

Synthesis of pseudo-disaccharide analogues of lipid A: haptens for the generation of antibodies with glycosidase activity towards lipid A

Berg, R.J.B.H.N. van den; Noort, D.; Marel, G.A. van der; Boom, J.H. van; Benschop, H.P.

Citation

Berg, R. J. B. H. N. van den, Noort, D., Marel, G. A. van der, Boom, J. H. van, & Benschop, H. P. (2002). Synthesis of pseudo-disaccharide analogues of lipid A: haptens for the generation of antibodies with glycosidase activity towards lipid A. *Journal Of Carbohydrate Chemistry*, *21*(3), 167-188. doi:10.1081/CAR-120004331

Version: Publisher's Version License: [Licensed under Article 25fa Copyright Act/Law \(Amendment Taverne\)](https://hdl.handle.net/1887/license:4) Downloaded from: <https://hdl.handle.net/1887/3151641>

Note: To cite this publication please use the final published version (if applicable).

Journal of Carbohydrate Chemistry

ISSN: 0732-8303 (Print) 1532-2327 (Online) Journal homepage:<https://www.tandfonline.com/loi/lcar20>

SYNTHESIS OF PSEUDO-DISACCHARIDE ANALOGUES OF LIPID A: HAPTENS FOR THE GENERATION OF ANTIBODIES WITH GLYCOSIDASE ACTIVITY TOWARDS LIPID A

Richard J.B.H.N. van den Berg , Daan Noort , Gijs A. van der Marel , Jacques H. van Boom & Hendrik P. Benschop

To cite this article: Richard J.B.H.N. van den Berg , Daan Noort , Gijs A. van der Marel , Jacques H. van Boom & Hendrik P. Benschop (2002) SYNTHESIS OF PSEUDO-DISACCHARIDE ANALOGUES OF LIPID A: HAPTENS FOR THE GENERATION OF ANTIBODIES WITH GLYCOSIDASE ACTIVITY TOWARDS LIPID A, Journal of Carbohydrate Chemistry, 21:3, 167-188, DOI: [10.1081/CAR-120004331](https://www.tandfonline.com/action/showCitFormats?doi=10.1081/CAR-120004331)

To link to this article: <https://doi.org/10.1081/CAR-120004331>

Published online: 20 Aug 2006.

 \overrightarrow{S} [Submit your article to this journal](https://www.tandfonline.com/action/authorSubmission?journalCode=lcar20&show=instructions) G

Article views: 114

 $\overline{\text{O}}$ [View related articles](https://www.tandfonline.com/doi/mlt/10.1081/CAR-120004331) \mathbb{Z}

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

J. CARBOHYDRATE CHEMISTRY, 21(3), 167–188 (2002)

SYNTHESIS OF PSEUDO-DISACCHARIDE ANALOGUES OF LIPID A: HAPTENS FOR THE GENERATION OF ANTIBODIES WITH GLYCOSIDASE ACTIVITY TOWARDS LIPID A

Richard J. B. H. N. van den Berg,¹ Daan Noort,¹ Gijs A. van der Marel,² Jacques H. van Boom,^{2,*} and Hendrik P. Benschop¹

¹Department of Chemical Toxicology, TNO Prins Maurits Laboratory, P.O. Box 45, NL-2280 AA Rijswijk, The Netherlands ² ²Leiden Institute of Chemistry, Gorlaeus Laboratories, University of Leiden, P.O. Box 9502, NL-2300 RA Leiden, The Netherlands

ABSTRACT

In order to develop a generic treatment of sepsis caused by infections with Gram-negative bacteria, a series of pseudo-disaccharide analogues of lipid A $(1-5)$ was synthesized. These adducts not only harbor a 2acylaminodideoxynojirimycin unit mimicking the transition state of the glycosidic hydrolysis, but also a 2-N, 3-O-diacylated glucosamine moiety capable of generating catalytic antibodies with more selective glycosidase properties towards lipid A.

INTRODUCTION

Endotoxins, the complex lipopolysaccharides (LPS) situated in the outer membrane of Gram-negative bacteria, are extremely potent toxins.^[1] Most of the biological activities of LPS reside in the small terminal disaccharide phospholipid moiety known

Copyright © 2002 by Marcel Dekker, Inc. www.dekker.com

^{*}Corresponding author. E-mail: j.boom@chem.leidenuniv.nl

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

168 VAN DEN BERG ET AL.

as lipid A.^[2] Therapeutic strategies under development aim at either preventing endotoxin interaction with host effector cells or interrupting endotoxin mediated signal transduction pathways. This objective can be attained in blocking the synthesis and binding of endotoxin, thus neutralizing its activity.^[3-5]

Our approach to suppress sepsis caused by Gram-negative bacteria is based on catalytic antibodies capable of degrading lipid A via hydrolysis of the interglycosidic bond resulting in the formation of non-toxic monosaccharides. Since catalytic antibodies are supposed to have catalytic activities with tailor-made specificities, these abzymes have many potential therapeutically applications. For example, Landry et al. succeeded in the generation of a catalytic antibody that was effective in detoxifying cocaine from the blood stream via hydrolysis of the benzoyl ester function.^[6-9] Based on this result it was envisioned that a generic treatment of sepsis caused by Gramnegative bacteria might be feasible using specific and selective glycosidase antibodies capable of degrading lipid A.

Several groups have reported the design and generation of antibodies with glycosidase activity.^[10-16] In general, haptens are based on iminocyclitol glycosidase inhibitors such as deoxynojirimycin and isofagomine, which in terms of polarity and shape resemble the transition state of the glycosidic cleavage reaction. The protonated endocyclic nitrogen atom of iminocyclitols mimics the electronic charge developing in the transition state formed during cleavage of the interglycosidic bond. In a previous paper $\left[17\right]$ we reported the preparation of 2-acylaminodideoxynojirimycin derivatives mimicking the transition state of the hydrolysis of the interglycosidic bond at the nonreducing end of lipid A. It turned out that monoclonal antibodies raised against these haptens showed promising glycosidase activity (results to be published).

It was envisaged, based on the early studies by $\text{Dong}^{[18]}$ as well as Yu,^[14] that conjugation of 2-N, 3-O-diacylated glucosamine derivatives to 2-acylaminodideoxynojirimycin units would afford haptens suitable to raise specific glycosidase antibodies towards lipid A. The latter can be achieved by anchoring the 2-N, 3-O-diacylated glucosamine units, which mimic the reducing part of lipid A, to the endocyclic nitrogen atom of the iminoglucitol moieties via a flexible linker.

We here report the synthesis of several pseudo-disaccharide analogues of lipid A (i.e. compounds 1–5, Figure 1) containing 2-acylaminodideoxynojirimycins as well as 2-N, 3-O-diacylated glucosamine units.

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

PSEUDO-DISACCHARIDE ANALOGUES OF LIPID A 169

RESULTS AND DISCUSSION

The easily accessible 3,4,6-tri-O-benzyl-2-N-benzyloxycarbonylamino-1,2,5-trideoxy-1,5-iminoglucitol $(6)^{[17]}$ was used as a starting compound for the preparation of iminocyclitols 11 and 12 (Scheme 1) mimicking the non-reducing part of lipid A. In the first step, the carboxymethyl linker in 7 was introduced by alkylation of the endocyclic nitrogen atom of 6 with tert-butyl bromoacetate under the agency of cesium carbonate in dimethyl formamide^[19] to give compound 7 in a yield of 95%. Selective removal of the benzyloxycarbonyl (Z) protecting group in 7 proceeded smoothly by hydrogenation over Degussa type palladium on carbon to give 8 in a quantitative yield. PyBOP mediated coupling^[20] of amine 8 with myristic acid and (R) -3-dodecanoyloxytetradecanoic acid^[21] gave the *N*-acylated derivatives 9 and 10, respectively, in good yield. Removal of the tert-butyl group in compounds 9 and 10 was readily effected by treatment with neat trifluoroacetic acid, affording building blocks 11 and 12 in 100% and 83% yield, respectively.

The individual hapten units 34, 35 and 36, mimicking the reducing part of lipid A, were obtained by subjecting D-glucosamine (13) to the sequence of reactions depicted in Scheme 2. Accordingly, D-glucosamine (13) was converted in two steps into Nbenzyloxycarbonyl protected glucosamine 15 ^[22] Acetonation of 15 with 2,2-dimethoxypropane and a catalytic amount of p-toluenesulfonic acid gave partially protected derivative 16 in a quantitative yield. Condensation of the secondary hydroxyl group in 16 with myristic acid under the agency of $DCC^{[23]}$ afforded 17 in a yield of 86%. Removal of the Z-protecting group in 17 by hydrogenation over palladium on carbon in ethyl acetate gave amine 18 in a quantitative yield. PyBOP mediated condensation of amine 18 with the individual acids $19-21$, of which the primary amine function was protected with the Z-group, afforded the corresponding derivatives $22-24$. At this stage, N-acylated compound 22 was transformed into the primary amino derivative 34 by following the four-step process as portrayed in Scheme 2. Thus, de-acetonation of compound 22 with trifluoroacetic acid in aqueous tetrahydrofuran, $[24]$ followed by

Scheme 1. Conditions: (i) tert-butyl bromoacetate, Cs_2CO_3 , DMF, 95%; (ii) EtOAc, 5% Pd/C, H₂, 100% ; (iii) myristic acid, PyBOP, DiPEA, DCM, 89%; (iv) (R)-3-HOOCCH₂CHO $(COC_{11}H_{23})C_{11}H_{23}$, PyBOP, DiPEA, DCM, 70%; (v) TFA (11, 100%), (12, 83%).

170 VAN DEN BERG ET AL.

Scheme 2. Conditions: (i) benzyl chloroformate, NaHCO₃, H₂O; (ii) HCl/MeOH (2%, w/w), 53%; (iii) 2,2-dimethoxypropane, toluene-4-sulfonic acid, acetone, DCM, 100%; (iv) myristic acid, DCC, DMAP, DCM, 86%; (v) 5% Pd/C, H2, 100%; (vi) 19/20/21, PyBOP, DiPEA, DCM, (22, 87%), (23, 96%), (24, 87%); (vii) TFA, THF/H2O, (4/1, v/v), (25, 99%), (26, 63%), (27, 87%); (viii) MsCl (TsCl), pyridine, (28, 80%), (29, 78%), (30, 79%); (ix) NaN₃, DMF, 80°C, (31, 84%), (32, 72%), (33, 73%) (x) triphenylphosphine (1.5 equiv), H2O (1.2 equiv), THF, (34, 100%), (35, 78%), (36, 52%).

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

PSEUDO-DISACCHARIDE ANALOGUES OF LIPID A 171

regio-selective sulfonylation of the primary hydroxyl group of diol 25 gave sulfonylate 28 in an overall yield of 80%. Treatment of the latter derivative with sodium azide in dimethyl formamide at elevated temperature afforded the 6-azidoglucosamine compound 31. Subsequent reduction of the resulting azido function in 31 under the agency of triphenylphosphine in aqueous tetrahydrofuran^[25] gave the requisite 6-amino derivative 34 in yield of 84% based on sulfonylate 28. In a similar way compounds 35 and 36 were attained by subjecting acetonides 23 and 24 to the same four-step process as described for the synthesis of 34.

Having the requisite building blocks 11, 12, 34–36 at hand, introduction of the required amide bond of the fully protected compounds 37–41 could be readily accomplished (see Scheme 3) using PyBOP as the condensation agent. For example, PyBOPmediated condensation of acid 11 and amine 34 proceeded smoothly to afford the fully protected hapten 37 in a yield of 96%. The identity and homogeneity of compound 37

Scheme 3. Conditions: (i) PyBOP, DiPEA, DCM, (37, 96%), (38, 96%), (39, 51%), (40, 88%), (41, 96%); (ii) 10% Pd/C, DMF (1, 100%), (2, 72%), (3, 54%), (4, 88%), (5, 65%).

172 VAN DEN BERG ET AL.

was fully ascertained by NMR spectroscopy and mass spectrometry. Similar yields were obtained by condensation of 11 with 35 and 12 with 34 as well as 35 to yield the fully protected haptens 38, 40 and 41, respectively. In contrast, the coupling of acid 11 with amine 36 to give amide 39 was in terms of yield not fully satisfactory, and may be ascribed to the increased lipophilicity of amine $36.$ 1 H, 13 C NMR and mass spectrometric data of the four conjugates 38–41 were in complete accordance with the proposed structures. In the final stage, haptens 37–41 were deprotected by hydrogenolysis over palladium on carbon in dimethyl formamide. Hydrogenolysis of compound 37 proceeded in near quantitative yield to give hapten 1, the identity and homogeneity of which was fully ascertained by NMR spectroscopy and mass spectrometry. The same results were obtained by hydrogenolysis of compounds 38 and 40 to yield the haptens 2 and 4, respectively. Unfortunately, unmasking of compounds 39 and 41 was rather sluggish and led to the isolation of haptens 3 and 5 in moderate yields. The disappointing outcome of the latter hydrogenolysis may also be due to the intrinsically high lipophilic nature of compounds 39 and 41 . The identity of compounds $2-5$ could be readily ascertained by mass spectrometry. Unfortunately, it turned out that the structure assignment of haptens $2-5$ was seriously hampered by the fact that NMR spectra^[26] could not be interpreted due to extensive line broadening.

