



Universiteit
Leiden
The Netherlands

Anisotropy, multivalency and flexibility-induced effects in colloidal systems

Verweij, R.W.

Citation

Verweij, R. W. (2021, May 27). *Anisotropy, multivalency and flexibility-induced effects in colloidal systems. Casimir PhD Series*. Retrieved from <https://hdl.handle.net/1887/3179461>

Version: Publisher's Version

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/3179461>

Note: To cite this publication please use the final published version (if applicable).

Cover Page



Universiteit Leiden



The handle <https://hdl.handle.net/1887/3179461> holds various files of this Leiden University dissertation.

Author: Verweij, R.W.

Title: Anisotropy, multivalency and flexibility-induced effects in colloidal systems

Issue Date: 2021-05-27

List of publications

In this thesis:

1. Conformations and diffusion of flexibly linked colloidal chains.

R.W. Verweij, P.G. Moerman, L.P.P. Huijnen, N.E.G. Ligthart, I. Chakraborty, J. Groenewold, W.K. Kegel, A. van Blaaderen and D.J. Kraft

Journal of Physics: Materials, in press (2021). doi:10.1088/2515-7639/abf571 In: Chapter 6.

2. Height distribution and orientation of colloidal dumbbells near a wall.

R.W. Verweij*, S. Ketsetzi*, J. Graaf and D.J. Kraft

Phys. Rev. E, 102, 062608 (2020). doi:10.1103/PhysRevE.102.062608 In: Chapter 2.

3. Flexibility-induced effects in the Brownian motion of colloidal trimers.

R.W. Verweij*, P.G. Moerman*, N.E.G. Ligthart, L.P.P. Huijnen, J. Groenewold, W.K. Kegel, A. van Blaaderen and D.J. Kraft

Phys. Rev. Research, 2, 033136 (2020). doi:10.1103/PhysRevResearch.2.033136 In: Chapter 5.

4. Colloid supported lipid bilayers for self-assembly.

M. Rinaldin*, R.W. Verweij*, I. Chakraborty and D.J. Kraft

Soft Matter, 15, 1345-1360 (2019). doi:10.1039/C8SM01661E In: Chapter 3.

Other:

5. Dumbbell impurities in 2D crystals of repulsive colloidal spheres induce particle-bound dislocations.

V. Meester, C. van der Wel, R.W. Verweij, G. Biondaro, D.J. Kraft

Under review (submitted Oct. 2020).

6. Micrometer-sized TPM emulsion droplets with surface-mobile binding groups.

C. van der Wel, G.L. van de Stolpe, R.W. Verweij, D.J. Kraft

J. Phys. Condens. Matter, 30, 094005 (2018). doi:10.1088/1361-648x/aaab22

7. Preparation of colloidal organosilica spheres through spontaneous emulsification.

C. van der Wel, R.K. Bhan, R.W. Verweij, H.C. Frijters, Z. Gong, A.D. Hollingsworth, S. Sacanna and D.J. Kraft

Langmuir, 33, 8174-8180 (2017). doi:10.1021/acs.langmuir.7b01398

8. Colloidal recycling: reconfiguration of random aggregates into patchy particles.

V. Meester, R.W. Verweij, C. van der Wel and D.J. Kraft

ACS Nano, 10, 4322-4329 (2016). doi:10.1021/acsnano.5b07901

*These authors contributed equally.

About the author

I was born on June 13th, 1992 in the Dutch village Heemskerk and grew up in the neighboring city of Beverwijk. I received my secondary education at the Gymnasium Felisenum in Velsen-Zuid, where I graduated cum laude in 2010. Next, I started my Bachelor studies in Physics at Leiden University. During my Bachelor studies, I additionally obtained a Propaedeutics degree in Computer Science and a Propaedeutics degree in Astronomy. Next to my studies, I worked as a tutor for high school students and as a web developer. I was a member of the student rowing association Asopos de Vliet. In 2014, I finished my Bachelor degree in Physics with the research project titled “Close-packed colloidal clusters” supervised by Vera Meester and Daniela Kraft.

Then, I started my Master studies in Physics at Leiden University, where I specialized in Biological and Soft Matter Physics. During this time, I was a member of the student canoe association Levitas. As part of my studies, I completed two research projects. The first project was titled “Simulation of diffusion-weighted MRI” and was supervised by Joor Arkesteijn, Frans Vos and Lucas van Vliet (Delft University). In this project, we collaborated with Farida Grinberg and Ezequiel Farrher from Forschungszentrum Jülich (Germany). For my second project titled “Synthesis of polymerizable emulsions and bulk synthesis of mobile clusters”, I was supervised by Casper van der Wel and Daniela Kraft. I obtained my Master degree in Physics in 2016.

Shortly afterwards, I started my PhD under the supervision of Daniela Kraft, which resulted in the present thesis on the effects of anisotropy, multivalency and flexibility on colloidal systems. In addition to the research presented here, I have supervised four students during their research projects. I was a Teaching Assistant for the “Diffusion” and “Experimental Physics” courses. I have attended the Han-sur-Lesse Winterschool for Physical Chemistry (Belgium) twice and I followed a Master course on “Deep learning and Neural networks”. Additionally, I attended several conferences to which I collaborated several poster presentations. I won the prize for the best poster at the Dutch Chemistry conference CHAINS (2017). I have given a talk during the International Soft Matter Conference (Edinburgh, UK, 2019). During my PhD studies, I collaborated with Melissa Rinaldin for Chapter 3, with Pepijn Moerman, Jan Groenewold, Willem Kegel and Alfons van Blaaderen (Utrecht University) for Chapters 5–6 and with Joost de Graaf (Utrecht University) and Stefania Ketzetzi for Chapter 2.

After my PhD studies, I will start as Modeler Air Quality at the *Rijksinstituut voor Volksgezondheid en Milieu* (RIVM, National Institute for Public Health and the Environment).

Acknowledgments

I am deeply grateful to a number of people that helped me during the course of my PhD studies. First, I would like to thank my supervisor Daniela Kraft for her excellent scientific advice, her overall support, enthusiasm and positive attitude: I could not have asked for a better PhD advisor. I thank Martin van Hecke for being my extra promotor and for useful discussions. I am grateful to the members of the Doctoral Committee for their critical assessment of my work.

Next, I would like to thank the people that have collaborated with me on parts of this thesis. I would like to thank Joost de Graaf (Utrecht University) and Stefania Ketsetzi for their contributions to Chapter 2. Joost, thank you for your insightful and speedy contributions, that often arrived late at night. Stefania, you have been a great office mate and I have enjoyed working together with you on this project. Thank you for all of your support and for being my paranympth. I would like to thank my other office mate, Melissa Rinaldin, not only for being a great colleague but also for her extensive contributions to Chapter 3. I am grateful to Indrani Chakraborty, who not only taught me how to make colloid supported lipid bilayers but also contributed additional data to Chapters 3, 6 and 7. I am heavily indebted to our collaborators from Utrecht University: Pepijn Moerman, Nathalie Ligthart, Willem Kegel, Jan Groenewold and Alfons van Blaaderen, who have made significant contributions to Chapters 5 and 6. Pepijn, thanks a lot for the pleasant collaboration and all of your input. I am grateful to Aleksandar Donev (NYU) and Brennan Sprinkle (NYU) for their advise on the simulations in Chapters 6 and 7. I thank Piotr Szymczak (University of Warsaw) for useful discussions about the center of diffusion of flexible objects.

During my PhD studies, I had the pleasure of supervising a number of students during their research projects: Max Ruckriegel, Loes Huijnen, Sarah Smolders and Nick Oikonomou. Max, thank you for being my first student, I have enjoyed working together. Loes, thank you for your great contributions to Chapters 5–7 and the nice collaboration. Sarah, thank you for paving the way for the measurements in Chapter 2. Nick, thank you for your exploratory experiments that led to Chapter 2, the interesting discussions on holographic microscopy and machine learning and the great collaboration. On that note, I would like to thank Vera Meester and Casper van der Wel, who have been fantastic supervisors during my own respective Bachelor and Master research projects and have inspired me to pursue a PhD in colloid science. I would like to express my gratitude to my high school teacher Winfried Appelman for showing me how exciting Physics can be. I am grateful to my university teacher Robert-Jan Kooman for enthusing me about Mathematics. I thank Kick Moors for his invaluable advise.

I thank the LION and Graduate School secretaries for their administrative support and want to express my deepest gratitude in particular to Daniëlle Duijn-ter Veer, for all her help and patience. I would like to thank the IT department for their support and am grateful for access to the Maris and ALICE computing clusters. I thank the

fine mechanical and electronic departments for their technical support.

I would like to thank all former and present members of the Leiden Soft Matter groups for their useful feedback during meetings but moreover, for the nice coffee breaks, long lunches, borrels and group outings. You have made it a pleasure to work each day, even on days when experimental results were scarce. I especially thank Rachel Doherty for keeping the lab running and for her experimental support. Finally, I am deeply grateful to Ali Azadbakht for building the Optical Tweezers setup and for teaching me how to use it.

Als laatste wil ik mijn vrienden bedanken voor hun geduld bij het aanhoren van mijn uitleg over mijn onderzoek en voor hun vriendschap. Ik wil mijn familie bedanken voor al hun steun. Wim en Fieneke Tesselaar, bedankt voor alle goede zorgen toen ik een tijdje bij jullie mocht inwonen, daarvoor en ook daarna. Ik bedank mijn tante Loes voor de logeerplek in Leiden en voor inspirerende gesprekken over studiekeuzes en onderzoek. Ook ben ik mijn opa Peter en oma Luck erg dankbaar, zij zijn heel nauw betrokken geweest bij mijn opvoeding en hebben mij altijd gesteund. Ik bedank mijn ouders Alexander & Désirée en mijn broertje Maurits voor al hun steun. In het bijzonder ben ik mijn moeder heel erg dankbaar, bij wie ik altijd terecht kon voor hulp en advies. Als laatste wil ik mijn partner Mark bedanken voor alles: dit proefschrift heeft ook van jou veel gevergd, maar je bent mij altijd blijven steunen.

