

 Very Important Paper

Tuning the Transglycosylation Reaction of a GH11 Xylanase by a Delicate Enhancement of its Thumb Flexibility

Kim Marneth,^[a] Hans van den Elst,^[b] Anneloes Cramer-Blok,^[a] Jeroen Codee,^[a] Hermen S. Overkleeft,^[b] Johannes M. F. G. Aerts,^[c] Marcellus Ubbink,^{*[a]} and Fredj Ben Bdira^{*[a]}

Glycoside hydrolases (GHs) are attractive tools for multiple biotechnological applications. In conjunction with their hydrolytic function, GHs can perform transglycosylation under specific conditions. In nature, oligosaccharide synthesis is performed by glycosyltransferases (GTs); however, the industrial use of GTs is limited by their instability in solution. A key difference between GTs and GHs is the flexibility of their binding site architecture. We have used the xylanase from *Bacillus circulans* (BCX) to study the interplay between active-site flexibility and transglycosylation. Residues of the BCX “thumb” were substituted to increase the flexibility of the enzyme binding site. Replacement of the highly conserved residue P116 with glycine shifted the balance of the BCX enzymatic reaction toward transglycosylation. The effects of this point mutation on the structure and dynamics of BCX were investigated by NMR spectroscopy. The P116G mutation induces subtle changes in the configuration of the thumb and enhances the millisecond dynamics of the active site. Based on our findings, we propose the remodelling of the GH enzymes glycon site flexibility as a strategy to improve the transglycosylation efficiency of these biotechnologically important catalysts.

the growth of beneficial intestinal microflora.^[1] Oligosaccharide derivatives have multiple therapeutic and cosmetic uses.^[2] To fulfil the market demand, significant efforts were dedicated to the development of synthesis strategies of oligosaccharides with high yield and low-cost. Synthesis by chemical methods is tedious because it requires multiple protection and deprotection steps and gives low yields.^[3] A greener approach is to use enzymes to obtain a high yield of oligosaccharides with defined stereochemistry. Enzymatic synthesis can be achieved by glycosyltransferase (GT); however, these enzymes are difficult to produce and unstable in solution, which makes them unsuitable for industrial use.^[4] An alternative is to use the ubiquitous family of glycoside hydrolases (GH).^[5]

Retaining GH hydrolyse the glycosidic linkage using the “Koshland” double displacement mechanism with a catalytic nucleophile and acid/base for catalysis.^[6] The glycosyl-enzyme adduct is a common intermediate in hydrolysis and transglycosylation reactions. During the deglycosylation step, a water molecule or an alcohol, respectively, act as nucleophilic acceptors in the positive subsites of the enzyme, after being activated by the acid/base residue of the catalytic dyad (Figure 1A).^[7] Therefore, the GH enzymes can perform carbohydrate synthesis and the balance between the two reactions is influenced by pH and temperature, as well as the availability of suitable acceptors like methanol and ethanol.^[8] However, the simultaneous presence of both activities of the enzymes represents a downside of this synthesis strategy. This bottleneck of the enzymatic carbohydrate synthesis was surmounted by the engineering of the GH enzymes to reduce or eliminate their hydrolytic activity.^[9] Multiple GH engineering strategies were adopted, including the substitution of the catalytic residues and the use of fluorinated substrates.^[8,10] Introduction of mutations in the positive subsites has improved the binding affinity of acceptors molecules and thus the transglycosylation efficiency of the GH enzymes.^[11]

Although GH and GT enzymes share similar active site architectures and topologies, they differ in flexibility. GT enzymes tend to be flexible, which is thought to be a requirement for the synthetic reaction.^[7b,12] On the contrary, the rigidity of the GH appears to be necessary for their hydrolytic function.^[13] In an attempt to gain new insight into the role of flexibility in transglycosylation, we used xylanase from *Bacillus circulans* (BCX). BCX is a globular GH11 member exhibiting a conserved β -jelly-roll fold.^[14] The enzyme is remarkably rigid due to an extensive hydrogen bond network, restricting dynamics on the pico- to millisecond NMR timescale.^[13b,15] We


Introduction


Oligosaccharides and their conjugates have found a wide range of pharmaceutical and biotechnological applications. For instance, in the food industry, oligosaccharides are used as prebiotics to improve the nutrition value of food by stimulating

[a] K. Marneth, A. Cramer-Blok, Prof. Dr. J. Codee, Prof. Dr. M. Ubbink, Dr. F. Ben Bdira
Department of Macromolecular Biochemistry, Leiden Institute of Chemistry
Einsteinweg 55, 2333 CC Leiden (The Netherlands)
E-mail: m.ubbink@chem.leidenuniv.nl
fredjbdira@gmail.com

