplitude are optional.

nal magnetization during the τ interval. After Fourier transformation, the integrated intensities of the two peaks are taken to be proportional to the Zeeman magnetizations of the appropriate sites at the end of the τ interval. The rotor-driven coherence transfer signals during the acquisition period are known not to affect peak integrals [26]. The experiment is repeated for a set of τ values, typically ranging from 0 to 50 ms. The difference between integrated peak amplitudes as a function of τ corresponds to the time course of difference Zeeman magnetization $\langle S_{1z} - S_{2z} \rangle (\tau)$. The data set is usually normalized so that the first point has unit amplitude.

The experimental Zeeman magnetization exchange data were collected with ^{1}H and ^{13}C $\pi/2$ pulse durations of 2.3 and 2.4 μs , respectively. In the results shown, the same rf field was used throughout the proton channel, corresponding to a nutation frequency of $|\omega_{\rm nut}^{\rm I}/2\pi| \approx 108$ kHz.

All experimental data were obtained on a modified Bruker MSL 200 spectrometer at a magnetic field of 4.7 T. A standard Bruker double bearing 4 mm rotor system ($\rm ZrO_2$ rotors, Kel-F end caps) was used. The sample spinning frequency was stabilized to ± 2 Hz using a home-built stabilization device. The CP contact interval was 2 ms.

3.3. Simulations

Numerical simulations of rotational resonance spectra and Zeeman magnetization exchange were performed using previously described procedures [3]. The calculations require the following parameters: (i) The principal values of the chemical shift tensors of the two sites; (ii) the isotropic *J*-coupling between the two spins; (iii) the through-space dipolar coupling between the spins; (iv) the relative orientations of the chemical shift tensors with respect to the internuclear vector; (v) relaxation parameters (see below).

The anisotropies of the chemical shift tensors have been derived from an analysis of the slow-spinning MAS spectrum [25]. The values used in the simulations are shown in Table 1. The chemical shift anisotropy (in deshielding units) is defined $\delta_{\rm aniso}^j = \delta_{zz}^j - \delta_{\rm iso}^j$, where $\delta_{\rm iso}^j$ is the isotropic chemical shift and the principal values are ordered as $|\delta_{zz}^j - \delta_{\rm iso}^j| > |\delta_{xx}^j - \delta_{\rm iso}^j| > |\delta_{yy}^j - \delta_{\rm iso}^j|$ [35]. The asymmetry pa-

Table 1
Parameters for the rotational resonance simulations of all-E[11,20- 13 C₂]-retinal, at a magnetic field of $B_0 = 4.7$ T

Parameters	all-E-[11,20- ¹³ C ₂]-	Notes
	retinal	
$(\delta_{\rm iso}^j - \delta_{\rm iso}^k)$ [ppm]	-121.2	a
$\delta_{\rm aniso}^{k}$ [ppm]	112.2	b
η^k	0.72	b
$\{\alpha_{\mathrm{PM}}^{k},eta_{\mathrm{PM}}^{k},\gamma_{\mathrm{PM}}^{k}\}$	$\{-31, 85, -87\}$	c
$\delta_{\rm aniso}^{j}$ [ppm]	15.5	b
$oldsymbol{\eta}^j$	1.0	b
$\{lpha_{ ext{PM}}^{j},eta_{ ext{PM}}^{j},\gamma_{ ext{PM}}^{j}\}$	{0, 57, 0}	c
$(b/2\pi)$ [Hz]	-293	d
J [Hz]	0	e
T_2^{SQ} [ms]	15.9	f
$T_2^{\rm ZQ}$ [ms]	11.0	g
Estimated internuclear		
distance (nm)		
from RR spectra	$0.301 \pm 0.008 \text{ nm}$	h
from RR magnetization exchange	$0.300 \pm 0.010 \text{ nm}$	i
from X-ray diffraction	$0.296 \pm 0.001 \text{ nm}$	