Bibliography

1. H. Míguez, S. M. Yang, and G. A. Ozin. Optical properties of colloidal photonic crystals confined in rectangular microchannels. *Langmuir*, 19(8):34793485, 2003.
2. M. He, J. P. Gales, É. Ducrot, Z. Gong, G.-R. Yi, S. Sacanna, and D. J. Pine. Colloidal diamond. *Nature*, 585(7826):524–529, 2020.
3. Y. Xia, Y. Yin, Y. Lu, and J. McLellan. Template-assisted self-assembly of spherical colloids into complex and controllable structures. *Adv. Funct. Mater.*, 13(12):907–918, 2003.
4. W. B. Rogers, W. M. Shih, and V. N. Manoharan. Using DNA to program the self-assembly of colloidal nanoparticles and microparticles. *Nat. Rev. Mater.*, 1(3):1–14, 2016.
5. E. Dickinson. Colloids in food: ingredients, structure, and stability. *Annu. Rev. Food Sci. Technol.*, 6:211–233, 2015.
6. D. P. Otto, A. Otto, and M. M. De Villiers. Differences in physicochemical properties to consider in the design, evaluation and choice between microparticles and nanoparticles for drug delivery. *Expert Opin. Drug Deliv.*, 12(5):763–777, 2015.
7. S. Scalia, P. M. Young, and D. Traini. Solid lipid microparticles as an approach to drug delivery. *Expert Opin. Drug Deliv.*, 12(4):583–599, 2015.
8. D. S. Kohane. Microparticles and nanoparticles for drug delivery. *Biotechnology and bioengineering*, 96(2):203–209, 2007.
9. A. M. Carmona-Ribeiro. Preparation and characterization of biomimetic nanoparticles for drug delivery. In *Nanoparticles in Biology and Medicine: Methods and Protocols*, pages 283–294. Humana Press, Totowa, NJ, 2012.
10. H. Löwen. Colloidal soft matter under external control. *J. Phys.: Condens. Matter*, 13(24): R415R432, 2001.
11. B. Carrasco and J. G. De La Torre. Hydrodynamic properties of rigid particles: Comparison of different modeling and computational procedures. *Biophys. J.*, 76(6):3044–3057, 1999. doi:10.1016/S0006-3495(99)77457-6.
12. J. Fung and V. N. Manoharan. Holographic measurements of anisotropic three-dimensional diffusion of colloidal clusters. *Phys. Rev. E*, 88(2):020302, 2013. doi:10.1103/PhysRevE.88.020302.
13. D. J. Kraft, R. Wittkowski, B. Ten Hagen, K. V. Edmond, D. J. Pine, and H. Löwen. Brownian motion and the hydrodynamic friction tensor for colloidal particles of complex shape. *Phys. Rev. E*, 88(5):050301, 2013. doi:10.1103/PhysRevE.88.050301.
14. A. Chakrabarty, F. Wang, K. Sun, and Q. H. Wei. Effects of translation-rotation coupling on the displacement probability distribution functions of boomerang colloidal particles. *Soft Matter*, 12(19):4318–4323, 2016. doi:10.1039/c6sm00568c.
15. S. Chemburu, K. Fenton, G. P. Lopez, and R. Zeineldin. Biomimetic Silica Microspheres in Biosensing. *Molecules*, 15(12):1932–1957, 2010.
16. M. Hoffmann, C. S. Wagner, L. Harnau, and A. Wittemann. 3D Brownian diffusion of submicron-sized particle clusters. *ACS Nano*, 3(10):3326–3334, 2009. doi:10.1021/nn900902b.
17. A. Einstein. Über die von der molekularkinetischen Theorie der Wärme geforderte Bewegung von in ruhenden Flüssigkeiten suspendierten Teilchen. *Annalen der Physik*, 322(8): 549–560, 1905. doi:10.1002/andp.19053220806.
18. W. Sutherland. LXXV. A dynamical theory of diffusion for non-electrolytes and the molecular mass of albumin. *London Edinburgh Philos. Mag. J. Sci.*, 9(54):781–785, 1905. doi:10.1080/14786440509463331.

19. J. Perrin. Mouvement brownien et réalité moléculaire. *Ann. Chim. Phys.*, 18:104, 1909.
20. J. N. Israelachvili. *Intermolecular and surface forces*. Academic press, 2011.
21. L. Teulon, Y. Hallez, S. Raffy, F. Guerin, E. Palleau, and L. Ressier. Electrostatic directed assembly of colloidal microparticles assisted by convective flow. *J. Phys. Chem. C*, 123(1):783–790, 2018.
22. V. A. Parsegian. *Van der Waals forces: a handbook for biologists, chemists, engineers, and physicists*. Cambridge University Press, 2005.
23. H. N. Lekkerkerker and R. Tuinier. Depletion interaction. In *Colloids and the depletion interaction*, pages 57–108. Springer, 2011.
24. L. Rossi, S. Sacanna, W. T. M. Irvine, P. M. Chaikin, D. J. Pine, and A. P. Philipse. Cubic crystals from cubic colloids. *Soft Matter*, 7(9):4139–4142, 2011.
25. S. Sacanna, L. Rossi, and D. J. Pine. Magnetic click colloidal assembly. *J. Am. Chem. Soc.*, 134(14):6112–6115, 2012.
26. G. I. Vega-Bellido, R. A. DeLaCruz-Araujo, I. Kretzschmar, and U. M. Córdova-Figueroa. Self-assembly of magnetic colloids with shifted dipoles. *Soft Matter*, 15(20):4078–4086, 2019.
27. A. Sánchez-Iglesias, M. Grzelczak, T. Altantzis, B. Goris, J. Perez-Juste, S. Bals, G. Van Tendeloo, S. H. Donaldson Jr, B. F. Chmelka, J. N. Israelachvili, et al. Hydrophobic interactions modulate self-assembly of nanoparticles. *ACS Nano*, 6(12):11059–11065, 2012.
28. Q. Chen, S. C. Bae, and S. Granick. Directed self-assembly of a colloidal kagome lattice. *Nature*, 469(7330):381–384, 2011.
29. S. Faber, Z. Hu, G. H. Wegdam, P. Schall, et al. Controlling colloidal phase transitions with critical casimir forces. *Nat. Commun.*, 4(1):1–6, 2013.
30. T. A. Nguyen, A. Newton, D. J. Kraft, P. G. Bolhuis, and P. Schall. Tuning patchy bonds induced by critical casimir forces. *Materials*, 10(11):1265, 2017.
31. J. Happel and H. Brenner. *Low Reynolds number hydrodynamics with special applications to particulate media*. Springer, 1983. doi:10.1007/978-94-009-8352-6.
32. H. Faxen. The resistance against the movement of a rigour sphere in viscous fluids, which is embedded between two parallel layered barriers. *Ann. Phys.*, 4:79–89, 1922.
33. K. Zahn, J. M. Méndez-Alcaraz, and G. Maret. Hydrodynamic interactions may enhance the self-diffusion of colloidal particles. *Phys. Rev. Lett.*, 79(1):175, 1997.
34. J. Rotne and S. Prager. Variational Treatment of Hydrodynamic Interaction in Polymers. *J. Chem. Phys.*, 50(11):4831–4837, 1969. doi:10.1063/1.1670977.
35. J. W. Swan and J. F. Brady. Simulation of hydrodynamically interacting particles near a no-slip boundary. *Physics of Fluids*, 19(11):113306, 2007. doi:10.1063/1.2803837.
36. A. Furukawa and H. Tanaka. Key role of hydrodynamic interactions in colloidal gelation. *Phys. Rev. Lett.*, 104(24):245702, 2010.
37. P. P. Lele, J. W. Swan, J. F. Brady, N. J. Wagner, and E. M. Furst. Colloidal diffusion and hydrodynamic screening near boundaries. *Soft Matter*, 7:6844–6852, 2011. doi:10.1039/C0SM01466D.
38. J. Ortega-Vinuesa, A. Martin-Rodriguez, and R. Hidalgo-Alvarez. Colloidal stability of polymer colloids with different interfacial properties: mechanisms. *J. Colloid Interface Sci.*, 184(1):259–267, 1996.
39. S. Flicker and S. Bike. Measuring double layer repulsion using total internal reflection microscopy. *Langmuir*, 9(1):257–262, 1993.
40. A. Dolan and S. F. Edwards. Theory of the stabilization of colloids by adsorbed polymer. *Proc. R. Soc. A*, 337(1611):509–516, 1974.

41. K. Ueno, A. Inaba, M. Kondoh, and M. Watanabe. Colloidal stability of bare and polymer-grafted silica nanoparticles in ionic liquids. *Langmuir*, 24(10):5253–5259, 2008.
42. E. B. Zhulina, O. V. Borisov, and V. A. Priamitsyn. Theory of steric stabilization of colloid dispersions by grafted polymers. *J. Colloid Interface Sci.*, 137(2):495–511, 1990.
43. J. Rädler, H. Strey, and E. Sackmann. Phenomenology and Kinetics of Lipid Bilayer Spreading on Hydrophilic Surfaces. *Langmuir*, 11(11):4539–4548, 1995. doi:10.1021/la00011a058.
44. P. S. Cremer and S. G. Boxer. Formation and Spreading of Lipid Bilayers on Planar Glass Supports. *The J. Phys. Chem. B*, 103(13):2554–2559, 1999.
45. R. P. Richter, R. Bérat, and A. R. Brisson. Formation of solid-supported lipid bilayers: an integrated view. *Langmuir*, 22(8):3497–3505, 2006.
46. Y. Jing, H. Trefna, M. Persson, B. Kasemo, and S. Svedhem. Formation of supported lipid bilayers on silica: relation to lipid phase transition temperature and liposome size. *Soft Matter*, 10(1):187–95, 2014.
47. J. Grdadolnik, F. Merzel, and F. Avbelj. Origin of hydrophobicity and enhanced water hydrogen bond strength near purely hydrophobic solutes. *Proc. Natl. Acad. Sci. U.S.A.*, 114(2):322–327, 2017.
48. G. M. Whitesides and B. Grzybowski. Self-Assembly at All Scales. *Science*, 295:1644–1225, 2002.
49. M.-P. Valignat, O. Theodoly, J. C. Crocker, W. B. Russel, and P. M. Chaikin. Reversible self-assembly and directed assembly of DNA-linked micrometer-sized colloids. *Proc. Natl. Acad. Sci. U.S.A.*, 102(12):4225–4229, 2005.
50. P. L. Biancaniello, A. J. Kim, and J. C. Crocker. Colloidal interactions and self-assembly using DNA hybridization. *Phys. Rev. Lett.*, 94(5):058302, 2005.
51. W. Li, H. Palis, R. Mérindol, J. Majimel, S. Ravaine, and E. Duguet. Colloidal molecules and patchy particles: complementary concepts, synthesis and self-assembly. *Chem. Soc. Rev.*, 49(6):1955–1976, 2020.
52. D. J. Kraft, W. S. Vlug, C. M. van Kats, A. van Blaaderen, A. Imhof, and W. K. Kegel. Self-assembly of colloids with liquid protrusions. *J. Am. Chem. Soc.*, 131(3):1182–1186, 2009.
53. É. Duguet, C. Hubert, C. Chomette, A. Perro, and S. Ravaine. Patchy colloidal particles for programmed self-assembly. *Comptes Rendus Chimie*, 19(1-2):173–182, 2016.
54. S. Sacanna, M. Korpics, K. Rodriguez, L. Colón-Meléndez, S.-H. Kim, D. J. Pine, and G.-R. Yi. Shaping colloids for self-assembly. *Nat. Commun.*, 4(1):1–6, 2013.
55. M. Hadorn and P. Eggenberger Hotz. DNA-Mediated Self-Assembly of Artificial Vesicles. *PLoS ONE*, 5(3):e9886, 2010.
56. Y. Zhang, A. McMullen, L. L. Pontani, X. He, R. Sha, N. C. Seeman, J. Brujic, and P. M. Chaikin. Sequential self-assembly of DNA functionalized droplets. *Nat. Commun.*, 8(1): 21, 2017.
57. D. Ortiz, K. L. Kohlstedt, T. D. Nguyen, and S. C. Glotzer. Self-assembly of reconfigurable colloidal molecules. *Soft Matter*, 10(20):3541–3552, 2014.
58. K. L. Kohlstedt and S. C. Glotzer. Self-assembly and tunable mechanics of reconfigurable colloidal crystals. *Phys. Rev. E*, 87(3), 2013.
59. J. D. Joannopoulos, P. R. Villeneuve, and S. Fan. Photonic crystals: putting a new twist on light. *Nature*, 386(6621):143–149, 1997.
60. Y. Lin, P. R. Herman, C. E. Valdivia, J. Li, V. Kitaev, and G. A. Ozin. Photonic band structure of colloidal crystal self-assembled in hollow core optical fiber. *Appl. Phys. Lett.*, 86(12):121106, 2005.