[b] H. van den Elst, Prof. Dr. H. S. Overkleeft
Department of Bio-organic Synthesis, Leiden Institute of Chemistry
Einsteinweg 55, 2333 CC Leiden (The Netherlands)

[c] Prof. Dr. J. M. F. G. Aerts
Department of Medical Biochemistry, Leiden Institute of Chemistry
Einsteinweg 55, 2333 CC Leiden (The Netherlands)

 Supporting information for this article is available on the WWW under <https://doi.org/10.1002/cbic.202000856>

 © 2021 The Authors. ChemBioChem published by Wiley-VCH GmbH. This is an open access article under the terms of the Creative Commons Attribution Non-Commercial NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

of this loop are in contact with the substrate at the -1 and -2 subsites of the enzyme binding cleft (Figure 1B).^[14] The thumb region displays a well-structured classical hairpin, containing a type I β -turn (residues 111–126 in BCX) with six internal hydrogen bonds (Figure 1B). Multiple molecular dynamics simulations on different members of GH11 xylanase suggested the flexibility of this region and a putative role in substrate binding and product dissociation.^[18b,19] However, in BCX, this region appears to be rigid with restricted dynamics on a wide range of NMR timescales.^[13b,15] This difference in the thumb flexibility between GH11 members appears to be due to the difference in length and sequence of the thumb, although the PSIXG sequence motif of the thumb tip is highly conserved.^[17b] Thus, we aimed to introduce mutations on different positions of the thumb to enhance the flexibility of BCX binding cleft at the glycon site and probe their effects on the enzyme-catalysed reactions. The first mutant was designed by substituting the residue P116 with a glycine. The almost complete conservation of this residue between all members of GH11 indicates its importance for the specific configuration of the thumb and the narrowing of the enzyme binding cleft for better selectivity towards non-decorated substrates.^[17a] The backbone carbonyl of P116 accepts a hydrogen bond from the O3 of the sugar substrate at the -1 subsite (Figure 1B). By replacing P116 with a glycine it is possible that the rigidity within the thumb loop will be diminished, increasing flexibility. The hydrogen-bonding pattern of the thumb is interrupted by N114 leading to a sharp bend in the thumb structure (Figure 1B). Additionally, it has been suggested that this residue represents a hinge point of the thumb in the GH11 family member, BsXynA.^[19] The configuration of the thumb is stabilized by a salt bridge between R73 of the palm and D119 of the thumb tip (Figure 1B). Thus, to trigger conformational freedom of the thumb, we also substituted N114 and R73 with glycine and alanine, respectively. In BsXynA another mutation, D11F, allowed the capture in the crystalline state of the thumb in an open conformation with a relocation of its tip by more than 15 Å.^[20] Thus, we reasoned that the substitution of D11 with a glycine might also promote flexibility by excluding the bulky side chain of the amino acid from the BCX aglycon site. Finally, T126 is a potential hinge point that links the thumb to the palm.^[19] Substitution of T126 with proline in BsXynA improved the hydrolysis activity, possibly by increasing the rigidity of the thumb.^[20] Thus, as a control variant, we substituted T126 with a proline in BCX.

Effects of the point mutations

To test whether the introduced mutations in BCX influenced the thermal stability, the melting temperature (T_m) was measured with a thermofluor assay.^[21] BCX WT has a T_m of 58 °C. The mutations reduced the T_m by 1 to 7 °C (Figure 2A). The most significant destabilization effect was observed for mutations P116G and R73 A, possibly due an increase in local flexibility. The melting temperature of mutant T126P is similar to the wild type, thus suggesting a minor effect on the protein

rigidity. All the mutants have a T_m above 50 °C, so their hydrolytic activity was measured at 40 °C, the optimum temperature for WT BCX, using an end-point enzymatic assay with the artificial substrate 4-methylumbelliferone-xylobioside (4MU-X2). Most mutants show a reduction of hydrolytic activity of 20 to 30%. However, for BCX P116G it is reduced by 90% (Figure 2A). These findings agree with previous reports in which the replacement of residues in BsXynA thumb by either glycine or proline, to modulate the flexibility, reduced the activity of the enzyme.^[20] Nevertheless, the activity enhancement observed for BsXynA T126P is not observed in BCX.

