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13C NMR Study of the Grafting of Maleic Anhydride onto Polyethylene, Polypropene, and Ethene–Propene Copolymers


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ABSTRACT: The reaction products from the radically initiated grafting of specifically 13C-enriched maleic anhydride ([2,3-13C2]MA) onto polyethylene, isotactic polypropene and ethene-propene copolymers in the melt and in solution are investigated using noise-decoupled and 1D inadequate 13C NMR spectroscopy. The sites of attachment and the structures of the grafts depend on (co)polymer composition. In random EPM, MA attaches to methylene and methine carbons in the backbone. In alternating EPM, MA attaches solely to polymer methines, indicating that (CH2)m sequences with m > 3 are needed for MA attachment to backbone methylene carbons. In the copolymers and in iPP, grafts are single succinic anhydride rings; in HDPE and LDPE short MA oligomers are also present. In polyolefins containing polypropene sequences, chain scission can yield structures in which the anhydride ring is attached to the chain terminus via a fully substituted double bond.

Introduction

Polyolefins functionalized with maleic anhydride (MA) exhibit enhanced adhesion to polar materials like polyamide, metals, and glass and are used as compatibilizing agents. For instance, the improvement of impact properties upon blending of polyamide-6 with various maleated polyolefins is associated with the formation of interfacial adhesion and the enhancement of mechanical properties. It is generally accepted that cross-linking in PE and PP may occur simultaneously with the graft reaction. EPMs may undergo both side reactions. Besides the exact site of attachment, an additional matter that has been speculated upon is the structure of the MA graft. A single, saturated succinic anhydride graft is usually suggested, but unsaturated, oligomeric, polymeric grafts, and a combination of single and oligomeric grafts have also been proposed.

The aims of this investigation are to elucidate the molecular structure of MA grafted PE, iPP, and EPMs and to characterize the mechanism of the peroxide initiated grafting. Genuine, essentially unperturbed materials are investigated using site specific isotope labeling and NMR spectroscopy. First, EPMs with varying propene contents, PE, and PP have been used as substrates in melt-grafting reactions to study the effect of (co)polymer composition on the chemistry of grafting. Subsequently, a selection of these materials was reacted with MA in a solution process to compare the two methods of functionalization.

Experimental Section

[2,3-13C2]MA was synthesized starting from [2,3-13C]acetic acid (Cambridge Isotopes Laboratories, 99% enriched). This was first converted to ethyl bromo[2-13C]acetate via the Hell–Volhard–Zelinsky reaction (Carboethoxy[13C]methyltriphenyolphosphonium bromide was prepared by treating the ethyl...
bromoacetic acid with triphenylphosphine. The Wittig salt thus obtained was deprotonated and coupled oxidatively in the presence of 0.5 equiv of triphenyl phosphite–ozone product to yield [2,3-13C2]-diethyl fumarate.21 This is hydrolyzed by refluxing in concentrated hydrochloric acid. The resulting [2,3-13C2]fumaric acid is isomerized and dehydrated to give [2,3-13C2]MA (approximately 98% doubly labeled) by heating in the presence of phosphorus pentoxide. The overall yield is 44% based on acetic acid. NMR and mass spectrometry did not reveal any significant dilution or scrambling of the labels. Full experimental details and spectral identification will be presented in a forthcoming publication.

The polymers used are HDPE (DSM, essentially linear), LDPE (DSM, containing 25 wt% octene), EPM (random, 66 wt% E, E:P 3:1), EPM (random, 57 wt% E, E:P 2:1), alt-EP (hydrogenated natural rubber, E:P 1:1), EPM-1 (random, 40 wt% E, E:P 1:1), BASF 2900NCX (random, 18 wt% E, E:P 1:3) (E denotes ethene, P propene), and isotactic PP (DSM).

Results and Discussion

All polyolefins investigated in this study are grafted in a small scale extruder. This mimics actual industrial processes accurately, and optimally with respect to the amount of expensive labeled material needed, allowing the investigation of the grafting process with atomic resolution without any disturbance from reaction conditions or workup.