61. E. Sackmann. Supported membranes: Scientific and practical applications. *Science*, 271(5245):43–48, 2007.
62. C. Madwar, G. Gopalakrishnan, and R. B. Lennox. Interfacing living cells and spherically supported bilayer lipid membranes. *Langmuir*, 31(16):4704–4712, 2015.
63. S. Mashaghi, T. Jadidi, G. Koenderink, and A. Mashaghi. Lipid Nanotechnology. *Int. J. Mol. Sci.*, 14:4242–4282, 2013.
64. M. Rinaldin, P. Fonda, L. Giomi, and D. J. Kraft. Geometric pinning and antimixing in scaffolded lipid vesicles. *Nat. Commun.*, 11(1):1–10, 2020.
65. P. Fonda, M. Rinaldin, D. J. Kraft, and L. Giomi. Interface geometry of binary mixtures on curved substrates. *Phys. Rev. E*, 98(3):032801, 2018.
66. F. Gentile, M. Moretti, T. Limongi, A. Falqui, G. Bertoni, A. Scarpellini, S. Santoriello, L. Maragliano, R. Proietti Zaccaria, and E. Di Fabrizio. Direct imaging of DNA fibers: the visage of double helix. *Nano Lett.*, 12(12):6453–6458, 2012.
67. Sponk. Difference DNA-RNA, 2010. URL commons.wikimedia.org/wiki/File:Difference_DNA_RNA-DE.svg.
68. M. P. Ball. Ligation, 2007. URL commons.wikimedia.org/wiki/File:Ligation.svg.
69. Y. Wang, Y. Wang, X. Zheng, É. Ducrot, J. S. Yodh, M. Weck, D. J. Pine, E. Ducrot, J. S. Yodh, M. Weck, and D. J. Pine. Crystallization of DNA-coated colloids. *Nat. Commun.*, 6:7253, 2015.
70. C. A. Mirkin, R. L. Letsinger, R. C. Mucic, and J. J. Storhoff. A DNA-based method for rationally assembling nanoparticles into macroscopic materials. *Nature*, 382(6592):607–609, 1996.
71. D. Nykypanchuk, M. M. Maye, D. Van Der Lelie, and O. Gang. DNA-guided crystallization of colloidal nanoparticles. *Nature*, 451(7178):549–552, 2008.
72. M. E. Leunissen, R. Dreyfus, R. Sha, T. Wang, N. C. Seeman, D. J. Pine, and P. M. Chaikin. Towards self-replicating materials of DNA-functionalized colloids. *Soft Matter*, 5(12):2422–2430, 2009. doi:10.1039/b817679e.
73. M. E. Leunissen and D. Frenkel. Numerical study of DNA-functionalized microparticles and nanoparticles: Explicit pair potentials and their implications for phase behavior. *The J. Chem. Phys.*, 134(8):084702, 2011.
74. N. Geerts and E. Eiser. DNA-functionalized colloids: Physical properties and applications. *Soft Matter*, 6(19):4647–4660, 2010.
75. L. Di Michele and E. Eiser. Developments in understanding and controlling self assembly of DNA-functionalized colloids. *Phys. Chem. Chem. Phys.*, 15(9):3115–3129, 2013. doi:10.1039/c3cp43841d.
76. L. Di Michele, F. Varrato, J. Kotar, S. H. Nathan, G. Foffi, and E. Eiser. Multistep kinetic self-assembly of DNA-coated colloids. *Nat. Commun.*, 4:2007, 2013.
77. M. L. Mansfield, L. Rakesh, and D. A. Tomalia. The random parking of spheres on spheres. *J. Chem. Phys.*, 105(8):3245–3249, 1996.
78. Y. Wang, Y. Wang, X. Zheng, É. Ducrot, M.-G. Lee, G.-R. Yi, M. Weck, and D. J. Pine. Synthetic strategies toward DNA-coated colloids that crystallize. *J. Am. Chem. Soc.*, 137(33):10760–10766, 2015.
79. H. R. Vutukuri, J. Stiefelhagen, T. Vissers, A. Imhof, and A. van Blaaderen. Bonding assembled colloids without loss of colloidal stability. *Adv. Mater.*, 24(3):412–416, 2012.
80. Y. Wang, Y. Wang, D. R. Breed, V. N. Manoharan, L. Feng, A. D. Hollingsworth, M. Weck, and D. J. Pine. Colloids with valence and specific directional bonding. *Nature*, 491(7422):51–55, 2012.

81. Z. Gong, T. Hueckel, G.-R. Yi, and S. Sacanna. Patchy particles made by colloidal fusion. *Nature*, 550(7675):234–238, 2017.
82. J. S. Oh, G.-R. Yi, D. J. Pine, et al. Photo-printing of faceted DNA patchy particles. *Proc. Natl. Acad. Sci. U.S.A.*, 117(20):10645–10653, 2020.
83. D. J. Kraft, J. Hilhorst, M. A. P. Heinen, M. J. Hoogenraad, B. Luijges, and W. K. Kegel. Patchy polymer colloids with tunable anisotropy dimensions. *JPCB*, 2011.
84. L. Feng, L.-L. Pontani, R. Dreyfus, P. Chaikin, and J. Brujic. Specificity, flexibility and valence of DNA bonds guide emulsion architecture. *Soft Matter*, 9(41):9816–9823, 2013.
85. I. Chakraborty, V. Meester, C. Van Der Wel, and D. J. Kraft. Colloidal joints with designed motion range and tunable joint flexibility. *Nanoscale*, 9(23):7814–7821, 2017. doi:10.1039/c6nr08069c.
86. T. Witten Jr and L. M. Sander. Diffusion-limited aggregation, a kinetic critical phenomenon. *Phys. Rev. Lett.*, 47(19):1400, 1981.
87. M. Kolb and J. Rémi. Chemically limited versus diffusion limited aggregation. *Journal de Physique Lettres*, 45(20):977–981, 1984.
88. Ved1123. Time reversible flow demonstration in a taylor-couette system, 2018. URL commons.wikimedia.org/wiki/File:Time_reversible_flow_demonstration_in_a_Taylor-Couette_system.png.
89. H. I. Andersson and F. Jiang. Forces and torques on a prolate spheroid: Low-reynolds-number and attack angle effects. *Acta Mechanica*, 230(2):431–447, 2019.
90. G. Bagheri and C. Bonadonna. On the drag of freely falling non-spherical particles. *Powder Technology*, 301:526–544, 2016.
91. Y. Han, A. M. Alsayed, M. Nobili, J. Zhang, T. C. Lubensky, and A. G. Yodh. Brownian motion of an ellipsoid. *Science*, 314(5799):626–630, 2006. doi:10.1126/science.1130146.
92. M. Von Smoluchowski. Zur kinetischen theorie der brownschen molekularbewegung und der suspensionen. *Annalen der Physik*, 326(14):756–780, 1906.
93. G. Stokes. On the effect of internal friction of fluids on the motion of pendulums. In *Transactions of the Cambridge Philosophical Society*, volume 9, chapter X, section IV, equation 126, page 51. Cambridge Philosophical Society, 1856.
94. V. Totlani and M. Chinnan. Effect of stabilizer levels and storage conditions on texture and viscosity of peanut butter. *Peanut Science*, 34(1):1–9, 2007.
95. A. Meunier. Friction coefficient of rod-like chains of spheres at very low Reynolds numbers. II. Numerical simulations. *Journal de Physique II*, 4(4):561–566, 1994. doi:10.1051/jp2:1994141.
96. K. Zahn, R. Lenke, and G. Maret. Friction coefficient of rod-like chains of spheres at very low Reynolds numbers. I. Experiment. *Journal de Physique II*, 4(4):555–560, 1994. doi:10.1051/jp2:1994146.
97. A. Chakrabarty, A. Konya, F. Wang, J. V. Selinger, K. Sun, and Q. H. Wei. Brownian motion of arbitrarily shaped particles in two dimensions. *Langmuir*, 30(46):13844–13853, 2014. doi:10.1021/la5037053.
98. L. Koenigs, M. Lisicki, and E. Lauga. The non-Gaussian tops and tails of diffusing boomerangs. *Soft Matter*, 13(16):2977–2982, 2017. doi:10.1039/c6sm02649d.
99. A. V. Butenko, E. Mogilko, L. Amitai, B. Pokroy, and E. Slutskin. Coiled to diffuse: Brownian motion of a helical bacterium. *Langmuir*, 28(36):12941–12947, 2012. doi:10.1021/la302056j.
100. I. Serdyuk, N. Zaccai, and J. Zaccai. *Methods in Molecular Biophysics: Structure, Dynamics, Function*. Cambridge University Press, 2007.