Although the trans-glycosylation activity has been reported for many GH families under specific conditions,^[8a] it remains unclear if it also occurs for GH11 members in the presence of oligosaccharides acceptors. Previously, transglycosylation was not observed in BCX at high concentrations of its natural and artificial substrates.^[22] Nevertheless, a minor transglycosylation activity was recently reported for an endoxylanase (Taxyl11) from *Trichoderma asperellum* ND-1.^[23] This activity was also described for the GH11 xylanase from *Thermobacillus xylanilyticus* in the presence of polyphenolic acceptors.^[11d] Polyphenyl β -oligoxylsides were synthesized by a GH11 extracted from *Bacillus* sp. KT12 culture filtrate. The balance between the hydrolysis and transglycosylation reactions of the enzyme was substantially altered by acidic pH and metallic ions.^[24]

To test whether BCX is capable of transglycosylation in the presence of oligosaccharides acceptors, the enzyme was incubated with 50 mM of *p*-nitrophenyl- β -D-xylobioside (PNP-X2) at 30% DMSO for 30 min at 30 °C. The oligosaccharides of the reaction mixture were analysed by TLC (Figure S1A in the Supporting Information), showing several bands that suggest the formation of products with different degree of polymerization (DP). Separation by ion exchange HPLC revealed that the reaction products comprised the X1, X2 and X3 oligosaccharides (Figure 2B). Although X2 might result from the hydrolysis of the glycosidic bond in PNP-X2, X3 and X1 can only occur from a transglycosylation reaction and/or the hydrolysis of newly formed products with $DP \geq 3$. The observed transglycosylation reaction of BCX might be due to the presence of DMSO, which is known to reduce the activity of water molecules, leading to the accumulation of the enzyme-glycosyl intermediate.^[8a] This promotes the transfer of oligosaccharides units from the glycon site to the acceptor in the aglycon site of the enzyme. Therefore, we have tested the transglycosylation activity of BCX with xylohexaose (X6) at 20 mM concentration in water (without DMSO). HPLC analysis identified oligosaccharides with DP7, DP8 and DP9 in the reaction mixture (Figure 2C). Also, shorter oligosaccharides (DP1, DP2 and DP3) are present due to the simultaneous hydrolysis activity of the enzyme. After establishing the presence of transglycosylation activity in BCX WT, we have tested the effect of the thumb mutations on this activity. Similar to their effects on the hydrolysis reaction, most of the thumb mutants did not have a major impact on the transglycosylation reaction of BCX (Figure 2B). However, for mutant P116G, an accumulation of longer oligosaccharides was observed, ranging from three to four xylose units when PNP-X2 is used as a substrate (Figure 2B). Additionally, the amount of