Structure of PE-g-MA. The top trace in Figure 1 shows the aliphatic part of the noise-decoupled 13C spectrum of HDPE grafted with [2,3-13C2]MA in the melt at 170°C with tert-butyl cumyl peroxide as the initiator. By comparison of this spectrum to that of HDPE grafted under identical conditions with unlabeled MA (data not shown) it is evident that the strong (truncated) signal at δ = 29.8 ppm is the natural abundance background of the HDPE backbone, while the signals at δ = 31.1 and 44.3 ppm are associated with the labels in the MA grafts. The 1D inadequate response of the same sample (Figure 1b) provides conclusive evidence for this assignment, since the natural abundance signal is strongly suppressed, while the adjacent labels give rise to large antiphase signals typical for this type of experiment. A splitting of ~34 Hz of the high-field label signal is observed, which is typical for a sp3–sp3 13C–13C one-bond coupling. 25 On the left hand side of the broad low-field signal small peaks separated by ~34 Hz can be distinguished. These signals are the doublet pair of an AB spin system, which is expected as the labeled resonances from the labels match that of the methylene protons of the MA grafted onto the PE backbone.

Peaks areas are estimated from the noise-decoupled spectra only. To determine the average lengths of the grafts for HDPE-g-MA 1D NMR spectra were recorded with a Bruker MSL 400 spectrometer at a temperature of 120°C. The spectra are referenced relative to TMS using the solvent signal as an internal standard. In various samples, T1 of the peaks were determined and then used to choose the appropriate recycle delay for the data acquisition. Noise-decoupled spectra were obtained using a short “tip” pulse and a recycle time of 2 s. 1D inadequate experiments were optimized for J = 34 Hz and spectra were acquired with a recycle time of 3 s, which is equal to 5 times the T1 of the labeled carbon atoms. The 13C-1H 2D J-resolved spectrum was recorded using the gated decoupling technique.

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Figure 1. Aliphatic part of (a) the noise-decoupled 13C NMR spectrum of HDPE grafted with [2,3-13C2]MA (the large backbone resonance at δ = 29.8 ppm is truncated) and (b) the 1D inadequate spectrum of the same material, in which the natural abundance resonance of the polymer chain is strongly reduced.

error. In the noise-decoupled spectra other than those from the PE-g-MA’s, the signals from the 13C-enriched sites are rather small relative to those from natural abundance sites and there is severe overlap between labeled and natural abundance signals; therefore average graft lengths could not be estimated for these materials.
which 2n + 1 are methines, for n + 1 MA rings in the graft (see structure I in Table 1). The $^{13}$C labels then give rise to a broad complex pattern due to the large number of $\mathcal{J}$ couplings, the occurrence of diasteromery, and the fact that the methine carbons have a small dispersion in chemical shifts. A signal at $\delta = 44 - 45$ ppm should indeed arise from oligomeric grafts, taking into account that the chemical shift of the methines in poly(MA) is about 45 ppm.27

Figure 2 shows the inadequate data for the various polyolefins grafted with [2,3-$^{13}$C$_2$]MA at 170 °C, either in the melt or in solution. The spectra are shown in magnitude mode for optimal visual clarity. They are placed in top-to-bottom order of decreasing E:P ratio, with the spectrum of each material grafted in solution placed in top-to-bottom order of decreasing E:P ratio, with the spectrum of each material grafted in solution below that of the same substrate grafted in the melt. Apparently, smaller amounts of [2,3-$^{13}$C$_2$]MA have grafted onto the polymer backbone in the melt-grafting experiments. For example, from the integrated responses it can be estimated that about 4 wt% [2,3-$^{13}$C$_2$]MA has grafted onto HDPE in solution and about 1 wt% in the melt-grafting experiments. For alt-EPM these numbers are about 3 wt% and about $3 \times 10^{-1}$ wt%, respectively. In the melt-grafted materials small residual peaks are observed due to incomplete double quantum filtering in the inadequate experiment and/or natural abundance spin pairs. These signals, in Figure 2 marked with an asterisk, are easily recognized by comparing the inadequate spectra with the noise-decoupled $^{13}$C spectra and can be assigned according to published data.28-29 In the spectrum of solution-grafted HDPE (Figure 2b), the expected response of the backbone methylene at $\delta = 29.8$ is invisibly small relative to label signals due to high levels of grafted [2,3-$^{13}$C$_2$]MA.