CONCLUSION

In summary, we have synthesized five pseudo-lipid A analogues as potential haptens for the generation of catalytic antibodies with glycosidase activity towards lipid A. These haptens contain a primary amino function in the N-acyl chain of the 2-N, 3-Odiacylated glucosamine units which will serve as a handle of anchoring haptens 1–5 to a carboxylic acid terminus of a carrier protein. The immunochemical evaluation of the haptens will be reported in due course.

EXPERIMENTAL

General Methods. Toluene (Merck) was distilled from P_2O_5 and stored over sodium wire. Dichloromethane and N,N-dimethylformamide were purchased from Biosolve Ltd. and freshly distilled from CaH₂. N,N-Diisopropylethylamine (Acros Chimica) was distilled from p-toluenesulfonyl chloride (60 g/L) and redistilled from potassium hydroxide pellets (40 g/L). Benzyl chloroformate, tert-butyl bromoacetate, cesium carbonate, N,N-dicyclohexylcarbodiimide, 4-(dimethylamino)pyridine, 2,2-dimethoxypropane, D-glucosamine hydrochloride, methanesulfonyl chloride, palladium on carbon (5%, Degussa E101 NO/W), sodium azide, tetrahydrofuran, toluene-p-sulfonic acid, toluene-p-sulfonyl chloride and triphenylphosphine were purchased form Fluka. PyBOP was purchased from NovaBiochem. Trifluoroacetic acid was purchased from Acros Chimica. 1 H NMR and 13 C NMR data were recorded with a Varian VXR-400S (399.9/ 100.6 MHz). ¹H and ¹³C chemical shifts are given in ppm (δ) relative to tetramethylsilane (δ =0.00), DMSO-d₅ (δ =2.525), DMSO-d₆ (δ =39.6) and CDCl₃ (δ =77.00) as internal standard. The purity of the compounds was established by ${}^{1}H$ NMR spectroscopy: >95% in all cases. Mass spectra were recorded with a VG Quattro II triple

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

PSEUDO-DISACCHARIDE ANALOGUES OF LIPID A 173

quadropole mass spectrometer (Fisons Instruments, Altrincham, UK). Column chromatography was performed on Silica gel 60 (220–440 mesh ASTM, Fluka). TLC analysis was performed with silica gel TLC plates (Fluka) with detection by UV absorption (254 nm) where applicable and charring with 20% H₂SO₄ in MeOH or ammonium molybdate (25 g/L) and ceric ammonium sulfate (10 g/L) in 20% H_2SO_4 . Prior to reactions that require anhydrous conditions, traces of water were removed by coevaporation with dry toluene. These reactions were conducted under dry argon atmosphere. Hydrogenations were executed at atmospheric pressure under an atmosphere of hydrogen gas maintained by an inflated balloon. Polytetrafluoroethylene (PFTE) filters were purchased from Alltech (Breda, The Netherlands).

3,4,6-Tri-O-benzyl-2-[(benzyloxycarbonyl)amino]-1,5-N-[(O-tert-butyl-carboxy**methyl)imino]-1,2,5-trideoxy-D-glucitol (7).** To a solution of 6 (1.20 g, 2.12 mmol) in DMF (20 mL) cesium carbonate (700 mg, 2.14 mmol) and tert-butyl bromoacetate (1.15 mL, 7.81 mmol) were added. The reaction mixture was stirred for 16 h at ambient temperature, after which TLC analysis indicated complete conversion of starting material into a compound with $R_f = 0.92$ (ethyl acetate/hexane, 1:1, v/v). The mixture was diluted with DCM (100 mL) and washed with aqueous NaOH (1 M, 50 mL). After drying over MgSO4, the organic layer was concentrated in vacuo. The crude product was purified by silica gel column chromatography. Elution was performed with DCM/ MeOH (100:0 \rightarrow 96.5:3.5, v/v). Yield 1.37 g (95%). ¹H NMR (CDCl₃): δ = 1.37 (s, 9H, CH₃, tBu), 2.71 (dd, 1H, H-1ax, J_{1ax,1eq} = 11.6 Hz, J_{1ax,2} = 8.9 Hz), 3.04 (br. s, 1H, H-5), 3.10 (br. d, 1H, H-1eq), 3.27 (d, 1H, Ha-acetyl, J = 17.7 Hz), 3.35 (br. t, 1H, H-3), 3.52 (dd, 1H, H-6, $J_{6.6'} = 10.5$ Hz, $J_{5.6} = 2.8$ Hz), 3.57 (d, 1H, Hb-acetyl, J = 17.7 Hz), 3.59 (t, 1H, H-4), 3.72 (dd, 1H, H-6', $J_{6.6'} = 10.5$ Hz, $J_{5.6'} = 3.9$ Hz), 3.79 (m, 1H, H-2), 4.40– 4.76 (m, $6H$, $3 \times CH_2$ Bn), 4.89 (s, 1H, NH), 5.05 (dd, $2H$, CH₂ Z), $7.18-7.39$ (m, 20H, CH-arom Bn/Z). ¹³C{¹H} NMR (CDCl₃): δ = 28.16 (CH₃ tBu), 50.88 (C-2), 53.38 (C-1), 55.08 (CH₂ tert-butyl acetate), 61.61 (C-5), 66.02 (C-6), 66.51 (CH₂ Z), 73.39, 73.66, 73.97 (3 × CH₂ Bn), 78.39 (C-4), 80.90 (Cq *t*Bu), 81.43 (C-3), 127.58–128.50 (CH-arom Bn/Z), 156.00 (C=O Z), 170.63 (C=O tert-butyl acetyl). ES-MS; m/z : 681.5, $[M + H]^{+}$; monoisotopic MW calculated for C₄₁H₄₈N₂O₇ = 680.35.

2-Amino-3,4,6-tri-O-benzyl-1,5-N-[(O-tert-butyl-carboxymethyl)imino]-1,2,5-trideoxy-D-glucitol (8). Pd/C (5%, Degussa type E101 NO/W, 100 mg) was added to a solution of 7 (109 mg, 0.160 mmol) in ethyl acetate (5 mL). Hydrogen was passed through the stirred mixture for 1 h, after which TLC analysis indicated the complete conversion of starting material into a compound with $R_f = 0.20$ (MeOH/DCM, 5:95, v/v). The mixture was passed over a short column containing a layer of glass wool and a layer of hyflo $^{(8)}$ and, finally, over a PTFE filter. Concentration of the filtrate in vacuo yielded 8 as a white solid (94 mg; 100%). ¹H NMR (CDCl₃): δ = 1.48 (s, 9H, CH₃, $t\text{Bu}$, 1.77 (br. s, 2H, NH₂), 2.72 (t, 1H, H-1ax), 2.92 (m, 3H, H-1eq, H-2, H-5), 3.16 $(t, 1H, H-3 J = 8.9 Hz)$, 3.30 (d, 1H, Ha-acetyl, J = 17.6 Hz), 3.51 (t, 1H, H-4, J = 9.1 Hz), 3.55 (dd, 1H, H-6), 3.59 (d, 1H, Hb-acetyl, J = 17.6 Hz), 3.72 (dd, 1H, H-6', $J_{6.6'} = 10.6$ Hz, $J_{5,6'} = 3.2$ Hz), 4.44–4.94 (m, 6H, $3 \times CH_2$ Bn), 7.18–7.35 (m, 15H, CH-arom Bn). ¹³C{¹H} NMR (CDCl₃): δ = 28.22 (CH₃ tBu), 52.64 (C-2), 54.43 (CH₂ tert-butyl acetate), 57.78 (C-1), 62.35 (C-5), 65.90 (C-6), 73.50, 74.71, 75.10 (3 \times CH₂ Bn), 79.67 (C-4), 81.00 (Cq tBu), 88.52 (C-3), 127.59–128.52 (CH-arom Bn), 137.70, 138.50,

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

174 VAN DEN BERG ET AL.

138.78 ($3 \times Cq$ Bn), 170.55 (C=O tert-butyl acetyl). ES-MS; m/z: 547.5, [M+H]⁺; monoisotopic MW calculated for $C_{33}H_{42}N_2O_5 = 546.31$.

3,4,6-Tri-O-benzyl-1,5-N-[(O-tert-butyl-carboxymethyl)imino]-2-(tetradecanoyl) amino-1,2,5-trideoxy-D-glucitol (9). To a stirred mixture of myristic acid (92 mg, 0.403 mmol), PyBOP (231 mg, 0.605 mmol) and DiPEA (76 μ L, 0.443 mmol) in DCM (10 mL) a solution of **8** (200 mg, 0.366 mmol) in DCM (10 mL) was added. After 30 min, TLC analysis indicated the complete conversion of starting material into a compound with $R_f = 0.89$ (MeOH/DCM, 5:95, v/v). The reaction mixture was diluted with DCM (100 mL) and washed with water $(1 \times 50 \text{ mL})$. After drying over MgSO₄, the organic layer was concentrated under reduced pressure. The crude product was purified by silica gel column chromatography. Elution was performed with hexane/ ethyl acetate (80:20 \rightarrow 60:40, v/v). Yield 228 mg (89%). ¹H NMR (CDCl₃): δ = 0.88 (t, 3H, CH₃ myristyl), 1.25 (m, 20H, $10 \times CH_2$ myristyl), 1.48 (s, 9H, $3 \times CH_3$ tBu), 1.49 (m, 2H, CH₂ myristyl), 1.94 (m, 2H, CH₂ myristyl), 2.57 (dd, 1H, H-1ax, J_{1ax,1eq} = 11.8 Hz, $J_{1ax,2} = 7.0$ Hz), 3.15 (m, 1H, H-5), 3.17 (dd, 1H, H-1eq, $J_{1ax,1eq} = 11.8$ Hz, $J_{1ea,2} = 3.8$ Hz), 3.29 (d, 1H, Ha-acetyl, J = 17.7 Hz), 3.45 (t, 1H, H-3), 3.52 (dd, 1H, H-6, $J_{6.6'} = 10.2$ Hz, $J_{5.6} = 4.9$ Hz), 3.54 (d, 1H, Hb-acetyl, J = 17.7 Hz), 3.60 (t, 1H, H-4, J = 6.1), 3.77 (dd, 1H, H-6', J_{6,6'} = 10.3 Hz, J_{5,6'} = 4.9 Hz), 4.00 (m, 1H, H-2), 4.41-4.70 $(m, 6H, 3 \times CH_2 Bn)$, 5.85 (d, 1H, NH, J_{2,NH} = 7.1 Hz), 7.24–7.35 (m, 15H, CH-arom Bn). ¹³C{¹H} NMR (CDCl₃): δ = 14.15 (CH₃ myristyl), 22.74–36.92 (CH₂ myristyl), 28.21 (CH3 tBu), 48.30 (C-2), 51.15 (C-1), 55.67 (CH2 acetyl), 61.51 (C-5), 66.28 (C-6), 72.93, 73.31, 73.39 (3 \times CH₂ Bn), 77.73 (C-4), 78.97(C-3), 80.80 (Cq tBu), 127.67– 128.59 (CH-arom Bn), 138.02, 138.41, 138.45 $(3 \times Cq$ Bn), 170.90 (C=O acetyl), 172.96 (C=O myristyl). ES-MS; m/z : 757.5, $[M+H]^+$; monoisotopic MW calculated for $C_{47}H_{68}N_2O_6 = 756.5$.

3,4,6-Tri-O-benzyl-1,5-N-[(O-tert-butylcarboxymethyl)imino]-2-[(R)-3-(dodecanoyloxytetradecanoyl)]amino-1,2,5-trideoxy-D-glucitol (10). (R)-3-Dodecanoyloxytetradecanoic acid (73 mg, 0.171 mmol) was coupled with compound 8 (94 mg, 0.172 mmol) as described for the preparation of compound 9. The crude product was purified by silica gel column chromatography; elution was performed with hexane/ethyl acetate $(80:20 \rightarrow 60:40, v/v)$. $R_f = 0.66$ (hexane/ethyl acetate, 2:1, v/v). Yield 115 mg (70%). ¹H NMR (CDCl₃): $\delta = 0.88$ (m, 6H, $2 \times CH_3$ acyloxyacyl), 1.25 (m, 34H, CH₂ acyloxyacyl), 1.40 (s, 9H, $3 \times CH_3$ tBu), 1.55 (m, 4H, $2 \times CH_2$ acyloxyacyl), 2.21 (m, 4H, $2 \times CH_2$ acyloxyacyl), 2.56 (dd, 1H, H-1ax, $J_{1ax,eq} = 11.7$ Hz, $J_{1ax,2} = 7.3$ Hz), 3.12 (m, 2H, H-1eq, H-5), 3.28 (d, 1H, Ha-acetyl, J = 17.8 Hz), 3.45 (t, 1H, H-3, J = 6.5 Hz), 3.53 (dd, 1H, H-6, $J_{6,6'} = 10.3$ Hz, $J_{5,6} = 3.6$ Hz), 3.55 (d, 1H, Hb-acetyl, J = 17.8 Hz), 3.60 (t, 1H, H-4, J = 6.3 Hz), 3.76 (dd, 1H, H-6', $J_{6.6'} = 10.3$ Hz, $J_{5.6'} = 5.0$ Hz), 4.00 (m, 1H, H-2), 4.40–4.71 (m, 6H, $3 \times CH_2$ Bn), 5.11 (m, 1H, CHO acyloxyacyl), 6.06 (d, 1H, NH, $J_{2,NH}$ = 7.5 Hz), 7.23–7.33 (m, 15H, CH-arom Bn). ¹³C{¹H} NMR (CDCl₃): δ = 14.12 (CH₃ acyloxyacyl), 22.70–41.65 (CH₂ acyloxyacyl), 28.17 (CH₃ tBu), 48.48 $(C-2)$, 51.30 $(C-1)$, 55.52 $(CH_2 \text{ acetyl})$, 61.43 $(C-5)$, 66.27 $(C-6)$, 71.21 (CHO acyloxyacyl), 72.92, 73.05, 73.34 $(3 \times CH_2 \text{ Bn})$, 77.64 $(C-4)$, 79.33 $(C-3)$, 80.77 (Cq) t Bu), 127.63–128.54 (CH-arom Bn), 137.98, 138.40, 138.45 ($3 \times Cq$ Bn), 169.43, 170.82, 173.12 $(3 \times C = O$ amide, ester). ES-MS; m/z : 954.67 $[M + H]^{+}$, monoisotopic MW calculated for $C_{59}H_{90}N_2O_8 = 955.63$.