- (a) Difference in isotropic chemical shifts (deshielding units) between the C20 site (index j) and C11 site (index k).
- (b) Chemical shift anisotropy parameters (deshielding units) for the C11 and C20 sites, as reported in Ref. [25].
- (c) Euler angles in degrees, defining the assumed chemical shift tensor orientations with respect to the molecular reference frame (chosen such that the *z*-axis is along the vector joining the C11 and C20 nuclei and the *x*-axis is in the (C11, C12, C20) plane.
- (d) Dipole–dipole coupling constant from the X-ray structure $r=0.296~\mathrm{nm}.$
- (e) The three-bond $^{13}C-^{13}C$ J coupling is ignored.
- (f) Single-quantum relaxation time constant estimated from the mean of the off-rotational resonance peak widths, at a proton decoupling field corresponding to $|\omega_{\rm nut}^I/2\pi| \approx 96$ kHz.
- (g) Zero-quantum relaxation time constant estimated from the single-quantum spectral peak widths, at a proton decoupling field corresponding to $|\omega_{\text{nut}}^{\text{I}}/2\pi| \approx 108 \text{ kHz}.$
- (h) Estimated internuclear distance from rotational resonance spectra of the 15% 13 C₂-labelled sample.
- (i) Estimated internuclear distance from rotor-driven Zeeman magnetization exchange on the 99% ¹³C₂-labelled sample.

rameter η^j of the chemical shift tensor is defined through $\eta^j = (\delta_{yy}^j - \delta_{xx}^j)/\delta_{\rm aniso}^j$. The superscripts j and k denote the C20 and C11 sites, respectively. The isotropic chemical shift frequency difference is given by $\omega_{\Delta}^{\rm iso} = \omega_{\rm iso}^j - \omega_{\rm iso}^k = \omega_0(\delta_{\rm iso}^j - \delta_{\rm iso}^k)$, where ω_0 is the signed Larmor frequency $\omega_0 = -\gamma B_0$ [36].

The Euler angles $\{\alpha_{PM}^j, \beta_{PM}^j, \gamma_{PM}^j\}$ define the relative orientation of the chemical shift tensor principal axis system P of site j and a molecular axis system M whose z-axis is defined as the internuclear

C11-C20 vector. The Euler angles are defined as in Ref. [37]. The Euler angles used in the simulation were calculated from the crystal structure geometry combined with plausible assumptions about the orientations of the chemical shift tensors with respect to the local functional groups. The principal axis system of the C11 site is defined such that the most shielded element is perpendicular to the (C10, C11, C12) plane, while the intermediate shielded element is along the C11-C12 bond. The orientation of the principal axis system of C20 site is defined so that the most shielded element is along the C13-C20 bond and the least shielded one is in the (C12, C13, C20) plane. These orientations conform to the consensus of single crystal studies in related compounds [38]. It should be emphasized that, at least for the n = 1 RR, the simulated behavior is insensitive to these chemical shift tensor orientations [25].

The three-bond *J*-coupling between the C11 and C20 sites was assumed to be negligible.

The rotational resonance peakshapes were simulated by evaluating the dynamics of the isolated spin-pair system, ignoring the transverse relaxation of the magnetization components. The simulated free induction decay was multiplied by an exponential damping function to take into account transverse relaxation effects [3]. This procedure is justifiable if the transverse relaxation time constants are the same for both sites [26]. The observed peak widths (full widths at half height) were ~ 20 Hz for both the C11 site and the C20 sites at a spinning frequency of $|\omega_r/2\pi| = 10.5$ kHz and a decoupler field strength corresponding to the proton nutation frequency $|\omega_{\text{nut}}|/2\pi| \approx 96$ kHz.

The magnetization exchange simulations require estimations of the zero-quantum relaxation time constant $T_2^{\rm ZQ}$. This parameter was estimated by making a further assumption that the transverse relaxation is caused by uncorrelated fluctuating random fields at the two sites. The zero-quantum relaxation rate constant is then equal to the sum of the single-quantum relaxation rate constants of the two sites [3]. In the present case, the peak widths (full-width-at-half-maximum) of 14 Hz for the C11 site and 15 Hz for the C20 site at $|\omega_{\rm r}/2\pi|=10$ kHz and $|\omega_{\rm nut}^{\rm I}/2\pi|\approx 108$ kHz allowed the estimate $T_2^{\rm ZQ}=(\pi(14+15))^{-1}=11.0$ ms. The simulations are insensitive to this parameter in the regime discussed here.