101. J. Garcia de la Torre. Hydrodynamics of segmentally flexible macromolecules. *Eur. Biophys. J.*, 5:307–322, 1994.
102. L. Gregory, K. G. Davis, B. Sheth, J. Boyd, R. Jefferis, C. Nave, and D. R. Burton. The solution conformations of the subclasses of human IgG deduced from sedimentation and small angle X-ray scattering studies. *Molecular Immunology*, 24(8):821–829, 1987. doi:10.1016/0161-5890(87)90184-2.
103. J. Yguerabide, H. F. Epstein, and L. Stryer. Segmental flexibility in an antibody molecule. *J. Mol. Biol.*, 51(3):573–590, 1970. doi:10.1016/0022-2836(70)90009-4.
104. P. Illien, T. Adeleke-Larodo, and R. Golestanian. Diffusion of an enzyme: The role of fluctuation-induced hydrodynamic coupling. *EPL*, 119(4):40002, 2017. doi:10.1209/0295-5075/119/40002.
105. W. A. Wegener. Center of Diffusion of Flexible Macromolecules. *Macromolecules*, 18(12): 2522–2530, 1985. doi:10.1021/ma00154a029.
106. W. A. Wegener. Bead models of segmentally flexible macromolecules. *J. Chem. Phys.*, 76 (12):6425–6430, 1982.
107. S. C. Harvey, P. Mellado, and J. G. De La Torre. Hydrodynamic resistance and diffusion coefficients of segmentally flexible macromolecules with two subunits. *J. Chem. Phys.*, 78 (4):2081–2090, 1983. doi:10.1063/1.444917.
108. S. C. Harvey. Transport properties of particles with segmental flexibility. I. Hydrodynamic resistance and diffusion coefficients of a freely hinged particle. *Biopolymers*, 18(5):1081–1104, 1979. doi:10.1002/bip.1979.360180506.
109. S. Mornet, O. Lambert, E. Duguet, and A. Brisson. The formation of supported lipid bilayers on silica nanoparticles revealed by cryoelectron microscopy. *Nano Lett.*, 5(2): 281–285, 2005.
110. S. A. Van Der Meulen and M. E. Leunissen. Solid colloids with surface-mobile DNA linkers. *J. Am. Chem. Soc.*, 135(40):15129–15134, 2013.
111. S. A. J. Van Der Meulen, G. V. Dubacheva, M. Dogterom, R. P. Richter, and M. E. Leunissen. Quartz crystal microbalance with dissipation monitoring and spectroscopic ellipsometry measurements of the phospholipid bilayer anchoring stability and kinetics of hydrophobically modified DNA oligonucleotides. *Langmuir*, 30(22):6525–6533, 2014.
112. S. A. J. Van der Meulen, G. Helms, and M. Dogterom. Solid colloids with surface-mobile linkers. *J. Phys.: Condens. Matter*, 27(23):233101, 2015.
113. A. L. Troutier and C. Ladavière. An overview of lipid membrane supported by colloidal particles. *Adv. Colloid Interface Sci.*, 133(1):1–21, 2007.
114. C. B. Carlson, P. Mowery, R. M. Owen, E. C. Dykhuizen, and L. L. Kiessling. Selective tumor cell targeting using low-affinity, multivalent interactions. *ACS chemical biology*, 2 (2):119–127, 2007.
115. D. Papakostas, F. Rancan, W. Sterry, U. Blume-Peytavi, and A. Vogt. Nanoparticles in dermatology. *Archives of dermatological research*, 303(8):533, 2011.
116. J. Li, X. Wang, T. Zhang, C. Wang, Z. Huang, X. Luo, and Y. Deng. A review on phospholipids and their main applications in drug delivery systems. *Asian J. Pharm. Sci.*, 10(2): 81–98, 2015.
117. S. Savarala, S. Ahmed, M. A. Ilies, and S. L. Wunder. Formation and Colloidal Stability of DMPC Supported Lipid Bilayers on SiO₂ Nanobeads. *Langmuir*, 26(14):12081–12088, 2010.
118. E. T. Castellana and P. S. Cremer. Solid supported lipid bilayers: From biophysical studies to sensor design. *Surf. Sci. Rep.*, 61(10):429–444, 2006.

119. F. Smallenburg, L. Filion, and F. Sciortino. Erasing no-man's land by thermodynamically stabilizing the liquid-liquid transition in tetrahedral particles. *Nat. Phys.*, 10(9):653–657, 2014.
120. H. Hu, P. S. Ruiz, and R. Ni. Entropy Stabilizes Floppy Crystals of Mobile DNA-Coated Colloids. *Phys. Rev. Lett.*, 120(4):048003, 2018.
121. S. M. Douglas, I. Bachelet, and G. M. Church. A logic-gated nanorobot for targeted transport of molecular payloads. *Science*, 335(6070):831–834, 2012.
122. A. E. Marras, L. Zhou, H.-J. Su, and C. E. Castro. Programmable motion of DNA origami mechanisms. *Proc. Natl. Acad. Sci. U.S.A.*, 112(3):713–718, 2015.
123. G. J. Lavella, A. D. Jadhav, and M. M. Maharbiz. A synthetic chemomechanical machine driven by ligand–receptor bonding. *Nano Lett.*, 12(9):4983–4987, 2012.
124. Y. Liu, J. Cheng, S. Fan, H. Ge, T. Luo, L. Tang, B. Ji, C. Zhang, D. Cui, Y. Ke, et al. Modular reconfigurable DNA origami: From two-dimensional to three-dimensional structures. *Angew. Chem.*, 59(51):23277–23282, 2020.
125. F. Wei, T. Zhong, Z. Zhan, and L. Yao. Self-assembled micro-nanorobots: From assembly mechanisms to applications. *ChemNanoMat*, 2020.
126. R. Lanfranco, P. K. Jana, L. Tunesi, P. Cicuta, B. M. Mognetti, L. Di Michele, and G. Bruylants. Kinetics of nanoparticlemembrane adhesion mediated by multivalent interactions. *Langmuir*, 35(6):2002–2012, 2019.
127. S.-H. Kim, J.-M. Lim, S.-K. Lee, C.-J. Heo, and S.-M. Yang. Biofunctional colloids and their assemblies. *Soft Matter*, 6(4):1092–1110, 2010. doi:10.1039/b920611f.
128. G. Wu, H. Cho, D. A. Wood, A. D. Dinsmore, and S. Yang. Confined assemblies of colloidal particles with soft repulsive interactions. *J. Am. Chem. Soc.*, 139(14):50955101, 2017. doi:10.1021/jacs.6b12975.
129. M. Han, J. K. Whitmer, and E. Luijten. Dynamics and structure of colloidal aggregates under microchannel flow. *Soft Matter*, 15(4):744–751, 2019. doi:10.1039/c8sm01451e.
130. E. Yildiz-Ozturk and O. Yesil-Celiktas. Diffusion phenomena of cells and biomolecules in microfluidic devices. *Biomicrofluidics*, 9(5):052606, 2015.
131. H. Serna, E. G. Noya, and W. T. Goźdz. The influence of confinement on the structure of colloidal systems with competing interactions. *Soft Matter*, 16:718, 2020.
132. P. Yang, A. H. Rizvi, B. Messer, B. F. Chmelka, G. M. Whitesides, and G. D. Stucky. Patterning porous oxides within microchannel networks. *Adv. Mat.*, 13(6):427–431, 2001.
133. R. Mondal and M. G. Basavaraj. Patterning of colloids into spirals via confined drying. *Soft Matter*, 15(16):3753–3761, 2020.
134. H.-J. Wu and M. A. Bevan. Direct measurement of single and ensemble average particle-surface potential energy profiles. *Langmuir*, 21(4):12441254, 2005.
135. H. Lorentz. *Adv. Theor. Phys.*, 1:23–33, 1907.
136. H. Faxen. Fredholm integral equations of hydrodynamics of liquids i. *Ark. Mat., Astron. Fys.*, 18:29–32, 1924.
137. H. Brenner. The slow motion of a sphere through a viscous fluid towards a plane surface. *Chem. Eng. Sci.*, 16:242–251, 1961.
138. A. J. Goldman, R. G. Cox, and H. Brenner. Slow viscous motion of a sphere parallel to a plane wall. I. Motion through a quiescent fluid. *Chem. Eng. Sci.*, 22:637–651, 1967.
139. N. A. Frej and D. C. Prieve. Hindered diffusion of a single sphere very near a wall in a nonuniform force field. *J. Chem. Phys.*, 98:7552, 1993. doi:10.1063/1.464695.
140. P. Sharma, S. Ghosh, and S. Bhattacharya. A high-precision study of hindered diffusion near a wall. *Appl. Phys. Lett.*, 97:104101, 2010. doi:10.1063/1.3486123.

141. S. A. Rogers, M. Lisicki, B. Cichocki, J. K. G. Dhont, and P. R. Lang. Rotational diffusion of spherical colloids close to a wall. *Phys. Rev. Lett.*, 109:098305, 2012. doi:10.1103/PhysRevLett.109.098305.
142. K. Huang and I. Szlufarska. Effect of interfaces on the nearby Brownian motion. *Nat. Commun.*, 6:8558, 2015. doi:10.1038/ncomms9558.
143. L. Lobry and N. Ostrowsky. Diffusion of Brownian particles trapped between two walls: Theory and dynamic-light-scattering measurements. *Phys. Rev. B*, 53:12050, 1996. doi:10.1103/PhysRevB.53.12050.
144. B. Lin, J. Yu, and S. A. Rice. Direct measurements of constrained Brownian motion of an isolated sphere between two walls. *Phys. Rev. E*, 62:3909, 2000. doi:10.1103/PhysRevE.62.3909.
145. E. R. Dufresne, D. Altman, and D. G. Grier. Brownian dynamics of a sphere between parallel walls. *Europhys. Lett.*, 53:264–270, 2001. doi:10.1209/epl/i2001-00147-6.
146. T. Benesch, S. Yiacoumi, and C. Tsouris. Brownian motion in confinement. *Phys. Rev. E*, 68:021401, 2003. doi:10.1103/PhysRevE.68.021401.
147. S. B. K Zembrzycki and T. A. Kowalewski. Analysis of wall effect on the process of diffusion of nanoparticles in a microchannel. *J. Phys.: Conf. Ser.*, 392:012014, 2012. doi:10.1088/1742-6596/392/1/012014.
148. S. L. Dettmer, S. Pagliara, K. Misiunas, and U. F. Keyser. Anisotropic diffusion of spherical particles in closely confining microchannels. *Phys. Rev. E*, 89:062305, 2014. doi:10.1103/PhysRevE.89.062305.
149. V. N. Michailidou, G. Petekidis, J. W. Swan, and J. F. Brady. Dynamics of concentrated hard-sphere colloids near a wall. *Phys. Rev. Lett.*, 102:068302, 2009. doi:10.1103/PhysRevLett.102.068302.
150. R. Pesché and G. Nägele. Stokesian dynamics study of quasi-two-dimensional suspensions confined between two parallel walls. *Phys. Rev. E*, 62:5432, 2000. doi:10.1103/PhysRevE.62.5432.
151. B. Eral, J. Oh, H. T. van den Ende, F. G. Mugele, and M. H. Duits. Anisotropic and hindered diffusion of colloidal particles in a closed cylinder. *Langmuir*, 22:16722–16729, 2010. doi:10.1021/la102273n.
152. B. Cui, H. Diamant, and B. Lin. Screened hydrodynamic interaction in a narrow channel. *Phys. Rev. Lett.*, 89:188302, 2002. doi:10.1103/PhysRevLett.89.188302.
153. J. Wang, C. F. Mbah, T. Przybilla, S. Englisch, E. Spiecker, M. Engel, and N. Vogel. Free energy landscape of colloidal clusters in spherical confinement. *ACS Nano*, 13:9005–9015, 2019. doi:10.1021/acsnano.9b03039.
154. J. T. Padding and W. J. Briels. Translational and rotational friction on a colloidal rod near a wall. *J. Chem. Phys.*, 132(5):054511, 2010. doi:10.1063/1.3308649.
155. T. Adeleke-Larodo, P. Illien, and R. Golestanian. Fluctuation-induced hydrodynamic coupling in an asymmetric, anisotropic dumbbell. *Eur. Phys. J. E*, 42(39):054511, 2019. doi:10.1140/epje/i2019-11799-5.
156. M. Haghghi, M. N. Tahir, W. Tremel, H.-J. Butt, and W. Steffen. Translational and rotational diffusion of gold nanorods near a wall. *J. Chem. Phys.*, 139:064710, 2013. doi:10.1063/1.4817405.
157. M. Lisicki, B. Cichocki, and E. Wajnryb. Near-wall diffusion tensor of an axisymmetric colloidal particle. *J. Chem. Phys.*, 145:034904, 2016. doi:10.1063/1.4958727.
158. S. Delong, F. B. Usabiaga, and A. Donev. Brownian dynamics of confined rigid bodies. *J. Chem. Phys.*, 143:144107, 2015. doi:10.1063/1.4932062.