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

PSEUDO-DISACCHARIDE ANALOGUES OF LIPID A 175

3,4,6-Tri-O-benzyl-1,5-N-[carboxymethylimino]-2-(tetradecanoyl)amino-1,2,5-trideoxy-D-glucitol (11). A solution of 9 (118 mg, 0.156 mmol) in TFA (5 mL) was stirred for 2 h at ambient temperature, after which TLC analysis indicated complete conversion of starting material into a compound with $R_f = 0.12$ (MeOH/DCM, 5:95, v/v). After concentration of the reaction mixture, the residue was coevaporated with toluene (3×5 mL). Yield 125 mg (quantitative). ¹H NMR (CDCl₃): $\delta = 0.88$ (t, 3H, CH_3 myristyl), 1.25 (m, 20H, CH₂ myristyl), 1.48 (br. s, 2H, CH₂ myristyl), 2.04 (m, 2H, CH2 myristyl), 2.95 (d, 1H, H-1ax), 3.26 (br. s, 2H, H-1eq, H-5), 3.45 (br. s, 1H, Ha-acetyl), 3.56 (br. s, 2H, H-3, H-6), 3.72 (m, 2H, Hb-acetyl, H-6'), 3.89 (m, 1H, H-4), 4.26 (br. s, 1H, H-2), 4.39–4.62 (m, 6H, $3 \times CH_2$ Bn), 7.09 (br. s, 1H, NH), 7.14– 7.27 (m, 15H, CH-arom Bn), 9.40 (br. s, 1H, COOH). ${}^{13}C[{^1H}]$ NMR (CDCl₃): δ = 14.15 (CH₃ myristyl), 22.73–36.68 (CH₂ myristyl), 46.32 (C-2), 50 (C-1), 54.46 (CH₂ acetyl), 61.08 (C-5), 64.88 (C-6), 73.10, 73.33, 73.49 (3 \times CH₂ Bn), 75.38 (C-4), 76.05(C-3), 127.82–128.71 (CH-arom Bn), 136.42–137.77 (Cq Bn), 165.08 (C = O carboxymethyl), 174.39 (C=O myristyl). ES-MS; m/z : 701.5, $[M+H]^+$; monoisotopic MW calculated for $C_{43}H_{60}N_2O_6 = 700.45$.

3,4,6-Tri-O-benzyl-1,5-N-[carboxymethylimino]-2-[(R)-3-(dodecanoyloxytetradecanoyl)]amino-1,2,5-trideoxy-D-glucitol (12). Compound 10 (115 mg, 0.120 mmol) was treated with TFA as described for the preparation of compound 11. The crude product was purified by silica gel column chromatography. Elution was performed with MeOH/DCM (0:100 \rightarrow 5:95, v/v). R_f = 0.12 (hexane/ethyl acetate, 1:2, v/v). Yield 90 mg (83%). ¹H NMR (CDCl₃): $\delta = 0.88$ (m, 6H, $2 \times CH_3$ acyloxyacyl), 1.25 (br. s, 34H, CH₂ acyloxyacyl), 1.56 (m, 4H, CH₂ acyloxyacyl), 2.23 (m, 4H, CH₂ acyloxyacyl), 2.60 (d, 1H, H-1ax), 3.21 (m, 2H, H-1eq, H-5), 3.40 (d, 1H, Ha-acetyl), 3.51 (m, 2H, H-3, H-6), 3.66 (m, 2H, Hb-acetyl, H-6'), 3.86 (m, 1H, H-4), 4.13 (br. s, 1H, H-2), 4.39–4.62 (m, 6H, 3-CH2 Bn), 5.05 (m, 1H, CHO acyloxyacyl), 6.70 (br. s, 1H, NH), 7.21–7.35 (m, 15H, CH-arom Bn). ¹³C{¹H} NMR (CDCl₃): δ =14.13 (CH₃) acyloxyacyl), 22.71-41.86 (CH₂ acyloxyacyl), 47.03 (C-2), 49.49 (C-1), 56.66 (CH₂ acetyl), 61.56 (C-5), 65.94 (C-6), 71.09 (CHO acyloxyacyl), 72.72, 73.12, 73.45 $(3 \times CH_2$ Bn), 75.83 (C-4), 76.08 (C-3), 127.69–128.70 (CH-arom Bn), 137.27, 137.46, 137.68 $(3 \times Cq \text{ Bn})$, 169.49, 171.96, 173.45 $(3 \times C = O \text{ acid}, \text{ amide}, \text{ester})$. ES-MS; m/z : 899.7, $[M + H]$ ⁺; monoisotopic MW calculated for $C_{55}H_{82}N_2O_8 = 898.6$.

Methyl 2-[(benzyloxycarbonyl)amino]-2-deoxy-a-D-glucopyranoside (15). To a cooled (0° C) solution of glucosamine hydrochloride (13) (20 g, 93 mmol) in water (400 mL) NaHCO₃ (14.4 g, 171 mmol) and benzyl chloroformate (17.4 mL, 104 mmol) were added. The mixture was stirred at ambient temperature for 16 h, after which the white crystalline residue (14) was filtered off, washed with cold acetone (-20° C) and dried. The white crystals were dissolved in acidic methanol (2% HCl, w/w) and refluxed for 7 h after which the reaction mixture was concentrated. The resulting residue was purified by silica gel column chromatography. Elution was performed with MeOH/DCM (10:90 \rightarrow 15:85, v/v). Yield 16 g (53%). R_f = 0.70 (MeOH/DCM, 15:85, v/v). ¹H NMR (DMSO-d₆): δ = 3.16 (m, 2H, H-4, H-5), 3.27 (s, 3H, OMe), 3.46 (m, 3H, H-2, H2-6), 3.67 (m, 1H, H-3), 4.51 (t, 1H, OH-6), 4.61 (d, 1H, H-1, J1,2 = 3.2 Hz), 4.76 (d, 1H, OH-3), 4.98 (d, 1H, OH-4), 5.04 (dd, 2H, CH₂ Z), 7.07 (d, 1H, NH, $J_{2,NH}$ =7.7 Hz), 7.31–7.42 (m, 5H, CH-arom Z). ¹³C{¹H} NMR (DMSO-d₆): δ =54.37

176 VAN DEN BERG ET AL.

 (OMe) , 55.95 (C-2), 60.89 (C-6), 65.34 (CH₂ Z), 70.65 (C-5), 70.81 (C-3), 72.72 (C-4), 98.09 (C-1), 127.78, 128.35 (CH-arom Z), 137.16 (Cq Z), 156.17 (C=O Z).

Methyl 2-[(benzyloxycarbonyl)amino]-2-deoxy-4,6-O-isopropylidene-a-D-glucopyranoside (16). To a mixture of 15 (16 g, 49 mmol) in dry acetone (200 mL) and DCM (150 mL) was added 2,2-dimethoxypropane (25 mL, 204 mmol) and p-toluene sulfonic acid (0.4 g, 2.1 mmol). The resulting mixture was stirred at ambient temperature and after 16 h, TLC analysis showed complete conversion of starting material into a compound with $R_f = 0.80$ (MeOH/DCM, 5:95, v/v). TEA (5 mL) and DCM (100 mL) were added and the mixture was washed with water (50 mL). The organic layer was dried (MgSO4) and concentrated. Purification of the crude product by silica gel column chromatography (elution with MeOH/DCM (0:100 \rightarrow 5:95, v/v)), yielded 18 g (quantitative) of a yellow oil. ¹H NMR (CDCl₃): $\delta = 1.42$ (s, 3H, CH₃ isopropylidene), 1.51 (s, 3H, CH3 isopropylidene), 2.84 (s, 1H, OH-3), 3.33 (s, 3H, OMe), 3.59 (m, 2H, H-4, H-5), 3.74 (m, 2H, H-3, H-6), 3.87 (m, 2H, H-2, H-6'), 4.68 (d, 1H, H-1, $J_{1,2} = 3.4$ Hz), 5.11 (s, 2H, CH₂ Z), 5.19 (d, 1H, NH, $J_{2,NH} = 8.7$ Hz), 7.32–7.36 (m, 5H, CH-arom Z). ¹³C{¹H} NMR (CDCl₃): $\delta = 19.17$, 29.14 (2 × CH₃ isopropylidene), 55.28 (OMe), 55.92 (C-2), 62.36 (C-6), 63.36 (C-5), 67.30 (CH₂ Z), 70.88 (C-3), 74.63 (C-4), 99.23 (C-1), 99.90 (Cq isopropylidene), 128.09–128.60 (CH-arom Z), 136.22 (Cq Z), 156.87 $(C = O Z)$.

Methyl 2-[(benzyloxycarbonyl)amino]-2-deoxy-4,6-O-isopropylidene-3-O-tetradecanoyl- α -D-glucopyranoside (17). To a solution of 16 (5.545 g, 15.10 mmol) in DCM was added DMAP (2.03 g, 16.6 mmol), myristic acid (3.79 g, 16.6 mmol) and DCC (3.43 g, 16.6 mmol). The reaction mixture was stirred for 18 h at ambient temperature. TLC analysis (MeOH/DCM, 1:99, v/v) showed complete conversion of starting material into a compound with $R_f = 0.91$. DCU was filtered off and the filter was washed with DCM $(3 \times 25 \text{ mL})$. DCM was concentrated and the crude product was purified by silica gel column chromatography. Elution was performed with MeOH/DCM $(0:100 \rightarrow 3:97, v/v)$. Yield: 7.49 g (86%) of a colorless oil. ¹H NMR (CDCl₃): $\delta = 0.88$ (t, 3H, CH₃ myristyl), 1.25 (br. s, 20H, CH₂ myristyl), 1.36 (s, 3H, CH₃ isopropylidene), 1.46 (s, 3H, CH3 isopropylidene), 1.54 (m, 2H, CH2 myristyl), 2.22 (m, 2H, CH2 myristyl), 3.36 (s, 3H, OMe), 3.69 (m, 2H, H-4, H-5), 3.76 (t, 1H, H-6), 3.87 (dd, 1H, H-6', $J_{5.6'} = 4.7$ Hz, $J_{6.6'} = 10.2$ Hz), 3.97 (m, 1H, H-2, $J_{1.2} = 3.7$ Hz, $J = 10.2$ Hz), 4.69 (d, 1H, H-1, $J_{1,2} = 3.7$ Hz), 5.06 (s, 2H, CH₂ Z), 5.12 (m, 2H, H-3, NH), 7.26–7.36 (m, 5H, CH-arom Z). ¹³C{¹H} NMR (CDCl₃): δ = 14.15 (CH₃ myristyl), 19.11, 29.11 (2 × CH₃ isopropylidene), 22.74–34.38 (CH2 myristyl), 54.60 (C-2), 55.32 (OMe), 62.51 (C-6), 63.86 (C-5), 66.93 (CH₂ Z), 70.36 (C-3), 72.15 (C-4), 99.43 (C-1), 99.73 (Cq isopropylidene), 128.03, 128.18, 128.55 (CH-arom Z), 136.40 (Cq Z), 156.01 (C=O Z), 173.83 (C=O ester).