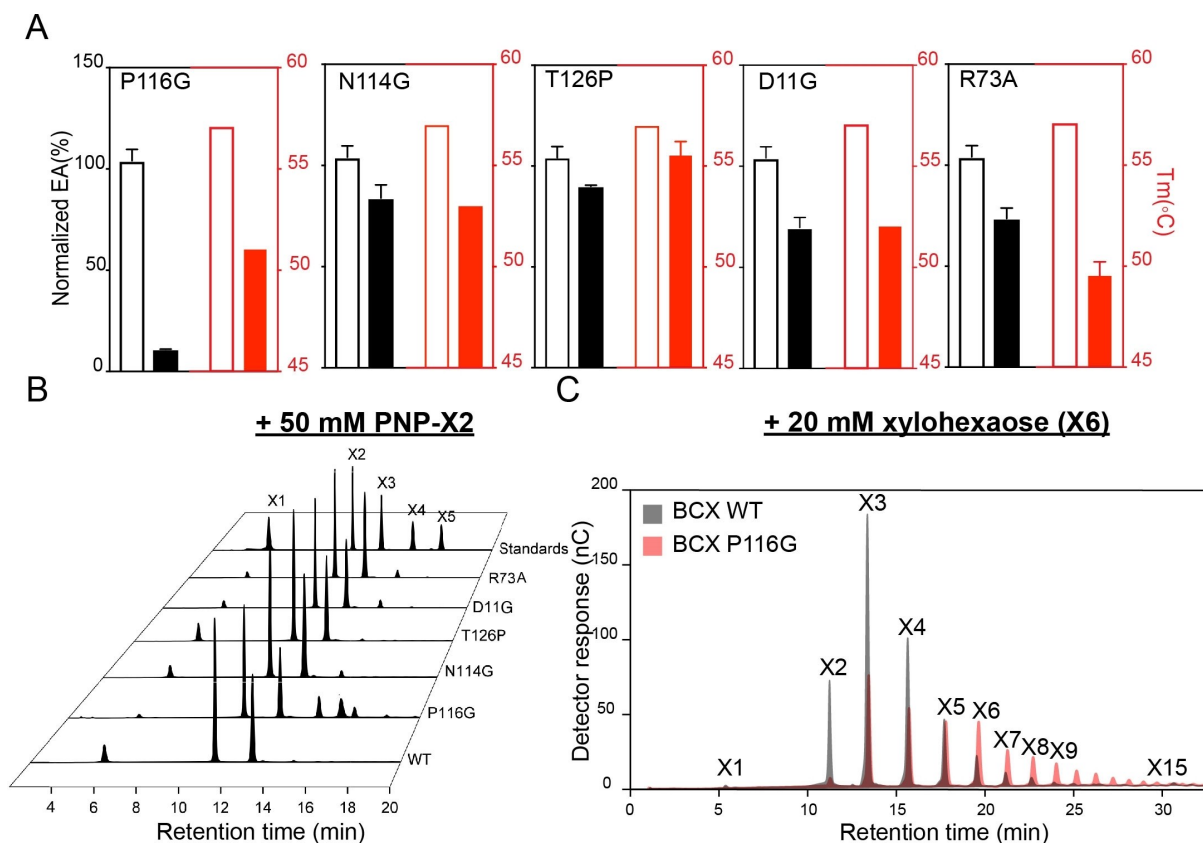


Figure 2. Effects of the thumb point mutations on BCX activity and stability. A) The end-point enzymatic assay with 4MU-X2 as a substrate (black) and melting temperatures (red) are depicted in percentage of BCX WT (open bars) vs. the mutants (filled bars); EA: relative enzymatic activity. B) Ion-exchange chromatograms of the transglycosylation products of the BCX variants. The reaction was conducted in the presence of 50 mM PNPX2 with 30% DMSO. A mixture of xylosides with different DP was used as standard. C) An overlay of the ion-exchange chromatograms of the BCX WT (grey) and the BCX P116G (red) transglycosylation products. The reaction was performed in the presence of 20 mM X6 in water (without DMSO) incubated for 30 min at 30 °C.

X1 and X2 in the reaction mixture, which are products of hydrolysis, is reduced. The HPLC analysis of the reaction mixture of BCX P116G in the presence of 20 mM X6 revealed a significant shift in the enzyme activity towards the synthesis of longer oligosaccharides (up to DP15), compared to the wild-type enzyme (Figure 2C). Clearly, the hydrolytic activity of this variant is reduced.

Effect of the P116G mutation on the protein structure and dynamics

The observed shift in activity specificity of BCX by the P116G substitution might be due to changes in the structure and dynamics of the enzyme. To investigate the effect of the point mutation on BCX structure, we recorded the $^1\text{H}, ^{15}\text{N}$ HSQC spectra of the wild type and the mutant at 30 °C and calculated the chemical shift perturbations (CSP). The P116G mutation results in small CSP localized in and around the thumb region with minor effects on residues of the fingers (Figure 3A). These CSP report on changes in the chemical environment of the backbone amides, which could be due to the proline substitution and/or subtle structural rearrangements. Pseudo-

contact shifts (PCS) are sensitive probes to structural changes within a protein. PCS of the BCX WT and the P116G mutant were obtained by introducing two cysteine residues (T109C/T111C)^[13b] for attachment of the paramagnetic tag CLANP-5-Yb³⁺ or the diamagnetic control tag CLANP-5-Lu³⁺.^[25] For both WT and mutant BCX more than 130PCS were obtained, by comparison of $^{15}\text{N}, ^1\text{H}$ HSQC spectra of paramagnetic and diamagnetic samples, which were used to determine the position, the size, and the orientation of the anisotropic component of magnetic susceptibility tensor ($\Delta\chi$) tensor. The data for WT BCX were fitted to the crystal structure with PDB ID 2bvj^[26] and the lanthanoid position was $\sim 8 \text{ \AA}$ from the double cysteines C α , in line with the previously reported distances (Figure 3A).^[25] The magnitudes of the axial and rhombic components of the tensor were determined to be $8.15 \pm 0.03 \times 10^{-32} \text{ m}^3$ and $2.36 \pm 0.04 \times 10^{-32} \text{ m}^3$, respectively. An excellent agreement between the experimental and the predicted PCS was obtained with a Q_s value [Eq. (S1)] of 0.024 (Figure S1B). These results obtained at 30 °C are in line with the reported ones at 20 °C.^[13b] Molecular dynamics (MD) simulations on BCX at different temperatures have suggested a temperature-dependent movement of the thumb region.^[27] A full opening of thumb was also captured in the crystalline state for the BsXynA