4. Results and discussion

4.1. Rotational resonance peakshapes

Expanded regions of the MAS spectrum of nat-RET at a spinning frequency $|\omega_r/2\pi| = 6.098$ kHz are shown in Fig. 3a. Most of the peaks may be assigned to the ¹³C sites according to the solution-state chemical shifts reported in Ref. [33], and the solid-state data reported in Ref. [34].

The data in Refs. [33,34] indicate nearly coincident ¹³C chemical shifts for the C19 and C20 sites, and for the C11, C12, and C6 sites. The measured shift frequency difference between the C11 and C20

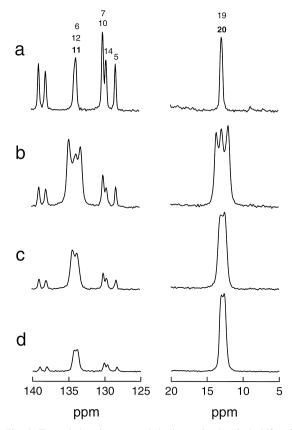


Fig. 3. Expanded regions around the isotropic chemical shifts of the C11 (left) and C20 (right) sites of the MAS spectra of all-*E*-retinal. (a) Natural abundance retinal, at a spinning frequency of $|\omega_r/2\pi| = 6.098$ kHz. (b) $15\%-^{13}\mathrm{C}_2$ -RET at the n=1 RR condition ($|\omega_r/2\pi| = 6.098$ kHz); (c) $15\%-^{13}\mathrm{C}_2$ -RET at the n=2 RR condition ($|\omega_r/2\pi| = 3.049$ kHz); (d) $15\%-^{13}\mathrm{C}_2$ -RET at the n=3 RR condition ($|\omega_r/2\pi| = 2.033$ kHz).

peaks in nat-RET is $\omega_{\Delta}^{\rm iso}/2\pi = 6.088$ kHz at a field of 4.7 T.

The MAS spectrum of the 15%- 13 C₂-RET sample, at the n=1 rotational resonance ($|\omega_r/2\pi|=6.098$ kHz) is shown in Fig. 3b. The prominent spectral features come from the 13 C labels at the C11 and C20 sites, broadened by the dipole–dipole coupling at the n=1 rotational resonance condition. Spectra at the n=2 rotational resonance ($|\omega_r/2\pi|=3.049$ kHz) and the n=3 rotational resonance ($|\omega_r/2\pi|=2.033$ kHz) are shown in Fig. 3c and d.

The NMR signals from the 15%-13C₂-RET sample come from three major classes of molecules: (i) molecules which have introduced ¹³C labels at both sites C11 and C20, and no naturally occurring ¹³C spins; (ii) molecules with no introduced ¹³C labels but a singly naturally occurring ¹³C spin; (iii) molecules containing a natural ¹³C spin as well as two artificial ¹³C labels. As discussed in Ref. [26]. the signal contributions from class (ii) and (iii) molecules may be eliminated, to a good approximation, by subtracting a suitably weighted spectrum of nat-RET, obtained under identical experimental conditions. Such "difference spectra" at the n = 1, 2, 3RR conditions are shown in Fig. 4. Simulations using the parameters in Table 1 are also shown. The measured and simulated rotational resonance peakshapes are in very good agreement. There are minor deviations at n = 2 and n = 3 RR (the experimental inner peak amplitudes being smaller than the simulated amplitudes). This may reflect a small change ($\sim 10 \text{ Hz}$) of the isotropic ¹³C chemical shift in the ¹³C₂-labelled molecules, due to the influence of the second ¹³C on the molecular vibrations and librations. Such effects are frequently observed in solution NMR [39,40].