Methyl 2-amino-2-deoxy-4,6-O-isopropylidene-3-O-tetradecanoyl- α -D-glucopyranoside (18). To a solution of 17 $(6.69 \text{ g}, 11.6 \text{ mmol})$ in EtOAc (100 mL) was added Pd/C (10%, 1.0 g). Hydrogen was passed through the stirred mixture for 66 h. TLC analysis showed complete conversion of starting material into a new product with R_f = 0.26 (MeOH/DCM, 1:99, v/v). The mixture was filtered over a PTFE filter. The filtrate was concentrated under reduced pressure. Yield: 5.19 g (quantitative) of a

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

PSEUDO-DISACCHARIDE ANALOGUES OF LIPID A 177

colorless oil. ¹H NMR (CDCl₃): δ = 0.88 (s, 3H, t, CH₃ myristyl), 1.25 (br. s, 20H, $CH₂$ myristyl), 1.36 (s, 3H, $CH₃$ isopropylidene), 1.45 (s, 3H, $CH₃$ isopropylidene), 1.64 (m, 2H, CH2 myristyl), 1.76 (m, 2H, NH2), 2.34 (q, 2H, CH2 myristyl), 2.86 (dd, 1H, H-2, $J_{1,2} = 3.6$ Hz, $J_{2,3} = 10.0$ Hz), 3.39 (s, 3H, OMe), 3.56 (d, 1H, H-4), 3.72 $(m, 2H, H-5, H-6), 3.87$ $(m, 1H, H-6), 4.71$ $(d, 1H, H-1, J_{1,2} = 3.6 \text{ Hz}), 5.05$ $(t, 1H,$ H-3). ¹³C{¹H} NMR (CDCl₃): $\delta = 14.11$ (CH₃ myristyl), 19.09, 29.13 (2 × CH₃ isopropylidene), 22.70–34.61 (CH2 myristyl), 55.29 (OMe), 55.54 (C-2), 62.64 (C-6), 63.83 (C-5), 72.50 (C-4), 73.76 (C-3), 99.52 (Cq isopropylidene), 101.37 (C-1), 173.63 $(C = O \text{ ester}).$

Methyl 2-[(6-benzyloxycarbonylamino)hexanoylamino]-2-deoxy-4,6-O-isopropylidene-3-O-tetradecanoyl- α -D-glucopyranoside (22). To a stirred mixture of 6-benzyloxycarbonylaminohexanoic acid (19; 516 mg, 1.95 mmol), PyBOP (1.522 g, 2.93 mmol) and DiPEA (367 µL, 2.15 mmol) in DCM (50 mL) was added 18 (0.913 g, 2.06 mmol) in DCM (10 mL). After 1 h, TLC analysis indicated the complete conversion of starting material into a product with $R_f = 0.57$ (hexane/ethyl acetate, 1:2, v/v). The reaction mixture was diluted with DCM (100 mL) and washed with water $(3 \times 50 \text{ mL})$. The organic layer was dried $(MgSO₄)$ and concentrated under reduced pressure. The crude product was purified by silica gel column chromatography. Elution was performed with hexane/ethyl acetate $(60:40 \rightarrow 40:60, v/v)$. Yield 1.172 g $(87%)$. ¹H NMR $(CDC1_3)$: $\delta = 0.88$ (t, 3H, CH₃ myristyl), 1.23–1.64 (m, 28H, CH₂ hexanoyl, myristyl), 1.37 (s, 3H, CH3 isopropylidene), 1.47 (s, 3H, CH3 isopropylidene), 2.11–2.35 (m, 4H, hexanoyl, myristyl), 3.17 (br. q, 2H, CH₂ hexanoyl), 3.35 (s, 3H, OMe), 3.66–3.79 (m, 3H, H-4, H-5, H-6), 3.87 (dd, 1H, H-6', $J_{5.6'} = 5.0$ Hz, $J_{6.6'} = 10.5$ Hz), 4.27 (m, 1H, H-2, $J_{1,2} = 3.7$ Hz, $J_{2,3} = 9.5$ Hz), 4.66 (d, 1H, H-1 $J_{1,2} = 3.7$ Hz), 4.95 (s, 1H, NH aminohexanoyl), 5.09 (s, 2H, CH₂ Z), 5.12 (t, 1H, H-3, $J_{2,3} = 9.5$ Hz, $J_{3,4} = 9.5$ Hz), 5.88 (s, 1H, NH), 7.28–7.35 (m, 5H, CH-arom Z). ¹³C{¹H} NMR (CDCl₃): δ = 14.08 (CH₃) myristyl), 22.66–40.82 (CH₂ hexanoyl, myristyl), 19.06, 26.23 (2 \times CH₃ isopropylidene), 52.51 (C-2), 55.23 (OMe), 62.43 (C-6), 63.73 (C-5), 66.55 (CH₂ Z), 70.38 (C-3), 71.94 (C-4), 99.07 (C-1), 99.73 (Cq isopropylidene), 128.03, 128.48 (CH-arom Z), 136.72 (Cq Z), 156.21 (C=O Z), 172.82, 174.26 ($2 \times C = O$ amide, ester).

Methyl 2-[(12-benzyloxycarbonylamino)dodecanoylamino]-2-deoxy-4,6-O-isopropylidene-3-O-tetradecanoyl- α -D-glucopyranoside (23). 12-Benzyloxycarbonylaminododecanoic acid 20 (0.642 g, 1.84 mmol) was coupled with compound 18 (0.815 g, 1.84 mmol) as described for the preparation of compound 22. The crude product was purified by silica gel column chromatography. Elution was performed with hexane/ ethyl acetate (90:10 \rightarrow 40:60, v/v). Yield 1.368 g (96%). ¹H NMR (CDCl₃): δ = 0.88 (t, 3H, CH₃ myristyl), 1.25 (m, 34H, CH₂ dodecanoyl, myristyl), 1.37 (s, 3H, CH₃ isopropylidene), 1.46 (s, 3H, CH₃ isopropylidene), 1.57 (m, 6H, CH₂ dodecanoyl, myristyl), $2.10-2.33$ (m, 4H, dodecanoyl, myristyl), 3.17 (q, $2H$, CH₂ dodecanoyl), 3.36 (s, 3H, OMe), 3.69–3.79 (m, 3H, H-4, H-5, H-6), 3.87 (dd, 1H, H-6'), 4.27 (m, 1H, H-2), 4.67 (d, 1H, H-1, $J_{1,2} = 3.6$ Hz), 4.82 (s, 1H, NH aminododecanoyl), 5.09 (s, 2H, CH2 Z), 5.13 (q, 1H, H-3), 5.82 (d, 1H, NH), 7.27–7.35 (m, 5H, CH-arom Z). ¹³C{¹H} NMR (CDCl₃): δ = 14.11 (CH₃ myristyl), 22.68–41.15 (CH₂ dodecanoyl, myristyl), 19.07, 29.07 (2 × CH₃ isopropylidene), 52.49 (C-2), 55.25 (OMe), 62.46 (C-6), 63.78 (C-5), 66.55 (CH₂ Z), 70.28 (C-3), 71.97 (C-4), 99.13 (C-1), 99.74 (Cq

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

178 VAN DEN BERG ET AL.

isopropylidene), 128.04, 128.49 (CH-arom Z), 136.5 (Cq Z), 156.5 (C=O Z), 173.14, 174.23 $(2 \times C = O \text{ amide, ester}).$

Methyl 2-[(R)-3-(6-benzyloxycarbonylamino)hexanoyloxytetradecanoylamino]-2 deoxy-4,6-O-isopropylidene-3-O-tetradecanoyl-a-D-glucopyranoside (24). (R)-3-(6- Benzyloxycarbonylamino)hexanoyloxytetradecanoic acid 21 (0.516 g, 1.95 mmol) was coupled with compound 18 (0.913 g, 2.06 mmol) as described for the preparation of compound 22. The crude product was purified by silica gel column chromatography. Elution was performed with hexane/ethyl acetate $(80:20 \rightarrow 30:70, v/v)$. Yield 1.172 g (87%). ¹H NMR (CDCl₃): δ =0.88 (t, 6H, CH₃ acyloxyacyl, myristyl), 1.25–1.67 (m, 48H, CH₂ acyloxyacyl, myristyl), 1.36 (s, 3H, CH₃ isopropylidene), 1.46 (s, 3H, CH₃ isopropylidene), $2.24 - 2.45$ (m, 6H, acyloxyacyl, myristyl), 3.17 (q, $2H$, CH₂ hexanoyl), 3.34 (s, 3H, OMe), 3.66–3.78 (m, 3H, H-4, H-5, H-6), 3.87 (dd, 1H, H-6'), 4.24 (m, 1H, H-2), 4.66 (d, 1H, H-1, J1,2 = 3.7 Hz), 4.92 (s, 1H, NH aminohexanoyl), 5.09 (m, 4H, CH2 Z, H-3, CHO acyloxyacyl), 5.97 (d, 1H, NH), 7.27–7.35 (m, 5H, CH-arom Z). ¹³C{¹H} NMR (CDCl₃): δ = 14.12 (CH₃ myristyl), 22.71–41.29 (CH₂ acyloxyacyl, myristyl), 19.10, 29.09 ($2 \times CH_3$ isopropylidene), 52.59 (C-2), 55.22 (OMe), 62.47 (C-6), 63.79 (C-5), 66.59 (CH2 Z), 70.27 (C-3), 71.09 (CHO acyloxyacyl), 72.05 (C-4), 98.98 (C-1), 99.77 (Cq isopropylidene), 128.05, 128.52 (CH-arom Z), 156.45 (C = O Z), 169.62, 172.82, 174.26 $(3 \times C = O \text{ amide, ester}).$

Methyl 2-[(6-benzyloxycarbonylamino)hexanoylamino]-2-deoxy-3-O-tetradecanoyl- α -D-glucopyranoside (25). To a stirred solution of 22 (0.801 g, 1.16 mmol) in THF/water (4:1, v/v; 25 mL) at 0° C was added TFA (1 mL). The resulting solution was allowed to warm to room temperature and left overnight. TLC analysis (ethyl acetate/ hexane, 2:1, v/v showed complete conversion of starting material into a compound with $R_f = 0.13$. The reaction mixture was concentrated under reduced pressure. The residue was diluted with diethyl ether (100 mL) and washed with water $(3 \times 50 \text{ mL})$. The organic layer was dried over $Na₂SO₄$, and the solvent was removed under reduced pressure. Purification of the residue by silica gel column chromatography with DCM/ ethanol (100:0 \rightarrow 93:7, v/v) yielded the desired diol 25 as a colorless oil (0.747 g; 99%). ¹H NMR (CDCl₃): δ = 0.88 (t, 3H, CH₃ myristyl), 1.24 (m, 22H, CH₂ hexanoyl, myristyl), 1.54 (m, 6H, CH₂ hexanoyl, myristyl), 2.13 (t, 2H, CH₂ hexanoyl, myristyl), 2.30 (m, 2H, CH2 hexanoyl, myristyl), 3.17 (q, 2H, CH2 hexanoyl), 3.37 (s, 3H, OMe), 3.65 (m, 1H, H-5, $J_{4.5} = 9.7$ Hz), 3.78 (t, 1H, H-4, $J_{3.4} = 9.5$ Hz, $J_{4.5} = 9.5$ Hz), 3.86 (s, 2H, H₂-6), 4.20 (m, 1H, H-2, J_{1,2} = 3.4 Hz, J_{2,NH} = 9.5 Hz, J_{2,3} = 10.5 Hz), 4.70 (d, 1H, H-1, $J_{1,2} = 3.4$ Hz), 4.99 (br. s, 1H, NH aminohexanoyl), 5.08 (br. s, 2H, CH₂ Z), 5.10 (t, 1H, H-3, $J_{2,3} = 10.5$ Hz, $J_{3,4} = 9.5$ Hz), 5.99 (d, 1H, NH, $J_{2,NH} = 9.2$ Hz), 7.27–7.36 (m, 5H, CH-arom Z). ¹³C{¹H} NMR (CDCl₃): δ = 14.11 (CH₃ myristyl), 22.71–40.83 (CH₂ hexanoyl, myristyl), 52.04 (C-2), 55.27 (OMe), 61.98 (C-6), 66.66 (CH₂ Z), 68.93 (C-4), 71.52 (C-5), 73.78 (C-3), 98.49 (C-1), 128.11, 128.53 (CH-arom Z), 136.68 (Cq Z), 156.55 (C=O Z), 173.14, 175.22 ($2 \times C = O$ amide, ester).

Methyl 2-[(12-benzyloxycarbonylamino)dodecanoylamino]-2-deoxy-3-O-tetradecanoyl- α -D-glucopyranoside (26). Acetonide 23 (1.368 g, 1.77 mmol) was treated with TFA as was described for the preparation of compound 25. Silica gel column chromatography with DCM/ethanol (100:0 \rightarrow 94:6, v/v) of the residue yielded the desired

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

PSEUDO-DISACCHARIDE ANALOGUES OF LIPID A 179

diol as a white solid (0.816 g; 63%). ¹H NMR (CDCl₃): δ = 0.88 (t, 3H, CH₃ myristyl), 1.25 (m, 34H, $CH₂$ dodecanoyl, myristyl), 1.48 (br. t, 2H, $CH₂$ dodecanoyl, myristyl) 1.56 (m, 4H, CH2 dodecanoyl, myristyl), 2.12 (m, 2H, CH2 dodecanoyl, myristyl), 2.31 $(m, 2H, CH₂ dodecanoyl, myristyl), 2.95 (br. s, 2H, 2 × OH) 3.17 (q, 2H, CH₂ dode$ canoyl), 3.38 (s, 3H, OMe), 3.67 (m, 1H, H-5, $J_{4,5} = 9.7$ Hz), 3.77 (t, 1H, H-4, $J_{3,4} = 9.3$, $J_{4.5} = 9.5$ Hz), 3.86 (d, 2H, H₂-6), 4.21 (m, 1H, H-2, $J_{1.2} = 3.6$ Hz, $J_{2,NH} = 9.4$ Hz, $J_{2,3} = 9.4$ Hz), 4.69 (d, 1H, H-1, $J_{1,2} = 3.6$ Hz), 4.78 (br. s, 1H, NH aminohexanoyl), 5.09 (br. s, 3H, CH₂ Z, H-3), 5.83 (d, 1H, NH, $J_{2,NH} = 9.3$ Hz), 7.27–7.36 (m, 5H, CH-arom Z). ¹³C{¹H} NMR (CDCl₃): δ = 14.17 (CH₃ myristyl), 22.75–36.78 (CH₂ dodecanoyl, myristyl), 41.20 (NCH₂) 51.92 (C-2), 55.31 (OMe), 62.22 (C-6), 66.66 (CH₂ Z), 69.21 (C-4), 71.45 (C-5), 73.82 (C-3), 98.53 (C-1), 126.81, 128.13, 128.72 (CH-arom Z), 136.72 (Cq Z), 156.49 (C=O Z), 173.34, 175.23 ($2 \times C = O$ amide, ester).