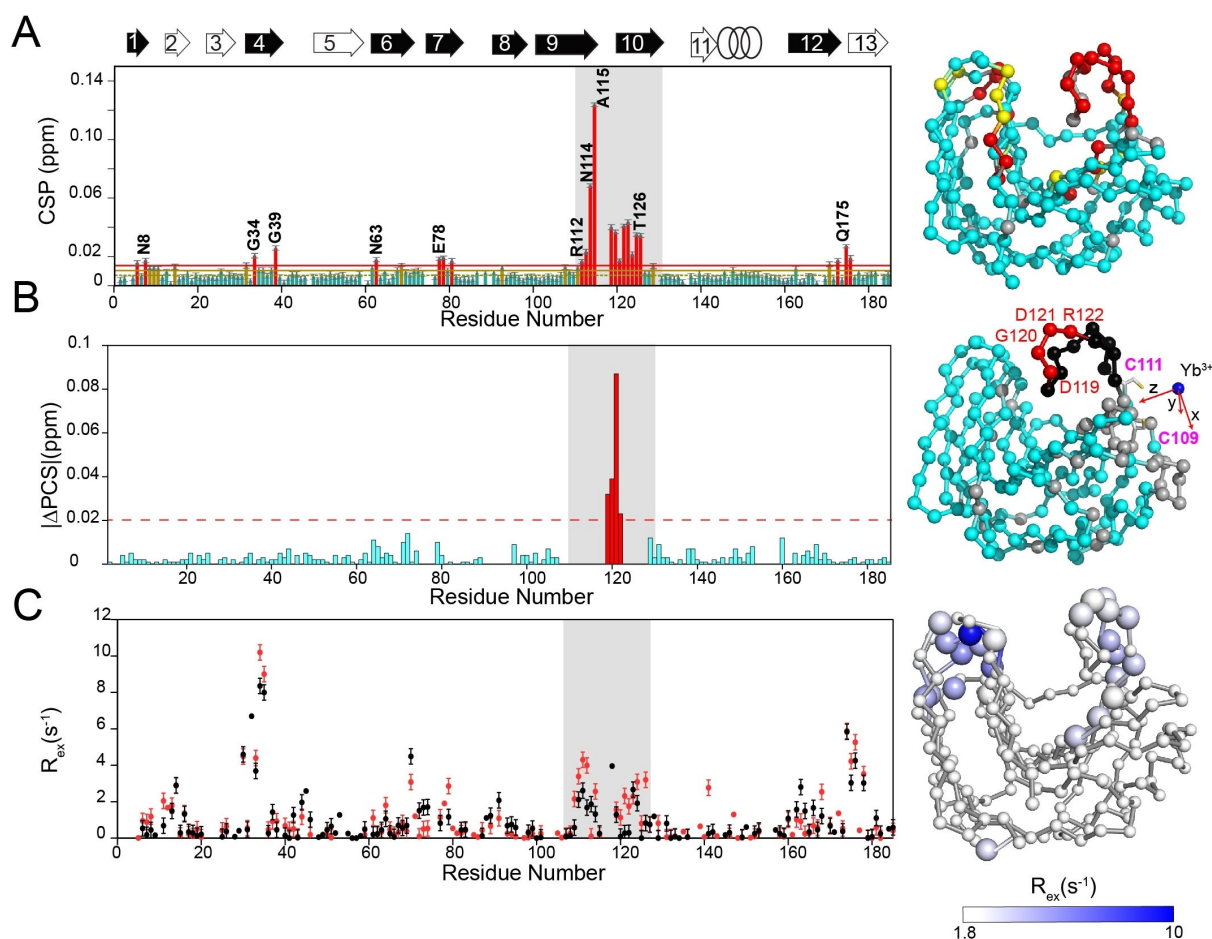


Figure 3. The effects of the P116G mutation on BCX structure and dynamics. A) Analysis of the weighted average CSP between BCX P116G and WT. Resonances with CSP more than one (yellow line) or two (red line) standard deviation(s) (SD) from the 10% trimmed mean (green dashed line) are labelled and shown by yellow, red, and green bars, respectively. Right: amide nitrogens with a CSP > 1 and 2 SD are shown in spheres on the BCX structure (PDB ID: 2BVV)^[26] and coloured in yellow and red, respectively. Nitrogens with no data are shown in grey. B) The $|\Delta\text{PCS}|$ between BCX P116G WT are plotted against residue number. $|\Delta\text{PCS}|$ values > 0.02 (red dashed line) are shown by red bars. Right: amide nitrogens of the residues with $|\Delta\text{PCS}| > 0.02$ are mapped on the BCX structure as red spheres and labelled. Nitrogens with not data in the thumb region and elsewhere are depicted in black and grey, respectively. The blue sphere indicates the position of the lanthanide ion (Yb^{3+}). The x -, y - and z -axes of the $\Delta\chi$ tensor are indicated by red arrows. The cysteine residues at which CLANP-5 was attached are shown as sticks and labelled. C) The experimental R_{ex} of BCX WT (black) and P116G (red) versus residue number. Right: the amide nitrogens of BCX P116G with $R_{\text{ex}} \geq 1.8 \text{ s}^{-1}$ are mapped onto the BCX structure, shown as large spheres and coloured with a white/blue gradient. Nitrogens without significant millisecond dynamics are shown as small spheres. Above the top panel, the BCX secondary structure is shown by black arrows for β -strands of sheet A and white ones for sheet B and the α -helix in rings. The thumb region is highlighted with a grey background in the panels.