To reveal the resolution of the determined distance by the rotational resonance peakshapes, a series of peakshape simulations at n=1 RR varying only the dipole–dipole coupling constant was carried out. Some results are shown in Fig. 5. The best overall match is for a dipole–dipole coupling constant $b/2\pi = -280$ Hz, corresponding to an internuclear distance r=0.301 nm. A 20 Hz deviation of the dipole–dipole coupling constant may easily be resolved. This translates into a rough confidence limit in the internuclear distance of ± 0.008 nm. The X-ray estimation of r=0.296 nm is within this

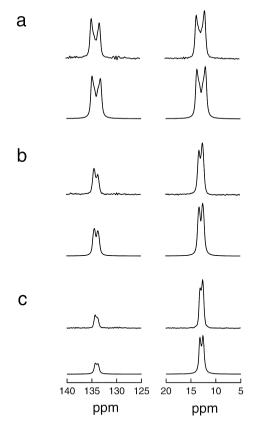


Fig. 4. Expanded regions around the isotropic chemical shifts of the C11 (left) and C20 (right) sites at n=1, 2, and 3 RR conditions ((a), (b), and (c) respectively). At each RR condition, the upper figures represent experimental rotational resonance spectra of 15%- 13 C₂-RET (with natural abundance spectra subtracted, see text); the lower figures are simulations using the parameters listed in Table 1.

confidence limit. A more rigorous estimation of the confidence limits could be obtained by statistical χ^2 analysis [41].

To illustrate the effect of the intermolecular interactions on the rotational resonance peakshapes, spectra were also recorded on the 99%- 13 C₂-RET sample at n = 1, 2, and 3 rotational resonances. Expanded regions around the isotropic chemical shifts of the C11 and C20 sites are displayed in Fig. 6.

At n=1 RR, the multiplets have a similar appearance as for the 15% doubly labelled sample. In particular, the C20 peak displays a prominent central feature in both the 15%- 13 C₂-RET and 99%- 13 C₂-RET sample. However, the origin of the central

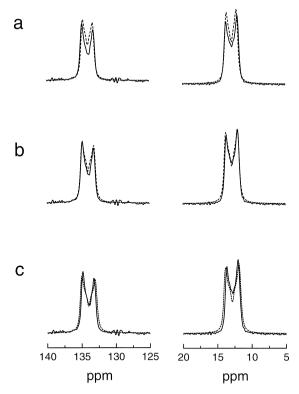


Fig. 5. Expanded regions around the isotropic chemical shifts of the C11 site (left) and the C20 site (right) of the n=1 rotational resonance difference spectra, compared with simulated spectra for the following values of dipole–dipole coupling constant $b/2\pi$: (a) -253 Hz, (b) -293 Hz, (c) -333 Hz. Solid lines represent the experimental spectra; dashed lines represent simulations.

feature in the C20 peak must be different in these two materials. In 15%-¹³C₂-RET, this peak could be attributed to isotopomers containing isolated ¹³C spins. Subtraction of the natural abundance spectrum essentially eliminates this peak. In the 99%-¹³C₂-RET sample, on the other hand, the singly ¹³C-labelled isotopomers are quite rare, arising mainly from incomplete labelling of the carbon sites in the starting materials. These isotopomers are too rare to account for the central feature in the C20 peak. Signals from ¹³C₂ isotopomers also contribute, but the relevant signals are severely broadened by rotational resonance effects. For example, consider the 11,19,20-¹³C₂ isotopomer. Since the isotropic chemical shifts of the C19 and C20 sites are almost coincident, and the C11...C20 and C11...C19 distances are comparable, the three-spin system in 11,19,20-¹³C₃-retinal is at "double rotational resonance" and gives broad signals for all sites. These broad signals cannot be responsible for the sharp central feature in the C20 peak.

Intermolecular spin–spin interactions provide an alternative explanation of the central feature in the C20 peak of the 99%-¹³C₂ labelled material. As remarked above, C11 sites on neighbouring molecules are rather close (0.384 nm). The strong intermolecular interactions of the C11 sites lead to enhanced transverse relaxation for spins in those sites. This enhanced transverse relaxation has an indirect decoupling effect on the C20 resonances, as described in Ref. [26]. Loosely speaking, the enhanced relaxation of the C11 coherences takes those spins out of rotational resonance, and narrows the C20 signals.