159. M. X. Fernandes and J. G. de la Torre. Brownian dynamics simulation of rigid particles of arbitrary shape in external fields. *Biophys J*, 83(6):3039–3048, 2002. doi:10.1016/S0006-3495(02)75309-5.
160. M. D. Carbajal-Tinoco, R. Lopez-Fernandez, and J. L. Arauz-Lara. Asymmetry in colloidal diffusion near a rigid wall. *Phys. Rev. Lett.*, 99:138303, 2007. doi:10.1103/PhysRevLett.99.138303.
161. J. Leach, H. Mushfique, S. Keen, R. D. Leonardo, G. Ruocco, J. M. Cooper, and M. J. Padgett. Comparison of faxéns correction for a microsphere translating or rotating near a surface. *Phys. Rev. E*, 79:026301, 2009. doi:10.1103/PhysRevE.79.026301.
162. S. Jeney, B. Lukic, J. A. Kraus, T. Franosch, and L. Forró. Anisotropic memory effects in confined colloidal diffusion. *Phys. Rev. Lett.*, 100:240604, 2008. doi:10.1103/PhysRevLett.100.240604.
163. E. Schäffer, S. F. Nørrelykke, and J. Howard. Surface forces and drag coefficients of microspheres near a plane surface measured with optical tweezers. *Langmuir*, 23:3654, 2007. doi:10.1021/la0622368.
164. N. Garnier and N. Ostrowsky. Brownian dynamics in a confined geometry. experiments and numerical simulations. *J. Phys. II*, 1:1221–1232, 1991. doi:10.1051/jp2:1991129.
165. P. Holmqvist, J. K. G. Dhont, and P. R. Lang. Colloidal dynamics near a wall studied by evanescent wave light scattering: Experimental and theoretical improvements and methodological limitations. *J. Chem. Phys.*, 126:044707, 2007. doi:10.1063/1.2431175.
166. T. Watarai and T. Iwai. Direct observation of submicron Brownian particles at a solidliquid interface by extremely low coherence dynamic light scattering. *Appl. Phys. Express*, 7: 032502, 2014. doi:10.7567/APEX.7.032502.
167. M. I. M. Feitosa and O. N. Mesquita. Wall-drag effect on diffusion of colloidal particles near surfaces: A photon correlation study. *Phys. Rev. A*, 44:6677, 1991. doi:10.1103/PhysRevA.44.6677.
168. K. H. Lan, N. Ostrowsky, and D. Sornette. Brownian dynamics close to a wall studied by photon correlation spectroscopy from an evanescent wave. *Phys. Rev. Lett.*, 57:17, 1986. doi:10.1103/PhysRevLett.57.17.
169. P. Holmqvist, J. K. G. Dhont, and P. R. Lang. Anisotropy of Brownian motion caused only by hydrodynamic interaction with a wall. *Phys. Rev. E*, 74:021402, 2006. doi:10.1103/PhysRevE.74.021402.
170. M. Lisicki, B. Cichocki, S. A. Rogers, J. K. G. Dhont, and P. R. Lang. Translational and rotational near-wall diffusion of spherical colloids studied by evanescent wave scattering. *Soft Matter*, 10:4312–4323, 2014. doi:10.1039/C4SM00148F.
171. Y. Kazoe and M. Yoda. Measurements of the near-wall hindered diffusion of colloidal particles in the presence of an electric field. *Appl. Phys. Lett.*, 99:124104, 2011. doi:10.1063/1.3643136.
172. D. Prieve. Measurement of colloidal forces with TIRM. *Adv. Colloid Interface Sci.*, 82: 93–125, 1999. doi:10.1016/S0001-8686(99)00012-3.
173. G. Volpe, T. Brettschneider, L. Helden, and C. Bechinger. Novel perspectives for the application of total internal reflection microscopy. *Opt. Express*, 17:23975–23985, 2009. doi:10.1364/OE.17.023975.
174. L. Liu, A. Woolf, A. W. Rodriguez, and F. Capasso. Absolute position total internal reflection microscopy with an optical tweezer. *Proc. Natl. Acad. Sci. U.S.A.*, 111:E5609–E5615, 2014. doi:10.1073/pnas.1422178112.

175. S.-H. Lee, Y. Roichman, G.-R. Yi, S.-H. Kim, S.-M. Yang, A. van Blaaderen, P. van Oostrum, and D. G. Grier. Characterizing and tracking single colloidal particles with video holographic microscopy. *Opt. Express*, 15:18275–18282, 2007. doi:10.1364/OE.15.018275.
176. L. Dixon, F. C. Cheong, and D. G. Grier. Holographic deconvolution microscopy for high-resolution particle tracking. *Opt. Express*, 19:16410, 2011. doi:10.1364/OE.19.016410.
177. D. S. Bolintineanu, G. S. Grest, J. B. Lechman, F. Pierce, S. J. Plimpton, and P. R. Schunk. Particle dynamics modeling methods for colloid suspensions. *Comput. Part. Mech.*, 1: 321356, 2014.
178. C. Middleton, M. D. Hannel, A. D. Hollingsworth, D. J. Pine, and D. G. Grier. Optimizing the synthesis of monodisperse colloidal spheres using holographic particle characterization. *Langmuir*, 35:66026609, 2019. doi:10.1021/acs.langmuir.9b00012.
179. J. Garcia-Sucerquia, W. Xu, S. K. Jericho, P. Klages, M. H. Jericho, and H. J. Kreuzer. Digital in-line holographic microscopy. *Appl. Opt.*, 45:836, 2006. doi:10.1364/AO.45.000836.
180. P. Marquet, B. Rappaz, P. J. Magistretti, E. Cuche, Y. Emery, T. Colomb, and C. Depeursinge. Digital holographic microscopy: a noninvasive contrast imaging technique allowing quantitative visualization of living cells with subwavelength axial accuracy. *Opt. Lett.*, 30:468–470, 2005. doi:10.1364/OL.30.000468.
181. C. B. Giuliano, R. Zhang, and L. G. Wilson. Digital inline holographic microscopy (DIHM) of weakly-scattering subjects. *J. Vis. Exp.*, 84:e50488, 2014. doi:10.3791/50488.
182. W. Xu, M. H. Jericho, I. A. Meinertzhausen, and H. J. Kreuzer. Digital in-line holography for biological applications. *Proc. Natl. Acad. Sci. U.S.A.*, 98:11301–11305, 2001. doi:10.1073/pnas.191361398.
183. L. E. Altman and D. G. Grier. Catch: Characterizing and tracking colloids holographically using deep neural networks. *J. Phys. Chem. B*, 124:16021610, 2020. doi:10.1021/acs.jpcb.9b10463.
184. J. Zhang, P. Zhan, Z. Wang, W. Zhang, and N. Ming. Preparation of monodisperse silica particles with controllable size and shape. *J. Mater. Res.*, 18(3):649–653, 2003. doi:10.1557/JMR.2003.0085.
185. W. S. Rasband. ImageJ, 1997-2018. URL imagej.nih.gov/ij/.
186. S. Barkley, T. G. Dimiduk, J. Fung, D. M. Kaz, V. N. Manoharan, R. McGorty, R. W. Perry, and A. Wang. Holographic microscopy with Python and HoloPy, 2018.
187. D. W. Mackowski and M. I. Mishchenko. Calculation of the T matrix and the scattering matrix for ensembles of spheres. *J. Opt. Soc. Am. A*, 13(11):2266–2278, 1996.
188. D. Allan, C. van der Wel, N. Keim, T. A. Caswell, D. Wieker, R. W. Verweij, C. Reid, Thierry, L. Grueter, K. Ramos, apiszcz, zoeith, R. W. Perry, F. Boulogne, P. Sinha, pfigliozzi, N. Bruot, L. Uieda, J. Katins, H. Mary, and A. Ahmadi. soft-matter/trackpy: Trackpy v0.4.2, 2019. URL doi.org/10.5281/zenodo.3492186.
189. J. C. Crocker and D. G. Grier. Methods of digital video microscopy for colloidal studies. *J. Colloid Interface Sci.*, 179(1):298–310, 1996. doi:10.1006/jcis.1996.0217.
190. Y. Gu and D. Li. The zeta-potential of glass surface in contact with aqueous solutions. *J. Colloid Interface Sci.*, 226(2):328 – 339, 2000. doi:10.1006/jcis.2000.6827.
191. S. Ketsetzi, J. de Graaf, and D. J. Kraft. Diffusion-based height analysis reveals robust microswimmer-wall separation. *Phys. Rev. Lett.*, (125):238001, 2020. doi:10.1103/PhysRevLett.125.238001.
192. H. Faxén. *Einwirkung der Gefäßwände auf den Widerstand gegen die Bewegung einer kleinen Kugel in einer zähen Flüssigkeit*. Uppsala Universitet, 1921.