D11F mutant.^[20] The occurrence of an open-closed motion of the thumb has also been proposed by several MD simulation studies on other GH11 members.^[17b,18b,19] However, the PCS data do not support such motion of the thumb in BCX and rather suggest that the conformation of the protein is the same at 20 and 30 °C (Figure S1B).^[13b] Similarly, it was found using PCS that in mimics of the structures of intermediates of the catalytic cycle no large changes in the backbone occurred either.^[13b] Thus, we suggest that a full opening in the thumb does not occur and is not required for the function of BCX.

For BCX P116G, several PCS of the amide nuclei of the thumb could not be determined due to line broadening caused by paramagnetic relaxation enhancement (PRE).^[28] The PCS of the thumb residues D119, G120, D121 and R122 could be measured, however (Figure 3B). The broadening suggests proximity in the spatial configuration of the mutated thumb

relative to the paramagnetic centre. The absolute values of the differences between the observed PCS of the BCX WT and P116G mutant ($|\Delta\text{PCS}|$) indicate a highly localized structural change in the thumb of the protein (Figures 3B and S1C). The rest of the backbone of the protein is unperturbed.

Structural motions in the millisecond timescale can influence substrate turnover by the enzyme in different ways. It can cause the exclusion of water molecules from the active site, optimize the position of either the substrate or the catalytic residues in the protein and assist the release of the product from the binding site.^[29] Therefore, to interrogate the effects of the P116G mutation on the millisecond dynamics of BCX, we performed a relaxation dispersion TROSY-CPMG experiment. Previously we have observed that BCX WT exhibits minor chemical exchange (R_{ex}) for some residues remote from the catalytic site.^[13b] The P116G mutation slightly enhanced the

millisecond chemical exchange of the enzyme binding cleft for residues of the thumb and fingers regions at the glycon site (Figure 3C).

In summary, these data indicate that the P116G mutation induces a structural change and somewhat enhanced millisecond dynamics localized to the thumb region at the glycon site of BCX. A conformational change in the loop may expose the hydrophobic interface between the thumb and the underlying palm, which could explain the large decrease in T_m .

Conclusion

GH enzymes are attractive tools in green chemistry for the synthesis of biotechnologically relevant oligosaccharides. Several approaches were used to improve their synthesis efficiency by focusing on the optimization of the reaction conditions (pH, T° , organic solvent, acceptors) and the enhancement of the acceptor affinity toward the enzymes aglycon sites. Although these strategies proved to be successful in many GH, the role of enzymes binding site dynamics and flexibility in transglycosylation remained unexplored.

In this report, we attempted to shed light on the interplay between fold flexibility and the trans-reaction using a GH11 xylanase from *B. circulans* (BCX) as a benchmark due to its distinctive fold rigidity. We found that the substitution of the conserved residue P116 of the thumb with a glycine induced subtle changes in the millisecond dynamics and conformation at the glycon site. The mutant also exhibits a tenfold reduction in hydrolysis activity, resulting in a shift in the balance between hydrolysis and transglycosylation activity toward the latter. It is possible that activation of the nucleophilic water molecule is less efficient, because of a slight displacement of the water molecule relative to the acid/base catalytic residue. The extensive network of ordered water molecules can transfer effects of the mutation throughout the active site. Alternatively, the glycosyl adduct, which is in direct contact with P116, might change position or conformation slightly, making the hydrolytic attack more difficult. Observations from the end-point enzymatic assay suggest that, with a xylose acceptor, the rate reduction might be less, resulting in a relatively larger transglycosylation and smaller hydrolysis activity. Nevertheless, the BCX P116G variant retains hydrolytic activity against its new synthesized products. Therefore, further investigations and protein design are required to eliminate this undesired reaction.

In conclusion, our study provides new insights into the possible role of fold flexibility in the trans-glycosylation reaction and propose the modulation of the rigid binding site characteristic of the GH enzymes as a strategy to engineer more efficient transglycosylation catalysts.

Conflict of Interest

The authors declare no conflict of interest.

Keywords: fold flexibility · glycosidases · NMR spectroscopy · transglycosylation