The C11 multiplet in the 99%- 13 C₂-RET spectrum at n = 1 RR also displays a sharp central feature. This is probably also due, at least in part, to inter-

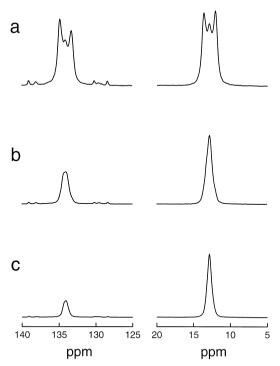


Fig. 6. Expanded regions around the isotropic chemical shifts of the C11 (left) and C20 sites of the rotational resonance spectra for the 99%- 13 C₂-RET sample at (a) n=1, (b) n=2, and (c) n=3 RR conditions.

molecular spin–spin interactions. Signals from the $11,12,20^{-13}C_3$ isotopomer may also contribute.

Intermolecular interaction effects on the rotational resonance peakshapes may also be noticed clearly at the n=2 and 3 rotational resonance. The small splittings observed in the peaks of $15\%^{-13}C_2$ -RET vanish for the case of $99\%^{-13}C_2$ -RET sample. Notice that the C20 peaks in $99\%^{-13}C_2$ -RET are narrower than in $15\%^{-13}C_2$ -RET. This is consistent with the "self-decoupling" mechanism indicated above, which is discussed in detail in Ref. [26].

4.2. Zeeman magnetization exchange

Experimental Zeeman magnetization exchange curves for $99\%^{-13}C_2$ -RET, at n = 1, 2, and 3 RR, as well as simulated curves, are shown in Fig. 7. The dipole–dipole coupling constant used in the simula-

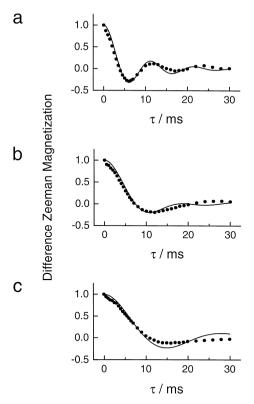


Fig. 7. Experimental and simulated Zeeman magnetization exchange curves for 99%- 13 C₂-RET sample at (a) n=1, (b) n=2, and (c) n=3 RR. Filled circles represent experimental data, solid lines indicate simulations.

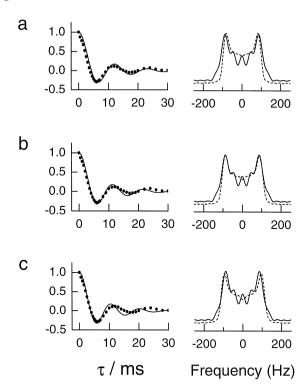


Fig. 8. Time-domain (left) and frequency-domain (right) experimental and simulated Zeeman magnetization exchange curves at the n=1 rotational resonance condition. Simulated curves are shown for the following dipole–dipole coupling constants: (a) $b/2\pi = -273$ Hz, (b) $b/2\pi = -293$ Hz, and (c) $b/2\pi = -313$ Hz. In the time-domain figures, filled circles represent experimental data, solid lines indicate simulations. In the frequency-domain figures, solid lines represent the experimental data and dashed lines indicate simulations.

tions is -293 Hz, corresponding to the X-ray internuclear distance of 0.296 nm. Other parameters used are listed in Table 1. The agreement between simulated and measured curves is acceptable, at least with respect to the initial decay. The discrepancies in the form of the curve may reflect the influence of intermolecular 13 C $^{-13}$ C interactions. Note that the n=1 curve is insensitive to the tensor orientations, so this cannot be held responsible for the deviations.