Methyl $2-[R]-3-(6-benzyloxycarbonylamino)hexanoyloxytetradecanoylamino]-2$ deoxy-3-O-tetradecanoyl- α -D-glucopyranoside (27). To a cooled (0^oC) solution of 24 (138 mg, 0.151 mmol) in DCM (1 mL) was added TFA (0.5 mL). After stirring for 2 h, TLC analysis indicated the complete conversion of starting material into a product with R_f = 0.27 (ethyl acetate/hexane, 2:1, v/v). The reaction mixture was concentrated and coevaporated with toluene $(2 \times 2$ mL). Silica gel column chromatography with hexane/ethyl acetate (50:50 \rightarrow 85:15, v/v) of the residue yielded the desired diol as a white solid (115 mg; 87%). ¹H NMR (CDCl₃): $\delta = 0.88$ (t, 6H, CH₃ acyloxyacyl, myristyl), $1.25-1.66$ (m, $40H$, CH_2 acyloxyacyl, myristyl), $1.51-1.66$ (m, $8H$, CH_2 acyloxyacyl, myristyl), $2.28-2.44$ (m, $6H$, $CH₂$, acyloxyacyl, myristyl), 2.68 (br. s, 1H, OH-6), 3.18 (q, 2H, CH2 hexanoyl), 3.35 (s, 3H, OMe), 3.49 (br. s, 1H, OH-4), 3.64 (m, 1H, H-5), 3.74 (t, 1H, H-4), 3.84 (d, 2H, H₂-6), 4.16 (m, 1H, H-2), 4.67 (d, 1H, H-1, $J_{1,2} = 3.5$ Hz), 5.02 (s, 1H, NH aminohexanoyl), 5.09 (m, 4H, CH₂ Z, H-3, CHO acyloxyacyl), 6.12 (d, 1H, NH), 7.28–7.35 (m, 5H, CH-arom Z). ${}^{13}C[{^{1}H}]$ NMR $(CDC1_3)$: $\delta = 14.09$ $(CH_3$ myristyl), 22.69–41.31 $(CH_2$ acyloxyacyl, myristyl), 52.06 (C-2), 55.20 (OMe), 62.08 (C-6), 66.63 (CH2 Z), 69.14 (C-4), 71.14 (CHO acyloxyacyl), 71.55 (C-5), 73.66 (C-3), 98.35 (C-1), 128.03, 128.06, 128.51 (CH-arom Z), 136.55 (Cq Z), 156.55 (C=O Z), 169.80, 172.81, 175.13 ($3 \times C = O$ amide, ester).

Methyl 2-[(6-benzyloxycarbonylamino)hexanoylamino]-2,6-dideoxy-6-O-mesyl-3-Otetradecanoyl- α -D-glucopyranoside (28). To a stirred solution of 25 (301 mg, 0.463) mmol) in dry pyridine (10 mL) was added mesyl chloride (1.5 equivalents, 60 μ L). After 24 h, methanol (2 mL) was added and the mixture was stirred for 0.5 h, after which the mixture was concentrated. The residue was dissolved in DCM (50 mL) and washed with water $(1 \times 25 \text{ mL})$. The organic layer was dried $(MgSO_4)$ and concentrated under reduced pressure. The crude product was purified by silica gel column chromatography. Elution was performed with hexane/ethyl acetate (70:30 \rightarrow 20:80, v/v). $R_f = 0.43$ (hexane/ethyl acetate, 1:2, v/v). Yield 269 mg (80%). ¹H NMR (CDCl₃): $\delta = 0.88$ (t, 3H, CH₃ myristyl), 1.25 (m, 24H, CH₂ hexanoyl, myristyl), 1.45–1.60 (m, 4H, CH₂ hexanoyl, myristyl), 2.13 (t, 2H, CH₂ hexanoyl, myristyl), 2.31 (m, 2H, CH₂ hexanoyl, myristyl), 3.05 (s, 3H, CH3 Ms), 3.15 (q, 2H, NCH2 hexanoyl), 3.39 (s, 3H, OMe), 3.69 (m, 1H, H-4), 3.81 (br. d, 1H, OH-4), 3.86 (m, 1H, H-5), 4.23 (m, 1H, H-2), 4.50 (q, 2H, H₂-6), 4.70 (d, 1H, H-1, J_{1,2}=3.6 Hz), 5.08 (m, 4H, H-3, CH₂ Z, NH aminohexanoyl), 5.99 (d, 1H, NH), 7.28-7.36 (m, 5H, CH-arom Z). ${}^{13}C[{^1}H]$ NMR

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

180 VAN DEN BERG ET AL.

 $(CDC1_3)$: $\delta = 14.02$ $(CH_3$ myristyl), 22.58–36.16 $(CH_2$ hexanoyl, myristyl), 37.41 (CH_3) Ms), 40.70 (CH₂N), 51.60 (C-2), 55.40 (OMe), 66.45 (CH₂ Z), 68.14 (C-4), 68.59 (C-6), 69.77 (C-5), 73.38 (C-3), 98.43 (C-1), 127.92–128.43 (CH-arom Z), 136.58 (Cq Z), 156.42 (C=O Z), 172.87, 174.92 ($2 \times C = O$ amide, ester).

Methyl 2-[(12-benzyloxycarbonylamino)dodecanoylamino]-2-deoxy-6-O-mesyl-3-Otetradecanoyl- α -D-glucopyranoside (29). Compound 29 (1.175 g, 1.60 mmol) was prepared as described for compound 28. $R_f = 0.68$ (hexane/ethyl acetate, 1:2, v/v). Yield 1.01 g (78%). ¹H NMR (CDCl₃): δ = 0.88 (t, 3H, CH₃ myristyl), 1.25 (m, 34H, CH₂ dodecanoyl, myristyl), 1.42–1.65 (m, 6H, CH2 dodecanoyl, myristyl), 2.12 (t, 2H, CH2 dodecanoyl, myristyl), 2.32 (m, 2H, $CH₂$ dodecanoyl, myristyl), 3.07 (s, 3H, $CH₃$ Ms), 3.17 (q, 2H, NCH2 dodecanoyl), 3.31 (br. d, 1H, OH-4), 3.39 (s, 3H, OMe), 3.70 (m, 1H, H-4, $J_{3,4} = 9.3$ Hz, $J_{4,5} = 9.7$ Hz), 3.86 (m, 1H, H-5, $J_{4,5} = 9.9$ Hz, $J_{5,6} = 6.7$ Hz, $J_{5,6'} = 3.3$ Hz), 4.23 (m, 1H, H-2, $J_{1,2}$ 3.6 Hz, $J_{2,3} = 9.4$ Hz), 4.51 (d, 2H, H₂-6), 4.70 (d, 1H, H-1, $J_{1,2} = 3.6$ Hz), 4.82 (br. s, 1H, NH aminohexanoyl) 5.09 (dd 3H, H-3, CH₂ Z,), 5.81 (d, 1H, NH, $J_{2,NH} = 8.9$ Hz), 7.27–7.35 (m, 5H, CH-arom Z). ¹³C{¹H} NMR (CDCl₃): δ = 14.12 (CH₃ myristyl), 22.69–36.67 (CH₂ dodecanoyl, myristyl), 37.59 $(CH_3 Ms)$, 41.14 (CH₂N), 51.63 (C-2), 55.52 (OMe), 66.57 (CH₂Z), 68.29 (C-4), 68.45 (C-6), 69.87 (C-5), 73.44 (C-3), 98.60 (C-1), 128.06, 128.50 (CH-arom Z), 136.70 (Cq Z), 156.43 (C=O Z), 173.12, 175.07 ($2 \times C = O$ amide, ester).

Methyl 2-[(R)-3-(6-benzyloxycarbonylamino)hexanoyloxytetradecanoylamino]-2 deoxy-3-O-tetradecanoyl-6-O-tosyl-a-D-glucopyranoside (30). Compound 27 (115 mg, 0.131 mmol) was treated with tosyl chloride (138 mg, 0.724 mmol) in pyridine (5 mL). After 96 h, methanol (2 mL) was added and the mixture was concentrated. Further work-up as described for compound 28. The crude product was purified by silica gel column chromatography. Elution was performed with hexane/ethyl acetate $(60:40 \rightarrow 35:65, v/v)$. $R_f = 0.84$ (hexane/ethyl acetate, 1:2, v/v). Yield 107 mg (79%). ¹H NMR (CDCl₃): $\delta = 0.88$ (t, 6H, CH₃ acyloxyacyl, myristyl), 1.25–1.49 (m, 40H, CH₂ acyloxyacyl, myristyl), 1.53 (m, $8H$, $CH₂$ acyloxyacyl, myristyl), 2.34 (m, $6H$, acyloxyacyl, myristyl), 2.44 (s, 3H, CH3 Ts), 3.13 (s, 1H, OH-4), 3.18 (q, 2H, CH2 hexanoyl), 3.29 (s, 3H, OMe), 3.59 (m, 1H, H-4), 3.79 (m, 1H, H-5), 4.14 (m, 1H, H-2), 4.30 (d, 2H, H₂-6), 4.59 (d, 1H, H-1, J_{1,2}=3.6 Hz), 4.94 (t, 1H, NH aminohexanoyl), 5.04 (dd, 1H, H-3), 5.07 (s, 2H, CH2 Z) 5.09 (m, 1H, CHO acyloxyacyl), 5.99 (d, 1H, NH), 7.27–7.80 (m, 9H, CH-arom Ts/Z). ¹³C{¹H} NMR (CDCl₃): δ = 14.11 $(CH₃$ acyloxyacyl), 21.65 (CH₃ Ts), 22.69–41.30 (CH₂ acyloxyacyl, myristyl), 51.72 (C-2), 55.35 (OMe), 66.63 (CH2 Z), 68.60 (C-4), 68.80 (C-6), 69.84 (C-5), 71.13 (CHO acyloxyacyl), 73.43 (C-3), 98.22 (C-1), 127.76–129.84 (CH-arom Z), 133.01, 136.68, 144.98 (Cq Ts, Z), 156.52 (C=O Z), 169.62, 172.76, 175.07 ($3 \times C = O$ amide, ester).

Methyl 6-azido-2-[(6-benzyloxycarbonylamino)hexanoylamino]-2,6-dideoxy-3-Otetradecanoyl- α -D-glucopyranoside (31). To a solution of 28 (269 mg, 0.369 mmol) in DMF (10 mL) was added sodium azide (36 mg, 0.55 mmol). After stirring for 4 h at 70C, TLC analysis indicated the complete conversion of starting material. The mixture was concentrated, diluted with DCM (50 mL) and washed with water (20 mL). The organic layer was dried (MgSO4) and concentrated under reduced pressure. The crude

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

PSEUDO-DISACCHARIDE ANALOGUES OF LIPID A 181

product was purified by silica gel column chromatography. Elution was performed with hexane/ethyl acetate (55:45 \rightarrow 30:70, v/v). $R_f = 0.76$ (hexane/ethyl acetate, 1:2, v/v). Yield 211 mg (84%). ¹H NMR (CDCl₃): δ = 0.88 (t, 3H, CH₃ myristyl), 1.25 (m, 22H, $CH₂$ hexanoyl, myristyl), 1.54 (m, 6H, $CH₂$ hexanoyl, myristyl), 2.12 (t, 2H, $CH₂$ hexanoyl, myristyl), 2.29 (m, 2H, CH₂ hexanoyl, myristyl), 3.17 (q, 2H, CH₂ hexanoyl), 3.41 (s, 3H, OMe), 3.46 (dd, 1H, H-6, $J_{5.6} = 6.2$ Hz, $J_{6.6'} = 13.2$ Hz), 3.57 (dd, 1H, H-6', $J_{5.6'} = 2.5$ Hz, $J_{6.6'} = 13.2$ Hz), 3.63 (t, 1H, H-4, $J_{3.4} = 9.3$ Hz, $J_{4.5} = 9.4$ Hz), 3.79 (m, 1H, H-5, $J_{4,5} = 9.4$ Hz, $J_{5,6} = 6.2$ Hz, $J_{5,6'} = 2.5$ Hz), 4.25 (m, 1H, H-2, $J_{1,2} = 3.6$ Hz, $J_{2,3} = 10.7$ Hz, $J_{2,NH} = 9.5$ Hz), 4.70 (d, 1H, H-1 $J_{1,2} = 3.6$ Hz), 4.92 (br. s, 1H, NH aminohexanoyl), 5.04 (dd, 1H, H-3, $J_{2,3} = 10.8$ Hz, $J_{3,4} = 9.1$ Hz), 5.08 (s, 2H, CH₂ Z), 5.87 (d, 1H, NH, $J_{2,NH} = 9.4$ Hz), 7.27–7.35 (m, 5H, CH-arom Z). ${}^{13}C[{^1H}]$ NMR $(CDC1_3)$: $\delta = 14.10$ $(CH_3$ myristyl), 22.68–40.83 $(CH_2$ hexanoyl, myristyl), 51.42 $(C-6)$, 51.64 (C-2), 55.40 (OMe), 66.63 (CH₂ Z), 69.76 (C-4), 71.15 (C-5), 73.97 (C-3), 98.44 (C-1), 128.05, 128.09, 128.52 (CH-arom Z), 136.67 (Cq Z), 156.49 (C = O Z), 172.73, 175.28 $(2 \times C = O \text{ amide, ester}).$