193. I. Brouwer, A. Giniatullina, N. Laurens, J. R. T. V. Weering, D. Bald, G. J. L. Wuite, and A. J. Groffen. Direct quantitative detection of Doc2b-induced hemifusion in optically trapped membranes. *Nat. Commun.*, 6:8387, 2015.
194. A. McMullen, M. Holmes-Cerfon, F. Sciortino, A. Y. Grosberg, and J. Brujic. Freely Jointed Polymers Made of Droplets. *Phys. Rev. Lett.*, 121(13):138002, 2018. doi:10.1103/PhysRevLett.121.138002.
195. C. L. Phillips, E. Jankowski, B. J. Krishnatreya, K. V. Edmond, S. Sacanna, D. G. Grier, D. J. Pine, and S. C. Glotzer. Digital colloids: reconfigurable clusters as high information density elements. *Soft Matter*, 10(38):7468–7479, 2014.
196. N. B. Schade, M. C. Holmes-Cerfon, E. R. Chen, D. Aronzon, J. W. Collins, J. A. Fan, F. Capasso, and V. N. Manoharan. Tetrahedral Colloidal Clusters from Random Parking of Bidisperse Spheres. *Phys. Rev. Lett.*, 110(14):148303, 2013.
197. T. Sugimoto and K. Sakata. Preparation of monodisperse pseudocubic α -Fe₂O₃ particles from condensed ferric hydroxide gel. *J. Colloid Interface Sci.*, 152(2):587–590, 1992.
198. V. Meester and D. J. Kraft. Spherical, Dimpled, and Crumpled Hybrid Colloids with Tunable Surface Morphology. *Langmuir*, 32(41):10668–10677, 2016.
199. C. Van der Wel, R. K. Bhan, R. W. Verweij, H. C. Frijters, Z. Gong, A. D. Hollingsworth, S. Sacanna, and D. J. Kraft. Preparation of colloidal organosilica spheres through spontaneous emulsification. *Langmuir*, 33(33):8174–8180, 2017.
200. R. P. Doherty and D. J. Kraft. One-pot surfactant-free synthesis of organosilica colloids with various surface functional groups. In preparation.
201. J. Appel, S. Akerboom, R. G. Fokkink, and J. Sprakel. Facile One-Step Synthesis of Monodisperse Micron-Sized Latex Particles with Highly Carboxylated Surfaces. *Macromol. Rapid Commun.*, 34(16):1284–1288, 2013. doi:10.1002/marc.201300422.
202. C. Van Der Wel, A. Vahid, A. Šarić, T. Idema, D. Heinrich, and D. J. Kraft. Lipid membrane-mediated attraction between curvature inducing objects. *Sci. Rep.*, 6(1):1–10, 2016. doi:10.1038/srep32825.
203. D. Axelrod, D. Koppel, J. Schlessinger, E. Elson, and W. Webb. Mobility measurement by analysis of fluorescence photobleaching recovery kinetics. *Biophys. J.*, 16(9):1055–1069, 1976.
204. J. D. Hunter. Matplotlib: A 2D graphics environment. *Comput. Sci. Eng.*, 9(3):90–95, 2007. doi:10.1109/MCSE.2007.55.
205. C. M. Van der Wel. *Lipid Mediated Colloidal Interactions*. PhD thesis, Casimir PhD Series, 2017.
206. R. Sarfati, J. Bławdziewicz, and E. R. Dufresne. Maximum likelihood estimations of force and mobility from single short Brownian trajectories. *Soft Matter*, 13(11):2174–2180, 2017.
207. D. Foreman-Mackey, D. W. Hogg, D. Lang, and J. Goodman. emcee : The MCMC Hammer. *Proc. Natl. Acad. Sci. U.S.A.*, 125(925):306–312, 2013. doi:10.1086/670067.
208. E. Sackmann. Supported Membranes: Scientific and Practical Applications. *Science*, 271 (5245):43–48, 1996. doi:10.1126/science.271.5245.43.
209. I. Gözen and A. Jesorka. Instrumental methods to characterize molecular phospholipid films on solid supports. *Anal. Chem.*, 84(2):822–838, 2012.
210. R. Machá and M. Hof. Lipid diffusion in planar membranes investigated by fluorescence correlation spectroscopy. *Biochim. Biophys. Acta*, 1798(7):1377–1391, 2010. doi:10.1016/j.bbamem.2010.02.014.

211. S. I. R. Castillo, S. Ouhajji, S. Fokker, B. H. Erné, C. T. W. M. Schneijdenberg, D. M. E. Thies-Weesie, and A. P. Philipse. Silica cubes with tunable coating thickness and porosity: From hematite filled silica boxes to hollow silica bubbles. *Microporous Mesoporous Mater.*, 195:75–86, 2014. doi:10.1016/j.micromeso.2014.03.047.
212. T. Cha, A. Guo, and X.-Y. Zhu. Formation of supported phospholipid bilayers on molecular surfaces: Role of surface charge density and electrostatic interaction. *Biophys. J.*, 90(4): 1270–1274, 2006.
213. P. G. De Gennes. Polymers at an interface; a simplified view. *Adv. Colloid Interface Sci.*, 27 (3-4):189–209, 1987.
214. S. Upadhyayula, T. Quinata, S. Bishop, S. Gupta, N. R. Johnson, B. Bahmani, K. Bozhilov, J. Stubbs, P. Jreij, P. Nallagatla, and V. I. Vullev. Coatings of polyethylene glycol for suppressing adhesion between solid microspheres and flat surfaces. *Langmuir*, 28(11): 5059–69, 2012.
215. C. Van der Wel, N. Bossert, Q. J. Mank, M. G. Winter, D. Heinrich, and D. J. Kraft. Surfactant-free colloidal particles with specific binding affinity. *Langmuir*, 33(38):9803–9810, 2017.
216. C. Van Der Wel, G. L. Van De Stolpe, R. W. Verweij, and D. J. Kraft. Micrometer-sized TPM emulsion droplets with surface-mobile binding groups. *J. Phys. Condens. Matter*, 30 (9), 2018.
217. C. A. Naumann, O. Prucker, T. Lehmann, J. Rühe, W. Knoll, and C. W. Frank. The polymer-supported phospholipid bilayer: Tethering as a new approach to substrate- membrane stabilization. *Biomacromolecules*, 3(1):27–35, 2002.
218. M. Tanaka and E. Sackmann. Polymer-supported membranes as models of the cell surface. *Nature*, 437(7059):656–663, 2005. doi:10.1038/nature04164.
219. M. Tanaka. Polymer-Supported Membranes: Physical Models of Cell Surfaces. *MRS Bulletin*, 31:513–520, 2006.
220. M. L. Wagner and L. K. Tamm. Tethered polymer-supported planar lipid bilayers for reconstitution of integral membrane proteins: silane-polyethyleneglycol-lipid as a cushion and covalent linker. *Biophys. J.*, 79(3):1400–1414, 2000. doi:10.1016/S0006-3495(00)76392-2.
221. M. A. Deverall, S. Garg, K. Ludtke, R. Jordan, J. Ruhe, and C. A. Naumann. Transbilayer coupling of obstructed lipid diffusion in polymer-tethered phospholipid bilayers. *Soft Matter*, 4:1899–1908, 2008.
222. R. Lipowsky. Bending of membranes by anchored polymers. *Europhys. Lett.*, 30(4):197, 1995.
223. P. L. Biancaniello, J. C. Crocker, D. A. Hammer, and V. T. Milam. DNA-mediated phase behavior of microsphere suspensions. *Langmuir*, 23(5):2688–2693, 2007.
224. G. Nordlund, R. Lonneborg, and P. Brzezinski. Formation of Supported Lipid Bilayers on Silica Particles Studied Using Flow Cytometry. *Langmuir*, 25(8):4601–4606, 2009.
225. A. Tan, A. Ziegler, B. Steinbauer, and J. Seelig. Thermodynamics of sodium dodecyl sulfate partitioning into lipid membranes. *Biophys. J.*, 83(3):1547–1556, 2002.
226. T. Drobek, N. D. Spencer, and M. Heuberger. Compressing PEG brushes. *Macromolecules*, 38(12):5254–5259, 2005.
227. F. Meng, G. H. M. Engbers, and J. Feijen. Polyethylene glycol-grafted polystyrene particles. *J. Biomed. Mater. Res. Part A*, 70(1):49–58, 2004.
228. O. Garbuzenko, Y. Barenholz, and A. Prieve. Effect of grafted PEG on liposome size and on compressibility and packing of lipid bilayer. *Chem. Phys. Lipids*, 135(2):117–129, 2005.