The LDPE is a copolymer of 1-octene (25 wt%) and ethene, and therefore it has as many tertiary carbon atoms in its chain as an EPM with a molar E:P ratio of 12:1. From the similarity of the signals at $\delta \approx 31.1$ and $\delta \approx 44.3$ ppm in the three PE spectra (Figure 2a–d) it is evident that MA attaches to similar sites in LDPE and HDPE, obviously extended polymethylene (CH$_2$)$_n$ sequences, in the form of both single anhydride rings and (short) oligomers, as represented by structure I in Table 1. Since the solution-grafted HDPE has been thoroughly extracted with boiling acetone, a good solvent for polyMA,27 it can be excluded that part of the label signals stem from oligo-MA that is not attached to PE. The overall mechanism of the grafting reaction is depicted in Figure 3.

From the integrated label signals in the noise-decoupled spectra of the grafted PEs, the values for n in structure I (Table 1) are estimated to be $0.8 \pm 0.1$ for HDPE grafted in the melt, $0.11 \pm 0.01$ for HDPE grafted in solution, and $0.30 \pm 0.02$ for melt-grafted LDPE. These values therefore indicate an average of about four dimeric grafts per every single graft for melt-grafted HDPE. Analogously, we deduce that there is an average of one dimeric graft per ten single ring grafts for solution-grafted HDPE and one dimeric graft per three monomeric units for melt-grafted LDPE. The higher values in the melt-grafting experiments can possibly be attributed to local high MA concentrations, due to limited solubility of MA in the polymer, which might favor MA homopolymerization. The predominance of single anhydride grafts in melt-grafted LDPE, when compared to melt-grafted HDPE, can probably be explained by the presence of some readily transferrable tertiary hydrogen atoms on the polymeric chain of LDPE, which can terminate the grafting reaction (vide infra). The width of the label responses in solution-grafted HDPE, in which predominantly single rings are present, and the absence of a prominent, neat AB pattern indicate that there is some dispersion in chemical shifts. For instance, clustering of grafts on the backbone caused by grafting of MA onto radical sites formed by intramolecular hydrogen abstraction by an appending succinyl radical may give rise to dispersed label signals. In contrast, for LDPE-g-MA, the relatively large width of the label responses could be simply associated with the many chemically different grafting sites on the backbone. Recently, Russell18 attributed the formation of oligomeric grafts during the radically initiated reaction between MA and low molecular weight aliphatic model compounds in solution at 60 and 80 °C to the fact that the reaction was carried out below the ceiling temperature of MA polymerization, which is estimated to be 108 °C for 1 M and 90 °C for 0.4 M monomer concentration.18 However, since we find

Table 1. $^{13}$C Chemical Shifts of the Methylene and Methine Carbons in the Anhydride Ring of MA-Grafted Polyolefins, the $^{13}$C–$^{13}$C $\mathcal{J}$ Coupling Constants between These Carbon Atoms, and the Associated Graft Structures