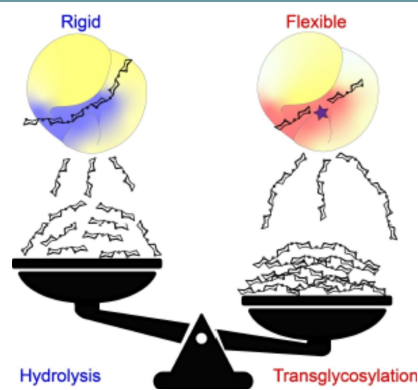
- [1] a) J. A. Linares-Pasten, A. Aronsson, E. Nordberg Karlsson, *Curr. Protein Pept. Sci.* **2018**, *19*, 48–67; b) J. Park, J. Yu, Y. Shin, H. Shin, S. Lee, K. Park, *J. Microbiol. Biotechnol.* **1992**, *20*, 237–242; c) T. Oku, S. Nakamura, *Pure Appl. Chem.* **2002**, *74*, 1253–1261.
- [2] a) W. von Rybinski, K. Hill, *Angew. Chem. Int. Ed. Engl.* **1998**, *37*, 1328–1345; b) L. Greffe, L. Bessueille, V. Bulone, H. Brumer, *Glycobiology* **2005**, *15*, 437–445; c) X. Vecino, J. Cruz, A. Moldes, L. Rodrigues, *Crit. Rev. Biotechnol.* **2017**, *37*, 911–923; d) C. Duarte, E. J. Gudiña, C. F. Lima, L. R. Rodrigues, *AMB express* **2014**, *4*, 40; e) E. J. Gudiña, E. C. Fernandes, J. A. Teixeira, L. R. Rodrigues, *RSC Adv.* **2015**, *5*, 90960–90968.
- [3] a) P. H. Seeberger, D. B. Werz, *Nature* **2007**, *446*, 1046–1051; b) D. B. Werz, *Methods Mol. Biol.* **2012**, 13–29.
- [4] P. Sears, C.-H. Wong, *Science* **2001**, *291*, 2344–2350.
- [5] a) L. F. Mackenzie, *J. Am. Chem. Soc.* **1998**, *120*, 5583–5585; b) D. H. Crout, G. Vic, *Curr. Opin. Chem. Biol.* **1998**, *2*, 98–111; c) R. M. Schmaltz, S. R. Hanson, C.-H. Wong, *Chem. Rev.* **2011**, *111*, 4259–4307; d) H.-Y. Feng, J. Drone, L. Hoffmann, V. Tran, C. Tellier, C. Rabiller, M. Dion, *J. Biol. Chem.* **2005**, *280*, 37088–37097.
- [6] D. Koshland Jr, *Biol. Rev.* **1953**, *28*, 416–436.
- [7] a) S. Withers, *Carbohydr. Polym.* **2001**, *44*, 325–337; b) V. Rojas-Cervellera, A. Ardèvol, M. Boero, A. Planas, C. Rovira, *Chem. Eur. J.* **2013**, *19*, 14018–14023.
- [8] a) N. H. Abdul Manas, R. Md. Illias, N. M. Mahadi, *Crit. Rev. Biotechnol.* **2018**, *38*, 272–293; b) S. M. Hancock, M. D. Vaughan, S. G. Withers, *Curr. Opin. Chem. Biol.* **2006**, *10*, 509–519.
- [9] a) L. F. Mackenzie, Q. Wang, R. A. J. Warren, S. G. Withers, *J. Am. Chem. Soc.* **1998**, *120*, 5583–5584; b) M. R. Hayes, J. Pietruszka, *Molecules* **2017**, *22*, 1434; c) C. Malet, A. Planas, *FEBS Lett.* **1998**, *440*, 208–212; d) M. Moracci, A. Trincone, G. Perugino, M. Ciaramella, M. Rossi, *Biochemistry* **1998**, *37*, 17262–17270.
- [10] B. Bissaro, P. Monsan, R. Fauré, M. J. O'Donohue, *Biochem. J.* **2015**, *467*, 17–35.
- [11] a) J. Durand, X. Biarnés, L. Watterlot, C. Bonzom, V. Borsenberger, A. Planas, S. Bozonnet, M. J. O'Donohue, R. Fauré, *ACS Catal.* **2016**, *6*, 8264–8275; b) P. Lundemo, E. N. Karlsson, P. Adlercreutz, *Appl. Microbiol. Biotechnol.* **2017**, *101*, 1121–1131; c) N. N. Aronson, B. A. Halloran, M. F. Alexeyev, X. E. Zhou, Y. Wang, E. J. Meehan, L. Chen, *Biosci. Biotechnol. Biochem.* **2006**, *70*, 243–251; d) C. Brusa, N. Bellay, D. Gérard, M. Muzard, M. Dauchez, R. Plantier-Royon, C. Rémond, *J. Biotechnol.* **2018**, *272*, 56–63.
- [12] a) J. Nyirenda, S. Matsumoto, T. Saitoh, N. Maita, N. N. Noda, F. Inagaki, D. Kohda, *Structure* **2013**, *21*, 32–41; b) P. Mark, M. J. Baumann, J. M. Eklöf, F. Gullfot, G. Michel, Å. M. Kallas, T. T. Teeri, H. Brumer, M. Czjzek, *Proteins* **2009**, *75*, 820–836; c) S. H. Light, L. A. Cahoon, K. V. Mahasenan, M. Lee, B. Boggess, A. S. Halavaty, S. Mobashery, N. E. Freitag, W. F. Anderson, *Structure* **2017**, *25*, 295–304; d) B. David, P. Arnaud, C. Tellier, Y.-H. Sanejouand, *Protein Eng. Des. Sel.* **2019**, *32*, 309–316.
- [13] a) A. White, D. Tull, K. Johns, S. G. Withers, D. R. Rose, *Nat. Struct. Biol.* **1996**, *3*, 149–154; b) F. B. Bdira, C. A. Waudby, A. N. Volkov, S. P. Schröder, E. Ab, J. D. Codée, H. S. Overkleef, J. M. Aerts, H. van Ingen, M. Ubbink, *Angew. Chem. Int. Ed.* **2020**, *59*, 20508–20514.
- [14] W. W. Wakarchuk, R. L. Campbell, W. L. Sung, J. Davoodi, M. Yaguchi, *Protein Sci.* **1994**, *3*, 467–475.
- [15] G. P. Connelly, S. G. Withers, L. P. McIntosh, *Protein Sci.* **2000**, *9*, 512–524.
- [16] G. J. Davies, K. S. Wilson, B. Henrissat, *Biotechnol. J.* **1997**, *321*, 557.
- [17] a) G. Paës, V. Tran, M. Takahashi, I. Boukari, M. J. O'Donohue, *Protein Eng. Des. Sel.* **2007**, *20*, 15–23; b) G. Paës, J.-G. Berrin, J. Beaugrand, *Biotechnol. Adv.* **2012**, *30*, 564–592.
- [18] a) N. N. Mhlongo, M. Ebrahim, A. A. Skelton, H. G. Kruger, I. H. Williams, M. E. Soliman, *RSC Adv.* **2015**, *5*, 82381–82394; b) G. Paës, J. Cortés, T. Siméon, M. J. O'Donohue, V. Tran, *Comput. Struct. Biotechnol. J.* **2012**, *1*, e201207001.
- [19] D. S. Vieira, R. J. Ward, *J. Mol. Model.* **2012**, *18*, 1473–1479.
- [20] A. Pollet, E. Vandermarliere, J. Lammertyn, S. V. Strelkov, J. A. Delcour, C. M. Courtin, *Proteins* **2009**, *77*, 395–403.
- [21] U. B. Ericsson, B. M. Hallberg, G. T. DeTitta, N. Dekker, P. Nordlund, *Anal. Biochem.* **2006**, *357*, 289–298.
- [22] M. L. Ludwiczek, M. Heller, T. Kantner, L. P. McIntosh, *J. Mol. Biol.* **2007**, *373*, 337–354.