To reveal the resolution of the determined distance, a series of simulated curves at n = 1 for a selection of dipole-dipole coupling constants is shown in Fig. 8. The comparison was made using both the direct time-domain magnetization exchange data (left), as well as Fourier transforms of the

magnetization exchange curves (right) [42]. The time-domain curves appear to give the subjectively best fit for a through-space coupling of $b/2\pi =$ -273 Hz (Fig. 8a, left plot). The frequency-domain curves, on the other hand, appear to give the subjectively best fit for a through-space coupling of $b/2\pi$ = -293 Hz, if one concentrates on the outer peak splitting, and ignores the discrepancies in the central part (Fig. 8b. right plot). The dipole-dipole coupling may therefore be estimated from the magnetization exchange curves to be $b/2\pi = -283 \pm 30$ Hz. This translates into an internuclear distance estimate of 0.300 + 0.010 nm, which is consistent with the X-ray diffraction value of 0.296 + 0.001 nm. A statistical v^2 analysis [41] would give a more rigorous estimate of the confidence limits.

It is difficult to compare the accuracy of the two methods directly (rotational resonance spectra against rotational resonance magnetization exchange), since the data in Figs. 5 and 8 were obtained on samples with different levels of isotope labelling. Magnetization exchange experiments on the 15%- 13 C₂-RET sample would clarify this issue.

5. Conclusions

The internuclear distance between the 13 C-labelled sites in all-E-[11,20- 13 C₂]-retinal was estimated using rotational resonance spectra and rotor-driven longitudinal magnetization exchange, in a magnetic field of $B_0 = 4.7$ T. The estimated distances of 0.301 ± 0.008 nm (for rotational resonance spectra of a 15% 13 C₂-labelled sample) and 0.300 ± 0.010 nm (for Zeeman magnetization exchange on a 99% 13 C₂-labelled sample) both agree well with the X-ray crystal structure.

Significant deviations between theoretical and experimental rotational resonance spectra were obtained when the 13 C₂-labelled molecules were not diluted in an unlabelled matrix. These deviations were attributed to intermolecular 13 C $^{-13}$ C interactions.

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References

- [1] D.P. Raleigh, M.H. Levitt, R.G. Griffin, Chem. Phys. Lett. 146 (1988) 71.
- [2] T. Gullion, J. Schaefer, J. Magn. Reson. 81 (1989) 479.
- [3] M.H. Levitt, D.P. Raleigh, F. Creuzet, R.G. Griffin, J. Chem. Phys. 92 (1990) 6347.
- [4] D.M. Gregory, D.J. Mitchell, J.A. Stringer, S. Kiihne, J.C. Shiels, J. Callahan, M.A. Mehta, G.P. Drobny, Chem. Phys. Lett. 246 (1995) 654.
- [5] A.C. deDios, E. Oldfield, Solid State Nucl. Magn. Reson. 6 (1996) 101.
- [6] M. Han, S.O. Smith, Biochemistry 34 (1995) 1425.
- [7] Y. Tomita, E.J. O'Connor, A. McDermott, J. Am. Chem. Soc. 116 (1994) 8766.
- [8] D.P. Weliky, R. Tycko, J. Am. Chem. Soc. 118 (1996) 8487.
- [9] K. Schmidt-Rohr, J. Am. Chem. Soc. 118 (1996) 7601.
- [10] X. Feng, Y.K. Lee, D. Sandström, M. Edén, H. Maisel, A. Sebald, M.H. Levitt, Chem. Phys. Lett. 257 (1996) 314.
- [11] E.R. Andrew, A. Bradbury, R.G. Eades, V.T. Wynn, Phys. Lett. 4 (1963) 99.
- [12] J. Heller, R. Larsen, M. Ernst, A.C. Kolbert, M. Baldwin, S.B. Prusiner, D.E. Wemmer, A. Pines, Chem. Phys. Lett. 251 (1996) 223.
- [13] A. McDermott, F. Creuzet, R.G. Griffin, Biochemistry 29 (1990) 5767.
- [14] F. Creuzet, A. McDermott, R. Gebhard, K. van der Hoff, M.B. Spijker-Assink, J. Herzfeld, M.H. Levitt, R.G. Griffin, Science 251 (1991) 783.
- [15] O.B. Peersen, S. Yoshimura, H. Hojo, S. Aimoto, S.O. Smith, J. Am. Chem. Soc. 114 (1992) 4332.
- [16] O.B. Peersen, S.O. Smith, Concepts Magn. Reson. 5 (1993) 303
- [17] K.V. Lakshmi, M. Auger, J. Raap, J. Lugtenburg, R.G. Griffin, J. Herzfeld, J. Am. Chem. Soc. 115 (1993) 8515.
- [18] A. McDermott, F. Creuzet, R. Gebhard, K. van der Hoef, M.H. Levitt, J. Herzfeld, J. Lugtenburg, R.G. Griffin, Biochemistry 33 (1994) 6129.
- [19] S.O. Smith, J. Hamilton, A. Salmon, B.J. Bormann, Biochemistry 33 (1994) 6327.
- [20] S.O. Smith, R. Jonas, M. Braiman, B.J. Bormann, Biochemistry 33 (1994) 6334.
- [21] J.M. Griffiths, T.T. Ashburn, M. Auger, P.R. Costa, R.G. Griffin, P.T. Lansbury, J. Am. Chem. Soc. 117 (1995) 3539.
- [22] O.B. Peersen, M. Groesbeek, S. Aimoto, S.O. Smith, J. Am. Chem. Soc. 117 (1995) 7228.
- [23] S.O. Smith, B.J. Bormann, Proc. Natl. Acad. Sci. U.S.A. 92 (1995) 488.