<table>
<thead>
<tr>
<th>material</th>
<th>$\delta$(CH$_2$) (ppm)</th>
<th>$\delta$(CH) (ppm)</th>
<th>$\mathcal{J}$ $^{13}$C–$^{13}$C (Hz)</th>
<th>structure</th>
</tr>
</thead>
<tbody>
<tr>
<td>HDPE-2[2,3-$^{13}$C$_2$]MA</td>
<td>31.1</td>
<td>44.3</td>
<td>33.7</td>
<td>I</td>
</tr>
<tr>
<td>LDPE-2[2,3-$^{13}$C$_2$]MA</td>
<td>31.1</td>
<td>44.2</td>
<td>33.9</td>
<td>I</td>
</tr>
<tr>
<td>EPM(3:1 E:P)-2[2,3-$^{13}$C$_2$]MA</td>
<td>31.2</td>
<td>44.2</td>
<td>33.9</td>
<td>I</td>
</tr>
<tr>
<td>EPM(2:1 E:P)-2[2,3-$^{13}$C$_2$]MA</td>
<td>31.2</td>
<td>48.5</td>
<td>33.9</td>
<td>II</td>
</tr>
<tr>
<td>alt-EPM-2[2,3-$^{13}$C$_2$]MA</td>
<td>31.2</td>
<td>48.6</td>
<td>34.2</td>
<td>II</td>
</tr>
<tr>
<td>EPM(1:1 E:P)-2[2,3-$^{13}$C$_2$]MA</td>
<td>31.1</td>
<td>44.2</td>
<td>34.2</td>
<td>II</td>
</tr>
<tr>
<td>EPM(1:3 E:P)-2[2,3-$^{13}$C$_2$]MA</td>
<td>31.1</td>
<td>48.3</td>
<td>34.2</td>
<td>II</td>
</tr>
<tr>
<td>iPP-2[2,3-$^{13}$C$_2$]MA</td>
<td>31.1</td>
<td>48.3</td>
<td>34.2</td>
<td>II</td>
</tr>
</tbody>
</table>

...
significant amounts of dimeric or oligomeric grafts in [2,3-13C2]MA-grafted PEs prepared in either the melt or in solution at 170 °C, our experiments do not provide support for a clear correlation between ceiling temperature and the occurrence of oligomeric grafts.

**Structure of EPM-g-MA.** In the spectra of the melt-grafted 3:1 and 2:1, and the solution-grafted 1:1 E:P random EPMs (Figure 2d, e, i, respectively) signals from labeled sites are found at the same chemical shifts as in the spectrum of grafted HDPE. Hence, these signals are attributed to the same structure as in HDPE-[2,3-13C2]MA (structure I in Table 1). The corresponding signal cannot be observed in the spectrum of melt-grafted random 1:1 E:P copolymer (Figure 2h), but this is most probably due to a low degree of grafting in the melt in this material.

The spectra of the melt-grafted 3:1 and 2:1 E:P random copolymers (Figure 2d, e, respectively), the melt-grafted alt-EPM (Figure 2f), the solution-grafted alt-EPM (Figure 2g), and the melt- and solution-grafted random 1:1 EP copolymers (Figure 2h, i, respectively) show a signal at δ = 48.6 ppm. This is close to the chemical shift of the methine of the anhydride ring (δ = 47.9 ppm) in the model compound (4-methylhept-4-yl)succinic anhydride.26 Therefore, this signal should arise from the methine group in succinic anhydride rings formed by the attachment of [2,3-13C2]MA to tertiary radicals on the polymer chain, as represented by structure II in Table 1. Since there is no response present at δ ≈ 45 ppm in the spectrum of alt-EPM-[2,3-13C2]MA grafted in either the melt or solution, which could be attributed to the methine in a succinic anhydride ring attached to a methylene, our results demonstrate that MA attaches predominantly to the tertiary carbon atoms of the alt-EPM backbone and that it attaches in the form of single anhydride rings, since oligomeric structures would have caused an additional response about δ = 45 ppm. The mechanism of grafting MA onto the random EPMs discussed above appears to be a combination of that of PE and alt-EPM, which are given in Figure 3. The responses from the methylene carbons in the anhydride rings attached to the various sites in EPM all coincide at ~31 ppm.