- [23] F. Zheng, L. Song, A. Basit, J. Liu, T. Miao, J. Wen, Y. Cao, W. Jiang, *Carbohydr. Polym.* **2020**, 237, 116121.
- [24] K. Chiku, J. Uzawa, H. Seki, S. Amachi, T. Fujii, H. Shinoyama, *Biosci. Biotechnol. Biochem.* **2008**, 72, 2285–2293.
- [25] P. H. Keizers, A. Saragliadis, Y. Hiruma, M. Overhand, M. Ubbink, *J. Am. Chem. Soc.* **2008**, 130, 14802–14812.
- [26] G. Sidhu, S. G. Withers, N. T. Nguyen, L. P. McIntosh, L. Ziser, G. D. Brayer, *Biochemistry* **1999**, 38, 5346–5354.
- [27] D. S. Vieira, L. Degrève, R. J. Ward, *Biochim. Biophys. Acta Gen. Subj.* **2009**, 1790, 1301–1306.
- [28] G. M. Clore, *Methods Enzymol.* **2015**, 564, 485–497.
- [29] K. Henzler-Wildman, D. Kern, *Nature* **2007**, 450, 964–972.

Manuscript received: December 19, 2020
Revised manuscript received: February 1, 2021
Accepted manuscript online: February 3, 2021
Version of record online: ■■■, ■■■■

COMMUNICATIONS

Shifting the balance: Glycoside hydrolases are attractive tools for the enzymatic synthesis of carbohydrates. We used the xylanase from *B. circulans* (BCX) to study the interplay between active-site flexibility and the transglycosylation reaction. A point mutation that enhances the flexibility and dynamics of BCX active site shifts the balance of the enzymatic reaction toward transglycosylation.



K. Marneth, H. van den Elst, A. Cramer-Blok, Prof. Dr. J. Codee, Prof. Dr. H. S. Overkleeft, Prof. Dr. J. M. F. G. Aerts, Prof. Dr. M. Ubbink, Dr. F. Ben Bdira**

1 – 8

Tuning the Transglycosylation Reaction of a GH11 Xylanase by a Delicate Enhancement of its Thumb Flexibility