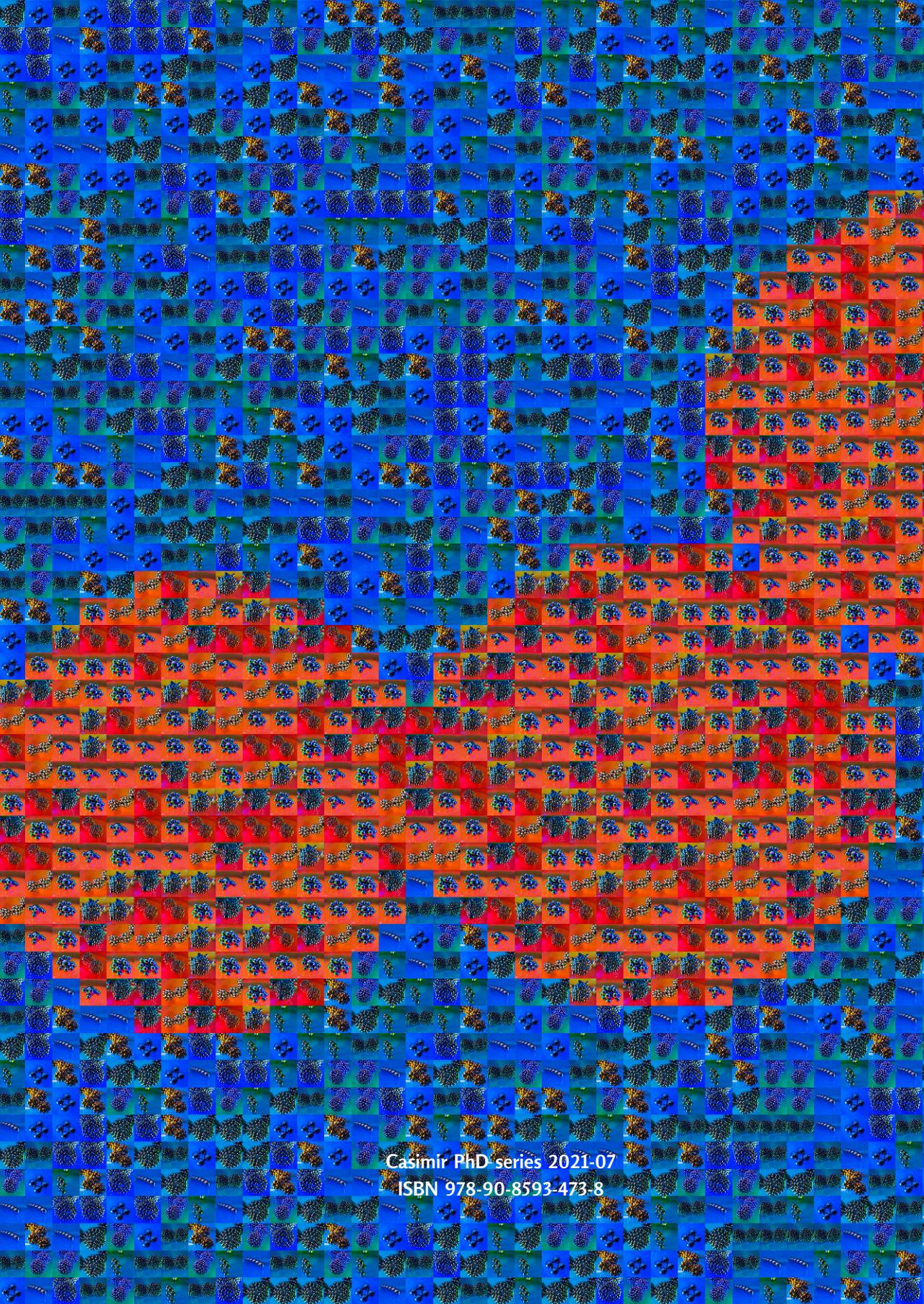
229. S. Angioletti-Uberti, P. Varilly, B. M. Mognetti, and D. Frenkel. Mobile linkers on DNA-coated colloids: Valency without patches. *Phys. Rev. Lett.*, 113(12):128303, 2014.
230. R. Lipowsky. Spontaneous tubulation of membranes and vesicles reveals membrane tension generated by spontaneous curvature. *Faraday Discuss.*, 161:305–331, 2013.
231. S. I. Nakano, M. Fujimoto, H. Hara, and N. Sugimoto. Nucleic acid duplex stability: Influence of base composition on cation effects. *Nucleic Acids Res.*, 27(14):2957–2965, 1999.
232. M. Marsh and A. Helenius. Virus entry: open sesame. *Cell*, 124(4):729–740, 2006.
233. C.-H. Heldin. Dimerization of cell surface receptors in signal transduction. *Cell*, 80(2):213–223, 1995.
234. M. Mammen, S.-K. Choi, and G. M. Whitesides. Polyvalent interactions in biological systems: implications for design and use of multivalent ligands and inhibitors. *Angew. Chem.*, 37(20):2754–2794, 1998.
235. L. L. Kiessling, J. E. Gestwicki, and L. E. Strong. Synthetic multivalent ligands as probes of signal transduction. *Angew. Chem.*, 45(15):2348–2368, 2006.
236. D. Nykypanchuk, M. M. Maye, D. Van Der Lelie, and O. Gang. DNA-based approach for interparticle interaction control. *Langmuir*, 23(11):6305–6314, 2007.
237. R. Dreyfus, M. E. Leunissen, R. Sha, A. V. Tkachenko, N. C. Seeman, D. J. Pine, and P. M. Chaikin. Simple quantitative model for the reversible association of DNA coated colloids. *Phys. Rev. Lett.*, 102(4):048301, 2009.
238. F. J. Martinez-Veracoechea and D. Frenkel. Designing super selectivity in multivalent nano-particle binding. *Proc. Natl. Acad. Sci. U.S.A.*, 108(27):10963–10968, 2011.
239. P. Varilly, S. Angioletti-Uberti, B. M. Mognetti, and D. Frenkel. A general theory of DNA-mediated and other valence-limited colloidal interactions. *J. Chem. Phys.*, 137(9):094108, 2012.
240. B. M. Mognetti, P. Cicuta, and L. Di Michele. Programmable interactions with biomimetic DNA linkers at fluid membranes and interfaces. *Rep. Prog. Phys.*, 82(11):116601, 2019.
241. J. S. Oh, M. He, G.-R. Yi, and D. J. Pine. High-density DNA coatings on carboxylated colloids by DMTMM- and azide-mediated coupling reactions. *Langmuir*, 36(13):3583–3589, 2020.
242. M. Rinaldin, R. W. Verweij, I. Chakraborty, and D. J. Kraft. Colloid supported lipid bilayers for self-assembly. *Soft Matter*, 15:1345–1360, 2019. doi:10.1039/C8SM01661E.
243. S. J. Bachmann, M. Petitzon, and B. M. Mognetti. Bond formation kinetics affects self-assembly directed by ligand–receptor interactions. *Soft Matter*, 12(47):9585–9592, 2016.
244. R. W. Verweij, P. G. Moerman, N. E. G. Ligthart, L. P. P. Huijnen, J. Groenewold, W. K. Kegel, A. van Blaaderen, and D. J. Kraft. Flexibility-induced effects in the Brownian motion of colloidal trimers. *Phys. Rev. Res.*, 2(3):033136, 2020. doi:10.1103/physrevresearch.2.033136.
245. E. B. Wilson. Probable inference, the law of succession, and statistical inference. *J. Am. Stat. Assoc.*, 22(158):209–212, 1927.
246. M. Newville, R. Otten, A. Nelson, A. Ingargiola, T. Stensitzki, D. Allan, A. Fox, et al. lmfit/lmfit-py 1.0.1, 2020.
247. P. Varilly, S. Angioletti-Uberti, et al. DNACC, 2015. URL github.com/patvarilly/DNACC.
248. S. Angioletti-Uberti, P. Varilly, B. M. Mognetti, A. V. Tkachenko, and D. Frenkel. Communication: a simple analytical formula for the free energy of ligand-receptor-mediated interactions. *J. Chem. Phys.*, (138):021102, 2013.
249. W. A. Kibbe. Oligocalc: an online oligonucleotide properties calculator. *Nucleic Acids Res.*, 35:W43–W46, 2007.

250. I. Chakraborty, D. J. Pearce, R. W. Verweij, S. C. Matysik, L. Giomi, and D. J. Kraft. Self-assembly dynamics of reconfigurable colloidal molecules. In preparation.
251. P. Mellado, A. Iniesta, F. Diaz, and J. Garcia de la Torre. Diffusion coefficients of segmentally flexible macromolecules with two subunits: A study of broken rods. *Biopolymers*, 27: 1771–1786, 1988.
252. S. Ishino, T. Yamagami, M. Kitamura, N. Kodera, T. Mori, S. Sugiyama, T. Ando, N. Goda, T. Tenno, H. Hiroaki, and Y. Ishino. Multiple interactions of the intrinsically disordered region between the helicase and nuclease domains of the archaeal Hef protein. *J. Biol. Chem.*, 289(31):21627–21639, 2014. doi:10.1074/jbc.M114.554998.
253. D. Burton. Structure and function of antibodies. *New Compr. Biochem.*, 17:1–50, 1987.
254. G. Barbato, M. Ikura, L. E. Kay, A. Bax, and R. W. Pastor. Backbone Dynamics of Calmodulin Studied by ¹⁵N Relaxation Using Inverse Detected Two-Dimensional NMR Spectroscopy: The Central Helix Is Flexible. *Biochemistry*, 31(23):5269–5278, 1992. doi:10.1021/bi00138a005.
255. D. E. Koshland. The KeyLock Theory and the Induced Fit Theory, 1995.
256. K. Nagasaka and H. Yamakawa. Dynamics of weakly bending rods: A trumbbell model. *J. Chem. Phys.*, 83(12):6480–6488, 1985. doi:10.1063/1.449548.
257. M. Fixman. Inclusion of Hydrodynamic Interaction in Polymer Dynamical Simulations. *Macromolecules*, 14(6):1710–1717, 1981. doi:10.1021/ma50007a019.
258. A. Z. Akcasu. Comments on the Diffusion Coefficient and First Cumulant. *Macromolecules*, 15(5):1321–1324, 1982. doi:10.1021/ma00233a021.
259. C. Riedel, R. Gabizon, C. A. Wilson, K. Hamadani, K. Tsekouras, S. Marqusee, S. Pressé, and C. Bustamante. The heat released during catalytic turnover enhances the diffusion of an enzyme. *Nature*, 517(7533):227–230, 2015. doi:10.1038/nature14043.
260. S. Sengupta, K. K. Dey, H. S. Muddana, T. Tabouillot, M. E. Ibele, P. J. Butler, and A. Sen. Enzyme molecules as nanomotors. *J. Am. Chem. Soc.*, 135(4):1406–1414, 2013. doi:10.1021/ja3091615.
261. L. Rundqvist, J. Ådén, T. Sparrman, M. Wallgren, U. Olsson, and M. Wolf-Watz. Noncooperative folding of subdomains in adenylate kinase. *Biochemistry*, 48(9):1911–1927, 2009. doi:10.1021/bi8018042.
262. D. Roitman. The elastic trumbbell model for dynamics of stiff chains. Rotational Dynamics of Small and Macromolecules. *Lecture Notes in Physics*, 293, 2005.
263. J. García de la Torre and B. Carrasco. Hydrodynamic properties of rigid macromolecules composed of ellipsoidal and cylindrical subunits. *Biopolymers*, 63(3):163–167, 2002. doi:10.1002/bip.10013.
264. A. Iniesta, F. G. Diaz, and J. García de la Torre. Transport properties of rigid bent-rod macromolecules and of semiflexible broken rods in the rigid-body treatment. Analysis of the flexibility of myosin rod. *Biophys. J.*, 54(2):269–275, 1988. doi:10.1016/S0006-3495(88)82956-4.
265. Y. Zhang, X. He, R. Zhuo, R. Sha, J. Brujic, N. C. Seeman, and P. M. Chaikin. Multivalent, multiflavored droplets by design. *Proc. Natl. Acad. Sci. U.S.A.*, 115(37):9086–9091, 2018. doi:10.1073/pnas.1718511115.
266. H. R. Vutukuri, A. F. Demirörs, B. Peng, P. D. Van Oostrum, A. Imhof, and A. Van Blaaderen. Colloidal analogues of charged and uncharged polymer chains with tunable stiffness. *Angew. Chem.*, 51(45):11249–11253, 2012. doi:10.1002/anie.201202592.
267. F. Montanarella, D. Urbonas, L. Chadwick, P. G. Moerman, P. J. Baesjou, R. F. Mahrt, A. Van Blaaderen, T. Stöferle, and D. Vanmaekelbergh. Lasing Supraparticles Self-Assembled from Nanocrystals. *ACS Nano*, 12(12):12788–12794, 2018. doi:10.1021/acsnano.8b07896.