**Structure of iPP-g-MA and P-Rich EPM-g-MA.** Figure 2l shows parts of the inadequate spectrum of iPP grafted in solution. It contains a signal at δ = 31.6 ppm and a broader signal at δ = 49.5 ppm. These are likely from single succinic anhydride rings that are formed when [2,3-13C2]MA reacts with a tertiary radical site on the iPP backbone (structure III in Table 1). The methine of the anhydride ring in iPP-g-[2,3-13C2]MA resonates at somewhat lower field when compared to the one in alt-EPM-g-[2,3-13C2]MA (δ = 48.6 ppm), most probably due to the proximity of methyl groups on the iPP backbone. De Roover et al.17 reported that PP-g-MA contains oligomeric or even polymeric grafts, exclusively attached to the chain ends of PP. However, such graft structures should give rise to signals at δ ≈ 45 ppm, which are not observed in our spectra (Figure 2k, l). Hence, in our opinion, the mechanism as proposed by Gaylord,7 describing the formation of oligomeric graft structures in PP-g-MA, is definitely not unique and is of minor importance for high temperature PP grafting. In addition, the presence of single anhydride rings attached to the PP chain end, formed by the addition of MA to secondary radicals produced by β-scission of iPP, can be excluded, taking into account that the chemical shift of the ring methine in the model compound (4-methylpent-2-yl)succinic anhydride is about 45.8 ppm.26

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**Figure 2.** Magnitude-calculated inadequate spectra of PE, PP, and EPM grafted with [2,3-13C2]MA at 170 °C in the melt and in solution. E:P (equivalent) ratio of the copolymer: (a) 1:0 (HDPE, melt); (b) 1:0 (HDPE, solution); (c) 12:1 (LDPE, 25 wt % octene, random, melt); (d) 3:1 (random, melt); (e) 2:1 (random, melt); (f) 1:1 (hydrogenated alt-EPM, melt); (g) 1:1 (hydrogenated alt-EPM, solution); (h) 1:1 (random, melt); (i) 1:1 (random, solution); (j) 1:3 (random, melt). Residual responses from unlabeled sites are marked with an asterisk.
Figure 3. Proposed mechanisms for the grafting of MA onto PE, alt-EPM, and iPP, and onto corresponding structures in copolymers. $I^*$ represents an initiator radical, $R$ a polymer chain, and $R^*$ a macroradical.

Grafting onto PE (sequences):

Grafting onto alt-EPM (sequences):

Grafting onto iPP (sequences):

The spectrum in Figure 2l shows two pairs of doublets centered at $\delta \approx 34.3$ ppm, in the spectral region of aliphatic carbons, and at 117.5 ppm, in the olefin region. Each doublet in both pairs has a splitting matching that of a doublet in the other pair, the values being 43.8 and 45.3 Hz, which are normal values for splittings caused by a one-bond $J_{CC}$ coupling. Two very similar, isomeric graft structures, with $E$ and $Z$ configurations around a double bond, can be formed when chain scission occurs near the site of attachment of an anhydride ring, which would explain the observation of two sets of corresponding doublets at similar chemical shifts (structure IV in Table 1). This mechanism, depicted in Figure 3, resembles the one describing the $\beta$-scission in PP, with the marked difference that in the present case a primary radical and a fully substituted double bond are finally created, while in the PP $\beta$ chain scission mechanism a secondary radical and a methyldiene double bond are formed. The spectrum of iPP grafted in the melt (Figure 2k) shows weak responses from labeled sites at the same chemical shifts as in the spectrum of solution-grafted iPP, revealing that in this material [2,3-$^{13}$C$_2$]MA grafts to the same sites.

Figure 4 shows the aliphatic part of a 2D-$J$-resolved spectrum of iPP-g-[2,3-$^{13}$C$_2$]MA. The signals at $\delta = 34.3$ ppm from the methylene in the anhydride ring pendant from the chain terminus by a double bond are two triplets due the combined effect of homonuclear $^{13}$C $J$ coupling and heteronuclear coupling with the protons directly attached to this carbon atom. The triplets are shifted in the $J$ domain with respect to each other due to the $^1H-^{13}C J$ coupling. It can be concluded that no proton is attached to the double bond, since this would cause a further splitting, which is not observed. This supports the proposed structure IV (Table 1), and hence the proposed mechanism as depicted in Figure 3.