Methyl 6-azido-2-[(12-benzyloxycarbonylamino)dodecanoylamino]-2,6-dideoxy-3- O -tetradecanoyl- α -D-glucopyranoside (32). Compound 29 (601 mg, 0.738 mmol) was treated with sodium azide as described for the preparation of compound 31. $R_f = 0.60$ (hexane/ethyl acetate, 1:2, v/v). Yield 404 mg (72%). ¹H NMR (CDCl₃): $\delta = 0.88$ (t, 3H, CH₃ myristyl), 1.25 (m, 34H, CH₂ dodecanoyl, myristyl), 1.51 (m, 6H, $CH₂$ dodecanoyl, myristyl), 2.12 (m, 2H, CH₂ dodecanoyl, myristyl), 2.30 (m, 2H, CH₂ dodecanoyl, myristyl), 3.16 (q, 2H, NCH2 dodecanoyl), 3.42 (s, 3H, OMe), 3.46 (m, 2H, H-6, OH-4), 3.57 (dd, 1H, H-6'), 3.62 (m, 1H, H-4), 3.80 (m, 1H, H-5), 4.25 (m, 1H, H-2), 4.71 (d, 1H, H-1, J1,2 = 3.7 Hz), 4.89 (br. s, 1H, NH dodecanoylamino), 5.05 (dd, 1H, H-3), 5.08 (br. s, 2H, CH₂ Z), 5.87 (d, 1H, NH J_{2,NH} = 9.4 Hz), 7.28–7.35 (m, 5H, CH-arom Z). ¹³C{¹H} NMR (CDCl₃): $\delta = 14.07$ (CH₃), 22.64–41.08 (CH₂) dodecanoyl, myristyl), 51.37 (C-6), 51.57 (C-2), 55.32 (OMe), 66.51 (CH₂ Z), 69.62 (C-4), 71.16 (C-5), 73.75 (C-3), 98.40 (C-1), 127.99, 128.44 (CH-arom Z), 136.64 (Cq Z), 156.42 (C=O Z), 173.06, 175.21 ($2 \times C = O$ amide, ester).

Methyl 6-azido-2-[(R)-3-(6-benzyloxycarbonylamino)hexanoyloxytetra-decanoylamino]-2,6-dideoxy-3-O-tetradecanoyl-a-D-glucopyranoside (33). Compound 30 (107 mg, 0.103 mmol) was treated with sodium azide as described for the preparation of compound 31. $R_f = 0.26$ (hexane/ethyl acetate, 2:1, v/v). Yield 68 mg (73%). ¹H NMR (CDCl₃): $\delta = 0.88$ (t, 6H, CH₃ acyloxyacyl, myristyl), 1.25 (m, 40H, CH₂ acyloxyacyl, myristyl), 1.56 (m, 8H, $CH₂$ acyloxyacyl, myristyl), 2.35 (m, 6H, $CH₂$ acyloxyacyl, myristyl), 3.05 (d, 1H, OH-4), 3.20 (t, 2H, hexanoyl), 3.39 (s, 3H, OMe), 3.45 (dd, 1H, H-6), 3.56 (dd, 1H, H-6'), 3.61 (t, 1H, H-4), 3.79 (m, 1H, H-5), 4.22 (m, 1H, H-2), 4.70 (d, 1H, H-1, J_{1,2}=3.6 Hz), 4.90 (br. s, 1H, NH aminohexanoyl), 5.08 (m, 4H, CH2 Z, H-3, CHO acyloxyacyl), 6.02 (d, 1H, NH), 7.27–7.35 (m, 5H, CHarom Z). ¹³C{¹H} NMR (CDCl₃): δ = 14.10 (CH₃ acyloxyacyl), 22.71–41.31 (CH₂ acyloxyacyl, myristyl), 51.40 (C-6), 51.72 (C-2), 55.39 (OMe), 66.66 (CH₂ Z), 69.78 (C-4), 69.89 (C-5), 71.15 (CHO acyloxyacyl), 73.75 (C-3), 98.28 (C-1), 128.06, 128.12, 128.54 (CH-arom Z), 136.62 (Cq Z), 156.5 (C=O Z), 169.66, 172.82, 175.26 $(3 \times C = O$ amide, ester).

182 VAN DEN BERG ET AL.

Methyl 6-amino-2-[(6-benzyloxycarbonylamino)hexanoylamino]-2,6-dideoxy-3-Otetradecanoyl- α -D-glucopyranoside (34). To a solution of 31 (491 mg, 0.727 mmol) in THF (73 mL) was added triphenylphosphine (286 mg, 1.09 mmol) and water (16 μ L, 0.871 mmol). The mixture was refluxed for 3.5 h, after which TLC analysis indicated the complete conversion of starting material into baseline material (hexane/ethyl acetate, 1:2, v/v). The reaction mixture was concentrated and the crude product was purified by silica gel column chromatography. Elution was performed with MeOH/ DCM/TEA $(10:89:1 \rightarrow 14:85:1, v/v/v)$. Yield 497 mg (100%) . ¹H NMR $(CDCl_3)$: δ = 0.88 (t, 3H, CH₃ myristyl), 1.25 (m, 22H, CH₂ hexanoyl, myristyl), 1.54 (m, 6H, $CH₂$ hexanoyl, myristyl), 2.12 (t, 2H, $CH₂$ hexanoyl, myristyl), 2.30 (m, 2H, $CH₂$ hexanoyl, myristyl), 2.77 (br. s, 3H, NH₂-6, OH-4), 3.03 (br. s, 2H, H₂-6), 3.17 (q, 2H, CH₂ hexanoyl), 3.36 (s, 3H, OMe), 3.60 (m, 1H, H-5), 3.64 (t, 1H, H-4, $J_{3,4} = 8.6$ Hz, $J_{4.5} = 8.6$ Hz), 4.18 (m, 1H, H-2, $J_{1.2} = 3.6$ Hz, $J_{2.3} = 10.8$ Hz, $J_{2.3} = 9.3$ Hz), 4.65 (d, 1H, H-1 $J_{1,2} = 3.6$ Hz), 4.95 (br. s, 1H, NH aminohexanoyl), 5.08 (m, 3H, CH₂ Z, H-3, $J_{2,3} = 9.7$ Hz, $J_{3,4} = 9.7$ Hz), 5.87 (d, 1H, NH, $J_{2,NH} = 9.3$ Hz), 7.28–7.36 (m, 5H, CHarom Z). ¹³C{¹H} NMR (CDCl₃): δ = 14.11 (CH₃ myristyl), 22.69–40.86 (CH₂ hexanoyl, myristyl), 43.74 (C-6), 51.93 (C-2), 55.17 (OMe), 66.59 (CH₂ Z), 71.09 (C-4), 71.34 (C-5), 73.71 (C-3), 98.43 (C-1), 128.06–128.60 (CH-arom Z), 136.74 (Cq Z), 156.45 (C=O Z), 172.69, 174.99 ($2 \times C = O$ amide, ester).

Methyl 6-amino-2-[(12-benzyloxycarbonylamino)dodecanoylamino]-2,6-dideoxy-3- O -tetradecanoyl- α -D-glucopyranoside (35). Compound 32 (914 mg, 1.20 mmol) was treated with triphenylphosphine as described for the preparation of compound 34. $R_f = 0.43$ (MeOH/DCM, 1:9, v/v). The crude product was purified by silica gel column chromatography. Elution was performed with MeOH/DCM/TEA (0:99.5:0.5 \rightarrow 10:89.5:0.5, v/v/v). Yield 698 mg (78%). ¹H NMR (CDCl₃): δ = 0.88 (t, 3H, CH₃ myristyl), 1.25 (m, 34H, CH2 hexanoyl, myristyl), 1.52 (m, 6H, CH2 hexanoyl, myristyl), 2.09 (m, 2H, CH2 hexanoyl, myristyl), 2.31 (m, 2H, CH2 hexanoyl, myristyl), 2.80 (br. s, 3H, NH2-6, OH-4), 3.03 (br. d, 2H, H2-6), 3.17 (q, 2H, CH2 hexanoyl), 3.37 (s, 3H, OMe), 3.55–3.67 (m, 2H, H-4, H-5), 4.19 (m, 1H, H-2), 4.66 (d, 1H, H-1, $J_{1,2} = 3.6$ Hz), 4.83 (s, 1H, NH aminohexanoyl), 5.09 (br. t, 3H, H-3, CH₂ Z), 5.81 (d, 1H, NH), 7.28–7.36 (m, 5H, CH-arom Z). ¹³C{¹H} NMR (CDCl₃): δ = 14.12 (CH₃) myristyl), 22.70-41.14 (CH₂ hexanoyl, myristyl), 43.59 (C-6), 51.86 (C-2), 55.19 (OMe), 66.55 (CH₂ Z), 71.00 (C-4), 71.17 (C-5), 73.55 (C-3), 98.45 (C-1), 128.05, 128.50 (CH-arom Z), 136.74 (Cq Z), 156.43 (C=O Z), 173.05, 174.95 ($2 \times C = O$ amide, ester).

Methyl 6-amino-2- $[(R)-3-(6-benzvloxvcarbonvlamino)hexanovloxv-tetradecanovl$ amino-2,6-dideoxy-3-O-tetradecanoyl-a-D-glucopyranoside (36). Compound 36 was prepared as described for compound 34. The crude product was purified by silica gel column chromatography. Elution was performed with MeOH/DCM/TEA (0:99:1 \rightarrow 9:90:1, v/v/v). Yield 35 mg (52%). ¹H NMR (CDCl₃): $\delta = 0.88$ (t, 6H, CH₃ acyloxyacyl, myristyl), 1.25 (m, 40H, CH₂ acyloxyacyl, myristyl), 1.55 (m, 8H, CH₂ acyloxyacyl, myristyl), 2.35 (m, 6H, CH2 acyloxyacyl, myristyl), 3.04–3.33 (m, 7H, OH-4, NH2-6, H2-6, NCH2 hexanoyl), 3.35 (s, 3H, OMe), 3.62–3.66 (m, 2H, H-4, H-5), 4.18 (m, 1H, H-2), 4.65 (d, 1H, H-1, $J_{1,2} = 3.5$ Hz), 4.95 (br. s, 1H, NH aminohexanoyl), 5.08 (m, 4H, CH2 Z, H-3, CHO acyloxyacyl), 6.00 (d, 1H, NH), 7.27–7.36 (m, 5H,

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

PSEUDO-DISACCHARIDE ANALOGUES OF LIPID A 183

CH-arom Z). ¹³C{¹H} NMR (CDCl₃): δ = 14.17 (CH₃ acyloxyacyl, myristyl), 22.74– 41.34 (CH2 acyloxyacyl, myristyl), 43.52 (C-6), 52.00 (C-2), 55.29 (OMe), 66.64 (CH2 Z), 70.43 (C-4), 71.15 (CHO acyloxyacyl), 71.49 (C-5), 73.32 (C-3), 98.31 (C-1), 128.11, 128.56 (CH-arom Z), 136.74 (Cq Z), 156.5 (C=O Z), 169.71, 172.79, 174.98 $(3 \times C = O$ amide, ester).

Compound 37. To a stirred mixture of 11 (30 mg, 0.428 mmol), PyBOP (25 mg, 0.480 mmol) and DiPEA (8.1 μ L, 0.480 mmol) in DCM (3.6 mL) a solution of amino sugar 34 (28 mg, 0.431 mmol) in DCM (2.8 mL) was added. After 1 h, TLC analysis indicated the complete conversion of starting material into a compound with $R_f = 0.62$ (MeOH/DCM, $7:93$, v/v). The reaction mixture was diluted with DCM (50 mL) and washed with water $(1 \times 20 \text{ mL})$. After drying over MgSO₄, the organic layer was concentrated under reduced pressure. The crude product was purified by silica gel column chromatography. Elution was performed with hexane/ethyl acetate $(40:60 \rightarrow$ 100:0, v/v). Yield 55 mg (96%). ¹H NMR (CDCl₃): $\delta = 0.88$ (m, 6H, CH₃ myristyl), 1.25 (br. s, 42H, $CH₂$ hexanoyl, myristyl), 1.52 (m, 8H, $CH₂$ hexanoyl, myristyl), 1.94 (m, 3H, CH2 hexanoyl, OH-4), 2.11 (t, 2H, CH2 hexanoyl), 2.27 (m, 2H, OCOCH₂), 2.41 (dd, 1H, H-1'ax, J_{1'ax,1'eq} = 12.5 Hz, J_{1'ax,2}' = 5.5 Hz), 2.90 (m, 1H, H-6a), 3.04 (br. s, 1H, H-5'), 3.10 (dd, 1H, H-1'eq, $J_{1'ax,1'eq} = 12.5$ Hz, $J_{1'eq,2'} = 3.0$ Hz), 3.17 (q, 2H, NCH2 hexanoyl), 3.29 (s, 3H, OMe), 3.39 (m, 3H, H-4, NCH2 acetyl), 3.52 (m, 1H, H-6'a), 3.55 (m, 1H, H-3'), 3.59 (m, 1H, H-5), 3.67 (t, 1H, H-4'), 3.83 (dd, 1H, H-6'b), 4.02 (m, 2H, H-2', H-6b), 4.12 (m, 1H, H-2), 4.45–4.67 (m, 7H, $3 \times CH_2$ Bn, H-1), 4.82 (br. s, 1H, NHCOO), 5.09 (s, 2H, CH2 Z), 5.11 (t, 1H, H-3), 5.77 (d, 1H, NH-2), 6.35 (br. d, 1H, NH-2'), 7.25–7.35 (m, 20H, CH-arom Bn/Z), 7.82 (m, 1H, NH-6). ¹³C{¹H} NMR (CDCl₃): δ = 14.14 (CH₃ 2 × myristyl), 22.72–38.84 (CH₂ hexanoyl, myristyl), 39.85 (C-6), 40.91 (NCH₂ hexanoyl), 47.56 (C-2'), 50.78 (C-1') 52.55 (C-2), 55.19 (OMe), 57.69 (CH₂ acetyl), 62.22 (C-5'), 66.44 (C-6'), 66.62 (CH₂ Z), 69.23 (C-4), 70.84 (C-5), 72.10 (C-3), 72.82, 73.35, 73.50 (3 \times CH₂ Bn), 76.93 (C-3'), 77.28 (C-4'), 98.62 (C-1), 127.80–128.64 (CH-arom Bn, Z), 136.75, 137.66, 137.83, 137.86 (4 × Cq Bn, Z), 156.75 (C=O Z), 172.59, 172.76, 173.18, 174.65 $(4 \times C = O$ amide, ester). ES-MS; m/z : 1332.9, $[M+H]^+$; monoisotopic MW calculated for $C_{78}H_{117}N_5O_{13} = 1331.86$.