268. F. G. Díaz and J. G. de la Torre. Viscoelastic properties of semiflexible macromolecules in solution: Brownian dynamics simulation of a trumbbell model. *Macromolecules*, 27(19): 5371–5376, 1994. doi:10.1021/ma00097a017.
269. B. Cichocki, M. Rubin, A. Niedzwiecka, and P. Szymczak. Diffusion coefficients of elastic macromolecules. *J. Fluid Mech.*, 878:R3, 2019. doi:10.1017/jfm.2019.652.
270. B. Carrasco and J. G. De La Torre. Improved hydrodynamic interaction in macromolecular bead models. *J. Chem. Phys.*, 111(10):4817–4826, 1999. doi:10.1063/1.479743.
271. V. Bloomfield, W. O. Dalton, and K. E. Van Holde. Frictional coefficients of multisubunit structures. I. Theory. *Biopolymers*, 5(2):135–148, 1967. doi:10.1002/bip.1967.360050202.
272. D. P. Filson and V. A. Bloomfield. Shell Model Calculations of Rotational Diffusion Coefficients. *Biochemistry*, 6(6):1650–1658, 1967. doi:10.1021/bi00858a011.
273. D. L. Ermak and J. A. McCammon. Brownian dynamics with hydrodynamic interactions. *J. Chem. Phys.*, 69(4):1352–1360, 1978. doi:10.1063/1.436761.
274. T. Lukka, J. Stewart, et al. FreeWRL: an X3D/VRML open source viewer for Windows, Linux, OSX and Android., 2017. URL <http://freewrl.sourceforge.net>.
275. H. Yamakawa. Transport properties of polymer chains in dilute solution: hydrodynamic interaction. *J. Chem. Phys.*, 53(1):436–443, 1970. doi:10.1063/1.1673799.
276. B. Carrasco and J. G. De La Torre. Improved hydrodynamic interaction in macromolecular bead models. *J. Chem. Phys.*, 111(10):4817–4826, 1999. doi:10.1063/1.479743.
277. G. D. Phillips. Translational drag coefficients of assemblies of spheres with higher-order hydrodynamic interactions. *J. Chem. Phys.*, 81(9):4046–4052, 1984. doi:10.1063/1.448148.
278. A. Ortega and J. García de la Torre. Equivalent radii and ratios of radii from solution properties as indicators of macromolecular conformation, shape, and flexibility. *Biomacromolecules*, 8(8):2464–2475, 2007. doi:10.1021/bm700473f.
279. P. Sharma, S. Ghosh, and S. Bhattacharya. *Nat. Phys.*, 4:960, 2008.
280. W. Weare, S. Reed, M. Warner, and J. Hutchison. Improved synthesis of small phosphine-stabilized gold nanoparticles. *J. Am. Chem. Soc.*, (122):12890–12891, 2000.
281. J. Piella, N. Bastús, and V. Puntes. Size-controlled synthesis of sub-10-nm citrate-stabilized gold nanoparticles and related optical properties. *Chem. Mater.*, (28):1066–1075, 2016.
282. H. J. Dyson and P. E. Wright. Intrinsically unstructured proteins and their functions. *Nat. Rev. Mol. Cell Biol.*, 6(3):197–208, 2005.
283. L. Makowski, D. J. Rodi, S. Mandava, D. D. Minh, D. B. Gore, and R. F. Fischetti. Molecular crowding inhibits intramolecular breathing motions in proteins. *J. Mol. Biol.*, 375(2):529–546, 2008.
284. A. Einstein. Über die von der molekularkinetischen Theorie der Wärme geforderte Bewegung von in ruhenden Flüssigkeiten suspendierten Teilchen. *Annalen der Physik*, 322(8): 549–560, 1905. doi:10.1002/andp.19053220806.
285. A. M. Mihut, B. Stenqvist, M. Lund, P. Schurtenberger, and J. J. Crassous. Assembling oppositely charged lock and key responsive colloids: A mesoscale analog of adaptive chemistry. *Sci. Adv.*, 3(9):e1700321, 2017.
286. J. Riseman and J. G. Kirkwood. The intrinsic viscosity, translational and rotatory diffusion constants of rod-like macromolecules in solution. *J. Chem. Phys.*, 18(4):512–516, 1950.
287. J. García De La Torre and B. Carrasco. Hydrodynamic properties of rigid macromolecules composed of ellipsoidal and cylindrical subunits. *Biopolymers*, 63(3):163–167, 2002. doi:10.1002/bip.10013.
288. R. R. Schmidt, J. H. Cifre, and J. G. de la Torre. Translational diffusion coefficients of macromolecules. *Eur. Phys. J. E*, 35(12):130, 2012.

289. T. Yamada, T. Yoshizaki, and H. Yamakawa. Transport coefficients of helical wormlike chains. 5. Translational diffusion coefficient of the touched-bead model and its application to oligo- and polystyrenes. *Macromolecules*, 25(1):377–383, 1992.
290. B. Sprinkle, E. B. van der Wee, Y. Luo, M. M. Driscoll, and A. Donev. Driven dynamics in dense suspensions of microrollers. *Soft Matter*, 16:7982–8001, 2020. doi:10.1039/D0SM00879F.
291. S. Delong, F. Balboa, B. Delmotte, B. Sprinkle, A. Donev, et al. Rigid Multiblobs in half-space (v. 427d241), 2020. URL github.com/stochasticHydroTools/RigidMultiblobsWall.
292. E. Wajnryb, K. A. Mizerski, P. J. Zuk, and P. Szymczak. Generalization of the Rotne-Prager-Yamakawa mobility and shear disturbance tensors. *J. Fluid Mech.*, 731:R3, 2013. doi:10.1017/jfm.2013.402.
293. F. Balboa Usabiaga, B. Delmotte, and A. Donev. Brownian dynamics of confined suspensions of active microrollers. *J. Chem. Phys.*, 146(13):134104, 2017. doi:10.1063/1.4979494.
294. P. A. Wiggins and P. C. Nelson. Generalized theory of semiflexible polymers. *Phys. Rev. E*, 73(3):031906, 2006.
295. P.-G. De Gennes and P.-G. Gennes. *Scaling concepts in polymer physics*. Cornell university press, 1979.
296. X. Wang and K. Wang. Calculation of the Flory exponent using the Renormalization theory. *Polym. J.*, 27:515518, 1995.
297. M. Doi, S. F. Edwards, and S. F. Edwards. *The theory of polymer dynamics*, volume 73. oxford university press, 1988.
298. J. P. Valleau. Distribution of end-to-end length of an excluded-volume chain. *J. Chem. Phys.*, 104(8):3071–3074, 1996.
299. M. Bishop and J. Clarke. Investigation of the end-to-end distance distribution function for random and self-avoiding walks in two and three dimensions. *J. Chem. Phys.*, 94(5):3936–3942, 1991.
300. J. M. Victor and D. Lhuillier. The gyration radius distribution of twodimensional polymer chains in a good solvent. *J. Chem. Phys.*, 92(2):1362–1364, 1990. doi:10.1063/1.458147.
301. J. Agudo-Canalejo and R. Golestanian. Diffusion and steady state distributions of flexible chemotactic enzymes. *Eur. Phys. J. ST*, 229(17):2791–2806, 2020.
302. T. McLeish. Polymers without beginning or end. *Science*, 297(5589):2005–2006, 2002. doi:10.1126/science.1076810.
303. M. Kapnistos, M. Lang, D. Vlassopoulos, W. Pyckhout-Hintzen, D. Richter, D. Cho, T. Chang, and M. Rubinstein. Unexpected power-law stress relaxation of entangled ring polymers. *Nat. Mat.*, 7(12):997–1002, 2008.
304. S. Liu, S. Murata, and I. Kawamata. DNA ring motif with flexible joints. *Micromachines*, 11(11):987, 2020.
305. J. D. Halverson, G. S. Grest, A. Y. Grosberg, and K. Kremer. Rheology of ring polymer melts: From linear contaminants to ring-linear blends. *Phys. Rev. Lett.*, 108(3):038301, 2012.
306. J. D. Halverson, J. Smrek, K. Kremer, and A. Y. Grosberg. From a melt of rings to chromosome territories: the role of topological constraints in genome folding. *Rep. Prog. Phys.*, 77(2):022601, 2014. doi:10.1088/0034-4885/77/2/022601.
307. J. D. Halverson, W. B. Lee, G. S. Grest, A. Y. Grosberg, and K. Kremer. Molecular dynamics simulation study of nonconcatenated ring polymers in a melt. II. Dynamics. *J. Chem. Phys.*, 134(20):204905, 2011. doi:10.1063/1.3587138.

308. E. W. Weisstein. Domino graph. URL mathworld.wolfram.com/DominoGraph.html. Accessed Jan. 2020.
309. J. C. Maxwell. On the calculation of the equilibrium and stiffness of frames. *London Edinburgh Philos. Mag. J. Sci.*, 27(182):294–299, 1864.
310. Y. Han, A. Alsayed, M. Nobili, and A. G. Yodh. Quasi-two-dimensional diffusion of single ellipsoids: Aspect ratio and confinement effects. *Phys. Rev. E*, 80(1):011403, 2009.
311. Z. Zeravcic, V. N. Manoharan, and M. P. Brenner. Size limits of self-assembled colloidal structures made using specific interactions. *Proc. Natl. Acad. Sci. U.S.A.*, 111(45):15918–15923, 2014.
312. E. V. Shevchenko, D. V. Talapin, C. B. Murray, and S. O'Brien. Structural characterization of self-assembled multifunctional binary nanoparticle superlattices. *J. Am. Chem. Soc.*, 128(11):3620–3637, 2006.
313. J.-H. Lee, J. P. Singer, and E. L. Thomas. Micro-/nanostructured mechanical metamaterials. *Adv. Mater.*, 24(36):4782–4810, 2012.
314. Y.-S. Cho, G.-R. Yi, S.-H. Kim, D. J. Pine, and S.-M. Yang. Colloidal clusters of microspheres from water-in-oil emulsions. *Chem. Mater.*, 17(20):5006–5013, 2005.
315. D. J. Kraft, J. Groenewold, and W. K. Kegel. Colloidal molecules with well-controlled bond angles. *Soft Matter*, 5(20):3823–3826, 2009.
316. L. Feng, R. Dreyfus, R. Sha, N. C. Seeman, and P. M. Chaikin. DNA patchy particles. *Adv. Mater.*, 25(20):2779–2783, 2013.
317. V. Meester and D. J. Kraft. Spherical, dimpled, and crumpled hybrid colloids with tunable surface morphology. *Langmuir*, 32(41):10668–10677, 2016.
318. W. K. Kegel, D. Breed, M. Elsesser, and D. J. Pine. Formation of anisotropic polymer colloids by disparate relaxation times. *Langmuir*, 22(17):7135–7136, 2006.
319. M. Rinaldin. *On the geometry of demixing: A study of lipid phase separation on curved surfaces*. PhD thesis, Casimir PhD Series, 2019.
320. P. A. Beales, J. Nam, and T. K. Vanderlick. Specific adhesion between DNA-functionalized “Janus” vesicles: size-limited clusters. *Soft Matter*, 7(5):1747–1755, 2011.
321. M. Wang, Z. Liu, and W. Zhan. Janus liposomes: Gel-assisted formation and bioaffinity-directed clustering. *Langmuir*, 34(25):7509–7518, 2018.
322. P. Swinkels, S. Stuij, Z. Gong, H. Jonas, B. van der Linden, P. Bolhuis, S. Sacanna, S. Woutersen, and P. Schall. Revealing pseudorotation and ring-opening reactions in colloidal organic molecules. Under review, 2021.
323. S. Sacanna, W. T. Irvine, P. M. Chaikin, and D. J. Pine. Lock and key colloids. *Nature*, 464 (7288):575–578, 2010.
324. K. Braun, A. Hanewald, and T. A. Vilgis. Milk emulsions: Structure and stability. *Foods*, 8(10):483, 2019.
325. A. Papadopoulou, J. Laucks, and S. Tibbits. From self-assembly to evolutionary structures. *Architectural Design*, 87(4):28–37, 2017.
326. Trazyanderson. Velcro photomicrograph, 2018. URL commons.wikimedia.org/wiki/File:Velcro_photomicrograph.jpg.



Casimir PhD series 2021-07

ISBN 978-90-8593-473-8