The spectrum of the 1:3 E:P EPM (Figure 2j) shows resonances at the same chemical shifts as the signals from the labeled sites in HDPE-g-[2,3-$^{13}$C$_2$]MA (Figure 2a,b), which are attributed to structure I (Table 1). Two weak resonances at $\delta = 48.2$ and $\delta = 49.8$ ppm can be recognized that are also observed in the data collected from the grafted 1:1 E:P EPM (Figure 2h,i) and from iPP-g-[2,3-$^{13}$C$_2$]MA (Figure 2k,l). These signals are therefore attributed to structures II and III (Table 1). The signal at $\delta \approx 34$ ppm is attributed to a methylene in a structure similar to that present in iPP-g-[2,3-$^{13}$C$_2$]-MA, i.e. succinic anhydride attached to a chain terminus by a double bond (structure IV in Table 1). The signal from the adjacent labeled site in the connecting double bond can be found at $\delta = 117$ ppm (not shown) but is less intense than in the spectrum of iPP-g-[2,3-$^{13}$C$_2$]MA, apparently due to a lower degree of this mode of grafting in this material. In the 1:3 E:P random copolymer there is a multitude of possible sites of attachment, and the combination of spectral features found in the spectra of HDPE-g-[2,3-$^{13}$C$_2$]MA, alt-EPM-g-[2,3-$^{13}$C$_2$]MA, and iPP-
should abstract a hydrogen atom from a secondary carbon atom in the backbone, creating a secondary radical. Since MA grafts readily onto secondary radicals, as in the case of PE, and no signals attributable to MA grafted onto a secondary backbone radical are observable in the spectrum of alt-EPM-MA, we conclude that there are no or few secondary radicals present in alt-EPM during radially initiated grafting of MA, and hence terminative hydrogen transfer from methine carbons does not involve a six-membered ring transition state. This is corroborated by the observation that in PE oligomeric grafts are present, while grafts occur solely as single rings in MA-grafted alt-EPM. Apparently, when radially initiated MA grafting is performed in either the melt or solution, the MA oligomerization and hydrogen transfer from methylenes proceed at similar rates, while hydrogen transfer from methine carbons proceeds faster than both.

The spectra of the materials grafted in the melt or the solution processes are very similar. This indicates that the structure and the sites of attachment of the grafts are largely independent of the method of functionalization. The major difference between the two methods is the higher level of grafted [2,3-\textsuperscript{13}C\textsubscript{2}]MA in the products from the solution process, but this can easily be rationalized by considering the larger MA to polyolefin ratio and the longer reaction times used there. Finally, we find no indication for MA acting as a cross-linker between two polymer chains, in the form of a single anhydride ring, as has been suggested.\textsuperscript{8} The overall mechanism accurately describing the grafting of MA onto polyolefins is depicted in Figure 3.

**Conclusion**

The structure of polyolefins grafted with MA in a melt and a solution process at 170 °C has been elucidated by \textsuperscript{13}C NMR spectroscopy in conjunction with specific isotope labeling of MA. By placing two adjacent \textsuperscript{13}C labels in the MA monomer, the inadequate technique can be successfully applied to suppress the intense resonances corresponding to the carbon atoms in the polymer backbone, thus showing only the signals of main interest when studying the grafting process.

MA attaches to the HDPE and LDPE in the form of single succinic anhydride rings as well as short oligomers. In (co)polymers with abundant tertiary hydrogen atoms, such as alt-EPM and iPP, MA grafts onto the polymer backbone chiefly in the form of single succinic anhydride rings. Polyethylene sequences (CH\textsubscript{2})\textsubscript{m}, with m > 3 are needed to allow grafting of MA onto a backbone methylene. In (co)polymers with high P content, i.e. iPP and 1:3 E-P copolymers, a chain scission reaction can occur, yielding an anhydride ring attached to the chain terminus via a double bond.

The structure and the site of attachment of the graft do not appear to depend on the method of functionalization, either in solution or under melt conditions. No evidence is found for MA acting as a cross-linking agent.

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**References and Notes**


Grafting of [2,3-13C2]MA onto PE, PP, and EPM

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