Compound 38. Compound 11 (22 mg, 31.4 μ mol) was coupled with compound 35 (24 mg, 32.7 µmol) as described for the preparation of compound 37. $R_f = 0.53$ (MeOH/ DCM, 5:95, v/v). Yield 42.8 mg (96%). ¹H NMR (CDCl₃): $\delta = 0.88$ (m, 6H, CH₃ myristyl), 1.25 (br. s, 52H, CH2 dodecanoyl, myristyl), 1.46 (m, 4H, CH2 dodecanoyl, myristyl), 1.55 (m, 8H, CH2 dodecanoyl, myristyl), 1.94 (q, 1H, OH-4), 2.10 (m, 2H, CH₂ dodecanoyl), 2.28 (m, 2H, OCOCH₂), 2.44 (dd, 1H, H-1'ax, $J_{1'ax,1'eq} = 12.4$ Hz, $J_{1'ax,2'} = 5.2$ Hz), 2.90 (m, 1H, H-6a, $J_{6a,6b} = 12.4$ Hz), 3.05 (br. s, 1H, H-5'), 3.10 (dd, 1H, H-1'eq, $J_{1'ax,1'ca} = 12.5$ Hz, $J_{1'ca,2'} = 3.1$ Hz), 3.18 (q, 2H, NCH₂ dodecanoyl), 3.29 (s, 3H, OMe), 3.38 (m, 3H, H-4, NCH2 acetyl), 3.52 (s, 1H, H-3'), 3.55 (m, 1H, H-6'a, $J_{5',6'3} = 3.3$ Hz, $J_{6a'6b'} = 10.3$ Hz), 3.60 (br. d, 1H, H-5, $J_{4.5} = 9.6$ Hz), 3.68 (t, 1H, H-4'), 3.84 (dd, 1H, H-6'b, $J_{5,6'b} = 6.2$ Hz, $J_{6'a,6'b} = 10.2$ Hz), 4.02 (m, 1H, H-6b), 4.07 (m, 1H, H-2'), 4.11 (m, 1H, H-2), 4.45–4.71 (m, 8H, H-1, NHCOO, $3 \times CH_2$ Bn, H-1), 5.09 (s, 2H, CH₂ Z), 5.12 (t, 1H, H-3, J = 10.4 Hz), 5.73 (d, 1H, NH-2), 6.36 (br. d, 1H, NH-2'), 7.22–7.35 (m, 20H, CH-arom Bn, Z), 7.85 (m, 1H, NH-6). $^{13}C(^{1}H)$ NMR (CDCl₃):

184 VAN DEN BERG ET AL.

 δ = 14.20 (CH₃ dodecanoyl, myristyl), 22.78–36.88 (CH₂ dodecanoyl, myristyl), 39.85 (C-6), 41.24 (NCH2 dodecanoyl), 47 (C-2'), 50 (C-1') 52.56 (C-2), 55.25 (OMe), 57.78 (CH₂ acetyl), 62 (C-5'), 66.47 (C-6'), 66.66 (CH₂ Z), 69.20 (C-4), 70.94 (C-5), 71.93 $(C-3)$, 72.86, 73.39, 73.52 $(3 \times CH_2 \text{ Bn})$, 77.30 $(C-3', C-4')$, 98.69 $(C-1)$, 127.86– 128.71 (CH-arom Bn/Z), 136.80, 137.87 (Cq Bn, Z), 156 (C=O Z), 173.04, 173.30, 174.69, $(C=O$ amide, ester). ES-MS; m/z : 1416.85, $[M+H]^+$; monoisotopic MW calculated for $C_{84}H_{129}-N_5O_{13}=1415.96$.

Compound 39. Compound 11 (10 mg, 14.3 μ mol) was coupled with compound 36 (13.7 mg, 15.7 µmol) as described for the preparation of compound 37. $R_f = 0.86$ (MeOH/DCM, 5:95, v/v). Yield 11.4 mg (51%). ¹H NMR (CDCl₃): $\delta = 0.88$ (t, 9H, CH3 acyloxyacyl, myristyl), 1.25 (m, 60H, CH2 acyloxyacyl, myristyl), 1.52 (m, 10H, CH₂ acyloxyacyl, myristyl), 1.94 (m, 3H, OH-4, CH₂ myristyl), 2.30 (m, 6H, NCH₂ hexanoyl, OCOCH₂), 2.42 (dd, 1H, H-1'ax), 2.88 (m, 1H, H-6a), 3.06 (br. s, 1H, H-5'), 3.11 (dd, 1H, H-1'eq), 3.18 (q, 2H, NCH₂ hexanoyl), 3.27 (s, 3H, OMe), 3.35 (m, 1H, H-4), 3.40 (s, 2H, NCH2 acetyl), 3.53 (m, 2H, H-3', H-6'a), 3.58 (m, 1H, H-5), 3.68 (t, 1H, H-4'), 3.83 (dd, 1H, H-6'b), 4.03 (m, 2H, H-2', H-6b), 4.10 (m, 1H, H-2), 4.45– 4.67 (m, 7H, $3 \times CH_2$ Bn, H-1), 4.85 (br. s, 1H, NHCOO), 5.10 (m, 4H, CH₂ Z, H-3, CHO acyloxyacyl), 5.93 (d, 1H, NH-2), 6.32 (br. d, 1H, NH-2'), 7.25–7.35 (m, 20H, CH-arom Bn, Z), 7.86 (m, 1H, NH-6). ¹³C{¹H} NMR (CDCl₃): $\delta = 14.20$ (CH₃) myristyl), 22.78–36.89 (CH2 acyloxyacyl, myristyl), 39.85 (C-6), 40.94 (C-1'), 41.35 (NCH₂ hexanoyl), 47.47 (C-2'), 52.61 (C-2), 55.19 (OMe), 57.81 (CH₂ acetyl), 62.16 (C-5'), 66.48 (C-6'), 66.67 (CH2 Z), 69.20 (C-4), 70.87 (C-5), 71.16 (CHO acyloxyacyl), 71.89 (C-3), 72.81, 73.40, 73.48 ($3 \times CH_2$ Bn), 76 (C-3'), 77.30 (C-4'), 98.48 (C-1), 127.85–128.71 (CH-arom Bn, Z), 136.77, 137.64, 137.85 (Cq Bn, Z), 156.49 $(C=O Z)$, 169.64, 172.80, 173.36, 174.67 $(4 \times C=O \text{ amide, ester}).$ ES-MS; m/z : 1559.00, $[M + H]^+$; monoisotopic MW calculated for C₉₂H₁₄₃N₅O₁₅ = 1558.05.

Compound 40. Compound 12 (30 mg, 0.334 mmol) was coupled with compound 34 (23 mg, 0.334 mmol) as described for the preparation of compound 37. $R_f = 0.48$ (hexane/ethyl acetate, 1:4, v/v). Yield 45 mg (88%). ¹H NMR (CDCl₃): δ 0.88 = (dt, 9H, CH3 acyloxycyl, myristyl), 1.25 (br. s, 56H, CH2 acyloxyacyl, hexanoyl, myristyl), 1.52 (m, 10H, CH2 acyloxyacyl, hexanoyl, myristyl), 1.86 (s, 1H, OH-4), 2.11 (t, 2H, NCH2 hexanoyl), 2.16–2.33 (m, 6H, CH2 acyloxyacyl, hexanoyl, myristyl), 2.40 (dd, 1H, H-1'ax, $J_{1'ax,1'ca} = 12.5$ Hz, $J_{1'ax,2'} = 5.8$ Hz), 2.96 (m, 1H, H-6a), 3.05 (br. s, 1H, H-5'), 3.10 (dd, 1H, H-1'eq, $J_{1'ax,1'eq} = 12.5$ Hz, $J_{1'eq,2'} = 3.4$ Hz), 3.17 (q, 2H, CH₂ hexanoyl), 3.29 (s, 3H, OMe), 3.37 (m, 1H, H-4), 3.40 (s, 2H, NCH2 acetyl), 3.54 (m, 2H, H-6a', H-3'), 3.59 (m, 1H, H-5), 3.67 (t, 1H, H-4'), 3.83 (dd, 1H, H-6b'), 4.03 (m, 2H, $H-2'$, $H-6b$), 4.12 (m, 1H, $H-2$), 4.44 -4.67 (m, $6H$, $3 \times CH_2$ Bn), 4.50 (d, 1H, $H-1$), 4.81 (s, 1H, NHZ), 5.09 (m, 4H, CH2 Z, H-3, CHO acyloxyacyl), 5.79 (d, 1H, NH-2), 6.55 (d, 1H, NH-2'), 7.25–7.35 (m, 20H, CH-arom Bn, Z), 7.87 (m, 1H, NH-6). ¹³C{¹H} NMR (CDCl₃): δ = 14.16 (CH₃, dodecanoyl, myristyl), 22.74–36.49 (CH₂ dodecanoyl, hexanoyl, myristyl), 39.78 (C-6), 40.91, 41.80, 41.92 (CH₂ myristyl, hexanoyl, dodecanoyl), 47.65 (C-2'), 50.93 (C-1'), 52.56 (C-2), 55.19 (OMe), 57.69 (CH2 acetyl), 62.20 (C-5'), 66.43 (C-6'), 66.66 (CH2 Z), 69.21 (C-4), 70.95 (C-5), 71.15 (CHO acyloxyacyl), 72.12 (C-3), 72.83, 72.99, 73.35 (3 × CH₂ Bn), 76 (C-3'), 77 (C-4'), 98.61 (C-1), 127.73–128.65 (CH-arom Bn, Z), 136.76, 137.80, 137.86, 137.89

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

PSEUDO-DISACCHARIDE ANALOGUES OF LIPID A 185

 $(4 \times Cq$ Bn, Z), 156.46 (C=O Z), 169.34, 172.64, 173.18, 173.32, 174.68 (5 \times C=O amide, ester). ES-MS; m/z : 1531.1, $[M + H]^{+}$; monoisotopic MW calculated for $C_{90}H_{139}N_5O_{15}= 1530.0.$

Compound 41. Compound 12 (20 mg, 22.2 μ mol) was coupled with compound 35 (18 mg, 24.5 µmol) as described for the preparation of compound 37. $R_f = 0.62$ (MeOH/ DCM, 5:95, v/v). Yield 34.5 mg (96%). ¹H NMR (CDCl₃): $\delta = 0.88$ (m, 9H, CH₃ dodecanoyl, myristyl), 1.25 (m, 68H, CH₂ acyloxyacyl, dodecanoyl, myristyl), 1.54 (m, 10H, $CH₂$ acyloxyacyl, dodecanoyl, myristyl), 2.10 (m, 2H, NCH₂ dodecanoyl), 2.23 (m, 6H, CH₂ acyloxyacyl, dodecanoyl, myristyl), 2.43 (dd, 1H, H-1'ax, J_{1'ax,1'eq} = 12.4 Hz, $J_{1'ax,2'} = 5.1$ Hz), 2.97 (m, 1H, H-6a, $J_{6a,6b} = 12.6$ Hz), 3.08 (br. s, 1H, H-5'), 3.11 (dd, 1H, H-1'eq, $J_{1'ax,1'eq} = 12.6$ Hz, $J_{1'eq,2'} = 2.8$ Hz), 3.18 (q, 2H, NCH₂ acyl), 3.29 (s, 3H, OMe), 3.39 (m, 3H, H-4, NCH2 acetyl), 3.55 (m, 2H, H-6a', H-3'), 3.60 (m, 1H, H-5, $J_{4,5} = 9.8$ Hz), 3.67 (t, 1H, H-4'), 3.84 (dd, 1H, H-6b', $J_{5',6b'} = 6.3$ Hz, $J_{6a',6b'} = 10.5$ Hz), 4.03 (m, 1H, H-6b, $J_{5,6b} = 4.2$ Hz, $J_{6a,6b} = 12.6$ Hz) 4.06 (m, 1H, H-2'), 4.12 (m, 1H, H-2, $J_{1,2} = 3.5$ Hz, $J_{2,3} = 9.8$ Hz), 4.42–4.78 (m, 8H, H-1, NHZ, $3 \times CH_2$ Bn), 5.08 (m, 4H, CH₂ Z, H-3, CHO acyloxyacyl), 5.74 (d, 1H, NH-2, $J_{2,NH} = 9.1$ Hz), 6.59 (d, 1H, NH-2', $J_{2'NH}$ = 6.3 Hz), 7.25–7.36 (m, 20H, CH-arom Bn, Z), 7.87 (m, 1H, NH-6). ¹³C{¹H} NMR (CDCl₃): δ = 14.20 (CH₃ 3 × dodecanoyl, myristyl), 22.78–36.87 (CH₂ acyloxyacyl, dodecanoyl, myristyl), 39.78 (C-6), 41.23 , 41.79 (CH₂ acyloxyacyl, dodecanoyl, myristyl), 47.79 (C-2'), (50, C-1'), 52.53 (C-2), 55.21 (OMe), 57.69 (CH2 acetyl), 62.20 (C-5'), 66.41 (C-6'), 66.66 (CH₂ Z), 69.18 (C-4), 71.03 (C-5), 71.13 (CHO acyloxyacyl), 71.95 (C-3), 72.83, 73.30, 73.35 (3 × CH₂ Bn), 77.09 (C-3'), 77.31 (C-4'), 98.61 (C-1), 127.74–128.68 (CH-arom Bn, Z), 136.80, 137.88, (Cq Bn, Z), 156.46 (C=O Z), 169.31, 173.02, 173.18, 173.25, 174.66 ($4 \times C = O$ amide, ester). ES-MS; m/z : 1615.00, $[M + H]^2$; monoisotopic MW calculated for $C_{96}H_{151}N_5O_{15} = 1614.12$.