- [24] P.T. Lansbury, P.R. Costa, J.M. Griffiths, E.J. Simon, M. Auger, K.J. Halverson, D.A. Kocisko, Z.S. Hendsch, T.T. Ashburn, R.G.S. Spencer, B. Tidor, R.G. Griffin, Nat. Struct. Biol. 2 (1995) 990.
- [25] P.J.E. Verdegem, M. Helmle, J. Lugtenburg, H.J.M. de Groot, J. Am. Chem. Soc. 119 (1997) 169.
- [26] M. Helmle, Y.K. Lee, P.J.E. Verdegem, T. Karlsson, H.J.M. de Groot, J. Lugtenburg, M.H. Levitt, submitted for publication
- [27] M. Groesbeek, J. Lugtenburg, J. Photochem. Photobiol. 56 (1992) 903.
- [28] T. Hamanaka, T. Mitsui, Acta Crystallogr., Sect. B 28 (1972) 214
- [29] S. Zhang, C.L. Czekaj, W.T. Ford, J. Magn. Reson. A 111 (1994) 87
- [30] G. Metz, X. Wu, S.O. Smith, J. Magn. Reson. A 110 (1994)
- [31] S. Hediger, B.H. Meier, N.D. Kurur, G. Bodenhausen, R.R. Ernst, Chem. Phys. Lett. 223 (1994) 283.
- [32] H. Geen, M.H. Levitt, G. Bodenhausen, Chem. Phys. Lett. 200 (1992) 350.

- [33] G. Englert, Helv. Chim. Acta 58 (1975) 2367.
- [34] G.S. Harbison, P.P.J. Mulder, H. Pardoen, J. Lugtenburg, J. Herzfeld, R.G. Griffin, J. Am. Chem. Soc. 107 (1985) 4809.
- [35] D.M. Grant, in: Encyclopedia of NMR, Vol. 2, Wiley, Chichester, England, 1996.
- [36] M.H. Levitt, J. Magn. Reson. 126 (1997) 164.
- [37] D.A. Varshalovich, A.N. Moskalev, V.K. Khersonskii, Quantum Theory of Angular Momentum, World Scientific, Singapore, 1988.
- [38] M. Mehring, Principles of High Resolution NMR in Solids, 2nd edn., Springer, Berlin, 1983.
- [39] H. Batiz-Hernandez, R.A. Bernheim, Prog. NMR Spectrosc. 3 (1967) 63.
- [40] C.J. Jameson, in: Encyclopedia of NMR, Vol. 4, Wiley, Chichester, England, 1996.
- [41] W.H. Press, B.P. Flannery, S.A. Teukolsky, W.T. Vetterling, Numerical Recipes. The Art of Scientific Computing. Cambridge Univ. Press, Cambridge, 1986.
- [42] N.C. Nielsen, F. Creuzet, R.G. Griffin, J. Magn. Reson. A 103 (1993) 245.