Compound 1. To a solution of 37 (10.4 mg, 7.54 μ mol) in DMF (0.5 mL), Pd/C (10%, 5 mg) was added. Hydrogen was passed through the stirred mixture for 46 h. After filtration of the mixture over a PTFE filter, the filtrate was concentrated under reduced pressure. Yield 7 mg quantitative. ¹H NMR (pyridine-d₅): δ =0.87 (m, 6H, $2 \times CH_3$ acyl), 1.24 (m, 43H, CH₂ acyl), 1.39 (m, 2H, CH₂ acyl), 1.53 (m, 2H, CH₂ acyl), 1.65 (m, 2H, CH₂ acyl), 1.82 (m, 4H, $2 \times CH_2$ acyl), 2.06 (m, 2H, CH₂ acyl), 2.18 (t, 1H, H-1'ax), 2.57–2.29 (m, 8H, CH2 acyl), 2.65 (m, 1H, H-5'), 3.05 (d, 1H, Hacetyl), 3.26 (t, 2H, CH2 acyl), 3.38 (dd, 1H, H-1'eq), 3.41 (s, 3H, OMe), 3.75 (m, 3H, H-acetyl, H-4', H-6a), 4.04 (dd, 1H, H-6b), 4.10 (dd, 1H, H-3'), 4.26 (t, 1H, H-4), 4.35 (t, 1H, H-6'a), 4.57 (m, 1H, H-5), 4.62 (m, 1H, H-2'), 4.86 (m, 1H, H-2), 4.97 (dd, 1H, H-6'b), 5.11 (d, 1H, H-1), 5.84 (dd, 1H, H-3), 8.65 (d, 1H, NH), 8.89 (d, 1H, NH'). ¹³C{¹H} NMR (pyridine-d₅): δ = 13.65 (CH₃ myristyl), 22.29–38.90 (CH₂ acyl), 40.62 $(C-1')$, 50.75 $(C-2')$, 51.76 $(C-2)$, 54.69 $(C-6)$, 55.16 (OMe) , 57.14 $(CH₂ acetyl)$, 59.73 (C-5'), 69.02 (C-5), 69.96 (C-4), 72.36 (C-6'), 73.07 (C-3), 73.56 (C-4'), 75.78 (C-3'), 99.12 (C-1), 166.63 (C=O ester), 172.71, 173.24, 173.31 ($3 \times C = O$ amide). ES-MS; m/z: 928.77, $[M + H]^{+}$; monoisotopic MW calculated for $C_{49}H_{93}N_{5}O_{11} = 927.69$.

Compound 2. Compound 38 (12.9 mg, 9.11 µmol) was treated as described for the preparation of compound 1. Yield 6.6 mg $(72%)$ of a white solid. ES-MS; mlz : 1012.54, $[M + H]^+$; monoisotopic MW calculated for $C_{55}H_{105}N_5O_{11} = 1011.78$.

186 VAN DEN BERG ET AL.

Compound 3. Compound 39 (1.9 mg, 1.2 μ mol) was treated as described for the preparation of compound 1. Yield 0.7 mg $(54%)$ of a white solid. ES-MS; m/z : 1154.98, $[M + H]^+$; monoisotopic MW calculated for $C_{63}H_{119}N_5O_{13} = 1153.88$.

Compound 4. Compound 40 (6.7 mg, 4.38 μ mol) was treated as described for the preparation of compound 1. Yield 4.3 mg $(88%)$ of a white solid. ES-MS; m/z : 1126.9, $[M+H]^+$; monoisotopic MW calculated for $C_{61}H_{115}N_5O_{13}=1125.85$.

Compound 5. Compound 41 (8.2 mg, 5.1 μ mol) was treated as described for the preparation of compound 1. Yield 4.0 mg $(65%)$ of a white solid. ES-MS; m/z : 1211.18, $[M + H]^+$; monoisotopic MW calculated for $C_{67}H_{127}N_5O_{13} = 1209.94$.

ACKNOWLEDGMENTS

We thank S.H. van Krimpen for recording the NMR spectra and A.G. Hulst for performing electrospray MS analysis.

REFERENCES

- 1. Rietschel, E.Th.; Brade, L.; Lindner, B.; Zähringer, U. Biochemistry of Lipopolysaccharides in Bacterial Endotoxic Lipopolysaccharides. In Molecular Biochemistry and Cellular Biology; Morrison, D.C., Ryan, J.L., Eds.; CRC: Boca Raton, Florida, 1992; Vol. I, 4–41, Chapter 1.
- 2. Galanos, C.; Lüderitz, O.; Rietschel, E.Th.; Westphal, O.; Brade, H.; Brade, L.; Freudenberg, M.A.; Schade, F.U.; Imoto, M.; Yoshimura, S.; Kusumoto, S.; Shiba, T. Synthetic and natural Escherichia coli free lipid A express identical endotoxic activities. Eur. J. Biochem. 1985 , 148 , $1-5$.
- 3. Chaby, R. Strategies for the control of LPS-mediated pathophysiological disorders. Drug Discovery Today 1999, 4, 209-221.
- 4. Opal, S.M.; Yu, R.L. Antiendotoxin strategies for the prevention and treatment of septic shock — new approaches and future directions. Drugs 1998, 55, 497– 508.
- 5. Wyckoff, T.J.O.; Raetz, C.R.H.; Jackman, J.E. Antibacterial and anti-inflammatory agents that target endotoxin. Trends Microbiol. **1998**, 6 , $154-159$.
- 6. Landry, D.W.; Zhoa, K.; Yang, G.X.Q.; Glickman, M.; Georgiadis, T.M. Antibody-catalyzed degradation of cocaine. Science 1993, 259, 1899– 1901.
- 7. Landry, D.W.; Yang, G.X.Q. Anti-cocaine catalytic antibodies a novel approach to the problem of addiction. J. Addict Dis. 1997 , 16 , $1-17$.
- 8. Mets, B.; Winger, G.; Cabrera, C.; Seo, S.; Jamdar, S.; Yang, G.; Zhao, K.; Briscoe, R.J.; Almonte, R.; Woods, J.H.; Landry, D.W. A catalytic antibody against cocaine prevents cocaine's reinforcing and toxic effects in rats. Proc. Natl. Acad. Sci. U.S.A. 1998, 95, 10176-10181.
- 9. Baird, T.J.; Deng, S.X.; Landry, D.W.; Winger, G.; Woods, J.H. Natural and artificial enzymes against cocaine. I. Monoclonal antibody 15A10 and the reinforcing effects of cocaine in rats. J. Pharmacol. Exp. Ther. 2000 , 295 , $1127 - 1134$.

©2002 Marcel Dekker, Inc. All rights reserved. This material may not be used or reproduced in any form without the express written permission of Marcel Dekker, Inc.

PSEUDO-DISACCHARIDE ANALOGUES OF LIPID A 187

- 10. Reymond, J.L.; Janda, K.D.; Lerner, R.A. Antibody catalysis of glycosidic bond hydrolysis. Angew. Chem., Int. Ed. Engl. 1991, 30, 1711–1713.
- 11. Suga, H.; Tanimoto, N.; Sinskey, A.J.; Masamune, S. Glycosidase antibodies induced to a half-chair transtition-state analog. J. Am. Chem. Soc. 1994, 116, 11197 – 11198.
- 12. Yu, J.H.; Hsieh, L.C.; Kochersperger, L.; Yonkovich, S.; Stephans, J.C.; Gallop, M.A.; Schultz, P.G. Progress toward an antibody glycosidase. Angew. Chem., Int. Ed. Engl. 1994, 33, 339–341.
- 13. Yu, J.; Choi, S.Y.; Lee, S.; Yoon, H.J.; Jeong, S.; Mun, H.; Park, H.; Schultz, P.G. Antibody-catalysed glycosyl transfer reactions from in vitro immunization. Chem. Commun. 1997, 1957– 1958.
- 14. Yu, J.; Choi, S.Y.; Moon, K.D.; Chung, H.H.; Yuon, H.J.; Jeong, S.; Park, H.; Schultz, P.G. A glycosidase antibody elicited against a chair-like transition state analog by in vitro immunization. Proc. Natl. Acad. Sci. U.S.A. 1998, 95, 2880-2884.
- 15. Shabat, D.; Sinha, S.C.; Reymond, J.L.; Keinan, E. Catalytic antibodies as probes of evolution: modeling of a primordial glycosidase. Angew. Chem., Int. Ed. Engl. 1996, 35, 2628– 2630.
- 16. Janda, K.D.; Lo, L.C.; Lo, C.H.L.; Sim, M.M.; Wang, R.; Wong, C.H.; Lerner, R.A. Chemical selection for catalysis in combinatorial antibody libraries. Science 1997, 275, 945– 948.
- 17. Van den Berg, R.J.B.H.N.; Noort, D.; Milder-Enacache, E.S.; Van der Marel, G.A.; Van Boom, J.H.; Benschop, H.P. Approach toward a generic treatment of Gram-negative infections: synthesis of haptens for catalytic antibody mediated cleavage of the interglycosidic bond in lipid A. Eur. J. Org. Chem. 1999, 2593 – 2600.
- 18. Dong, W.L.; McCabe, K.W.; Bols, M.; Sierks, M.R. Catalytic antibodies for carbohydrate. Protein Eng. 1995, 8 (Suppl. S), 58.
- 19. Van den Broek, L.A.G.M.; Vermaas, D.J.; Heskamp, B.M.; Van Boeckel, C.A.A.; Tan, M.C.A.A.; Bolscher, J.G.M.; Ploegh, H.L.; Van Kemenade, F.J.; De Goede, R.E.Y.; Miedema, F. Chemical modification of azasugars, inhibitors of Nglycoprotein-processing glycosidases and of HIV-I infection — review and structure-activity-relationships. Recl. Trav. Chim. Pays-Bas. 1993, 112, 82-94.
- 20. Coste, J.; Le-Nguyen, D.; Castro, B. PYBOP a new peptide coupling reagent devoid of toxic by-product. Tetrahedron Lett. 1990, 31, 205-208.
- 21. Kiso, M.; Tanaka, S.; Fujita, M.; Fujishima, Y.; Ogawa, Y.; Ishida, H.; Hasegawa, A. Synthesis of the optically active 4-O-phosphono-D-glycosamine derivatives related to the nonreducing sugar subunit of bacterial lipid A. Carbohydr. Res. 1987, 162, 127– 140.
- 22. Foster, A.B.; Horton, D.; Stacey, M. Amino-sugar and related compounds. Part II. Observations on the acidic hydrolysis of derivatives of 2-amino-2-deoxy-Dglucose (D-glucosamine). J. Chem. Soc. 1957 , $81-86$.
- 23. Neises, B.; Steglich, W. Einfaches verfahren zur veresterung von carbonsäuren. Angew. Chem. **1978**, 90, 556-557.
- 24. Leblanc, Y.; Fitzsimmons, B.J.; Adams, J.; Perez, F.; Rokach, J. The total synthesis of 12-HETE and 12,20-diHETE. J. Org. Chem. 1986, 51, 789-793.
- 25. Vaultier, M.; Knouzi, N.; Carrie, R. Reduction d'azides en amines primaires par

188 VAN DEN BERG ET AL.

une methode generale utilisant la reaction de staudinger. Tetrahedron Lett. 1983, 24, 763 –764.

26. NMR spectrometry of compounds $2-5$ was performed with CDCl₃, methanol-d₄ and pyridine- d_5 as solvents at ambient temperature.

Received November 5, 2001 Accepted January 28, 2002